

#### **BIOGRAPHICAL SKETCH**

NAME	POSITION TITE	POSITION TITLE		
Jingjia Han	Ph.D. Cand	Ph.D. Candidate		
eRA COMMONS USER NAME (credential, e.g., agency login)				
EDUCATION/TRAINING (Begin with baccalaureate or other initia	l professional education,	such as nursing, an	d include postdoctoral training.)	
INSTITUTION AND LOCATION	DEGREE (if applicable)	YEAR(s)	FIELD OF STUDY	
Sichuan University, Chengdu, P.R.China	BSc	07/2004	Biomedical Engineering	
Drexel University, Philadelphia, U.S.A	PhD	07/2010 (expected)	Biomedical Engineering	

#### A. Positions and Honors

### **Positions**

09/2005—Present

Ph.D. Drexel University, School of Biomedical Engineering, Science & Health Systems

# **Other Experiences**

Research Experiences

Sep. 2005-present

Drexel University, School of Biomedical Engineering, Science & Health Systems

Cellular Tissue Engineering Lab

Philadelphia, PA, USA

Co-electrospun PLGA-gelatin-elastin (PGE) scaffold as a non-thrombogenic scaffold for vascular tissue engineering

By systematically adjusting the relative ratio of PLGA and gelatin, I co-electrospinned tertiary blends of poly-(lactide-co-glycolide) (PLGA), gelatin, and elastin (PGE). The morphology and mechanical properties of the blend PGE fibers was characterized by scanning electron microscopy (SEM) and tensile tests (Instron) respectively. The suitability of the composites as vascular scaffolds was assessed in vitro by evaluating their interactions with human endothelial cell line (EA.hy926, ECs), and bovine aortic smooth muscle cells (SMCs) in terms of cell morphology and cellular in-growth. My data indicate that PGE blend fibers supported ECs and SMCs attachment and proliferation with small variances in cell morphology and cytoskeletal spreading for different PGE compositions initially but no differences upon confluence. In addition, histology data demonstrated the unique ability of PGE scaffolds to support EC monolayer formation on the surface while concomitantly fostering penetration of SMC cells into the PGE scaffold. To genetically understand how PGE scaffold interacts with ECs as a prerequisite for a non-thrombogenic endothelium formation, I used a focused PCR array (SA Bioscience/Qiagen) to compare differential genes expression of extracellular matrix (ECM) and adhesion related genes in human aortic endothelial cell (HAEC) monolayer cultured on PGE321 with that on gelatin-coated surface. My results indicate that HAECs seeded on PGE321 and on gelatin shared a similar expression pattern of ECM and adhesion



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molecules and HAEC monolayer has a significant lower E-selectin gene expression on PGE321 upon confluence. Further, the mRNA expression level of tissue factor (TF) on HAEC monolayers cultured on various natural proteins and synthetic biomaterials were compared using qRT-PCR. HAEC monolayers (24hr post-seeding) on PGE321 expressed the same TF mRNA level as on both gelatin and fibronectin (FN) coated surfaces, at both basal level and TNF $\alpha$ -induced level respectively. This demonstrates that PGE321 is a potential non-thrombogenic substrate for endothelium formation thus may be useful scaffold for application in small diameter vascular graft.

#### Aug.2004-Jun.2005

Sichuan University, College of Polymer Science & Engineering

Research Assistant, Biomaterial & Artificial Organ Lab

Chengdu, Sichuan, China

Fabricated nano-hydroxyapatite (HAp) and polylactide (PLA) *in vitro* using wet solution method as a potential artificial biomimetic bone material.

(Co-Project with DIKANG Base of Biomaterials, DIKANG Group, China)

### Sep.2003-Jun.2004

Sichuan University, College of Polymer Science & Engineering

Research Assistant, Biomaterial & Artificial Organ Lab Synthesize high molecular weight polyurethane hydrogeneous control of the control of t

Chengdu, Sichuan, China

Synthesize high molecular weight polyurethane hydrogels with creation of a new type of chain extender: O, O- Bis (2-hydroxy ethyl)-ethylenedicarbamate (EC-DE) in application of wound dressing. A series of polyurethanes (PU) by methods of two-shot process or one-shot process by using EC-DE or Bis (2-hydroxy ethyl) carbamate (EC-AE) as chain extender, 4,4'-diphenylmenthane diisocyanate (HDI) as hard segment and Polyethylene glycol (PEG) or poly(tetrahydrofuran) (PTMG) as soft segment were synthesized and characterized with Fourier Transform Infrared Spectroscopy (FTIR) and Gel Permeation Chromatography (GPC). (Co-Project with Baikang Medical Products Ltd. Conba Group, China)

## Teaching Experiences

Drexel University, School of Biomedical Engineering, Science & Health Systems

01/2006-03/2006, Teaching Assistant, Introduction of Polymers

01/2007-03/2007, Teaching Assistant, Introduction of Polymers

04/2007-06/2007, Teaching Assistant, the Body Synthetic

01/2008-03/2008, Teaching Assistant, Tissue Engineering II

04/2008-06/2008, Teaching Assistant, Biomaterials and Tissue Engineering III

06/2008-09-2008, Teaching Assistant, the Body Synthetic

09/2008-12/2008, Teaching Assistant, Tissue Engineering I

01/2009-03/2009, Teaching Assistant, Tissue Engineering II

04/2009-06/2009, Teaching Assistant, Biomaterials and Tissue Engineering III

09/2009-12/2009, Teaching Assistant, Tissue Engineering I

01/2010-03/2010, Teaching Assistant, Tissue Engineering II



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**Honors** 

2000-2003 Consecutive Scholarship (First-class) from Sichuan University 2000-2003 Consecutive Awards of "Sichuan University Excellent Student"

2004 Award of "Excellent Graduate of Sichuan Province"

2005-2007 Provost Fellowship, Drexel University 2005-present Calhoun Fellowship, Drexel University

# B. Peer-reviewed publications

- 1. Han J, Li M, Chen X, Lelkes PI, Co-electrospun of PLGA, gelatin and elastin blend nanofibers as potential scaffolds for vascular grafts. In preparation.
- 2. Han J, Lelkes PI. Electrospun fibrous scaffold as a non-thrombogenic substrate for endothelium formation. In Preparation.
- 3. Guo Y, Li M, Mylonakis A, Han J, MacDiarmid AG, Chen X, Lelkes PI, Wei Y, Electroactive oligoaniline-containing self-assembled monolayers for tissue engineering applications, *Biomacromolecules*, 2007, 8(10), pp. 3025-34.

## **Book Chapter**

1. Lelkes, PI, Li, M, Perets, A, Lin, L, Han, J, and Woerdeman, DL. (2008) Electrospinning of natural proteins for tissue engineering scaffolding in: Handbook of Natural-based Polymers for Biomedical Applications Rui L.Reis editor), Woodhead Publishing Ltd.

## C. Research Support

Title: Co-electrospun of PLGA, Gelatin and Elastin Blend Nanofibers as Non-thrombogenic Scaffolds for Vascular grafts

This project focuses on testing the feasibility of developing a non-thrombogenic scaffold for small diameter vascular grafts based on electrospun tertiary blend scaffolds.

Sources: a Calhoun Fellowship (JH) and a Provost Fellowship (JH)

Partial Support by:

Title: LCL Grafts (P.I. Lelkes, PI) \$ 100, 000 direct cost

Source: Coulter Foundation, Feasibility Study application, Dates: 04/1/2007-03/31/2008

This grant focuses on testing the feasibility of developing novel small diameter vascular grafts.

#### D. Hobbies

(Additionally, though this is not supported by the NIH biosketch format), I like travelling, cooking, gymnatstics, calligraphy and photography as the coming one...