

# Microbial communities as resilient systems: from one to one billion

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July 28, 2016  
MBL Microbial Diversity

# *Resilience*

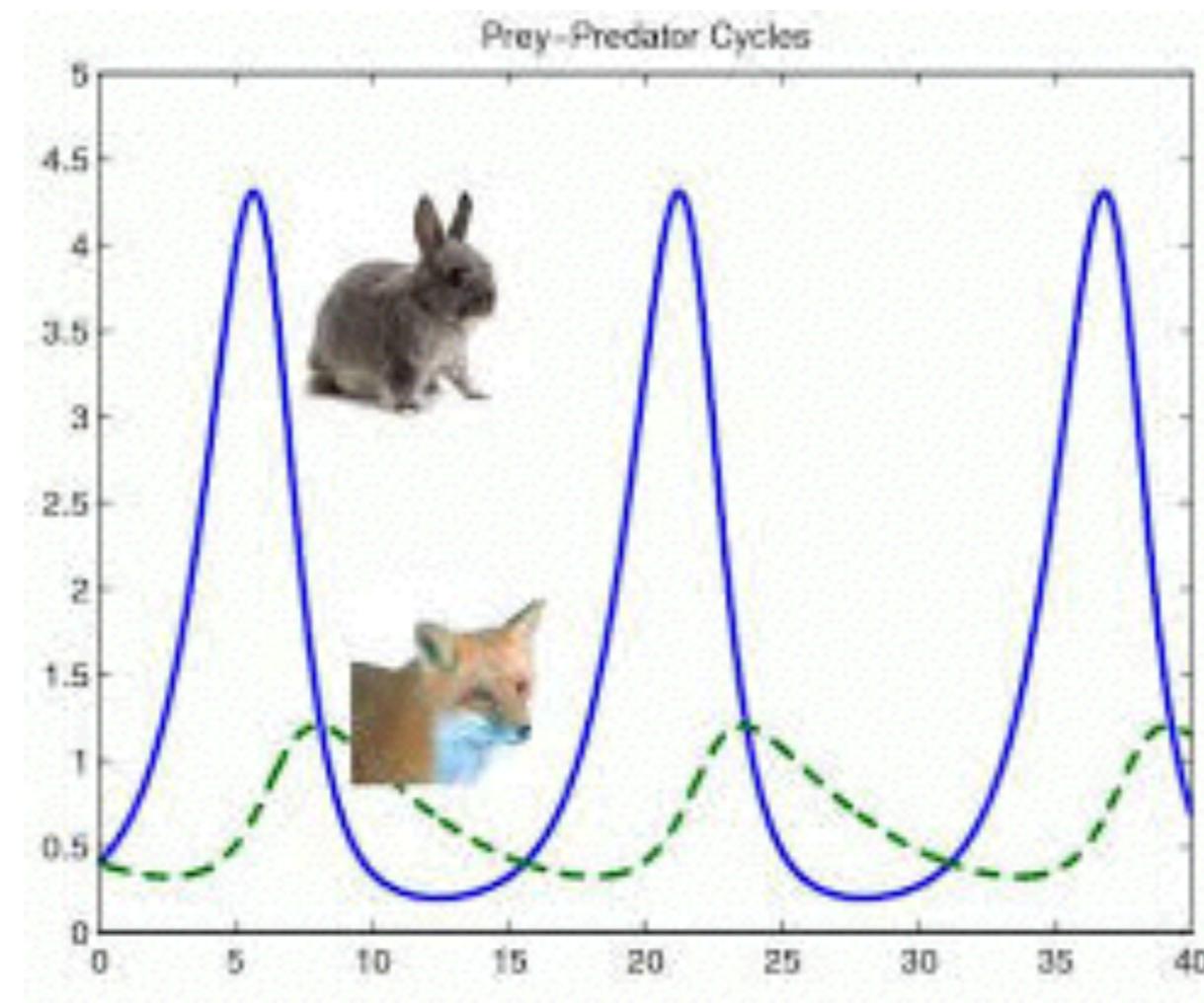
1. Interacting parts make up a system.
2. The system has some adaptive capacity.

The system has the ability to withstand, recover, or reorganize in response to environmental perturbations.

## Resilient communities

Community is maintained but the system structure may not be

# Lotka-Volterra interspecific competition (1932)



# Communities of microbes & their interactions

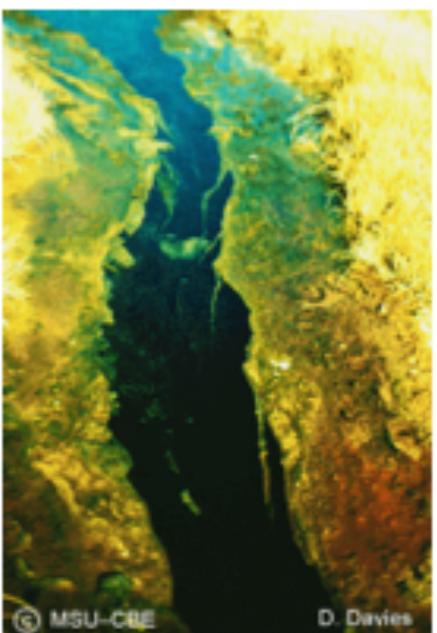
Putting the informatics back into bio

Single-species *Shewanella oneidensis* biofilm

The role of SoxR in response to phenazines

Microbial communities and greenhouse gas fluxes in response to agricultural land management

# Bacteria live in communities called biofilms



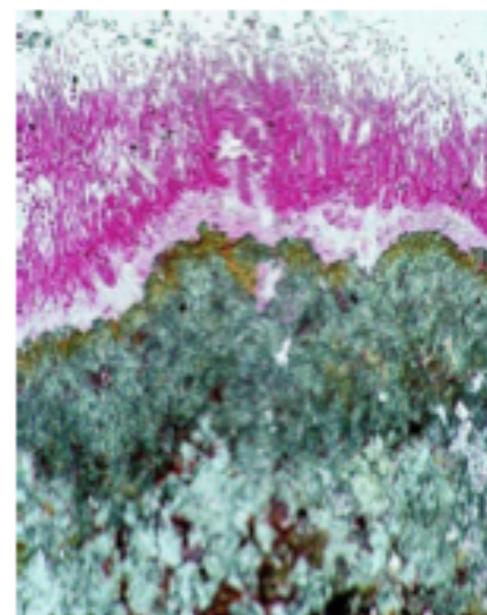
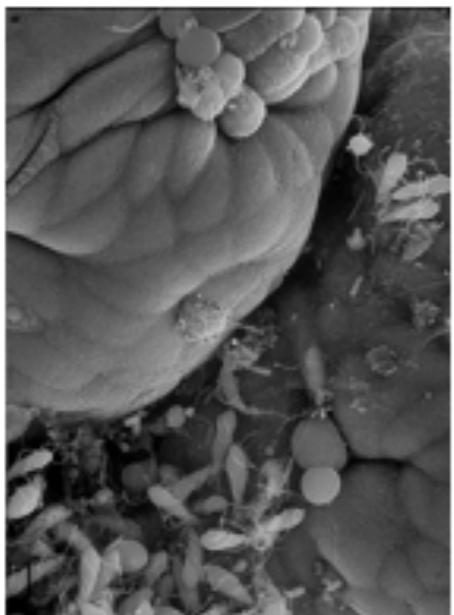
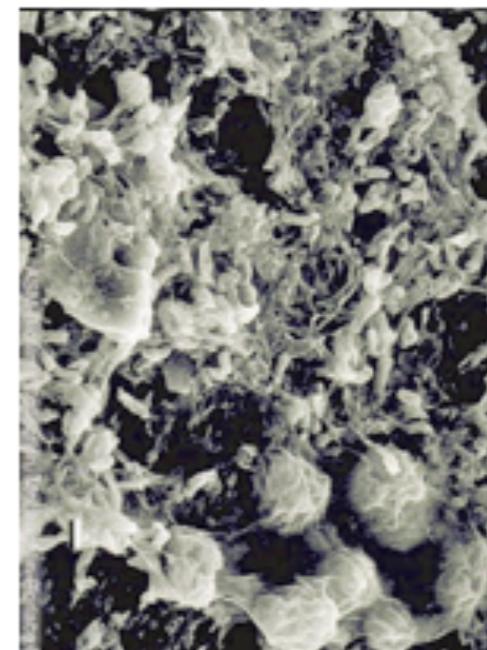
© MSU-CBE

D. Davies



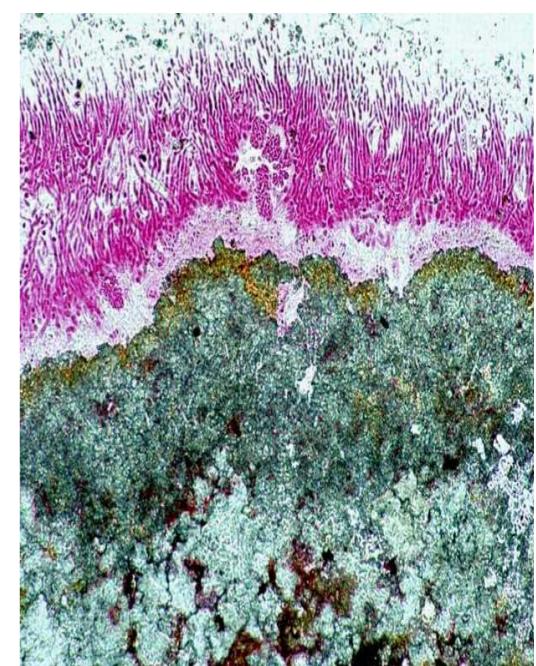
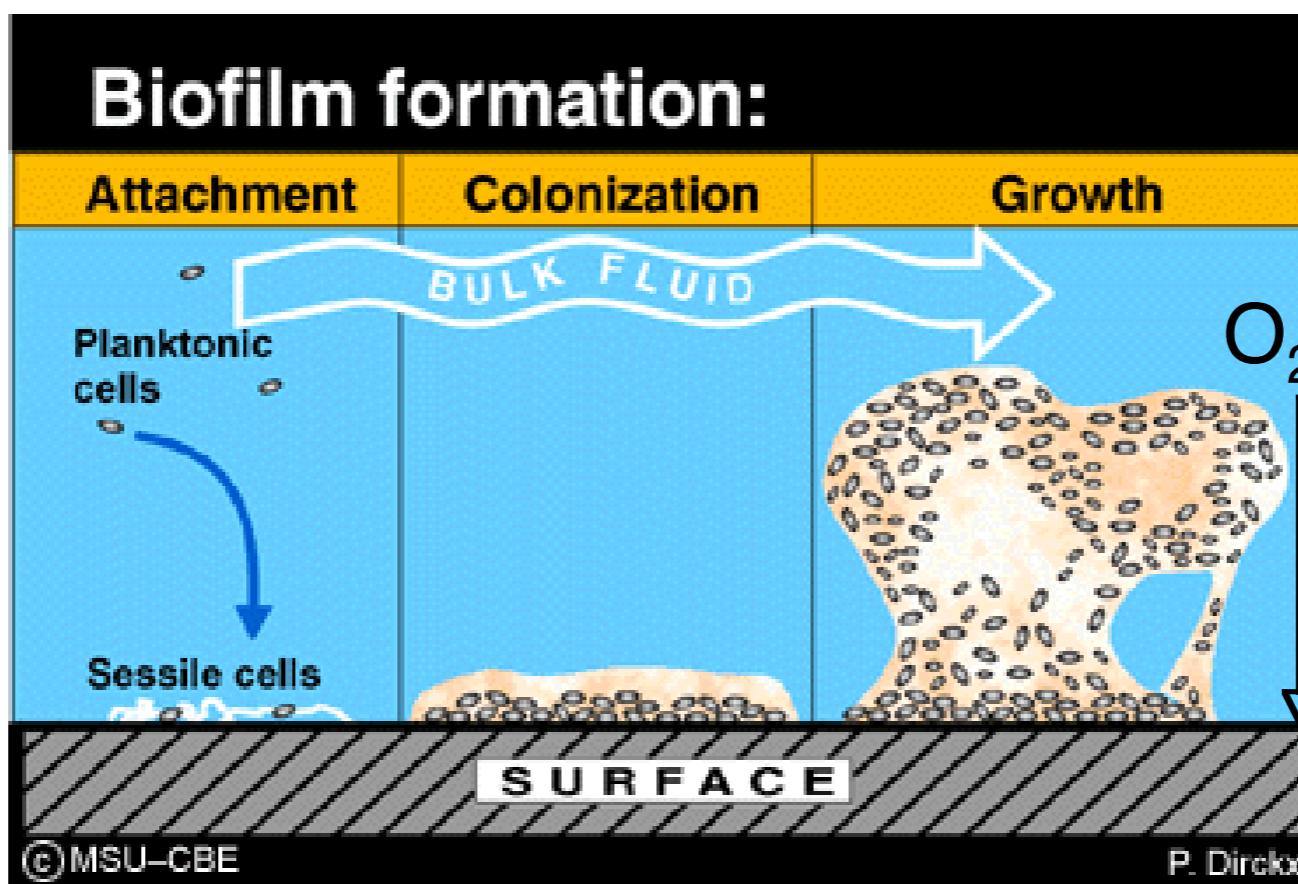
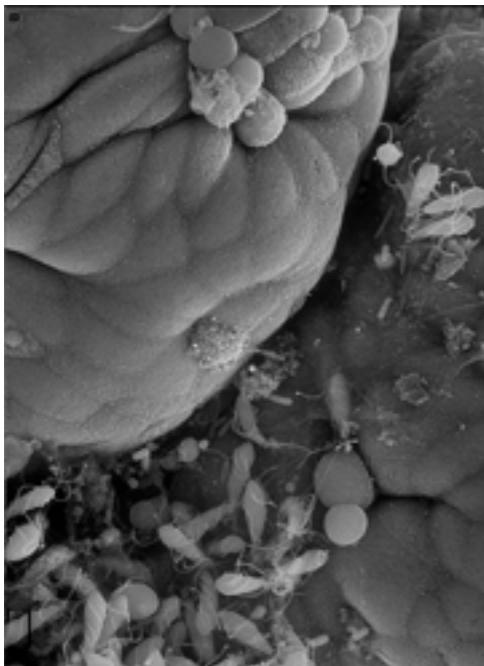
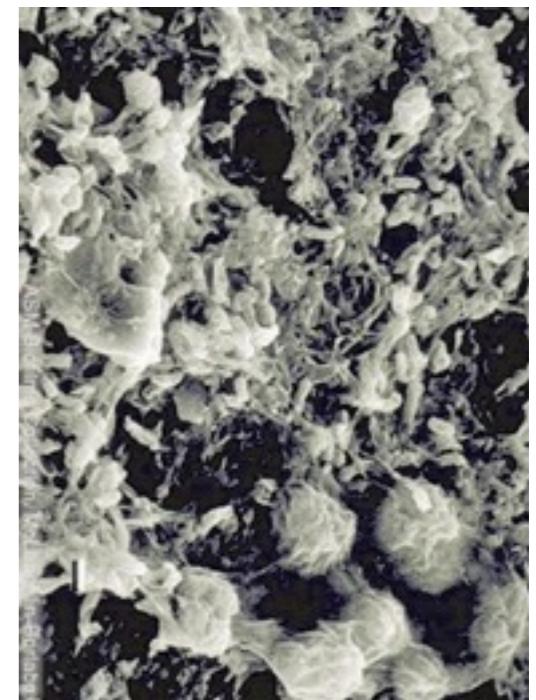
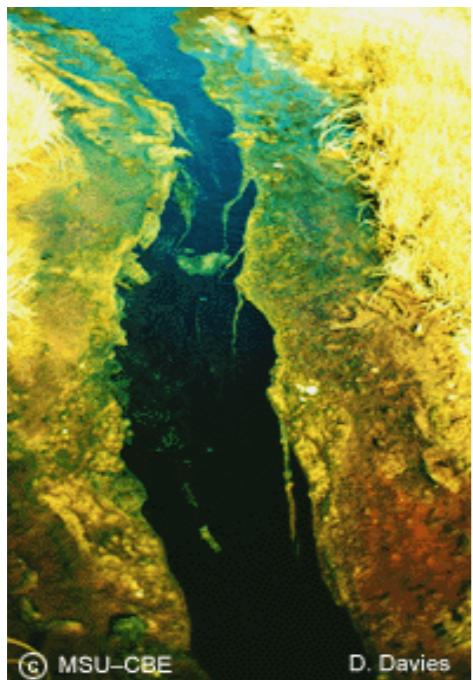
JD Ruby & VF Gerencser

Courtesy, ASM

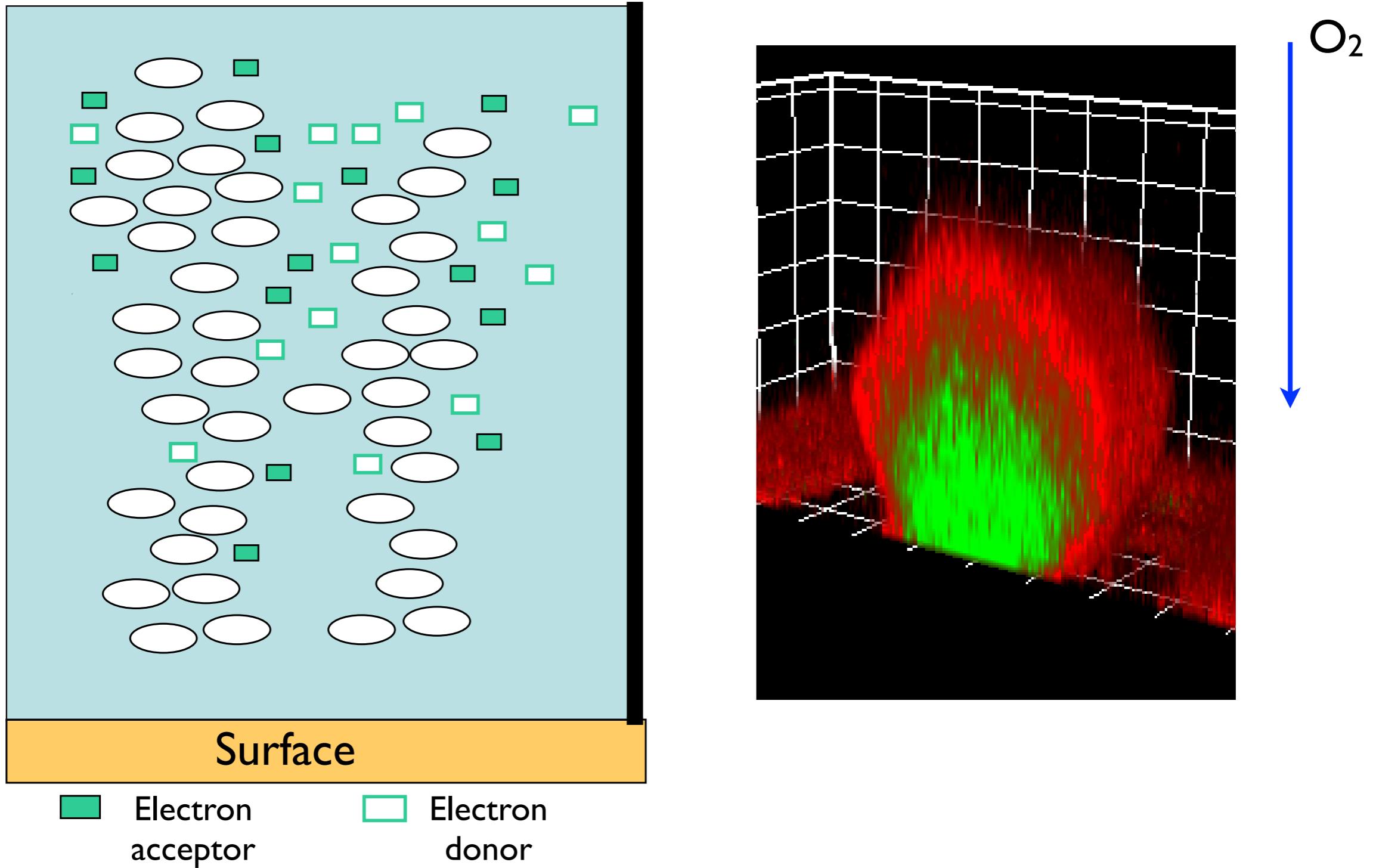


# Biofilms

## Spatially stratified microbial communities



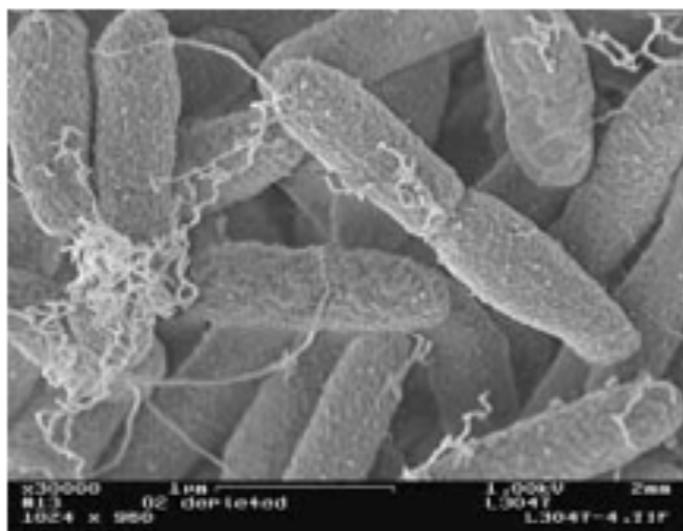
# Heterogeneity within biofilms



# Spatial organization in biofilms

- What is the spatial and temporal stratification of growth and metabolic states within a biofilm?
- Does availability of nutrients affect growth and metabolic stratification and can this stratification respond and change dynamically?

# *Shewanella oneidensis* MR-I



- Facultative anaerobe
- Versatile respiration
- Forms biofilms
- Metabolism well-characterized
- Genome sequenced
- Manipulated genetically

Image: PNNL

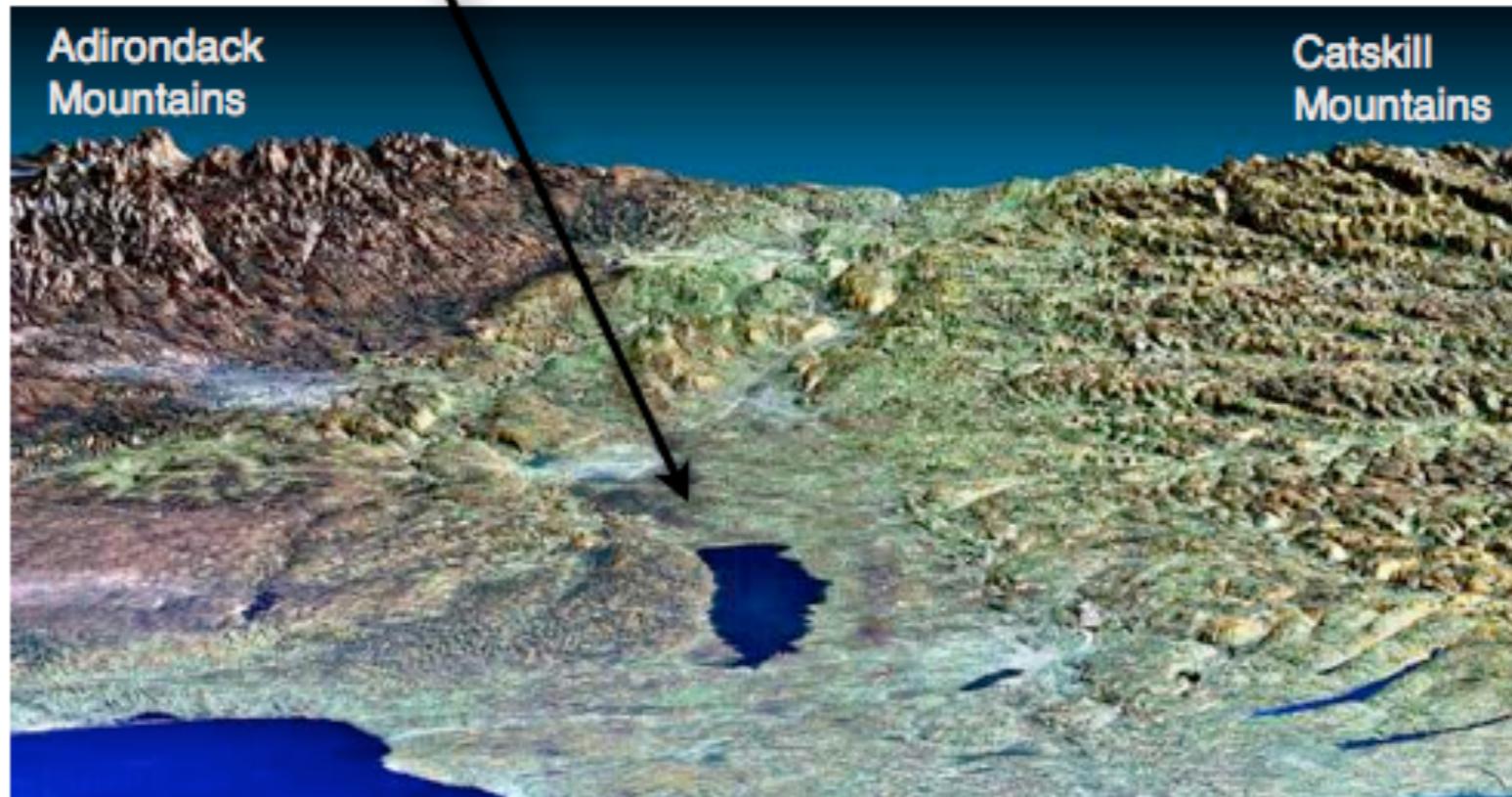
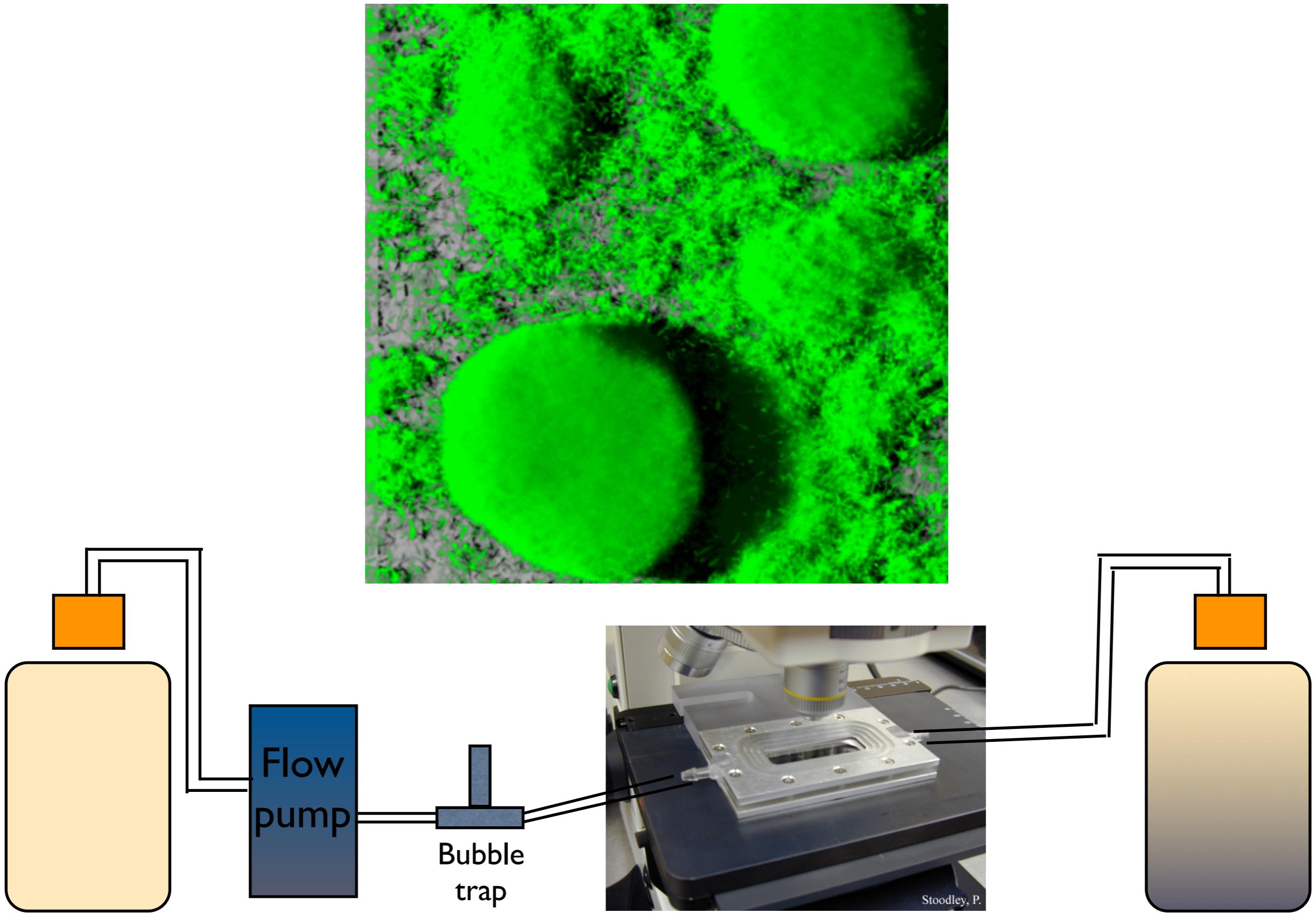
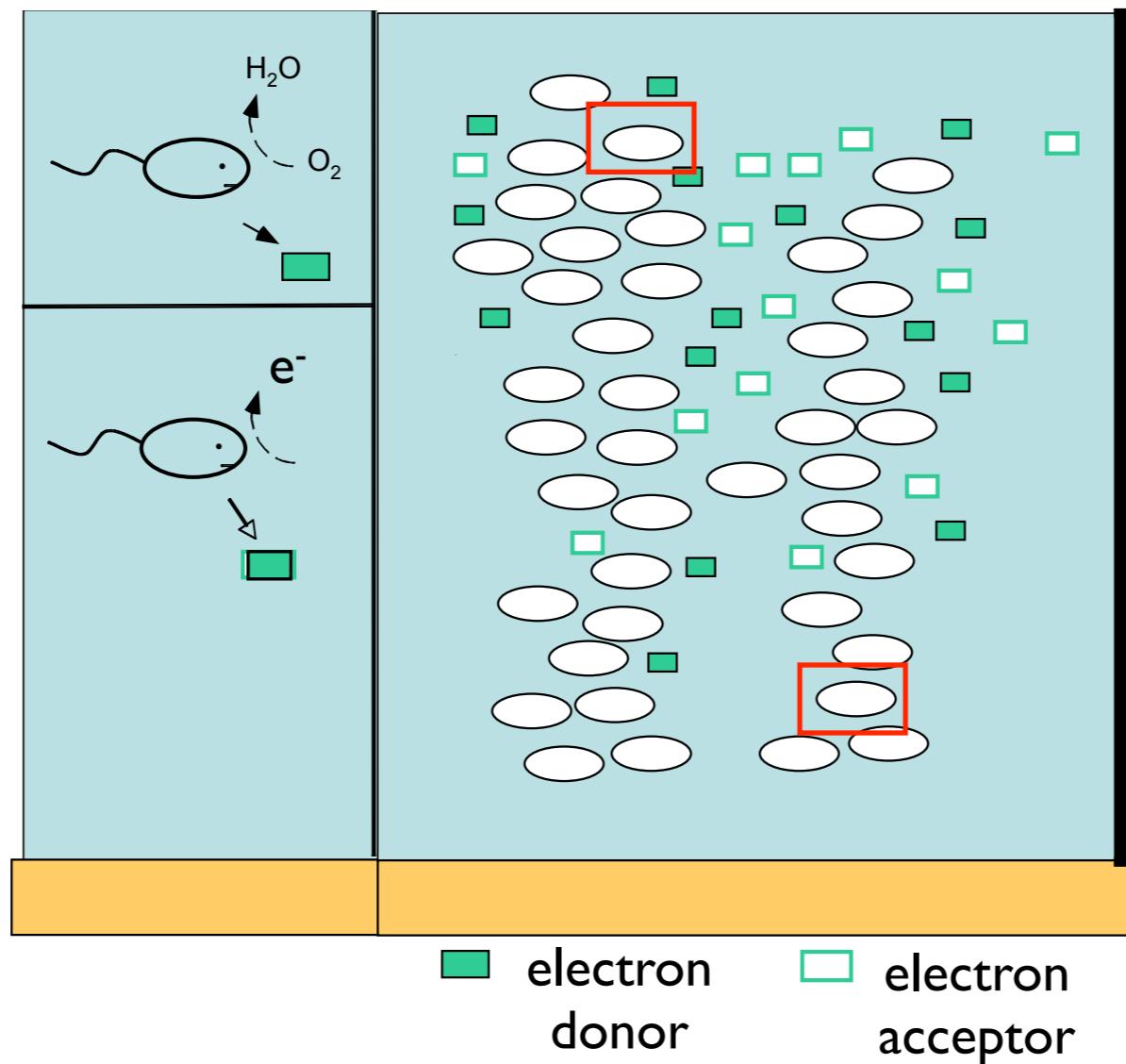


Photo: NASA

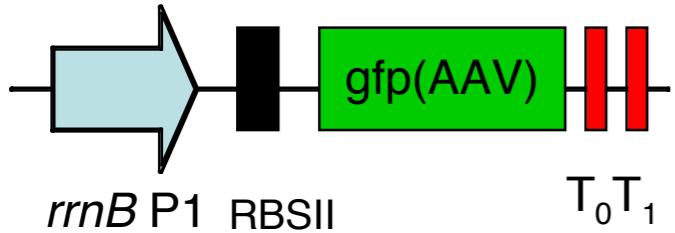
# Experimental setup



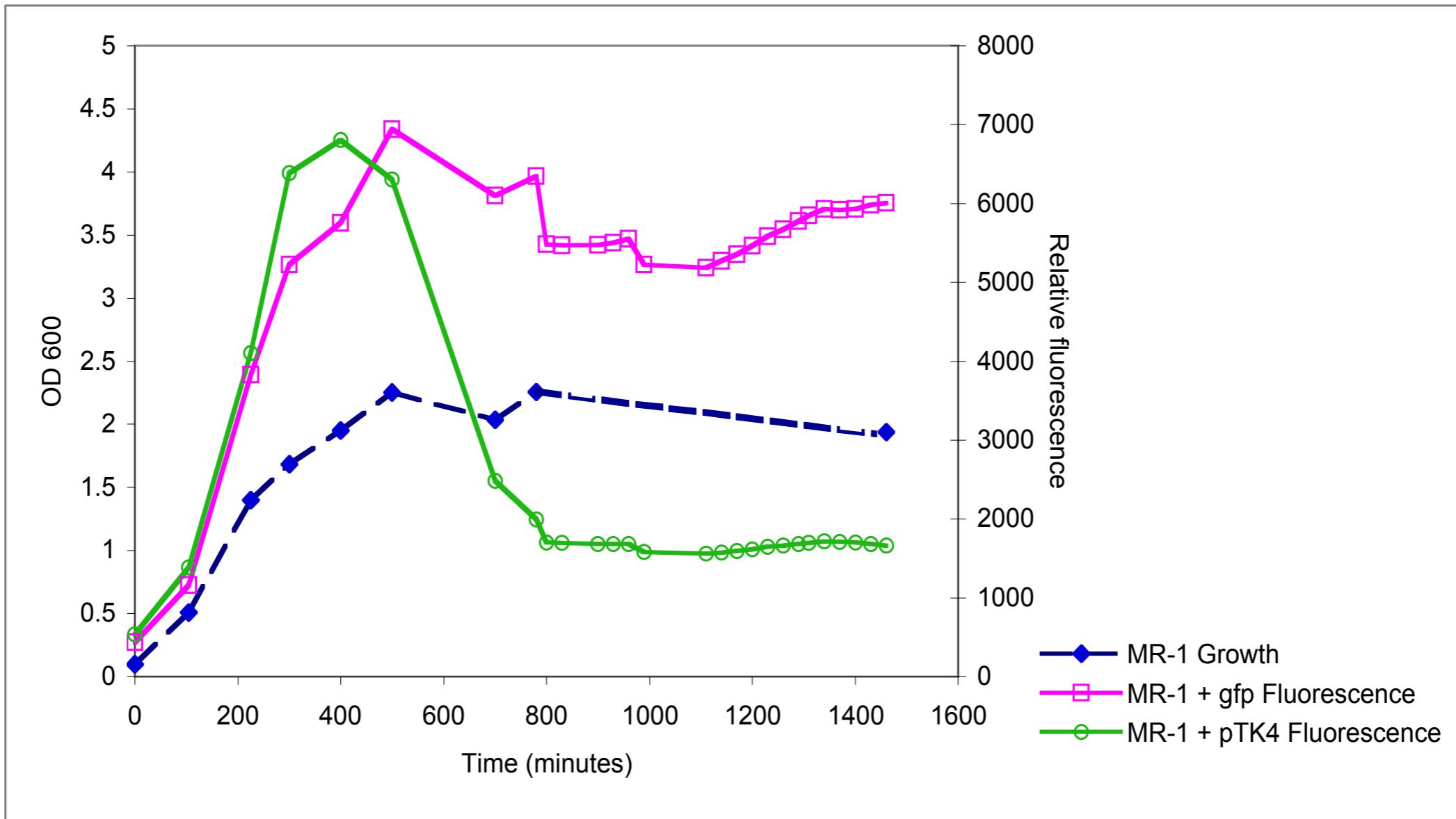
# What is the growth state of cells in a biofilm?



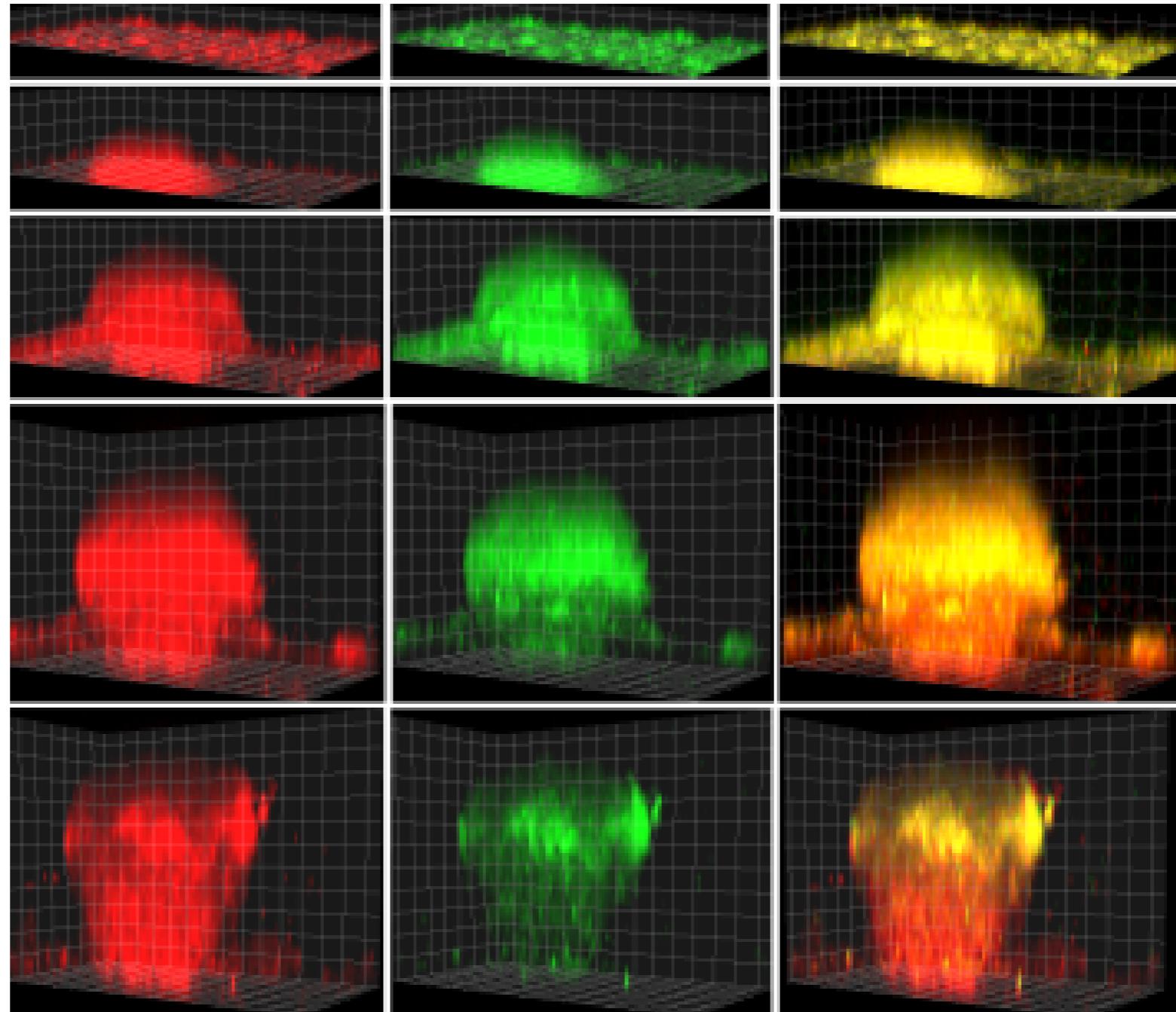
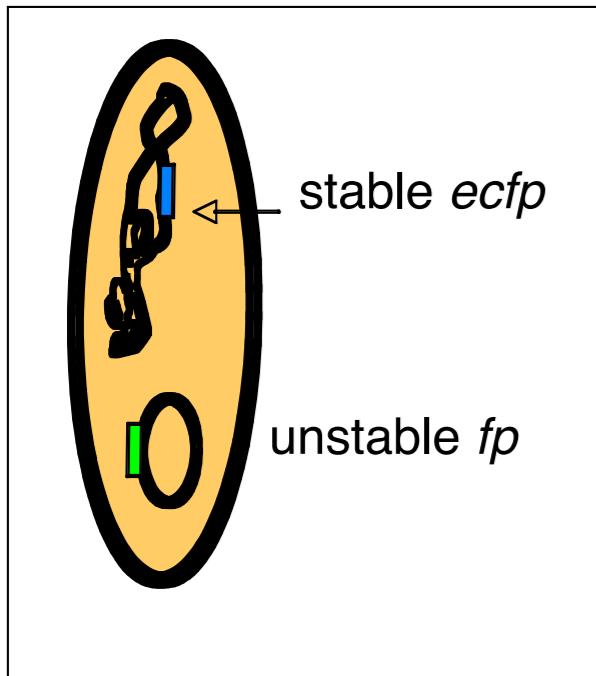
# *In vivo* reporter of growth activity



A reporter system for ribosomal RNA expression is correlated with growth activity



# Cells have variable growth state at different regions of the biofilm



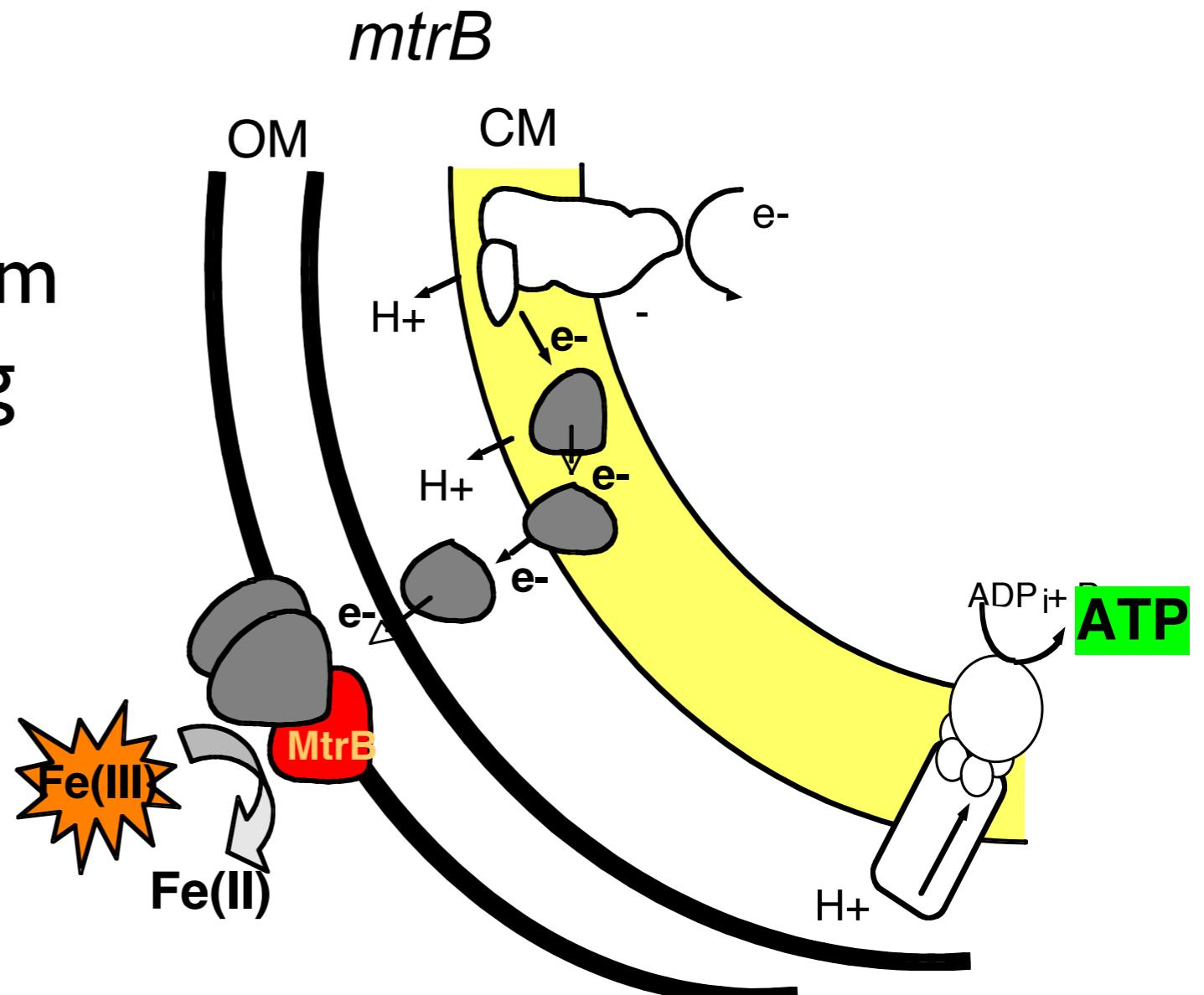
All cells

Growth

Overlay

# Are cells responding to local microenvironments?

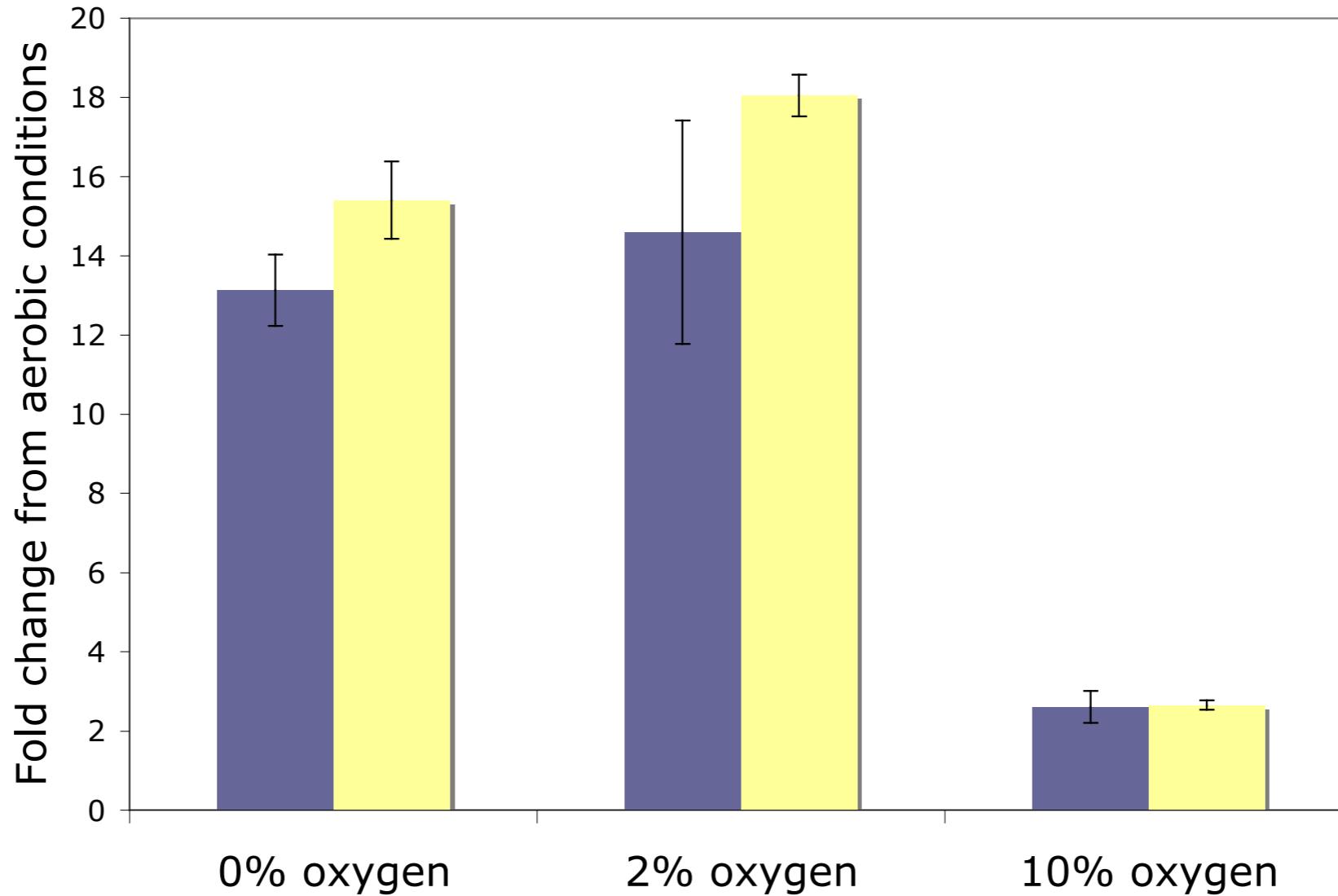
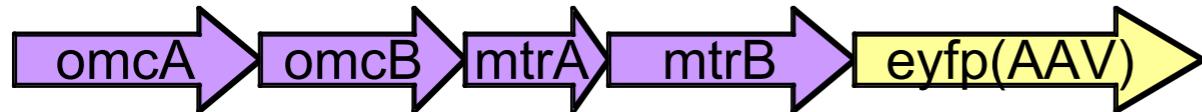
A reporter for metabolism linked to oxygen sensing



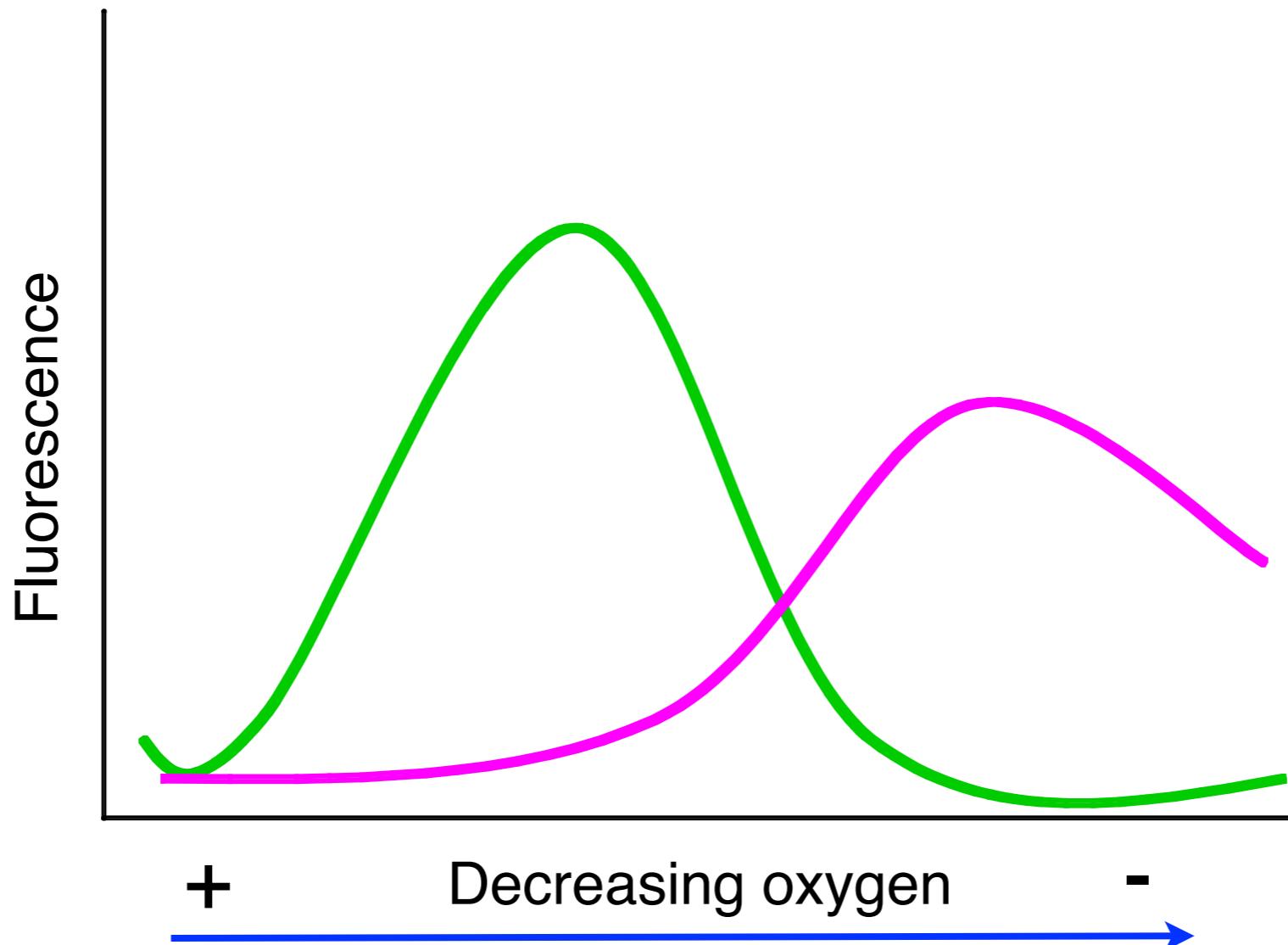
*mtrB* expression implies 2 things:

- 1  $O_2$  status within a biofilm
- 2 Active metabolism under low  $O_2$  conditions

# YFP is expressed from *mtrB* under anaerobic conditions



# Expectations for expression under different oxygen conditions

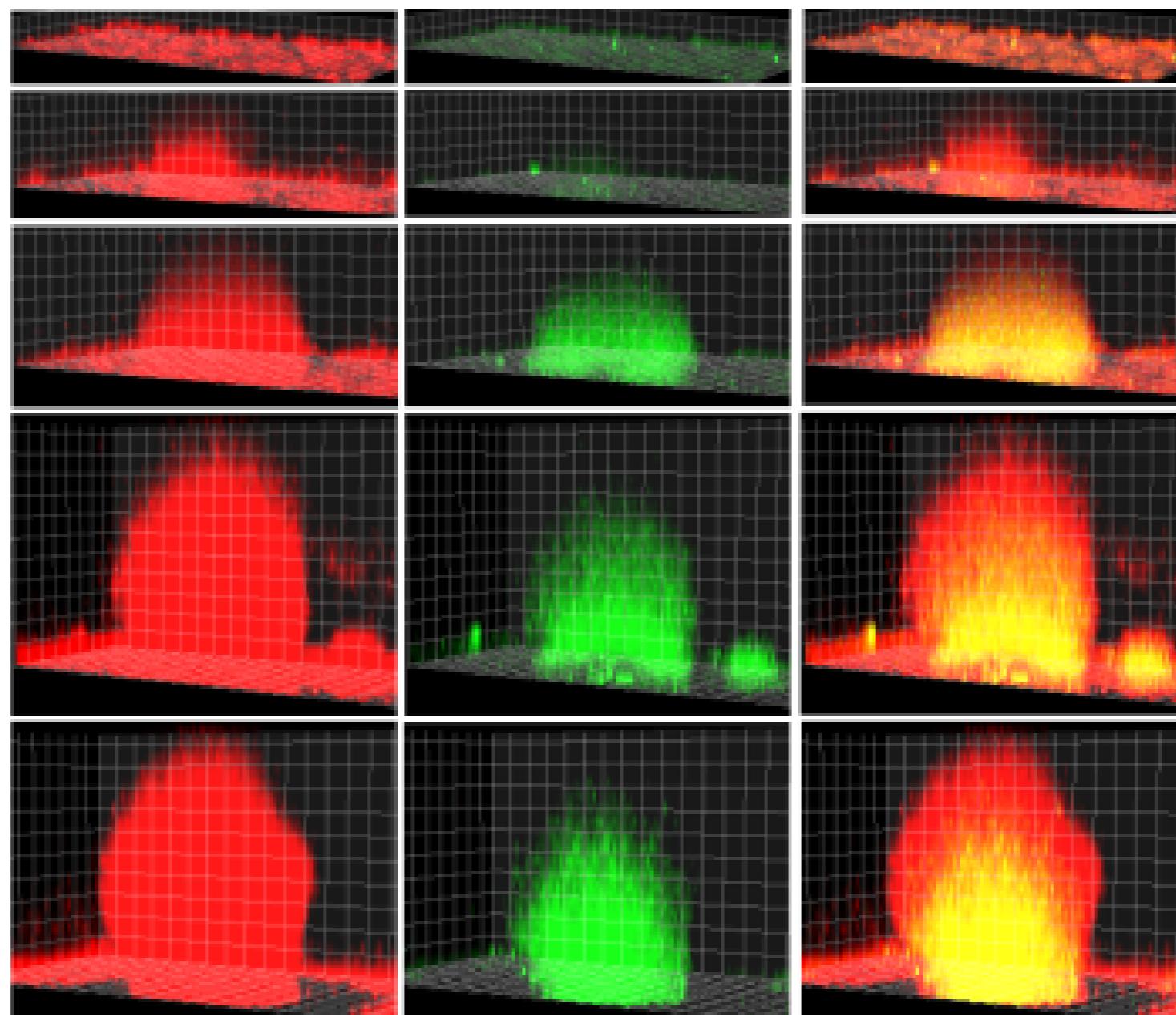


*rrnB P1* (growth activity)

*mtrB* (anaerobic metabolism)



# Distribution of *mtrB* expression indicating response that changes with oxygen conditions

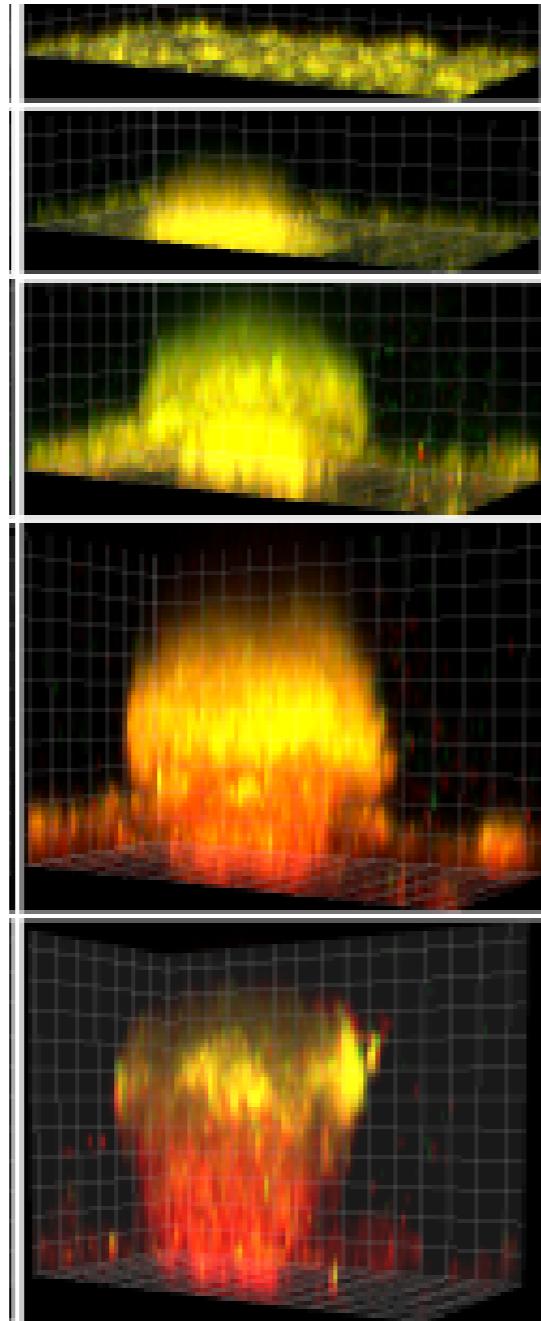


All cells

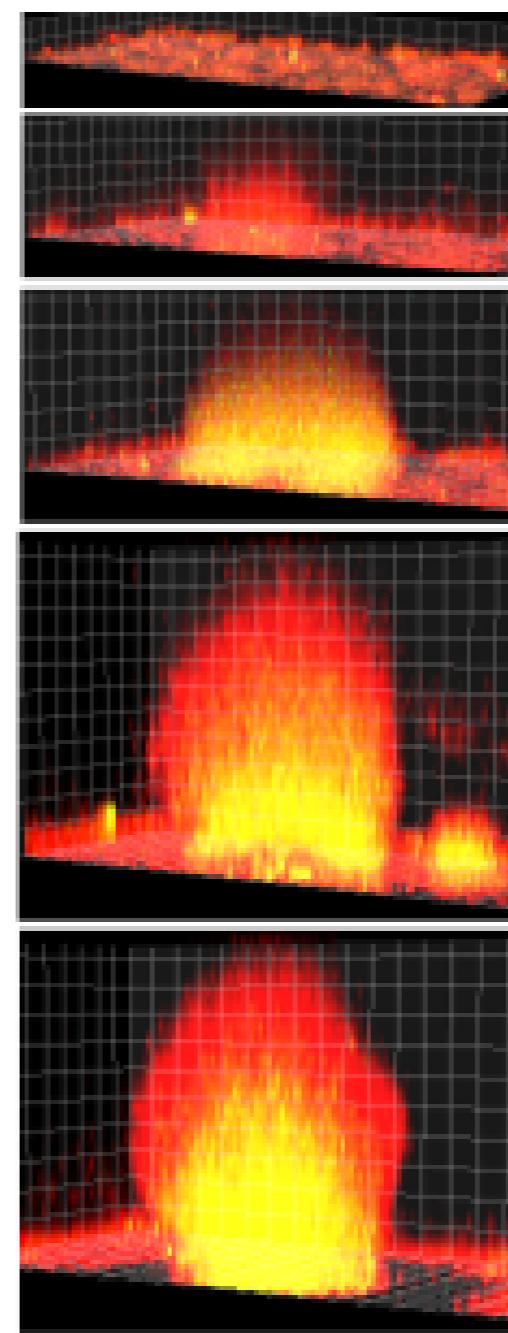
*mtrB*

Overlay

# Cells have variable growth state and metabolism at different regions of the biofilm



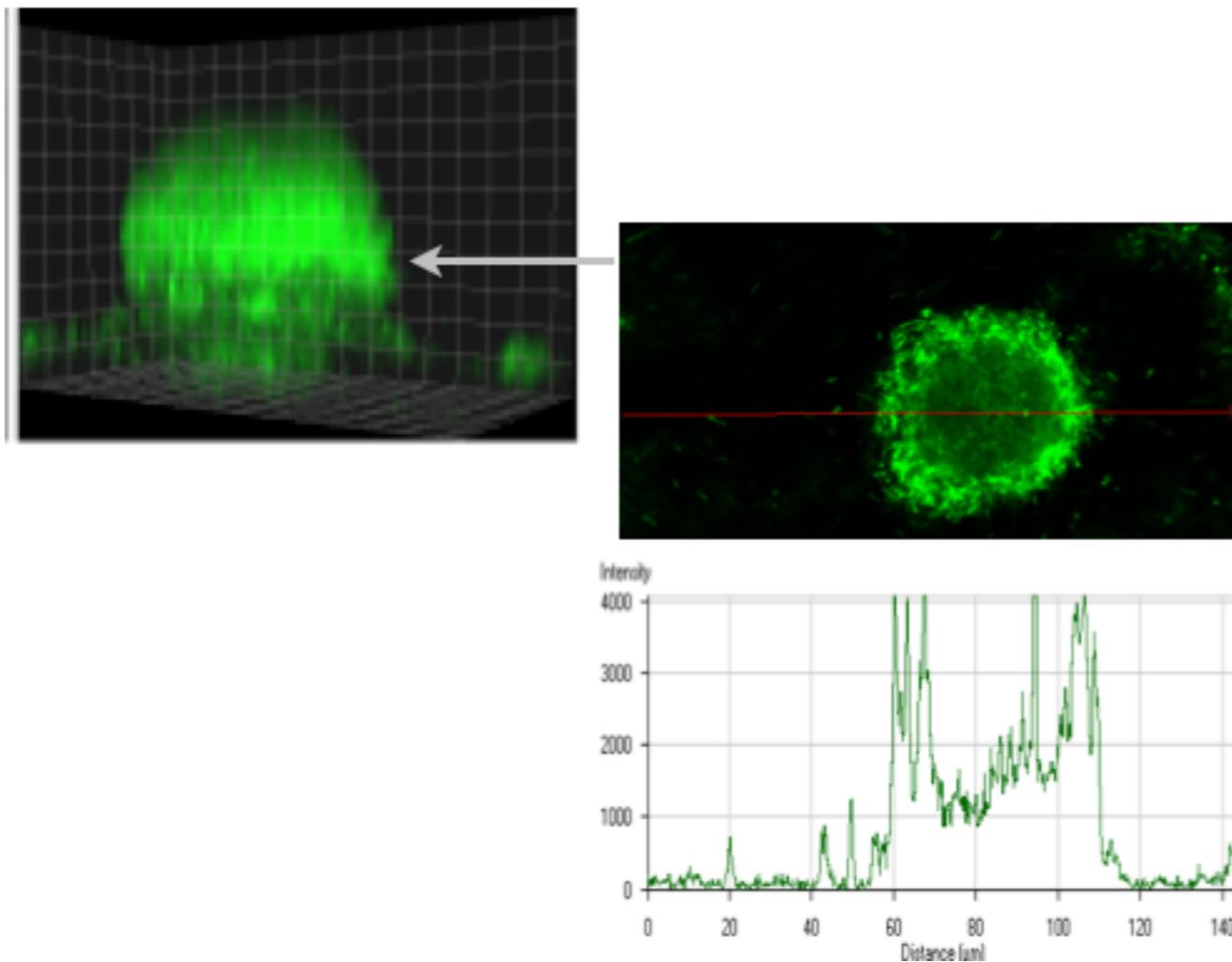
Growth activity



Oxygen metabolism

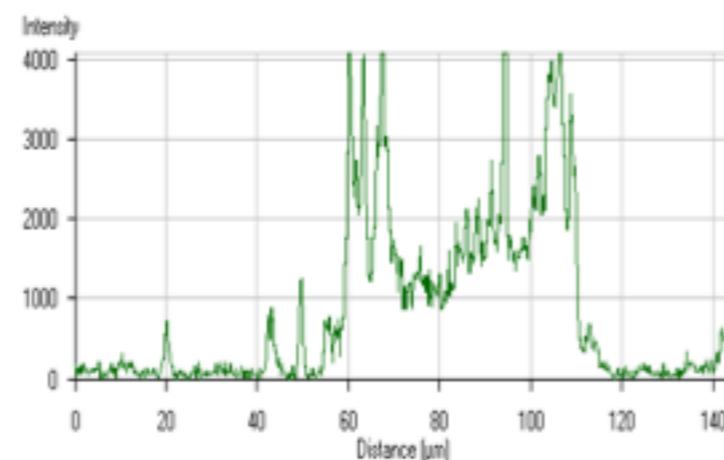
# Quantitative analysis of fluorescence distribution and intensity

*"When you employ the microscope, shake off all prejudice, nor harbor any favorite opinions; for, if you do, 'tis not unlikely fancy will betray you into error, and make you see what you wish to see."* - The Microscope Made Easy, Henry Baker, "Cautions in Viewing Objects", 1742



- extract fluorescent intensity profile
- average 9-pixel bins
- plot fluorescence intensity versus distance from center
- bin structures into representative sizes and average

# Quantitative analysis of fluorescence distribution and intensity

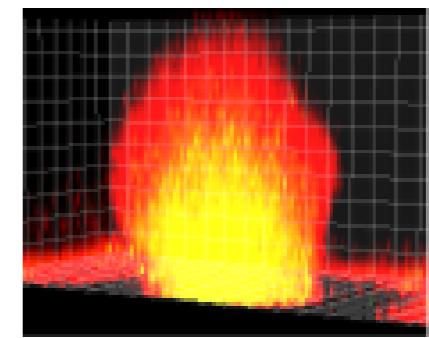
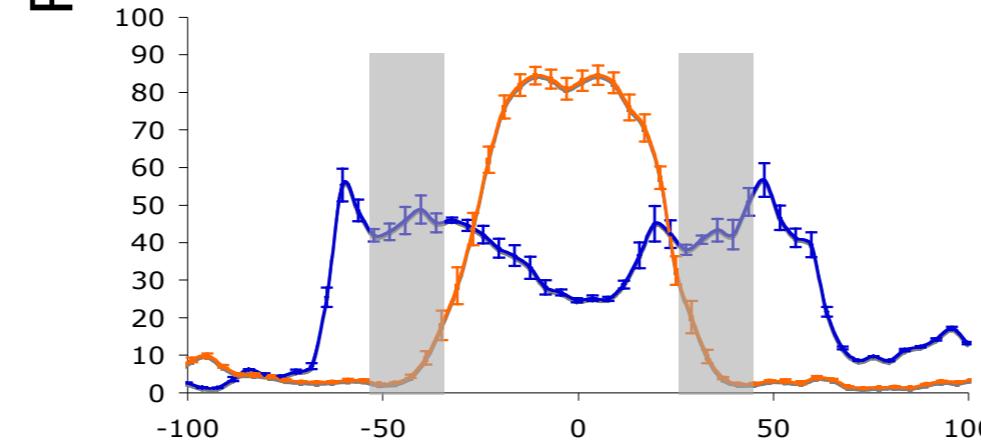
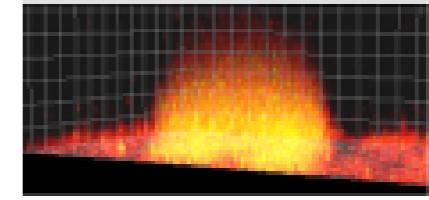
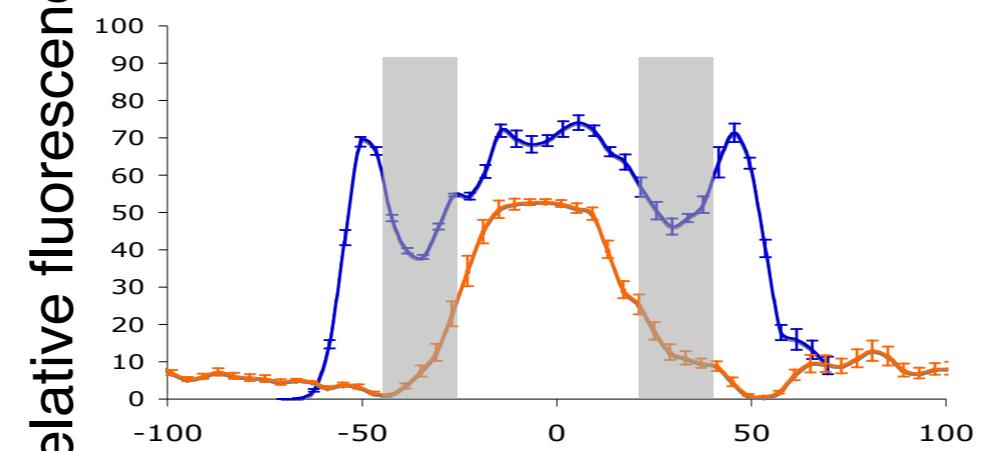
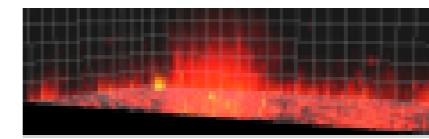
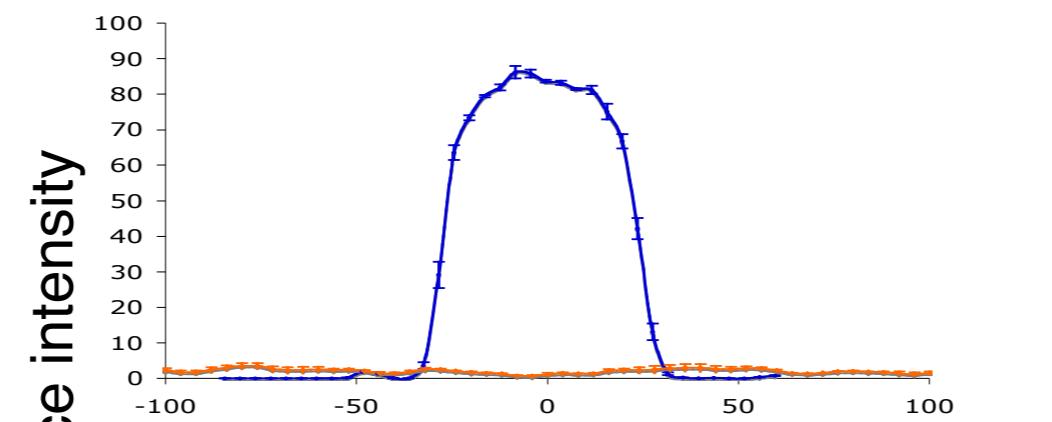
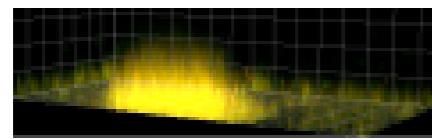


Intensity  
plot binned

Intensity  
plot distance  
from center

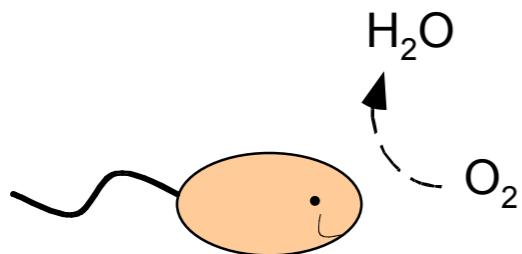
Intensity  
plot distance  
from center  
averaged

# Patterns of growth and activity correlate with colony size



— *rrnB* P1 — *mtrB* □ EPS

# Does oxygen availability affect biofilm stratification?



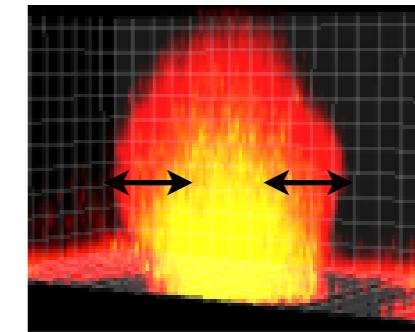
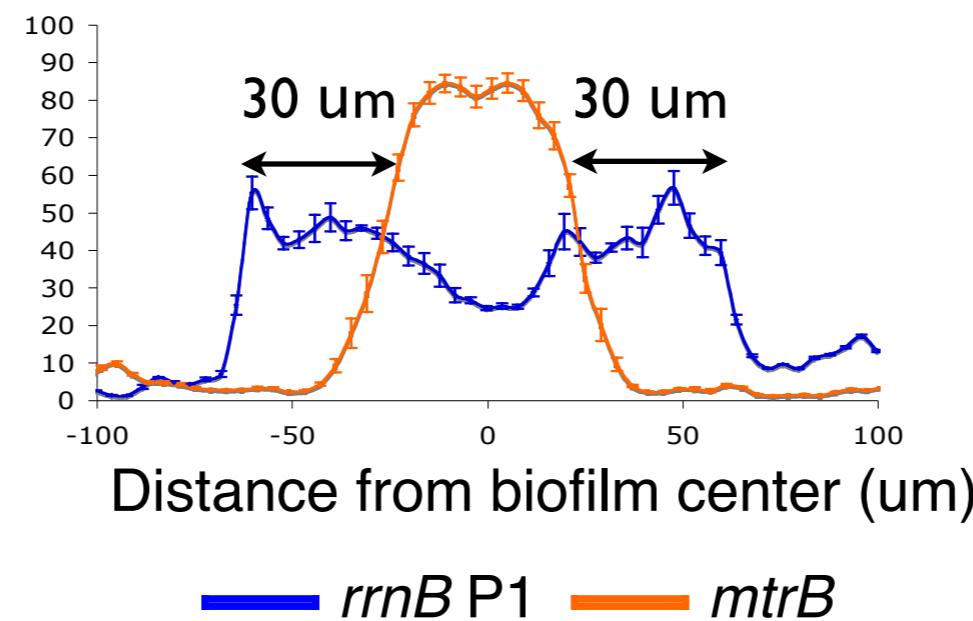
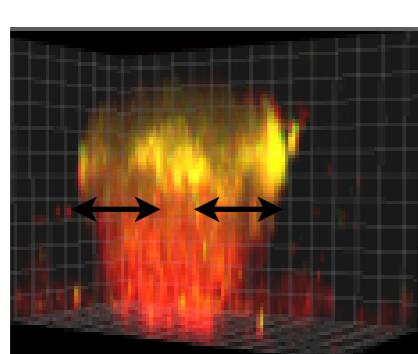
Rate of  $O_2$  consumption by  
*Shewanella* under stationary phase  
conditions =  $5.53\text{mg O}_2/\text{L*s}$

Use a reaction/diffusion equation  $R_{\min} = (6*D_e*S_0/K_0)^{1/2}$

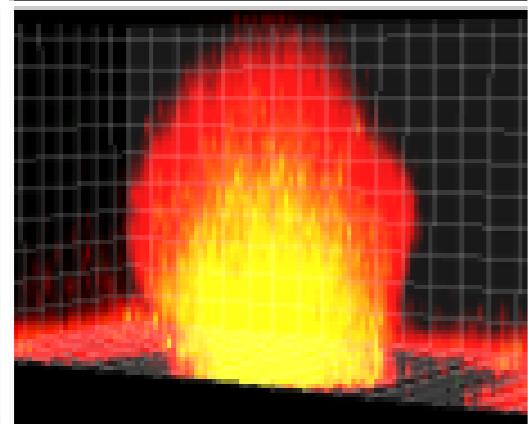
Predict that oxygen will diffuse  
 $\sim 30\text{um}$  into the biofilm

# Oxygen diffusion correlates with growth activity

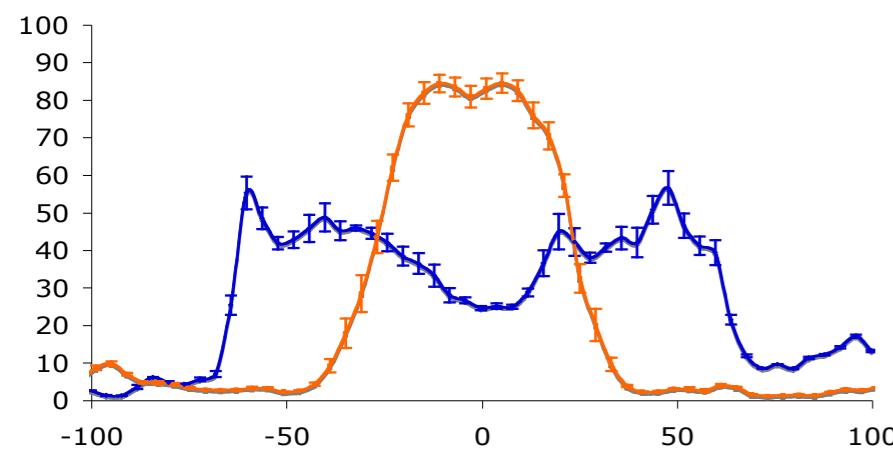
Predict that oxygen will diffuse ~30um into the biofilm



# Summary



Cell growth and metabolism is heterogeneous within the biofilm. Growth can be decoupled from metabolism.



Quantitative profiles show that patterns of growth and metabolism are consistent and predictable based on structure size and oxygen concentrations

Cells are dynamically regulating their response to different environmental conditions

**SoxR**

# SoxR responses differ between bacteria

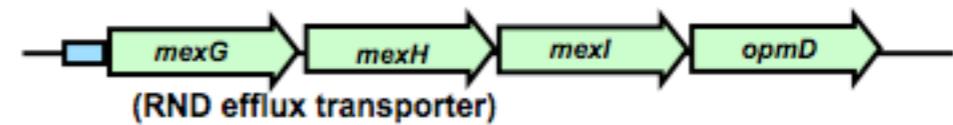
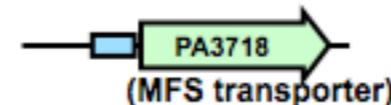
*E. coli*



$\Delta\text{soxR}$ : sensitive to paraquat

Superoxide response

*P. aeruginosa*



$\Delta\text{soxR}$ : insensitive to paraquat

Phenazine response

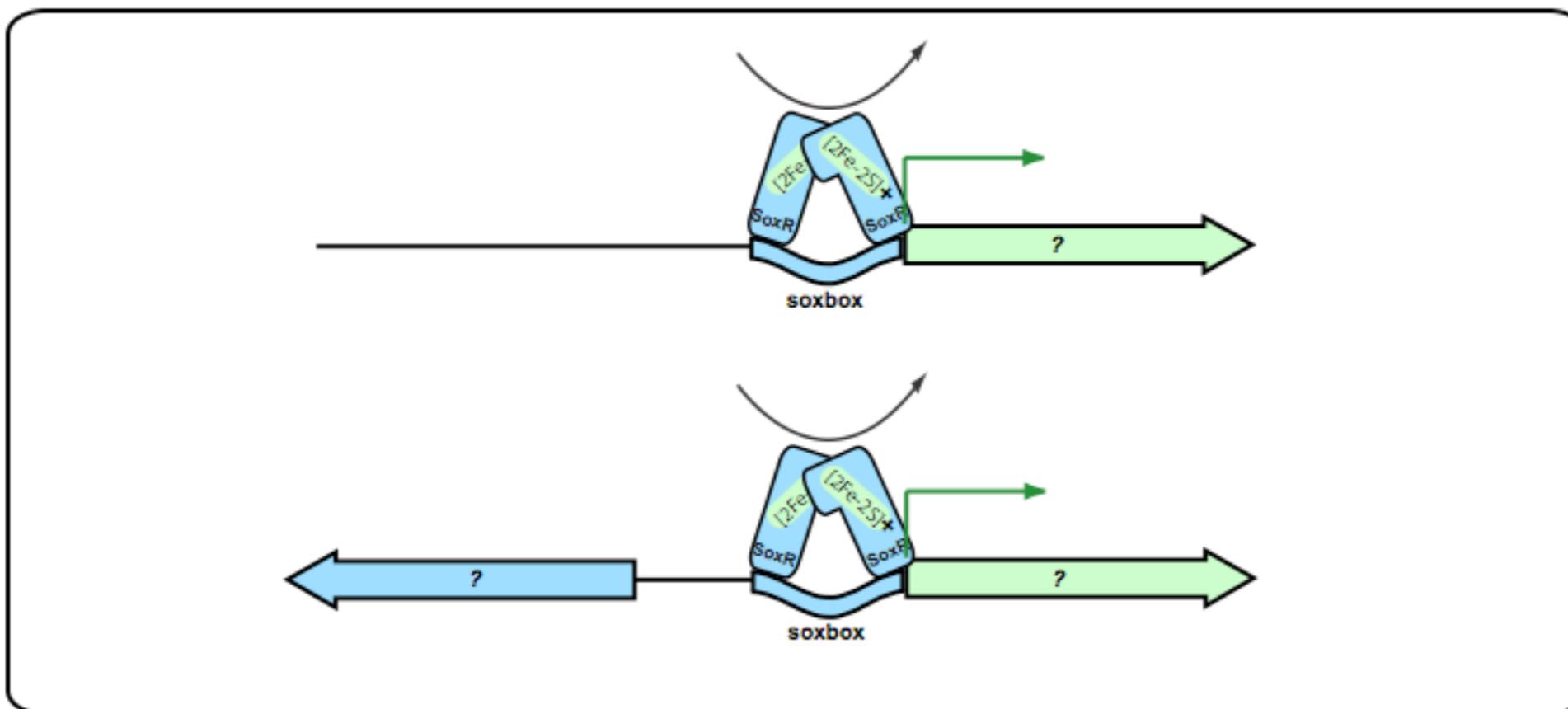
Kobayashi et al., J. Biochem., 2004

Palma et al., Infection and Immunity, 2005

Lars Dietrich

# How ubiquitous is SoxR and what types of genes does it regulate?

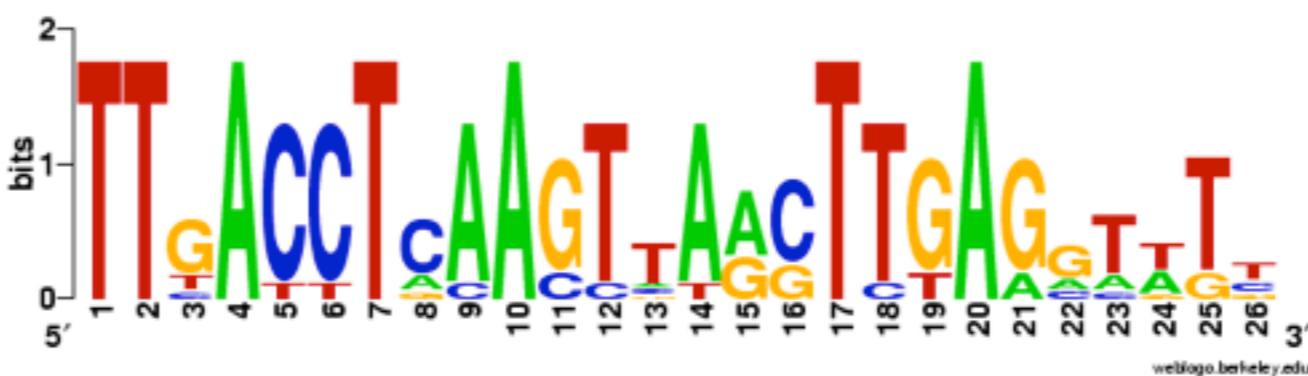
## Pseudomonads



"We cannot exclude the possibility that a completely new oxidative defense system may have evolved in *Pseudomonas* species, thus rendering SoxR-dependent system obsolete or taking over the control of genes that were formerly SoxR regulon." Park et al 2006

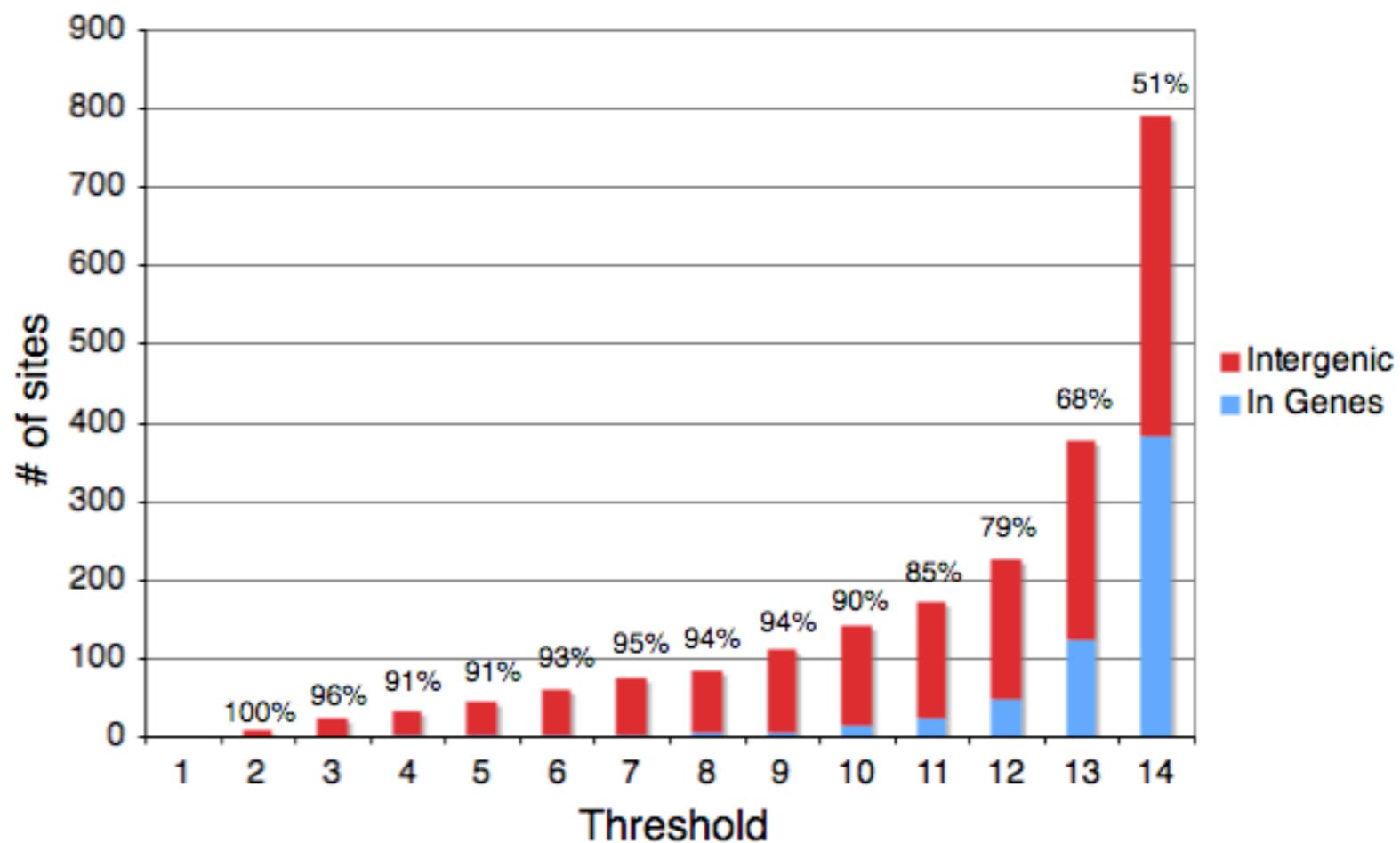
# Bioinformatics approach to finding SoxR regulated genes

|   |                              |
|---|------------------------------|
| <i>Pseudomonas aeruginosa</i> PAO1<br>PA2274 (from Pseudomonas.com) | TTGACCTCAAGTTGCTTGAGGTTT     |
| <i>Pseudomonas aeruginosa</i> PAO1<br>PA4205 (from Pseudomonas.com) | TTGACCTCAACTTAAC TTGAGGTTT   |
| <i>Pseudomonas aeruginosa</i> PAO1<br>PA3718 (from Pseudomonas.com) | TTTACCTCAAGTTAAC TTGAGCTATC  |
| <i>Escherichia coli</i><br>from Kobayashi_Tagawa2004)               | TTTACCTCAAGTTAAC TTGAGGAATT  |
| <i>Xanthomonas axonopodis</i><br>from Kobayashi_Tagawa2004          | TTGACCTCAACTTAGGTTGAGGCAGG   |
| <i>Chromobacterium axonopodis</i><br>from Kobayashi_Tagawa2004      | TTGACTTCAAGTTAAC TTGAAC TTTG |

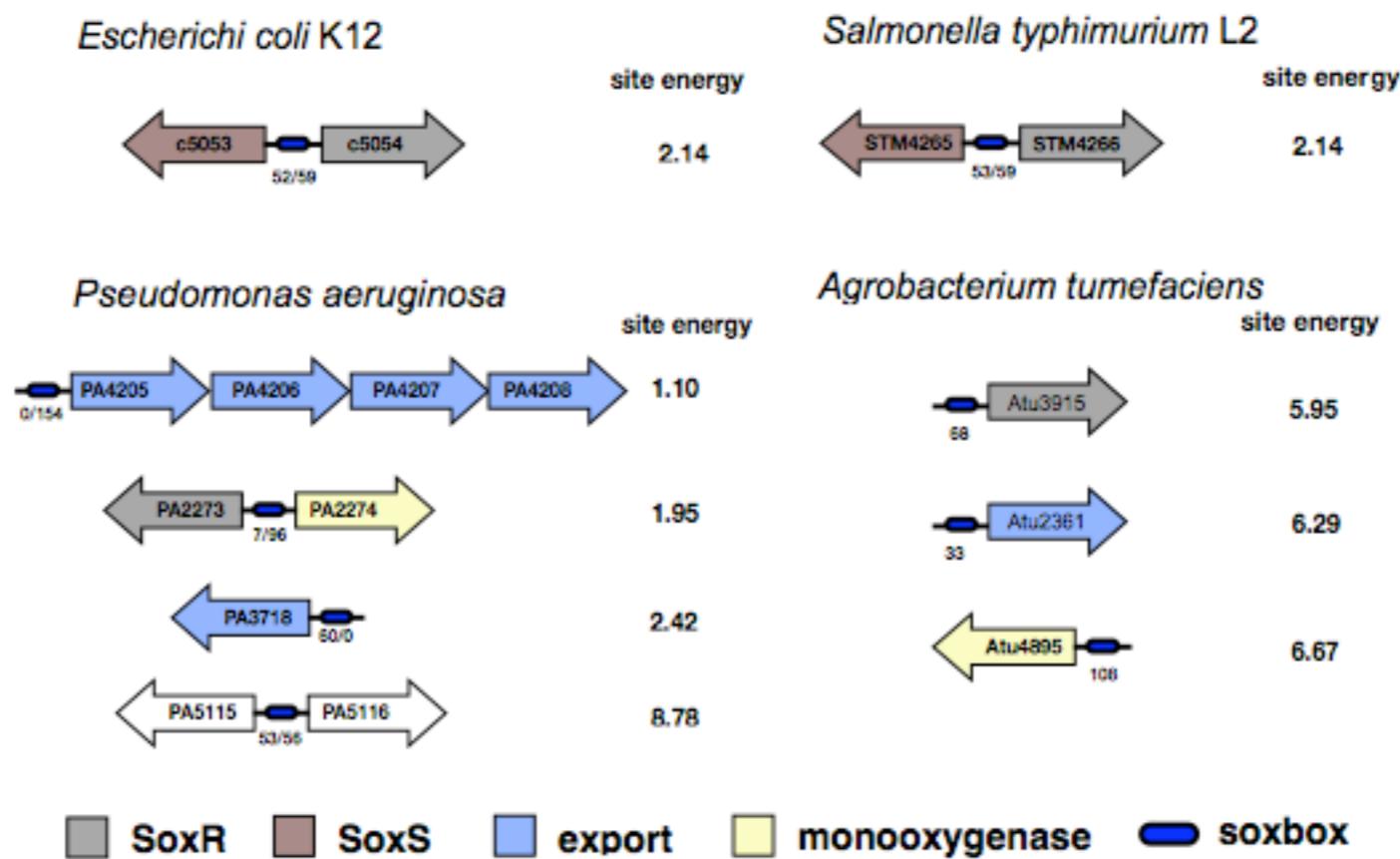


# Reliability of the PWM

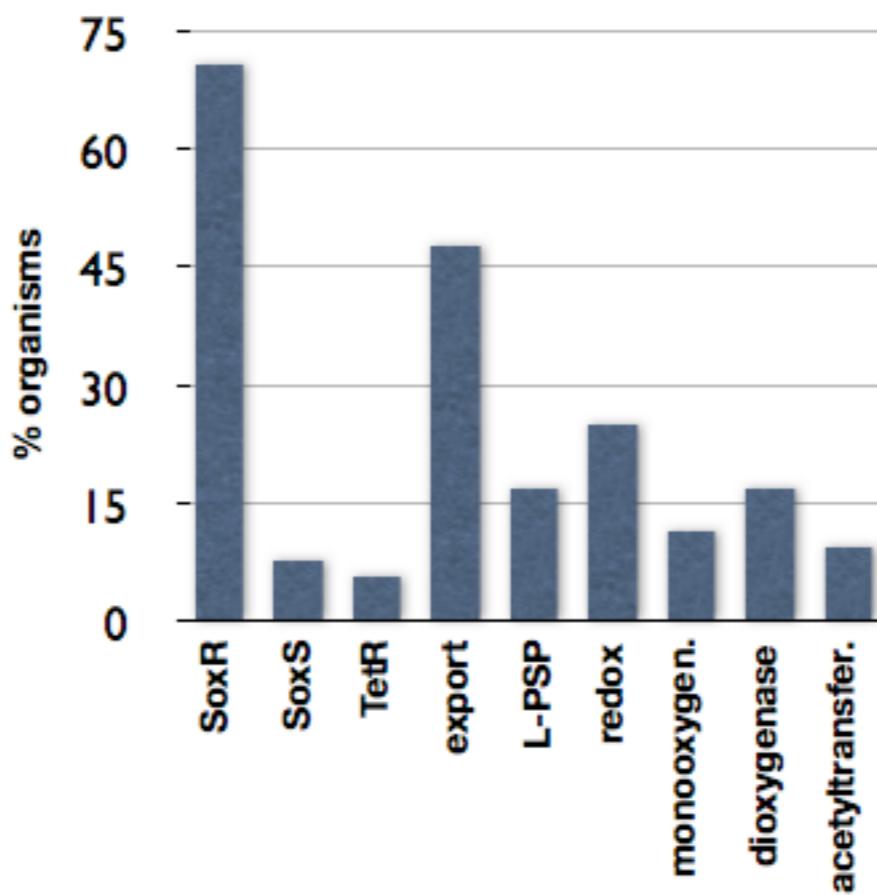
## Intergenic versus in genes



# Predictions of soxboxes match data from the literature

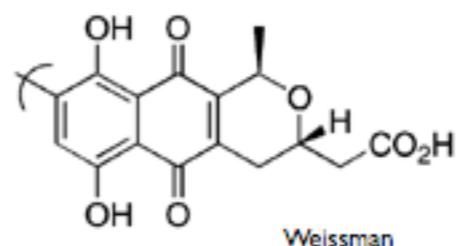


# Genes not associated with superoxide stress are more highly represented



# Testing soxbox predictions in *Streptomyces coelicolor*

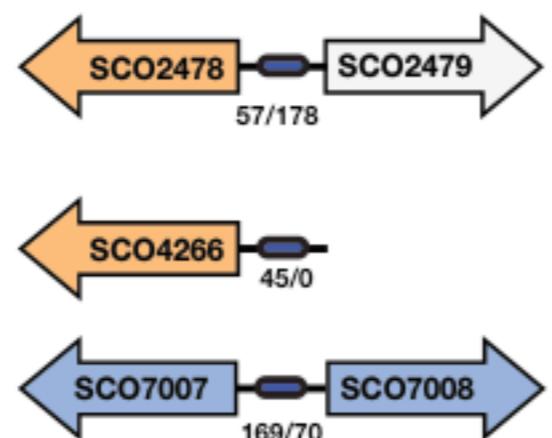
## Actinorhodin



Weissman

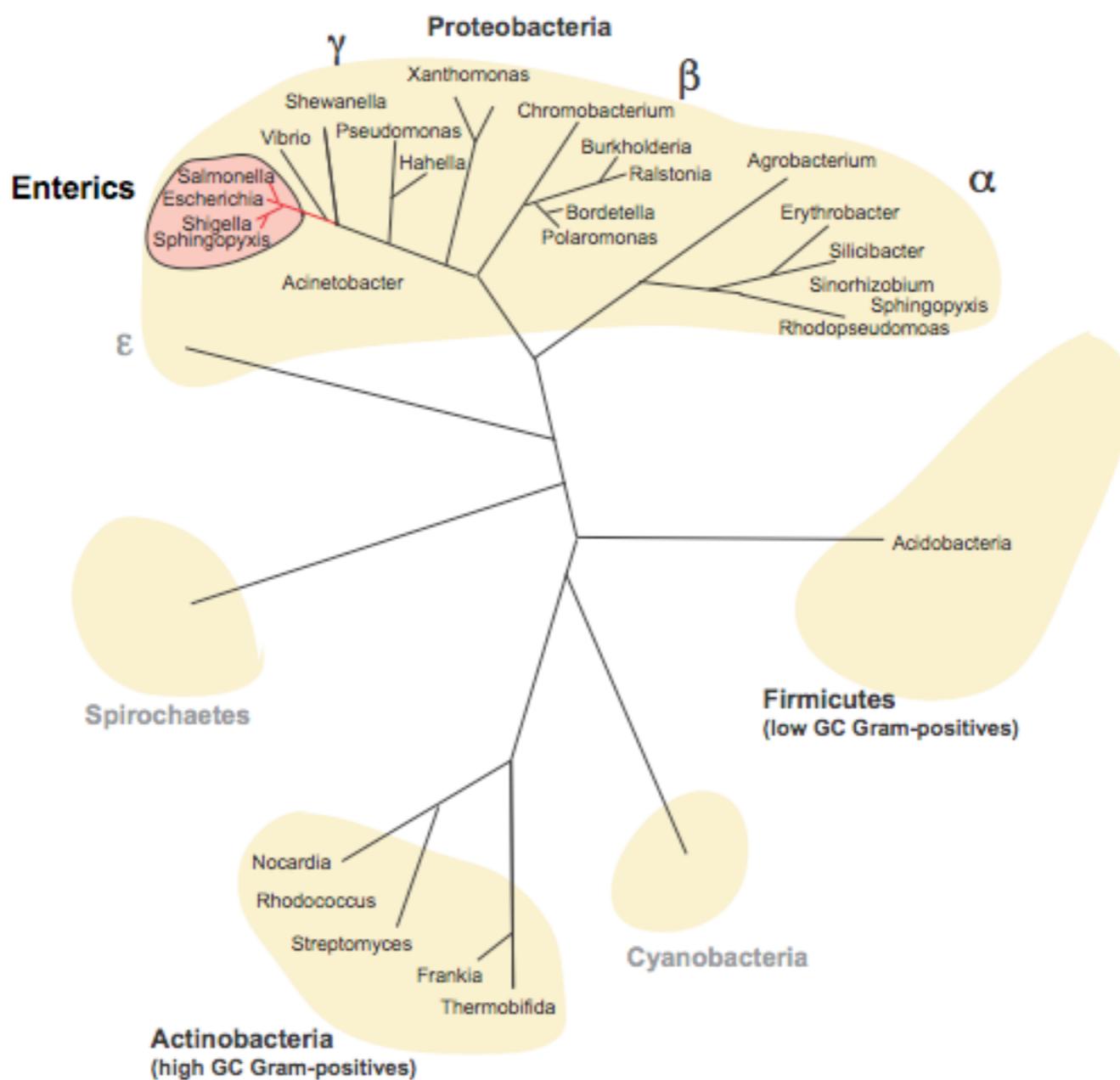
In the presence of actinorhodin: **wild type** versus  $\Delta\text{act}$

### *Streptomyces coelicolor*



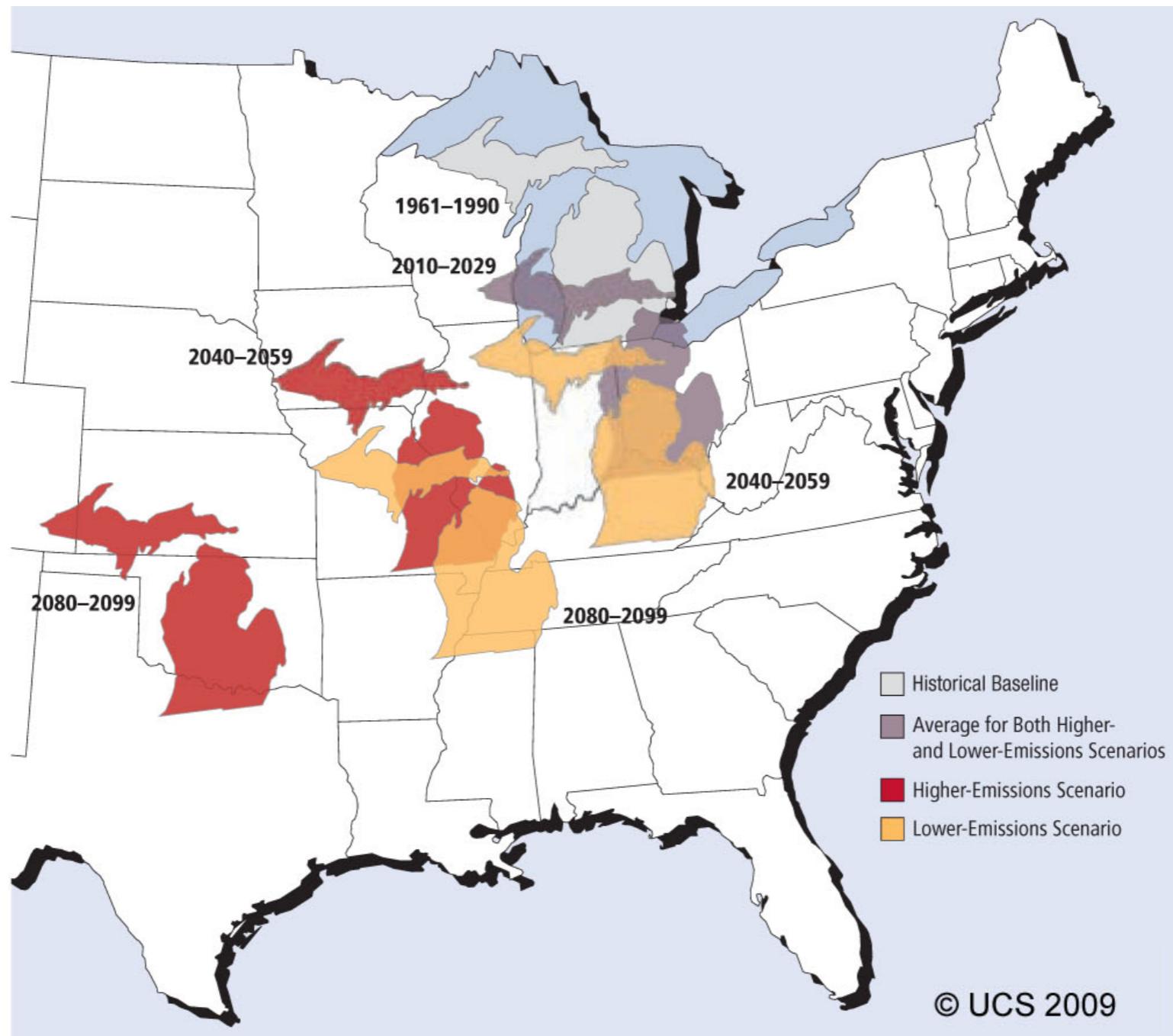
|                                 | Site energy | Regulation   |
|---------------------------------|-------------|--------------|
| SCO2478 ← - SCO2479 →<br>57/178 | 5.78        | 18x ↑ 1.7x ↓ |
| SCO4266 ← - SCO4266 →<br>45/0   | 8.53        | 53x ↑        |
| SCO7007 ← - SCO7008 →<br>169/70 | 8.75        | 11x ↑ -      |

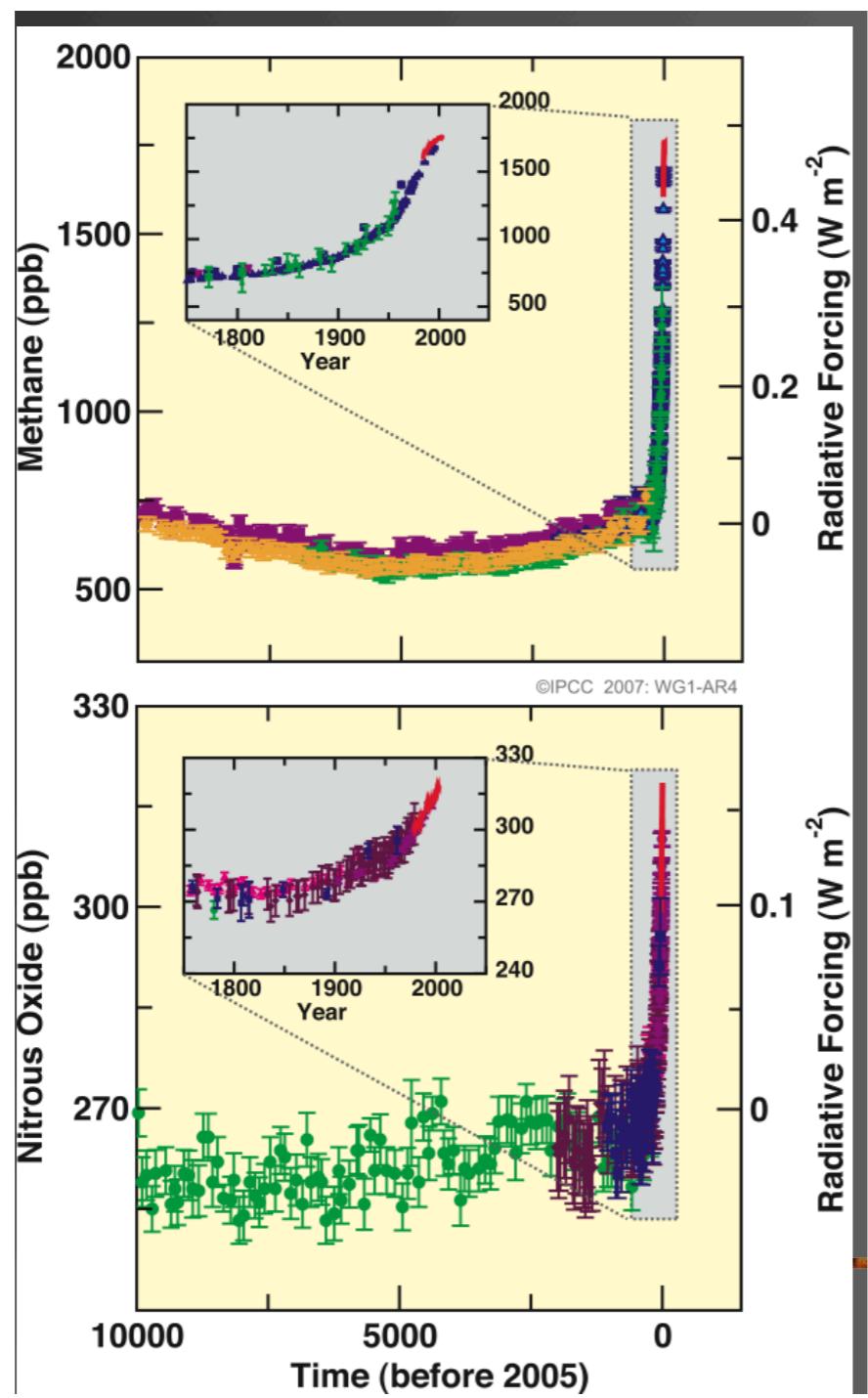
# SoxRS paradigm seems limited to enterics



# The effects of land use change on microbial communities and greenhouse gas fluxes

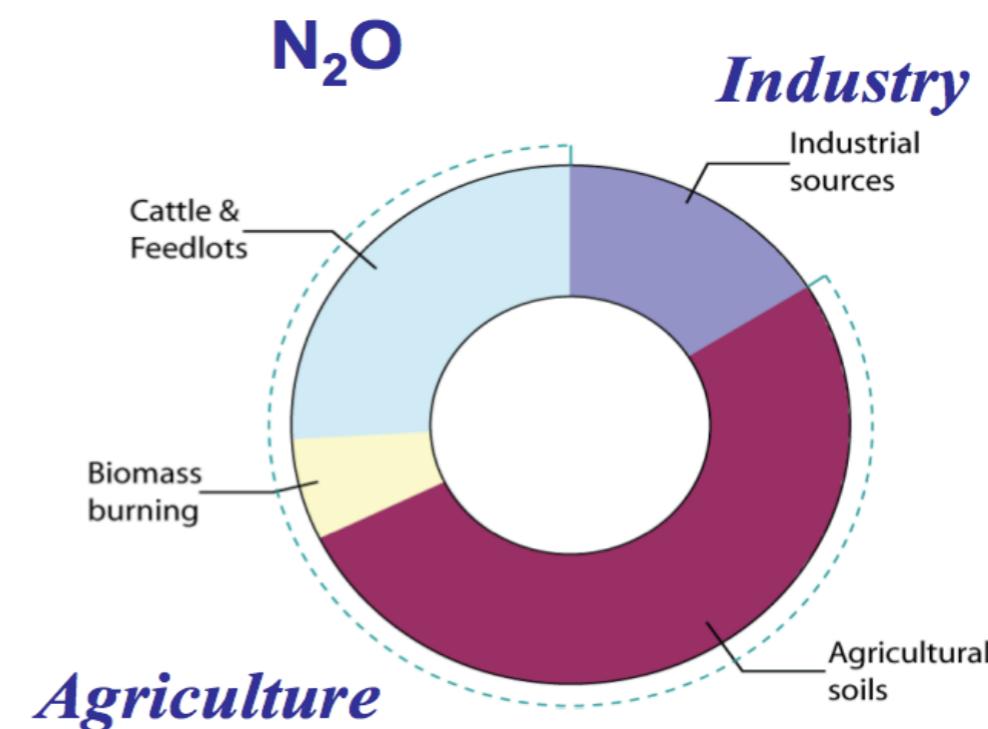
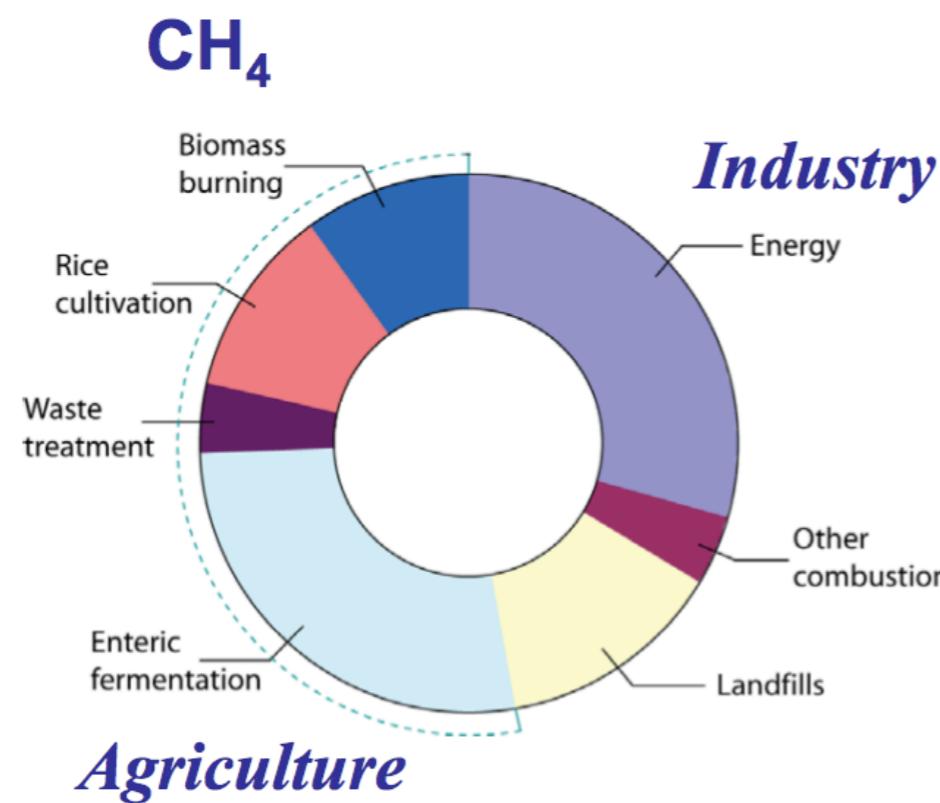
# Climate change is happening



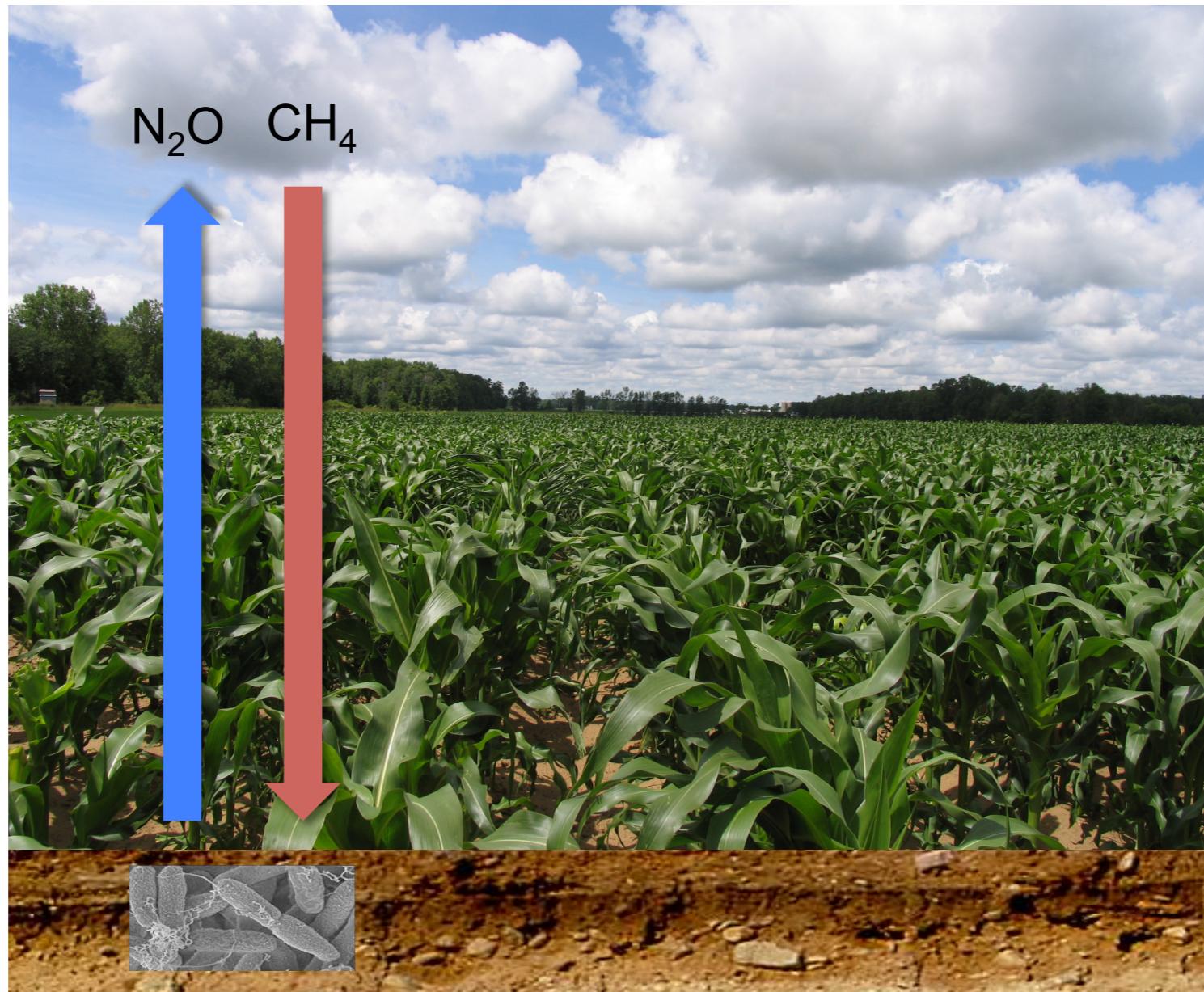


- ❖ Dramatic increases in  $\text{CH}_4$  and  $\text{N}_2\text{O}$  in the last 200 years
- ❖ High Global Warming Potential (GWP) due to time in the atmosphere and infrared absorption

# Much of this shift is due to land use change and intensive agriculture



# Microbes are the primary mediators of methane and nitrous oxide



How do microbial communities change with land management?

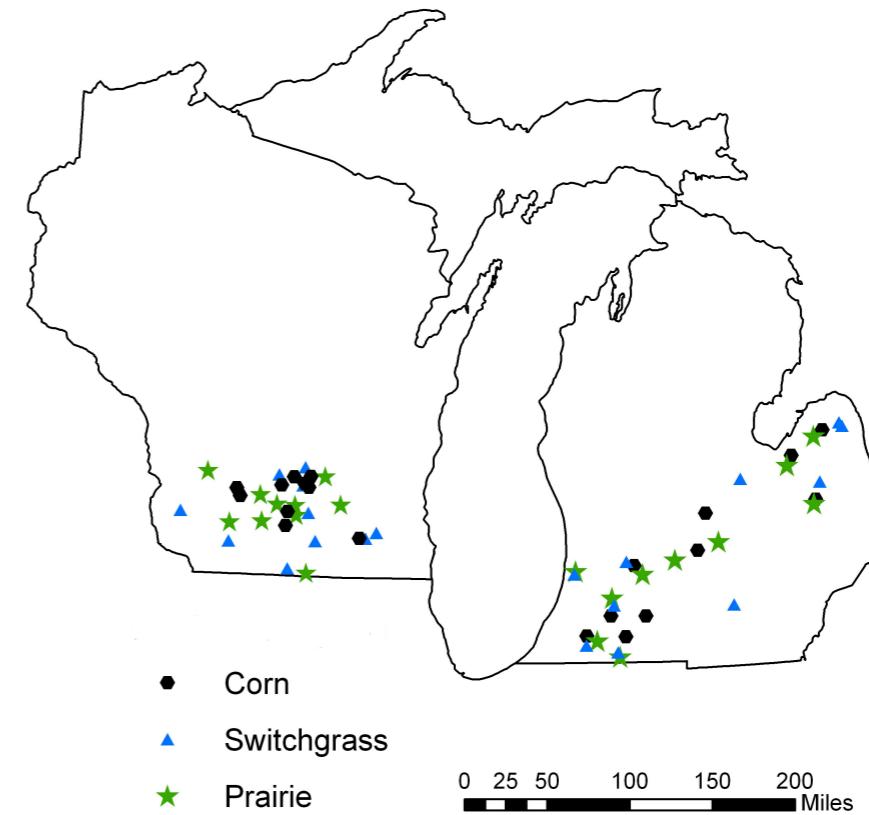
What is the relationship between denitrifying bacteria and N<sub>2</sub>O in agricultural soils?

# Experiment sites

Kellogg Biological Station LTER



GLBRC Extensive Sites



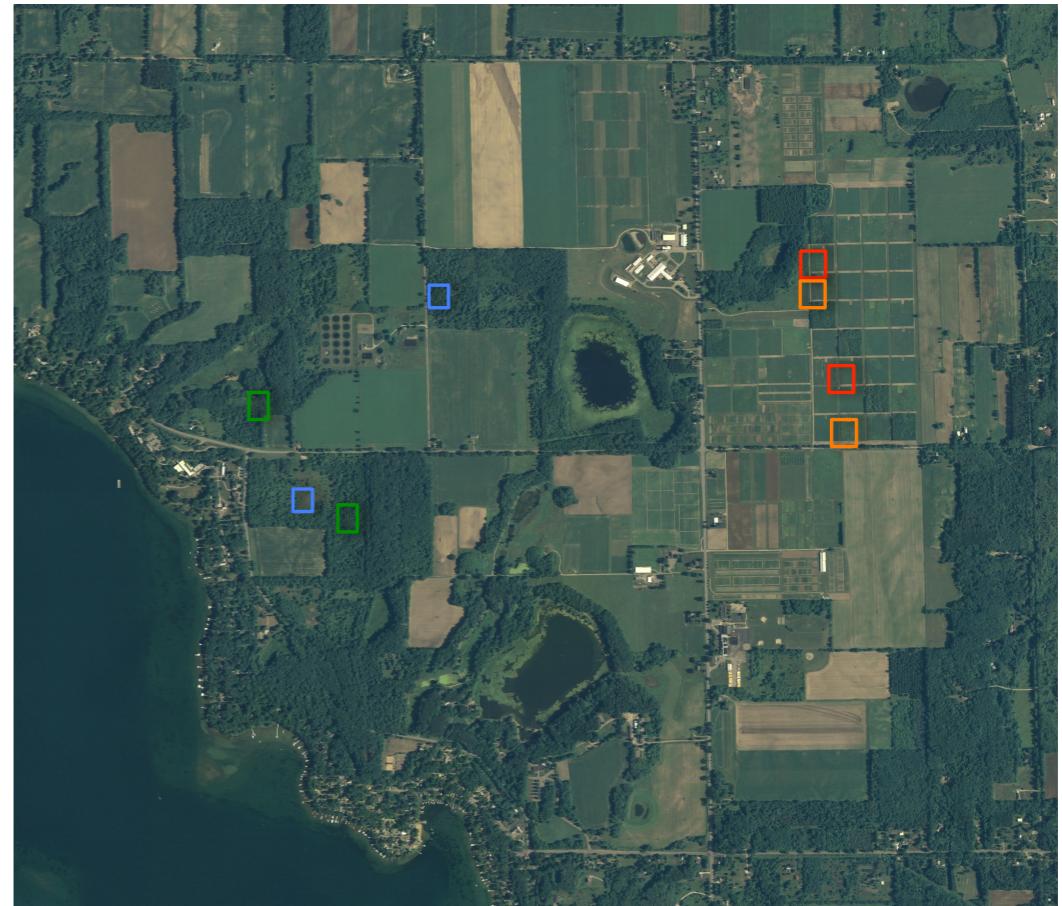
- Corn
- ▲ Switchgrass
- ★ Prairie

0 25 50 100 150 200 Miles

# How do microbial communities change with land management?

Kellogg Biological Station LTER

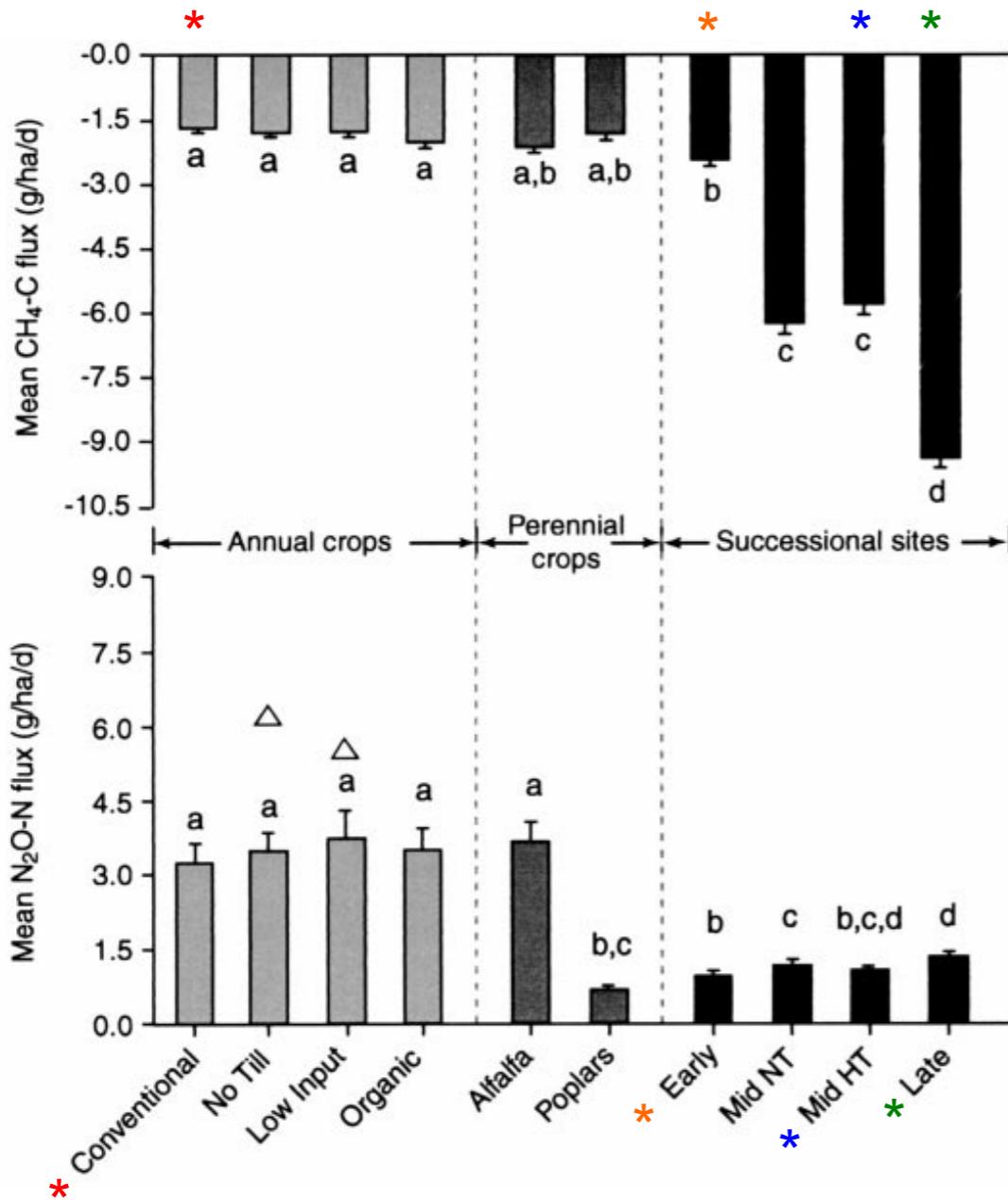
- ✖ Vicente Gomez-Alvarez
- ✖ Tom Schmidt



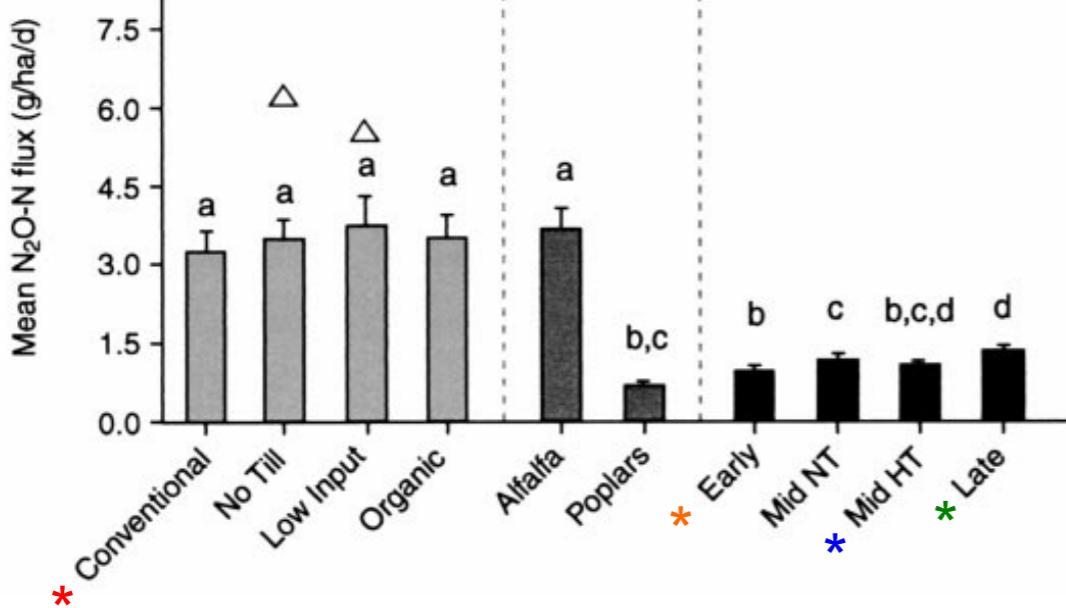
# How do microbial communities change with land management?

Kellogg Biological Station LTER

Methane



Nitrous Oxide

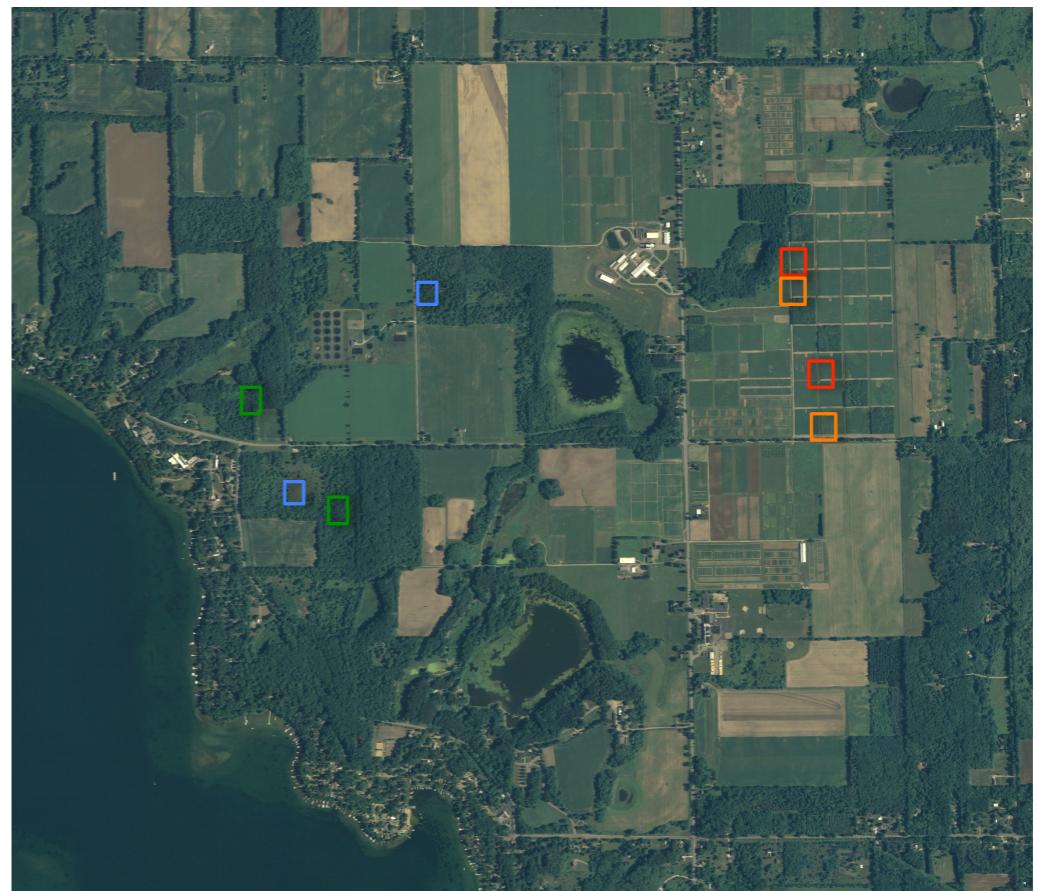


- AG      Conventional Agriculture
- ES      Early Successional
- SF      Successional Forest
- DF      Deciduous Forest

# Sampling design

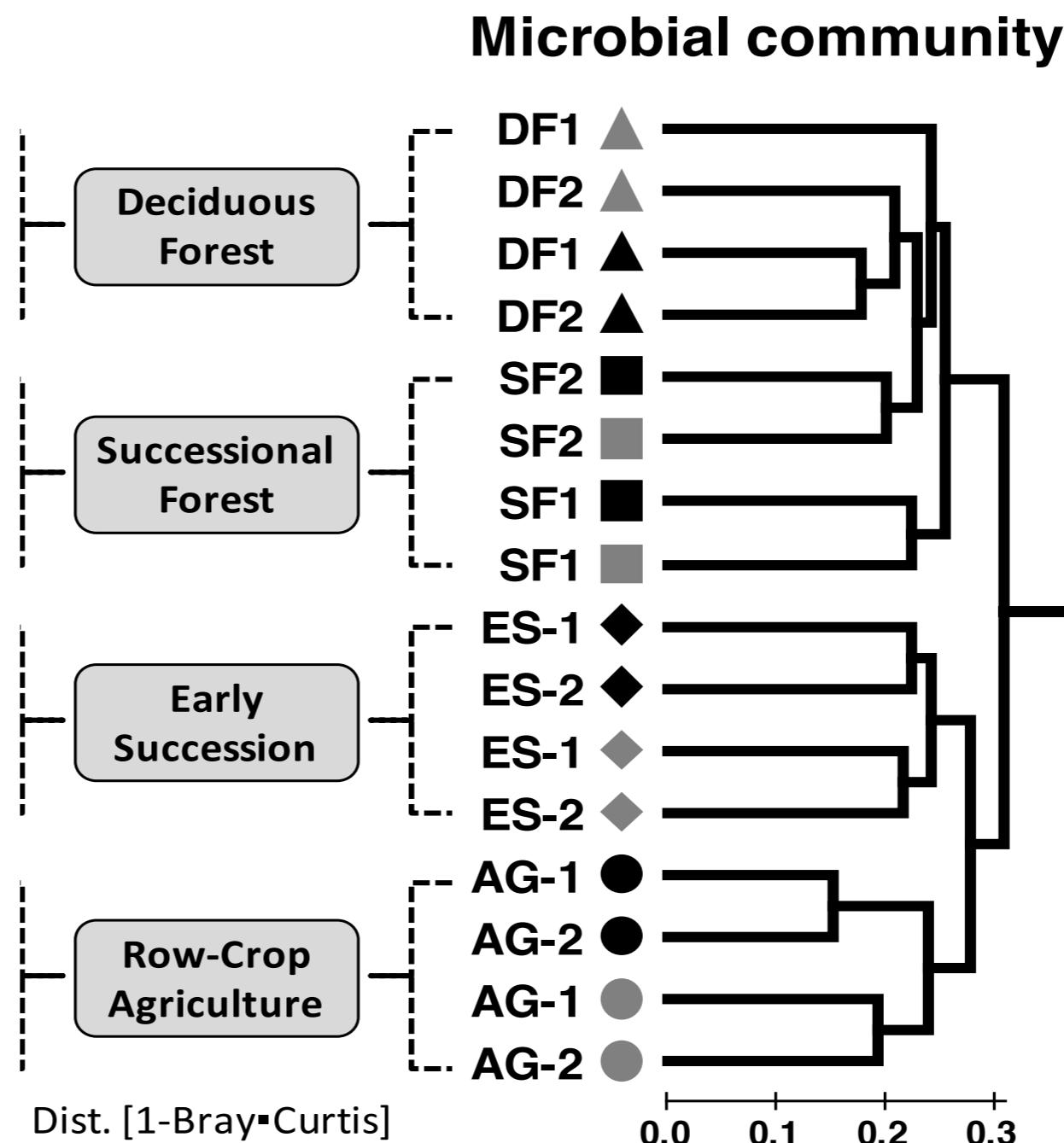
Sampled two plots of 4 treatments  
In two years – 2008 and 2009

- 10 cm cores
- Soil from 5 flags pooled and sieved
- Soil frozen after being collected
- Microbial community analysis
- Soil analysis

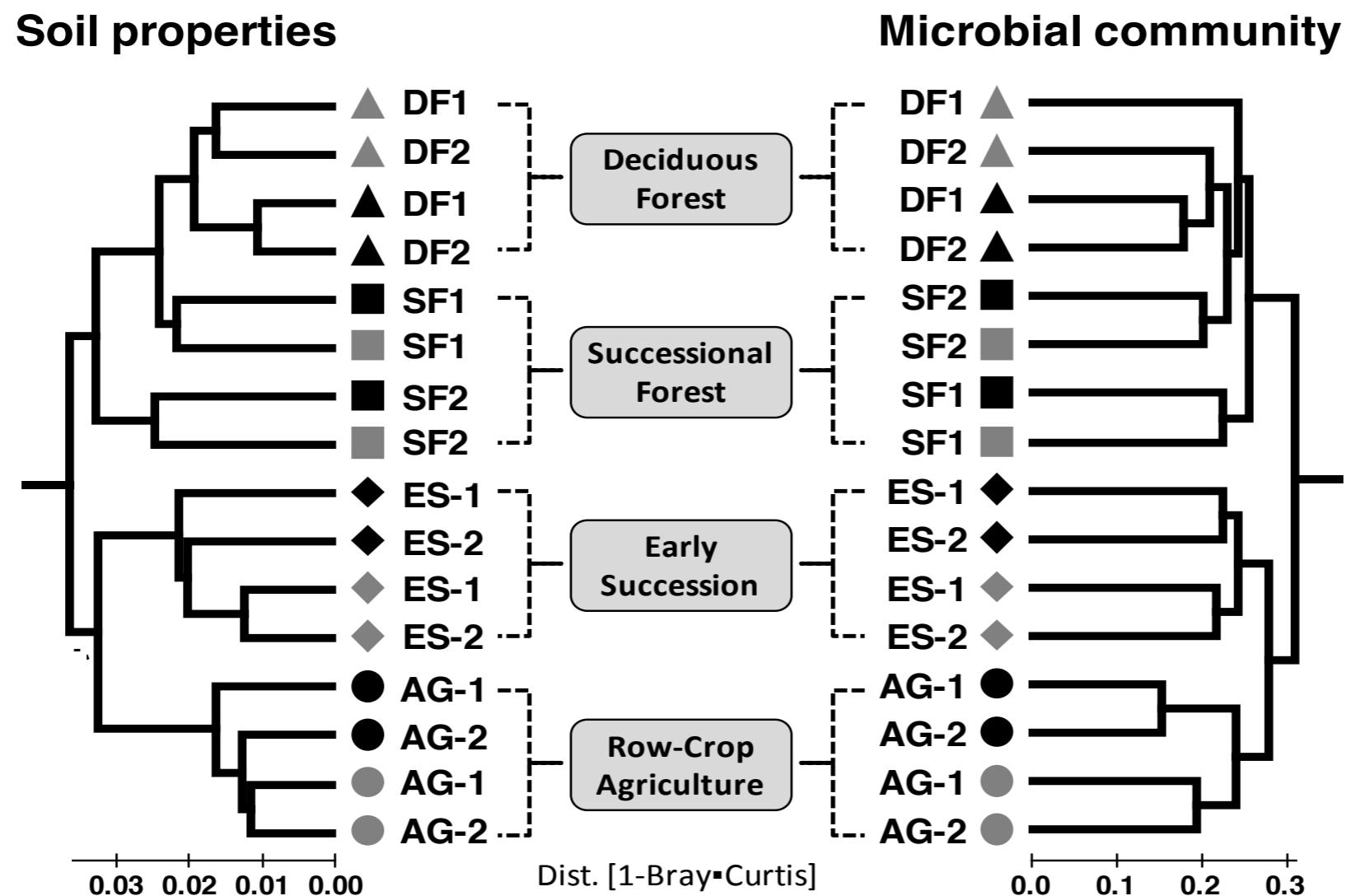


- AG Conventional Agriculture
- ES Early Successional
- SF Successional Forest
- DF Deciduous Forest

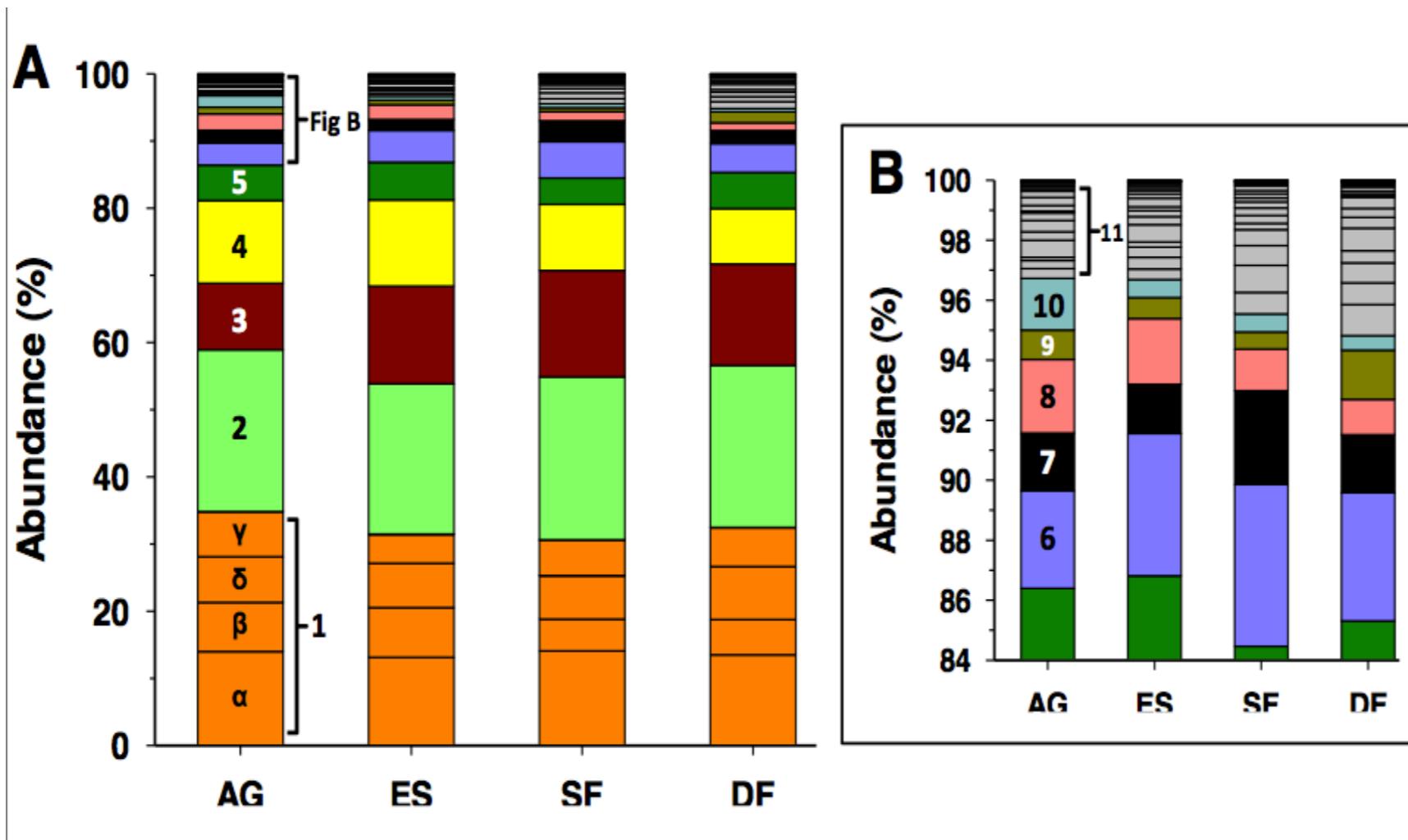
# Taxonomic composition of bacterial communities changes with treatment



# Biogeochemistry and bacterial community change concomitantly

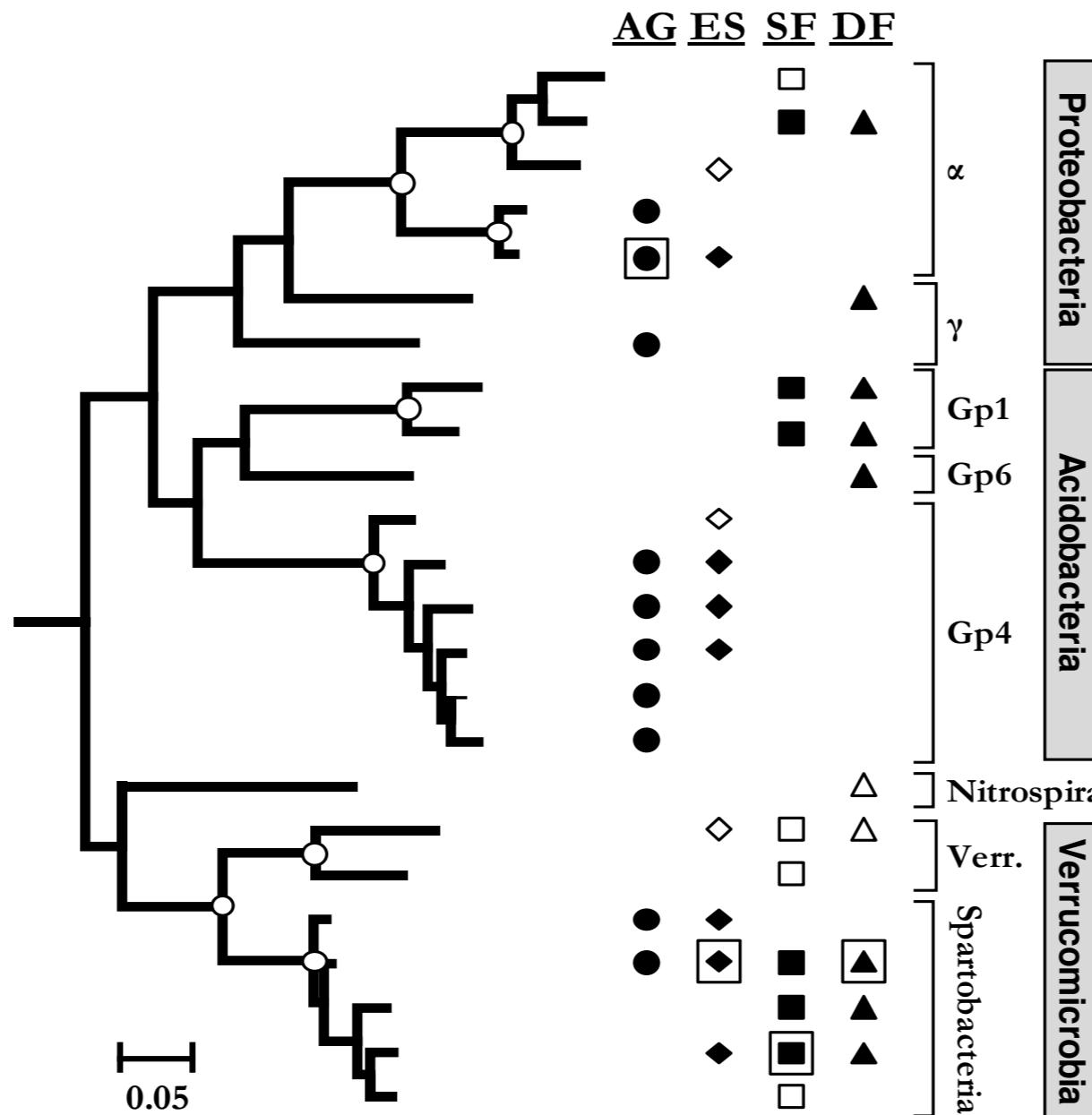


# Who's there and differentiating communities



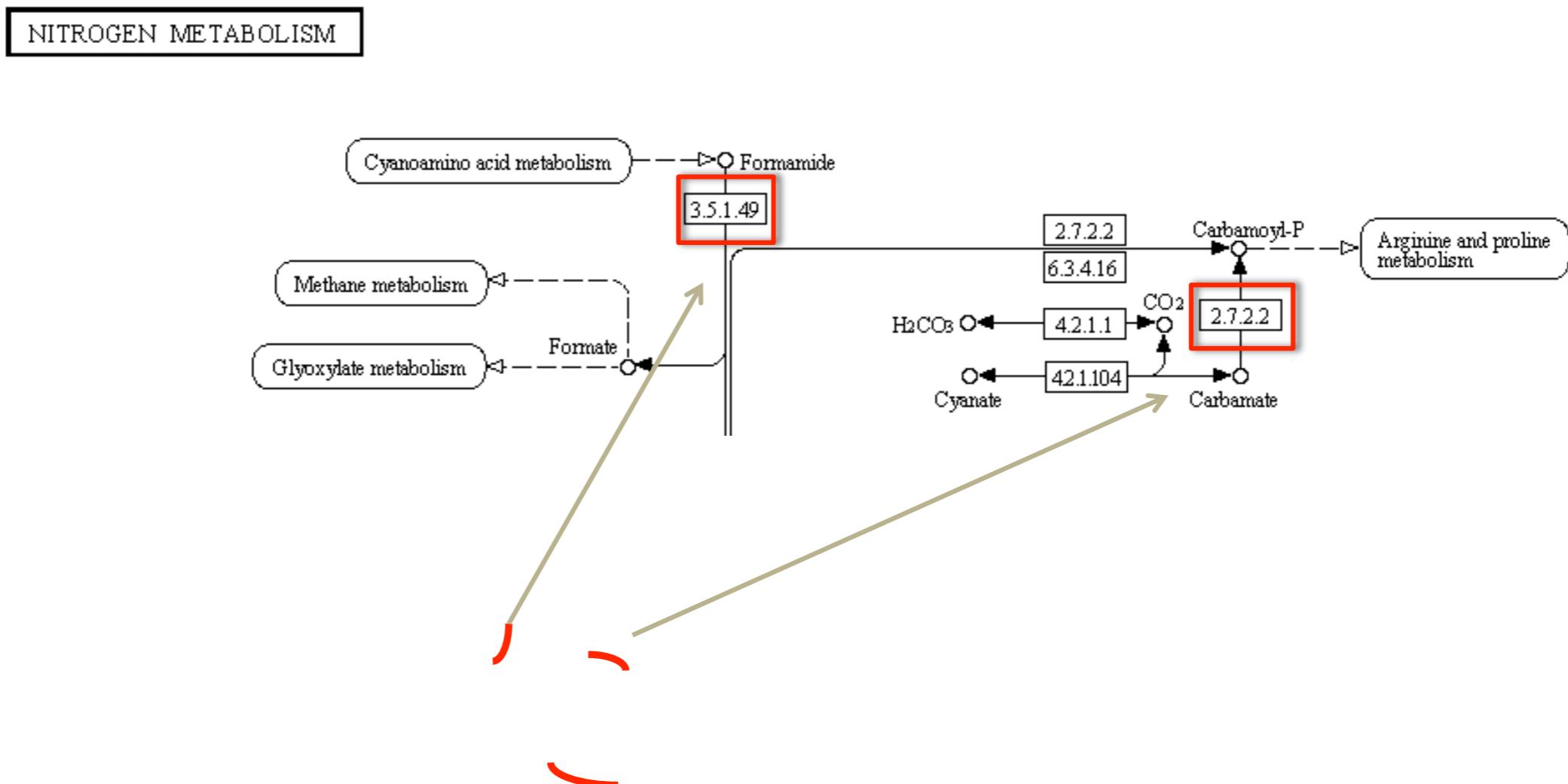
- (1) Proteobacteria [class added], (2) Acidobacteria, (3) Verrucomicrobia, (4) Actinobacteria, (5) Bacteroidetes, (6) Planctomycetes, (7) Chloroflexi, (8) Gemmatimonadetes, (9) Nitrospirae, (10) Firmicutes, and (11) 30 additional phyla corresponding to less than 4% of the total distribution.

## Shifts within phyla differentiate communities



Metagenomics is terrible, but why we use it anyway

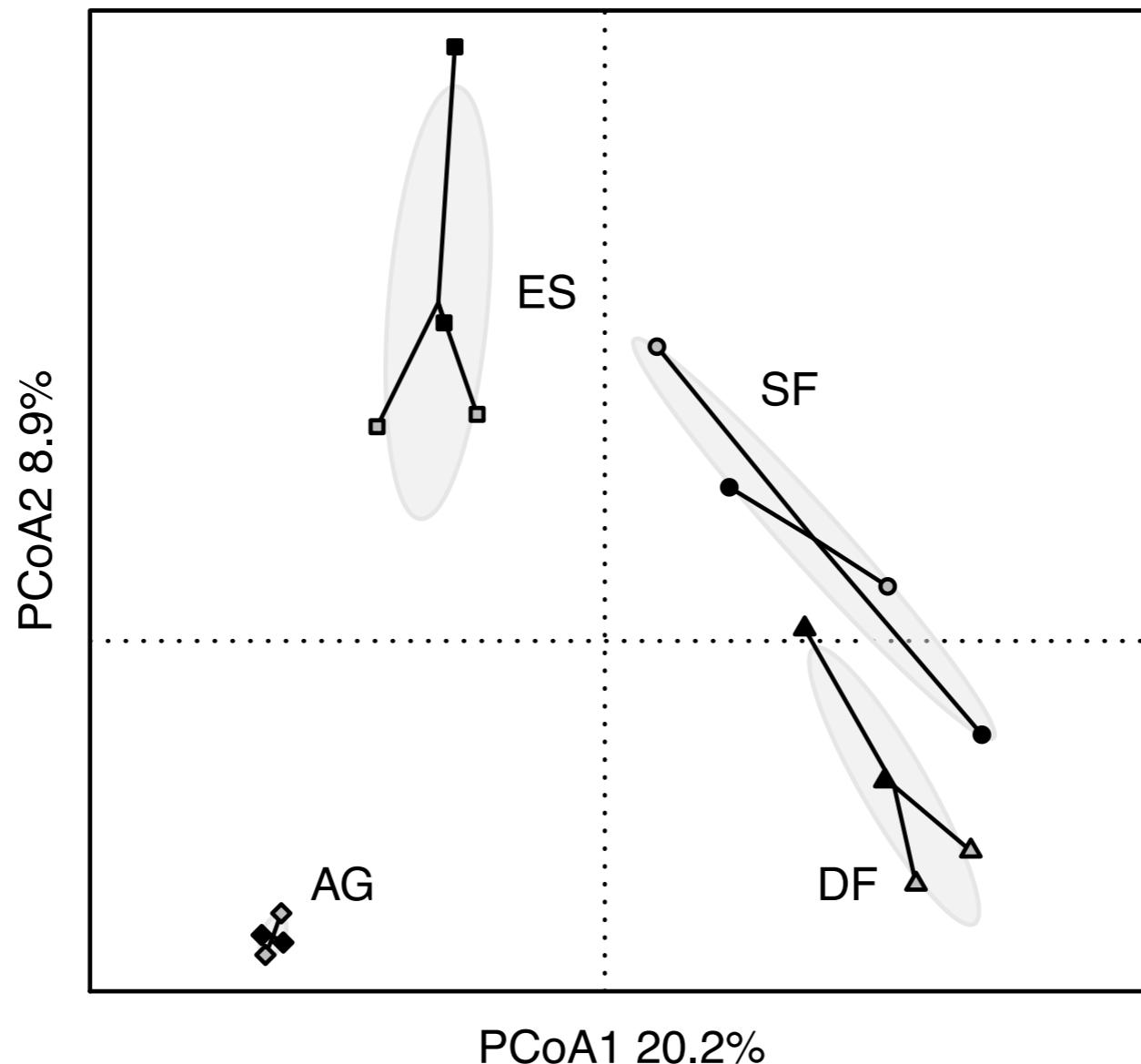
# Annotate shotgun reads



# Matrix of normalized gene abundance by treatment

| Function  | Subsystem                   | T1R1 |      |      |      | T1R2 |      |      |      | T1R3 |      |      |      |
|---|-----------------------------|------|------|------|------|------|------|------|------|------|------|------|------|
|   |                             | 2008 | 2008 | 2009 | 2009 | 2008 | 2008 | 2009 | 2009 | 2008 | 2008 | 2009 | 2009 |
| (R)-citramalate synthase (EC 2.3.1.182)   | Amino Acids and Derivatives | 84   |      | 103  |      |      |      |      |      | 53   |      |      |      |
| 1,2-dihydroxy-3-keto-5-methylthiopentene dioxygenase (EC 1.13.11.54)              | Amino Acids and Derivatives | 9    |      | 4    |      |      |      |      |      | 8    |      |      |      |
| 1-pyrroline-4-hydroxy-2-carboxylate deaminase (EC 3.5.4.22)                       | Amino Acids and Derivatives | 21   |      | 17   |      |      |      |      |      | 17   |      |      |      |
| 2,3,4,5-tetrahydropyridine-2,6-dicarboxylate N-acetyltransferase (EC 2.3.1.89)    | Amino Acids and Derivatives | 28   |      | 23   |      |      |      |      |      | 21   |      |      |      |
| 2,3,4,5-tetrahydropyridine-2,6-dicarboxylate N-succinyltransferase (EC 2.3.1.117) | Amino Acids and Derivatives | 71   |      | 72   |      |      |      |      |      | 42   |      |      |      |
| 2,3-diketo-5-methylthiopentyl-1-phosphate enolase                                 | Amino Acids and Derivatives | 9    |      | 10   |      |      |      |      |      | 6    |      |      |      |
| 2,3-diketo-5-methylthiopentyl-1-phosphate enolase-phosphatase (EC 3.1.3.77)       | Amino Acids and Derivatives | 9    |      | 10   |      |      |      |      |      | 5    |      |      |      |
| 2,4-diaminopentanoate dehydrogenase (EC 1.4.1.12)                                 | Amino Acids and Derivatives | 1    |      | 0    |      |      |      |      |      | 0    |      |      |      |
| 2,4-dihydroxyhept-2-ene-1,7-dioic acid aldolase (EC 4.1.2.-)                      | Amino Acids and Derivatives | 62   |      | 42   |      |      |      |      |      | 38   |      |      |      |
| 2-Amino-2-deoxy-isochorismate synthase (EC 4.1.3.-)                               | Amino Acids and Derivatives | 2    |      | 8    |      |      |      |      |      | 0    |      |      |      |
| 2-Oxobutyrate dehydrogenase E1 (EC:1.2.4.1)                                       | Amino Acids and Derivatives | 0    |      | 1    |      |      |      |      |      | 2    |      |      |      |
| 2-Oxobutyrate oxidase, putative   | Amino Acids and Derivatives | 11   |      | 6    |      |      |      |      |      | 5    |      |      |      |
| 2-amino-3,7-dideoxy-D-threo-hept-6-ulosonate synthase (EC 2.5.1.-)                | Amino Acids and Derivatives | 0    |      | 1    |      |      |      |      |      | 0    |      |      |      |
| 2-amino-3-carboxymuconate-6-semialdehyde decarboxylase (EC 4.1.1.45)              | Amino Acids and Derivatives | 52   |      | 52   |      |      |      |      |      | 43   |      |      |      |
| 2-amino-3-ketobutyrate coenzyme A ligase (EC 2.3.1.29)                            | Amino Acids and Derivatives | 107  |      | 97   |      |      |      |      |      | 53   |      |      |      |
| 2-amino-4-ketopentanoate thiolase, beta subunit                                   | Amino Acids and Derivatives | 1    |      | 0    |      |      |      |      |      | 0    |      |      |      |
| 2-aminomuconate deaminase (EC 3.5.99.5)   | Amino Acids and Derivatives | 2    |      | 0    |      |      |      |      |      | 0    |      |      |      |
| 2-aminomuconate semialdehyde dehydrogenase (EC 1.2.1.32)                          | Amino Acids and Derivatives | 6    |      | 3    |      |      |      |      |      | 3    |      |      |      |
| 2-hydroxy-3-keto-5-methylthiopentenyl-1-phosphate phosphatase                     | Amino Acids and Derivatives | 3    |      | 3    |      |      |      |      |      | 1    |      |      |      |
| 2-hydroxy-3-keto-5-methylthiopentenyl-1-phosphate phosphatase related protein     | Amino Acids and Derivatives | 3    |      | 2    |      |      |      |      |      | 1    |      |      |      |

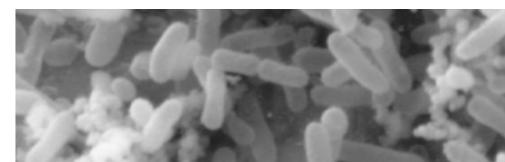
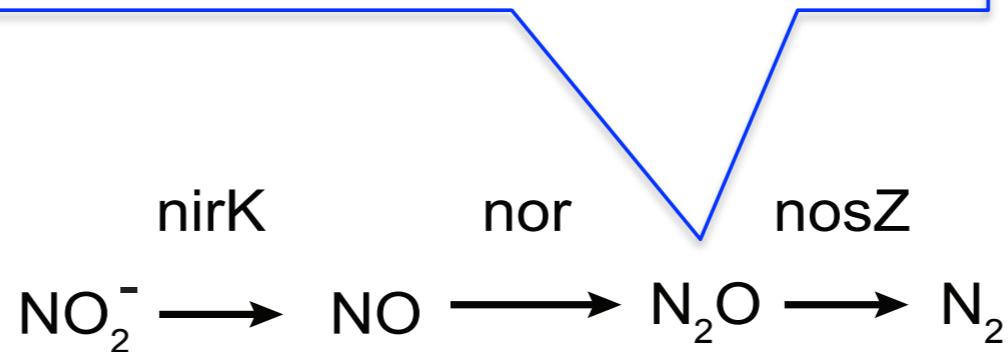
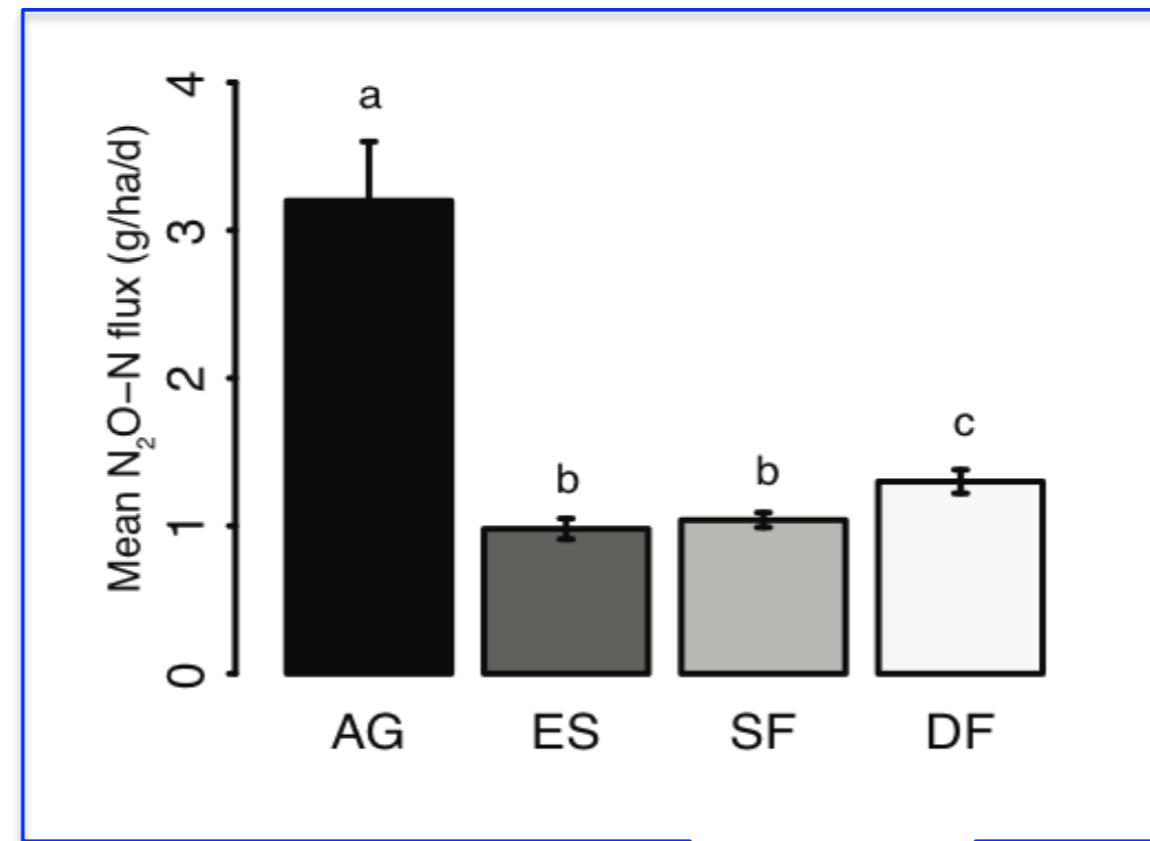
# Functional potential changes with land management



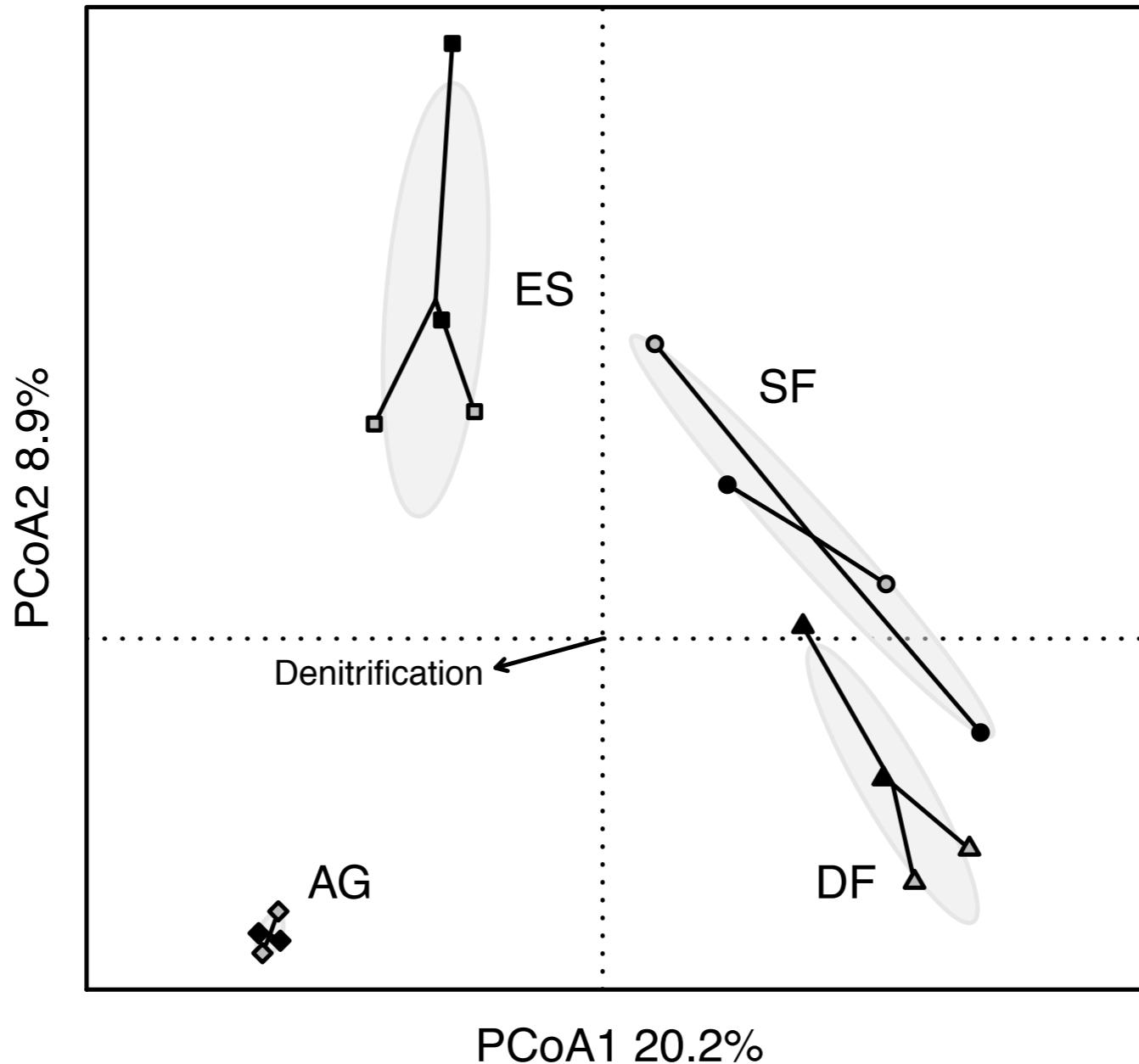
# How do microbial communities change with land management?

- ✖ Bacterial community taxonomic composition and functional potential changes with land use
- ✖ Communities change concomitantly with soil biogeochemistry
- ✖ Community composition differs in agriculture from forested sites

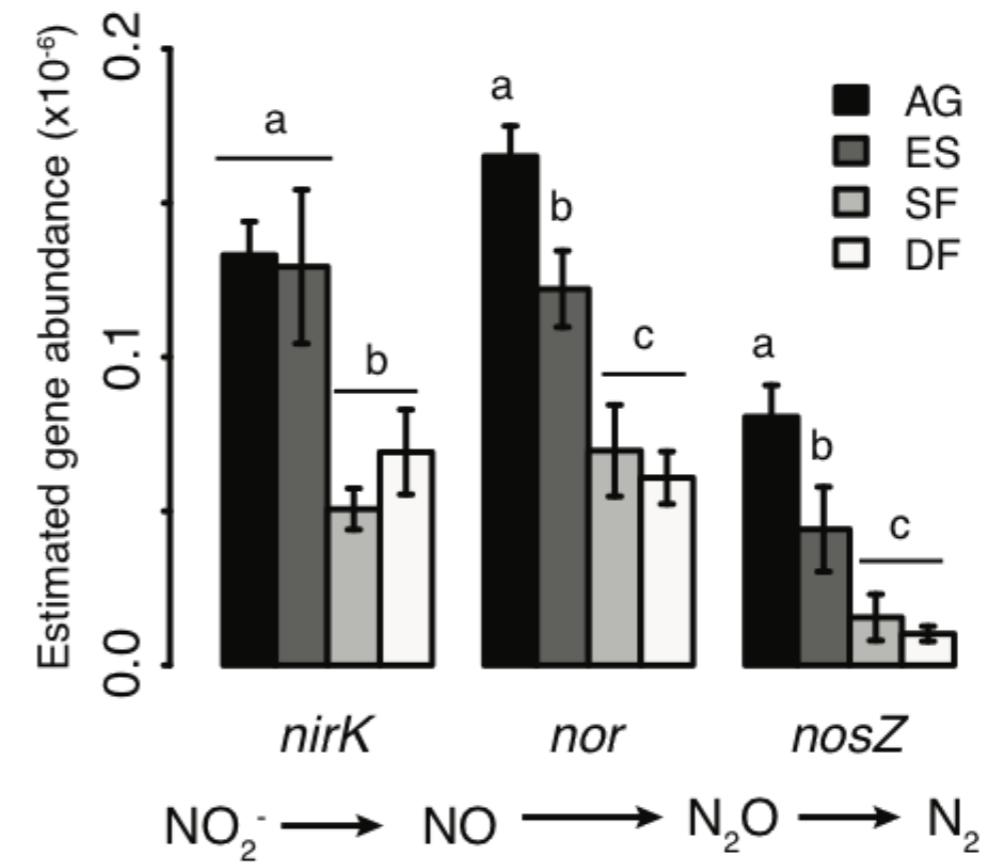
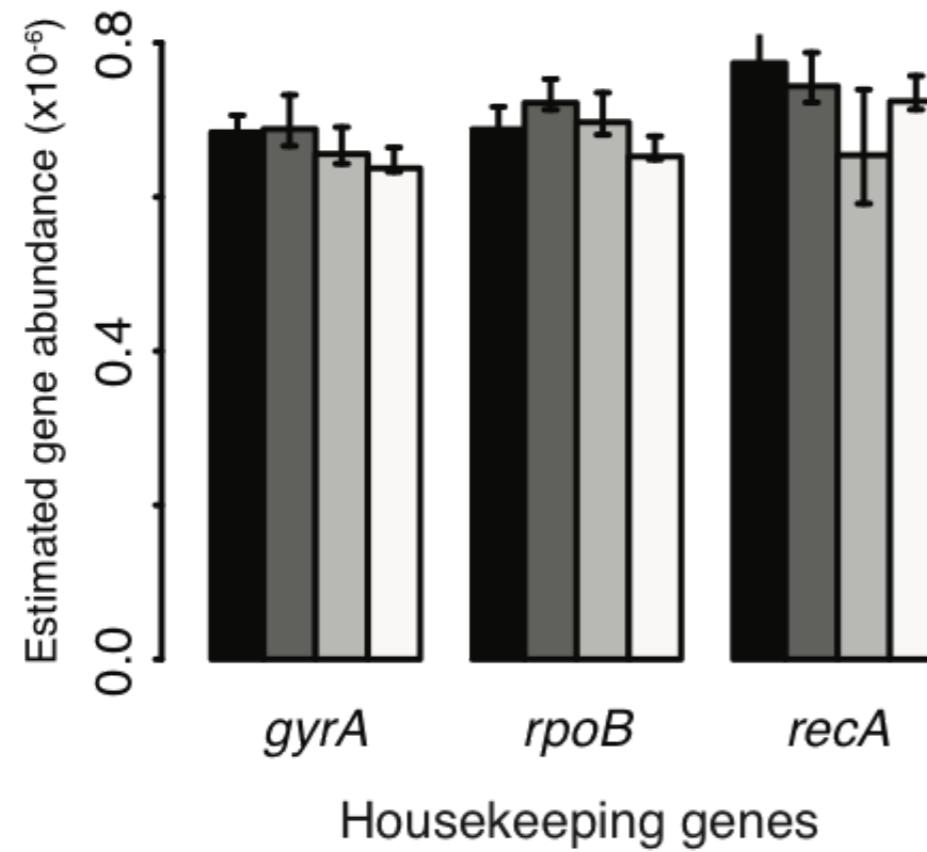
# Denitrifying microbes



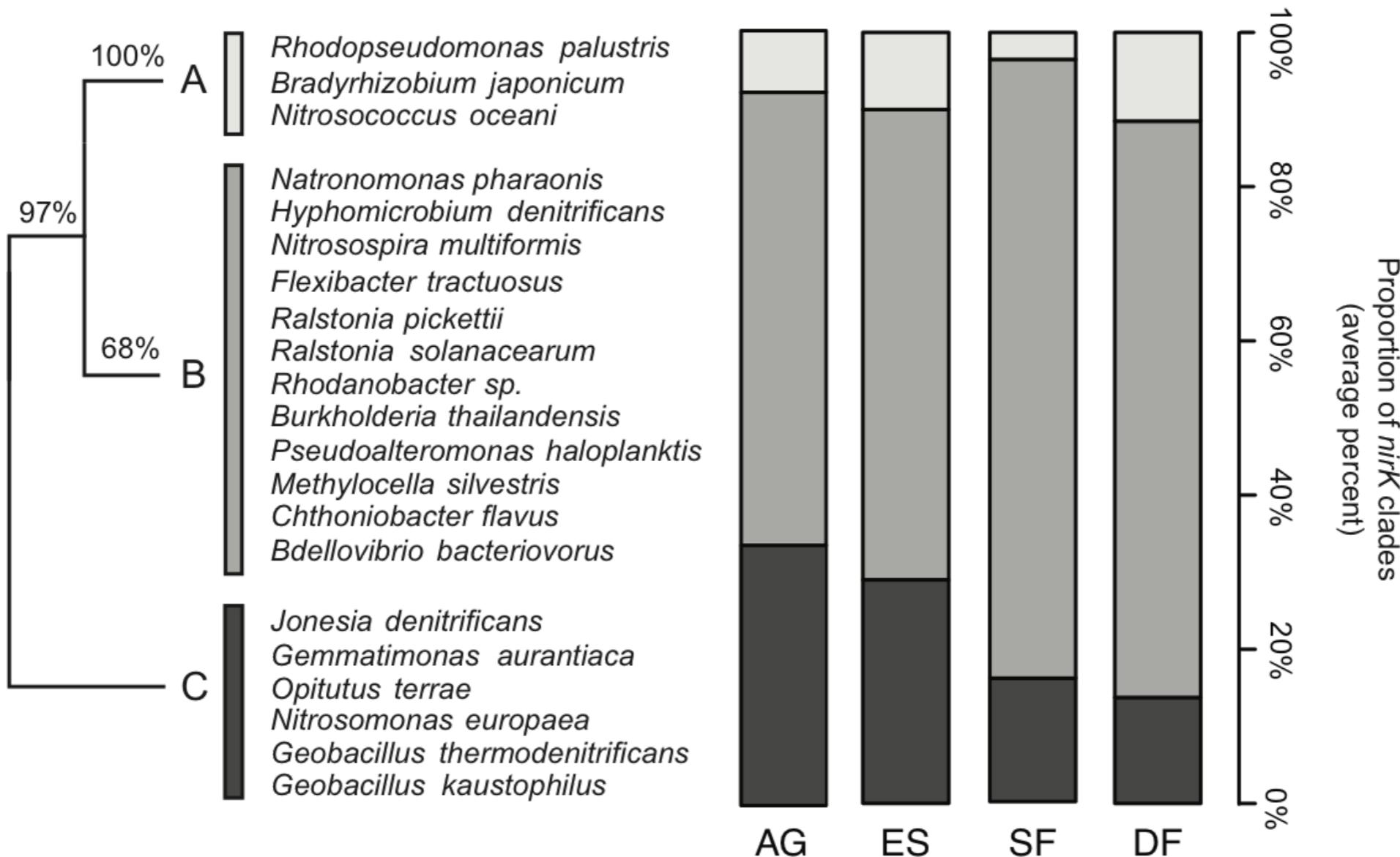
# Nitrogen metabolism contributes to the differentiation of communities



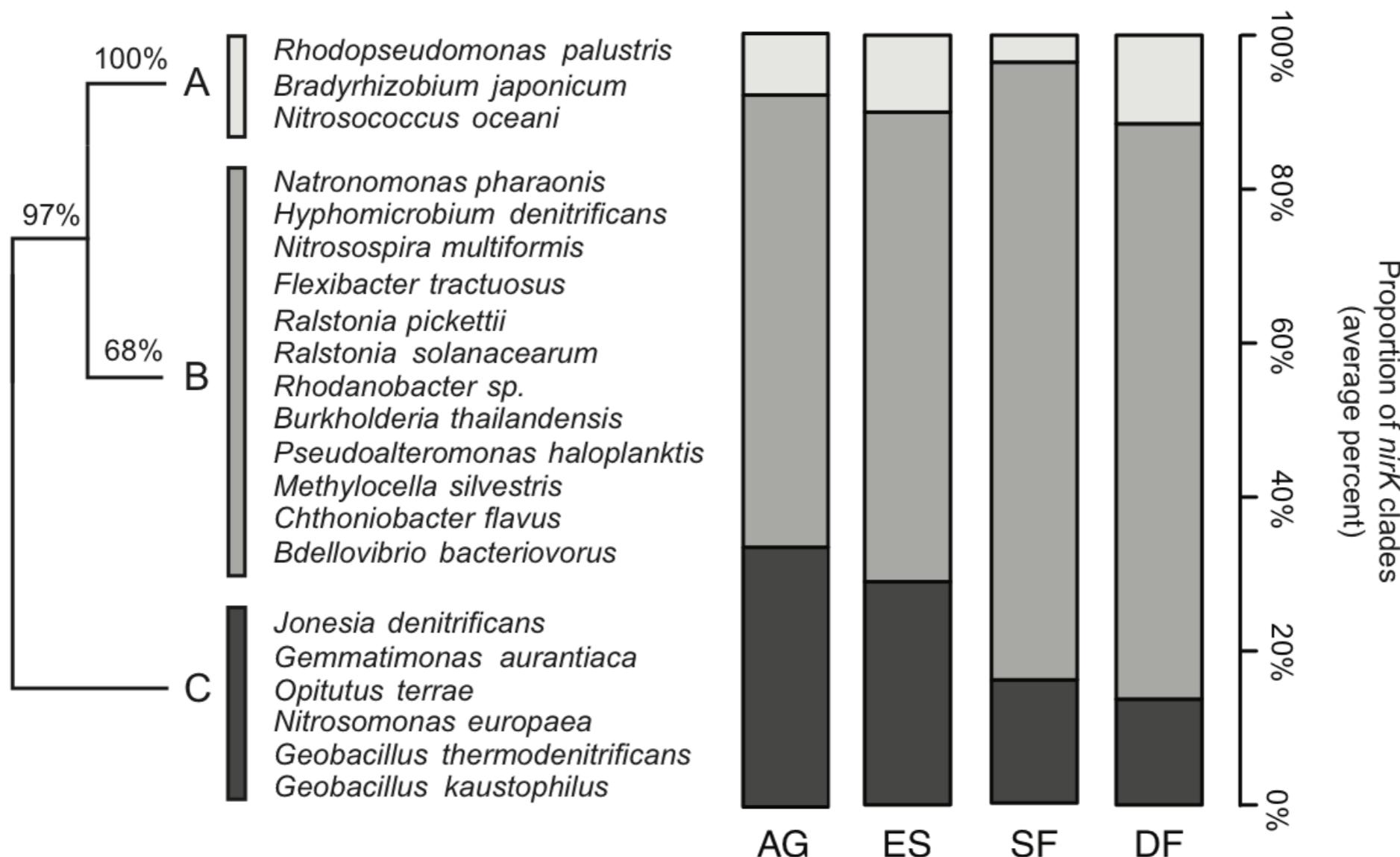
# More denitrification potential in Ag soils



# Denitrifier composition also changes

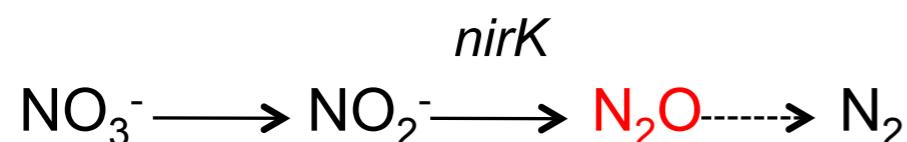


# High denitrifier diversity



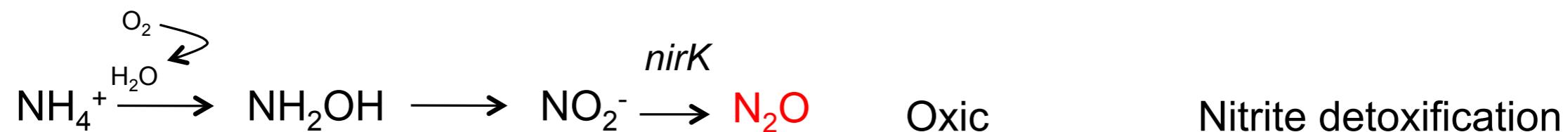
# Types of denitrification

## Heterotrophic denitrification

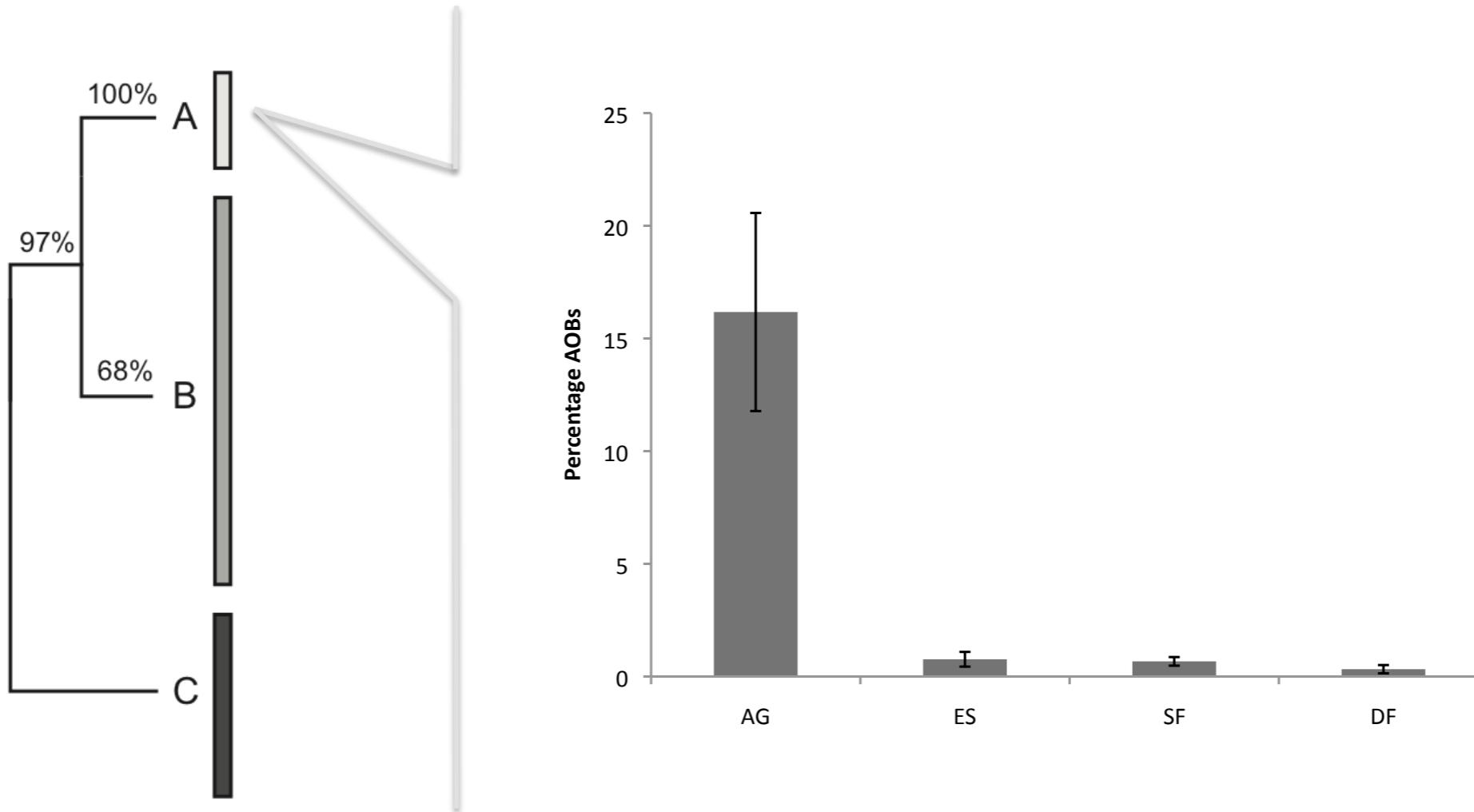


| Conditions | Function   |
|------------|------------|
| Anoxic     | Energetics |

## Autotrophic denitrification (AOBs)

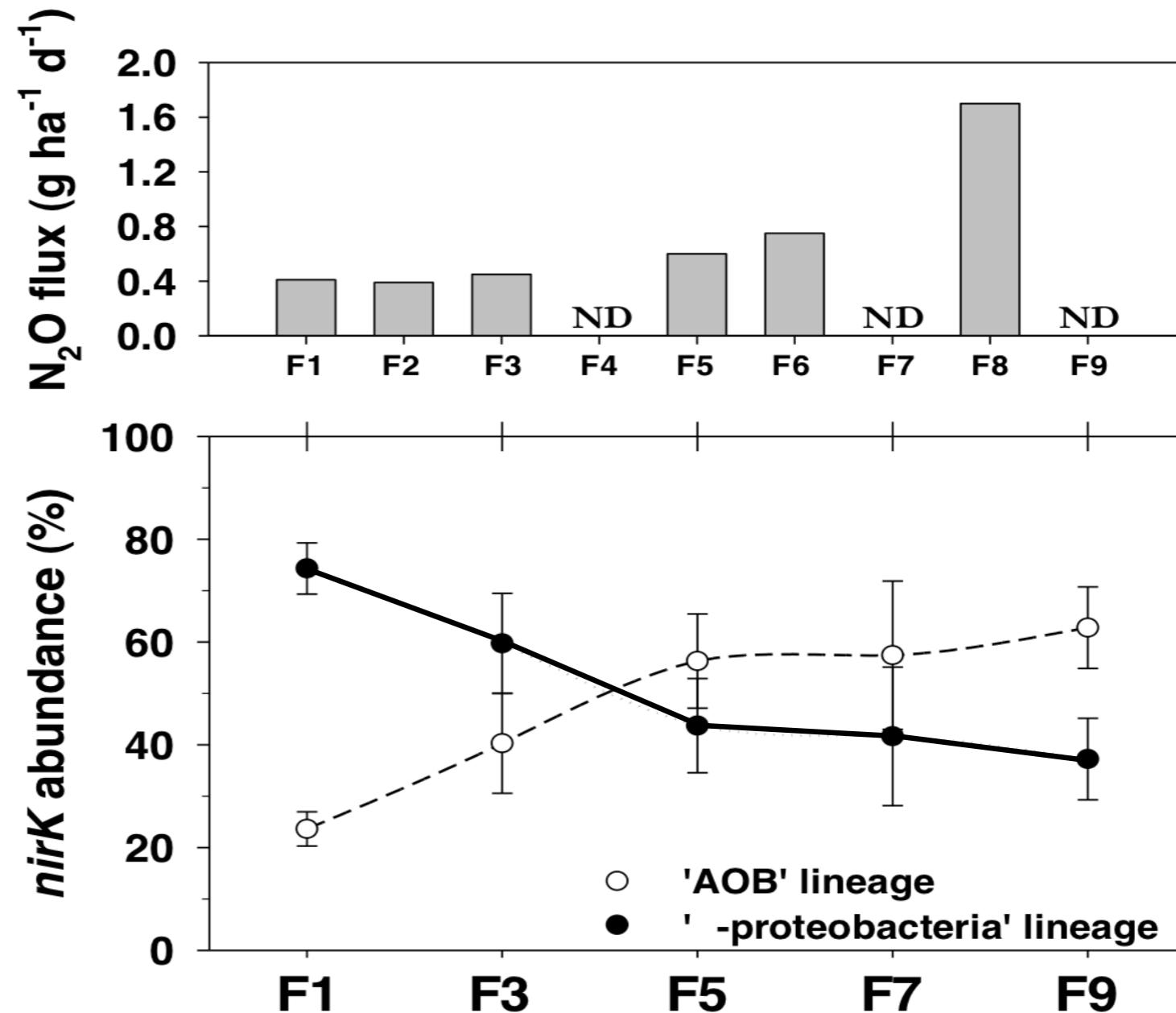
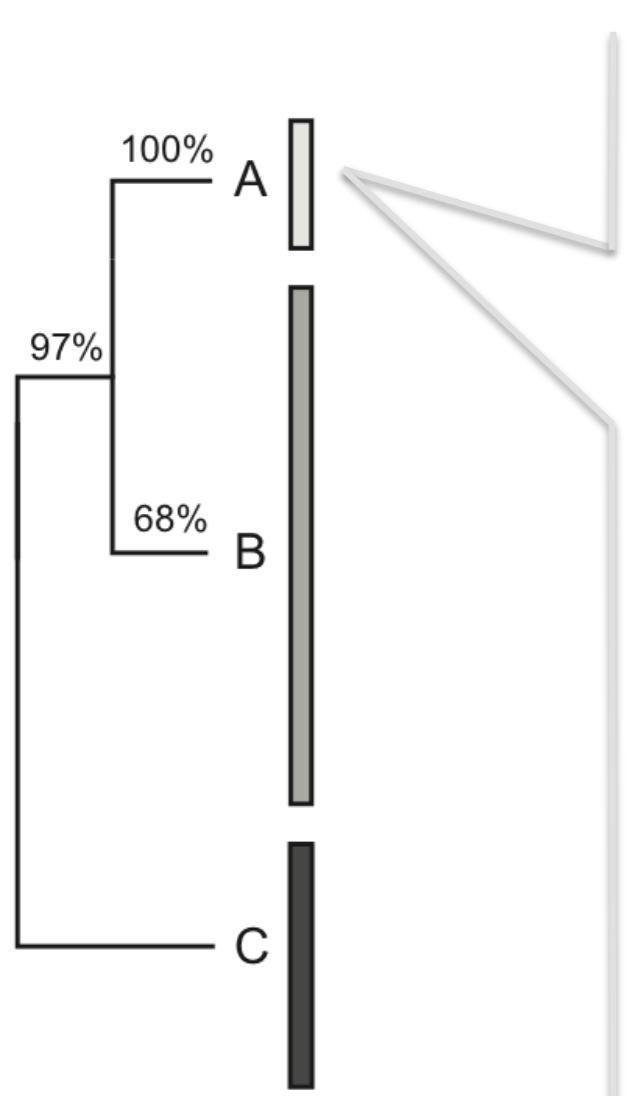


## AOB proportion increased significantly in AG

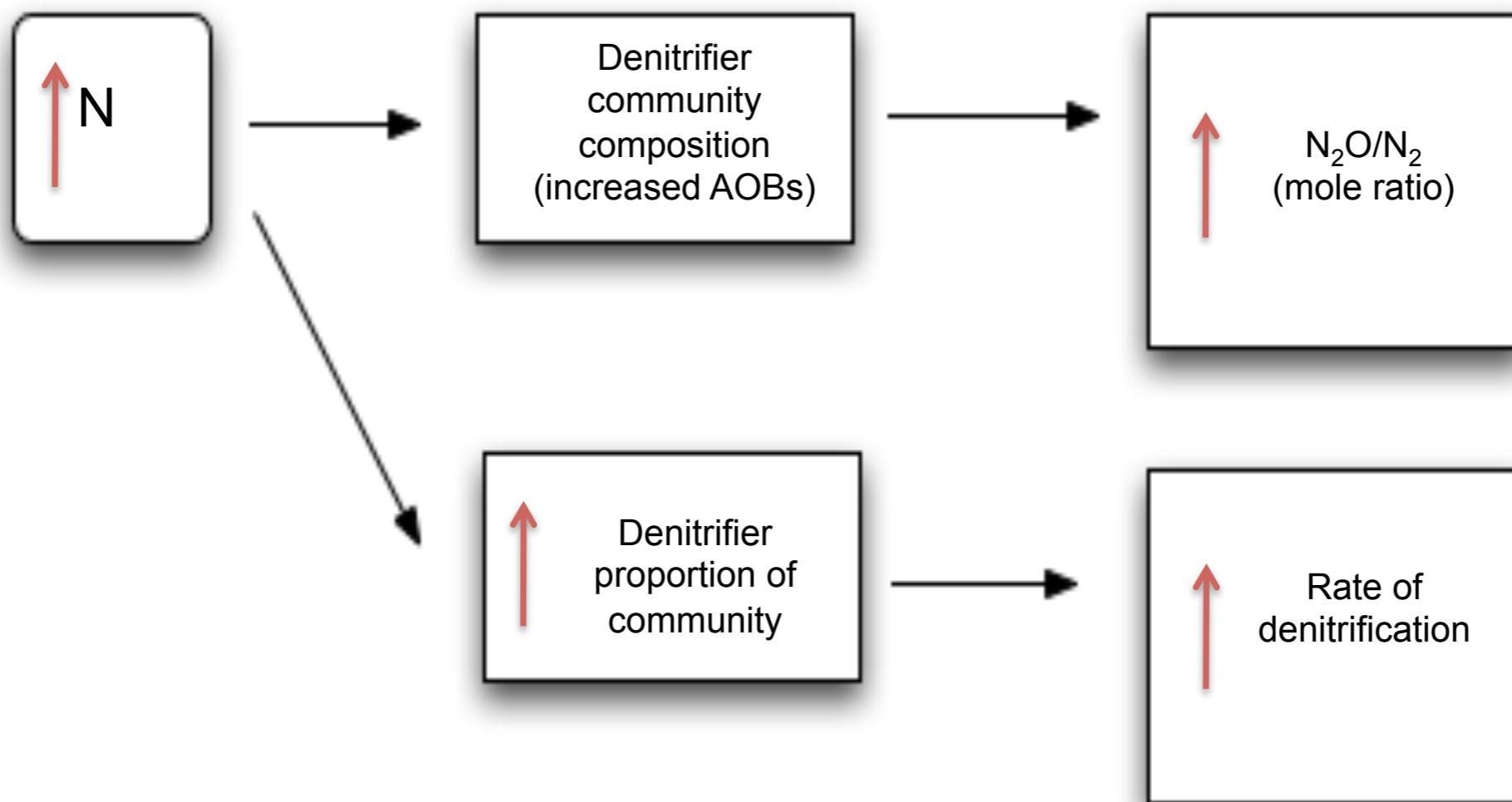


# AOB percentage correlates with fertilization

30

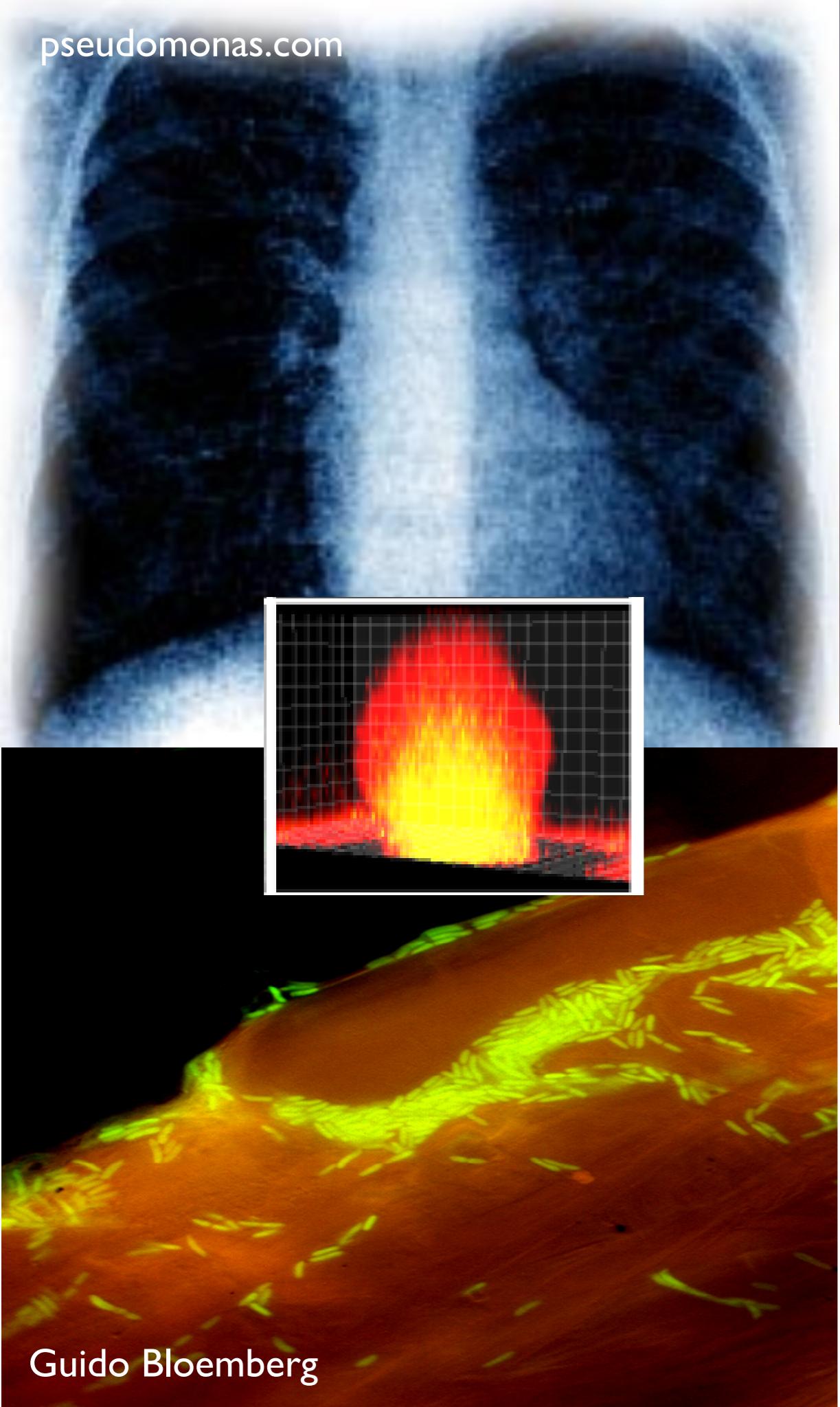


# Microbial communities in agriculture are poised for denitrification



# What is the relationship between denitrifying bacteria and N<sub>2</sub>O in agricultural soils ?

- Denitrifier abundance and composition changes with agricultural management. Communities more diverse than previously thought and poised for denitrification.
- May be potential for microbial mediation



Guido Bloemberg



Filtralite

# Acknowledgments

Dr. Dianne Newman

Dr. Barbara Wold

The Newman & Wold Labs

Doug Lies

Lars Dietrich



# Dr. Tom Schmidt



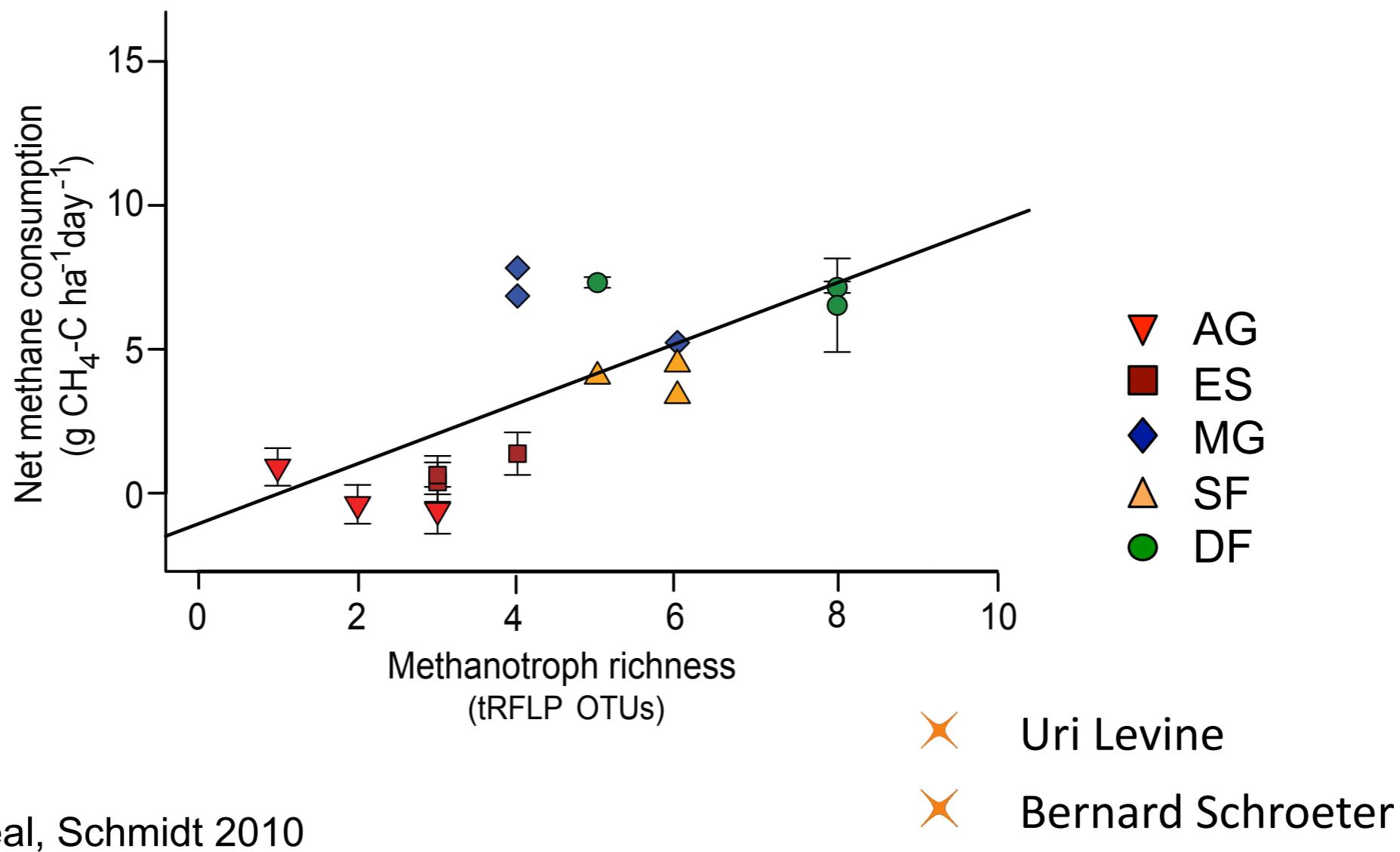
- ✖ Soil team
  - Vicente Gomez-Alvarez
  - Uri Levine
  - Keara Towery
  - Bernard Schroeter
  - John Dover
  - Zarraz Lee
  - Brendan O'Neill
  - Ben Roller
  - Tom Schmidt

- ✖ Funding
  - ✖ DOE GLBRC
  - ✖ NSF
  - ✖ NSF Postdoctoral Fellowship



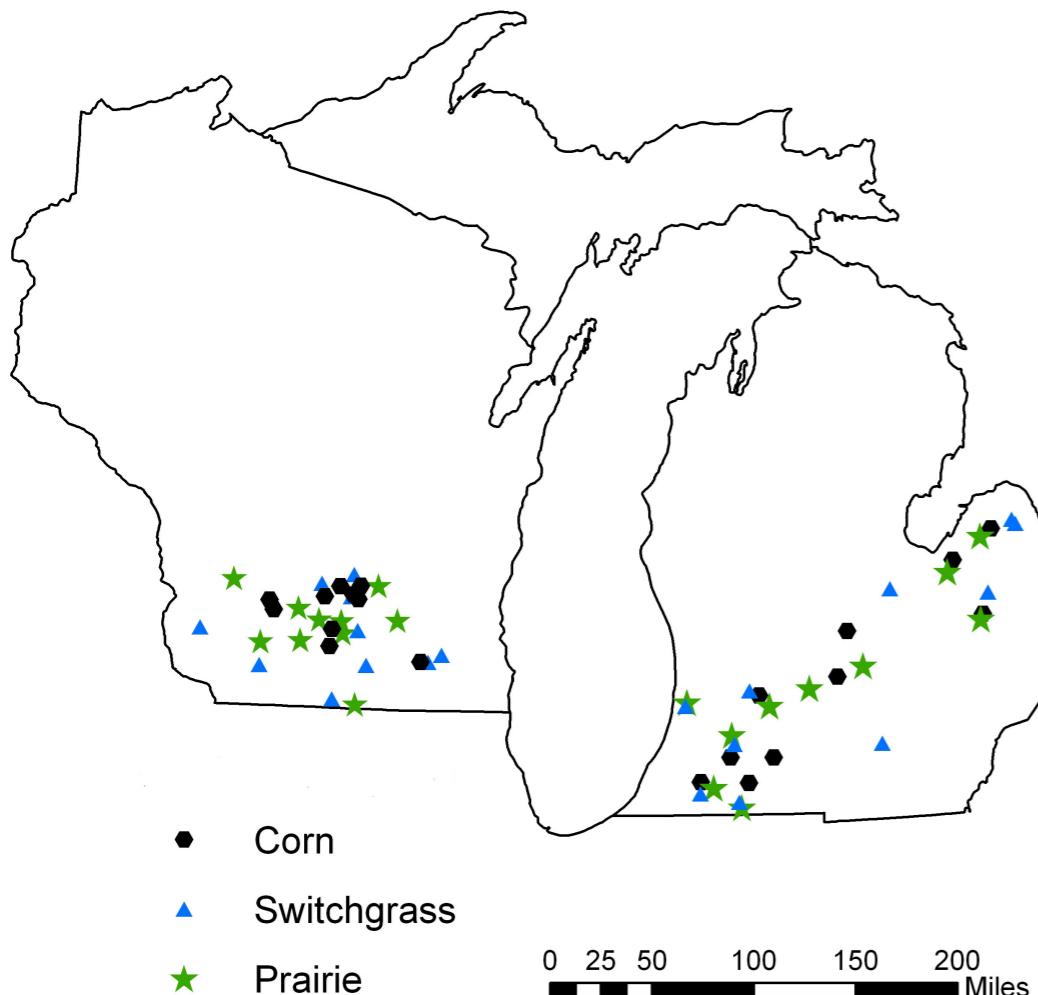


# Methanotroph richness correlates with methane consumption



# Methane in biofuel crops

GLBRC Extensive Sites



✖ Clone libraries

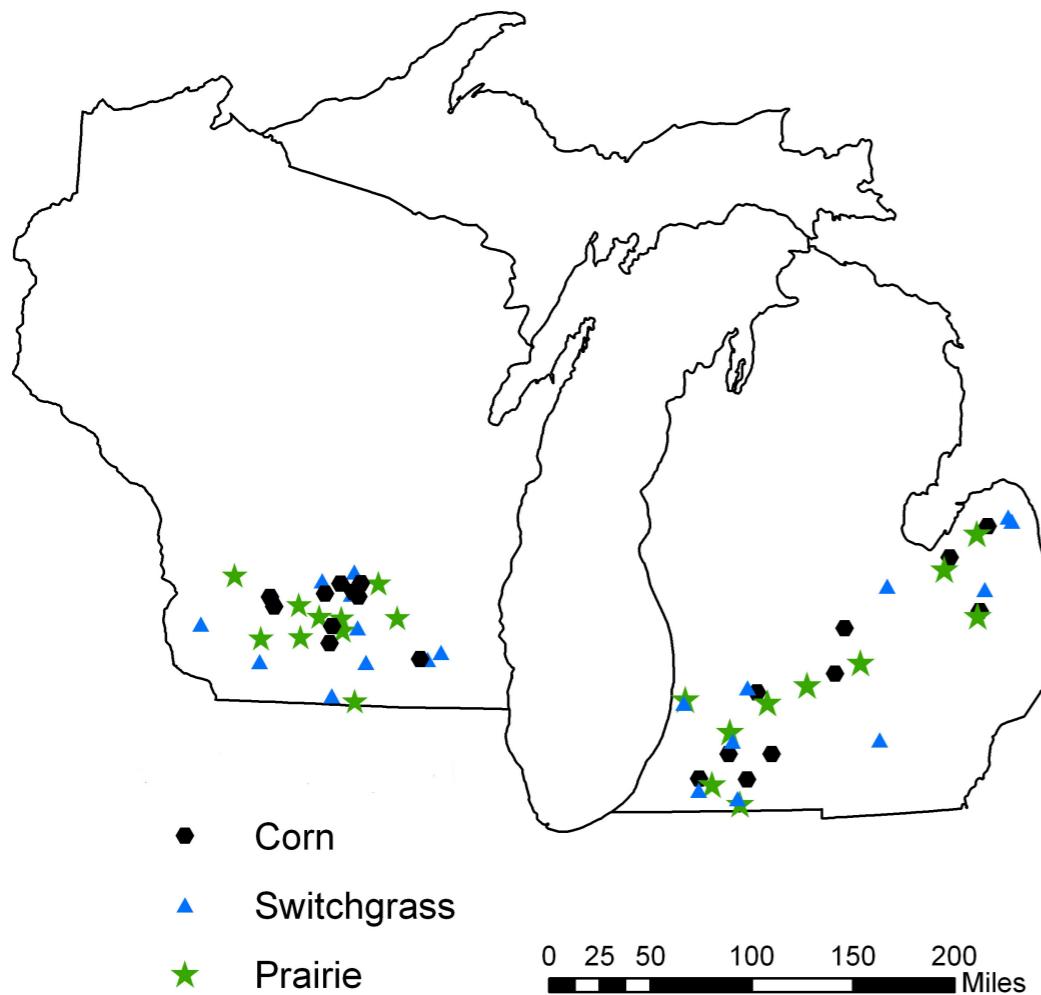
✖ Classification of *pmoA* / *amoA*

GFClassify –  
Ribosomal Database Project  
Jordan Fish

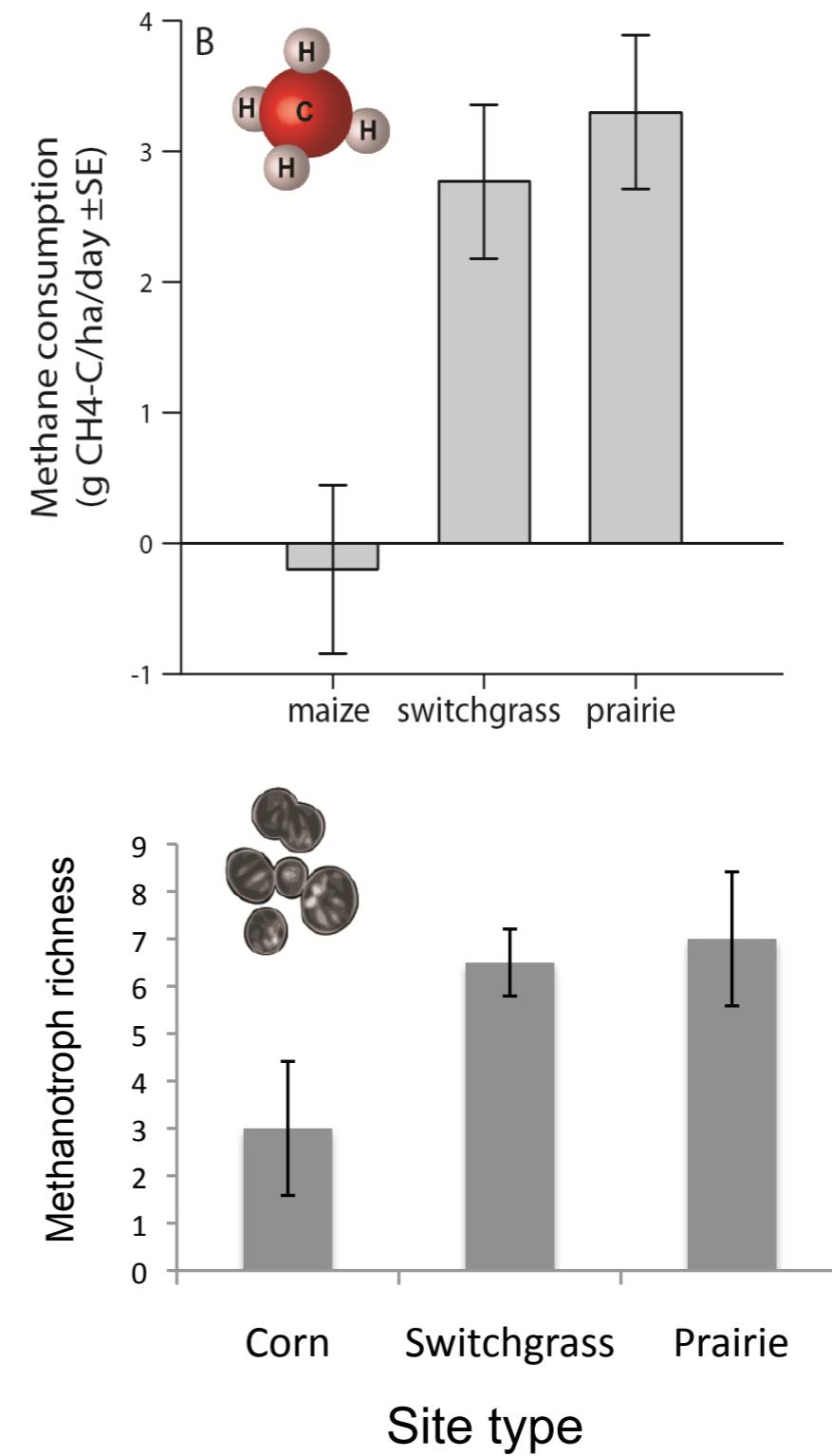
✖ Leilei Ruan and G. Philip Robertson

# Methane in biofuel crops

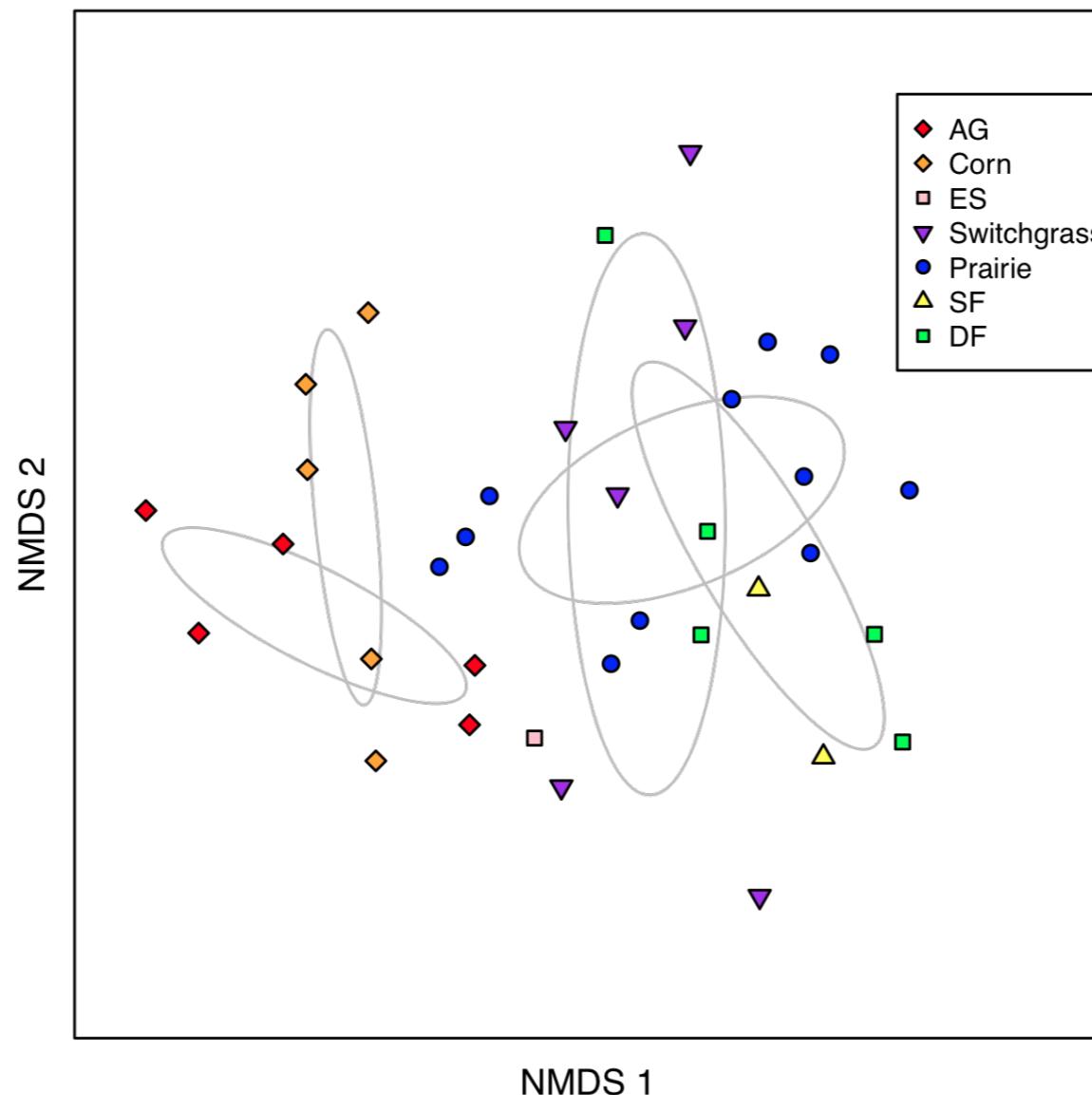
GLBRC Extensive Sites

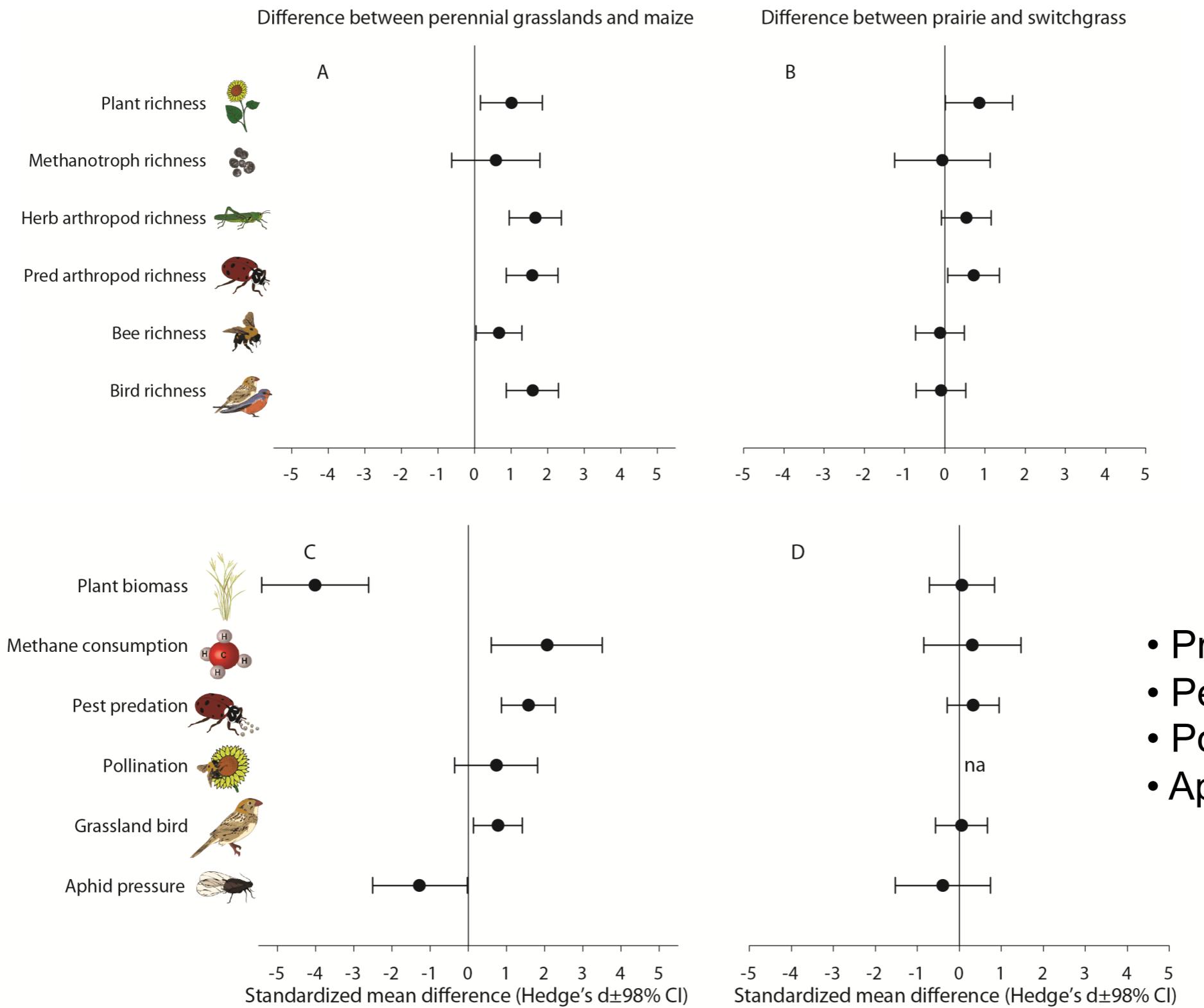


✗ Leilei Ruan and G. Philip Robertson



# Methanotroph composition changes with agricultural management





- Productive
- Perennial
- Polycultures
- Appropriate Placement

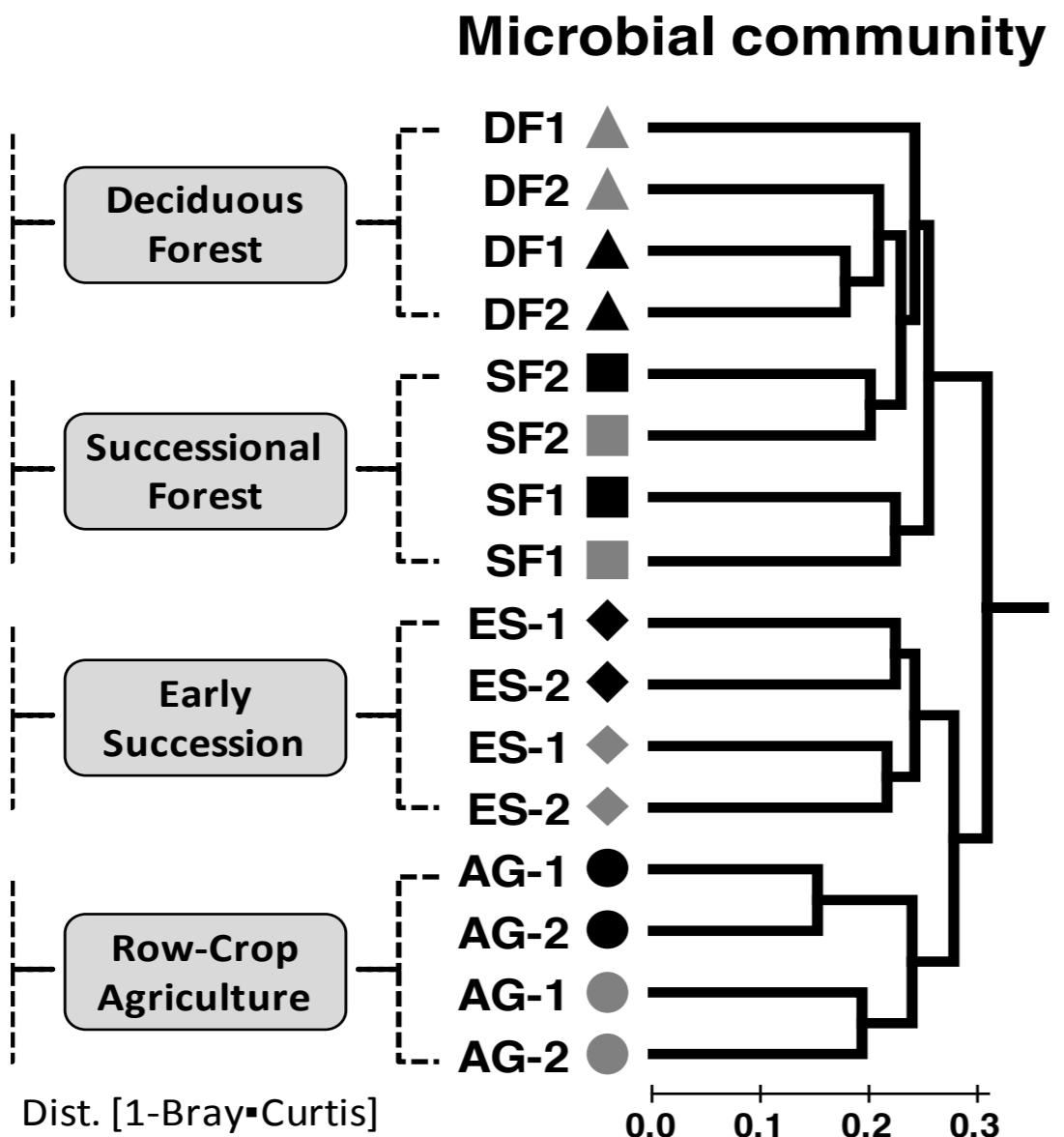
Werling et al, submitted

## **What is the relationship between denitrifying bacteria and N<sub>2</sub>O, and methane oxidizing bacteria and CH<sub>4</sub> in upper Midwest soils?**

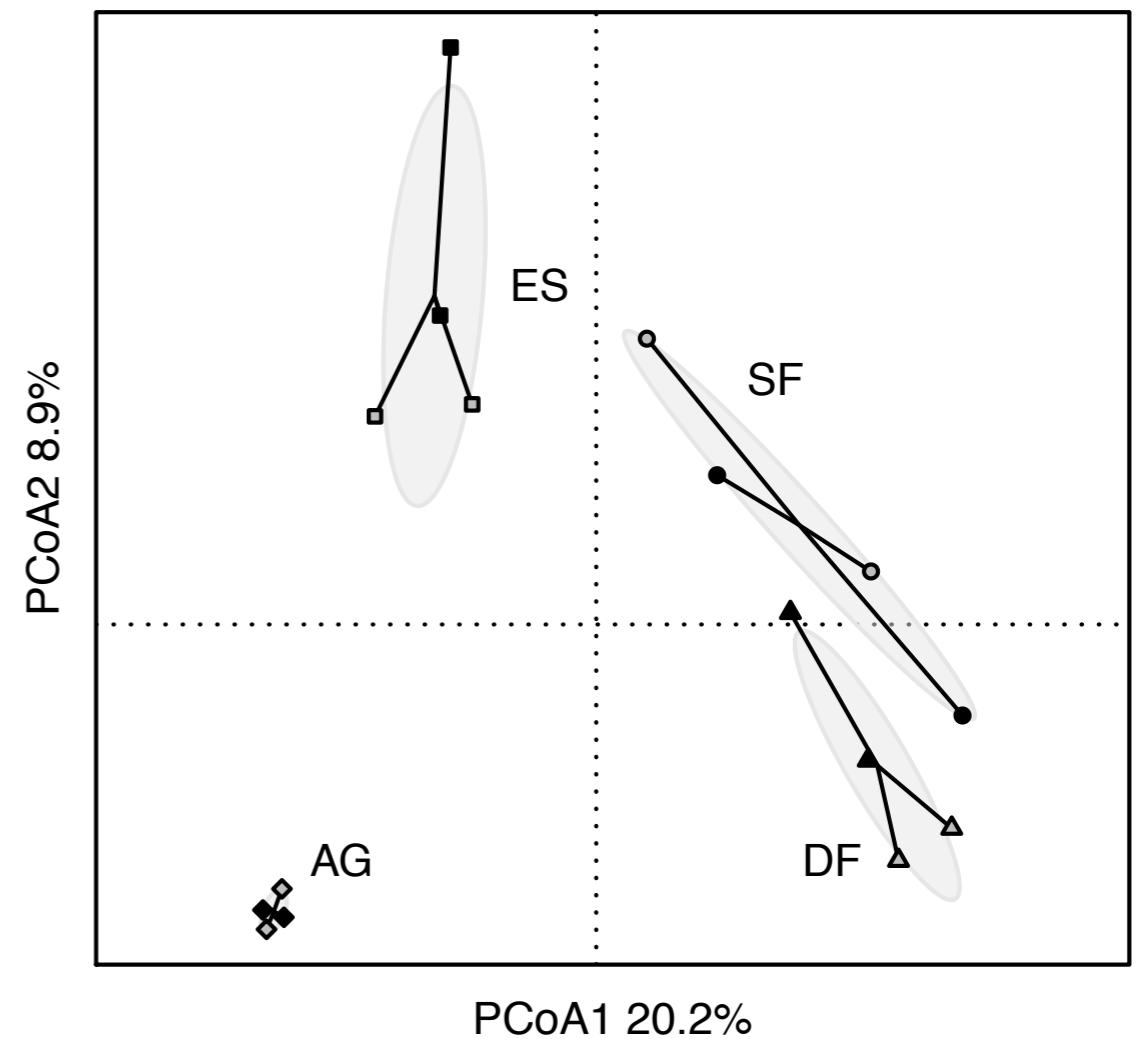
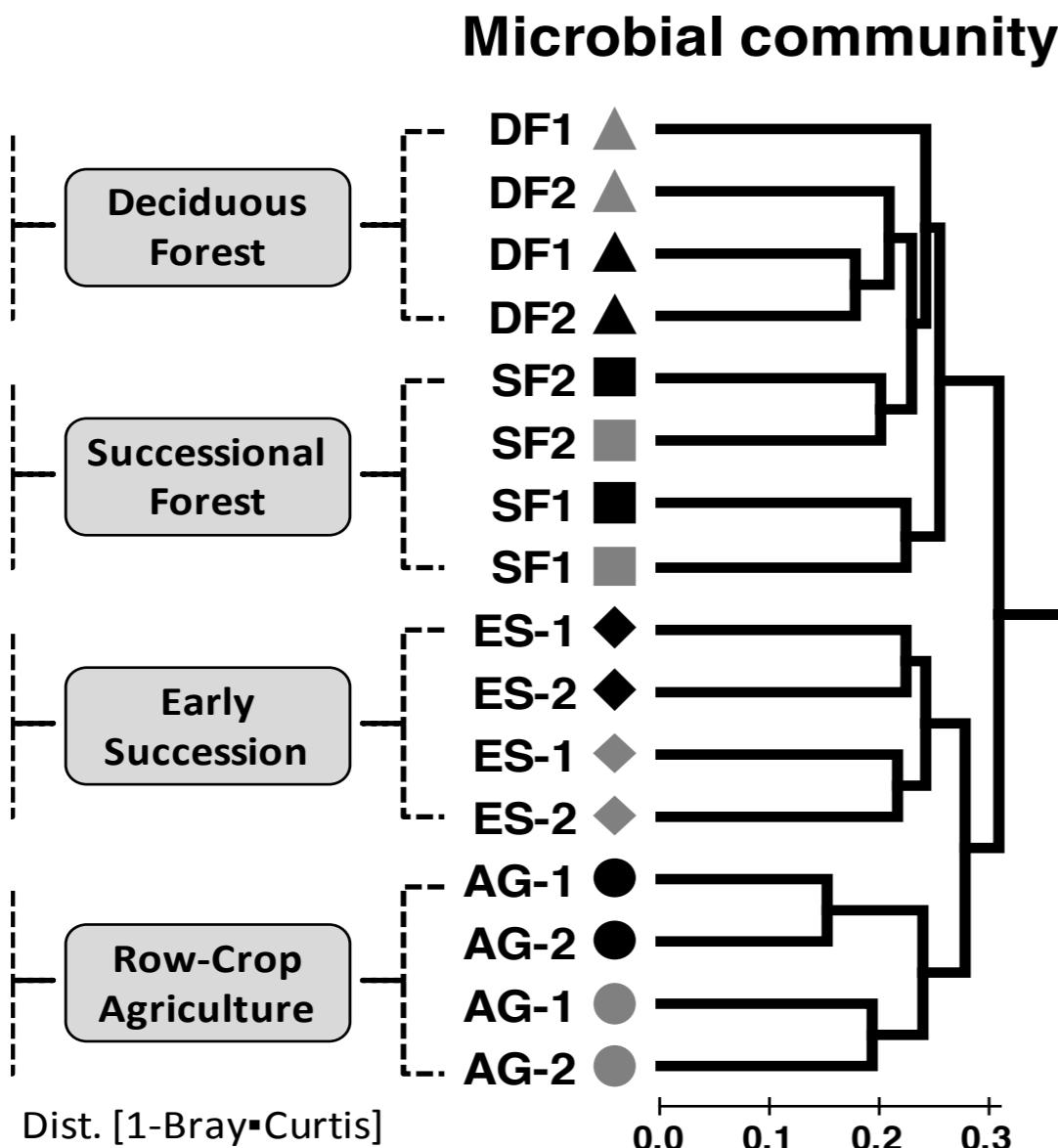
- ✖ Denitrifier abundance and composition changes with agricultural management.  
Communities more diverse than previously thought and poised for denitrification.
- ✖ Methanotroph richness and composition also changes with agricultural management and correlates with methane consumption
- ✖ May be potential for microbial mediation

**How long does it take for a community to recover from agriculture? Do all groups recover in the same way?**

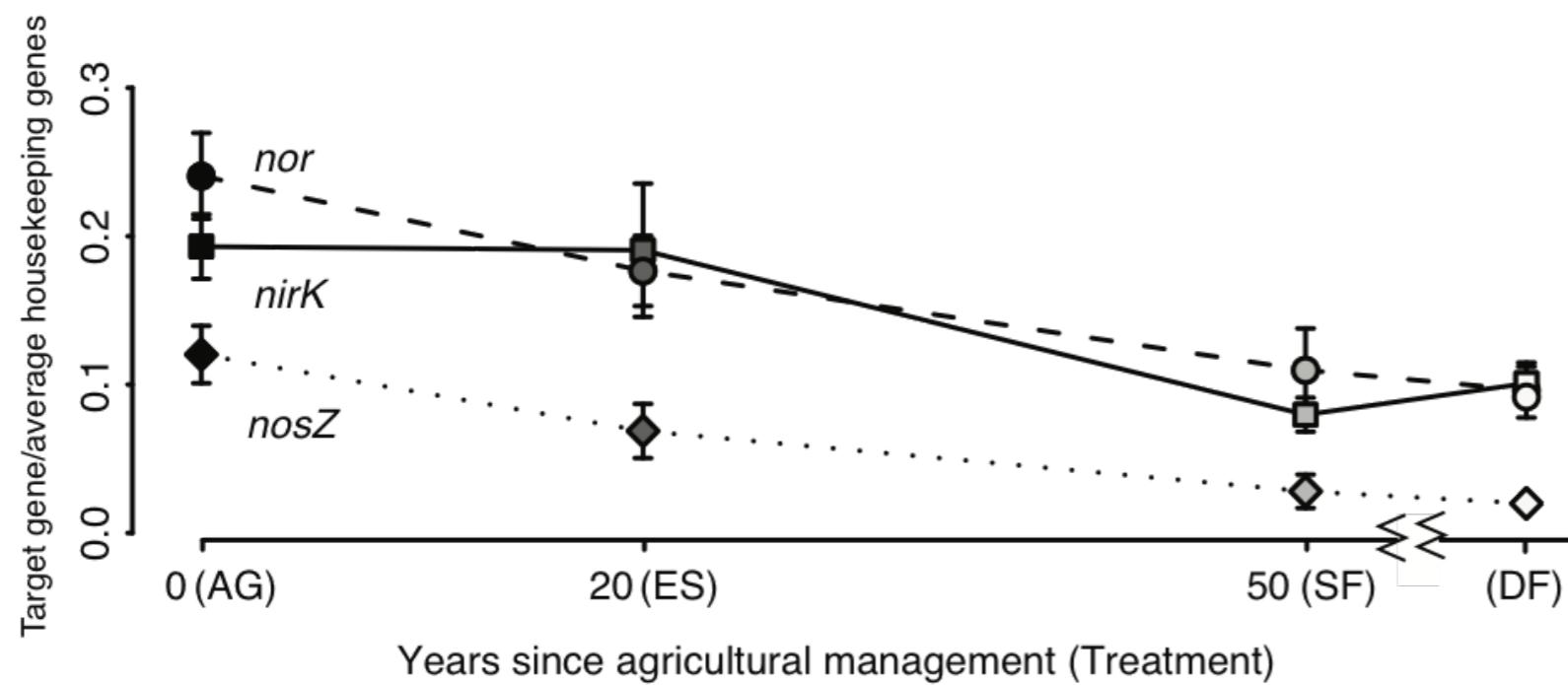
# More than 20 year recovery for bacterial community



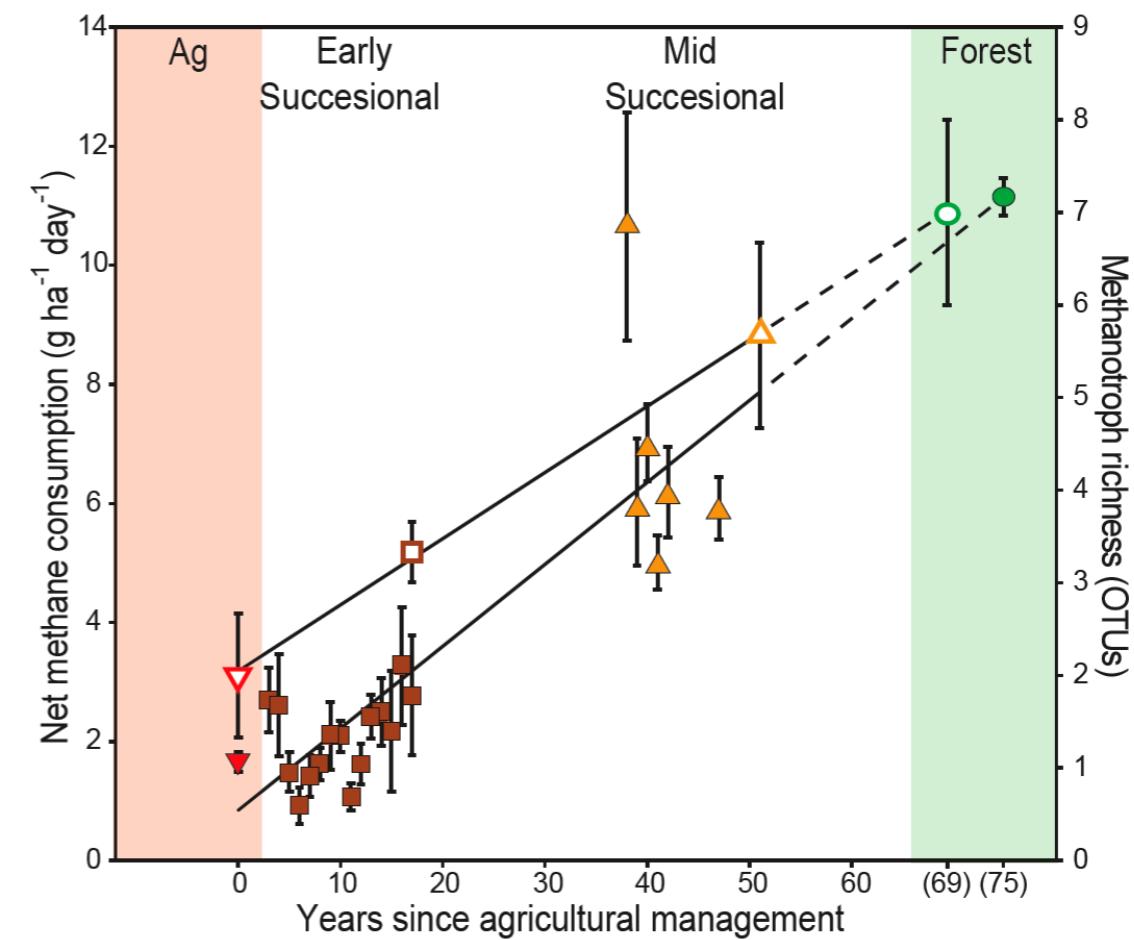
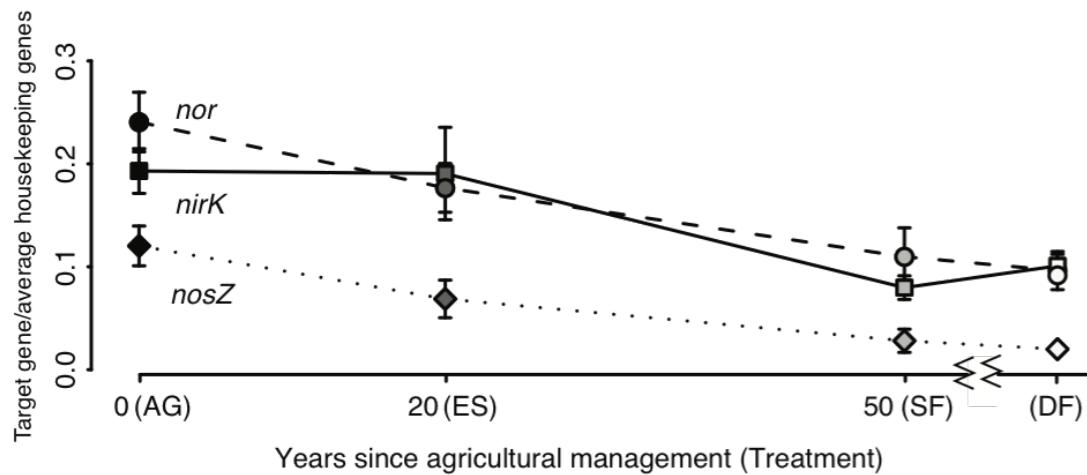
# More than 20 year recovery for bacterial community



## More than 20 year recovery for denitrifiers



# Methanotrophs take more than 40 years to recover



# Properties contributing to resilience

1. Responsive, regulatory feedbacks

Single-species *Shewanella oneidensis* biofilm

2. Diversity and redundancy

Soil microbial communities and greenhouse gas fluxes

3. Modular networks

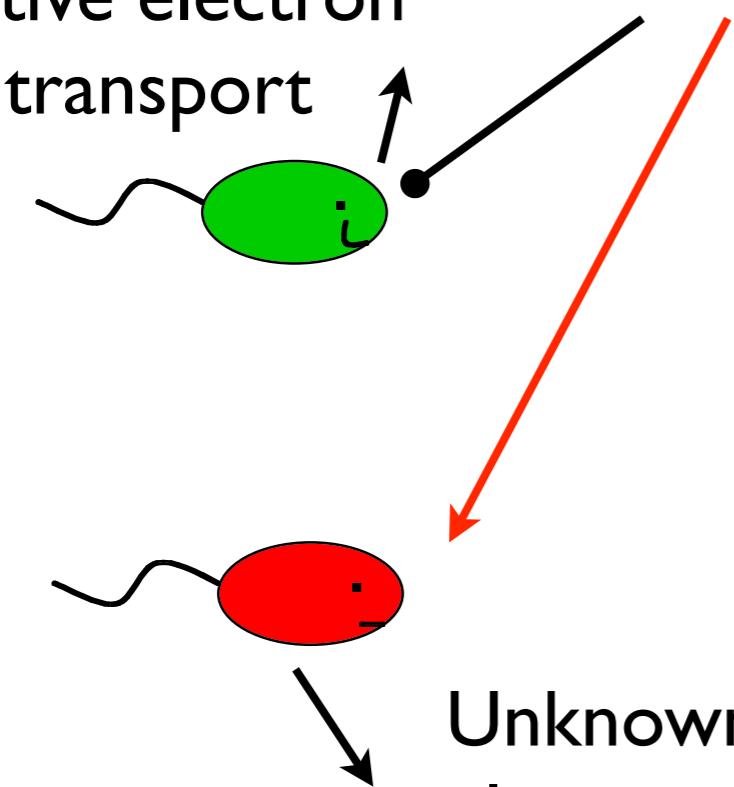
Future directions

# Live/Dead stain may not be accurate

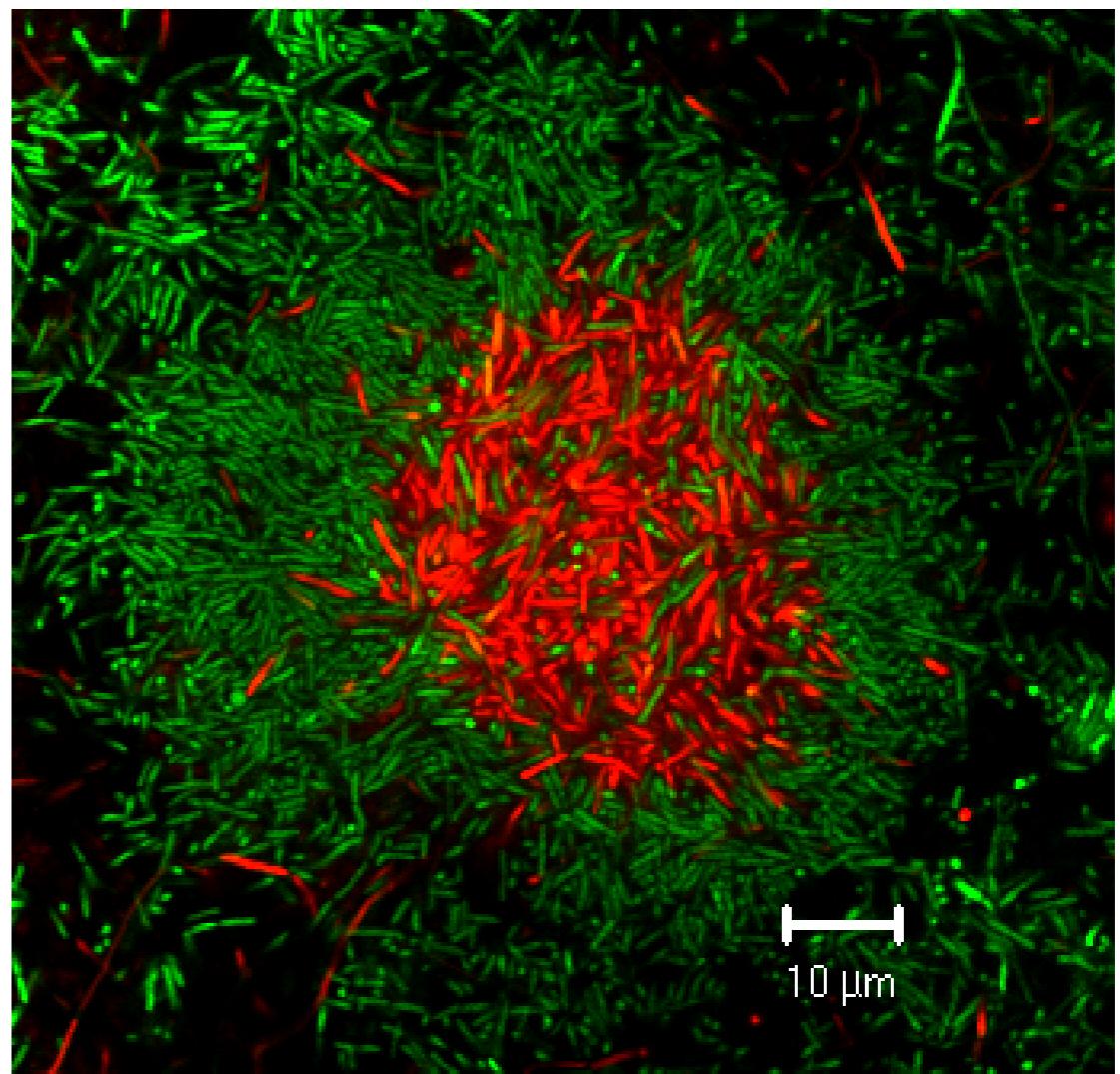
Dead stain

Propidium iodide

Active electron transport

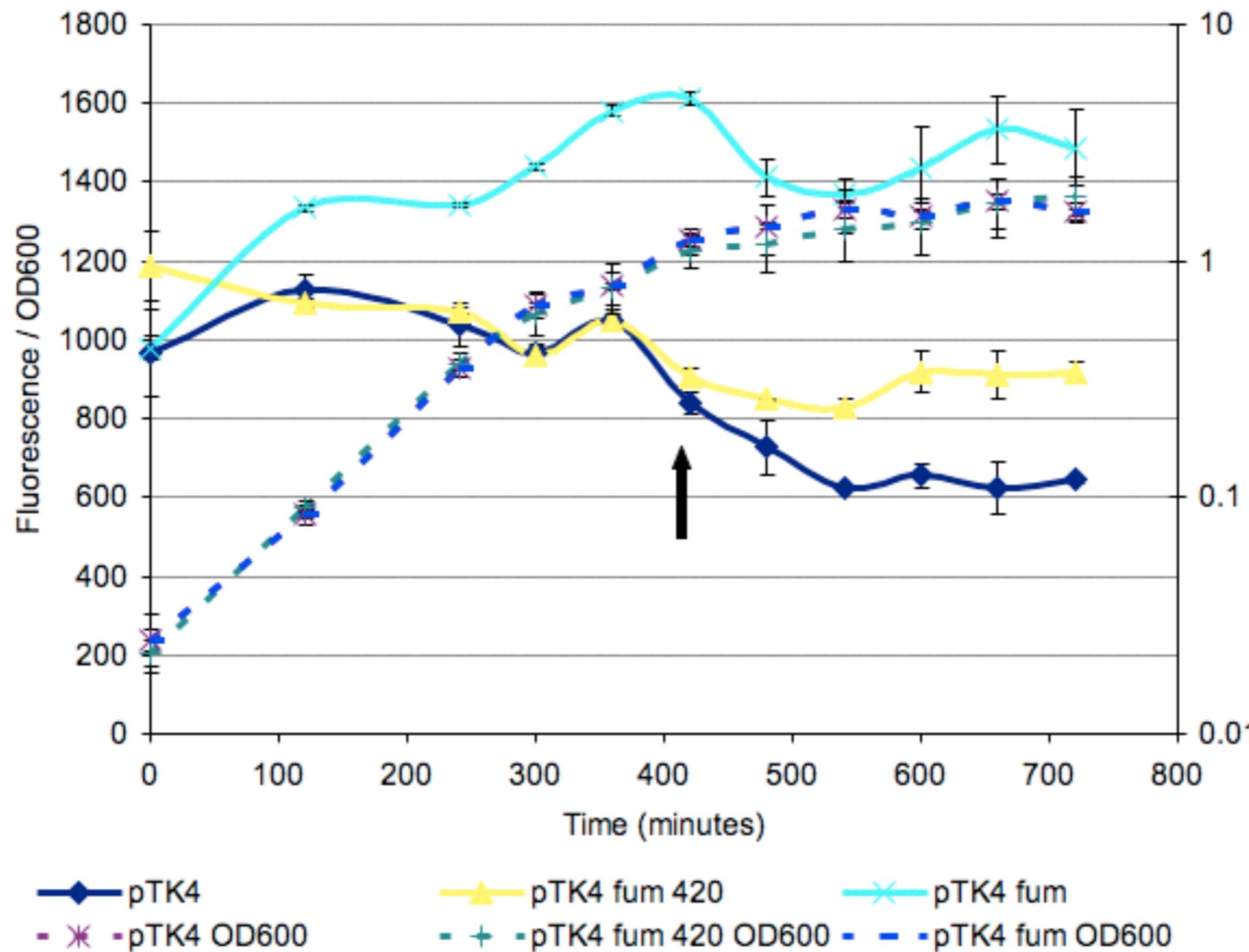


Unknown levels of electron transport

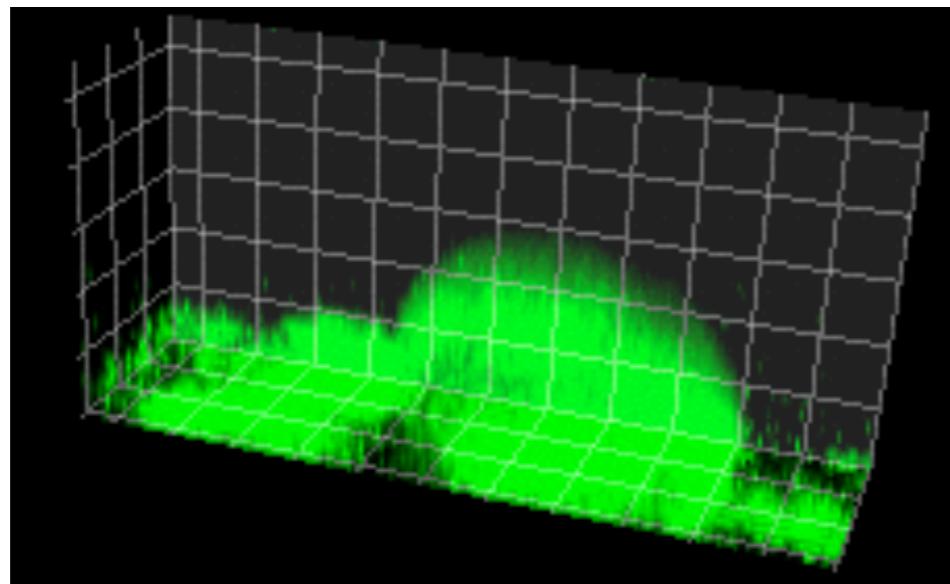


Teal et al, 2006

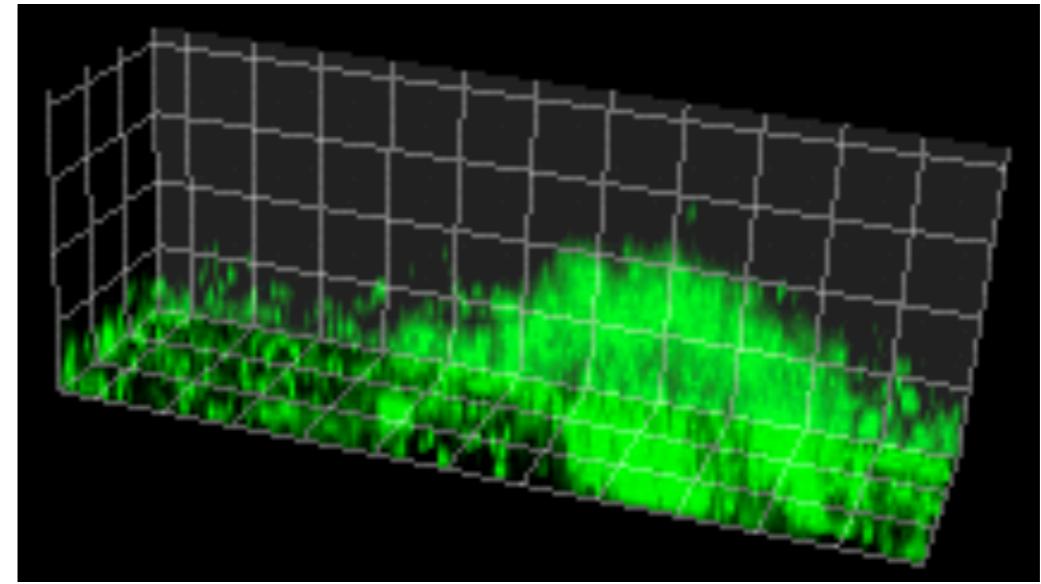
# Addition of an alternative electron acceptor affects cell growth state



# Addition of an electron donor does not have the same effect



Before



20mM lactate

Colony size not significantly different, but intensity is lower due to decreased number of cells