

Supplementary Information I : An application of CaN modelling to the Barents Sea trophic network

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Contents

1	Introduction	1
2	Installing RCaNmodel	2
3	The RCaNmodel file for the Barents Sea ecosystem	2
3.1	Observational data-series	3
3.2	Constraints	5
4	The polytope behind a CaNmod object	5
4.1	Structure of the polytope	6
4.2	Suitability of the polytope	6
4.3	Bounds of the polytope	8
4.4	2D-slices of the polytope	8
5	Sampling the polytope	12
5.1	Sampling	12
5.2	Convergence of the sample chains	13
6	Visualisation of the model results	16
6.1	Empirical distributions of biomass and fluxes	17
6.2	Diet composition	19
6.3	Growth and density-dependence	20
6.4	Trophic functional relationships	21
6.5	Satiation	22
6.6	Feeding and Growth	23
6.7	Pair-plots	24
6.8	Top-Down and Bottom-Up controls	25

1 Introduction

We present here an example of RCaNmodel study for the Barents Sea ecosystem. The CaN model has the same structure as the model presented in the main text of the article: same trophic structure, same trophic parameters, same set of time series of observations (Lindstrøm, Planque, and Subbey (2017)). Due to data access restriction, the values in the observation times series were slightly modified and do not correspond to the original measurements. Most of the original data can be provided on demand.

The network is illustrated in figure 1.

```
knitr::include_graphics("BarentsNetwork.png")
```

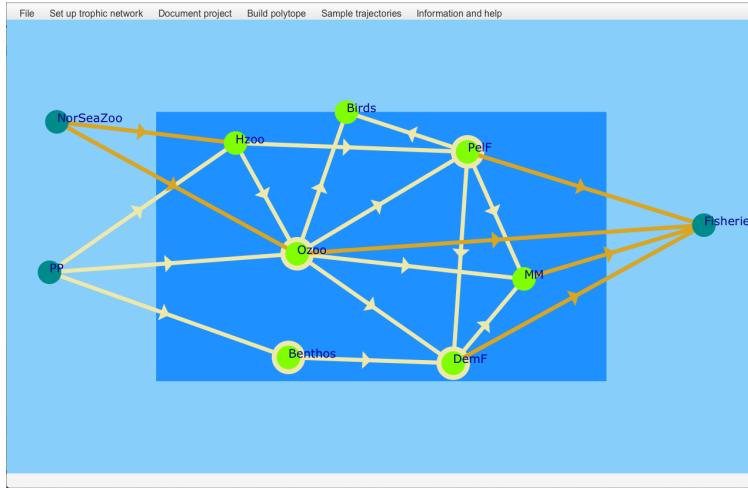


Figure 1: The Barents Sea tropic network. The dashed-line rectangle separate components that are within the model domain (herbivorous and omnivorous zooplankton, benthos, pelagic and demersal fish, mammals and birds) from those that are outside (primary producers, Norwegian Sea zooplankton and fisheries)

The RCaN-file, constructed with RCaNconstructor, is attached to this supplementary information. The R-Markdown file, used to generate this supplementary material, is also attached.

We first provide information for the installation of RCaNmodel and RCaNconstructor. We then provide a short description of the content of the Barents Sea RCaN-file. Finally, we go through the main steps of the modelling which follow the RCaN file construction: (a) build the polytope, (b) check the polytope status, (c) if required, revise the model by adding and removing constraints, (d) sample the polytope, (d) analyze the samples.

2 Installing RCaNmodel

RCaNmodel requires R (version 3.2+) to be installed.

The R library RCaNmodel can be downloaded from <https://github.com/inrae/RCaNmodel>. This is done by following installation instructions given in the ReadMe file of the package .

```
# if (!require("RCaNmodel")) {
#   require(devtools)
#   devtools::install_github(
#     "https://github.com/inrae/RCaNmodel.git",
#     subdir="RCaNmodel",
#     dependencies = TRUE)
# }
```

Then, the library can be loaded

```
library(RCaNmodel) #the RCaNmodel package itself
```

3 The RCaNmodel file for the Barents Sea ecosystem

The food web structure, biological parameters, observational data-series and model constraints are all provided in the RCaN file in the .xlsx format. A RCaN file can be edited using a spreadsheet editor (e.g. libre office,

Table 1: Components

Component	Inside	AssimilationE	Digestibility	OtherLosses	Inertia	Satiation	RefugeBiomass
PP	0	0.00	0.65	0.00	0.00	0.0	0.0000
Hzoo	1	1.00	0.90	8.40	7.58	128.0	0.2300
Ozoo	1	1.00	0.90	5.50	3.10	42.0	0.1300
Benthos	1	0.94	0.60	1.50	0.74	25.2	0.6600
PelF	1	0.90	0.90	2.85	0.90	13.5	0.0250
DemF	1	0.93	0.85	1.65	0.25	5.5	0.0230
MM	1	1.00	0.00	5.50	0.11	10.9	0.0034
Birds	1	0.84	0.00	60.00	0.81	123.0	0.0001
NorSeaZoo	0	0.00	0.00	0.00	0.00	0.0	0.0000
Fisheries	0	0.00	0.00	0.00	0.00	0.0	0.0000

open office, MS Excel) or with the RCaNconstructor. The use of RCaNconstructor is recommended as it ensures that names of model entities (species, fluxes and observations) are consistent across the different worksheets of the RCaN-file.

```
NAMEFILE <- 'annexC_BarentsSeaDummy.xlsx'
```

Importing an RCaN-file in the R environment and building the corresponding polytope are done with one single RCaNmodel command:

```
POLYTOPE <- buildCaN(NAMEFILE)
```

The buildCaN function reads four worksheets from the RCaN-file and uses the information to construct the model, i.e. build the corresponding polytope. The information in these four worksheets pertains to components, fluxes, observational data-series and constraints. These are as follows:

3.0.1 Components

Components correspond to the tropho-species in the systems (figure 1). They are specified in the “Components & input parameter” of the RCaN file, with their names (column Component), a flag to indicate whether they are inside or outside the model domain (column Inside) and their biological parameters. The biological parameters are used in the CaN master equation (which expresses mass conservation and the relationship between fluxes and biomass) and to implement implicit constraints (inertia, satiation and refuge biomass).

The above information is stored in the dataframe ‘components_param’ in the POLYTOPE object:

```
kable(POLYTOPE$components_param[c(1:8)], caption='Components') %>%
  kable_styling(latex_options = c("striped", "scale_down"), font_size = 8)
```

3.0.2 Fluxes

Fluxes correspond to the transfer of biomass between food-web components (i.e., arrows in figure 1). They are specified in the “Fluxes” worksheet of the RCaN-file, identified by a name (column Flux), a source component (column From) and a sink component (column To) and a flag that indicates whether the flux is trophic or not (column Trophic). Non-trophic fluxes are typically import/export to/from the model domain and fisheries. The fluxes information is stored in the dataframe ‘flux_def’ in the POLYTOPE object.

```
kable(POLYTOPE$fluxes_def, caption='Fluxes') %>%
  kable_styling(latex_options = c("striped", "scale_down"), font_size = 8)
```

3.1 Observational data-series

Observational time-series in the Barents Sea are derived from monitoring programs, mainly for plankton and fish species. In this example, the original data compiled by the ICES Working Group on the Integrated

Table 2: Fluxes

Flux	From	To	Trophic
PP_Ozoo	PP	Ozoo	1
PP_Hzoo	PP	Hzoo	1
PP_Benthos	PP	Benthos	1
Hzoo_Ozoo	Hzoo	Ozoo	1
Hzoo_PelF	Hzoo	PelF	1
Ozoo_Ozoo	Ozoo	Ozoo	1
Ozoo_PelF	Ozoo	PelF	1
Ozoo_DemF	Ozoo	DemF	1
Ozoo_MM	Ozoo	MM	1
Ozoo_Birds	Ozoo	Birds	1
Benthos_Benthos	Benthos	Benthos	1
Benthos_DemF	Benthos	DemF	1
PelF_PelF	PelF	PelF	1
PelF_DemF	PelF	DemF	1
PelF_MM	PelF	MM	1
PelF_Birds	PelF	Birds	1
DemF_DemF	DemF	DemF	1
DemF_MM	DemF	MM	1
NorSeaZoo_Hzoo	NorSeaZoo	Hzoo	0
NorSeaZoo_Ozoo	NorSeaZoo	Ozoo	0
PelF_Fisheries	PelF	Fisheries	0
DemF_Fisheries	DemF	Fisheries	0
MM_Fisheries	MM	Fisheries	0
Ozoo_Fisheries	Ozoo	Fisheries	0

Table 3: Observations

Table 4: Constraints defined by user

ID	Constraint	Time-range	Action	Comment
C01	PF_Huso >= PP_Oomo + PP_Benthos <= Prod_Sat * 1.5	1998-2018	1	Max primary production
C02	PF_Huso >= PP_Chees + PP_Benthos <= Prod_Sat * 1.5	1998-2018	1	Min primary production
C03	PP_Chees >= PP_Chees + PP_Benthos <= Prod_Sat * 1.5	1998-2018	1	Max primary production when no data is available
C04	PP_Huso >= PP_Oomo + PP_Benthos >= 5000000	1998-2018	1	Min primary production when no data is available
C05	NoRestrZone_Oomo >= 2 * Prod_Sat	1998-2018	1	From input of ecosystem production from the Norwegian Sea
C06	NoRestrZone_Fisheries <= Prod_Sat	1998-2018	1	From input of ecosystem production
C08	PPB_Fisheries <= Prod_Sat * 1.1	1998-2018	1	Max marine fisheries catches
C09	DensP_Fisheries >= Dens_harvestings	1998-2018	1	Min densities harvestings
C10	DensP_Fisheries <= Dens_harvestings	1998-2018	1	Min densities harvestings
C11	MM_Fisheries >= MM_harvestings * 1.1	1998-2018	1	Min marine fisheries harvestings
C12a	MM_Fisheries <= MM_harvestings * 1.1 + 1.5	1998-1991	1	Max marine fisheries harvestings prior to 1994 - seal catches should be to index 1500
C12b	MM_Fisheries <= MM_harvestings * 1.1 + 1.5	1992-2018	1	Max marine fisheries harvestings prior to 1994 - seal catches should be to index 1500
C13	Oomo_Oseeria <= Oseeria_LocIndex	1998-2018	1	Min ecosystem healthiness
C14	Oomo_Oseeria <= Oseeria_LocIndex * 1.5	1998-2018	1	Min ecosystem healthiness
C15	PF_Oseeria <= PF_Oseeria * 1.5	1998-2018	1	Min ecosystem healthiness
C16	PF_Oseeria <= PF_Oseeria * 1.5	1998-2018	1	Min ecosystem healthiness
C17	Benthos <= 66 * 1000 * Prod_Sat * Leverage * 1.5	1998-2018	1	Max marine benthos biomass
C18	Benthos <= 66 * 1000 / 2	1998-2018	1	Absolute min benthos biomass
C19	MM_Benthos <= 0.3 * 1000 / 2	1998-2018	1	Absolute min benthos biomass
C20	MM >= 0.1 * 1000 / 2	1998-2018	1	Absolute min marine biomass
C21	MM >= 0.0007 * 1000 / 2	1998-2018	1	Absolute min marine biomass
C22	BioAbd >= 0.0007 * 1000 / 2	1998-2018	1	Absolute min benthos biomass
C23	Benthos <= Benthos_Biomass * 2	1998-2018	1	Max marine benthos biomass
C24	PF_Benthos <= Benthos_Biomass * 2	1998-2018	1	Max marine benthos biomass
C25a	Oomo <= Omo_Biomass * 2	(1998-2001, 2002-2017)	1	Max marine ecosystem healthiness
C25b	Oomo <= Omo_Biomass * 2	(2018-2019)	1	Max marine ecosystem healthiness
C26	Oomo <= Omo_Biomass * 2	1998-2018	1	Max marine ecosystem healthiness
C27a	PP_Benthos <= Pelagics * 1.4	(1998-1992, 1995-2000, 2004-2015, 2017-2019)	1	Min absolute pelagic biomass
C27b	PP_Benthos <= Pelagics * 1.4	(1993-1994, 2001-2004, 2006-2010)	1	Max pelagic biomass
C28	PP_Benthos <= Pelagics * 1.4	(1998-1993, 1997-2010)	1	Min absolute pelagic biomass prior to blue whiting data available
C29	PP_Benthos <= Pelagics * 1.4	(1998-1993, 1997-2010)	1	Min pelagic biomass constraint incompatible with inertia in 1996
C30	DensP <= Dens_harvestings * 1.5	1998-2018	1	Min densities harvestings
C31	PP_Dens <= Dens_harvestings * 1.5 * GladiatorsCod + GladiatorsCod * Dens_harvestings / 2	1998-2018	1	Min consumption of demersals by demersals
C32	PP_Dens <= Dens_harvestings * 1.5 * GladiatorsCod + GladiatorsCod * PolarizedCod + PolarizedCod * Herring2Cod	1998-2018	1	Min consumption of demersals by demersals
C33	Oomo_Dens >= ShrimpsCod * KelpCod + AmphipodsCod * KelpCod + HerringCod + PolarizedCod + Cod2Cod + HaddockCod + Gobies2Cod + 4W2Cod	1998-2018	1	Min consumption of ecosystem production by demersals
C34	BioAbd >= 0.007 * 1000 * (DensP + PP_Benthos + DensP * Demer) <= 2 * (Other2Cod + AmphipodsCod + KelpCod + ShrimpsCod + CapelinCod + HerringCod + PolarizedCod + Cod2Cod + HaddockCod + Gobies2Cod + 4W2Cod)	1998-2018	0	One constraint to illustrate incompatibility

Ecosystem Assessment for the Barents Sea (ICES (2020)) have been altered to comply with data access restrictions (not all data is publicly available). They are stored in the “Input time-series” sheet in the RCaN-file. Each row indicate a time-step (year) and each column correspond to a particular data-series. The first row is a header with the name of each date-series. The observational data-series information is stored in the dataframe ‘series’ in the POLYTOPE object:

```
kable(POLYTOPE$series, caption="Observations")%>%
  kable_styling(latex_options = c("striped", "scale_down"))
```

3.2 Constraints

Explicit constraints that relate food-web components, fluxes and observations can be specified in the RCaN file. These are written in the form of inequalities or equalities and are provided in the worksheet “Constraints”. A constraint can be set to apply to every year or to selected years only. A constraint can be set to active/inactive (column ‘active’), which is practical for testing different combination of constraints without the need for writing/deleting the constraints in the worksheet. The constraint information is stored in the dataframe ‘constraints’ in the POLYTOPE object.

```
kable(POLYTOPE$constraints[,1:5],caption='Constraints defined by user') %>%
  kable_styling(latex_options = c("striped", "scale_down"), font_size = 8)
```

4 The polytope behind a CaNmod object

In section 3, we used the `buildCaN` function to read the RCaN file, but also to construct the corresponding polytope. The polytope is a multidimensional geometrical object that corresponds to the “space of possible food-web trajectories”. The dimension of the polytope is defined by the number of fluxes times the number of time-steps (years) + the number of components. The different elements of the CaNmod object constructed

using the buildCaN function are as follows:

```
names(POLYTOPE)

## [1] "components_param" "species"           "fluxes_def"      "flow"
## [5] "series"            "ntstep"             "aliases"         "dynamics"
## [9] "data_series_name" "constraints"       "H"                "N"
## [13] "Nend"              "A"                 "AAll"            "C"
## [17] "CAll"              "v"                 "vAll"            "L"
## [21] "b"                 "bAll"              "symbolic_enviro"
```

Matrices C and A, and vectors b and v correspond to the matrices of active constraints that define the polytope $A X \leq b$ and $C X = v$ (AAll, CAll, bAll and vAll are similar but include all constraints, active or not). Matrices N, H, F and L are matrices that are used to describe the dynamics of the system, more specifically, they describe how biomasses at any time can be derived from flows, using the relationship provided in the supplementary material of Planque and Mullon Planque and Mullon (2020).

4.1 Structure of the polytope

The polytope is defined inside a space with a high number of dimensions.

For the Barents sea example, we have:

```
paste("number of inequalities:", nrow(POLYTOPE$A))

## [1] "number of inequalities: 2593"
paste("number of equalities:", nrow(POLYTOPE$C))

## [1] "number of equalities: 64"
paste("number of parameters (dimensions):", ncol(POLYTOPE$C))

## [1] "number of parameters (dimensions): 775"
```

The inequalities and equalities result from both implicit and explicit constraints, each line corresponds to a constraint and a specific year. The number of parameters is the dimension of the polytope.

4.2 Suitability of the polytope

The polytope contains the set of possible food-web trajectories. If some constraints are incompatible, the polytope will be empty, signifying that it is not possible to find any food-web trajectories that can simultaneously satisfy all constraints. When there are too few constraints, the polytope can be unbounded. In such case, some of the fluxes can take an unbounded range of values, a somehow unrealistic situation. Checking the polytope is an important step. Users can adopt different strategy: some users try first to specify as many constraints as possible with the risk of constructing an empty polytope. They then need to relax some of the model constraints in order to find possible solutions. Others tend to start with a reduced set of constraints with the risk of having an unbounded polytope. They then need to add extra constraints to derive possible and plausible food-web trajectories. Both approaches have their pros and cons. Here, we focus on how to identify potential issues with the polytope and the possible ways to resolve them so that the polytope is suitable for sampling. As a recommendation: it is often a good strategy to start working with a limited number of years to limit the dimension of the polytope and decrease computation time. Moreover, it is often convenient to add a temporary constraint that specifies an upper limits to the sum of all flows; it ensures to have a bounded polytope and to carry out some preliminary explorations.

4.2.1 First step: checking the polytope status

The R command `checkPolytopeStatus` returns the current status of the polytope. In the Barents Sea example, the polytope is 'OK' which means that it is non-empty and bounded. In other words, there is a limited set of

possible trajectories.

```
checkPolytopeStatus(POLYTOPE)
```

```
## [1] "polytope ok"
```

In some situations, the function may return ‘empty polytope’ or ‘unbounded polytope’. It might then be necessary to add/activate new constraints or remove/disactivate existing constraints.)

4.2.2 Toggling constraints

Constraints can be activated/deactivate in the RCaN-file, using the ‘active’ switch in the ‘Constraints’ worksheet. In addition, it is possible to activate/deactivate constraints directly within R, once the RCaN file has been loaded. We illustrate this below, using the command *toggleConstraint*:

```
POLYTOPE <- quiet(toggleConstraint(POLYTOPE,
                                      c("C01", "C02", "C03", "C04", "C05", "C06",
                                         "C07", "C08", "C09", "C10", "C11", "C12",
                                         "C15", "C16", "C19", "C21")))
```

In this example, we removed constraints C01 to C12 and C15, C16, C19 and C21, which bound the input (primary production), the output of the system (fisheries) and the maximum biomass of top predators (birds and marine mammals).

```
checkPolytopeStatus(POLYTOPE)
```

```
## [1] "polytope not bounded"
```

As a result, the polytope is now unbounded. We therefore reactivate the constraint with the ‘*toggleConstraint*’ command.

```
POLYTOPE <- quiet(toggleConstraint(POLYTOPE,
                                      c("C01", "C02", "C03", "C04", "C05", "C06",
                                         "C07", "C08", "C09", "C10", "C11", "C12",
                                         "C15", "C16", "C19", "C21")))
```

4.2.3 Finding incompatible constraints

We artificially add a new constraint which is incompatible with the existing ones to illustrate a situation where the polytope is empty.

```
POLYTOPE<-quiet(toggleConstraint(POLYTOPE, "C35"))
```

From section 3.2, we can see that constraint C35 (the biomass of birds should exceed 33.6) is not compatible with constraints C21 (the biomass of birds should NOT exceed 44.8), i.e. the two can not be fulfilled simultaneously. As a consequence, checking the polytope status returns ‘empty polytope’:

```
checkPolytopeStatus(POLYTOPE)
```

```
## [1] "empty polytope"
```

The RCaNmodel function ‘findingIncompatibleConstr’ is then useful to identify which constraints are incompatible:

```
options(width = 60)
incomp <- findingIncompatibleConstr(POLYTOPE)
```

```
...
```

```
## [1] "###polytope is ok when following constraints are relaxed:"
```

```
## [1] " C35 : 1988, C35 : 1989, C35 : 1990, C35 : 1991, C35 : 1992, C35 : 1993, C35 : 1994, C35
```

```
## [1] "####Those constraints seem incompatible with:"
```

```

## [[1]]
## [1] " C35 : 1988: C21 : 1988, C35 : 1988, C35 : 1989, C35 : 1990, C35 : 1991, C35 : 1992, C35
##
## [[2]]
## [1] " C35 : 1989: C21 : 1989, C35 : 1988, C35 : 1989, C35 : 1990, C35 : 1991, C35 : 1992, C35
##
## [[3]]
...

```

The function returns that constraint C35 is problematic (see `####polytope` is ok when following constraints are relaxed) and that it is incompatible with constraints C21 (“`####Those constraints seem incompatible with:`”).

In such situation, it is necessary to relax some of the above constraints (C35, C21) to obtain a non-empty polytope. The choice of the constraint to relax is left to the modeller, based on domain knowledge.

In the following, we revert to a model version in which constraint C35 is inactive and the polytope is OK:

```
quiet(POLYTOPE<-toggleConstraint(POLYTOPE, "C35"))
```

4.3 Bounds of the polytope

To obtain a first overview of the polytope, it is useful to compute its limits in all dimensions. This can be achieved with the RCaNmodel function `getAllBoundsParam`. The output is a dataframe with a row per parameter/dimension. It is also possible to compute the bounds for a small set of dimensions only. This can be done using the function `getBoundParam`. This is illustrated below for the two dimensions corresponding to the flux from Omnivorous Zooplankton to Birds in 2017 and 2018:

```
getBoundParam(POLYTOPE, which(colnames(POLYTOPE$A)=="Ozoo_Birds[2017]"))
```

```

##      min      max
## 0.000 2666.667

```

```
getBoundParam(POLYTOPE, which(colnames(POLYTOPE$A)=="Ozoo_Birds[2018]"))
```

```

##      min      max
## 0.000 2666.667

```

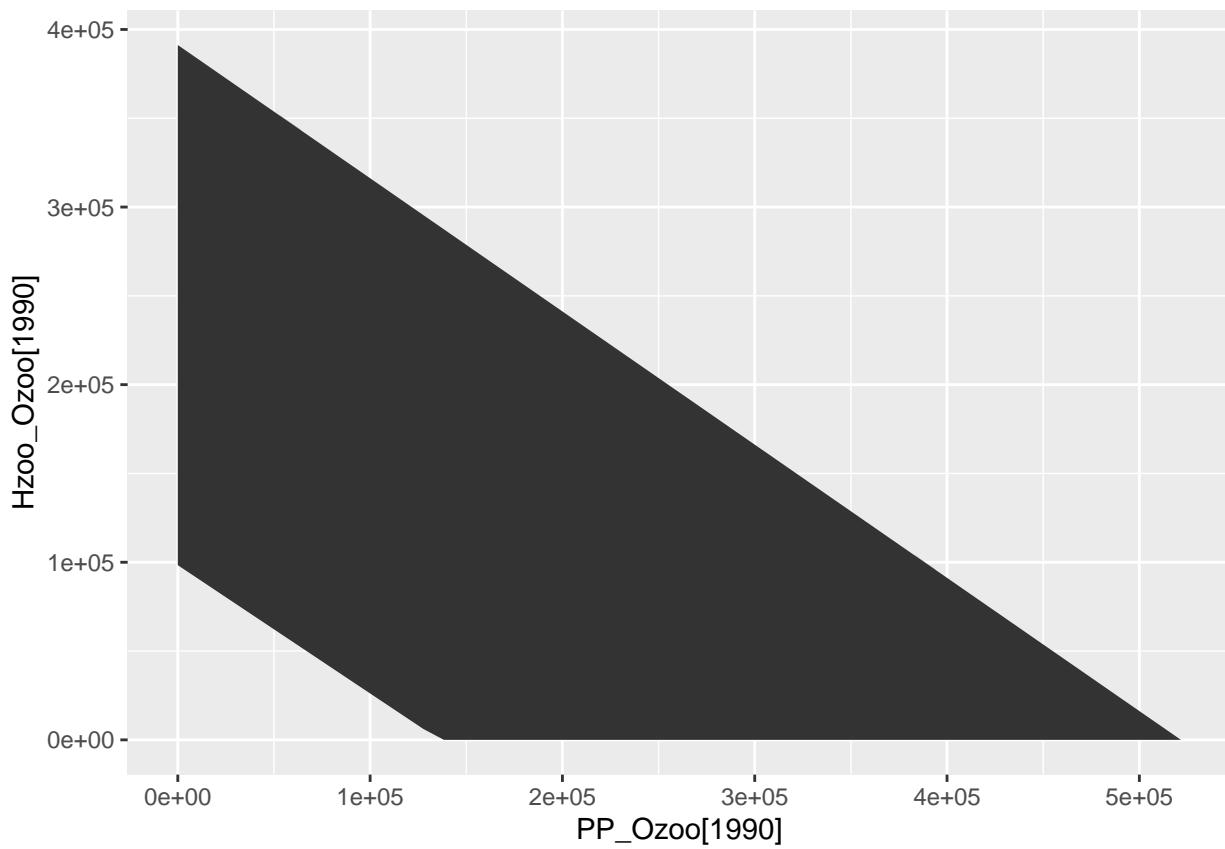
4.4 2D-slices of the polytope

2-dimensional pictures of the polytope are useful to visualise how the bounds of different fluxes are jointly constrained. These are obtained using the RCaNmodel function `plotPolytope2D`:

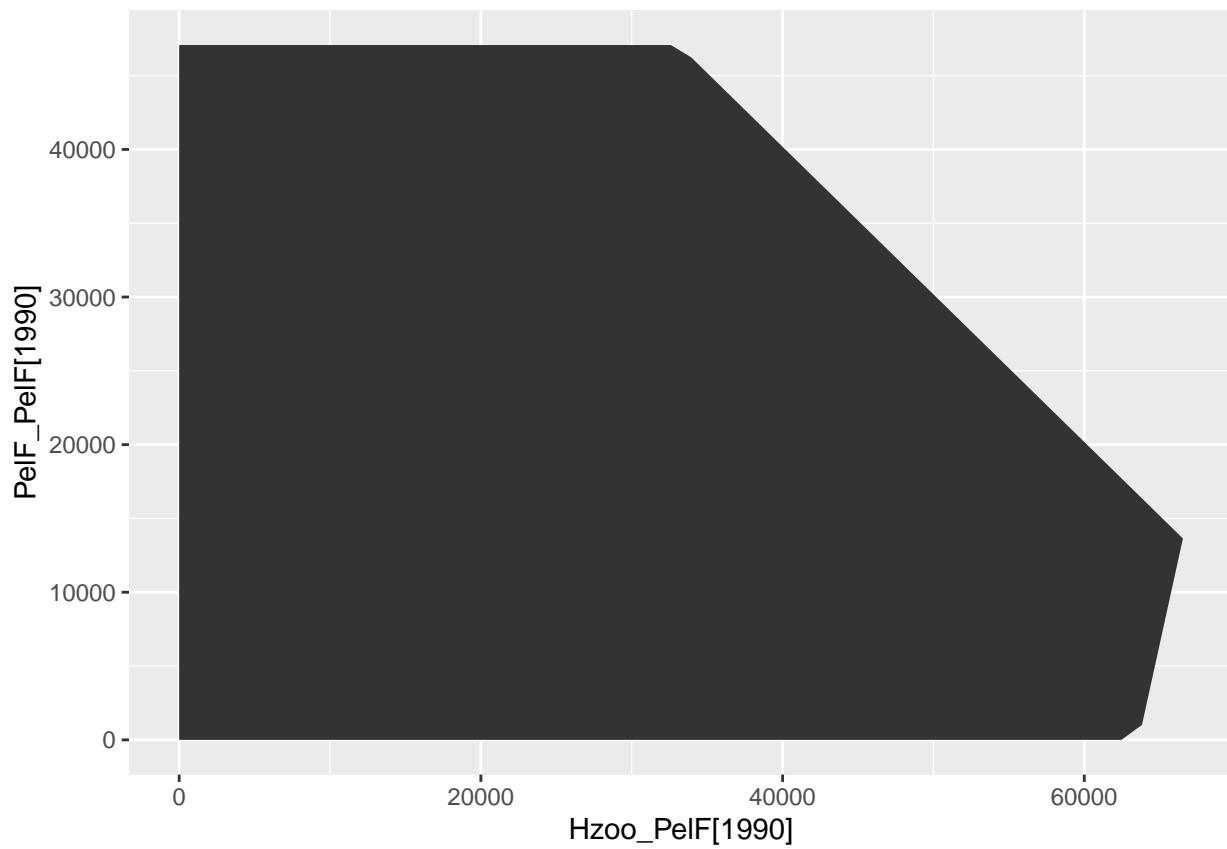
```

start_time <- Sys.time()
plotPolytope2D(POLYTOPE,
               c('PP_Ozoo[1990]', 'Hzoo_Ozoo[1990]'),
               progressBar=FALSE)

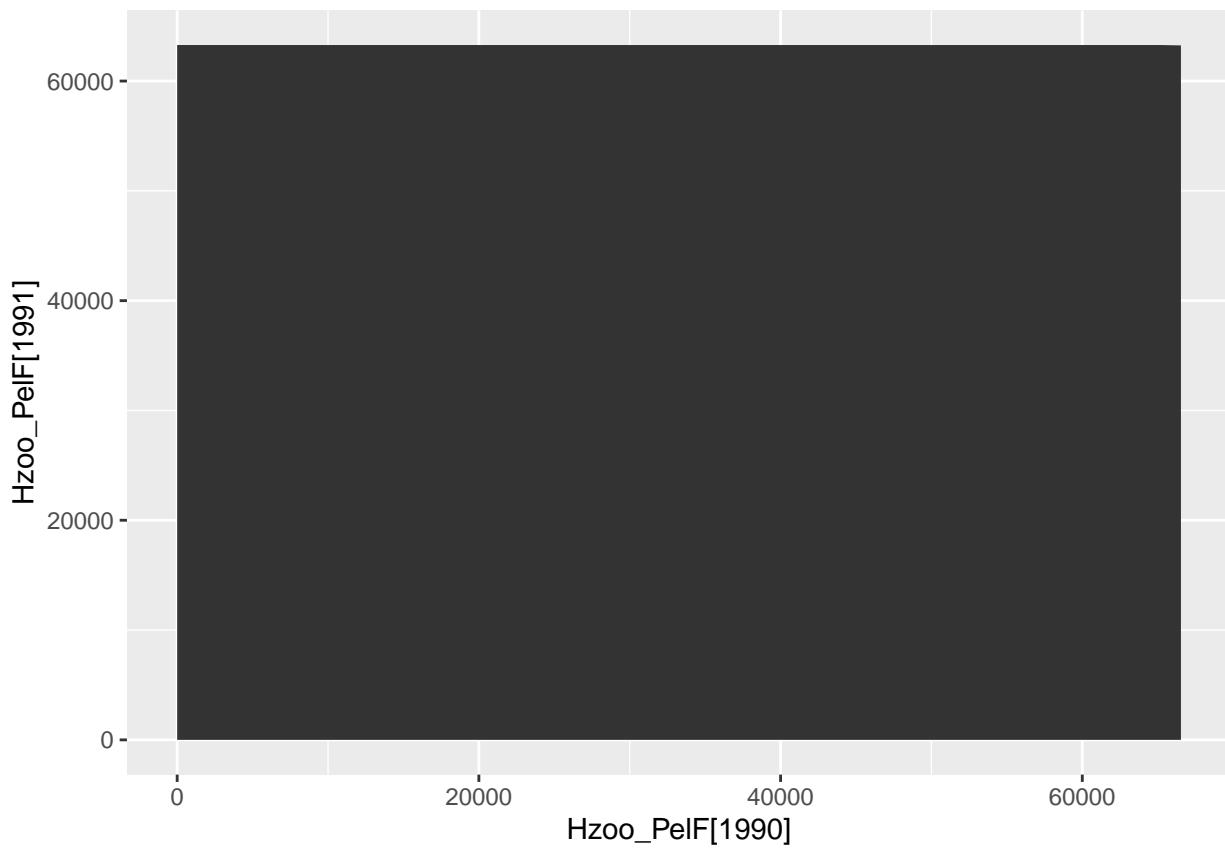
```



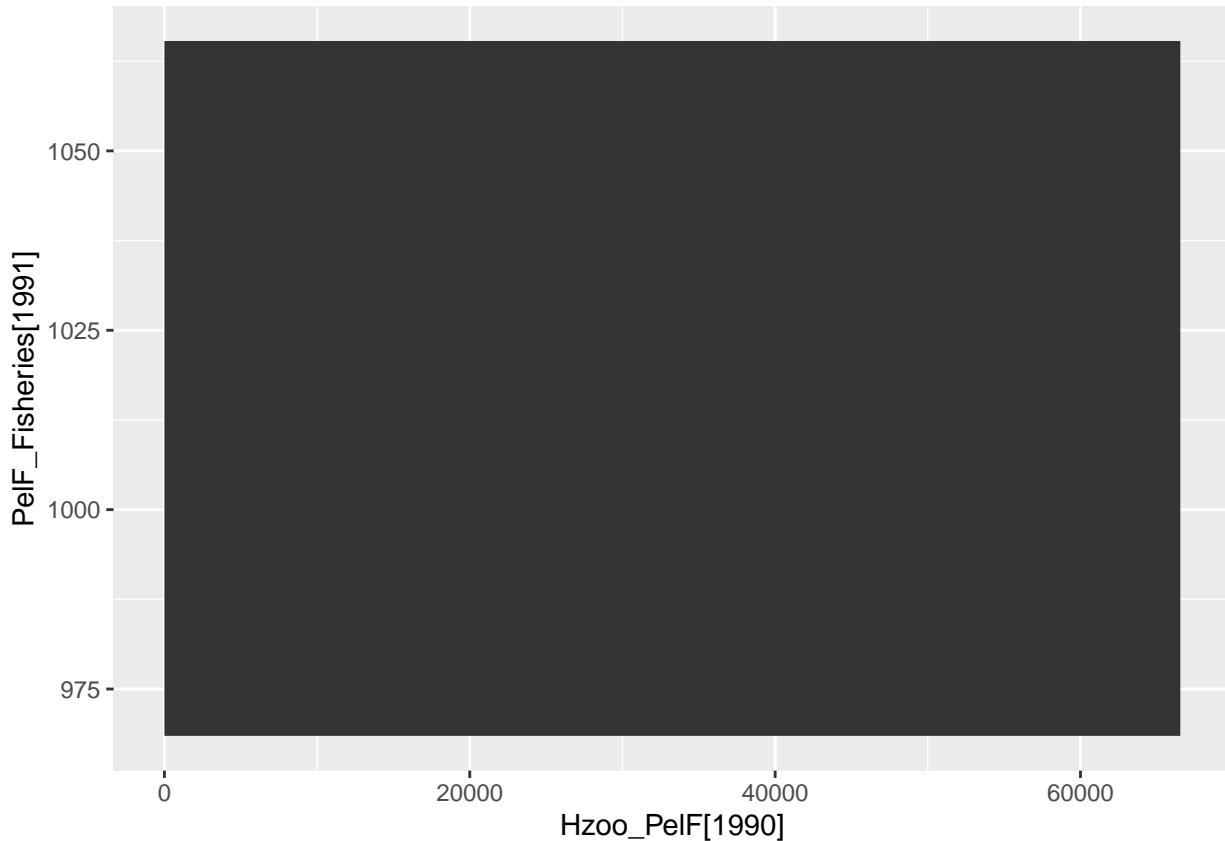
```
plotPolytope2D(POLYTOPE,
                 c('Hzoo_PelF[1990]', 'PelF_PelF[1990]'),
                 progressBar=FALSE)
```



```
plotPolytope2D(POLYTOPE,
                 c('Hzoo_PelF[1990]', 'Hzoo_PelF[1991]'),
                 progressBar=FALSE)
```



```
plotPolytope2D(POLYTOPE,
                 c('Hzoo_PelF[1990]', 'PelF_Fisheries[1991]'),
                 progressBar=FALSE)
```



```
end_time <- Sys.time()
end_time - start_time

## Time difference of 3.809366 mins
```

5 Sampling the polytope

5.1 Sampling

The RCaNmodel function *sampleCaN* is used to sample of the polytope, i.e. to sample several random possible trajectories of the food-web dynamics. The output of this command is a mcmc.list object (defined in package coda). The function parameters include:

- * N: the number of samples
- * thin: the thinning, i.e. the number of samples that are left out from the output
- * nchains: the number of sampling chains
- * ncore: the number of CPU cores used
- * method: one of gibbs (default) or hitandrun

With N=100, thin=10, and nchain=2 the *sampleCaN* function will sample two chains of 1,000 values and return one output every 10, that is a total of 200 valid samples

```
if (file.exists("sampleCaN.rdata")){
  load("sampleCaN.rdata")
} else {
  system.time(SAMPLE <- sampleCaN(POLYTOPE, N=100, thin=10, nchain=2, ncore=2))
  save(SAMPLE, file = "sampleCaN.rdata")
}
```

5.2 Convergence of the sample chains

Check the mixing of the Monte Carlo Marko chain algorithm that is used to build the sample can be achieved with commands from the coda library.

```
summ <- summary(SAMPLE$mcmc)
n = 31
kable( head(cbind.data.frame(summ$statistics, summ$quantiles) %>%
  select('Mean', 'SD', '2.5%', '50%', '97.5%'),
  n=n),
  caption=paste("summary of", n, "parameter distributions"),
  digits=0) %>%
kable_styling(latex_options = c("striped", "scale_down"))
```

5.2.1 Gelman diagnostic

Gelman and Rubin (1992) proposed a general approach to asses the convergence of MCMC outputs in which $m > 1$ parallel chains are run with starting values that are overdispersed relative to the posterior distribution. Convergence is diagnosed when the chains have ‘forgotten’ their initial values, and the output from all chains is indistinguishable. The gelman.diag diagnostic is applied to a single variable from the chain. It is based a comparison of within-chain and between-chain variances, and is similar to a classical analysis of variance.

```
gelman.diag(SAMPLE$mcmc[,c('PP_Benthos[1990]',
  'Ozoo_Birds[1990]',
  'DemF_Fisheries[1990]',
  'PelF_DemF[1990]')],
  multivariate = FALSE)

## Potential scale reduction factors:
##
##          Point est. Upper C.I.
## PP_Benthos[1990]      1.03    1.06
## Ozoo_Birds[1990]      1.04    1.21
## DemF_Fisheries[1990]   1.04    1.20
## PelF_DemF[1990]       1.06    1.26
```

Theoretically a statistics below 1.05 suggests an appropriate convergence.

5.2.2 Autocorrelation function

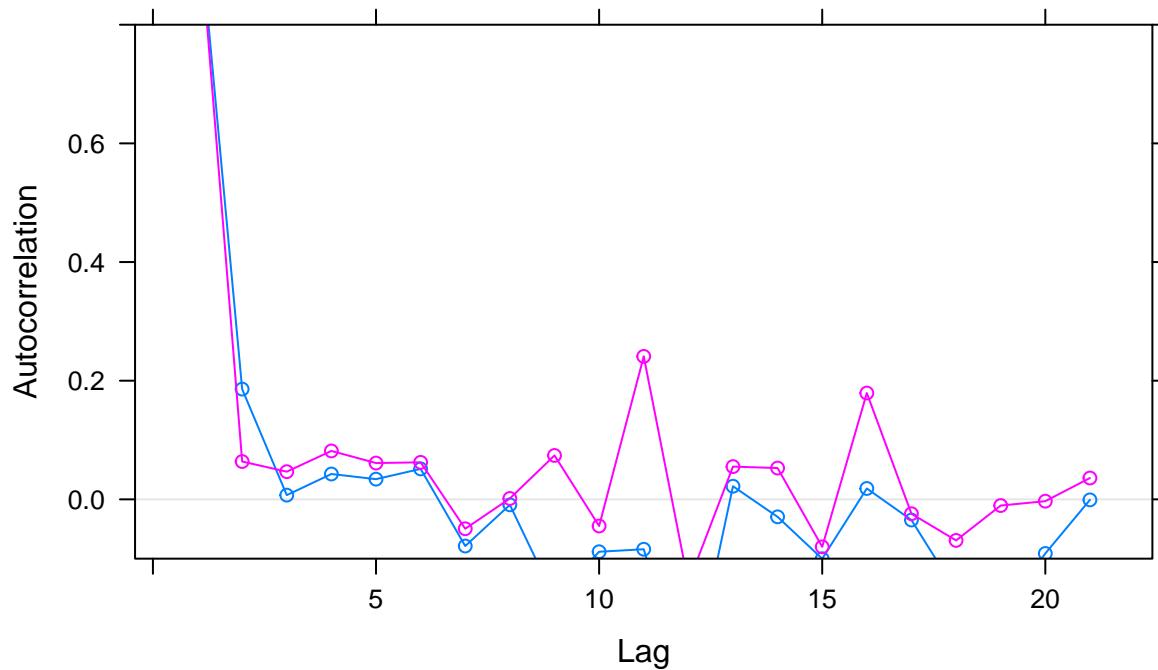
The *acfplot* function from the coda library shows the autocorrelation function of sampling chains. It is illustrative of the independence of successive samples. If the samples are not serially correlated (which is desired) the autocorrelation should remain around zero (except for the first value which is always 1). Otherwise, it might be worthwhile to increase the thinning during RCaNmodel sampling and/or to discard the first samples.

```
acfplot(SAMPLE$mcmc[, 'PP_Benthos[1990)'],
  ylim=c(-0.1,0.8),
  main='PP_Benthos[1990]')
```

Table 5: summary of 31 parameter distributions

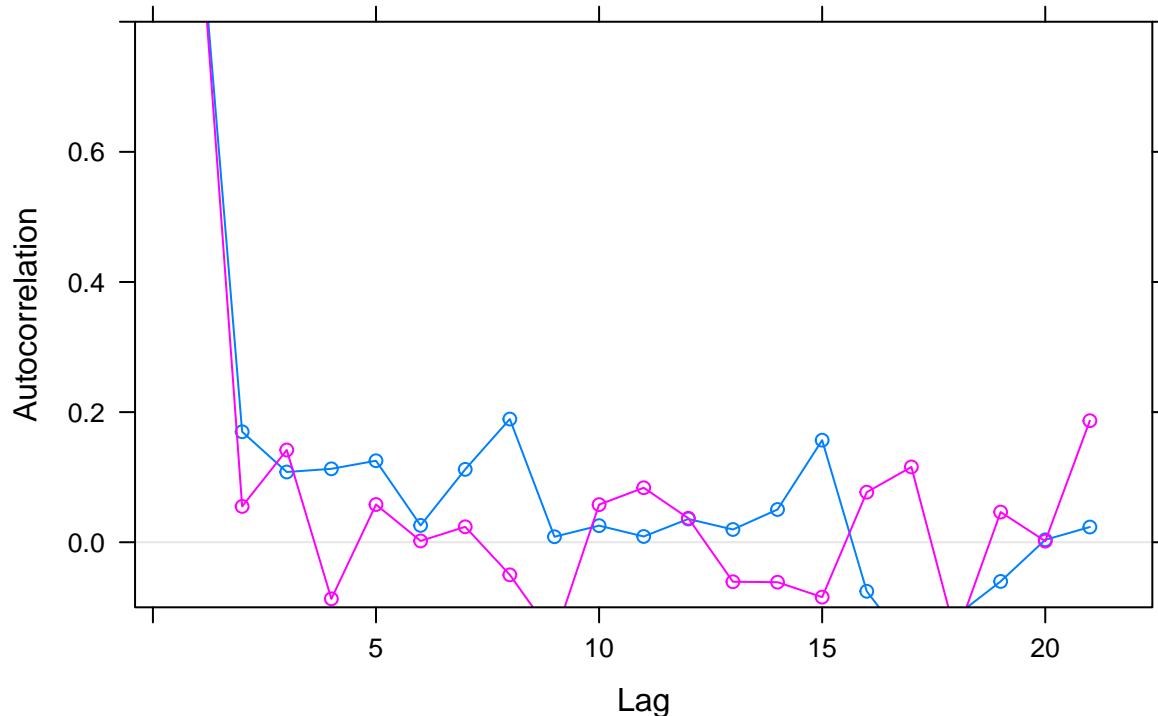
	Mean	SD	2.5%	50%	97.5%
PP_Ozoo[1988]	214808	133334	17269	191936	494022
PP_Hzoo[1988]	865830	230974	470279	850205	1344889
PP_Benthos[1988]	659391	227324	282740	649583	1096390
Hzoo_Ozoo[1988]	150207	91959	7901	137421	333707
Hzoo_PelF[1988]	7929	4651	420	7910	16060
Ozoo_Ozoo[1988]	1147095	683535	99058	1056117	2622754
Ozoo_PelF[1988]	8130	4656	600	7775	17055
Ozoo_DemF[1988]	2717	662	1791	2605	4220
Ozoo_MM[1988]	6043	1702	2463	6471	8669
Ozoo_Birds[1988]	1057	662	44	1029	2229
Benthos_Benthos[1988]	368841	277017	13122	300944	950440
Benthos_DemF[1988]	882	654	50	732	2388
PelF_PelF[1988]	2071	1604	54	1698	6231
PelF_DemF[1988]	1516	527	740	1454	2646
PelF_MM[1988]	1976	1571	57	1551	5730
PelF_Birds[1988]	883	605	26	773	2013
DemF_DemF[1988]	599	320	223	516	1297
DemF_MM[1988]	499	336	20	442	1276
NorSeaZoo_Hzoo[1988]	12800	0	12800	12800	12800
NorSeaZoo_Ozoo[1988]	3200	0	3200	3200	3200
PelF_Fisheries[1988]	0	0	0	0	0
DemF_Fisheries[1988]	888	25	847	891	928
MM_Fisheries[1988]	1	0	0	1	2
Ozoo_Fisheries[1988]	55	2	52	55	57
PP_Ozoo[1989]	82979	52060	7915	79744	186820
PP_Hzoo[1989]	459605	141233	209739	444772	713586
PP_Benthos[1989]	926577	308764	411940	903412	1511172
Hzoo_Ozoo[1989]	80289	38522	10231	81098	147290
Hzoo_PelF[1989]	14209	7982	1572	15063	28555
Ozoo_Ozoo[1989]	510648	299139	37509	486652	1087051
Ozoo_PelF[1989]	12748	8220	199	11067	29727

PP_Benthos[1990]



```
acfplot(SAMPLE$mcmc[, 'Ozoo_Birds[1990]' ],  
        ylim=c(-0.1,0.8),  
        main='Ozoo_Birds[1990]')
```

Ozoo_Birds[1990]

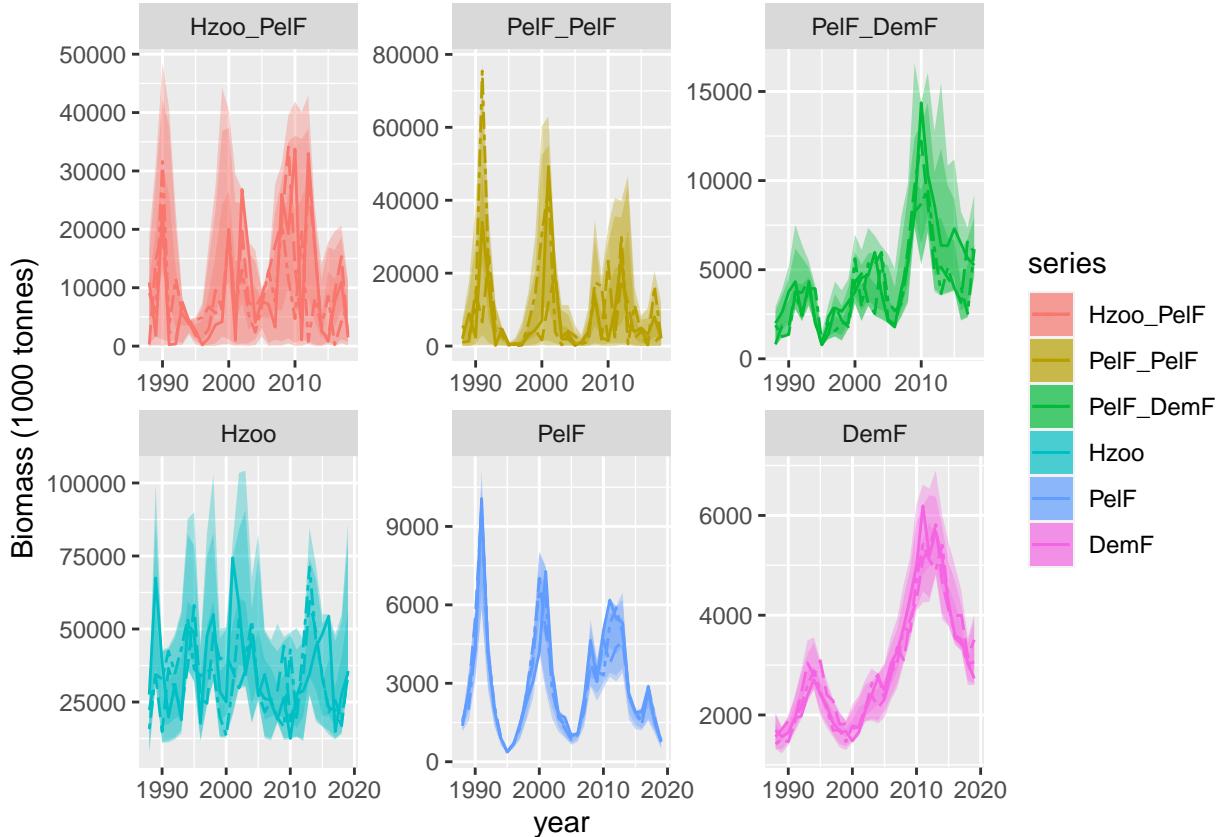


6 Visualisation of the model results

The graphical representation of the possible dynamics of the food-web is an important output of the model. The RCaNmodel function *ggSeries* produces standardised plots of the time-series of components and/or fluxes. By default, for each component, 3 randomly picked samples/trajectories are plotted (plain, dashed and dotted lines) together with the envelopes containing 100%, 95% and 50% of the sampled trajectories.

For components, we get:

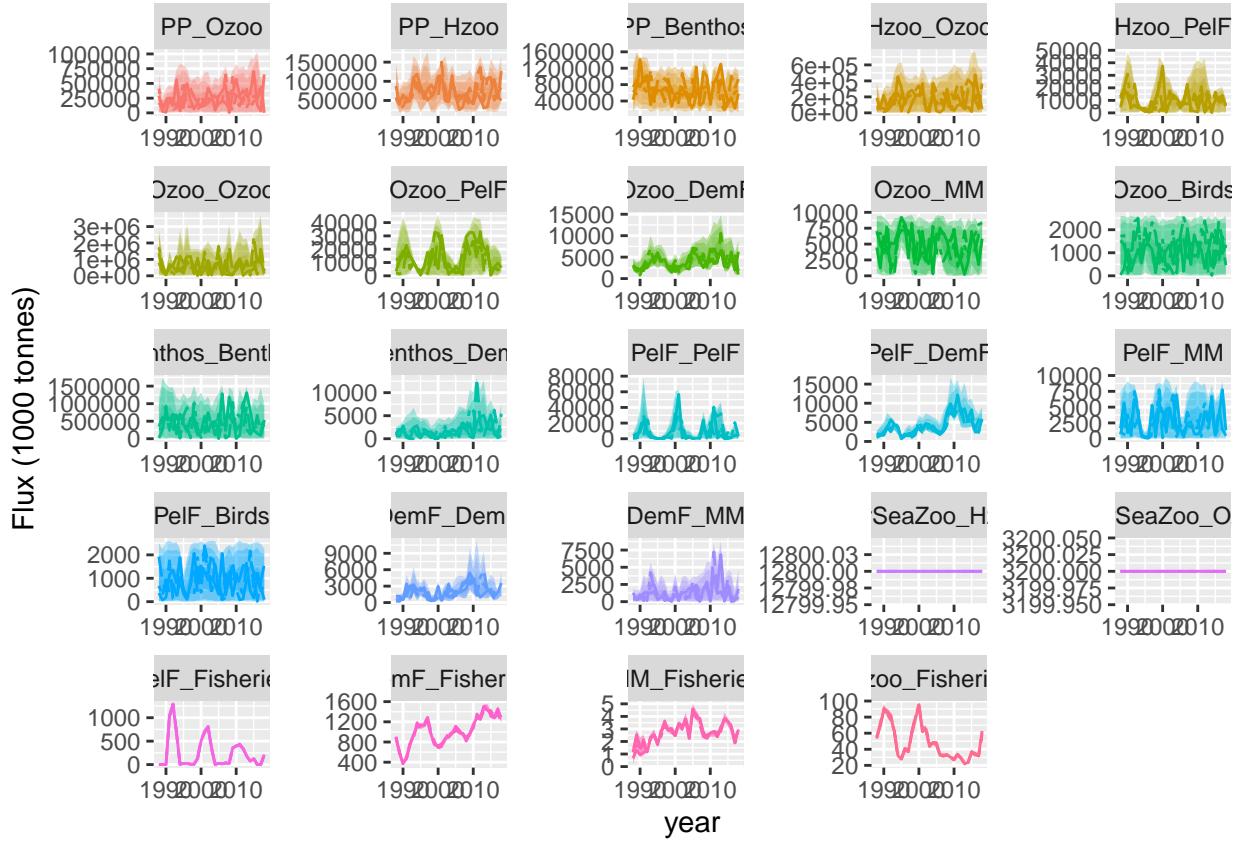
```
ggSeries(SAMPLE,c('Hzoo_PelF','PelF_PelF','PelF_DemF','Hzoo','PelF','DemF'))+
  ylab("Biomass (1000 tonnes)")
```



In this example we observe different situations for the different species: for pelagic fish (PelF) and demersal fish (DemF) the constraints imposed in relation to the observational data-series constrain the trajectories of biomass which are all situated within a narrow band. On the other hand, there is little information provided to constrain directly the trajectories of marine mammal and Birds which leads to large variability between the different samples.

The dynamics of fluxes can be plotted in a similar fashion:

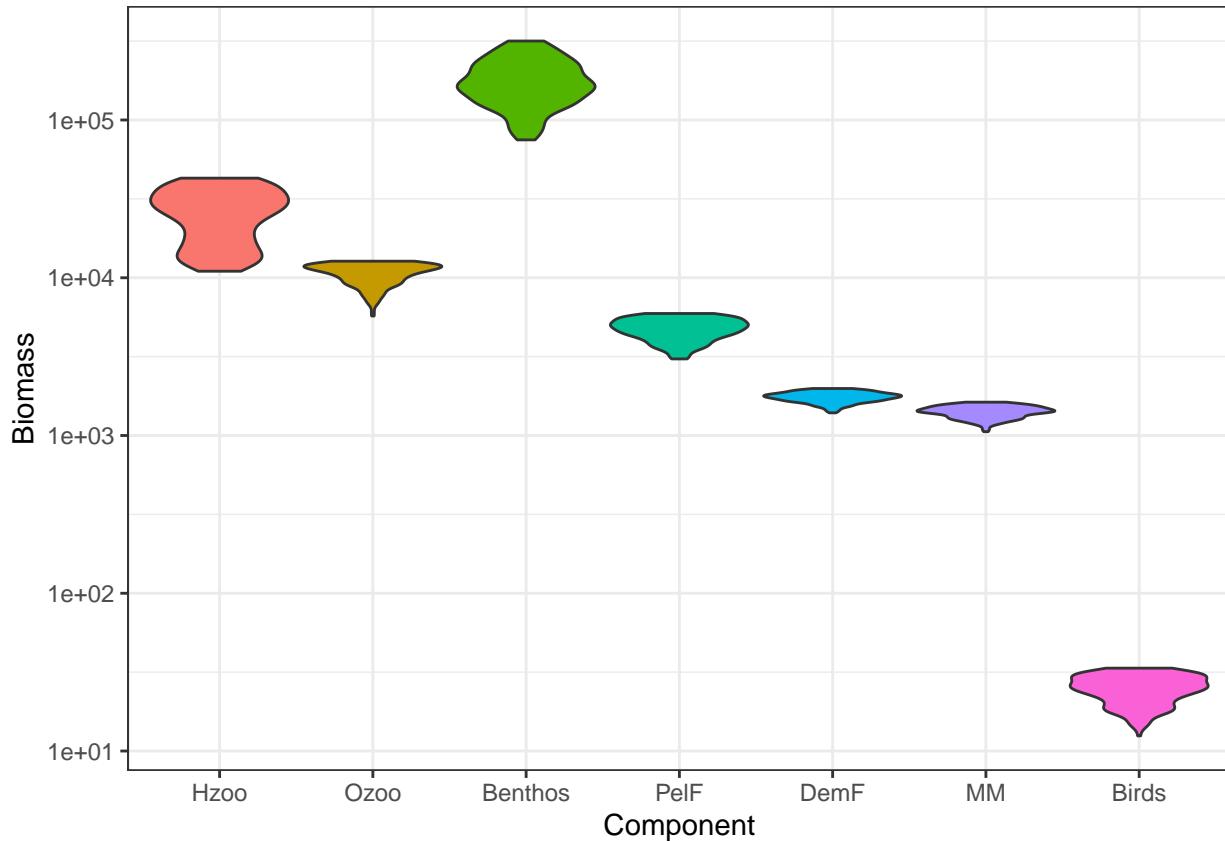
```
ggSeries(SAMPLE, POLYTOPE$flow, TRUE) +
  theme(legend.position = "none") +
  ylab("Flux (1000 tonnes)")
```



6.1 Empirical distributions of biomass and fluxes

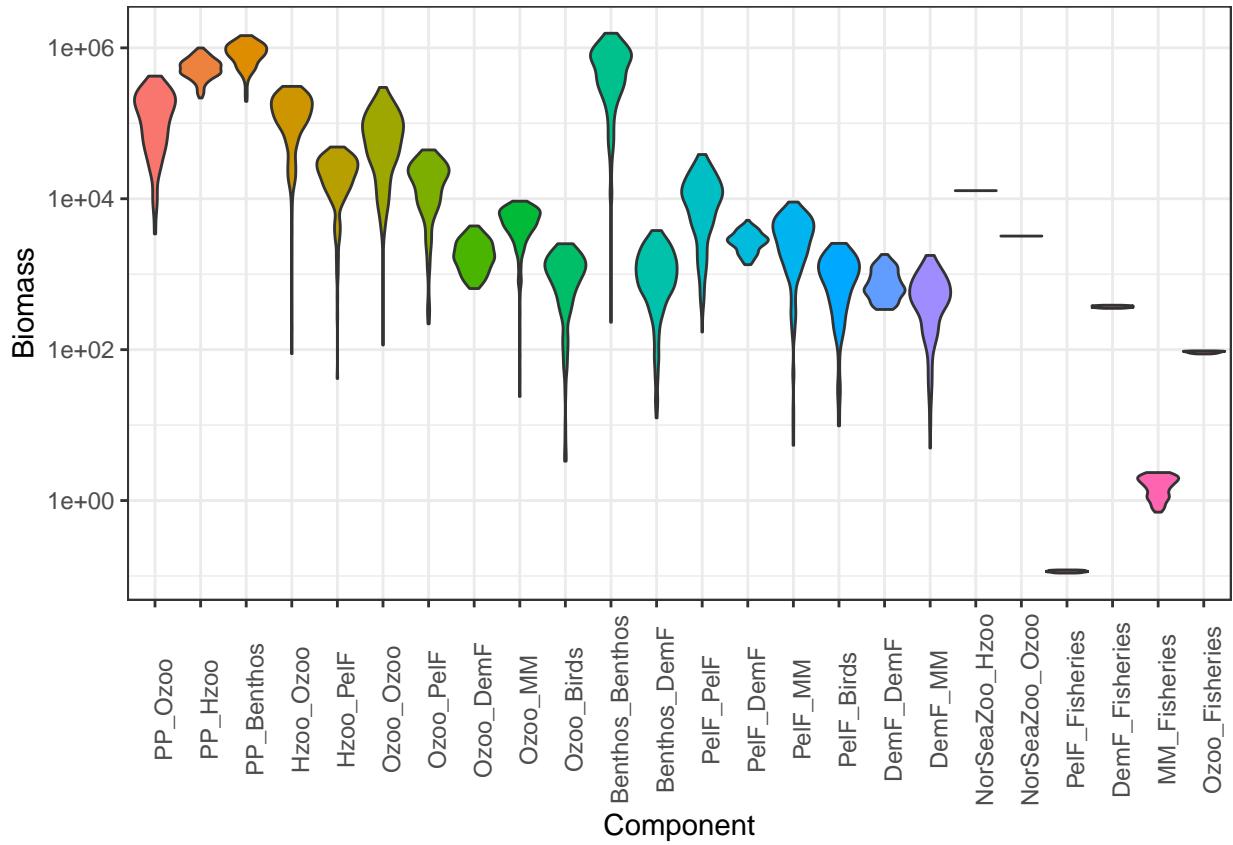
Exploring the distribution of biomass and fluxes across samples and across years is also a useful output of the model. The RCaNmodel function *ggViolin* produces standardised violin plots of these distributions. By default, these are presented on a log scale which allows for comparison between biomass or fluxes that can span several orders of magnitude. For example, the distribution of species biomass for the year 1990 is obtained as follows:

```
ggViolin(SAMPLE,POLYTOPE$species,year=1990,TRUE) +
  xlab('Component') +
  ylab('Biomass')
```



The violin plots of all fluxes in 1990 are produced as follows::

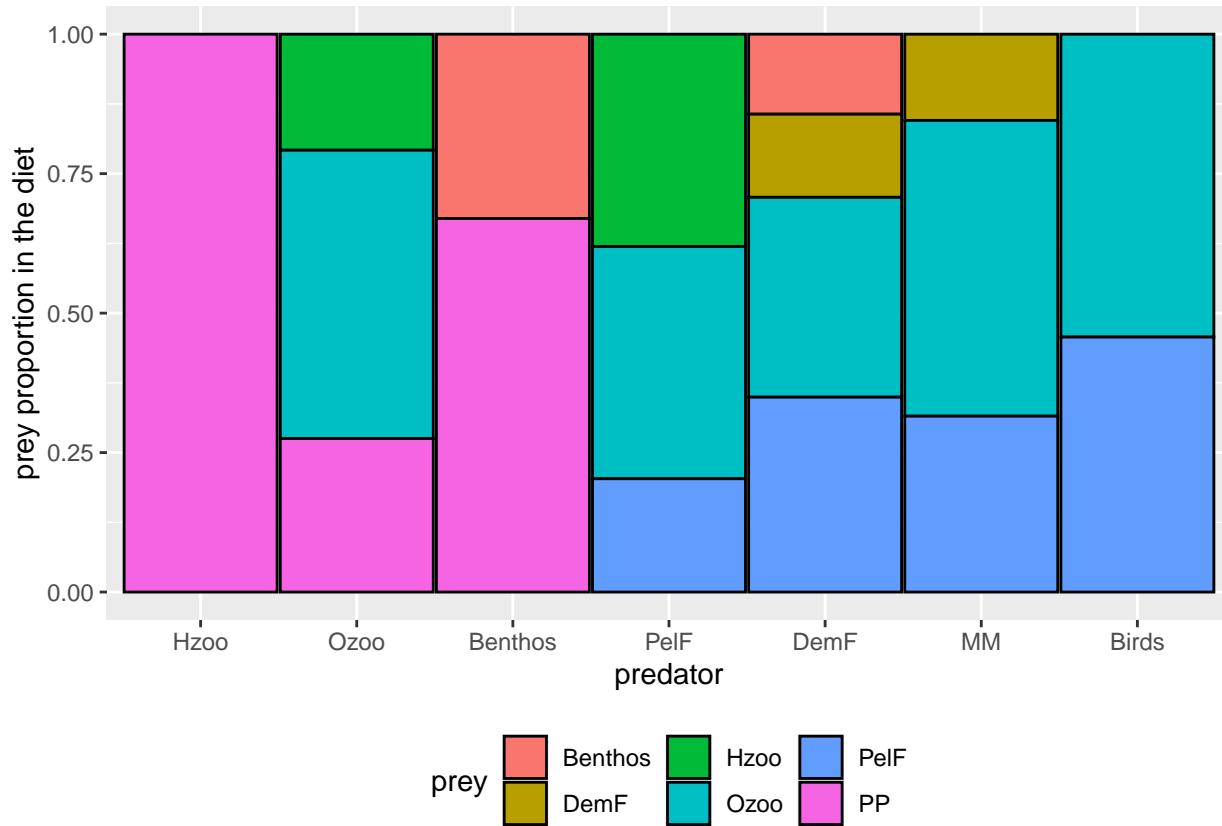
```
ggViolin(SAMPLE,POLYTOPE$flow,year=1990,TRUE) +
  xlab('Component') +
  ylab('Biomass') +
  theme(axis.text.x=element_text(angle=90))
```



6.2 Diet composition

The diet matrix is a key parameter of many trophic models, such as e.g. Ecopath (Christensen and Walters (2004)). In CaN models, diet is non deterministic, in the sense that trophic fluxes are constrained but not determined by biomass. Diets are thus emerging from the model and it is interesting to explore the diets of individual species. A convenient way is to use the RCaNmodel function *ggDiet* to draw the average diet for each predators:

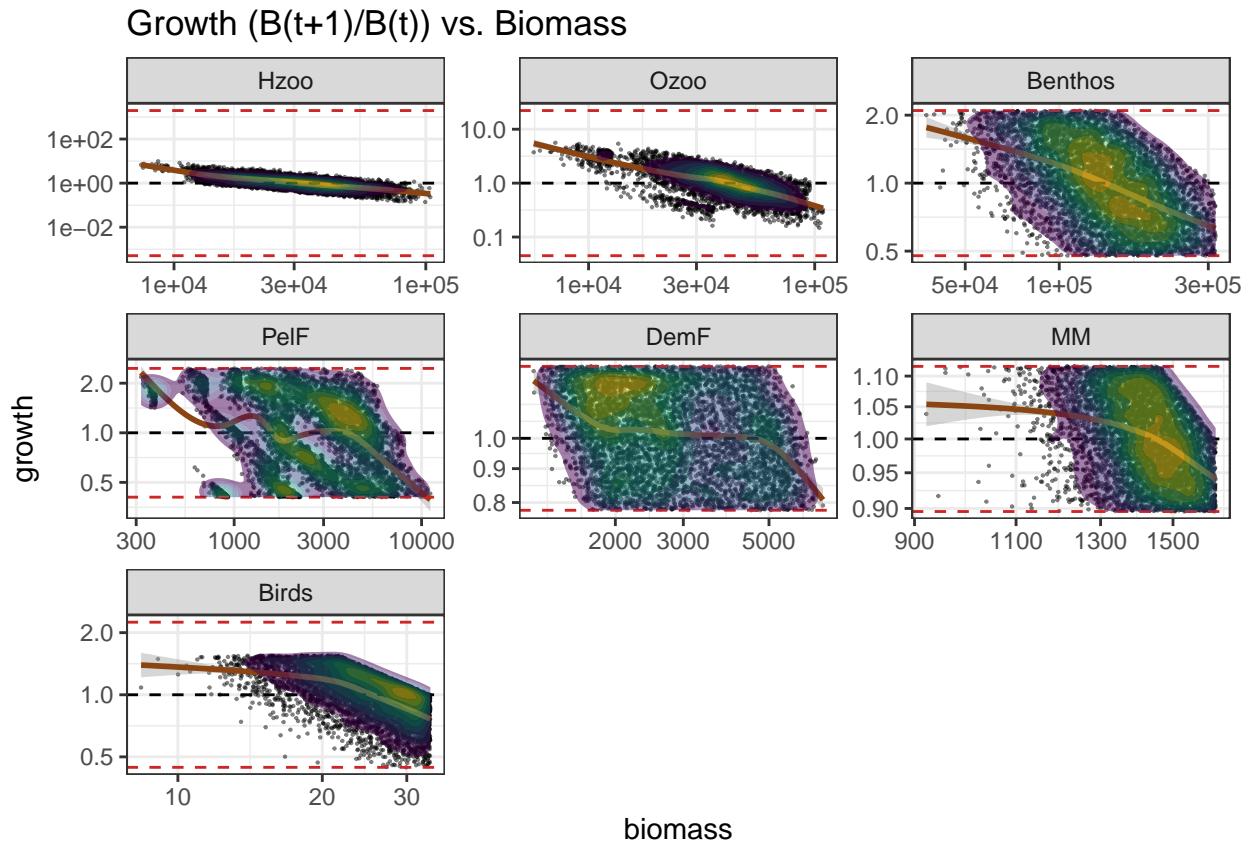
```
ggDiet(SAMPLE, POLYTOPE$species) + ylab('prey proportion in the diet')
```



6.3 Growth and density-dependence

Density-dependence usually reflects how growth is related to population size. In RCaNmodel trajectories this can be expressed as growth in biomass in relation to biomass. The RCaNmodel function *ggGrowth* plots the growth rate (measured as the ratio of biomass at time $t + 1$ over the biomass at time t) as a function of the biomass (at time t). Plotting density-dependence for all species is done as follows:

```
ggGrowth(SAMPLE, POLYTOPE$species)
```

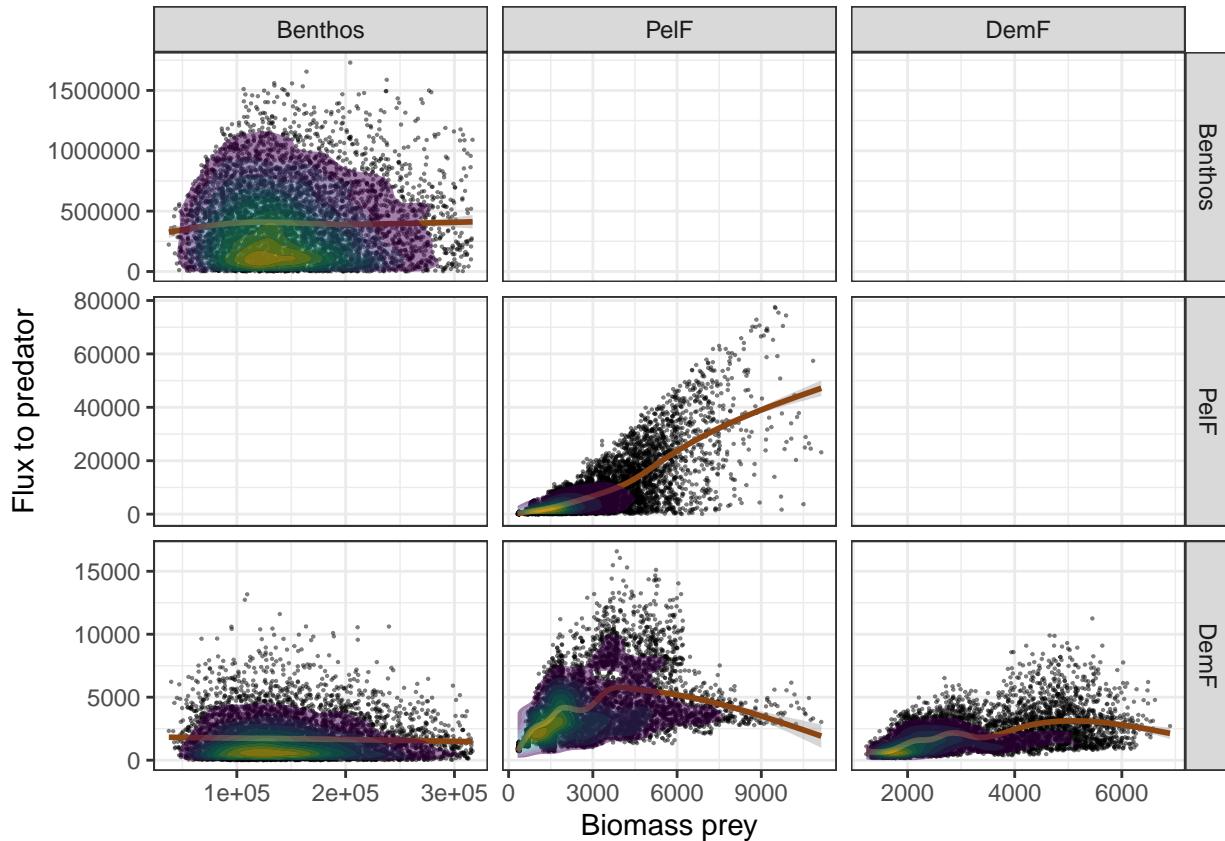


This illustrates how density-dependence of the food-web components can emerge from the food-web dynamics. The upper and lower red-dotted lines visualise the minimum and maximum growth rates set by the inertia constraints for each component. The thick lines are smoothers which highlight the possible underlying relationship between growth and biomass.

6.4 Trophic functional relationships

The RCaNmodel function *ggTrophicRelation* plots the empirical trophic functional relationships (TFRs) between the biomass of a prey and the quantity of that prey consumed by a predator. Below is the command for plotting the TFR between benthos, pelagic fishes and demersal fishes in the Barents Sea example.

```
ggTrophicRelation(SAMPLE, POLYTOPE$species[3:5])
```

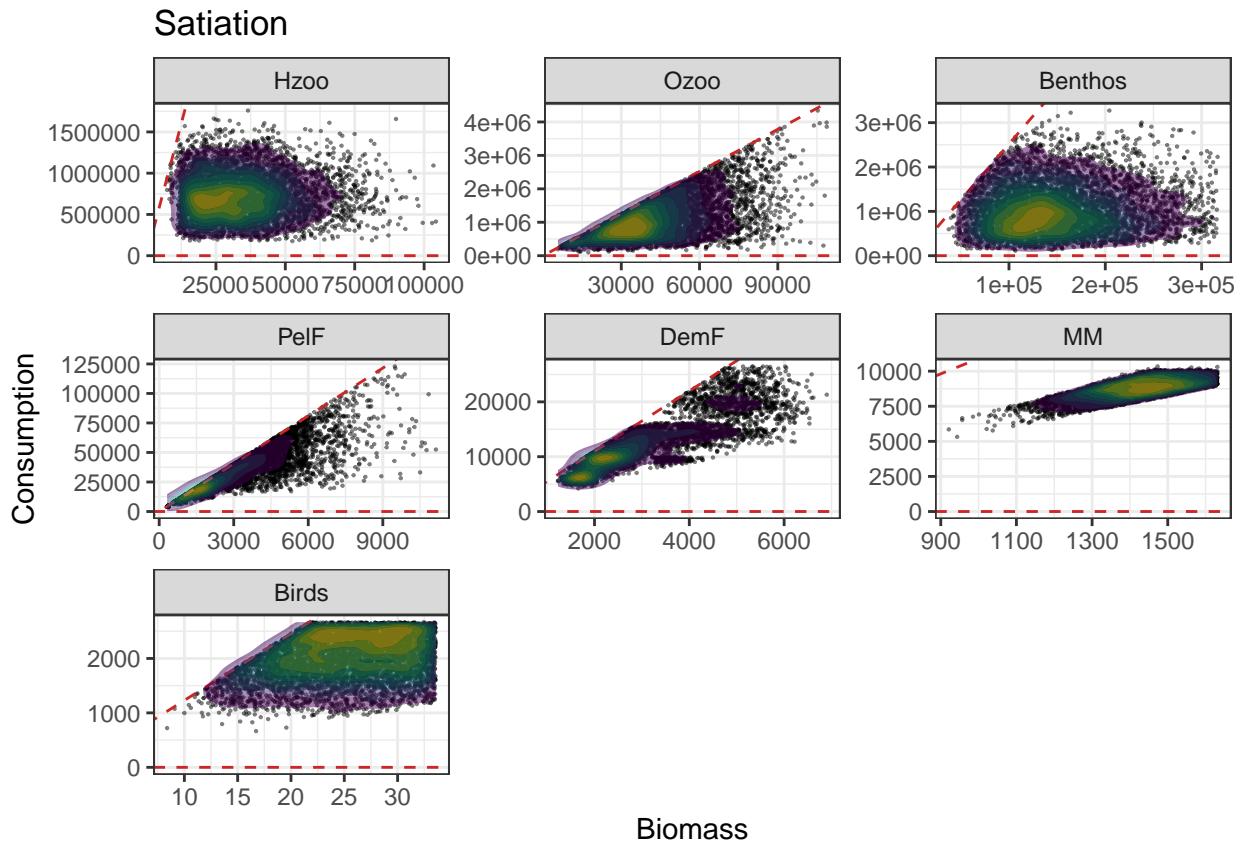


Above, we see that the predation of benthos by demersal fishes does not increase with the biomass of benthos, suggesting that benthos is not a limiting factor in the diet of demersal fishes.

6.5 Satiation

The function *ggSatiation* plots the total biomass of prey eaten by a predator, as a function of the biomass of the predator. In addition the plot includes an indication (upper red dotted line) of the maximum possible consumption rate, derived from the satiation parameter of the predator species. An example is illustrated below for all model components and over the entire sampling period. It shows that marine mammals never reach their satiation. On the other hand, demersal fish are often closed to satiation, suggesting that they are not food limited.

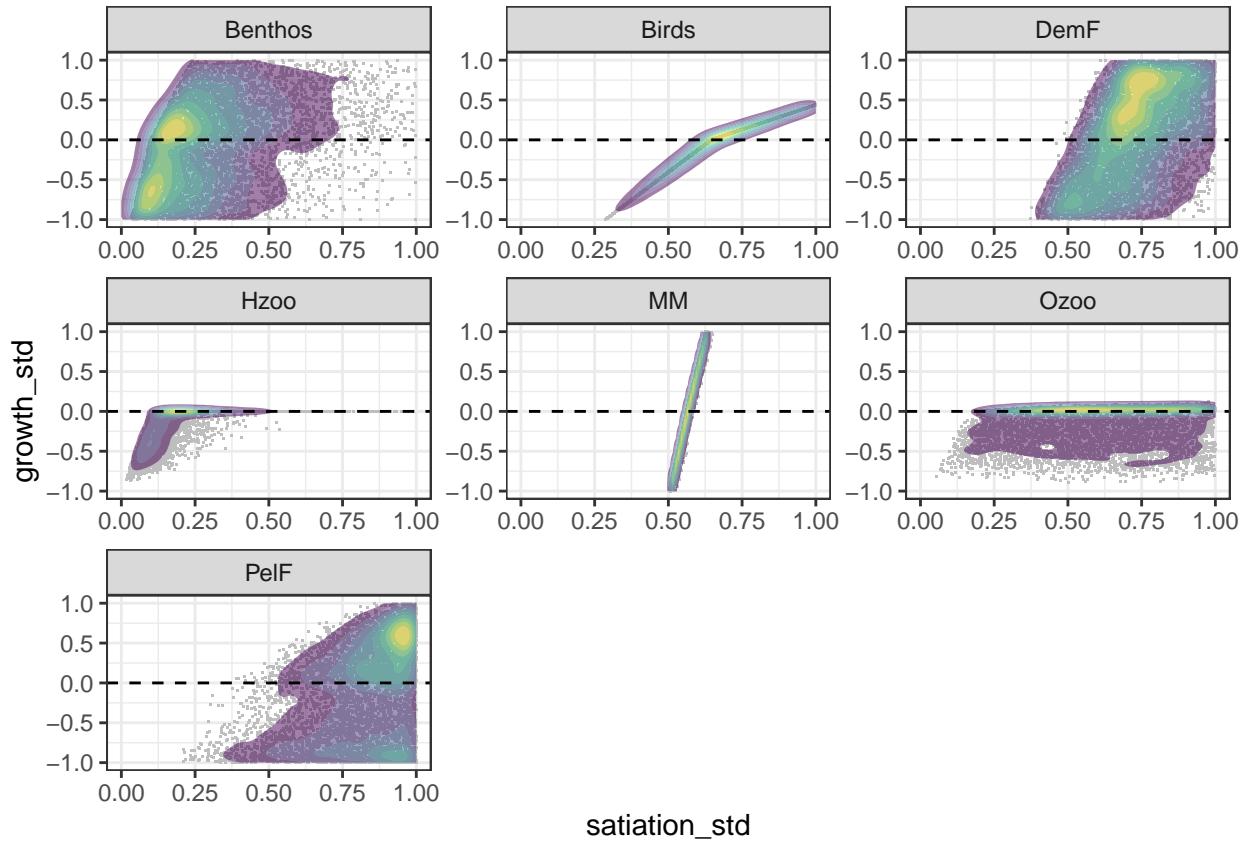
```
ggSatiation(SAMPLE, POLYTOPE$species)
```



6.6 Feeding and Growth

The RCaNmodel function `ggSatiatInertia` is used to explore the relationships between population growth and feeding. The plot shows standardized total consumption(0=no feeding, 1=feeding to satiation) as a function of standardized population growth/mortality (-1 = maximum mortality, 0 = no change in biomass, 1 = maximum growth). A positive relationship between growth and satiation is indicative of bottom up control.

```
ggSatiatInertia(SAMPLE, POLYTOPE$species)
```

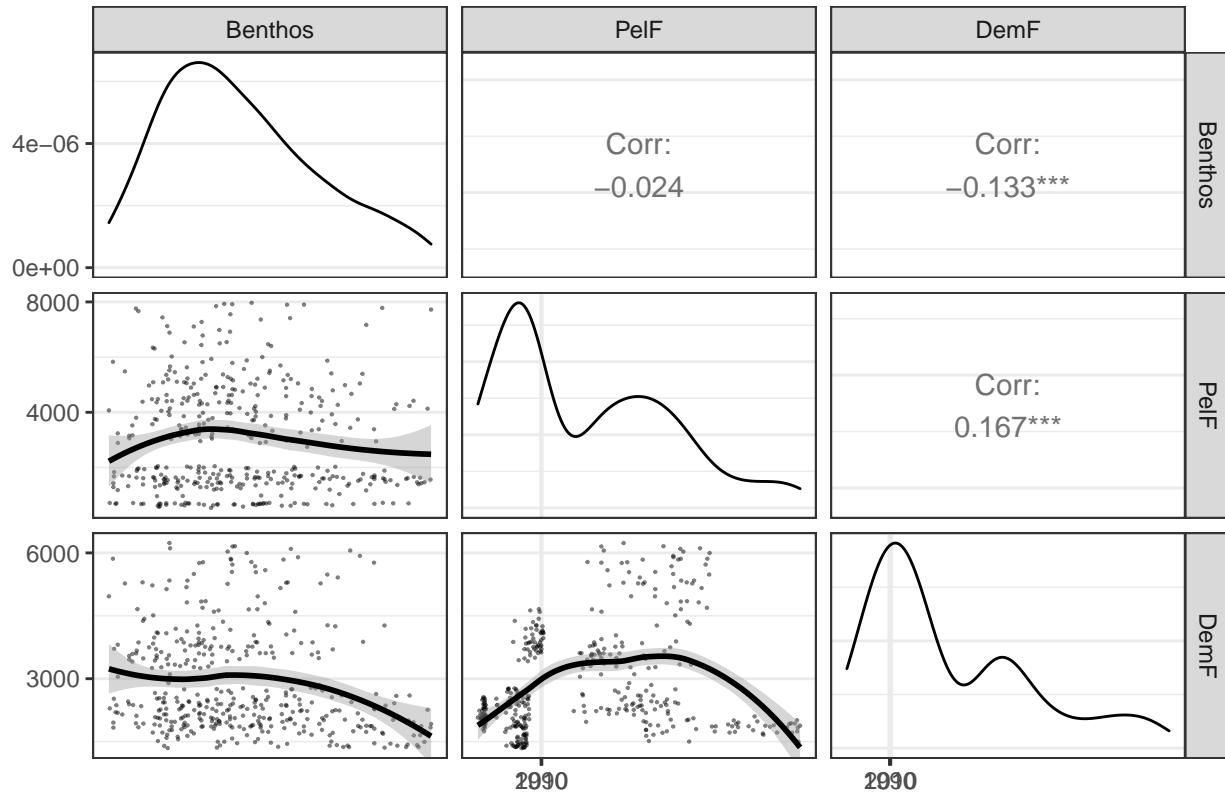


6.7 Pair-plots

The function `ggPairsBiomass` is used to explore the relationships between the biomass of different food-web components/species. The plot shows the individual component density function in the diagonal, the scatterplots of one species against the other in the lower triangle and the Kendall correlation coefficient in the upper triangle. An example is illustrated below for three model components and over the entire sampling period. The function parameter `frac` indicates the fraction of the samples which will be plotted. This is useful for large sample size which can lead to overloaded figures.

```
ggPairsBiomass(SAMPLE,c("Benthos","PelF","DemF"),logscale=FALSE, frac = .25) + #we only 25% of the points
  scale_y_continuous(n.breaks=3)+
  scale_x_continuous(breaks=c(1990,2010))
```

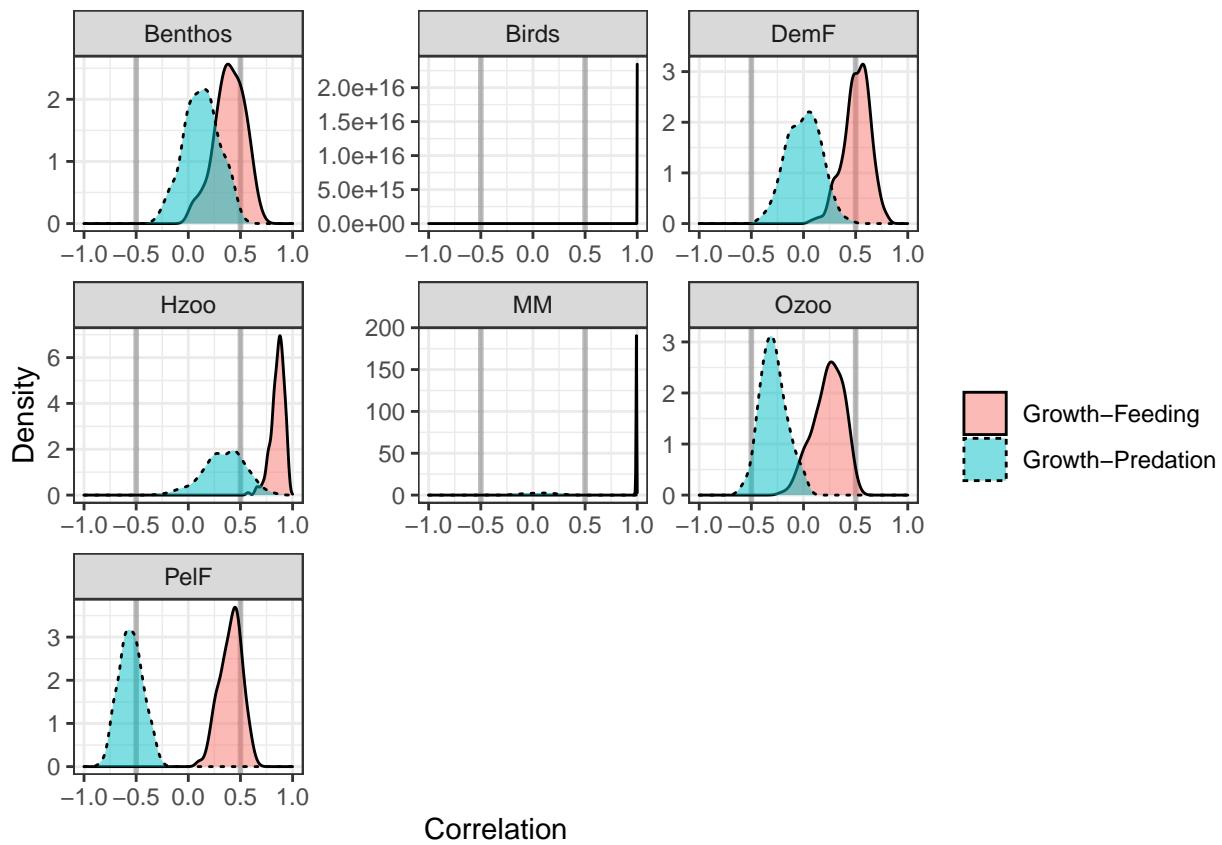
Species pair-plot



6.8 Top-Down and Bottom-Up controls

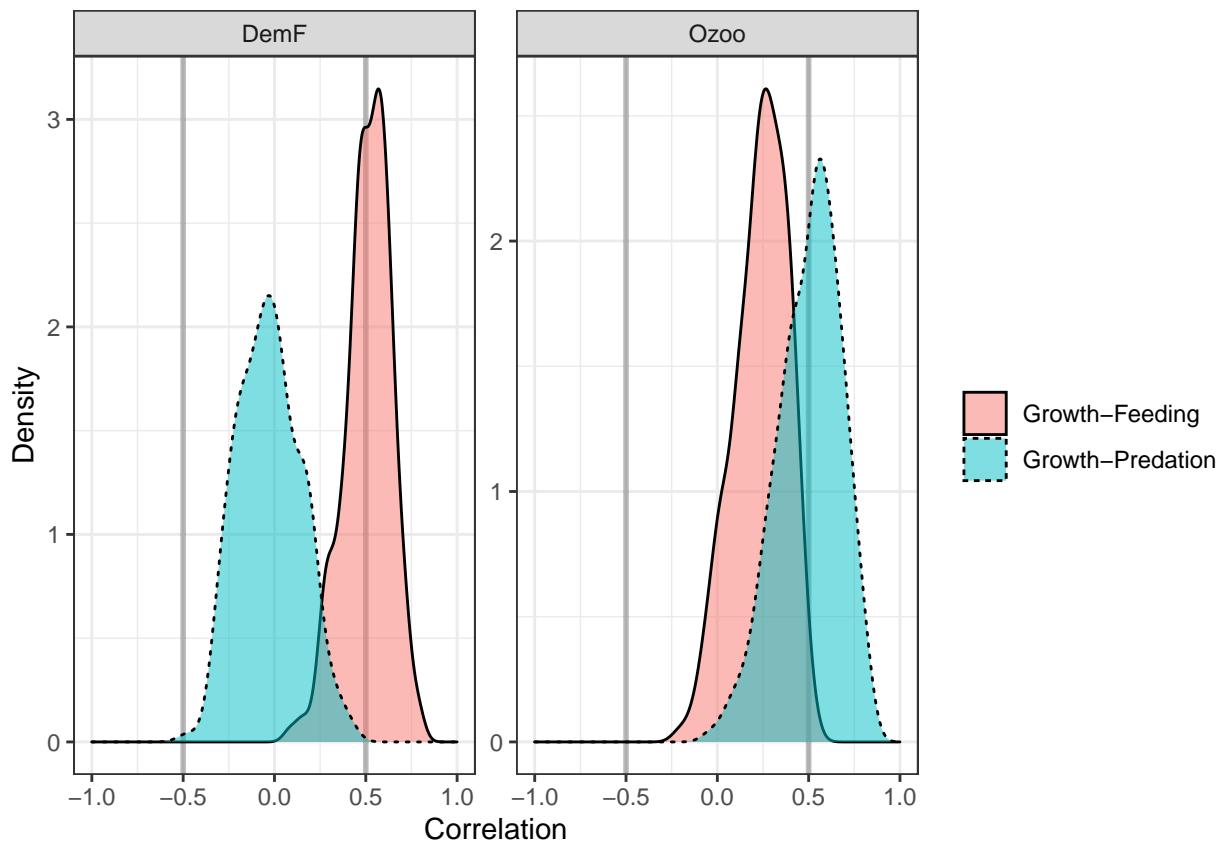
One important aspect of food-web dynamics is the presence of possible trophic controls (Bottom-up or Top-Down). We define here bottom-up as a situation in which the growth of a species/component is positively correlated to the consumption by this species. Similarly, Top-down control is defined as a situation in which the growth of a species/component is negatively correlated to the predation/fishing on this species. The RCaNmodel function `ggTopDownBottomUp` plots the density function of the correlation coefficients between growth and consumption/predation:

```
ggTopDownBottomUp(SAMPLE)
```



This function display the correlation between the growth of the species with both (1) its total ingestion of prey per unit of biomass (in red) and (2) the how much it is predated per unit of biomass (in green). A red density plot tending to 1 indicates that growth is correlated to feeding, and therefore a bottom up control. Conversely, a green density plot tending towards minus one indicates that growth is negatively correlated to the predation and suggests a top down control. It is possible to select which fluxes are accounted for in this diagram (for example to exclude non-trophic fluxes):

```
# we restrict the analysis to Ozoo and DemF and limit to the predation by
#respectively DemF/PelF and MM
ggTopDownBottomUp(SAMPLE,
  species=list(Ozoo=c("DemF", "PelF"),
  DemF=c("MM")))
```



- Christensen, Villy, and Carl J Walters. 2004. "Ecopath with Ecosim: Methods, Capabilities and Limitations." *Ecological Modelling*, Placing Fisheries in their Ecosystem Context, 172 (2): 109–39. <https://doi.org/10.1016/j.ecolmodel.2003.09.003>.
- ICES. 2020. "Working Group on the Integrated Assessments of the Barents Sea (WGIBAR)." Journal Article. *ICES Scientific Reports* 2 (30): 212pp. <https://doi.org/10.17895/ices.pub.5998>.
- Lindstrøm, U., B. Planque, and Sam Subbey. 2017. "Multiple Patterns of Food Web Dynamics Revealed by a Minimal Non Deterministic Model." Journal Article. *Ecosystems* 20: 163–82. <https://doi.org/10.1007/s10021-016-0022-y>.
- Planque, B., and C. Mullon. 2020. "Modelling Chance and Necessity in Natural Systems." Journal Article. *ICES Journal of Marine Science* 77 (4): 1573–88. <https://doi.org/10.1093/icesjms/fsz173>.