# AUTOMATIC CELL SEGMENTATION TO ANALYSE SINGLE CELL HETROGENEITY BASED ON MORPHOLOGICAL FEATURES





#### **OBJECTIVE**

- To perform automatic segmentation and extract multiple morphological features from cells using MATIab.
- To elucidate the single cell heterogeneity among the cells based on shape and morphological features.
- To compare between single cell and overlapped cells in a image and infer the changes.

# BACKGROUND



#### **Problem Identification**

ECM stiffness may cause heterogeneity

0.03

0.3

# INTRODUCTION



#### **INTRODUCTION**

- The ensemble behaviors of cell population may not represent the behaviors of any individual cell.
- Behavior of each and every cell in a population is important to understand the accurate behavior of cells.
- The diverse collection of cells harboring distinct molecular signatures will have differential levels of sensitivity to treatment.
- So, it is essential to identify the differences that are important to differentiate between the cells.

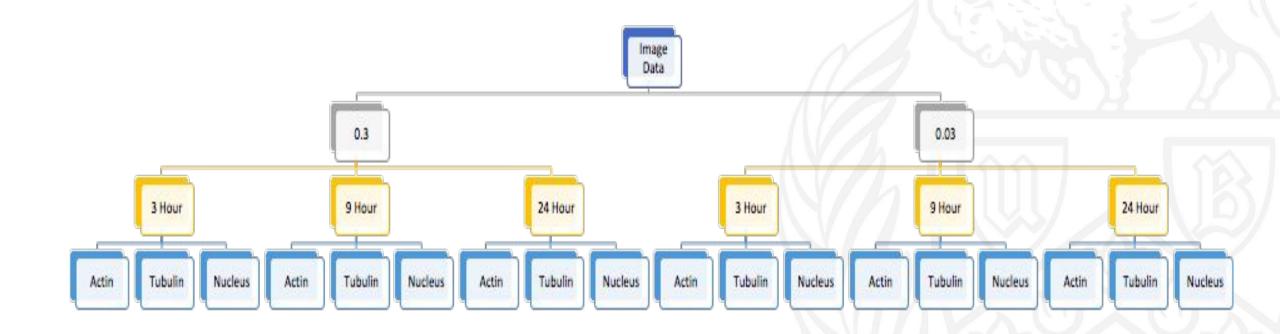
# DATASET DESCRIPTION



#### **Dataset**

- Our dataset consists of images of two types of cells, Actin and Tubulin, nucleus at two different stress levels, 0.03 and 0.3.
- The stress levels are the mechanical stress experienced by the cells.
- The cells in 0.03 corresponds to healthy cells and at 0.3 are the diseased cells.
- Both the healthy and diseased state cells were monitored at three different times, 3 hours, 9 hours, and 24 hours

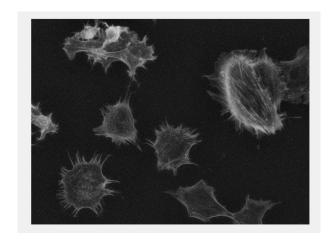
#### Data Framework



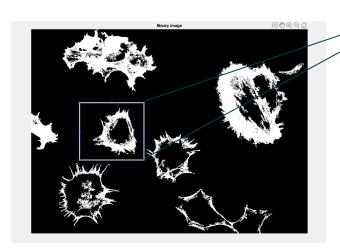
## WORK FLOW

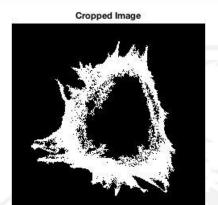


#### WORKFLOW

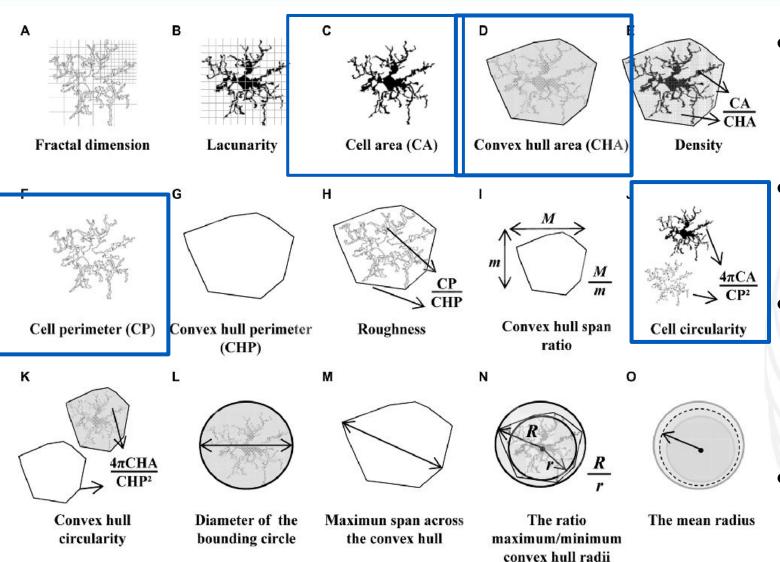








#### University at Buffalo The State University of New York

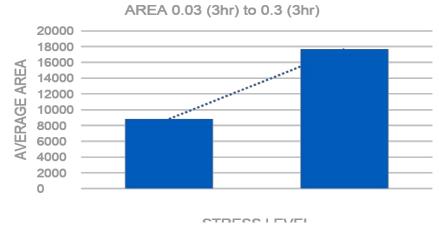


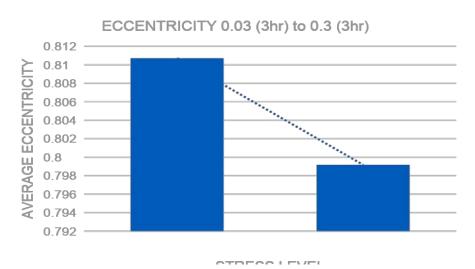
- Area: The area of the particle is calculated as the sum of the areas of each individual pixel within the borders of the cell.
- Perimeter: The perimeter of the particle, P, is defined as the total length of the object boundary.
- Solidity: Solidity, S, is the measurement of the overall concavity of a particle. It is defined as the image area, A, divided by the convex hull area, Ac.
- ratio of the distance between the foci of the ellipse and its major axis length. The value is between 0 and 1.

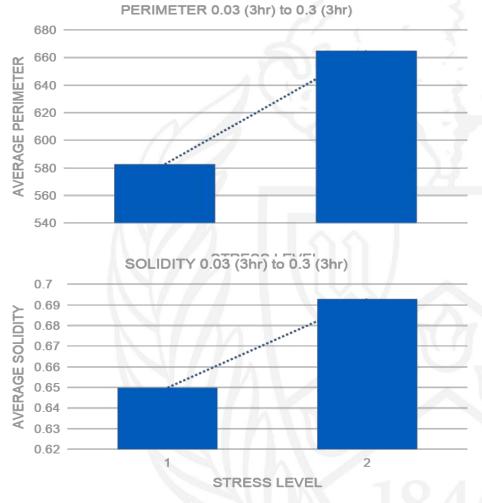
# ANALYSIS



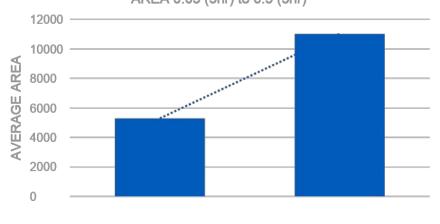
#### AVERAGE VALUE COMPARISON [ACTIN @ 3HR]





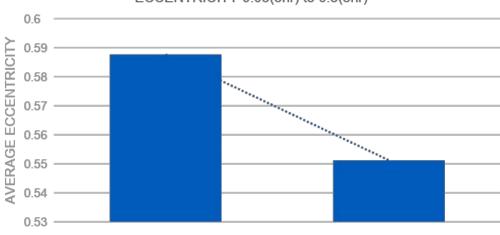


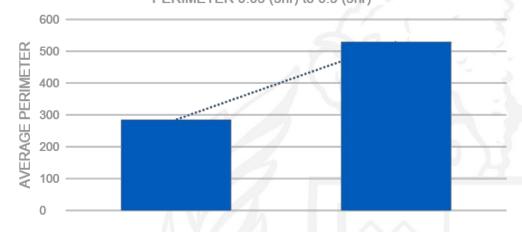
### AVERAGE VALUE COMPARISON TUBULIN @ 3 HR]



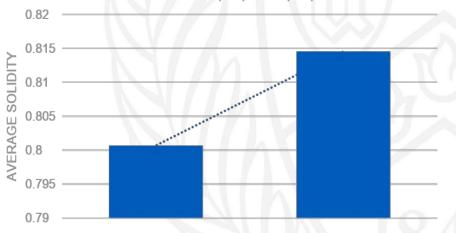


CTDECC LEVE

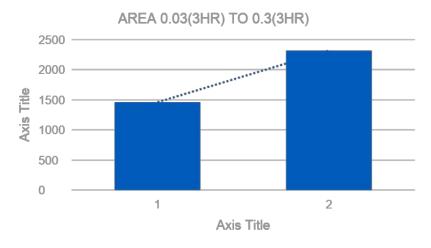


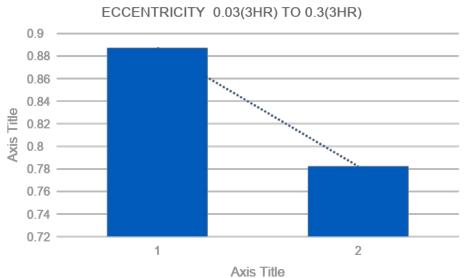


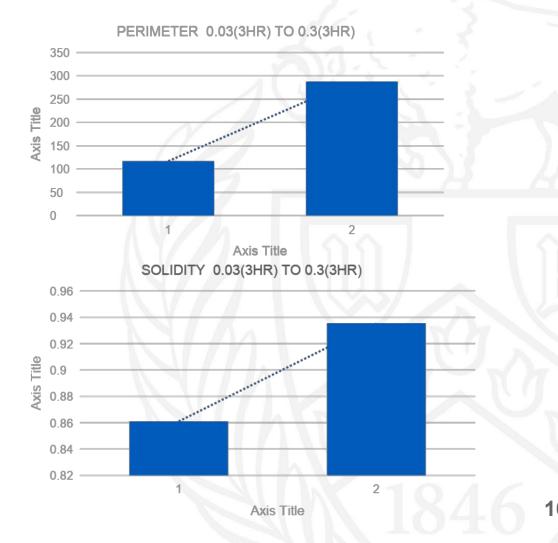




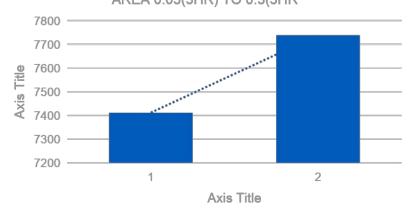
#### AVERAGE VALUE COMPARISON [NUCLEUS @ 3 HR]

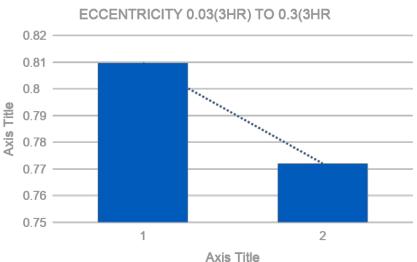


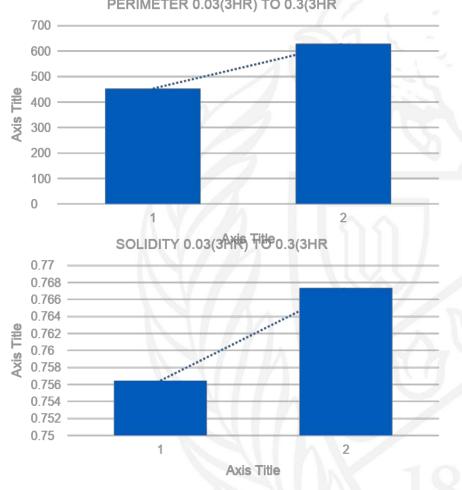




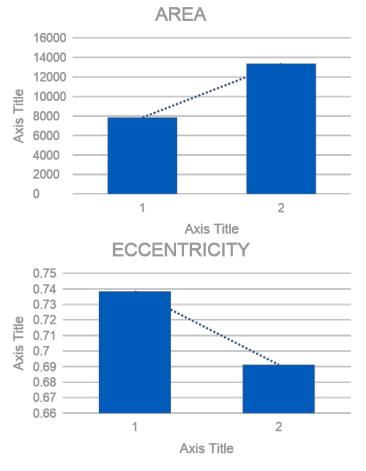
## AVERAGE VALUE COMPARISON [ACTIN @ 9 HR]

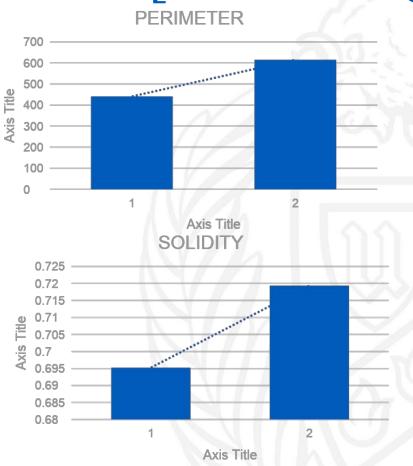




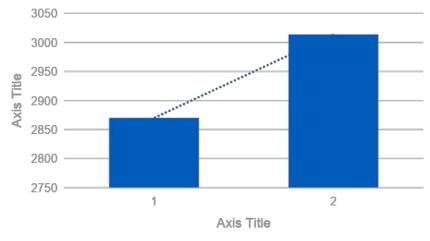


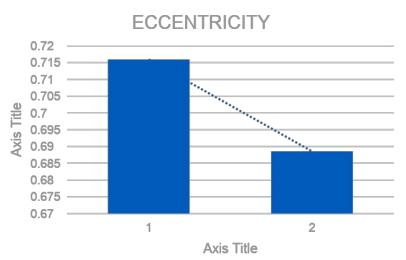
#### AVERAGE VALUE COMPARISON [TUBULIN @ 9 HR]

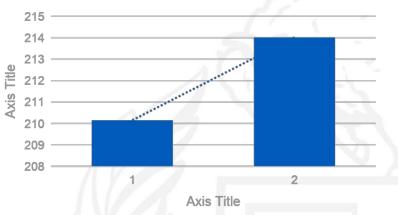


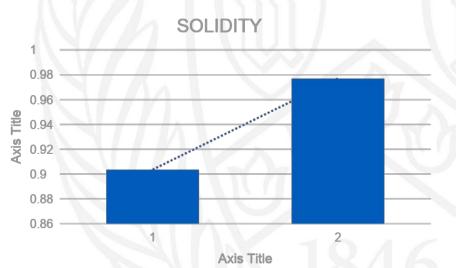


#### AVERAGE VALUE COMPARISON [NUCLEUS @ 9 HR]

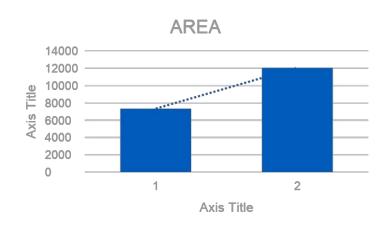


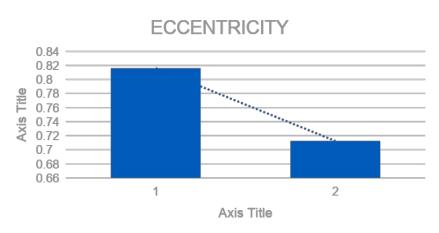


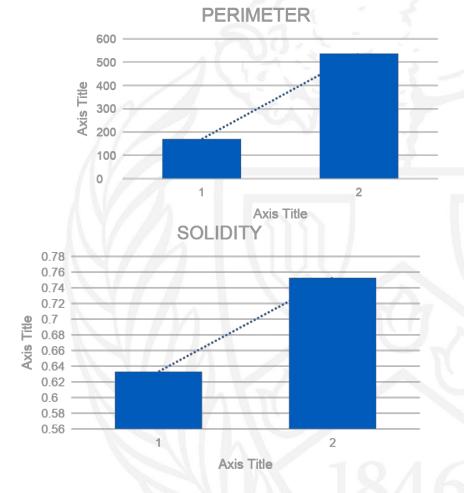




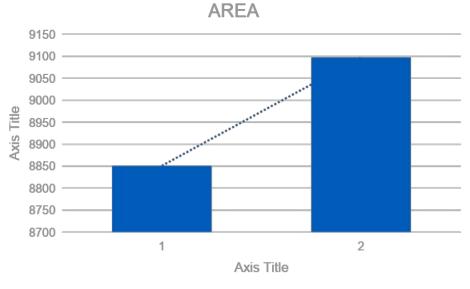
#### AVERAGE VALUE COMPARISON [ACTIN @ 24 HR]

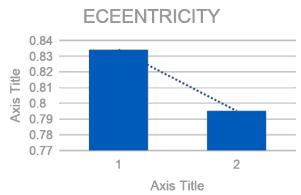


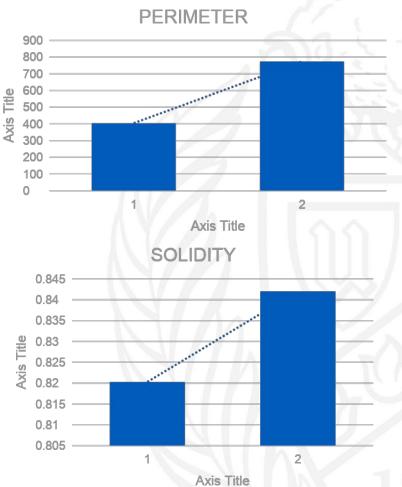




#### AVERAGE VALUE COMPARISON [TUBULIN @ 24 HR]





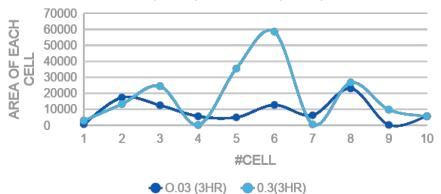


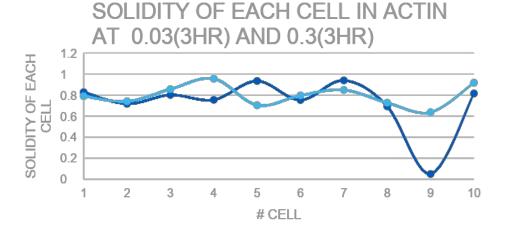
# SINGLE CELL VALUE IN ONE IMAGE COMPARISON



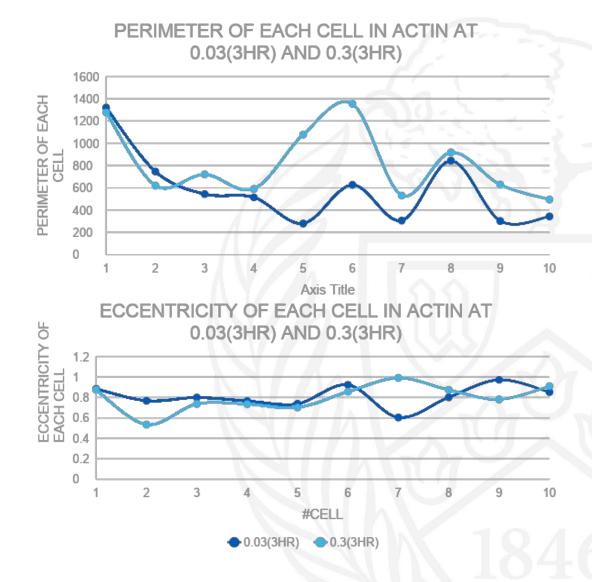
### ACTIN @ 3HR AREA OF EACH CELL IN ACTIN AT

AREA OF EACH CELL IN ACTIN AT 0.03(3HR) AND 0.3(3HR)



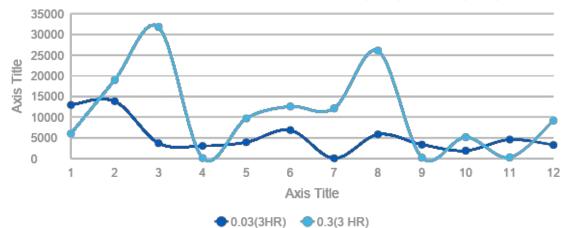


• 0.03(3HR)
• 0.3(3HR)

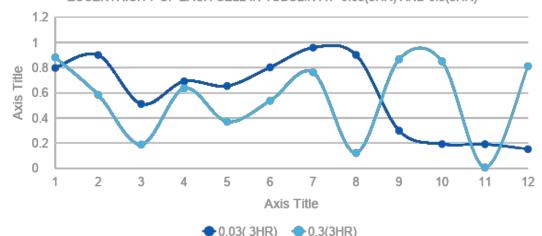


#### TUBULIN @ 3 HR

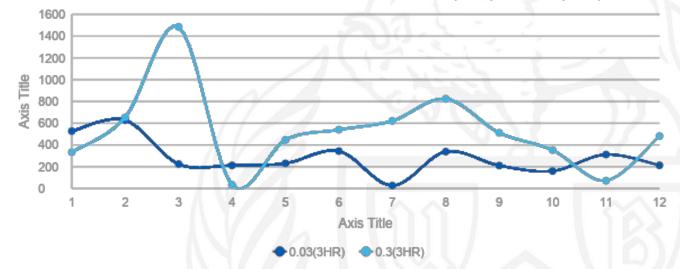




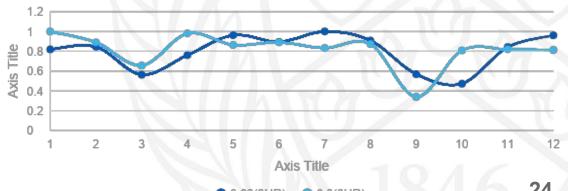
#### ECCENTRICITY OF EACH CELL IN TUBULIN AT 0.03(3HR) AND 0.3(3HR)



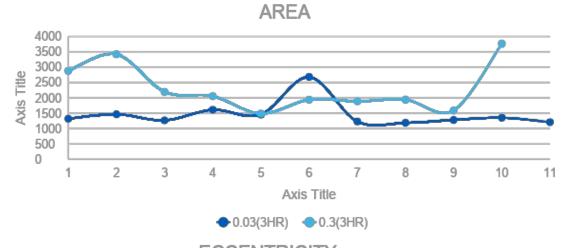
#### PERIMETER OF EACH CELL IN TUBULIN AT 0.03(3HR) AND 0.3(3HR)

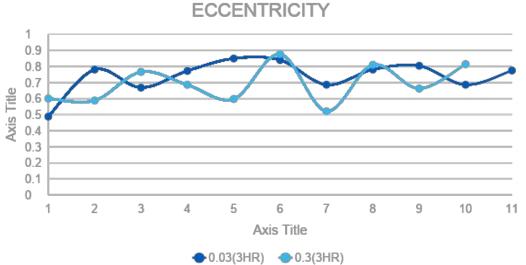


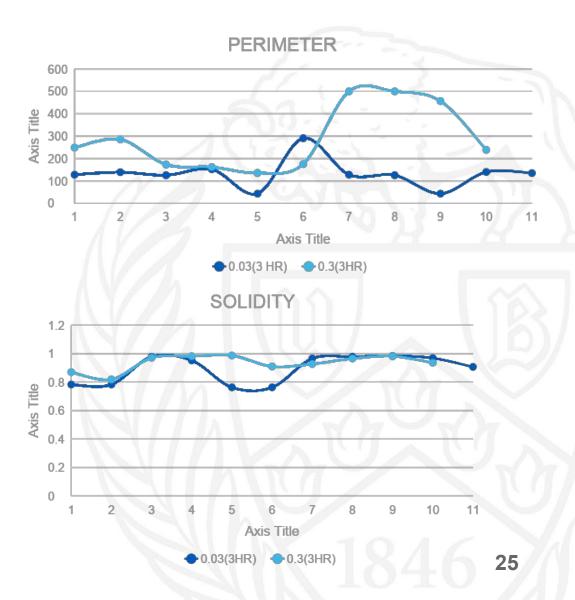
#### SOLIDITY OF EACH CELL INT UBULIN AT 0.03(3HR) AND 0.3(3HR)

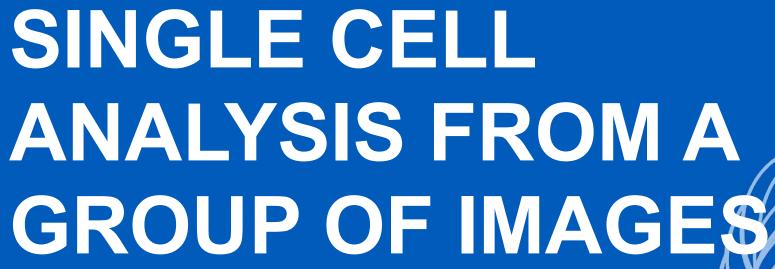


#### NUCLEUS @ 3 HR

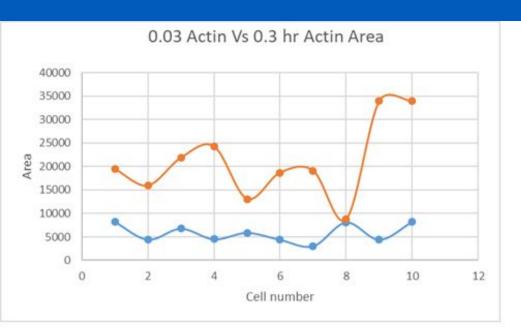


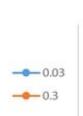


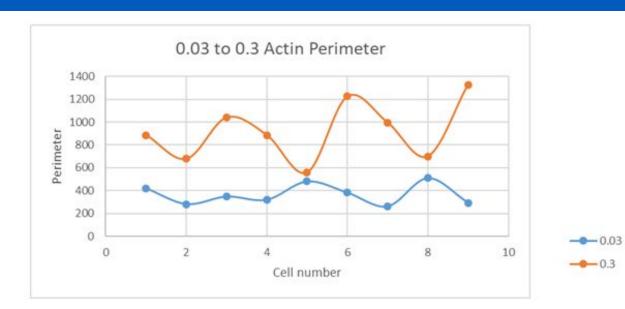


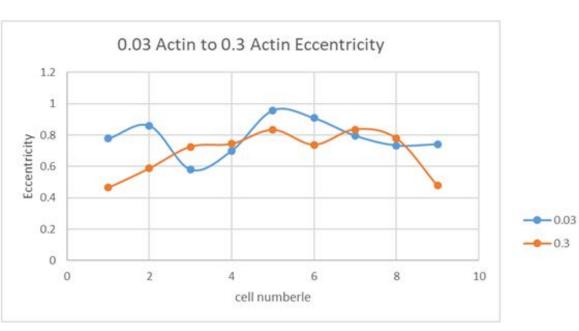


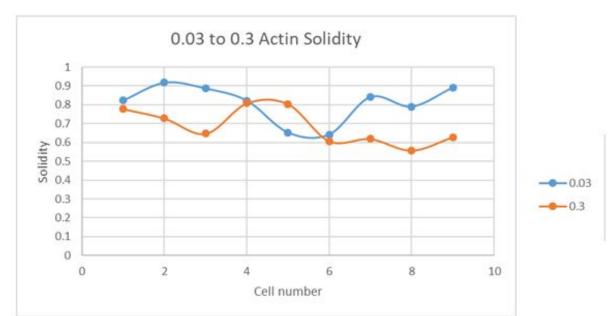




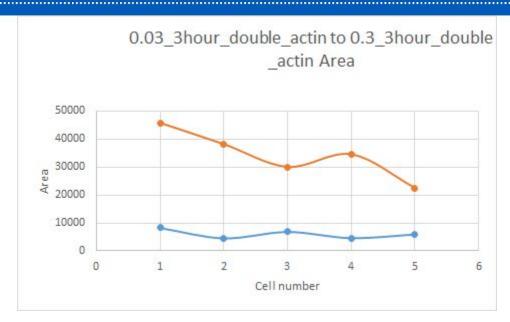


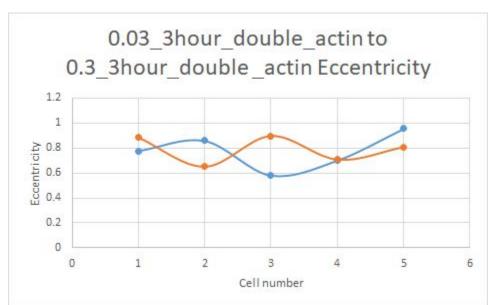


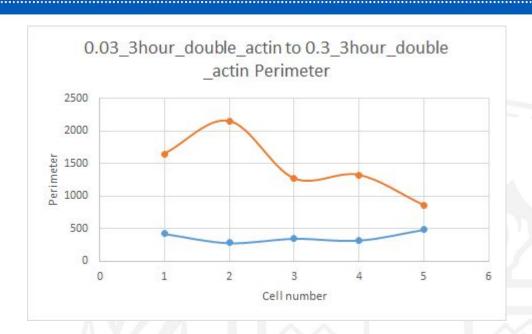


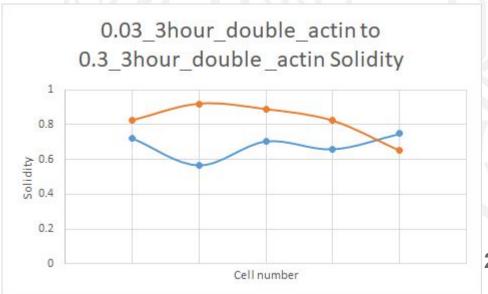


#### University at Buffalo The State University of New York

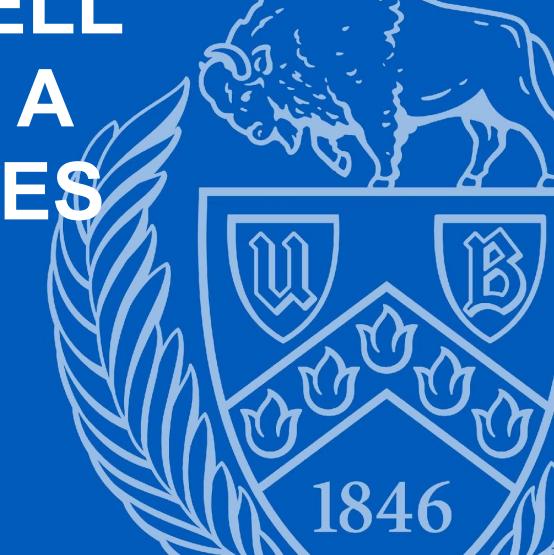




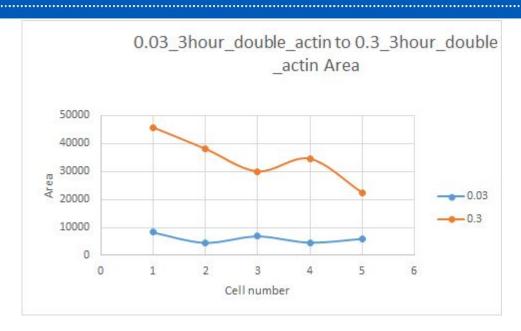


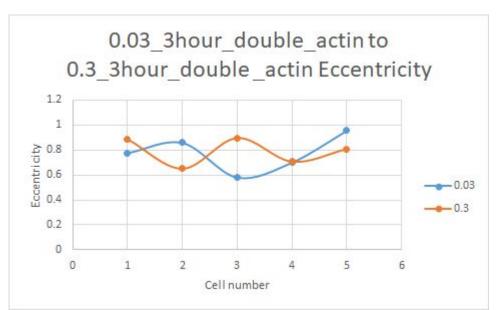


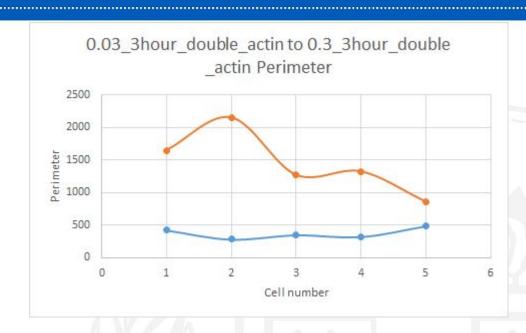


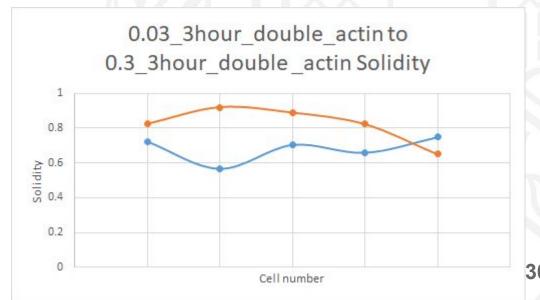


#### University at Buffalo The State University of New York









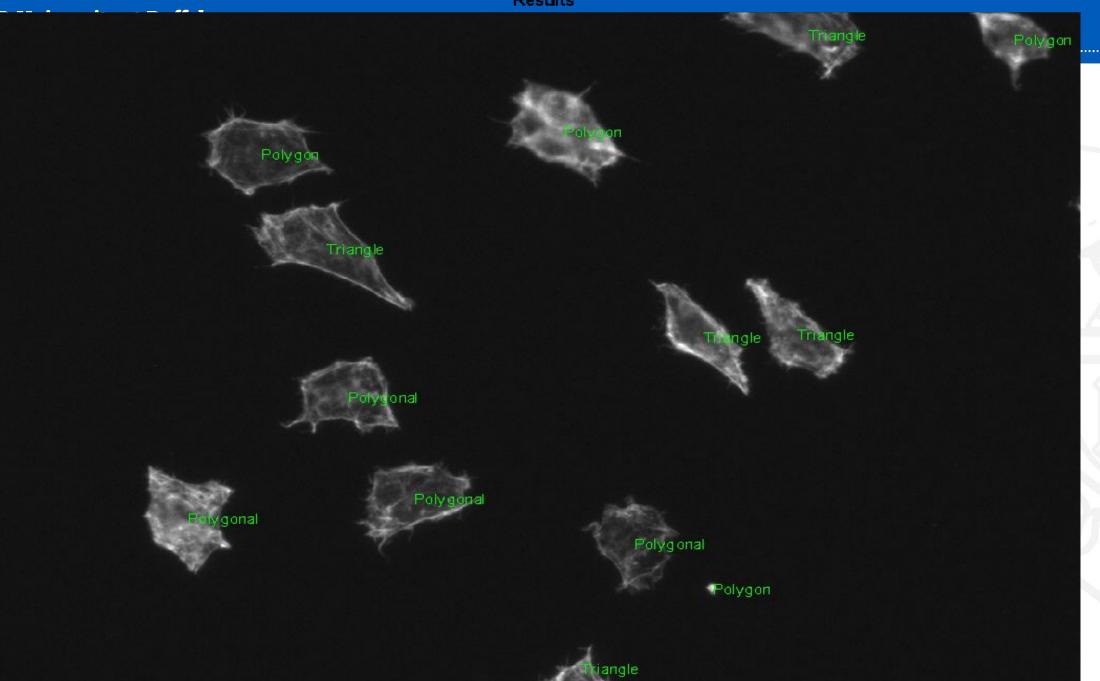
# DETERMINING THE SHAPE EACH CELL



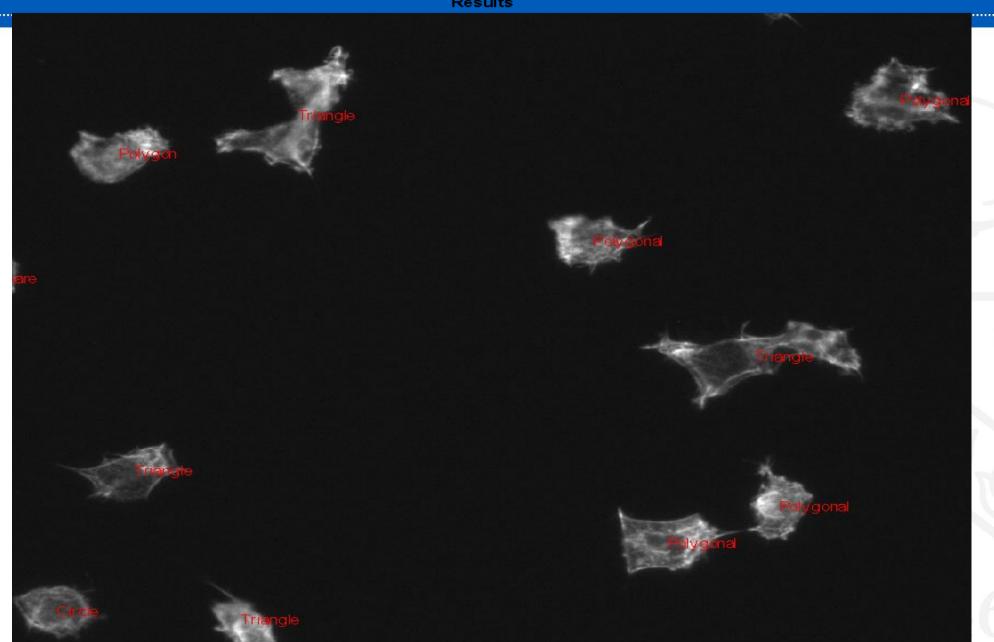
#### **Circularity**

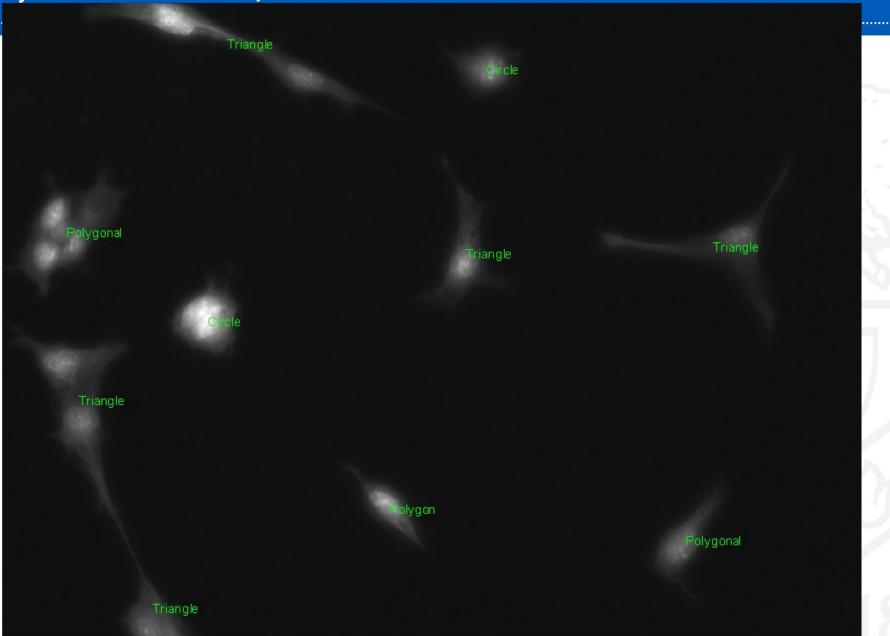
- Circularity is defined as the degree to which the particle is similar to a circle, taking into consideration the smoothness of the perimeter.
- Circularity is a measurement of both the particle form and roughness.
- Smooth circle a particle, the lower the circularity value.
- Circularity is a dimensionless value.





#### Results





# INFERENCE



#### CONCLUSIONS

#### **INFERENCE 1:**

- Average morphological feature values of all the cells in a single image has a higher variability at 0.3 stress level as compared to 0.03, irrespective of the time point and the type of cytoskeleton.
- Gradual increase in the morphological feature value can be explained as the time of each observation increases.

#### **INFERENCE 2:**

- Single cell in the same group shows varied morphological features, i.e. cells represent single cell heterogeneity.
- Each cell should be studied individually to elucidate the changes in the morphological features at various stress levels.

#### **INFERENCE 3:**

- Varied morphological features of single cell obtained from the image data set show single cell heterogeneity.
- This concludes that each cell in an image should be treated individually

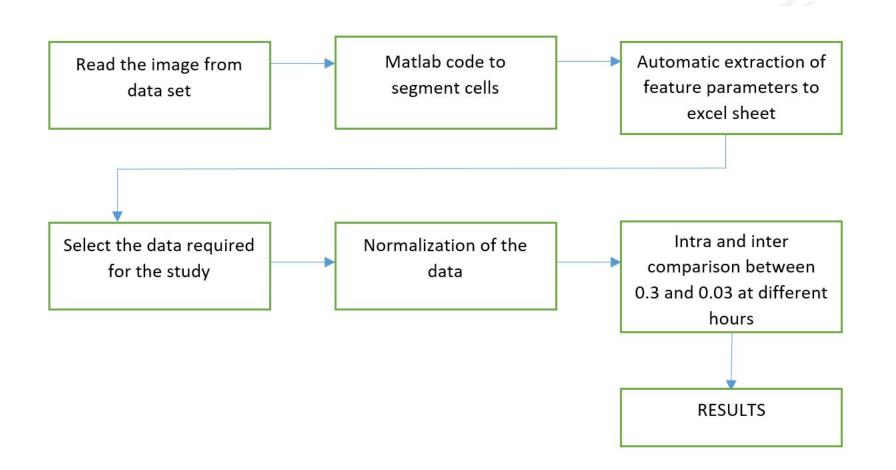
#### **INFERENCE 4:**

- All the cells in actin, nucleus and tubulin images show different shapes such as circle, polygon, hexagonal and triangle.
- This strengthens the existence of single cell heterogeneity.

#### **INFERENCE 5:**

• The single cell and overlapped cell show the same behavior when subjected to different stress at different time points.

PROCEDURE:



# FUTURE SCOPES



 Implementing machine learning algorithm to automatically segment the cells and classify the features.

• To study the effect of cell textures on single cell homogeneity.

# REFERENCES



ISO, ISO 13322-1, "Particle size analysis – Image analysis methods – Part 1: Static image analysis methods," 2004. 10.

ISO, ISO 13322-2, "Particle size analysis – Image analysis methods – Part 2: Dynamic image analysis methods," 2016.

Malvern Instruments Ltd, Morphologi G3 User Manual, 2010.

. Faria, Pons, Feyo De Azevedo, Rocha, & Vivier, "Quantification of the Morphology of Sucrose Crystals by Image Analysis", Powder Technology, 133, 2015.

Schmid, Dvorak, Müller, & Müssig, "Characterizing Flock Fibers using Quantitative Image Analysis", Flock, 30, 2009.