# 03\_01 NEB Protocol for TdT Tailing Reaction

#### FREITAG, 14.5.2021

#### **Goal-Setting**

• Tailing a primer with particular dNTPs using TdT

### Terms / abbreviations

- dNTP = Deoxynucleoside triphosphate
- TdT = Terminal deoxynucleotidyl transferase

#### Risk areas

• If spilled, always wipe surface with alcohol



## Required materials and / or information

- Chemicals:
  - o 2.5 mM CoCl<sub>2</sub> solution, NEB
  - o 5.0 pmol DNA Primer
    - 5 pmol in 50  $\mu$ L is equivalent to 0.1 pmol/ $\mu$ L = 100 nM
  - o 10x TdT Reaction Buffer, NEB
    - 50 mM Potassium acetate, 20 mM Tris-acetate, 10 mM Magnesium acetate, pH 7.9 at 25 °C
  - o 10 mM dNTPs, ThermoFisher
  - o Nuclease free water, ThermoFisher
  - TdT (20 U/μL), NEB
    - 50 mM KPO<sub>4,</sub> 100 mM NaCl, 1.43 mM 2-Mercaptoethanol, 0.1 % (v/v) Triton X-100, 50 % (v/v) Glycerol, pH 7.3 at 25 °C
- Material:
  - o Gloves
  - o PCR tube rack
  - o PCR tubes, Sarstedt (autoclaved)
  - o Pipettes, Eppendorf
  - o Trash bags, Th. Geyer GmbH & Co. KG

### Templates, devices, software

None

# **Preliminary work**

• Thawing frozen ingredients

# **Operation**

- 1. Prepare the following reaction mixture
- 2. Add dNTPs and primer last
  - a. This will start the incubation immediatly
  - b. Stop the time
- 3. Incubate at X °C for X min (varying depending on experiment)

NEB TdT reaction mixture				
	А	В	С	D
1	Component	Stock concentration	Desired concentration	Volume [mL]
2	H2O	-	-	38.5
3	TdT Buffer	10x	1x	5
4	CoCl2	2.5 mM	0.25 mM	5
5	TdT	20 U/μL	0.2 U/μL	0.5
6	dNTP	10 mM	100 μΜ	0.5
7	Primer	10 μΜ	100 nM	0.5
8	total			50

# **Disposal**

• Autoclave trash bags, discard in S1 waste

## **Troubleshooting**

- Wear gloves to reduce the risk of DNAse and RNAse contamination
- Always keep TdT cooled until usage

# Follow-up work

- 03\_02 Heat Inactivation of TdT Reaction
- 03\_03 EDTA Inactivation of TdT Reaction
- If materials are empty care about new order