

## Repeated measures, simple methods

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## Plan for this week

## 09\_Repeated measures

Monday Statistical inference, and the t-test

Tuesday Simple and Multiple regression

Wednesday ANOVA, ANCOVA, and linear models

Thursday Categorical data, Writing statistical reports,  
Logistic regression

Friday Repeated measurements, Principal Component  
Analysis

# Overview of this module

## 09\_Repeated measures

- 1 The repeated measurements setup
  - Aspirin Example
  - Activity of rats
- 2 Separate analysis for each time-point
  - Example: rats data
- 3 Analysis of a summary statistic
  - Example: rats data
- 4 Random effects model - simple version
  - Example: rats data
- 5 Random effects model - advanced version
- 6 Pros and cons of simple approaches

# Aim of this module

## 09\_Repeated measures

- Present simple methods for dealing with repeated measurements - and a few more advanced applications
- Easy to use, and a lot better than pretending to have independent observations
- Useful even after more advanced models are presented
- See how to specify these models in R

# Overview

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# The repeated measurements setup

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- Several "individuals"
- Several measurements on each individual
- Two measurements on the same individual might be correlated
- Might even be highly correlated if "close" and less correlated if "far apart"

# The repeated measurements setup

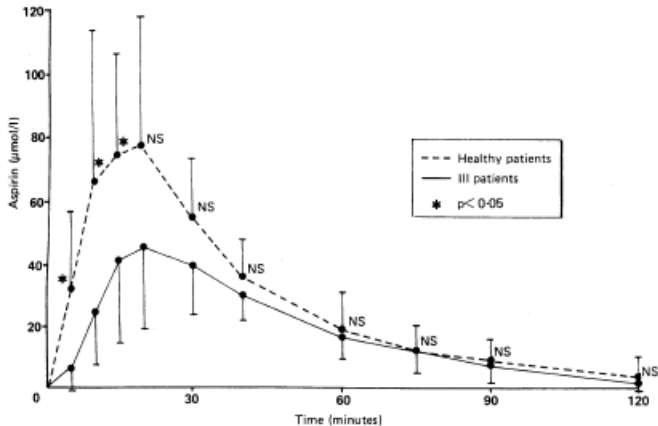
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- Several "individuals"
- Several measurements on each individual
- Two measurements on the same individual might be **correlated**
- Might even be highly correlated if "**close**" and less correlated if "**far apart**"
- **Typical example** (Matthews et al 1990):
  - Two groups of patients (ill/healthy)
  - Treated with aspirin
  - The aspirin concentration is measured in the blood 10 times during 2 hours

*To pretend all observations are independent can lead to wrong conclusions*

# Traditional figure of means at each time point

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# What is the aim of the study?

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It is always important to consider the aim of the study!

## 1 To describe pattern over time

- Do we have a change over time?
- Linear or curve?
- Is the pattern the same for all groups?

## 2 Can we see a difference between groups?

- Is the difference the same at all time points?
- Is the difference in the levels?
- Is the difference in the trend?

# Problems using the traditional presentation of the data

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- Comparing the groups at each time point.
  - Not efficient
  - Tests are not independent, carried out on the same individuals
  - Difficult to interpret
- The pattern over time
  - This can only be studied if we use that each individual has several observations

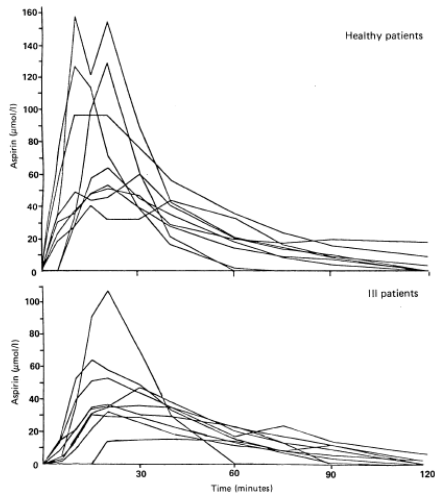
# Take care when using average curves

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- Important structures can be overlooked if you start with the average curves.
- You cannot see the variation over time.
- Always make a plot of the individual observations.
- Only use the average if the patterns are similar, i.e. we only see shifts up and down.

## Individual curves

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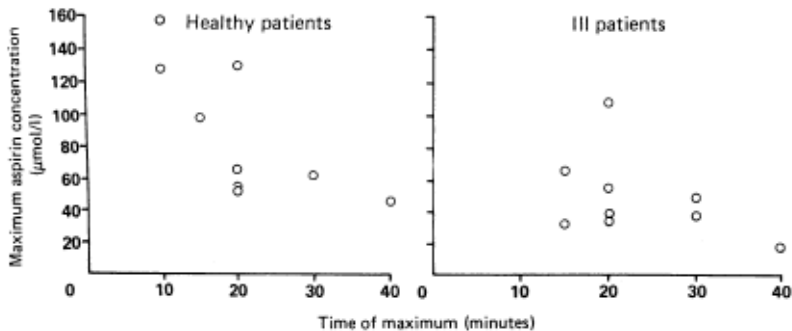
# Analysis of summary statistics

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- Choose a single measure to **summarize** the individual curves
- This again reduces the data set to **independent** observations
- **Popular choices of summary measures** - "relevant" feature extraction:
  - Total increase (last point minus first point)
  - Area under curve (AUC)
  - Maximum or minimum point
  - Average over time
  - Slope in regression with time
  - (or higher order polynomial coefficients)
- Good method with few and easily checked assumptions
- Information may be lost
- Important to choose a **good summary measure**

# Summary Statistics

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## Analysis of aspirin study

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TABLE 1—*Analysis of data from aspirin study*

	Area under curve	Maximum concentration
<i>Healthy patients (n = 9)</i>		
Arithmetic mean (SD) ( $\mu\text{mol/l}$ )	26.5 (8.8)	86.0 (41.5)
Geometric mean ( $\mu\text{mol/l}$ )	25.4	77.8
<i>Ill patients (n = 9)</i>		
Arithmetic mean (SD) ( $\mu\text{mol/l}$ )	17.5 (5.0)	46.7 (26.3)
Geometric mean ( $\mu\text{mol/l}$ )	16.8	41.2
Ratio of geometric means	1.52	1.89
95% Confidence interval	1.11 to 2.08	1.14 to 3.13
<i>t</i> Test	2.83 (df = 16)	2.66 (df = 16)
p Value	0.01	0.02

# Example: Activity of rats

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Summary of experiment:

- 3 treatments: 1, 2, 3 (concentrations)
- 10 cages per treatment
- 10 months
- The response is activity ( $\log(\text{count})$ ) of intersections of light beam during 57 hours)



# The rats data

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```
rats <- read.csv("rats.txt")

# make treatment and cage factors
rats$treatm <- factor(rats$treatm)
rats$cage <- factor(rats$cage)

# make two versions of the time variable
# - one quantitative and one factor
rats$monthQ <- rats$month
rats$month <- factor(rats$month)

summary(rats)
str(rats)
```

In this example we have repeated measurements from each cage.

# Data in two formats, long

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This is the format we want. One response (here lnc) in each line

```
##      treatm cage month      lnc
## 1         1     1     1 9.9323
## 2         1     1     2 9.6447
## 3         1     1     3 9.7628
## 4         1     1     4 9.6014
## 5         1     1     5 9.3227
## 6         1     1     6 9.2463
## 7         1     1     7 9.0739
## 8         1     1     8 9.2077
## 9         1     1     9 9.1670
## 10        1     1    10 8.8319
```

# Data in two formats, wide

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One line for each cage.

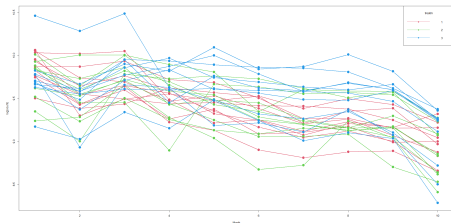
```
##      treatm cage   lnc.1   lnc.2   lnc.3   lnc.4   lnc.5   lnc.6   lnc.7   lnc.8
## 1         1     1  9.9323  9.6447  9.7628  9.6014  9.3227  9.2463  9.0739  9.2077
## 11        1     3 10.0547  9.7384  9.6928  9.4676  9.5297  9.3837  9.2052  9.2582
## 21        1     5  9.7448  9.4278  9.5995  9.2614  9.3744  9.0857  9.1068  9.1732
## 31        1     7  9.8660  9.8687  9.9329  9.7183  9.6861  9.5047  9.2622  9.2699
## 41        1     9  9.9552  9.5536  9.4930  9.4369  9.4520  9.4147  9.4107  9.3751
##      lnc.9 lnc.10
## 1  9.1670  8.8319
## 11 9.0681  8.8753
## 21 8.9907  8.9995
## 31 9.1555  9.1584
## 41 9.2475  9.0435
```

Not usually the format wanted for analysis but adapted to plots and group averages.

# The rats data - plotting the individual patterns

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```
plot(rats$monthQ,rats$lnrc,xlab="Months",
     ylab="log(count)",type="n")
for(i in 1:dim(rats2)[1]){
  lines(1:10,rats2[i,-(1:2)],type="b",col=rats2$treatm[i]+1,
       pch=16,lwd=2,cex=2)
}
legend("topright",legend=1:3,col=2:4,pch=16,lty=2,title="treatm")
```



# The rats data - plotting the average patterns

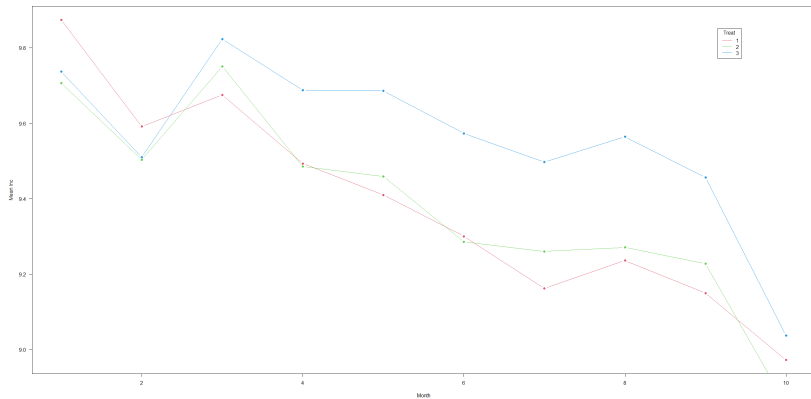
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```
Mean_data <- aggregate(rats$lnrc, by = list(rats$month, rats$treatm),
                        mean)
names(Mean_data) <- c("month", "treatm", "Meanlnrc" )
#Plot the means
Grp1<-subset(Mean_data, treatm==1)
Grp2<-subset(Mean_data, treatm==2)
Grp3<-subset(Mean_data, treatm==3)

plot(as.numeric(Grp1$month), Grp1$Meanlnrc, type = "b", pch=16,
     xlab = "Month", ylab = "Mean lnrc", las = 1, col= 2)
lines(as.numeric(Grp2$month), Grp2$Meanlnrc, type = "b",
     col = 3, pch=16)
lines(as.numeric(Grp3$month), Grp3$Meanlnrc, type = "b",
     col = 4, pch=16)
legend(locator(1), # we will place it with a mouse click
     legend = c("1", "2", "3"),
     title = "Treat", lty = c(2,2,2), col= 2:4)
```

# The rats data - plotting the average patterns

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# Separate analysis for each time-point (continued)

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- Select a fixed time point
- The observations at that time (one from each individual) are independent
- Do a **separate** analysis for the observations at that time
- This is not wrong, but (possibly) a lot of **information is wasted**
- This can be done for several time-points, but
  - Difficult to reach a **coherent** conclusion
  - Sub-tests are not independent
  - Tempting to select time-points supporting our preference
  - Mass significance: If many tests are carried out at 5% level some might be significant by chance. (Bonferroni correction: Use significance level  $0.05/n$  instead of 0.05)



# Separate analysis of rats data

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```
#use the by function to make 10 tests
byMonth <- by(rats, rats$monthQ,
              function(x) anova(lm(lnc ~ treatm, data = x)))
#The largest effect at month 8
byMonth[[8]]

## Analysis of Variance Table
##
## Response: lnc
##           Df Sum Sq Mean Sq F value Pr(>F)
## treatm      2  0.649   0.324    7.29 0.0029 **
## Residuals  27  1.201   0.044
```

# Separate analysis of rats data

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- The model at each month is:

$$\ln c_i = \mu + \alpha(\text{treat}_i) + \varepsilon_i, \quad \varepsilon_i \sim \text{i.i.d. } N(0, \sigma^2), \quad i = 1 \dots 30$$

- The result of the ten tests for no treatment effect:

Month	1	2	3	4	5	6	7	8	9	10
F-value	1.22	0.27	1.02	2.30	3.87	4.10	4.70	7.29	4.09	0.88

Compare with  $F_{95\%;2,27} = 3.35$  or  $F_{99.5\%;2,27} = 6.49$  if Bonferroni correction is used

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# Rats data analyzed via summary measure

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- The log of the total activity in each cage is chosen as summary measure  $\ln\text{Tot} = \log(\text{Total count})$
- The one way ANOVA model becomes:

$$\ln\text{Tot}_i = \mu + \alpha(\text{treatm}_i) + \varepsilon_i, \quad \varepsilon_i \sim \text{i.i.d. } N(0, \sigma^2), \quad i = 1 \dots 30$$

- Notice the simplicity of the model and the relative **few assumptions**

# Rats data analyzed via a summary measure – log total activity

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```
rats$count <- exp(rats$lncl)
DataTot <- aggregate(rats$count, by = list(rats$cage,
                                           rats$treatm), sum)
names(DataTot) <- c("cage", "treatm", "Tot")
head(DataTot)
```

```
##      cage treatm      Tot
## 1      1      1 124864
## 2      3      1 131277
## 3      5      1 110166
## 4      7      1 145418
## 5      9      1 128819
## 6     11      1 136214
```

## log total activity (contd.)

## 09\_Repeated measures

```
reg1 <- lm(log(Tot) ~ treatm, data = DataTot); summary (reg1)
```

```
## Coefficients:
```

```
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 11.73354    0.05409  216.91  <2e-16 ***
## treatm2     -0.00567    0.07650   -0.07   0.941
## treatm3      0.16728    0.07650    2.19   0.038 *
```

```
##
```

```
## Residual standard error: 0.171 on 27 degrees of freedom
```

```
## Multiple R-squared:  0.196, Adjusted R-squared:  0.137
```

```
## F-statistic:  3.3 on 2 and 27 DF,  p-value: 0.0522
```

```
anova(reg1)
```

```
## Analysis of Variance Table
```

```
##
```

```
## Response: log(Tot)
```

```
##              Df Sum Sq Mean Sq F value Pr(>F)
## treatm        2  0.193  0.0965    3.3  0.052 .
```

```
## Residuals    27  0.790  0.0293
```

# Rats data analyzed via a summary measure - slopes

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```
# byCage is a regression with different slope and intercept
# for each cage. Saves the coefficients
byCage <- coef(lm(lnc ~ -1 + cage + monthQ:cage, data = rats))
slope1 <- data.frame(matrix(byCage, nrow=30, byrow=F))

names(slope1) <- c("Intercept", "Slope")

#We also need info about cage and treat
SlopeData <- cbind(DataMean[, 1:2], slope1)
head(SlopeData)
```

##	cage	treatm	Intercept	Slope
## 1	1	1	9.9685	-0.107163
## 2	3	1	10.0411	-0.111593
## 3	5	1	9.6981	-0.076675
## 4	7	1	10.0831	-0.098336
## 5	9	1	9.8067	-0.066990
## 6	11	1	9.7310	-0.043147

# Rats data analyzed via a summary measure - plotting slopes

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```
# Individual slopes:
fit <- unlist(by(rats, rats$cage,
                function(x) fitted.values(lm(lnc ~ monthQ, data=x))))
names(fit) <- NULL

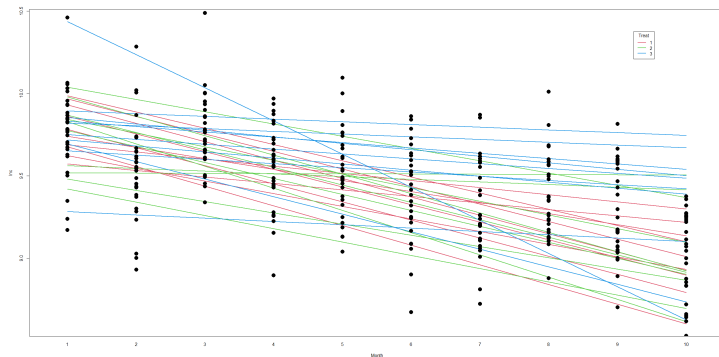
#plotting the linear fit by cage
interaction.plot(rats$monthQ, rats$cage, fit,
                xlab="Month", ylab="lnc", legend=F,
                col=as.numeric(SlopeData$treatm)+1, lty=1, lwd=2)

lines(rats$monthQ, rats$lnc, type="p", pch=16, cex=2)
legend(locator(1), # we will place it with a mouse click
      legend = c("1", "2", "3"), title = "Treat",
      lty = c(1, 1, 1), lwd=2, col= 2:4)
```



# Rats data analyzed via a summary measure - plotting slopes

09\_Repeated measures



# Rats data analyzed via a summary measure - slopes (contd)

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```
reg3 <- lm(Slope ~ treatm, data = SlopeData)
summary(reg3)
```

## Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
## (Intercept)	-0.0879	0.0142	-6.20	1.2e-06 ***
## treatm2	0.0100	0.0200	0.50	0.620
## treatm3	0.0355	0.0200	1.77	0.088 .

##

## Residual standard error: 0.0448 on 27 degrees of freedom

## Multiple R-squared: 0.11, Adjusted R-squared: 0.0439

## F-statistic: 1.67 on 2 and 27 DF, p-value: 0.208

# Rats data analyzed via a summary measure - slopes (contd)

09\_Repeated measures

```
reg3 <- lm(Slope ~ treatm, data = SlopeData)
summary(reg3)

## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  -0.0879     0.0142   -6.20  1.2e-06 ***
## treatm2       0.0100     0.0200    0.50   0.620
## treatm3       0.0355     0.0200    1.77   0.088 .
##
## Residual standard error: 0.0448 on 27 degrees of freedom
## Multiple R-squared:  0.11, Adjusted R-squared:  0.0439
## F-statistic: 1.67 on 2 and 27 DF,  p-value: 0.208
```

- Treat 1 has been chosen as reference with estimated slope -0.09.
- Treat 2 is very similar to Treat 1 with slope  $-0.09 + 0.01 = -0.08$
- Treat 3 has slope closer to 0:  $-0.09 + 0.04 = -0.05$

# Rats data analyzed via a summary measure - slopes (contd)

09\_Repeated measures

```
anova(reg3)

## Analysis of Variance Table
##
## Response: Slope
##           Df Sum Sq Mean Sq F value Pr(>F)
## treatm      2 0.0067  0.00334    1.67   0.21
## Residuals  27 0.0542  0.00201
```

From the one-way analysis of variance we do not see a significant difference in the slopes between the three groups.

# Rats data analyzed via a summary measure - plotting slopes

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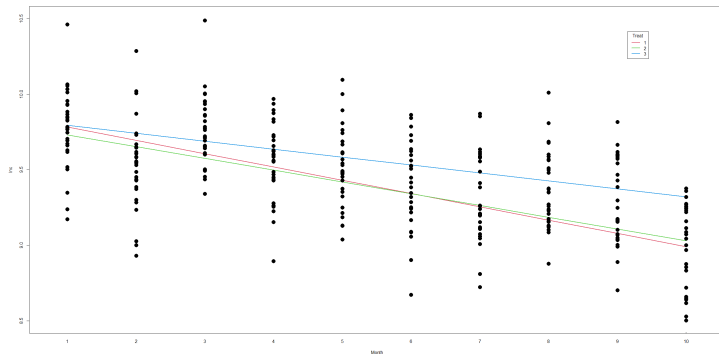
```
# Treatment average slopes:
fit2 <- unlist(by(rats, rats$treatm,
                 function(x) fitted.values(lm(lnc ~ monthQ, data=x))))
names(fit2) <- NULL

#plotting the linear fit by treatment
interaction.plot(rats$monthQ, rats$treatm, fit2,
                 xlab="Month", ylab="lnc", legend=F,
                 col=2:4, lty=1, lwd=2, ylim=c(8.5,10.5))

lines(rats$monthQ, rats$lnc, type="p", pch=16, cex=2)
legend(locator(1),
       legend = c("1","2","3"), title = "Treat",
       lty = c(1,1,1), lwd=2, col= 2:4)
```

# Rats data analyzed via a summary measure - plotting slopes

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# Rats data analyzed via a summary measure - plotting curves

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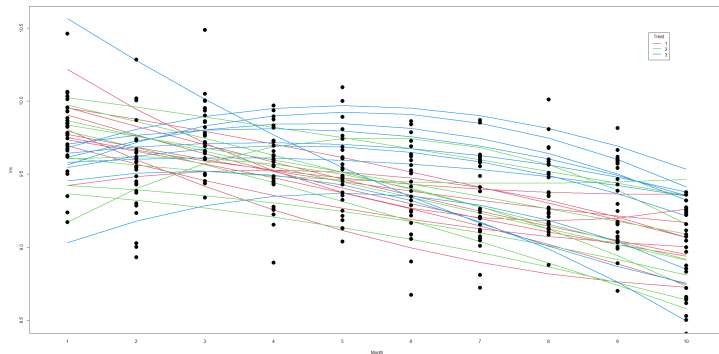
```
# Individual curves: Look similar apart from one of the blue
rats$monthQ2 <- rats$monthQ^2
fit3 <- unlist(by(rats, rats$cage,
  function(x) fitted.values(lm(lnc ~ monthQ+monthQ2, data=x))))
names(fit3) <- NULL
```

```
#plotting the fit by cage
interaction.plot(rats$monthQ, rats$cage, fit3,
  xlab="Month", ylab="lnc", legend=F,
  col=as.numeric(SlopeData$treatm)+1, lty=1, lwd=2,)

lines(rats$monthQ, rats$lnc, type="p", pch=16, cex=2)
legend(locator(1),
  legend = c("1", "2", "3"), title = "Treat",
  lty = c(1, 1, 1), lwd=2, col= 2:4)
```

# Rats data analyzed via a summary measure - plotting curves

09\_Repeated measures





# Rats data analyzed via a summary measure - plotting curves

09\_Repeated measures

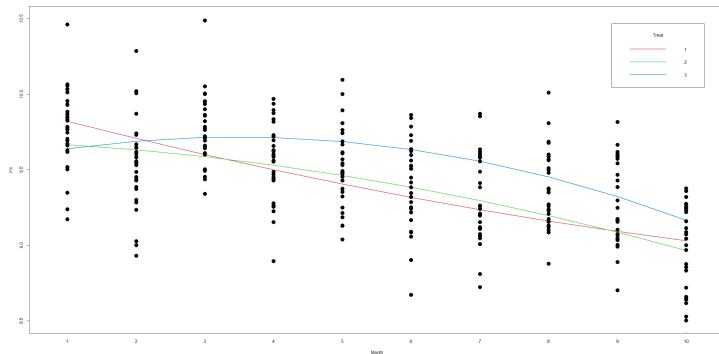
```
# Treatment average curves:
fit4 <- unlist(by(rats, rats$treatm,
  function(x) fitted.values(lm(lnc ~ monthQ+monthQ2, data=x))))
names(fit4) <- NULL

#plotting the fit by treatment
interaction.plot(rats$monthQ, rats$treatm, fit4,
  xlab="Month", ylab="lnc", legend=F,
  col=2:4, lty=1, lwd=2, ylim=c(8.5,10.5))

lines(rats$monthQ, rats$lnc, type="p", pch=16, cex=2)
legend(locator(1),
  legend = c("1", "2", "3"), title = "Treat",
  lty = c(1,1,1), lwd=2, col= 2:4)
```

# Rats data analyzed via a summary measure - plotting curves

09\_Repeated measures



# Rats data analyzed via a summary measure - curvatures

09\_Repeated measures

```
byCage2 <- coef(lm(lnc ~ -1 + cage + monthQ:cage +
                  monthQ2:cage, data = rats))
#I want a data frame, one row for each cage
curve <- data.frame(matrix(byCage2, nrow=30, byrow=F))
names(curve) <- c("Intercept", "monthQ", "monthQ2")
#We also need info about cage and treat
CurveData <- cbind(DataMean[, 1:2], curve)
head(CurveData)
```

##	cage	treatm	Intercept	monthQ	monthQ2
## 1	1	1	10.0465	-0.146171	0.0035462
## 2	3	1	10.0886	-0.135343	0.0021591
## 3	5	1	9.8302	-0.142721	0.0060042
## 4	7	1	10.0237	-0.068641	-0.0026996
## 5	9	1	9.8664	-0.096853	0.0027148
## 6	11	1	9.8445	-0.099881	0.0051576

# Rats data analyzed via a summary measure - curvatures (contd)

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Each estimate of the curve is used as the response

```
reg4 <- lm(monthQ2 ~ treatm, data = CurveData)
summary(reg4)
```

```
## Coefficients:
```

```
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  0.00320    0.00298   1.08  0.29185
## treatm2      -0.00854    0.00421  -2.03  0.05237 .
## treatm3      -0.01608    0.00421  -3.82  0.00071 ***
```

```
##
```

```
## Residual standard error: 0.00941 on 27 degrees of freedom
```

```
## Multiple R-squared:  0.351, Adjusted R-squared:  0.303
```

```
## F-statistic:  7.3 on 2 and 27 DF,  p-value: 0.00291
```

- For Treat 1 the estimated curve is positive (nearly zero 0.003)
- For Treat 2 the estimated curve is negative (0.003-0.009=-0.006)
- For Treat 2 the estimated curve is also negative (0.003-0.016=-0.013)

# Rats data analyzed via a summary measure - curvatures (contd)

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```
anova(reg4)

## Analysis of Variance Table
##
## Response: monthQ2
##           Df Sum Sq Mean Sq F value Pr(>F)
## treatm      2 0.00129  0.000647    7.3 0.0029 **
## Residuals  27 0.00239  0.000089
```

When using the curvature as the response we get a significant difference between the three groups ( $p = 0.003 < 0.05$ )

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# Random effects model - simple version

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- This model uses **all observations** instead of reducing to one observation per individual
- We can test the **time-by-treatment interaction**
- Add “individual” as a random effect
- Makes measurements on same individual correlated
- Unfortunately **equally correlated** no matter if they are “close” or “far apart”
- Can be considered first step in modeling the actual covariance structure
- Usually only good for short series

# Rats data analyzed via random effects approach

09\_Repeated measures

- The model can now be enhanced to:

$$\text{ln}c_i = \mu + \alpha(\text{treat}_i) + \beta(\text{month}_i) + \gamma(\text{treat}_i, \text{month}_i) + d(\text{cage}_i) + \varepsilon_i,$$

- The covariance structure of this model is:

$$\text{cov}(y_{i_1}, y_{i_2}) = \begin{cases} 0 & , \text{ if } \text{cage}_{i_1} \neq \text{cage}_{i_2} \text{ and } i_1 \neq i_2 \\ \sigma_d^2 & , \text{ if } \text{cage}_{i_1} = \text{cage}_{i_2} \text{ and } i_1 \neq i_2 \\ \sigma_d^2 + \sigma^2 & , \text{ if } i_1 = i_2 \end{cases}$$



# Rats data analyzed via random effects approach

09\_Repeated measures

- The model can now be enhanced to:

$$\ln c_i = \mu + \alpha(\text{treat}_i) + \beta(\text{month}_i) + \gamma(\text{treat}_i, \text{month}_i) + d(\text{cage}_i) + \varepsilon_i,$$

- The covariance structure of this model is:

$$\text{cov}(y_{i_1}, y_{i_2}) = \begin{cases} 0 & , \text{ if } \text{cage}_{i_1} \neq \text{cage}_{i_2} \text{ and } i_1 \neq i_2 \\ \sigma_d^2 & , \text{ if } \text{cage}_{i_1} = \text{cage}_{i_2} \text{ and } i_1 \neq i_2 \\ \sigma_d^2 + \sigma^2 & , \text{ if } i_1 = i_2 \end{cases}$$

One can think of the model as:

$\log(\text{count}) = \text{"mean"} + \text{"between cage variation"} + \text{"within cage variation"}$

# Multilevel model structure

09\_Repeated measures

- This model is also called a two-level model.

Level	1	2
Unit	Each separate obs.	Cages
Variation	Within cage $\sigma^2$	Between cage $\sigma_d^2$
Covariates	month month:treatm	treatm

# Model synonyms

## 09\_Repeated measures

- Two-level model
- Mixed model with random subject level
- Mixed model with random intercept
- Model with compound symmetry correlation structure
- Model with exchangeable correlation structure

# Compound symmetry

## 09\_Repeated measures

The model implies that all observations from the same cage are correlated with the intra-class correlation:

$$\text{Corr}(Y_{i_1}, Y_{i_2}) = \frac{\text{cov}(Y_{i_1}, Y_{i_2})}{\sqrt{\text{var}(Y_{i_1})}\sqrt{\text{var}(Y_{i_2})}} = \frac{\sigma_d^2}{\sigma_d^2 + \sigma^2}$$

Here  $\text{cage}_{i_1} = \text{cage}_{i_2}$  and  $i_1 \neq i_2$ .

We are not taking the distance between observations into account.

Maybe too simple? Perhaps observations close in time are more similar than observations far apart.

This correlation structure is called **exchangeable** or **compound symmetry**.

# Potential mistakes when leaving the random effect out

09\_Repeated measures

Level	Unit	Covariates
1	Each separate obs.	month month:treatm
2	Cages	treatm

If a random effect is present:

- Potential bias in the mean (ignoring that observations go together)
- Estimates on **Level 1** may have too much variation. Ignoring pairs → too high p-values. **Effects can be overlooked**
- Estimates on **Level 2** may have too little variation → too small p-values. **“Noise” can become an effect**

# Rats data analyzed via random effects approach in R

09\_Repeated measures

```
library(nlme)
model1 <- lme(lnc ~ month + treatm + month:treatm,
              random = ~1 | cage, data = rats)
anova(model1)
```

	numDF	denDF	F-value	p-value
## (Intercept)	1	243	85525	<.0001
## month	9	243	46	<.0001
## treatm	2	27	3	0.0557
## month:treatm	18	243	2	0.0059

# Rats data analyzed via random effects approach in R

09\_Repeated measures

```
library(nlme)
model1 <- lme(lnc ~ month + treatm + month:treatm,
              random = ~1 | cage, data = rats)
anova(model1)
```

##	numDF	denDF	F-value	p-value
## (Intercept)	1	243	85525	<.0001
## month	9	243	46	<.0001
## treatm	2	27	3	0.0557
## month:treatm	18	243	2	0.0059

So we have a significant interaction between treatment and month.

## Table of fixed effects

## 09\_Repeated measures

```
#This time summary gives a lot of output!
#summary(model1)
```

```
#A table of fixed effects estimates
summary(model1)$tTable
```

##	Value	Std.Error	DF	t-value	p-value
## (Intercept)	9.874280	0.080856	243	122.12192	4.1653e-220
## month2	-0.282870	0.087062	243	-3.24908	1.3213e-03
## month3	-0.199010	0.087062	243	-2.28585	2.3124e-02
## month4	-0.381285	0.087062	243	-4.37948	1.7692e-05
## month5	-0.464289	0.087062	243	-5.33288	2.2101e-07
.....					
## month7:treatm3	0.472180	0.123124	243	3.83500	1.6005e-04
## month8:treatm3	0.465124	0.123124	243	3.77770	1.9913e-04
## month9:treatm3	0.443817	0.123124	243	3.60464	3.7919e-04
## month10:treatm3	0.202159	0.123124	243	1.64192	1.0190e-01



## Table of random effects

09\_Repeated measures

```

#The estimates random effects
VarCorr(model1)

##      cage = pdLogChol(1)
##              Variance StdDev
## (Intercept) 0.027478 0.16577
## Residual    0.037899 0.19468

#intra class correlation
0.027478/(0.027478+0.037899)

## [1] 0.4203

```

$$\hat{\sigma}_d^2 = 0.03 \text{ and } \hat{\sigma}^2 = 0.04$$

# Overview

## 09\_Repeated measures

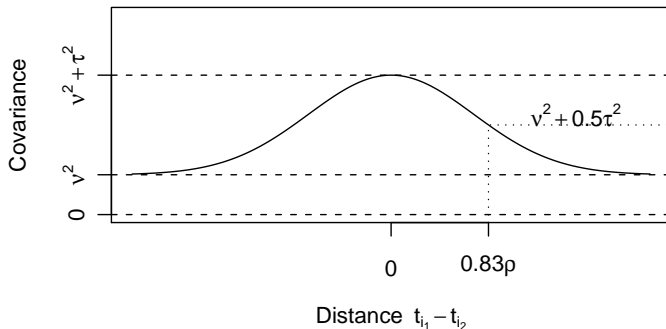
- 1 The repeated measurements set up
  - Aspirin Example
  - Activity of rats
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# Advanced Covariance Structures: Gaussian spatial correlation

09\_Repeated measures

- **Spatial** covariance structures, depending on “how far” observations are apart.
- Gaussian decline of dependency:

$$\text{Cov}(Y_{i_1}, Y_{i_2}) = \begin{cases} 0 & , \text{ if } \text{indiv}_{i_1} \neq \text{indiv}_{i_2} \text{ and } i_1 \neq i_2 \\ \nu^2 + \tau^2 \exp\left\{\frac{-(t_{i_1} - t_{i_2})^2}{\rho^2}\right\} & , \text{ if } \text{indiv}_{i_1} = \text{indiv}_{i_2} \text{ and } i_1 \neq i_2 \\ \nu^2 + \tau^2 + \sigma^2 & , \text{ if } i_1 = i_2 \end{cases}$$



# Rats data via spatial Gaussian correlation model

09\_Repeated measures

- Model for the entire observational vector:

$$\begin{aligned}
 \mathbf{Y} &\sim N(\boldsymbol{\mu}, \mathbf{V}), \quad \text{where} \\
 \mu_i &= \mu + \alpha(\text{treat}_i) + \beta(\text{month}_i) + \gamma(\text{treat}_i, \text{month}_i), \quad \text{and} \\
 V_{i_1, i_2} &= \begin{cases} 0 & , \text{ if } \text{cage}_{i_1} \neq \text{cage}_{i_2} \text{ and } i_1 \neq i_2 \\ \nu^2 + \tau^2 \exp\left\{ \frac{-(\text{month}_{i_1} - \text{month}_{i_2})^2}{\rho^2} \right\} & , \text{ if } \text{cage}_{i_1} = \text{cage}_{i_2} \text{ and } i_1 \neq i_2 \\ \nu^2 + \tau^2 + \sigma^2 & , \text{ if } i_1 = i_2 \end{cases}
 \end{aligned}$$

- R code for model implementation:

```

analysis<-lme(lnc~month+treatm+month:treatm,
random=~1|cage,
correlation=corGaus(form=~as.numeric(month)|cage,nugget=T),
data=rats)

```

## 09\_Repeated measures

- Partial R output:

Random effects:

Formula: ~1 | cage

(Intercept) Residual

StdDev: 0.1404056 ( $= \hat{\nu}$ ) 0.2171559 ( $= \sqrt{\hat{\sigma}^2 + \hat{\tau}^2}$ )

Correlation Structure: Gaussian spatial correlation

Formula: ~as.numeric(month) | cage

Parameter estimate(s):

range nugget

2.3863954 ( $= \hat{\rho}^2$ ) 0.2186744 ( $= \hat{\sigma}^2 / (\hat{\sigma}^2 + \hat{\tau}^2)$ )

Number of Observations: 300

Number of Groups: 30

- Notice the R parametrization of the variance parameters

## Other spatial correlation structures

09\_Repeated measures

- R has a number of build-in correlation structures. A few examples:

correlation=	Name	Correlation term
sp(gau)(t)	Gaussian	$\tau^2 \exp\left\{\frac{-(t_{i_1}-t_{i_2})^2}{\rho^2}\right\}$
sp(exp)(t)	exponential	$\tau^2 \exp\left\{\frac{- t_{i_1}-t_{i_2} }{\rho}\right\}$
ar(1)	autoregressive(1)	$\tau^2 \rho^{ i_1-i_2 }$
un	unstructured	$\tau_{i_1,i_2}^2$

# How to select the correlation structure - Stationarity

09\_Repeated measures

- For all the listed covariance structures, the value of  $Cov(Y_t, Y_{t+u})$  **does not depend on the time  $t$** ; only on the time difference  $u$ .
- We say that the error proces  $\varepsilon_t$  is weakly stationary of order 2;
- weakly because the stationarity is defined from moments, and not distributions (for Gaussian processes this is the same though);
- of order 2 because the stationarity is defined through 2nd order moments (variance, covariance).

# The semi-variogram

## 09\_Repeated measures

- The semi-variogram plots  $\gamma(u) = \frac{1}{2}V(Y_t - Y_{t+u})$ ,  $u > 0$ :

$$\begin{aligned}
 \gamma(u) &= \frac{1}{2}V(Y_t - Y_{t+u}) \\
 &= \frac{1}{2}(V(Y_t) + V(Y_{t+u}) - 2Cov(Y_t, Y_{t+u})) \\
 &= V(Y_t) - Cov(Y_t, Y_{t+u}) \\
 &= \nu^2 + \tau^2 + \sigma^2 - \nu^2 - \tau^2\lambda(u) \\
 &= \sigma^2 + \tau^2(1 - \lambda(u)),
 \end{aligned}$$

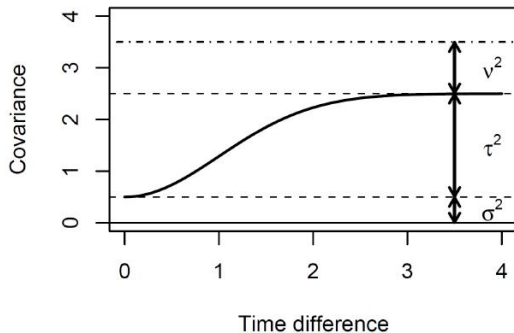
where  $\lambda(u) = \exp(-u^2/\rho^2)$ ,  $\exp(-u/\rho)$ ,  $\rho^u$  for spatial Gaussian, exponential and AR1 correlation structures, respectively.



# The semi-variogram

## 09\_Repeated measures

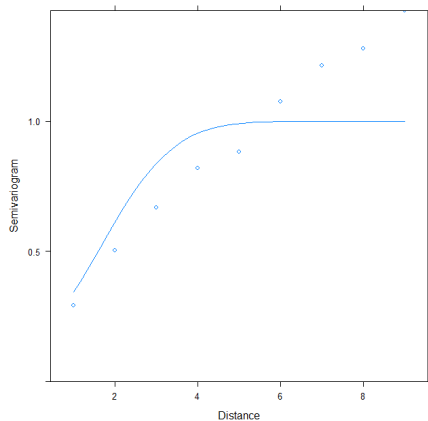
- Theoretical semi-variogram, Gaussian correlation structure:
- Plot of  $\sigma^2 + \tau^2(1 - \lambda(u))$ , where  $\lambda(u) = \exp(-u^2/\rho^2)$ :



# The semi-variogram

09\_Repeated measures

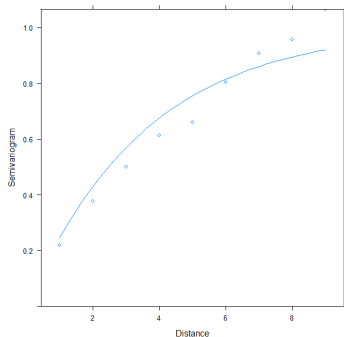
```
plot(Variogram(analysis, form=~monthQ|cage, data=rats))
```



# The semi-variogram, exponentially decreasing correlation

09\_Repeated measures

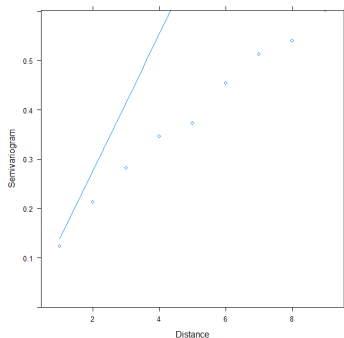
```
analysis2<-lme(lnc~month+treatm+month:treatm, random=~1|cage,
               correlation=corExp(form=~monthQ|cage,nugget=T),
               data=rats)
plot(Variogram(analysis2,form=~monthQ|cage,data=rats))
```



# The semi-variogram, linearly decreasing correlation

09\_Repeated measures

```
analysis3<-lme(lnc~month+treatm+month:treatm, random=~1|cage,
               correlation=corLin(form=~monthQ|cage),
               data=rats)
plot(Variogram(analysis3,form=~monthQ|cage,data=rats))
```



# The semi-variogram, summing up

09\_Repeated measures

- The semi-variogram **compares a theoretical and empirical correlation structure**. Similarity indicates a good model.
- For the *rats data*, the semi-variogram indicates that the optimal correlation structure seems to be an exponentially decreasing spatial correlation structure, rather than the standard spatial Gaussian structure.

# The rats data, partial final model output

09\_Repeated measures

- Random effects:

Formula: ~1 | cage

(Intercept)      Residual

StdDev:    0.07828533 ( $= \hat{\nu}$ ) 0.2510369 ( $= \sqrt{\hat{\sigma}^2 + \hat{\tau}^2}$ )

Correlation Structure: Exponential spatial correlation

Formula: ~as.numeric(month) | cage

Parameter estimate(s):

range                      nugget

3.556503e+00 ( $= \hat{\rho}$ ) 3.341370e-08 ( $= \hat{\sigma}^2 / (\hat{\sigma}^2 + \hat{\tau}^2)$ )

Number of Observations: 300

Number of Groups: 30

# Overview

## 09\_Repeated measures

- 1 The repeated measurements set up
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# Pros and cons of approaches

09\_Repeated measures

## Separate analysis for each time-point

- + Not wrong
- Can be confusing
- Difficult to reach coherent conclusion
- In general not very informative

## Analysis of summary statistic

- + Good method with few and easily checked assumptions
- Important to choose good summary measure(s)

## Random effects approach - simple version

- + Good method for short series
- + Uses all observations
- Usually not good for long series

## Random effects approach - qadvanced version

- + Works for short and long series
- + Uses all observations
- Requires appropriate choice of covariance decay function



# Overview of this module

## 09\_Repeated measures

- 1 The repeated measurements setup
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# Exercise: Histamin in dogs

09\_Repeated measures

In an experiment with 16 dogs the blood histamine concentration was measured 0, 1, 3, and 5 minutes after injection of morphine or trimethaphane. Before injection the dogs were classified into two groups according to their level of histamine (intact or depleted).

```
histamin <- read.table("histamin.txt", header=T, sep="," , dec=".")
histamin$dog <- factor(histamin$dog)
histamin$minQ <- histamin$min
histamin$min <- factor(histamin$min)
histamin$TRT <- factor(paste(histamin$treatm, ":", histamin$level,
                             sep=""))
#summary(histamin)
```

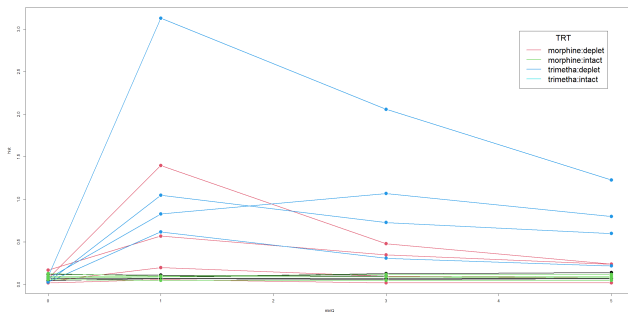
## Histamin in dogs

## 09\_Repeated measures

```
plot(histamin$minQ,histamin$hist,xlab="minQ",ylab="hist",pch="")
for(i in 1:64){
  temp<-histamin[histamin$dog==i,]
  lines(temp$minQ,temp$hist,col=(1:4)[temp$TRT[1]],lwd=2,type="b",
        pch=16,cex=2)
}
legend(locator(1),
       legend = levels(histamin$TRT), title = "TRT",
       lty = c(1,1,1,1),lwd=2,col= 2:5,cex=2)
```

## Histamin in dogs

## 09\_Repeated measures



# Histamin in dogs - questions

09\_Repeated measures

First of all use the TRT factor for the analysis (defining 4 groups of dogs).

- 1 Make some plots of the data, for instance one line for each dog (maybe colored differently in each TRT group).
- 2 Analyze these data using one or more of “the simple methods”.
- 3 How would you approach making a conclusion for the “real” treatment: morphine vs. trimetha?