# 4. R6K-dCas9-sgeGFP-M library

### 4.1 Obtain R6K-dCas9-eGFP-M-1 fragment by PCR

The target fragment is located on the R6K-dCas9-eGFP plasmid, and the amplified target fragment is obtained by PCR. The PCR system and procedure are showed below.

PCR system (50 μL)		
pSC101-dCas9-eGFP	10 ng	
pCas-vec-GG-F=0615	2 μL	
pCas9-vec-GG-R=0615	2 μL	
2 x Mix	25 μL	
DDW	Το 50 μL	

Table 1

The PCR products are detected by agarose gel electrophoresis, and the correct target fragment is 5015bp. We obtained the correct target fragment, and the sample is purified.

### 4.2 Obtain R6K-dCas9-eGFP-M-2 fragment by PCR

The target fragment is located on the R6K-dCas9-eGFP plasmid, and the amplified target fragment is obtained by PCR. The PCR system and procedure are showed below.

PCR system (50 μL)		
pSC101-dCas9-eGFP	10 ng	
2-F	$2~\mu L$	
M2-R	$2~\mu L$	
2 x Mix	25 μL	
DDW	To 50 μL	

Table 2

The PCR products are detected by agarose gel electrophoresis, and the correct target fragment is 4348bp. We obtained the correct target fragment, and the sample is purified.

#### 4.3 Gibson connection

The pSC101-dCas9-eGFP-M-1 fragment and R6K-dCas9-eGFP-M-2 fragment are connected by Goldengate connection method, and the connection system is as follows.

Connection system	(10 µL)
R6K-dCas9-eGFP-M-1	1 μL
R6K-dCas9-eGFP-M-2	1 μL
Cutsmart	1 μL
T4 Buffer	1 μL
BsaI	0.5 μL
T4 ligase	0.2 μL
DDW	3.3 μL

Table 3

## 4.4 Colony PCR

After the petri dish is incubated at 37°C for 12 hours, 48 colonies were selected on the plate. The

colony PCR system and procedure were as follows.

PCR system (10 μL)		
pSC101-dCas9-eGFP-M	1 μL	
link-GG-F=0615	0.2 μL	
link-GG-F=0615	0.2 μL	
2 x Mix	5 μL	
DDW	3.6 μL	

Table 4

The PCR products were detected by agarose gel electrophoresis, and the results were as follows. The correct target fragment is about 1711bp, and The positive rate was approximately 80%. We collectively transferred the positive monoclonal colonies identified through colony PCR into a single LB medium for Extended cultivation, and the plasmids were put forward as a mutation library.