

Review

Pre-Clinical Detection of Alzheimer's Disease Using FDG-PET, with or without Amyloid Imaging

Lisa Mosconi^{a,*}, Valentina Berti^{a,b}, Lidia Glodzik^a, Alberto Pupi^b, Susan De Santi^a and Mony J. de Leon^{a,c}

^a*Department of Psychiatry, New York University School of Medicine, New York, NY, USA*

^b*Department of Clinical Neurophysiology, Nuclear Medicine Unit, University of Florence, Italy*

^c*Nathan Kline Institute, Orangeburg, NY, USA*

Accepted 16 January 2010

Abstract. The development of prevention therapies for Alzheimer's disease (AD) would greatly benefit from biomarkers that are sensitive to subtle brain changes occurring in the preclinical stage of the disease. Early diagnostics is necessary to identify and treat at risk individuals before irreversible neuronal loss occurs. *In vivo* imaging has long been used to evaluate brain structural and functional abnormalities as predictors of future AD in non-demented persons. Prior to development of amyloid- β ($A\beta$) tracers for positron emission tomography (PET), the most widely utilized PET tracer in AD was 2-[¹⁸F]fluoro-2-Deoxy-D-glucose (FDG) PET. For over 20 years, FDG-PET has been used to measure cerebral metabolic rates of glucose (CMRglc), a proxy for neuronal activity, in AD. Many studies have shown that CMRglc reductions occur early in AD, correlate with disease progression, and predict histopathological diagnosis. This paper reviews reports of clinical and preclinical CMRglc reductions observed in association with genetic and non-genetic risk factors for AD. We then briefly review brain $A\beta$ PET imaging studies in AD and discuss the potential of combining symptoms-sensitive FDG-PET measures with pathology-specific $A\beta$ -PET to improve the early detection of AD.

Keywords: Amyloid- β , cerebral metabolic rate of glucose (CMRglc), normal aging, positron emission tomography, preclinical detection

INTRODUCTION

Alzheimer's disease (AD) is becoming an increasingly important reason for concern for healthcare and society. AD is the most common form of dementia, affecting approximately 10% of individuals 65 years of age, with the prevalence doubling every 5 years up to

age 80, above which the prevalence exceeds 40% [1]. In 2007, there were more than 26.6 million people affected by AD in the world [2]. The prevalence of AD is estimated to be further increasing in the next few years, as the baby-boomers generation ages [3]. The main reason for increasing prevalence in AD is the lack of disease-modifying treatments. Once disease-modifying drugs become available, they will likely be most effective if administered early in the course of disease, before irreversible brain damage has occurred. Therefore, another major problem in AD is the lack of diagnostic markers, especially for the early stages of disease when clinical symptoms are not clearly ex-

*Correspondence to: Lisa Mosconi, PhD, Department of Psychiatry, NYU School of Medicine 560 First Avenue, New York, NY 10016, USA. Tel.: +1 212 263 3255; Fax: +1 212 263 3279; E-mail: lisa.mosconi@nyumc.org.

pressed. Detection of preclinical pathological modifications in the brain is likely the key to identify individuals bound to develop AD.

Brain imaging, among other techniques, is a promising tool for the early detection of AD. Changes in brain histopathology, and consequently in its structure and function, are known to precede the clinical manifestations of disease by many years. These modifications can be visualized *in vivo* using brain imaging modalities. In chronological order, computerized tomography (CT) came first, followed by magnetic resonance (MR), and positron emission tomography (PET) with functional tracers such as 2- ^{18}F fluoro-2-Deoxy-D-glucose (FDG), and to a lesser extent with receptor ligands. Not yet 10 years ago, PET tracers for fibrillar amyloid- β ($\text{A}\beta$), the principal constituent of AD senile plaques, have been developed, which made detection of AD pathology *in vivo* a reality. Presently, many AD clinical and research centers perform $\text{A}\beta$ PET imaging, and an effort has been made in the United States to receive FDA approval for amyloid PET ligands in the diagnosis of AD and other dementias. However, the petition was rejected considering that detection of $\text{A}\beta$ would not be specific for an AD diagnosis, based on the observation that many elderly with $\text{A}\beta$ plaques never develop dementia and that other dementias share similar pathological substrates [4]. Much remains to be learned about fibrillar $\text{A}\beta$ as a biomarker for AD, especially at the early stages of disease. Prior to development of $\text{A}\beta$ PET tracers, FDG-PET was the most widely used PET technique in AD. For over 20 years, FDG-PET has been used to measure cerebral metabolic rates of glucose (CMRglc), a proxy for neuronal activity, in clinical AD patients and in at risk individuals. The present paper reviews FDG-PET findings in the early detection of AD, and discusses the value of performing FDG-PET with or without amyloid imaging.

WHY WE NEED BRAIN IMAGING FOR THE EARLY DETECTION OF AD

AD is a neurodegenerative disorder with insidious onset and progressive declines in memory, attention, and language [5]. Currently, the provisional diagnosis of AD remains based on clinical history, neurological examination, cognitive testing, and structural neuroimaging, while the definitive diagnosis of AD is based on the postmortem detection of specific pathological lesions: $\text{A}\beta$ plaques in the extracellular space and blood vessels, intracellular neurofibrillary tangles (NFT), and

neuronal and synaptic loss in specific brain regions [6]. There are no tests for the definitive diagnosis of AD *in vivo*. Ironically, we ultimately define the disease with pathological criteria, but we have hardly any information in this regard during life. As a result, patients may be misdiagnosed with AD when in fact they have another dementia or may be left undiagnosed [7]. The lack of standardized diagnostic tests for AD greatly limits the potential for an accurate diagnosis, and even more so, for early detection.

The problem is not what to measure, but how to measure it. Once the 'how' is resolved, the next question is when – how early in life can we detect clear-cut signs of an ongoing neurodegenerative process distinct from normal aging. Neurodegeneration in AD is estimated to begin 20–30 years before the clinical manifestations of disease become evident [8–11]. According to a popular theoretical model in AD, the "amyloid cascade hypothesis" [12], during this preclinical phase, $\text{A}\beta$ plaques and NFT load increase, causing synapse loss and neuronal death. In light of recent findings that $\text{A}\beta$ fibrils do not appear to be the main promoter of neuronal degeneration [12], the amyloid hypothesis was reformulated by stating that $\text{A}\beta$ oligomers confer neurotoxicity to neurons by disrupting nerve signaling pathways in AD [13,14].

While the causes of AD are being investigated, consensus exists as to where neurodegeneration strikes first in AD. The medial temporal lobes (MTL, i.e., hippocampus, transentorhinal/entorhinal cortex, and subiculum), which are critically involved in the neural control of memory functions, are most vulnerable to AD pathology [9,11,15–17]. The posterior cingulate, parieto-temporal, and frontal cortices become affected later in the course of disease, in keeping with progression of clinical symptoms [9,11,15–17]. The local and distant effects of AD pathology on tissue physiology impair neuronal function in these vulnerable regions [18], causing cognitive impairment and dementia [19]. While postmortem staging is based on cross-sectional detection of different patterns of anatomical involvement across subjects with different levels of dementia severity, longitudinal imaging studies enabled us to characterize the temporal progression of these regional brain deficits in the same individual, as discussed below.

WHY WE HAVE BEEN USING FDG-PET IMAGING

The early appearance of pathological lesions and the progressive nature of cognitive deterioration in AD in-

dicating a great need for developing biological markers of disease, sensitive to early, longitudinal changes. Until 10 years ago, when $A\beta$ PET imaging was developed, other technologies had to be used to measure surrogate markers of AD pathology. The most readily available techniques were magnetic resonance imaging (MRI), which is used to measure structural tissue loss (i.e., atrophy), and FDG-PET to measure the functional effects of neuronal activity at the tissue level.

A growing list of observations has highlighted the importance of FDG-PET as a tool to distinguish AD from other dementias, predict and track decline from normal cognition to AD, and to identify individuals at risk for AD prior to the onset of cognitive symptoms. What we have learned from over 30 years of FDG-PET research in AD is that, first of all, AD is characterized by a specific regional pattern of CMRglc reductions. AD patients show consistent CMRglc deficits in the parieto-temporal areas [20,21], posterior cingulate cortex (PCC) [22], and MTL [23]. As the disease progresses, frontal association cortices become involved, while cerebellum, striatum, basal ganglia, primary visual, and sensorimotor cortices remain preserved [21,23]. The extent and regional distribution of hypometabolism may vary across subjects, and hemispherical asymmetries are often noted [23,24], especially at the early stages of AD. Asymmetries are often detected in clinical practice and may be attributed to co-morbidity factors (e.g., vascular brain disease) or compensatory mechanisms (e.g., neuroplasticity), which would not be easily revealed by AD pathology imaging. This *in vivo* pattern of hypometabolism is found in the vast majority of clinically diagnosed AD patients and in over 85% pathologically confirmed AD cases [21].

CMRglc is highly correlated with clinical disabilities in dementia [25]. Clinical AD symptoms essentially never occur without CMRglc decreases, the extent of which is related to the severity of cognitive impairment [26–29]. Moreover, despite some overlap, the characteristic AD-pattern of CMRglc reductions yields high sensitivity in distinguishing AD from controls [30,31], from other neurodegenerative dementias, such as frontotemporal (FTD) and Lewy body dementia (DLB) [21,31], and from cerebrovascular disease [32]. In a large multi-center study of normal (NL), AD, FTD, and DLB subjects, individual FDG-PET scans were processed using automated voxel-based methods to generate disease-specific patterns of regional FDG uptake [31]. These standardized disease-specific PET patterns correctly classified 95% AD, 92% DLB, 94%

FTD, and 94% NL [31]. The method yielded high discrimination accuracy in patients with mild dementia as well as moderate-to-severe dementia [31]. Altogether, these findings support the use of FDG-PET in the differential diagnosis of the major neurodegenerative dementing disorders.

Second, CMRglc reductions on FDG-PET precede the onset of AD symptoms in predisposed individuals, in both genetic early-onset and late-onset AD forms. FDG-PET findings in preclinical AD are summarized in Table 1. Presymptomatic persons carrying autosomal dominant genetic mutations associated with early onset familial AD (EOFAD, onset age < 65 yrs) show the typical AD pattern of hypometabolism compared to age-matched mutation non-carriers [33–35]. FDG-PET abnormalities were observed up to 13 years prior to the onset of symptoms in EOFAD subjects [35]. While findings in EOFAD may not apply to the more common forms of late-onset AD, studies of patients with mild cognitive impairment (MCI) have reported similar evidence for presymptomatic CMRglc reductions. Among MCI patients, those presenting with more pronounced, or more AD-like, CMRglc reductions decline to AD at higher rates than those who do not show hypometabolism [22,36–38]. CMRglc reductions in MCI predict future AD with 75%–100% accuracy [22,36–41]. While early and late onset AD may or may not share a common pathology [12], FDG-PET studies have identified a similar outcome pattern of hypometabolism that appears to be a prodromal “metabolic signature” of AD independent of the age at onset of disease. More studies are needed to examine and compare the mechanisms underlying CMRglc reductions in the early and late onset AD.

A few FDG-PET studies showed an even earlier prediction capacity at the normal stages of cognition. By monitoring progression to MCI and AD among cognitively normal (NL) elderly, these studies showed that CMRglc reductions precede the onset of dementia by many years [42–45], and predict cognitive decline from NL cognition to MCI/AD with over 80% accuracy [42, 43]. The decliners to MCI and AD showed greater rates of CMRglc reductions as compared to the non-decliners [38,42–45]. Progressive CMRglc reductions were observed years in advance of clinical symptoms in a clinico-pathological series of subjects followed with longitudinal *in vivo* FDG-PET scans from normal cognition to the clinical diagnosis and to post-mortem confirmation of AD [45].

More work is needed to establish how early FDG-PET deficits become detectable in the course of dis-

Table 1
FDG- and PIB-PET findings in preclinical AD

At-risk group	Control group	FDG-PET findings vs. controls	References	PIB-PET findings vs. controls	References
Presymptomatic Early-onset Familial AD	Mutation Non-carriers	Cross-sectional – Whole brain hypometabolism – Parieto-temporal, PCC, frontal cortex, and MTL hypometabolism Longitudinal – Greater CMRglc declines over time	33	– Higher PIB retention in striatum	71, P 105 106
NL decliners to MCI and to AD	Stable NL	Cross-sectional (base- line data predicts clinical change) Longitudinal – MTL hypometabolism when NL – Parieto-temporal and PCC hypometabolism at time of decline to MCI/AD – Greater CMRglc declines over time	42 43	N.A. N.A. N.A.	
MCI decliners to AD	Stable MCI	Cross-sectional (base- line data predicts clinical change) Longitudinal – Parieto-temporal, PCC and frontal cortex hypometabolism – Greater CMRglc declines over time	22 38	– Higher PIB retention in PCC and frontal cortex – Higher cortical FDDNP binding – Increases in cortical FDDNP binding	80 89 89
NL with Subjective Memory Complaints	NL without Subjective Memory Complaints	Cross-sectional Longitudinal N.A.	48	N.A.	
NL ApoE-4 Carriers	NL ApoE-4 Non- carriers	Cross-sectional Longitudinal – Parieto-temporal, PCC, thalamus, and frontal cortex hypometabolism – Greater CMRglc declines over time	49 51,52	– Higher PIB retention in frontal, tem- poral, PCC/precuneus, parietal cortex and basal ganglia N.A.	83
NL Kibra CC carriers	NL Kibra CT and TT carriers	Cross-sectional Longitudinal – PCC/Precuneus hypometabolism N.A.	55	N.A. N.A.	
NL with a 1 st degree family history of late onset AD	NL with negative family history of AD	Cross-sectional Longitudinal – Parieto-temporal, PCC, frontal cortex, and MTL hypometabolism in NL with AD mothers as compared to those with AD fathers and to those with no parents with AD – Greater CMRglc declines over time in NL with AD mothers as compared to those with AD fathers and to those with no parents with AD	56 57	N.A. N.A.	

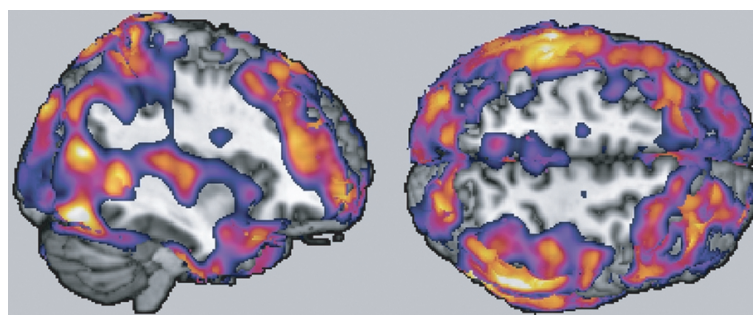


Fig. 1. Brain regions showing reduced CMRglc on FDG-PET in cognitively normal individuals with a maternal family history of AD as compared to demographically matched subjects with a paternal family history and with a negative family history of AD [57]. Statistical parametric maps showing regions of hypometabolism in NL individuals with a maternal history of AD are displayed on a purple-to-yellow color coded scale at $P < 0.001$. Figure shows the left lateral and superior views of a 3D volume-rendered MRI.

ease. Nonetheless, published studies show that non-demented individuals with reduced CMRglc are at increased risk for developing AD, which supports the use of FDG-PET in the early detection of AD.

CMRglc deficits resembling those in clinical AD patients have been observed in NL individuals at clinical or genetic risk for AD. With respect to AD risk established on clinical grounds, cognitively normal individuals with subjective memory complaints are regarded as a group at increased risk for dementia [46, 47]. On FDG-PET, middle-age to old normal individuals with subjective memory complaints showed CMRglc reductions in AD-vulnerable brain regions as compared to demographically matched individuals with no such complaints [48]. With respect to genetic risk factors for late-onset AD, many studies have shown that non-demented individuals carrying an apolipoprotein E (ApoE) $\epsilon 4$ allele have CMRglc reductions as compared to ApoE $\epsilon 4$ non-carriers [49–53]. CMRglc deficits in NL ApoE $\epsilon 4$ carriers are progressive, correlate with reductions in cognitive performance [49, 52], and occur in young adulthood [53]. Likewise, NL carriers of the KIBRA CC haplotype, a risk factor for memory impairment in late life [54], showed CMRglc reductions as compared to low-risk KIBRA TT and CT carriers [55].

First degree relatives of AD patients also appear to be at high risk for late onset AD. In particular, a maternal history of AD was shown to affect brain metabolism in NL individuals [56, 57]. CMRglc deficits in AD-vulnerable regions were observed in NL with a maternal family history of AD as compared to those with a paternal history and those with no family history of AD (Fig. 1) [56]. Interestingly, NL with AD fathers did not show CMRglc abnormalities [56]. Over a 2-year period, NL individuals with an AD mother showed progressive declines in regional CMRglc compared to

those with no parents with AD as well as to those with an AD father [57]. The genetic mechanisms that underlie maternally inherited CMRglc reductions are under investigation [58]. More studies are needed to replicate these first reports and to identify the genetic factors involved in hypometabolism in preclinical AD.

Overall, the major strengths of FDG-PET in AD can be summarized as: high sensitivity to distinguish AD from controls and from other neurodegenerative diseases, and individuals at higher versus lower AD risk, and good quantitative and topographical correlation with clinical progression. However, a major limitation to most of the above FDG-PET studies is the absence of postmortem data. Doubt remains as to whether clinical symptoms and CMRglc reductions are due to AD pathology or to other causes. Using clinical diagnosis as the gold-standard may lead to erroneously include patients with a dementia other than AD in the AD group, and vice versa. In asymptomatic subjects showing hypometabolism, CMRglc deficits may develop for reasons other than AD, and not all subjects showing hypometabolism will necessarily decline to AD. Here is where, in our opinion, imaging of AD pathology plays an essential role.

WHEN FDG-PET ALONE IS NOT ENOUGH, AND THE ADVENT OF AMYLOID PET TRACERS

Several PET tracers for $A\beta$ plaques have been developed in the last few years. The best known tracers are *N*-methyl- ^{11}C 2-(4'-methylaminophenyl)-6-hydroxybenzothiazole, aka Pittsburgh Compound-B (PIB) [59], 4-N- ^{11}C -methylamino-4'-hydroxystilbene (SB13) [60], 2-(1-96-(2- ^{18}F -fluoroethyl)(methyl)

amino)-2-naphthyl)ethylidene)malononitrile (FDDNP) [61], and more recently the *trans*-4-(*N*-methyl-amino)-4'-{2-[2-(2-[18F]fluoro-ethoxy)-ethoxy]-ethoxy}-stilbene (BAY94-9172) [62]. Among these tracers, PIB is the most widely utilized and best characterized in terms of tracer kinetics, modeling, and analytic methods. PIB binds to fibrillar A β plaques with high affinity [63]. Several PIB-PET studies demonstrated significant PIB retention in AD patients as compared to controls, mostly in the frontal cortex, parieto-temporal, PCC/precuneus, occipital lobes, thalamus, and striatum [63–67], consistent with the known pattern of A β plaques deposition observed at postmortem. Significant PIB retention is found in over 90% clinically diagnosed AD patients, in as many as 60% of MCI [65,64, 66–70], and 30% of NL elderly [67]. PIB-PET showed higher A β load in asymptomatic and symptomatic individuals carrying presenilin-1 (PSEN1) and A β PP mutations as compared to controls [71]. PIB retention was especially high in the striatum of mutation carriers as compared to controls and to sporadic AD patients [71]. These findings suggest that the striatum may be more affected in early onset AD and the neocortex in the late onset AD forms.

Interestingly, patients can be easily dichotomized as showing either significant (PIB+) or absent PIB retention (PIB-), but hardly show intermediate levels [65,64, 66–70]. This could facilitate interpretation of PIB-PET scans for clinical use. The presence of a PIB+ pattern has been shown to improve the differential diagnosis of AD from FTD and from Parkinson's disease [67, 72]. However, significant PIB retention is observed in DLB [64], and in patients with cerebral amyloid angiopathy (CAA) [73]. The impact of vascular amyloid on PIB signal is particularly relevant in view of using PIB, or other A β tracers, in the early detection of AD. A study showed that PIB is not specific for dense, classical plaques [74], but rather binds to a family of amyloid substrates ranging from diffuse plaques to plaques in the vascular system (i.e., CAA) [75].

Vascular pathology is common in the elderly, and it is not known how much of the PIB retention observed in NL elderly is due to vascular A β deposits. Moreover, it is known from postmortem studies that typical amyloid and NFT lesions are found in both demented and non-demented individuals [10,11,76–78]. Non-demented cases with substantial AD pathology are often described as a 'preclinical' AD group given the absence of cognitive abnormalities [10,78]. This observation brings up the important, and often overlooked, discrepancy between AD-pathology and AD-

dementia [79]. Those NL and MCI showing an AD-like PIB pattern (and therefore A β pathology) are conceivably at higher risk for developing AD-dementia as compared to individuals without brain A β pathology. However, having A β plaques does not equal to being at a 'pre-dementia stage', and the prognostic value of increased A β load on PET has to be established. Since A β imaging is a relatively new technique, there are not enough published longitudinal PIB-PET studies to draw conclusions on its preclinical value in AD (Table 1). A few studies in MCI have shown that, at baseline, those MCI who later declined to AD showed higher PIB retention as compared to the non-decliners [80–82]. There are no published PIB-PET reports in NL individuals declining to AD. A recent PIB-PET study in middle aged to old NL individuals showed significantly higher PIB retention in NL ApoE ϵ 4 carriers compared to non-carriers [83]. Although the predictive value of PIB abnormalities in the asymptomatic ApoE ϵ 4 carriers is not known, PIB measures may be useful to discriminate NL individuals at higher versus lower risk for AD [83].

However, the correlation between PIB retention and cognition is generally fairly weak [84], consistent with the notion that A β plaques distribution does not correlate with clinical symptoms in AD [85]. Additionally, the few published longitudinal PIB-PET papers indicate a lack of progression of PIB uptake in NL, MCI, and AD [86–88]. AD patients apparently reach a plateau in PIB retention, despite progression of their clinical symptoms and worsening of hypometabolism on FDG-PET [86]. Jack and colleagues [87] examined longitudinal PIB-PET in NL, MCI, and AD and showed that the rate of PIB change did not differ by clinical group. The lack of longitudinal progression suggests that PIB deposition could be an early event during aging and disease. Otherwise, lack of change suggests that PIB and similar tracers may not be the best option for longitudinal studies (discussed below). In this respect, FDDNP appears to have an advantage over PIB, since FDDNP studies showed some longitudinal effects [61]. FDDNP binds both A β fibrils and NFT, and shows a cortical binding pattern similar to PIB, and additionally binds to the MTL [61,89]. Moreover, MCI patients showed intermediate FDDNP levels between NL and AD, demonstrating finer grading than PIB measures [89]. FDDNP uptake was highly correlated with scores on memory and global cognition [89]. Although limited by the small sample, longitudinal progression effects were reported for 3 non-demented subjects that deteriorated over 2 years, including one subject that

declined from NL to MCI, and 2 MCI that converted to AD [89]. The major limitation to using FDDNP is the low specific to non-specific binding ratio of the tracer [61,89] which makes these scans difficult to interpret for clinical use.

USING FDG-PET WITH OR WITHOUT AMYLOID IMAGING FOR THE EARLY DETECTION OF AD

Given the low specificity of FDG-PET for AD pathology, the addition of amyloid PET tracers may be useful in the early detection of AD. An effective strategy to increase the preclinical diagnostic accuracy would be to combine the sensitivity of FDG-PET with pathology-specific $A\beta$ measures.

For clinical purposes, amyloid-PET appears to be most useful to distinguish AD from non-amyloid dementias, such as FTD. Such capacity may prove particularly useful at the mild stages of dementia, when symptoms are not fully expressed, and FDG-PET scans may not show clear-cut regional metabolic abnormalities. Amyloid imaging would be suitable to rule out AD in the presence of a PIB- scan, since a demented patient without brain $A\beta$ cannot have AD-dementia by definition. However, amyloid imaging may not be sufficient to rule in AD. If a patient with uncertain diagnosis is PIB+, it would not be possible to distinguish between AD, DLB, and CAA based on PIB alone.

Diagnosis of AD at early stages of dementia would be more problematic because many NL elderly with brain $A\beta$ deposits never develop dementia in life [90]. Amyloid imaging is necessary for the early detection of $A\beta$ pathology, but may not be sufficient to make an early diagnosis of AD-dementia. Histology studies have shown that, among individuals with AD pathology, what differentiates demented from non-demented subjects is the presence of neuronal loss. In general, non-demented subjects with AD pathology do not show neuronal loss at postmortem, while demented subjects with AD pathology show decreases in neuronal number and volume [90]. These findings indicate that neuronal degeneration is a stronger predictor of dementia than AD pathology [90]. Therefore, functional tracers like FDG-PET, whose signal correlates well with cognitive impairment, may be needed to appreciate the extent to which $A\beta$ is affecting brain function. Non-demented individuals showing increased $A\beta$ load and reduced CMRglc would be the ideal target population for prevention studies in AD (Fig. 2).

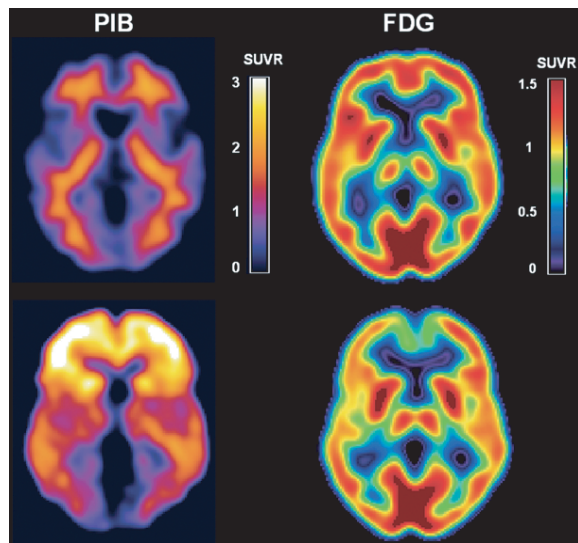


Fig. 2. Coregistered PIB and FDG-PET scans in 2 representative cognitively normal individuals at conceivably low risk for AD (top row: negative PIB and normal FDG uptake), and at high risk for AD (bottom row: positive PIB and reduced FDG uptake) based on PET imaging findings. Cerebral-to-cerebellar PIB Standardized Uptake Value ratios (SUVR) are displayed for each modality using a color coded scale.

FDG-PET studies have shown preclinical CMRglc abnormalities in individuals in their 40's [53]. It is not known how early in the course of disease $A\beta$ depositions can be detected. Except for the known presence of amyloid deposits in young individuals with Down's syndrome [91,92], $A\beta$ plaques are more prevalent in brains of individuals older than age 50 years [93,94]. It was hoped that $A\beta$ imaging would facilitate the study of the time course of amyloid deposition in brain. However, people appear to either have substantial brain $A\beta$ or not and remain relatively unchanged over time. This could be due to the fact that $A\beta$ deposition is a very early event in AD. Should this be the case, then amyloid tracers may be more useful for longitudinal examination of younger individuals with minimal tracer uptake, who may still show progression effects. Otherwise, lack of effects could be due to technical issues, such as the intrinsically low spatial resolution of PET scanners, or to the fact that PIB uptake reflects the presence of $A\beta$ fibrils, but not fibrils' dimension or growth [95,98]. Ever since the first validation studies [96,97], PIB analysis has been based on simplified reference tissue models from receptor studies, which treat $A\beta$, a polymer, as if it were a receptor [98]. There is a conceptual difference between imaging the density of $A\beta$ fibril polymers and neuronal receptors. Based on this observation, the concept of $A\beta$ molecular imaging probes was introduced

as a new paradigm that goes beyond classic binding potential parameters to include binding characteristics to polymeric peptide aggregates [95,99]. This would ideally increase resolving power in characterizing the progression of A β , especially for subjects presenting with substantial uptake at the first examination.

The exact role played by A β plaques in AD is not clear. Recent studies have shown that A β dimers and oligomers, not plaques, promote neuronal degeneration [12,13,100,101]. A β plaques represent a fraction of total A β in the brain that has been condensed and neutralized, and no longer contributes to neurotoxicity [101]. Measurement of soluble A β is needed to correctly estimate risk for developing AD, but tracers for soluble A β are not available. Measurement of fibrillar A β , as achieved with PET, could be seen in two opposing ways: either as an index of how much soluble A β the brain has been dealing with, and therefore as a sign of increased risk, or as an index of how well the brain has been getting rid of toxic A β , and therefore as a sign that the brain is strong enough to cope with the bad A β . There is evidence that highly educated AD patients show increased PIB uptake and lower CMRglc as compared to patients with low education, but with a similar degree of cognitive impairment [102, 103]. The results support the hypothesis that 'cognitive reserve' influences the association between A β pathology and cognition [102,103]. The question of whether A β predisposes to AD-dementia will be answered once treatment against fibrillar A β becomes available.

Finally, in the enthusiasm of being able to image A β *in vivo*, NFT have been somewhat neglected. While the relationship between these two abnormal proteins in AD is still under investigation [85,100], the diagnosis of AD remains based on the presence of both plaques, NFT, and neuronal loss with a specific neuroanatomy. Imaging NFT is particularly important as NFT progression follows the expected pattern of regional involvement based on clinical symptoms [11], and unlike A β plaques, NFT load correlates with cognitive impairment in AD [104]. *In vivo* imaging of NFT is still under development.

In conclusion, much research has been done with FDG-PET in AD since the first studies in the 1980s. Thanks to the technique's sensitivity to progression effects, FDG-PET is a candidate modality for detecting functional brain changes in early AD. Technical improvements, particularly the enhanced resolution of modern PET-CT and HRRT scanners, have led to increased anatomical accuracy, providing the possibility to detect energetic changes within the neuro-vascular

unit, as well as to identify "specific" patterns of cortical and subcortical hypometabolism to distinguish AD from other dementias at an early stage [107]. Nonetheless, there remains a great need to increase preclinical diagnostic specificity.

It is possible that the combination of dementia-sensitive CMRglc with pathology-specific A β and NFT imaging would improve the early, differential diagnosis of AD. Additional validation studies are needed before A β PET imaging can enter into clinical practice, and more longitudinal studies are necessary to establish the limits and strengths PET for early diagnosis of AD. Accurate characterization of the extent and nature of brain damage in individual patients, based on converging evidence from different biomarkers, will likely play an important role in the prediction of subjects' clinical course. Other potential benefits include the selection of individualized treatment plans and screening of patients with more uniform underlying pathology for targeted research and drug trials. Hopefully, continued technological progress will one day allow us to image all aspects of AD pathology *in vivo*, at proper microscopic resolution, without the need for invasive procedures.

ACKNOWLEDGMENTS

This study was supported by NIH/NIA grants AG032554 and AG13616, NIH/NCRR grant M01-RR0096, the Alzheimer's Association, and an Anonymous foundation.

Authors' disclosures available online (<http://www.j-alz.com/disclosures/view.php?id=290>).

REFERENCES

- [1] Kukull WA, Higdon R, Bowen JD, McCormick WC, Teri L, Schellenberg GD, van Belle, Jolley, Larson EB (2002) Dementia and Alzheimer disease incidence: A prospective cohort study. *Arch Neurol* **59**, 1737-1746.
- [2] Brookmeyer R, Johnson E, Ziegler-Graha K, Arrighi HM (2007) Forecasting the global burden of Alzheimer's disease. *Alzheimers Dementia* **3**, 186-191.
- [3] Hebert LE, Scherr PA, Bienias JL, Bennett DA, Evans DA (2004) State-specific projections through 2025 of Alzheimer disease prevalence. *Neurology* **62**, 1645.
- [4] Talan J (2008) Neuroimaging tracers for AD detection not yet ready for prime time, FDA Panel advises. *Neurol Today* **8**, 7-8.
- [5] McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM (1984) Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology* **34**, 939-944.

- [6] Mirra SS, Heyman A, McKeel D, Sumi SM, Crain BJ, Brownlee LM, Fogel FS, Hughes JP, van Belle, G, Berg L (1991) The consortium to establish a registry for Alzheimer's disease (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. *Neurology* **41**, 479-486.
- [7] Hebert LE, Scherr PA, Bienias JL, Bennett DA, Evans DA (2003) Alzheimer disease in the US population: prevalence estimates using the 2000 census. *Arch Neurol* **60**, 1119-1122.
- [8] Braak H, Braak E (1996) Development of Alzheimer-related neurofibrillary changes in the neocortex inversely recapitulates cortical myelogenesis. *Acta Neuropathol* **92**, 197-201.
- [9] Delacourte A, David JP, Sergeant N, Buee L, Wattez A, Vermeersch P, Ghazali F, Fallet-Bianco C, Pasquier F, Lebert F, Petit H, Di Menza C (1999) The biochemical pathway of neurofibrillary degeneration in aging and Alzheimer's disease. *Neurology* **52**, 1158-1165.
- [10] Morris JC, Storandt M, McKeel DW, Rubin EH, Price JL, Grant EA, Berg L (1996) Cerebral amyloid deposition and diffuse plaques in "normal" aging: Evidence for presymptomatic and very mild Alzheimer's disease. *Neurology* **46**, 707-719.
- [11] Braak H, Braak E (1991) Neuropathological staging of Alzheimer-related changes. *Acta Neuropathol* **8**, 239-259.
- [12] Selkoe DJ (1997) Alzheimer's disease: genotypes, phenotype, and treatments. *Science* **275**, 630-631.
- [13] Lambert MP, Barlow AK, Chromy BA, Edwards C, Freed R, Liosatos M, organ TE, Rozovsky I, Tronner B, Viola KL, Wals P, Zhang C, Finch CE, Krafft GA, Klein WL (1998) Diffusible, nonfibrillar ligands derived from Abeta1-42 are potent central nervous system neurotoxins. *Proc Natl Acad Sci U S A* **95**, 6448-6453.
- [14] Lue LF, Kuo YM, Roher AE, Brachova L, Shen Y, Sue L, Beach T, Kurth JH, Rydel RE, Rogers J (1999) Soluble amyloid beta peptide concentration as a predictor of synaptic change in Alzheimer's disease. *Am J Pathol* **155**, 853-862.
- [15] Arriagada PV, Marzloff K, Hyman BT (1992) Distribution of Alzheimer-type pathologic changes in nondemented elderly individuals matches the pattern in Alzheimer's disease. *Neurology* **42**, 1681-1688.
- [16] Giannakopoulos P, Hof PR, Mottier S, Michel JP, Bouras C (1994) Neuropathological changes in the cerebral cortex of 1258 cases from a geriatric hospital: retrospective clinicopathological evaluation of a 10-year autopsy population. *Acta Neuropathol* **87**, 456-468.
- [17] Ulrich J (1985) Alzheimer changes in nondemented patients younger than sixty-five: Possible early stages of Alzheimer's disease and senile dementia of Alzheimer type. *Ann Neurol* **17**, 273-277.
- [18] Morrison JH, Hof PR (1997) Life and death of neurons in the aging brain. *Science* **278**, 412-419.
- [19] Hyman BT, Van Hoesen GW, Damasio AR, Barnes CL (1984) Alzheimer's disease: cell-specific pathology isolates the hippocampal formation. *Science* **225**, 1168-1170.
- [20] Mazziotta JC, Phelps ME (1986) Positron Emission Tomography studies of the brain. In: Phelps ME, Mazziotta JC, Schelbert H, editors. *Positron Emission Tomography & Autoradiography: Principles & Applications for the Brain & Heart*. New York: Raven Press, pp. 493-579.
- [21] Silverman DHS, Small GW, Chang CY, Lu CS, Kung de Aburto MA, Chen W, Czernin J, Rapoport SI, Pietrini P, Alexander GE, Schapiro MB, Jagust WJ, Hoffman JM, Welsh-Bohmer KA, Alavi A, Clark CM, Salmon E, de Leon MJ, Mielke R, Cummings JL, Kowell AP, Gambhir SS, Hoh CK, Phelps ME (2001) Positron emission tomography in evaluation of dementia: Regional brain metabolism and long-term outcome. *JAMA* **286**, 2120-2127.
- [22] Minoshima S, Giordani B, Berent S, Frey KA, Foster NL, Kuhl DE (1997) Metabolic reduction in the posterior cingulate cortex in very early Alzheimer's disease. *Ann Neurol* **42**, 85-94.
- [23] Mosconi L (2005) Brain glucose metabolism in the early and specific diagnosis of Alzheimer's disease. *Eur J Nucl Med* **32**, 486-510.
- [24] Kim EJ, Cho SS, Jeong Y, Park KC, Kang SJ, Kang E, Kim SE, Lee KH, Na DL (2005) Glucose metabolism in early onset versus late onset Alzheimer's disease: an SPM analysis of 120 patients. *Brain* **128**, 1790-1801.
- [25] Blass JP (2002). Alzheimer's disease and Alzheimer's dementia: distinct but overlapping entities. *Neurobiol Aging* **23**, 1077-1084.
- [26] Grady CL, Haxby JV, Schlageter NL, Berg G, Rapoport SI (1986) Stability of metabolic and neuropsychological asymmetries in dementia of the Alzheimer type. *Neurology* **36**, 1390-1392.
- [27] Haxby JV, Grady CL, Koss E, Horwitz B, Heston L, Schapiro M, Friedland R, Rapoport SJ (1990) Longitudinal study of cerebral metabolic asymmetries and associated neuropsychological patterns in early dementia of the Alzheimer type. *Arch Neurol* **47**, 753-760.
- [28] Desgranges B, Baron J-C, De La Sayette V, Petit-Taboué MC, Benali K, Landeau B, Lechevalier B, Eustache F (1998) The neural substrates of memory systems impairment in Alzheimer's Disease a PET study of resting brain glucose utilization. *Brain* **121**, 611-631.
- [29] Brown AM, Sheu RK, Mohs R, Haraoutian V, Blass JP (2001) Correlation of the clinical severity of Alzheimer's disease with an aberration in mitochondrial DNA (mtDNA). *J Mol Neurosci* **16**, 41-48.
- [30] Herholz K, Salmon E, Perani D, Baron JC, Holthoff V, Frölich L, Schönknecht P, Ito K, Mielke R, Kalbe E, Zündorf G, Delbeuck X, Pelati O, Anchisi D, Fazio F, Kerrouche N, Desgranges B, Eustache F, Beuthien-Baumann B, Menzel C, Schröder J, Kato T, Arahata Y, Henze M, Heiss WD (2002) Discrimination between Alzheimer dementia and controls by automated analysis of multicenter FDG PET. *Neuroimage* **17**, 302-316.
- [31] Mosconi L, Tsui WH, Herholz K, Pupi A, Drzezga A, Lucignani G, Reiman EM, Holthoff V, Kalbe E, Sorbi S, Diehl-Schmid J, Pernecky R, Mariani C, Caselli R, Beuthien-Baumann B, Kurz A, Minoshima S, de Leon MJ (2008) Multi-center standardized FDG-PET diagnosis of Mild Cognitive Impairment, Alzheimer's disease and other dementias. *J Nucl Med* **49**, 390-398.
- [32] Szelies B, Mielke R, Herholz K, Heiss W-D (1994) Quantitative topographical EEG compared to FDG PET for classification of vascular and degenerative dementia. *Electroencephalogr Clin Neurophysiol* **91**, 131-139.
- [33] Kennedy AM, Newman SK, Frackowiak RS, Cunningham VJ, Roques P, Stevens J, Neary D, Bruton CJ, Warrington EK, Rossor MN (1995) Chromosome 14 linked familial Alzheimer's disease. A clinico-pathological study of a single pedigree. *Brain* **118**, 185-205.
- [34] Kennedy AM, Frackowiak RSJ, Newman SK, Bloomfield PM, Seaward J, Roques P, Stevens J, Neary D, Bruton CJ, Warrington EK, Rossor MN (1995) Deficits in cerebral glucose metabolism demonstrated by positron emission tomography in individuals at risk of familial Alzheimer's Disease. *Neurosci Lett* **186**, 17-20.

- [35] Mosconi L, Sorbi S, de Leon MJ, Li Y, Nacmias B, Myoung PS, Tsui W, Bessi V, Fayyaz M, Caffarra P, Pupi A (2006) Hypometabolism exceeds atrophy in presymptomatic early-onset Familial Alzheimer's disease. *J Nucl Med* **47**, 1778-1786.
- [36] Chetelat G, Desgranges B, De La Sayette V, Viader F, Eustache F, Baron JC (2003) Mild cognitive impairment: Can FDG-PET predict who is to rapidly convert to Alzheimer's disease? *Neurology* **60**, 1374-1377.
- [37] Mosconi L, Perani D, Sorbi S, Herholz K, Nacmias B, Holthoff V, Salmon E, Baron JC, Padovani A, Borroni B, Franceschi M, Bracco L, Pupi A (2004) MCI conversion to dementia and the APOE genotype: a prediction study with FDG-PET. *Neurology* **63**, 2332-2340.
- [38] Drzezga A, Lautenschlager N, Siebner H, Riemenschneider M, Willech F, Minoshima S, Schwaiger M, Kurz A (2003) Cerebral metabolic changes accompanying conversion of mild cognitive impairment into Alzheimer's disease: a PET follow-up study. *Eur J Nucl Med Mol Imaging* **30**, 1104-1113.
- [39] Herholz K, Nordberg A, Salmon E, Perani D, Kessler J, Mielke R, Halber M, Jelic V, Almkvist O, Collette F, Alberoni M, Kennedy A, Hasselbalch S, Fazio F, Heiss WD (1999) Impairment of neocortical metabolism predicts progression in Alzheimer's disease. *Dement Geriatr Cogn Dis* **10**, 494-504.
- [40] Drzezga A, Grimmer T, Riemenschneider M, Lautenschlager N, Siebner H, Alexopoulos P, Minoshima S, Schwaiger M, Kurz A (2005) Prediction of individual outcome in MCI by means of genetic assessment and 18F-FDG PET. *J Nucl Med* **46**, 1625-1632.
- [41] Anchisi D, Borroni B, Franceschi M, Kerrouche N, Kalbe E, Beuthien-Beumann B, Cappa S, Lenz O, Ludecke S, Marccone A, Mielke R, Ortelli P, Padovani A, Pelati O, Pupi A, Scarpini E, Weisenbach S, Herholz K, Salmon E, Holthoff V, Sorbi S, Fazio F, Perani D (2005) Heterogeneity of brain glucose metabolism in mild cognitive impairment and clinical progression to Alzheimer disease. *Arch Neurol* **62**, 1728-1733.
- [42] de Leon MJ, Convit A, Wolf OT, Tarshish CY, De Santi S, Rusinek H, Tsui W, Kandil E, Scherer AJ, Roche A, Imossi A, Thorn E, Bobinski M, Caraos C, Lesbre P, Schlyer D, Poirier J, Reisberg B, Fowler J (2001) Prediction of cognitive decline in normal elderly subjects with 2-[18F]fluoro-2-deoxy-D-glucose/positron-emission tomography (FDG/PET). *Proc Natl Acad Sci U S A* **98**, 10966-10971.
- [43] Mosconi L, De Santi S, Li J, Tsui WH, Li Y, Boppana M, Laska E, Rusinek H, de Leon MJ (2008) Hippocampal hypometabolism predicts cognitive decline from normal aging. *Neurobiol Aging* **29**, 676-692.
- [44] Jagust WJ, Gitcho A, Sun F, Kuczynski B, Mungas D, Haan M (2006) Brain imaging evidence of preclinical Alzheimer's disease in normal aging. *Ann Neurol* **59**, 673-681.
- [45] Mosconi L, Mistur R, Switalski R, Tsui WH, Glodzik L, Brys M, Li Y, Pirraglia E, De Santi S, Reisberg B, Wisniewski T, de Leon MJ (2009) Longitudinal changes in brain glucose metabolism from normal cognition to pathologically verified Alzheimer's disease. *Eur J Nucl Med Mol Imaging* **36**, 811-822.
- [46] Geerlings MI, Jonker C, Bouter LM, Ader HJ, Schmand B (1999) Association between memory complaints and incident Alzheimer's disease in elderly people with normal baseline cognition. *Am J Psychiat* **156**, 531-537.
- [47] Reisberg B, Prichep L, Mosconi L, John ER, Glodzik-Sobanska L, Boksay I, Monteiro I, Torossian C, Vedvyas A, Ashraf N, Jamil IA, de Leon MJ (2008) The pre-Mild Cognitive Impairment, Subjective Cognitive Impairment stage of Alzheimer's disease. *Alzheimers Dementia* **4**, S98-S108.
- [48] Mosconi L, De Santi S, Brys M, Tsui WH, Pirraglia E, Glodzik-Sobanska L, Rich KE, Switalski R, Mehta PD, Pratico D, Zinkowski R, Blennow K, de Leon MJ (2008) Hypometabolism and altered CSF markers in normal ApoE E4 carriers with subjective memory complaints. *Biol Psychiatry* **63**, 609-618.
- [49] Small GW, Mazziotta JC, Collins MT, Baxter LR, Phelps ME, Mandelkern MA, Kaplan A, La Rue A, Adamson CF, Chang L (1995) Apolipoprotein E type 4 allele and cerebral glucose metabolism in relatives at risk for familial Alzheimer disease. *JAMA* **273**, 942-947.
- [50] Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, Thobodu SN, Osborne D (1996) Preclinical evidence of Alzheimer's disease in persons homozygous for the E4 allele for apolipoprotein E. *N Engl J Med* **334**, 752-758.
- [51] Small GW, Ercoli LM, Silverman DHS, Huang SC, Komo S, Bookheimer S, Lavretsky H, Miller K, Siddarth P, Rasgon NL, Mazziotta JC, Saxena S, Wu HM, Mega MS, Cummings JL, Saunders AM, Pericak-Vance MA, Roses AD, Barrio JR, Phelps ME (2000) Cerebral metabolic and cognitive decline in persons at genetic risk for Alzheimer's disease. *Proc Natl Acad Sci U S A* **97**, 6037-6042.
- [52] Reiman EM, Caselli RJ, Chen K, Alexander GE, Bandy D, Frost J (2001) Declining brain activity in cognitively normal apolipoprotein E epsilon 4 heterozygotes: A foundation for using positron emission tomography to efficiently test treatments to prevent Alzheimer's disease. *Proc Natl Acad Sci U S A* **98**, 3334-3339.
- [53] Reiman EM, Chen K, Alexander GE, Caselli RJ, Bandy D, Osborne D, Saunders AM, Hardy J (2004) Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia. *Proc Natl Acad Sci U S A* **101**, 284-289.
- [54] Papassotiropoulos A, Stephan DA, Huentelman MJ, Hoernndli FJ, Craig DW, Pearson JV, Huynh KD, Brunner F, Corneveaux J, Osborne D, Wollmer MA, Aerni A, Coluccia D, Hänggi J, Mondadori CR, Buchmann A, Reiman EM, Caselli RJ, Henke K, de Quervain DJ (2006) Common kibra alleles are associated with human memory performance. *Science* **314**, 475-478.
- [55] Corneveaux JJ, Liang WS, Reiman EM, Webster JA, Myers AJ, Zismann VL, Joshupura KD, Pearson JV, Hu-Lince D, Craig DW, Coon KD, Dunckley T, Bandy D, Lee W, Chen K, Beach TG, Mastroeni D, Grover A, Ravid R, Sando SB, Aasly JO, Heun R, Jessen F, Kölsch H, Rogers J, Hutton ML, Melquist S, Petersen RC, Alexander GE, Caselli RJ, Papassotiropoulos A, Stephan DA, Huentelman MJ (2010) Evidence for an association between KIBRA and late-onset Alzheimer's disease. *Neurobiol Aging* **31**, 901-909.
- [56] Mosconi L, Brys M, Switalski R, Mistur R, Glodzik L, Pirraglia E, Tsui WH, De Santi S, de Leon MJ (2007) Maternal family history of Alzheimer's disease predisposes to reduced brain glucose metabolism. *Proc Natl Acad Sci U S A* **104**, 19067-19072.
- [57] Mosconi L, Mistur R, Glodzik L, Pirraglia E, Tsui WH, De Santi S, de Leon MJ (2009) Declining brain glucose metabolism in normal individuals with a maternal history of Alzheimer's. *Neurology* **72**, 513-520.
- [58] Mosconi L, Berti V, Swerdlow RH, Mistur R, Pupi A, Duara R, de Leon MJ (2010) Maternal transmission of Alzheimer's

- disease: Prodromal metabolic phenotype and the search for genes. *Human Genom* **4**, 170-193.
- [59] Mathis CA, Bacskai BJ, Kajdasz ST, McLellan ME, Frosch MP, Hyman BT, Holt DP, Wang Y, Huang GF, Debnath, ML Klunk WE (2002) A lipophilic thioflavin-T derivative for positron emission tomography (PET) imaging of amyloid in brain. *Bioorganic Med Chem Lett* **12**, 295-298.
- [60] Ono M, Wilson A, Nobrega J, Westaway D, Verhoeff P, Zhuang ZP, Kung MP, Kung HF (2003) ¹¹C-labeled stilbene derivatives as Abeta-aggregate-specific PET imaging agents for Alzheimer's disease. *Nucl Med Biol* **30**, 565-571.
- [61] Agdeppa ED, Kepe V, Liu J, Flores-Torres S, Satyamurthy N, Petric A, Cole GM, Small GW, Huang SC, Barrio JR (2001) Binding characteristics of radiofluorinated 6-dialkylamino-2-naphthylethylidene derivatives as positron emission tomography imaging probes for beta-amyloid plaques in Alzheimer's disease. *J Neurosci* **21**, 1-5.
- [62] Rowe CC, Ackermann U, Browne W, Mulligan R, Pike KL, O'Keefe G, Tochon-Danguy H, Chan G, Berlangieri SU, Jones G, Dickinson-Rowe KL, Kung HP, Zhang W, Kung MP, Skovronsky D, Dyrks T, Holl G, Krause S, Friebe M, Lehman L, Lindemann S, Dinkelborg LM, Masters CL, Villenaghe VL (2008) Imaging of amyloid beta in Alzheimer's disease with (18)F-BAY94-9172, a novel PET tracer: proof of mechanism. *Lancet Neurol* **7**, 129-135.
- [63] Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, Holt DP, Bergström M, Savitcheva I, Huang GF, Estrada S, Ausén B, Debnath ML, Barletta J, Price JC, Sandell J, Lopresti BJ, Wall A, Koivisto P, Antoni G, Mathis CA, Långström B (2004) Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. *Ann Neurol* **55**, 306-319.
- [64] Rowe CC, Ng S, Ackermann U, Gong SJ, Pike K, Savage G, Cowie TF, Dickinson KL, Maruff P, Darby D, Smith C, Woodward M, Merory J, Tochon-Danguy H, O'Keefe G, Klunk WE, Mathis CA, Price JC, Masters CL, Villenaghe VL (2007) Imaging {beta}-amyloid burden in aging and dementia. *Neurology* **68**, 1718-1725.
- [65] Kemppainen N, Aalto S, Wilson I, Nagren K, Nägren K, Helin S, Brück A, Oikonen V, Kailajärvi M, Scheinin M, Viitanen M, Parkkola R, Rinne JO (2006) Voxel-based analysis of PET amyloid ligand [¹¹C]PIB uptake in Alzheimer disease. *Neurology* **67**, 1575-1580.
- [66] Pike KE, Savage G, Villenaghe VL, Ng S, Moss SA, Maruff P, Mathis C, Klunk WE, Masters CL, Rowe CC (2007) Beta-amyloid imaging and memory in non-demented individuals: evidence for preclinical Alzheimer's disease. *Brain* **130**, 2837-2844.
- [67] Mintun MA, LaRossa GN, Sheline YIM, Dence CSM, Lee SYP, Mach RHP, Klunk WE, Mathis CA, DeKosky ST, Morris JC (2006) [¹¹C]PIB in a nondemented population: Potential antecedent marker of Alzheimer disease. *Neurology* **67**, 446-452.
- [68] Kemppainen NM, Aalto S, Wilson IA, Nagren K, Helin S, Bruck A, Oikonen V, Kailajärvi M, Scheinin M, Viitanen M, Parkkola R, Rinne JO (2007) PET amyloid ligand [¹¹C]PIB uptake is increased in mild cognitive impairment. *Neurology* **68**, 1603-1606.
- [69] Li Y, Rinne JO, Mosconi L, Tsui W, Pirraglia E, Rusinek H, De Santi S, Kemppainen N, Nagren K, Kim BC, de Leon MJ (2008) Regional analysis of FDG and PIB-PET images in normal aging, mild cognitive impairment and Alzheimer's disease. *Eur J Nucl Med Mol Imaging* **35**, 2169-2181.
- [70] Jack CR, Jr., Lowe VJ, Senjem ML, Weigand SD, Kemp BJ, Shiung MM, Knopman DS, Boeve BF, Klunk WE, Mathis CA, Petersen RC (2008) ¹¹C PiB and structural MRI provide complementary information in imaging of Alzheimer's disease and amnesic mild cognitive impairment. *Brain* **131**, 665-680.
- [71] Klunk WE, Price JC, Mathis CA, Tsopelas ND, Lopresti BJ, Ziolkowski SK, Bi W, Hoge JA, Cohen AD, Ikonomic MD, Saxton JA, Snitz BE, Pollen DA, Moonis M, Lippa CF, Swearer JM, Johnson KA, Rentz DM, Fischman AJ, Aizenstein HJ, DeKosky ST (2007) Amyloid deposition begins in the striatum of presenilin-1 mutation carriers from two unrelated pedigrees. *J Neurosci* **27**, 6174-6184.
- [72] Johansson A, Savitcheva I, Forsberg A, Engler H, Langstrom B, Nordberg A, Asmark H (2008) [¹¹C]-PIB imaging in patients with Parkinson's disease: preliminary results. *Parkinsonism Relat Disord* **14**, 345-347.
- [73] Johnson KA, Gregas M, Becker JA, Kinnecom C, Salat DH, Moran EK, Smith EE, Rosand J, Rentz DM, Klunk WE, Mathis CA, Price JC, Dekosky ST, Fischman AJ, Greenberg SM (2007) Imaging of amyloid burden and distribution in cerebral amyloid angiopathy. *Ann Neurol* **62**, 229-234.
- [74] Mathis CA, Wang Y, Klunk W (2004) Imaging [beta]-amyloid plaques and neurofibrillary tangles in the aging human brain. *Curr Pharm Design* **10**, 1469-1492.
- [75] Lockhart A, Lamb JR, Osredkar T, Sue LI, Joyce JN, Ye L, Libri V, Leppert D, Beach TG (2007) PIB is a non-specific imaging marker of amyloid-beta (A{beta}) peptide-related cerebral amyloidosis. *Brain* **130**, 2607-2615.
- [76] Crystal HA, Dickson D, Davies P, Masur D, Grober E, Lipton RB (2000) The relative frequency of "dementia of unknown etiology" increases with age and is nearly 50% in nonagenarians. *Arch Neurol* **57**, 713-719.
- [77] Crystal H, Dickson D, Fuld P, Masur D, Scott R, Mehler M, Masdue J, Kawas C, Aronson M, Wolfson L (1988) Clinicopathologic studies in dementia: nondemented subjects with pathologically confirmed Alzheimer's disease. *Neurology* **38**, 1682-1687.
- [78] Petersen RC, Doody R, Kurz A, Mohs RC, Morris JC, Rabins PV, Ritchie K, Rossor M, Thal L, Winblad B (2001) Current concepts in mild cognitive impairment. *Arch Neurol* **58**, 1985-1992.
- [79] Blass JP (2002) Alzheimer's disease and Alzheimer's dementia: distinct but overlapping entities. *Neurobiol Aging* **23**, 1077-1084.
- [80] Forsberg A, Engler H, Almkvist O, Blomqvist G, Hagman G, Wall A, Ringheim A, Långström B, Nordberg A (2008) PET imaging of amyloid deposition in patients with mild cognitive impairment. *Neurobiol Aging* **29**, 1456-1465.
- [81] Okello A, Koivunen J, Edison P, Archer HA, Turkheimer FE, Nagren K, Bullock R, Walker Z, Kennedy A, Fox NC, Rossor MN, Rinne JO, Brooks DJ (2009) Conversion of amyloid positive and negative MCI to AD over 3 years. An ¹¹C-PIB PET study. *Neurology* **73**, 754-760.
- [82] Wolk DA, Price JC, Saxton JA, Snitz BE, James JA, Lopez OL, Aizenstein HJ, Cohen AD, Weissfeld LA, Mathis CA, Klunk WE, DeKosky ST (2009) Amyloid imaging in mild cognitive impairment subtypes. *Ann Neurol* **65**, 557-568.
- [83] Reiman EM, Chen K, Liu X, Bandy D, Yu M, Lee W, Ayutyanont N, Keppler J, Reeder SA, Langbaum JB, Alexander GE, Klunk WE, Mathis CA, Price JC, Aizenstein HJ, DeKosky ST, Caselli RJ (2009) Fibrillar amyloid-beta burden in cognitively normal people at 3 levels of genetic risk

- for Alzheimer's disease. *Proc Natl Acad Sci U S A* **106**, 6820-6825.
- [84] Jagust WJ (2009) Mapping brain beta-amyloid. *Curr Opin Neurol* **22**, 356-361.
- [85] Mesulam MM (1999) Neuroplasticity failure in Alzheimer's disease: bridging the gap between plaques and tangles. *Neuron* **24**, 521-529.
- [86] Engler H, Forsberg A, Almkvist O, Blomquist G, Larsson E, Savitcheva I, Wall A, Ringheim A, Långström B, Nordberg A (2006) Two-year follow-up of amyloid deposition in patients with Alzheimer's disease. *Brain* **129**, 2856-2866.
- [87] Jack CR Jr, Lowe VJ, Weigand SD, Wiste HJ, Senjem ML, Knopman DS, Shiung MM, Gunter JL, Boeve BF, Kemp BJ, Weiner M, Petersen RC; Alzheimer's Disease Neuroimaging Initiative (2009) Serial PIB and MRI in normal, mild cognitive impairment and Alzheimer's disease: implications for sequence of pathological events in Alzheimer's disease. *Brain* **132**, 1355-1365.
- [88] Klunk WE, Mathis CA, Price JC, Lopresti BJ, DeKosky ST (2006) Two-year follow-up of amyloid deposition in patients with Alzheimer's disease. *Brain* **129**, 2805-2807.
- [89] Small GW, Kepe V, Ercoli LM, Siddarth P, Bookheimer SY, Miller KJ, Lavretsky H, Burggren AC, Cole GM, Vinters HV, Thompson PM, Huang SC, Satyamurthy N, Phelps ME, Barrio JR (2006). PET of brain amyloid and tau in mild cognitive impairment. *N Engl J Med* **355**, 2652-2663.
- [90] Price JL, Ko AI, Wade MJ, Tsou SK, McKeel DW, Morris JC (2001) Neuron number in the entorhinal cortex and CA1 in preclinical Alzheimer disease. *Arch Neurol* **58**, 1395-1402.
- [91] Masters CL, Simms G, Weinman NA, Multhaup G, McDonald B, Beyreuther K (1985) Amyloid plaque core protein in Alzheimer disease and Down syndrome. *Proc Natl Acad Sci U S A* **82**, 4245-4249.
- [92] Isacson O, Seo H, Lin L, Albeck D, Granholm AC (2002) Alzheimer's disease and Down's syndrome: roles of APP, trophic factors and ACh. *Trends Neurosci* **25**, 79-84.
- [93] Price JL, Morris JC (1999) Tangles and plaques in nondemented aging and "preclinical" Alzheimer's disease. *Ann Neurol* **45**, 358-368.
- [94] Davies L, Wolska B, Hilbich C, Multhaup G, Martins R, Simms G, Bayreuther K, Masters CL (1988) A4 amyloid protein deposition and the diagnosis of Alzheimer's disease: prevalence in aged brains determined by immunocytochemistry compared with conventional neuropathologic techniques. *Neurology* **38**, 1688-1693.
- [95] Shoghi-Jadid K, Barrio JR, Kepe V, Huang SC (2006) Exploring a mathematical model for the kinetics of beta-amyloid molecular imaging probes through a critical analysis of plaque pathology. *Mol Imaging Biol* **8**, 151-162.
- [96] Price JC, Klunk WE, Lopresti BJ, Lu X, Hoge JA, Ziolk SK, Holt DP, Meltzer CC, DeKosky ST, Mathis CA (2005) Kinetic modeling of amyloid binding in humans using PET imaging and Pittsburgh Compound-B. *J Cereb Blood Flow Metab* **25**, 1528-1547.
- [97] Lopresti BJ, Klunk WE, Mathis CA, Hoge JA, Ziolk SK, Lu X, Meltzer CC, Schimmel K, Tsopelas ND, DeKosky ST, Price JC (2005) Simplified quantification of Pittsburgh Compound B amyloid imaging PET Studies: a comparative analysis. *J Nucl Med* **46**, 1959-1972.
- [98] Logan J, Fowler JS, Volkow ND, Wang GJ, Ding YS, Alexoff DL (1996) Distribution volume ratios without blood sampling from graphical analysis of PET data. *J Cereb Blood Flow Metab* **16**, 834-840.
- [99] Shoghi-Jadid K, Barrio JR, Kepe V, Wu HM, Small GW, Phelps ME, Huang SC (2005) Imaging beta-amyloid fibrils in Alzheimer's disease: a critical analysis through simulation of amyloid fibril polymerization. *Nucl Med Biol* **32**, 337-351.
- [100] Haass C, Selkoe D (2007) Soluble protein oligomers in neurodegeneration: lessons from the Alzheimer's amyloid beta-peptide. *Nat Rev Mol Cell Biol* **8**, 101-112.
- [101] Hardy J, Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* **297**, 353-356.
- [102] Kempainen NM, Aalto S, Karrasch M, Nagren K, Savisto N, Oikonen V, Vitanen M, Parkkola R, Rinne JO. (2008) Cognitive reserve hypothesis: Pittsburgh Compound B and fluorodeoxyglucose positron emission tomography in relation to education in mild Alzheimer's disease. *Arch Neurol* **63**, 112-118.
- [103] Roe CM, Mintun MA, D'Angelo D, Xiong C, Grant EA, Morris JC (2008) Alzheimer disease and cognitive reserve: variation of education effect with carbon 11-labeled Pittsburgh Compound B uptake. *Arch Neurol* **65**, 1467-1471.
- [104] Powell MR, Smith GE, Knopman DS, Parisi JE, Boeve BF, Petersen RC, Ivnik RJ (2006) Cognitive measures predict pathologic Alzheimer disease. *Arch Neurol* **63**, 865-868.
- [105] Koivunen J, Verkkoniemi S, Aalto S, Paetau A, Ahonen JP, Viitanen M, Rinne JO (2008). PET amyloid ligand [¹¹C]PIB uptake shows predominantly striatal increase in variant Alzheimer's disease. *Brain* **131**, 1845-1853.
- [106] Remes AM, Laru L, Tuominen H, Aalto S, Kempainen N, Mononen H, Nagren K, Parkkola R, Rinne JO (2008) Carbon 11-labeled Pittsburgh Compound B positron emission tomographic amyloid imaging in patients with APP locus duplication. *Arch Neurol* **65**, 540-544.
- [107] Sestini S, Castagnoli A, Mansi L (2010) The new FDG brain revolution: the neurovascular unit and the default network. *Eur J Nucl Med Molec Imaging* **37**, 913-916.