

# Day 2 – 04 Full Heatmap (Exercises Solution)

## Seminar reference solution

This solution notebook offers one way to complete the full heatmap exercise. Feel free to tweak palettes, ordering, or filtering thresholds to suit your teaching needs.

### 1. Packages, paths, helpers

```
suppressPackageStartupMessages({
  library(ComplexHeatmap)
  library(circlize)
  library(viridisLite)
})
subset_path <- file.path('..', 'data', 'dataset3_subset.csv')
long_path   <- file.path('..', 'data', 'dataset3_subset_long.csv')
pdf_path    <- file.path('..', 'pdf', '04_full_heatmap_exercise.pdf')
na_color    <- '#dcdcdc'
```

### 2. Load data and NA report

```
wide_df <- read.csv(subset_path, check.names = FALSE, stringsAsFactors = FALSE)
long_df <- read.csv(long_path, check.names = FALSE, stringsAsFactors = FALSE)
cat('Wide rows x cols:', nrow(wide_df), ncol(wide_df), '\n')

## Wide rows x cols: 71 67

cat('Long rows x cols:', nrow(long_df), ncol(long_df), '\n')

## Long rows x cols: 4544 7

na_total <- sum(is.na(long_df$value))
cat('NA count (value):', na_total, '\n')

## NA count (value): 443

if (na_total > 0) {
  na_table <- with(long_df, tapply(value, list(mouse_id, day), function(x) sum(is.na(x))))
  print(na_table)
}

##           0  4  9 14 18 23 30 37 44 49 53 58 63 67 72 79
## 1683      0  1  2  3  4  4  2  0  1  2  1  1  1  0  1  1
## 1688     15  1  1  2  3  0  1  1  0  0  0  1  0  0  1  1
## 1692      0 47 24  0 46 20  6  0  1  9 38  0  0 28  0  0
## 1699      0 23 37  0 24 13  0  0  1  1 23 13  1 23 13  1
```

### 3. Matrix + metadata

```
sample_cols <- setdiff(names(wide_df), c('Genome','snp_id','Position'))
mat <- as.matrix(wide_df[, sample_cols])
mode(mat) <- 'numeric'
rownames(mat) <- paste(wide_df$Genome, wide_df$snp_id, sep = ' | ')

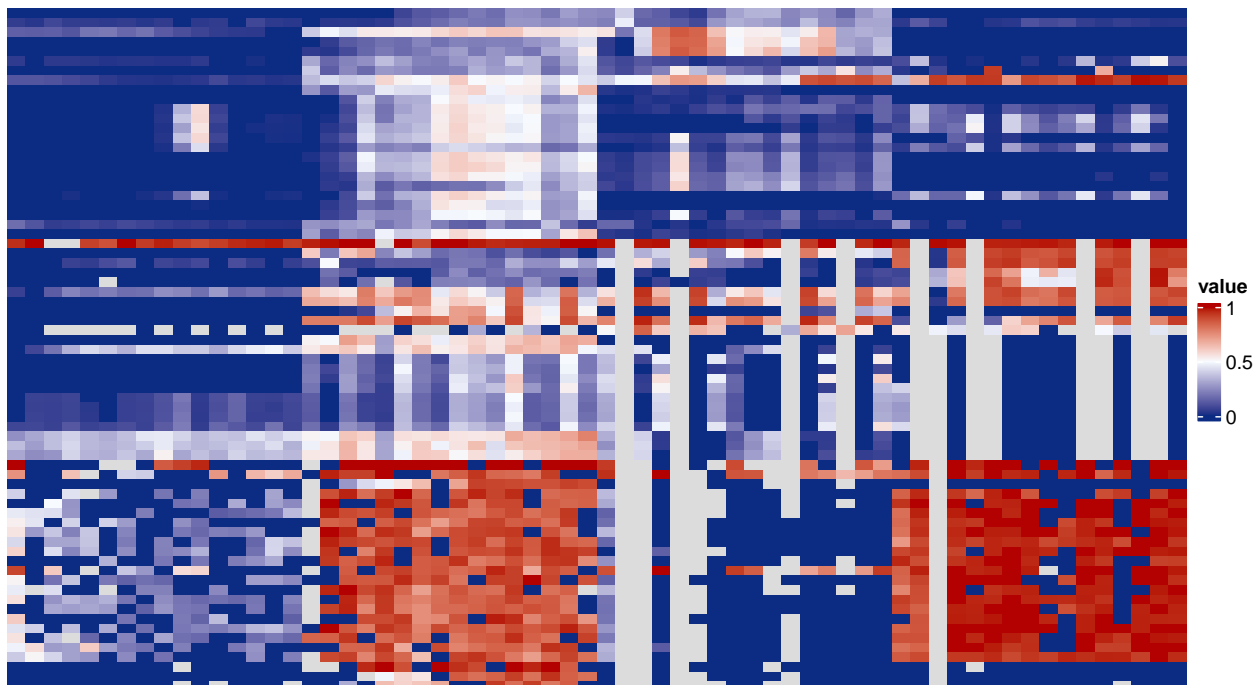
sample_meta <- unique(long_df[, c('mouse_id','day','treatment_group')])
sample_meta$sample_id <- paste(sample_meta$mouse_id, sample_meta$day, sep='-')
sample_meta <- sample_meta[match(colnames(mat), sample_meta$sample_id), ]
stopifnot(identical(colnames(mat), sample_meta$sample_id))
```

### 4. Baseline heatmap + palettes

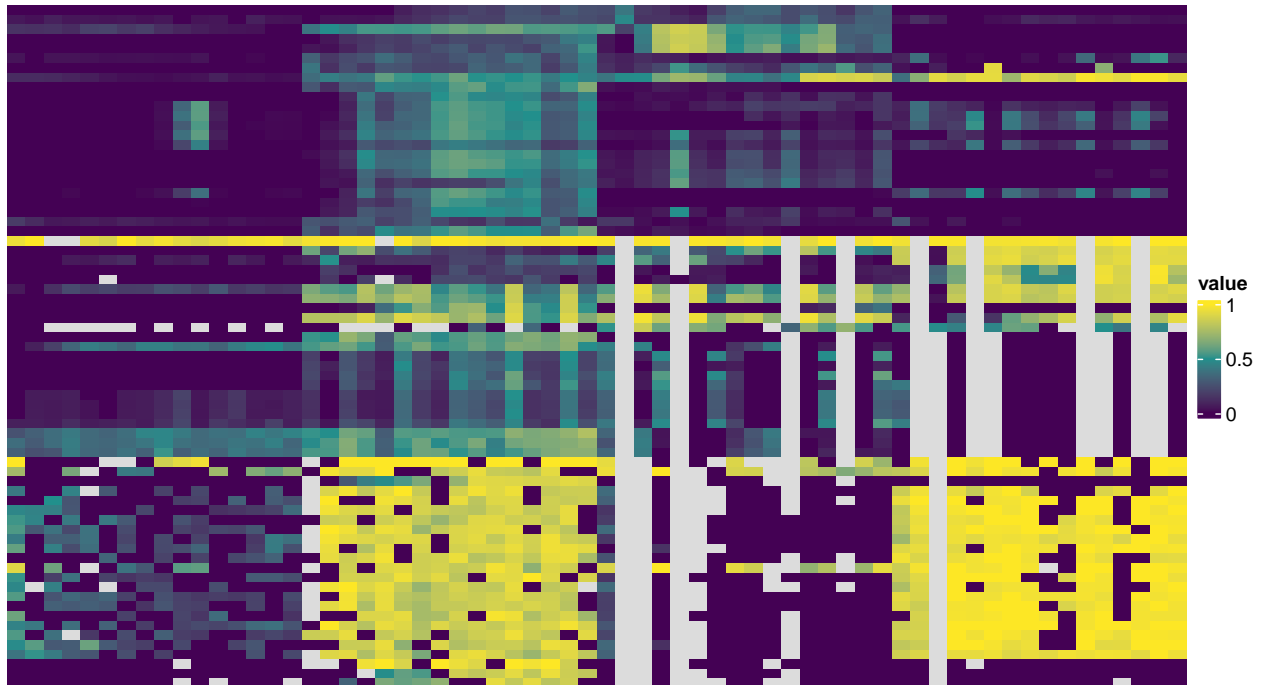
```
mins <- min(mat, na.rm = TRUE)
maxs <- max(mat, na.rm = TRUE)
mids <- (mins + maxs) / 2
palette_blue_red <- circlize::colorRamp2(c(mins, mids, maxs), c('#0c2c84','#f7fbff','#b30000'))
palette_viridis <- circlize::colorRamp2(c(mins, mids, maxs), viridisLite::viridis(3))

ht_blue <- Heatmap(mat, name = 'value', col = palette_blue_red, na_col = na_color,
  cluster_rows = FALSE, cluster_columns = FALSE,
  show_row_names = FALSE, show_column_names = FALSE)
ht_viridis <- Heatmap(mat, name = 'value', col = palette_viridis, na_col = na_color,
  cluster_rows = FALSE, cluster_columns = FALSE,
  show_row_names = FALSE, show_column_names = FALSE)

ht_blue
```



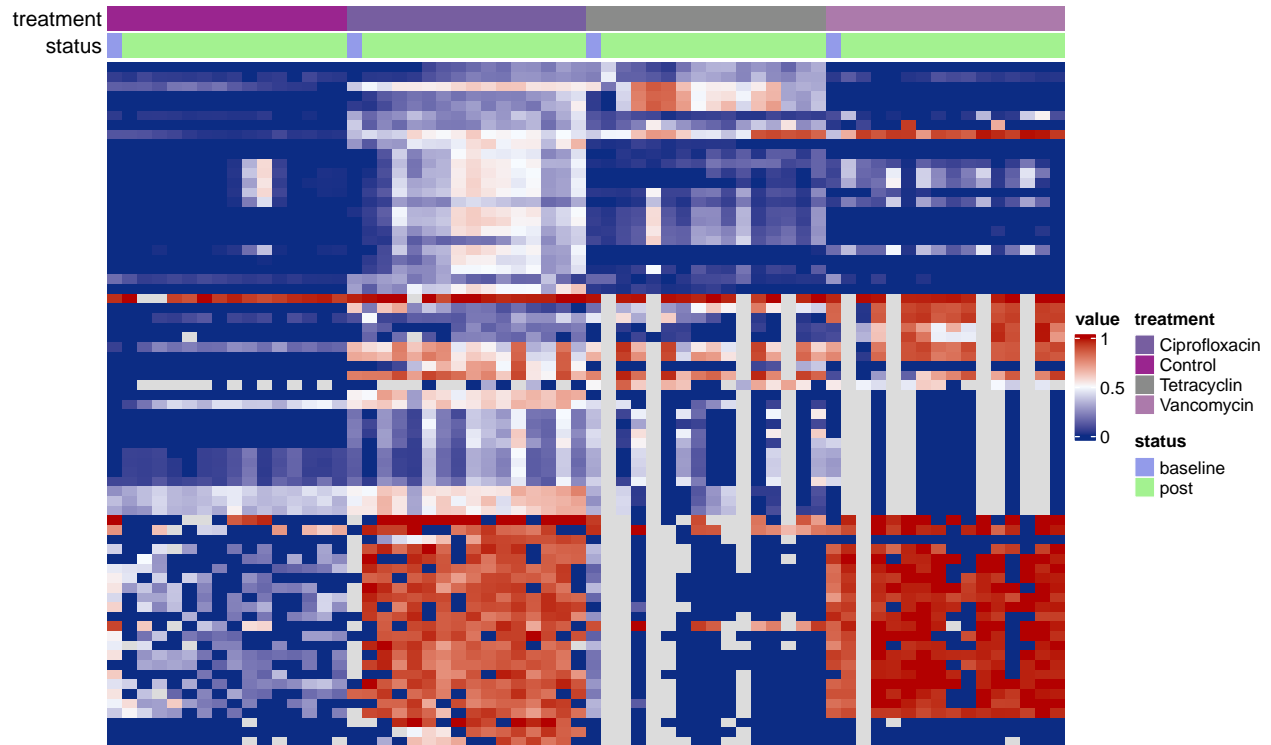
ht\_viridis



## 5. Annotations + ordering

```
sample_meta$post_ab <- ifelse(sample_meta$day == 0, 'baseline', 'post')
col_ann <- HeatmapAnnotation(
  treatment = sample_meta$treatment_group,
  status = sample_meta$post_ab,
  annotation_name_side = 'left'
)
order_idx <- order(sample_meta$mouse_id, sample_meta$day)
mat_ordered <- mat[, order_idx]
col_ann_ordered <- col_ann[order_idx]

Heatmap(
  mat_ordered,
  name = 'value',
  col = palette_blue_red,
  top_annotation = col_ann_ordered,
  cluster_rows = FALSE,
  cluster_columns = FALSE,
  show_row_names = FALSE,
  show_column_names = FALSE,
  na_col = na_color
)
```



## 6. Annotation enhancements

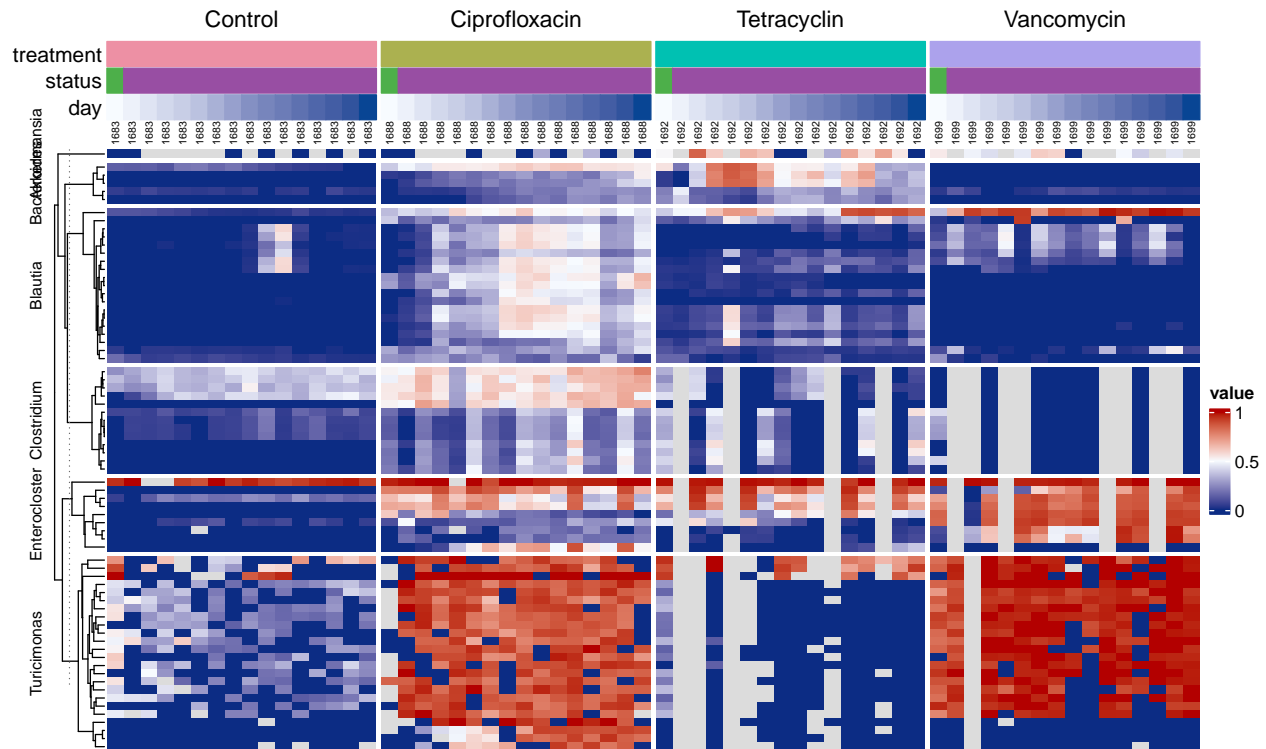
```
treatment_levels <- unique(sample_meta$treatment_group)
treatment_cols <- grDevices::hcl.colors(length(treatment_levels), palette = "Set2")
names(treatment_cols) <- treatment_levels
status_cols <- c(baseline = '#4daf4a', post = '#984ea3')
day_col_fun <- circlize::colorRamp2(range(sample_meta$day), c('#f7fbff', '#084594'))
col_ann_rich <- HeatmapAnnotation(
  treatment = anno_simple(sample_meta$treatment_group, col = treatment_cols),
  status = anno_simple(sample_meta$post_ab, col = status_cols),
  day = anno_simple(sample_meta$day, col = day_col_fun),
  mouse = anno_text(sample_meta$mouse_id, rot = 90, gp = grid::gpar(fontsize = 6)),
  annotation_name_side = 'left'
)
col_ann_rich_ordered <- col_ann_rich[order_idx]
column_split <- factor(sample_meta$treatment_group[order_idx], levels = treatment_levels)
row_split <- factor(wide_df$Genome, levels = unique(wide_df$Genome))
row_titles <- sub('_.*', '', levels(row_split))

Heatmap(
  mat_ordered,
  name = 'value',
  col = palette_blue_red,
  top_annotation = col_ann_rich_ordered,
  cluster_rows = TRUE,
  cluster_columns = FALSE,
  column_split = column_split,
  show_row_names = FALSE,
  show_column_names = FALSE,
```

```

na_col = na_color,
row_split = row_split,
row_title = row_titles,
row_title_gp = grid::gpar(fontsize = 9)
)

```

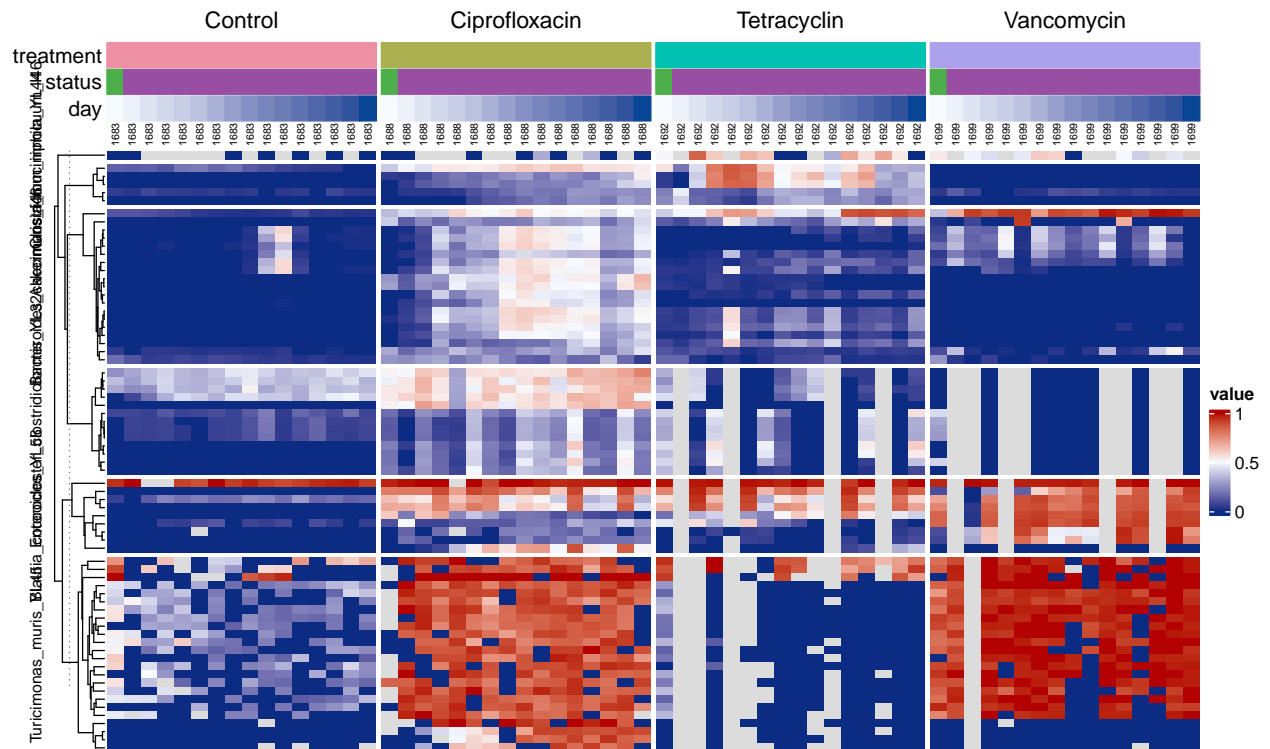


## 7. Row variance filter

```

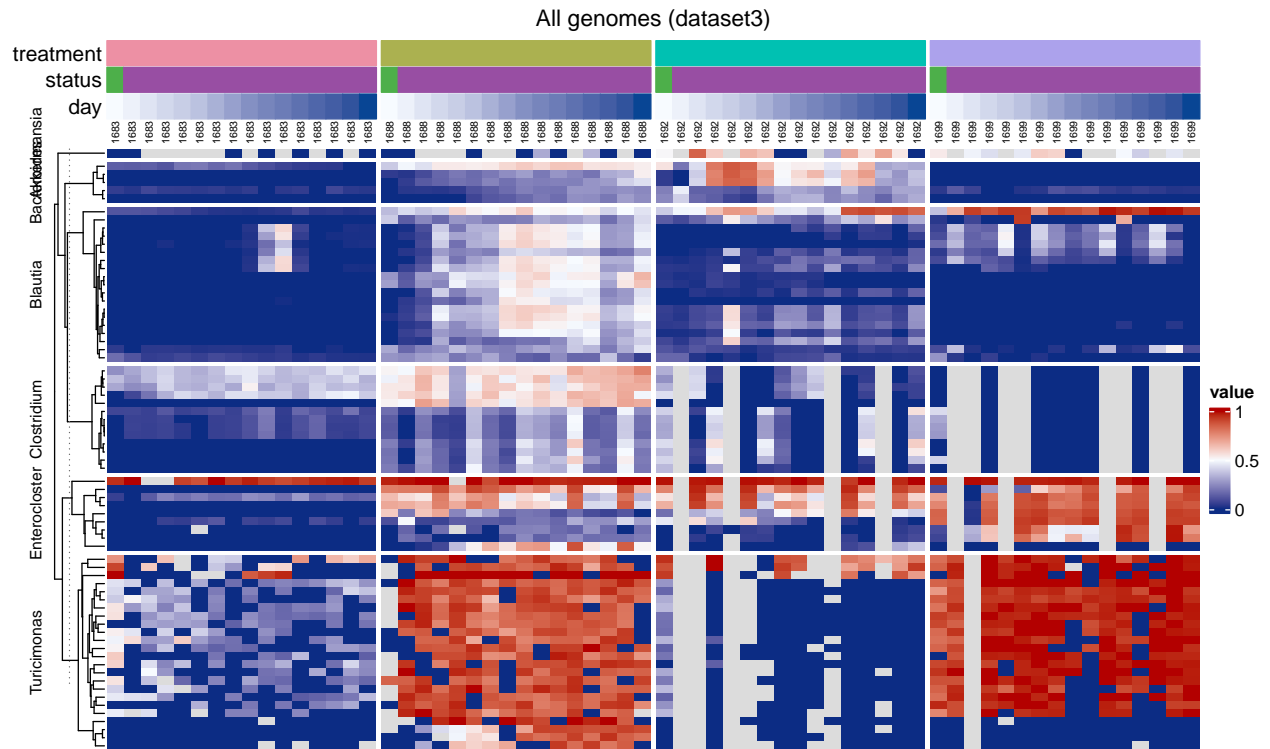
row_var <- apply(mat, 1, var, na.rm = TRUE)
keep_idx <- order(row_var, decreasing = TRUE)[seq_len(min(100, nrow(mat)))]
mat_topvar <- mat[keep_idx, order_idx]
row_split_top <- droplevels(row_split[keep_idx])
Heatmap(
  mat_topvar,
  name = 'value',
  col = palette_blue_red,
  top_annotation = col_ann_rich_ordered,
  cluster_rows = TRUE,
  cluster_columns = FALSE,
  column_split = column_split,
  show_row_names = FALSE,
  show_column_names = FALSE,
  na_col = na_color,
  row_split = row_split_top,
  row_title_gp = grid::gpar(fontsize = 9)
)

```



## 8. Final heatmap + PDF export

```
ht_final <- Heatmap(
  mat_ordered,
  name = 'value',
  col = palette_blue_red,
  top_annotation = col_ann_rich_ordered,
  cluster_rows = TRUE,
  cluster_columns = FALSE,
  column_split = column_split,
  show_row_names = FALSE,
  show_column_names = FALSE,
  na_col = na_color,
  row_split = row_split,
  row_title = row_titles,
  row_title_gp = grid::gpar(fontsize = 9),
  column_title = 'All genomes (dataset3)'
)
draw(ht_final)
```



```
pdf(pdf_path, width = 11, height = 7)
draw(ht_final)
dev.off()
cat('Saved heatmap to', pdf_path, '\n')
```

## 9. Notes

- Palette choice: the blue-white-red ramp emphasizes deviations from mid values.
- Column ordering by mouse/day reveals antibiotic pulses more clearly than the CSV default.
- Variance filtering is helpful when presenting in class; it trims the figure to the most dynamic SNPs and speeds up PDF export.
- Extra annotations (day gradient + mouse labels + row splits) plus column splits by treatment mirror the annotation tutorial and keep the story tied to the experimental design.