1 Identification and Exploration of Pharmacological pyroptosis-related

2 **Biomarkers of Ulcerative Colitis**

Kaiwei Chen†, Shipeng Shang†, Shengnan Yu, Luwen Cui, Shangyong Li‡, Ningning
 He‡

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- 7 School of Basic Medicine, Qingdao Medical College, Qingdao University, Qingdao
- 8 266003, China.

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- † Those authors contributed equally.
- ‡Corresponding author: Prof. Shangyong Li; Email: lisy@qdu.edu.cn
- 12 Prof. Ningning He; Email: heningning@qdu.edu.cn

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Abstract

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Ulcerative colitis (UC) is a a type of chronic inflammatory bowel disease (IBD). Its 17 etiology is unclear. Much evidence suggests that the abnormal intestinal epithelial 18 19 cells (IECs) deathdeath of abnormal intestinal epithelial cells (IECs) leads to intestinal barrier disruption, and the subsequent inflammatory response plays an important vital 20 role in UC. Pyroptosis is a form of programmed inflammatory cell death, and the role 21 of pyroptosis in UC etiology remains to be explored. In this study, 10 hub genes in 22 pyroptosis were identified This study identified 10 hub genes in pyroptosis by gene 23 expression profiles obtained from the GSE87466 dataset. Meanwhile, the biomarkers 24 were screened according based onto gene significance (GS) & and module 25 membership (MM) in-through the Weighted Gene Co-Expression Network Analysis 26 27 (WGCNA) analysis. The following analysis indicated those hub genes were were closely related associated withto the UC progression and therapeutic drugs response. 28 The single-cell RNA (scRNA) sequencing data from UC patients in-within the 29 30 GSE162335 dataset indicated that macrophages were most related to the occurrence 31 of pyroptosis. Finally, the expression of hub genes and response to the therapeutic drug (5-aminosalicylic acid, 5-ASA) were verified in dextran sulphate sulfate sodium 32 (DSS) induced colitis mice. Our study identified IL1B is as the key critical 33 34 pyroptosis-related biomarker in UC, and t. The crosstalk between macrophages 35 pyroptosis and IECs pyroptosis may play an important essential role in UC, which 36 deservesing further exploration.

Keywords: Ulcerative colitis; pyroptosis; Transcriptome; *IL1B*; DSS-induced colitis

Introduction

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Ulcerative colitis (UC) is a chronic inflammatory bowel disease (IBD) of unclear etiology, that usually beginsning in the rectum and subsequently spreads spreading to the colonic mucosa (1). The incidence rate of UC is 9 to 20 cases per 100,000 person-years, and the prevalence rate is 156 to 291 cases per 100,000 people, which has become constituting a global burden (2). The life quality of life of UC patients is commonly impaired by due to diarrhea, abdominal pain, bloody stools, and the increased elevated risks of colon and rectal cancer (3). A variety of Various factors have been implicated in the developingment of UC, such as dysregulated immune response, gut microbial dysbiosis, genetic susceptibility, and environmental influences (4). UC patients cannot cannot be completely wholly cured and often take long-term medication (5). Currently, tThere is a widely accepted theory that a vicious cycle of intestinal barrier disruption, cell death, and subsequent inflammatory response liesrests at the heart of chronic inflammation (6). The continuous monolayer of IECs—is, the first barrier against microbial and environmental pressure, performs implements a critical innate immune function (7). Remarkably, the abnormal death of large numbers of IECs is commonly observed in preclinical models of UC patients with UC (8). Moreover, intestinal barrier dysfunction can be detected by confocal endoscopy before the occurrence of intestinal lesions, which canthereby predicting the recurrence of IBD (9). Pyroptosis, also known ascalled inflammatory cell necrosis, is a new form of programmed inflammatory cell death (10). The canonical pyroptosis pathway relies depends on caspase 1 for the cleavage of gasdermin D (GSDMD). In contrast, whereas the non-canonical pathway relies on caspase 4/5/11. After cleavage, the N-terminal end of GSDMD forms develops a transmembrane pore, which releasinges inflammatory cytokines such as IL-1\beta and IL-18, and interferes with ion and water regulation, ultimately leading tocausing intense inflammation and cell death (11). This pattern of cell death accompanied by <u>a strong robust</u> inflammatory response <u>is becomes</u> a double-edged sword for the host (10). An over-activated inflammatory response helps

the host defend against pathogenic infections, but may could also cause 68 69 various inflammatory diseases, including such as sepsis (12) and gout (13). During the active phase of UC, many several different forms of cell death, including 70 71 pyroptosis, are widely activated (14). The development of UC may could be related to 72 pyroptosis of IECs and the release of inflammatory factors, t. The relationship between the two has attracted extensive significant interest (15). For example instance, it has 73 been reported that IL-36β, a member of the IL-36 subfamily of the IL-1 family, 74 75 enhances increases the pathology of DSS-induced colitis in mice by promoting enhancing Th2 responses in LPL while decreasing Foxp3+ Treg responses (16). 76 However, current molecular mechanisms related associated withto pyroptosis in UC 77 are still-lacking in research. Therefore, exploration of its regulatory mechanisms and 78 79 gene expression characteristics will help to reveal the etiology of UC and provideing its regulatory mechanisms and gene expression characteristics will help understand 80 the etiology of UC and provide a new perspective for on UC treatment. 81 In this study, using GSE87466 dataset as the base data, hub genes associated with 82 83 pyroptosis in UC were identified by Weighted Gene Co-Expression Network Analysis (WGCNA) combined gene expression matrix using the GSE87466 dataset as the base 84 data. After validation by GSE92415, GSE107499, GSE59071, GSE73661, and 85 GSE46451 datasets, the expression change of hub genes after different drug 86 87 treatments wereas further observed after different drug treatments to revealdemonstrate the role of pyroptosis in UC. Subsequently, by single-cell RNA analysis 88 of the GSE162335 dataset, we explored the expression patterns of hub genes in the 89 90 macrophage clusters most associated with pyroptosis of immune cells by single-cell 91 RNA analysis of the GSE162335 dataset, and. We identified macrophage clusters with having different roles in UC pyroptosis as a way to further reveal the mechanisms 92

MATERIALS AND METHODS

associated with related to pyroptosis in UC.

Datasets and Preprocessing

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96 Gene expression profiles and corresponding clinical data of UC were obtained

retrieved from the Gene Expression Omnibus (GEO) database (http://www.ncbi.nlm.nih.gov/geo). Information of on all the datasets in this study was is shown described in Table 1. Descriptions of The patient demographic characteristics for all the datasets are available in Supplementary Material 1. Pyroptosis-related (PRGs) were obtained procured from the Uniprot genes database (https://www.uniprot.org/), MSigDB database (https://www.gsea-msigdb.org/), and previous studies in the literature (17, 18), see (Supplementary Table S1) for more details.

Table 1 Information for all the datasets in this study

Dataset	Platform	Title
GSE87466	GPL13158	[HT_HG-U133_Plus_PM] Affymetrix HT HG-U133+ PM Array Plate
GSE92415	GPL13158	[HT_HG-U133_Plus_PM] Affymetrix HT HG-U133+ PM Array Plate
GSE107499	GPL15207	[PrimeView] Affymetrix Human Gene Expression Array
GSE59071	GPL6244	[HuGene-1_0-st] Affymetrix Human Gene 1.0 ST Array
GSE73661	GPL6244	[HuGene-1_0-st] Affymetrix Human Gene 1.0 ST Array
GSE46451	GPL10558	Illumina HumanHT-12 V4.0 expression beadchip
GSE162335	GPL20301	Illumina HiSeq 4000

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Differential Expression Analysis

The dataset GSE87466 containing 87 UC patients and 21 healthy individuals, was used_utilized_to identify the differentially expressed genes (DEGs) between the UC and healthy controls (19). The DEGs were screened with_having_a threshold of P-value<0.05 &_and_|log2FC|>1 by using the "limma" package. Visualization of DEGs was performDEGs were visualized using the R packages "ggplot2" and "pheatmap".."

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Construction of Co-Expression Network and Identification of

Modules

In order to reduce the computational effort of the whole network and maintain the characteristics of scale-free topological network structure, tThe 3,945 variant genes were screened (Supplementary Table S2) with a coefficient of variation > 0.08 to

reduce the computational effort of the whole network and maintain the characteristics of a scale-free topological network structure. A weighted gene co-expression network was constructed using-with the R package "WGCNA" for the screened-3,945 variant genes. First, the sample data with abnormal gene expression values were filtered by using hierarchical clustering, and the "pickSoftThreshold" function was used-utilized to evaluate assess the appropriate soft threshold β. Subsequently, the Pearson correlation coefficients of the gene was were calculated determined, the weighted adjacency matrix was constructed with using a soft threshold β of 13, and the adjacency matrix was converted into a the topological overlap matrix (TOM). The minimum module size cutoff was set to 30, and the same modules were merged with using a threshold value of 0.2. Genes with similar expression patterns were grouped into within the same module. Module eigengene (ME) was calculated to represent the overall level of gene expressiongene expression level within inside the module and used to identify modules highly-significantly correlated to with the disease.

Identification of Pyroptosis-Related Hub Genes and Biomarkers

The hub gene was obtained retrieved by taking the overlapping of PRGs, DEGs, and genes from the module with having the highest relevance to UC in WGCNA analysis. Biomarkers were screened from hub genes based depending on the criteria of gene significance (GS) > 0.2 & module membership (MM) > 0.8 in the WGCNA analysis (20). The Receiver Operating Characteristic (ROC) analysis of the hub genes using was performed with the R package "pROC"..."

Immune Cell Infiltration Estimation

The relative abundance of immune cells in—within the colonic mucosal tissue of the UC and healthy controls were evaluated assessed using the CIBERSORT algorithm (https://cibersort.stanford.edu/) (21). The differences between the two groups of immune cells were—was compared using—with Student's t-test and visualized by a "ggboxplot"—." The correlation between each infiltrating immune cells and the

relationship between hub genes and immune cells was visualized by using the "corrplot" package.

Single-Cell Analysis

The dataset GSE162335 was contained single-cell sequencing data of immune cells		
from the CD45+ colonic lamina propria of the 11 UC patients (22). The "Seurat"		
package was <u>used-utilized</u> for subsequent data processing (23). The low-expressing		
cells and genes were filtered, while ensuring that the percentage of mitochondria per		
cell was below 5% & and the features of genes was were below 6,000 and above 500.		
Finally, a total of 18,375 cells were obtained identified. After normalization using the		
"LogNormalize" method, 3,000 highly variable genes (HVG) were identified using		
with the "vst" method. PCA was applied to identify significant principal components		
based on the expression of HVG. Thus, and 25 PCs were selected for t-SNE analysis		
with having a resolution of "2" to identify the different clusters. "FindAllMarkers"		
function (logfc.threshold = 0.25) was applied incorporated to identify DEGs in each		
cluster_and marker gene for each cluster with using avg_log2FC>1.		
Using the "BlueprintEncodeData" dataset from the R package "celldex" as a a		
reference, each cluster was initially annotated by with "SingleR" The R package		
"scHCL" was used to further annotate the cluster of interest (24), with using the		
Human Cell Landscape (http://bis.zju.edu.cn/HCL/) & the scRNASeqDB		
(<u>https://bioinfo.uth.edu/scrnaseqdb/</u>) databases as <u>a-the</u> secondary reference.		
The R package "AUCell" was used to assess evaluate the response of single cells to		
PRGs (25), and the "Aucell_explorethreshold" function was used to determine the		
threshold for identifying gene set active cells. Then, the AUC score of each cell were		
was mapped to the t-SNE embedding using with "ggplot2" for visualization. The		

Functional Annotation and Pathway Enrichment Analysis

GO annotations of genes from the R package "org.Hs.eg.db" (version 3.1.0) was

used_utilized_as background. The selected genes were mapped to the background set, and performed_the_enrichment analysis was performed_using the R package ""clusterProfiler"—" (version 3.14.3). P < 0.05 & FDR < 0.25 were considered statistically significant.

Animal experiment

The experimental mice and standard rodent chow food were purchased obtained from Jinan Pengyue Laboratory Animal Breeding Company (Jinan, China). The Eexperimental procedures were reviewed and approved by the Ethics Committee of the Medical College of Qingdao University (QDU-AEC-2022314). C57BL/6J mice (18-20 g) were randomized into 3-three groups (n=6) after 1-one week of acclimation. NC group: no extra treatment for 7seven days; DSS group: free drinking water containing 2.5% DSS during 7-seven days; 5-aminosalicylic acid (5-ASA) group: the free drinking water containing with 2.5% DSS for 7-seven days while 5-ASA (400 mg/kg/day) were was administered by gavage. At the end of the experiment, the entire colon was excised, and the length of colon colon length was measured determined. The collected colon tissue was stored at -80°C for further analysis.

Histopathological analysis

The collected colonic tissues were fixed in 10% formalin overnight,—. The fixed tissues were embedded in paraffin and sliced after dehydration—in gradient concentrations of alcohol after dehydration at a thickness of 5 μm. Stained with hematoxylin and eosin (H&E) were observed using with a 200x magnification (E100, Nikon, Tokyo, Japan) and an imaging system (DS-U3, Nikon, Tokyo, Japan). Histological—As previously described, the histological score for H&E staining was assessed in a blinded fashion as previously described (27).

RNA extraction and quantitative real_real_time PCR (RT-qPCR) analysis

The extraction of total RNA and reverse transcription were performed according based onto the kit instructions of SparkJade (Jinan, China). The RT-qPCR primers are depicted were shown in **Supplementary Table S3**. The gene expression levels were normalized using with GAPDH, and the relative quantification of gene expression was calculated determined using through the $2^{-\Delta\Delta Ct}$ method.

Enzyme-linked immunosorbent assay (ELISA)

In At the end of the experiment, mice were anesthetized by with chloral hydrate and separated from the serum by centrifugation (3500 rpm, 4°C, 30 min) after picking out the eyeball. The ELISA kit was purchased from ABclonal (#RK00027, Wuhan, China). The serum levels of ____in the serum was were measured according based onto the instruction of the ELISA kit.

Statistical Analysis

All the statistical analysis analyses was were performed by using the R software (version 4.1.1). All the data are expressed as mean \pm SE. Unpaired Student's Student's t-test was used for to compare two comparison groups, and .-One-way ANOVA was used for three and more group comparisons groups, and Tukey_HSD was used for inter-comparison between multiple groups. This e-above statistical analysis was performed with using the R package "ggpubr" and "stats"..." P < 0.05 was considered as statistically different significant.

Result

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Research Design Summary

The flow chart was is shown in Figure 1. Firstly, DEGs were screened between the UC patient and the healthy controls from the GEO database the GEO database screened DEGs between the UC patient and the healthy controls. Subsequently, pyroptosis-related hub genes were identified using through DEGs combined with genes of the key module in WGCNA analysis and PRGs. Six genes were screened as biomarkers among hub genes based on GS and MM values, six genes were screened as biomarkers from the hub genes. and ROC analysis was used to assess determine their diagnostic values. The obtained hub genes were validated from two aspects. Next, the differences of in hub genes between UC/healthy controls, mucosal lesional/nonlesional group, and active/inactive/healthy controls were validated based depending on three datasets obtained from the GEO database, respectively. The pattern of changes in the hub gene after treatment with 5-ASA, IFX (infliximab), and VDZ (vedolizumab) was evaluated assessed. Infiltrating immune cells in UC patients was were analyzed using with CIBERSORT. Finally, the expression pattern of hub genes was evaluated evaluated in immune cells from the colonic lamina propria of UC patients at the single-cell level-and. Verification of The hub gene expression was verified usingby animal experiments.

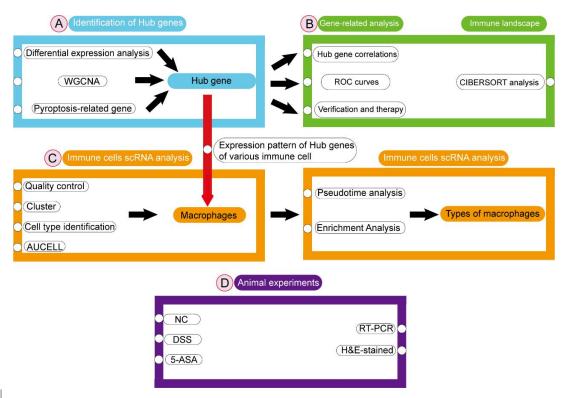


Figure 1. Flowchart of bioinformatics <u>analysis analyses</u> in this study.

Identification of DEGs and Functional Annotation and Pathway

Enrichment of DEGs

Correlation analysis of the GSE87466 dataset showed revealed stronger intra-group correlations for the UC group (Figure S1A-B). Using P-value<0.05 & |log2FC|>1 as the threshold, wWe identified 1,247 DEGs, including 843 upregulated genes and 404 downregulated genes, using P-value<0.05 & |log2FC|>1 as the threshold (Supplementary Table S4). The up and down-down-regulation distributions of DEGs were shown in the volcano plot (Figure 2A). Heatmap showed the expression patterns of DEGs and relative consistency within groups. Upregulated DEGs showed a positive correlation with the UC group and a negative correlation with healthy controls, while downregulated DEGs showed revealed the opposite (Figure 2B).

To better understand the functions of DEGs, GO and KEGG enrichment analysis was performed to understand the functions of DEGs better. GO terms of biological process (BP) showed described that DEGs were mainly primarily enriched in immunomodulation and immune response, such as response to chemical; and immune system processes; immune response (Figure 2C). Moreover, the enriched KEGG

pathway showed that a series of pathways related associated withto the inflammatory response were was activated, including such as Cytokine-cytokine receptor interaction, B cell receptor signaling pathway, Chemokine signaling pathway, and the IL-17 signaling pathway (Figure 2D). More nNotably, DEGs were also enriched by pyroptosis-related inflammatory pathways, such-includingas the NOD-like receptor signaling pathway, NF-kappa B signaling pathway, Toll-like receptor signaling pathway, and TNF signaling pathway. The enrichment results of these DEGs showed revealed that various external stimuli induced a series of immune responses, including such as the activation of some inflammatory pathways associated with pyroptosis, which indicated activating some inflammatory pathways associated with pyroptosis, indicating that pyroptosis is widely activated in UC. And Moreover, the enrichment results of these DEGs also predicted a damaged intestinal barrier in among UC patients. Intestinal barrier function consists has of three main components: mechanicalbarrier, ecological barrier and immune barrier, ecological, and immune. The disruption of the immune barrier often causes leads to functional impairment of the other two. As we-mentioned above, these terms focusing on immune responses and inflammatory pathways indicate that the immune barrier is severely impaired inamong UC patients and t. This damage further causes disruption of disrupts the mechanical barrier, such as including the abnormal death of IECs.

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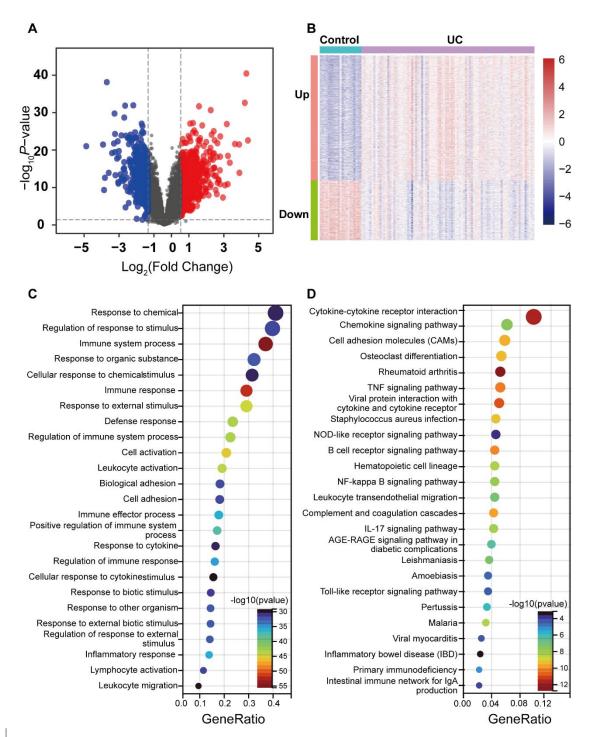


Figure 2. Identification of DEGs between the UC group and control groups in the dataset and enrichment analysis. (A) Volcano maps of DEGs between UC and control, where red dots represent the upregulated genes, blue dots represent the downregulated genes, and gray dots represent no differential gene. (B) Heatmap of DEGs in UC and control. The red indicates high expression.

(C) GO enrichment analysis of DEGs. (D) KEGG pathway enrichment analysis of DEGs.

WGCNA Construction and Key Modules Identification

The co-expression network was constructed developed to identify the most relevant modules for UC based on the expression of 3,945 CV genes. The 108 samples were clustered, and the two outlier data were removed (Figure S1C). A scale-independent topological network (soft threshold 13 scale-free R^2 0.87) and the mean connectivity network were established (Figure 3A). We obtained 11 gene modules by through hierarchical clustering and module merging (Figure 3B). Among the sem, black (r = 0.52, P = 1e-8), blue (r = 0.53, P = 4e-9), and turquoise (r = 0.75, P = 3e-20) modules were highly significantly correlated with UC (Figure 3C). The turquoise module, which containing a total of containing 1,186 genes, was selected as the feature module for UC dbased epending on the correlation coefficient and P-value, (Supplementary Table S5). Thus, 354 genes were screened as module hub genes from the turquoise module based depending on GS > 0.2 & MM > 0.8 (Figure 3D).

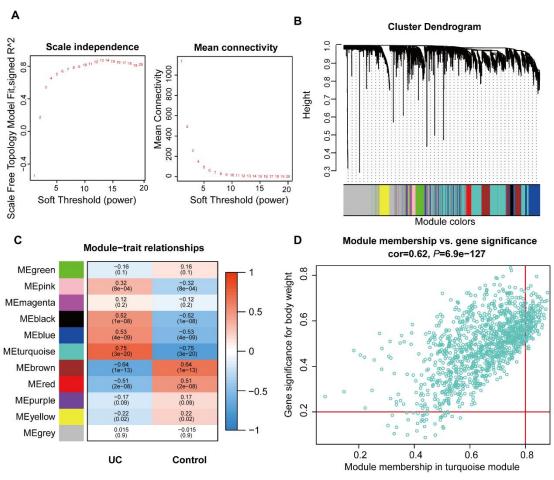


Figure 3. Identification of the key modules and genes that relate to associated with UC

by WGCNA. (A) Estimation of the scale independence index of the 1–20 soft threshold power and determiningation of the mean connectivity of the 1–20 soft threshold power. (B) Module clustering dendrogram derived from the 1-tom matrix. The different color bands represent different modules. (C) Correlations between the different various modules and traits. The number of each module represents depicts the correlation coefficient with the trait, and the color of the module ranges from red to green, representing showing from high to low correlation. (D) Scatter plot of the turquoise module genes. The vertical coordinate represents the GS score for each gene, and the horizontal coordinate represents the MM score for each gene.

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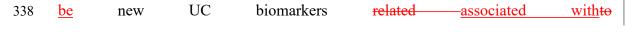
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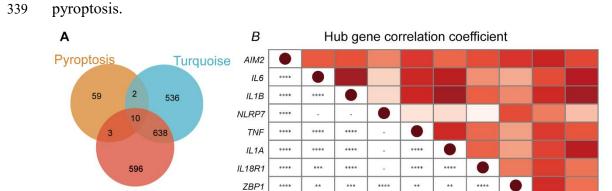
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Identification of Pyroptosis -Related Hub Genes and Biomarkers

We performed GO enrichment analysis to analyze 74 PRGs and discovered observed that these genes were enriched with interleukin-1-related biological processes inaddition to along with pyroptosis (Figure S2). This suggested indicated that pyroptosis may could be closely related associated withto interleukin 1. The 10 hub genes (AIM2, IL6, IL1B, NLRP7, TNF, IL1A, IL18R1, ZBP1, GZMB, TREM1) were obtained from the overlap of DEGs, PRGs, and turquoise module genes (Figure 4A). NLRP7 and ZBP1 were weakly-poorly interrelated with other hub genes, and IL1B had the highest correlation with IL6, IL1A, and TREM1, based depending on the expression patterns of these hub genes (Figure 4B). In order tTo identify new UC biomarkers related to pyroptosis, the overlapping parts of turquoise module hub genes, and DEGs, PRGs were taken as new biomarkers (Figure 4C). A total of 6-six genes (IL1B, IL18R1, ZBP1, AIM2, GZMB, TREM1) were obtained, respectively. To verify the diagnostic significance of the six biomarkers, ROC analysis was performed to verify the diagnostic significance of the six biomarkers (Figure 4D-I). The AUC values for all the genes were greater more significant than 0.75, with IL1B (AUC 0.97, 95%CI) having with the largest AUC value and IL18R1 (AUC 0.91, 95%CI) having showing the smallest AUC value. This indicated that IL1B, IL18R1, ZBP1, AIM2, GZMB, TREM1 may as and TREM1 could





GZMB

DEG

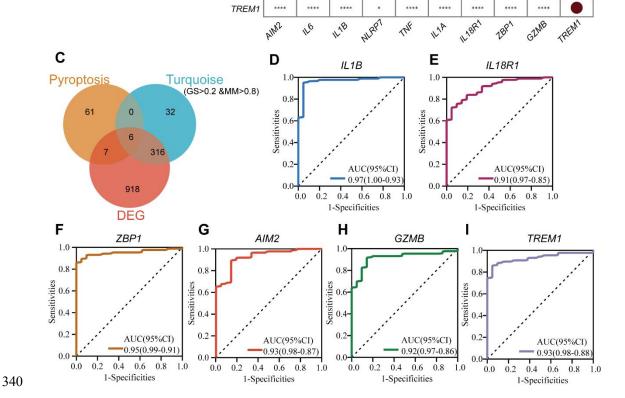


Figure 4. Identification of pyroptosis-related hub genes and biomarkers in UC. (A) Overlapping genes generated by the intersection of turquoise module genes, DEGs and PRGs showed by venn diagram The Venn diagram shows the overlapping genes generated by the intersection of turquoise module genes, DEGs, and PRGs. (B) Expression correlation matrix of each PRGs hub gene in UC. (C) Overlapping genes generated by the intersection of turquoise module hub genes, DEGs and PRGs showed by venn diagram The Venn diagram shows the overlapping genes generated by the intersection of turquoise module hub genes, DEGs, and PRGs. ROC (rReceiver operating characteristic (ROC) curves for IL1B(D), IL18R1(E), ZBP1(F), AIM2(G),

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Verification of Pyroptosis-Related Hub Genes

The 10 hub genes were further validated by using the GSE92415 dataset (27). It showed that aAll the hub genes were statistically significantly different in UC and healthy controls (P < 0.001, Figure 5A). It was verified that Moreover, all the hub genes were statistically significantly different between lesional and nonlesional groups from the GSE107499 dataset (Figure 5B). In addition, we compared the differences in hub genes between activated UC groups, inactivated UC groups, and healthy controls by the GSE59071 dataset (28). Interestingly, only IL1B showed a statistical difference among the activateOnly IL1B showed a statistical difference among the activated UC groups, inactivated UC groups, and healthy controls (Figure 5C). All the hub genes except IL1B were not statistically significantly different in healthy controls and inactivated UC groups includeds IL6 and TNF, common pro-inflammatory factors. Subsequently, we explored four other pro-inflammatory factors, including IL8, IL17A, IL18, and IL33, were explored, all of which are were thought to be closely associated with the development of UC disease. although. However, their role in pyroptosis remains unexplored. Interestingly, t The expression pattern of IL33 was similar to IL1B, with statistically significant differences between all three groups (Figure S3). The relationship between IL33 and UC may could also deserve to be focused on on in the future (29). In conclusion Therefore, IL1B was probably the more vital of the hub genes and and it showed strong correlationtrongly correlated with other hub genes.

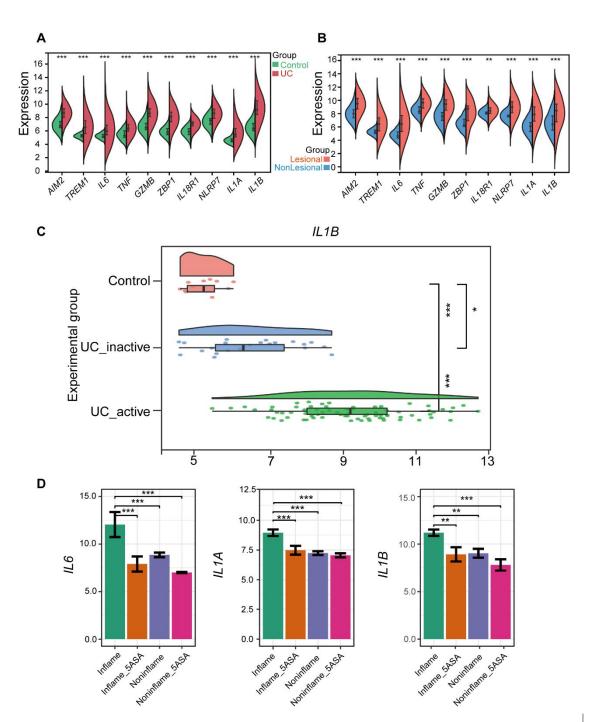


Figure 5. Verification of PRGs hub genes in UC and Evaluation of 5-ASA for the treatment of the UC pyroptosis. (A) Expression—The violin plot showed the expression of 10 hub genes in the colonic mucosa of the UC group and the control group in GSE92415 of 10 hub genes in the colonic mucosa of UC group and control group in GSE92415 showed by violin plot. (B) Expression of 10 hub genes in the lesional and nonlesional colonic mucosa of UC patients in GSE107499 showed by violin plot Comp violin plot showed the expression of 10 hub genes in the lesional and nonlesional colonic mucosa of UC patients in GSE107499. (C) The expression—of—

IL1B in the colonic mucosa of control, inactive and active UC patients in GSE59071-showed by raincloud plotraincloud plot showed the expression of IL1B in the colonic mucosa of control, inactive and active UC patients in GSE59071. (D) 5-ASA alleviates rectal mucosal damage in UC patients by regulating-controlling IL6, IL1A, and IL1B in the RPGs hub genes. The relative expression levels of IL6, IL1A, and IL1B within the rectal mucosa of inflame (inflammation but no drug), inflame_5-ASA (inflammation and treated with 5-ASA), non-inflame (non- inflammation and no drug), and non-inflame_5-ASA (non- inflammation and treated with 5-ASA). *P < 0.05, **P < 0.01 and ***P < 0.001.

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Drugs Improve Rectal and Colonic Mucosal Damage in UC Patients

by Reducing Pyroptosis-Related Hub Genes

Subsequently, we explored the influence of drugs on changes in the expression of pyroptosis-related hub genes was explored using GSE46451 and GSE73661 (30). 5-ASA was the drug of choice for mild-to-moderate UC (31). IFX, adalimumab, and golimumab that target to TNF-α, VDZ that target to α4β7 integrin, and ustekinumab that target to TNF-α, VDZ that target α4β7 integrin, and ustekinumab that target IL-12 and IL-23 were are the five most common biologics approved for the treatment of ing UC, and they were recommended as the first-line treatment for moderate-to-severe UC (32). It showed revealed that there was no difference in the expression of AIM2, TREM1, TNF, GZMB, ZBP1, IL18R1, and NLRP7 before and after 5-ASA effect on inflamed or non-inflamed rectal mucosa in vitro of UC patients in vitro, suggesting indicating that 5-ASA may have no effect onnot affect these 7-seven RPGs. (Figure S4). However, the expression levels of *IL6*, *IL1A*, and *IL1B* were significantly reduced <u>decreased</u> in the inflamed rectal mucosa after receiving 5-ASA, n. Tot only that, these three genes also showed depicted significant differences between inflamed rectal mucosa and non-inflamed rectal mucosa (Figure 5D). Meanwhile, the expression of hub genes in the colonic mucosa of patients with with active UCactive UC was significantly decreased after IFX treatment. Moreover, we found no difference - between pre-IFX treatment and non-responders, but a significant difference was observed between after-IFX treatment and non-responders. Most importantly, except for IL1A, the expression levels of the other 9-nine hub genes were restored to the levels of healthy controls, except for IL1A (Figure 6A). The Eexpression patterns of pyroptosis-related hub genes before and after VDZ treatment of patients with active UC were almost identical similar to IFX (Figure 6B). Therefore, we speculated that IFX and VDZ probably could have reduced the occurrence of pyroptosis while treating UC_{7-.}5-ASA has been used to treat mild UC for nearly three decades, while biologics such as IFX₇ and VDZ were often used to treat patients with moderate and severe UC. Based on the above-mentioned changabovementionede pattern of hub genes, we speculated that the progression of the UC condition might-could be associated with pyroptosis.

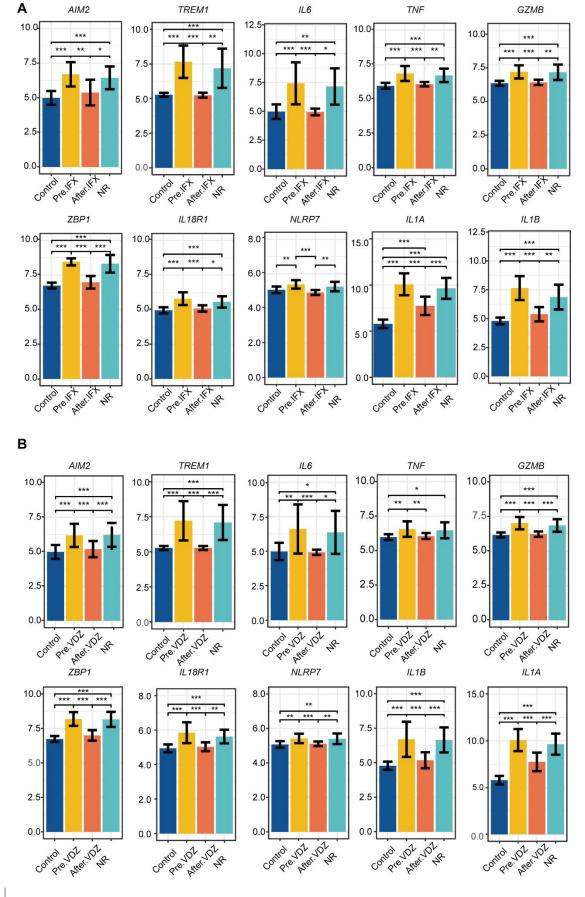


Figure 6. IFX and VDZ reduces impaired colonic mucosa of UC patients by

regulating the PRGs hub gene. (A, B) The relative expression levels of *AIM2*, *TREM1*, *IL6*, *TNF*, *GZMB*, *ZBP1*, *IL18R1*, *NLRP7*, *IL1A*₂ and *IL1B* in the colonic mucosa of control, Pre.IFX (UC patients before IFX therapy), After.IFX (UC patients in remission after IFX therapy) and NR (UC patients not n-responding to IFX therapy).

429 *P < 0.05, **P < 0.01 and ***P < 0.001.

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Immune Infiltration Analysis

Infiltration immune cell of UC calculated by CIBERSORT algorithmThe CIBERSORT algorithm determined the infiltration of immune cells of UC. The was used to evaluated the differences in immune cell abundance between the UC group and healthy controls were evaluated, and samples with P > 0.05 were filtered. The correlation between the 22 immune cells was is shown in Figure 7A. Mast cells activated and NK cells resting had the highest positive correlation (r = 0.58). T cells follicular helper and B cells naive also showed-revealed a positive correlation (r = 0.51). Meanwhile, there is was a negative correlation between T cells follicular helper and macrophages M2, mast cells resting, and mast cells activated (r = -0.52). Figure 7B showed demonstrates the difference in the abundance of immune cell infiltration between the UC group and the healthy control groups. The levels of T cells CD4 memory activated (P < 0.001), T cells follicular helper (P < 0.01), macrophages M0 (P < 0.001), macrophages M1 (P < 0.01), dendritic cells activated (P < 0.001), mast cells activated (P < 0.01), eosinophils (P < 0.05), and neutrophils (P < 0.01) levels were significantly higher in the UC group than healthy controls. On the other hand, healthy controls had showed higher levels of NK cells activated (P < 0.001), macrophages M2 (P < 0.001), and mast cells resting (P < 0.001). Moreover, T cells CD4 memory activated, T cells follicular helper, macrophages M1, and neutrophils showed positively correlated positive correlation with all the hub genes, a. Among them, neutrophils showed had the highest correlation (Figure 7C). Macrophages M2 showed negative correlationnegatively correlated with all the hub genes (P < 0.05). AIM2 had showed the highest negative correlation with macrophage M2, IL1B, and

TREM1 had depicted the highest positive correlation with Neutrophils. These results indicated that a severe immune imbalance occurred in the colon of UC patients, and the relationship between the different types of macrophages and UC was more notable.

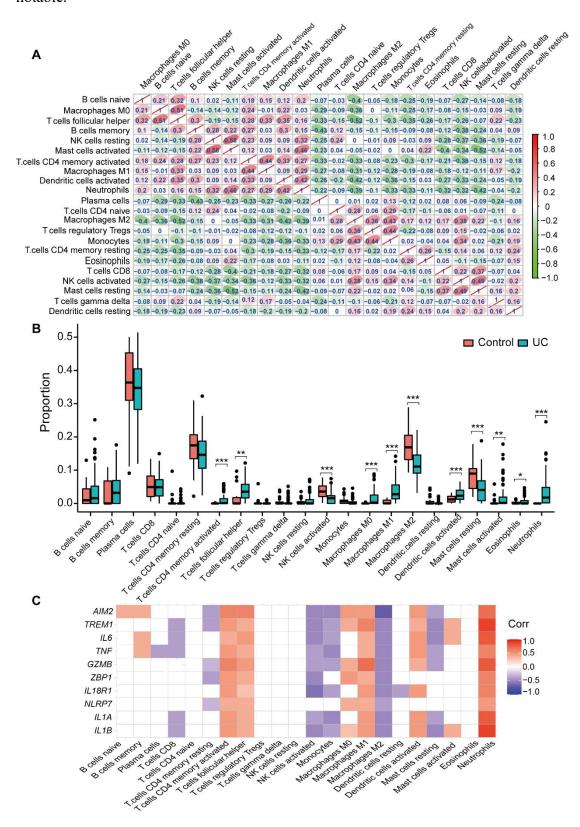


Figure 7. CIBERSORT probes the immune-infiltration landscape of UC. (A) Interrelationship The interrelationship between different infiltrating immune cells. The red to green indicates the change from high to low correlation. (B) Boxplot shows the difference in colonic immune cell infiltration colonic immune cell infiltration difference between the control and UC groups. Blue represents the UC group; red represents the Control group. (C) The correlation between the hub gene and the immune cell. Blank area means the Ssignificant level is higher than 0.05 *P < 0.05, **P < 0.01 and ***P < 0.001.

Immune Cell scRNA Analysis of Colonic Lamina Propria in Inflamed

UC

The GSE162335 dataset was used_utilized_for single_cell analysis. After filtering, 18,375 immune cells from the colonic lamina propria of inflamed UC patients were obtainedcollected. Expression_The expression_characteristics of the sample was are shown in Figure 8. We normalized the data and identified 3000 highly variable genes (HVG) using "VST", __, of which the top 10 HVG were shown in Figure 8D. Principal component analysis was performed and heatmap of the top 10 PCs with signature genes were are shown in the Figure S5. 37 Thirty-seven different was identified using the t-distributed stochastic neighbor embedding (t-SNE) method (Figure 8E), and the marker gene for each cell cluster werewere identified using the t-distributed stochastic neighbor embedding (t-SNE) method (Figure 8E), and the marker gene for each cell cluster is available from in Supplementary Table S6.

Seven immune cells were identified by using different annotation methods, including B-cells, CD4+ T-cells, CD8+ T-cells, Fibroblasts, Hematopoietic Stem Cells (HSC), Macrophages, and NK cells (Figure 9A). B cells and CD4+ T-cells were the most numerous, while macrophages could bwere significantly divided into two clusters.

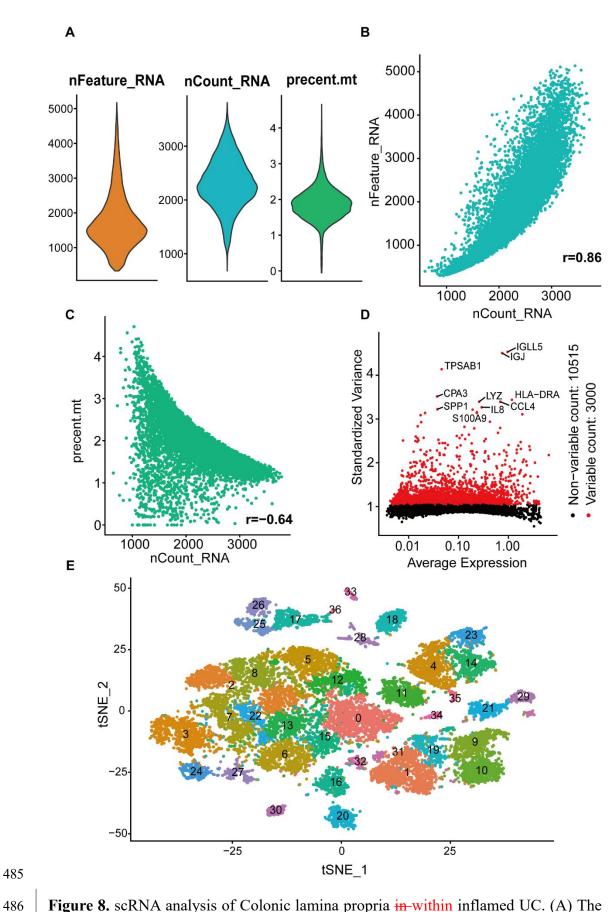


Figure 8. scRNA analysis of Colonic lamina propria in-within inflamed UC. (A) The genes (features), counts, and mitochondrial gene percentages of the sample. (B)

Correlation between genes and counts in <u>the</u> sample. (C) Correlation between genes and mitochondrial gene percentage in <u>the</u> sample. (D) The gene scatter plot with the top10 highly variable genes. The red dots represent <u>the</u> highly variable genes, and the black dots represent other genes. (E) t-SNE projection of 18,375 Immune cells. 18,375 Immune cells.

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Macrophages Highly Correlated with Pyroptosis

First, the relationship of between immune cells and PRGs was explored, and we found that NK cells and macrophages had high AUC values (Figure 9B-C). Combining with the results of the previous immune infiltration analysiprevious immune infiltration analysis results, we hypothesized that macrophages could be more closely related to pyroptosis. Therefore, we performed a KEGG analysis of DEGs in macrophages. The results showed DEGs were mainly enriched in inflammatory diseases and inflammatory signaling pathways, alsothat DEGs were mainly enriched in inflammatory diseases and signaling pathways and involved phagosome and lysosome (Figure S6). We also analyzed the expression distribution of 10 hub genes in—within various immune cells (Figure 10A). The results showed revealed that TNF was more widely distributed, and NLRP7 was less distributed among the seven immune cells. More interestingly, aAlmost all hub genes were expressed in macrophages, especially IL1B, as the more critical hub gene, wasthe more critical hub gene, strongly expressed (Figure 10B-C). This indicated that macrophages may could have a vital role in pyroptosis. Similarly, the high expression of GZMB in NK cells was also noteworthysignificant.

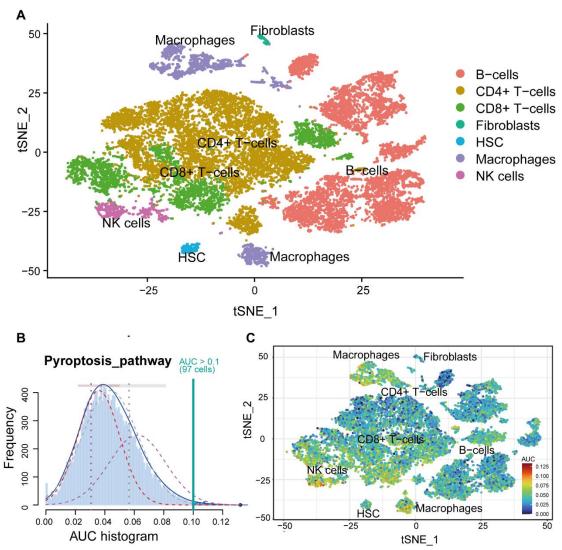


Figure 9. The Ccell annotation results and PRGs scores of colonic laminas propria immune cell types in inflamed UC. (A) The results of cell annotation with -Ddifferent Immune immune cell types are were colored with distinctively colorsed. (B) Score A score of 74 screened PRGs. The threshold was chosen as 0.10. (C) t-SNE plots of RPGs score in all the cell types. Macrophages and NK cells express more genes and exhibit higher AUC values.

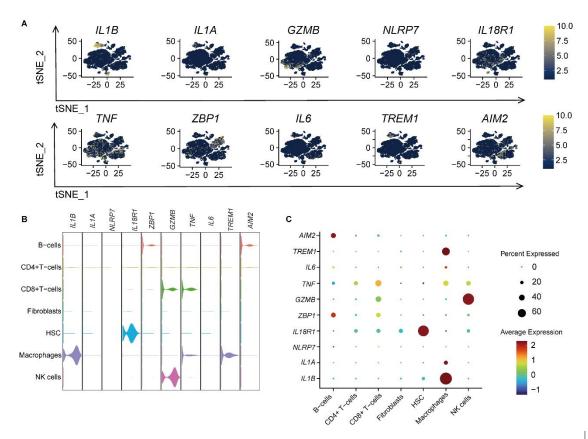


Figure 10. 10-Ten RPGs hub gene expression patterns in different immune cells. (A)

The t-SNE plot shows the distribution of hub genes. (B) The Violin plot showed the expression levels of hub genes. (C) Dot The dot plot showed depicted the expression patterns of hub genes.

Pseudotime Analysis and Enrichment Analysis Identification of

Pyroptosis Related-Macrophages Clusters in UC

To further reveal the relationship between macrophages and pyroptosis, wwe performed a pseudotime analysis of the five macrophage clusters, including cluster17, 20, 25, 26, and 28, to reveal the relationship between macrophages and pyroptosis. The analysis-research showed that almost all cells were projected onto two branches of a trunk, where clusters 17, 20, and 28 were located at one pole each, and clusters 25, and 26 were on the evolutionary line (Figure 11A, B). Based on the expression of pyroptosis-related hub genes, cluster17 and -20 were defined as IL1B+IL1A-IL6-, cluster 25 were—was defined—described as IL1B+IL1A+IL6+, cluster 26 were—was defined—designated as IL1B+IL1A+IL6-, and cluster 28 were—was defined as

IL1B-IL1A-IL6 cell group (Figure 11C). The expression of 74 RPGs in five cellular clusters was evaluated assessed (Figure 11D). Almost all the RPGs were not expressed in cluster 28, which also predicted that cluster 28 was probably not related to pyroptosis. Clusters 17, and 20 had similar hub gene expression patterns, but cluster 17 expressed more RPGs than cluster 20. To determine the molecular characteristics of the macrophages related with pyroptosis, GO enrichment analysis of unique DEGs from each cluster were was preformed performed to determine the molecular characteristics of the macrophages related to pyroptosis (Supplementary Table S7). Clusters 17, 20, 25, and 26 identified 33, 120, 96, and 99 DEGs (Figure 11E). GO terms of BP were are shown in Figure S7. Enrichment analysis of cluster17 indicated this cluster was involved in a number ofseveral regulatory processes, including the positive regulation of immune response, regulation of cell adhesion, positive regulation of cell adhesion regulation, and the immune system process (Figure S7A). Therefore, we named cluster17 as "Immunomodulatory macrophages"..." The GO terms in cluster 20 focused on the negative regulatory processes of cells, and cluster 20 was named termed ""negative regulation macrophages"_"(Figure S7B). The trajectory analysis of cluster25 showed revealed that it was located between cluster 17, and 20, and enrichment analysis showed that it was mainly focused on the response process to external stimuli and immunity, <u>; so thus, it was named as ""response macrophages" (Figure S7C).</u> Finally, cluster26 was named as a ""transporter and secretory macrophages" (Figure **S7D**). These results suggested indicated that macrophages related to pyroptosis may could be classified into four types and perform different functions in the colonic lamina propria of inflamed UC patients. Therefore, Eexploring the relationship between macrophages and pyroptosis may contribute facilitateto a better understanding of the occurrence of pyroptosis occurrence in UC.

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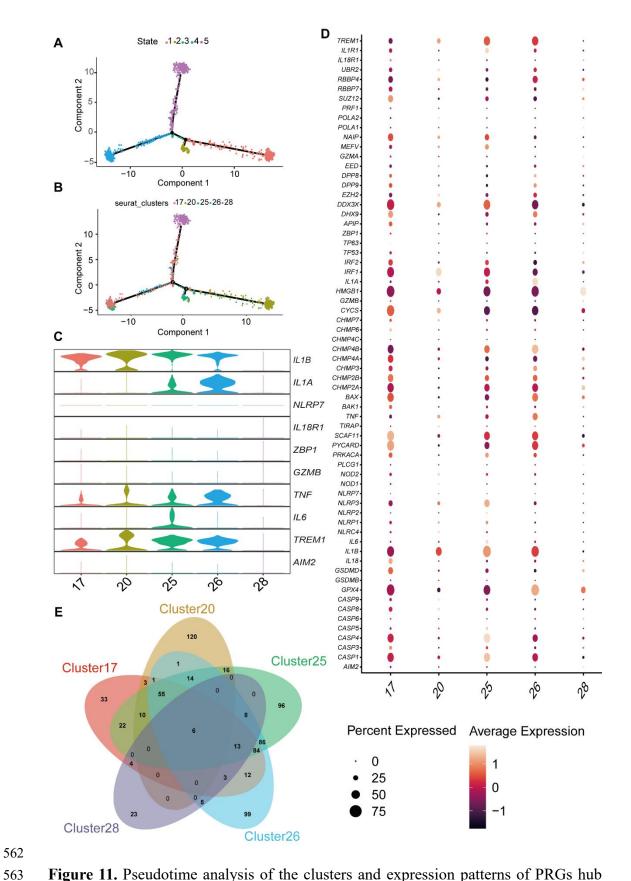


Figure 11. Pseudotime analysis of the clusters and expression patterns of PRGs hub genes in among different macrophage clusters. (A, B) Monocle pseudotime trajectory shows the progression of macrophages. (C) The Violin plot demonstrates the

expression levels of hub genes in five cluster macrophages. (D) Dot-The dot plot shows the expression pattern of 74 RPGs in five cluster macrophages. (E) The venn-diagram shows-depicts the DEGs of within each cluster macrophage.

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Animal experiments to verify Hub gene expression levels

The DSS-induced acute colitis was used utilized to verify the gene expression of hub genes (Figure 12A). The results show-indicate that DSS-induced colitis in mice with bloody stools, which was relieved by using 5-ASA treatment (Figure 12B). DSS-induced colitis caused weight loss and shorter colonic length in mice, and 5-ASA could slow suppress down the weight loss and colon shortening (Figure 12C-D). The symptoms that commonly occurringred during UC, such asincluding bloody stools and shortening of the colon, are evidence of increased enhanced abnormal death of IECs. To further observe the damage of intestinal mucosa during UC, wWe performed histological staining and examination of colon sections of mice to observe the damage of intestinal mucosa during UC.,—and wCompared with the NC group, we found a series of features such as disorganized mucosal structure, inflammatory cell infiltration, crypt detachment, and IECs death in colon sections of mice in the DSSe observed a series of features such as disorganized mucosal structure, inflammatory cell infiltration, crypt detachment and IECs death in colon sections of mice in the DSS group compared with the NC group. In contrast, 5-ASA treatment showed a protective effect on colonic mucosal structures (Figure 12E). All of tThereis is strong evidence that UC is highly correlated with the death of IECs, disruption of the intestinal barrier, and that intestinal barrier disruption, and therapeutic drugs can alleviate these symptoms. Consistently Therefore, inflammatory factor (TNF-α) in serum of DSS-DSS-induced mice were was increased, while 5-ASA intervention reduced decreased the elevation of TNF-α (Figure 12F). As shown in Figure 12G-O shows, that the relative expression levels of hub genes in DSS-induced colitis mice were are consistent with bioinformatics analysis. The expression of most genes was elevated in the DSS group. Genes with inconsistent expression maycould

be due to the species differences between mice and humans; *NLRP7* has been shown to beis associated with intestinal diseases such as colon cancer (33), but only a human-specific gene is that not found in rodents (34). *ZBP1*, a key-critical innate sensor that recognizes and binds Z-RNA structures produced mainly by various viruses, triggers different forms of cell death (35), yet. However, microorganisms with such structures may could be lacking in the pathogenic factors of DSS-induced colitis mice (36). A further findingoutcome was that 5-ASA significantly downregulated only one gene, *IL1B*, and had nowithout any significant effect on other genes; It suggesting suggests that there may be no relationship between the relief of colitis by 5-ASA and the regulation of pyroptosis, consistent with our previous prediction findings.

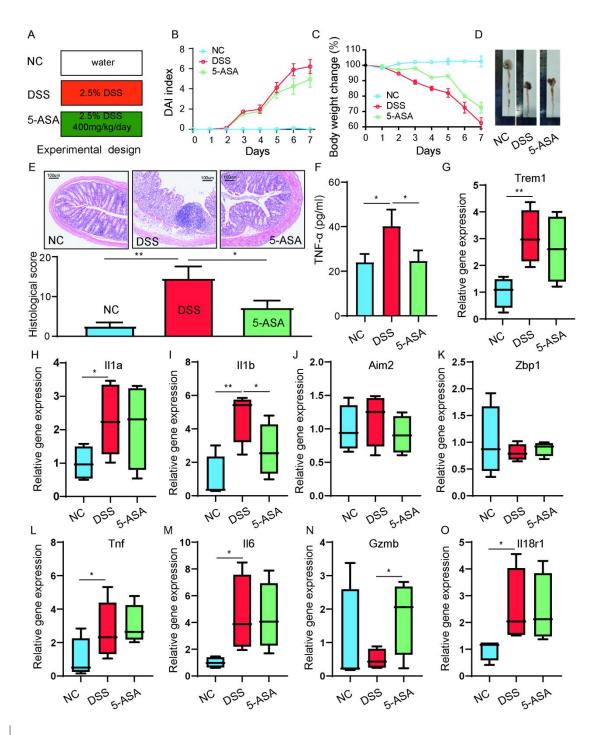


Figure 12. Animal experiments to verify the Hub gene expression levels. (A) Experimental design. (B) DAI index. (C) Body weight change. (D) Length of colons. (E) H&E-stained colon tissue. (F) Serum TNF- α was determined using with ELISA. The relative gene expression levels of *Trem1* (G), *Il1a* (H), *Il1b* (I), *Aim2* (J), *Zbp1* (K), *Tnf* (L), *Il6* (M), *Gzmb* (N), and *Il18r1* (O) were determined using with RT-PCR. *P < 0.05, **P < 0.01 and ***P < 0.001.

DISCUSSION

618	UC is a common chronic inflammatory intestinal disease of the intestine. The life
619	quality of UC patients is seriously severely impaired due to its recurrent and incurable
620	characteristics (37). At present, although some progress has been made in the
621	treatment of Ithough some progress has been made in treating UC, further mitigation
622	of recurrence and eradication remains a global challenge. Pyroptosis, as-a new form of
623	cell death discovered in recent years recently, has shown great promise in UC (38). For
624	example, One has beena reported study claims that DSS potentiates NLRP3
625	inflammasome activation by modulating regulating the KCa3.1 potassium channel in
626	a mouse model of colitis, This provides direct evidence for the role of pyroptosis in
627	UC (39). In another study, the authors <u>discovered</u> that
628	Trans-10-Hydroxy-2-Decenoic Decanoic Aacid could treat UC by inhibiting
629	pyroptosis, while also enhancing the barrier function of the colon (40). However,
630	there is still a lack of reports on concerning the detailed mechanism of Pyroptosis in
631	UC. Our study attempts to explore the role of pyroptosis in UC by identification
632	of explores the role of pyroptosis in UC by identifying the relevant hub genes and
633	immune cells, t. This will provide a new perspective for on the pathogenesis and
634	treatment of UC.
635	A total of 1,247 DEGs and 1,186 module genes were obtained by using differential
636	analysis—and WGCNA analysisanalyses. The results of the enrichment
637	analysienrichment analysis s showedrevealed that these genes were mainly primarily
638	involved in inflammatory pathways related to pyroptosis-related
639	inflammatory pathways, including NF-kappa B signaling pathway, NOD-like receptor
640	signaling pathway, Toll-like receptor signaling pathway, and TNF signaling pathway.
641	This suggested that there is a close relationship between UC and pyroptosis.
642	Subsequently, we identified 10 pyroptosis-related hub genes (AIM2, IL6, IL1B,
643	NLRP7, TNF, IL1A, IL18R1, ZBP1, GZMB, TREM1) were identified. AIM2 is a

cytoplasmic sensor of double-stranded DNA from pathogens or damaged organelles. It recruits ASC and caspase-1 to form-develop the AIM2 inflammasome, which activatesing caspase-1 and the processing of processing IL-1\beta and IL-18, causing leading to pyroptosis (41). Innate immune sensor Z-DNA binding protein 1 (ZBPI) asis the apical sensor of fungal infection and regulates controls the NLRP3 inflammasome to participate in pyroptosis (42). Notably, __iIt has been reported that AIM2 in complex with pyrin and ZBP1 can be engaged in pyroptosis, apoptosis, and necroptosis simultaneously simultaneously engage in pyroptosis, apoptosis, and necroptosis, this which is probably a new novel research direction (43). IL1B, IL1A, IL6, and TNF are important pro-inflammatory cytokines in pyroptosis (44). IL18R1 is the receptor for pro-inflammatory cytokine IL18 and is responsible for the binding of IL18 (45). The interleukin-1 cytokine family plays a key crucial role in maintaining intestinal barrier integrity and inflammatory responses, but their specific functions remains are controversial. More and more Various studies suggest-indicate that they playtheir a dual role in inflammation and homeostasis in vivo (46). GZMB catalyzes the cleavage of gasdermin-E (GSDME) and releases the pore-forming fraction of GSDME, thereby triggering pyroptosis (47). TREM1 amplifies inflammatory signals involving Toll-like receptors and NOD-like receptors, and in this waywhich mediates the exacerbation of pyroptosis (48). NLRP7 is related associated withto innate immune signaling and affects macrophage polarization, but its exact role remains is unclearsomewhat controversial. With In recent the research in recent years, different pathways of pyroptosis have been identified (49). However, the specific mechanism of pyroptosis in UC remains poorly deciphered understood. Based on the 10 hub genes identified, we hypothesized that there may could be two main pyroptosis pathways in UC., Firstly the AIM2 inflammasome-mediated caspase-1-dependent classical pyroptosis pathway was formed developed after dsDNA recognition by AIM2 (50), and the caspase-independent pyroptosis pathway was mediated controlled by GZMB (51). AIM2, TREM1, LI1B, GZMB, IL18R1, ZBP1 were identified as and ZBP1 were diagnostic biomarkers. The results of ROC analysis indicated that IL1B has the greatest-most significant potential as a biomarker.

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We observed the expression patterns of hub genes in within the lesional/nonlesional, active/inactive/control groups by different datasets, and aAll the results showed significant differences except for inactive/control groups, Aand among the inactive/control groups, IL1B became the only hub gene with significant differences. IL-1β is a powerful potent pro-inflammatory factor that plays a key-crucial role in pyroptosis. The synthesis of pro-IL-1 β is stimulated in response to an inflammatory signal, and t. Then when a large number of, when many inflammatory vesicles activate caspase 1, pro-IL-1β is cleaved to form synthesize IL-1β, and subsequently, caspase-1 cleaves the GSDMD to form-create the 22 kDa C-terminus (C-GSDMD) and 31 kDa N-terminal (N-GSDMD). N-GSDMD completes by forming forming pores in the plasma membrane and mitochondria to release IL-1\beta and IL-18, t. Thus, IL1B is a critical part of pyroptosis. There have been many reports Many reports have revealed that targeting IL1B can effectively alleviate UC (52). Combined with the previous ROC analysis analyses, we speculate that IL1B is the most crucial gene in the pyroptosis of UC. A characteristic feature of UC is that 80-90% of patients have alternated between active and inactive intervals (53). The significant difference between the active and inactive phases of the hub gene may could indicated the increased elevated expression of the hub gene in patients, resulting causingin damage to the intestinal mucosa triggered by pyroptosis in the IECs and an increase in pro-inflammatory factors, causing leading to recurrent disease (54). And Thus, focusing on the development of pyroptosis development is probably a new idea way of thinking to avoiding the recurrence of UC and an important crucial direction for future treatment direction. Moreover, we also explored the effects of some common UC drugs on hub genes to better reveal the significance of pyroptosis in UCreveal the significance of pyroptosis in UC better. 5-ASA and biologics are common UC drugs but showed showed different effects on the hub gene. 5-ASA reduced the expression levels of IL1B, IL1A, and $IL6_7$ but had no effect ondid not affect other hub genes. Biologics such as IFX₇ and VDZ significantly reduced decreased all hub genes. For 30 years, 5-ASA has

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been the treatment of choice for against mild UC (55). However, a recent study has showned that compared to biologic agents such as IFX and VDZ, 5-ASA was not helpful in the treatment ofdid not help treat patients with moderate to severe UC, especially in controlling inflammation (56). Therefore, we speculated that the progression of UC disease may could be related associated withto the degree of pyroptosis, and patients with severe UC may be subjected to a higher degree of intestinal mucosa invasion by due to pyroptosis. , and tTherefore, 5-ASA is not effective in treating patients with having severe UC. In conclusion, we verified that both UC drugs had a different effect on the pyroptosis-related hub gene and reduced decreased its expression while alleviating UC. The link between pyroptosis and the progression of UC deserves more-in-depth exploration, especially, the expression of relevant genes/proteins need. The expression of relevant genes/proteins needs to be verified by using the tissue of from the patients. By rReviewing the research literature using with UC clinical samples, we found that many previous studies reported the test results of the hub genes in patient tissues. For example instance, IL-1β in serum and tissues of UC patients was measured by ELISA, and the results showed depicted that the expression of IL-1β in UC patients was significantly higher than that-in healthy controls (57, 58). In addition, the flow cytometry analysis showed revealed that the number of cells expressing GZMB and TREM1 in the intestinal mucosa of UC patients significantly increased significantly (59, 60). Using western blot analysis, the expression of AIM2 in active UC is higher than that in inactive UC(28). These are consistent with the results of from our bioinformatics analysis analyses. To further explore the dysregulation of inflammatory cells in UC, an immune Infiltration analysis was performed An immune Infiltration analysis was performed to explore the dysregulation of inflammatory cells in UC. The results showed that, immune cells were severely dysregulated in within the colonic tissue of UC patients. Neutrophils, Macrophages M0, M1, Dendritic cells activated, T cells CD4 memory resting, and T cells follicular helper were positively correlated associated with hub genes. The results of sSingle-cell analysis showed that 9 nine hub genes were distributed in macrophages except for NLRP7, and IL1B was strongly expressed. In

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addition, macrophages were identified by AUCell evaluation to show deliver the highest degree of response to the pyroptosis gene set. By increasing enhancing the resolution, we identified 5-five clusters of macrophages. Among them, fFour clusters are were closely related associated withto pyroptosis and may perform different biological functions. The Ppro-inflammatory and anti-inflammatory functions of macrophages play have a key crucial role in the development of UC, and many drugs alleviate UC by targeting the macrophages (61, 62). However, the interaction between different immune cell types still needsrequires further exploration. More importantly, the alleviation of UC by inhibiting macrophage pyroptosis has been reported described in recent years (15). It was also shown that the Lactic Acid-Producing Probiotic Saccharomyces cerevisiae inhibits the hyperactivation of the NLRP3 inflammasome and downstream caspase-1 pathway in-among macrophages, thereby inhibiting the process of pyroptosis in macrophages. When macrophage pyroptosis was inhibited, the colonic mucosal barrier was strengthened, the immune response in the intestine was decreased, whiledecreased, and histological damage was also restored. Abnormal death of IECs leads to causes damage of to the intestinal mucosa, and this structural weakening of the intestinal mucosal barrier is an early event in the UC pathogenesis of UC (54). Pyroptosis is an important necessary forms of death for IECs, and a. A series of pro-inflammatory factors secreted during pyroptosis, such asincluding IL-18, and IL-1\beta, can disrupt the integrity of the intestinal mucosal barrier (63). Therefore, excessive death of IECs and the secretion of pro-inflammatory factors caused due toby excessive activation of pyroptosis are important essential factors in disrupting the intestinal barrier, which may could be a critical mechanism in the UC pathogenesis of UC. After the disruption of the intestinal barrier, microorganisms in the gut invade into the intestine causing a series of immuneresponses while macrophages themselvdisrupting the intestinal barrier, microorganisms in the gut invade the intestine, causing a series of immune responses while macrophages undergo pyroptosis. The pro-inflammatory factors released in pyroptosis may could worsen the immune response in the intestine and lead to more

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death of IECs, which further disrupts the intestinal barrier and tfurther disrupting the intestinal barrier. This may be one of the reasons why UC is incurable and recurrent.

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CONCLUSION

Ten pyroptosis-pyroptosis-related hub genes in UC were identified, and the expression pattern of hub genes was validated. The effect of the existing UC treatment drugs on the gene expression of hub genes were was explored, and IL1B was identified as a the predictor for drug response and marker for the active state of UC. Combining single cell analysis and immune infiltration, we determined identified that macrophages as the most relevant immune cell type in the UC progression of UC. Our study explored the molecular mechanisms of the pyroptosis process in UC and. We proposed that the crosstalk regarding pyroptosis between macrophages and IECs regarding pyroptosis is probablycould be responsible for the incurability and recurrence of UC. The IL1B-macrophage-pyroptosis relationship chain provides a new perspective for on the pathogenesis and treatment of UC.

Supplementary data

Supplementary data are available online.

Acknowledgement

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Author contributions

- N.H and S.L conceived the study and performed data analysis. K.C. and S.S. collected
- data and performed data analysis. S.Y and L.C performed animal experiments and
- related tests. N.H and S.L supervised the project. K.C., S.S., S.L and N.H. wrote the
- 790 manuscript. All authors approved the final manuscript.

Data Availability Statement

- The datasets presented in this study can be found in online repositories. These datasets
- were freely downloaded from GEO database.

794 Code Availability Statements

- 795 The code for data analysis have been uploaded to Github
- 796 (https://github.com/illusion621/Identification-and-Exploration-of-Pharmacological-py
- 797 roptosis-related-Biomarkers-of-Ulcerative-Coliti

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References

- 1. Ordás I, Eckmann L, Talamini M, Baumgart DC, Sandborn WJ. Ulcerative Colitis.
- 801 Lancet (London, England) (2012) 380(9853):1606-19. Epub 2012/08/24. doi:
- 802 10.1016/s0140-6736(12)60150-0.
- 2. Ungaro R, Mehandru S, Allen PB, Peyrin-Biroulet L, Colombel JF. Ulcerative
- 804 Colitis. Lancet (London, England) (2017) 389(10080):1756-70. Epub 2016/12/05. doi:
- 805 10.1016/s0140-6736(16)32126-2.
- 806 3. Ozaki R, Kobayashi T, Okabayashi S, Nakano M, Morinaga S, Hara A, et al.
- 807 Histological Risk Factors to Predict Clinical Relapse in Ulcerative Colitis with
- 808 Endoscopically Normal Mucosa. Journal of Crohn's & colitis (2018) 12(11):1288-94.
- 809 Epub 2018/06/26. doi: 10.1093/ecco-jcc/jjy092.
- 4. Xavier RJ, Podolsky DK. Unravelling the Pathogenesis of Inflammatory Bowel
- 811 Disease. Nature (2007) 448(7152):427-34. Epub 2007/07/27. doi:
- 812 10.1038/nature06005.
- 813 5. Rozich JJ, Dulai PS, Fumery M, Sandborn WJ, Singh S. Progression of Elderly
- 814 Onset Inflammatory Bowel Diseases: A Systematic Review and Meta-Analysis of
- 815 Population-Based Cohort Studies. Clin Gastroenterol Hepatol (2020)
- 816 18(11):2437-47.e6. Epub 2020/03/07. doi: 10.1016/j.cgh.2020.02.048.
- 817 6. Patankar JV, Becker C. Cell Death in the Gut Epithelium and Implications for
- 818 Chronic Inflammation. Nature reviews Gastroenterology & hepatology (2020)
- 819 17(9):543-56. Epub 2020/07/12. doi: 10.1038/s41575-020-0326-4.

- 7. Okumura R, Takeda K. Roles of Intestinal Epithelial Cells in the Maintenance of
- 821 Gut Homeostasis. Experimental & molecular medicine (2017) 49(5):e338. Epub
- 822 2017/05/27. doi: 10.1038/emm.2017.20.
- 823 8. Iwamoto M, Koji T, Makiyama K, Kobayashi N, Nakane PK. Apoptosis of Crypt
- Epithelial Cells in Ulcerative Colitis. The Journal of pathology (1996) 180(2):152-9.
- 825 Epub 1996/10/01. doi:
- 826 10.1002/(sici)1096-9896(199610)180:2<152::Aid-path649>3.0.Co;2-y.
- 9. Baxt LA, Xavier RJ. Role of Autophagy in the Maintenance of Intestinal
- 828 Homeostasis. Gastroenterology (2015) 149(3):553-62. Epub 2015/07/15. doi:
- 829 10.1053/j.gastro.2015.06.046.
- 830 10. Kovacs SB, Miao EA. Gasdermins: Effectors of Pyroptosis. Trends in cell
- biology (2017) 27(9):673-84. Epub 2017/06/18. doi: 10.1016/j.tcb.2017.05.005.
- 11. Kayagaki N, Stowe IB, Lee BL, O'Rourke K, Anderson K, Warming S, et al.
- 833 Caspase-11 Cleaves Gasdermin D for Non-Canonical Inflammasome Signalling.
- Nature (2015) 526(7575):666-71. Epub 2015/09/17. doi: 10.1038/nature15541.
- 835 12. Wu C, Lu W, Zhang Y, Zhang G, Shi X, Hisada Y, et al. Inflammasome
- 836 Activation Triggers Blood Clotting and Host Death through Pyroptosis. Immunity
- 837 (2019) 50(6):1401-11.e4. Epub 2019/05/12. doi: 10.1016/j.immuni.2019.04.003.
- 838 13. Szekanecz Z, Szamosi S, Kovács GE, Kocsis E, Benkő S. The Nlrp3
- 839 Inflammasome Interleukin 1 Pathway as a Therapeutic Target in Gout. Archives of
- 840 biochemistry and biophysics (2019) 670:82-93. Epub 2019/02/03. doi:
- 841 10.1016/j.abb.2019.01.031.
- 14. Marchiando AM, Shen L, Graham WV, Edelblum KL, Duckworth CA, Guan Y, et
- al. The Epithelial Barrier Is Maintained by in Vivo Tight Junction Expansion During
- 844 Pathologic Intestinal Epithelial Shedding. Gastroenterology (2011)
- 845 140(4):1208-18.e1-2. Epub 2011/01/18. doi: 10.1053/j.gastro.2011.01.004.
- 15. Sun S, Xu X, Liang L, Wang X, Bai X, Zhu L, et al. Lactic Acid-Producing
- 847 Probiotic Saccharomyces Cerevisiae Attenuates Ulcerative Colitis Via Suppressing
- 848 Macrophage Pyroptosis and Modulating Gut Microbiota. Front Immunol (2021)
- 849 12:777665. Epub 2021/12/14. doi: 10.3389/fimmu.2021.777665.

- 850 16. Zhu J, Xu Y, Li Z, Liu S, Fu W, Wei Y. Interleukin-36β Exacerbates Dss-Induce
- Acute Colitis Via Inhibiting Foxp3(+) Regulatory T Cell Response and Increasing
- The Cell Response. International immunopharmacology (2022) 108:108762. Epub
- 853 2022/04/19. doi: 10.1016/j.intimp.2022.108762.
- 17. Ye Y, Dai Q, Qi H. A Novel Defined Pyroptosis-Related Gene Signature for
- Predicting the Prognosis of Ovarian Cancer. Cell death discovery (2021) 7(1):71.
- 856 Epub 2021/04/09. doi: 10.1038/s41420-021-00451-x.
- 857 18. Wei R, Li S, Yu G, Guan X, Liu H, Quan J, et al. Deciphering the
- 858 Pyroptosis-Related Prognostic Signature and Immune Cell Infiltration Characteristics
- 859 of Colon Cancer. Front Genet (2021) 12:755384. Epub 2021/10/30. doi:
- 860 10.3389/fgene.2021.755384.
- 19. Li K, Strauss R, Ouahed J, Chan D, Telesco SE, Shouval DS, et al. Molecular
- 862 Comparison of Adult and Pediatric Ulcerative Colitis Indicates Broad Similarity of
- 863 Molecular Pathways in Disease Tissue. Journal of pediatric gastroenterology and
- 864 nutrition (2018) 67(1):45-52. Epub 2018/02/06. doi:
- 865 10.1097/mpg.000000000001898.
- 20. Tang J, Kong D, Cui Q, Wang K, Zhang D, Gong Y, et al. Prognostic Genes of
- 867 Breast Cancer Identified by Gene Co-Expression Network Analysis. Front Oncol
- 868 (2018) 8:374. Epub 2018/09/27. doi: 10.3389/fonc.2018.00374.
- 21. Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, et al. Robust
- 870 Enumeration of Cell Subsets from Tissue Expression Profiles. Nature methods (2015)
- 871 12(5):453-7. Epub 2015/03/31. doi: 10.1038/nmeth.3337.
- 22. Devlin JC, Axelrad J, Hine AM, Chang S, Sarkar S, Lin JD, et al. Single-Cell
- 873 Transcriptional Survey of Ileal-Anal Pouch Immune Cells from Ulcerative Colitis
- 874 Patients. Gastroenterology (2021) 160(5):1679-93. Epub 2020/12/29. doi:
- 875 10.1053/j.gastro.2020.12.030.
- 23. Hao Y, Hao S, Andersen-Nissen E, Mauck WM, 3rd, Zheng S, Butler A, et al.
- Integrated Analysis of Multimodal Single-Cell Data. Cell (2021) 184(13):3573-87.e29.
- 878 Epub 2021/06/02. doi: 10.1016/j.cell.2021.04.048.
- 879 24. Han X, Zhou Z, Fei L, Sun H, Wang R, Chen Y, et al. Construction of a Human

- 880 Cell Landscape at Single-Cell Level. Nature (2020) 581(7808):303-9. Epub
- 881 2020/03/28. doi: 10.1038/s41586-020-2157-4.
- 25. Lu Y, Li K, Hu Y, Wang X. Expression of Immune Related Genes and Possible
- Regulatory Mechanisms in Alzheimer's Disease. Front Immunol (2021) 12:768966.
- 884 Epub 2021/11/23. doi: 10.3389/fimmu.2021.768966.
- 885 26. Wang L, He T, Liu J, Tai J, Wang B, Zhang L, et al. Revealing the Immune
- 886 Infiltration Landscape and Identifying Diagnostic Biomarkers for Lumbar Disc
- 887 Herniation. Front Immunol (2021) 12:666355. Epub 2021/06/15. doi:
- 888 10.3389/fimmu.2021.666355.
- 889 27. Sandborn WJ, Feagan BG, Marano C, Zhang H, Strauss R, Johanns J, et al.
- 890 Subcutaneous Golimumab Induces Clinical Response and Remission in Patients with
- 891 Moderate-to-Severe Ulcerative Colitis. Gastroenterology (2014) 146(1):85-95; quiz
- 892 e14-5. Epub 2013/06/06. doi: 10.1053/j.gastro.2013.05.048.
- 28. Vanhove W, Peeters PM, Staelens D, Schraenen A, Van der Goten J, Cleynen I, et
- al. Strong Upregulation of Aim2 and Ifi16 Inflammasomes in the Mucosa of Patients
- 895 with Active Inflammatory Bowel Disease. Inflammatory bowel diseases (2015)
- 896 21(11):2673-82. Epub 2015/08/28. doi: 10.1097/mib.000000000000535.
- 897 29. Yuan C. Il-33 in Autoimmunity; Possible Therapeutic Target. International
- 898 immunopharmacology (2022) 108:108887. Epub 2022/06/23. doi:
- 899 10.1016/j.intimp.2022.108887.
- 30. Arijs I, De Hertogh G, Lemmens B, Van Lommel L, de Bruyn M, Vanhove W, et
- 901 al. Effect of Vedolizumab (Anti-A4β7-Integrin) Therapy on Histological Healing and
- 902 Mucosal Gene Expression in Patients with Uc. Gut (2018) 67(1):43-52. Epub
- 903 2016/11/02. doi: 10.1136/gutjnl-2016-312293.
- 31. Le Berre C, Roda G, Nedeljkovic Protic M, Danese S, Peyrin-Biroulet L. Modern
- 905 Use of 5-Aminosalicylic Acid Compounds for Ulcerative Colitis. Expert opinion on
- 906 biological therapy (2020) 20(4):363-78. Epub 2019/09/10. doi:
- 907 10.1080/14712598.2019.1666101.
- 908 32. Bhattacharya A, Osterman MT. Biologic Therapy for Ulcerative Colitis.
- Gastroenterology clinics of North America (2020) 49(4):717-29. Epub 2020/10/31.

- 910 doi: 10.1016/j.gtc.2020.08.002.
- 33. Li B, Qi ZP, He DL, Chen ZH, Liu JY, Wong MW, et al. Nlrp7 Deubiquitination
- 912 by Usp10 Promotes Tumor Progression and Tumor-Associated Macrophage
- Polarization in Colorectal Cancer. J Exp Clin Cancer Res (2021) 40(1):126. Epub
- 914 2021/04/12. doi: 10.1186/s13046-021-01920-y.
- 915 34. Amoushahi M, Sunde L, Lykke-Hartmann K. The Pivotal Roles of the Nod-Like
- Receptors with a Pyd Domain, Nlrps, in Oocytes and Early Embryo Development[†].
- 917 Biology of reproduction (2019) 101(2):284-96. Epub 2019/06/16. doi:
- 918 10.1093/biolre/ioz098.
- 919 35. Yang D, Liang Y, Zhao S, Ding Y, Zhuang Q, Shi Q, et al. Zbp1 Mediates
- 920 Interferon-Induced Necroptosis. Cellular & molecular immunology (2020)
- 921 17(4):356-68. Epub 2019/05/12. doi: 10.1038/s41423-019-0237-x.
- 36. Balachandran S, Mocarski ES. Viral Z-Rna Triggers Zbp1-Dependent Cell Death.
- 923 Current opinion in virology (2021) 51:134-40. Epub 2021/10/25. doi:
- 924 10.1016/j.coviro.2021.10.004.
- 925 37. Cosnes J, Gower-Rousseau C, Seksik P, Cortot A. Epidemiology and Natural
- 926 History of Inflammatory Bowel Diseases. Gastroenterology (2011) 140(6):1785-94.
- 927 Epub 2011/05/03. doi: 10.1053/j.gastro.2011.01.055.
- 928 38. Wu G, Zhang D, Yang L, Wu Q, Yuan L. Microrna-200c-5p Targets Nima Related
- 929 Kinase 7 (Nek7) to Inhibit Nod-Like Receptor 3 (Nlrp3) Inflammasome Activation,
- 930 Mode-K Cell Pyroptosis, and Inflammatory Bowel Disease in Mice. Molecular
- 931 immunology (2022) 146:57-68. Epub 2022/04/22. doi:
- 932 10.1016/j.molimm.2022.03.121.
- 933 39. Zeng B, Huang Y, Chen S, Xu R, Xu L, Qiu J, et al. Dextran Sodium Sulfate
- Potentiates Nlrp3 Inflammasome Activation by Modulating the Kca3.1 Potassium
- Channel in a Mouse Model of Colitis. Cellular & molecular immunology (2022)
- 936 19(8):925-43. Epub 2022/07/08. doi: 10.1038/s41423-022-00891-0.
- 937 40. Huang S, Tao R, Zhou J, Qian L, Wu J. Trans-10-Hydroxy-2-Decenoic Acid
- 938 Alleviates Dextran Sulfate Sodium-Induced Colitis in Mice Via Regulating the
- 939 Inflammasome-Mediated Pyroptotic Pathway and Enhancing Colonic Barrier

- 940 Function. Molecular nutrition & food research (2022) 66(12):e2100821. Epub
- 941 2022/04/05. doi: 10.1002/mnfr.202100821.
- 942 41. Wang B, Tian Y, Yin Q. Aim2 Inflammasome Assembly and Signaling. Adv Exp
- 943 Med Biol (2019) 1172:143-55. Epub 2019/10/20. doi: 10.1007/978-981-13-9367-9 7.
- 42. Kuriakose T, Man SM, Malireddi RK, Karki R, Kesavardhana S, Place DE, et al.
- 25 Zbp1/Dai Is an Innate Sensor of Influenza Virus Triggering the Nlrp3 Inflammasome
- and Programmed Cell Death Pathways. Science immunology (2016) 1(2). Epub
- 947 2016/12/06. doi: 10.1126/sciimmunol.aag2045.
- 948 43. Lee S, Karki R, Wang Y, Nguyen LN, Kalathur RC, Kanneganti TD. Aim2 Forms
- a Complex with Pyrin and Zbp1 to Drive Panoptosis and Host Defence. Nature (2021)
- 950 597(7876):415-9. Epub 2021/09/03. doi: 10.1038/s41586-021-03875-8.
- 951 44. Li LL, Dai B, Sun YH, Zhang TT. The Activation of Il-17 Signaling Pathway
- Promotes Pyroptosis in Pneumonia-Induced Sepsis. Ann Transl Med (2020) 8(11):674.
- 953 Epub 2020/07/04. doi: 10.21037/atm-19-1739.
- 45. Sergi B, Penttila I. Interleukin 18 Receptor. J Biol Regul Homeost Agents (2004)
- 955 18(1):55-61. Epub 2004/08/25.
- 46. Kanai T, Kamada N, Hisamatsu T. Clinical Strategies for the Blockade of Il-18 in
- 957 Inflammatory Bowel Diseases. Current drug targets (2013) 14(12):1392-9. Epub
- 958 2013/05/09. doi: 10.2174/13894501113149990006.
- 959 47. Liu Y, Fang Y, Chen X, Wang Z, Liang X, Zhang T, et al. Gasdermin E-Mediated
- 960 Target Cell Pyroptosis by Car T Cells Triggers Cytokine Release Syndrome. Science
- 961 immunology (2020) 5(43). Epub 2020/01/19. doi: 10.1126/sciimmunol.aax7969.
- 962 48. Bouchon A, Facchetti F, Weigand MA, Colonna M. Trem-1 Amplifies
- 963 Inflammation and Is a Crucial Mediator of Septic Shock. Nature (2001)
- 964 410(6832):1103-7. Epub 2001/04/27. doi: 10.1038/35074114.
- 965 49. Zhai Z, Yang F, Xu W, Han J, Luo G, Li Y, et al. Attenuation of Rheumatoid
- 966 Arthritis through the Inhibition of Tumor Necrosis Factor-Induced Caspase
- 3/Gasdermin E-Mediated Pyroptosis. Arthritis & rheumatology (Hoboken, NJ) (2022)
- 968 74(3):427-40. Epub 2021/09/05. doi: 10.1002/art.41963.
- 969 50. Sharma BR, Karki R, Kanneganti TD. Role of Aim2 Inflammasome in

- 970 Inflammatory Diseases, Cancer and Infection. European journal of immunology (2019)
- 971 49(11):1998-2011. Epub 2019/08/03. doi: 10.1002/eji.201848070.
- 51. Zhang Z, Zhang Y, Xia S, Kong Q, Li S, Liu X, et al. Gasdermin E Suppresses
- 973 Tumour Growth by Activating Anti-Tumour Immunity. Nature (2020)
- 974 579(7799):415-20. Epub 2020/03/20. doi: 10.1038/s41586-020-2071-9.
- 975 52. Yin Q, Pi X, Jiang Y, Ren G, Liu Z, Liu H, et al. An Immuno-Blocking Agent
- 976 Targeting Il-1β and Il-17a Reduces the Lesion of Dss-Induced Ulcerative Colitis in
- 977 Mice. Inflammation (2021) 44(5):1724-36. Epub 2021/04/21. doi:
- 978 10.1007/s10753-021-01449-4.
- 979 53. Blonski W, Buchner AM, Lichtenstein GR. Treatment of Ulcerative Colitis.
- 980 Current opinion in gastroenterology (2014) 30(1):84-96. Epub 2013/11/29. doi:
- 981 10.1097/mog.0000000000000031.
- 982 54. van der Post S, Jabbar KS, Birchenough G, Arike L, Akhtar N, Sjovall H, et al.
- 983 Structural Weakening of the Colonic Mucus Barrier Is an Early Event in Ulcerative
- 984 Colitis Pathogenesis. Gut (2019) 68(12):2142-51. Epub 2019/03/28. doi:
- 985 10.1136/gutinl-2018-317571.
- 986 55. Cottone M, Renna S, Modesto I, Orlando A. Is 5-Asa Still the Treatment of
- 987 Choice for Ulcerative Colitis? Current drug targets (2011) 12(10):1396-405. Epub
- 988 2011/04/07. doi: 10.2174/138945011796818126.
- 989 56. Balram B, Joshi H, Wong K, Kroeker KI, Dieleman LA, Halloran BP, et al.
- 990 Concomitant 5-Aminosalicylate Therapy in Moderate-to-Severe Ulcerative Colitis
- Patients Escalated to Infliximab Is Not Beneficial. Digestive diseases and sciences
- 992 (2021) 66(11):3985-92. Epub 2020/11/14. doi: 10.1007/s10620-020-06704-6.
- 993 57. Feng J, Zhu Y, Chen L, Wang M. Clinical Significance of Microrna-146a in
- Patients with Ulcerative Colitis. Annals of clinical and laboratory science (2020)
- 995 50(4):463-7. Epub 2020/08/23.
- 58. Carter MJ, Jones S, Camp NJ, Cox A, Mee J, Warren B, et al. Functional
- 997 Correlates of the Interleukin-1 Receptor Antagonist Gene Polymorphism in the
- 998 Colonic Mucosa in Ulcerative Colitis. Genes and immunity (2004) 5(1):8-15. Epub
- 999 2004/01/22. doi: 10.1038/sj.gene.6364032.

- 1000 59. Cupi ML, Sarra M, Marafini I, Monteleone I, Franzè E, Ortenzi A, et al. Plasma
- 1001 Cells in the Mucosa of Patients with Inflammatory Bowel Disease Produce Granzyme
- 1002 B and Possess Cytotoxic Activities. Journal of immunology (Baltimore, Md : 1950)
- 1003 (2014) 192(12):6083-91. Epub 2014/05/20. doi: 10.4049/jimmunol.1302238.
- 1004 60. Brynjolfsson SF, Magnusson MK, Kong PL, Jensen T, Kuijper JL, Håkansson K,
- et al. An Antibody against Triggering Receptor Expressed on Myeloid Cells 1 (Trem-1)
- Dampens Proinflammatory Cytokine Secretion by Lamina Propria Cells from Patients
- 1007 with Ibd. Inflammatory bowel diseases (2016) 22(8):1803-11. Epub 2016/06/01. doi:
- 1008 10.1097/mib.0000000000000822.
- 1009 61. Wu MM, Wang QM, Huang BY, Mai CT, Wang CL, Wang TT, et al. Dioscin
- 1010 Ameliorates Murine Ulcerative Colitis by Regulating Macrophage Polarization.
- 1011 Pharmacological research (2021) 172:105796. Epub 2021/08/04. doi:
- 1012 10.1016/j.phrs.2021.105796.
- 1013 62. Wei YY, Fan YM, Ga Y, Zhang YN, Han JC, Hao ZH. Shaoyao Decoction
- 1014 Attenuates Dss-Induced Ulcerative Colitis, Macrophage and Nlrp3 Inflammasome
- 1015 Activation through the Mkp1/Nf-Kb Pathway. Phytomedicine (2021) 92:153743.
- 1016 Epub 2021/09/29. doi: 10.1016/j.phymed.2021.153743.
- 1017 63. Bergmann S, von Buenau B, Vidal YSS, Haftek M, Wladykowski E, Houdek P, et
- 1018 al. Claudin-1 Decrease Impacts Epidermal Barrier Function in Atopic Dermatitis
- 1019 Lesions Dose-Dependently. Sci Rep (2020) 10(1):2024. Epub 2020/02/08. doi:
- 1020 10.1038/s41598-020-58718-9.