## Statistical and mathematical modelling for seroepidemiological data of tropical infectious diseases

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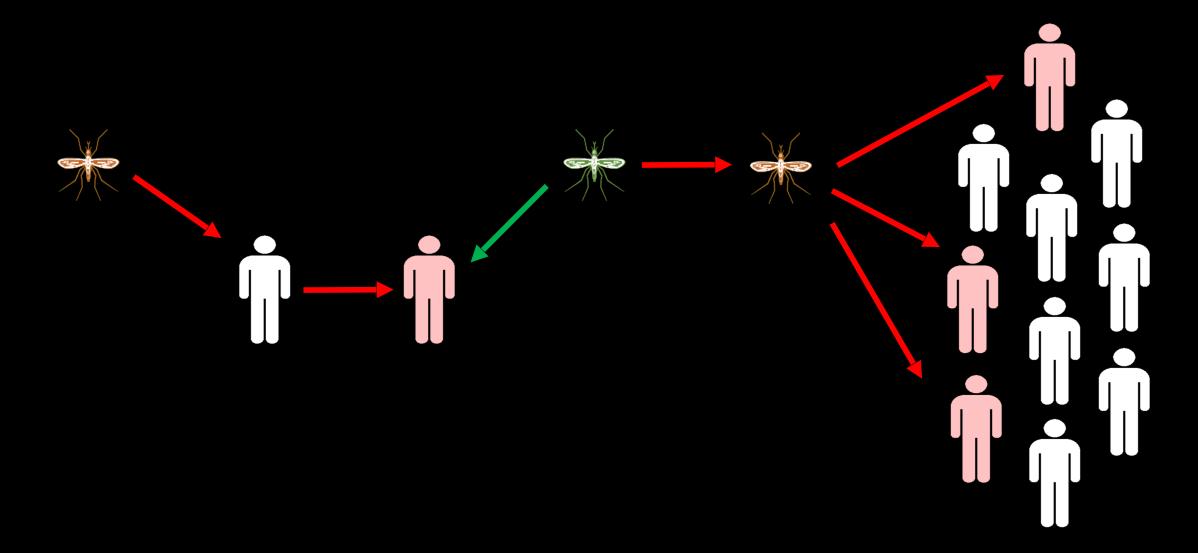
## Workshop content

- 1. Defining seropositivity (using two-Gaussian mixture models) and estimating seroprevalence
- 2. Estimating seroconversion rate (using reversible catalytic models)
- 3. Calculating sample size for controlling precision of seroconversion rate

#### GitHub:

https://github.com/immune-stats/Workshop\_Malaria\_PALOP\_2025

## How to measure malaria transmission?



### How to measure malaria transmission?

- 1. Prevalence of infection or parasite rate (non-informative when disease transmission intensity is low)
- 2. Entomological inoculation rate (trick to estimate)
- 3. Seroprevalence (prevalence of exposure)
- 4. Seroconversion rate (proxy of transmission intensity)

## Seroepidemiology

## **Epidemiology**

Seroepidemiology

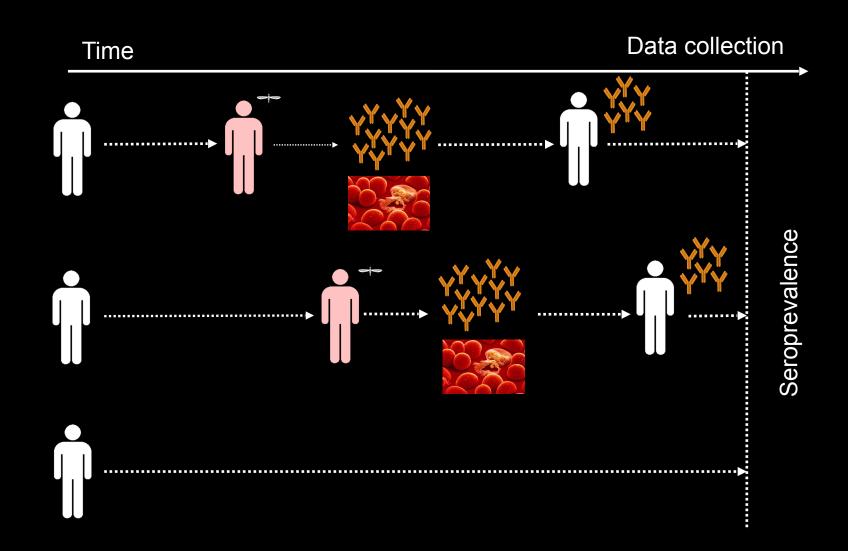
Immunology

**Statistics** 

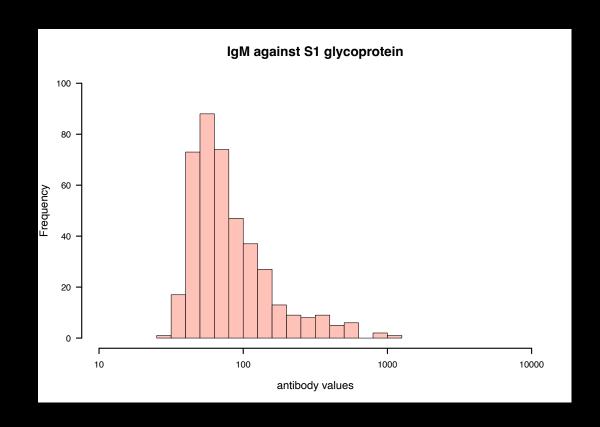
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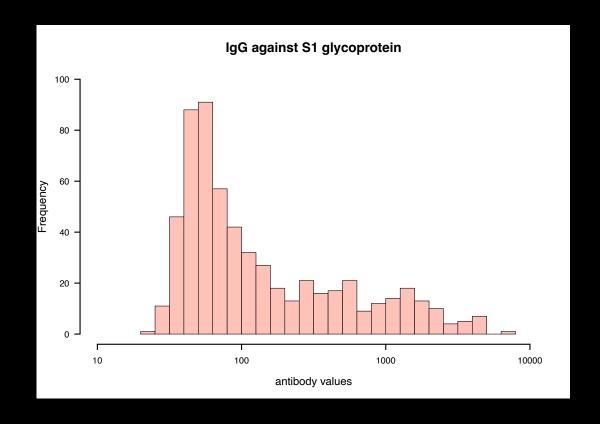
## Defining seropositivity (using two-Gaussian mixture models) and estimating seroprevalence

## Basic principle



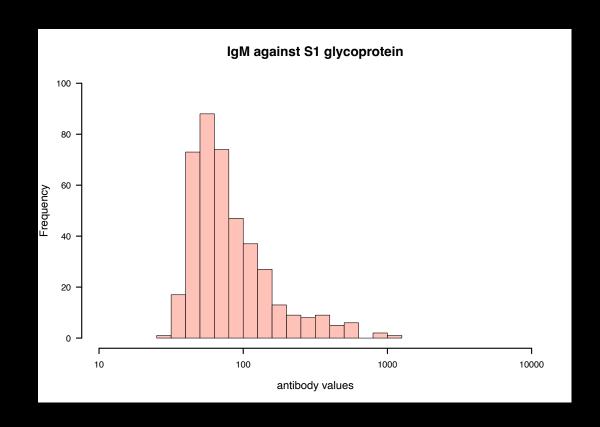
### Antibody data are intrinsically quantitative

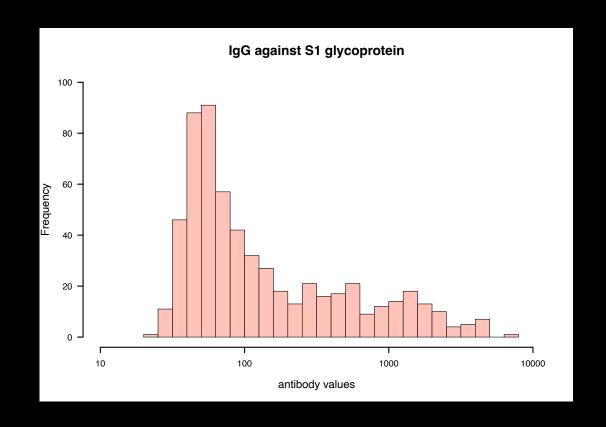




Rosado et al (2020). Serological signatures of SARS-CoV-2 infection: Implications for antibody-based diagnostics. medRxiv 2020.05.07.20093963.

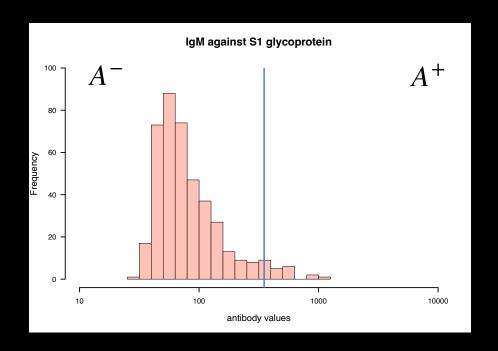
### Who are the seropositive individuals?

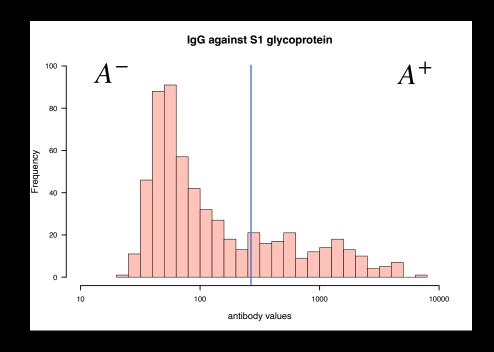


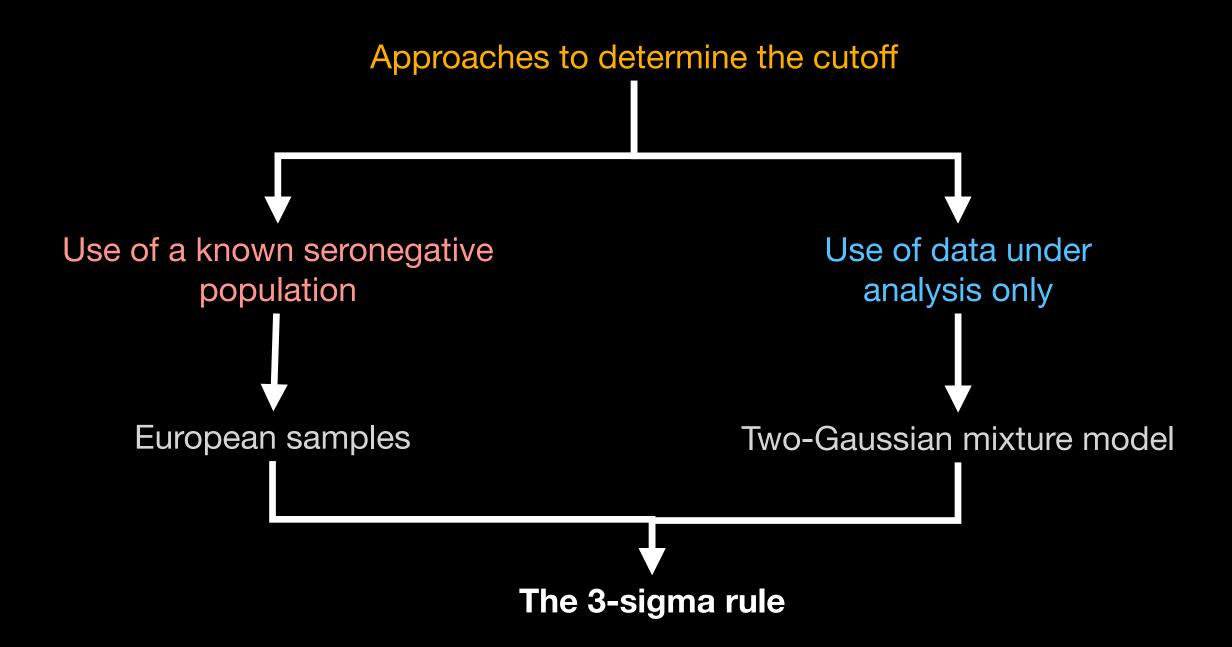


Rosado et al (2020). Serological signatures of SARS-CoV-2 infection: Implications for antibody-based diagnostics. medRxiv 2020.05.07.20093963.

### How to determine the cut-off?







## Approaches to determine the cutoff Use of data under Use of a known seronegative population analysis only European samples Two-Gaussian mixture model The 3-sigma rule

### The 3-sigma rule

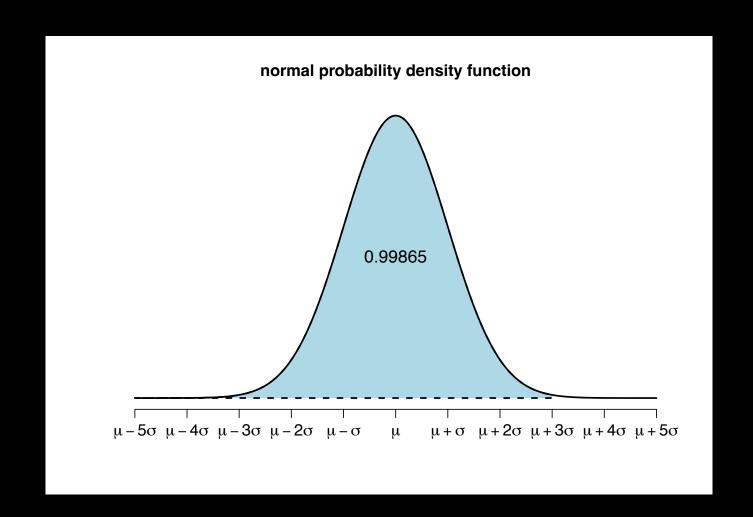
$$\mu_{A^{-}} = E\left[X \mid A^{-}\right]$$

$$\sigma_{A^{-}} = \sqrt{Var\left[X|A^{-}\right]}$$

Seronegative, if  $X_i \leq \mu_{A^-} + 3\sigma_{A^-}$ 

Seropositive, otherwise

### The link to the Normal distribution



## Approaches to determine the cutoff Use of data under Use of a known seronegative population analysis only Two-Gaussian mixture model Pre-pandemic samples The 3-sigma rule

## Two-Gaussian mixture models

$$f_X(x) = (1 - \pi) f_{N(\mu_{S^-}, \sigma_{S^-})}(x) + \pi f_{N(\mu_{S^+}, \sigma_{S^+})}(x)$$

Definition of 
$$S^- \Rightarrow \mu_{S^-} < \mu_{S^-}$$

In general:

$$f_X(x) = \sum_{i=1}^k \pi_i f_{N(\mu_i, \sigma_i)}(x)$$
 where  $\sum_{i=1}^k \pi_i = 1$ 

## Estimation of the model by maximum likelihood method

EM (Expectation-Maximization) Algorithm

- 1. Start with initial estimates for the parameters
- 2.E-Step calculate the probability of each individual belonging to a given subpopulation according to estimates at 1.
- 3.M-Step re-estimate the parameters using these probabilities and repeat the E-step with these new estimates
- 4. Stop with the increment in the log-likelihood is below a given tolerance error.

Calculate the cutoff for seropositivity according to  $\hat{\mu}_{S^-}$  and  $\hat{\sigma}_{S^-}$ 

Package mixtools



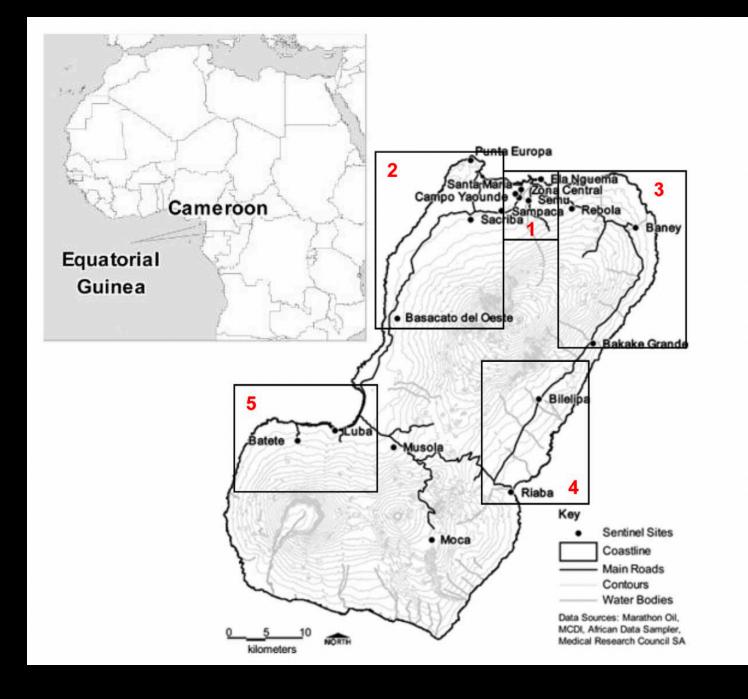
## time!



## Serological Markers Suggest Heterogeneity of Effectiveness of Malaria Control Interventions on Bioko Island, Equatorial Guinea

Jackie Cook<sup>1</sup>, Immo Kleinschmidt<sup>2</sup>, Christopher Schwabe<sup>3</sup>, Gloria Nseng<sup>4</sup>, Teun Bousema<sup>1</sup>, Patrick H. Corran<sup>1</sup>, Eleanor M. Riley<sup>1</sup>, Chris J. Drakeley<sup>1</sup>\*

1 Department of Immunology and Infection, London School of Hygiene and Tropical Medicine, London, United Kingdom, 2 Department of Infectious Disease Epidemiology, London School of Hygiene and Tropical Medicine, London, United Kingdom, 3 Medicinal Care Development International, Silver Spring, Maryland, United States of America, 4 Ministry of Health and Social Welfare, Malabo, Equatorial Guinea



**Table 1.** Demographic characteristics of the study population.

		% [n]						
		Malabo N = 2328	North West N=1749	North East N = 1323	South East N = 700	South West N = 588	Other** N = 699	Total N = 7387
Age (years)	0–1	14.1 [324]	10.5 [182]	10.4 [137]	12.2 [85]	10.0 [58]	7.5 [52]	11.4 [838]
	1–5	21.1 [458]	18.0 [312]	19.8 [261]	14.6 [102]	16.8 [97]	15.2 [106]	18.6 [1363]
	5–15	26.3 [605]	30.6 [531]	30.1 [396]	21.0 [146]	24.0 [139]	28.7 [200]	27.5 [2017]
	15–90	38.6 [890]	41.0 [712]	39.8 [524]	52.2 [364]	49.2 [285]	48.6 [338]	42.5 [3113]
Sex	Female	61.2 [1410]	54.2 [932]	61.1 [805]	55.8 [389]	58.4 [338]	54.8 [382]	58.2 [4256]
House recently sprayed <sup>1</sup>	Yes	74.2 [1580]	81.2 [1306]	85.6 [1076]	81.7 [519]	89.5 [477]	87.9 [574]	81.2 [5532]
Slept under ITN <sup>2</sup>	Yes	82.6 [1629]	68.0 [988]	65.8 [797]	63.3 [404]	73.1 [385]	71.4 [449]	72.4 [4652]
Parasite positive	Yes	14.8 [300]	27.0 [374]	7.9 [94]	21.7 [135]	18.6 [97]	12.1 [75]	16.9 [1075]

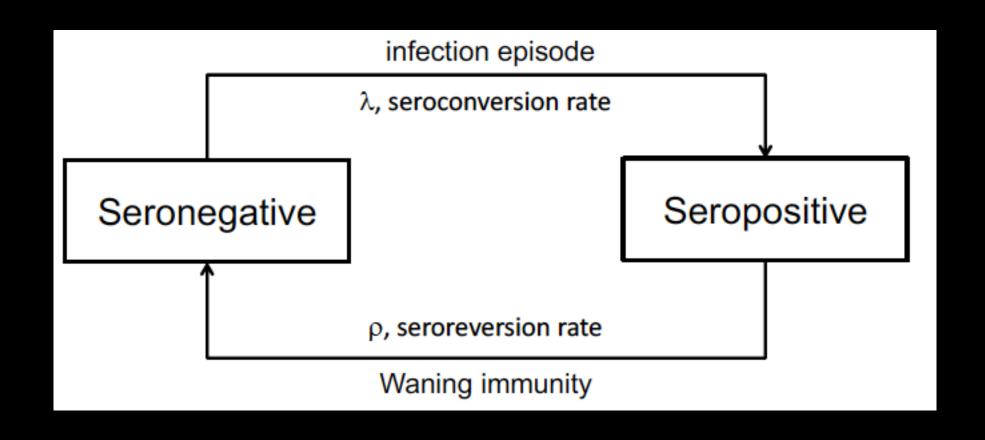
<sup>&</sup>lt;sup>1</sup>- within the previous 6 months. <sup>2</sup>- on the night before the survey.

<sup>\*\*</sup>Moca and Musola kept separate due to their high altitude. doi:10.1371/journal.pone.0025137.t001

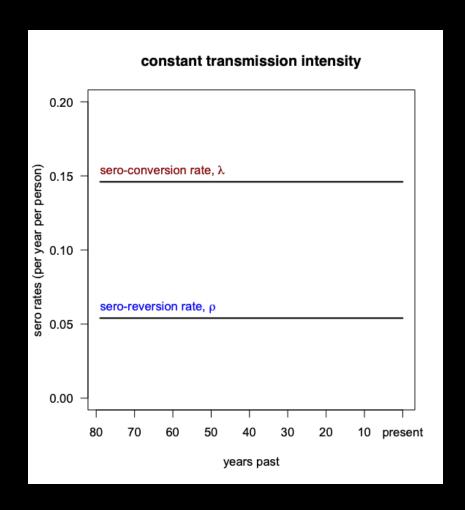
#### Variables

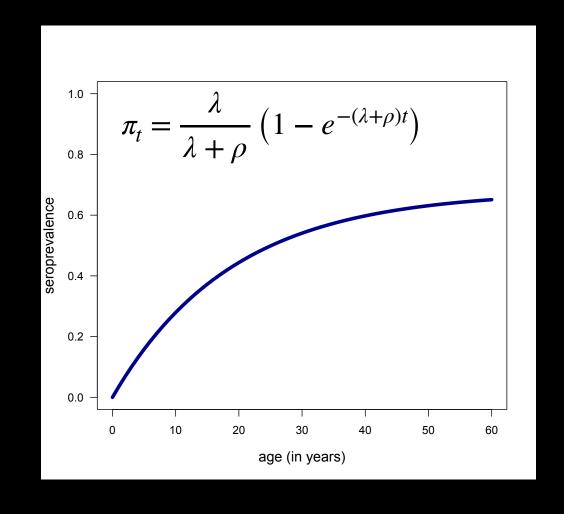
# 2. Estimating seroconversion rate (using reversible catalytic models)

## Reversible catalytic models

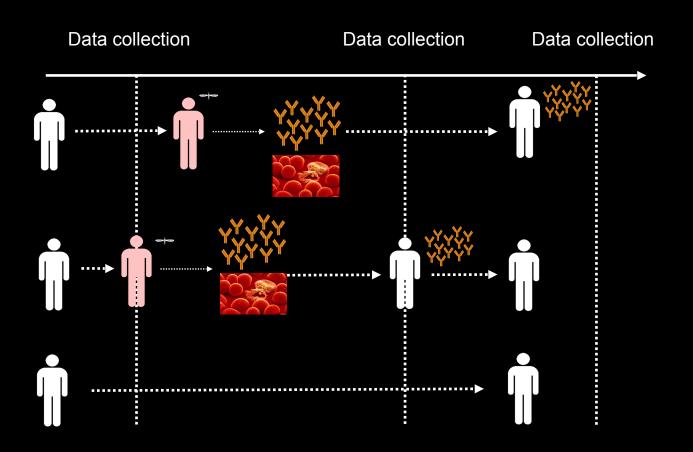


## Constant transmission intensity





## Longitudinal surveys



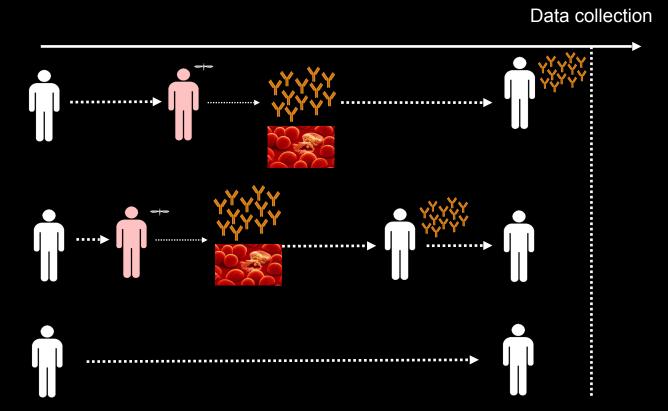
#### Statistical information: ++++

Direct observation of serological transitions

#### Execution difficulty: ++++

Time consuming
Sampling intensive
Participation adherence/drop-outs

## Cross-sectional surveys



#### Statistical information: ++

No direct observation of serological transitions Age as proxy of time

#### Execution difficulty: ++

Easy to engage participation Quick sampling

## Longitudinal versus cross-sectional surveys

Type of Study	Seroconversion rate	Seroreversion rate
Longitudinal	<b>0.021</b> (0.001-0096)	<b>0.163</b> (0.001,0.729)
Cross-sectional	<b>0.023</b> (0.001,0.052)	<b>0.0001</b> (0.001,0.255)

Seroreversion rate is difficult to be estimated! So it is often fixed at 0 or an good estimate, say 0.017.



#### Fixed seroreversion rate at 0

$$\rho = 0 \Rightarrow \pi_t = 1 - e^{-\lambda t}$$

$$\Rightarrow \log 1 - \pi_t = -\lambda t$$

$$\Rightarrow \log(-\log(1-\pi_t)) = \log \lambda + \log t$$

Complementary log-log model

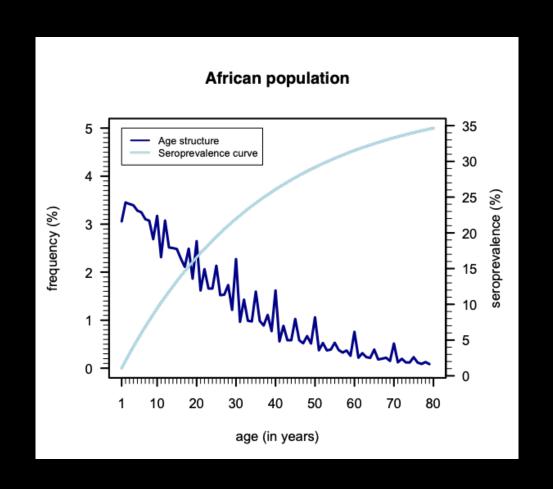


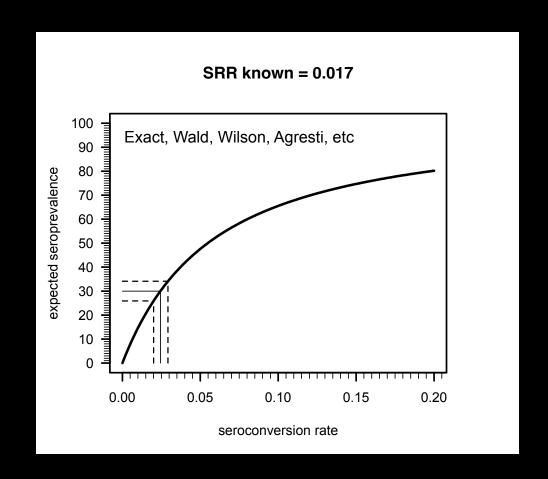
## time!

3.

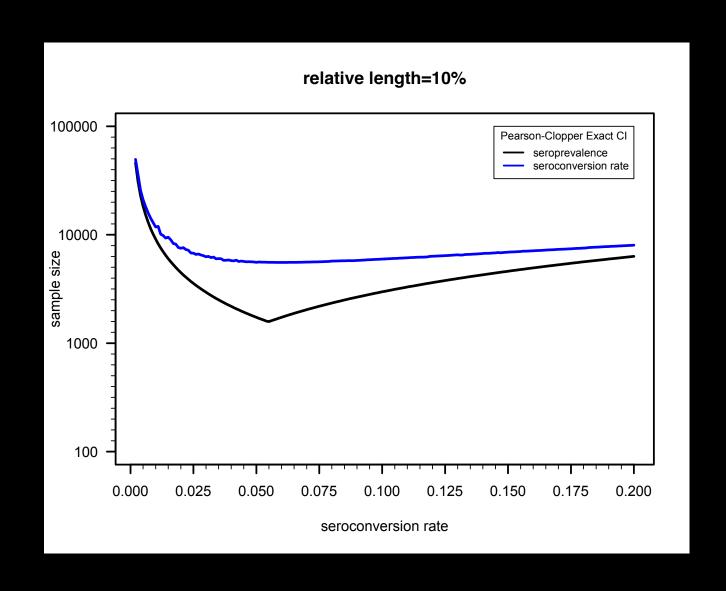
## Calculating sample size for controlling precision of seroconversion rate estimate

## Sample size calculation for seroconversion rate





## Sample size calculation for seroconversion rate



## Sample size calculation in practice

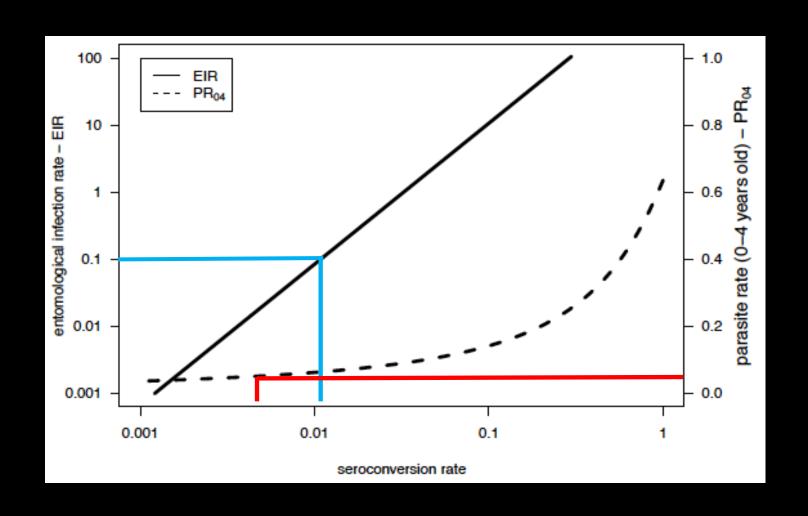
- 1. Desired precision
- 2. Antibody with known seroreversion rate
- 3. Transmission intensity of the population
- 4. Age structure associated with sampling scheme
- 5. Type of confidence interval to be used

## Identification of transmission intensity

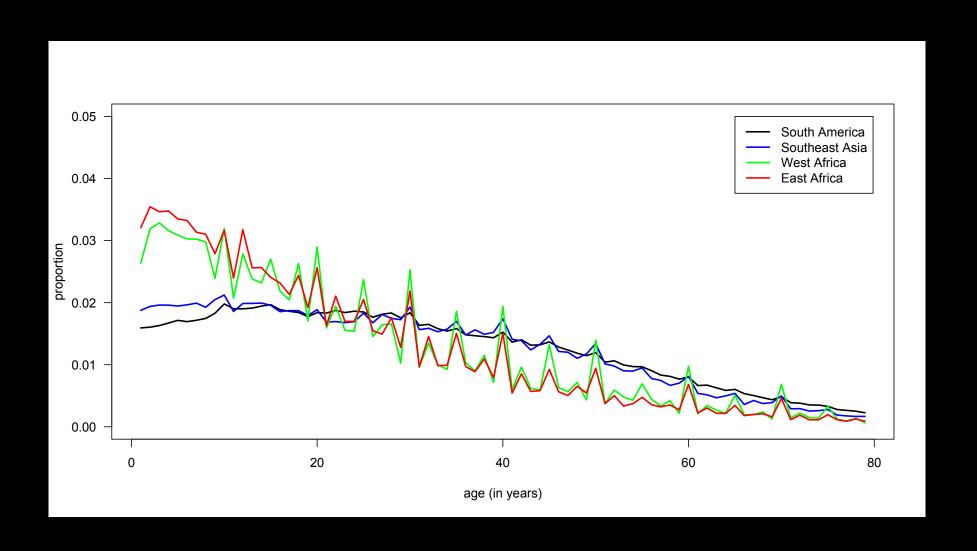
Table 1 Expected relationship between EIR, PR<sub>04</sub>, SCR and SP in African (AFR), Southeast Asian and South American (SEA + SA) populations where seroreversion rate was fixed at 0.017

			Seroprevalence		
EIR	PR <sub>04</sub>	SCR	AFR	SEA + SA	
0.01	0.050	0.0036	0.057	0.073	
0.10	0.073	0.0108	0.156	0.195	
1.00	0.119	0.0324	0.365	0.437	
10.0	0.231	0.0969	0.647	0.720	
100.0	0.625	0.2900	0.860	0.896	
			3.300		

## Identification of transmission intensity



## Identification of age structure



## Type of confidence interval

#### **Pearson-Clopper exact**

Coverage higher than nominal confidence level

#### Wald

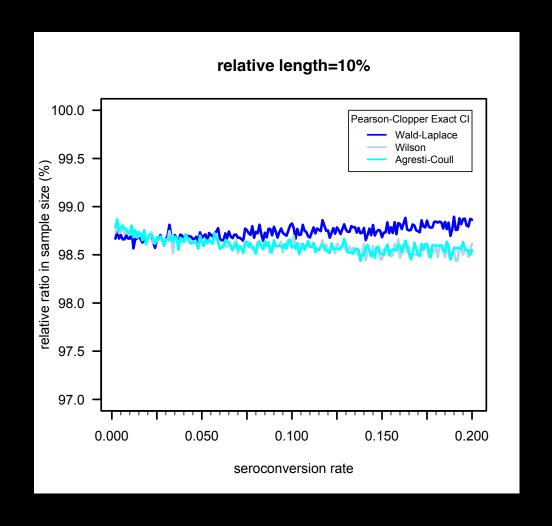
Degenerate when x=0 or x=n Overshooting

#### Wilson

Good coverage in extreme probabilities

#### **Agresti-Coull**

Overshooting
Better coverage than the exact CI





## time!

#### References

Cook J, Kleinschmidt I, Schwabe C, et al. Serological markers suggest heterogeneity of effectiveness of malaria control interventions on Bioko Island, equatorial Guinea. PLoS One. 2011;6(9):e25137. doi:10.1371/journal.pone.0025137

Sepúlveda N, Paulino CD, Drakeley C. Sample size and power calculations for detecting changes in malaria transmission using antibody seroconversion rate. Malar J. 2015;14:529. Published 2015 Dec 30. doi:10.1186/s12936-015-1050-3

Sepúlveda N, Stresman G, White MT, Drakeley CJ. Current Mathematical Models for Analyzing Anti-Malarial Antibody Data with an Eye to Malaria Elimination and Eradication. J Immunol Res. 2015;2015:738030. doi:10.1155/2015/738030