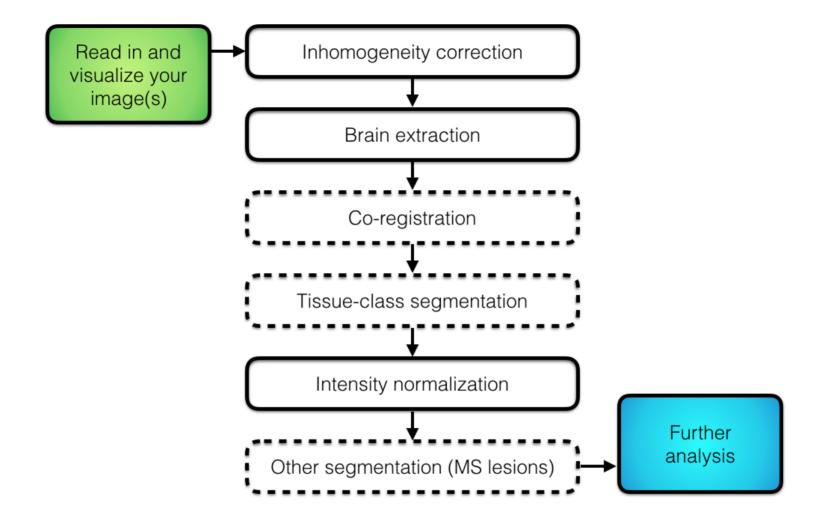
# Tissue-Class Segmentation

## **Overall Pipeline**



#### Image segmentation

- · We are often interested in subdividing or segmenting the brain into meaningful biological regions of interest (ROIs) for an analysis.
- Examples: tissue segmentation, segmentation of gray matter structures, segmentation of pathology (MS lesions, tumors, ...)
- We will perform 3-class tissue segmentation in R using fslr and ANTSR:
  - Cerebrospinal fluid (CSF)
  - Gray Matter (GM)
  - White Matter (WM)

#### **Loading Data**

• Let's read in the training T1 and brain mask for subject 05 (not 01!).

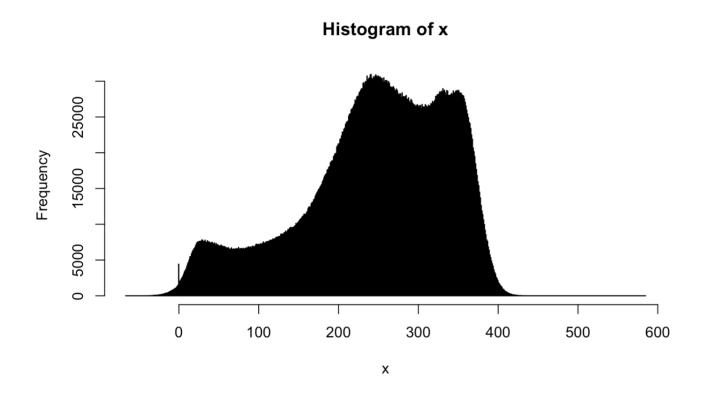
```
library(ms.lesion)
library(neurobase)
all_files = get_image_filenames_list_by_subject(
    group = "training",
    type = "coregistered")
files = all_files$training05 # NOT training subject 1!
t1 = readnii(files["T1"])
rt1 = robust_window(t1)
mask = readnii(files["Brain_Mask"])

run_mask = t1 > 100
dd_orig = drop_empty_dim(run_mask, keep_ind = TRUE)
```

## Tissue Segmentation: Large Outliers

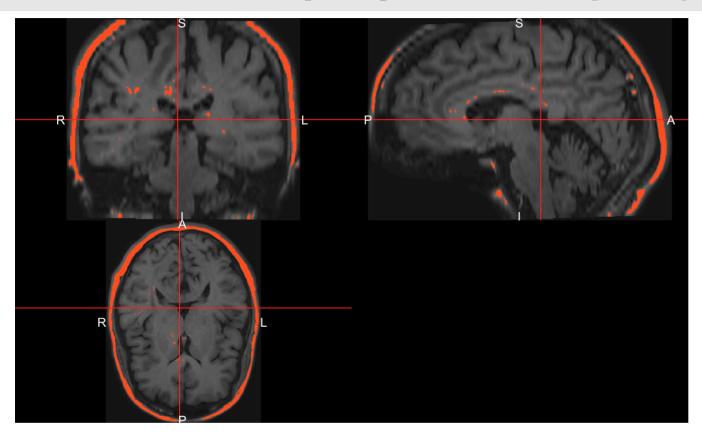
- · Many tissue class segmentations are based on k-means clustering.
- These methods can be skewed by large outliers.

```
hist(t1, mask = mask, breaks = 2000); text(x = 800, y = 3000, "outliers!")
```



#### Where are the outliers?

```
ortho2(rt1, t1 > 400, xyz = xyz(t1 > 400)) # xyz - cog of a region
```

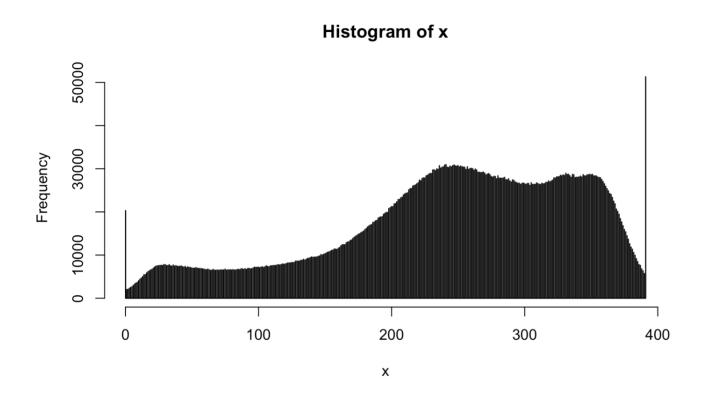


## Cleaning up values: Masking the image

```
t1[ t1 < 0 ] = 0
t1 = mask_img(t1, mask)
rt1 = robust_window(t1)</pre>
```

## What does the histogram look like now?

hist(rt1, mask = mask, breaks = 2000);



#### Tissue Segmentation using FSL FAST

- FAST is based on a hidden Markov random field model and an Expectation-Maximization algorithm (Zhang, Brady, and Smith 2001).
- · It jointly produces a bias field corrected image and a probabilistic tissue segmentation.
- More robust to noise and outliers than finite mixture model-based methods that do not incorporate spatial information.

The fslr function fast calls fast from FSL. The --nobias option tells FSL to not perform inhomogeneity correction (N4 already performed in ANTSR).

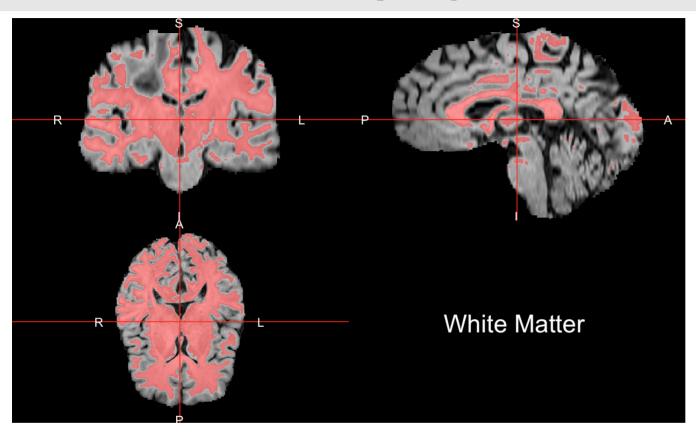
#### **FAST Results**

FAST assumes three tissue classes and produces an image with the three labels, ordered by increasing within-class mean intensities. In a T1 image, this results in:

- · Level 1: CSF
- · Level 2: Gray Matter
- · Level 3: White Matter

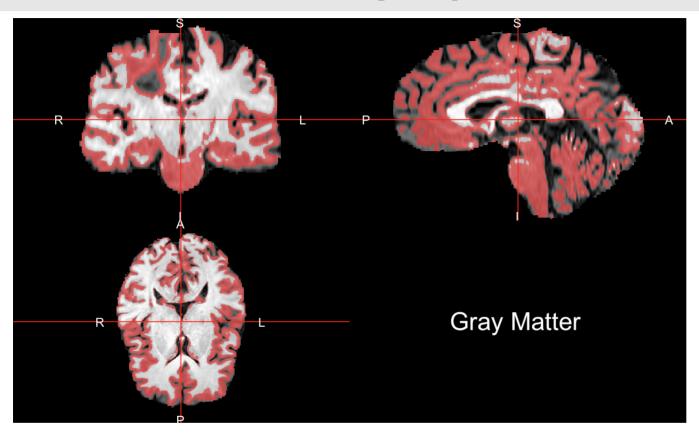
#### **FAST: White Matter**

```
ortho2(rt1, t1fast == 3, col.y = alpha("red", 0.5), text = "White Matter")
```



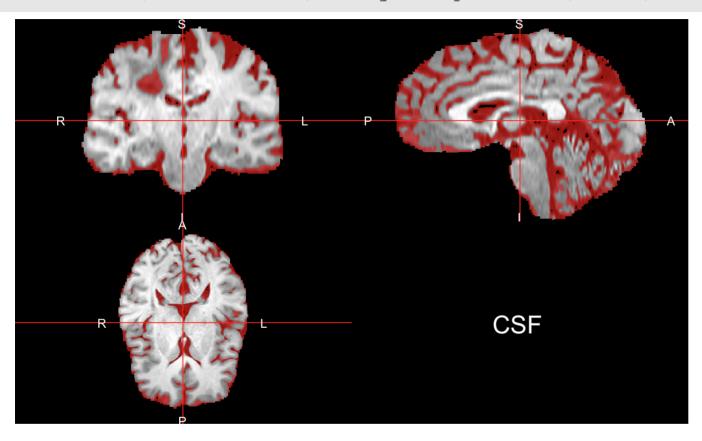
## **FAST: Gray Matter**

```
ortho2(rt1, t1fast == 2, col.y = alpha("red", 0.5), text = "Gray Matter")
```



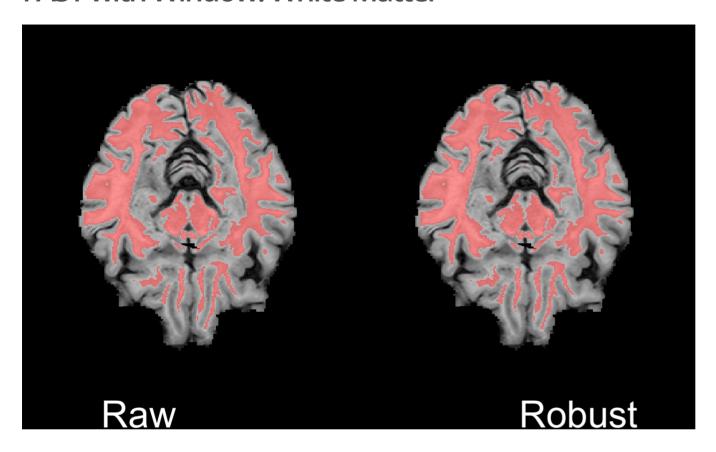
## **FAST: CSF**

ortho2(rt1, t1fast == 1, col.y = alpha("red", 0.5), text = "CSF")

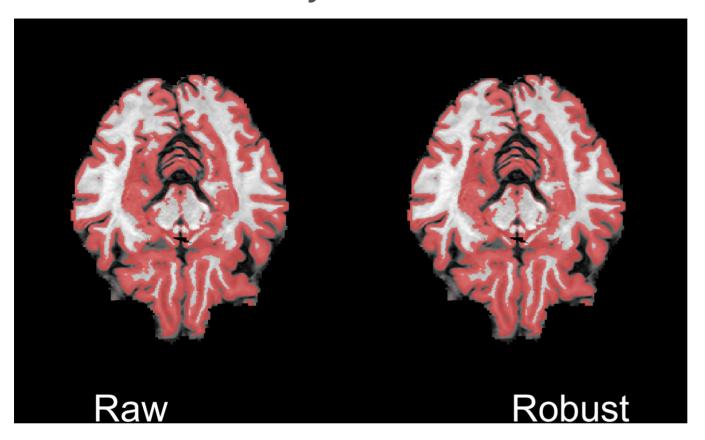


## Removing large values: Is there an effect?

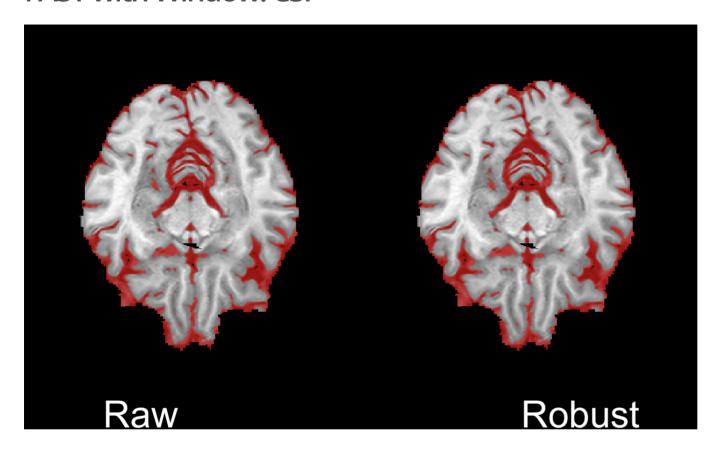
## FAST with Window: White Matter



# FAST with Window: Gray Matter



## **FAST** with Window: CSF



#### **FAST Results**

- · Overall the results look good
  - Not much difference after dampening outliers using robust\_window

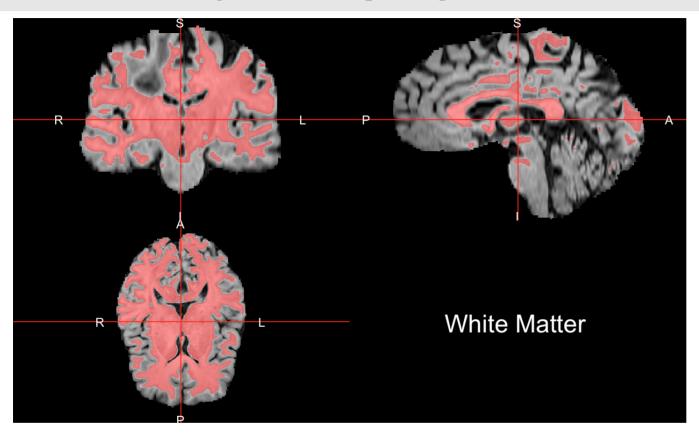
#### Tissue Segmentation using ANTsR, extrantsr

- Uses Atropos (Avants et al. 2011)
  - 3D K-means clustering + a Markov random field
- The extrantsr::otropos function works with nifti objects
  - calls ANTSR::atropos function

```
t1_otropos = otropos(a = t1, x = mask) # using original data
t1seg = t1 otropos$segmentation
```

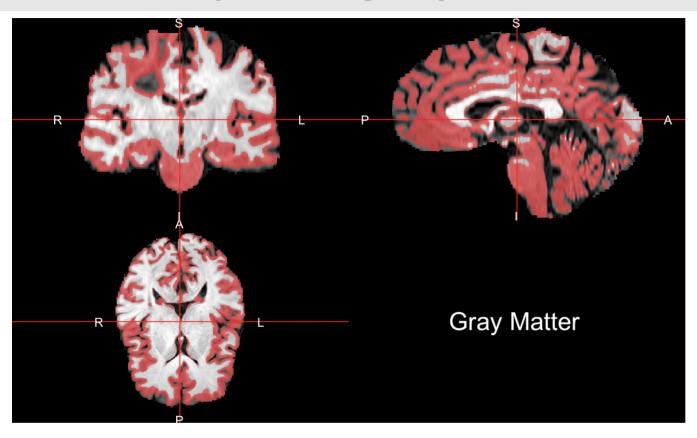
## **Atropos: White Matter**

```
ortho2(rt1, t1seg == 3, col.y = alpha("red", 0.5), text = "White Matter")
```



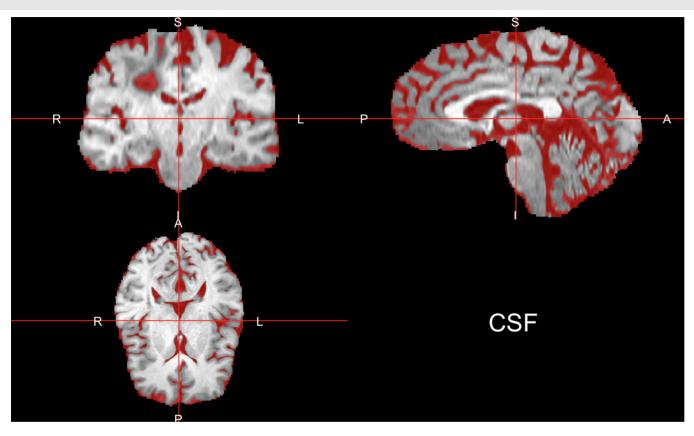
## **Atropos: Gray Matter**

```
ortho2(rt1, t1seg == 2, col.y = alpha("red", 0.5), text = "Gray Matter")
```



## Atropos: CSF

```
ortho2(rt1, t1seg == 1, col.y = alpha("red", 0.5), text = "CSF")
```



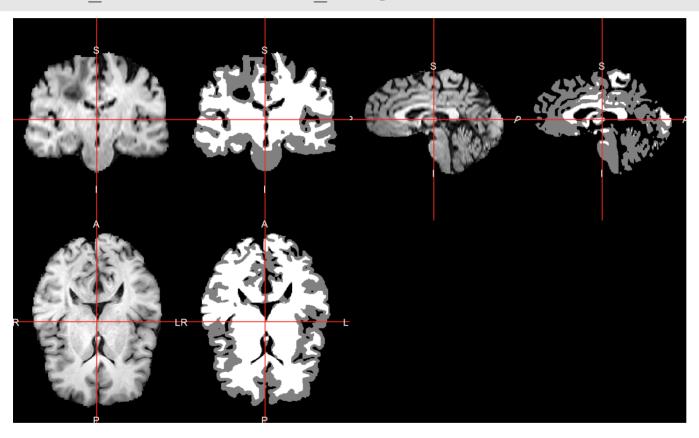
## **Default Atropos Results**

- · Overall the results do not look good
  - The k-means clustering is affected by large outliers
- We will try using robust\_window

## **Atropos using Windowing**

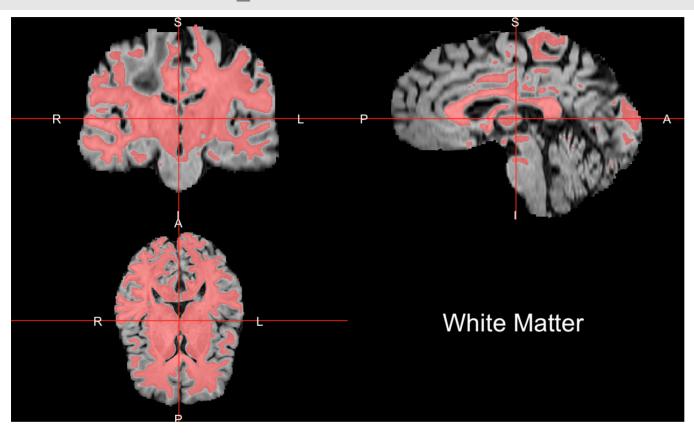
```
robust_t1_otropos = otropos(a = rt1, x = mask) # using robust
robust_t1seg = robust_t1_otropos$segmentation
```

double ortho(rt1, robust t1seg)



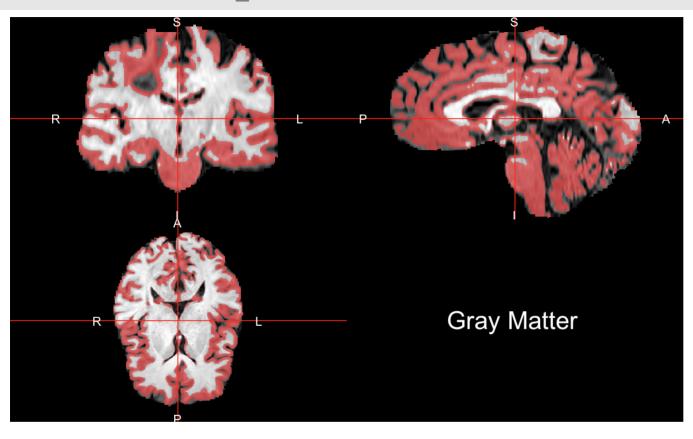
## Atropos with Window: White Matter

```
ortho2(rt1, robust_t1seg == 3, col.y = alpha("red", 0.5), text = "White Matter")
```



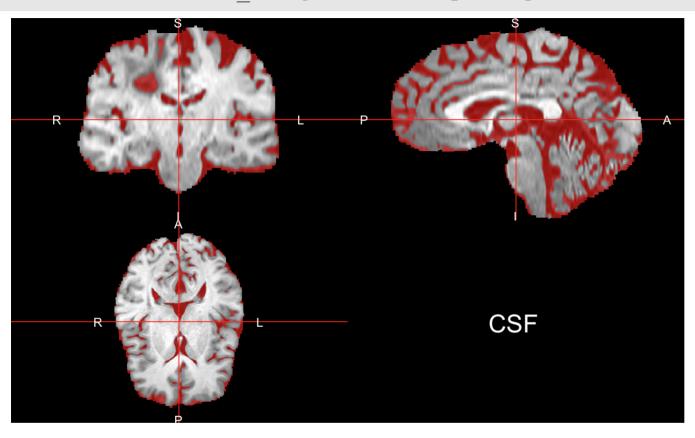
## Atropos with Window: Gray Matter

```
ortho2(rt1, robust_t1seg == 2, col.y = alpha("red", 0.5), text = "Gray Matter")
```



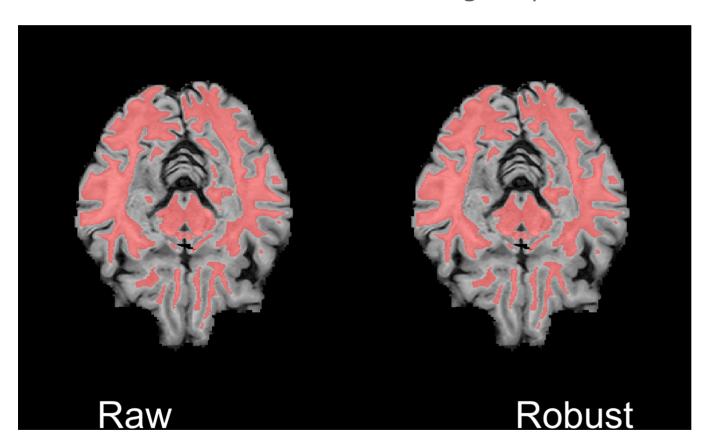
## Atropos with Window: CSF

```
ortho2(rt1, robust_t1seg == 1, col.y = alpha("red", 0.5), text = "CSF")
```

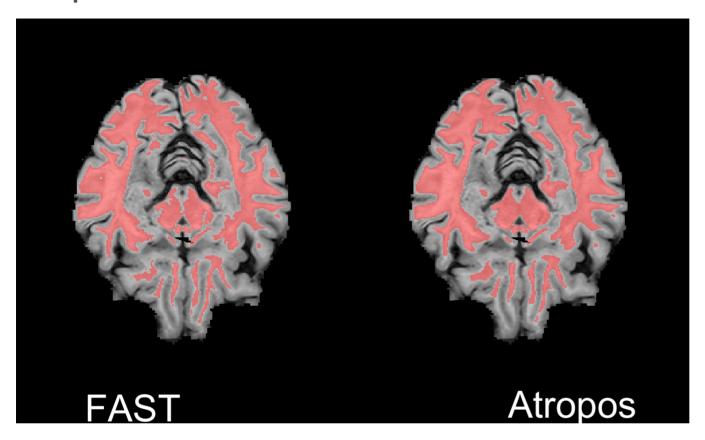


## **Atropos with Window Results**

- · Overall the results look like they reasonably separate the classes
  - No ground truth
- · Winsorizing large outliers aided the k-means clustering
  - Results much better than running Atropos on the raw data



# Atropos WM vs. FAST WM



#### Estimating the Volume of Each Class

We can create a table which will count the number of voxels in each category:

## Estimating the Volume of Each Class

By multiplying by the voxel resolution (in cubic centimeters) using the voxres function, we can get volumes

## Website

http://johnmuschelli.com/imaging\_in\_r

#### References

Avants, Brian B, Nicholas J Tustison, Jue Wu, Philip A Cook, and James C Gee. 2011. "An Open Source Multivariate Framework for N-Tissue Segmentation with Evaluation on Public Data." 9 (4). Springer:381–400.

Zhang, Yongyue, Michael Brady, and Stephen Smith. 2001. "Segmentation of Brain MR Images Through a Hidden Markov Random Field Model and the Expectation-Maximization Algorithm." 20 (1):45–57. http://ieeexplore.ieee.org/xpls/abs\_all.jsp?arnumber=906424.