

Flux Variability Analysis



Learning Objectives

- Explain alternate optimal solutions,
- · Explain flux variability analysis,
- Demonstrate the ability to use of flux variability analysis.



Lesson Outline

- Alternate Optimal Solutions
- Flux Variability Analysis
- FVA Examples



Phenotypes

- Phenotype = A phenotype (from Greek phainein, meaning "to show", and typos, meaning "type") is the composite of an organism's observable characteristics or traits, such as its morphology, development, biochemical or physiological properties, phenology, behavior, and products of behavior.
- Silent phenotypes have the same overall external cellular function but are based on different underlying reaction networks.

https://en.wikipedia.org/wiki/Phenotype



Alternate Equivalent Optimal Solutions

- The flux distributions calculated by FBA are often not unique. In many cases, it is necessary for a biological system to achieve the same objective value by using alternate equivalent optimal pathways, creating phenotypically different alternate optimal solutions (silent phenotypes).
- Requires the Mixed Integer Linear Programming (MILP) solver
- For large models there can be a very large number of alternate equivalent optimal solutions.

Same objective value for all alternate flux vectors

Maximize the objective function

$$\longrightarrow Z = \sum_{i} c_{i} v_{i}^{k} = \mathbf{c} \cdot \mathbf{v}^{k}$$

with the following constraints

Up to *n* alternate flux vectors

$$\frac{d\mathbf{x}}{dt} = \mathbf{S} \cdot \mathbf{v}^k = \mathbf{0}$$

$$\alpha_j \le v_j^k \le \beta_j$$

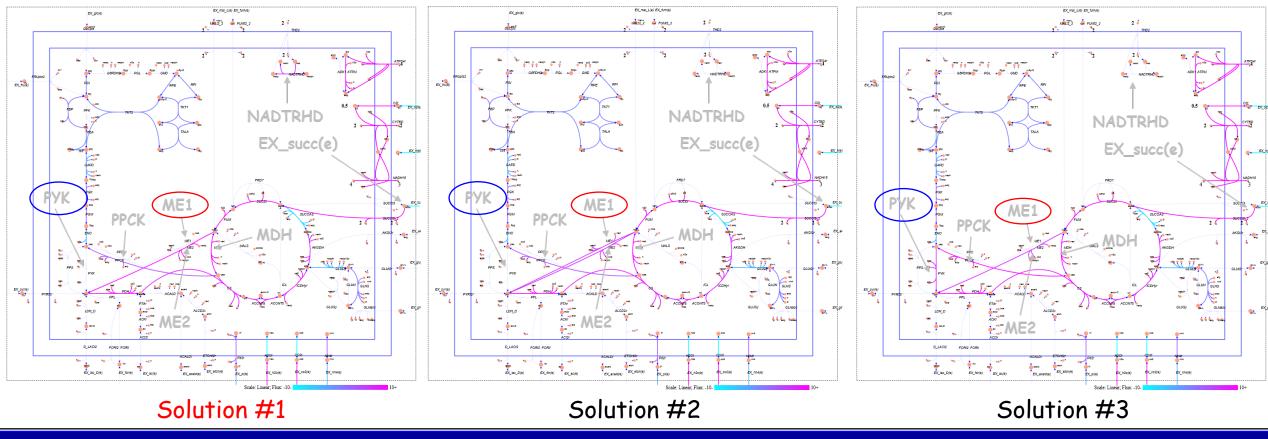
$$1 \le k \le n$$

 $\Rightarrow v^1, v^2, \dots, v^n$ all have the same value of Z

Reed, J. L. & Palsson, B. Ø. Genome-scale in silico models of *E. coli* have multiple equivalent phenotypic states: assessment of correlated reaction subsets that comprise network states. Genome Res. 14, 1797-1805 (2004).



Alternate Optimal Solutions





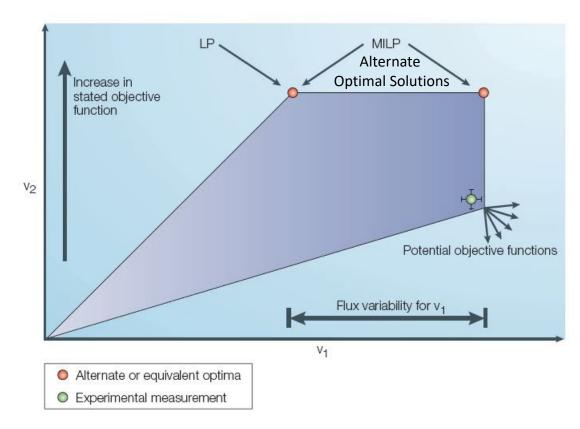
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Flux Variability Analysis

- This method identifies the allowable range of flux values through a given reaction by finding the maximum and minimum possible fluxes through the particular reaction for a given maximum objective value.
- All reactions under test have the same objective value
- This analysis method begins by finding the optimal value of the objective function for a given set of constraints and then optimizes for the minimum and maximum flux values for each reaction (1 + 2n) optimizations where n is the number of reactions).
- A method that can be used to identify alternate optimal pathways.



Price, N. D., J. L. Reed, et al. (2004). "Genome-scale models of microbial cells: evaluating the consequences of constraints." Nature reviews. Microbiology 2(11): 886-897.

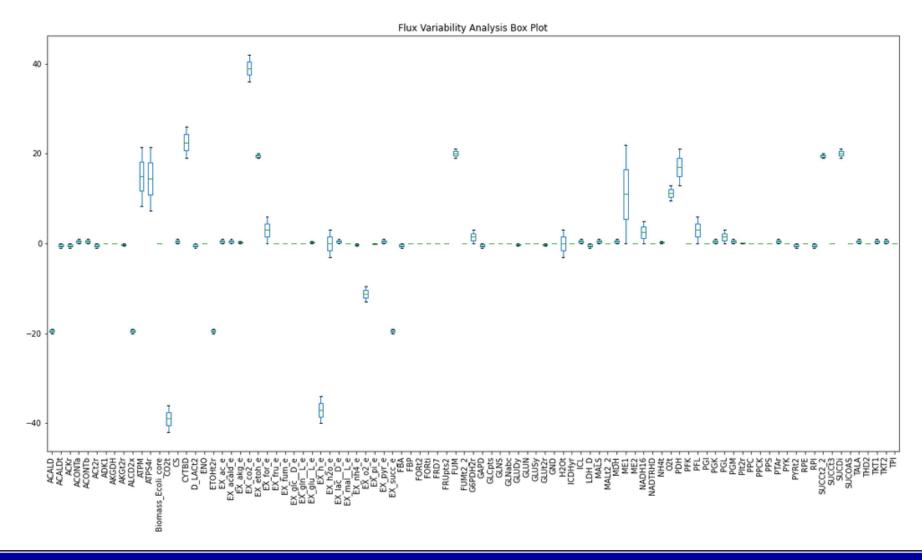


FVA Data

	minimum	maximum
ACALD	-1.204692	0.000000
ACALDt	-1.204692	0.000000
ACKr	-1.842471	0.000000
ACONTa	6.685508	8.730759
ACONTb	6.685508	8.730759
ACt2r	-1.842471	0.000000
ADK1	0.000000	5.780234
AKGDH	0.039160	7.869660
AKGt2r	-0.728419	0.000000
ALCD2x	-1.044067	0.000000
ATPM	8.390000	16.220500



FVA Box Plot





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- → · FVA Examples



FVA Lecture Examples

FVA_Lecture_Examples.ipynb

Maximum Growth FVA

Set Python environment and load the model

```
In [1]: import cobra.test
        import pandas as pd
        pd.set option('display.max rows', 500)
        model = cobra.test.create test model("textbook")
         #model = cobra.io.load matlab model('./e coli core.mat')
        Academic license - for non-commercial use only - expires 2022-10-10
        Using license file C:\Users\hinton\qurobi.lic
        Set the simulation conditions
In [2]: from cobra.flux analysis import flux variability analysis
        model.reactions.EX qlc D e.lower bound = -0 # Must be set to zero if not a carbon source
        model.reactions.EX succ e.lower bound = -20
         #model.reactions.ME1.bounds = [6.49242,6.49242]
         #model.reactions.ME1.bounds = [0,0]
        Run flux variability analysis
In [3]: fva=flux variability analysis (model, model.reactions, loopless = True, fraction of optimum=1.0)
        fva.round(3)
```



Bordbar et al. BMC Systems Biology 2011, **5**:110 http://www.biomedcentral.com/1752-0509/5/110



RESEARCH ARTICLE

Open Access

iAB-RBC-283: A proteomically derived knowledgebase of erythrocyte metabolism that can be used to simulate its physiological and patho-physiological states

Aarash Bordbar, Neema Jamshidi and Bernhard O Palsson*

Bordbar, A., N. Jamshidi, et al. (2011). "iAB-RBC-283: A proteomically derived knowledge-base of erythrocyte metabolism that can be used to simulate its physiological and patho-physiological states." BMC systems biology 5: 110.



iAB_RBC_283 Model Attributes

iAB_RBC_283_Model_Attributes.ipynb

Set the environment

```
In [1]: import cobra.test
   import pandas as pd
   import numpy as np
   import pandas as pd
   import escher
   from escher import Builder
   from cobra.flux_analysis import flux_variability_analysis
   import matplotlib.pyplot as plot
   from cobrapy_bigg_client import client
   pd.set_option('display.max_rows', 1000)
   pd.set_option('display.width',1000)
   pd.set_option('display.max_colwidth',None)
```

Load the red blood cell model iAB RBC 283.json

```
In [2]: model = client.download_model('iAB_RBC_283', save=False) # Loading the model to the simulation

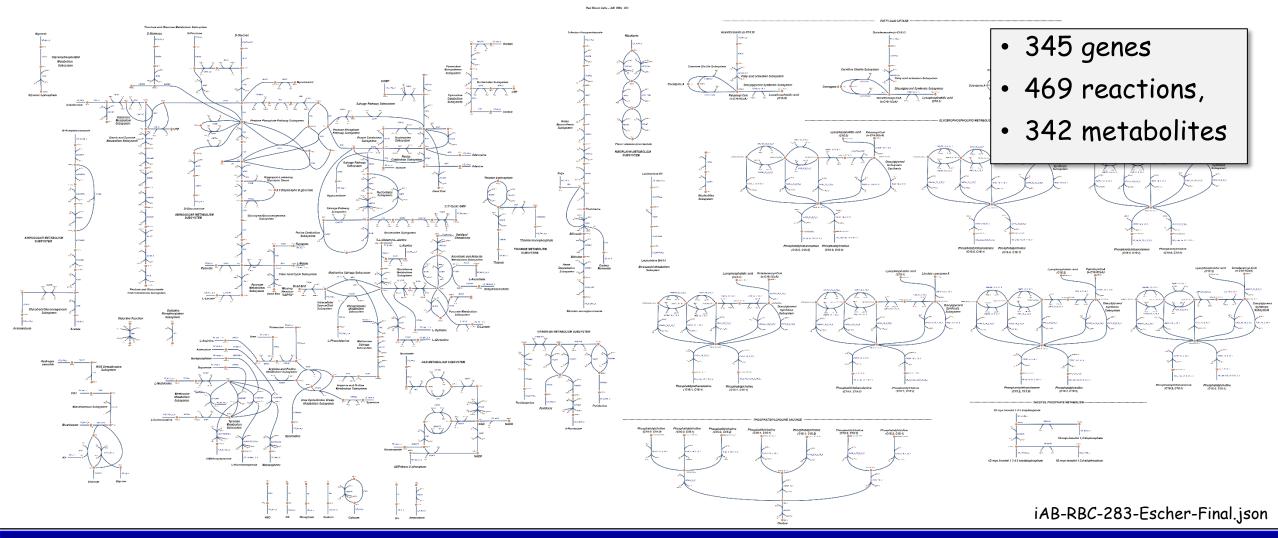
Set parameter Username
Academic license - for non-commercial use only - expires 2022-10-10
```

Model Attribute Summary

```
In [3]: model
```



Escher Map: iAB_RBC_283.metabolism

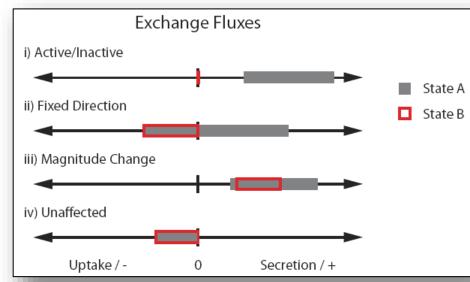




Analyzing iAB-RBC-283 as a Functional Biomarker

- The Morbid Map from the Online Mendelian Inheritance in Man (OMIM) and the DrugBank were downloaded from their respective databases. The enzyme names in iAB-RBC-283 were cross-referenced against the database entries to determine morbid single nucleotide polymorphisms (SNPs) in erythrocyte proteins and drugs with protein targets in the erythrocyte. The morbid SNPs that did not have sole pathological effects in the erythrocyte were classified using the Merck Manual.
- Just as FVA can be used to assess the function of a network under a particular set of constraints, it can also be used to assess the changes in function and thus has applications for characterizing disease states and identifying biomarkers. When simulating a morbid SNP or a drug inhibited enzyme, the lower and upper bound constraints on the affected reaction is set to zero. FVA is then used to characterize the exchange reactions under morbid SNP or drug treated conditions and then compared to the normal state.
- A reaction was considered to be confidently altered if the change in the minimum or maximum flux was 40% of the total flux span. The flux span is defined as the absolute difference between the original (unperturbed) maximum and minimum fluxes.

Bordbar, A., N. Jamshidi, et al. (2011). "iAB-RBC-283: A proteomically derived knowledge-base of erythrocyte metabolism that can be used to simulate its physiological and patho-physiological states." BMC systems biology 5: 110.



There are four major differences that can occur for an exchange reaction in two different states: i) the reaction is either active (non-zero minimum or maximum flux) or inactive (zero minimum and maximum flux), ii) the exchange becomes fixed in one direction (uptake or secretion only), iii) there is a magnitude change in exchange, iv) the reaction is unaffected and is the same for both states.



OMIM Database

OMIM®	Online Mendelian Inheritance in Man [®] An Online Catalog of Human Genes and Genetic Disorders Updated 12 June 2015			
	Search			
Advanced Search : OMIM, Clinical Synopses, Gene Map Need help? : Example Searches, OMIM Search Help, OMIM Tutorial Mirror sites : us-east.omim.org, europe.omim.org				

http://omim.org/

Amberger J, Bocchini CA, Scott AF, Hamosh A: McKusick's Online Mendelian Inheritance in Man (OMIM). Nucleic Acids Res 2009, , 37 Database: D793-6.

	OMIM Advanced Search							
	Search OMIM		Search					
	Sort by: Relevance ○ Date updated ○ Date created Entries per page: 10 ✓							
	Search in:	Only Records With:	MIM Number Prefix:					
	☐ Mim Number	☐ Allelic Variants	* gene with known sequence					
	☐ Title	\square Clinical Synopsis	\square + gene with known sequence and phenotype					
	□Text	\square Gene Map Locus	\square # phenotype description, molecular basis known					
	\square Allelic Variants		$\square\%$ mendelian phenotype or locus, molecular basis unknown					
	\square Clinical Synopsis	Only Records without:	\square none - other, mainly phenotypes with suspected mendelian basis					
	☐ Contributors ☐ Gene Map Locus							
Chromosome:								
ı	□1 □2 □3	□ 4 □ 5 □ 6 □ :	□8 □9 □10 □11 □12					
ı	□ 13 □ 14 □ 15	□ 16 □ 17 □ 18 □	9 □ 20 □ 21 □ 22 □ X □ Y					
	☐ Mitochondrial ☐ Autosomal		□ Unknown					
	Created From: YYYY/MM/DD To: present Updated From: YYYY/MM/DD To: present Note: Entries created before June 2, 1986 have a creation date of June 2, 1986.							
	Clear Search	1						



Detected SNPs and FVA Results for SNP Perturbations

OMIM	Location	Pathology	Gene Protein Name			
		Acatalasemia (3)				
115500	-	CAT				
	9p21-p12 Acromesomelic dysplasia, Maroteaux type, 602875 (3) NPR2, ANPRB, AMDM					
608958	20q13.11	ADA				
609712	1q21	Adenosine triphosphate, elevated, of erythrocytes, 102900 (3)	PKLR, PK1			
180297	6p21.1-p11	RHAG, RH50A				
606224	7p15-p14	NT5C3, UMPH1, PSN1				
608313	3 6q23 Argininemia, 207800 (3) ARG1					
173335	335 6q22-q23 Arterial calcification, generalized, of infancy, 208000 (3) ENPP1, PDNP1, NPPS, M6S1, PCA1, ARHR2 350 Xq22-q24 Arts syndrome, 301835 (3) PRPS1, CMTX5, DFNX1, DFN2 350 1p32 CPT II deficiency, lethal neonatal, 608836 (3) CPT2 352 11q13 CPT deficiency, hepatic, type IA, 255120 (3) CPT1A 350 1p32 CPT deficiency, hepatic, type II, 600649 (3) CPT2					
311850						
600650						
600528						
600650						
154550	15g22-gter Carbohydrate-deficient glycoprotein syndrome, type lb. 602579 (3) MPI, PMI1					
182500	OMIM is a comprehensive, authoritative compendium of human genes and					
311850	genetic phe	notypes that is freely available and updated o	daily OMIM is			
607672	authored and edited at the McKusick-Nathans Institute of Genetic Medicine, Johns Hopkins University School of Medicine, under the					
600179						
601785						
612732						
609414	2q35	Corneal fleck dystrophy, 121850 (3)	PIKFYVE, PIP5K3			
191740	2q37	Crigler-Najjar syndrome, type I, 218800 (3)	UGT1A1, UGT1, GNT1, BILIQTL1			
191740						
244050	Deafness, X-linked 1, 304500 (3) PRPS1, CMTX5, DFNX1, DFN2					

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Affected Protein	ADA	AHCY	AK1
Affected Reaction	ADA	AHC	ADK1
Varying Exchanges	EX_ade(e)	EX_3moxtyr(e)	EX_35cgmp(e)
	EX_hxan(e)	EX_adrnl(e)	EX_3moxtyr(e)
	EX_nh4(e)	EX_dopa(e)	EX_adrnI(e)
		EX_hcyst-L(e)	EX_camp(e)
		EX_mepi(e)	EX_chol(e)
		EX_met-L(e)	EX_dopa(e)
		EX_normete-L(EX_etha(e)
		EX_nrpphr(e)	EX_glyc(e)
			EX_hcyst-L(e)
A noostice .		danad	EX_hdca(e)
	A reaction was considered to be confidently altered		
if the change in the			EX_met-L(e)
minimum or	maximun	n flux	EX_nac(e)
was 40% of	the tota	l flux	EX_ncam(e)
			EX_normete-L(e)
span. The flux span is defined as the absolute difference between the original (unperturbed) maximum and minimum fluxes.			EX_nrpphr(e)
			EX_ocdcea(e)
			EX_orot(e)
			EX_spmd(e)
			EX_sprm(e)
			EX_thm(e)
			EX_thmmp(e)
			EX_uri(e)



DrugBank Database



Open Data Drug & Drug Target Database

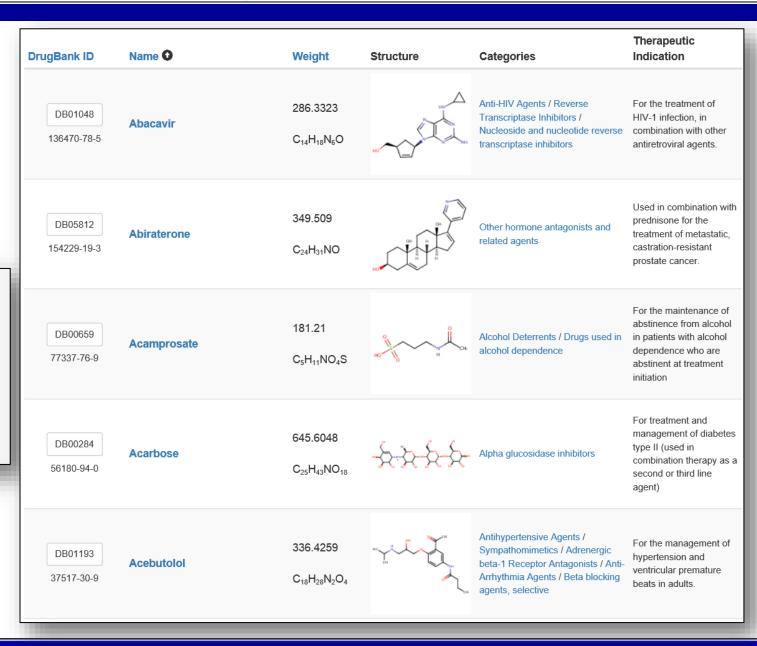
DrugBank Version 4.2

The DrugBank database is a unique bioinformatics and cheminformatics resource that combines detailed drug (i.e. chemical, pharmacological and

pharmaceutical) data with comprehensive drug target (i.e. sequence, structure, and pathway) information. The database contains 7759 drug entries including 1600 FDA-approved small molecule drugs, 160 FDA-approved biotech (protein/peptide) drugs, 89 nutraceuticals and over 6000 experimental drugs. Additionally, 4282 non-redundant protein (i.e. drug target/enzyme/transporter/carrier) sequences are linked to these drug entries. Each DrugCard entry contains more than 200 data fields with half of the information being devoted to drug/chemical data and the other half devoted to drug target or protein data.

http://www.drugbank.ca/

Wishart DS, Knox C, Guo AC, Cheng D, Shrivastava S, Tzur D, Gautam B, Hassanali M: DrugBank: a knowledgebase for drugs, drug actions and drug targets. Nucleic Acids Res 2008, , 36 Database: D901-6.





Drug Impacts on Erythrocytes

RBC_FVA_Drugs.ipynb

Set the environment

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    import numpy as np
    import pandas as pd
    import escher
    from escher import Builder
    from cobra.flux_analysis import flux_variability_analysis
    import matplotlib.pyplot as plot
    from cobrapy_bigg_client import client
    pd.set_option('display.max_rows', 1000)
    pd.set_option('display.width',1000)
    pd.set_option('display.max_colwidth',None)
```

Download the red blood cell model iAB RBC 283.json from the BIGG database to insure it is the latest version

```
In [2]: model = client.download_model('iAB_RBC_283', save=False) # Loading the model to the simulation
Academic license - for non-commercial use only - expires 2022-10-10
Using license file C:\Users\hinton\gurobi.lic
```

The model summary using the default conditions of the model

```
In [3]: model.summary()
```



Lesson Outline

- Alternate Optimal Solutions
- Flux Variability Analysis
- FVA Examples



Reflective Questions

- 1. What are alternate optimal solutions?
- 2. What is the relationship between alternate optimal solutions and a cell's phenotype?
- 3. What are silent phenotypes?
- 4. How many alternate optimal solutions can there be for a given phenotype?
- 5. How many alternate optimal solutions can there be for a carbon source?
- 6. Do aerobic/anaerobic conditions impact the number alternate optimal solutions?
- 7. Does the choice of objective function impact the number alternate optimal solutions?
- 8. What is flux variability analysis?
- 9. What is the relationship between the value of the objective function and the flux values calculated through flux variability analysis?
- 10. How is flux variability analysis related to alternate optimal flux vectors?
- 11. Does flux variability analysis identify the specific alternate optimal solutions?
- 12. What is the value of knowing which reactions carry flux, which reactions carry no flux, and which reactions span a range of flux values?



References

- 1. Reed, J. L. & Palsson, B. Ø. Genome-scale in silico models of *E. coli* have multiple equivalent phenotypic states: assessment of correlated reaction subsets that comprise network states. Genome Res. 14, 1797-1805 (2004).
- 2. Price, N. D., J. L. Reed, et al. (2004). "Genome-scale models of microbial cells: evaluating the consequences of constraints." Nature reviews. Microbiology 2(11): 886-897.
- 3. Bordbar, A., N. Jamshidi, et al. (2011). "iAB-RBC-283: A proteomically derived knowledge-base of erythrocyte metabolism that can be used to simulate its physiological and patho-physiological states." BMC systems biology 5: 110.
- 4. AmbergerJ, BocchiniCA, Scott AF, HamoshA: McKusick'sOnline Mendelian Inheritance in Man (OMIM). Nucleic Acids Res 2009, 37 Database: D793-6.
- 5. WishartDS, Knox C, Guo AC, Cheng D, ShrivastavaS, TzurD, GautamB, HassanaliM: DrugBank: a knowledgebase for drugs, drug actions and drug targets. Nucleic Acids Res 2008, , 36 Database: D901-6.
- Mahadevan, R. and C. H. Schilling (2003). "The effects of alternate optimal solutions in constraint-based genome-scale metabolic models." Metabolic engineering 5(4): 264-276
- 7. Phalakornkule, C. et al. A MILP-based flux alternative generation and NMR experimental design strategy for metabolic engineering. Metab. Eng. 3, 124-137 (2001).
- 8. Lee, S., Phalakornkule, C., Domach, M. M. & Grossmann, I. E. Recursive MILP model for finding all the alternate optima in LP models for metabolic networks. Comp. Chem. Eng. 24, 711-716 (2000).