FUMA Practical 2025

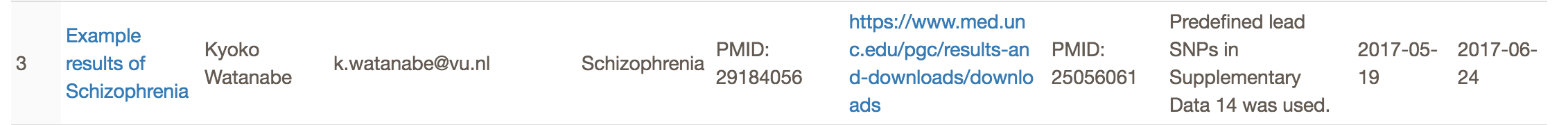
[FUMA GWAS](http://fuma.ctglab.nl/) uses information from publicly available annotation and mapping datasets (see <https://fuma.ctglab.nl/tutorial#celltype> – Data sets)

We will work with results from published GWAS studies on either Neuroticism or Schizophrenia – you can choose which trait to work on.

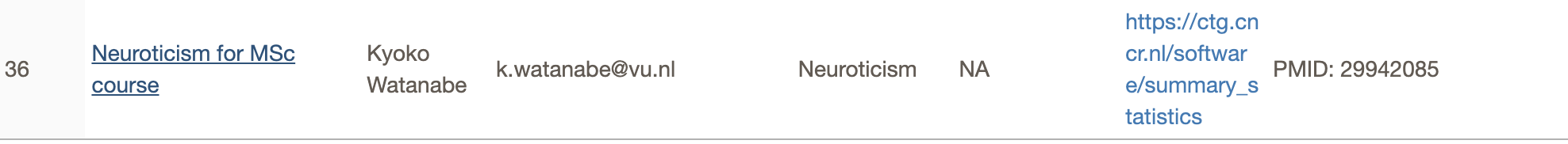
* Schizophrenia (SCZ; [Ripke et al., 2014](https://www.ncbi.nlm.nih.gov/pubmed/25056061))
* Neuroticism (NEU; [Nagel et al., 2018](https://www.nature.com/articles/s41588-018-0151-7.pdf))

*Finding your data*

* Go to: <http://fuma.ctglab.nl/>
* Click ‘Tutorial’ (top of the page) to see a schematic overview of what you can do with FUMA. If something’s unclear during the practical, first consult the tutorial, it’s probably explained in there!
* We will use previously published GWAS results that have already been submitted to FUMA by Kyoko Watanabe. Click ‘Browse Public Results’, and scroll down to the trait of your choice (Schizophrenia Ripke et al. or Neuroticism Nagel et al.). E.g., if you chose schizophrenia, you should click this line:



* For Neuroticism, choose ‘Example results of Neuroticism’.



You may get an AJAX warning that you can ignore.

**Before you start answering question,**

1. **Quickly read through the tutorial page to get an idea of what FUMA can do**
2. **Then click through the results and try to connect the results with the aspects of FUMA outlined in the lecture/tutorial to get a better idea how the results were generated. If you don’t understand how results were generated, check the Tutorial Tab for methods descriptions.**

*Section: Summary of results*

The questions below concern the ‘Summary of results’ section in FUMA.

Due to privacy concerns the publicly available data (as used in this practical) is sometimes slightly different from the data on which published results are based. E.g., the personal genomics company 23andMe shares data, but researchers are not allowed to include the 23andMe participants in any data they make publicly available.

* As you saw in the lecture, SNPs located close to each other often show similar association signal (e.g., if SNP A is strongly associated then SNP B that lies right next to it will likely also show fairly strong association, i.e. low *P*-value). This has to do with the concept of linkage disequilibrium (LD).

*Q1: What LD threshold does FUMA apply to define independent significant SNPs? (see Watanabe et al. (2017) or tutorial).*

*Section: Summary of Results*

Navigate to the Summary of Results tab.

* SNPs in exonic regions potentially affect the expression of the gene or the function of the gene product (protein).

*Q2: How many SNPs are annotated to exonic regions in your example results (schizophrenia or neuroticism)?* ***[Hint: hover over bar plot in Summary of Results Tab****]*

* FUMA established genomic risk loci, based on the independent associations. If you scroll down you see four bar plots, showing information for each genomic locus.

*Q3: What is the largest genomic locus? (Copy-paste the y-axis label)*

*Q4: Does this locus also contain the most SNPs? If not, which locus does?*

*Q5: Why does the number of genes physically located within a locus is not always equal to the number of genes that is mapped to the locus?*

*Section: Genome-wide plots*

The questions below concern the ‘Genome-wide plots’ section in FUMA.

* Below the SNP-level Manhattan plot you find the gene Manhattan plot. This figure shows the gene associations.

*Q6: Why is the threshold for significance different from that in the SNP Manhattan plot?*

* Scrolling down you see a histogram, showing whether 53 types of tissue are enriched for the genes associated to the trait of interest. In other words; this informs you on whether the identified genes are primarily expressed in a specific tissue type.

*Q7: Is there a tissue type that shows significant enrichment? If so, which one(s)? Is this what you would expect, given the trait of interest?*

*Section: Results*

The questions below concern the ‘Results’ section in FUMA (note that this section has several tabs).

The ‘Genomic risk loci’ tab lists all genomic loci, and provides information on the location, the number of SNPs, the lowest SNP *P*-value etc. Moreover, if you click a locus, an interactive regional plot appears below the table (you can, for example, click on a SNP for info on that particular SNP). Essentially this is a zoomed in version of the Manhattan plot, with each dot representing a SNP.

Depending on your trait of interest, create a regional plot for genomic locus number:

* Schizophrenia: 97
* Neuroticism: 73

[**If the plot does not appear, try refreshing the page and selecting the locus again**]

*Q8: What is the rsID of the top lead SNP? Which chromosome is it on? How many SNPs are in LD with this lead SNP? What is the minor allele frequency? [****Hint: try hovering over the lead SNP****]*

Now click the ‘Plot’ button on the bottom to open a new window, showing the same regional plot with additional more information. Try zooming, scrolling and clicking a bit to see what happens.

[**There may be a JobQuery error. Press OK. Hopefully this will be fixed soon**]

*Q9: What is the nearest mapped gene to the lead SNP? [****Hint: Click on the lead SNP****]*

* The plot below shows the CADD score. Since GWAS often identifies many SNPs, we need a criterion to prioritize which ones to study further.

*Q10: Why is the CADD score helpful for this? Which SNP has the highest CADD score in this locus? Is this below or above the threshold mentioned in Watanabe et al. (2017)?*

*Q11: Was the SNP with the highest CADD score genome-wide significant?*

*Q12: Are there any eQTLs in this locus? How specific are the eQTLs to a single tissue type?*

*Q13: Based on these results, which gene would you recommend to study in more detail? Explain why.*

*Chromatin interactions*

* In the ‘Results’ section, click ‘Chromatin interactions’. If you scroll down you find circos plots, showing eQTL and chromatin interactions per chromosome (read the info text on the circos plots!). As mentioned earlier, these techniques allow SNPs to be mapped to genes that are further apart.

[**Hint: left click on the plot -> open image in new tab**]

*Q14: In the circos plots, do you find examples of these long-range interactions, where genes are implied through interactions with (physically) distant regions? What do the different layers and colors mean?*

*Section: GENE2FUNC*

There exist many collections of genes that have something in common; gene sets. Sometimes these are composed by experts, e.g. listing all genes that are involved in a specific biological process. In FUMA, you can find those results in the ‘Gene sets’ section. We will have a look at the ‘GWAS catalog reported genes’ (second-last), gene sets based on association with a large variety of traits (listed in the GWAS catalog).

*Q15: What is the top 3 gene sets reported for your trait? Does this surprise you?*

**This is the end of the practical. If you have finished early, there are some bonus questions to think about and discuss below, along with some further reading.**

**Bonus Questions:**

* **How do gene-set enrichment analyses in MAGMA and GENE2FUNC compare?**
* **What are some limitations of FUMA?**
* **What are the assumptions behind the MAGMA gene property analysis implemented in FUMA?**
* **What is a limitation of using genes prioritised by FLAMES and PoPs as input for GENE2FUNC?**
* **What are some methods beyond FUMA to prioritise specific variant, genes, and pathways for experimental testing?**
* **Is every eQTL annotated in FUMA causally influencing gene expression?**
* **When finding multiple significant tissues in the tissue enrichment analyses, what are some follow-up steps you could perform?**

**Further reading**

*FUMA:*

* [Watanabe (2017)](file:////Users/kyoko/Documents/VU/CTGLab/Lectures/2018_11_FUMApractice/−%2509Watanabe%20(2017)%20-%20Functional%20mapping%20and%20annotation%20of%20genetic%20associations%20with%20FUMA) - Functional mapping and annotation of genetic associations with FUMA
* [Watanabe (2019)](https://www.nature.com/articles/s41467-019-11181-1) – FUMA cell type

*Studies from which data was used:*

* [Ripke et al. (2014)](https://www.ncbi.nlm.nih.gov/pubmed/25056061) - Biological insights from 108 schizophrenia-associated genetic loci
* [Nagel et al., 2018](https://www.nature.com/articles/s41588-018-0151-7.pdf) - Meta-analysis of genome-wide association studies for neuroticism in 449,484 individuals identifies novel genetic loci and pathways

Other tools

* [de Leeuw et al. (2015)](https://doi.org/10.1371/journal.pcbi.1004219) - MAGMA: Generalized Gene-Set Analysis of GWAS Data
* [Schipper et al. (2025)](https://doi.org/10.1038/s41588-025-02084-7) - Prioritizing effector genes at trait-associated loci using multimodal evidence (FLAMES)
* [Weeks et al. (2023)](https://doi.org/10.1038/s41588-023-01443-6) - Leveraging polygenic enrichments of gene features to predict genes underlying complex traits and diseases (PoPs)