

## Tutorial for *PinMol* Mac version (contact us by email if you have questions–[icatrina@gmail.com](mailto:icatrina@gmail.com)):

### 1. Generate input file using *mfold*:

<http://unafold.rna.albany.edu/?q=mfold/RNA-Folding-Form>

Paste/upload your transcript sequence in FASTA format, and run an immediate (<800 bases) OR batch job (800-9,000 bases).

The screenshot shows the 'RNA Folding Form' interface. At the top, it says 'M. Zuker' and provides a brief description of the web server. Below this, there's a section for 'Enter sequence name:' with a text box containing 'oskar-RC'. A large text area for the sequence is present, with a sample sequence of 'oskar-RC' pasted in. Below the sequence area are buttons for 'Format Sequence', 'Clear Constraints', and 'Check Constraints'. At the bottom, there's a section for 'Enter constraint information' with a list of four options for specifying constraints.

### 2. Input file: Once the *mfold* job is completed, save the ss-count file as an **ASCII/ANSI** text file using the *Notepad*, *Atom* or *MS Word* software. When clicking the “ss-count ” link (red arrow), a new window opens with the ss-count information.

Select the whole text (Win: CTRL+A; Mac: Command+A), copy/paste it into a text editor, and then save it (e.g. “oskarRC\_sscount\_31str.txt”).

The screenshot shows the 'Output' page. It displays the energy dot plot for 'oskar-RC' and provides links for downloading the results. A red arrow points to the 'View ss-count information' link. Below this, there's a section for 'View Individual Structures' with three structures listed, each with its initial ΔG and file formats.

The screenshot shows the 'ss-count information' page. It displays a list of 31 structures, each with its initial ΔG and file formats. The list is scrollable, showing structures from 2811 to 2849.

If you use *MS Word*, after the text is pasted, use:  
*File>Save As>Simple Text>Save>Other encoding>US ASCII*

3. Run the *PinMol* software from a terminal window and follow the instructions. Each step is described below with the entered text/values highlighted with red boxes.
- A. Run software from command prompt.
  - B. Input the file path for the ss-count file saved at step 2.  
**Note:** Previous files will be overwritten so it is advised to create a new folder if you wish to keep those files.
  - C. Input the length of probe, between 18 and 26 bases (e.g. “22”).  
**Note:** this tutorial shows 14 and 28 as probe length limits, which were used to identify the optimum values of 18 and 26 (see our publication for more details).
  - D. Input target region. If the full length target OR a limited target region is considered, enter the nucleotide number for the first nucleotide (“1” OR the number for the desired start nucleotide) and last nucleotide (maximum possible – for *oskar-RC* mRNA “2,869” OR the desired number for the end nucleotide).  
**Note:** *PinMol* provides the total number of nucleotides of the RNA target.
  - E. Input desired number of probes (e.g. “50” or maximum possible if smaller than 50, which is listed in the output – for this *oskar-RC* example is “616”, which is the default value if a larger number than the maximum possible is entered).
  - F. Select whether a user-provided BLAST analysis should be taken into consideration (e.g. “n”).

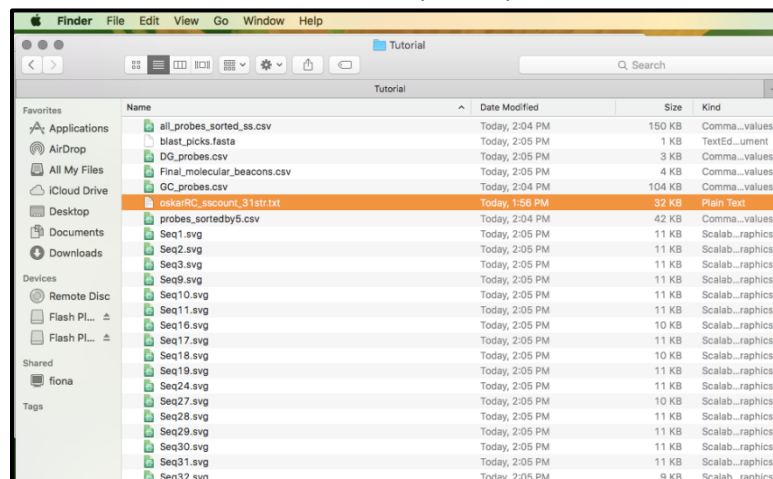
The beginning (green box) and end (blue box) of the output data is presented below.



**4. Analyze results.** The output data is saved in the folder containing the input file along with the following files, which are listed in alphabetical order (the input file is highlighted in orange in the screenshot shown below). We used “probe” to refer to the region of the molecular beacon that is complementary to the target region.

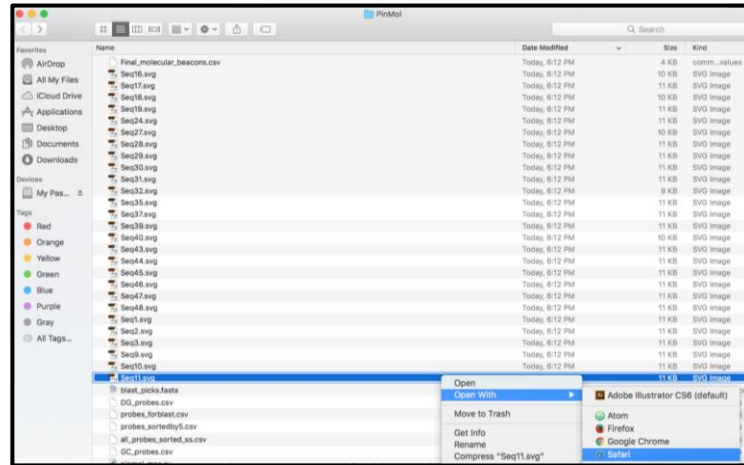
- a. **“all\_probes\_sorted\_ss.csv”** – all possible probe sequences sorted in descending order by ss-count fraction – “1” for fully single stranded and “0” for fully double stranded, as predicted in the input file.
- b. **“blast\_picks.fasta”** – probe sequences that should be used for BLAST analysis (see tutorial file with BLAST analysis).
- c. **“DG\_probes.csv”** – probes that meet both the GC (between 31 and 55%) and the energetic criteria,  $\Delta G_{unimol} > -2.5$  kcal/mol and  $\Delta G_{bimol} > -7.5$  kcal/mol – see point f below and our publication for more information.
- d. **“GC\_probes.csv”** – probes that meet the GC content criteria (between 31 and 55%, also see publication for more information).
- e. **“Final\_molecular\_beacons.csv”** – the output data, as seen in the terminal window.
- f. **“probes\_sortedby5.csv”** – probes sorted by five criteria: in descending order by ss-count fraction,  $\Delta G_{unimol}$ ,  $\Delta G_{bimol}$  and probe’s GC percentage, and in ascending order by  $\Delta G_{duplex}$ . Where the ss-count fraction is described above in point a;  $\Delta G_{unimol}$  is the predicted free energy of uni- or intra-molecular folding of the probe sequence;  $\Delta G_{bimol}$  is the predicted free energy of bi- or inter-molecular folding of the probe sequence;  $\Delta G_{duplex}$  is the predicted free energy of hybridization of the probe sequence with a complementary RNA sequence.
- g. **“Seqi.svg”** – the drawing of the structure of each of the final molecular beacons.

**Note:** some molecular beacons were discarded (e.g. 4 through 8), because they were highly structured or do not fold into a hairpin shape.



## 5. Visualize molecular beacon results using a browser.

Open “Seqi.svg” files to view drawing of the final molecular beacons (**red** circle indicates probe region). Each “Seqi.svg” file corresponds to the “i” sequence listed for all sorted probes (see point 4f above).



The screenshot shows a Microsoft Excel spreadsheet titled 'probs\_samethy1.csv'. The spreadsheet contains a list of protein sequences and their properties. The columns are labeled as follows:

- Base number**: A column of integers ranging from 2125 to 2138.
- NCBI**: A column of NCBI accession numbers, such as 'U00558AAAGCGAAGCAACUUG'.
- Protein name**: A column of protein names, such as 'U00558AAAGCGAAGCAACUUG'.
- Molecular weight**: A column of values representing molecular weight, such as '68.1'.
- pI**: A column of values representing the isoelectric point, such as '3.8'.
- Digestion**: A column of values representing the number of digestion sites, such as '12'.

The data is organized into rows, with some rows highlighted in blue. The spreadsheet is titled 'probs\_samethy1.csv'.

