# 05\_03\_Seamless-cloning-reaction

#### **DIENSTAG, 1.6.2021**

## **Goal-Setting**

• Cloning of an insert and transformation into competent cells

### **Terms / abbreviations**

- cPCR = Colony polymerase chain reaction
- RT = Room temperature

#### Risk areas

None

## Required materials and / or information

- Chemicals:
  - o Chemical competent cells (E. coli C3040I)
  - o Desired insert
  - o pMA\_217 vector
- Materials:
  - 1.5 mL tubes, Eppendorf
  - Agar plates (Ampicillin 100 μg/mL)
  - GeneArt<sup>™</sup> Seamless Cloning and Assembly Enzyme Mix, Thermo Fisher Scientific
  - Ice
  - o Pipettes, Eppendorf
  - o S.O.C. Medium, Thermo Fisher Scientific or self-made
  - o Timer

## Templates, devices, software

• ThermoMixer C, Eppendorf

#### **Preliminary work**

- 05\_01\_Preparation-of-S.O.C.-medium
- 06\_03\_Casting-agar-plates

#### **Operation**

- 1. Pipette 5 µL enzyme mix into a cup (on ice)
- 2. The insert/vector ratio is 4:1. The amount of the insert is 200 ng. In addition, 50 ng of the vector is pipetted to the enzyme mix. The volume should not exceed  $5 \mu$ L.
- 3. Incubate for 20 min at RT and incubate for 5 min on ice afterwards
- 4. Add 50 µL of chemical competent cells (E. coli C3040I)
- 5. Incubate on ice for 20 min
- 6. Heat shock at 42 °C for 45 s and incubate on ice for 3 min afterwards
- 7. Add 300 µL S.O.C. medium
- 8. Incubate for 30 min at 37 °C and 300 rpm
- 9. Plating out the medium on an agar plate and incubate them for 12-16 h at 37 °C

# **Troubleshooting**

• Expected heights of the bands after cPCR for pMA\_217 vector:

Uncut vector: 763 bpRe-ligated vector: 190 bp

o Vector with control insert: 732 bp

○ Vector with AT-rich insert: 190 + ~insert length bp

## Follow-up work

None