ELUCIDATING SELF-ASSEMBLY AND ANTIMICROBIAL STRATEGIES OF SYNTHETIC PEPTIDES: AN IN SILICO INVESTIGATION

Irene Marzuoli

RANDALL CENTRE OF CELL AND MOLECULAR BIOPHYSICS

KING'S COLLEGE LONDON



This dissertation is submitted for the degree of Doctor of Philosophy

September 2019



Declaration

This dissertation describes work I have carried out between October 2016 and September 2019 at the Randall Centre of King's College London, under the supervision of Professor Franca Fraternali (first supervisor) and Dr. Chris D. Lorenz (second supervisor).

This dissertation contains material appearing in the following articles:

• ...

In addition to the above, I have contributed to the following publications during the course of my PhD:

• ...

This dissertation is my own work and contains nothing which is the outcome of work done in collaboration with others, except as specified in the text and acknowledgements. It has not been submitted in whole or in part for any degree or diploma at this or any other university.

Irene Marzuoli September 2019

Acknowledgements

. . .

Summary

Elucidating self-assembly and antimicrobial strategies of synthetic peptides an in silico investigation

Irene Marzuoli King's College London

... ..

Contents

1	Introduction		
	1.1	Drug delivery: challenges and solutions	14
	1.2	Antimicrobial resistance	19
	1.3	Alternative antibiotic strategies: antimicrobial peptides	26
	1.4	Closing the circle: an antimicrobial drug delivery vehicle	43
	App	pendices	48
	A.1	On the derivation of the GP predictive distribution	48
\mathbf{B}^{i}	bliog	graphy	49

List of Figures

1.1	Figures a. to k. adapted from: a. [11], a. [11], a. [11], a. [11], a.	
	[11], a. [11], a. [11], a. [11], a. [11], a. [11], a. [11]	13

THEORY stays to experiment as experiment stays to nature. And science stays to technology as technology stays to life.

To navigate this huge gap is the scientist call, in an effort to bring the extremes closer, giving a model of how nature functions, or to enrich the space in between, inventing new realities. Technology bridges every day this very gap: every consequence of our abstract thinking is technology, an hidden layer of inductions and deductions which brings us from abstract principle to solutions. Computers, the ultimate technology, are emulating increasingly better and more efficiently this process, sparing us the awareness of the complex mechanisms which thread the problem to its solution. It was in the past two centuries that we witnessed such an evolution of techniques, inventions and machineries that we can now exploit years of theoretical thinking by using tools in practical problems to eventually modify nature in every daily activities. If in the past nature was the mystery and the human intervention on it was simple to understand, now on the contrary many basic principles of the physics laws are clear to most of us but human inventions became increasingly complex, condensing centuries of discoveries in simple, efficient tools. We use planes, drugs and the internet not because we perfectly understand how they work, but because we trust the collective knowledge we, as humans, have accumulated so far.

Out of the many fields at service of the humanity wellness, the most challenging and still far away from being exhausted is the understanding and manipulation of the human body and mind. While machines - in the broadest sense possible - can perform actions in our place, they can't yet think and

live on their own. It is striking how we are finally scratching the understanding of these two entities, mind and body, in the same historical moment, at a point where computers imitates the human reasoning [1?] and biological materials are turned into semi functional organs [2]. However, we are far from completing the jigsaw of knowledge on these topic. On the contrary with the progression of the techniques available to investigate various fields, we realise how vast is the space to be explored. It is then a logical consequence that the modern scientist is becoming more and more specialised, drifting away from the comprehensive knowledge owned by scientists up to two centuries ago; but exactly because the full picture is challenging, every project aimed at understanding an aspect of these enormously vast themes, no matter how tiny the subject is, is involved in a network of efforts, in the hope and trust that piecewise knowledge can build a unique and organic corpus.

This thesis places itself in the domain of understanding how the human body works - how the (non) equilibrium of life is possible and how human intervention can be possible. The tiny and narrow topic it covers wants to explore one possible way in which we help the body to heal itself and to defend itself against external malicious agents. To correct those processes that go wrong means life, and we are biologically and emotionally pushed towards actions that prolong and improve life and have always looked at ways of curing ourselves. But the body evolved in the past blind to reason, on the contrary taking advantage of multiple defences and barrier which secured it from the failure of the reasoning, and it is know to us often a mystery, as we struggle to understand many of its components, letting aside the whole picture.

Quite blindly then we developed in the past a medicinal science which managed to be of service to the human envelope, in what was a remarkable game of trial and error resorting to magic first and to our intelligence last [?]. The risk and inevitable failures tracing the path were a necessary toll to the utmost necessity of keeping healthy, safe and - ultimately - alive. In a history resembling the evolution of technology, we resorted to nature for beneficial molecules [?], which we called drugs in the initial fuzziness existing between healing and deadly, but then humans started identifying the beneficial principle in these natural remedies [?], and ultimately to produce new molecules [?]. The increasing understanding of how we worked pointed out the many challenges a drug has to withstand to be efficient. And this knowledge poses

question: if so many barriers prevent a drug from entering the body, how can infectious agents have found a way to our cells? And if we want to fight those ones instead, why the body cannot recognise these helpful molecules as beneficial ones and is fighting them instead? Are they perhaps damaging for us as well, in some way we have not yet understood?

Luckily, if knowledge has brought awareness of the complexity of the machine our body is, it started bringing also solutions. We do now have drugs [?], we know how to selectively deliver some of them [?]. We have disinfectants [?], and we have antibiotics to fight pathogens [3]. We do know what we are composed of [?], and how this material rearranges in organelle, cells, organs. And we know some of the mechanisms concerting these parts together [?].

We are finally moving in the direction of the magic bullet envisioned a century ago by Nobel Prize Paul Ehrlich, who dreamed of a 'personalised and tailored drug' able to target specific molecular defects while being harmless - if not beneficial - to the other cells [4]. Such success would condense in a tiny amount of space a century of efforts in understanding the human body. But we still miss many pieces of information as the more we zoom in, the less each single researcher can monitor at once and the more we realize is there to be discovered.

In this prospect, looking at how one particular molecule behaves with respect to a particular environment, as this thesis does, using a simplified theory (sometimes the only possible) is certainly a tiny fragment of knowledge added to the world of science. But, joint to the other scientific output from the community, it is a necessary, meaningful and promising fragment.

This introduction is meant to give an overview of the many different challenges the fields of medicine and bioengineering have faced in recent years, challenges that have arisen the interest for self-assembling antimicrobial peptides. Antimicrobial peptides were not a primary source of interest in these fields as other materials and concepts were deemed more suitable to solve the tasks coming along the way. It is therefore important to clarify the landscape of such other solutions and approaches to understand and value why a change in the research focus has come to age. Figure 1.1 provides a work flow of this introductory chapter to help the reader in identifying the sections of interest.

Motivations of the work: a graphical abstract

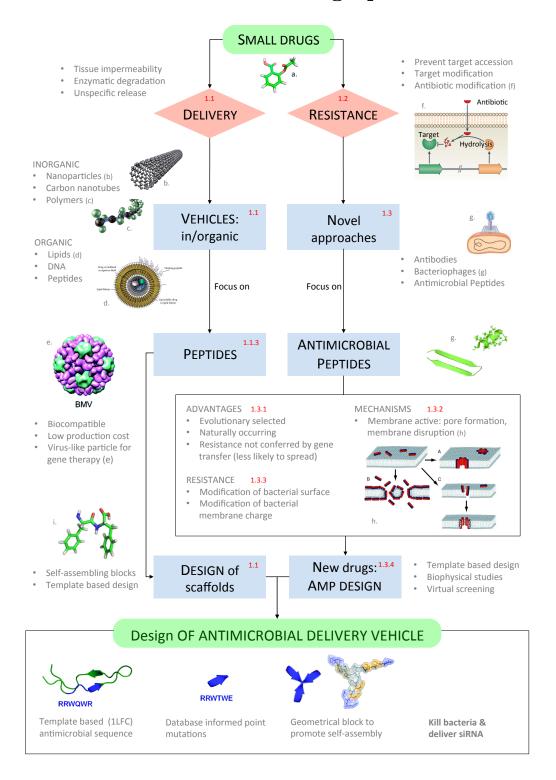


Figure 1.1: Figures a. to k. adapted from: a. [11], a. [11]

1.1 Drug delivery: challenges and solutions

1.1.1 Environmental challenges of drug delivery

The problem of drug delivery is an excellent example of the hurdles existing between a theoretical reasoning and nature: a new drug is usually designed to affect a specific target. Even if in silico experiment can prove its efficacy, its usefulness is bound to its ability to cross the many barriers dividing the inoculation site from the very target inside the human body. To reach the aimed organ, drug molecules must be compatible with the different cellular environments they cross but be preferentially retained and act only on the ones they are designed for. This implies a subtle balance between a disruptive activity on one side, and harmlessness on the other, least the compound is recognised as dangerous and disposed of by the efficient immune and reticuloendothelial systems of the body which aim at neutralise every exogenous substance.

As en example, the trip of an orally administered "free" drug, i.e. an active molecule without any aiding delivery agent, passes through the digestive system, with its challenging acidic environment and limited permeation across the intestinal epithelium, and from there to the blood stream [5, 6]. Then the drug diffuses in the tissues flanking the blood vessels naturally depleting its concentration downstream [7], so that regions further away in the line have less chances of getting a sizeable dose, which implies that high drug concentrations might be needed as starting point to efficiently target every organ.

However, this naive picture of a drug diffusing in the body is complicated by the impermeability of specific tissues: the brain for example, one of the most delicate organs in the body, is well protected from the attack of external agents by the blood brain barrier (BBB), which allows the passage of small molecules only (< 400-500 Da, while standard 'small molecule drugs' go up to 900 Da) of high lipid solubility [7, 8]. Other tissues, like tumoral ones, are instead poorly vasculated, reducing the chances of delivery at their interior [8].

Finally, during their journey to reach the receptor, enzyme or organelle they are meant for, drugs must not be captured and disposed by the immune systems. However, inorganic small molecules are not mimetic by themselves, i.e. they often do not resemble the ones naturally present in the body, and this brings uncertainty on how they would interact with organic molecules. Generally, as soon as they reach the blood stream they are coated by a protein

corona based on their shape and charge [7]. Such modifications are often difficult to predict and can disrupt of decrease significantly the efficacy of the compound as they modify the way drugs are recognised and absorbed by the target.

For all the above reasons, research has focussed on developing systems to assist the delivery of drugs [6, 8, 9]. A mimetic carrier can not only improve delivery, but also be designed to selectively bind to particular tissues or to trigger the drug release after a given time or upon changes in environmental variables (for example pH) to reduce drug concentration in non targeted regions. A stand alone field of research has then focussed on the development of delivery vehicles irrespective from the quest for disease targeting compounds. The optimised products of the two efforts can then be paired according to the condition to address.

At present, many molecules have been successfully employed to build drug vehicles: inorganic metals, polymers, lipids and proteins are all suitable for the aim and offer a range of different physico-chemical characteristics useful to target different regions [10]. A brief (and non exhaustive) overview of some them is meaningful to point out the broad variety and exoticity of structures which can be useful in the medical world, sometimes unexpectedly.

1.1.2 Inorganic materials for small drugs delivery

Metal nanoparticles In the range of inorganic compounds, golden nanoparticle demonstrated to be remarkable for tumour treatment: first of all, they can be customised in shape and size (down to a 10 nm radius), coated with biologically active moieties or made less visible to the immune system by conjugation to a poly-ethyleneglycol (PEG) polymer layer [11]. Moreover they possess optical properties that allow them to be tracked inside the body and they can be thermally stimulated to trigger the release of drug, favour the penetration through the cell membrane or perform thermal therapy, disrupting the cells nearby [12]. At present, there are mixed evidence about their toxicity [12] and doubts have been raised on the long term effects of metallic fragments in the body. For that reason, only a few golden nanoparticle based compounds have made to the clinical stage so far [11] but, given their high and still unexplored potential, they continue to be a primary interest of the medical community and a very active research field.

Carbon nanotubes Similarly, carbon nanotubes have been used for biomedical applications as they have a high loading efficiency thanks to their high surface area and easy interaction with biomolecules through van der Waals, π - π stacking or hydrophobic effect [13]. Therefore they are easy to functionalise through conjugation to extra organic groups to increase their biocompatibility; and have potential for targeted drug release upon change in environmental pH [14].

Polymers Polymers are another large class of inorganic molecules functionalised for the benefit of medicine: for example PEG has already been mentioned as aid to make golden nanoparticles bio-compatible. Indeed, thanks to its high hydrophilicity, it is a clinically approved molecule widely used to mimetise structures (e.g. inorganic, peptides) which in turn carry a drug [15]. or as a stand alone carrier system, as it has a high drug payload [16]. As each of their monomer constituent can be either hydrophilic or hydrophobic, the great strength of polymers is their flexibility. They can be engineered to assemble in many different structures,[17]; they can trigger a sustained drug release by swelling slowly in water,[18] or undergo sol-gel phase transition upon specific changes in the environment [16]. Finally, research has also focussed on improving their biodegradability [19] or in making polymers a bioactive compound itself [20].

1.1.3 Organic materials for small drugs delivery

A somehow opposite approach for designing drug vehicles to the use of inorganic material consists in using molecules similar to the ones present in the body, in an effort to exploit already available biocompatible materials and reduce toxicity [21]. In this category fall lipids, DNA and peptides.

Lipids Lipids are the main constituents of the cell membrane and as such they represent a mimetic material. They come with a great variety, enhanced by the many species produced synthetically. The components selected for drug delivery are usually taken from the biological lipidome, but their composition differs from the cellular membrane one, and possibly includes synthetic molecules, to tune their release properties and enable them to survive the delivery journey [22]. They can encapsulate efficiently both hydrophobic

or hydrophilic drugs, arranging themselves in micelles structures (monolayer spheres with the hydrophobic tails facing the interior) or in liposomes (bilayer spheres with a water filled core) [23], with many of them overcoming the clinical stage and currently approved for cancer and infections [24, 25].

DNA scaffolds Similarly, many DNA scaffolds have been tested for smart delivery: DNA origami is nowadays an established technique to build three dimensional customised solids [26], and the nanometric knowledge about their constituents makes possible fine tuning them for a triggered release of the content [27]. First studies proved them successful in delivering anticancer agents [28, 29], however they are very sensitive to cellular environment and this, united with high production costs, prevented them constitute a viable class of carriers so far.

Peptidic scaffolds Another widely used and trustworthy mimetic vehicle comes, quite surprisingly, from the world of pathogens: viruses have co-evolved with humans, to be able to penetrate into cells where they complete their reproductive cycle [30] Therefore their capsid, the peptidic shell encapsulating the genome, is highly suitable for cell penetration. The first application sought historically was to employ genome free viruses to stimulate and train the natural immune response against the respective genome-loaded ones, creating viral vaccines - in a similar fashion to what already done with the inoculation of dead bacteria to counteract the infections caused from them [31]. Later in the history, their potential as cargo carrier was pursued first by modifying their genetic material to include sequences beneficial for the host cell and prevent the infectious duplication at the same time. In particular the adeno-associated virus (AAV) has been widely studied [32] as it triggers a low immune response [33], and the first AAV viral therapy has been finally approved a few years ago [34]. To fully exploit the potential of a peptidic carrier many efforts have focussed on synthesising in vitro gene-free capsids, either as they appear in nature [35] or designing artificial building blocks, which assemble in so called Virus-Like particles (VLPs), to help overcoming the reaction stimulated by specific viral capsids to which the immune system is (already) sensible to.

Among their advantages, peptides present biocompatibility, a low production cost and a tunable bioactivity thanks to their chemical diversity, which

help in tailor the assembly toward the target of interest [36]. Moreover, the variety of amino acid available makes possible to load peptidic structures with both hydrophilic and hydrophobic drugs, according to their amino acid composition [37, 38]. Similarly to other delivery vehicles, the surface of such particles can be functionalised with additional molecules to improve the target selectivity and increase biocompatibility, while the peptidic scaffold grants robustness to the structure. Therefore, VLPs loaded with drugs can be tuned for an efficient intra cellular release [38]. The easy manipulation of peptidic structure derives from the fact that proteins are a fundamental component of the human body, so that there is a vast literature on their interactions with membranes, cell receptors and in general biological components, from which the design for novel materials can take inspiration to employ building block sensible to particular triggers within the body.

A step further in engineering peptidic structures is represented by the design of self-assembling functional structures from first principles, i.e. to exploit the physico chemical characteristics of peptides, regardless their resemblance of viral capsids. Indeed self-assembling peptides can form nanostructures ranging from nanoparticles to nanotubes, nanofibers, nanorods and hydrogels [36, 37]. The assembly is modulated by the peptide length and its hydrophobic or hydrophilic character: on one end of the length scale, phenylalanine dipeptides were designed with inspiration of a pathogenic process towards molecular self-assembly [39] and were shown to self-assemble in a multiscale process producing nanotubes able to load drug molecules [40]. The relatively small diphenylalanine building block is non the less complex has it bears two charged termini (as the process is observed at neutral pH), and two aromatic hydrophobic rings, therefore the dipeptide is driven towards assembly by the hydrophobic forces acting on the phenylalanine side chains.

In a different approach, longer sequences can be employed to guide the formation of the local structure, as they organise spatially in well studied motives (the secondary structure) with a known interaction among themselves. The two typical secondary structures, α -helices and β -sheets, appear in sequences of about 20 or more amino acids length and are both amphiphatic, thus promoting the assembly between the hydrophobic faces of different copies of the same structure. With the appearance of a secondary structure, more complex building blocks can be designed, to tune the shape into the ones needed for

the supramolecular organisation of interest [41]. Again, the knowledge of many protein structures [42] give us insight in how the small structure can hierarchically assemble into larger ones - however the challenge and outlook often goes in the direction of synthesising exotic, non natural, novel geometries [43, 44].

1.2 Antimicrobial resistance

The previous brief review on drug carriers rotates around the paradigm that a drug is a small molecule inorganic compound (of mass up to 900 Da) which targets a specific molecule of a specific target of a mammal or bacterial cell. In this light, the ultimate goal of the delivery vehicle is to carry the drug to the site of action where it can interfere with the processes it is assigned to. Very often the target of interest of small molecule drugs are proteins: out of the 695 small drugs approved by FDA (the American Food and Drug Administration agency) to target human molecules, 667 acts on proteins. Similarly, 189 of the 198 small drugs approved to treat pathogens have a protein as their target [45]. It must be noticed however that the identification of an unambiguous drug target poses challenges in many cases, especially when the drug binds to a protein complex or to a number of closely related gene products [45].

In presenting the aforementioned figures, the data were naturally split among the drugs which target human molecules, "repairing" some faulty process in the human body, or the ones active against bacteria, which "disrupts" the bacterium life cycle in order to kill or prevent the reproduction of the pathogen. It appears evident that the pool of drugs available to the second purpose are in consistently lower number than the ones addressing human molecules. This comes from the nature of the action they perform: molecules targeting human proteins need to be highly specific to avoid interference with other proteins or with healthy cells, and in a sufficient number to address the variety of diseases affecting the human body. Antibiotic must be non-toxic for human cells as well, i.e. their target must not be shared between mammal and bacterial cells ?]. but there is a less stringent requirement in their selectivity on different bacterial species. On the contrary, it is often useful to have a broad-spectrum compound. This cross-species efficacy and non-toxic property is obtained thanks to the evolutionary relationship among bacterial species, and between bacteria and humans: while the first are closely related,

and therefore share homologous proteins with very similar structures, humans have less architectures in common with them, allowing for a resilience against bacteria-targeting drugs [?]. Of course the set of bacterial species is very diverse and the cross-species effectiveness of some drugs does not extend to the whole bacterial population. This demonstrates to be a positive feature, given the large amount of beneficial bacteria that live in symbiosis with the human body (especially in the gut [?]) and that must be preserved for an optimal wellness.

In the framework described above, it is understandable that the first time research on antibiotics was satisfied with the development of a handful of potent, broad-spectrum compounds. Penicillin, the first of them, was isolated from a mould in 1928 by Alexander Fleming. It acts inhibiting the formation of peptidoglycan cross-links in the bacterial cell wall (for a review of bacterial cell membrane structure the reader can refer to Section – and the relative references). This inhibition is achieved through binding to the enzyme DD-transpeptidase responsible for the catalysis of such cross-link [?]. As foreseen from Fleming himself in his Nobel Prize acceptance speech, bacteria can become immune to penicillin, and this is specifically achieved by either production of penicillase, an enzyme that degrades penicillin, or by subtle changes in the structure of the penicillin-binding proteins to prevent penicillin binding or again by removal of the drug outside of the cell through specially re-purposed efflux pumps that they use to release substances from the cell.

1.2.1 Course of antimicrobial resistance

This mechanism is not an exceptional characteristic of penicillin, and many drugs lost their effectiveness against some bacteria since their discovery till nowadays. Indeed the antibiotic landscape is a dynamic entity in which newly discovered ones enter while other exit after having been exploited for years.

In the first stages of the insurgence of antimicrobial resistance (AMR), less effectiveness of a drug means that some strains of bacteria are not damaged by the standard doses of the drug as they possess some natural occurring mutations in their genome which promote an escape mechanism which invalidate the drug effectiveness [46, 47]. Usually only a small population of bacteria is resistant, and it can be killed never the less by augmenting the dose of the drug. However, the natural course of AMR states that the resistant popula-

tion will replicate faster that the peers of the same species because it is more fit in an environment challenged by the presence of the drug. It is noteworthy that this fitness might not be optimal in a natural drug-free environment - and indeed the wild population has not been selected for that genotype - but under the pressure derived from the treatment, other characteristics result more advantageous. In the short time scale it is usually sufficient to increase the doses of a drug to re-gain efficiency against the target, but it has been observed that a species resistant to a drug can usually adapt to higher doses of the same [?]. Moreover, high drug doses are not always applicable due to the severe side effects they are connected to [?].

The spread of resistance between bacterial cells and even species is very effective as bacteria are able to exchange genetic material with other individuals via small rings of DNA in a process called conjugation [?]. In this way the advantageous characters which promote resistance spread across individuals and species with an innate resistance can transfer to other ones their mechanisms of resilience to a particular drug. Therefore, despite AMR is an evolutionary mechanism, the fast pace at which bacteria replicates, their enormous population (in terms of individuals), and the relative easy horizontal gene transfer through conjugation place the insurgence of resistance well within the human lifespan time scale [?].

It is then clear that resistance is a very complex problem which depends on many variables: the casual appearance of resistant individuals, the transfer of information between them, the relatively larger fitness of resistant individuals and the dosage and time line of the drug administration. Many mathematical models have been implemented to understand the issue [48, 49], but it is known that some particular strategies of drug administration are worse than other, favouring the proliferation of resistant bugs. One example is the underdosage of antibiotics: a low drug load is likely to harm but not kill pathogens, in particular to promote the fitness of resistant ones. In a sort of "gym" or "vaccination" process for bacteria, an underdosed drug would kill the weakest individuals but strengthen the resistant population, which would now be fitted to the challenges of a higher dosage [?]. Similarly, the abuse of antibiotics puts an high pressure on the pathogenic populations, which is desirable but at the same time can induce a faster emergence of escape mechanisms [?]. In this context it must be noticed that many drugs are bacteriostatic agent as opposed

to bactericidal: i.e. they prevent the bacterium growth rather than kill it, as they are meant to control the bacteria and slow down the damage while host defence mechanisms to eradicate them. It is then clear that a high dosage of a bactericidal agent may to extinguish the bacterial population and eradicate the disease, but for bacteriostatic drugs, once they are removed, bacteria can usually start again their reproduction cycle.

It is noteworthy that abuse of antibiotics can take many forms: a part from the drugs used to treat human related diseases, the agricultural and breeding sectors are constantly using antibiotics to keep their products secure form illness. This results in large quantities of drugs to be released in the soil and water, which ultimately reach humans in underdosed quantities. Diseases of plants and animals are different from the ones affecting humans, however some drugs are effective on many bacteria including the one affecting humans. Therefore the widespread use of antibiotic for animals or plants can ultimately lead to train resistant bacteria in humans [?]. Additionally, the diseases can cross species: this means that an extra care must be taken in the treatment of non human bacteria least to promote resistant ones which can at a point cross species and affect us [?].

The complexity and severity of the issue is such that it has been raised to the status of national emergency in several countries, including UK, as we are leaving the century in which antibiotics were discovered, to enter a phase in which we count the number of the ones loosing efficacy [50].

REDO: SAY THAT IT MAKES SENSE THAT SOME BEHAVIOURS TRAINS BETTER RESISTANT BACTERIA, AND THAT THIS IS A SOCIAL EMERGENCY??

1.2.2 Mechanisms of antimicrobial resistance to small drugs

Antimicrobial resistance can manifest through many different mechanisms, as highlighted in the example of the penicillin resistant bacteria. In particular, resistance mechanisms fall into three main groups: a first group minimises intracellular concentration of the antibiotic preventing penetration of maximising efflux; a second one modifies the antibiotic target by genetic mutation or post-translational modification; finally a third group inactivates the antibiotic by hydrolysis or modification of the drug molecule [47].

Prevention of access to target One possible mechanism of defence bacteria employ against antibiotics is to prevent the access to the target. This is performed either preventing the drug influx or promoting its quick efflux in the eventuality it has entered the cell.

Regarding drug influx, not all the molecules can enter the cell permeating the membrane, and this holds particularly for hydrophilic antibiotics tackling Gram-negative bacteria: indeed, compared with Gram-positive ones, Gram-negative bacteria are intrinsically less permeable because of the structure of the additional outer membrane [51], therefore hydrophilic molecules are imported into the cell through outer-membrane porin proteins [52, 53]. The major porins of most Enterobacteriaceae are thought to be non-specific channels [54], therefore replacing porins with more selective channels or down regulating their expression would limit the intake of the drug. This last mechanism is well established and contribute to resistance to many different drugs in Gram-negative bacteria, including newer drugs such as carbapenems and cephalosporins, for which resistance is usually mediated by enzymatic degradation [55–59]. Finally, in E. coli exposed to carbapenems, not only the porin expression is down-regulated, but also the genes coding for porins are heavily mutated, suggesting that changes in the porin structure can enhance their selectivity and reduce the drug influx [57, 60, 61].

A strategy complementary to prevent drug influx is to dispose of the drug efficiently once it has invaded the cell. Bacterial efflux pumps transport many antibiotics out of the cell, and they constitute a major hurdle for the treatment of Gram-negative bacteria as opposed to Gram-positive ones. Indeed, many of the drugs effective of the latter are evacuated by the former through efflux pumps; in particular, multidrug resistance (MDR) efflux pumps can transport a wide range of structurally dissimilar substrates. All bacteria can produce their own MDR pumps [62–65]. moreover it has been shown that the genes encoding for some of them have been transferred to plasmids and thus can be transferred to other bacterial species, disseminating resistance [66]. The over expression of efflux pump seen in multidrug-resistant bacteria is often due to mutation in the regulatory network controlling it, either in the local or the global regulators [67]. Increased expression of efflux pumps can also occur as a result of induction in response to environmental signals and in conditions in which their function is required [68–70].

Change or modification of the antibiotic target Most antibiotics bind to the target with high affinity and therefore specificity. Small modifications of the target structure can disrupt an efficient binding of the antibiotic, still allowing the target to maintain its normal function. Preservation the target from the action of the drug can be reached by either mutation or protection of the binding site.

In the first case, a casual mutation would provide such minimal required change and the resistant population would spread according to its improved fit. An example is the development of resistance to linezolid in S. pneumoniae and S. aureus: this drugs targets the 23S rRNA ribosomal subunit of Gram-positive bacteria which is encoded by multiple, identical copies of its gene. The use of linezolid has selected first a population with a mutation in one of the copies, which has afterwards passed to other copies via recombination, generating a population favouring the synthesis of the mutant subunit [71, 72]. Other examples include mutations occurred by transformation, i.e. uptake of DNA from the environment as in the case of penicillin resistant S. pneumoniae, which is conferred by a penicillin-binding protein gene included in the genome by recombination with DNA from the closely related species Streptococcusmitis; or by acquisition of a gene homologous to the original target as in methicillinresistant S. aureus, which acquired the staphylococcal cassette chromosome mec element [73]: this gene allows the synthesis of the PBP2 protein which enable cell wall synthesis despite the native PBP is inhibited by the antibiotic [74].

The second mechanism includes ways of protecting the target from binding of the drug via addition of chemical groups to the target after its synthesis, and thus they do not require mutations at the genetic level. Methylation is an important process which triggers resistance: for example, under the pressure of macrolides, lincosamines and streptogramins, the 16S rRNA subunit is methylated and thus the drug-binding site altered [75]. Similarly, it has found that specific methylation of A2503 in the 23S rRNA subunit confer resistance to many drugs (phenicols, pleuromutilins, streptogramins, lincosamides and oxazolidonones) that target nearby regions [76]. In a different mechanism, quinolone resistance can be conferred by a gene coding for a pentapeptide repeat proteins (PRPs), which binds to topoisomerase IV and DNA gyrase promoting the release o the drug and rescuing the normal function of topoiso-

merase [77].

Direct modification of antibiotics Finally, bacteria can modify or destroy drugs to prevent their action, usually by either hydrolysis or by transfer of a chemical group. The enzyme-catalysed modification of antibiotics is a major mechanism of antibiotic resistance: the very first example being penicillinase (a β -lactamase) which destroy penicillin [78]. Since this discovery, thousands of enzymes have been identified that can degrade and modify antibiotics of different classes, such as β -lactams, aminoglycosides, phenicals and macrolides [79–82]. These enzymes co-evolved together with the newly developed drugs which bacteria are exposed to, to include in their spectrum of action new compounds of similar composition: for example the first β -lactmases evolved in broad spectrum ones active against the new β -lactams antibiotics, up to the emergence of isolates resistant to all the drugs in the β -lactam class [83]. This localised emergence of resistance constitutes a serious problem as these mechanisms are usually effective in spreading resistance to the whole bacterial population in a short period of time [81, 83, 84]. Moreover, as hinted in the section 1.2.1, the inefficacy of one class of drugs brings inevitably to a more massive use of other compounds (for example carbapenem in replacement of β -lactams), to which bacteria develop resistance (in the example above, developing the so called carbapenemases to hydrolyse the drug) [85–87].

The addition of chemical groups to vulnerable sites on the antibiotic molecule by bacterial enzymes is another mechanism to block the action of the drug, as it prevents the binding to its target protein due to steric hindrance. Different enzyme can complete such task, transferring many groups as acyl, phosphate, nucleotidyl or ribitoyl [88]. Antibiotics constitute by large molecules with many exposed hydroxyl and amide groups are particularly susceptible to this modification. An example class of such antibiotics is aminoglycoside (in which streptomycin is included), which can be modified by three classes of enzyme, grouped according to the chemical moiety added: acetyltransferases, phosphotransferases and nucleotidyltransferases [89]. A recent development reports the discovery of a genetic island in Campylobacter coli isolated from broiler chickens in China coding for six of these enzymes at once, including members of all three classes: the expression of such genes would then confer resistance to many antibiotic of the aminoglycoside class [90].

All together, the recent progress in understanding the mechanisms of antimicrobial resistance has helped in directing the development of new drugs, in particular the modification and improvement of existing compounds to escape the resistance developed by bacteria. This in turn has highlight some clinical strategies, such as the use of combined therapies, to counteract an early development of resistance. However, the problem persists and more knowledge needs to be gather for a complete understanding and the possible development of resistance-free compounds.

1.3 Alternative antibiotic strategies: antimicrobial peptides

In the landscape sketched above it is evident that the development of novel drugs is of crucial importance. Even more beneficial would be to have at disposal a new paradigm for their design, in order to attack pathogens in a completely novel way, avoiding to target pathways which are recognised to easily lead to the development of antimicrobial resistance.

A possible solution is the use antibodies, bacteriophages or antimicrobial peptides instead of small molecules [91]. Regarding antibodies, the development of pathogen-specific monoclonal antibodies (mAb) is an emerging area of research. They can be employed for example for immunisation though serum therapy, i.e. exposing the patient to the serum of an individual already immunised. Such passive immunization has been used for the treatment of bacterial infections well before the discovery and development of antibiotics, but has since then been overshadowed by the use of small-molecule compounds. The second class mentioned, bacteriophages, are viruses which infect bacteria and archea rather than eukarya. They are effective as they can be used both in natural environmental reservoirs and in humans and are usually highly specific for one bacterial strain. Both these strategies have been only partially explored so far, bringing potential for new therapies. Phage therapy is also promising in terms of promoting a low resistance development: indeed phages and bacteria have been coexisting since a long time - in evolutionary scale - and the formers

are never the less effective against the latter, suggesting that their mechanism of attack is weakly prone to provoke the insurgence of resistance.

But are antimicrobial peptides the focus of this thesis: we have already highlighted the importance of peptides as tunable structural elements of drug delivery vehicles. However, they can have a role against bacteria as drug themselves when their sequence possesses some specific characteristics: such sequences, capable of damaging and/or killing bacteria, are referred to as antimicrobial peptides. The following paragraphs will explore their characteristics, modes of action and the response of bacteria against them: indeed it is crucial to understand the complexity of the picture of what is already known versus the questions that are still open. This holds in particular when the investigation proceeds by the use of simplified models, as meaningful results can proceed only from a sensible modelling of the problem.

1.3.1 Host-defence, membrane active peptides

Antimicrobial peptides (AMPs) are naturally produced by the human body and more in general by mammal, either as stand-alone sequences or embedded in larger proteins, as first weak and broad-spectrum defence against bacteria [92–95]. Thus this pool of molecules has been selected though evolution to be active against pathogens, suggesting that they are not prone to provoke resistance reactions in the microbes they attack.

To exploit their potential and engineer AMP-like molecules, a careful characterisation and classification of such peptides must be done. This task has been carried on in the past decades but it is not trivial, so that up to date there are many peptides with ascertained antimicrobial activity for which the mode of action is still not fully understood [?]. However, some general characteristics of these sequences and the mechanisms they employ have emerged. Unsurprisingly, AMPs are heterogeneous in shape, targets and mode of action. The size can vary between 6 and 59 amino acids [96]: despite being small with respect to the average size of a protein in the human body, these macromolecules are hundreds of times larger than small molecule drugs and as such they penetrate and act on bacteria differently.

The most common target of AMPs is the bacterial membrane. Many of them cause disruption of the physical integrity of the microbial membrane while others translocate into the cytoplasm to act on intracellular targets, and

the combination of the two is not uncommon either [97]. In general, it is widely accepted that membrane interaction is a key factor for the direct antimicrobial activity of AMPs [92, 98]. The determinant driving the interaction between the two is the positive charge that many AMPs presents, opposed to the negative charge of bacterial membrane [99, 100]. It is striking that such simple mechanism based on the common presence of a certain number of negatively charged lipids holds across many bacterial species despite the great variability found in their membrane composition. Based on the differences in their cell envelope structure, bacteria are classified into two macro families, Gram-positive and Gram-negative. In Gram-positive bacteria, the cytoplasmic membrane is surrounded by a thick peptidoglycan layer, while for Gram-negative bacteria this membrane (which assumes the name of internal one) is surrounded by a thin peptidoglycan layer as well as an outer membrane [101]. The cytoplasmic membranes of both Gram-positive and Gram-negative bacteria are rich in phospholipids like phosphatidylglycerol, cardiolipin, and phosphatidylserine, which have negatively charged headgroups, highly attractive for positively charged AMPs, and this is often sufficient to promote the preferential interaction between this membrane and the peptides.

The fact that AMPs tackle negatively charged membranes is crucial for their selectivity, i.e. the fact that they do not disrupt the cells they are produced from [102]. Indeed, mammalian cells have a different membrane composition with respect to the bacterial one, in particular the mammalian membrane is rich in the zwitterionic phospholipids phosphatidylethanolamine, phosphatidylcholine, and sphingomyelin, providing a neutral net charge [103, 104. Strictly speaking, some negatively charged lipids are present in some cell types, however they are located in the inner leaflet, while the zwitterionic phospholipids are more abundant in the outer leaflet, giving an asymmetric composition [?]. This structures promotes weaker interaction between AMPs and the mammalian cell membrane with respect to the bacterial membrane as the former is driven mainly by hydrophobic interactions, while the latter by electrostatic ones. Furthermore, the mammalian cell membrane has a high cholesterol content [105, 106], which is proposed to stabilise the membrane enhancing its fluidity, so that it is more able to accommodate the perturbations caused by AMPs [107]. Finally, bacterial cells have a typical transmembrane potential - the difference of electrostatic potential between the inside and the outside environment - between -130 and -150 mV, while mammalian cells between -90 and -110 mV [105, 108, 109]. Given that a potential generates an electric field across the membrane, the higher the potential, the higher the electric field pointing from outside to inside the cell. A field in such direction pushes cationic compounds on the outside of the membrane toward the membrane itself. Therefore a stronger transmembrane potential may promote an enhanced - and thus disruptive - interaction with bacterial cells, contributing to the selectivity of AMPs between bacteria versus mammals [105].

1.3.2 Common mechanisms of action of AMPs

Investigating the perturbation and disruption of a bacterial membrane by antimicrobial peptides is a key point of this work, therefore it is important to highlight the mechanisms known so far through which AMPs reach this outcome. As already mentioned, many AMPs have a positive charge which facilitates the binding to the membrane via charge-charge recognition; accordingly, Arginine and Lysine residues are usually abundant in AMPs sequences. However, the disruptive action takes place through the interaction of the AMP with the hydrophobic core of the membrane, therefore many AMPs contain hydrophobic aromatic residues, especially Tryptophan, which favours the anchoring to the membrane core [110]. Overall, AMPs resort often to adopt an amphiphatic structure to segregate the hydrophilic from the hydrophobic amino acids and thus act at the interface between membrane and solution. It is interesting to notice that some of them fold into the active structure only nearby the membrane, as they can expose their hydrophobic components to face it, while in solution these ones are preferentially buried inside to be screened from the solvent [92].

The folds adopted by AMPs are both α -helix or β -sheet rich structures. Amphiphatic α -helices present a charged side which is tailored to face outward towards the phospholipid head groups and an hydrophobic ones which is favourably buried into the acyl chains core. In the initial phases of the interaction, the peptide lies parallel to the membrane with the two faces in the respective favourable regions, while subsequent rearrangements bring the helix axis to form an angle with the membrane plane, and finally to insert deeper into the lipid core, often spanning the full membrane thickness [109]. Structures rich in β -sheets include β -hairpins, which again show an amphiphatic

distribution of residues and, similarly to helices, insert within the membrane after a first flat approach. The final insertion arrangement depends on the peptide characteristics and length, the presence of kinks in its structure in case of helices, and the interactions with other copies of the peptide.

Transmembrane pore formation Several models have been proposed to describe the exact mechanisms of AMPs penetration after they bind to the cytoplasmatic membrane, and how this leads to membrane permeabilization. [92, 96, 111] At low peptide to lipid ratio, the favourable configuration is represented by peptides lying parallel to the membrane plane as described previously, [112] but an increase in peptide concentration triggers the transition to an inserted state where the main axis of the peptide is perpendicular to the membrane. The organisation of AMPs inside the membrane core can assume different configurations, and a few models have been outlined to describe them.

The "barrel-stave" model proposes that AMPs insert perpendicularly into the bilayer. Recruitment of peptides in the same area results in the formation of a transmembrane pore with a central lumen. The walls of the pore are constituted by the hydrophilic face of the peptides, while their hydrophobic side is interacting with the lipid tails around the pore. This model is adopted for example by the α -helical AMP alamethicin, which forms voltage-dependent ion channels by aggregation of four to six molecules [112–115]. According to the "toroidal" pore model instead, the insertion of peptides forces the phospholipid to bend continuously from one leaflet to the other, resulting in a pore defined by both peptides and phospholipids head groups. The toroidal model differs from the barrel-stave model as the peptides are always associated with the lipid head groups even when they are perpendicularly inserted in the lipid bilayer. The "toroidal" pore mechanism is induced by the α -helical magainins, protegrins and melittin [112, 116, 117], and leads to more extensive membrane perturbation, as the lipids must rearrange around the pore [113]. As a comparison between the two models, alamethic in induced barrel-stave pores have an inner and outer diameters of 1.8 nm and 4.0 nm respectively [115, 118], while magainin-induced toroidal pores are larger and can vary in their size, with an inner diameter of 3.05.0 nm and an outer diameter of 7.08.4 nm, involving about 4 to 7 magainin monomers and 90 lipid molecules [119, 120]. Finally, in the "carpet" model, the accumulation of AMPs on the surface of the membrane, laying parallel to it, causes tension in the bilayer. Subsequently the membrane is disrupted by peptides in a detergent-like manner, leading to the formation of micelles [121, 122]. The critical threshold concentration triggers a cascade effect, in which formation of the first disruption in the membrane allows the penetration of other AMPs in the inner side of the bilayer. The cooperation between peptides on both sides of the lipid membrane enhance the AMP induced curvature on the membrane causing accelerated disruption [123]. The "carpet" model mechanism is again observed for peptides presenting an α -helical structure, generally with two to more helices connected by short loops (like cecropin [124] or ovispirin [125]).

The prevalence of examples with an helical structure for the above models derives from the fact that the understanding of how helical AMPs function is often easier than the one of β -sheet rich structures. Indeed, helices have a well defined fold (at least in the membrane environment), a compact structure, and often a clear segregation of complementary patches that can attract other copies of the peptide and thus promote the self-assembly process necessary for pore formation. On the contrary, many β -sheet AMPs have a more flexible structure, diversifying their mechanisms of action [?].

Alternative mechanisms of action AMPs rich in β -sheets can be divided into β -hairpins and peptides from the defensin family [92]. Many representative of the former class disrupt bacterial membranes via formation of toroidal pores: as an example, porcine peptide protegrin I triggers the toroidal pore formation assembling into a β -barrel structure when in contact with anionic membranes. However, it folds into β -sheet aggregates on the surface of cholesterol containing membranes, thus acting selectivity on bacterial membranes only [126]. Defensing permeabilise the membrane as well but their mechanisms are not as well explored [107, 127, 128]. Some members of the family form transmembrane pores on planar bilayer when a physiologically relevant negative potential is applied to the membrane, [129] while others like sapecin from Sarcophaga peregrina form oligomers in phospholipid vesicles [130]. Although various descriptions of membrane damage have been reported, and include ion channels, transmembrane pores and extended rupture of the membrane, they are likely related, being a modulation of a similar acting principle [131].

Finally, many other non-lytic mechanisms are suggested for β -sheet AMPs: defensin A from P. terramovae reduces the cytoplasmic potassium concentration, partially depolarising the inner membrane; tachyplesin from horseshoe crabs is able to bind to the minor groove of DNA, interfering DNA protein interactions [132], and bovine lactoferricin can act synergistically with other antimicrobial agents by affecting the transmembrane potential and protonmotive force, resulting in inhibition of ATP-dependent multi-drug efflux pumps [133]. Moreover, after translocation within the cell, bovine lactoferricin can also inhibit DNA, RNA and protein synthesis. Section – will treat in detail the functioning of lactoferricin distinguishing its role as membrane active peptide versus intra-cellular targeting compound: indeed, many works have focussed on locating the section of the sequence performing the antimicrobial activity [134–137] to understand whether it retains its activity regardless of the fold. This investigation, together with similar ones conducted on other AMPs [?], provided the first minimal functioning antimicrobial blocks, promoting the understanding of how AMPs work in general and boosting the design of tailored AMPs from specific blocks.

1.3.3 Mechanisms of resistance to AMPs

Antimicrobial peptides were introduced in this review as a class of new drugs and a possible solution to the crisis of antimicrobial resistance. Any new drug entering the pool of the clinically approved compounds is at least temporary - a solution to the problem of resistance to known antibiotics. It must be clarified that bacteria can develop resistance to AMPs too, therefore they are not a definitive solution to such problem; never the less, it is generally not based on dedicated resistance genes that are conferred by horizontal gene transfer, as in the case of many antibiotics resistance mechanism [138, 139]. Therefore, a certain increase of resistance after exposure to the drug is to be expected (MIC creep), but it is less likely to spread quickly to other species.

Some of the mechanisms of resistance to AMPs are similar to the ones employed by bacteria to counteract small molecule drugs, for example the overexpression of efflux pumps to dispose of the AMP; or prevention to accession to the targets via proteolytic degradation of the peptide by extracellular enzymes or sequestration by the bacterial biofilm matrix; while others tackle the specific action of an AMP on the cell membrane, and prevent it by mod-

ification of the surface or the cytoplasmic membrane. Table 1.1, from Ref. [140] gathers these mechanisms offering examples for each of them in both Gram-positive and Gram-negative bacteria, and in the following paragraph some of them are explained in more details, with the omission of efflux pump, for which the principle is very similar to what explained in Section 1.2.2 for the resistance to small molecules antibiotics.

Mechanism	Gram-positive bacteria	Gram-negative bacteria
Extracellular proteins	Proteolytic degradation Sequestration	Proteolytic degradation
Exopolymers	PIA, PGA	Alginate, polysialic acid
Surface modification	Repulsion (D-alanylation of TA) Steric hindrance (L-rhamnosylation of WTA) Lipid II modification	Repulsion (lipid A phosphate modification) Increased OM rigidity (lipid A acylation) O-antigen of LPS
Cytoplasmic membrane alteration	Charge repulsion (PG amino-acylation)	Increased IM rigidity (PG acylation)

Table 1.1: Overview of bacterial resistance mechanisms against antimicrobial peptides. Adapted from Ref. [140]

Proteolitic degradation and sequestration (TAKE OFF???) The first defence of bacteria against AMPs are the proteins secreted on the extracellular side of the membrane, as some of them, the proteases, are able to degrade peptides and thus AMPs. For example, staphylococci secrete various metalloproteases such as aureolysin and SepA, and serine endopeptidases such as the V8 protease, which are known to degrade linear AMPs such as the human cathelicidin LL-37 [141, 142]; while group A Streptococcus produces instead a cysteine protease able to disrupt many host AMPs, including LL-37 and beta-defensins [143–146]. It is interesting to notice that linear AMPs are more easily degraded than the ones with non-linear structures containing disulfide bonds[138] such as defensins [147]. However, some bacteria have evolved proteases able to degrade even these AMPs with increased stability;

for example protein OmpT from the omptin family contributes to resistance in E. coli. being able to degrade the AMP protamine [148] which is thought to conform to a nonlinear structure involving three disulfide bonds [149]. AMPs can be shielded from the action of proteases by binding to proteins such as extracellular actin, preventing the access of degradative proteases while still maintaining its antimicrobial activity [150]. Finally other more complex mechanisms of degradation are possible, like the ones occurring in the intracellular environment after the AMP is being imported by specific transport proteins [151–153] or the exploitation by bacteria of host enzymes with AMP-degrading activity, in particular of host immune response related proteins [154].

Another process relevant for the neutralisation of AMPs happening in the extracellular environment is the sequestration of the peptides: as an example, staphylokinase is one of the most prominent extracellular AMP-sequestering molecules [155, 156], and inactivates α -defensin binding to them.

Biofilms Bacteria can resist AMPs by organising into specialized structures known as biofilms. These structures are formed by sessile bacteria adhering to a surface in organized manner that allows the circulation of nutrients [157]. Bacteria in a biofilm secretes an extracellular matrix with adhesion and protection functions. This matrix includes various compounds as cellulose, teichoic acids, proteins, lipids and nucleic acids [158] and it is effective in conferring resistance to antibiotics and AMPs. In some cases a resistance 1000 times as great as their planktonic form can be observed [159, 160]. This is done by repulsion and/or capture of aMPs by mainly exopolysaccharid or capsular polysaccharides. For example polysaccharide intercellular adhesin (PIA) produced by S. aureus and a variety of other bacteria is responsible for the resistance to both cationic AMPs (HBD-3, LL-37) and anionic dermcidin [161, 162]: this is done by deacetylation of PIA, which increases its positive net charge, thus repelling more efficiently cationic CAMPs, but perhaps also increased sequestration of dermcidin, as well as forming a mechanical barrier for both of them [162].

In other cases it is structural hindrance as well as electrostatic trapping that prevent cationic AMPs to penetrate the biofilm (as in observed for polymyxin B, HNP-1,HBD-1, lactoferrin and protamine on K. pneumoniae, S. pneumoniae or P. aeruginosa) [163, 164].

Finally, it is important to note that AMPs are being tested as alterna-

tives to traditional antibiotics in the treatment of biofilm-associated infection. Indeed in this type of infections (where bacteria are growing slowly) it is advantageous to have a bactericidal agent as opposed to a bacteriostatic one targeting fast-growing bacteria, as often traditional antibiotics are [165, 166]. However, biofilm-intrinsic AMP resistance make also the use of AMPs for the treatment of biofilm infections challenging [167, 168].

Surface remodelling As mentioned in the previous two paragraphs, the bacterial cell envelope environment constitutes a major impediment for AMPs activity. Even if a peptide reaches the bacterial envelope intact, bacteria can modify the characteristics of their surface to prevent its efficient action. Grampositive and Gram-negative bacteria put in place different strategies to do that, according to their distinct cell envelopes. In particular, the target of such modifications are the teichoic acids (TA) in Gram-positive cell wall and lipopolysaccharide (LPS) in the Gram-negative outer membrane.

For example, D-Alanylation of TA adds a positive charge to it, reducing the attraction of cationic AMPs, as observed in Staphylococcus and others for many AMPs including protegrins [169–171]. This in turn increases the cell wall density, reducing the surface permeability [171]. For Gram-negative bacteria (like P. aeruginosa), the LPS negative charge is increased by addition of different amine-containing molecules [172, 173] or by removing phosphate lipids, which have a negative charge, from lipid A, one of the constitutive moieties of LPS [161, 174]. Another target of AMPs in Gram-positive bacteria is the bacterial peptidoglycan precursor, lipid II, which has a key role in the formation of the cell wall, thus many bacteria use a modified version of it. The best known case is the replacement of its terminal D-alanine with D-lactate or D-serine [175] to avoid the action of the glycopeptide vancomycin, which works binding to the D-Ala-D-Ala terminal moieties of the precurson, preventing cross linking of molecules between them and thus the cell wall synthesis [176].

Gram-negative bacteria instead can enhance the rigidity of the outer membrane to reduce permeability to AMPs via addition of extra acyl chains into lipid A [177, 178]. Finally, the long polysaccharide chain of LPS (called O-antigen) makes this class of bacteria particularly resilient to the action of AMPs [179] indeed both the LPS core and the O-antigen were proven to promote AMP resistance in B. cenocepacia and Brucella abortus using mutants

that lack the respective sugar structures in LPS [180, 181].

Surface modification to counteract the AMPs activity occur very often also in the cytoplasmic membrane as it is the target of many antimicrobial peptides. In the eventuality that the AMP successfully passes the cell wall and reaches this membrane, it is attracted to its surface by the negative charge of the lipids composing it. In particular, the cytoplasmic membrane is rich in phosphatidylglycerol (PG) and diphosphatidylglycerol (DPG, also called cardiolipin) lipids: their negative charge can be masked by amino-acylation of the PG head group, so that the final compound repels AMPs through electrostatic interaction [182]. Usually the group added is a Lysine [183], but Alanine is commonly chosen as well [184].

Finally, also the rigidity of the cytoplasmic membrane can be enhanced, by an increase of saturated acyl chains and has been proven to confer resistance [185, 186], though the precise mechanisms underlying the connections are still partially unclear.

It must be highlighted that often resistant bacteria employs many of the aforementioned strategies at the same time, for example the modification of the surface charge together with the modification of other membrane components for a decreased recognition and augmented rigidity [187].

1.3.4 Principles of AMP design

It was already highlighted in Section 1.3.2 that the classification of AMPs provided knowledge on which characteristics a sequence must have to perform an antimicrobial function. At the present state of the art, several databases exist gathering AMPs and subclasses of them, like membrane active, biofilm active or haemolytic peptides [?]. Based on the increasing amount of data, it is now possible to identify five features which, comprehensively, discriminate AMPs with respect to non antimicrobial peptides:

• Structure: as mentioned before, both α -helical and β -sheet rich AMPs exist, as well as mixed structures. Short helix (~ 22 amino acids) [?] and short β -sheet (~ 10 amino acids) [?] are particularly common and the structural difference is reflected in slightly different mechanisms of actions. When screening a peptide to identify its AMP-likeness, it must

be considered that some sequences may rearrange in proximity of the membrane, thus their structure in solution does not reflect their active conformation.

- Charge: AMPs are charged moieties. Usually they present positive charge (up to ~ +10 e), but there are examples of anionic ones [?]. Their potency is related to the amount of charge each unit possesses, however an increased charge may promote haemolytic activity as well.
 [?]
- **Hydrophobicity**: together with charged amino acids, AMPs contain also hydrophobic residues, usually with abundance of aromatic chains and specifically Tryptophan. Indeed membrane active peptides must insert into the lipid core of membranes, which is an hydrophobic environment, therefore having such residues help them in anchoring in such region.
- Amphipathicity: to host both the charged and hydrophobic residues, most AMPs organise themselves in an amphiphatic structure, i.e. the two types of amino acids side chains are located on the opposite side of the peptide. The usefulness of this segregation in the anchoring and pore formation mechanism has been explained in Section 1.2.2.
- Solubility AMPs needs to have a good solubility to prevent aggregation in the aqueous environment they float in before arriving to the membrane. Indeed aggregation would impede their optimal interaction with the membrane.
- Sequence motifs: finally a long debate exist on whether the effectiveness of AMPs is related to particular sequence motifs or only to the overall amino acid composition. Statistical methods are trying to extract relevant pattern from the databases available, however the details of the structure-activity relationship are still unclear for most AMPs.

Based on the above features, it is possible to make prediction on whether an amino acid sequence is antimicrobial or not. Several online servers are available to evaluate the antimicrobial properties of user provided sequences: the output is generally a score of how likely the peptide is to have such function. —(name

server) provides also an indication on whether the peptide is considered more active against Gram-positive or Gram-negative bacteria. However, it is still impossible to foresee the precise efficacy and mechanism of action of an AMP from its sequence only.

The knowledge of such structure-activity relationship would be beneficial to find new, better performing AMPs. Indeed design of new AMP sequences aims at improving some specific characteristics:

- specificity against particular bacterial species;
- **stability** against the action of protease, thus allowing a longer residence time in the body;
- low cytotoxicity at the therapeutic dose required (so an high therapeutic index).

The need for such improved peptides lies in the fact that in their natural form they constitute the first broad spectrum defence our body employs against infectious bacteria and are often of mild potency. However, foreseeing their application as future drugs, it is desirable to tailor them to fulfil different criteria according to the infection to treat. Several methodological approaches to AMP design are possible, and they can be grouped in three main lines: template based studies, biophysical studies and virtual screening [188].

Template based studies The main idea behind template based methods consists in employing existing antimicrobial sequences and modifying them in the direction of more potency or less toxicity. The most widely explored templates are cecropin, magainin and protegrin for their short sequences and because the action and structure has been well characterised [189–192].

Ideally, an amino acid scanning of all the residues in an AMP provides information on the role of each of them thus prompting at the most suitable mutations. High-throughput methods allow nowadays for such thorough investigation in the case of short AMPs [193, 194]; similarly, less resource consuming Alanine scannings point at the most critical residues on which all the mutations can be tested [195–197]. Earlier studies focused on a simpler approach to enhance charge and amphiphilicity of the peptides, as these characteristics are deemed crucial in their effectiveness (see the paragraph above) [189–192].

Finally, the addition of acyl moieties have been shown to improve the performances of AMPs, as these can provide the necessary hydrophobic domains that, together with charged amino acids, allow an amphiphatic structure in short peptides [198–200]. The above methods however can not take into account the interplay between two residues, while the paired mutation of two of them can give optimal results with respect to a single intervention. Furthermore there is little to no information on the three dimensional structure of the mutated peptide. Without such information, it is difficult to extract general rules on why some mutations work better and often the results of these studies give indeed enhanced AMP, but cannot be generalised to other sequences. Only recently a structure based approach has been developed to integrate structural information on template based models and design peptides active against many bacterial lines at the same time [201].

An example of template design combining chemical and case specific structural information comes from Jiang et al. [202] who designed AMPs with improved selectivity for bacterial membrane. Starting from a synthetic broad spectrum AMP with high toxicity, the positioning of positively charged residues at the centre of the non polar face of the amphipathic α -helix reduced its haemolytic activity while improving its therapeutic index. This proves that charge and structure features do affect the antimicrobial activity, but more work needs to be done to obtain generalised design rules.

Another approach consists in focussing on minimal antimicrobial blocks: several investigations proved the importance of single residues and their intercalated pattern in natural and designed AMPs. In particular, natural AMPs are rich in Tryptophan and Arginine residues [110], while synthetic ones have been produced with Lysines in combination with Leucine, or Arginine with Valine to produce amphipathic helices [203]. Furthermore, polyarginine are long known for being cell penetrating peptides [204].

An effort to extract principles from these examples is represented by text based model where amino acids constitutes the letters and patterns occurring in natural AMPs the grammar rules [?]. This approach can benefit of the improving size of peptide database, together with the advancement in text mining technology and dedicated machine learning algorithms, bringing to the streamlined selection of promising sequence to investigate further [?].

In general, the advantage of template based methods is in the reduced number of sequences to test, with decrease cost, as only a subspace of them is explored, namely the ones close to the original template.

Biophysical studies Biophysical studies aim at understanding the functioning of AMPs by investigating their structure. Free energy perturbation, molecular dynamics (MD) simulations and thermodynamics calculations can all provide knowledge on how the three dimensional arrangement of residues is important to allow their functional role. Contrary to sequence based methods, these techniques give an insight into the mechanism of action of an AMP: free energy perturbation allows for example to pinpoint the interactions that stabilise a structure, while molecular dynamics simulations can show why a particular residue is favourably binding to the membrane. Some mechanistic hypothesis can be deduced from the analysis of many AMPs sequences, but it is only zooming into the details that these hypothesis can be confirmed. Moreover, structural information is crucial to discriminate cases in which similar sequence stretches behave differently due to the environment around them, as this information is necessarily lost in a sequence only approach.

The drawbacks of such techniques lay in their computational cost. All of them can approach systems with a limited size, and simulations can access short (microseconds) time scales preventing the reproduction of phenomena of the order of millisecond (a detailed overview of the state of the art, advantages and drawback of MD simulations will be given in Chapter –). For these reasons, such techniques have been applied to fewer systems in comparison to sequence based screenings, and only few mutations have been tested and compared in silico. Therefore the strength of biophysical studies does not lay in the power of analysing large dataset, but rather in the fact that, as they exploit the whole information available (sequence, structure, chemistry), they can single out the interactions that are crucial for a mechanism, clarifying whether they are peculiar of a given local environment or they can be maintained elsewhere. In this respect, they provide a generalisable knowledge applicable to different systems and thus to the design of novel AMPs at the atomic level.

An example of how MD simulations shed light on AMP-membrane interactions is given by the protegrine peptide: porcine protegrin is a β -hairpin AMP which is though to act through pore formation. A model for the de-

tailed steps of such mechanism was obtained by simulations and proposed a non trivial process of electrostatic attraction to the anionic membrane followed by dimerisation and subsequent insertion into the membrane. Finally, inserted peptides form large aggregates that lead to transmembrane pores formation [?]. Steps further were taken for ovispirin [?], indolicidin [?] and temporin [205] sequences, designing and testing, computationally and then experimentally, mutants of the original sequence with improved activity and decreased haemolytic activity.

Virtual screening Contrary to biophysical assay, virtual screening methods are employed to analyse a large number of sequence, when an experimental or computational test of all of them would be prohibitive. The key concept of these methods consists in the identifications of some descriptors which allow to predict the potency of the sequence: from the analysis of a database of AMP with known activity, a model is created and used to score novel synthetic sequences.

These methods are witnessing new popularity due to the recent evolution of machine learning (ML) techniques: if originally they relied on regression methods, in the past three decades artificial neural network have been extensively applied to the problem (for a historically informed review see Table 1 in Ref. [188]). Machine learning appears particularly suitable to the task as the potency of AMPs is certainly determined by the combination of many factors, and it is difficult to properly weight them and identify the predominant ones in each context.

Therefore ML algorithm are trained on a set of AMPs sequences labelled by their potency, where the sequence (or each amino acids) can be characterised by many different properties: partial charge, hydrophobicity, amphiphilicity and molecular weight are the most intuitive ones; but also by experimental measures of pK (the logarithm of the dissociation constant), pI (the isoelectric point), nuclear magnetic resonance data and chromatographic indices retention time in a given chromatography column, octanolwater partition fraction or circular dichroism. Finally, other theoretically computed features as van der Waals surface area and hardness (the energy required to remove the outermost electron) can be considered as well. The more the input features to consider, the more expensive is to train the model, but an higher accuracy can

be meet: the power of such approach is exactly in the identification of relevant features traditionally overlooked. At the same time though, the output is likely providing complex descriptors (i.e. combinations of many features), thus of difficult interpretation. This is why the step of features selection is important and more than one model is trained to identify the minimum set giving satisfactory agreement with the data.143,144 In principle, having complex descriptors is not a problem as rather than guide the design, they can be used to scores all possible combinatorial sequences of the desired length to identify the best ones. In practise this can be difficult even for such an in silico screening because of their number: as a meter, all the possible amino acid combinations on a 10 residues sequence are of the order of 10¹¹. A possible solution is constituted by evolutionary search methods in the sequence space: analogously to an energy minimization process in space, single amino acid mutations are attempted and the fitness of the new sequence computed, based on the model generated via ML. The move is accepted if it proceeds toward an improved fitness, i.e. a maximum in the fitness landscape.

Another obstacle to these procedures is given by the fact that the more features one wants to consider, the more sequences need to be given as input to the algorithm: nowadays, high-throughput synthesis methods, together with surrogate measures of bacterial killing, such as lipid vesicle experiments 147 or the diminished energy dependent luminescence of bacteria constitutively expressing luciferase 90 (rather that Minimum Inhibitory Concentration assays 148), allow for quick screening of many of those. This was employed by Cherkasov et al. [?], as they assessed the antimicrobial properties of thousands of 9 residues sequences and trained a neural network on the outcome, and then scored novel sequences with good accuracy as proved by experiments. More recently, new studies are exploring novel machine learning approaches, with the aim of selecting sequences for experimental assays [206]; moreover, some work suggests the combined use of machine learning and molecular dynamics for an improved selection of interesting peptides.

1.3.5 Clinical applications

Antimicrobial peptides have been studies for many year, however the push to capitalise them to get compounds viable for the clinical stage has been delayed by many factors, including high production costs, lack of interest in the face of more potent small molecules which were deemed more economically advantageous by pharmaceutical companies. The constant creeping of AM resistance though has focussed more effort on this class of compounds, mainly from small biopharmaceutical companies and at present several of them are in clinical trials, in phase 1 or 2 [?].

The two major problems encountered so far for the AMPs sequences in trial are their liability to proteolytic degradation, and their unknown toxicology profile when administered systemically [?]. For the last reason in particular, many of them in are trial for topical use only against skin infection, while they are deemed unsuitable for internal administration. Design of novel AMPs can be tailored to improve the liability to degradation, for example introducing D-amino acids, non natural amino acid analogues of opposite chirality, which, with appropriate formulations, are mimetic to the immune system [??], and ML protocols can help in pre-screening its toxicity.

Overall, antimicrobial peptides remains a promising tool to counteract infections and as their design is still - comparatively - in its infancy, there is room to explore novel applications and synthesise improved sequences apt to get to the clinical stage.

1.4 Closing the circle: an antimicrobial drug delivery vehicle

Twice in this introduction peptide design has been brought to the reader's attention. First, design can obtain self-assembling building blocks for the formation of delivery scaffolds. Second, it can produce antimicrobial peptides with improved potency or selectivity, and/or reduced toxicity. As design is not bound to the natural rules, it can foresee and imagine multifunctional materials which are not observed in nature. In particular, the introduction above poses the question of whether it is possible to engineer peptides able to perform both functions at once.

Such self-assembling antimicrobial compounds would have a twofold interest for medical applications. First of all, self-assembly is functional to the antimicrobial activity: many AMP sequences have a weak potency, and only an high (critical) concentration can trigger the bactericidal mechanism. This is intuitive in the case of the carpet model strategy (see Section 1.3.2), where

AMPs lay homogeneously on the surface of the bacterial membrane and breaks it upon sufficient coverage of the area. Also the barrel-stave and toroidal pore models rely on the mutual interaction between peptides to maintain the pore edges. As a general rule, as AMPs are in general positively charged, the presence of many copies of a sequence enhance the local electric field and charge imbalance, which are critical to the membrane stability. Therefore the local AMP concentration is crucial to initiate the membrane disruption and a self-assembling sequence will amplify the AMP effects. Second, in order for the assembly to be able to perform the additional delivery function, it must be able to either organise in a tailored structure, rather than an amorphous aggregate (for example a capsule able to host a drug), or to co-assemble with the cargo of interest.

Out of all the possible applications, the most promising is perhaps the use of such vehicles to deliver drugs for metabolic or genetic diseases: while the cargo tackle a defect of the host system, the vehicle can counteract the proliferation of bacteria. This is particularly important in situation where the host immune response is weakened and thus infections normally harmless can spread and cause damage. At this point it must be noticed that the cargo is not bound to be a small molecule roduced the concept of gene-therapy. Up to this stage, delivery vehicles were mentioned in the context of small drug delivery, but the panel of active and beneficial molecules we can deliver to the body and which need a proper coating is much broader. In section – we briefly mentioned novel

And ultimately, it would be possible to co-assemble AMPs with other agents (either genetic material or drug) to perform delivery.

say it is interesting to do gene therapy (and drug delivery) It must be noticed that delivery systems are designed also to assist molecules other than small drugs. Indeed in the past forty years many organic biomolecules proved to have medical potential, extending the research of drugs beyond the small molecule paradigm.

[Design: merging of the two properties together] [What: rationale for exploring this direction]

1.4.1 Choice of the AMP lactoferricin

1.4.2 A viable systems: experimental background

[How/methodology: in general why experiments are limited] [My case, what we do know and how MD can help] [How: introduction on in general how MD can help (but methods details in the relative chapter)]

simulation mammal membrane [207] SELF assembly

Bits The specificity of these antibiotics allows generally to selectively target the desire species of bacteria, however it has the drawback that small modification in the target can easily invalidate the effect of the drug.

The typical time at which this resilient population appears varies greatly according to the mechanism targeted by the drug, but also and especially by how this is administered, resulting in the modern ways of treating infections a particularly fertile ground to train drug-resistant bacteria. Key reasons of the acceleration of the mechanism are the more extensive - and sometimes unjustified - use of antibiotics, both in the doses used for patients and in the world wide coverage that nowadays can, luckily, been reached. It must not be forgotten that a large portion, if not the majority, of antibiotics do not come from human treatments but actually from agriculture and breeding. This increased pressure allow for a fast spread of resistance, but the emergency comes from the parallel lack of new discoveries of novel drugs. The few discovered and approve are kept as last resources and used carefully, both to prevent insurgence of resistance as long as possible and for their collateral effects: indeed, the inefficacy of other drugs has pushed pharmaceutical industry to resort to compounds discarded in the first screening because too toxic for the human body.

This point - whether and how it is possible to engineer peptides for specific functions - will be a central theme of this work. The engineering process must always be connected with the function the carrier has to perform, and its relation with its cargo: the drug.

we know that some enter the membrane because there are the cell penetrating peptides so use database [start chapter on this]

Gandhi2014 for nanocarrier for siRNA/miRNA

Appendices

A.1 On the derivation of the GP predictive distribution

This appendix gives a sketch of the procedure by which Eq. (??) is obtained, which substantially relies on the properties of multivariate Gaussian distributions. For full details on this one can consult the excellent Refs. [?] and [?].

Bibliography

- [1] Artificial intelligence: Go master Lee Se-dol wins against AlphaGo program BBC News. URL https://www.bbc.co.uk/news/technology-35797102.
- [2] Rossi, G., Manfrin, A. & Lutolf, M. P. Progress and potential in organoid research. *Nature Reviews Genetics* **19**, 671–687 (2018). URL http://www.nature.com/articles/s41576-018-0051-9.
- [3] Hopkins ABX Guide. URL http://www.hopkins-abxguide.org/.
- [4] Strebhardt, K. & Ullrich, A. Paul Ehrlich's magic bullet concept: 100 years of progress. *Nature Reviews Cancer* **8**, 473–480 (2008). URL http://www.nature.com/articles/nrc2394.
- [5] Masaoka, Y., Tanaka, Y., Kataoka, M., Sakuma, S. & Yamashita, S. Site of drug absorption after oral administration: Assessment of membrane permeability and luminal concentration of drugs in each segment of gastrointestinal tract. European Journal of Pharmaceutical Sciences 29, 240–250 (2006). URL https://www.sciencedirect.com/science/article/pii/S0928098706001709?via{%}3Dihub.
- [6] Mitragotri, S., Burke, P. A. & Langer, R. Overcoming the challenges in administering biopharmaceuticals: formulation and delivery strategies. Nature Reviews Drug Discovery 13, 655–672 (2014). URL http://www.nature.com/articles/nrd4363.
- [7] Krol, S. Challenges in drug delivery to the brain: Nature is against us. Journal of Controlled Release 164, 145-155 (2012). URL https://www.sciencedirect.com/science/article/pii/S0168365912003999.

[8] Pattni, B. S. & Torchilin, V. P. Targeted Drug Delivery Systems: Strategies and Challenges. 3-38 (Springer, Cham, 2015). URL http://link.springer.com/10.1007/978-3-319-11355-5{_}1.

- [9] Jain, S. & Edwards, M. J. Advances and challenges in the development of drug delivery systems A European perspective (2016). URL https://www.semanticscholar.org/paper/Advances-and-challenges-in-the-development-of-drug-Jain-Edwards/51a16060d641078aac675e3a9c2ab8e7b1effbfa.
- [10] Hughes, G. A. Nanostructure-mediated drug delivery. Nanomedicine: Nanotechnology, Biology and Medicine 1, 22-30 (2005). URL https://www.sciencedirect.com/science/article/pii/S1549963405000122?via{%}3Dihub.
- [11] Singh, P. et al. Gold Nanoparticles in Diagnostics and Therapeutics for Human Cancer. International journal of molecular sciences 19 (2018). URL http://www.ncbi.nlm.nih.gov/pubmed/29986450http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC6073740.
- [12] Boisselier, E. & Astruc, D. Gold nanoparticles in nanomedicine: preparations, imaging, diagnostics, therapies and toxicity. *Chemical Society Reviews* 38, 1759 (2009). URL http://xlink.rsc.org/?DOI=b806051g.
- [13] Erol, O. et al. Recent Advances in Bioactive 1D and 2D Carbon Nanomaterials for Biomedical Applications. Nanomedicine:

 Nanotechnology, Biology and Medicine (2017). URL http://www.ncbi.nlm.nih.gov/pubmed/28552644http://linkinghub.elsevier.com/retrieve/pii/S1549963417300898.
- [14] Depan, D., Shah, J. & Misra, R. Controlled release of drug from folate-decorated and graphene mediated drug delivery system: Synthesis, loading efficiency, and drug release response. *Materials Science and Engineering: C* 31, 1305–1312 (2011). URL https://www.sciencedirect.com/science/article/pii/S0928493111001159.
- [15]Lammers, T. et al. Simultaneous delivery of doxorubicin tumors in vivo and gemcitabine to using prototypic polymeric drug carriers. **30**, 3466 - 3475(2009).*Biomaterials*

URL http://www.ncbi.nlm.nih.gov/pubmed/19304320https://linkinghub.elsevier.com/retrieve/pii/S0142961209002324.

- [16] Liechty, W. B., Kryscio, D. R., Slaughter, B. V. & Peppas, N. A. Polymers for drug delivery systems. Annual review of chemical and biomolecular engineering 1, 149-73 (2010). URL http://www.ncbi.nlm.nih.gov/pubmed/22432577http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3438887.
- [17] Kawakatsu, T. Statistical Physics of Polymers: an Introduction (Springer Berlin Heidelberg, 2004).
- [18] Nicolas, J., Mura, S., Brambilla, D., Mackiewicz, N. & Couvreur, P. Design, functionalization strategies and biomedical applications of targeted biodegradable/biocompatible polymer-based nanocarriers for drug delivery. *Chem. Soc. Rev.* 42, 1147–1235 (2013). URL http://xlink.rsc.org/?DOI=C2CS35265F.
- [19] Nair, L. S. & Laurencin, C. T. Biodegradable polymers as biomaterials. Progress in Polymer Science 32, 762-798 (2007). URL https://www.sciencedirect.com/science/article/pii/S0079670007000664.
- [20] Rao, S. H., Harini, B., Shadamarshan, R. P. K., Balagangadharan, K. & Selvamurugan, N. Natural and synthetic polymers/bioceramics/bioactive compounds-mediated cell signalling in bone tissue engineering. *International journal of biological macromolecules* 110, 88–96 (2018). URL http://www.ncbi.nlm.nih.gov/pubmed/28917940.
- [21] Yoo, J.-W., Irvine, D. J., Discher, D. E. & Mitragotri, S. Bio-inspired, bioengineered and biomimetic drug delivery carriers. *Nature Reviews Drug Discovery* **10**, 521–535 (2011). URL http://www.nature.com/articles/nrd3499.
- [22] Yingchoncharoen, P., Kalinowski, D. S. & Richardson, D. R. Lipid-Based Drug Delivery Systems in Cancer Therapy: What Is Available and What Is Yet to Come. *Pharmacological reviews* **68**, 701–87 (2016). URL http://www.ncbi.nlm.nih.gov/pubmed/27363439http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4931871.

[23] Bunker, A., Magarkar, A. & Viitala, T. Rational design of liposomal drug delivery systems, a review: Combined experimental and computational studies of lipid membranes, liposomes and their PEGylation. BBA - Biomembranes 1858, 2334–2352 (2016). URL http://dx.doi.org/10.1016/j.bbamem.2016.02.025.

- [24] Pattni, B. S., Chupin, V. V. & Torchilin, V. P. New Developments in Liposomal Drug Delivery. *Chemical Reviews* **115**, 10938–10966 (2015). URL http://pubs.acs.org/doi/10.1021/acs.chemrev.5b00046.
- [25] Jain, S. & Pillai, J. Bacterial membrane vesicles as novel nanosystems for drug delivery. *International journal of nanomedicine* 12, 6329-6341 (2017). URL http://www.ncbi.nlm.nih.gov/pubmed/28919737http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC5587191.
- [26] Linko, V., Ora, A. & Kostiainen, M. A. DNA Nanostructures as Smart Drug-Delivery Vehicles and Molecular Devices. *Trends in Biotechnology* 33, 586-594 (2015). URL https://www.sciencedirect.com/science/article/pii/S0167779915001614.
- [27] Douglas, S. M., Bachelet, I. & Church, G. M. A Logic-Gated Nanorobot for Targeted Transport of Molecular Payloads. Science 335, 831– 834 (2012). URL http://www.sciencemag.org/cgi/doi/10.1126/ science.1214081.
- [28] Zhang, Q. et al. DNA Origami as an <i>In Vivo</i> Drug Delivery Vehicle for Cancer Therapy. ACS Nano 8, 6633-6643 (2014). URL http://pubs.acs.org/doi/10.1021/nn502058j.
- [29] Jiang, Q. et al. DNA Origami as a Carrier for Circumvention of Drug Resistance. Journal of the American Chemical Society 134, 13396–13403 (2012). URL http://pubs.acs.org/doi/10.1021/ja304263n.
- [30] Lobo, F. P. et al. Virus-Host Coevolution: Common Patterns of Nucleotide Motif Usage in Flaviviridae and Their Hosts. PLoS ONE 4, e6282 (2009). URL http://dx.plos.org/10.1371/journal.pone. 0006282.

[31] Lauer, K. B., Borrow, R. & Blanchard, T. J. Multivalent and Multipathogen Viral Vector Vaccines. *Clinical and Vaccine Immunology* **24**, e00298–16 (2017). URL https://cvi.asm.org/content/24/1/e00298-16.

- [32] Daya, S. & Berns, K. I. Gene therapy using adeno-associated virus vectors. Clinical microbiology reviews 21, 583-93 (2008). URL http://www.ncbi.nlm.nih.gov/pubmed/18854481http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2570152.
- [33] Büning, H. & Schmidt, M. Adeno-associated Vector Toxicity-To Be or Not to Be? *Molecular therapy: the journal* of the American Society of Gene Therapy 23, 1673-1675 (2015). URL http://www.ncbi.nlm.nih.gov/pubmed/26606658http://www. pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4817949.
- [34] Smalley, E. First AAV gene therapy poised for landmark approval. *Nature Biotechnology* **35**, 998–999 (2017). URL http://www.nature.com/articles/nbt1117-998.
- [35] Wu, W., Hsiao, S. C., Carrico, Z. M. & Francis, M. B. Genome-free viral capsids as multivalent carriers for taxol delivery. *Angewandte Chemie (International ed. in English)* 48, 9493-7 (2009). URL http://www.ncbi.nlm.nih.gov/pubmed/19921725http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2919582.
- [36] Fan, T., Yu, X., Shen, B. & Sun, L. Peptide Self-Assembled Nanostructures for Drug Delivery Applications. *Journal of Nanomaterials* **2017**, 1–16 (2017). URL https://www.hindawi.com/journals/jnm/2017/4562474/.
- [37] Habibi, N., Kamaly, N., Memic, A. & Shafiee, H. Self-assembled peptide-based nanostructures: Smart nanomaterials toward targeted drug delivery. *Nano Today* 11, 41–60 (2016). URL https://www.sciencedirect.com/science/article/pii/S1748013216300081.
- [38] Ma, Y., Nolte, R. J. & Cornelissen, J. J. Virus-based nanocarriers for drug delivery. *Advanced Drug Delivery Reviews* **64**, 811–825

(2012). URL https://www.sciencedirect.com/science/article/pii/S0169409X12000087.

- [39] Yan, X., Zhu, P. & Li, J. Self-assembly and application of diphenylalanine-based nanostructures. *Chemical Society Reviews* **39**, 1877 (2010). URL http://xlink.rsc.org/?DOI=b915765b.
- [40] Silva, R. F., Araújo, D. R., Silva, E. R., Ando, R. A. & Alves, W. A. 1-Diphenylalanine Microtubes As a Potential Drug-Delivery System: Characterization, Release Kinetics, and Cytotoxicity. *Langmuir* 29, 10205–10212 (2013). URL http://pubs.acs.org/doi/10.1021/1a4019162.
- [41] King, N. P. *et al.* Accurate design of co-assembling multi-component protein nanomaterials. *Nature* **510** (2014). URL https://www.nature.com/nature/journal/v510/n7503/full/nature13404.html.
- [42] Berman, H. M. et al. The protein data bank. Nucleic Acids Research, 28: 235-242 (2000). URL http://www.rcsb.org/.
- [43] Yeates, T. O. Protein assembles into Archimedean geometry. Nature 569, 340-342 (2019). URL http://www.nature.com/articles/d41586-019-01407-z.
- [44] Malay, A. D. *et al.* An ultra-stable gold-coordinated protein cage displaying reversible assembly. *Nature* **569**, 438–442 (2019). URL http://www.nature.com/articles/s41586-019-1185-4.
- [45] Santos, R. et al. A comprehensive map of molecular drug targets. Nature Reviews Drug Discovery 16, 19-34 (2017). URL http://www.nature.com/articles/nrd.2016.230.
- [46] Kapoor, G., Saigal, S. & Elongavan, A. Action and resistance mechanisms of antibiotics: A guide for clinicians. *Journal of anaesthesiology, clinical pharmacology* **33**, 300–305 (2017). URL http://www.ncbi.nlm.nih.gov/pubmed/29109626http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC5672523.
- [47] A Blair, J. M., Webber, M. A., Baylay, A. J., Ogbolu, D. O. & V Piddock, L. J. Molecular mechanisms of antibiotic resistance. *Nature Publishing*

- Group 13 (2014). URL https://www.nature.com/nrmicro/journal/v13/n1/pdf/nrmicro3380.pdf.
- [48] Birkegård, A. C., Halasa, T., Toft, N., Folkesson, A. & Græsbøll, K. Send more data: a systematic review of mathematical models of antimicrobial resistance. *Antimicrobial resistance and infection control* 7, 117 (2018). URL http://www.ncbi.nlm.nih.gov/pubmed/30288257http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC6162961.
- [49] Niewiadomska, A. M. et al. Population-level mathematical modeling of antimicrobial resistance: a systematic review. BMC Medicine 17, 81 (2019). URL https://bmcmedicine.biomedcentral.com/articles/10.1186/s12916-019-1314-9.
- [50] O 'neill, J. TACKLING DRUG-RESISTANT INFECTIONS GLOBALLY: FINAL REPORT AND RECOMMENDATIONS THE REVIEW ON ANTIMICROBIAL RESISTANCE (2016). URL https://amr-review.org/sites/default/files/160525{_}Finalpaper{_}withcover.pdf.
- [51] Delcour, A. H. Outer membrane permeability and antibiotic resistance. Biochimica et Biophysica Acta (BBA) Proteins and Proteomics 1794, 808-816 (2009). URL https://www.sciencedirect.com/science/article/pii/S1570963908003592?via{%}3Dihub.
- [52] Vargiu, A. V. & Nikaido, H. Multidrug binding properties of the AcrB efflux pump characterized by molecular dynamics simulations. *Proceedings of the National Academy of Sciences of the United States of America* **109**, 20637–42 (2012). URL http://www.ncbi.nlm.nih.gov/pubmed/23175790http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3528587.
- [53] Kojima, S. & Nikaido, H. Permeation rates of penicillins indicate that Escherichia coli porins function principally as nonspecific channels. *Proceedings of the National Academy of Sciences* **110**, E2629–E2634 (2013). URL http://www.ncbi.nlm.nih.gov/pubmed/23798411http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3710850http://www.pnas.org/cgi/doi/10.1073/pnas.1310333110.

[54] Tran, Q.-T., Williams, S., Farid, R., Erdemli, G. & Pearlstein, R. The translocation kinetics of antibiotics through porin OmpC: Insights from structure-based solvation mapping using WaterMap. *Proteins:* Structure, Function, and Bioinformatics 81, 291–299 (2013). URL http://doi.wiley.com/10.1002/prot.24185.

- [55] Tamber, S. & Hancock, R. E. W. On the mechanism of solute uptake in Pseudomonas. Frontiers in bioscience: a journal and virtual library 8, s472–83 (2003). URL http://www.ncbi.nlm.nih.gov/pubmed/12700103.
- [56] Baroud, M. et al. Underlying mechanisms of carbapenem resistance in extended-spectrum β-lactamase-producing Klebsiella pneumoniae and Escherichia coli isolates at a tertiary care centre in Lebanon: role of OXA-48 and NDM-1 carbapenemases. International Journal of Antimicrobial Agents 41, 75–79 (2013). URL https://linkinghub.elsevier.com/retrieve/pii/S0924857912003470.
- [57] Lavigne, J.-P. et al. An adaptive response of Enterobacter aerogenes to imipenem: regulation of porin balance in clinical isolates.

 International Journal of Antimicrobial Agents 41, 130-136 (2013).

 URL http://www.ncbi.nlm.nih.gov/pubmed/23280442https:
 //linkinghub.elsevier.com/retrieve/pii/S0924857912004219.
- etOutbreak [58]Poulou, Α. al.Caused by an Ertapenem-Resistant. CTX-M-15-Producing Klebsiella Sepneumoniae quence Type 101 Clone Carrying an OmpK36 Porin Journal of Clinical Microbiology 51, 3176–3182 (2013). ant. URL http://www.ncbi.nlm.nih.gov/pubmed/23850951http: //www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC3811621http://jcm.asm.org/cgi/doi/10.1128/JCM.01244-13.
- [59] Wozniak, A. et al. Porin alterations present in non-carbapenemase-producing Enterobacteriaceae with high and intermediate levels of carbapenem resistance in Chile. Journal of Medical Microbiology 61, 1270–1279 (2012). URL http://www.ncbi.nlm.nih.gov/pubmed/22700549http://jmm.microbiologyresearch.org/content/journal/jmm/10.1099/jmm.0.045799-0.

[60] Novais, Â. et al. Spread of an OmpK36-modified ST15 Klebsiella pneumoniae variant during an outbreak involving multiple carbapenem-resistant Enterobacteriaceae species and clones. European Journal of Clinical Microbiology & Infectious Diseases 31, 3057–3063 (2012). URL http://www.ncbi.nlm.nih.gov/pubmed/22706513http://link.springer.com/10.1007/s10096-012-1665-z.

- [61] Tangden, T., Adler, M., Cars, O., Sandegren, L. & Lowdin, E. Frequent emergence of porin-deficient subpopulations with reduced carbapenem susceptibility in ESBL-producing Escherichia coli during exposure to ertapenem in an in vitro pharmacokinetic model.

 Journal of Antimicrobial Chemotherapy 68, 1319-1326 (2013).
 URL http://www.ncbi.nlm.nih.gov/pubmed/23478794https://academic.oup.com/jac/article-lookup/doi/10.1093/jac/dkt044.
- [62] Floyd, J. L., Smith, K. P., Kumar, S. H., Floyd, J. T. & Varela, M. F. LmrS Is a Multidrug Efflux Pump of the Major Facilitator Superfamily from Staphylococcus aureus. *Antimicrobial Agents and Chemotherapy* 54, 5406-5412 (2010). URL http://aac.asm.org/cgi/doi/10.1128/AAC.00580-10.
- [63] Hu, R.-M., Liao, S.-T., Huang, C.-C., Huang, Y.-W. & Yang, T.-C. An Inducible Fusaric Acid Tripartite Efflux Pump Contributes to the Fusaric Acid Resistance in Stenotrophomonas maltophilia. *PLoS ONE* 7, e51053 (2012). URL https://dx.plos.org/10.1371/journal.pone.0051053.
- [64] Kim, C. et al. The Mechanism of Heterogeneous Beta-Lactam Resistance in MRSA: Key Role of the Stringent Stress Response. PLoS ONE 8, e82814 (2013). URL http://dx.plos.org/10.1371/journal.pone. 0082814.
- [65] Ogawa, W., Onishi, M., Ni, R., Tsuchiya, T. & Kuroda, T. Functional study of the novel multidrug efflux pump KexD from Klebsiella pneumoniae. *Gene* 498, 177–182 (2012). URL https://linkinghub.elsevier.com/retrieve/pii/S0378111912002107.
- [66] Dolejska, M., Villa, L., Poirel, L., Nordmann, P. & Carattoli, A. Complete sequencing of an IncHI1 plasmid encod-

ing the carbapenemase NDM-1, the ArmA 16S RNA methylase and a resistance-nodulation-cell division/multidrug efflux pump. Journal of Antimicrobial Chemotherapy 68, 34-39 (2013). URL http://www.ncbi.nlm.nih.gov/pubmed/22969080https://academic.oup.com/jac/article-lookup/doi/10.1093/jac/dks357.

- [67]Abouzeed, Y. M., Baucheron, S. & Cloeckaert, ramR efflux-mediated mutations involved in multidrug resistance Salmonella enterica serovar Typhimurium. Antimicroin agents and chemotherapy **52**, 2428 - 34(2008).URL http://www.ncbi.nlm.nih.gov/pubmed/18443112http://www. pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2443889.
- [68] Baucheron, S. et al. Bile-mediated activation of the acrAB and tolC multidrug efflux genes occurs mainly through transcriptional derepression of ramA in Salmonella enterica serovar Typhimurium.

 Journal of Antimicrobial Chemotherapy 69, 2400-2406 (2014).

 URL http://www.ncbi.nlm.nih.gov/pubmed/24816212https://
 academic.oup.com/jac/article-lookup/doi/10.1093/jac/dku140.
- [69] Nikaido, E., Shirosaka, I., Yamaguchi, A. & Nishino, K. Regulation of the AcrAB multidrug efflux pump in Salmonella enterica serovar Typhimurium in response to indole and paraquat. *Microbiology* 157, 648– 655 (2011). URL http://mic.microbiologyresearch.org/content/ journal/micro/10.1099/mic.0.045757-0.
- [70] Hirakawa, H., Inazumi, Y., Masaki, T., Hirata, T. & Yamaguchi, A. Indole induces the expression of multidrug exporter genes in Escherichia coli. *Molecular Microbiology* 55, 1113-1126 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15686558http://doi.wiley.com/10.1111/j.1365-2958.2004.04449.x.
- [71] Billal, D. S., Feng, J., Leprohon, P., Légaré, D. & Ouellette, M. Whole genome analysis of linezolid resistance in Streptococcus pneumoniae reveals resistance and compensatory mutations. *BMC Genomics* **12**, 512 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/22004526http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=

- PMC3212830http://bmcgenomics.biomedcentral.com/articles/10.1186/1471-2164-12-512.
- [72] Gao, W. et al. Two Novel Point Mutations in Clinical Staphylococcus aureus Reduce Linezolid Susceptibility and Switch on the Stringent Response to Promote Persistent Infection. PLoS Pathogens 6, e1000944 (2010). URL http://dx.plos.org/10.1371/journal.ppat.1000944.
- [73]Shore, A. C. et al. Detection of Staphylococcal Cassette Chro-<i><mec</i> Type XICarrying Highly Divergent <i>mecA</i>, <i><math>mecI</i>, <i><math>mecR1</i>, <i>blaZ</i>>, and <i>ccr</i> Genes in Human Clinical Isolates of Clonal Complex 130 Methicillin-Resistant <i>Staphylococcus aureus</i>. Antimicrobial Agents and Chemotherapy 55, 3765–3773 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/21636525http: //www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC3147645http://aac.asm.org/lookup/doi/10.1128/AAC.00187-11.
- [74] Katayama, Y., Ito, T. & Hiramatsu, K. A new class of genetic element, staphylococcus cassette chromosome mec, encodes methicillin resistance in Staphylococcus aureus. *Antimicrobial agents and chemotherapy* 44, 1549-55 (2000). URL http://www.ncbi.nlm.nih.gov/pubmed/10817707http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC89911.
- [75] Kumar, N. et al. Crystal structure of the transcriptional regulator Rv1219c of <i>Mycobacterium tuberculosis</i>. Protein Science 23, 423-432 (2014). URL http://www.ncbi.nlm.nih.gov/pubmed/24424575http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3970893http://doi.wiley.com/10.1002/pro.2424.
- [76] Long, K. S., Poehlsgaard, J., Kehrenberg, C., Schwarz, S. & Vester, B. The Cfr rRNA Methyltransferase Confers Resistance to Phenicols, Lincosamides, Oxazolidinones, Pleuromutilins, and Streptogramin A Antibiotics. *Antimicrobial Agents and Chemotherapy* **50**, 2500–2505 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/16801432http:

- //www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC1489768http://aac.asm.org/cgi/doi/10.1128/AAC.00131-06.
- [77] Vetting, M. W. et al. Structure of QnrB1, a Plasmid-mediated Fluoroquinolone Resistance Factor. Journal of Biological Chemistry 286, 25265-25273 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/21597116http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3137097http://www.jbc.org/lookup/doi/10.1074/jbc.M111.226936.
- [78] Abraham, E. P. & Chain, E. An enzyme from bacteria able to destroy penicillin. 1940. Reviews of infectious diseases 10, 677-8 (1988). URL http://www.ncbi.nlm.nih.gov/pubmed/3055168.
- [79] Livermore, D. Defining an extended-spectrum β-lactamase. Clinical Microbiology and Infection 14, 3-10 (2008). URL http://www.ncbi.nlm.nih.gov/pubmed/18154524https://linkinghub.elsevier.com/retrieve/pii/S1198743X14604717.
- [80] Nordmann, P., Poirel, L., Walsh, T. R. & Livermore, D. M. The emerging NDM carbapenemases. *Trends in Microbiology* **19**, 588-595 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/22078325https://linkinghub.elsevier.com/retrieve/pii/S0966842X11001776.
- [81] Voulgari, Ε., Poulou, Α., Koumaki, V. &Tsakris, Α. Carbapenemase-producing <i>Enterobacteriaceae</i> the storm is finally here. how will timely detection help us fight back? Future Microbiology 8, 27–39 (2013). URL http://www.ncbi.nlm.nih.gov/pubmed/23252491https: //www.futuremedicine.com/doi/10.2217/fmb.12.130.
- [82] Woodford, N., Turton, J. F. & Livermore, D. M. Multiresistant Gramnegative bacteria: the role of high-risk clones in the dissemination of antibiotic resistance. FEMS Microbiology Reviews 35, 736-755 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/21303394https://academic.oup.com/femsre/article-lookup/doi/10.1111/j.1574-6976.2011.00268.x.

[83] Woodford, N. & Johnson, A. P. Global spread of antibiotic resistance: the example of New Delhi metallo-β-lactamase (NDM)-mediated carbapenem resistance. Journal of Medical Microbiology 62, 499–513 (2013). URL http://www.ncbi.nlm.nih.gov/pubmed/23329317http://www.microbiologyresearch.org/content/journal/jmm/10.1099/jmm.0.052555-0.

- Lynch, J. P., Clark, N. M. & Zhanel, G. G. Evolution of antimicrobial resistance Enterobacteriaceae among (focus on extended spectrum β -lactamases and carbapenemases). Expert Opinion on Pharmacotherapy 14, 199–210 (2013). URL http://www.ncbi.nlm.nih.gov/pubmed/23321047http://www. tandfonline.com/doi/full/10.1517/14656566.2013.763030.
- [85] Queenan, A. M. & Bush, K. Carbapenemases: the Versatile-Lactamases. Clinical Microbiology Reviews 20, 440-458 (2007). URL http://cmr.asm.org/cgi/doi/10.1128/CMR.00001-07.
- [86] Queenan, A. M., Shang, W., Flamm, R. & Bush, K. Hydrolysis and Inhibition Profiles of -Lactamases from Molecular Classes A to D with Doripenem, Imipenem, and Meropenem.

 Antimicrobial Agents and Chemotherapy 54, 565-569 (2010).

 URL http://www.ncbi.nlm.nih.gov/pubmed/19884379http:
 //www.pubmedcentral.nih.gov/articlerender.fcgi?artid=
 PMC2798497http://aac.asm.org/cgi/doi/10.1128/AAC.01004-09.
- [87] Tzouvelekis, L. S., Markogiannakis, A., Psichogiou, M., Tassios, P. T. & Daikos, G. L. Carbapenemases in Klebsiella pneumoniae and Other Enterobacteriaceae: an Evolving Crisis of Global Dimensions. Clinical Microbiology Reviews 25, 682-707 (2012). URL http://www.ncbi.nlm.nih.gov/pubmed/23034326http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC3485753http://cmr.asm.org/cgi/doi/10.1128/CMR.05035-11.
- [88] Wright, G. Bacterial resistance to antibiotics: Enzymatic degradation and modification. *Advanced Drug Delivery Reviews* 57, 1451–1470 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/

- 15950313https://linkinghub.elsevier.com/retrieve/pii/S0169409X05000980.
- [89] Norris, A. L. & Serpersu, E. H. Ligand promiscuity through the eyes of the aminoglycoside <i>N</i> 3 acetyltransferase IIa. *Protein Science* 22, 916-928 (2013). URL http://www.ncbi.nlm.nih.gov/pubmed/23640799http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3719086http://doi.wiley.com/10.1002/pro.2273.
- [90] Qin, S. et al. Identification of a Novel Genomic Island Conferring Resistance to Multiple Aminoglycoside Antibiotics in Campylobacter coli. Antimicrobial Agents and Chemotherapy 56, 5332 (2012). URL https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3457361/.
- [91] Mantravadi, P., Kalesh, K., Dobson, R., Hudson, A. & Parthasarathy, A. The Quest for Novel Antimicrobial Compounds: Emerging Trends in Research, Development, and Technologies. *Antibiotics* 8, 8 (2019). URL http://www.ncbi.nlm.nih.gov/pubmed/30682820http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC6466574http://www.mdpi.com/2079-6382/8/1/8.
- [92] Nguyen, L. T., Haney, E. F. & Vogel, H. J. The expanding scope of antimicrobial peptide structures and their modes of action. *Trends in Biotechnology* **29**, 464-472 (2011). URL https://www.sciencedirect.com/science/article/pii/S0167779911000886?via{%}3Dihub.
- [93] Bahar, A. A. & Ren, D. Antimicrobial peptides. *Pharmaceuticals* (Basel, Switzerland) **6**, 1543-75 (2013). URL http://www.ncbi.nlm.nih.gov/pubmed/24287494http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3873676.
- [94] Mahlapuu, M., Håkansson, J., Ringstad, L. & Björn, C. Antimicrobial Peptides: An Emerging Category of Therapeutic Agents. Frontiers in Cellular and Infection Microbiology 6, 194 (2016). URL http://www.ncbi.nlm.nih.gov/pubmed/28083516http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC5186781.

[95] Zhang, L.-j. & Gallo, R. L. Antimicrobial peptides. Current Biology 26, R14-R19 (2016). URL https://www.sciencedirect.com/science/article/pii/S0960982215014098{#}!

- [96] Brogden, K. A. Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria? *Nature Reviews Microbiology* **3**, 238–250 (2005). URL http://www.nature.com/articles/nrmicro1098.
- [97] Hancock, R. E. W. & Sahl, H.-G. Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies. *Nature Biotechnology* **24**, 1551–1557 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/17160061http://www.nature.com/articles/nbt1267.
- [98] Malmsten, M. Interactions of Antimicrobial Peptides with Bacterial Membranes and Membrane Components. *Current topics in medicinal chemistry* **16**, 16–24 (2016). URL http://www.ncbi.nlm.nih.gov/pubmed/26139113.
- [99] Zhang, L., Rozek, A. & Hancock, R. E. Interaction of cationic antimicrobial peptides with model membranes. *The Journal of biological chemistry* **276**, 35714–22 (2001). URL http://www.ncbi.nlm.nih.gov/pubmed/11473117.
- [100] Schmitt, P., Rosa, R. D. & Destoumieux-Garzón, D. An intimate link between antimicrobial peptide sequence diversity and binding to essential components of bacterial membranes. *Biochimica et Biophysica Acta (BBA) Biomembranes* 1858, 958–970 (2016). URL http://www.ncbi.nlm.nih.gov/pubmed/26498397http://linkinghub.elsevier.com/retrieve/pii/S0005273615003430.
- [101] Lin, T.-Y. & Weibel, D. B. Organization and function of anionic phospholipids in bacteria. *Applied Microbiology and Biotechnology* **100**, 4255–4267 (2016). URL http://link.springer.com/10.1007/s00253-016-7468-x.
- [102] Glukhov, E., Stark, M., Burrows, L. L. & Deber, C. M. Basis for selectivity of cationic antimicrobial peptides for bacterial versus mammalian membranes. *The Journal of biological chemistry* **280**, 33960–7 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/16043484.

[103] Spector, A. A. & Yorek, M. A. Membrane lipid composition and cellular function. *Journal of lipid research* **26**, 1015–35 (1985). URL http://www.ncbi.nlm.nih.gov/pubmed/3906008.

- [104] van Meer, G., Voelker, D. R. & Feigenson, G. W. Membrane lipids: where they are and how they behave. Nature reviews. Molecular cell biology 9, 112-24 (2008). URL http://www.ncbi.nlm.nih.gov/pubmed/18216768http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2642958.
- [105] Yeaman, M. R. & Yount, N. Y. Mechanisms of Antimicrobial Peptide Action and Resistance. *Pharmacological Reviews* **55**, 27–55 (2003). URL http://www.ncbi.nlm.nih.gov/pubmed/12615953http://pharmrev.aspetjournals.org/cgi/doi/10.1124/pr.55.1.2.
- [106] Lai, Y. & Gallo, R. L. AMPed up immunity: how antimicrobial peptides have multiple roles in immune defense. *Trends in Immunology* **30**, 131–141 (2009). URL https://www.sciencedirect.com/science/article/pii/S1471490609000052.
- [107] Zasloff, M. Antimicrobial peptides of multicellular organisms. *Nature* 415, 389-395 (2002). URL http://www.ncbi.nlm.nih.gov/pubmed/11807545http://www.nature.com/articles/415389a.
- [108] Matsuzaki, K. Control of cell selectivity of antimicrobial peptides. Biochimica et Biophysica Acta (BBA) - Biomembranes 1788, 1687–1692 (2009). URL https://www.sciencedirect.com/science/article/pii/S0005273608003076.
- [109] Ebenhan, T., Gheysens, O., Kruger, H. G., Zeevaart, J. R. & Sathekge, M. M. Antimicrobial peptides: their role as infection-selective tracers for molecular imaging. *BioMed research international* 2014, 867381 (2014). URL http://www.ncbi.nlm.nih.gov/pubmed/25243191http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4163393.
- [110] Chan, D. I., Prenner, E. J. & Vogel, H. J. Tryptophan- and arginine-rich antimicrobial peptides: Structures and mechanisms of action. Biochimica et Biophysica Acta (BBA) - Biomembranes 1758, 1184–1202

(2006). URL http://www.ncbi.nlm.nih.gov/pubmed/16756942http://linkinghub.elsevier.com/retrieve/pii/S0005273606001404.

- [111] Toke, O. Antimicrobial peptides: New candidates in the fight against bacterial infections. *Biopolymers* **80**, 717–735 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/15880793http://doi.wiley.com/10.1002/bip.20286.
- [112] Yang, L., Harroun, T. A., Weiss, T. M., Ding, L. & Huang, H. W. Barrel-stave model or toroidal model? A case study on melittin pores. *Biophysical journal* 81, 1475-85 (2001). URL http://www.ncbi.nlm.nih.gov/pubmed/11509361http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC1301626.
- [113] Bertelsen, K., Dorosz, J., Hansen, S. K., Nielsen, N. C. & Vosegaard, T. Mechanisms of Peptide-Induced Pore Formation in Lipid Bilayers Investigated by Oriented 31P Solid-State NMR Spectroscopy. *PLoS ONE* 7, e47745 (2012). URL http://dx.plos.org/10.1371/journal.pone. 0047745.
- [114] Lee, M.-T., Chen, F.-Y. & Huang, H. W. Energetics of Pore Formation Induced by Membrane Active Peptides . Biochemistry 43, 3590–3599 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15035629https://pubs.acs.org/doi/10.1021/bi036153r.
- [115] Spaar, A., Münster, C. & Salditt, T. Conformation of peptides in lipid membranes studied by x-ray grazing incidence scattering. Biophysical journal 87, 396-407 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15240474http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC1304361.
- [116] Matsuzaki, K., Murase, O., Fujii, N. & Miyajima, K. An Antimicrobial Peptide, Magainin 2, Induced Rapid Flip-Flop of Phospholipids Coupled with Pore Formation and Peptide Translocation . Biochemistry 35, 11361-11368 (1996). URL http://www.ncbi.nlm.nih.gov/pubmed/8784191https://pubs.acs.org/doi/10.1021/bi960016v.

[117] Hallock, K. J., Lee, D.-K. & Ramamoorthy, A. MSI-78, an analogue of the magainin antimicrobial peptides, disrupts lipid bilayer structure via positive curvature strain. *Biophysical journal* 84, 3052-60 (2003). URL http://www.ncbi.nlm.nih.gov/pubmed/12719236http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC1302867.

- [118] He, K., Ludtke, S. J., Huang, H. W. & Worcester, D. L. Antimicrobial peptide pores in membranes detected by neutron in-plane scattering. *Biochemistry* 34, 15614–8 (1995). URL http://www.ncbi.nlm.nih.gov/pubmed/7495788.
- [119] Matsuzaki, K. et al. Relationship of Membrane Curvature to the Formation of Pores by Magainin 2 . Biochemistry 37, 11856–11863 (1998). URL https://pubs.acs.org/doi/10.1021/bi980539y.
- [120] Matsuzaki, K., Sugishita, K.-i., Harada, M., Fujii, N. & Miyajima, K. Interactions of an antimicrobial peptide, magainin 2, with outer and inner membranes of Gram-negative bacteria. *Biochim*ica et Biophysica Acta (BBA) - Biomembranes 1327, 119-130 (1997). URL https://www.sciencedirect.com/science/article/ pii/S0005273697000515?via{%}3Dihub.
- [121] Shai, Y. Mechanism of the binding, insertion and destabilization of phospholipid bilayer membranes by α-helical antimicrobial and cell non-selective membrane-lytic peptides. Biochimica et Biophysica Acta (BBA) Biomembranes 1462, 55–70 (1999). URL https://www.sciencedirect.com/science/article/pii/S000527369900200X?via{%}3Dihub.
- [122] Ladokhin, A. S. & White, S. H. Detergent-like' permeabilization of anionic lipid vesicles by melittin. *Biochimica et Biophysica Acta* (BBA) Biomembranes 1514, 253-260 (2001). URL https://www.sciencedirect.com/science/article/pii/S0005273601003820.
- [123] Oren, Z. & Shai, Y. Mode of action of linear amphipathic α-helical antimicrobial peptides. Biopolymers 47, 451–463 (1998). URL http://www.ncbi.nlm.nih.gov/pubmed/10333737http://doi.wiley.com/10.1002/{%}28SICI{%}291097-

- 0282{%}281998{%}2947{%}3A6{%}3C451{%}3A{%}3AAID-BIP4{%}3E3. 0.CO{%}3B2-F.
- [124] Gazit, E., Boman, A., Boman, H. G. & Shai, Y. Interaction of the mammalian antibacterial peptide cecropin P1 with phospholipid vesicles. *Biochemistry* 34, 11479–88 (1995). URL http://www.ncbi.nlm.nih.gov/pubmed/7547876.
- [125] Yamaguchi, S. et al. Orientation and Dynamics of an Antimicrobial Peptide in the Lipid Bilayer by Solid-State NMR Spectroscopy. Biophysical Journal 81, 2203-2214 (2001). URL http://www.ncbi.nlm.nih.gov/pubmed/11566791http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC1301692https://linkinghub.elsevier.com/retrieve/pii/S0006349501758687.
- [126] Tang, M. & Hong, M. Structure and mechanism of β-hairpin antimicrobial peptides in lipid bilayers from solid-state NMR spectroscopy. Molecular BioSystems 5, 317 (2009). URL http://xlink.rsc.org/?D0I=b820398a.
- [127] Lehrer, R. I. Primate defensins. *Nature Reviews Microbiology* **2**, 727-738 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15372083http://www.nature.com/articles/nrmicro976.
- [128] Fujii, G., Eisenberg, D. & Selsted, M. E. Defensins promote fusion and lysis of negatively charged membranes. Protein Science 2, 1301-1312 (1993). URL http://www.ncbi.nlm.nih.gov/pubmed/8401215http: //www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC2142441http://doi.wiley.com/10.1002/pro.5560020813.
- [129] Kagan, B. L., Selsted, M. E., Ganz, T. & Lehrer, defensin Antimicrobial peptides form voltage-dependent ionpermeable channels in planar lipid bilayer membranes. Proceedings of the National Academy of Sciences 87, 210–214 (1990). URL http://www.ncbi.nlm.nih.gov/pubmed/1688654http: //www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC53231http://www.pnas.org/cgi/doi/10.1073/pnas.87.1.210.

[130] Takeuchi, K. et al. Channel-forming membrane permeabilization by an antibacterial protein, sapecin: determination of membrane-buried and oligomerization surfaces by NMR. The Journal of biological chemistry 279, 4981–7 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/14630928.

- [131] Dathe, M. & Wieprecht, T. Structural features of helical antimicrobial peptides: their potential to modulate activity on model membranes and biological cells. *Biochimica et biophysica acta* **1462**, 71–87 (1999). URL http://www.ncbi.nlm.nih.gov/pubmed/10590303.
- [132] Yonezawa, A., Kuwahara, J., Fujii, N. & Sugiura, Y. Binding of tachyplesin I to DNA revealed by footprinting analysis: significant contribution of secondary structure to DNA binding and implication for biological action. *Biochemistry* **31**, 2998–3004 (1992). URL http://pubs.acs.org/doi/abs/10.1021/bi00126a022.
- [133] Gifford, J. L., Hunter, H. N. & Vogel, H. J. Lactoferricin. Cellular and Molecular Life Sciences 62, 2588-2598 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/16261252http://link.springer.com/10.1007/s00018-005-5373-z.
- [134] Tomita, M., Takase, M., Bellamy, W. & Shimamura, S. A review: the active peptide of lactoferrin. *Acta paediatrica Japonica: Overseas edition* **36**, 585–91 (1994). URL http://www.ncbi.nlm.nih.gov/pubmed/7825467.
- [135] Hwang, P. M., Zhou, N., Shan, X., Arrowsmith, C. H. & Vogel, H. J. Three-Dimensional Solution Structure of Lactoferricin B, an Antimicrobial Peptide Derived from Bovine Lactoferrin . Biochemistry 37, 4288-4298 (1998). URL http://www.ncbi.nlm.nih.gov/pubmed/9521752http://pubs.acs.org/doi/abs/10.1021/bi972323m.
- [136] Schibli, D. J., Hwang, P. M. & Vogel, H. J. The structure of the antimicrobial active center of lactoferricin B bound to sodium dodecyl sulfate micelles. *FEBS Letters* **446**, 213–217 (1999). URL http://doi.wiley.com/10.1016/S0014-5793{%}2899{%}2900214-8.

[137] Nguyen, L. T., Schibli, D. J. & Vogel, H. J. Structural studies and model membrane interactions of two peptides derived from bovine lactoferricin. *Journal of Peptide Science* 11, 379-389 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/15635665http://doi.wiley.com/10.1002/psc.629.

- [138] Peschel, A. & Sahl, H.-G. The co-evolution of host cationic antimicrobial peptides and microbial resistance. *Nature Reviews Microbiology* 4, 529–536 (2006). URL http://www.nature.com/articles/nrmicro1441.
- [139] Juhas, M. Horizontal gene transfer in human pathogens. *Critical Reviews* in *Microbiology* 41, 101–108 (2015). URL http://www.tandfonline.com/doi/full/10.3109/1040841X.2013.804031.
- [140] Joo, H.-S., Fu, C.-I. & Otto, M. Bacterial strategies of resistance to antimicrobial peptides. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences* **371** (2016). URL http://www.ncbi.nlm.nih.gov/pubmed/27160595http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4874390.
- [141] Sieprawska-Lupa, M. et al. Degradation of human antimicrobial peptide LL-37 by Staphylococcus aureus-derived proteinases. Antimicrobial agents and chemotherapy 48, 4673-9 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15561843http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC529204.
- [142] Teufel, P. & Götz, F. Characterization of an extracellular metalloprotease with elastase activity from Staphylococcus epidermidis. *Journal of Bacteriology* **175**, 4218 (1993). URL https://www.ncbi.nlm.nih.gov/pmc/articles/PMC204852/.
- [143] Schmidtchen, A., Frick, I.-M., Andersson, E., Tapper, H. & Björck, L. Proteinases of common pathogenic bacteria degrade and inactivate the antibacterial peptide LL-37. *Molecular Microbiology* 46, 157–168 (2002). URL http://doi.wiley.com/10.1046/j.1365-2958.2002.03146.x.
- [144] Barańska-Rybak, W., Sonesson, A., Nowicki, R. & Schmidtchen, A. Glycosaminoglycans inhibit the antibacterial activity of LL-37 in biological fluids. *Journal of Antimicrobial Chemotherapy* **57**, 260–

- 265 (2006). URL http://academic.oup.com/jac/article/57/2/260/804780/Glycosaminoglycans-inhibit-the-antibacterial.
- [145] Nelson, D. C., Garbe, J. & Collin, M. Cysteine proteinase SpeB from Streptococcus pyogenes a potent modifier of immunologically important host and bacterial proteins. *Biological Chemistry* **392**, 1077–88 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/22050223https://www.degruyter.com/view/j/bchm.2011.392.issue-12/bc.2011. 208/bc.2011.208.xml.
- [146] Frick, I.-M. et al. Constitutive and Inflammation-Dependent Antimicrobial Peptides Produced by Epithelium Are Differentially Processed and Inactivated by the Commensal <i>Finegoldia magna</i> and the Pathogen <i>Streptococcus pyogenes</i> The Journal of Immunology 187, 4300–4309 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/21918193http://www.jimmunol.org/lookup/doi/10.4049/jimmunol.1004179.
- [147] Selsted, M. E. & Harwig, S. S. Determination of the disulfide array in the human defensin HNP-2. A covalently cyclized peptide. *The Journal of biological chemistry* **264**, 4003–7 (1989). URL http://www.ncbi.nlm.nih.gov/pubmed/2917986.
- [148] Stumpe, S., Schmid, R., Stephens, D. L., Georgiou, G. & Bakker, E. P. Identification of OmpT as the protease that hydrolyzes the antimicrobial peptide protamine before it enters growing cells of Escherichia coli. *Journal of bacteriology* **180**, 4002–6 (1998). URL http://www.ncbi.nlm.nih.gov/pubmed/9683502http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC107389.
- [149] Biegeleisen, K. The probable structure of the protamineDNA complex. Journal of Theoretical Biology 241, 533-540 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/16442565https://linkinghub.elsevier.com/retrieve/pii/S0022519305005473.
- [150] Sol, A. et al. Actin enables the antimicrobial action of LL-37 peptide in the presence of microbial proteases. The Journal of biological chemistry 289, 22926–41 (2014). URL

- http://www.ncbi.nlm.nih.gov/pubmed/24947511http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4132794.
- [151] Groisman, E. A., Parra-Lopez, C., Salcedo, M., Lipps, C. J. & Heffron, F. Resistance to host antimicrobial peptides is necessary for Salmonella virulence. Proceedings of the National Academy of Sciences of the United States of America 89, 11939 (1992). URL https://www.ncbi.nlm.nih.gov/pmc/articles/PMC50673/.
- [152] Parra-Lopez, C., Baer, M. T. & Groisman, E. A. Molecular genetic analysis of a locus required for resistance to antimicrobial peptides in Salmonella typhimurium. *The EMBO journal* 12, 4053–62 (1993). URL http://www.ncbi.nlm.nih.gov/pubmed/8223423http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC413698.
- [153] Mason, K. M., Munson, R. S., Jr. & Bakaletz, L. O. A Mutation in the sap Operon Attenuates Survival of Nontypeable Haemophilus influenzae in a Chinchilla Model of Otitis Media. *Infection and Immunity* 73, 599 (2005). URL https://www.ncbi.nlm.nih.gov/pmc/articles/ PMC538956/.
- [154] Taggart, C. C. et al. Inactivation of Human -Defensins 2 and 3 by Elastolytic Cathepsins. The Journal of Immunology 171, 931-937 (2003). URL http://www.ncbi.nlm.nih.gov/pubmed/12847264http://www.jimmunol.org/cgi/doi/10.4049/jimmunol.171.2.931.
- [155] Bokarewa, M. & Tarkowski, A. Human α-defensins neutralize fibrinolytic activity exerted by staphylokinase. Thrombosis and Haemostasis 91, 991–999 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15116261http://www.thieme-connect.de/DOI/DOI?10.1160/TH03-11-0696.
- [156] Jin, T. et al. Staphylococcus aureus Resists Human Defensins by Production of Staphylokinase, a Novel Bacterial Evasion Mechanism. The Journal of Immunology 172, 1169–1176 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/14707093.
- [157] Costerton, J. W., Stewart, P. S. & Greenberg, E. P. Bacterial biofilms: a common cause of persistent infections. *Science (New York, N.Y.)*

284, 1318-22 (1999). URL http://www.ncbi.nlm.nih.gov/pubmed/10334980.

- [158] Jolivet-Gougeon, A. & Bonnaure-Mallet, M. Biofilms as a mechanism of bacterial resistance. Drug Discovery Today: Technologies 11, 49-56 (2014). URL http://www.ncbi.nlm.nih.gov/pubmed/24847653https://linkinghub.elsevier.com/retrieve/pii/S1740674914000043.
- [159] Nickel, J. C., Ruseska, I., Wright, J. B. & Costerton, J. W. Tobramycin resistance of Pseudomonas aeruginosa cells growing as a biofilm on urinary catheter material. *Antimicrobial agents and chemotherapy* 27, 619–24 (1985). URL http://www.ncbi.nlm.nih.gov/pubmed/3923925http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC180108.
- [160] Mah, T. F. & O'Toole, G. A. Mechanisms of biofilm resistance to antimicrobial agents. *Trends in microbiology* **9**, 34–9 (2001). URL http://www.ncbi.nlm.nih.gov/pubmed/11166241.
- [161] Wang, X., Preston, J. F. & Romeo, T. The pgaABCD locus of Escherichia coli promotes the synthesis of a polysaccharide adhesin required for biofilm formation. *Journal of bacteriology* **186**, 2724–34 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15090514http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC387819.
- [162] Vuong, C. et al. Polysaccharide intercellular adhesin (PIA) protects Staphylococcus epidermidis against major components of the human innate immune system. Cellular microbiology 6, 269–75 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/14764110.
- [163] Campos, M. A. et al. Capsule polysaccharide mediates bacterial resistance to antimicrobial peptides. Infection and immunity 72, 7107-14 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/15557634http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC529140.
- [164] Llobet, E., Tomas, J. M. & Bengoechea, J. A. Capsule polysaccharide is a bacterial decoy for antimicrobial peptides. *Microbiology* 154, 3877–3886

(2008). URL http://www.ncbi.nlm.nih.gov/pubmed/19047754http://mic.microbiologyresearch.org/content/journal/micro/10. 1099/mic.0.2008/022301-0.

- [165] Batoni, G., Maisetta, G., Lisa Brancatisano, F., Esin, S. & Campa, M. Use of Antimicrobial Peptides Against Microbial Biofilms: Advantages and Limits. *Current Medicinal Chemistry* 18, 256-279 (2011). URL http://www.eurekaselect.com/openurl/content.php?genre=article{&}issn=0929-8673{&}volume=18{&}issue=2{&}spage=256.
- [166] Strempel, N., Strehmel, J. & Overhage, J. Potential Application of Antimicrobial Peptides in the Treatment of Bacterial Biofilm Infections. Current Pharmaceutical Design 21, 67-84 (2014). URL http://www.eurekaselect.com/openurl/content.php?genre=article{&}issn=1381-6128{&}volume=21{&}issue=1{&}spage=67.
- [167] Joo, H.-S. & Otto, M. Molecular Basis of In Vivo Biofilm Formation by Bacterial Pathogens. *Chemistry & Biology* **19**, 1503-1513 (2012). URL http://www.ncbi.nlm.nih.gov/pubmed/23261595http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3530155https://linkinghub.elsevier.com/retrieve/pii/S1074552112004231.
- [168] Di Luca, M., Maccari, G. & Nifosì, R. Treatment of microbial biofilms in the post-antibiotic era: prophylactic and therapeutic use of antimicrobial peptides and their design by bioinformatics tools. *Pathogens and Dis*ease 70, 257–270 (2014). URL https://academic.oup.com/femspd/ article-lookup/doi/10.1111/2049-632X.12151.
- [169] Peschel, A. et al. Inactivation of the dlt operon in Staphylococcus aureus confers sensitivity to defensins, protegrins, and other antimicrobial peptides. The Journal of biological chemistry 274, 8405–10 (1999). URL http://www.ncbi.nlm.nih.gov/pubmed/10085071.
- [170] Fabretti, F. et al. Alanine Esters of Enterococcal Lipoteichoic Acid Play a Role in Biofilm Formation and Resistance to Antimicrobial Peptides. Infection and Immunity 74, 4164–4171 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/16790791http:

- //www.pubmedcentral.nih.gov/articlerender.fcgi?artid=
 PMC1489678http://iai.asm.org/cgi/doi/10.1128/IAI.00111-06.
- [171] Saar-Dover, R. et al. D-Alanylation of Lipoteichoic Acids Confers Resistance to Cationic Peptides in Group B Streptococcus by Increasing the Cell Wall Density. PLoS Pathogens 8, e1002891 (2012). URL http://www.ncbi.nlm.nih.gov/pubmed/22969424http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC3435245http://dx.plos.org/10.1371/journal.ppat.1002891.
- [172] Moskowitz, S. M., Ernst, R. K. & Miller, S. I. PmrAB, a two-component regulatory system of Pseudomonas aeruginosa that modulates resistance to cationic antimicrobial peptides and addition of aminoarabinose to lipid A. *Journal of bacteriology* **186**, 575–9 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/14702327http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC305751.
- [173] Gunn, J. S. et al. PmrA-PmrB-regulated genes necessary for 4-aminoarabinose lipid A modification and polymyxin resistance. Molecular microbiology 27, 1171–82 (1998). URL http://www.ncbi.nlm.nih.gov/pubmed/9570402.
- [174] Wang, X., McGrath, S. C., Cotter, R. J. & Raetz, C. R. H. Expression cloning and periplasmic orientation of the Francisella novicida lipid A 4'-phosphatase LpxF. *The Journal of biological chemistry* **281**, 9321-30 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/16467300http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2758525.
- [175] Bugg, T. D. H. et al. Molecular basis for vancomycin resistance in Enterococcus faecium BM4147: biosynthesis of a depsipeptide peptidoglycan precursor by vancomycin resistance proteins VanH and VanA. Biochemistry 30, 10408–10415 (1991). URL http://pubs.acs.org/doi/abs/10.1021/bi00107a007.
- [176] Brötz, H. et al. Role of lipid-bound peptidoglycan precursors in the formation of pores by nisin, epidermin and other lantibiotics. *Molecular microbiology* 30, 317–27 (1998). URL http://www.ncbi.nlm.nih.gov/pubmed/9791177.

[177] Guo, L. et al. Lipid A acylation and bacterial resistance against vertebrate antimicrobial peptides. Cell 95, 189–98 (1998). URL http://www.ncbi.nlm.nih.gov/pubmed/9790526.

- R. E. et al. Transfer of palmitate from phospho-[178] Bishop, outer membranes of gram-negative baclipids to lipid A in TheEMBOjournal**19**, 5071 - 80(2000).URL teria. http://emboj.embopress.org/cgi/doi/10.1093/emboj/19.19. 5071http://www.ncbi.nlm.nih.gov/pubmed/11013210http://www. pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC302101.
- [179] Silhavy, T. J., Kahne, D. & Walker, S. The Bacterial Cell Envelope. Cold Spring Harbor Perspectives in Biology 2, a000414-a000414 (2010). URL http://www.ncbi.nlm.nih.gov/pubmed/20452953http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC2857177http://cshperspectives.cshlp.org/lookup/doi/10.1101/cshperspect.a000414.
- [180] Loutet, S. A., Flannagan, R. S., Kooi, C., Sokol, P. A. & Valvano, M. A. A complete lipopolysaccharide inner core oligosaccharide is required for resistance of Burkholderia cenocepacia to antimicrobial peptides and bacterial survival in vivo. *Journal of bacteriology* 188, 2073–80 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/16513737http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC1428139.
- [181] Allen, C. A., Adams, L. G. & Ficht, T. A. Transposon-derived Brucella abortus rough mutants are attenuated and exhibit reduced intracellular survival. *Infection and immunity* 66, 1008-16 (1998). URL http://www.ncbi.nlm.nih.gov/pubmed/9488389http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC108009.
- [182] Peschel, A. et al. <i>Staphylococcus aureus</i> Resistance to Human Defensins and Evasion of Neutrophil Killing via the Novel Virulence Factor Mprf Is Based on Modification of Membrane Lipids with l-Lysine. The Journal of Experimental Medicine 193, 1067–1076 (2001). URL http://www.ncbi.nlm.nih.gov/pubmed/11342591http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=

PMC2193429http://www.jem.org/lookup/doi/10.1084/jem.193.9.1067.

- [183] Thedieck, K. et al. The MprF protein is required for lysiny-lation of phospholipids in listerial membranes and confers resistance to cationic antimicrobial peptides (CAMPs) on Listeria monocytogenes. Molecular Microbiology 62, 1325–1339 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/17042784http://doi.wiley.com/10.1111/j.1365-2958.2006.05452.x.
- [184] Klein, S. et al. Adaptation of <i>Pseudomonas aeruginosa</i>to various conditions includes tRNA-dependent formation of alanyl-phosphatidylglycerol. *Molecular Microbiology* **71**, 551–565 (2009). URL http://doi.wiley.com/10.1111/j.1365-2958.2008.06562.x.
- [185] Band, V. & Weiss, D. Mechanisms of Antimicrobial Peptide Resistance in Gram-Negative Bacteria. Antibiotics 4, 18-41 (2014).

 URL http://www.ncbi.nlm.nih.gov/pubmed/25927010http:
 //www.pubmedcentral.nih.gov/articlerender.fcgi?artid=
 PMC4410734http://www.mdpi.com/2079-6382/4/1/18.
- [186] Kumariya, R., Sood, S. K., Rajput, Y. S., Saini, N. & Garsa, A. K. Increased membrane surface positive charge and altered membrane fluidity leads to cationic antimicrobial peptide resistance in Enterococcus faecalis. *Biochimica et biophysica acta* 1848, 1367–75 (2015). URL http://www.ncbi.nlm.nih.gov/pubmed/25782727.
- [187] Raetz, C. R., Reynolds, C. M., Trent, M. S. & Bishop, R. E. Lipid A Modification Systems in Gram-Negative Bacteria. Annual Review of Biochemistry 76, 295-329 (2007). URL http://www.ncbi.nlm.nih.gov/pubmed/17362200http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC2569861http://www.annualreviews.org/doi/10.1146/annurev.biochem.76.010307.145803.
- [188] Fjell, C. D., Hiss, J. A., Hancock, R. E. W. & Schneider, G. Designing antimicrobial peptides: form follows function. *Nature Reviews Drug Discovery* 11, 37 (2011). URL http://www.nature.com/doifinder/10.1038/nrd3591.

[189] Wiradharma, N. et al. Synthetic cationic amphiphilic α-helical peptides as antimicrobial agents. Biomaterials 32, 2204–2212 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/21168911https://linkinghub.elsevier.com/retrieve/pii/S0142961210015097.

- [190] Huang, Y., Huang, J. & Chen, Y. Alpha-helical cationic antimicrobial peptides: relationships of structure and function. *Protein & Cell* 1, 143-152 (2010). URL http://www.ncbi.nlm.nih.gov/pubmed/21203984http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4875170http://link.springer.com/10.1007/s13238-010-0004-3.
- [191] Pag, U. et al. Analysis of in vitro activities and modes of action of synthetic antimicrobial peptides derived from an α-helical sequence template'. Journal of Antimicrobial Chemotherapy 61, 341-352 (2008). URL https://academic.oup.com/jac/article-lookup/doi/10.1093/jac/dkm479.
- [192] Wang, J. et al. High specific selectivity and Membrane-Active Mechanism of the synthetic centrosymmetric α-helical peptides with Gly-Gly pairs. Scientific reports 5, 15963 (2015). URL http://www.nature.com/articles/srep15963http://www.ncbi.nlm.nih.gov/pubmed/26530005http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4632126.
- [193] Hilpert, K., Volkmer-Engert, R., Walter, T. & Hancock, R. E. W. High-throughput generation of small antibacterial peptides with improved activity. *Nature Biotechnology* **23**, 1008-1012 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/16041366http://www.nature.com/articles/nbt1113.
- [194] Hilpert, K. et al. Sequence Requirements and an Optimization Strategy for Short Antimicrobial Peptides. Chemistry & Biology 13, 1101–1107 (2006). URL http://www.ncbi.nlm.nih.gov/pubmed/17052614https://linkinghub.elsevier.com/retrieve/pii/S107455210600336X.

[195] Migoń, D. et al. Alanine Scanning Studies of the Antimicrobial Peptide Aurein 1.2. Probiotics and Antimicrobial Proteins 1–13 (2018). URL http://link.springer.com/10.1007/s12602-018-9501-0.

- [196] Grieco, P. et al. Alanine scanning analysis and structure-function relationships of the frog-skin antimicrobial peptide temporin-1Ta. Journal of Peptide Science 17, 358-365 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/21337476http://doi.wiley.com/10.1002/psc.1350.
- [197] Xie, J. et al. Potent effects of amino acid scanned antimicrobial peptide Feleucin-K3 analogs against both multidrug-resistant strains and biofilms of Pseudomonas aeruginosa. Amino Acids 50, 1471–1483 (2018). URL http://www.ncbi.nlm.nih.gov/pubmed/30136030http://link.springer.com/10.1007/s00726-018-2625-4.
- [198] Radzishevsky, I. S. et al. Effects of acyl versus aminoacyl conjugation on the properties of antimicrobial peptides. Antimicrobial agents and chemotherapy 49, 2412-20 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/15917541http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC1140510.
- [199] Serrano, G. N., Zhanel, G. G. & Schweizer, F. Antibacterial Activity of Ultrashort Cationic Lipo-Peptides. Antimicrobial Agents and Chemotherapy 53, 2215-2217 (2009).

 URL http://www.ncbi.nlm.nih.gov/pubmed/19237652http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=

 PMC2681537http://aac.asm.org/cgi/doi/10.1128/AAC.01100-08.
- [200] Avrahami, D. & Shai, Y. A New Group of Antifungal and Antibacterial Lipopeptides Derived from Non-membrane Active Peptides Conjugated to Palmitic Acid. *Journal of Biological Chemistry* **279**, 12277–12285 (2004). URL http://www.ncbi.nlm.nih.gov/pubmed/14709550http://www.jbc.org/lookup/doi/10.1074/jbc.M312260200.
- [201] Liu, S., Bao, J., Lao, X. & Zheng, H. Novel 3D Structure Based Model for Activity Prediction and Design of Antimicrobial Peptides. Scientific Reports 8, 11189 (2018). URL http://www.nature.com/articles/ s41598-018-29566-5.

[202] Jiang, Z., Vasil, A. I., Gera, L., Vasil, M. L. & Hodges, R. S. Rational Design of α-Helical Antimicrobial Peptides to Target Gramnegative Pathogens, Acinetobacter baumannii and Pseudomonas aeruginosa: Utilization of Charge, Specificity Determinants,' Total Hydrophobicity, Hydrophobe Type and Location as Design Para. Chemical Biology & Drug Design 77, 225–240 (2011). URL http://www.ncbi.nlm.nih.gov/pubmed/21219588http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid= PMC3063396http://doi.wiley.com/10.1111/j.1747-0285.2011. 01086.x.

- [203] Deslouches, B. et al. De Novo Generation of Cationic Antimicrobial Peptides: Influence of Length and Tryptophan Substitution on Antimicrobial Activity. Antimicrobial Agents and Chemotherapy 49, 316-322 (2005). URL http://www.ncbi.nlm.nih.gov/pubmed/15616311http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC538858http://aac.asm.org/cgi/doi/10.1128/AAC.49.1.316-322.2005.
- [204] Schmidt, N., Mishra, A., Lai, G. H. & Wong, G. C. Arginine-rich cell-penetrating peptides. FEBS Letters 584, 1806–1813 (2010). URL http://doi.wiley.com/10.1016/j.febslet.2009.11.046.
- [205] Farrotti, A. et al. Molecular Dynamics Simulations of the Host Defense Peptide Temporin L and Its Q3K Derivative: An Atomic Level View from Aggregation in Water to Bilayer Perturbation. *Molecules* 22, 1235 (2017). URL http://www.mdpi.com/1420-3049/22/7/1235.
- [206] Lee, E. Y., Lee, M. W., Fulan, B. M., Ferguson, A. L. & Wong, G. C. L. What can machine learning do for antimicrobial peptides, and what can antimicrobial peptides do for machine learning? *Interface Focus* 7, 20160153 (2017). URL http://rsfs.royalsocietypublishing.org/lookup/doi/10.1098/rsfs.2016.0153.
- [207] Ingólfsson, H. I. et al. Lipid Organization of the Plasma Membrane. Journal of the American Chemical Society 136, 14554-14559 (2014). URL http://pubs.acs.org/doi/10.1021/ja507832e.