

IreneChang_A03_DataExploration.Rmd

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OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Rename this file `<FirstLast>_A03_DataExploration.Rmd` (replacing `<FirstLast>` with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

TIP: If your code extends past the page when knit, tidy your code by manually inserting line breaks.

TIP: If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse, lubridate), and upload two datasets: the ECOTOX neonicotinoid dataset (`ECOTOX_Neonicotinoids_Insects_raw.csv`) and the Niwot Ridge NEON dataset for litter and woody debris (`NEON_NIWO_Litter_massdata_2018-08_raw.csv`). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the subcommand to read strings in as factors.

```
getwd()
```

```
## [1] "/home/guest/EDA/EDA-Spring2023"
```

```
library(tidyverse)
library(lubridate)
Neonics <- read.csv("./Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv", stringsAsFactors = TRUE)
Litter <- read.csv("./Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv", stringsAsFactors = TRUE)
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: You might want to look at how effective neonicotinoids are on insects at different stages of their life. Did the use of a neonicotinoid lead to the mortality of a specific class on insects? What was the toxicity of the chemical used? A person may also be interested in whether or not they have negative externalities on the environment, ie. soil quality, but this was not studied in this dataset.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: You would be interested in studying litter and woody debris that falls on the ground to better understand the chemical makeup (such as the carbon and nitrogen concentrations) of the debris over time and how changes may affect decomposition and plant productivity. Increased nitrogen in debris can potentially be associated with soil carbon sequestration.

4. How is litter and woody debris sampled as part of the NEON network? Read the `NEON_Litterfall_UserGuide.pdf` document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. Clear definition of litter and woody debris. For example, litter is defined as “dropped from the forest canopy and has a butt end diameter <2cm and a length of <50cm”. 2. Each sample is sorted by functional group. 3. Then each sample is dried and weighed.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
dim(Litter) #188 rows and 19 columns
```

```
## [1] 188 19
```

```
dim(Neonics) #4623 rows and 30 columns
```

```
## [1] 4623 30
```

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
summary(Neonics$Effect)
```

```
##      Accumulation      Avoidance      Behavior      Biochemistry
##           12           102           360           11
##      Cell(s)      Development      Enzyme(s) Feeding behavior
##           9           136           62           255
##      Genetics      Growth      Histology      Hormone(s)
##          82           38           5           1
##      Immunological      Intoxication      Morphology      Mortality
##          16           12           22           1493
##      Physiology      Population      Reproduction
##           7           1803           197
```

Answer: The most common effects that are studied are mortality (1493 observations), population (1803), and behavior (360). These may be of interest to understand what effect the insecticide had on the mortality, population, and behavior of insects. Changes in these effects are informative to the effectiveness of the insecticide.

- Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed. [TIP: The `sort()` command can sort the output of the summary command...]

```
sort(summary(Neonics$Species.Common.Name))
```

```
##      Ant Family      Apple Maggot
##           9
##      Glasshouse Potato Wasp      Lacewing
##          10
##      Southern House Mosquito      Two Spotted Lady Beetle
##          10
##      Spotless Ladybird Beetle      Braconid Parasitoid
##          11
##      Common Thrip      Eastern Subterranean Termite
##          12
##      Jassid      Mite Order
##          12
##      Pea Aphid      Pond Wolf Spider
##          12
##      Armoured Scale Family      Diamondback Moth
##          13
##      Eulophid Wasp      Monarch Butterfly
##          13
##      Predatory Bug      Yellow Fever Mosquito
##          13
##      Corn Earworm      Green Peach Aphid
##          14
##      House Fly      Ox Beetle
##          14
##      Red Scale Parasite      Spined Soldier Bug
##          14
##      Western Flower Thrips Hemlock Woolly Adelgid Lady Beetle
```

##	15	16
##	Hemlock Wooly Adelgid	Mite
##	16	16
##	Onion Thrip	Araneoid Spider Order
##	16	17
##	Bee Order	Egg Parasitoid
##	17	17
##	Insect Class	Moth And Butterfly Order
##	17	17
##	Oystershell Scale Parasitoid	Black-spotted Lady Beetle
##	17	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Codling Moth	Flatheaded Appletree Borer
##	19	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Argentine Ant	Beetle
##	21	21
##	Mason Bee	Mosquito
##	22	22
##	Citrus Leafminer	Ladybird Beetle
##	23	23
##	Spider/Mite Class	Tobacco Flea Beetle
##	24	24
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Ground Beetle Family
##	25	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ladybird Beetle Family
##	29	30
##	Parasitoid	Braconid Wasp
##	30	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Sweetpotato Whitefly	Aphid Family
##	37	38
##	Cabbage Looper	Buff-tailed Bumblebee
##	38	39
##	True Bug Order	Sevenspotted Lady Beetle
##	45	46
##	Beetle Order	Snout Beetle Family, Weevil

##		47		47
##	Erythrina Gall Wasp		Parasitoid Wasp	
##		49		51
##	Colorado Potato Beetle		Parastic Wasp	
##		57		58
##	Asian Citrus Psyllid		Minute Pirate Bug	
##		60		62
##	European Dark Bee		Wireworm	
##		66		69
##	Euonymus Scale		Asian Lady Beetle	
##		75		76
##	Japanese Beetle		Italian Honeybee	
##		94		113
##	Bumble Bee		Carniolan Honey Bee	
##		140		152
##	Buff Tailed Bumblebee		Parasitic Wasp	
##		183		285
##	Honey Bee		(Other)	
##		667		670

Answer: The 6 most commonly studied species are: Other (670), Honey Bee (667), Parasitic Wasp (285), Buff Tailed Bumblebee (183), Carniolan Honey Bee (152), Bumble Bee (140), and Italian Honeybee (113) if not including other. These species all come from the same Hymenoptera family. They may be of interest over other insects to see if the effects of insecticides differentiate among different bee species. It may also be that they would like to specifically target this family with the insecticide, so they are testing the effects it has within the hymenoptera family.

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` column in the dataset, and why is it not numeric?

```
class(Neonics$Conc.1..Author.)
```

```
## [1] "factor"
```

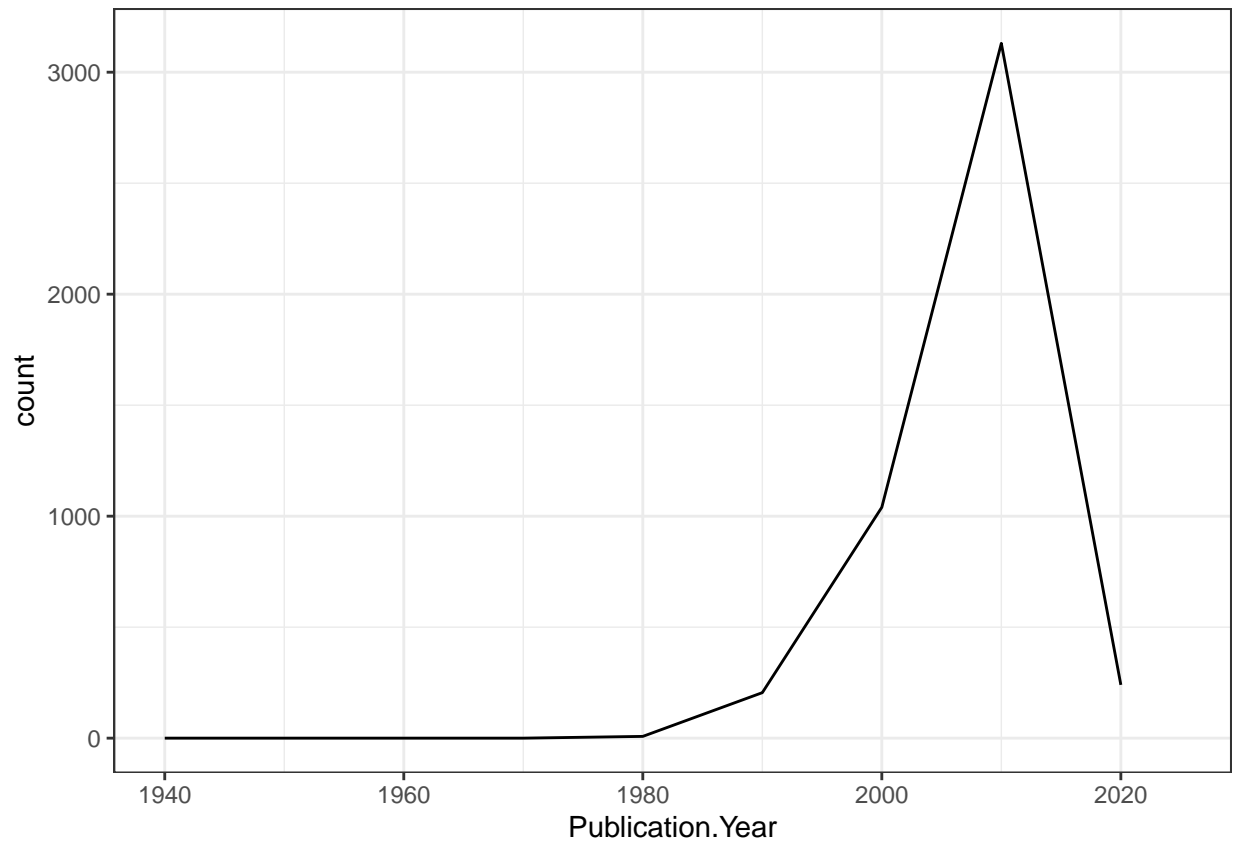
Answer: The class is a factor. These can be used to represent categorical data and are stored as integers, which makes them easy for statistical analysis and plotting.

Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics, aes(x=Publication.Year)) +
  geom_freqpoly(binwidth=10)+
  xlim(c(1940,2025))+
  theme_bw()
```

```
## Warning: Removed 2 rows containing missing values ('geom_path()').
```

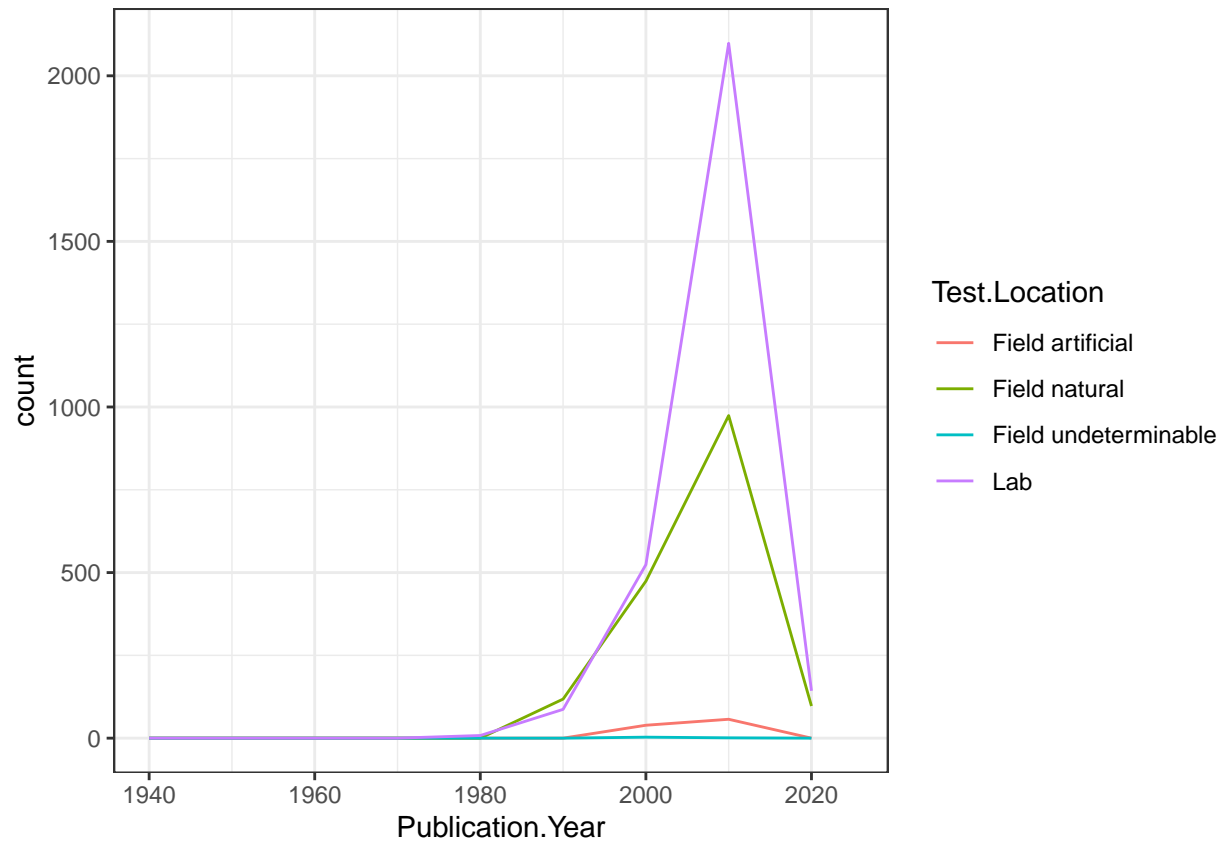


*#if you have a smaller binwidth, the buckets of data that you are putting it in is smaller
 #set the limits by data and making it into a vector
 #theme_bw() makes it so that the color scheme of the chart is black and white*

10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
ggplot(Neonics, aes(x=Publication.Year, color=Test.Location)) +
  geom_freqpoly(binwidth=10)+
  xlim(c(1940,2025))+
  theme_bw()
```

Warning: Removed 8 rows containing missing values (‘geom_path()’).



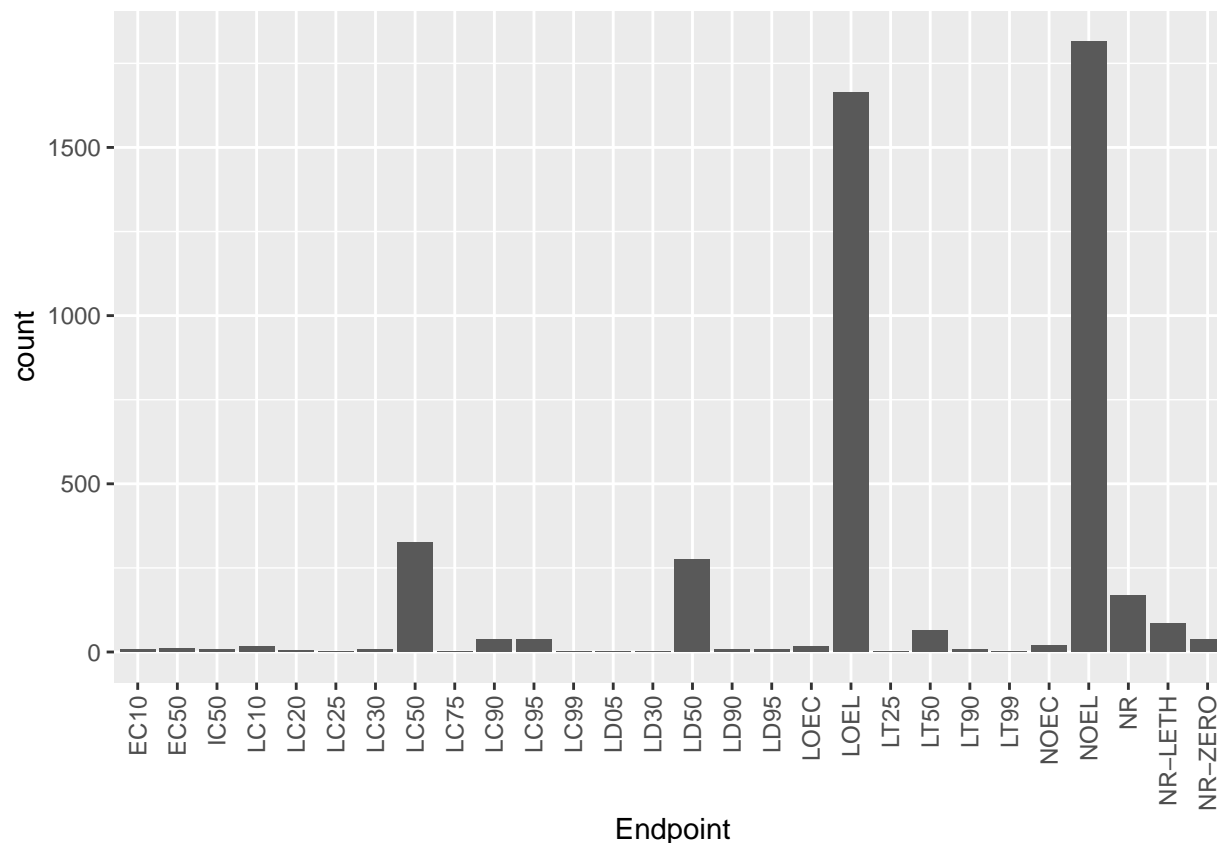
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common is lab, though there was a brief time between 1980 to 1995 where Field natural was more common. The number of observations goes up drastically around 1995, and the lab test location becomes extremely common and field natural rises, but not to the extent that lab does.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

[**TIP:** Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics, aes(x=Endpoint)) +
  geom_bar() +
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))
```



Answer: The most common end points are NOEL and LOEL. For LOEL, the database use was terrestrial and is the lowest-observable effect level: on the lowest dose producing effects that were significantly different from the response of controls. For NOEL, the database was terrestrial and is the was the no observable effect level: the highest dose producing effects not significantly different from responses from the control.

Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate) #this is a factor.
```

```
## [1] "factor"
```

```
CD.Collection.Date <- ymd(Litter$collectDate)
class(CD.Collection.Date)
```

```
## [1] "Date"
```

```
unique(Litter$collectDate)
```

```
## [1] 2018-08-02 2018-08-30
## Levels: 2018-08-02 2018-08-30
```


13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

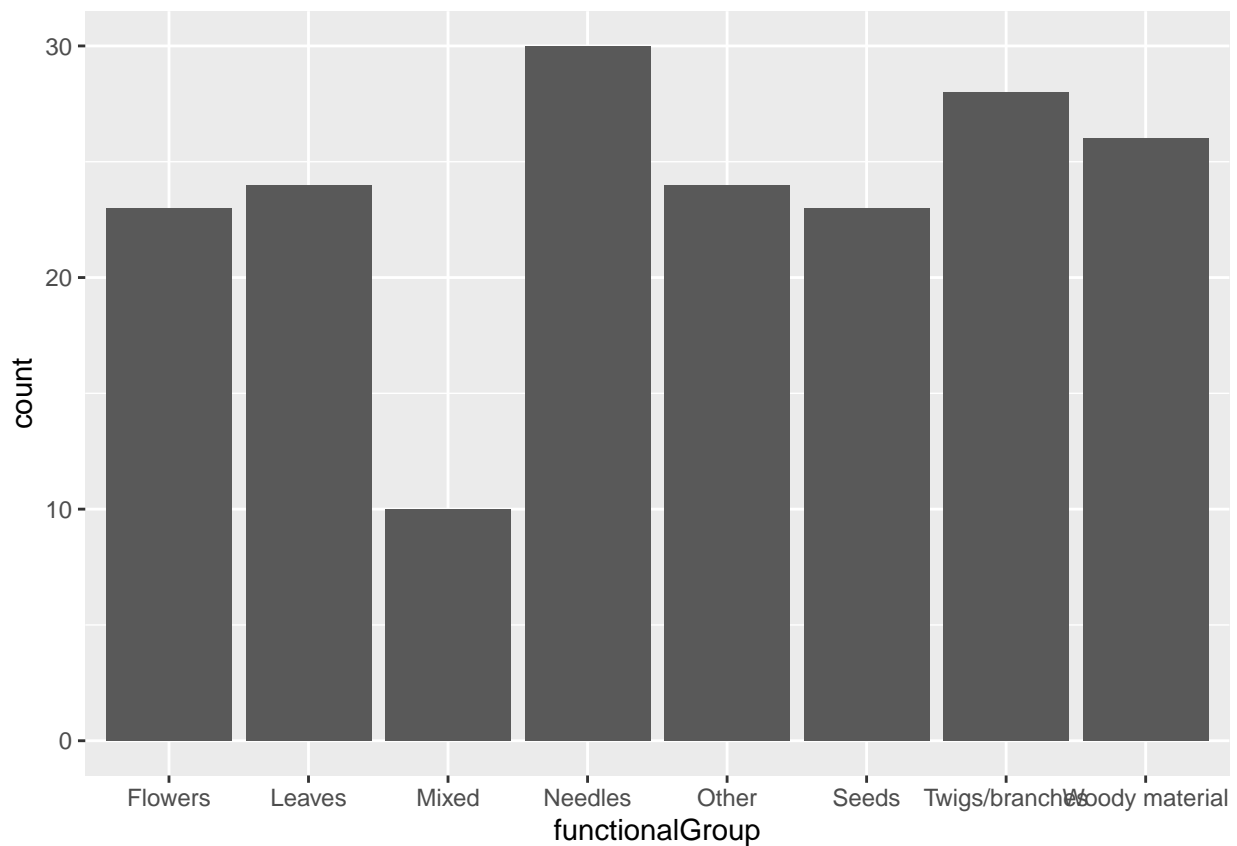
```
unique(Litter$plotID)
```

```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051  
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057  
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

Answer: The `unique` function will get rid of duplicates in the data.

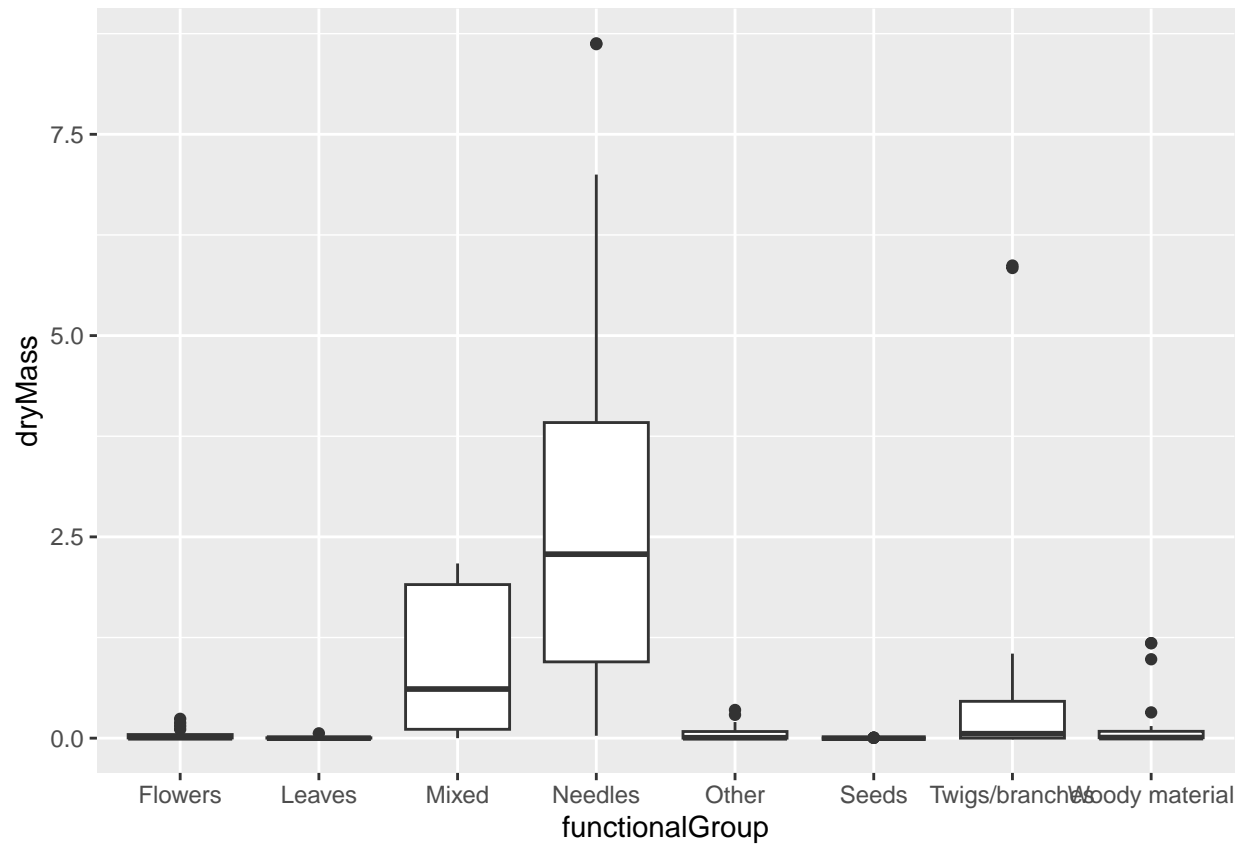
14. Create a bar graph of `functionalGroup` counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter, aes(x=functionalGroup)) +  
geom_bar()
```

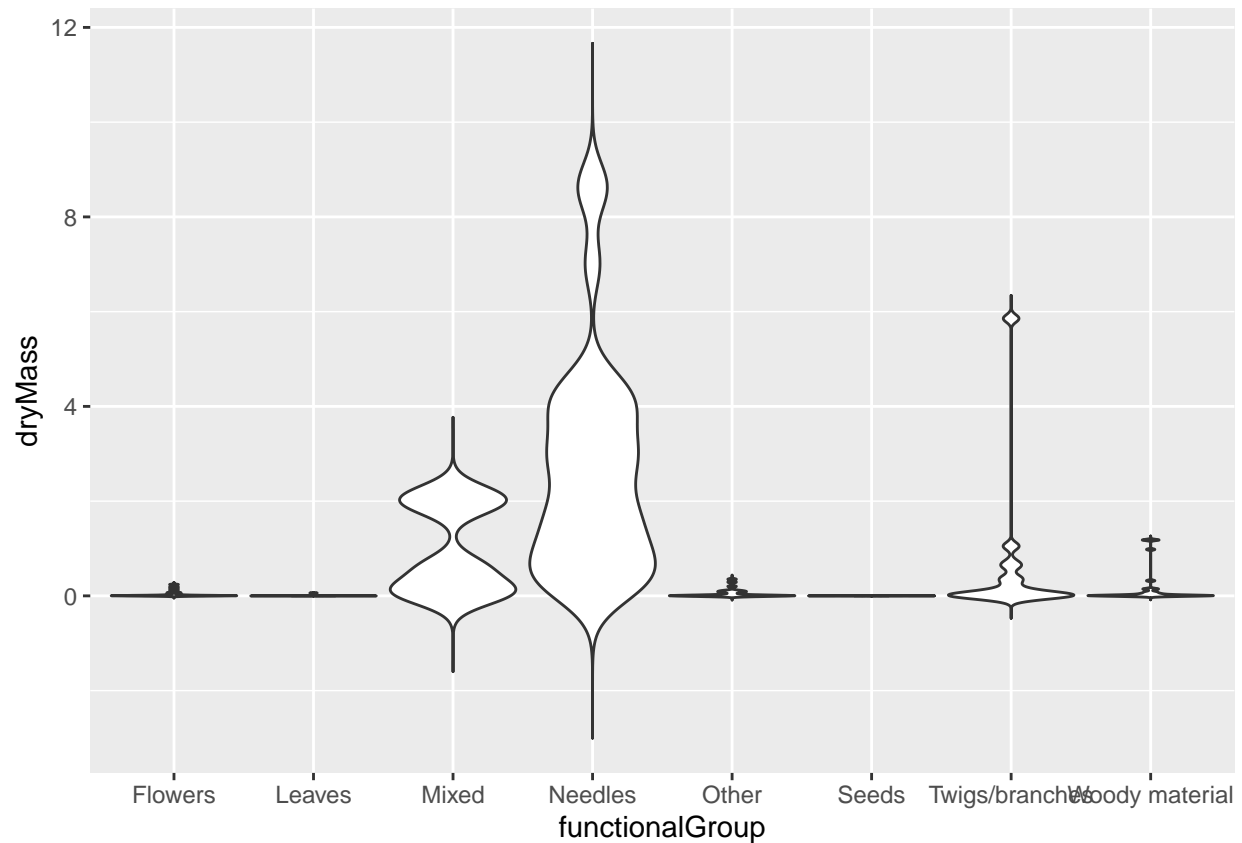


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
ggplot(Litter, aes(x=functionalGroup, y=dryMass)) +  
geom_boxplot()
```



```
ggplot(Litter, aes(x=functionalGroup, y=dryMass)) +  
geom_violin(scale = "width", trim = FALSE, adjust = 0.5)
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: The boxplot is more effective because it gives a more clear picture of the summary statistics. In general for the violin plot, you need enough data for it to be effective and unless setting parameters for the violin plot, there is little data you can obtain from first glance.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles

#check output