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# Insects as a Source of Sustainable Proteins

## Seema Patel

Bioinformatics and Medical Informatics Research Center, San Diego State University, San Diego, CA, United States

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### 2.1 INTRODUCTION

The importance of dietary proteins cannot be underestimated (Pihlanto and Korhonen, 2003). Proteins form the enzymes, transcription factors, antibodies, neurotransmitters, hormones, and a whole lot of other vital body components. Protein deficiency can lead to a gamut of illnesses (Bhutta and Sadiq, 2012), and undernutrition is killing millions of people annually. So far, most of the starvation and famine conditions are confined to developing countries. But, as climate change is causing erratic weather patterns, and the human population is poised to exceed 9 billion by 2050, global food insecurity looms on the horizon. At this rate of human population growth, conventional protein production from livestock, poultry, and fish cannot keep up (van Huis, 2013), so novel and affordable sources of proteins are being searched for. As sustainable meat substitutes, options like soybean, algae, insects, mycoproteins, among others, have been explored (Smetana et al., 2015). Each of these alternatives has their pros and cons. Particularly, insects, generally considered pests for agriculture and vectors of zoonotic diseases, have emerged as a prospective solution (Ramaswamy, 2015;

Nadeau et al., 2015; van Huis et al., 2015). In the past half-decade, insects have drawn immense interest from diversified hierarchies.

Humankind consuming certain arthropods is not new. Especially crustaceans, which encompass crabs, lobsters, prawns, and shrimps, are considered as delicacies (Hadley, 2006). Krill (Euphausia superba), a marine crustacean, is being harvested for its oil, rich in omega-3 fatty acids, choline, and antioxidant astaxanthin (Maki et al., 2009; Barros et al., 2014). Aquaculture along with fishery, is a billion dollar sector. Bee (Apis sp.) products as honey, pollen, and propolis are dietary supplements (Al-Hariri, 2011; Rossano et al., 2012; Silva-Carvalho et al., 2014; Patel, 2016a). The cochineal insects (Dactylopius coccus)-derived dye carmine (the pigment carminic acid) is used in food processing (Voltolini et al., 2014). Apiculture (Verde, 2014) and cochineal insect farming (De León-Rodríguez et al., 2006; Ramos-Elorduy et al., 2011) are popular traditional practices. Also, crickets and mealworms are reared to feed zoo animals and birds (McClements et al., 2003). Silkworm (Bombyx mori, Antheraea assamensis, etc.) culture or sericulture, to harvest silk, is an age-old practice, and is a major part of some economies (Takeda, 2009; Tikader et al., 2013). Cantharidin, a topical vesicant from blister beetles (Cantharis vesicatoria), has been traditionally used to treat warts, calluses, and other skin conditions. Also, it is currently being used as a sexual stimulant (Torbeck et al., 2014). The horseshoe crab (Limulus polyphemus) hemolymph is harvested for multiple biomedical applications (Hurton et al., 2005).

In certain ethnic cultures and some Oriental countries, insects have been integral part of diet. But overall global consumption is minimal and often considered a taboo, insects being regarded as filthy and disease carriers (Sidali et al., 2018). But with changing times and the escalating need for additional protein sources, insects are being viewed in a new light. Their merits are outnumbering the demerits. Insects are rich in protein (40-75 g/100 g dry weight), and other micronutrients (Verkerk et al., 2007). Their "feed conversion ratio" is high, and turnover time is low (Premalatha et al., 2011; Nowak et al., 2016). Insect husbandry releases less greenhouse gases, while requiring less water and agricultural land (Van Huis and Dunkel, 2016). Black soldier fly (Hermetia illucens) can grow on organic materials, which is an added advantage as wastes are accumulating and their disposal is an additional problem (Wang and Shelomi, 2017). Insect farming is energy-efficient compared to livestock farming or aquaculture (Grau et al., 2017). Edible insects can be reared on organic byproducts (Lang and Barling, 2013). So, insect farming and entomophagy practices by humans are being promoted (Gahukar, 2011; Nadeau et al., 2015). Several countries are allocating funds and resources for insect-based protein food developments. While insects as the sources of protein and their relevance as human food are indisputable, some strong hiccups lie in the path of their popularization. The most important hurdles are "repulsion towards arthropods," which is psychological (Caparros Megido et al., 2016), and "allergenicity," which is of clinical significance. This chapter delves into the present status of insect-based protein foods, enumerates the roadblocks in the path of insects emerging as "sustainable protein source," and discusses the prospects.

### 2.2 ETHNIC AND MODERN ENTOMOPHAGY PRACTICES

The class Insecta includes the orders Coleoptera, Orthoptera, Hemiptera, Hymenoptera, Lepidoptera, Isoptera, Ephemeroptera, Odonata, and Mantodea (Chakravorty et al., 2011, 2013).

Humans have consumed insects since the early days of evolution and before as their other primate ancestors. During the hunter-gatherer stage of mankind, insects were a major source of nutrition (Morris, 2008). Insects have been part of subsistence diets for millennia (Pal and Roy, 2014). They have served as an emergency food, as a staple, and as delicacies (Johnson, 2010). Until now, entomophagy, the practice of consuming insects, has been followed across the world (Raubenheimer and Rothman, 2013). Several ethnic food habits encompass insects (Bodenheimer, 1951; Costa-Neto and Dunkel, 2016). As per one report, about 1700 insects species are consumed globally (Chakravarthy et al., 2016), while as per another report, 2086 species are consumed by 3071 ethnic groups (Ramos-Elorduy, 2009). As per another report, 527 different insects are consumed across 36 countries in Africa, 29 insects in Asia, and 23 in the America. The eggs, larvae, pupae, and adults of several insects are consumed by frying, stewing, drying, smoking, steaming, blanching, and roasting (Chen et al., 2009). Snacks, beverage, and condiments are popular modes of insect consumption.

Fried honey bees (Apis sp.) are deemed a delicacy in parts of China (Hartmann et al., 2015). The black chafer beetle (Holotrichia parallela Motschulsky) and black ant (Polyrhachis vicina Roger) are traditionally consumed in China. In Cambodia, roasted crickets are a popular delicacy (Walia et al., 2018). The Japanese consume hachinoko (boiled wasp larvae), sangi (fried silk moth pupae), zazamushi (aquatic insect larvae), semi (fried cicada), and inago (fried grasshopper) (Nonaka, 2010; Césard et al., 2015). Dried Tenebrio molitor, Oxya chinensis sinuosa, B. mori, Protaetia brevitarsis seulensis, and Verlarifictorus asperses are consumed in Korea (Kim et al., 2017). About 164 insect species are consumed in Thailand (Boulidam, 2010; Hanboonsong, 2014). Rural Filipinos consume migratory locust, field crickets, mole crickets, carpenter ant eggs, coconut beetles grubs, June beetles, and katydids (Adalla and Cervancia, 2010). In Laos, weaver ant eggs, bamboo worms, crickets, and wasps are consumed (Barennes et al., 2015). In Borneo, more than 80 species of insects, including honey bee brood, grasshoppers and sago grubs, crickets, rice bugs, cicadas, termites, ants, and beetles, are consumed by the Kadazandusun, Murut, and Rungus people (Chung, 2008). The Vedda tribal people of Sri Lanka consume bee brood and larvae of Apis dorsata, Apis cerana, and Apis florea (Nandasena et al., 2010). Tribal people inhabiting the North Eastern part of India, as in Assam and Manipur, consume several insects to supplement their diet (Nath et al., 2005; Shantibala et al., 2014). In Manipur, they ingest Lethocerus indicus, Laccotrephes maculatus, Hydrophilus olivaceous, Cybister tripunctatus, and Crocothemis servilia (Shantibala et al., 2014). Green weaver ant (Oecophylla smaragdina) is consumed as a condiment. The edibility of 18 species of insects in the Kolhapur region of India has been reported (Sathe, 2015).

Emperor moth (*Gonimbrasia belina*) caterpillar (mopane worm) is consumed in Southern Africa (Okezie et al., 2010). African palm weevil (*Rhychophorus phoenicis*) larvae are consumed by some tribes (Elemo et al., 2011). The Azande and Mangbetu people of Congo (van Huis, 2017) and inhabitants of Limpopo Province, South Africa (Netshifhefhe et al., 2018) consume termites. In Nigeria, the termite (*Macrotermes natalensis*), African cricket (*Brachytrupes membranaceus*), and pallid emperor moth (*Cirina forda*) are popular insect foods (Agbidye et al., 2009). The Pangwe people in Guinea, Gabon, and Cameroon gather aquatic larvae of dragonflies. People living around Lake Victoria consume black ants (*Carebara vidua* Smith) (Ayieko et al., 2012).

Native American and Latin American tribes consume a large variety of insects (Navarro et al., 2010). Tukanoan Indians in the Northwest Amazon consumed over 20 species of insects which included beetle larvae (*Rhynchophorus*), ants (*Atta*), termites (*Syntermes*), and caterpillars (from the families Noctuidae and Saturniidae) (Dufour and Dufour, 1987). In Mexico, roasted ants, lime and chile cricket, grasshoppers (chapulines), mescal worms (gusanos de maguey), and insect eggs (escamoles) are traditional snacks. The leafcutter ant (*Atta laevigata*, *Atta cephalotes*, and *Atta sexdens*) is eaten in parts of Colombia and Brazil. Brazil, Colombia, Ecuador, Mexico, Peru and Venezuela are countries where insect consumption is prevalent.

Regional differences in insect-eating practices have been seen in the United States (Schrader et al., 2016).

Australian Aborigines have been consuming Bogong moth (*Agrotis infusa*), larvae of cossid moth (*Xyleutes leucomochla*), honeypot ant (*Melophorus bagoti* Lubbock), and carpenter ant (*Camponotus* spp.) (Yen, 2010; Warrant et al., 2016). Tribes in the New Guinea island, collect oviposits of the beetle *Rhynchophorus ferrugineus papuanus* from the sago palm (*Ponzetta and Paoletti*, 1997).

In the Carnia region of Italy, *Zygaena* moths are eaten as a seasonal delicacy. In the putrid cheese Casu Marzu, a traditional Sardinian sheep milk cheese, live larvae of cheese fly (*Piophila casei*) are present (Manca et al., 2015).

A review on termites records 43 species being used in the human diet (Figueirêdo et al., 2015). People consume locally-available insects to supplement their diet for hunger and taste, but insects are rich in nutrients as well. Insects are dense sources of carbohydrates, proteins, fats, minerals, and vitamins (Gahukar, 2011). They are particularly abundant in protein, containing 40–75 g/100 g dry weight (Verkerk et al., 2007). Also, insect meat has more polyunsaturated fatty acid than conventional meat (Van Huis, 2016). The minerals obtained from eating insects include K, Ca, P, Mg, Fe, Mn, and Zn. Organic compounds include 9-octadecenoic acid, ethyl oleate, cholesterol and n-hexadecanoic acid. Some other fatty compounds include hexadecanoic acid, ethyl ester, linoleic acid, ethyl oleate, oleic acid, and cholesta-3, 5-diene (Shen et al., 2006). A study found that the lipids from three Orthopterans, Acheta domesticus, Conocephalus discolor, and Chorthippus parallelus, contain much higher amounts of essential fatty acids than those of T. molitor larvae. A. domesticus and C. discolor contain linoleic acid in major quantities, while C. parallelus contains  $\alpha$ -linolenic acid in major quantities (Paul et al., 2017). Fig. 2.1 shows some of the edible insect preparations. Not only interspecies biochemical compositions differ, but differences exist in the various stages of a species' life cycle, as the egg, larvae, pupae, and adult forms contain very different biochemical profiles.

# 2.3 ISSUES WITH INSECTS-BASED PROTEIN AND THE SOLUTIONS FOR THEM

Food neophobia prevents people from ingesting new things (Demattè et al., 2014). Consuming a new plant-based food is easier than that of a fauna-based food. When it comes to insects, disgust is a major factor (Hamerman, 2016; Menozzi et al., 2017), and neophobia is high (Caparros Megido et al., 2016). People in Western countries have had



FIGURE 2.1 Edible insect preparations.

access to other source of proteins, so they have not had to ingest insects. As a result, they are not conditioned to consume insects with ease, and consider insect eating as culturally inappropriate (Tan et al., 2016). Experts predict that controlled exposure to insect-based foods can help overcome the repulsion. Chopping insects into ready-to-eat preparations obtained a better response than the whole forms (Caparros Megido et al., 2016). Whole forms evoke rejection because of the characteristic cuticle, antennae, wings, strong odor, crawling, filthy habitats, etc. Insects use their pheromones to communicate and for defense, so emit unpleasant smell (Suwannapong and Benbow, 2011). Ground insect powder was used as an additive in cookies, crackers, and protein bars (Smetana et al., 2016). Based on a study involving response towards cricket flour-based chips, it was derived that the exposure to processed insect products can increase consumer willingness to taste, compared to unprocessed insects (Hartmann and Siegrist, 2016). In a study conducted in Australia, incorporating insects into familiar products like biscuits or cookies gained better appeal (Wilkinson et al., 2018). In a survey among the Belgian population, the participants mostly showed neophobia, but also partial willingness for insect preparations with flavor and crisp texture (Caparros Megido et al., 2014). People who do not consume pork or beef can consume their derivative gelatin. Similarly, while blended in a known food, insect extract can obtain better acceptance. Japanese relish sushi and sashimi and the Hawaiians consume ahi-poke where fish is largely raw. Western consumers are showing interest towards them, though infections of parasitic helminth (roundworm, tapeworm, and fluke) from the raw fish consumption have been reported (Kaneko and Medina, 2009). Sea urchin roe is considered a gourmet food, though it has been associated with IgE-mediated allergy (Rodriguez et al., 2007). Palm civet coffee or kopi luwak is an expensive coffee, even though the bean is collected from the droppings of civet cat (Onishi, 2010). Such examples of seemingly gross yet popular food abound. Approval of insects as food is an acquired taste, so transitioning into insectivore requires time and exposure (Hartmann and Siegrist, 2016). Furthermore, food habits are culturally-ingrained,

reflected by halal (Islam), kosher (Jewish), and vegetarian (Hindus and Jainism). So, acceptance of insects is sure to be sluggish. So far, insect farming and marketing as food, is highly unregulated. Legislatives supporting their safety and nutrition can make them popular (Van Huis and Dunkel, 2016).

Anaphylaxis is a life-threatening allergic condition, which can be triggered by insects (Kemp and Lockey, 2002). Case studies of anaphylaxis after the consumption of chapuline have been documented (Sokol et al., 2017). Hymenoptera (bees, wasps, hornets, yellowjackets, and ants) sting-caused allergy, has been observed (Przybilla and Ruëff, 2010). House dust mites (Dermatophagoides pteronyssinus), cockroaches (Blatella germanica, Periplaneta americana), and moths have allergens that can provoke IgE-mediated hypersensitivity in atopic individuals (Arlian, 2002; Kim and Hong, 2007; Okezie et al., 2010). These allergens can be serine proteases (trypsin, chymotrypsin, collagenase) (Wan et al., 2001; Sudha et al., 2008; Dumez et al., 2014), aspartic proteases, chitinases, calycin, troponin, tropomyosin, arylophorin, glutathione-S-transferases, and chitin (Arlian, 2002; Hindley et al., 2006; Jeong et al., 2006; Kim and Hong, 2007; Reese et al., 1999), among others. Mosquitoes have salivary gland proteins such as D7 protein family, adenosine deaminase, serpin, and apyrase (Doucoure et al., 2013). The saliva and secretions of flea, grain weevil, black fly, bluebottle fly, horse fly, bedbugs, cockroach, head louse, butterfly caterpillars, or silkworm have immunogenic components as well (Buczylko et al., 2015). The medical potential of antimicrobial peptides (AMP) from insects have been explored (Patel and Akhtar, 2017). The  $\alpha$ -helical AMPs melittin, cecropin, abaecin, moricin, formaecin, and ponericin have been studied regarding their pathological and antibacterial roles (Chen and Lin-Shiau, 1985; Sousa et al., 2013). The peptides may have potential to inhibit microbes, but also have human cytotoxicity as well (Tonk and Vilcinskas, 2017). Also, insect protease inhibitors can inhibit human chymotrypsin, elastase, and plasmin (Wan et al., 2013; Negulescu et al., 2015). Some of these insects are not edible, but they share the same protein repertoire, which indicates the immunogenicity of the edible insects. Insects have storage proteins which are important for their metamorphosis and egg production (Telang et al., 2002). Storage proteins are synthesized in fat bodies and then secreted into the hemolymph. These vital proteins are hexameric glycoproteins which include hemocyanin, arylphorin, etc. Arylphorin is a major cockroach allergen (Arruda et al., 2001; Kim et al., 2003) and it possesses mitogenic properties as well (Hakim et al., 2007). Hexamerin1B, another insect storage protein in Gryllus bimaculatus (field cricket), can induce allergy (Srinroch et al., 2015; Pener, 2016). Not only insects, but plants also contain storage proteins which have exerted allergenicity towards humans. Peanut vicilin allergen Ara h 1, cashew vicilin-like Ana o 1 (Wang et al., 2002), conglutin beta in Lupinus sp. (Goggin et al., 2008), 7S and 11S globulins of soybean are all some of the allergenic storage proteins in plant-based foods (Shewry and Jones, 2006). However, it seems plant-origin storage proteins are easier to inactivate by cooking, compared to those of insects.

Lipocalin family of  $\beta$ -barrel proteins are transport proteins with allergenicity (Chudzinski-Tavassi et al., 2010). Insect lipocalins induce IgE production in atopic individuals. Nitrophorins are salivary heme proteins, from a lipocalin homology family, that are present in blood-sucking insects, which transfer nitric oxide (NO) to the victim, causing vasodilatation, sequestering histamine, and inhibiting blood coagulation (Ascenzi et al., 2002).

Insect exoskeleton chitin is an immunotoxin and a neurotoxin which, if regularly exposed to, can cause profound illnesses (Patel and Goyal, 2017; Patel et al., 2017). Even the inhalation of chitin particles can provoke the immune, neural, and endocrine systems. Chitin is the substrate of host chitinase, which is crucial for inhibiting infectious agents, but can damage host tissues as well. Enzymes work in cascade, so it can unleash a range of other aberrant enzyme activity. It would not be wrong to regard chitin as an endocrine disruptor or estrogenic agent. It is a top-grade inflammatory agent, and likely to induce excess cytochrome oxidase production. Aromatase is a cytochrome P450 enzyme that converts androgens (C19) into estrogens (C18) (Patel, 2017a). Estrogen is mitogenic and proliferative in nature and may cause breast cancer, prostate cancer, ovarian cancer, gastric cancer, pituitary cancer, polycystic ovary syndrome, diabetes, endometriosis, osteoporosis, Alzheimer's disease, and schizophrenia, among others (Patel et al., 2018).

Some moths and butterflies, such as Burnet moths (*Zygaena filipendulae*), contain toxic hydrogen cyanide in their tissues (*Zagrobelny and Møller*, 2011), and monarch butterfly (*Danaus plexippus*) larvae contain the cardiac glycoside cardenolide (*Petschenka and Agrawal*, 2015). Cardenolide manipulates the Na(+)/K(+)-ATPases, which can affect blood pressure and electrolyte balance (*Patel*, 2016b).

These allergens and venoms can provoke the human immune system in a myriad of ways. They can disrupt cell membranes, cleave tight junction proteins between the epithelial cells, manipulate the cytoskeleton, and induce cytokine proliferation (Chapman et al., 2007; Navarro-Garcia et al., 2010; Kempkes et al., 2014; Zhang et al., 2014). The immune activation can cause asthma, dermatitis, urticaria, sinusitis, rhinitis, otitis, etc. (Arshad, 2010), or even anaphylaxis (Asokananthan et al., 2002; Macan et al., 2003; Ichikawa et al., 2009; Ahmed et al., 2010). Insect venom causing mastocytosis (Bonadonna et al., 2010) has been documented. Females seem to be particularly hypersensitive to insect allergens due to their higher level of estrogen and better immune defense (Patel et al., 2018). Also, individuals with inflammatory diseases are more likely to show sensitivity toward the allergens as the host proteins are highly glycosylated and they recognize the insect proteins. It is an evolutionary adaptation to protect the stressed body from further threats by escalating immune surveillance. Like methylation of proteins is akin to an off switch, glycosylation, due to the presence of N- and O-linked oligosaccharides, alters protein properties. Reports are emerging that hyperglycosylation prolongs the circulation of coagulation factor IX (Bolt et al., 2012).

In silico analyses have revealed that all living organisms, viruses to humans and in between all, possess some evolutionarily conserved protein domains (Patel, 2017b). These motifs are pivotal in "offense and defense," and so are strictly conserved. Some of such virulent protein domains in insects include chitin-binding domain (ChtBD3) (Patel et al., 2017). The allergens often cause cross-reactivity, so the sensitive individuals should not consume insects (Pier and Lomas, 2017). Incidences of allergic reactions on consumption of silkworm pupae, cicadas, and crickets have been reported in China (Feng et al., 2018). Caterpillars, sago worm, locust, grasshopper, bee, etc., have also been associated with insect allergy (de Gier and Verhoeckx, 2018). The allergen proteins are so stable that even thermal processing and digestion cannot eliminate all of them (de Gier and Verhoeckx, 2018). So, food processing of the insects ought to be rigorous and the products should have warning labels like other allergy-inducing foods. Insect allergy is also a cause of

occupational health problems. Systemic allergic reactions in beekeepers has been documented (Ludman and Boyle, 2015). Post-hire asthma instances among insect-rearing workers have been observed (Suarthana et al., 2012). Those rearing the insects ought to be aware of the risks (Pener, 2016). Both potential consumers and breeders ought to be cognizant of the risks of embracing insects, as the immune sensitization may not manifest immediately, but after a certain titer of IgE antibodies are formed. Once the antibodies are formed, and inflammasomes are activated, it takes years to restore the immune system homeostasis (Patel and Meher, 2016). Even after exposure to the allergens is discontinued, the health hazards continue to occur

Also, insects are vectors of human pathogens such as viruses (Campos et al., 2015), bacteria (Hager et al., 2006), protozoa (Takeo et al., 2009), fungi, and nematodes. Food pathogens such as *Staphylococcus aureus*, *Bacillus cereus*, *Clostridium perfringens*, enterohemorrhagic *Escherichia coli*, *Listeria monocytogenes*, and *Salmonella* spp. have been found associated with crickets (Walia et al., 2018). Insects being protein dense often serve as substrates for fungal growth. Dried insects in Zambia have been detected with aflatoxin-producing fungi. Aflatoxins elaborated by *Aspergillus flavus* are carcinogenic (Barrett, 2005). In the study, above 10 µg/kg aflatoxin was detected in *Gynanisa maja*, *Gonimbrasia zambesina*, and *Macrotermes falciger* (Kachapulula et al., 2018). The insect families Muscidae, Glossinidae, Culicidae, and Phlebotominae are vectors for diarrhea, myiasis, sleeping sickness, filariasis, malaria, leishmaniases, and bartonellosis, among others (Burgess, 2010).

So, a complete risk assessment of the edible insects-based food candidates must be carried out, before recommending them for mass consumption (Belluco et al., 2015; Grabowski and Klein, 2016). In fact, the European Union (EU) has published an edible insect food safety guide (Robinson, 2015). Other publications also have discussed the risk and safety aspects (Marshall et al., 2016).

Fig. 2.2 presents the merits and demerits of insect-based foods.

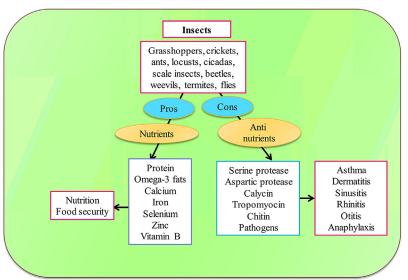


FIGURE 2.2 The merits and demerits of insect-based food.

### 2.4 LOOKING AHEAD AND DISCUSSION

Despite the hurdles in the path, insects as a sustainable food source are an optimistic prospect (Sun-Waterhouse et al., 2016). Food futurists believe that insects as an alternative protein source will claim a major niche in the next few years. The UN FAO is recommending the consumption of insects to mitigate food insecurity (Costa-Neto and Dunkel, 2016; Nowak et al., 2016). The US Department of Agriculture (USDA) is willing to fund edible insect research (LIGMAN, 2015). Entomoculturing and entomophagy is part of the "One World — One Health" (OWOH) movement as well (Yates-Doerr, 2015). United Nations (UN) and NGOs are promoting insect-based food availability in regions with food insecurity. Undernutrition-afflicted countries like Congo can benefit from insect-based foods. In a pilot study, school children were provided with biscuits containing 10% cricket powder (Homann et al., 2017). Another study in Nigeria determined the protein quality of common edible insects such as moth caterpillar, termite, cricket, and grasshopper, finding them to be substitutes for dietary protein (Oibiokpa et al., 2018).

China and Thailand have numerous insect farms, both small and medium enterprises, and industrial scale farms. Thailand is a leader at insect farming, with an estimated 20,000 food insect farms, mostly rearing crickets and palm weevils (Hanboonsong et al., 2013). Cricket farming in Thailand, Cambodia, Lao People's Democratic Republic (Lao PDR), the Democratic Republic of the Congo (DRC), and Kenya was studied, and in most countries their infancy status was reported (Halloran et al., 2018). In Latin America, chapuline farming is common, given its popularity as a food. The European Commission (EC) is also promoting the inclusion of insects in food (Finke et al., 2015). The Netherlands is open to the integration of insects in food (House, 2016) and insect farming practices are rising (De Goede et al., 2013). The Dutch populace are being motivated to include crickets, worms, and caterpillars as nutritious protein sources (Tagliabue, 2011). Canada-based Entomo Farms raises crickets and mealworms for food. The North American Coalition for Insect Agriculture (NACIA), comprises stakeholders who arrange conferences to promote insect agriculture. Many startups in California (United States) are selling insect-based food products. In fact, insect farming has been present in almost every continent, as animals in captivity and pets are fed with them. Now, with the new wave of insect-based food consumption by humans, the safety and hygiene dimensions are being emphasized, and human-grade insect farms are being set up. The edible insect business is now a more than \$20 million industry (Hoffman, 2014). Entrepreneurs across the world are attempting to rear edible insects and to take them mainstream (Costa-Neto and Dunkel, 2016). Insects in the mass rearing facilities are prone to viral diseases, which requires adequate knowledge to avert the problem (Maciel-Vergara and Ros, 2017). To make insects into edible products, they are subjected to sun drying, freeze-drying, grinding, defatting, and acid hydrolysis (Kim et al., 2016). Each processing technique is likely to affect the biochemical composition of the insect biomass differently. In a study of sun-dried edible black ant, out of the 28 organic components, five were lost while four were formed. Fatty acids, aldehyde, and alkanes appeared during the sun drying (Li et al., 2009).

For the economic viability of insects in the food sector, better rearing, harvesting, and processing tools and techniques ought to be developed (Rumpold and Schlüter, 2013). Diet optimization for the insects can enhance their biomass. As genetic engineering has

improved crop yield (Khan et al., 2013), the development of less immunogenic and more meat-containing edible insects by genetic manipulation can be very economical.

Nowadays, interested consumers are purchasing edible insects online. Mixed bag of grasshoppers, crickets, silk worms, and sago worms are marketed as energy snacks like granola bars. Cricket powder is sold and recommended to be mixed with flour for baking purposes. Chocolate-coated roasted crickets or mealworms, and insect lollipops are some of the innovative products being marketed. Even global leaders like Amazon are selling these products. Although at present there are only a handful of insect-based food traders, such as ecoEat, Don Bugito Prehispanic Snackeria, Mercado Mio, Newport Jerky Company, Hotlix, Merci Mercado, Candy Crate, Thailand Unique, EntoVida, and Meat Maniac, soon new establishments are likely to emerge. Renowned chefs are serving insect-laced recipes like cricket-sprinkled salads, mealworm lettuce wraps, cricket fried rice, mescal worm tacos, fried dragonflies, ant egg tostada, mealworm-peppered noodles, silkworm powder-flavored broth, etc. Cookbooks, featuring the culinary uses of insects are being published (Linn, 2014). Experimenting with insects has just begun and possibilities abound. In a study, sausage fortified with mealworm larvae and silkworm pupae was evaluated (Kim et al., 2016). In an insect-eating festival held in the Philippines, various agricultural pest insects were processed into chayote bars. rice cakes, burgers, sandwich fillers, etc., and presented to people (Sabado and Aguanta, 2014). In a festival in Switzerland, pizzas were served topped with mealworms and beetle larvae (Wallace, 2010). Even countries like the Czech Republic are showing interest in including insects in the diet (Bednarova et al., 2010).

Human societies are appearance-conscious. The same insect product that will be normally dismissed as repulsive might attract attention if it is claimed that it prevents obesity, skin wrinkles, hair loss, or restores fertility. Similarly, anticancer, antidiabetic, and immunity building claims can raise the acceptance of insect-based foods. It will not be surprising if traders adopt this path for profits. Silkworm moth (*B. mori* L.) larvae was being evaluated as an animal protein source for astronauts on space missions (Tong et al., 2011). Apart from silkworm, the hawkmoth, the drugstore beetle, and the termite are considered candidates for space agriculture (Katayama et al., 2008).

It appears that compared to the whole insects or their crushed forms, there is lower aversion and better prospects for acceptance for bioactives extracted from them. Insects elaborate proteins and AMP for immune defense (Slocinska et al., 2008; Ezzati-Tabrizi et al., 2013), which might be exploited as dietary supplements. Lipid extracted from meadow grasshopper (C. parallelus) was found to have an interesting fatty acid composition (Paul et al. 2015). However, the immunogenicity or immunosuppressive aspect of the extracted components must be addressed as not all proteins are benign and healthpromoting, but can be perceived as antigens. Astaxanthin from krill is known to be an antioxidant. However, recent studies have revealed that astaxanthin by virtue of its oxidative stressor elimination might resolve immunopathology, but can exert an immune depressive effect (Dhinaut et al., 2017). Other studies have shown how astaxanthin feeding suppresses the expression of inflammatory cytokines, including nuclear factor (NF)-κB, tumor necrosis factor (TNF)- $\alpha$ , and interleukin (IL)-1 $\beta$ . While insects no doubt are proteinrich, the ingestion of inadequately prepared insects can cause immune activation and inflammatory diseases. The FDA and other regulatory bodies ought to keep a tab on unscrupulous merchandise and marketing.

Every new application of an object requires imparting education on the merits and demerits. In every element, there is a trade-off. If the risks are too high, the element is not worth applying for the new use. Some of the merits and demerits have been discussed earlier. Host factors are different, so the response to a substance can vary. Insects are allergenic as discussed previously. Therefore not all human beings are expected to fare well after insect consumption. The way a lot of people cannot consume seafood, shrimps, and crabs for hypersensitivity reasons (Lopata et al., 2010), some will not be fit to consume insects. However, different processing techniques might neutralize the allergens and make them amenable for mass consumption. In fact, if the allergenicity issue can be taken care of, insects can be a superfood, a way better substitute to canned, frozen, food additivelaced packaged foods. Ethnic people have been consuming them for a long time, and are thriving. To urban people in developed countries, insects are a novelty. Those who can savor shrimps or clams should not have a psychological issue with the consumption of insects. Apart from a sustainable protein source, insect farming can lower the burden on the environment, and can provide livelihood opportunities to many (Halloran et al., 2017). By choice, few people would have consumed insects, but protein requirements can lead to their consumption. Food insecurity looms, but the situation is still under control. Maybe in the next few decades, the availability of nutritious foods will be challenging, and insects will have to be coerced onto the human dietary platter. So, this time period ought to be directed towards the safety evaluation of insects, developing facilities, and processing the edible insects into human consumption-worthy food objects. In fact, future meat options are likely to include cultured meat and imitation meat, apart from insect meat (Alexander et al., 2017). Table 2.1 presents a list of insects that are or have been consumed in different regions of the world. This is a fraction of the actual list, as the exact picture is elusive and exhaustive.

TABLE 2.1 Some Common Edible Insects and Their Common Names

No.	Scientific Name	Common Name
1.	Acheta domesticus	House cricket
2.	Agrotis infusa	Bogong moth
3.	Anabrus simplex	Mormon cricket
4.	Apis spp.	Honey bees
5.	Arsenura armada	Giant silk moth
6.	Atractomorpha psittacina	Slant-faced grasshopper
7.	Bombyx mori	Silkworm moth
8.	Brachytrupes membranaceus	Tobacco cricket
9.	Brontispa longissimi	Coconut leaf beetles
10.	Camponotus spp.	Jet-black carpenter ants
11.	Carebara vidua Smith	Black ant
12.	Cirina forda	Pallid emperor moth

(Continued)

TABLE 2.1 (Continued)

No.	Scientific Name	Common Name
13.	Chorthippus parallelus	Meadow grasshopper
14.	Coloradia pandora	Pandora moth
15.	Corcyra cephalonica	Rice moth
16.	Crocidolomia pavonana	Cabbage worm
17.	Crocothemis servilia	Scarlet skimmer
18.	Cybister tripunctatus	Three-punctured diving beetle
19.	Ephydra hians	Alkali fly
20.	Gonimbrasia zambesina	Bulls eye silk moth
	Gonimbrasia belina	Emperor moth
21.	Gryllotalpa africana	Mole crickets
22.	Gynanisa maja	Speckled emperor
23.	Hermetia illucens	Black soldier fly
24.	Holotrichia parallela Motschulsky	Black chafer beetle
25.	Hydrophilus olivaceous	Water scavenger beetle
26.	Hypopta agavis	Tequila worm
27.	Laniifera cyclades	Nopal worm
28.	Latebraria amphipyroides	-
29.	Leptocorisa oratorius	Rice bug
30.	Lethocerus indicus	Giant water bugs
31.	Liometopum apiculatum	_
32.	Locusta migratoria	Locusts
33.	Macrotermes falciger	Termites
	Macrotermes natalensis	
	Macrotermes michaelseni	
34.	Melophorus bagoti	Honeypot ant
35.	Odontotermes sp.	Termites
36.	Oecophylla smaragdina	Red weaver ant
37.	Omphisa fuscidentalis	Bamboo borers
38.	Oxya chinensis	Chinese grasshopper
39.	Patanga succincta	Bombay locusts
40.	Piophila casei	Cheese fly

(Continued)

TABLE 2.1 (Continued)

No.	Scientific Name	Common Name
41.	Polyrhachis vicina Roger	Black ant
42.	Protaetia brevitarsis seulensis	White-spotted flower chafer
43.	Rhynchophorus ferrugineus	Asiatic palm weevil
	Rhynchophorus phoenicis	African palm weevil
44.	Schistocerca gregaria	Desert locust
45.	Scotinophara coarctata	Malaysian black bug
46.	Sphenarium sp.	Grasshoppers
47.	Tribolium castaneum	Red flour beetle
48.	Verlarifictorus aspersus	_
49.	Xyleutes leucomochla	Cossid moth
50.	Zygaena sp.	Burnet moth

### **CONCLUSION**

Food trends keep evolving, and if certain impediments in the path of mass adoption of insect diet can be addressed, insects can become a substantial part of the human diet. Going by the significant spike in the number of publications on the potential of insects as human food, the rising interest and fund allocations, can be gauged. Research projects and partnerships are being forged and entrepreneurs are establishing insect farms and facilities. The interest in insects as human food has been stirred in academia, the food sector, as well as among the general public (Evans et al., 2015). It can be anticipated that insects can be a regular part of the food production chain in the next few years. Surveys, food fests, and symposia are being held to create consumer awareness and to familiarize people to the emerging novel edible protein source (Liu and Zhao, 2018). Chefs, insect farmers, and nutrition experts are collaborating to formulate novel insect-based food preparations. Despite the hurdles and genuine concerns, opinions are cohesive that insects have the potential to be used as a functional food source. If the above-voiced concerns can be resolved, and nutritional analyses optimized, insects can be a sustainable protein food source, and can generate an alternative food industry with less pressure on environmental resources.

#### References

Adalla, C.B., Cervancia, C.R., 2010. Philippine edible insects: a new opportunity to bridge the protein gap of resource-poor families and to manage pests. RAP Publication, 2010/02, pp. 151–160.

Agbidye, F.S., Ofuya, T.I., Akindele, S.O., 2009. Some edible insect species consumed by the people of Benue State, Nigeria. Pakistan J. Nutr. 8, 946–950. Available from: https://doi.org/10.3923/pjn.2009.946.950.

- Ahmed, A., Minhas, K., Namood-E-Sahar, et al., 2010. In silico identification of potential American cockroach (*Periplaneta americana*) Allergens. Iran J. Public Health 39, 109–115.
- Al-Hariri, M.T., 2011. Propolis and its direct and indirect hypoglycemic effect. J. Family Community Med. 18, 152–154. Available from: https://doi.org/10.4103/2230-8229.90015.
- Alexander, P., Brown, C., Arneth, A., et al., 2017. Could consumption of insects, cultured meat or imitation meat reduce global agricultural land use? Glob. Food Security 15, 22–32.
- Arlian, L.G., 2002. Arthropod allergens and human health. Annu. Rev. Entomol. 47, 395–433. Available from: https://doi.org/10.1146/annurev.ento.47.091201.145224.
- Arruda, L.K., Ferriani, V.P., Vailes, L.D., et al., 2001. Cockroach allergens: environmental distribution and relationship to disease. Curr. Allergy Asthma Rep. 1, 466–473.
- Arshad, S.H., 2010. Does exposure to indoor allergens contribute to the development of asthma and allergy? Curr. Allergy Asthma Rep. 10, 49–55. Available from: https://doi.org/10.1007/s11882-009-0082-6.
- Ascenzi, P., Nardini, M., Bolognesi, M., Montfort, W.R., 2002. Nitrophorins: lipocalin-based heme proteins transporting nitric oxide. Biochem. Mol. Biol. Educ. 30, 68–71.
- Asokananthan, N., Graham, P.T., Stewart, D.J., et al., 2002. House dust mite allergens induce proinflammatory cytokines from respiratory epithelial cells: the cysteine protease allergen, Der p 1, activates protease-activated receptor (PAR)-2 and inactivates PAR-1. J. Immunol. 169, 4572–4578.
- Ayieko, M.A., Kinyuru, J.N., Ndong'a, M.F., Kenji, G.M., 2012. Nutritional value and consumption of black ants (*Carebara vidua* Smith) from the lake Victoria region in Kenya. Adv. J. Food Sci. Technol. 4, 39–45.
- Barennes, H., Phimmasane, M., Rajaonarivo, C., 2015. Insect consumption to address undernutrition, a national survey on the prevalence of insect consumption among adults and vendors in Laos. PLoS One. Available from: https://doi.org/10.1371/journal.pone.0136458.
- Barrett, J.R., 2005. Liver cancer and aflatoxin: new information from the Kenyan Outbreak. Environ. Health Perspect. 113, A837.
- Barros, M.P., Poppe, S.C., Bondan, E.F., 2014. Neuroprotective properties of the marine carotenoid astaxanthin and omega-3 fatty acids, and perspectives for the natural combination of both in krill oil. Nutrients 6, 1293–1317. Available from: https://doi.org/10.3390/nu6031293.
- Bednarova, M., Borkovcova, M., Zornikova, G., Zeman, L., 2010. Insect as food in Czech Republic. In: MENDELNET 2010, Mendel University in Brno, pp. 674–682.
- Bhutta, Z.A., Sadiq, K., 2012. Protein deficiency. In: Encyclopedia of Human Nutrition, pp 111-115.
- Bodenheimer, F.S., 1951. Australia. Insects as Human Food. Springer, Dordrecht, pp. 70-136.
- Bolt, G., Bjelke, J.R., Hermit, M.B., et al., 2012. Hyperglycosylation prolongs the circulation of coagulation factor IX. J. Thromb. Haemost. 10, 2397–2398.
- Bonadonna, P., Zanotti, R., Müller, U., 2010. Mastocytosis and insect venom allergy. Curr. Opin. Allergy Clin. Immunol. 10, 347–353.
- Boulidam, S., 2010. Edible insects in a Lao market economy. In: Forest Insects as Food: Humans Bite Back. Proceedings of a Workshop on Asia-Pacific Resources and Their Potential for Development, Chiang Mai, Thailand, 19–21 February 2008, pp. 131–140.
- Buczylko, K., Majsiak, E., Wagner, A., 2015. Allergy to non-hymenoptera insects. Alerg ASTMA Immunol. 20, 170–177.
- Burgess, I.F., 2010. Insects and health. Perspect. Public Health 130, 12.
- Campos, G.S., Bandeira, A.C., Sardi, S.I., 2015. Zika Virus Outbreak, Bahia, Brazil. Emerg. Infect. Dis. 21, 1885–1886. Available from: https://doi.org/10.3201/eid2110.150847.
- Caparros Megido, R., Sablon, L., Geuens, M., et al., 2014. Edible insects acceptance by belgian consumers: promising attitude for entomophagy development. J. Sens. Stud. 29, 14–20. Available from: https://doi.org/10.1111/joss.12077.
- Caparros Megido, R., Gierts, C., Blecker, C., et al., 2016. Consumer acceptance of insect-based alternative meat products in Western countries. Food Qual. Prefer. 52, 237–243. Available from: https://doi.org/10.1016/j. foodqual.2016.05.004.
- Césard, N., Komatsu, S., Iwata, A., 2015. Processing insect abundance: trading and fishing of zazamushi in Central Japan (Nagano Prefecture, Honshū Island). J. Ethnobiol. Ethnomed. 11, 78. Available from: https://doi.org/10.1186/s13002-015-0066-7.
- Chakravarthy, A.K., Jayasimha, G.T., Rachana, R.R., Rohini, G., 2016. Insects as human food. In: Economic and Ecological Significance of Arthropods in Diversified Ecosystems: Sustaining Regulatory Mechanisms, Springer, Singapore, pp. 133–146.

- Chakravorty, J., Ghosh, S., Meyer-Rochow, V., 2011. Practices of entomophagy and entomotherapy by members of the Nyishi and Galo tribes, two ethnic groups of the state of Arunachal Pradesh (North-East India). J. Ethnobiol. Ethnomed. 7, 5. Available from: https://doi.org/10.1186/1746-4269-7-5.
- Chakravorty, J., Ghosh, S., Meyer-Rochow, V.B., 2013. Comparative survey of entomophagy and entomotherapeutic practices in six tribes of eastern Arunachal Pradesh (India). J. Ethnobiol. Ethnomed. 9, 50. Available from: https://doi.org/10.1186/1746-4269-9-50.
- Chapman, M.D., Wünschmann, S., Pomés, A., 2007. Proteases as Th2 adjuvants. Curr. Allergy Asthma Rep. 7, 363–367.
- Chen, C.-C., Lin-Shiau, S.-Y., 1985. Mode of inhibitory action of melittin on Na + -K + -ATPase activity of the rat synaptic membrane. Biochem. Pharmacol. 34, 2335–2341. Available from: https://doi.org/10.1016/0006-2952 (85)90791-9.
- Chen, X., Feng, Y., Chen, Z., 2009. Common edible insects and their utilization in China: invited review. Entomol. Res. 39, 299–303.
- Chudzinski-Tavassi, A.M., Carrijo-Carvalho, L.C., Waismam, K., et al., 2010. A lipocalin sequence signature modulates cell survival. FEBS Lett. 584, 2896–2900. Available from: https://doi.org/10.1016/j.febslet.2010.05.008.
- Chung, A.Y.C., 2008. An overview of edible insects and entomophagy in Borneo. In: Forest Insects as Food: Humans Bite. FAO Regional Office for Asia and the Pacific, Bangkok, pp. 1–9.
- Costa-Neto E.M., Dunkel F.V., 2016. Insects as food: history, culture, and modern use around the world. In: Insects as Sustainable Food Ingredients. Academic Press, pp. 29–60 (Chapter 2).
- De Goede D.M., Erens J., Kapsomenou E., Peters M., 2013. Large scale insect rearing and animal welfare. In: The Ethics of Consumption: The Citizen, the Market and the Law. Wageningen Academic Publishers, Wageningen, pp. 236–242.
- De León-Rodríguez, A., González-Hernández, L., Barba de la Rosa, A.P., et al., 2006. Characterization of volatile compounds of mezcal, an ethnic alcoholic beverage obtained from *Agave salmiana*. J. Agric. Food Chem. 54, 1337–1341. Available from: https://doi.org/10.1021/jf052154 + .
- Demattè, M.L., Endrizzi, I., Gasperi, F., 2014. Food neophobia and its relation with olfaction. Front. Psychol. 5, 127. Available from: https://doi.org/10.3389/fpsyg.2014.00127.
- Dhinaut, J., Balourdet, A., Teixeira, M., et al., 2017. A dietary carotenoid reduces immunopathology and enhances longevity through an immune depressive effect in an insect model. Sci. Rep. Available from: https://doi.org/10.1038/s41598-017-12769-7.
- Doucoure, S., Cornelie, S., Patramool, S., et al., 2013. First screening of *Aedes albopictus* immunogenic salivary proteins. Insect. Mol. Biol. 22, 411–423. Available from: https://doi.org/10.1111/imb.12032.
- Dufour, D.L., Dufour, L., 1987. Insects as food: a case study from the northwest Amazon. Am. Anthropol. 89, 383–397. Available from: https://doi.org/10.1525/aa.1987.89.2.02a00070.
- Dumez, M.-E., Herman, J., Campizi, V., et al., 2014. Orchestration of an uncommon maturation cascade of the house dust mite protease allergen quartet. Front. Immunol. 5, 138. Available from: https://doi.org/10.3389/fimmu.2014.00138.
- Elemo, B.O., Elemo, G.N., Makinde, M., Erukainure, O.L., 2011. Chemical evaluation of African palm weevil, *Rhychophorus phoenicis*, larvae as a food source. J. Insect Sci. 11, 1–6. Available from: https://doi.org/10.1673/031.011.14601.
- Evans, J., Alemu, M.H., Flore, R., et al., 2015. "Entomophagy": an evolving terminology in need of review. J. Insects Food Feed 1, 293–305. Available from: https://doi.org/10.3920/JIFF2015.0074.
- Ezzati-Tabrizi, R., Farrokhi, N., Talaei-Hassanloui, R., et al., 2013. Insect inducible antimicrobial peptides and their applications. Curr. Protein Pept. Sci. 14, 698–710. Available from: https://doi.org/10.2174/1389203711209070620.
- Feng, Y., Chen, X.M., Zhao, M., et al., 2018. Edible insects in China: utilization and prospects. Insect Sci. 25, 184–198.
- de Figueirêdo, R.E.C.R., Vasconcellos, A., Policarpo, I.S., Alves, R.R.N., 2015. Edible and medicinal termites: a global overview. J. Ethnobiol. Ethnomed. 11, 1.
- Finke, M.D., Rojo, S., Roos, N., et al., 2015. The European Food Safety Authority scientific opinion on a risk profile related to production and consumption of insects as food and feed. J. Insects Food Feed 1, 245–247. Available from: https://doi.org/10.3920/JIFF2015.x006.
- Gahukar, R.T., 2011. Entomophagy and human food security. Int. J. Trop. Insect Sci. 31, 129–144. Available from: https://doi.org/10.1017/S1742758411000257.

- de Gier, S., Verhoeckx, K., 2018. Insect (food) allergy and allergens. Mol. Immunol. 100, 82–106. Available from: https://doi.org/10.1016/j.molimm.2018.03.015.
- Goggin, D.E., Mir, G., Smith, W.B., et al., 2008. Proteomic analysis of lupin seed proteins to identify conglutin β as an allergen, Lup an 1. J. Agric. Food Chem. 56, 6370–6377. Available from: https://doi.org/10.1021/jf800840u.
- Grabowski, N.T., Klein, G., 2016. Microbiology of processed edible insect products—results of a preliminary survey. J. Food Microbiol. 243, 103–107.
- Grau, T., Vilcinskas, A., Joop, G., 2017. Sustainable farming of the mealworm *Tenebrio molitor* for the production of food and feed. Z. Nat. Sect. C 72, 337–349. Available from: https://doi.org/10.1515/znc-2017-0033.
- Hindley, J., Wunschmann, S., Satinover, S., et al., 2006. Bla g 6: a troponin C allergen from *Blattella germanica* with IgE binding calcium dependence. J. Allergy Clin. Immunol. 117, 1389–1395. Available from: https://doi.org/10.1016/j.jaci.2006.02.017.
- Hadley, C., 2006. Food allergies on the rise? Determining the prevalence of food allergies, and how quickly it is increasing, is the first step in tackling the problem. EMBO Rep. 7, 1080–1083. Available from: https://doi.org/10.1038/sj.embor.7400846.
- Hager, A.J., Bolton, D.L., Pelletier, M.R., et al., 2006. Type IV pili-mediated secretion modulates Francisella virulence. Mol. Microbiol. 62, 227–237. Available from: https://doi.org/10.1111/j.1365-2958.2006.05365.x.
- Hakim, R.S., Blackburn, M.B., Corti, P., et al., 2007. Growth and mitogenic effects of arylphorin in vivo and in vitro. Arch. Insect. Biochem. Physiol. 64, 63–73. Available from: https://doi.org/10.1002/arch.20155.
- Halloran, A., Roos, N., Hanboonsong, Y., 2017. Cricket farming as a livelihood strategy in Thailand. Geogr. J. 183, 112–124. Available from: https://doi.org/10.1111/geoj.12184.
- Halloran, A., Megido, R.C., Oloo, J., et al., 2018. Comparative aspects of cricket farming in Thailand, Cambodia, Lao People's Democratic Republic, Democratic Republic of the Congo and Kenya. J. Insects Food Feed 4, 1–14. Available from: https://doi.org/10.3920/JIFF2017.0016.
- Hamerman, E.J., 2016. Cooking and disgust sensitivity influence preference for attending insect-based food events. Appetite 96, 319–326. Available from: https://doi.org/10.1016/j.appet.2015.09.029.
- Hanboonsong, Y., 2014. Farming insects for food in Thailand. RAP Publication, pp. 116-122.
- Hanboonsong, Y., Jamjanya, T., Durst, P.B., 2013. Six-legged livestock: edible insect farming, collecting and marketing in Thailand. Food and Agriculture Organization of the United Nations, Regional Office for Asia and the Pacific, Bangkok.
- Hartmann, C., Siegrist, M., 2016. Becoming an insectivore: results of an experiment. Food Qual. Prefer. 51, 118–122. Available from: https://doi.org/10.1016/j.foodqual.2016.03.003.
- Hartmann, C., Shi, J., Giusto, A., Siegrist, M., 2015. The psychology of eating insects: a cross-cultural comparison between Germany and China. Food Qual. Prefer. 44, 148–156. Available from: https://doi.org/10.1016/j. foodqual.2015.04.013.
- Hoffman, A., 2014. Inside the edible insect industrial complex. In: Fast Co. Available from: <a href="https://www.fastcompany.com/3037716/inside-the-edible-insect-industrial-complex">https://www.fastcompany.com/3037716/inside-the-edible-insect-industrial-complex</a>.
- Homann, A.M., Ayieko, M.A., Konyole, S.O., Roos, N., 2017. Acceptability of biscuits containing 10% cricket (*Acheta domesticus*) compared to milk biscuits among 5–10-year-old Kenyan schoolchildren. J. Insects Food Feed 3, 95–103. Available from: https://doi.org/10.3920/JIFF2016.0054.
- House, J., 2016. Consumer acceptance of insect-based foods in the Netherlands: academic and commercial implications. Appetite 107, 47–58. Available from: https://doi.org/10.1016/j.appet.2016.07.023.
- Hurton, L., Berkson, J., Smith, S., 2005. Estimation of total hemolymph volume in the horseshoe crab *Limulus poly-phemus*. Mar. Freshw. Behav. Physiol. 38, 139–147. Available from: https://doi.org/10.1080/10236240500064354.
- Ichikawa, S., Takai, T., Yashiki, T., et al., 2009. Lipopolysaccharide binding of the mite allergen Der f 2. Genes Cells 14, 1055–1065. Available from: https://doi.org/10.1111/j.1365-2443.2009.01334.x.
- Jeong, K.Y., Hong, C.-S., Yong, T.-S., 2006. Allergenic tropomyosins and their cross-reactivities. Protein Pept. Lett. 13, 835–845.
- Johnson, D.V., 2010. The contribution of edible forest insects to human nutrition and to forest management. In: Forest Insects as Food: Humans Bite Back. FAO Regional Office for Asia and the Pacific, Bangkok, pp. 5–22.
- Kachapulula, P.W., Akello, J., Bandyopadhyay, R., Cotty, P.J., 2018. Aflatoxin contamination of dried insects and fish in Zambia. J. Food Prot. 81, 1508–1518. Available from: https://doi.org/10.4315/0362-028X.JFP-17-527.

- Kaneko, J.J., Medina, L.B., 2009. Risk of parasitic worm infection from eating raw fish in Hawai'i: a physician's survey. Hawaii Med. J. 68, 227–229.
- Katayama, N., Ishikawa, Y., Takaoki, M., et al., 2008. Entomophagy: a key to space agriculture. Adv. Sp. Res. 41, 701–705. Available from: https://doi.org/10.1016/j.asr.2007.01.027.
- Kemp, S.F., Lockey, R.F., 2002. Anaphylaxis: a review of causes and mechanisms. J. Allergy Clin. Immunol. 110, 341–348.
- Kempkes, C., Buddenkotte, J., Cevikbas, F., et al., 2014. Role of PAR-2 in neuroimmune communication and itch. In: Carstens, E., Akiyama, T. (Eds.). CRC Press/Taylor & Francis.
- Khan, M.H., Lone, A.A., Wani, S.H., 2013. Genetic engineering for crop improvement. J. Plant Sci. Res. 29, 5.
- Kim, C.-W., Hong, C.-S., 2007. Allergy to miscellaneous household arthropods. Protein Pept. Lett. 14, 982-991.
- Kim, H.W., Setyabrata, D., Lee, Y.J., et al., 2016. Pre-treated mealworm larvae and silkworm pupae as a novel protein ingredient in emulsion sausages. Innov. Food Sci. Emerg. Technol. 38, 116–123. Available from: https://doi.org/10.1016/j.ifset.2016.09.023.
- Kim, S., Hwang, S.K., Dwek, R.A., et al., 2003. Structural determination of the N-glycans of a lepidopteran arylphorin reveals the presence of a monoglucosylated oligosaccharide in the storage protein. Glycobiology 13, 147–157. Available from: https://doi.org/10.1093/glycob/cwg023.
- Kim, S.-K., Weaver, C.M., Choi, M.-K., 2017. Proximate composition and mineral content of five edible insects consumed in Korea. CyTA J. Food 15, 1–4. Available from: https://doi.org/10.1080/19476337.2016.1223172.
- Ligman, K., 2015. Grass-whoppers and yummy stinkbugs. Newsweek Glob. 164, 48-50.
- Lang, T., Barling, D., 2013. Nutrition and sustainability: an emerging food policy discourse. Proc. Nutr. Soc. 72, 1–12.
- Li, D., Sihamala, O., Bhulaidok, S., Shen, L., 2009. Changes in the organic compounds following sun drying of edible black ant (*Polyrhachis vicina* Roger). Acta Aliment. 38, 493–501. Available from: https://doi.org/10.1556/AAlim.38.2009.4.9.
- Linn, S.E., 2014. The insect cookbook: food for a sustainable planet review. FlaEntomol. 99, 157–158. Available from: https://doi.org/10.1016/j.cell.2016.01.041.
- Liu, C., Zhao, J., 2018. Insects as a novel food. In: Reference Module in Food Science. Available from: http://dx.doi.org/10.1016/B978-0-08-100596-5.21782-4.
- Lopata, A.L., O'Hehir, R.E., Lehrer, S.B., 2010. Shellfish allergy. Clin. Exp. Allergy 40, 850-858.
- Ludman, S.W., Boyle, R.J., 2015. Stinging insect allergy: current perspectives on venom immunotherapy. J. Asthma Allergy 8, 75–86. Available from: https://doi.org/10.2147/JAA.S62288.
- Macan, J., Plavec, D., Kanceljak, B., Milkovic-Kraus, S., 2003. Exposure levels and skin reactivity to German cockroach (*Blattella germanica*) in Croatia. Croat. Med. J. 44, 756–760.
- Maciel-Vergara, G., Ros, V.I.D., 2017. Viruses of insects reared for food and feed. J. Invertebr. Pathol. 147, 60–75. Available from: https://doi.org/10.1016/j.jip.2017.01.013.
- Maki, K.C., Reeves, M.S., Farmer, M., et al., 2009. Krill oil supplementation increases plasma concentrations of eicosapentaenoic and docosahexaenoic acids in overweight and obese men and women. Nutr. Res. 29, 609–615. Available from: https://doi.org/10.1016/j.nutres.2009.09.004.
- Manca, G., Porcu, A., Ru, A., et al., 2015. Comparison of  $\gamma$ -aminobutyric acid and biogenic amine content of different types of ewe's milk cheese produced in Sardinia, Italy. Ital. J. Food Saf. 4, 4700. Available from: https://doi.org/10.4081/ijfs.2015.4700.
- Marshall, D.L., Dickson, J.S., Nguyen, N.H., 2016. Ensuring food safety in insect based foods: mitigating microbiological and other foodborne hazards. In: Insects as Sustainable Food Ingredients Production, Processing and Food Applications, 223–253.
- McClements, R.D., Lintzenich, B.A., Boardman J., 2003. A zoo-wide evaluation into the current feeder insect supplementation program at The Brookfield Zoo. In: Proceedings of Nutr Advis Gr Fifth Conference Zoo Wildlife Nutrition, pp. 54–59.
- Menozzi, D., Sogari, G., Veneziani, M., et al., 2017. Eating novel foods: an application of the Theory of Planned Behaviour to predict the consumption of an insect-based product. Food Qual. Prefer. 59, 27–34. Available from: https://doi.org/10.1016/j.foodqual.2017.02.001.
- Morris, B., 2008. Insects as food among hunter-gatherers. Anthropol. Today 24, 6–8. Available from: https://doi.org/10.1111/j.1467-8322.2008.00558.x.
- Nadeau, L., Nadeau, I., Franklin, F., Dunkel, F., 2015. The potential for entomophagy to address undernutrition. Ecol. Food Nutr. 54, 200–208. Available from: https://doi.org/10.1080/03670244.2014.930032.

- Nandasena M.R.M.P., Disanayake D.M.S.K., Weeratunga L., 2010. Sri Lanka as a potential gene pool of edible insects. In: Forest Insects as Food: Humans Bite Back. Proceedings of a Workshop on Asia-Pacific Resources and Their Potential for Development, Chiang Mai, Thailand, 19–21 February 2008, pp. 161–164.
- Nath, H.K.D., Gogoi, R., Gogoi, G., 2005. Insects as human food with special reference to Assam and Northeast India. Asian Agrihist. 9, 119–127.
- Navarro, J.C.A., Prado, S.M.C., Cárdenas, P.A., et al., 2010. Pre-historic eating patterns in Latin America and protective effects of plant-based diets on cardiovascular risk factors. Clinics (Sao Paulo) 65, 1049–1054. Available from: https://doi.org/10.1590/s1807-59322010001000022.
- Navarro-Garcia, F., Sonnested, M., Teter, K., 2010. Host—toxin interactions involving EspC and pet, two serine protease autotransporters of the Enterobacteriaceae. Toxins (Basel) 2, 1134—1147. Available from: https://doi.org/10.3390/toxins2051134.
- Negulescu, H., Guo, Y., Garner, T.P., et al., 2015. A Kazal-type serine protease inhibitor from the defense gland secretion of the subterranean termite *Coptotermes formosanus* Shiraki. PLoS One 10, e0125376. Available from: https://doi.org/10.1371/journal.pone.0125376.
- Netshifhefhe, S.R., Kunjeku, E.C., Duncan, F.D., 2018. Human uses and indigenous knowledge of edible termites in Vhembe District, Limpopo Province, South Africa. S. Afr. J. Sci. 114, 1–2.
- Nonaka, K., 2010. Cultural and commercial roles of edible wasps in Japan. In: Forest Insects as Food: Humans Bite Back. FAO Regional Office for Asia and the Pacific, Bangkok, pp 123–130.
- Nowak, V., Persijn, D., Rittenschober, D., Charrondiere, U.R., 2016. Review of food composition data for edible insects. Food Chem. 193, 39–46. Available from: https://doi.org/10.1016/j.foodchem.2014.10.114.
- Oibiokpa, F.I., Akanya, H.O., Jigam, A.A., et al., 2018. Protein quality of four indigenous edible insect species in Nigeria. Food Sci. Hum Wellness . Available from: https://doi.org/10.1016/j.fshw.2018.05.003.
- Okezie, O.A., Kgomotso, K.K., Letswiti, M.M., 2010. Mopane worm allergy in a 36-year-old woman: a case report. J. Med. Case Rep. 4, 42. Available from: https://doi.org/10.1186/1752-1947-4-42.
- Onishi, N., 2010. From dung to coffee brew with no aftertaste. New York Times.
- Pal, P., Roy, S., 2014. Edible Insects: future of human food—a review. Int. Lett. Nat. Sci. 26, 1–11. Available from: https://doi.org/10.18052/www.scipress.com/ILNS.26.1.
- Patel, S., 2016a. Emerging adjuvant therapy for cancer: propolis and its constituents. J. Diet Suppl. 13, 245–268. Available from: https://doi.org/10.3109/19390211.2015.1008614.
- Patel, S., 2016b. Plant-derived cardiac glycosides: role in heart ailments and cancer management. Biomed. Pharmacother. 84, 1036–1041. Available from: https://doi.org/10.1016/j.biopha.2016.10.030.
- Patel, S., 2017a. Disruption of aromatase homeostasis as the cause of a multiplicity of ailments: a comprehensive review. J. Steroid Biochem. Mol. Biol. 168, 19–25. Available from: https://doi.org/10.1016/j.jsbmb.2017.01.009.
- Patel, S., 2017b. Pathogenicity-associated protein domains: the fiercely-conserved evolutionary signatures. Gene Rep. 7, 127–141. Available from: https://doi.org/10.1016/j.genrep.2017.04.004.
- Patel, S., Akhtar, N., 2017. Antimicrobial peptides (AMPs): the quintessential "offense and defense" molecules are more than antimicrobials. Biomed. Pharmacother. 95, 1276–1283. Available from: https://doi.org/10.1016/j. biopha.2017.09.042.
- Patel, S., Goyal, A., 2017. Chitin and chitinase: role in pathogenicity, allergenicity and health. Int. J. Biol. Macromol. 97, 331–338. Available from: https://doi.org/10.1016/j.ijbiomac.2017.01.042.
- Patel, S., Meher, B.R., 2016. A review on emerging frontiers of house dust mite and cockroach allergy research. Allergol. Immunopathol. (Madr). Available from: https://doi.org/10.1016/j.aller.2015.11.001.
- Patel, S., Rauf, A., Meher, B.R., 2017. In silico analysis of ChtBD3 domain to find its role in bacterial pathogenesis and beyond. Microb. Pathog. 110, 519–526. Available from: https://doi.org/10.1016/j.micpath.2017.07.047.
- Patel, S., Homaei, A., Raju, A.B., Meher, B.R., 2018. Estrogen: the necessary evil for human health, and ways to tame it. Biomed. Pharmacother. 102, 403–411. Available from: https://doi.org/10.1016/j.biopha.2018.03.078.
- Paul, A., Frederich, M., Uyttenbroeck, R., et al., 2015. Food compounds from meadow grasshoppers. Troisièmes Journées Sci. l'Agro-Alimentaire A4 - Assoc. Méditarranéenne des Ind. Agro-Alimentaires A0.
- Paul, A., Frederich, M., Megido, R.C., et al., 2017. Insect fatty acids: a comparison of lipids from three Orthopterans and *Tenebrio molitor* L. larvae. J. Asia Pac. Entomol. 20, 337–340. Available from: https://doi.org/10.1016/j.aspen.2017.02.001.
- Pener, M.P., 2016. Allergy to crickets: a review. J. Orthoptera Res. 25, 91–95. Available from: https://doi.org/10.1665/034.025.0208.

- Petschenka, G., Agrawal, A.A., 2015. Milkweed butterfly resistance to plant toxins is linked to sequestration, not coping with a toxic diet. Proc. Biol. Sci. 282, 20151865. Available from: https://doi.org/10.1098/rspb.2015.1865.
- Pier, J., Lomas, J., 2017. P083 anaphylaxis to crickets. Ann. Allergy Asthma Immunol. 119, S34. Available from: https://doi.org/10.1016/j.anai.2017.08.124.
- Pihlanto, A., Korhonen, H., 2003. Bioactive peptides and proteins. Adv. Food Nutr. Res. 47, 175-276.
- Ponzetta, M.T., Paoletti, M.G., 1997. Insects as food of the Irian Jaya populations. Ecol. Food Nutr. 36, 321–346. Available from: https://doi.org/10.1080/03670244.1997.9991522.
- Premalatha, M., Abbasi, T., Abbasi, T., Abbasi, S.A., 2011. Energy-efficient food production to reduce global warming and ecodegradation: the use of edible insects. Renew. Sustain. Energy Rev. 15, 4357–4360. Available from: https://doi.org/10.1016/j.rser.2011.07.115.
- Przybilla, B., Ruëff, F., 2010. Hymenoptera venom allergy. J. Dtsch. Dermatol. Ges. 8, 114–27-30. Available from: https://doi.org/10.1111/j.1610-0387.2009.07125.x.
- Ramaswamy, S.B., 2015. Setting the table for a hotter, flatter, more crowded earth: insects on the menu? J. Insects Food Feed 1, 171–178. Available from: https://doi.org/10.3920/JIFF2015.0032.
- Ramos-Elorduy, J., 2009. Anthropo-entomophagy: cultures, evolution and sustainability. Entomol. Res. 39, 271–288.
- Ramos-Elorduy, J., Moreno, J.M.P., Vázquez, A.I., et al., 2011. Edible Lepidoptera in Mexico: geographic distribution, ethnicity, economic and nutritional importance for rural people. J. Ethnobiol. Ethnomed. 7, 2. Available from: https://doi.org/10.1186/1746-4269-7-2.
- Raubenheimer, D., Rothman, J.M., 2013. Nutritional ecology of entomophagy in humans and other primates. Annu. Rev. Entomol. 58, 141–160. Available from: https://doi.org/10.1146/annurev-ento-120710-100713.
- Reese, G., Ayuso, R., Lehrer, S.B., 1999. Tropomyosin: an invertebrate pan-allergen. Int. Arch. Allergy Immunol. 119, 247–258. Available from: https://doi.org/10.1159/000024201.
- Robinson, N., 2015. First EU edible insect food safety guide published. Food Manuf.
- Rodriguez, V., Bartolomé, B., Armisén, M., Vidal, C., 2007. Food allergy to *Paracentrotus lividus* (sea urchin roe). Ann. Allergy Asthma Immunol. 98, 393–396. Available from: https://doi.org/10.1016/S1081-1206(10)60888-5.
- Rossano, R., Larocca, M., Polito, T., et al., 2012. What are the proteolytic enzymes of honey and what they do tell us? A fingerprint analysis by 2-D zymography of unifloral honeys. PLoS One 7, e49164. Available from: https://doi.org/10.1371/journal.pone.0049164.
- Rumpold, B.A., Schlüter, O.K., 2013. Potential and challenges of insects as an innovative source for food and feed production. Innov. Food Sci. Emerg. Technol. 17, 1–11. Available from: https://doi.org/10.1016/j. ifset.2012.11.005.
- Sabado, E.M., Aguanta L.M., 2014. Promoting entomophagy through insect-eating festivals. http://www.uplb.edu.ph. Sathe, T.V., 2015. Insects for human diet from Kolhapur region, India. Int. J. Pharm. Biol. Sci. 6, B519—B527.
- Schrader, J., Oonincx, D.G.A.B., Ferreira, M.P., 2016. North American entomophagy. J. Insects Food Feed 2, 111–120. Available from: https://doi.org/10.3920/JIFF2016.0003.
- Shantibala, T., Lokeshwari, R.K., Debaraj, H., 2014. Nutritional and antinutritional composition of the five species of aquatic edible insects consumed in Manipur, India. J. Insect. Sci. 14, 14. Available from: https://doi.org/10.1093/jis/14.1.14.
- Shen, L., Li, D., Feng, F., Ren, Y., 2006. Nutritional composition of *Polyrhachis vicina* Roger (edible Chinese black ant). Songklanakarin J. Sci. Technol. 28, 107–114.
- Shewry, P.R., Jones, H.D., 2006. Developing allergen-free foods by genetic manipulation. In: Managing Allergens in Food. CRC Press. Woodhead Publishing, Cambridge, pp. 147–158.
- Belluco, S., Losasso, C., Maggioletti, M., Alonzi, C., Ricci, A., Paoletti, M.G., 2015. Edible insects: a food security solution or a food safety concern? Anim. Front. 5 (2), 25–30. Available from: https://doi.org/10.2527/af.2015-0016.
- Sidali, K.L., Pizzo, S., Garrido-Pérez, E.I., Schamel, G., 2018. Between food delicacies and food taboos: a structural equation model to assess Western students' acceptance of Amazonian insect food. Food Res. Int. 115, 83–89.
- Silva-Carvalho, R., Miranda-Gonçalves, V., Ferreira, A.M., et al., 2014. Antitumoural and antiangiogenic activity of *Portuguese propolis* in in vitro and in vivo models. J. Funct. Foods 11, 160–171. Available from: https://doi.org/10.1016/j.jff.2014.09.009.
- Slocinska, M., Marciniak, P., Rosinski, G., 2008. Insects antiviral and anticancer peptides: new leads for the future. Protein Pept. Lett. 15, 578–585. Available from: https://doi.org/10.2174/092986608784966912.

- Smetana, S., Mathys, A., Knoch, A., Heinz, V., 2015. Sustainability of meat substitutes: a path to future foods? In: 29th EFFoST International Conference Proceedings, pp. 126–131.
- Smetana, S., Palanisamy, M., Mathys, A., Heinz, V., 2016. Sustainability of insect use for feed and food: Life Cycle Assessment perspective. J. Clean. Prod. 137, 741–751. Available from: https://doi.org/10.1016/j.jclepro.2016.07.148.
- Sokol, W.N., Wünschmann, S., Agah, S., 2017. Grasshopper anaphylaxis in patients allergic to dust mite, cockroach, and crustaceans: is tropomyosin the cause? Ann. Allergy Asthma Immunol. 119, 91–92. Available from: https://doi.org/10.1016/j.anai.2017.05.007.
- Sousa, P.C.P., Brito, T.S., Freire, D.S., et al., 2013. Vasoconstrictor effect of Africanized honeybee (*Apis mellifera* L.) venom on rat aorta. J. Venom Anim. Toxins Incl. Trop. Dis. 19, 24. Available from: https://doi.org/10.1186/1678-9199-19-24.
- Srinroch, C., Srisomsap, C., Chokchaichamnankit, D., et al., 2015. Identification of novel allergen in edible insect, *Gryllus bimaculatus* and its cross-reactivity with *Macrobrachium* spp. allergens. Food Chem. 184, 160–166. Available from: https://doi.org/10.1016/j.foodchem.2015.03.094.
- Suarthana, E., Shen, A., Henneberger, P.K., et al., 2012. Post-hire asthma among insect-rearing workers. J. Occup. Environ. Med. 54, 310–317. Available from: https://doi.org/10.1097/JOM.0b013e31823fe098.
- Sudha, V.T., Arora, N., Gaur, S.N., et al., 2008. Identification of a serine protease as a major allergen (Per a 10) of *Periplaneta americana*. Allergy 63, 768–776. Available from: https://doi.org/10.1111/j.1398-9995.2007.01602.x.
- Sun-Waterhouse, D., Waterhouse, G.I.N., You, L., et al., 2016. Transforming insect biomass into consumer wellness foods: a review. Food Res. Int. 89, 129–151. Available from: https://doi.org/10.1016/j.foodres.2016.10.001.
- Suwannapong, G., Benbow, M.E., 2011. The biology of insect odors: sources and olfaction. In: The Biology of Odors. first ed., Logan E. Weiss, Jason M. Atwood (Eds.), Nova Science, pp.153–184.
- Tagliabue, J., 2011. Dutch try to change "Ick" to "Yum" for insect dishes. New York Times.
- Takeda, S., 2009. Sericulture. In: Encyclopedia of Insects, pp. 912–914. Available from: https://doi.org/10.1016/B978-0-12-374144-8.X0001-X.
- Takeo, S., Hisamori, D., Matsuda, S., et al., 2009. Enzymatic characterization of the *Plasmodium vivax* chitinase, a potential malaria transmission-blocking target. Parasitol. Int. 58, 243–248. Available from: https://doi.org/10.1016/j.parint.2009.05.002.
- Tan, H.S.G., Fischer, A.R.H., van Trijp, H.C.M., Stieger, M., 2016. Tasty but nasty? Exploring the role of sensory-liking and food appropriateness in the willingness to eat unusual novel foods like insects. Food Qual. Prefer. 48, 293–302. Available from: https://doi.org/10.1016/j.foodqual.2015.11.001.
- Telang, a, Buck, N., Wheeler, D., 2002. Response of storage protein levels to variation in dietary protein levels. J. Insect. Physiol. 48, 1021–1029. Available from: https://doi.org/10.1016/S0022-1910(02)00190-7.
- Tikader, A., Vijayan, K., Saratchandra, B., 2013. Muga silkworm, *Antheraea assamensis* (Lepidoptera: Saturniidae)—an overview of distribution, biology and breeding. Eur. J. Entomol. 110, 293–300. Available from: https://doi.org/10.14411/eje.2013.096.
- Tong, L., Yu, X., Liu, H., 2011. Insect food for astronauts: gas exchange in silkworms fed on mulberry and lettuce and the nutritional value of these insects for human consumption during deep space flights. Bull. Entomol. Res. 101, 613–622. Available from: https://doi.org/10.1017/S0007485311000228.
- Tonk, M., Vilcinskas, A., 2017. The medical potential of antimicrobial peptides from insects. Curr. Top. Med. Chem. 17, 554–575. Available from: https://doi.org/10.2174/1568026616666160713123654.
- Torbeck R., Pan M., DeMoll E., Levitt J., 2014. Cantharidin: a comprehensive review of the clinical literature. Dermatol. Online J. 20, pii: 13030/qt45r512w0.
- van Huis, A., 2013. Potential of insects as food and feed in assuring food security. Annu. Rev. Entomol. 58, 563–583. Available from: https://doi.org/10.1146/annurev-ento-120811-153704.
- van Huis, A., 2017. Cultural significance of termites in sub-Saharan Africa. J. Ethnobiol. Ethnomed. 13, 8. Available from: https://doi.org/10.1186/s13002-017-0137-z.
- van Huis, A., Berry, E., Dernini, S., et al., 2015. Edible insects contributing to food security? Agric. Food Security 4, 20. Available from: https://doi.org/10.1186/s40066-015-0041-5.
- Van Huis, A., 2016. Edible insects are the future? Proc. Nutr. Soc. 75, 294–305.
- Van Huis, A., Dunkel, F.V., 2016. Edible insects: a neglected and promising food source. In: Sustainable Protein Sources, pp. 341–355 (Chapter 21). http://dx.doi.org/10.1016/B978-0-12-802778-3.00021-4.
- Verde, M.M., 2014. Apiculture and food safety. Cuba J. Agric. Sci. 48, 25–31.

- Verkerk, M.C., Tramper, J., van Trijp, J.C.M., Martens, D.E., 2007. Insect cells for human food. Biotechnol. Adv. 25, 198–202.
- Voltolini, S., Pellegrini, S., Contatore, M., et al., 2014. New risks from ancient food dyes: cochineal red allergy. Eur. Ann. Allergy Clin. Immunol. 46, 232–233.
- Walia, K., Kapoor, A., Farber, J.M., 2018. Qualitative risk assessment of cricket powder to be used to treat undernutrition in infants and children in Cambodia. Food Control 92, 169–182.
- Wallace, A., 2010. Insect toppings. Sci. World 67, 22.
- Wan, H., Winton, H.L., Soeller, C., et al., 2001. The transmembrane protein occludin of epithelial tight junctions is a functional target for serine peptidases from faecal pellets of *Dermatophagoides pteronyssinus*. Clin. Exp. Allergy 31, 279–294.
- Wan, H., Lee, K.S., Kim, B.Y., et al., 2013. A spider-derived Kunitz-type serine protease inhibitor that acts as a plasmin inhibitor and an elastase inhibitor. PLoS One 8, e53343. Available from: https://doi.org/10.1371/journal.pone.0053343.
- Wang, F., Robotham, J.M., Teuber, S.S., et al., 2002. Ana o 1, a cashew (*Anacardium occidental*) allergen of the vicilin seed storage protein family. J. Allergy Clin. Immunol. 110, 160–166. Available from: https://doi.org/10.1067/mai.2002.125208.
- Wang, Y.-S., Shelomi, M., 2017. Review of black soldier fly (*Hermetia illucens*) as animal feed and human food. Foods 6, 91. Available from: https://doi.org/10.3390/foods6100091.
- Warrant, E., Frost, B., Green, K., et al., 2016. The Australian Bogong moth *Agrotis infusa*: a long-distance nocturnal navigator. Front. Behav. Neurosci 10, 77. Available from: https://doi.org/10.3389/fnbeh.2016.00077.
- Wilkinson, K., Muhlhausler, B., Motley, C., et al., 2018. Australian consumers' awareness and acceptance of insects as food. Insects. Available from: https://doi.org/10.3390/insects9020044.
- Yates-Doerr, E., 2015. The world in a box? Food security, edible insects, and "One World, One Health" collaboration. Soc. Sci. Med. 129, 106–112. Available from: https://doi.org/10.1016/j.socscimed.2014.06.020.
- Yen, A.L., 2010. Edible insects and other invertebrates in Australia: future prospects. For insects as food humans bite back 65. In: Proceedings of a Workshop on Asia-Pacific Resources and Their Potential for Development, Chiang Mai, Thailand. ISBN: 978-92-5-106488-7.
- Zagrobelny, M., Møller, B.L., 2011. Cyanogenic glucosides in the biological warfare between plants and insects: the Burnet moth-Birds foot trefoil model system. Phytochemistry 72, 1585–1592. Available from: https://doi.org/10.1016/j.phytochem.2011.02.023.
- Zhang, H., Zeng, X., He, S., 2014. Evaluation on potential contributions of protease activated receptors related mediators in allergic inflammation. Mediators Inflamm. 2014, 829068. Available from: https://doi.org/ 10.1155/2014/829068.