Fithian et al. (2014) NSW

```
library(maptools)
#> Loading required package: sp
#> Checking rgeos availability: FALSE
        Note: when rgeos is not available, polygon geometry
                                                                 computations in maptools depend on gpcl
#>
        which has a restricted licence. It is disabled by default;
        to enable gpclib, type gpclibPermit()
library(ppjsdm)
library(raster)
library(sf)
#> Linking to GEOS 3.6.2, GDAL 2.2.3, PROJ 4.9.3
library(spatstat)
#> Loading required package: spatstat.data
#> Loading required package: nlme
#>
#> Attaching package: 'nlme'
#> The following object is masked from 'package:raster':
#>
       getData
#> Loading required package: rpart
#> spatstat 1.63-0
                         (nickname: 'Space camouflage')
#> For an introduction to spatstat, type 'beginner'
#> Attaching package: 'spatstat'
#> The following objects are masked from 'package:raster':
#>
       area, rotate, shift
remove(list = ls())
source("../R/get_nsw.R")
set.seed(1)
```

This vignette explains how to use the ppjsdm package with the NSW dataset from Fithian et al. (2014). We begin by loading the data with only the most prevalent species.

```
number_of_species <- 2 # Includes the most prevalent species from the plot

nsw <- get_nsw(prevalent = number_of_species)
configuration <- nsw$configuration
window <- nsw$window
covariates <- nsw$covariates</pre>
```

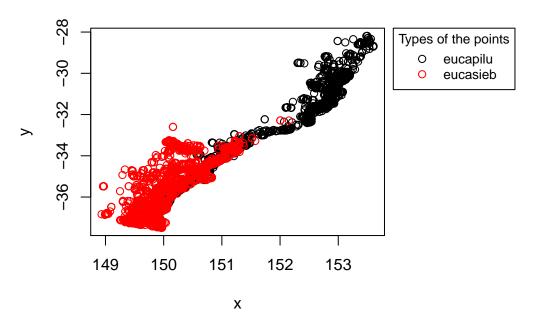
Some of the covariates give NA values at some locations in the configuration, so we have to remove these points

```
for(covariate in append(covariates, list(window))) {
  ok <- inside.owin(x_coordinates(configuration), y_coordinates(configuration), covariate)</pre>
```

The point configuration is plotted below.

```
print(configuration)
#> An S3 object representing a configuration.
#>
#> Number of points: 4913.
par(mar = c(5, 4, 4, 13) + 0.1)
plot(configuration, window = window)
```

Points in the configuration



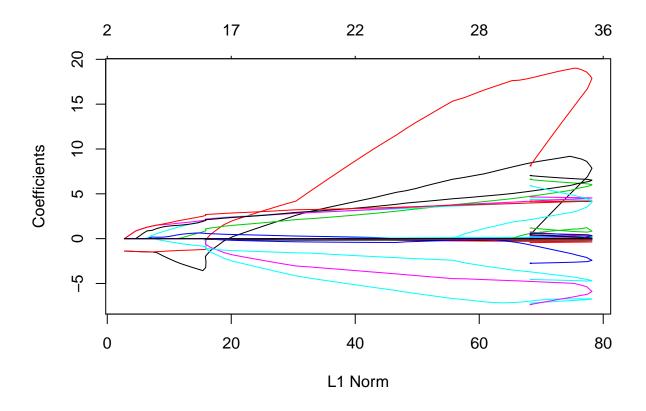
The matrix radii defined below models interaction radii within a species (on the diagonal), and between species (outside the diagonal).

```
short_range <- matrix(0.0005, number_of_species, number_of_species)
medium_range <- diag(0.0005, number_of_species)
long_range <- medium_range + diag(0.001, number_of_species)

model <- "Geyer"
medium_range_model <- "Geyer"</pre>
```

Fitting the model to the dataset is then quite easy.

```
short_range = short_range,
                     medium_range = medium_range,
                     long_range = long_range,
                     use_glmnet = TRUE)
#> 39 x 1 sparse Matrix of class "dgCMatrix"
#> (Intercept) .
#> log_lambda1 6.691550e+00
#> log_lambda2 1.440687e+01
#> alpha_1_1 6.684091e+00
#> alpha_1_2
            6.242880e-02
             4.386132e+00
#> alpha_2_2
4.173006e+00
#> gamma_2_2
#> bc02_1
             -1.067341e-01
#> bc02_2
               4.301501e-01
#> bc04_1
              -7.186190e+00
#> bc04_2
             -7.358519e+00
#> bc05_1
              1.901697e-02
           -4.409896e-01
#> bc05_2
#> bc12 1
              7.228977e-03
#> bc12_2
#> bc14_1
              2.655619e-01
              2.086023e-01
#> bc14_2
#> bc21_1
              3.428696e-01
#> bc21_2
              4.368892e-01
#> bc32_1
              7.312548e-01
              1.189161e+00
#> bc32_2
              5.876227e-01
#> bc33_1
#> bc33_2
              5.910044e+00
#> mvbf_1
              -1.518058e-02
#> mvbf_2
              -1.549144e-01
           -3.832016e-v2
-2.326913e-02
-2.747255e+00
#> rjja_1
#> rjja_2
#> rsea_1
             -4.546395e+00
#> rsea_2
#> rugg_1
              -2.205302e-02
#> rugg_2
             -7.174033e-02
#> subs_1
              1.603860e-02
#> subs_2
#> twmd_1
             -1.864165e-04
#> twmd_2
              -3.094481e-04
#> twmx_1
               -1.767750e-05
#> twmx_2
               -6.435004e-05
plot(fit$complete)
```



plot(fit\$cv)

