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# Modelling the transmission dynamics of dengue in the presence of *Wolbachia*



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#### ABSTRACT

Use of the bacterium Wolbachia is an innovative new strategy designed to break the cycle of dengue transmission. There are two main mechanisms by which Wolbachia could achieve this: by reducing the level of dengue virus in the mosquito and/or by shortening the host mosquito's lifespan. However, although Wolbachia shortens the lifespan, it also gives a breeding advantage which results in complex population dynamics. This study focuses on the development of a mathematical model to quantify the effect on human dengue cases of introducing Wolbachia into the mosquito population. The model consists of a compartment-based system of first-order differential equations; seasonal forcing in the mosquito population is introduced through the adult mosquito death rate. The analysis focuses on a single dengue outbreak typical of a region with a strong seasonally-varying mosquito population. We found that a significant reduction in human dengue cases can be obtained provided that Wolbachia-carrying mosquitoes persist when competing with mosquitoes without Wolbachia. Furthermore, using the Wolbachia strain WMel reduces the mosquito lifespan by at most 10% and allows them to persist in competition with non-Wolbachia-carrying mosquitoes. Mosquitoes carrying the WMelPop strain, however, are not likely to persist as it reduces the mosquito lifespan by up to 50%. When all other effects of Wolbachia on the mosquito physiology are ignored, cytoplasmic incompatibility alone results in a reduction in the number of human dengue cases. A sensitivity analysis of the parameters in the model shows that the transmission probability, the biting rate and the average adult mosquito death rate are the most important parameters for the outcome of the cumulative proportion of human individuals infected with dengue.

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#### 1. Introduction

Dengue is a viral vector-borne disease which is a major public health concern in many tropical and subtropical regions worldwide. It is estimated that around two-thirds of the world's population lives in dengue-endemic regions, and approximately 50–100 million cases occur each year [1]. Due to a high rate of asymptomatic cases, this is an underestimate of the actual number of cases, with a recent estimate being approximately three times this number [2]. Individuals with non-haemorrhagic dengue show symptoms such as high fever, severe headache and vomiting, which last for 2–7 days [1]. If they have the more severe form, dengue haemorrhagic fever (DHF), those symp-

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toms above are followed, after 3–7 days by rapid breathing, bleeding gums and fatigue [1].

Historically, four serotypes of virus that cause dengue have been identified, namely DEN1, DEN2, DEN3, DEN4, although recently potential fifth strain has been discovered [3]. An infected individual gains lifelong immunity to the serotype they are exposed to, but only temporary immunity to the other strains. If they are reinfected with a different serotype, they then have a higher chance of developing the more severe forms of dengue, dengue haemorrhagic fever or dengue shock syndrome. This effect is related to the "antibody dependent enhancement" [4]. This makes the development of vaccines against dengue very challenging as they must protect against all five strains simultaneously, however some progress is being made [5,6].

Dengue is transmitted to humans via the bite of an infectious mosquito. When dengue infects a mosquito, it requires a period of time to replicate and disseminate throughout the insect before it can be transmitted to susceptible humans: this period is called the extrinsic incubation period (EIP), and ranges between 5 and 15 days

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[7]. The closer the mosquito lifespan is to the EIP, the smaller the chance that dengue will be transmitted. There are two mosquito species responsible for the transmission of dengue, *Aedes aegypti* and *Aedes albopictus*. Of these, *Aedes aegypti* is the most competent vector. It is highly anthropophilic, preferring to obtain its blood meals from humans, and prefers artificial water containers such as pots, rain-water containers and discarded tyres for breeding sites, which are most available in areas where humans live. The lifespan of *Aedes aegypti* varies depending on environmental factors such as temperature, humidity and rainfall [8], which leads to fluctuations of the mosquito population in regions where these factors are seasonal.

In the past, a number of strategies for controlling dengue by reducing the mosquito population have been implemented, but they are largely unsustainable [9–12]. For example, the use of insecticides becomes ineffective as mosquitoes develop resistance to particular insecticides [9,10]. The removal of mosquito breeding sites has also been carried out but is often unsustainable since it has to be repeated often, as has been found in Cairns and Singapore [11,12]. These problems highlight a critical need for a novel and sustainable strategy to break the cycle of dengue transmission.

An innovative approach for controlling dengue is introducing the *Wolbachia* bacterium into *Aedes aegypti* mosquitoes [13–17]. *Wolbachia* naturally infects an estimated 66% of insects [18], including *Aedes albopictus*, but is not endemic in *Aedes aegypti*. There are two strains of *Wolbachia* that can be introduced into *Aedes aegypti*, *WMel* and *WMelPop* [15,16], with different effects that are further discussed in Section 2.2.2. *Wolbachia* can reduce dengue transmission via two mechanisms. First, *Wolbachia* inhibits viral replication and dissemination in the dengue vector [13], and, consequently, the ability of *Wolbachia*-carrying mosquitoes to transmit dengue viruses is reduced. Secondly, *Wolbachia* reduces the lifespan of the mosquitoes [14,15] and as a result this may be close to, or less than, the EIP, and so there is less time for the mosquito to transmit dengue to susceptible humans.

This strategy will only be effective against dengue if the Wolbachia-carrying mosquitoes can compete successfully with Wolbachia-free ones and therefore persist in the wild. Cytoplasmic incompatibility (CI) is a feature of Wolbachia infection that is important for population invasion and persistence, since it gives Wolbachia-carrying females a reproductive advantage [19,20]. CI results in Wolbachia-carrying females being able to reproduce successfully when mating with either Wolbachia-free or Wolbachia-carrying males, whereas non-Wolbachia-carrying females can only reproduce successfully when mating with non-Wolbachia-carrying males. Moreover, non-Wolbachia-carrying females produce an embryo upon mating with Wolbachia-carrying males, but the embryo is not viable and dies, thereby effectively blocking reproduction. CI therefore leads to complex dynamics for the mosquito population in the presence of Wolbachia, and so also affects the dynamics of dengue transmission.

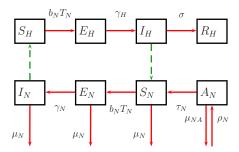
To study the population dynamics of Wolbachia-carrying mosquitoes, several mathematical models have been developed [21–24]. Hancock et al. [21] developed a mathematical model to explore the host population dynamics and Wolbachia infection frequency with both single and multiple introductions ("seeding") of Wolbachia-carrying mosquitoes. The same authors also developed a metapopulation model to explore the spatial dynamics and found that spatial variation in the density-dependent competition has an effect on the spread of Wolbachia infection [22]. Chan and Kim [23] developed a mathematical model incorporating both slow and fast dispersal situations. They found that temperature affects the speed of Wolbachia invasion and that the death rate of Wolbachia-carrying mosquitoes influences their persistence. Ndii et al. [24] developed a mathematical model of mosquito population dynamics by including the effect of CI in the mating function and competition for resources in the aquatic stage. They found parameter ranges where Wolbachiacarrying mosquitoes persist, and showed that the steady state where Wolbachia-carrying mosquitoes alone persist only exists when maternal transmission is perfect. They showed that there is a stable steady state where Wolbachia-free and Wolbachia-carrying mosquito populations can coexist, and this was also found by Chan and Kim [23]. Therefore, throughout the paper we refer to 'absence' and 'presence' of Wolbachia, wherein non-Wolbachia-carrying mosquitoes persist in both cases. The body of research above suggests that it is possible for Wolbachia-carrying mosquitoes to persist in competition with non-Wolbachia-carrying mosquitoes, and hence to reduce the transmission of dengue to humans. The question that then arises is to what extent Wolbachia infection can reduce human dengue cases. In this paper, a mathematical model for dengue transmission between mosquitoes and humans with underlying Wolbachia mosquito population dynamics is developed to quantify the effect of Wolbachia on dengue transmission in a human population. We determine the possible effectiveness of using Wolbachia as a dengue intervention, but not its relative worth compared to other control measures.

A number of mathematical models have been developed to understand the transmission dynamics of dengue (for example, see [25-33]) and a review of different dengue models was conducted by Andraud et al. [34]. However, only a few mathematical models for dengue transmission with an underlying Wolbachia-carrying mosquito population exist [32,33]. Hughes and Britton [32] analysed dengue reduction for situations when only non-Wolbachia-carrying mosquitoes persist and when only Wolbachia-carrying mosquitoes persist, and did not present the situations when both populations persist. Hancock et al. [33] developed a model for vector-borne pathogen dynamics in the presence of Wolbachia-carrying mosquitoes. However, this model was not specifically developed for dengue transmission dynamics and its aim was to investigate the effect of releasing more Wolbachia-carrying male mosquitoes. In this paper, we develop a mathematical model for dengue with underlying seasonal mosquito population dynamics in the presence of Wolbachia-carrying mosquitoes. Unlike the model of Hughes and Britton [32] and Hancock et al. [33], here we consider seasonality using a sinusoidal forcing function and we also include competition for resources in the aquatic stage. Seasonal effects are included because they are appropriate for regions with a strong seasonal climate, such as Far North Queensland in Australia, which has distinct wet and dry periods: this is where the Wolbachia field trials are being conducted [16]. We study a single dengue outbreak over the course of 31 weeks, using data from 2008 [35]. This allows us to make a number of simplifying assumptions, including having only a single strain of dengue and ignoring human migration and demographics. An exploration of key parameter ranges is conducted to quantify the level of dengue reduction obtained from a Wolbachia intervention and to study the persistence of Wolbachia-carrying mosquitoes over possible parameter ranges.

The remainder of this paper is organised as follows. In Section 2 we describe the model in the absence of *Wolbachia*, the model in the presence of *Wolbachia* and the sensitivity analysis methodology. The results are presented in Section 3. Finally, discussion and conclusions are given in Section 4.

#### 2. Methods

In this section methods are presented that are later used to investigate the *Wolbachia* intervention. Firstly, a model of dengue transmission with humans and a seasonal mosquito population is constructed. Secondly, the model is adapted to include a *Wolbachia*-carrying mosquito population complete with the impact of cytoplasmic incompatibility on the population dynamics. Thirdly, the methodology for the parameter sensitivity analysis is presented, which is used to determine which parameters most influence the cumulative number of human infections.



**Fig. 1.** Schematic of the model in the absence of *Wolbachia*-carrying mosquitoes, Eqs. (1)–(8). Solid lines are population progression lines and dashed lines are disease transmission lines. The subscript H is for the human population and N is for the non-*Wolbachia* mosquito population. The compartments are 'S' for susceptible, 'E' for exposed to dengue but not yet infectious, 'I' for infectious, 'R' for recovered, and 'A' for the aquatic phase of the mosquito life cycle. The transition rates between compartments are shown next to the progression lines and are described in the text.

#### 2.1. Model in the absence of Wolbachia

#### 2.1.1. Model formulation

A mathematical model for the transmission dynamics of dengue between humans and mosquitoes in the absence of Wolbachia is developed. This model assumes homogeneous and well-mixed populations and is for one dengue strain only. The model comprises human and vector (mosquito) populations, with births and seasonally-forced deaths for the mosquito population. Since we are considering only a single outbreak with a duration of less than 1 year, human migration and demographics are not influential and hence omitted. As the ratio between male and female mosquitoes is approximately 1.02:1 [36], the number of male and female mosquitoes are taken to be the same. This model only explicitly considers female mosquitoes as they are responsible for dengue transmission, since only the female mosquito takes a blood meal from humans as part of their reproductive cycle. Male mosquitoes are implicitly included, since only half of the aquatic mosquitoes mature to be adult female susceptible mosquitoes. A schematic representation of the model is given in Fig. 1, with the subscript H for the human population and N for the non-Wolbachia-carrying mosquito population.

The human population is divided into four subpopulations, namely Susceptible  $(\hat{S}_H)$ , Exposed  $(\hat{E}_H)$ , Infectious  $(\hat{I}_H)$  and Recovered  $(\hat{R}_H)$ , where the hat is used to denote fully dimensional state variables. Furthermore, since we only consider a single outbreak with a time span on the order of a year, a constant human population size is assumed  $N_H = \hat{S}_H + \hat{I}_H + \hat{I}_H + \hat{R}_H$ , with no demography. The mosquito population is divided into subpopulations of Aquatic ( $\hat{A}_N$ , combining the egg, larval and pupal stages), Susceptible  $(\hat{S}_N)$ , Exposed  $(\hat{E}_N)$  and Infectious  $(\hat{I}_N)$ . The total adult female mosquito population is  $\hat{F}_N = \hat{S}_N + \hat{E}_N + \hat{I}_N$ . The subscript N is to denote non-Wolbachia-carrying mosquitoes and is used here for consistency and to differentiate from Wolbachiacarrying mosquitoes in later sections. We group eggs, larvae and pupae into one compartment as they are not involved in the transmission of dengue. Without loss of generality, they can be represented by a single death rate and a single maturation rate for the purpose of modelling the dengue transmission dynamics. No "recovered" class is required for mosquitoes as they remain infected for the rest of their life. The populations of both human and mosquitoes are converted to proportions by letting  $S_H = \hat{S}_H/N_H$ ,  $E_H = \hat{E}_H/N_H$ ,  $I_H = \hat{I}_H/N_H$ ,  $R_H = \hat{R}_H/N_H$ and  $A_N = \hat{A}_N/K$ ,  $S_N = \hat{S}_N/K$ ,  $E_N = \hat{E}_N/K$ , and  $I_N = \hat{I}_N/K$ . Since the carrying capacity, K, is related to the number of available breeding sites, which depends on the number of humans, we have  $K \propto N_H$ , and hence  $K = LN_H$ , where L is the ratio of the carrying capacity to the total human population. The model, with populations as proportions, is then governed by the system of differential equations,

$$\frac{dS_H}{dt} = -b_N T_N L I_N S_H,\tag{1}$$

$$\frac{dE_H}{dt} = b_N T_N L I_N S_H - \gamma_H E_H,\tag{2}$$

$$\frac{dI_H}{dt} = \gamma_H E_H - \sigma I_H,\tag{3}$$

$$\frac{dR_H}{dt} = \sigma I_H,\tag{4}$$

$$\frac{dA_N}{dt} = \rho_N \frac{F_N}{2} \left( 1 - A_N \right) - (\tau_N + \mu_{NA}) A_N, \tag{5}$$

$$\frac{dS_N}{dt} = \tau_N \frac{A_N}{2} - \left(b_N T_N I_H + \mu_N(t)\right) S_N,\tag{6}$$

$$\frac{dE_N}{dt} = (b_N T_N I_H) S_N - (\gamma_N + \mu_N(t)) E_N$$
(7)

and

$$\frac{dI_N}{dt} = \gamma_N E_N - \mu_N(t) I_N,\tag{8}$$

where  $S_H + E_H + I_H + R_H = 1$ . Since the probability of transmission from a non-Wolbachia mosquito to a human is the same as the converse, we also let  $T_{HN} = T_{NH} = T_N$ . The variation in the adult mosquito death rate is strongly influenced by environmental factors such as temperature, humidity, and rainfall [8] and is therefore sinusoidally forced, according to

$$\mu_N(t) = \mu_{N0} \left( 1 - \eta \cos \left( \frac{2\pi \left( t + \omega \right)}{365} \right) \right), \tag{9}$$

where  $\eta$  is the strength of seasonal forcing in the adult death rate, and  $\mu_{N0}$  is the average adult death rate, t is time and  $\omega$  is the phase shift, which is used to align the cosine function with the seasonal factors in Far North Queensland. Forcing more than one term in this model is unnecessary since the mosquito population is explicitly modelled. The mosquito population size is most sensitive to the death rate, and hence this parameter is chosen for the forcing, resulting in appropriate seasonal fluctuations in the adult mosquito population. Since the mating function is dependent on the population size, this in turn results in appropriate seasonal fluctuations in the aquatic population.

After being bitten by an infectious mosquito, humans become exposed (but not yet infectious) at a rate of  $b_N T_N \hat{l}_N/N_H$ , which is then non-dimensionalised to  $b_N T_N L l_N$  (Eq. (1) and (2)), where  $b_N$  is the successful biting rate and  $T_N$  is the transmission probability from non-*Wolbachia*-carrying mosquitoes to humans and vice versa. These exposed humans then become infectious at a rate of  $\gamma_H$  and then recover from dengue at a rate of  $\sigma$ .

The aquatic mosquito population increases as the adult mosquitoes mate and breed, but the population is limited by carrying capacity K through the logistic term

$$\rho_N \frac{\hat{F}_N \hat{M}_N}{\hat{M}_N + \hat{F}_N} \left( 1 - \frac{\hat{A}_N}{K} \right) .$$

Since there are equal numbers of male and female mosquitoes  $\hat{M}_N = \hat{F}_N$ , and with the scaling this becomes  $\rho_N F_N (1-A_N)/2$  in Eq. (5). The aquatic population dies at a rate of  $\mu_{NA}$  and mature into susceptible female mosquitoes at a rate of  $\tau_N$ , where only half of the maturing aquatics are female. Susceptible mosquitoes progress to the exposed class after biting infectious humans at a rate of  $b_N T_N \hat{I}_H / N_H$ , which non-dimensionalises to  $b_N T_N I_H$  in Eqs. (6) and (7). They then become infectious at a rate of  $\gamma_N$  (Eq. (8)), where  $1/\gamma_N$  is the extrinsic incubation period.

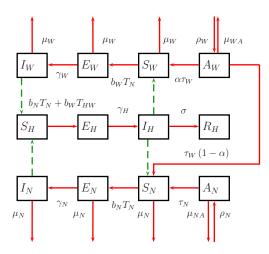


Fig. 2. Schematic of the model in the presence of Wolbachia-carrying mosquitoes, Eqs. (11)-(22). Solid lines are population progression lines and dashed lines are disease transmission lines. The subscript H is for the human population, N is for the non-Wolbachia-carrying mosquito population and W is for the Wolbachia-carrying mosquito population. The compartments are 'S' for susceptible, 'E' for exposed to dengue but not yet infectious, 'I' for infectious, 'R' for recovered, and 'A' for the aquatic phase of the mosquito life cycle. The transition rates between compartments are shown next to the progression lines and are described in the text.

#### 2.1.2. Data and parameter estimation

Cairns is the largest city in the region of Australia where Aedes aegypti are present and local dengue transmission occurs there [37,38]. It is also where the Wolbachia field trials, which began in 2011, are being conducted [16]. In summer 2008/2009, there was a DEN3 outbreak in Cairns [35] and we use data from this outbreak to estimate the parameter values of transmission probability  $(T_N)$ , amplitude of seasonality  $(\eta)$  and seasonal phase  $(\omega)$  for the baseline model, i.e., in the absence of *Wolbachia*, represented by Eqs. (1)–(8). The other parameter values were obtained from the literature and are given in Table 2. The Cairns data covers the period from 2nd November 2008 to 31st May 2009 and was extracted from Fig. 2 of the paper by Ritchie et al. [35]. As our model is formulated as a proportion of the population, each data point is divided by 150,000, which was the approximate population of Cairns in 2008 [39].

We minimise the sum of the squared error between the model and data, which is given by

RSS = 
$$\sum_{i=1}^{n} (y_i - f_i(x))^2$$
, (10)

where  $y_i$  is the total proportion of human dengue cases up to the ith week from the observed data, and  $f_i(x)$  is the total proportion of human dengue cases up to the ith week from the model simulations. The "Isqnonlin" built-in function in MATLAB is then used to estimate the  $T_N$ ,  $\eta$  and  $\omega$  parameter values.

#### 2.2. Model in the presence of Wolbachia

## 2.2.1. Model formulation

Similarly to the model above, the Wolbachia-carrying mosquito population is divided into subpopulations of Aquatic (A<sub>W</sub>, again comprised of eggs, larvae and pupae), Susceptible  $(S_W)$ , Exposed  $(E_W)$  and Infectious ( $I_W$ ) mosquitoes, where  $S_W + E_W + I_W = F_W$  and these have been non-dimensionalised by carrying capacity K. The subscript W is to denote Wolbachia and to differentiate between Wolbachia-free and Wolbachia-carrying mosquitoes. The model comprises 12 compartments in total, for the two mosquito and human populations, and a schematic representation is given in Fig. 2.

The model is governed by the following system of differential

$$\frac{dS_H}{dt} = -b_N T_N L I_N S_H - b_W T_{HW} L I_W S_H,\tag{11}$$

$$\frac{dE_H}{dt} = b_N T_N L I_N S_H + b_W T_{HW} L I_W S_H - \gamma_H E_H, \tag{12}$$

$$\frac{dI_H}{dt} = \gamma_H E_H - \sigma I_H,\tag{13}$$

$$\frac{dR_H}{dt} = \sigma I_H,\tag{14}$$

$$\frac{dA_N}{dt} = \rho_N \frac{F_N^2}{2(F_N + F_W)} (1 - (A_N + A_W)) - (\tau_N + \mu_{NA}) A_N, \tag{15}$$

$$\frac{dS_N}{dt} = \tau_N \frac{A_N}{2} + (1 - \alpha)\tau_W \frac{A_W}{2} - (b_N T_N I_H + \mu_N(t))S_N, \tag{16}$$

$$\frac{dE_N}{dt} = b_N T_N I_H S_N - (\gamma_N + \mu_N(t)) E_N, \tag{17}$$

$$\frac{dI_N}{dt} = \gamma_N E_N - \mu_N(t) I_N,\tag{18}$$

$$\frac{dA_W}{dt} = \rho_W \frac{F_W}{2} (1 - (A_N + A_W)) - (\tau_W + \mu_{WA}) A_W, \tag{19}$$

$$\frac{dS_W}{dt} = \tau_W \alpha \frac{A_W}{2} - (b_W T_N I_H + \mu_W(t)) S_W, \tag{20}$$

$$\frac{dE_W}{dt} = b_W T_N I_H S_W - (\gamma_W + \mu_W(t)) E_W$$
 (21)

$$\frac{dI_W}{dt} = \gamma_W E_W - \mu_W(t) I_W. \tag{22}$$

In this model, the rates at which a human becomes exposed are now different to the model in the absence of Wolbachia, as seen in Eq. (1). Here, a susceptible human becomes exposed after being bitten by either non-Wolbachia or Wolbachia-carrying infectious mosquitoes at a rate of  $b_N T_N LI_N$  and  $b_W T_{HW} LI_W$ , respectively (see Eqs. (11) and (12)), where  $b_W$  is the successful biting rate of Wolbachia-carrying mosquitoes and T<sub>HW</sub> is the transmission probability from Wolbachiacarrying mosquitoes to humans. Note that the transmission probability from humans to Wolbachia-carrying mosquitoes is assumed to be equal to that of humans to non-Wolbachia mosquitoes, so that  $T_{WH} =$  $T_N$ , while the transmission probability of dengue from mosquitoes to human differs between Wolbachia- and non-Wolbachia-carrying mosquitoes (see Section 2.2.2 for explanation).

For the mosquito populations, the effects of cytoplasmic incompatibility and imperfect maternal transmission are included. The effect of CI is included by considering different mating functions, i.e., the non-Wolbachia-carrying females can only reproduce when mating with Wolbachia-carrying males, giving  $\rho_N F_N M_N/P$ , where  $P = F_N + M_N + F_W + M_W$ . Since the ratio of male to female mosquitoes is taken to be 1:1, this is reduced to  $\rho_N F_N^2/(2(F_N + F_W))$ (see Eq. (15)). Aquatic Wolbachia-carrying mosquitoes are produced when Wolbachia-carrying females mate with both Wolbachia-free and *Wolbachia*-carrying males, giving  $\rho_W F_W (M_N + M_W) / P$ , which simplifies to  $\rho_W F_W / 2$  (Eq. (19)). Wolbachia-carrying aquatic mosquitoes mature into *Wolbachia*-carrying adults at a rate of  $\tau_W$ , with a proportion,  $\alpha$ , of them becoming Wolbachia-carrying adults and  $(1 - \alpha)$  becoming non-Wolbachia-carrying adults (see Eqs. (16) and (20)), to capture the effect of imperfect maternal transmission of Wolbachia [15,40]. As with the non-Wolbachia-carrying mosquitoes, the death rate of Wolbachia-carrying adult mosquitoes varies seasonally according to Eq. (9).

**Table 1** Relationships between non-Wolbachia and Wolbachia related parameters. c, d, f and g are the ratios of the reproductive rate, transmission probability, death rate and the biting rate of Wolbachia-carrying mosquitoes to non-Wolbachia rates, respectively.

Parameter	Relationship		
$\rho_{W}$	$c\rho_N$		
$T_{HW}$	$dT_N$		
$\mu_W$	$f\mu_N$		
$b_W$	$gb_N$		

#### 2.2.2. Wolbachia parameters

In this section, model parameters relating to *Wolbachia* are presented and discussed. Most of these are given in terms of non-*Wolbachia* ones, following the conventions in the literature. The relationships between *Wolbachia* and non-*Wolbachia* parameters are given in Table 1.

The reproductive rate of Wolbachia-carrying mosquitoes is generally lower than for non-Wolbachia-carrying ones (c < 1), significantly so for WMelPop and marginally so for the WMel strain. This is because WMelPop decreases the viability of eggs [20,41], whereas WMel does not have a significant effect on them [15]. The death rate of Wolbachia-carrying mosquitoes is higher than non-Wolbachia ones (f > 1) because Wolbachia reduces the mosquito lifespan [15,20]. WMel and WMelPop reduce the lifespan of mosquitoes up to 10% and 50% respectively [15,20]. Wolbachia also inhibits viral replication and dissemination in the mosquitoes [13,15]. This results in a lower dengue viral load in the Wolbachia-carrying mosquitoes, and to reflect this, we set the transmission probability from infectious Wolbachia-carrying mosquitoes to humans to less than that for Wolbachia-free mosquitoes (d < 1). Additionally, Wolbachia causes a condition known as bendy proboscis [42], which inhibits feeding and lowers the successful biting rate (g < 1). This lower biting rate also captures the effect that due to the viral replication inhibition, some Wolbachia-carrying mosquitoes are effectively not infected with dengue, and so the overall transmission rate from humans is lower (that is,  $b_W T_N < b_N T_N$ ).

For the parameter values used in this paper, there are only two realistic stable states, which are that only non-*Wolbachia*-carrying mosquitoes persist, and that both populations persist (Fig. 3). An additional stable state does exist where only the *Wolbachia*-carrying mosquitoes persist, but this requires perfect maternal transmission of *Wolbachia*, which may not be realistic [15]. This state was also found by Ndii et al. [24] for an autonomous system. When both populations persist, the proportion of *Wolbachia*-carrying mosquitoes is around 86%, which compares well with the 90% observed in Hoffmann et al. [16].

#### 2.2.3. Measurement of Wolbachia effect on dengue

A measure is needed to assess the impact of the *Wolbachia* intervention on dengue transmission, which we do by comparing the total number of human dengue cases in the absence and presence (i.e., persistence) of *Wolbachia*-carrying mosquitoes. The relative effect is expressed as a percentage, given by

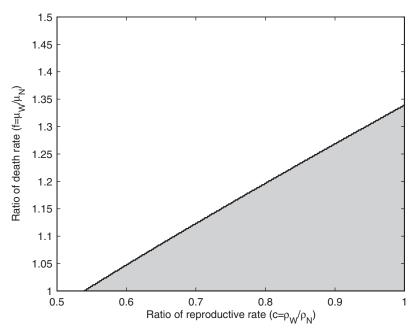
$$\kappa = 100 \times \left(\frac{H_A - H_P}{H_A}\right) \%, \tag{23}$$

where *H* is the final attack proportion of the human population, with subscripts to denote the absence (A) and presence (P) of *Wolbachia*.

#### 2.3. Sensitivity analysis

A global sensitivity analysis is performed using the standard combination of Latin Hypercube Sampling (LHS) and Partial Rank Correlation Coefficient (PRCC) multivariate analysis [43–45]. LHS is a stratified Monte Carlo sampling method, where the random parameter distributions are divided into *N* equal probability intervals and samples taken from each [43–45], where *N* is the sample size. Each interval of each parameter is sampled only once without replacement, and the entire range of each parameter is explored [43–45]. Parameters are sampled from a triangular probability distribution because we expect the values close to the peak of the triangular distribution pattern are those that are more likely to occur. The minimum, maximum and expected values are given in Table 2.

PRCC is an efficient method for measuring the nonlinear but monotonic relationship between inputs and the model outcome of interest [43-45]. In this paper, the inputs are the parameters as well as the



**Fig. 3.** A plot of the persistence, shown by the shaded region, of the *Wolbachia*-carrying mosquito population, over a range of the *Wolbachia* reproductive and death rates, expressed as ratios of the non-*Wolbachia* rates. In the unshaded region only non-*Wolbachia*-carrying mosquitoes persist.

initial number of exposed humans, while the model outcome is the cumulative proportion of infectious individuals, which is the solution of

$$\frac{dC_{IH}}{dt} = \gamma_H E_H.$$

The ranges of the input parameters are available in the literature and only samples of the parameter values that result in the persistence of mosquitoes are included in the calculation. The PRCC is computed for the full length of 31 weeks. The statistical significance test for each PRCC value is performed according to:

$$t = \text{PRCC}\sqrt{\frac{N-2}{1 - \text{PRCC}^2}}.$$

The most significant parameters are those for which a small change in value leads to a significant change in the output, that is the cumulative number of infectious humans.

#### 3. Results

In this section the model simulation and sensitivity analysis results are presented for both models. The governing systems of differential equations are integrated using MATLAB's inbuilt routine "ode45", with the parameter values given in Table 2. We run the model until the mosquito population reaches the periodic stable state, before the infected humans are introduced into the population on 2nd November. That is, the transient dynamics of the introduction of the *Wolbachia* mosquitoes into the system are not considered. In Far North Queensland, dengue is not endemic, hence dengue outbreaks occur as dengue cases are introduced into the population. For both models, the initial conditions for the human population are  $E_{H0} = 2/(1.5 \times 10^5)$ ,  $I_{H0} = 0 = R_{H0}$  and  $S_{H0} = 1 - E_{H0} - I_{H0} - R_{H0}$ .

#### 3.1. Model in the absence of Wolbachia

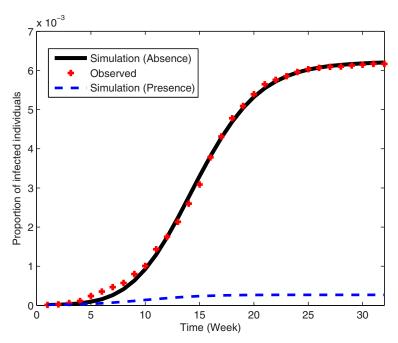
In this section we consider the model in the absence of *Wolbachia*, Eqs. (1)–(8), as described in Section 2.1, which serves as a baseline model for comparison with the *Wolbachia* intervention. Most of the parameters are obtained from the literature, as per references in Table 2. The remaining three parameters,  $T_N$ ,  $\eta$  and  $\omega$ , are optimised using MATLAB's lsqnonlin function. We constrain the optimisation

by physical limits, that is, between 0 and 1 for  $T_N$  since it is a probability, 0 to 365 for seasonal phase  $\omega$ , and 0 to 1 for  $\eta$  so that the death rate  $\mu_N$  remains positive at all times. We fit the parameters as follows. The model is run to its periodic stable state and then two exposed humans are introduced. The sum of square errors is calculated using the cumulative 31-week proportional weekly data of Ritchie et al. [35]. The parameters are then optimised using MATLAB's 1sqnonlin function, with a final sum of squares error equal to  $3 \times 10^{-7}$ ; the resulting parameter values  $T_N$ ,  $\eta$  and  $\omega$  are given in Table 2. To assess the goodness-of-fit we calculated the  $\chi^2$  statistic for the model residuals using MATLAB's chi2gof function. This returned a p-value of 0.3713, and so we fail to reject the hypothesis that the residuals are from a normal distribution. There is some systematic bias in the residuals around the start of the outbreak, evident in Fig. 4, where the observed outbreak rises slightly faster than the model. While model embellishments could be added to account for this, here we consider that the standard SEIR model of infectious disease is sufficient for our intended purpose: comparing the outbreak dynamics in the absence and presence of Wolbachia.

For the sensitivity analysis, 5200 runs are performed to assess the model's sensitivity to the parameters. The parameter ranges used are given in Table 2. The range of initially-exposed humans is taken to be between one and five individuals. This is realistic as only a small number of initial cases triggers the outbreak [35]. The changes in parameter sensitivity over time are common for SEIR models due to the changes in disease dynamics over time, as seen in Figs. 5 and 6. Fig. 5 shows that, for most of the time period, the most influential parameters are the transmission probability  $(T_N)$ , the biting rate  $(b_N)$ and the average adult death rate ( $\mu_{N0}$ ), where the latter has a negative correlation. The phase,  $\omega$ , influences the outbreak size by shifting the peaks and troughs of the mosquito population around; an outbreak can take off around the peak. Because of the sinusoidal nature of the seasonality, the correlation between  $\omega$  and the outbreak size changes sign over time. The parameter  $\gamma_H$  determines the progression rate of humans from the exposed to infectious class. In early times, when introducing exposed individuals, this parameter drives an increase in infectious humans. If this parameter is high, the initial introduced cases will quickly move to the infectious class. As the epidemic takes off, the cumulative number of infectious individuals are determined more by the biting rate  $b_N$ , successful transmission probability  $T_N$  and so  $\gamma_H$  declines in importance. After the initial introduction of cases in early times, as the epidemic takes off, the dynamics of the mosquito

**Table 2**Parameter descriptions, values and sources for both models. The *Wolbachia*-related parameters are for the *WMel* strain. Further explanation of the parameter values is given in the text.

Symbol	Description	Min	Expected	Max	Unit	Source
$T_N$	Transmission probability	0	0.2614	1	N/A	Fitted
$b_N$	Biting rate	0	0.63	1	$day^{-1}$	[46]
$\mu_{N0}$	Adult mosquito death rate	1/30	1/14	1/10		[8]
$d = T_{HW}/T_N$	Ratio of transmission probability W cf. non-W	0	0.5	1	N/A	[13]
$\sigma$	Recovery rate	1/14	1/5	1/3	$day^{-1}$	[47]
$g = b_W/b_N$	Ratio of biting rates W cf. non-W	0	0.95	1	N/A	[42]
$ au_W$	Maturation rate of W	1/12	1/10	1/8	$day^{-1}$	[8]
ω	Phase	0	20.61	365	day	Fitted
$f = \mu_W/\mu_N$	Ratio of death rate W cf. non-W	1	1.1	1.25	N/A	[15,20]
$\rho_N$	Reproductive rate	1	1.25	2.5	$day^{-1}$	[24]
α	Maternal transmission	0.85	0.9	1	N/A	[15,24,40]
$c = \rho_W/\rho_N$	Ratio of reproductive rate W cf. non-W	0.7	0.95	1	N/A	[15]
γw	Progression rate from exposed to infectious	1/12	1/10	1/8	$day^{-1}$	[48]
$ au_N$	Maturation rate	1/17	1/10	1/6	$day^{-1}$	[8]
$\mu_{WA}$	Aquatic death rate	1/20	1/14	1/7	$day^{-1}$	[8]
η	Seasonality amplitude	0	0.6228	1	N/A	Fitted
γн	Progression rate from exposed to infectious human	1/7	1/5.5	1/4	$day^{-1}$	[47]
$\mu_{NA}$	Death rate of aquatic non-W	1/20	1/14	1/7	$day^{-1}$	[8]
$\gamma_N$	Progression from exposed to infectious non-W	1/12	1/10	1/8	$day^{-1}$	[48]
$L = K/N_H$	Ratio of carrying capacity cf. total human population		3		N/A	[48]
E <sub>H0</sub>	Initial exposed human	1	2	5	N/A	



**Fig. 4.** Plots of observed data and the output of the fitted model (*Wolbachia* absent), Eqs. (1)–(8)), as well as the model in the presence of *Wolbachia* (Eqs. (11)–(22)). The data covers the time period 2nd November 2008 to 31st May 2009, extracted from Fig. 2 of Ritchie et al. [35]. The parameter values are given in Table 2. The initial human subpopulations are  $E_{H0} = 2/(1.5 \times 10^5)$ ,  $I_{H0} = R_{H0} = 0$  and  $S_{H0} = 1 - E_{H0} - I_{H0} - R_{H0}$ , for both models. The initial mosquito subpopulations for the model in the absence of *Wolbachia* are  $A_{N0} = 0.8210$ ,  $S_{N0} = 1.2634$  and  $E_{M0} = I_{M0} = 0$ , and for the model in the presence of *Wolbachia* they are  $A_{N0} = 0.0138$ ,  $S_{N0} = 0.1326$ ,  $A_{W0} = 0.7535$ ,  $S_{N0} = 0.9400$  and  $E_{M0} = I_{M0} = E_{W0} = I_{W0} = 0$ . The initial proportion of exposed individuals are introduced after the mosquito population reaches the stable periodic state.

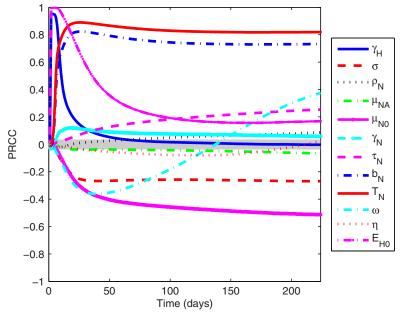


Fig. 5. Plot of the PRCC over time of the model in the absence of Wolbachia. The PRCC is calculated with respect to cumulative number of infectious individuals. The grey area indicates the region where the PRCC is not significantly different from zero (significance level 0.01), using 5200 samples.

population have a larger influence on the disease dynamics. When there are many susceptible mosquito in the population, there will be many infectious mosquitoes and hence many infected humans. An increase in the number of susceptible mosquitoes is regulated by the parameter  $\tau_N$  and so this parameter is impactful in the later period after the epidemic has taken off. As expected, the cumulative number of infectious individuals is most sensitive to the initially-exposed humans ( $E_{H0}$ ) at early times, since they are immediately added to this output. However, for the range considered,  $E_{H0}$  does not change the proportion of dengue reduction due to the introduction of Wolbachia-carrying mosquitoes.

#### 3.2. Model in the presence of Wolbachia

In this section the effect of *Wolbachia*-carrying mosquitoes on the dengue dynamics is investigated. Variations in parameter values are explored to quantify the persistence of *Wolbachia* and its effects on human dengue cases. The initial conditions for the two mosquito populations are  $A_{N0} = 0.0138$ ,  $S_{N0} = 0.1326$ ,  $E_{N0} = I_{N0} = 0$ ,  $A_{W0} = 0.7535$ ,  $S_{W0} = 0.9400$ , and  $E_{W0} = 0 = I_{W0}$ , which are obtained by running the model to the periodic stable state with no dengue. Since the *WMel* strain of *Wolbachia* is used in the Cairns field experiments [16], we use its expected parameter values in our model.

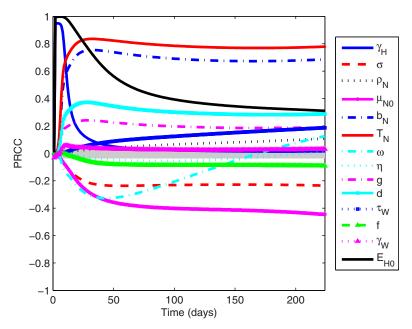
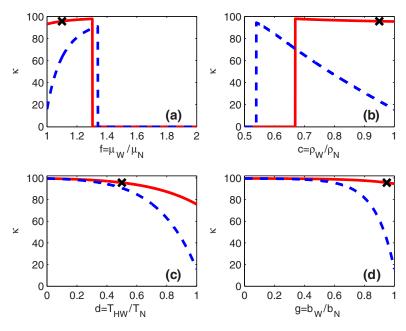


Fig. 6. Plot of PRCC over time for the model in the presence of Wolbachia. For clarity we show only those parameters that have a PRCC outside the range (-0.05, 0.05). The grey area indicates the region in which the PRCC is not significantly different from zero (significance level 0.01), using 5190 samples.



**Fig. 7.** Plot of the relative effect,  $\kappa$  from Eq. (23), on human dengue cases in the absence and presence of *Wolbachia*-carrying mosquitoes against the ratios of (a) adult death rate  $(f = \mu_W/\mu_N)$ , (b) reproductive rate  $(c = \rho_W/\rho_N)$ , (c) transmission probability  $(d = T_{HW}/T_N)$  and (d) biting rate  $(g = b_W/b_N)$ . In each case the other ratios are set to the expected values for *WMel* from Table 2 (solid lines), or 1 (dashed lines). Crosses mark the case where all ratios are set to their expected *WMel* values.

#### 3.2.1. Sensitivity analysis

The results of the sensitivity analysis are similar to those of the model in the absence of *Wolbachia*, and are shown in Fig. 6. The transmission probability  $(T_N)$ , the biting rate  $(b_N)$  and the average adult mosquito death rate  $(\mu_{N0})$  are the most influential parameters on the model outcome. Furthermore, in early times, the cumulative number of infectious individuals is sensitive to  $E_{H0}$  and  $\gamma_H$ . An exploration of the ratios of transmission probability,  $d=T_{HW}/T_N$ , biting rate,  $g=b_W/b_N$ , and death rate,  $f=\mu_W/\mu_N$ , is also carried out, as they relate to the most influential parameters. We also explore the ratio of reproductive rate,  $c=\rho_W/\rho_N$ , to obtain information regarding its effect on dengue and on the parameter range where *Wolbachia*-carrying mosquitoes persist.

#### 3.2.2. Parameter exploration

Although we are using values for c, d, f and g from the literature, the provided values are generally qualitative descriptions, or from lab trials which may not be representative of what happens in the field, and hence the correct parameter values may differ. Therefore, in this section we vary these parameters one at a time to determine their effects on human dengue cases, as measured by the relative effect given in Eq. (23). Thus, we quantify the effect of introducing Wolbachia-carrying mosquitoes on dengue transmission for a range of realistic parameter values. There are two scenarios being considered, and their resulting effects on dengue cases are shown in Fig. 7. The first uses the expected values for WMel for the non-varied Wolbachia parameters (the solid line in the figure). The second scenario

uses the non-Wolbachia-carrying mosquito values (i.e., taking the ratios in Table 1 equal to one), allowing us to explore the effect of cytoplasmic incompatibility on the dengue transmission dynamics, should Wolbachia have no other effect on the mosquito physiology. This second scenario is represented by the dashed line in the figure.

The vertical lines in Fig. 7(a) and (b), for f and c, respectively, denote the boundary between persistence ( $\kappa > 0$ ) and non-persistence  $(\kappa = 0)$  for the Wolbachia-carrying mosquitoes. For f, the ratio of death rates, both scenarios show that as the death rate of Wolbachiacarrying mosquitoes increases, the relative effect on dengue also increases up to the point where the Wolbachia-carrying mosquitoes no longer persist, around f = 1.30 for the solid line and f = 1.34 for the dashed line. The default values, with f = 1.1, are identified on the figure by a cross, showing that WMel should reduce human dengue cases by approximately 96%. Interestingly, in the second scenario, when c = d = f = g = 1, we see that even if all of the Wolbachia parameters are equal to their non-Wolbachia counterparts, the human dengue cases are still reduced by approximately 16%. This occurs due to the combination of CI and competition, with the total female mosquito population  $(F_N + F_W)$  being reduced. The CI means there is a different mating function for the Wolbachia-free and Wolbachia-carrying mosquitoes, and in particular that no non-Wolbachia-carrying offspring are produced from the combination of a non-Wolbachia female and Wolbachia-carrying male (there is effectively a  $0 \times F_N F_W$  term in Eq. (15)). This difference in the mating functions combined with the competition in the aquatic phase results in a reduced total mosquito population, and hence less dengue. This provides the new insight that CI alone affects the dengue transmission dynamics.

In Fig. 7(b), for the solid line with expected *WMel* values, when the reproductive rate of *Wolbachia*-carrying mosquitoes is too low compared to *Wolbachia*-free mosquitoes, the *Wolbachia*-carrying mosquitoes no longer persist, and hence have no effect on human dengue cases. This occurs at approximately c=0.66 for the expected parameter values (solid line), and c=0.52 for d=f=g=1 (dashed line). When *Wolbachia*-carrying mosquitoes do persist, the effect on human dengue decreases with increasing c since the total number of mosquitoes increases, despite Cl. However, this effect is negligible for the expected parameter values (solid line), with over 90% reduction in dengue for all values where *Wolbachia*-carrying mosquitoes persist.

In Fig. 7(c), the results show that even if  $d\approx 1$  (transmission is not directly affected by Wolbachia), a reduction in human dengue cases of approximately 76% is still obtained, due to the other effects of Wolbachia on mosquito physiology. In addition, as shown in Fig. 7(d), a 90% reduction in human dengue cases is obtained even though the biting rate of Wolbachia-carrying mosquitoes is close to that of non-Wolbachia mosquitoes ( $g\approx 1$ ).

### 4. Discussion and conclusions

We have developed a mathematical model for the presence of a Wolbachia-carrying mosquito population in order to quantify the effects of such an intervention on human dengue cases. Our model incorporates seasonal forcing through the adult mosquito death rate, the effect of cytoplasmic incompatibility, and competition for resources in the aquatic stage. This model considers only a single dengue serotype with the mosquito population in periodic stable state, appropriate to the study of a single dengue outbreak. It is assumed that half of the mosquito population mature into female adults, that a single introduction event occurs, and that there is no initial dengue immunity in the human population. These effects will have an impact on the dengue transmission dynamics, with initial dengue immunity reducing the relative effect of the Wolbachia introduction due to smaller outbreak sizes. However, the purpose of this model was to determine an indicative effect of the introduction of Wolbachia-carrying mosquitoes on the dengue transmission dynamics. Extensions to the

model which consider the effect of immunity in the human population are the subject of future work.

A model without seasonality can only fit the data for far North Queensland in one of two ways. Firstly, nearly all humans need to be previously infected and are now recovered ( $R_{H0} \approx 1$ ), which is not realistic. Secondly, all mosquitoes can die out, which mimics seasonal effects for a single year. However, unlike in a model which includes seasonal effects, the mosquito population cannot re-emerge. Exploration of key parameter ranges was performed to determine the level of dengue reduction due to *Wolbachia* and the persistence of *Wolbachia*-carrying mosquitoes, which is important since definite values are not known. A global sensitivity analysis was used to determine the most influential parameters.

We found that using the Wolbachia strain WMel, which reduces the mosquito lifespan by at most 10%, allows Wolbachiacarrying mosquitoes to persist. This result is compatible with what has been suggested by Walker et al. [15] and the results from field experiments [16]. Furthermore, Fig. 7 provides the new insight that cytoplasmic incompatibility is an important biological factor. CI not only gives a reproductive advantage to Wolbachia-carrying female mosquitoes, but also influences the transmission dynamics of dengue, through a reduced mosquito population. The release of more Wolbachia-carrying males than females is being considered by Hancock et al. [33], and their analysis may be important in considering the effect of cytoplasmic incompatibility on dengue transmission dynamics, and the possible feminisation of the Wolbachia-carrying mosquito population. However, our results suggest that with equal numbers of males and females, Wolbachia-carrying mosquitoes can persist, and hence a reduction in dengue human cases can be obtained. Furthermore, the ratios of transmission probability,  $d = T_{HW}/T_N$ , and of biting rate,  $g = b_W/b_N$ , determine the level of reduction in human dengue cases. In contrast, the ratios of death rate,  $f = \mu_W/\mu_N$ , and of reproductive rate,  $c = \rho_W/\rho_N$ , determine the persistence of Wolbachia-carrying mosquitoes. This result is corroborated by the results of the sensitivity analysis. Additionally, the biting and reproductive rates are linked, since female mosquitoes take blood meals for reproduction. Here we treat them as independent for the purpose of exploring the parameter space and note that the relative effect is

In conclusion, a one-strain dengue mathematical model incorporating *Wolbachia*-carrying mosquitoes has been developed for a single outbreak. We quantify the effect that *Wolbachia* will have on human dengue cases, once the mosquito populations have reached their periodic stable states. *Wolbachia* comes with a fitness cost to *Aedes aegypti* mosquitoes, and we explore the parameter values for which *Wolbachia* will persist in the mosquito population. Our results show that for realistic *WMel* parameter values, *Wolbachia*-carrying mosquitoes can greatly reduce the transmission of dengue. We also show that in the absence of any other effect on the mosquito physiology by *Wolbachia*, cytoplasmic incompatibility alone affects the transmission of dengue, reducing the human cases.

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#### References

 World Health Organisation, Dengue and severe dengue Fact sheet No. 117, 2012, http://www.who.int/mediacentre/factsheets/fs117/en/index.html (Accessed 17 February 2013).

- [2] S. Bhatt, P.W. Gething, O.J. Brady, J.P. Messina, A.W. Farlow, et al., The global distribution and burden of dengue, Nature 496 (2013) 1476–4687. 10.1038/nature12060.
- [3] D. Normile, First New Dengue Virus Type in 50 Years, 2013, http://news.science mag.org/health/2013/10/first-new-dengue-virus-type-50-years (Accessed 31 October 2013).
- [4] N.M. Ferguson, C.A. Donnelly, R.M. Anderson, Transmission dynamics and epidemiology of dengue: insights from age stratified sero-prevalence surveys, Philos. Trans. R. Soc. Lond. B: Biol. Sci. 354 (1384) (1999) 757–768. 10.1098/rstb.1999.0428.
- [5] D.P. Webster, J. Farrar, S. Rowland-Jones, Progress towards a dengue vaccine, Lancet Infect. Dis. 9 (11) (2009) 678–687. 10.1016/S1473-3099(09)70254-3.
- [6] U. Thisyakorn, C. Thisyakorn, Latest developments and future directions in dengue vaccines, Ther. Adv. Vaccines 2 (1) (2014) 3–9. 10.1177/2051013613507862.
- [7] M. Chan, M.A. Johansson, The incubation periods of dengue viruses, PLoS One 7 (11) (2012) e50972. 10.1371/journal.pone.0050972.
- [8] H.M. Yang, M.L.G. Macoris, K.C. Galvani, M.T.M. Andrighetti, D.M.V. Wanderley, Assessing the effects of temperature on the population of *Aedes aegypti*, the vector of dengue, Epidemiol. Infect. 137 (2009) 1188–1202. 10.1017/S0950268809002040.
- [9] N. Grisales, R. Poupardin, S. Gomez, I. Fonseca-Gonzalez, H. Ranson, A. Lenhart, Temephos resistance in *Aedes aegypti* in Colombia compromises dengue vector control, PLoS Negl. Trop. Dis. 7 (9) (2013) e2438. 10.1371/journal.pntd.0002438.
- [10] I.R. Montella, A.J. Martins, P.F. Viana-Medeiros, J.B.P. Lima, I.A. Braga, D. Valle, Insecticide resistance mechanisms of Brazilian Aedes aegypti populations from 2001 to 2004, Am. J. Trop. Med. Hyg. 77 (3) (2007) 467–477.
- [11] S.A. Ritchie, J.N. Hanna, S.L. Hills, J.P. Piispanen, W.J.H. McBride, A. Pyke, R.L. Spark, Dengue control in North Queensland, Australia: case recognition and selective indoor residual spraying, Dengue Bull. 26 (2002) 7–12.
- [12] E. Ooi, K. Goh, D.J. Gubler, Dengue prevention and 35 years of vector control in Singapore, Emerg. Infect. Dis. 12 (6) (2006) 887–893.
- [13] G. Bian, Y. Xu, P. L., Y. Xie, Z. Xi, The endosymbiotic bacterium Wolbachia induces resistance to dengue virus in Aedes aegypti, PLoS Pathog. 6 (4) (2010) e1000833. 10.1371/journal.ppat.1000833.
- [14] C.J. McMeniman, R.V. Lane, B.N. Cass, A.W.C. Fong, M. Sidhu, Y.F. Wang, S.L. O'Neill, Stable introduction of a life-shortening *Wolbachia* infection into the mosquito *Aedes aegypti*, Science 323 (5910) (2009) 141–144. 10.1126/science.1165326.
- [15] T. Walker, P.H. Johnson, L.A. Moreira, I. Iturbe-Ormaetxe, F.D. Frentiu, et al., The WMel Wolbachia strain blocks dengue and invades caged Aedes aegypti populations, Nature 476 (2011) 450-453. 10.1038/nature10355.
- [16] A.A. Hoffmann, B.L. Montgomery, J. Popovici, I. Iturbe-Ormaetxe, P.H. Johnson, et al., Successful establishment of Wolbachia in Aedes populations to suppress dengue transmission, Nature 476 (2011) 454–457. 10.1038/nature10356.
- [17] L.A. Moreira, I. Iturbe-Ormaetxe, J.A. Jeffery, G. Lu, A.T. Pyke, et al., Wolbachia symbiont in Aedes aegypti limits infection with dengue, chikungunya, and plasmodium, Cell 139 (2009) 1268–1278.
- [18] K. Hilgenboecker, P. Hammerstein, A. Telschow, J.H. Werren, How many species are infected with Wolbachia? A statistical analysis of current data, FEMS Microbiol. Lett. 281 (2) (2008) 215–220. 10.1111/j.1574-6968.2008.01110.x.
- [19] M. Turelli, Cytoplasmic incompatibility in populations with overlapping generations, Evolution 64 (2010) 232–241. 10.1111/j.1558-5646.2009.00822.x.
- [20] H.L. Yeap, P. Mee, T. Walker, A.R. Weeks, S.L. O'Neill, et al., Dynamics of the "Popcorn" Wolbachia infection in outbred *Aedes aegypti* informs prospects for mosquito vector control, Genetics 187 (2) (2011) 583–595. 10.1534/genetics.110.122390.
- [21] P.A. Hancock, S.P. Sinkins, H.C.J. Godfray, Population dynamic models of the spread of Wolbachia, Am. Nat. 177 (3) (2011) 323–333.
- [22] P.A. Hancock, H.C.J. Godfray, Modelling the spread of Wolbachia in spatially heterogeneous environments, J. R. Soc. Interface 9 (76) (2012) 3045–3054. 10.1098/rsif.2012.0253.
- [23] M.H.T. Chan, P.S. Kim, Modelling a Wolbachia invasion using a slow fast dispersal reaction diffusion approach, Bull. Math. Biol. 75 (9) (2013) 1501–1523. 10.1007/s11538-013-9857-y.
- [24] M.Z. Ndii, R.I. Hickson, G.N. Mercer, Modelling the introduction of Wolbachia into Aedes aegypti to reduce dengue transmission, ANZIAM J. 53 (2012) 213–227. 10.1017/S1446181112000132.
- [25] M. Andraud, N. Hens, P. Beutels, A simple periodic-forced model for dengue fitted to incidence data in Singapore, Math. Biosci. 244 (1) (2013) 22–28. 10.1016/j.mbs.2013.04.001.

- [26] L. Esteva, C. Vargas, Analysis of a dengue disease transmission model, Math. Biosci. 150 (2) (1998) 131–151. 10.1016/S0025-5564(98)10003-2.
- [27] L. Esteva, C. Vargas, Influence of vertical and mechanical transmission on the dynamics of dengue disease, Math. Biosci. 167 (1) (2000) 51–64. 10.1016/S0025-5564(00)00024-9.
- [28] S.M. Garba, A.B. Gumel, M.R. Abu Bakar, Backward bifurcations in dengue transmission dynamics, Math. Biosci. 215 (1) (2008) 11–25. 10.1016/j.mbs.2008.05.002.
- [29] T. McLennan-Smith, G.N. Mercer, Complex behavior in a dengue model with a seasonally varying vector population, Math. Biosci. 248 (2014) 22–30. 10.1016/j.mbs.2013.11.003.
- [30] A.K. Supriatna, E. Soewono, S.A. van Gils, A two-age-classes dengue transmission model, Math. Biosci. 216 (1) (2008) 114–121. 10.1016/j.mbs.2008.08.011.
- [31] N. Bacaer, Approximation of the basic reproduction number  $R_0$  for vector-borne diseases with a periodic vector population, Bull. Math. Biol. 69 (3) (2007) 1067–1091. 10.1007/s11538-006-9166-9.
- [32] H. Hughes, N. Britton, Modelling the use of Wolbachia to control dengue fever transmission, Bull. Math. Biol. 75 (5) (2013) 796–818. 10.1007/s11538-013-9835-4.
- [33] P.A. Hancock, S.P. Sinkins, H.C.J. Godfray, Strategies for introducing Wolbachia to reduce transmission of mosquito-borne diseases, PLoS Negl. Trop. Dis. 5 (4) (2011) e1024. 10.1371/journal.pntd.0001024.
- [34] M. Andraud, N. Hens, C. Marais, P. Beutels, Dynamic epidemiological models for dengue transmission: a systematic review of structural approaches, PLoS One 7 (11) (2012) e49085. 10.1371/journal.pone.0049085.
- [35] S.A. Ritchie, A.T. Pyke, S. Hall-Mendelin, A. Day, C.N. Mores, R.C. Christofferson, D.J. Gubler, S.N. Bennett, A.F. van den Hurk, An explosive epidemic of denv-3 in Cairns, Australia, PLoS One 8 (7) (2013) e68137. 10.1371/journal.pone.0068137.
- [36] J. Arrivillaga, R. Barrera, Food as a limiting factor for *Aedes aegypti* in water storage containers, J. Vector Ecol. 29 (1) (2004) 11–20.
- [37] J.S. Mackenzie, J.T. La Brooy, L. Hueston, A.L. Cunningham, Dengue in Australia, J. Med. Microbiol. 45 (3) (1996) 159–161. 10.1099/00222615-45-3-159.
- [38] R.C. Russell, C.E. Webb, C.R. Williams, S.A. Ritchie, Mark release recapture study to measure dispersal of the mosquito *Aedes aegypti* in Cairns, Queensland, Australia, Med. Vet. Entomol. 19 (4) (2005) 451–457. 10.1111/j.1365-2915.2005.00589.x.
- [39] Q.T. Cairns Regional Council, Trade, Population and Dwelling Profile, 2008, http://www.dsdip.qld.gov.au/resources/map/population-housing-factsheets/cairns-regional-council.pdf (Accessed 23 September 2013).
- [40] A.A. Hoffmann, M. Turelli, L.G. Harshman, Factors affecting the distribution of cytoplasmic incompatibility in Drosophila simulans, Genetics 126 (4) (1990) 933–948.
- [41] C.J. McMeniman, S.L. O'Neill, A virulent Wolbachia infection decreases the viability of the dengue vector Aedes aegypti during periods of embryonic quiescence, PLoS Negl. Trop. Dis. 4 (7) (2010) e748. 10.1371/journal.pntd.0000748.
- [42] A.P. Turley, L.A. Moreira, S.L. O'Neill, E.A. McGraw, Wolbachia infection reduces blood-feeding success in the dengue fever mosquito, Aedes aegypti, PLoS Negl. Trop. Dis. 3 (9) (2009) e516. 10.1371/journal.pntd.0000516.
- [43] S. Marino, I.B. Hogue, C.J. Ray, D.E. Kirschner, A methodology for performing global uncertainty and sensitivity analysis in systems biology, J. Theor. Biol. 254 (1) (2008) 178–196. 10.1016/j.jtbi.2008.04.011.
- [44] J. Wu, R. Dhingra, M. Gambhir, J.V. Remais, Sensitivity analysis of infectious disease models: methods, advances and their application, J. R. Soc. Interface 10 (86) (2013). 10.1098/rsif.2012.1018.
- [45] S.M. Blower, M. Dowlatabadi, Sensitivity and uncertainty analysis of complex models of disease transmission: an HIV model, as an example, Int. Stat. Rev. 62 (1994) 229–243.
- [46] T.W. Scott, P.H. Amerasinghe, A.C. Morrison, L.H. Lorenz, G.G. Clark, D. Strickman, P. Kittayapong, J.D. Edman, Longitudinal studies of *Aedes aegypti* (Diptera: Culicidae) in Thailand and Puerto Rico: blood feeding frequency, J. Med. Entomol. 37 (1) (2000) 89–101.
- [47] D.J. Gubler, Dengue and dengue hemorrhagic fever, Clin. Microbiol. Rev. 11 (3) (1998) 480–496.
- [48] G. Chowel, P. Diaz-Duenas, J.C. Miller, A.A. Velazco, J.M. Hyman, P.W. Fenimore, C. Castillo-Chaves, Estimation of the reproduction number of dengue fever from spatial epidemic data, Math. Biosci. 208 (2007) 571–589.