

# SNPs

## inputs

```
In [1]: source('jupyterFunctions_broadCellType.R')
        source('jupyterFunctions_perCellType.R')
```

```
In [2]: CT <- 'SNPs'
        data_prefix <- paste(sep='', '../data/', CT, '/')
        ATAC_pxCT_norm <- readRDS(paste(sep='', data_prefix, 'ATAC_pxc_afc_SNPpeak
s_pxCT_norm.rds')) #should get all peaks here!
        chosenSNPs <- readRDS(paste(sep='', data_prefix, 'ATAC_chosenSNPs.rds'))
```

```
In [3]: ATAC_colors <- readRDS('../data/misc/ATAC_class_colors.rds')
```

```
In [15]: save_dir <- NA #'../output/' #or NA if don't want to save
```

## SNP matrix

```
In [5]: class_order <- c('TA-0', 'TA-4', 'TA-1', 'TA-2', 'TA-3',
                        'BA-3', 'BA-4', 'BA-2', 'BA-5', 'BA-0', 'BA-1',
                        'MA-0', 'MA-2', 'MA-4', 'MA-1', 'MA-3',
                        'EA-2', 'EA-3', 'EA-0', 'EA-1',
                        'SA-1', 'SA-2', 'SA-0', 'SA-3')
        if(!all(class_order %in% colnames(ATAC_pxCT_norm))) stop('not all classes in pxCT')
        if(!all(class_order %in% names(ATAC_colors))) stop('not all classes in colors')

        if(!all(unname(chosenSNPs) %in% rownames(ATAC_pxCT_norm))) stop('not all chosen SNPs in pxCT')
```

In [6]: #Fig 7d

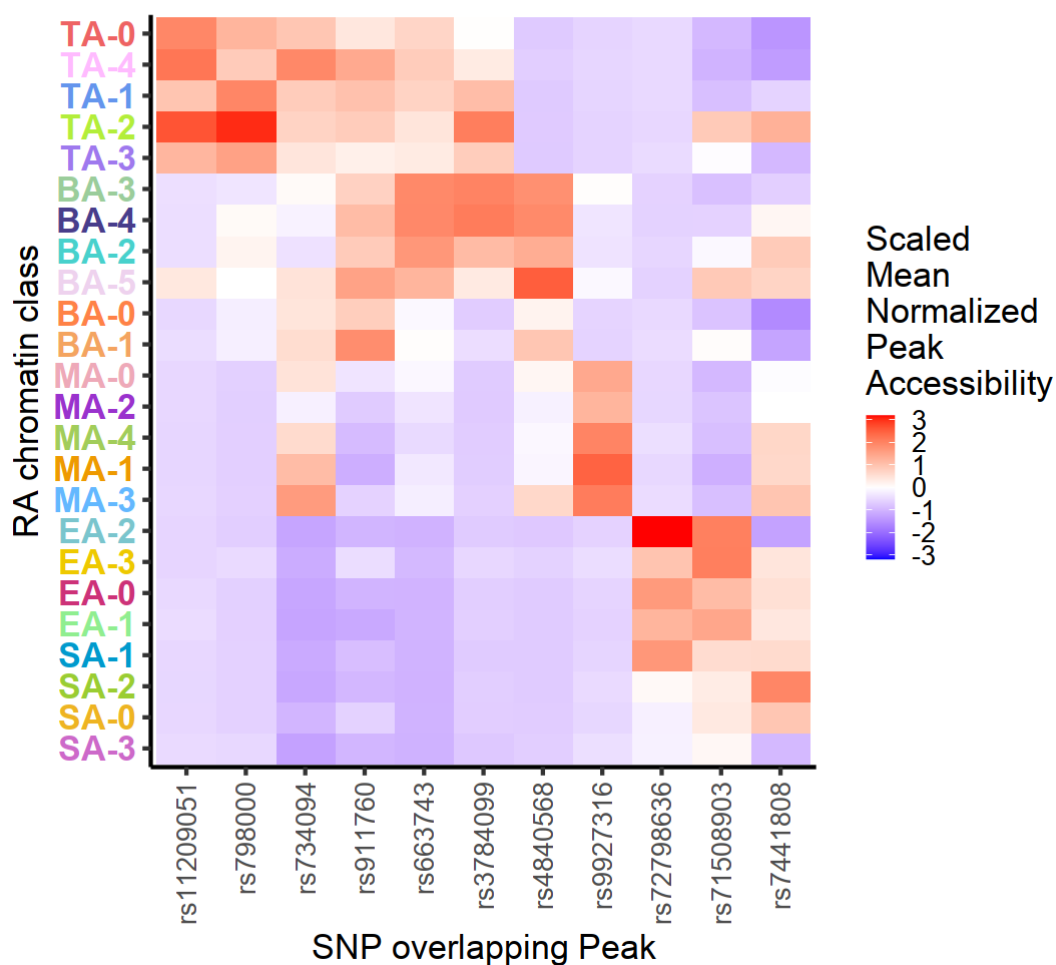
```

ATAC_pxCT_norm_subset_scaled <- scalePeak_forHeatmap(names(chosenSNPs),c
lass_order,chosenSNPs,ATAC_pxCT_norm)

options(repr.plot.height=9,repr.plot.width=9)
g <- pseudobulk_scaled_heatmap(ATAC_pxCT_norm_subset_scaled,'SNP overlap
ping Peak','RA chromatin class',
                                'Scaled\nMean\nNormalized\nPeak\nAccessib
ility',
                                plotTit='Scaled Mean Normalized Peak Acce
ssibility\nof ATAC cells by RA chromatin class',
                                scale_lim=max(abs(ATAC_pxCT_norm_subset_s
caled)),clustColors=ATAC_colors)
print(g)
if(!is.na(save_dir)) ggsave(file=paste(sep='',save_dir,'ATAC_SNP_markerP
eak_heatmap.png'),
                             plot=g,units='in',height=9,width=9,dpi=600)

```

Scaled Mean Normalized Peak Accessibility  
of ATAC cells by RA chromatin class



```
In [7]: class_overlaps <- as.data.frame(rev(apply(ATAC_pxCT_norm_subset_scaled,
1,function(x){paste(names(x[x>1])),collapse=', ')})),
stringsAsFactors=FALSE)
colnames(class_overlaps) <- c('classOL')
class_overlaps
```

A data.frame: 11 × 1

	classOL
	<chr>
rs11209051	TA-3, TA-2, TA-4, TA-0
rs798000	TA-3, TA-2, TA-1, TA-0
rs734094	MA-3, MA-1, TA-4
rs911760	BA-1, BA-5, BA-4, TA-1, TA-4
rs663743	BA-5, BA-2, BA-4, BA-3
rs3784099	BA-2, BA-4, BA-3, TA-2, TA-1
rs4840568	BA-5, BA-2, BA-4, BA-3
rs9927316	MA-3, MA-1, MA-4, MA-2, MA-0
rs72798636	SA-1, EA-1, EA-0, EA-2
rs71508903	EA-1, EA-0, EA-3, EA-2
rs7441808	SA-2, TA-2

## Wilcoxon tests

```
In [8]: classCT_conv <- c('stromal','endothelial','myeloid','Bplasma','Tcell')
names(classCT_conv) <- c('S','E','M','B','T')

snps_gathered <- gather_SNPs(ATAC_pxCT_norm_subset_scaled,chosenSNPs,classCT_conv)
```

```
In [9]: within <- snps_gathered[which(snps_gathered$rsID=='rs11209051' & snps_gathered$cellType=='Tcell'), 'norm_pxCTmean_scale']
without <- snps_gathered[which(snps_gathered$rsID=='rs11209051' & snps_gathered$cellType!='Tcell'), 'norm_pxCTmean_scale']

wilcox.test(within, without, alternative = "greater")
```

Warning message in wilcox.test.default(within, without, alternative = "greater"):

"cannot compute exact p-value with ties"

Wilcoxon rank sum test with continuity correction

data: within and without

W = 95, p-value = 0.0004165

alternative hypothesis: true location shift is greater than 0

```
In [10]: within <- snps_gathered[which(snps_gathered$rsID=='rs4840568' & snps_gathered$cellType=='Bplasma'), 'norm_pxCTmean_scale']
without <- snps_gathered[which(snps_gathered$rsID=='rs4840568' & snps_gathered$cellType!='Bplasma'), 'norm_pxCTmean_scale']

wilcox.test(within, without, alternative = "greater")
```

Wilcoxon rank sum test

data: within and without

W = 107, p-value = 1.486e-05

alternative hypothesis: true location shift is greater than 0

```
In [11]: within <- snps_gathered[which(snps_gathered$rsID=='rs798000' & snps_gathered$cellType=='Tcell'), 'norm_pxCTmean_scale']
without <- snps_gathered[which(snps_gathered$rsID=='rs798000' & snps_gathered$cellType!='Tcell'), 'norm_pxCTmean_scale']

wilcox.test(within, without, alternative = "greater")
```

Wilcoxon rank sum test

data: within and without

W = 95, p-value = 2.353e-05

alternative hypothesis: true location shift is greater than 0

```
In [12]: within <- snps_gathered[which(snps_gathered$rsID=='rs9927316' & snps_gathered$cellType=='myeloid'), 'norm_pxCTmean_scale']
without <- snps_gathered[which(snps_gathered$rsID=='rs9927316' & snps_gathered$cellType!='myeloid'), 'norm_pxCTmean_scale']

wilcox.test(within, without, alternative = "greater")
```

Warning message in wilcox.test.default(within, without, alternative = "greater"):

"cannot compute exact p-value with ties"

Wilcoxon rank sum test with continuity correction

data: within and without

W = 95, p-value = 0.0004165

alternative hypothesis: true location shift is greater than 0

```
In [13]: within <- snps_gathered[which(snps_gathered$rsID=='rs734094' & (snps_gathered$cellType %in% c('myeloid', 'Tcell'))),
                                     'norm_pxCTmean_scale']
without <- snps_gathered[which(snps_gathered$rsID=='rs734094' & !(snps_gathered$cellType %in% c('myeloid', 'Tcell'))),
                                     'norm_pxCTmean_scale']

wilcox.test(within, without, alternative = "greater")
```

Wilcoxon rank sum test

data: within and without

W = 131, p-value = 4.946e-05

alternative hypothesis: true location shift is greater than 0

## Session Info

In [14]: `sessionInfo()`

```
R version 3.6.1 (2019-07-05)
Platform: x86_64-conda_cos6-linux-gnu (64-bit)
Running under: Red Hat Enterprise Linux Server release 6.5 (Santiago)

Matrix products: default
BLAS/LAPACK: /PHShome/kew47/miniconda3/lib/R/lib/libRblas.so

locale:
[1] en_US.UTF-8

attached base packages:
[1] grid      stats      graphics  grDevices  utils      datasets  methods
[8] base

other attached packages:
[1] repr_1.0.1      gridExtra_2.3    scales_1.1.1     viridis_0.5.
1
[5] viridisLite_0.3.0 ggrepel_0.8.2     ggtrastr_0.2.3    stringr_1.4.
0
[9] ROCR_1.0-7      gplots_3.0.1.1    Rmisc_1.5.1      plyr_1.8.6
[13] lattice_0.20-41 gtools_3.8.2      tidyr_1.0.3      Matrix_1.2-1
8
[17] ggplot2_3.3.0

loaded via a namespace (and not attached):
[1] Rcpp_1.0.4.6      vipor_0.4.5       pillar_1.4.4
[4] compiler_3.6.1    bitops_1.0-6      base64enc_0.1-3
[7] tools_3.6.1       digest_0.6.25     uuid_0.1-2
[10] jsonlite_1.7.1    evaluate_0.14     lifecycle_0.2.0
[13] tibble_3.0.1      gtable_0.3.0      pkgconfig_2.0.3
[16] rlang_0.4.8       IRdisplay_0.7.0   IRkernel_1.0.2.9000
[19] beeswarm_0.2.3    withr_2.2.0       dplyr_1.0.2
[22] caTools_1.18.0    generics_0.0.2    vctrs_0.3.5
[25] tidyselect_1.1.0  glue_1.4.0        R6_2.4.1
[28] ggbeeswarm_0.6.0  gdata_2.18.0      pbdZMQ_0.3-3
[31] farver_2.0.3      purrr_0.3.4       magrittr_1.5
[34] ellipsis_0.3.1    htmltools_0.4.0   colorspace_1.4-1
[37] labeling_0.3      KernSmooth_2.23-15 stringi_1.4.6
[40] munsell_0.5.0     crayon_1.3.4      Cairo_1.5-10
```

In [ ]: