

First task

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Introduction

This work will present data analysis from a research that studied the enhancer's at *IGF2* differential methylation association with abnormal dopamine synthesis in major psychosis (Pai et al., 2019).

Our samples were taken from the prefrontal cortex isolated neurons from patients with schizophrenia and bipolar disorder.

The study analysed data from individuals diagnosed with schizophrenia, bipolar disorder and controls (29, 26 and 27 individuals respectively). In the analysis, study controlled for age, sex, post-mortem interval, genetic ancestry (determined by genotyping the same individuals).

Experiment design

The experiment design was multi-omics study with 55 cases (with schizophrenia or bipolar disorder) and 27 controls.

Objective of the research

According to authors, schizophrenia and bipolar disorder have got characteristic of periods of psychosis. The main objective of the research was to gather epigenomic profiling data to get a more accurate model of neuronal dysregulation in diseases with periods of psychosis.

Biological targets of the research

Researchers intended to look for specific patterns of DNA methylation in isolated neurons from the frontal cortex of individuals that had diseases.

- IGF2 - insulin growth factor 2 protein
- *IGF2* - IGF2 gene
- *Igf2* - enhancer of *IGF2*
- TH - tyrosine hydroxylase protein
- dopamine - a neuromodulatory molecule
- psychosis - an abnormal condition of the mind that results in difficulties determining what is real and what is not real

Results received

Authors found a strong association between methylation of *Igf2* and TH synthesis. TH is the bottleneck enzyme that is responsible for dopamine synthesis. If enhancer *Igf2* is hypomethylated, levels of TH are higher, which determines the higher production of dopamine. Apparently, dopamine is responsible for psychosis in the mental disorders of interest.

Additional information

Schizophrenia and bipolar disorder patients are consistently hypomethylated at *IGF2* locus when compared to controls. This locus remained significantly hypomethylated even after accounting lifestyle-related variables of smoking and anti-psychotic use.

The reaction chain of interest of the research (upward arrows show elevated expression or synthesis of the protein, product, or effect):

Hypomethylation of *Igf2* \rightarrow \uparrow IGF2 \rightarrow \uparrow TH \rightarrow \uparrow dopamine \rightarrow \uparrow psychosis

Data preparation

Sample keys heading is made of the following columns names:

- *id* - an identifier of the sample
- *sentrrix_id* - Illumina's Sentrix BeadChip identifier (13 unique values) (National Institutes of Health, n.d.)
- *sentrrix_row* - row number in the Sentrix array
- *sentrrix_col* - column number in the Sentrix array
- *basename* - sample identifier in the research (joined values in a format: `[id]_[sentrrix_id]_R0[sentrrix_row]C0[sentrrix_id]`)
- *tissue_bank_id* - an identifying number of the tissue bank from which the sample was taken
- *tissue_bank* - the literal identifier of the tissue bank
- *tissue* - a tissue type from which the sample was taken
- *cell_type* - a cell type found in the sample
- *donor* - an integer number that identifies the donor of the sample (82 unique values)
- *pmi* - a post-mortem interval, unknown values were labeled as NA
- *race* - race of the donor (white, black, hispanic, or unknown (NA))
- *sex* - gender of the donor
- *diagnosis* - an experimental group of the donor (bipolar, schizophrenia, or control)
- *age* - age of the donor (years)

As it was noted in the article, there were 100 records in the sample keys dataset.

Calculating detection p-values

Getting detection p-value for each score of DNA modification. These p-values determine whether the measured intensity can be distinguished from the background.

All values that have got p-value higher than 0.01 are considered as bad and all samples that have more than 1% of bad detection p-values should be removed.

Although, in our data, none of the samples had more than 1% of bad values, therefore no sample was removed.

Predicting sample sex

This stage estimates sample sex based on methylation data.

Number of females and males after estimation matched original data (25 female and 75 male).

Converted ‘M’ and ‘F’ notation to ‘male’ and ‘female’.

No mismatches between real and estimated sex were found.

Data normalisation

According to the documentation of *minfi* package (Fortin & Hansen, n.d.), *preprocessFunnorm()* function is recommended for known large-scale differences (for example, cancer/normal) or between-tissue studies. Our chosen data spans only over one cell type of one tissue, therefore it was decided to opt for different normalisation methods.

Authors (Pai et al., 2019) noted that they used noob normalisation followed by the quantile one. Quantile normalisation performs processing of Type I and Type II array design differences. Whereas, *preprocessIllumina()* normalisation has only background subtraction and control normalisation implemented. Therefore, we decided to choose *preprocessSWAN()* normalisation, since this method performs within-array normalisation correction for technical differences between Type I and Type II array designs.

Filtering position data by detection p-values

There were 5835 positions found that had p-value higher than 0.01 in 1% of the samples. These positions were removed from the dataset. After this procedure, we have 861001 positions in each sample.

Removing methylation loci positions

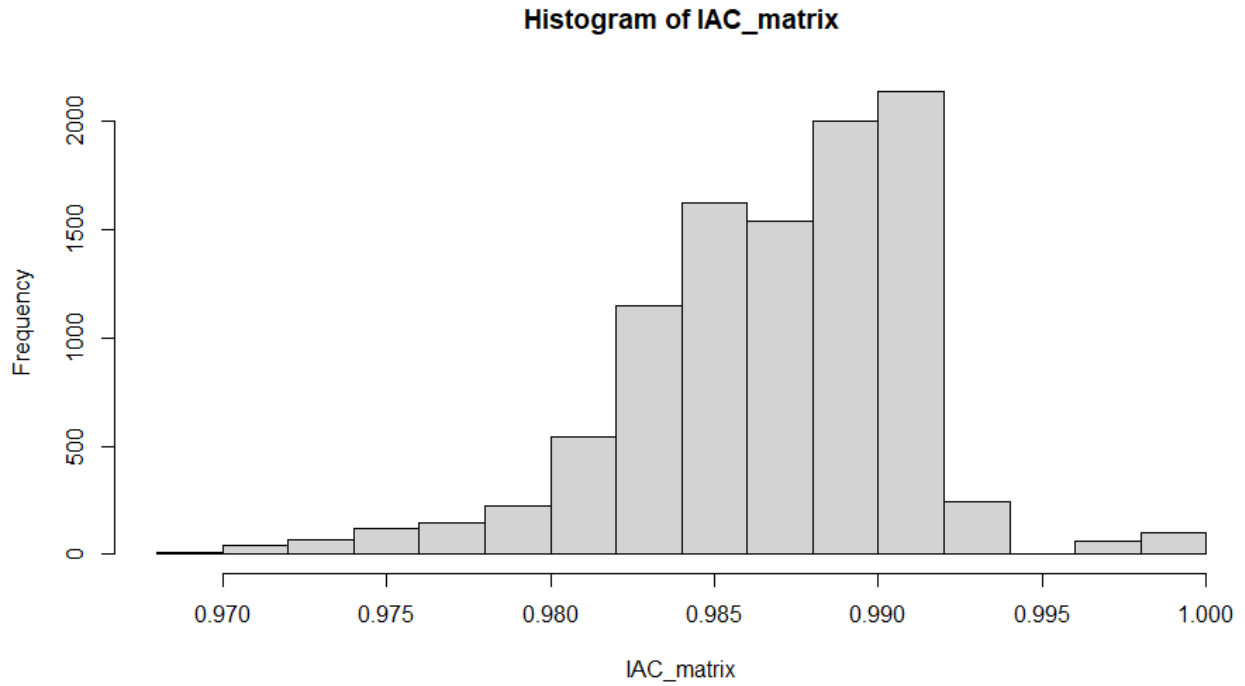
Removed 2918 methylation loci, that do not contain “CG” nucleotide pair (CH probes) or are close to DNR polymorphisms. After removal data contains 858083 positions in each sample.

Making three different data objects

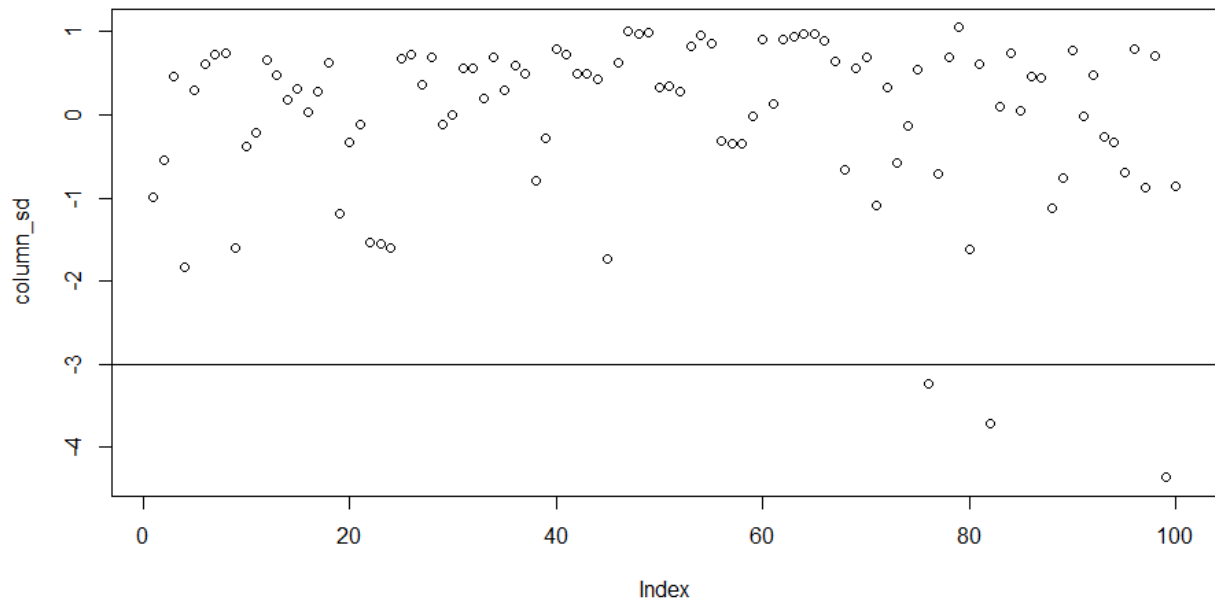
Generated DNA modification score matrix, information about main matrix samples and information about main matrix positions and saved this data into files for later manipulations.

Interarray correlation outliers elimination

Identification and removal of samples with divergent modification scores.

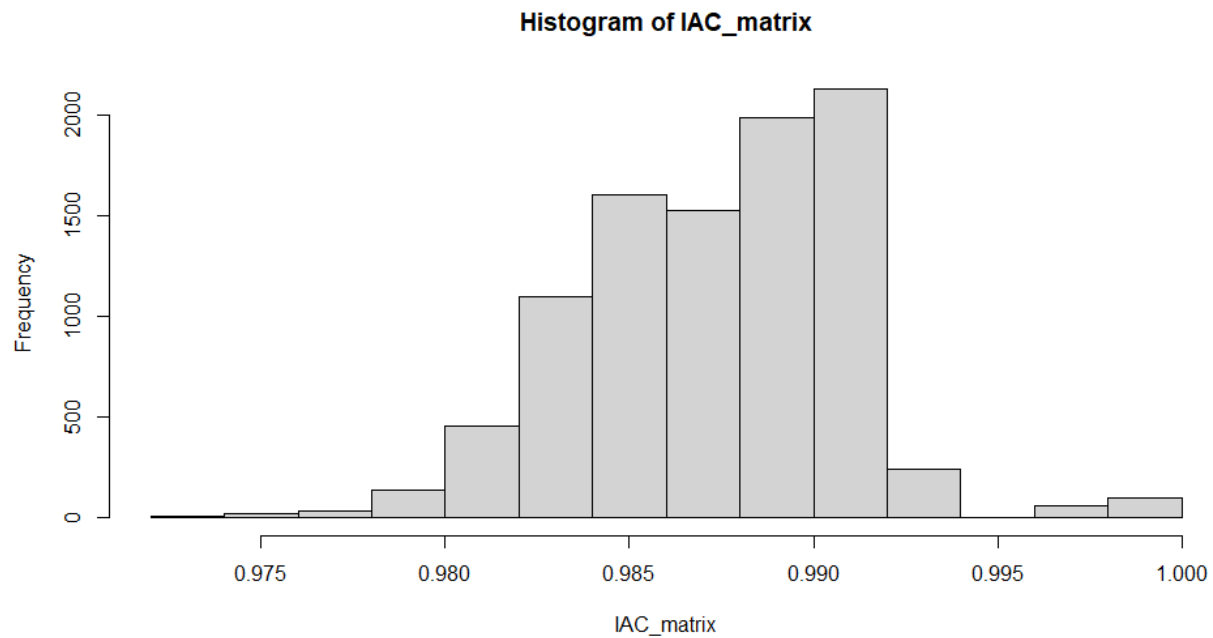


Histogram identifies that our dataset contains values which distort the overall distribution. For further investigation, standard deviation from mean in each sample was calculated.

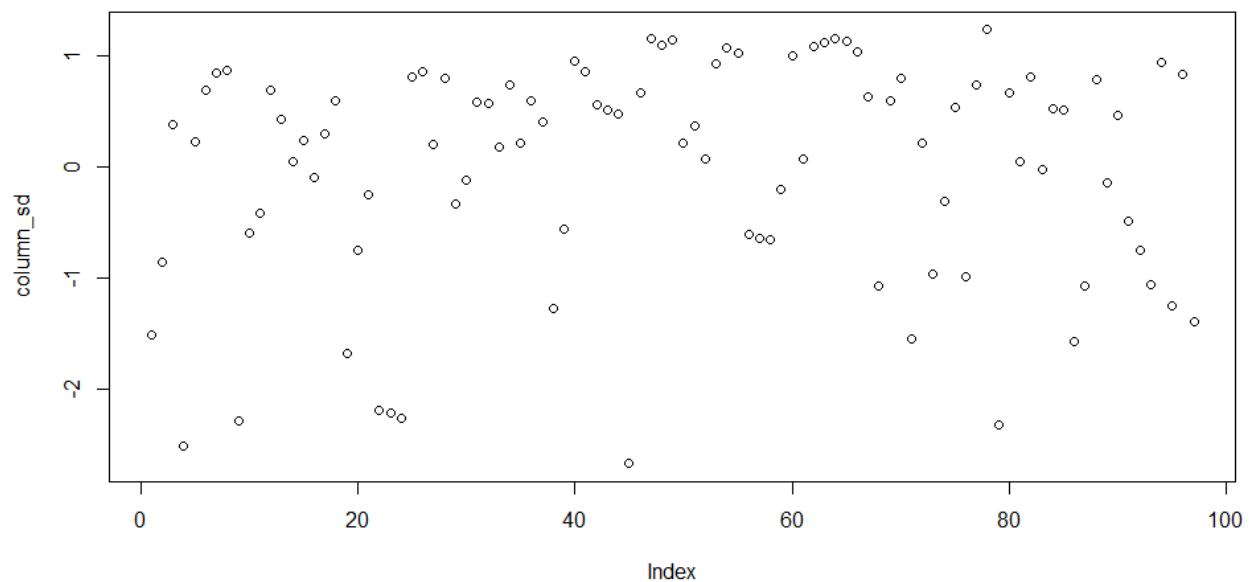


Scatter plot of each sample (column) standard deviation from mean visually highlights data outliers (under -3 limit).

Algorithm identified and removed 3 outliers: “GSM3059462_200590490031_R08C01”
 “GSM3059520_200357150067_R08C01” “GSM3059454_200590490031_R01C01”.



There is a visible difference on the left side of the histogram. This change indicated that we correctly removed distorting values.

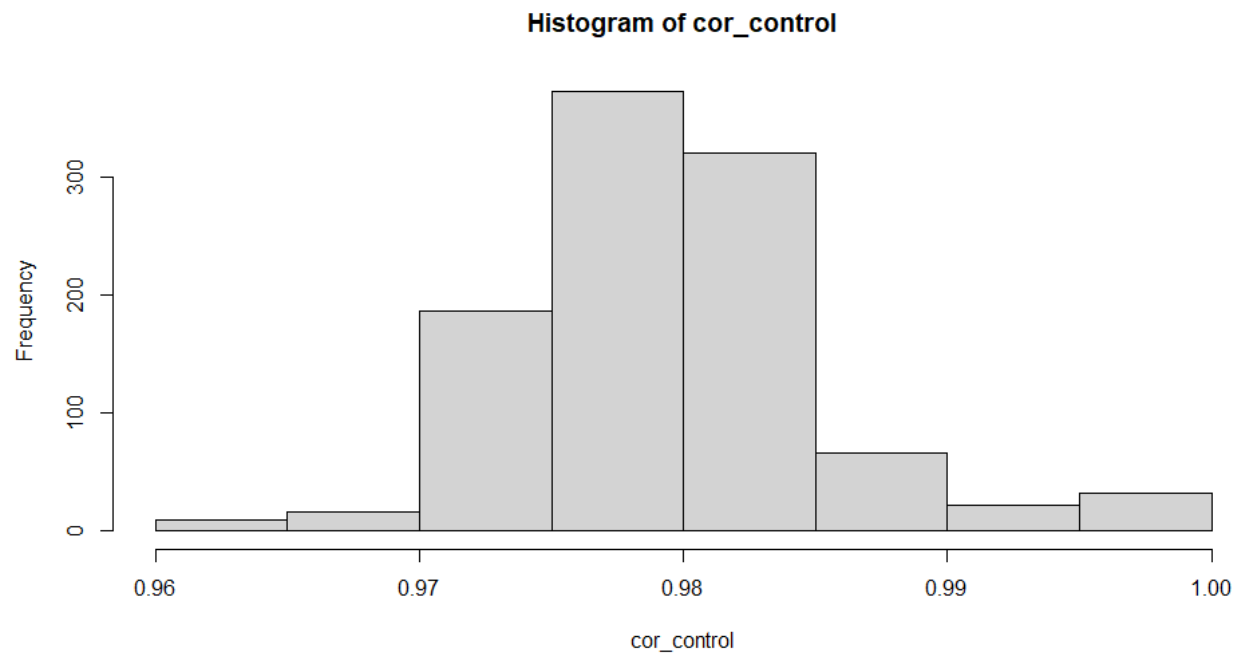


No outliers are left in recalculated scatter plot.

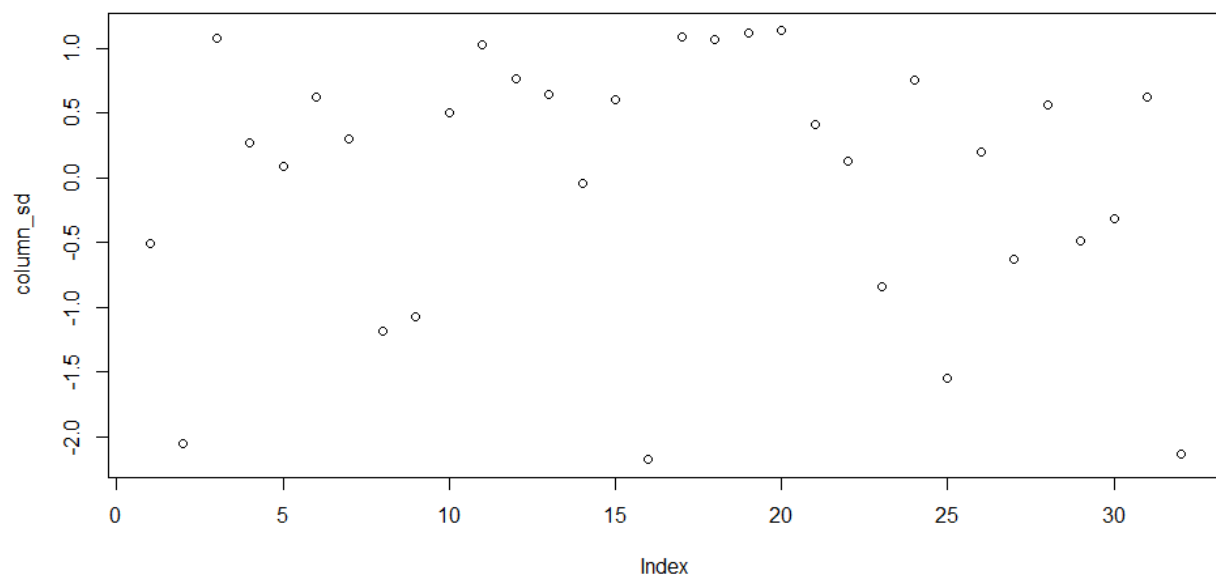
Quality control

After all data manipulations, our set has 97 samples with 861001 positions.

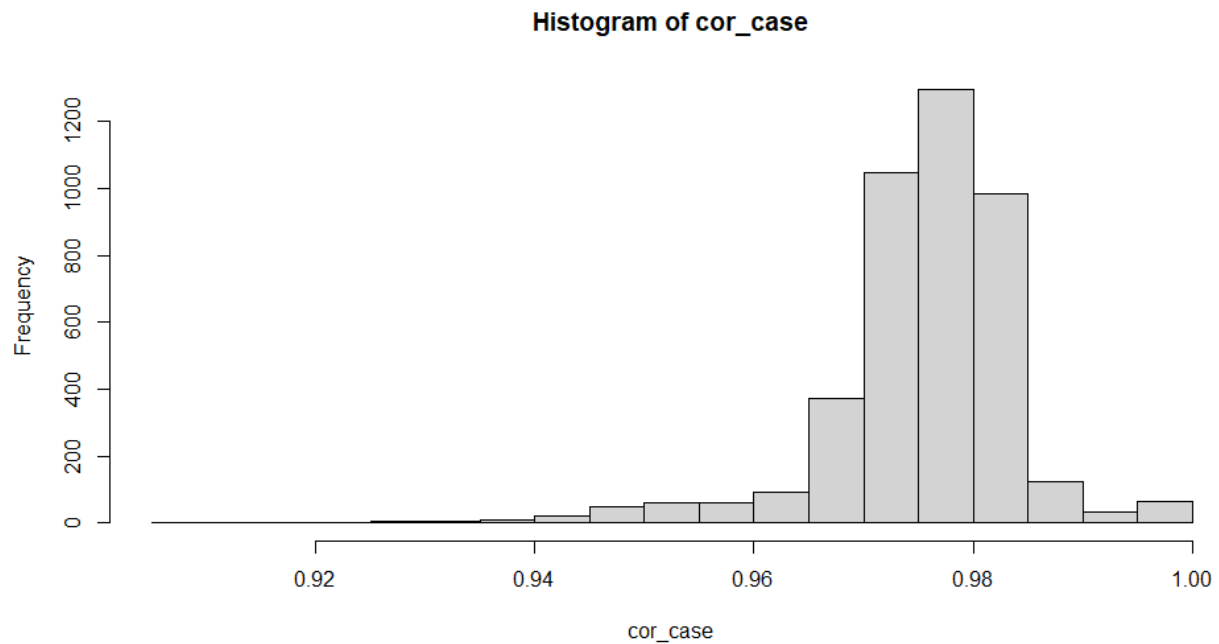
Data for quality control was separated into case (65) and control (32). Our main goal is to check if distortions in methylation data exist.



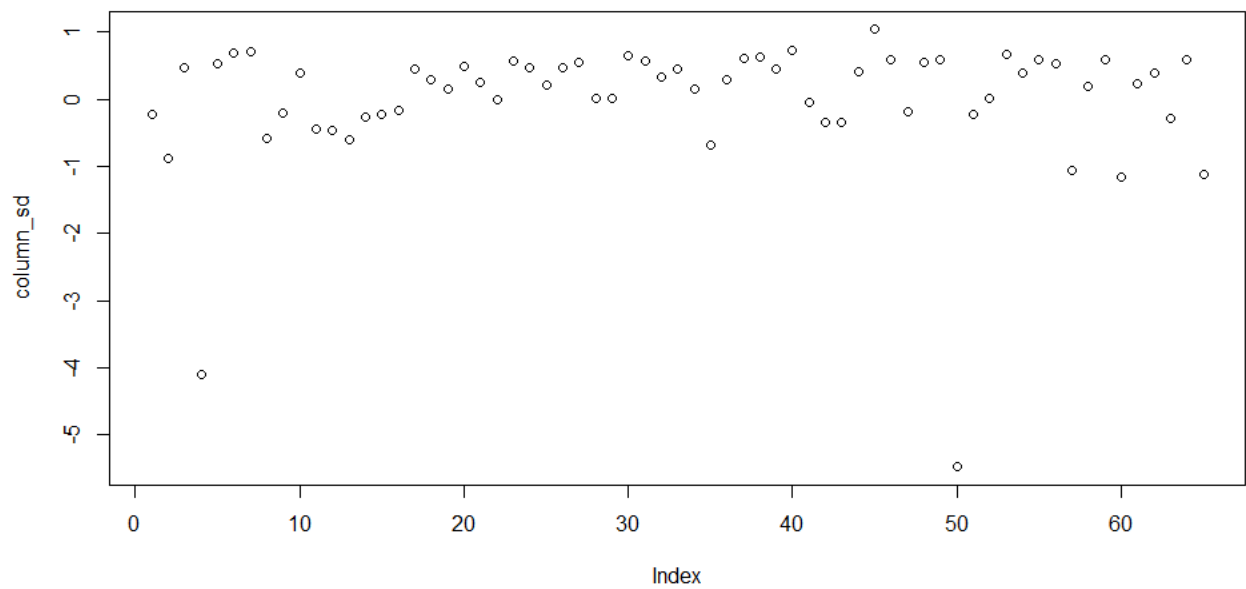
Histogram represents pair-to-pair correlation in control methylation data.



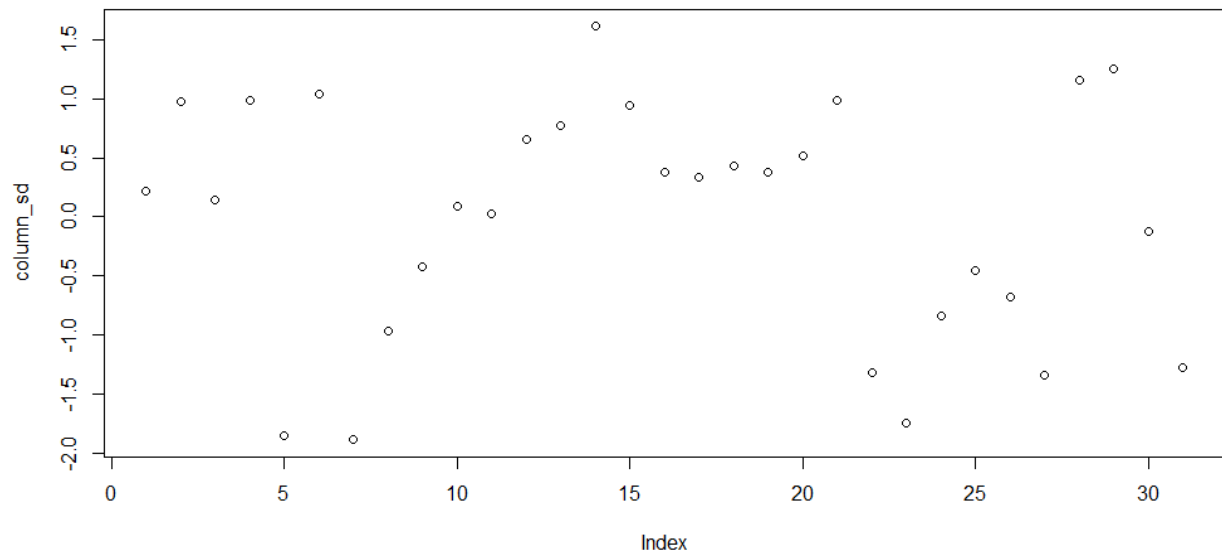
We can indicate both from histogram and scatter plot that data is distributed normally and there is no need for data removal.



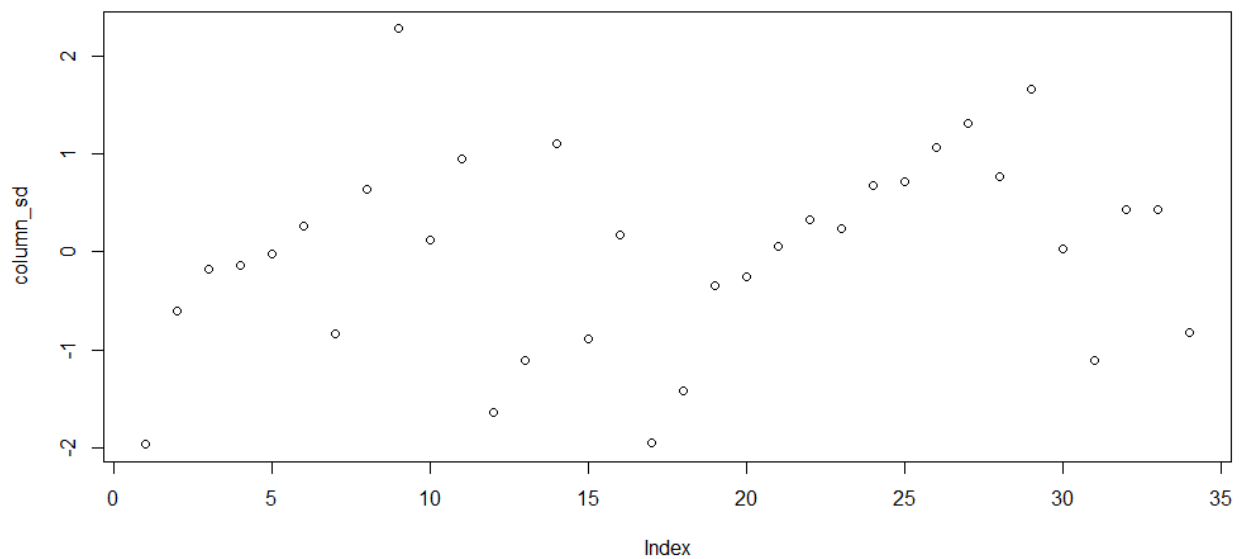
Case methylation data indicates, that our data has distorted values.



Scatter plot demonstrates two outliers in standard deviation from methylation mean. For further analysis we separated case data into “bipolar” and “schizophrenia” cases.



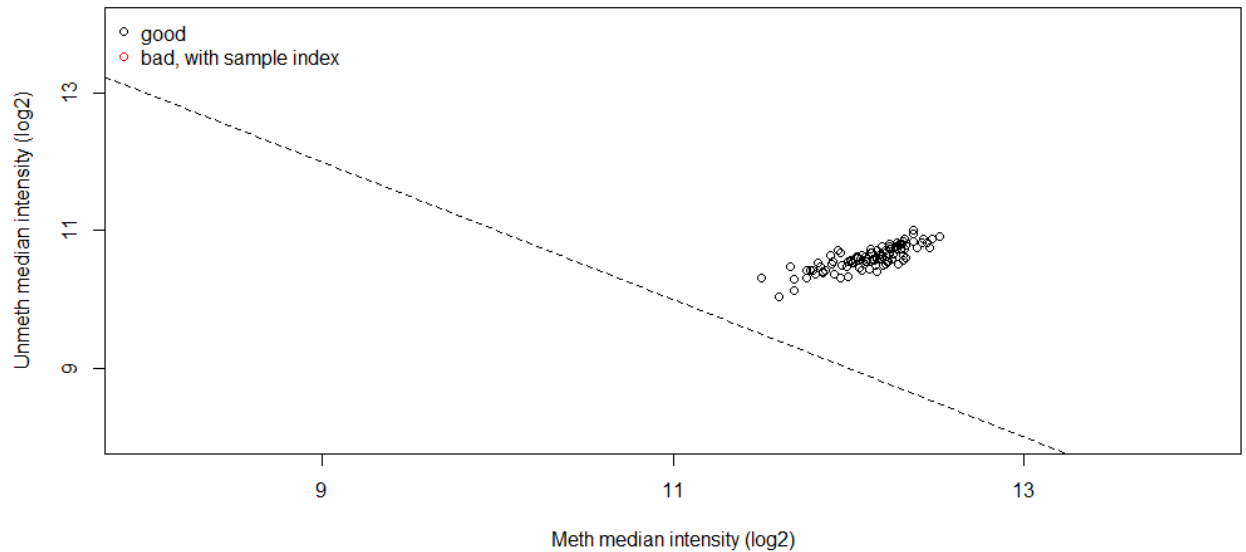
Bipolar cases does not show any big fluctuations from methylation mean value.



Schizophrenia cases also does not show any wide variations from methylation mean value.

This separated each case data indicated that there is no need to remove any data.

Estimated sample-specific quality control for methylation data with getQC, addQC and plotQC functions.



PlotQC demonstrates, that bad samples does not exist in our data set.

Saving data

Saved data after all manipulations into *GSE112179_clear.rds* file.

References

- Fortin, J.-P., & Hansen, K. D. (n.d.). Analysis of 450k data using minfi. *Dim*, 485512, 6.
- National Institutes of Health, N. C. I. at the. (n.d.). *Sentrix® BeadChip and BeadArray technology (illumina, inc.) / innovative molecular analysis technologies (IMAT)*. <https://imat.cancer.gov/about-imat/outputs-and-achievements/individual-technologies-and-platforms/sentrix%C2%AE-beadchip-and>
- Pai, S., Li, P., Killinger, B., Marshall, L., Jia, P., Liao, J., Petronis, A., Szabó, P. E., & Labrie, V. (2019). Differential methylation of enhancer at IGF2 is associated with abnormal dopamine synthesis in major psychosis. *Nature Communications*, 10(1), 1–12.