First task

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Introduction

This work will present data analysis from a research that studied the enchancer's at *IGF2* differential methylation association with abnormal dopamine synthesis in major psychosis (Pai et al., 2019).

Our samples were taken from the prefrontal cortex isolated neurons in schizophrenia and bipolar disorder.

Study analysed data from individuals diagnosed with schizophrenia, bipolar disorder and controls (29, 26 and 27 individuals, respectively). In the analysis study controlled for age, sex, post-mortem interval, genetic ancestry (determined by genotyping the same individuals).

Experiment design

The experiment design was multi-omics study with 55 cases (with schizophrenia or bipolar disorder) and 27 controls.

Objective of the research

According to authors, schizophrenia and bipolar disorder have got characteristic of periods of psychosis. The main objective of the research was to gather epigenomic profiling data to get a more accurate model of neuronal dysregulation in diseases with periods of psychosis.

Biological targets of the research

Researchers intended to look for specific patters of DNA methylation in isolated neurons from the frontal cortex of individuals that had diseases.

- IGF2 insulin growth factor 2 protein
- IGF2 IGF2 gene
- Igf2 enhancer of IGF2
- TH tyrosine hydroxylase protein
- dopamine a neuromodulatory molecule
- psychosis an abnormal condition of the mind that results in difficulties determining what is real and what is not real

Results received

Authors found a strong association between methylation of Igf2 and TH synthesis. TH is the bottleneck enzyme that is responsible for dopamine synthesis. If enhancer Igf2 is hypomethylated, levels of TH are higher, which determines the higher production of dopamine. Apparently, dopamine is responsible for psychosis in the mental disorders of interest.

Additional information

Schizophrenia and bipolar disorder patients are consistently hypomethylated at *IGF2* locus when compared to controls. This locus remained significantly hypomethylated even after accounting lifestyle-related variables of smoking and anti-psychotic use.

The reaction chain of interest of the research (upward arrows show elevated expression or synthesis of the protein, product, or effect):

Hypomethylation of $Igf2 \rightarrow \uparrow IGF2 \rightarrow \uparrow TH \rightarrow \uparrow dopamine \rightarrow \uparrow psychosis$

Data preparation

```
BiocManager::install("IlluminaHumanMethylationEPICanno.ilm10b4.hg19")
library(IlluminaHumanMethylationEPICanno.ilm10b4.hg19)
```

Sample keys heading is made of the following columns names:

- *id* an identifier of the sample
- sentrix_id Illumina's Sentrix BeadChip identifier (13 unique values) (National Institutes of Health, n.d.)
- sentrix row row number in the Sentrix array
- sentrix col column number in the Sentrix array
- basename sample identifier in the research (joined values in a format: [id]_[sentrix_id]_R0[sentrix_row]C0[sentrix_id])
- tissue bank id an identifying number of the tissue bank from which the sample was taken
- tissue bank the literal identifier of the tissue bank
- tissue a tissue type from which the sample was taken
- cell type a cell type found in the sample
- donor an integer number that identifies the donor of the sample (82 unique values)
- pmi a post-mortem interval (time units?????????), unknown values were labeled as NA
- race race of the donor (white, black, hispanic, or unknown (NA))
- sex gender of the donor
- diagnosis an experimental group of the donor (bipolar, schizophrenia, or control)
- age age of the donor (years)

As it was noted in the article, there were 100 records in the sample keys dataset.

Calculating detection p-values

Getting detection p-value for each score of DNA modification. These p-values determine whether the measured intensity can be distinguished from the background.

All values that have got p-value higher than 0.01 are considered as bad and all samples that have more than 1% of bad detection p-values should be removed.

Although, in our data, none of the samples had more than 1% of bad values, therefore no sample was removed.

Predicting sample sex

This stage estimates sample sex based on methylation data.

Number of females and males after estimation matched original data (25 female and 75 male).

Converted 'M' and 'F' notation to 'male' and 'female'.

No mismatches between real and estimated sex were found.

Data normalisation

According to the documentation of *minfi* package (Fortin & Hansen, n.d.), *preprocessFunnnorm()* function is recommended for known large-scale differences (for example, cancer/normal) or between-tissue studies. Our chosen data spans only over one cell type of one tissue, therefore it was decided to opt for different normalisation methods.

Authors (Pai et al., 2019) noted that they used noob normalisation followed by the quantile one. Quantile normalisation performs processing of Type I and Type II array design differences. Whereas, preprocessIllumina() normalisation has only background subtraction and control normalisation implemented. Therefore, we decided to choose preprocessSWAN() normalisation, since this method performs within-array normalisation correction for technical differences between Type I and Type II array designs.

Filtering position data by detection p-values

There were 5835 positions found that had p-value higher than 0.01 in 1% of the samples. These positions were removed from the dataset. After this procedure, we have 861001 positions in each sample.

Removing methylation loci positions

Removed 2918 methylation loci, that do not contain "CG" nucleotide pair (CH probes) or are close to DNR polymorphisms. After removal data contains 858083 positions in each sample.

Making three different data objects

Generated DNA modification score matrix, information about main matrix samples and information about main matrix positions and saved this data into files for later manipulations.

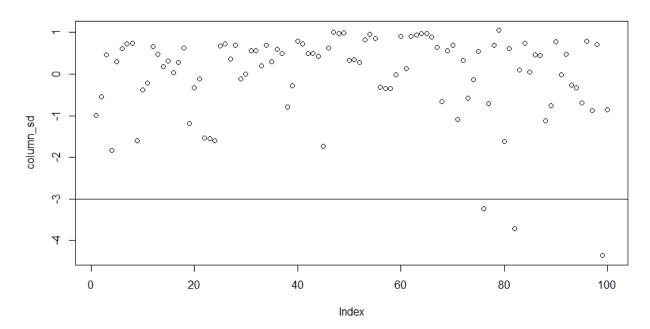
Histogram of IAC matrix

Interarray correlation outliers elimination

Identification and removal of samples with divergent modification scores.

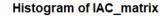
Histogram identifies, that in our data set exists distribution distorting values. For further investigation

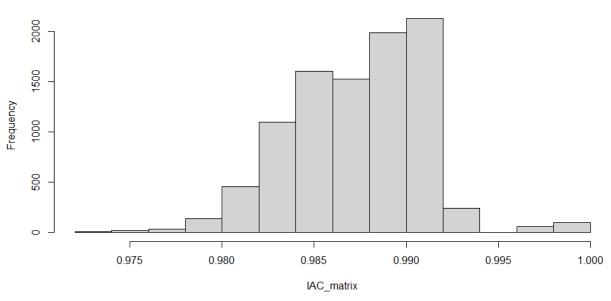
standard deviation from mean in each sample was calculated.



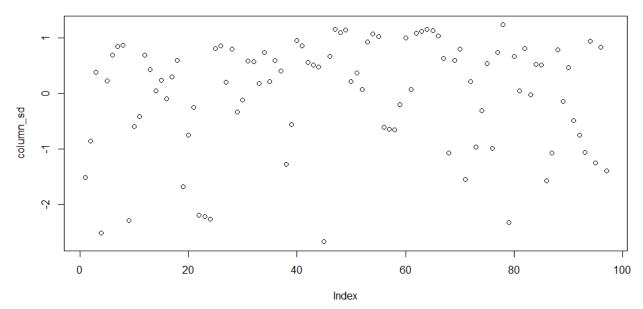
Scatter plot of each sample (column) standard deviation from mean visually highlights data outliers (under -3 limit).

Algorithm identified and removed 3 outliers: " $GSM3059462_200590490031_R08C01$ " " $GSM3059520_200357150067_R08C01$ " " $GSM3059454_200590490031_R01C01$ ".





There is visible difference on the left side of the histogram. This change indicated that we correctly removed distorting values.

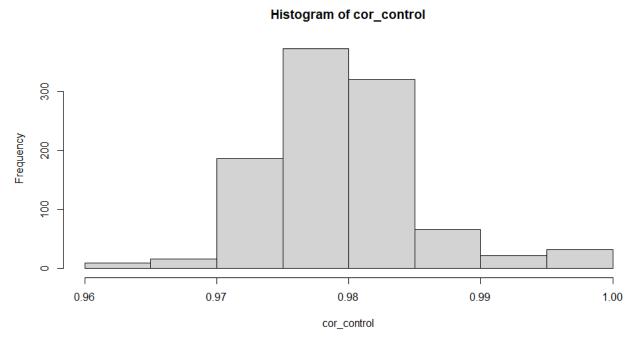


No outliers are left in recalculated scatter plot.

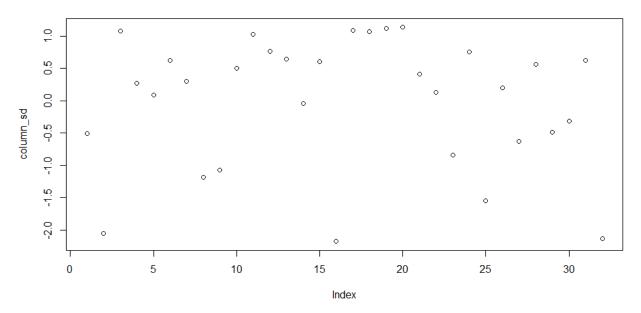
Quality control

After all data manipulations, our set has 97 samples with 861001 positions.

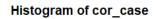
Data for quality control was separated into case (65) and control (32). Our main goal is to check if there exists distortions in methylation data.

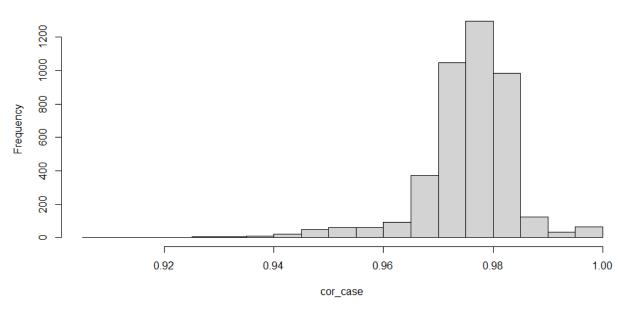


Histogram represents pair-to-pair correlation in control methylation data.

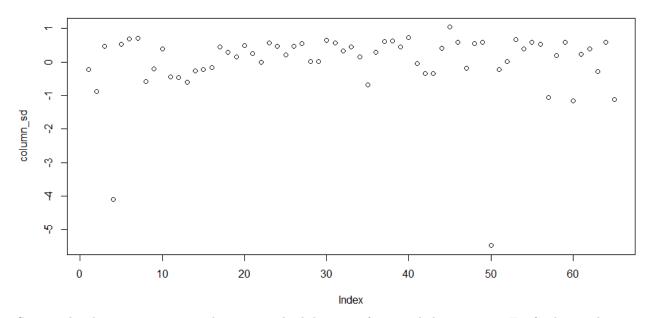


We can indicate both from histogram and scatter plot, that data is distributed mostly normal and there is no need for data removal.

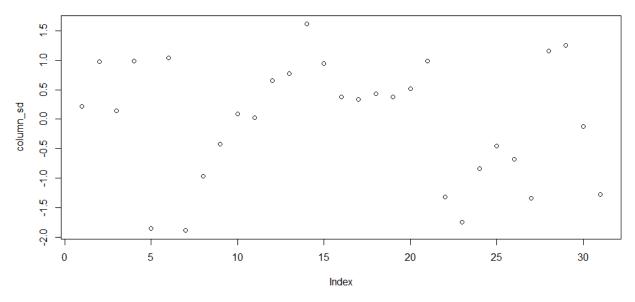




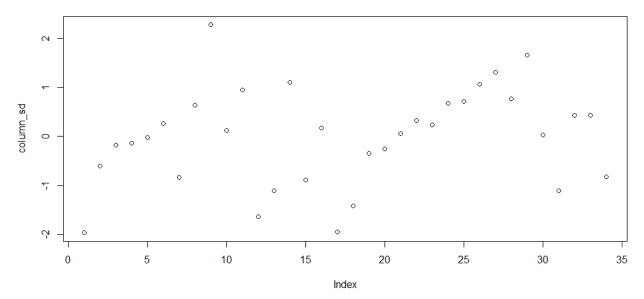
Case methylation data indicates, that our data has distorted values.



Scatter plot demonstrates two outliers in standard deviation from methylation mean. For further analysis we separated case data into "bipolar" and "schizophrenia" cases.



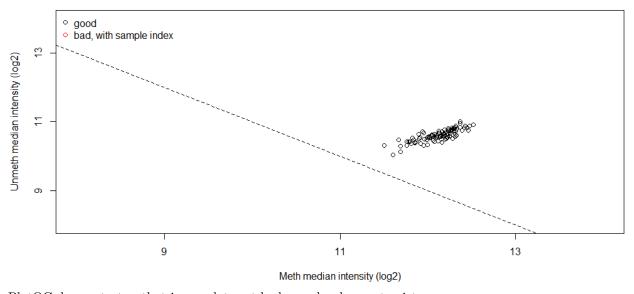
Bipolar cases does not show any big fluctuations from methylation mean value.



Schizophrenia cases also does not show any wide variations from methylation mean value.

This separated each case data indicated, that there is no need to remove any data.

Estimated sample-specific quality control for methylation data with getQC, addQC and plotQC functions.



PlotQC demonstrates, that in our data set bad samples does not exist.

Saving data

Saved data after all manipulations into GSE112179_clear.rds file.

References

Fortin, J.-P., & Hansen, K. D. (n.d.). Analysis of 450k data using minfi. Dim, 485512, 6.

- National Institutes of Health, N. C. I. at the. (n.d.). Sentrix® BeadChip and BeadArray technology (illumina, inc.) / innovative molecular analysis technologies (IMAT). https://imat.cancer.gov/about-imat/outputs-and-achievements/individual-technologies-and-platforms/sentrix%C2%AE-beadchip-and
- Pai, S., Li, P., Killinger, B., Marshall, L., Jia, P., Liao, J., Petronis, A., Szabó, P. E., & Labrie, V. (2019). Differential methylation of enhancer at IGF2 is associated with abnormal dopamine synthesis in major psychosis. *Nature Communications*, 10(1), 1–12.