

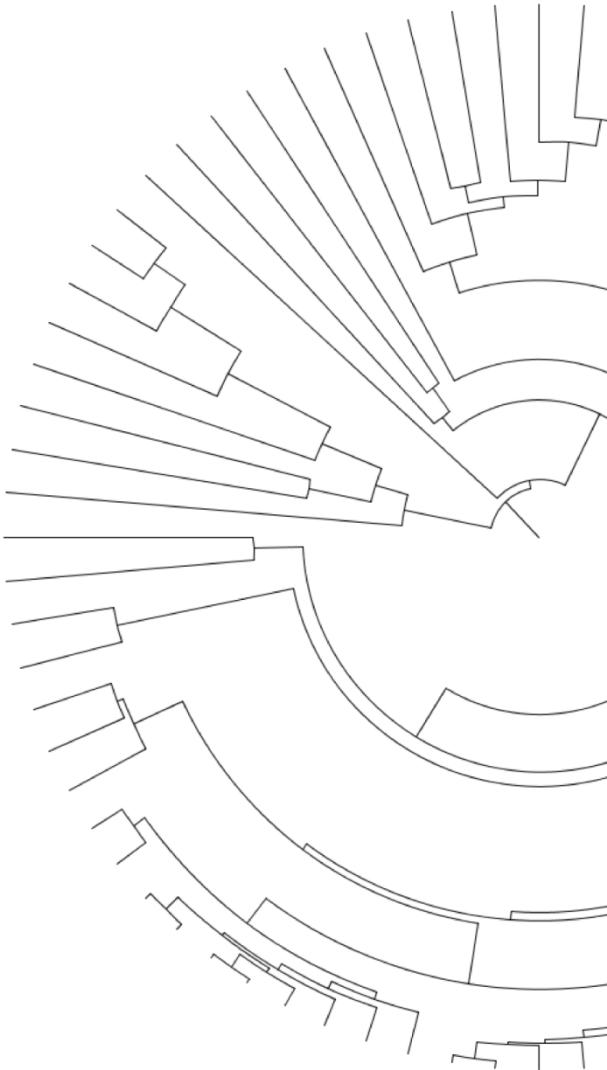
Large-scale gene family analysis of 76 Arthropods

i5K webinar / September 5, 2018

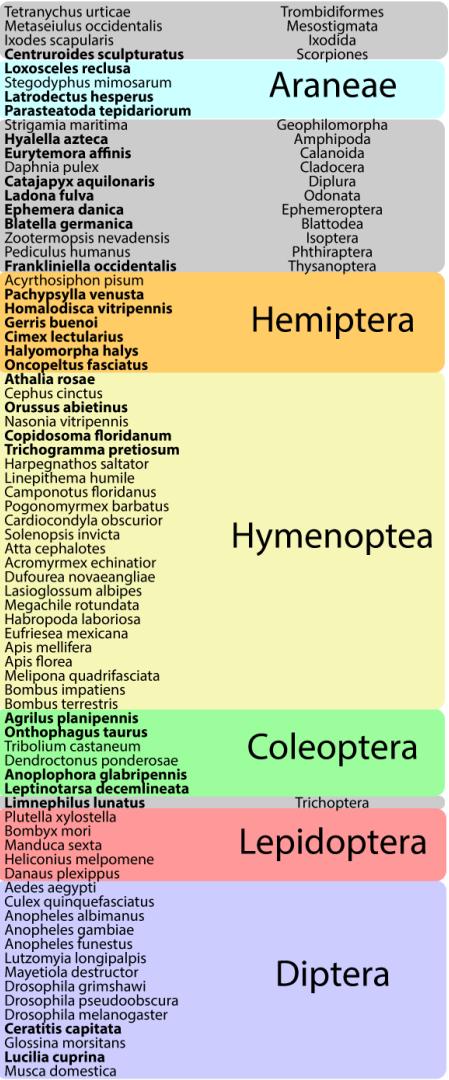
Gregg Thomas

 @greggwcthomas

Indiana University



76 species



The genomic basis of Arthropod diversity

<https://www.biorxiv.org/content/early/2018/08/04/382945>

Why and how did we do it?

21 orders

76 species

Tetranychus urticae
Metaseiulus occidentalis
Ixodes scapularis
Centruroides sculpturatus
Loxosceles reclusa
Stegodyphus mimarorum
Latrodectus hesperus
Parasteatoda tepidariorum
Strigamia maritima
Hyailella azteca
Eurytemora affinis
Daphnia pulex
Catajapyx aquilonaris
Ladona fulva
Ephemera danica
Blaetilia germanica
Zootermopsis nevadensis
Pediculus humanus
Frankliniella occidentalis
Acyrthosiphon pisum
Pachypsylus venusta
Homalodisca vitripennis
Gerris buenoi
Cimex lectularius
Halymomorpha halys
Oncomeltus fasciatus
Athalia rosae
Cephus cinctus
Orussus abietinus
Nasonia vitripennis
Copidosoma floridanum
Trichogramma pretiosum
Harpagnothos saltator
Linepithema humile
Camponotus floridanus
Pogonomyrmex barbatus
Cardiomyrmex obscurior
Solenopsis invicta
Atta cephalotes
Acromyrmex echinatior
Dufourea novaeangliae
LasioGLOSSUM albipes
Megachile rotundata
Habropoda laboriosa
Eufriesea mexicana
Apis mellifera
Apis florea
Melipona quadrifasciata
Bombus impatiens
Bombus terrestris
Agrilus planipennis
Oncophagus taurus
Tribolium castaneum
Dendroctonus ponderosae
Anoplophora glabripennis
Leptinotarsa decemlineata
Limnephilus lunatus
Plutella xylostella
Bombyx mori
Manduca sexta
Heliconius melpomene
Danaua plexippus
Aedes aegypti
Culex quinquefasciatus
Anopheles albimanus
Anopheles gambiae
Anopheles funestus
Lutzomyia longipalpis
Mayetiolia destructor
Drosophila grimshawi
Drosophila pseudoobscura
Drosophila melanogaster
Ceratitis capitata
Glossina morsitans
Lucilia cuprina
Musca domestica

Trombidiformes
Mesostigmata
Ixodida
Scorpiones

Araneae

Geophilomorpha
Amphipoda
Calanoida
Cladocera
Diplura
Odonata
Ephemeroptera
Blattodea
Isoptera
Phthiraptera
Thysanoptera

Hemiptera

21 orders

Hymenoptera

Coleoptera

Trichoptera

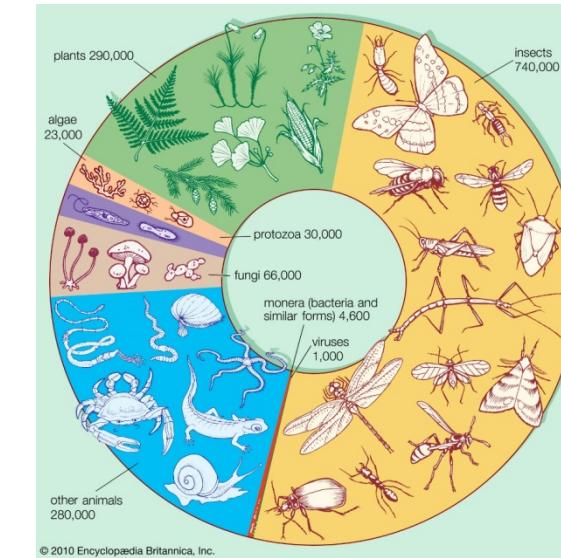
Lepidoptera

Diptera

The genomic basis of Arthropod diversity

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Why and how did we do it?



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Why and how did we do it?



FiveThirtyEight

Politics Sports Science & Health Economics Culture

MAY 2, 2017 AT 10:00 AM

The Bugs Of The World Could Squish Us All

And we'd deserve it.

By [Maggie Koerth-Baker](#)

Filed under [Science Question From A Toddler](#)



<https://fivethirtyeight.com/features/the-bugs-of-the-world-could-squish-us-all/>

Arthropods exhibit vast phenotypic diversity

Aedes aegypti
(Diptera)



Bombus terrestris *Latrodectus hesperus*
(Hymenoptera) (Araneae)



Whole-genome sequencing reveals vast molecular differences

...AAGGCCA...

Aedes aegypti
(Diptera)



...AAGTCCA...

Bombus terrestris *Latrodectus hesperus*
(Hymenoptera) (Araneae)



...AAGTCCA...

(Araneae)



Nucleotide substitutions

Whole-genome sequencing reveals vast molecular differences

4

4

2

Gene copy number variation

...AAGGCCA...

...AAGTCCA...

...AAGTCCA...

Nucleotide substitutions

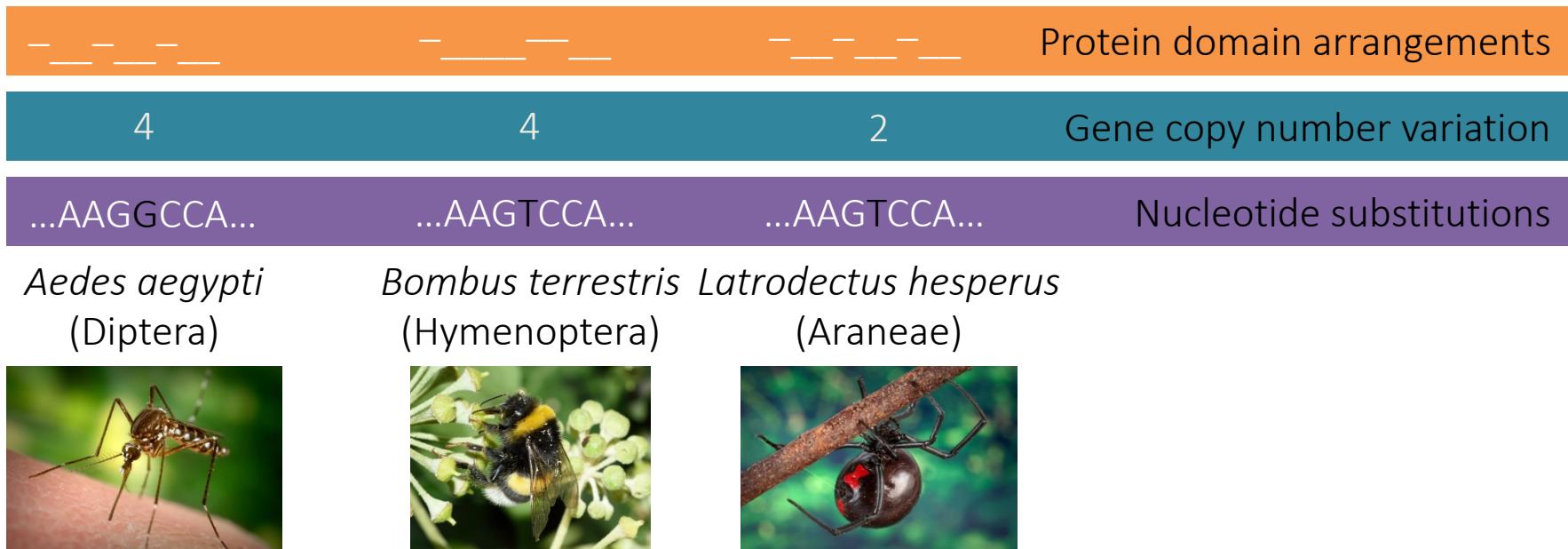
Aedes aegypti
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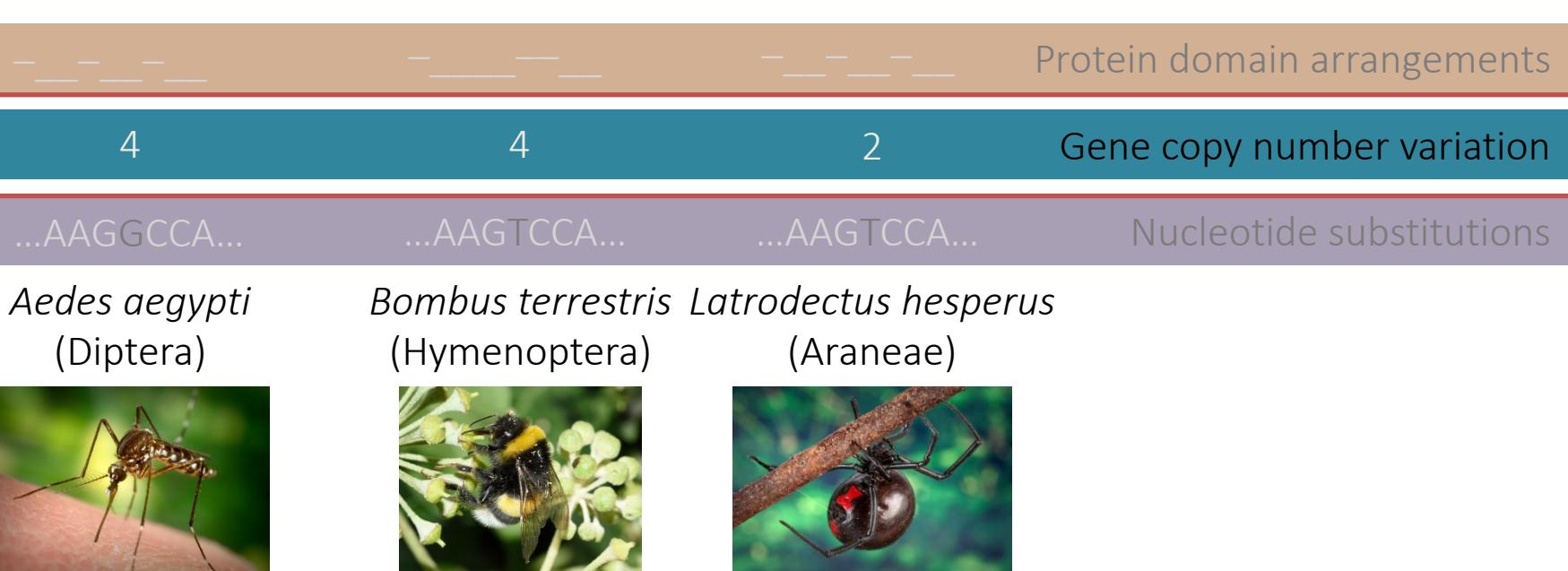
Bombus terrestris *Latrodectus hesperus*
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Whole-genome sequencing reveals vast molecular differences



Whole-genome sequencing reveals vast molecular differences



Whole-genome sequencing reveals vast molecular differences



...AAGGCCA...

Aedes aegypti
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Bombus terrestris *Latrodectus hesperus*
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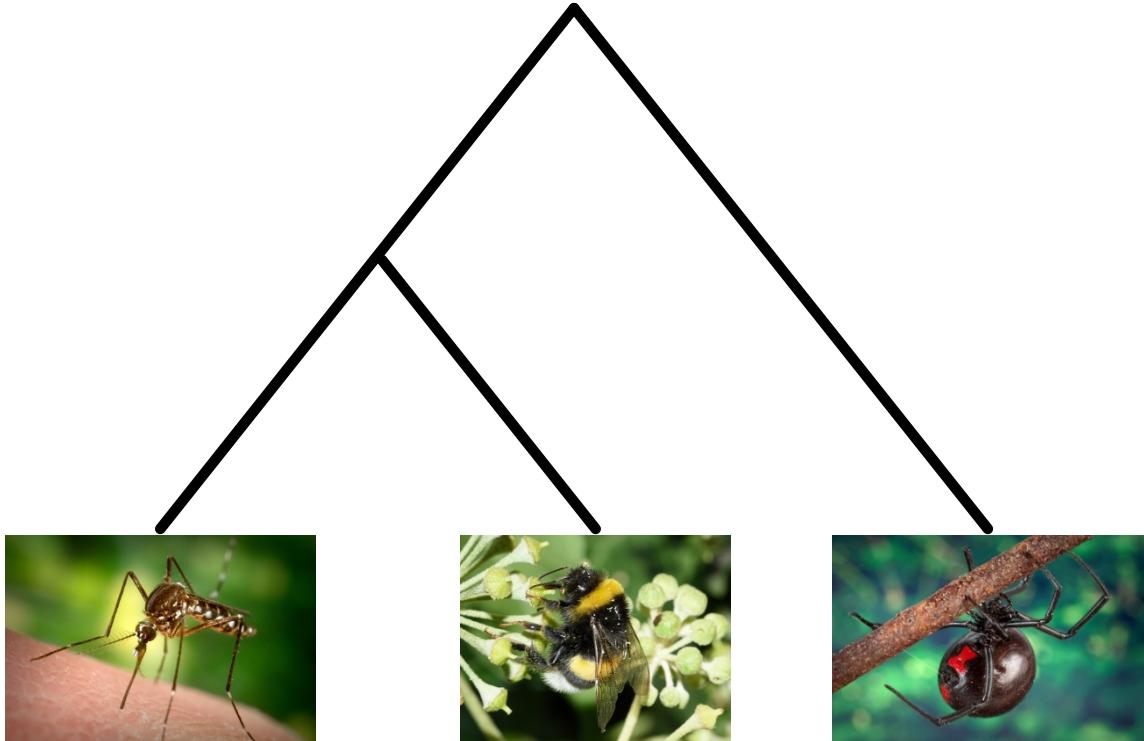
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Latrodectus hesperus
(Araneae)



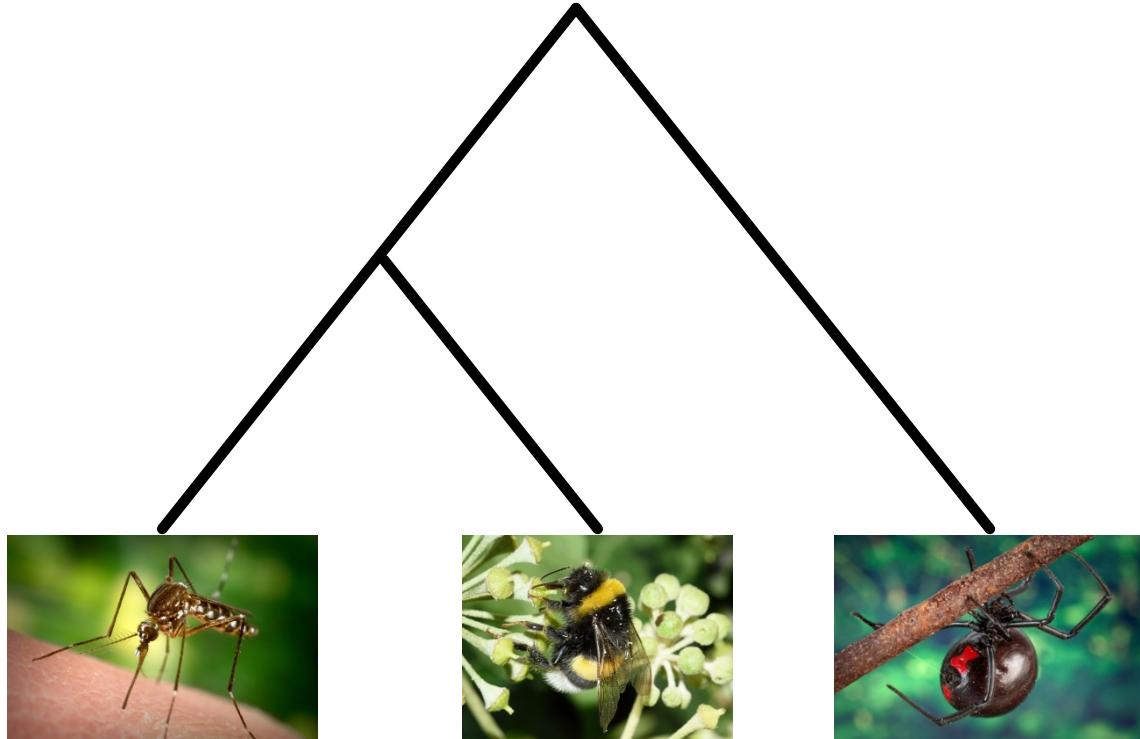
How can we begin to understand what changes are interesting or important?

Phylogenies act as a framework for asking these types of questions



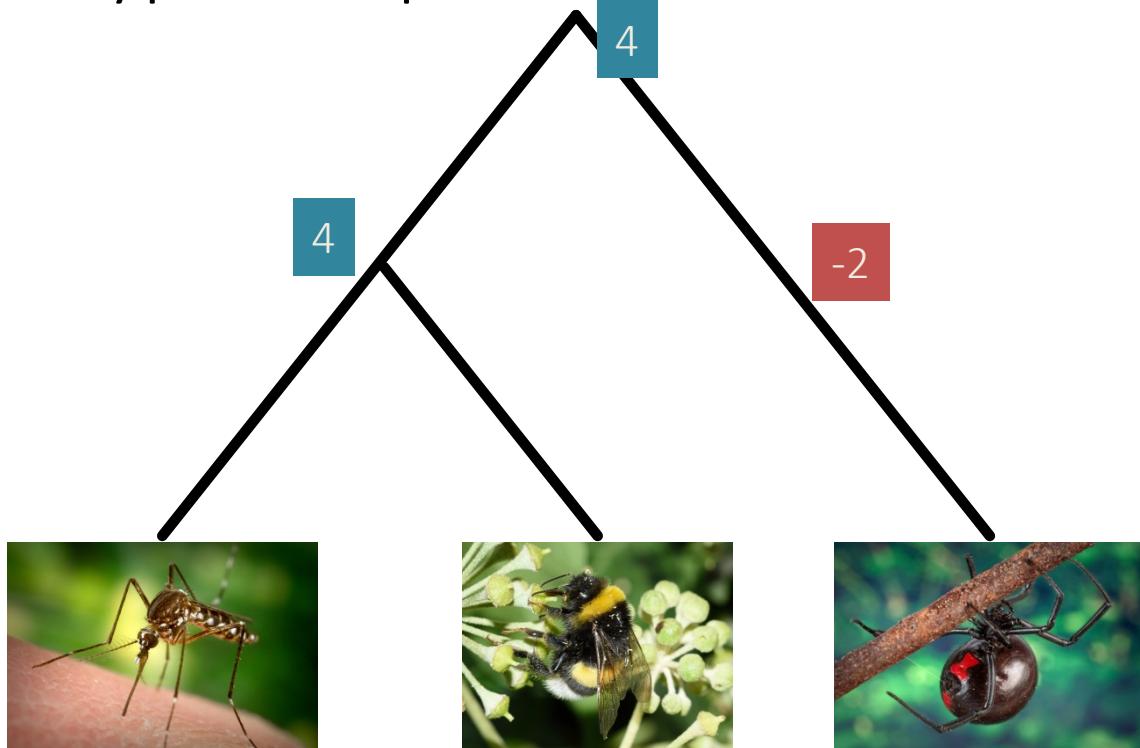
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How can we begin to understand what changes are interesting or important?

Which molecular changes lead to interesting phenotypic differences?

1. Sequence and annotate many genomes

Stephen Richards
Monica Poelchau

Which molecular changes lead to interesting phenotypic differences?

1. Sequence and annotate many genomes
2. Determine orthology of sequences

Stephen Richards
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Rob Waterhouse
Evgeny Zdobnov
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Which molecular changes lead to interesting phenotypic differences?

1. Sequence and annotate many genomes
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3. Determine phylogeny of species

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3. Determine phylogeny of species
4. Map orthology onto phylogeny to reconstruct the evolutionary history of all loci

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Rob Waterhouse
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Panagiotis Ioannidis

Elias Dohmen
Karl Glastad
Yiyuan Li

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Today's topics

1. Determining the Arthropod phylogeny
2. Reconstructing ancestral gene counts
3. Using the i5k gene family web site

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2. Reconstructing ancestral gene counts
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1) Predict orthogroups

1	C A A T G C G A A A T G C G B A A T G C G A A A T G C G
---	--

4	C A T A T A T C A A A T A T T A B A T A
---	---

2	C T T C A A G A A T T A A G B A T T A C G
---	---

5	C C G A A A A C G A T C A B C G T T C A A C G A C A B C G A C A
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2) Select single-copy groups

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3) Align each group

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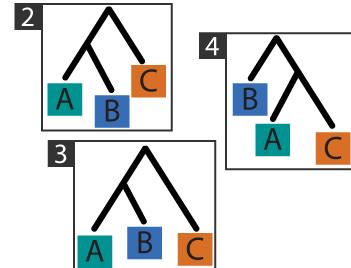
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4) Infer gene trees



1) Predict orthogroups

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3	CCGAAA ACGATCA BCGTTCA	CGAAG AATTAAG BATTACG

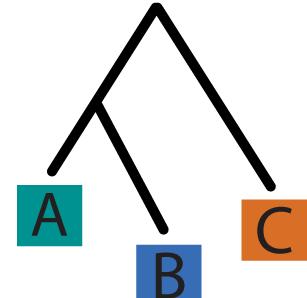
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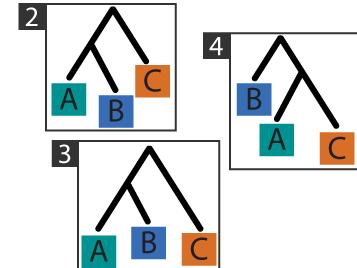
3) Align each group

2	CTTCAAG AATTAAG BATTACG
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5) Infer species tree



4) Infer gene trees



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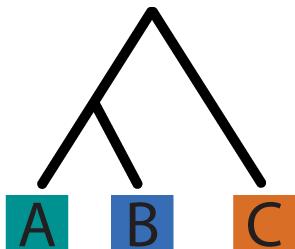
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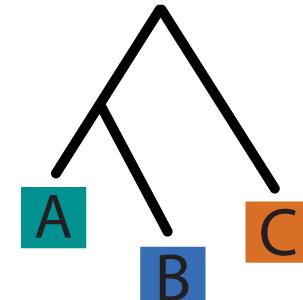
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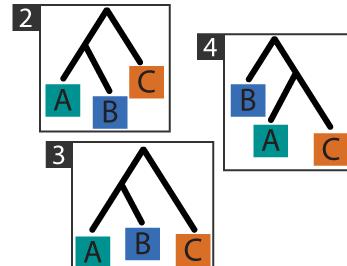
6) Scale branch lengths with fossil calibrations



5) Infer species tree



4) Infer gene trees



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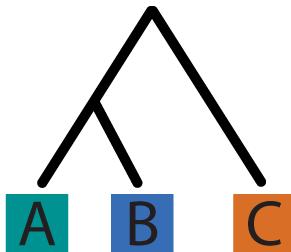
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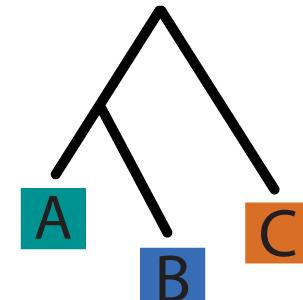
3) Align each group

2	CTTCAAG AATTAAG BATTACG
3	CCGAA-A ACGATCA BCGTTCA
4	CATATCA AATATTAA BATA ---

6) Scale branch lengths with fossil calibrations



5) Infer species tree



4) Concatenate alignments

2	CTTCAAG	CCGAA	-AATATCA
3	AATTAAG	CGATCA	AATATTAA
4	BATTACG	CGTTCA	AATA---

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1	CAATGCG AAATGCG BAATGCG AAATGCG	CATATCA AATATTAA BATA
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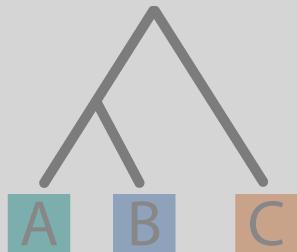
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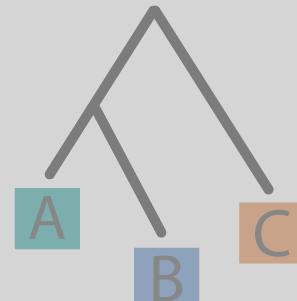
What are the details of these 6 steps in the context of the i5k project?



6) Scale branch lengths with fossil calibrations



5) Infer species tree



4) Concatenate alignments

2	CTTCAAGCGAA-AATATCA
3	AATTAAGCGATCAATATTAA
4	BATTACGCGTTCAATA---

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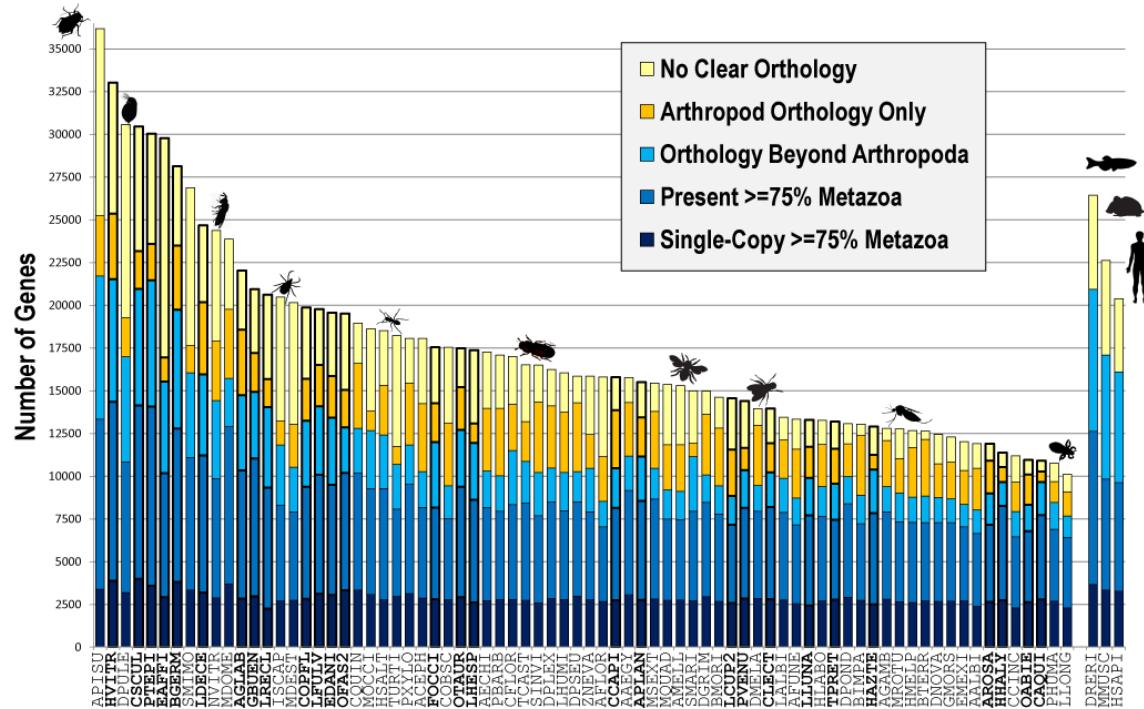
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1) Predict orthogroups – OrthoDB



Rob Waterhouse
Evgeny Zdobnov
Panagiotis Ioannidis

38,195 ortho-groups across 76 arthropod species

(See i5k webinar from Feb. 1, 2017: <http://i5k.github.io/webinar>)

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2) Select single-copy orthogroups

How many single-copy orthologs in our 38,195 groups?



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How many single-copy orthologs in our 38,195 groups?

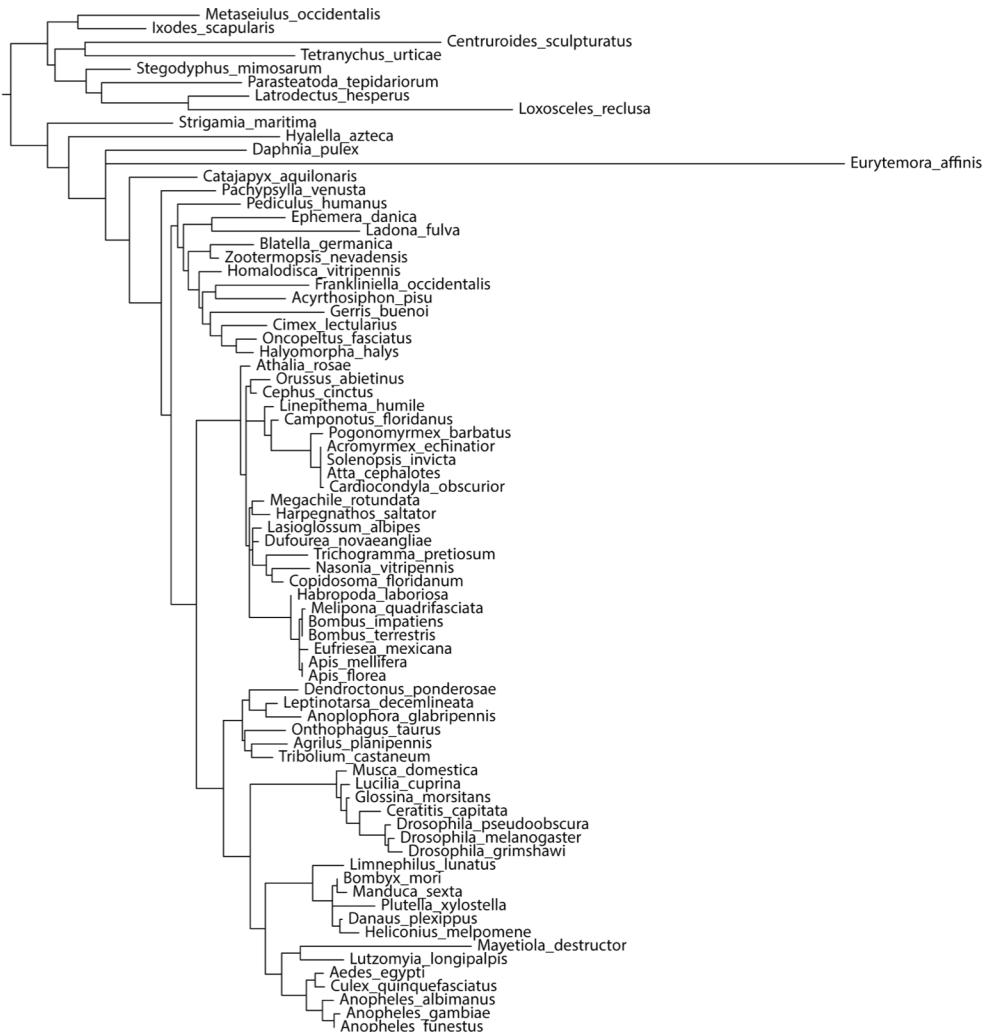
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1 family single copy in all but one species
(2 copies in *Plutella xylostella*)



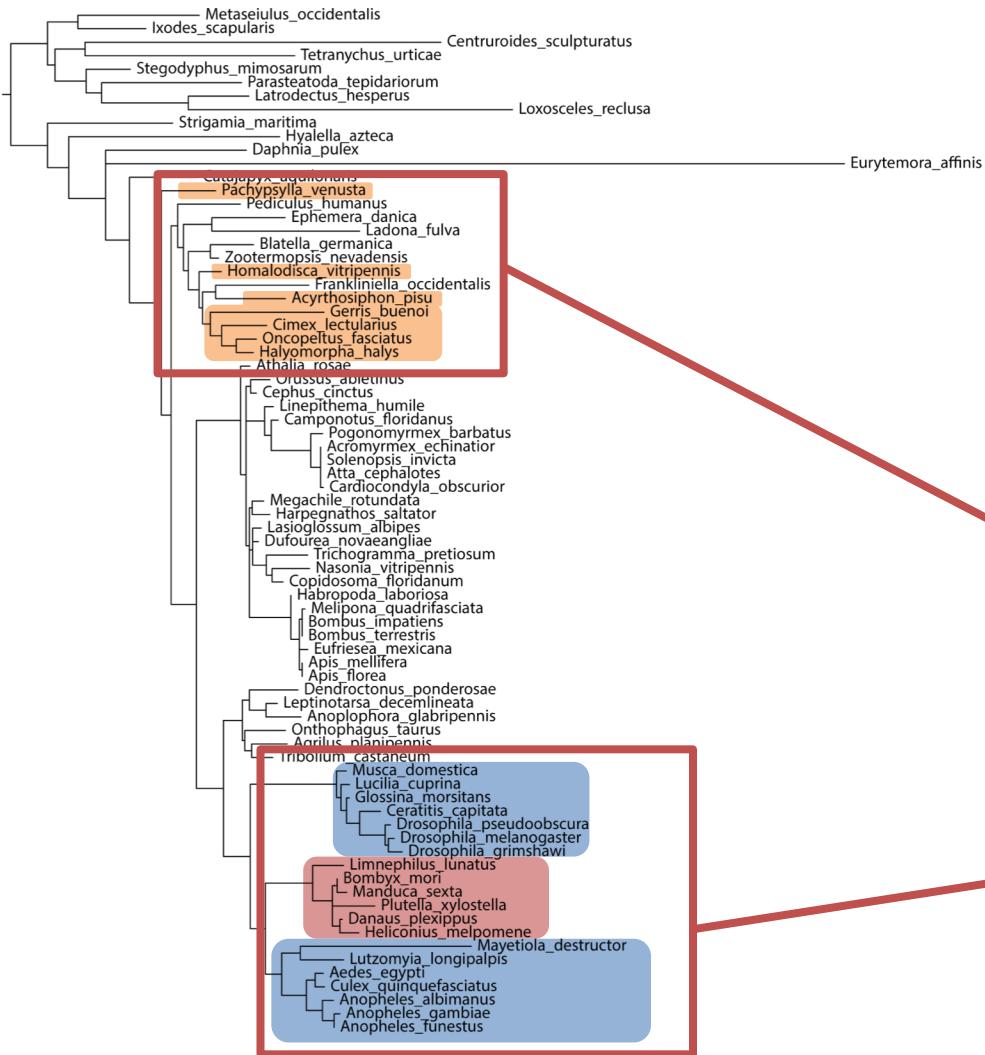
EOG8DFS3J

Single-copy in all but one species
(2 copies in *Plutella xylostella*)





EOG8DFS3J



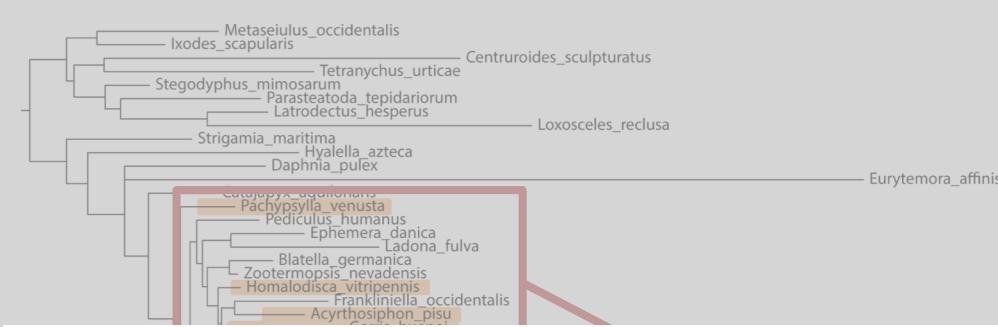
Single-copy in all but one species
(2 copies in *Plutella xylostella*)

Problem: Hemiptera not monophyletic

Problem: Lepidoptera and Trichoptera nested within Diptera



EOG8DFS3J



Single-copy in all but one species

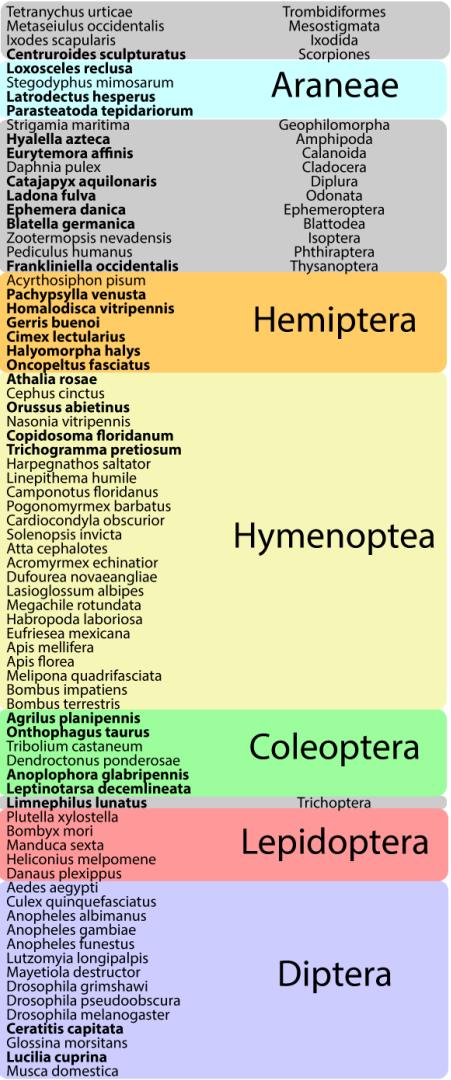
How can we turn our species rich data into sequence rich data?

monophyletic



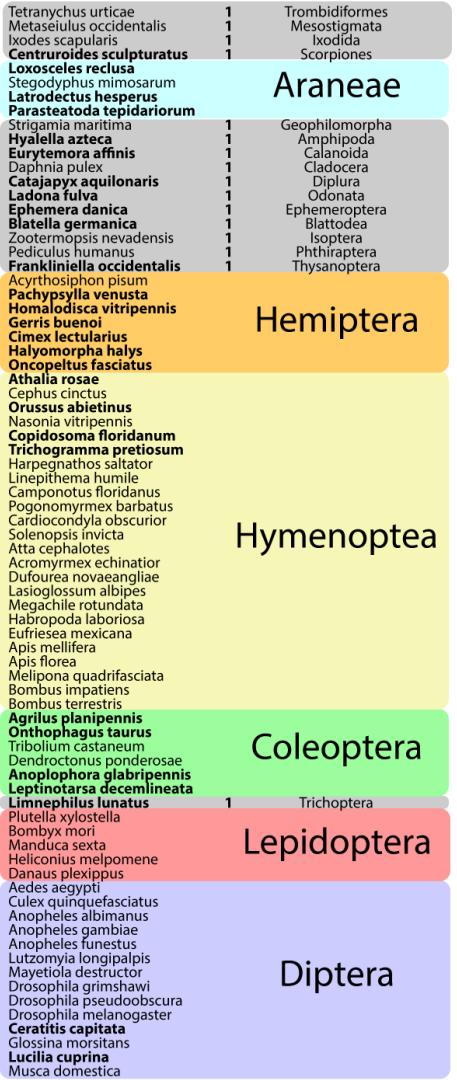
Problem: Lepidoptera and Trichoptera nested within Diptera

76 species



Construct a backbone
tree among orders
rather than species

76 species



Construct a backbone tree among orders rather than species

"Single-copy" orthologs are now those that:

1. Are single-copy in ALL the orders represented by a single species.

76 species

Tetranychus urticae	1	Trombidiformes
Metaseiulus occidentalis	1	Mesostigmata
Ixodes scapularis	1	Ixodida
Centruroides sculpturatus	1	Scorpiones
Loxosceles reclusa	0	
Stegodyphus mirmarum	0	
Latrodectus hesperus	1	
Parasteatoda tepidariorum	0	
Strigamia maritima	1	Geophilomorpha
Hyalella azteca	1	Amphipoda
Eurytemora affinis	1	Cladocera
Daphnia pulex	1	Diplura
Catajapyx aquilonaris	1	Odonata
Ladona fulva	1	Ephemeroptera
Ephemerella danica	1	Blaetodea
Biatella germanica	1	Isoptera
Zootermopsis nevadensis	0	Phthiraptera
Pediculus humanus	0	Thysanoptera
Frankliniella occidentalis	1	
Acyrtosiphon pisum	0	
Pachypsylva venusta	0	
Homalodisca vitripennis	0	
Gerris buenoi	0	
Cimex lectularius	0	
Halymomphala halys	0	
Oncopeltus fasciatus	0	
Atthalia rosae	0	
Cephus cinctus	0	
Orussus abietinus	0	
Nasonia vitripennis	0	
Copidosoma floridanum	0	
Trichogramma pretiosum	1	
Harpegnathos saltator	0	
Linepithema humile	0	
Campponotus floridanus	0	
Pogonomyrmex barbatus	0	
Cardiomyrmex obscurior	0	
Solenopsis invicta	0	
Atta cephalotes	0	
Acromyrmex echinatior	0	
Dufourea novaeangliae	0	
Lasioglossum albipes	0	
Megachile rotundata	0	
Habropoda laboriosa	0	
Eufriesea mexicana	0	
Apis mellifera	0	
Apis florea	0	
Melipona quadrifasciata	0	
Bombus impatiens	0	
Bombus terrestris	0	
Agrilus planipennis	0	
Onthophagus taurus	0	
Tribolium castaneum	0	
Dendroctonus ponderosae	0	
Anoplophora glabripennis	0	
Leptinotarsa decemlineata	1	Trichoptera
Limnephilus lunatus	1	
Plutella xylostella	0	
Bombyx mori	0	
Manduca sexta	0	
Heliconius melpomene	0	
Danaus plexippus	0	
Aedes aegypti	0	
Culex quinquefasciatus	0	
Anopheles albimanus	0	
Anopheles gambiae	0	
Anopheles funestus	0	
Lutzomyia longipalpis	0	
Mayetiolia destructor	0	
Drosophila grimshawi	0	
Drosophila pseudoobscura	0	
Drosophila melanogaster	0	
Ceratitis capitata	0	
Glossina morsitans	0	
Lucilia cuprina	0	
Musca domestica	0	

Construct a backbone tree among orders rather than species



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76 species

Tetranychus urticae	1	Trombidiformes
Metaseiulus occidentalis	1	Mesostigmata
Ixodes scapularis	1	Ixodida
Centruroides sculpturatus	1	Scorpiones
Loxosceles reclusa	0	
Stegodyphus mimosarum	0	
Latrodectus hesperus	1	
Parasteatoda tepidariorum	0	
Strigamia maritima	1	Geophilomorpha
Hyalella azteca	1	Amphipoda
Eurytemora affinis	1	Cladocera
Daphnia pulex	1	Diplura
Catajapyx aquilonaris	1	Odonata
Ladona fulva	1	Ephemeroptera
Ephemerella danica	1	Blaetodea
Biatella germanica	1	Isoptera
Zootermopsis nevadensis	1	Phthiraptera
Pediculus humanus	1	Thysanoptera
Frankliniella occidentalis	1	
Acyrtosiphon pisum	0	
Pachypsylla venusta	0	
Homalodisca vitripennis	0	
Gerris buenoi	0	
Cimex lectularius	0	
Halymomphala halys	0	
Oncopeltus fasciatus	0	
Atthalia rosae	0	
Cephus cinctus	0	
Orussus abietinus	0	
Nasonia vitripennis	0	
Copidosoma floridanum	0	
Trichogramma pretiosum	1	
Harpegnathos saltator	0	
Linepithema humile	0	
Campponotus floridanus	0	
Pogonomyrmex barbatus	0	
Cardiomyrmex obscurior	0	
Solenopsis invicta	0	
Atta cephalotes	0	
Acromyrmex echinatior	0	
Dufourea novaeangliae	0	
Lasioglossum albipes	0	
Megachile rotundata	0	
Habropoda laboriosa	0	
Eufriesea mexicana	0	
Apis mellifera	0	
Apis florea	0	
Melipona quadrifasciata	0	
Bombus impatiens	0	
Bombus terrestris	0	
Agrilus planipennis	0	
Onthophagus taurus	0	
Tribolium castaneum	0	
Dendroctonus ponderosae	0	
Anoplophora glabripennis	0	
Leptinotarsa decemlineata	1	Trichoptera
Limnephilus lunatus	1	
Plutella xylostella	0	
Bombyx mori	0	
Manduca sexta	0	
Heliconius melpomene	1	
Danaus plexippus	0	
Aedes aegypti	0	
Culex quinquefasciatus	0	
Anopheles albimanus	0	
Anopheles gambiae	0	
Anopheles funestus	0	
Lutzomyia longipalpis	1	
Mayetiolia destructor	0	
Drosophila grimshawi	0	
Drosophila pseudoobscura	0	
Drosophila melanogaster	0	
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Loxosceles reclusa	0	
Stegodyphus mimosarum	0	
Latrodectus hesperus	1	
Parasteatoda tepidariorum	0	
Strigamia maritima	1	Geophilomorpha
Hyalella azteca	1	Amphipoda
Eurytemora affinis	1	Cladocera
Daphnia pulex	1	Diplura
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Pediculus humanus	1	Thysanoptera
Frankliniella occidentalis	1	
Acyrtosiphon pisum	0	
Pachypsylva venusta	1	
Homalodisca vitripennis	0	
Gerris buenoi	0	
Cimex lectularius	0	
Halymomphala halys	1	
Oncopeltus fasciatus	0	
Atthalia rosae	0	
Cephus cinctus	0	
Orussus abietinus	0	
Nasonia vitripennis	0	
Copidosoma floridanum	0	
Trichogramma pretiosum	1	
Harpegnathos saltator	0	
Linepithema humile	0	
Campponotus floridanus	0	
Pogonomyrmex barbatus	0	
Cardiomyrmex obscurior	0	
Solenopsis invicta	0	
Atta cephalotes	0	
Acromyrmex echinatior	0	
Dufourea novaeangliae	0	
Lasioglossum albipes	0	
Megachile rotundata	0	
Habropoda laboriosa	0	
Eufriesia mexicana	0	
Apis mellifera	6	
Apis florea	6	
Melipona quadrifasciata	4	
Bombus impatiens	5	
Bombus terrestris	5	
Agrilus planipennis	0	
Onthophagus taurus	0	
Tribolium castaneum	1	
Dendroctonus ponderosae	0	
Anoplophora glabripennis	0	
Leptinotarsa decemlineata	0	
Limnephilus lunatus	1	Trichoptera
Plutella xylostella	0	
Bombyx mori	0	
Manduca sexta	0	
Heliconius melpomene	1	
Danaus plexippus	0	
Aedes aegypti	0	
Culex quinquefasciatus	0	
Anopheles albimanus	0	
Anopheles gambiae	1	
Anopheles funestus	0	
Lutzomyia longipalpis	0	
Mayetiolia destructor	0	
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Lucilia cuprina	0	
Musca domestica	0	

76 species

Hemiptera

Hymenoptera

Coleoptera

Lepidoptera

Diptera

21 orders

Araneae

Geophilomorpha
Amphipoda
Calanoida
Cladocera
Diplura
Odonata
Ephemeroptera
Blaetodea
Isoptera
Phthiraptera
Thysanoptera

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Stegodyphus mirmarum	0	
Latrodectus hesperus	1	
Parasteatoda tepidariorum	0	
Strigamia maritima	1	Geophilomorpha
Hyalella azteca	1	Amphipoda
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Catajapyx aquilonaris	1	Odonata
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Frankliniella occidentalis	1	
Acythosiphon pisum	0	
Pachypsylla venusta	0	
Homalodisca vitripennis	0	
Gerris buenoi	0	
Cimex lectularius	0	
Halymonoma halys	0	
Oncopeltus fasciatus	0	
Atthalia rosae	0	
Cephus cinctus	0	
Orussus abietinus	0	
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Copidosoma floridanum	0	
Trichogramma pretiosum	1	
Harpegnathos saltator	0	
Linepithema humile	0	
Campponotus floridanus	0	
Pogonomyrmex barbatus	0	
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Dufourea novaeangliae	0	
Lasioglossum albipes	0	
Megachile rotundata	0	
Habropoda laboriosa	0	
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Construct a backbone tree among **orders** rather than **species**

Phylum	# Orders	# single-copy orthologs
Arthropoda	21	150



Construct a backbone tree among orders rather than species

Phylum	# Orders	# single-copy orthologs
Arthropoda	21	150

Then use single-copy orthologs from the 6 multi-species orders to construct order-level trees

Order	# Species	# single-copy orthologs
Araneae	4	1627
Hemiptera	7	2053
Hymenoptera	24	2121
Coleoptera	6	3880
Lepidoptera	5	3660
Diptera	14	1324

1) Predict orthogroups

1	CAATGCG AAATGCG BAATGCG AAATGCG	CATATCA AATATTAA BATA
2	CTTCAAG AATTAAG BATTACG	
3	CCGAAA ACGATCA BCGTTCA	

2) Select single-copy groups

1	CAATGCG AAATGCG BAATGCG AAATGCG	
4	CATATCA AATATTAA BATA	
5	CCGAAA ACGATCA BCGTTCA	

3) Align each group

2	CTTCAAG AATTAAG BATTACG
3	CCGAAA-A ACGATCA BCGTTCA
4	CATATCA AATATTAA BATA - - -



3) Align each group

Phylum	# Orders	# single-copy orthologs
Arthropoda	21	150
Order	# Species	# single-copy orthologs
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Two alignment programs:

1. MUSCLE
2. PASTA

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2) Select single-copy groups

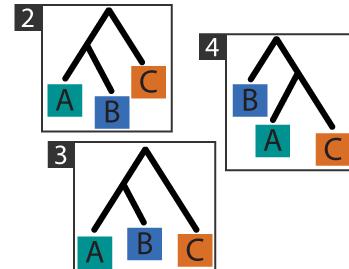
1	CAATGCG AAATGCG BAATGCG AAATGCG	CATATCA AATATTAA BATA
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4) Infer gene trees





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RAxML:

with PROTGAMMAJTTF
amino acid substitution
model



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RAxML:

with PROTGAMMAJTTF
amino acid substitution
model

Topologies largely
insensitive to substitution
model

1) Predict orthogroups

1	CAATGCG AAATGCG BAATGCG AAATGCG	CATATCA AATATTAA BATA
2	CTTCAAG AATTAAG BATTACG	C CGAAA ACGATCA BCGTTCA
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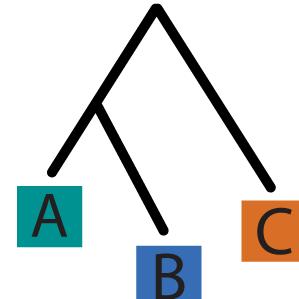
2) Select single-copy groups

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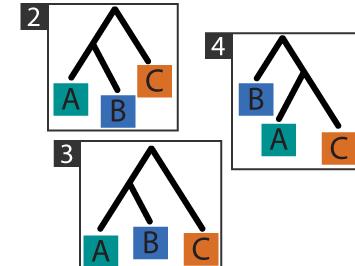
3) Align each group

2	CTTCAAG AATTAAG BATTACG
3	CCGAAA-A ACGATCA BCGTTCA
4	CATATCA AATATTAA BATA - - -

5) Infer species tree



4) Infer gene trees





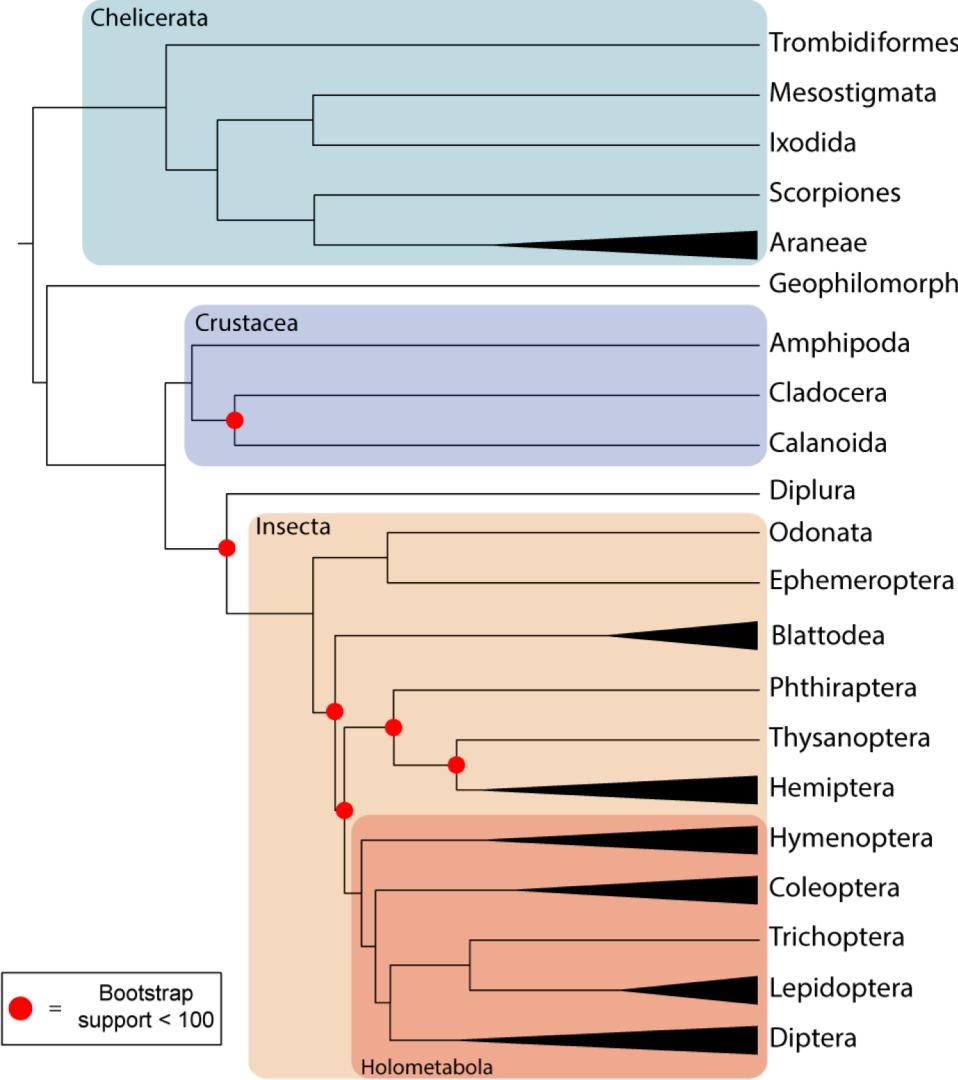
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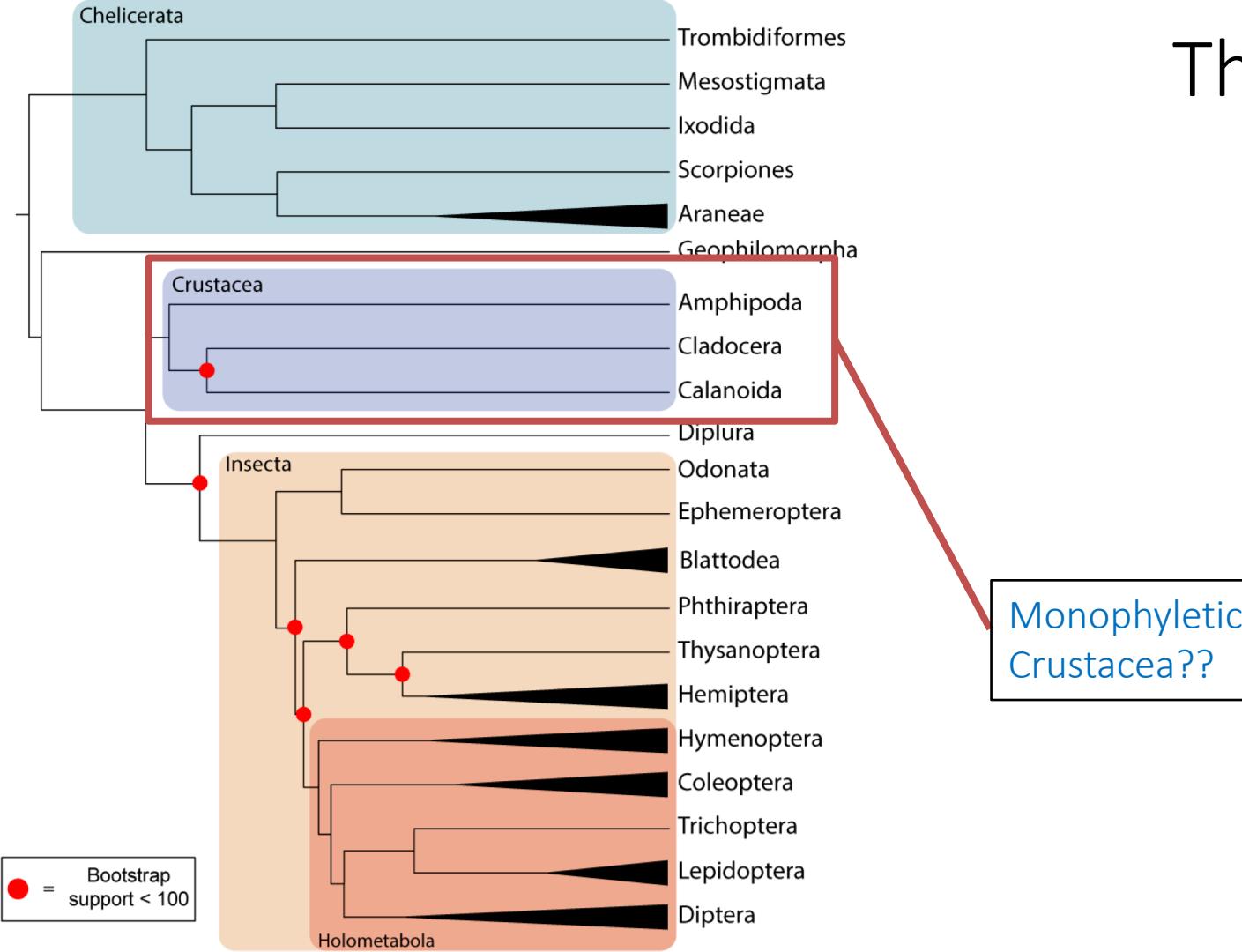
Three species tree methods:

1. Average consensus
2. Concatenation
3. ASTRAL

The backbone phylogeny



The backbone phylogeny





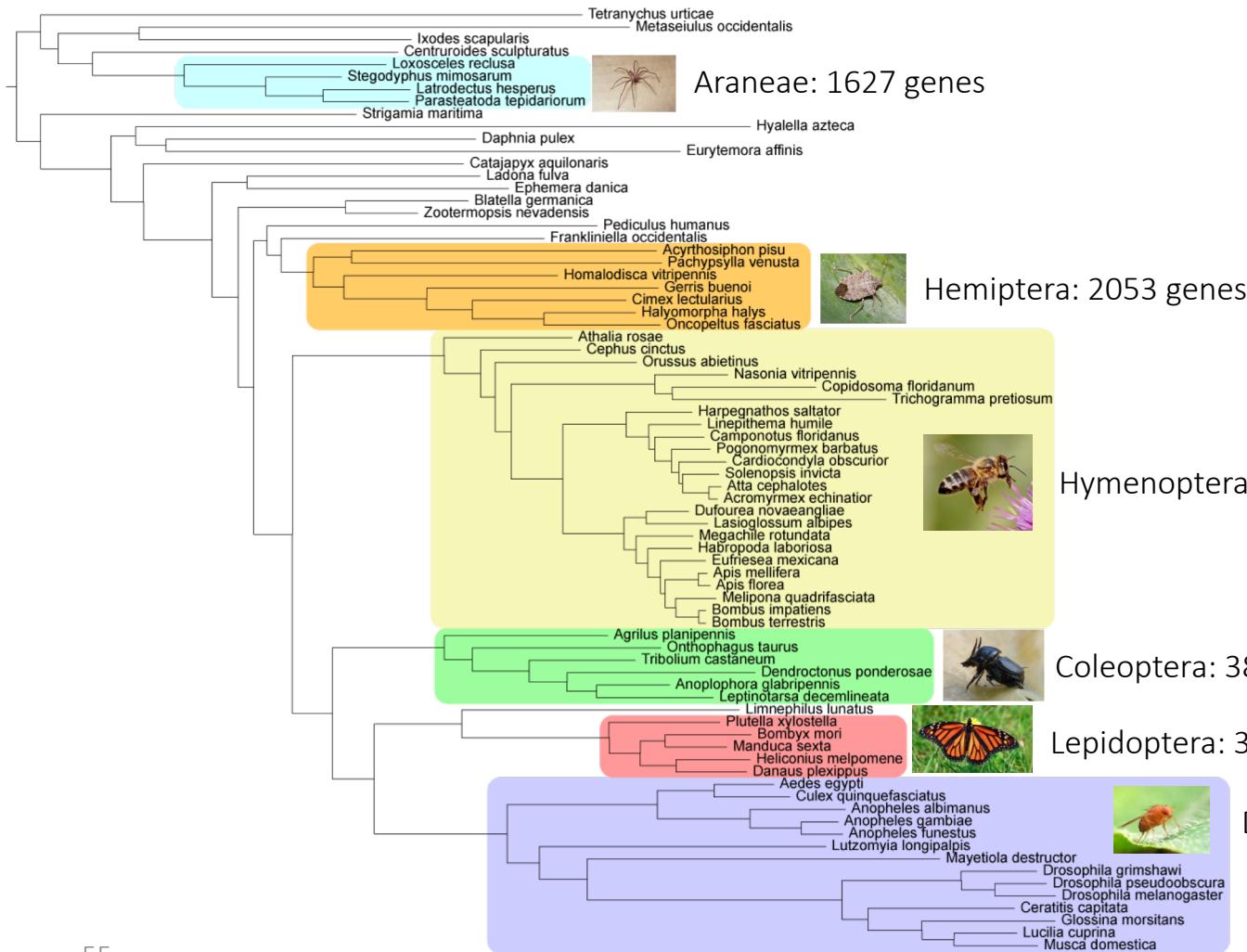
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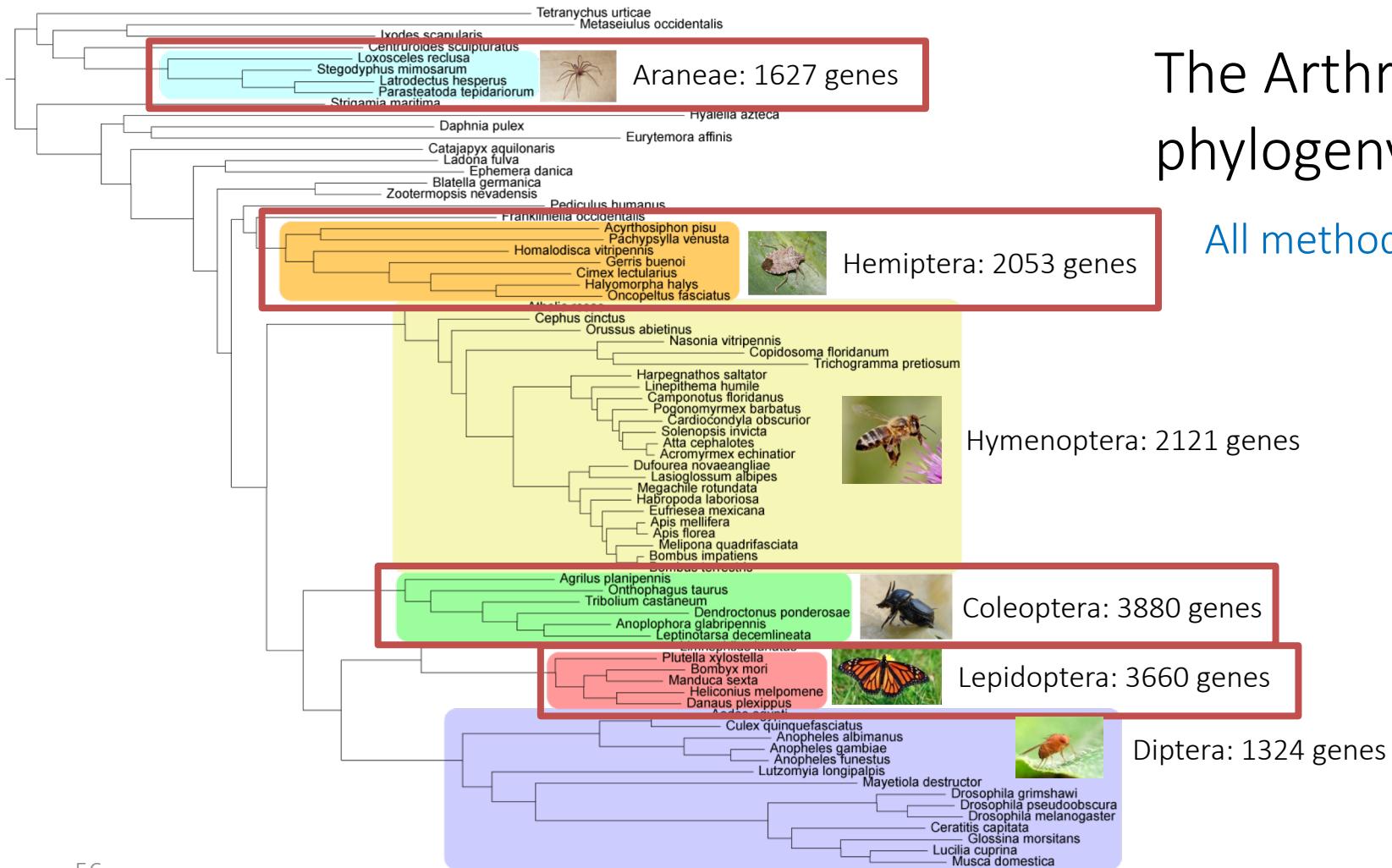
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The Arthropod phylogeny



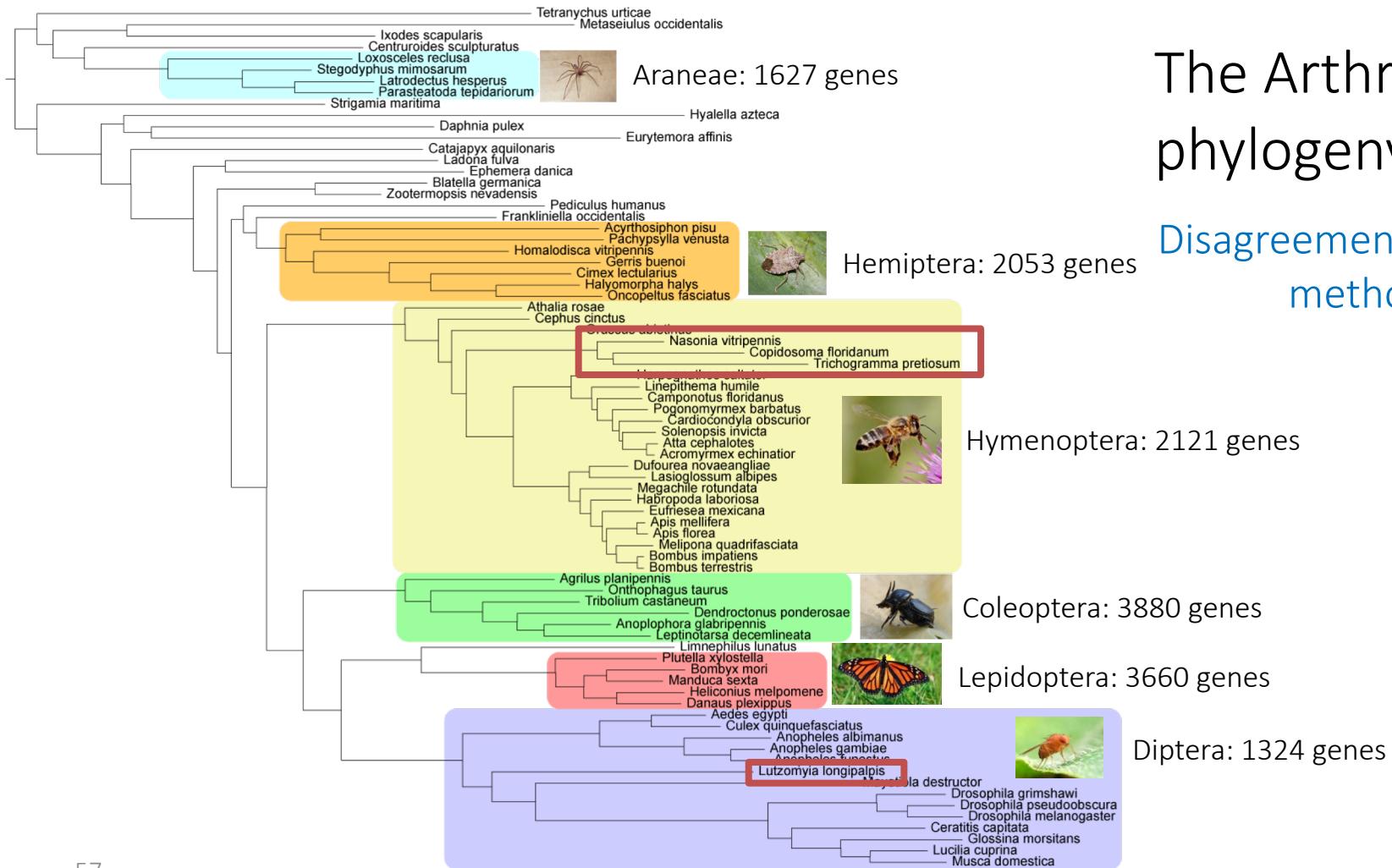
The Arthropod phylogeny

All methods agree ✓



The Arthropod phylogeny

Disagreement between methods 



1) Predict orthogroups

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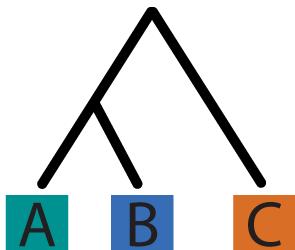
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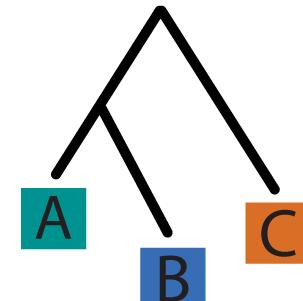
3) Align each group

2	CTTCAAG AATTAAG BATTACG
3	CCGAAA-A ACGATCA BCGTTCA
4	CATATCA AATATTAA BATA - - -

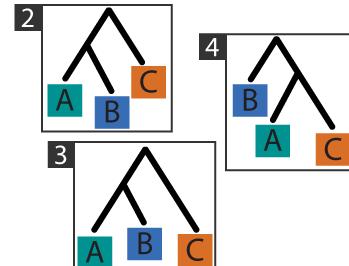
6) Scale branch lengths with fossil calibrations



5) Infer species tree



4) Infer gene trees



6) Scale branch lengths with fossil calibrations



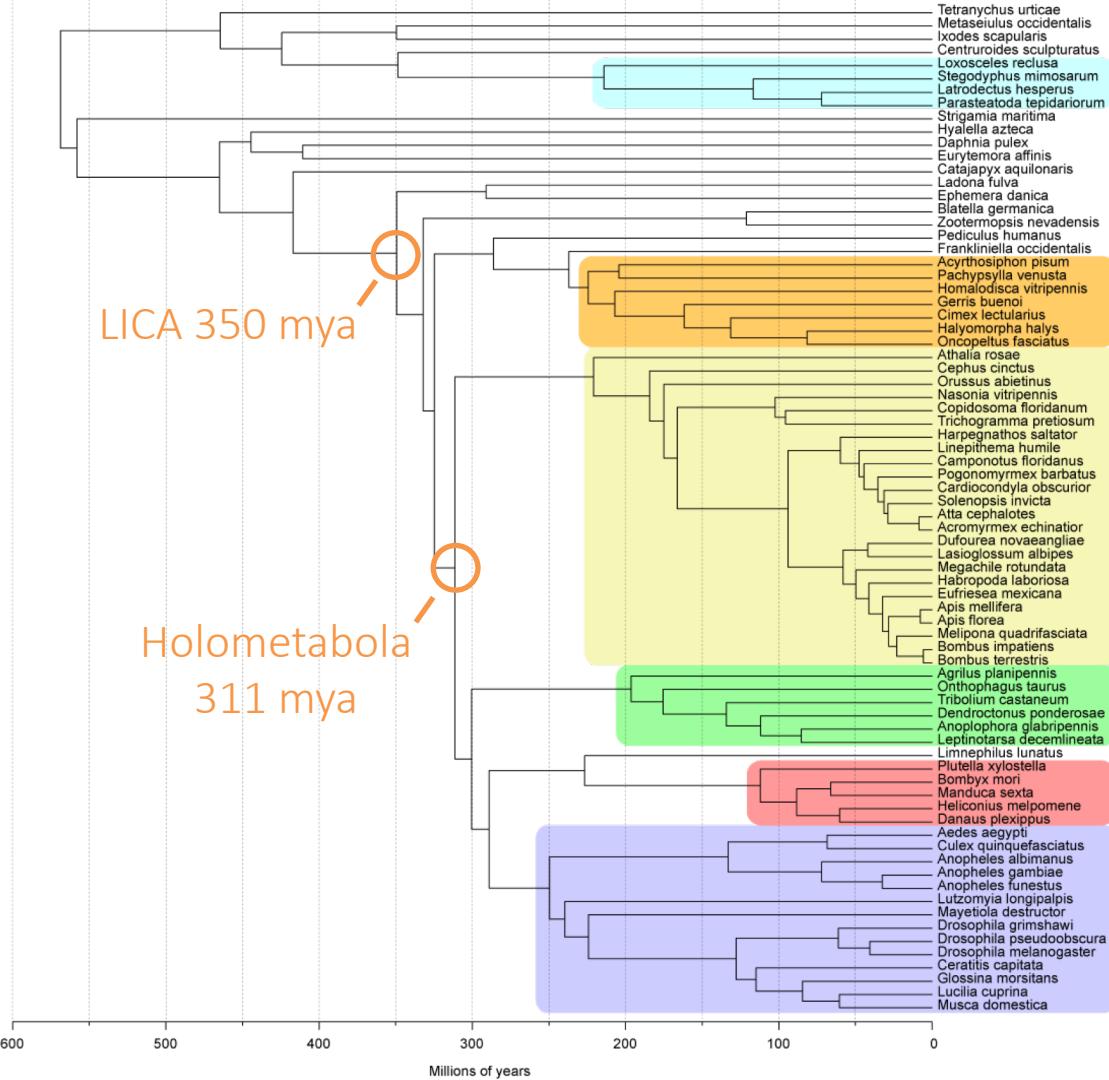
Crown group	Node	Min time	Max time
Euarthropoda	75	514	636.1
Arachnida	74	432.6	636.1
Parasitiformes	72	98.17	514
Mandibulata	67	514	636.1
Multicrustacea	64	487	636.1
Pterygota	62	322.83	521
Paleoptera	1	319.9	521
Neoptera	61	319.9	411
Blattodea	2	130.3	411
Eumetabola	60	319.9	411
Condylgnatha	58	306.9	411
Hemiptera	57	306.9	411
Holometabola	51	313.7	411
Hymenoptera	25	226.4	411
Aparaglossata	50	313.7	411
Coleoptera	30	208.5	411
Mecopterida	49	271.8	411
Amphiesmenoptera	35	195.31	411
Lepidoptera	34	129.41	411
Diptera	48	240.5	411

Order	Node	Min time	Max time
Hymenoptera	HY25	89.9	93.9
Hymenoptera	HY13	23	28.4

Use r8s to smooth the tree:

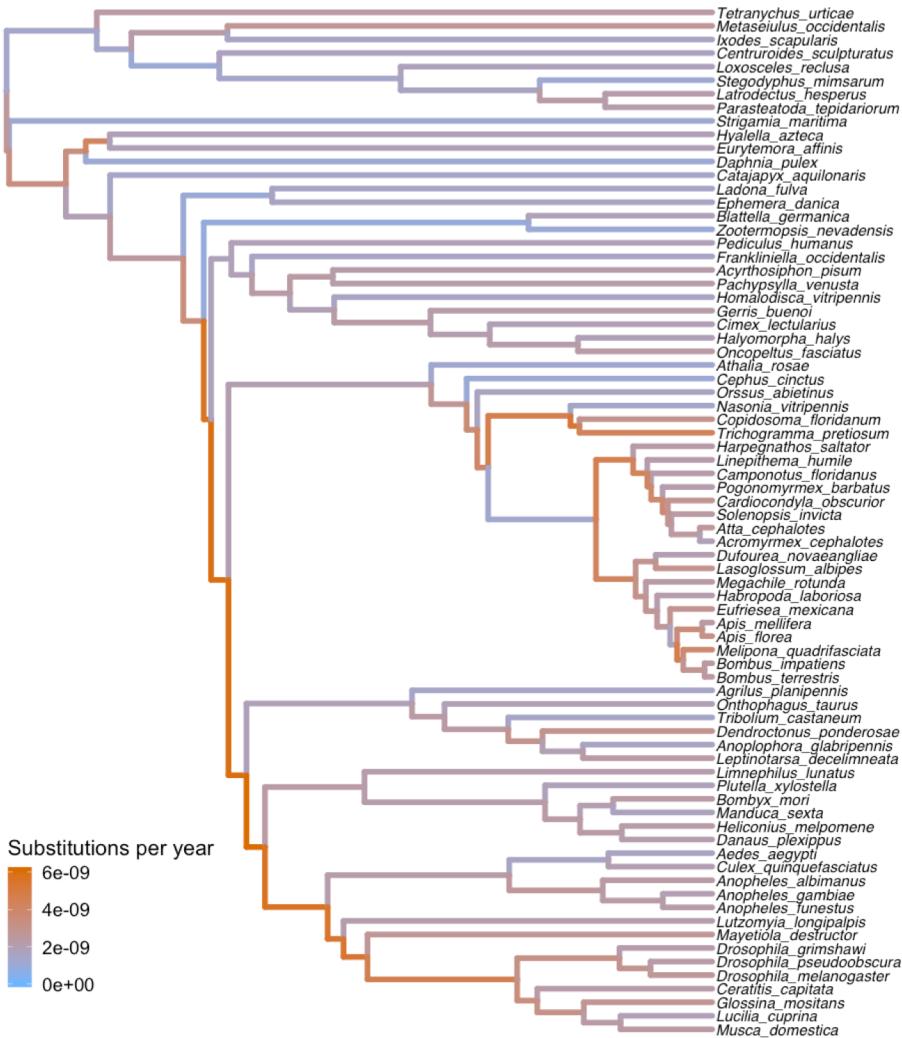
Penalized likelihood method
to correlate rates of
evolution among branches

Arthropod Time Tree



Arthropod Time Tree

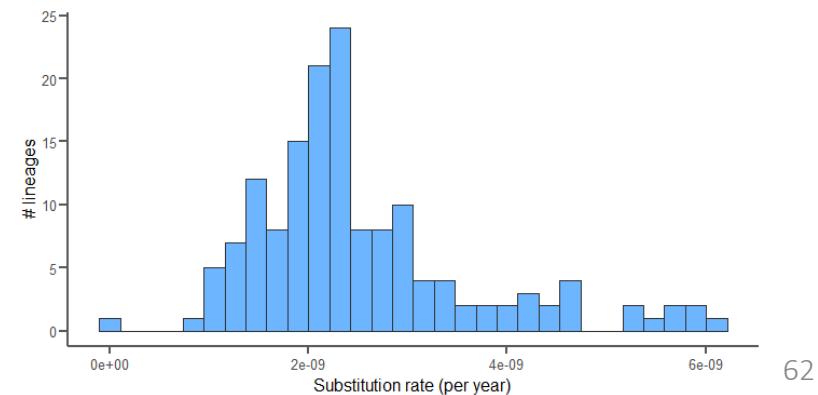
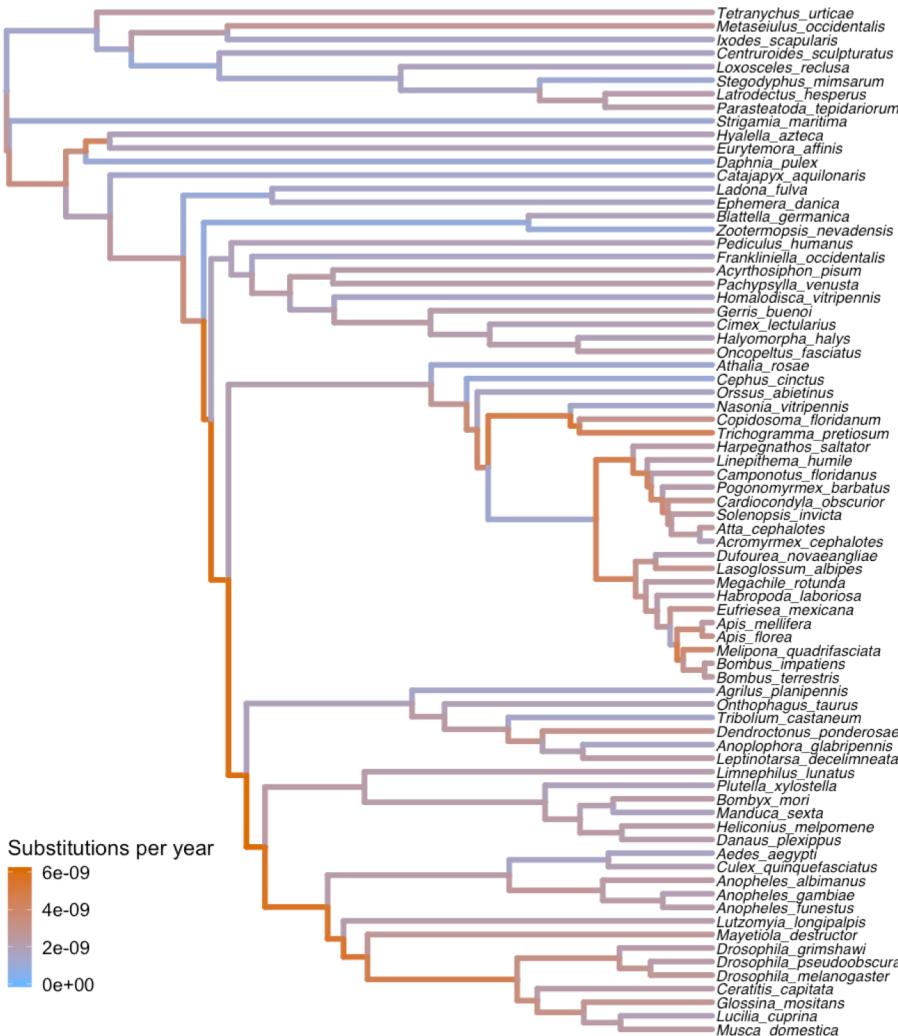
The branches of the ML tree can be scaled by time to infer substitution rates



Arthropod Time Tree

The branches of the ML tree can be scaled by time to infer substitution rates

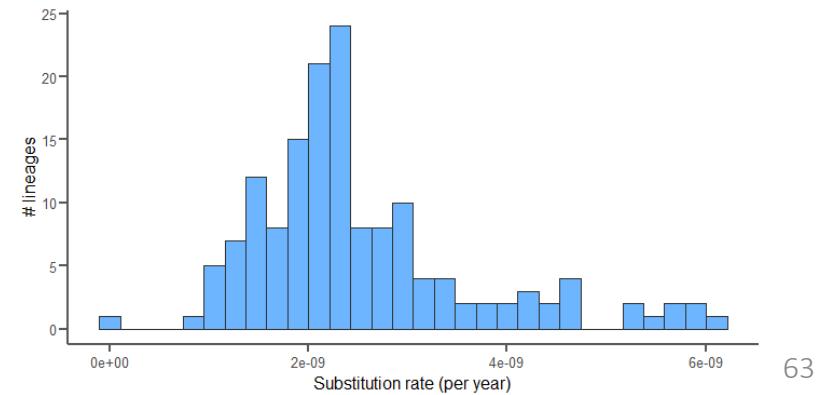
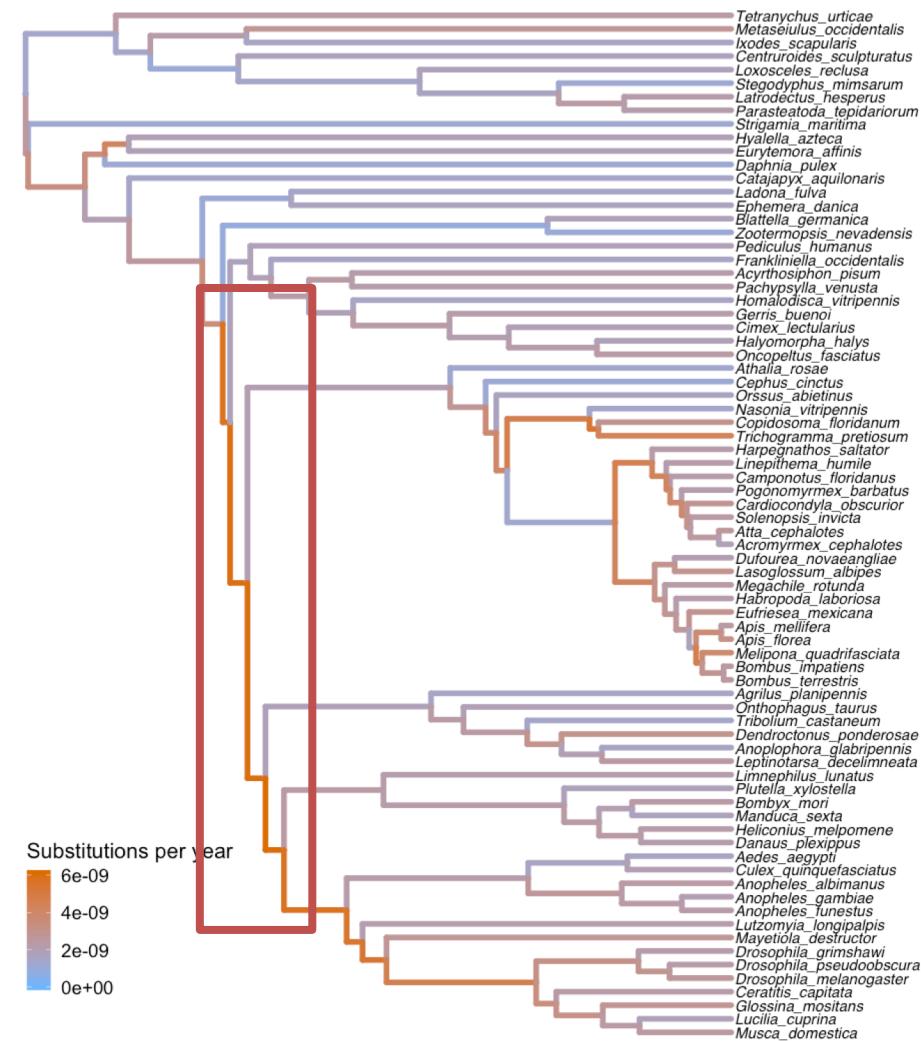
Rates are mostly consistent across arthropods



Arthropod Time Tree

The branches of the ML tree can be scaled by time to infer substitution rates

Rates are mostly consistent across arthropods



Today's topics

1. Determining the Arthropod phylogeny
2. Reconstructing ancestral gene counts
3. Using the i5k gene family web site

1) Predict orthogroups

1	C A A T G C G A A A T G C G B A A T G C G A A A T G C G
---	--

4	C A T A T A T C A A A T A T T A B A T A
---	---

2	C T T C A A G A A T T A A G B A T T A C G
---	---

5	C C G A A A A C G A T C A B C G T T C A A C G A C A B C G A C A
---	---

3	C C G A A A A C G A T C A B C G T T C A
---	---

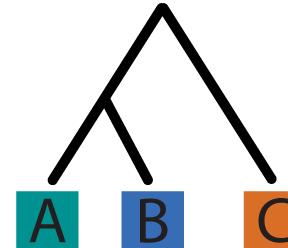
6	C G G C A A T A G G C A T
---	------------------------------

1) Predict orthogroups

1	<table border="1"><tr><td>C</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr><tr><td>A</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr><tr><td>B</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr><tr><td>A</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr></table>	C	A	A	T	G	C	G	A	A	A	T	G	C	G	B	A	A	T	G	C	G	A	A	A	T	G	C	G							
C	A	A	T	G	C	G																														
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A	A	A	T	G	C	G																														
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	A	A	T	T	A	A																														
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	A	C	G	A	T	C																														
B	C	G	T	T	C	A																														
4	<table border="1"><tr><td>C</td><td>A</td><td>T</td><td>A</td><td>T</td><td>C</td><td>A</td></tr><tr><td></td><td>A</td><td>A</td><td>T</td><td>T</td><td>T</td><td>A</td></tr><tr><td>B</td><td>A</td><td>T</td><td>A</td><td></td><td></td><td></td></tr></table>	C	A	T	A	T	C	A		A	A	T	T	T	A	B	A	T	A																	
C	A	T	A	T	C	A																														
	A	A	T	T	T	A																														
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5	<table border="1"><tr><td>C</td><td>C</td><td>G</td><td>A</td><td>A</td><td>A</td><td>A</td></tr><tr><td>A</td><td>C</td><td>G</td><td>A</td><td>T</td><td>C</td><td>A</td></tr><tr><td>B</td><td>C</td><td>G</td><td>T</td><td>T</td><td>C</td><td>A</td></tr><tr><td>A</td><td>C</td><td>G</td><td>A</td><td>C</td><td>A</td><td></td></tr><tr><td>B</td><td>C</td><td>G</td><td>A</td><td>C</td><td>A</td><td></td></tr></table>	C	C	G	A	A	A	A	A	C	G	A	T	C	A	B	C	G	T	T	C	A	A	C	G	A	C	A		B	C	G	A	C	A	
C	C	G	A	A	A	A																														
A	C	G	A	T	C	A																														
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C	G	G	C	A	A	T																														
	A	G	G	C	A	T																														



2) Infer time tree



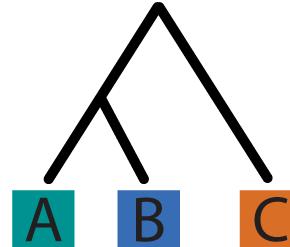
1) Predict orthogroups

1	<table border="1"><tr><td>C</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr><tr><td>A</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr><tr><td>B</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr><tr><td>A</td><td>A</td><td>A</td><td>T</td><td>G</td><td>C</td><td>G</td></tr></table>	C	A	A	T	G	C	G	A	A	A	T	G	C	G	B	A	A	T	G	C	G	A	A	A	T	G	C	G							
C	A	A	T	G	C	G																														
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2	<table border="1"><tr><td>C</td><td>T</td><td>T</td><td>C</td><td>A</td><td>A</td><td>G</td></tr><tr><td></td><td>A</td><td>A</td><td>T</td><td>T</td><td>A</td><td>A</td></tr><tr><td>B</td><td>A</td><td>T</td><td>T</td><td>A</td><td>C</td><td>G</td></tr></table>	C	T	T	C	A	A	G		A	A	T	T	A	A	B	A	T	T	A	C	G														
C	T	T	C	A	A	G																														
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C	G	G	C	A	A	T																														
	A	G	G	C	A	T																														

3) Construct gene count matrix

	A	B	C
1	2	1	1
2	1	1	1
3	1	1	1
4	1	1	1
5	2	2	1
6	1	0	1

2) Infer time tree



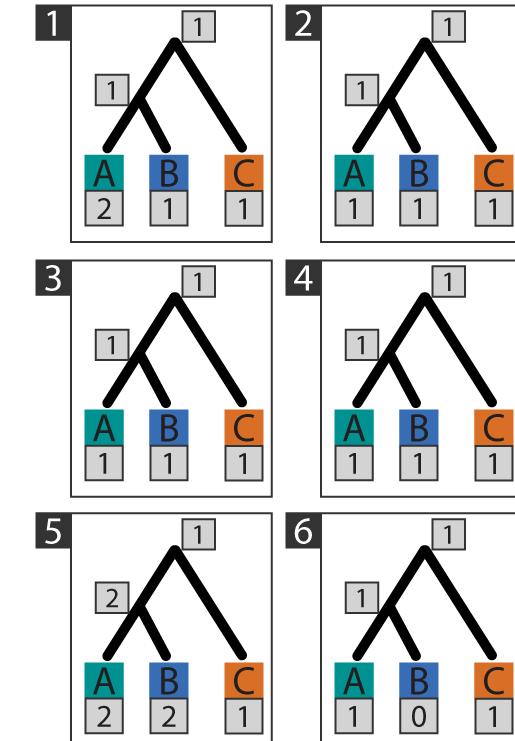
1) Predict orthogroups

1	CAATGCG AAATGCG BAATGCG AAATGCG	CATATCA AATATTAA BATA
2	CTTCAAG AATTAAG BATTACG	CCGAAA ACGATCA BCGTTCA ACGACA BCGACA
3	CGAAA ACGATCA BCGTTCA	GGCAAT AGGCAT
4		
5		
6		

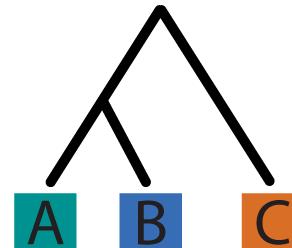
3) Construct gene count matrix

	A	B	C
1	2	1	1
2	1	1	1
3	1	1	1
4	1	1	1
5	2	2	1
6	1	0	1

4) Infer ancestral gene counts



2) Infer time tree



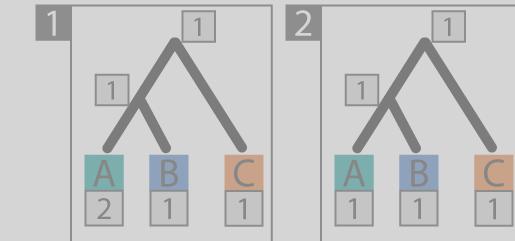
1) Predict orthogroups

1	CAATGCG AAATGCG BAATGCG AAATGCG
2	CTTCAAG AATTAAG BATTACG
3	CGAAA ACGATCA BCGTTCA
4	CATATCA AATATTAA BATA
5	CCGAAA ACGATCA BCGTTCA ACGACA BCGACA
6	CGGCAAT AGGCAT

3) Construct gene count matrix

	A	B	C
1	2	1	1
2	1	1	1
3	1	1	1
4	1	1	1
5	2	2	1
6	1	0	1

4) Infer ancestral gene counts



Family	AAEGY	AALBI	ACEPH	AECHI	AFLOR	AFUNE	AGAMB	AGLAB	AMELL	APISU	APLAN	AROSA	BGERM	BIMPA	BMORI	BTERR	CAQUI	CCAPI	CCINC	CFLOR	CLECT	COBSC	COPFL	CQUIN	CSCUL	DGRIM
EG08003Z0	1	1	1	1	1	1	2	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
EG08003Z1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
EG08003Z2	0	0	0	0	0	0	0	0	0	0	10	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0
EG08003Z3	0	1	1	1	1	0	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1
EG08003Z4	0	0	1	1	1	0	0	1	1	1	1	1	1	1	1	1	1	1	0	2	0	0	3	0	0	0
EG08003Z5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
EG08003Z6	0	0	2	1	1	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0
EG08003Z7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
EG08003Z8	1	1	1	1	1	1	1	2	1	3	1	2	1	1	1	1	1	1	1	1	1	1	1	1	1	1
EG08003Z9	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
EG08003ZB	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
EG08003ZC	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
EG08003ZD	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

A B C

A B C A B C

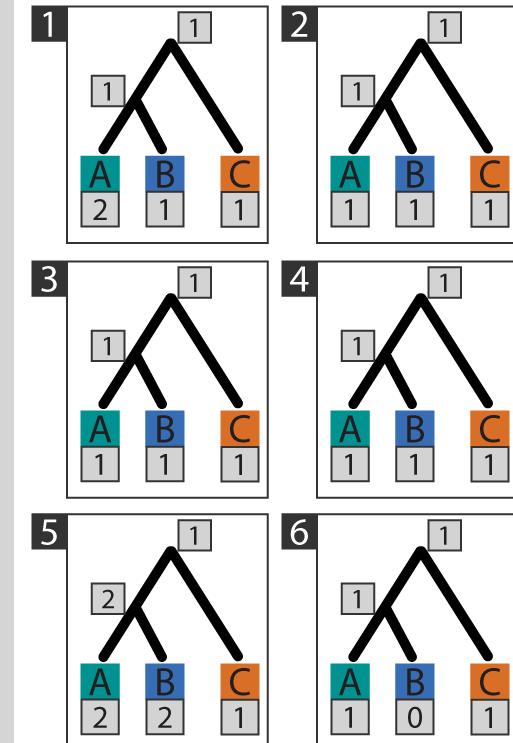
1) Predict orthogroups

1	CAATGCG AAATGCG BAATGCG AAATGCG
2	CTTCAAG AATTAAG BATTACG
3	CCGAAA ACGATCA BCGTTCA
4	CATATCA AATATTAA BATA
5	CCGAAA ACGATCA BCGTTCA ACGACA BCGACA
6	GGCAAT AGGCAT

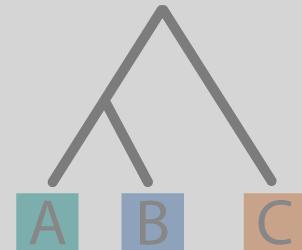
3) Construct gene count matrix

	A	B	C
1	2	1	1
2	1	1	1
3	1	1	1
4	1	1	1
5	2	2	1
6	1	0	1

4) Infer ancestral gene counts



2) Infer time tree



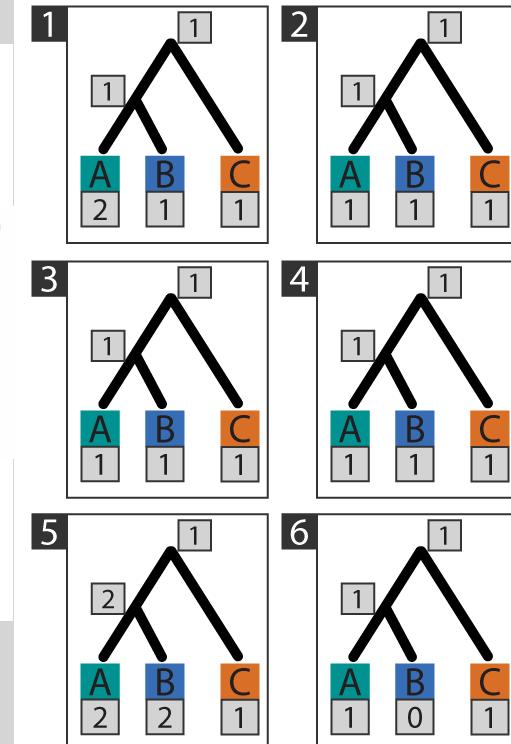
1) Predict orthogroups

1	CAATGCG	
2	TTTCAAG	
3	AAATGCG	
4	CATATCA	
5	AAATTAA	
6	AAATGCG	
7	BAAATGCG	
8	BATA	

3) Construct gene count matrix

	A	B	C
1	2	1	1
2	1	1	1

4) Infer ancestral gene counts



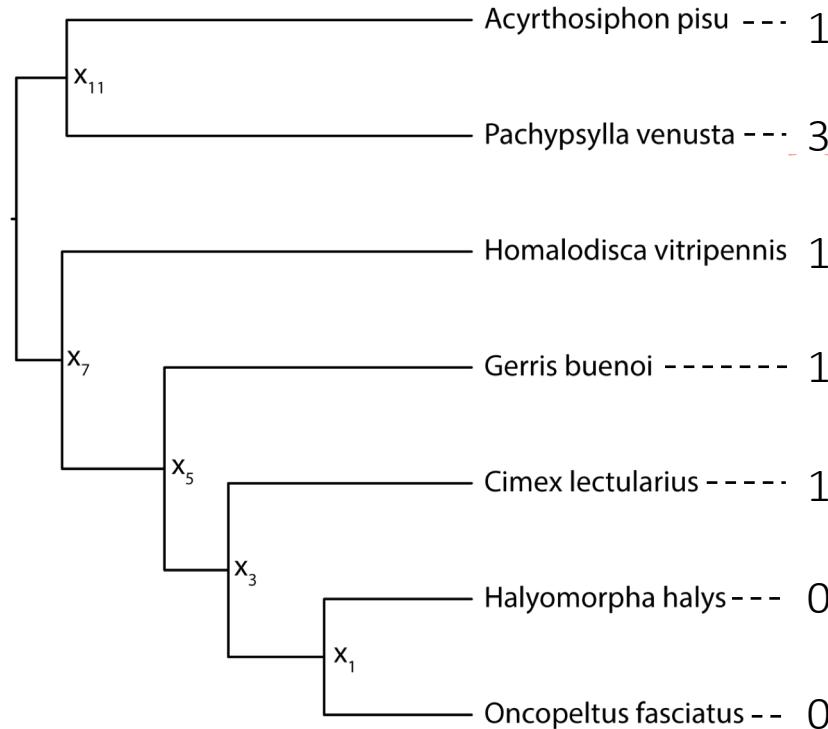
Ancestral gene counts inferred
with:

1. Maximum likelihood (CAFE) for the 6 multi-species orders
2. Parsimony (Dupliphy) for all nodes

A B C

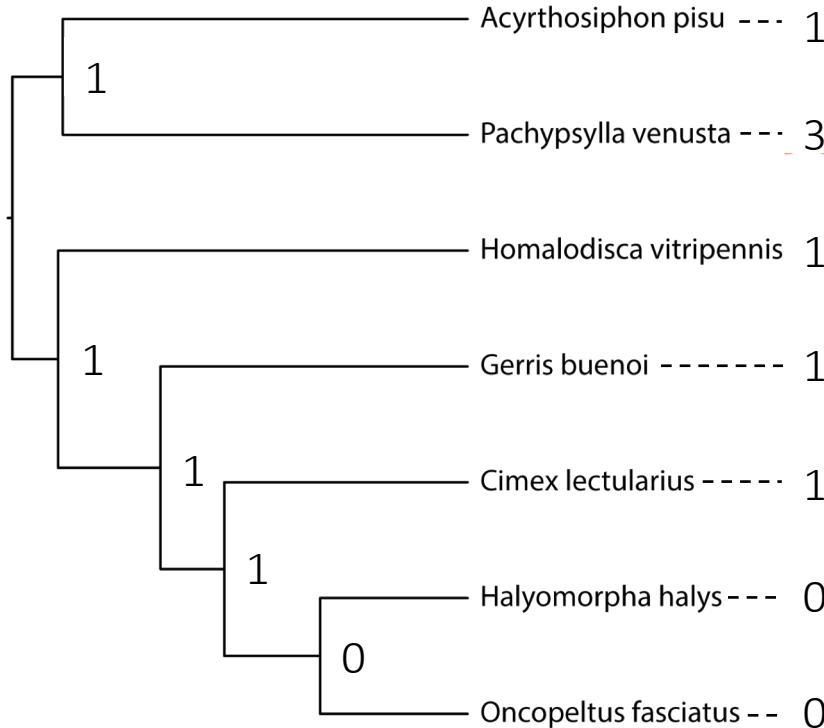


Ancestral gene counts: Example



Tips: observed variables
 x_i : hidden variables

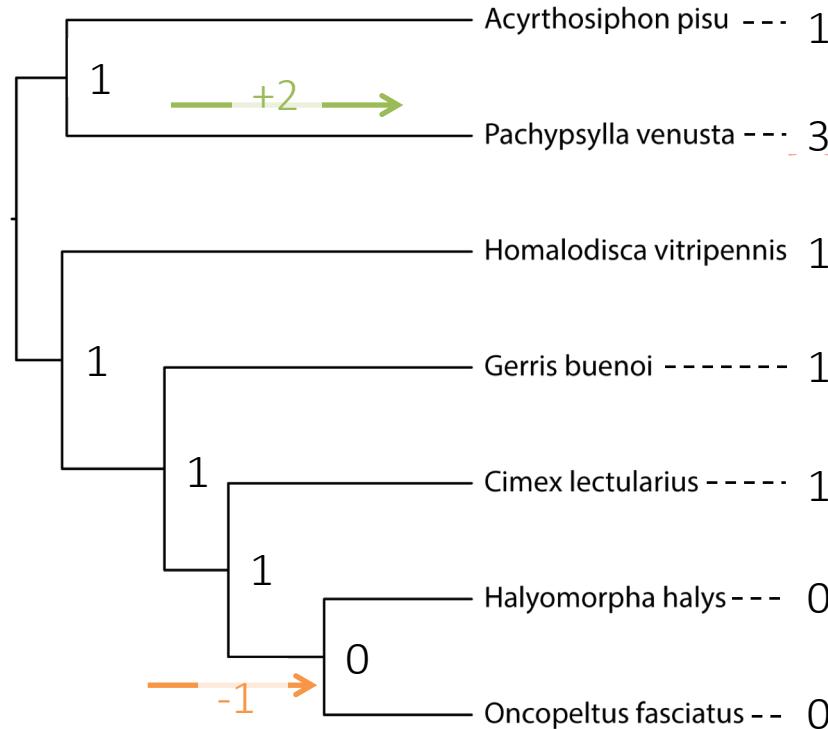
Ancestral gene counts: Example



Tips: observed variables
 x_i : hidden variables

Our goal is to infer the states of the internal nodes of the tree

Ancestral gene counts: Example

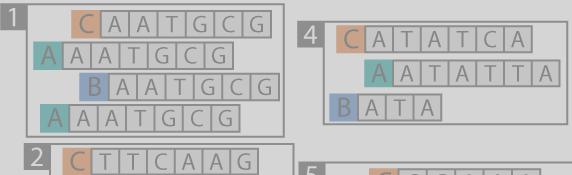


Tips: observed variables
 x_i : hidden variables

Our goal is to infer the states of the internal nodes of the tree

Then we can count changes along each lineage

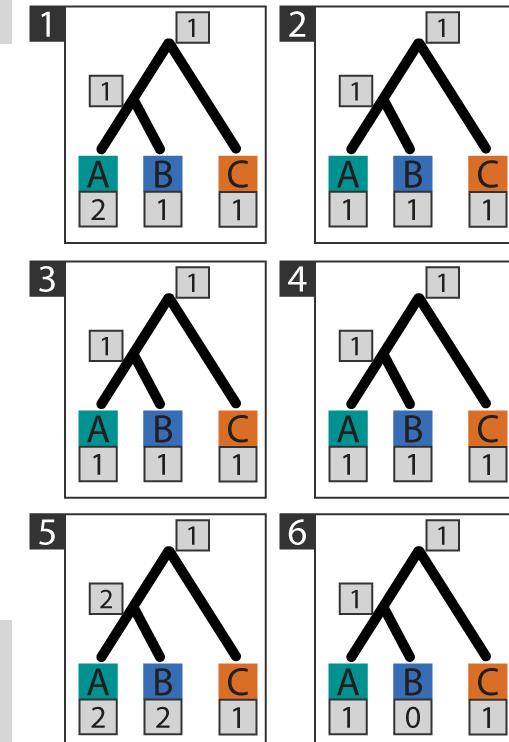
1) Predict orthogroups



3) Construct gene count matrix

	A	B	C
1	1	2	1
2	2	1	1

4) Infer ancestral gene counts

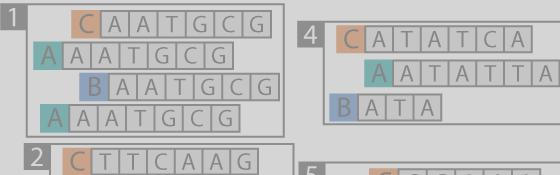


With ancestral gene counts we can:

1. Infer rates of gene gain/loss
2. Count gene gains and losses and check for rapid changes on every lineage
3. Estimate gene counts in extinct ancestors

A B C

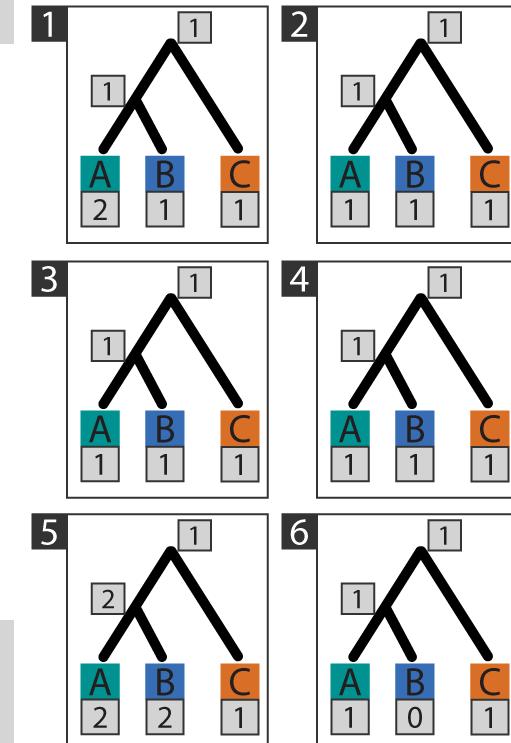
1) Predict orthogroups



3) Construct gene count matrix

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1	2	1	1
2	1	1	1

4) Infer ancestral gene counts



With ancestral gene counts we can:

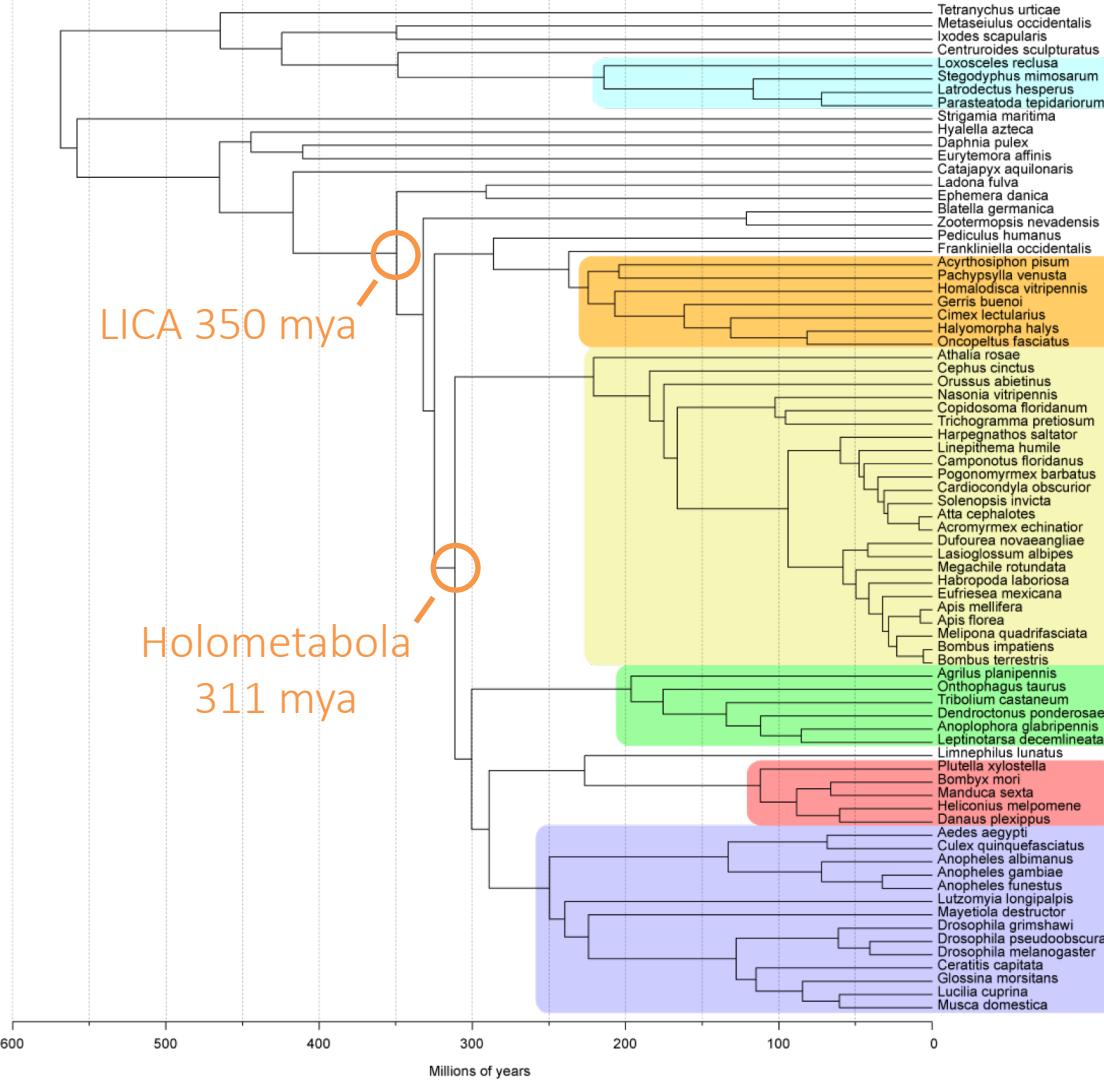
1. Infer rates of gene gain/loss
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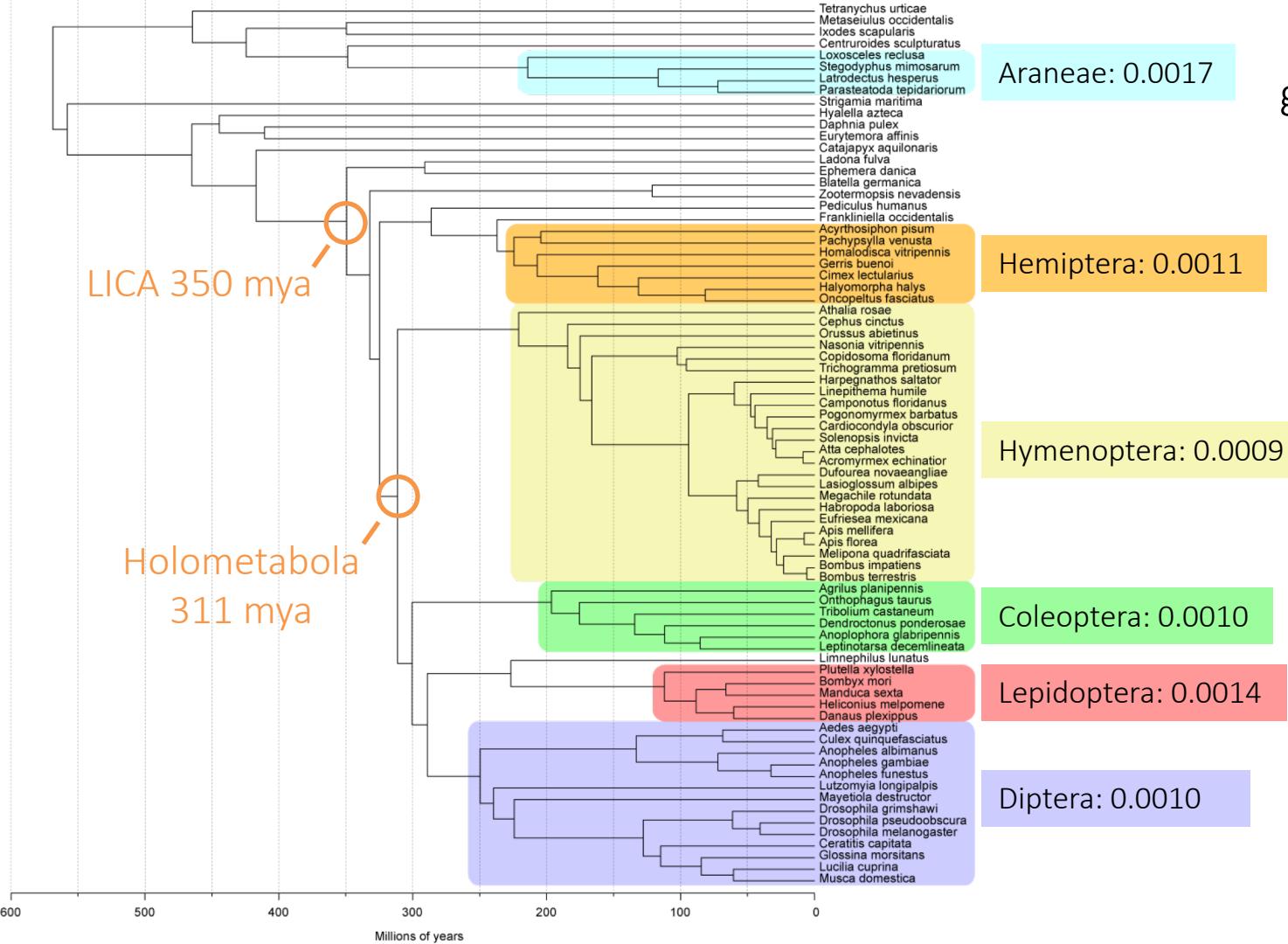
A B C

Arthropod Time Tree

LICA 350 mya

Holometabola
311 mya

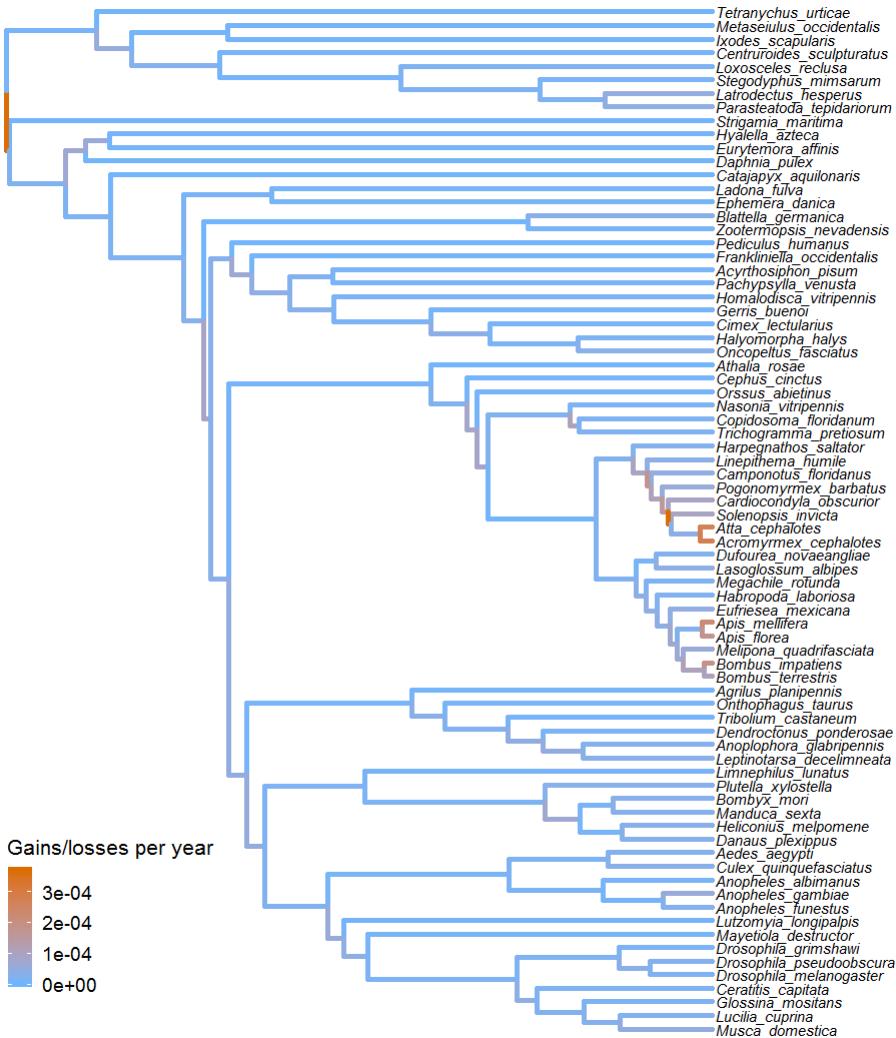




Rates of gene gain/loss between orders are largely consistent

Arthropod Time Tree

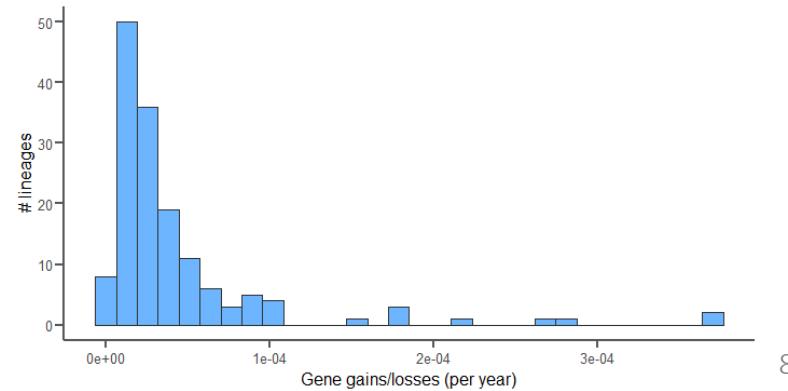
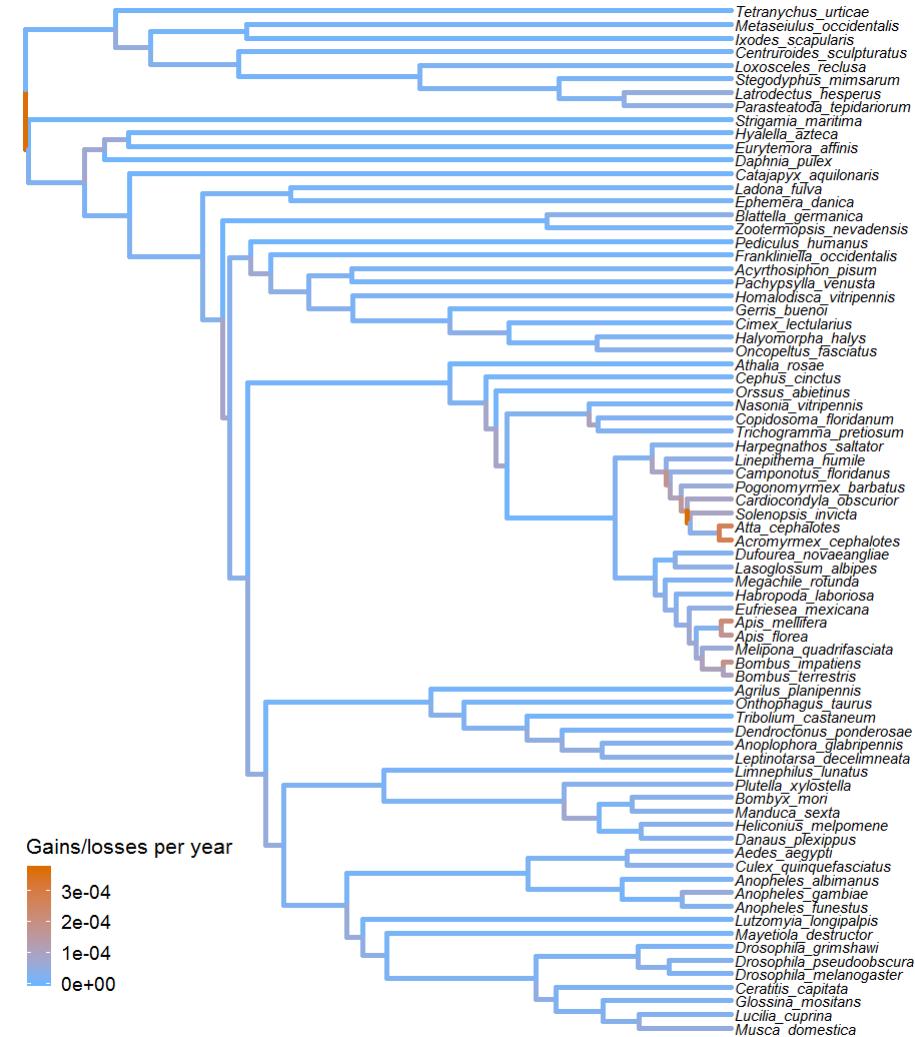
The branches of the ML tree can be scaled by time to infer lineage specific gain/loss rates



Arthropod Time Tree

The branches of the ML tree can be scaled by time to infer lineage specific gain/loss rates

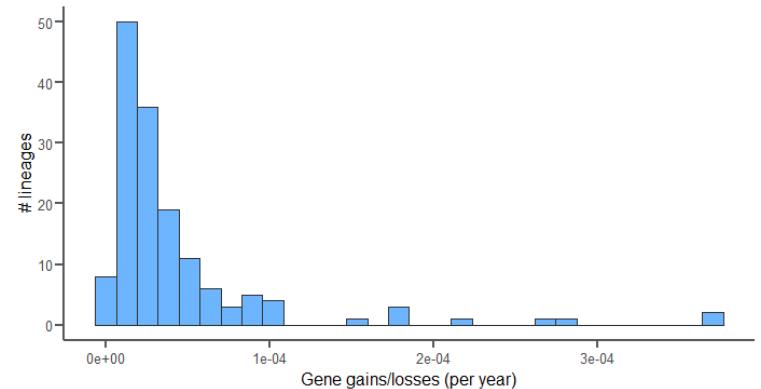
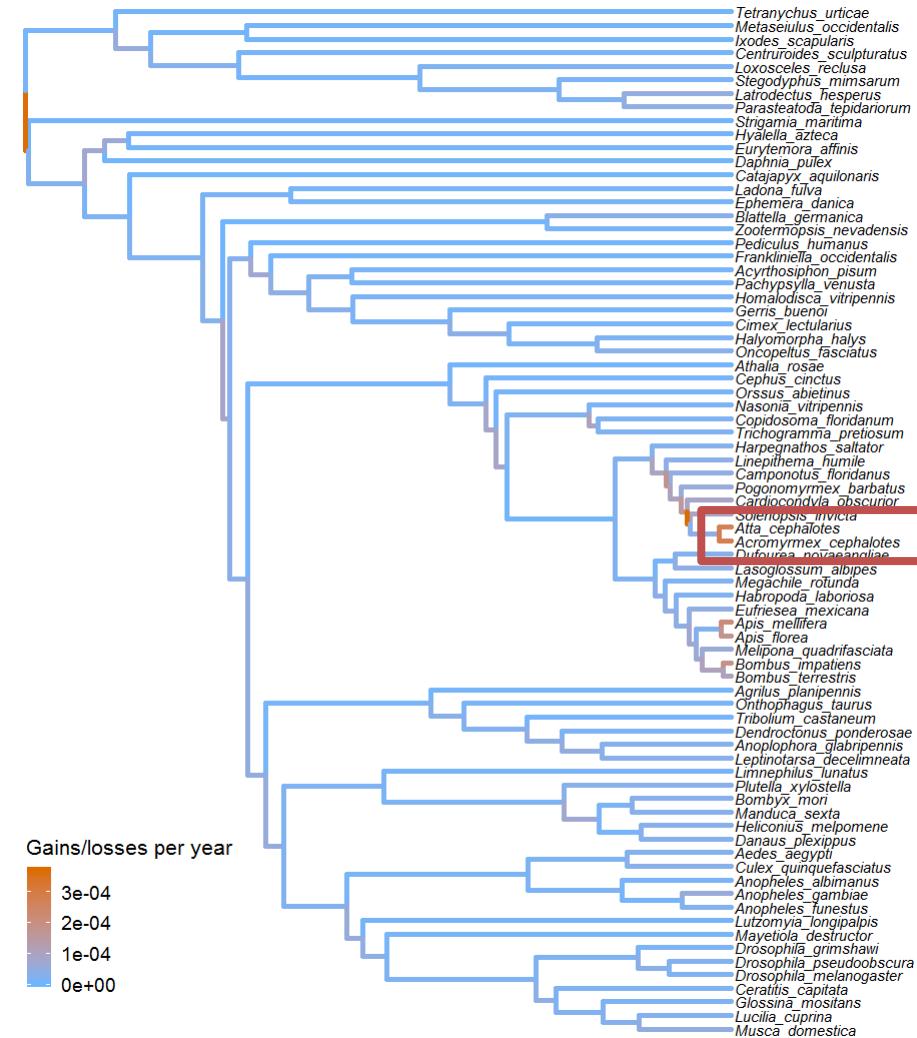
Rates are mostly consistent across arthropods

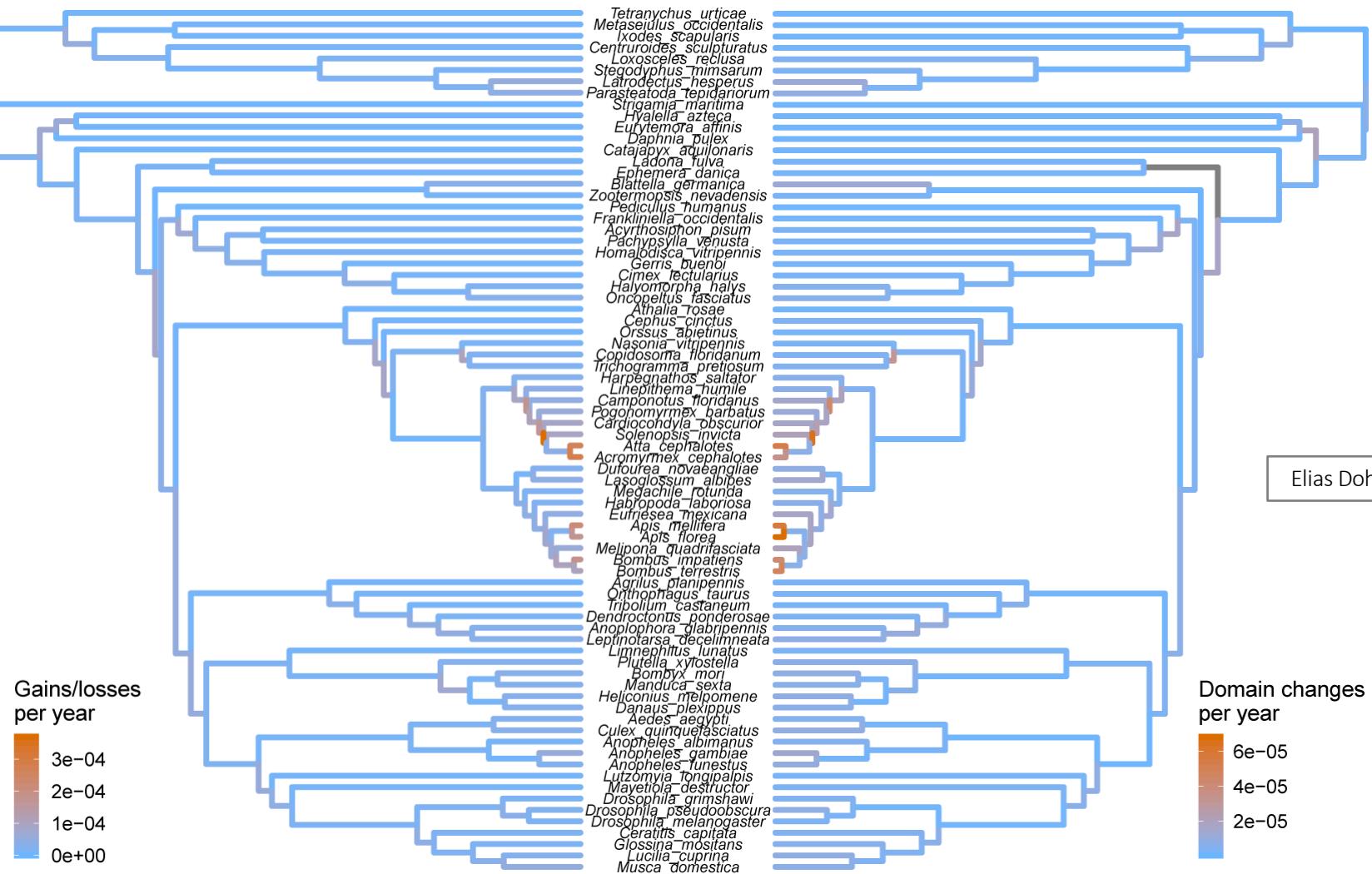


Arthropod Time Tree

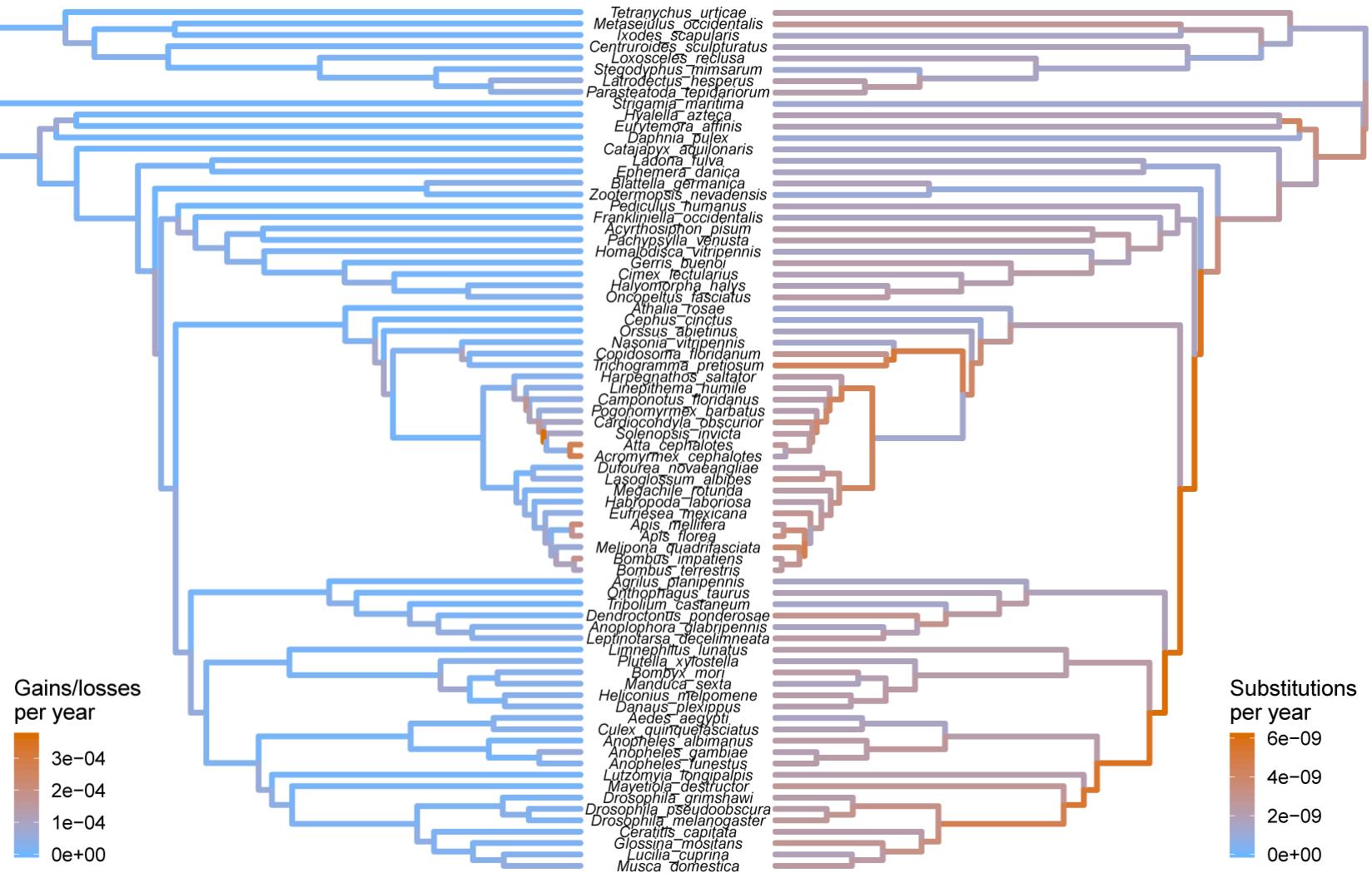
The branches of the ML tree can be scaled by time to infer lineage specific gain/loss rates

Accelerated rates in leafcutter ants

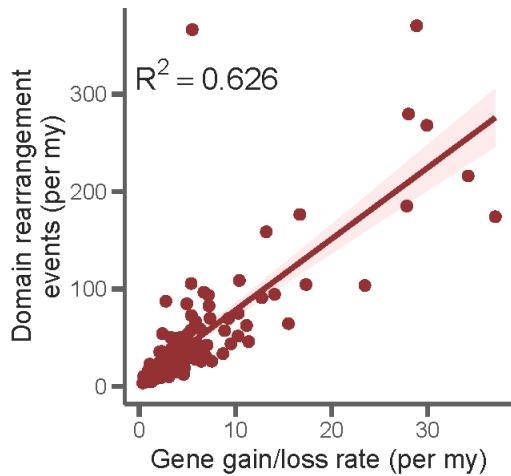




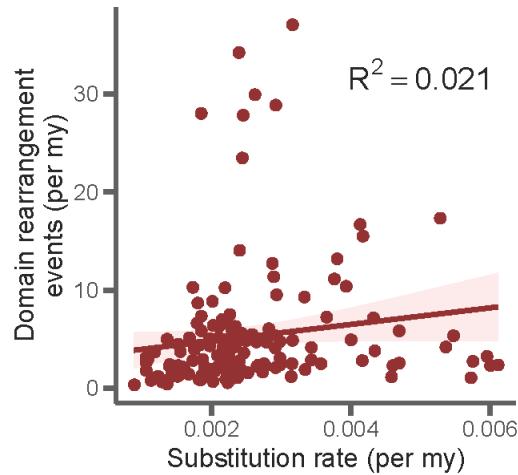
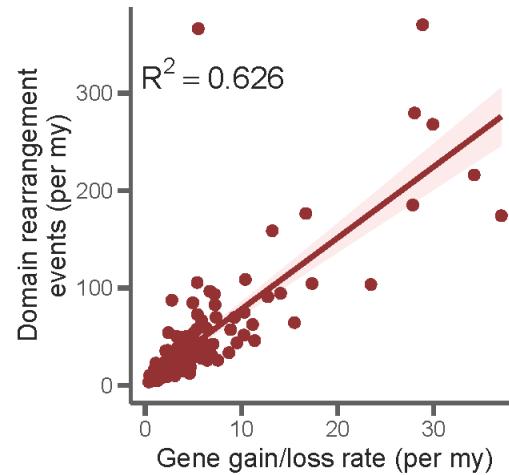
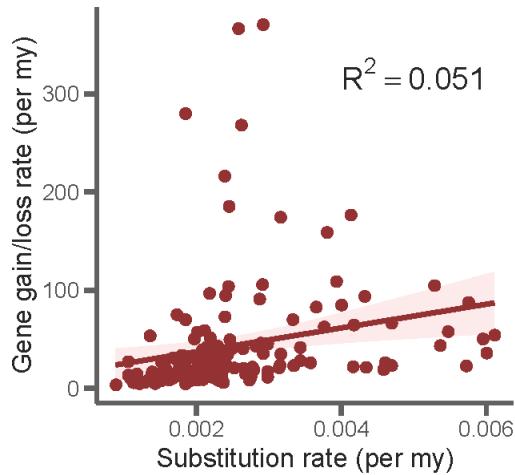
Elias Dohmen



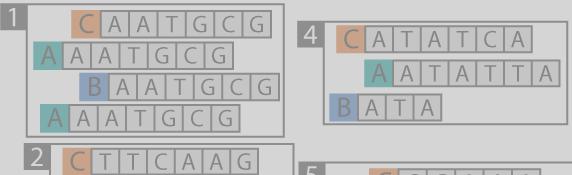
Gene gain and loss rates are correlated with protein domain rearrangements



Neither are correlated with substitution rate



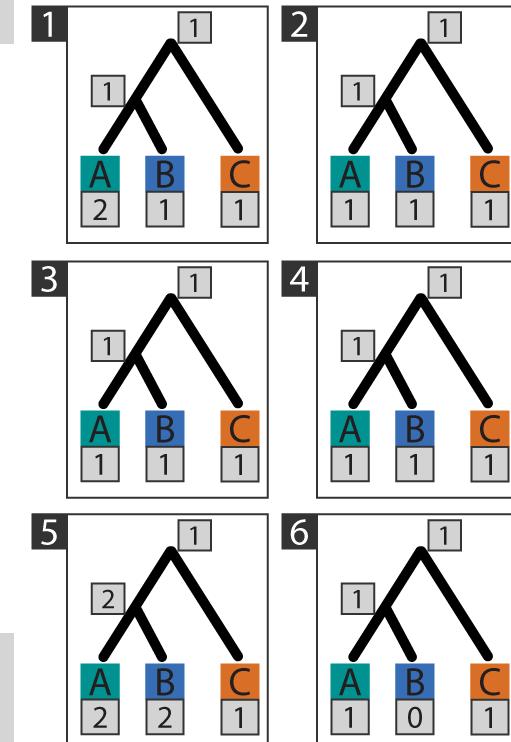
1) Predict orthogroups



3) Construct gene count matrix

	A	B	C
1	2	1	1
2	1	1	1

4) Infer ancestral gene counts

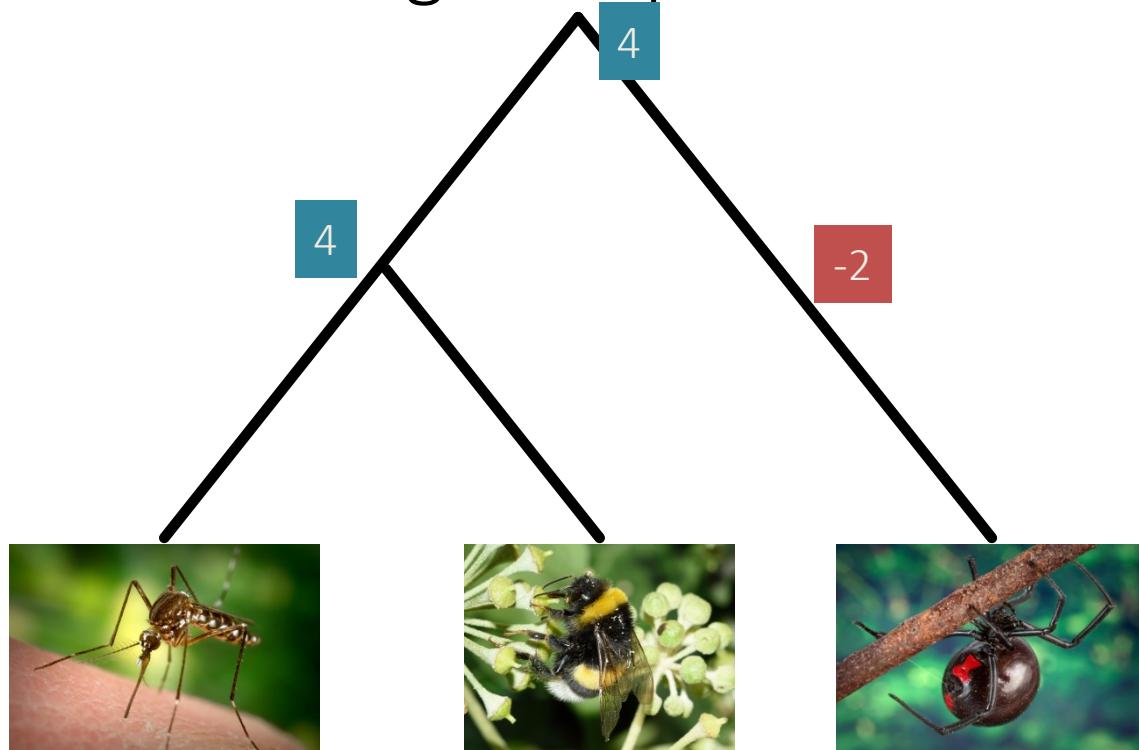


With ancestral gene counts we can:

1. Infer rates of gene gain/loss
2. Count gene gains and losses and check for rapid changes on every lineage
3. Estimate gene counts in extinct ancestors

A B C

What specific gene family changes are interesting or important?



4

4

2

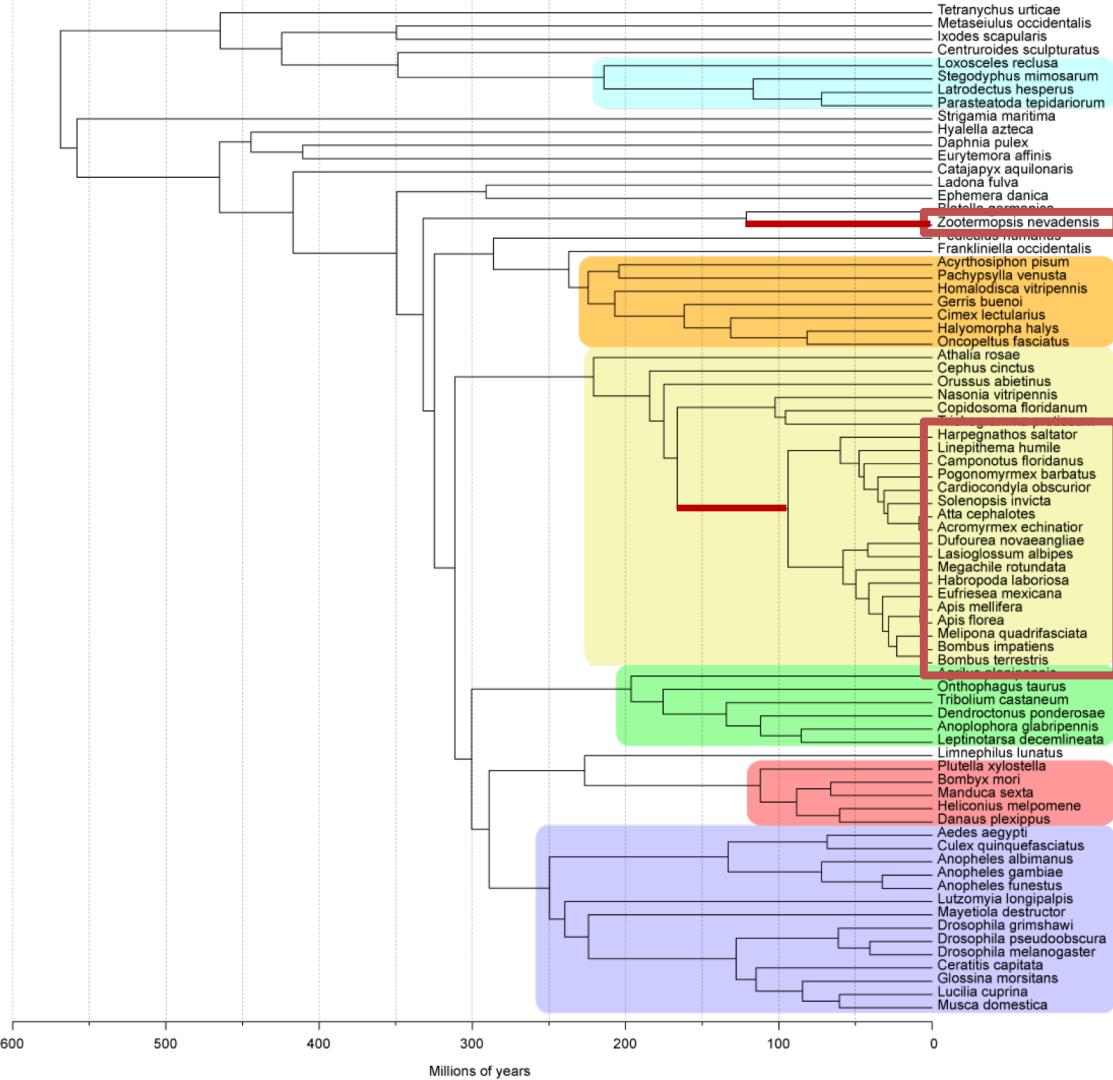
Gene copy number variation

Common gene family changes among eusocial insects

Termites



Bees & ants



Common gene family changes among eusocial insects



Termites



Bees & ants

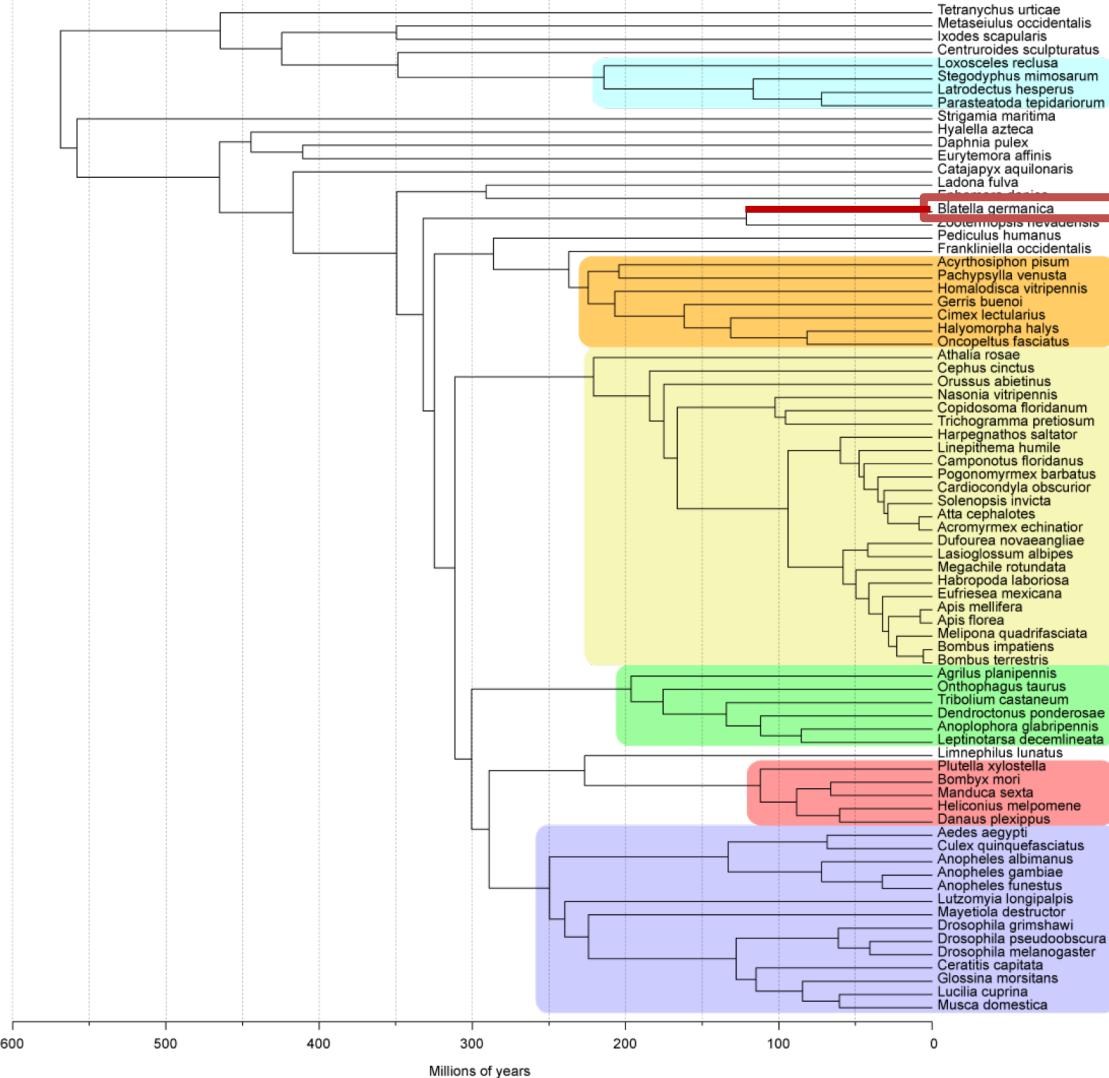


- 41 enriched functional terms in BOTH groups
- Olfactory reception and odorant binding

Largest number of gene family changes

German cockroach





Largest number of gene family changes

German cockroach

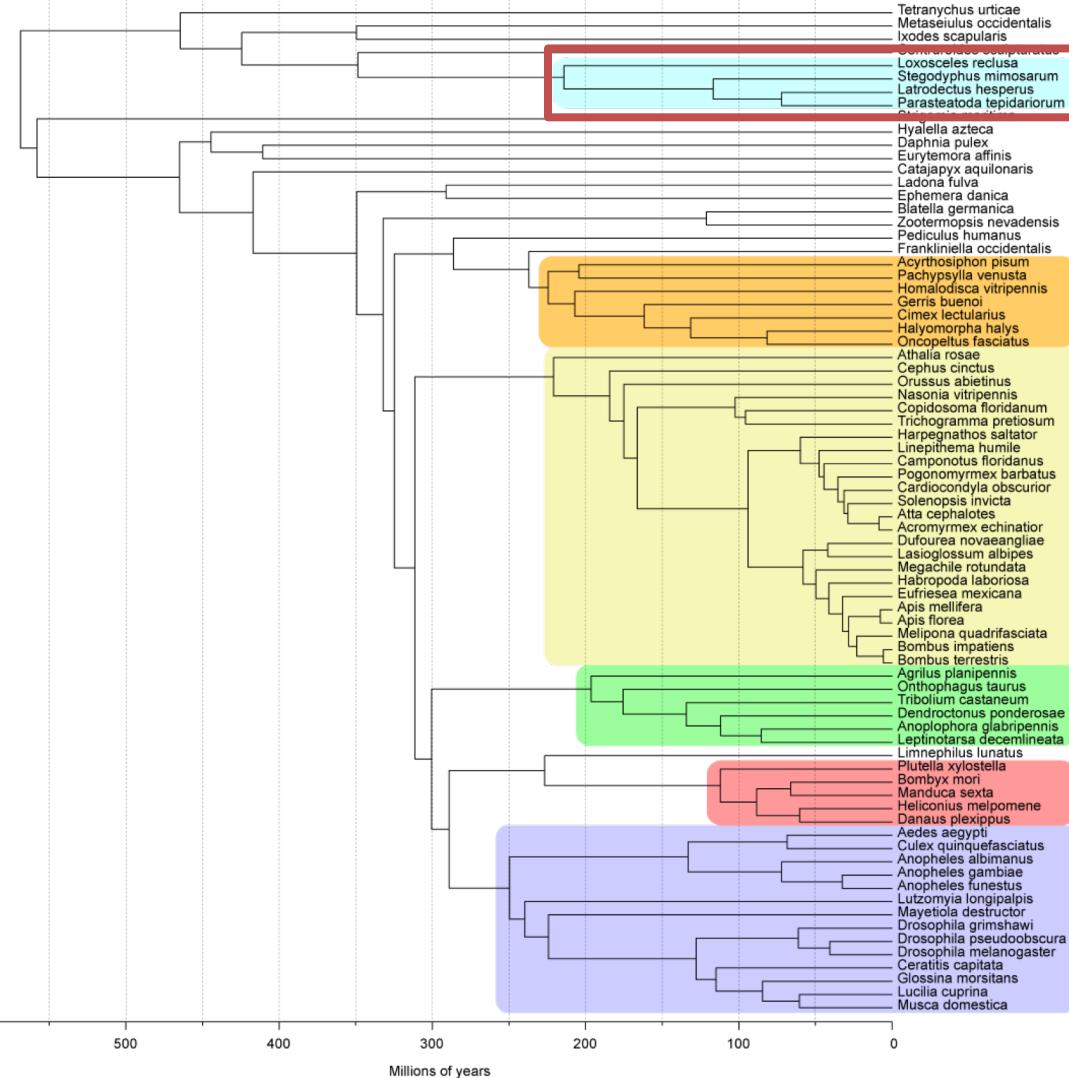


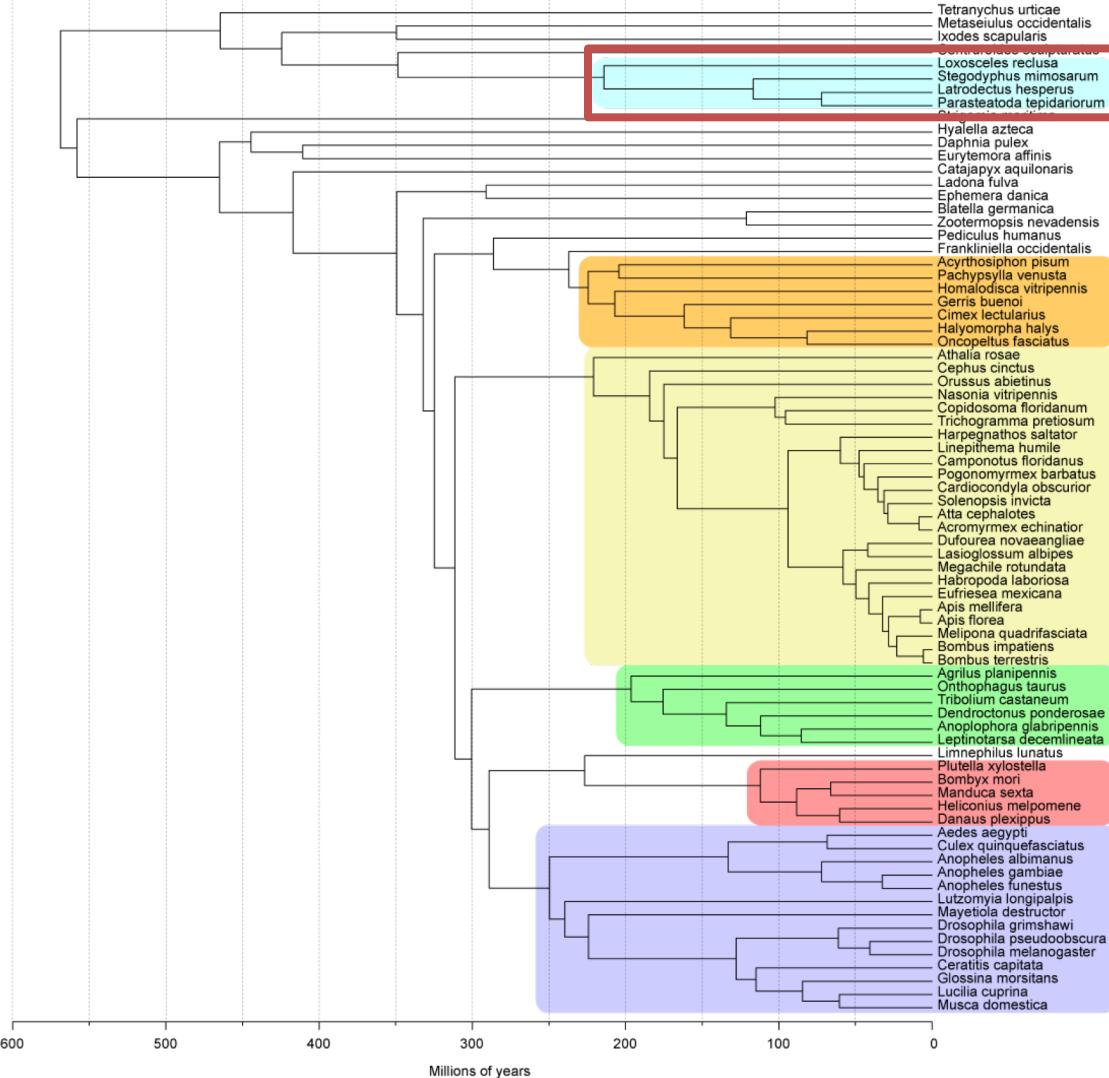
- Most RAPID gene family changes
- Major expansion of chemosensory genes
- Most protein domain rearrangements

Spider silk and venom gene families



Araneae





Spider silk and venom gene families



Araneae



- 10 rapidly expanding gene families within Araneae related for silk or venom
- High rate of protein domain emergences, including some related to venom

Jessica Garb

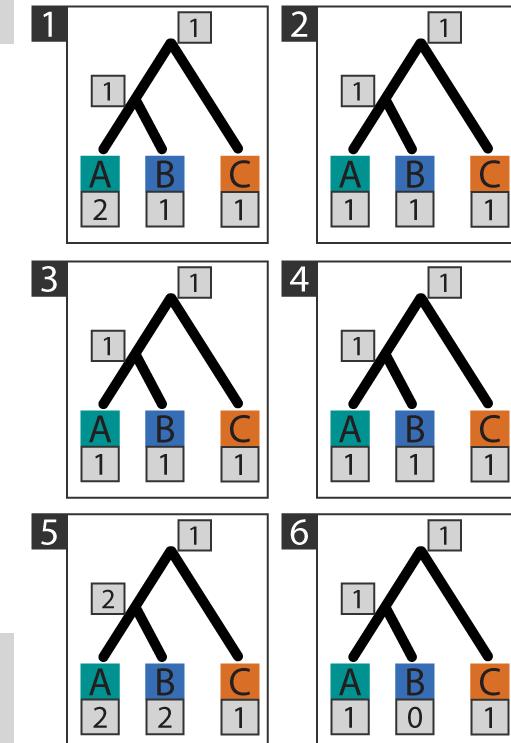
1) Predict orthogroups

1	CAATGCG
2	AAATGCG
3	BAATGCG
4	AAATGCG
5	CTTCAAG
6	CATAATCA
7	AATATTAA
8	BATA

3) Construct gene count matrix

	A	B	C
1	2	1	1
2	1	1	1

4) Infer ancestral gene counts

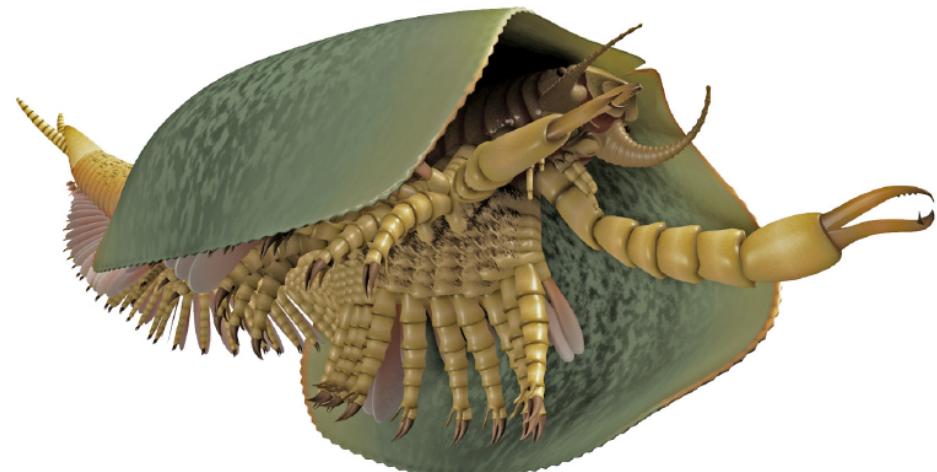
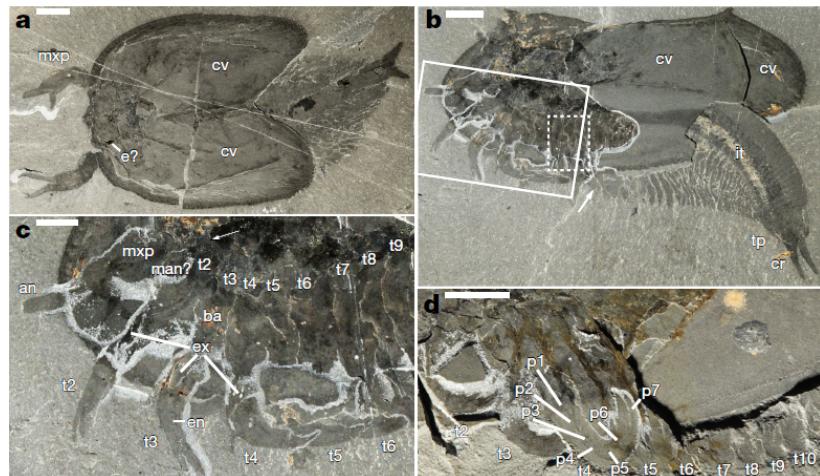


With ancestral gene counts we can:

1. Infer rates of gene gain/loss
2. Count gene gains and losses and check for rapid changes on every lineage
3. Estimate gene counts in extinct ancestors

A B C

What did the ancestral insect (LICA) look like?

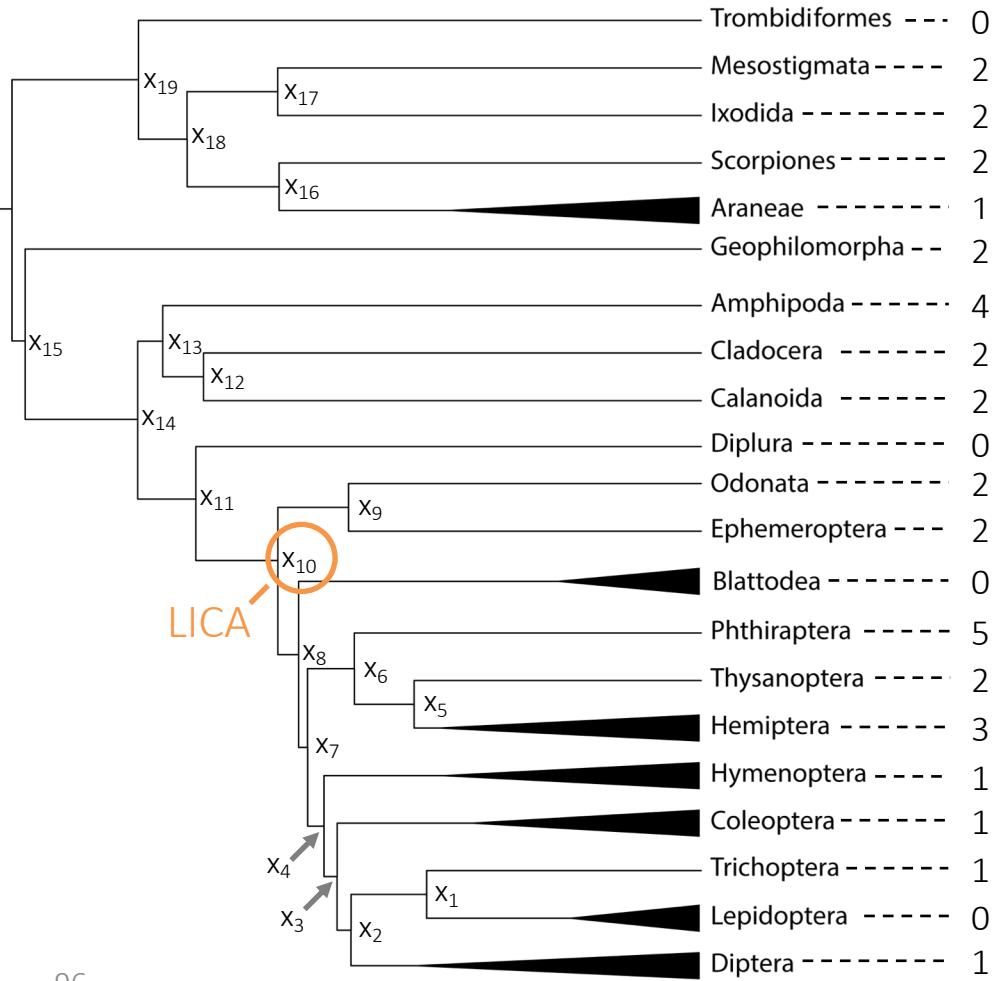


Burgess Shale fossils illustrate the origin of the
mandibulate body plan

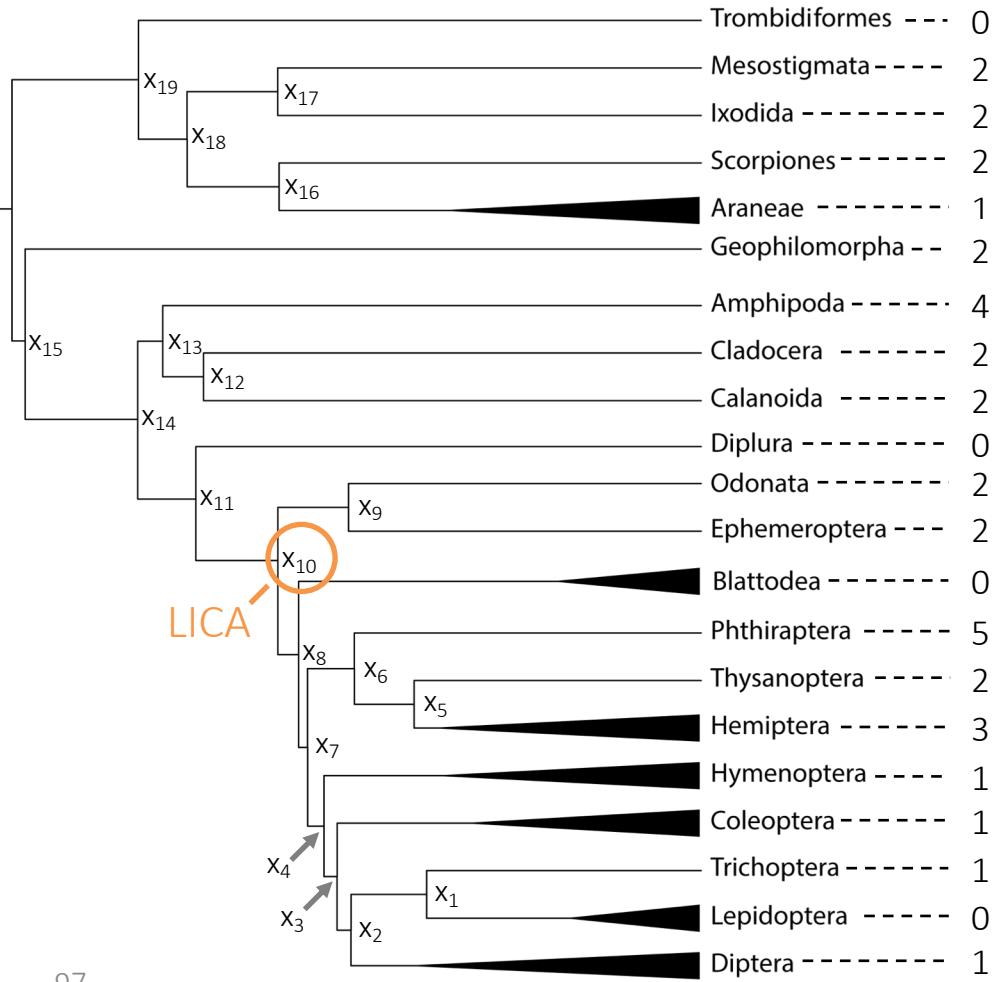
Cédric Aria^{1,2†} & Jean-Bernard Caron^{1,2,3}

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How can we infer
characteristics of the
genome of LICA?

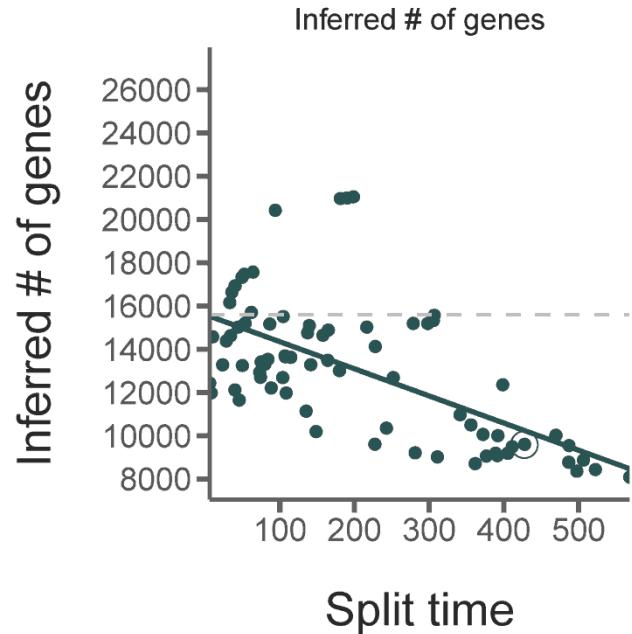


How can we infer characteristics of the genome of LICA?

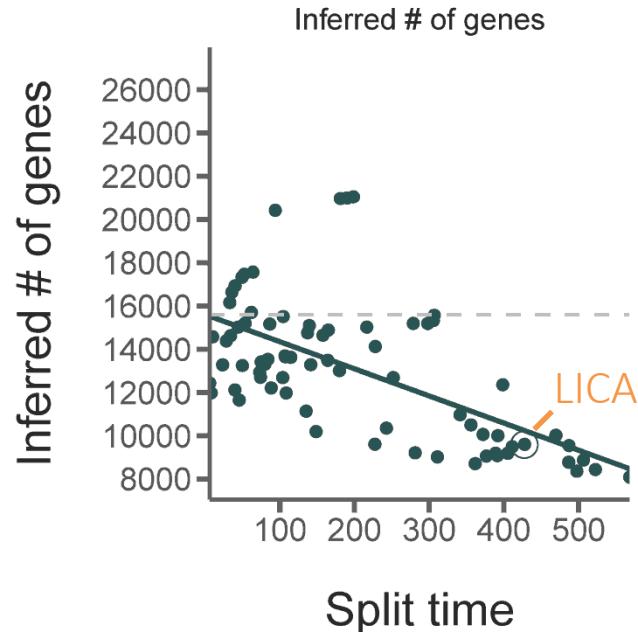


How many genes were present in the LICA genome?

Ancestral genome sizes are underestimated due to extinctions

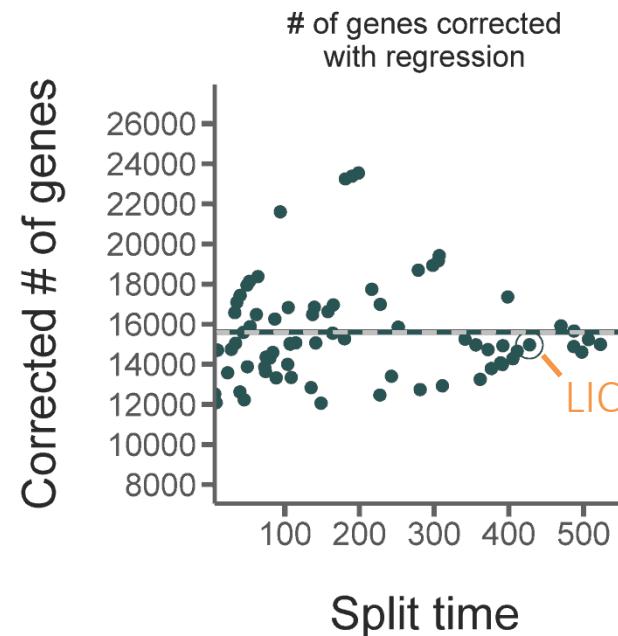
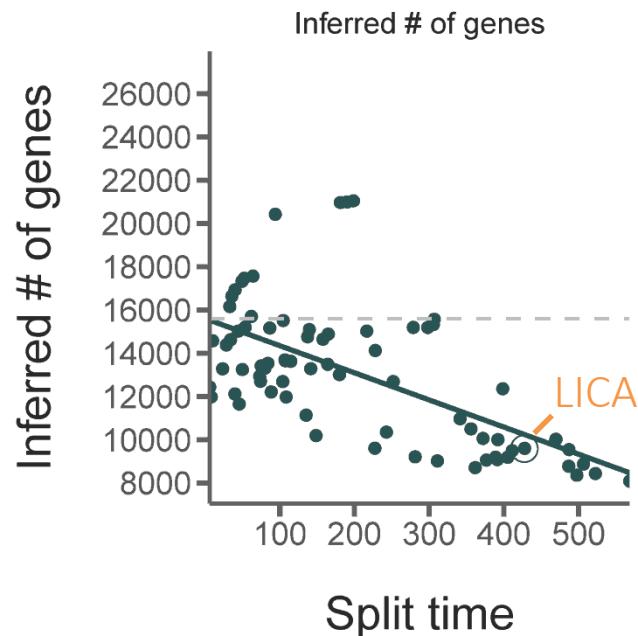


Ancestral genome sizes are underestimated due to extinctions

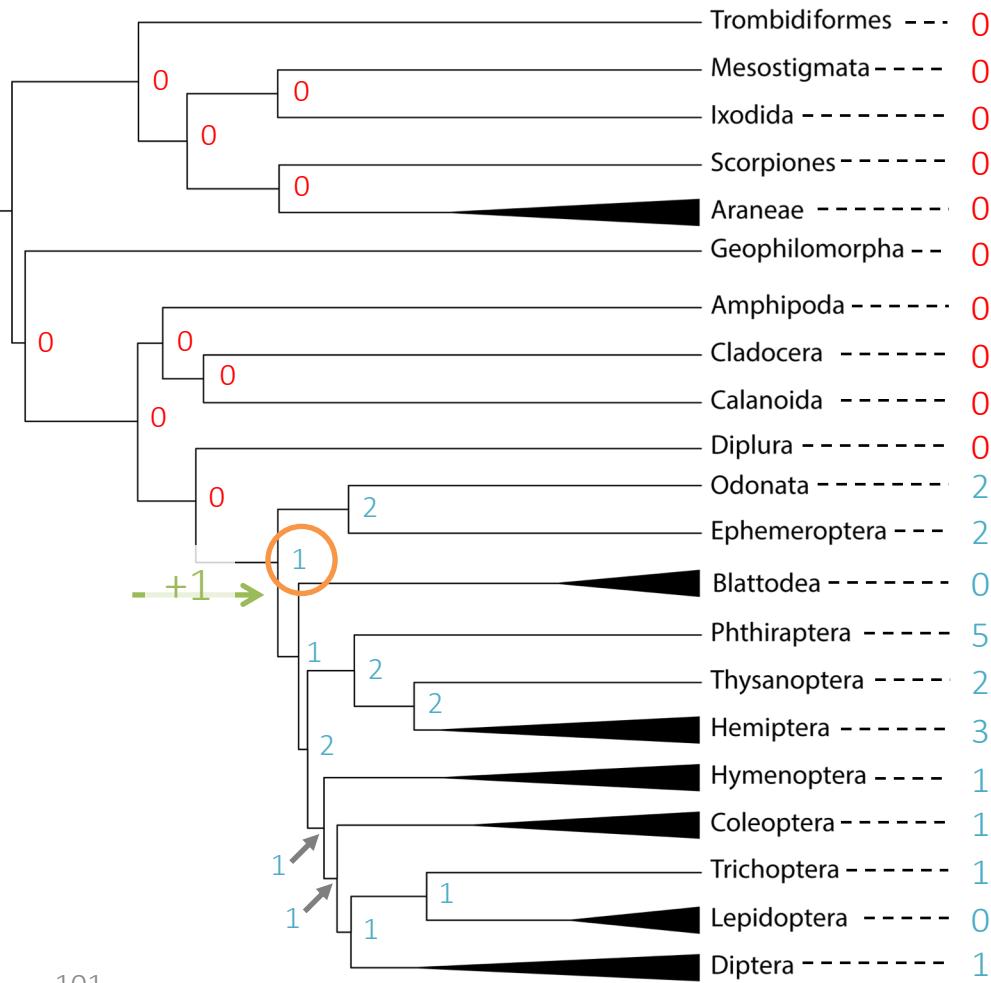


Estimated: 9,601 genes

Ancestral genome sizes are underestimated due to extinctions

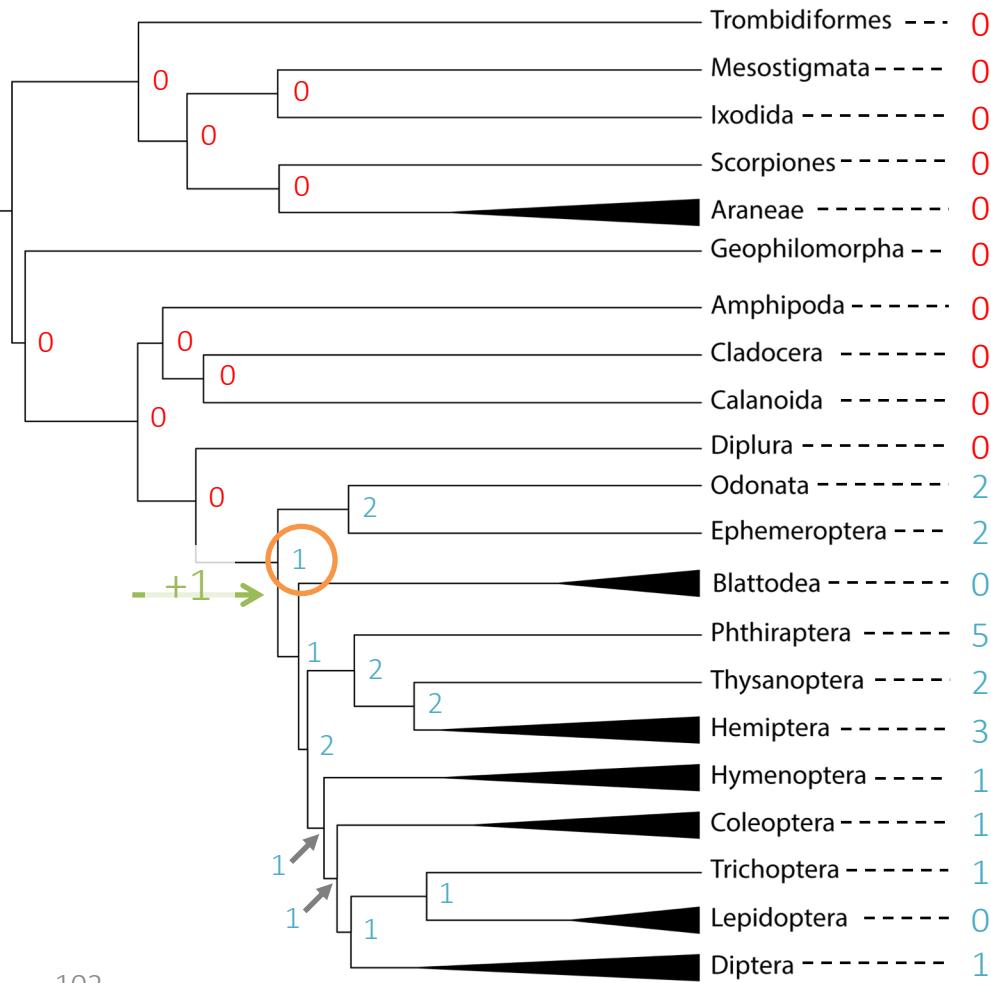


How can we infer characteristics of the genome of LICA?



Which families were 'born' during the transition to insects?

How can we infer characteristics of the genome of LICA?



Which families were 'born' during the transition to insects?

147 emergent insect families

Emergent insect families correspond to insect lifestyle adaptations

Changes in exoskeleton development

7 chitin and cuticle production families

Emergent insect families correspond to insect lifestyle adaptations

Changes in exoskeleton development

Ability to sense in a terrestrial environment

7 chitin and cuticle production families

1 visual learning and behavior family
2 odorant binding families
5 families involved in neural activity

Emergent insect families correspond to insect lifestyle adaptations

Changes in exoskeleton development

Ability to sense in a terrestrial environment

Unique development

7 chitin and cuticle production families

1 visual learning and behavior family
2 odorant binding families
5 families involved in neural activity

1 larval behavior family
4 imaginal disk development families

Emergent insect families correspond to insect lifestyle adaptations

Changes in exoskeleton development

Ability to sense in a terrestrial environment

Unique development

Flight

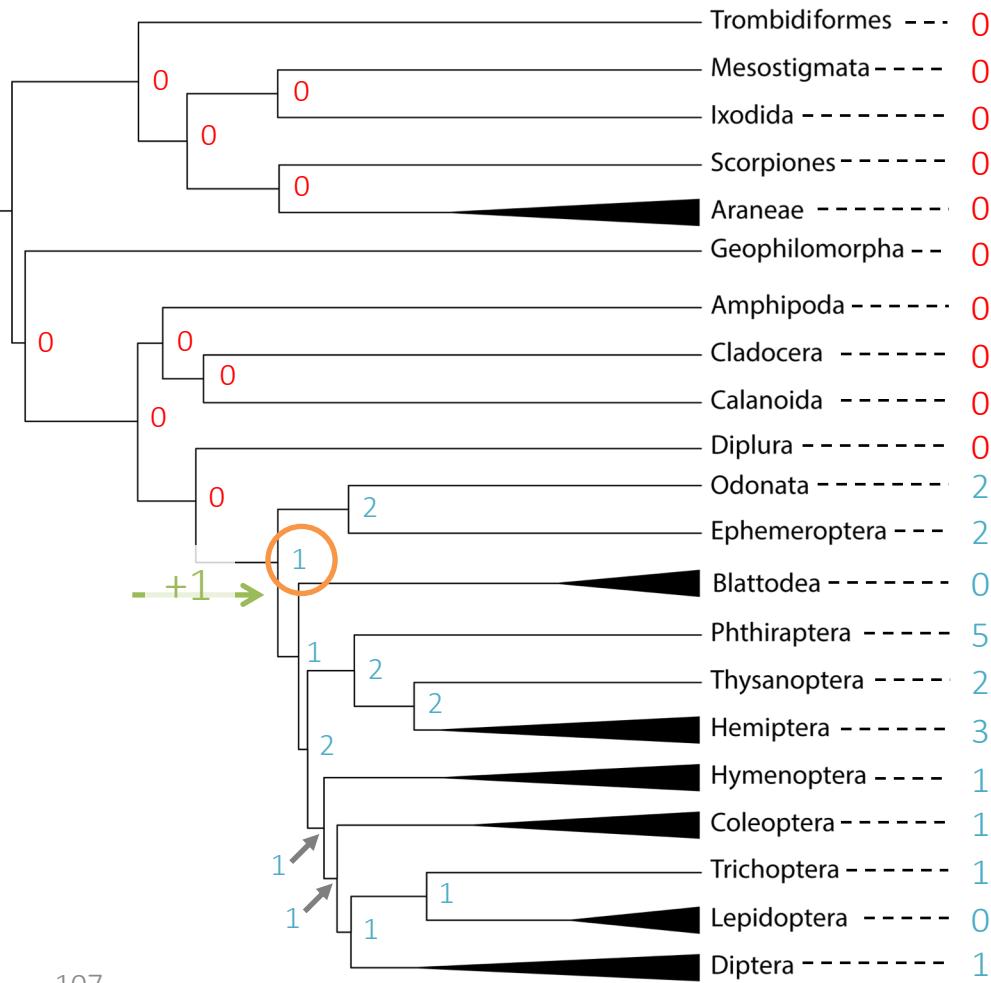
7 chitin and cuticle production families

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1 larval behavior family
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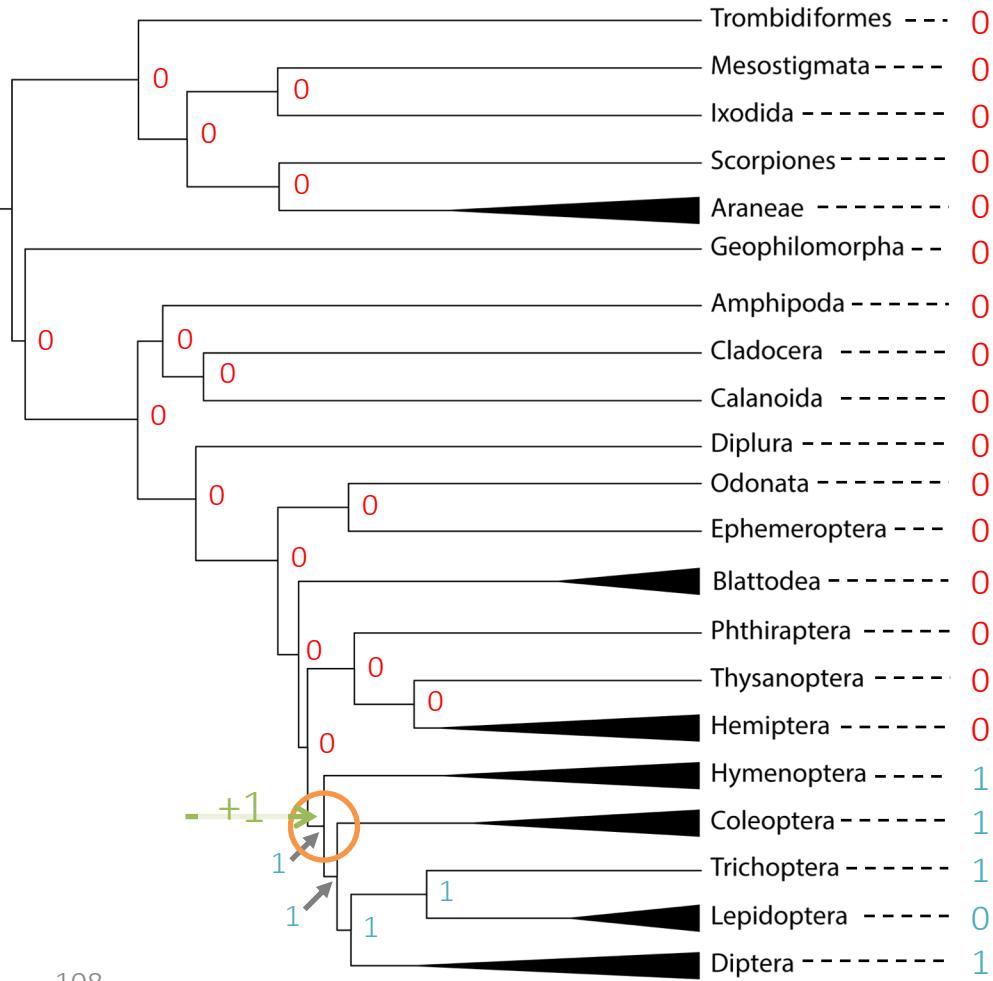
3 wing morphogenesis families

How can we infer
characteristics of the
genome of LICA?



Which families were ‘born’
during the
transition to insects?

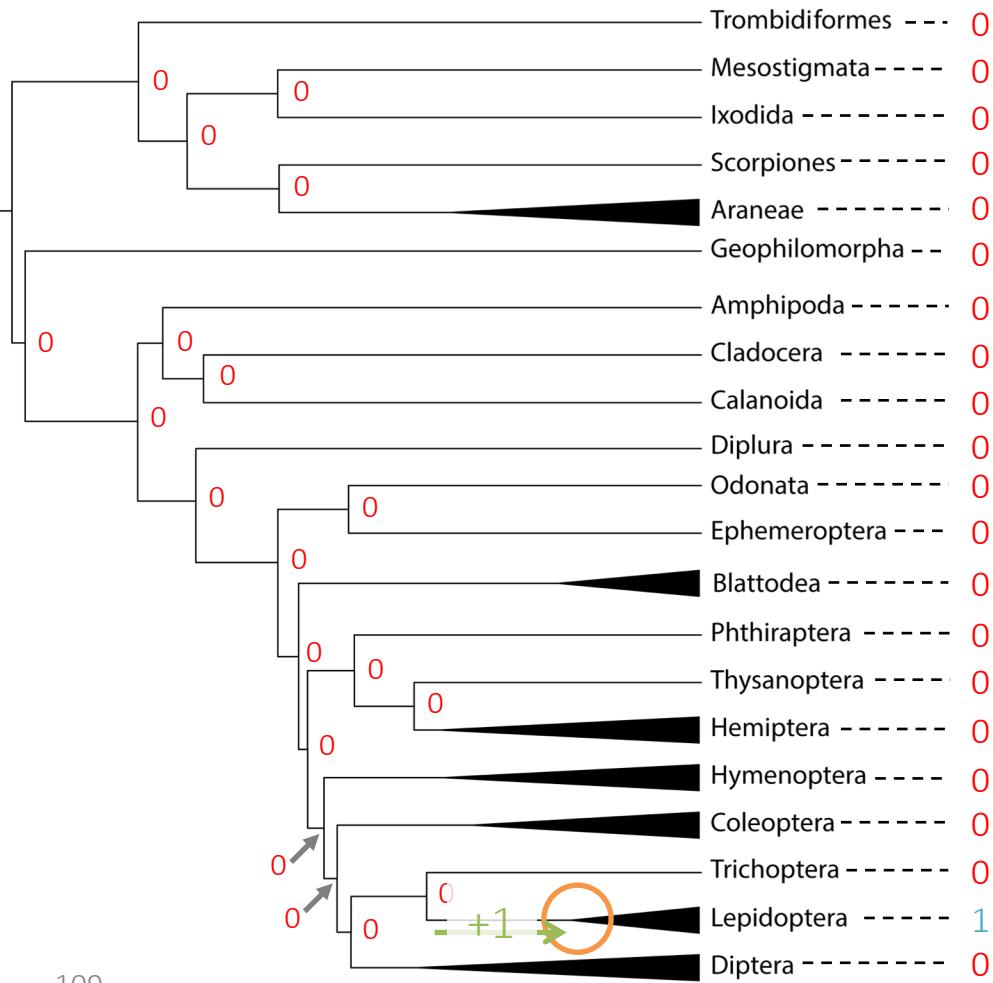
How can we infer
characteristics of the
genome of LICA?



Which families were 'born'
during the
~~transition to insects?~~
transition to Holometabola?

Only 10 emergent Holometabola
gene families

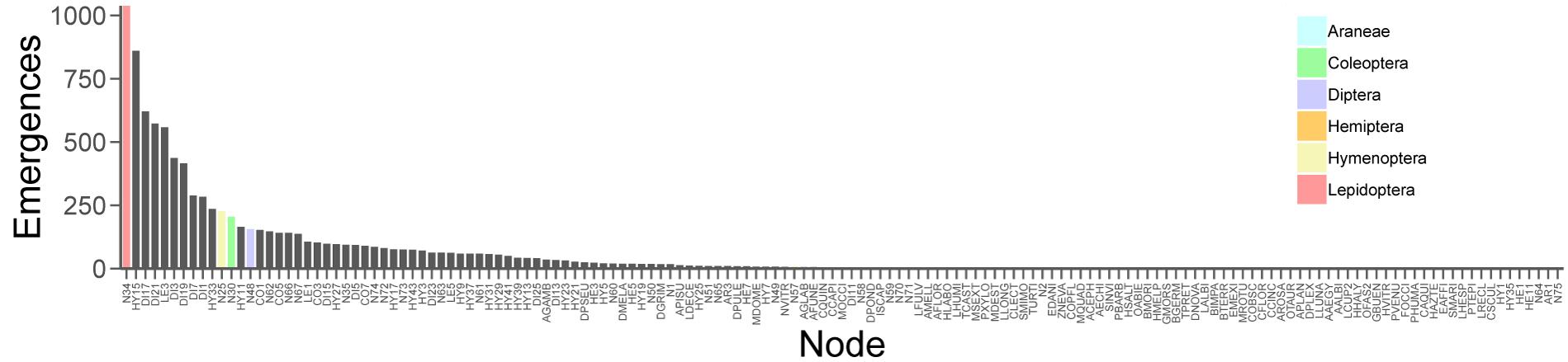
How can we infer
characteristics of the
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Which families were 'born'
during the
~~transition to insects?~~
~~transition to Holometabola?~~
~~transition to Lepidoptera?~~

1,038 emergent Lepidopteran
gene families

Lepidoptera has the most emergent gene families

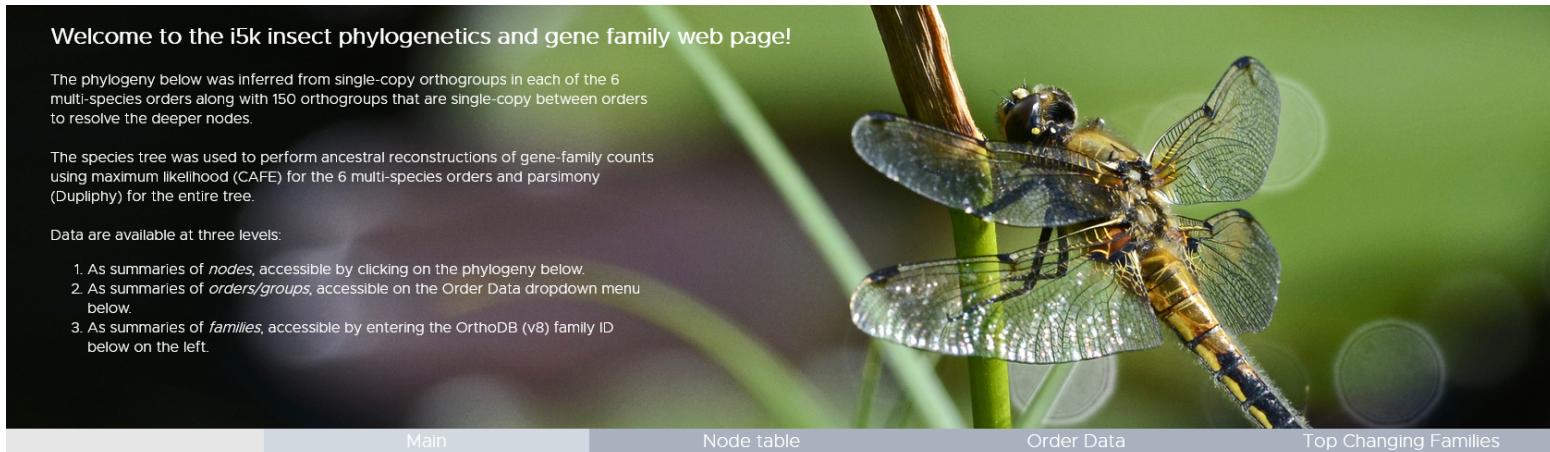


Today's topics

1. Determining the Arthropod phylogeny
2. Reconstructing ancestral gene counts
3. Using the i5k gene family web site

All data has been made available in our online tool

<https://i5k.gitlab.io/ArthroFam/>



 i5k

Jump to page

Enter an OrthoDB family ID or node ID to go to that page.

OrthoDB ID or node ID

GO TO PAGE

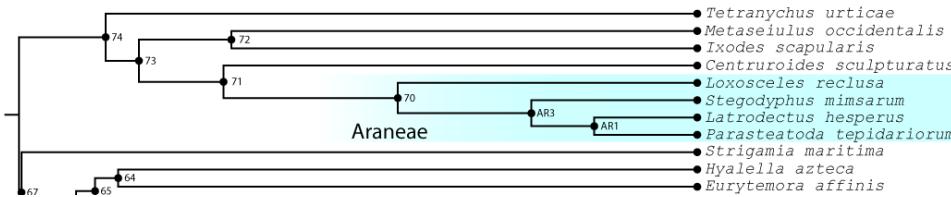
Function search

Download summaries for all nodes as:

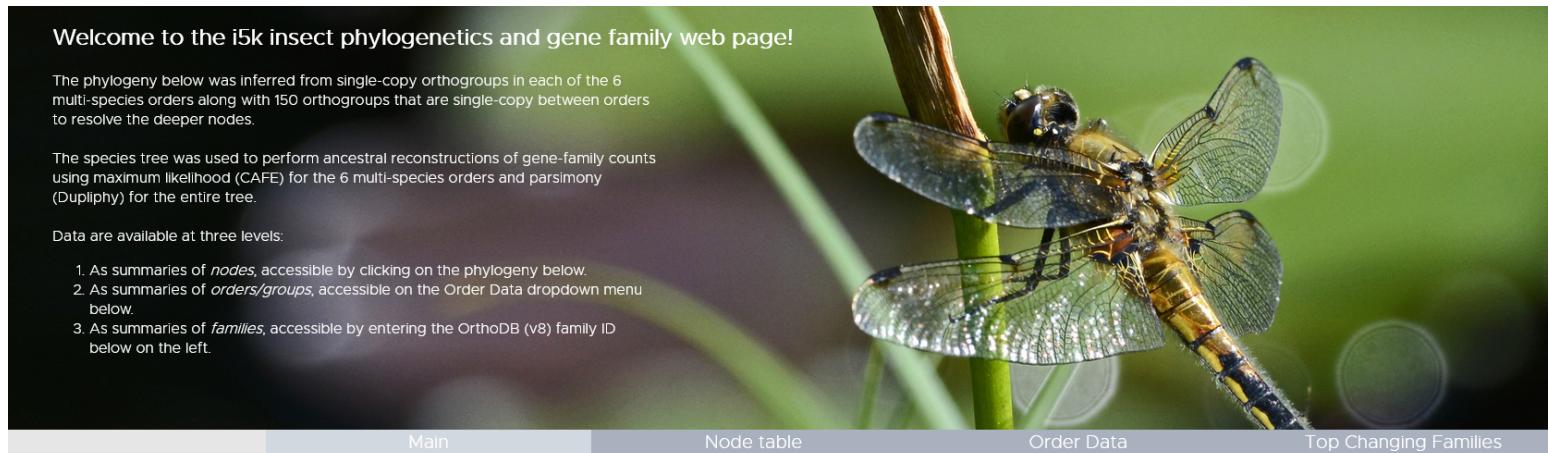
CSV file

Excel spreadsheet

Arthropod phylogeny -- Click on a node to go to that page.



Working search functions temporarily available at: <https://cgi.soic.indiana.edu/~grthomas/i5k-web/main.html>



 i5k

Jump to page

Enter an OrthoDB family ID or node ID to go to that page.

OrthoDB ID or node ID

GO TO PAGE

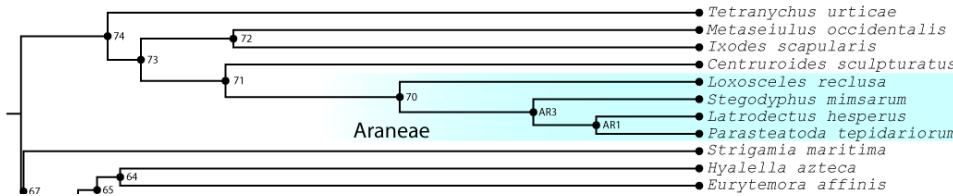
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Arthropod phylogeny -- Click on a node to go to that page.



Demo

Acknowledgements

- Matthew Hahn
 - Stephen Richards
 - Rob Waterhouse
 - Jessica Garb
 - Elias Dohmen
 - Ariel Chipman
- The i5k community
- The Hahn lab + Clara Boothby



i5k website:

<http://i5k.github.io/>

Gene family website:

<https://i5k.gitlab.io/ArthroFam/>