DNA manipulation.

PCR was performed with 2x Taq PCR MasterMix (Solarbio®, Beijing, China). Plasmid DNA was isolated with the TIANprep Mini Plasmid Kit (TIANGEN, Bejing, China). DNA fragments were purifified from agarose gels by using the Universal DNA Purufucation Kit (TIANGEN, Bejing, China) or the Monarch® DNA Gel Extraction Kit (NEB). DNA sequencing and primer synthesis were carried out by Sangon Biotech (Shanghai, China).

Plasmid construction. The plasmids and primers used in this study are listed in the supplemental material.

- 1) To construct pTarget-luxAB, the coding regions of luxAB, p15A, and AmpR are amplified from the genome of Fg-1, a plasmid from 2021OUC-China iGEM and Cloned UpB_4A3m with three different primer pairs 138F/2488R, p15A-R/p15A-F, AmpR-R/AmpR-F, respectively. The J23119 promoter is added to PCR products of luxAB by primers138F and J23119R, resulting in J&luxAB. Oligonucleotides are designed to contribute flanking homologous regions to adjacent DNA fragments of 20-25bp in length, resulting in p15A-homo and AmpR-Homo.
- 2) A new pEvolvR plasmid expressing enCas9-Poll3M-TBD with gRNA targeting luxAB is constructed. Three fragments pEA, pEB and pEC with flanking homologous regions to adjacent DNA fragments of 20-25bp are amplifified from pEvolvR-enCas9-Poll3M-TBD with primer pairs 11615R/8425F, 8450R/2601F, 2627R/11662F, respectively. Double-stranded gRNA is gained after DNA annealing, and it is modified by the promoter with the primer pair——. Similarly, then homologous regions are added to produce J&gRNA-Homo.
- 3) Two plasmids were both constructed using the Seamless cloning Master Mix provided by Sangon Biotech (Shani, China). The plasmids pTarget-luxAB and pEvolvR were cotransformed into E. coli strainDH5 α , following the specification steps provided.

The supplemental material

The supplemental material		
Oligonucleotides used in this study		
138F	ATCTTGGATTACTTTTTGTTCTTTA	
2488R	CAAATTGAAAATGAATCTCTCGAACTT	
p15A-R	CCAGGAAGATACTTAACAGGG	
p15A-F	GGTGCTACATTTGAAGAGATA	
AmpR-R	TGTCCCTAGTGCTTGGATTCT	
AmpR-F	AAATGTGCGCGGAACCCCTAT	
AmpR-R-Homo2	ACCTAGGACTGAGCTGTCAATGTCCCTAGTGCTTGGATTCT	
p15A-R-Homo2	AACAAATAGGGGTTCCGCGCACATTTCCAGGAAGATACTTAACAGGG	
p15A-F-Homo	GGAATTTAAAGAACAAAAAGTAATCCAAGATGgtgctacatttgaagaga	
J23119R	TTGACAGCTAGCTCAGTCCTAGGTATAATGCTAGCtttctcctctttCAAATTGAAAATGAATCTCTCGAACTT	
11615R	CGAAGGTGAGCCAGTGTGAC	
8425F	AGCGTCAGACCCCGTAGAAAA	
8450R	TGATCTTTTCTACGGGGTCTGA	
2601F	ATCAACAGGCTCTCAGACTACGAC	
2627R	CACGTCGTAGTCTGAGAGCCTGT	
11662F	AGCAAGTTAAAATAAGGCTAGTCC	
gRNA protospacers used in this study		
gRNA1		
J&gRNA1-Homo		

Plasmids used in this study

pEvolvR-enCas9-P oli3M-TBD

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