WT

Marginal Blastozone vs Other, B region (middle) attempt 1

```
library(edgeR)
```

Read in Data

Read in raw count data per gene. Add checknames to FALSE because it was making the columns unique.

```
counts <- read.delim("../sam2countsResults.tsv",check.names=FALSE,row.names=1)

#check the file
head(counts)
summary(counts)
colnames(counts)
#need to convert NA to 0 counts
counts[is.na(counts)] <- 0</pre>
```

Subset per DE expirement

I am going to start by subsetting the particular treatments I am looking at. In this case I am going to get rid of wtbmbr8 and wtbother1.4, because their count are very low and this could be the reason I am getting the errors from attempt 1.

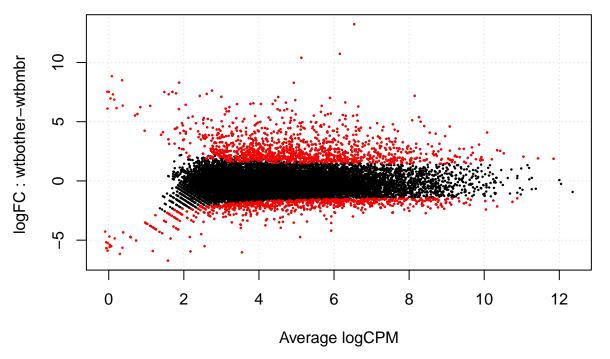
```
colnames(counts)
```

```
##
    [1] "tf2ambr1"
                         "tf2ambr3"
                                          "tf2ambr4"
                                                           "tf2ambr6"
                         "tf2aother2"
##
    [5] "tf2aother1"
                                                           "tf2aother7"
                                          "tf2aother4"
   [9] "tf2bmbr2"
                         "tf2bmbr5"
                                          "tf2bmbr6"
                                                           "tf2bother1"
## [13] "tf2bother3"
                         "tf2bother4"
                                          "tf2bother6"
                                                           "tf2cmbr1.4"
## [17] "tf2cmbr3"
                         "tf2cmbr6"
                                          "tf2cmbr7"
                                                           "tf2cother2"
## [21] "tf2cother5"
                         "tf2cother6"
                                          "tf2cother7"
                                                          "wtambr2"
## [25] "wtambr4"
                         "wtambr5"
                                                           "wtaother5"
                                          "wtaother1"
## [29] "wtaother6"
                                                           "wtbmbr2"
                         "wtaother7"
                                          "wtaother8"
                                          "wtbmbr8"
## [33]
       "wtbmbr3"
                         "wtbmbr6"
                                                           "wtbother1.4"
## [37] "wtbother3"
                         "wtbother5"
                                          "wtbother8"
                                                           "wtcmbr10"
## [41] "wtcmbr1.4.6"
                         "wtcmbr2"
                                          "wtcmbr3"
                                                           "wtcmbr7"
## [45] "wtcmbr9"
                         "wtcother1.3.4" "wtcother2"
                                                           "wtcother6"
wtbregion <- counts[,c(32:34, 37:39)]
head(wtbregion)
```

```
## wtbmbr2 wtbmbr3 wtbmbr6 wtbother3 wtbother5 wtbother8
## Solyc00g005040.2.1 2 4 3 8 0 3
## Solyc00g005050.2.1 20 5 18 25 0 14
```

```
## Solyc00g005060.1.1
                           1
                                            1
                                                       0
                                                                 0
                                                                            0
## Solyc00g005070.1.1
                           14
                                   6
                                            12
                                                       6
                                                                 2
                                                                            4
## Solyc00g005080.1.1
                           25
                                            27
                                                      29
                                   15
                                                                 0
                                                                           11
## Solyc00g005150.1.1
                            0
                                    0
                                             3
                                                       2
                                                                 0
                                                                            2
colnames(wtbregion)
## [1] "wtbmbr2"
                                            "wtbother3" "wtbother5" "wtbother8"
                   "wtbmbr3"
                                "wtbmbr6"
group <- c(rep("wtbmbr", 3), rep("wtbother", 3))</pre>
d <- DGEList(counts=wtbregion,group=group)</pre>
d$samples
##
                group lib.size norm.factors
## wtbmbr2
               wtbmbr 1355352
## wtbmbr3 wtbmbr 1213142
                                           1
## wtbmbr6
               wtbmbr 1598917
                                           1
## wtbother3 wtbother 1076939
                                           1
## wtbother5 wtbother 200587
                                           1
## wtbother8 wtbother
                        499487
cpm.d <- cpm(d)</pre>
d \leftarrow d[rowSums(cpm.d>5)>=3,]
d <- estimateCommonDisp(d, verbose=T) #No error this time.
## Disp = 0.3968 , BCV = 0.6299
d <- calcNormFactors(d)</pre>
d <- estimateCommonDisp(d)</pre>
DEtest <- exactTest(d,pair=c("wtbmbr","wtbother"))</pre>
head(DEtest$table)
##
                        logFC logCPM
                                         PValue
## Solyc00g005050.2.1 0.8282 3.960 4.221e-01
## Solyc00g005070.1.1 -0.0464 3.194 1.000e+00
## Solyc00g005080.1.1 0.1422 4.260 1.000e+00
## Solyc00g005440.1.1 -0.6585 4.723 4.205e-01
## Solyc00g005840.2.1 3.8416 6.975 2.742e-06
## Solyc00g006470.1.1 1.7425 8.663 1.898e-02
sum(DEtest$table$PValue<.05)</pre>
## [1] 1985
summary(decideTestsDGE(DEtest,p.value=.05))
      [,1]
## -1
         64
## 0 13894
        502
## 1
```

```
sig.genes <- rownames(DEtest$table[DEtest$table$PValue<0.05,])
plotSmear(d,de.tags=sig.genes)</pre>
```



Subset by all the ones with a significant score

```
results.sig <- subset(DEtest$table, DEtest$table$PValue < 0.05)
```

What are the genes that are misexpressed? For this we need to add some annotation

```
annotation1<- read.delim("../ITAG2.3_all_Arabidopsis_ITAG_annotations.tsv", header=FALSE) #Changed to
colnames(annotation1)<- c("ITAG", "SGN_annotation")
annotation2<- read.delim ("../ITAG2.3_all_Arabidopsis_annotated.tsv")
annotation <- merge (annotation1, annotation2, by =1,1, all.x=TRUE)
head(annotation)
results.annotated <- merge(results.sig,annotation,by.x="row.names",by.y="ITAG",all.x=T,sort=F)</pre>
```

Write table with results

```
write.table(results.annotated, "wtbmbr_wtbother_DE1.attempt2.txt", sep="\t", row.names=F)
```