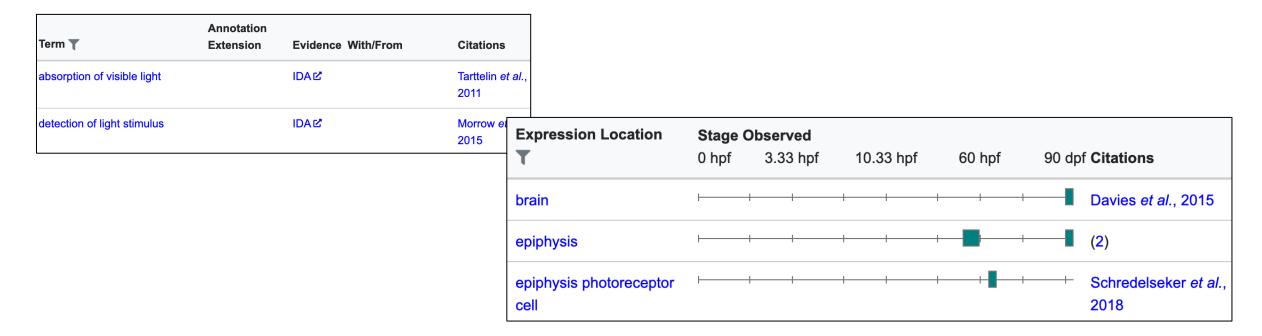
Zebrafish Anatomical Term Enrichment

ZFIN annotation

- As well as annotating genes with GO terms, ZFIN associate genes with anatomical terms (based on literature review and in situ images submitted directly)
- For example, rhodopsin (https://zfin.org/ZDB-GENE-990415-271)



Anatomical Hierarchy

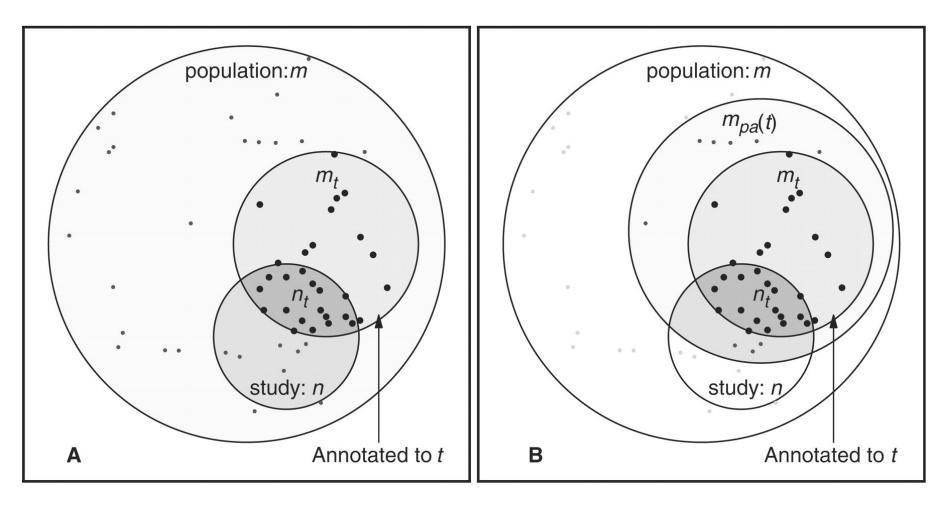
 The zebrafish anatomy (ZFA) terms form a tree-like hierarchy, where each term can have children and parents

```
* ZFA:0009127 photoreceptor cell
  * ZFA:0009154 eye photoreceptor cell
  * ZFA:0009262 retinal cone cell
  * ZFA:0009275 retinal rod cell
  * ZFA:0009219 visible light photoreceptor cell
  * ZFA:0009220 photopic photoreceptor cell
        * ZFA:0009222 blue sensitive photoreceptor cell
        * ZFA:0009223 green sensitive photoreceptor cell
        * ZFA:0009224 red sensitive photoreceptor cell
        * ZFA:0009221 UV sensitive photoreceptor cell
```

Anatomical Enrichment

- To check for enrichment of terms, can't just check each term as if they are independent of each other
 - They aren't, so you'll get spurious enrichments for some child terms of terms that are significantly enriched
- Instead check for enrichment of a term in the context of the genes annotated to the term's parents

Anatomical Enrichment



From Grossman et al, 2007

Files

- Need zfin.txt file linking ZFIN gene IDs to ZFA terms: https://funcgen2022.buschlab.org/downloads/zfin.txt
- And zfa.obo file describing all ZFA terms: https://funcgen2022.buschlab.org/downloads/zfa.obo
- http://ontologizer.de/ is software for testing enrichment of ontology terms
- Need Ontologizer.jar file: https://funcgen2022.buschlab.org/downloads/Ontologizer.jar
- All files are also available on "penelopeprime"

How Files Made

- wget https://raw.githubusercontent.com/cerivs/zebrafish-anatomical-ontology/master/zfa.obo
- wget http://zfin.org/downloads/phenoGeneCleanData_fish.txt
- wget http://zfin.org/downloads/wildtype-expression_fish.txt
- wget http://ontologizer.de/cmdline/Ontologizer.jar
- cut -f3,4,8,13,17 phenoGeneCleanData_fish.txt | grep ZDB-GENE- |
 awk '{ print \$1 "\t" \$2 "\n" \$1 "\t" \$3 "\n" \$1 "\t" \$4 "\n" \$1
 "\t" \$5 }' | grep ZFA: > zfin1.tmp
- cut -f1,4,6 wildtype-expression fish.txt | grep ZDB-GENE- | awk '{
 print \$1 "\t" \$2 "\n" \$1 "\t" \$3 }' | grep ZFA: > zfin2.tmp
- sort -u zfin1.tmp zfin2.tmp | awk '{ print "ZFIN\t" \$1 "\t" \$1
 "\t\t" \$2 "\tRef\tND\t\tC\t" \$1
 "\t\tgene_product\ttaxon:7955\t20220929\tZFIN" }' > zfin.txt
- rm phenoGeneCleanData_fish.txt wildtype-expression_fish.txt zfin1.tmp zfin2.tmp

More Files

 Also need a file listing all the detectable Ensembl IDs (the population set):

```
awk -F"\t" '$3 != "NA"' Amp.counts.tsv | cut -
f1 | grep ENS > population.tsv
```

- And a file listing all the significant Ensembl IDs (the study set):

 awk -F"\t" '\$3 < 0.05' Amp.counts.tsv | cut -f1

 | grep ENS > study.tsv
- Then use BioMart to convert Ensembl IDs to ZFIN IDs to make population.zfin.tsv and study.zfin.tsv

Best population set?

- Population set should be all the genes that are detectable
- Do NOT just use all the genes in the genome (which some online tools default to)
- Instead either:
 - Use all the genes that are above some threshold number of counts (with 1 count not being unreasonable)
 - If you're using DESeq2, use all the genes that haven't been subjected to independent filtering (i.e. those that don't have "NA" for their adjusted p-value)
 - See
 <u>http://bioconductor.org/packages/devel/bioc/vignettes/DESeq2/inst/doc/DESeq2.ht</u>

 <u>ml#independent-filtering-of-results</u> for more info on independent filtering

Run Ontologizer

- java -jar Ontologizer.jar -a zfin.txt -g zfa.obo -c Parent-Child-Union -m Bonferroni -p population.zfin.tsv -s study.zfin.tsv
- Will produce a file called table-study.zfin-Parent-Child-Union-Bonferroni.txt containing all the ZFA terms, including significantly enriched ones

Ontologizer Output

• cut -f1,11,13 table-study.zfin-Parent-Child-Union-Bonferroni.txt | awk '\$2 < 0.05'

Thank You

Any Questions?