# compgen2021: Week 1 exercises

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```
library(mosaic)
## Registered S3 method overwritten by 'mosaic':
     method
##
##
     fortify.SpatialPolygonsDataFrame ggplot2
##
## The 'mosaic' package masks several functions from core packages in order to add
## additional features. The original behavior of these functions should not be affected by this.
##
## Attaching package: 'mosaic'
## The following objects are masked from 'package:dplyr':
##
##
       count, do, tally
## The following object is masked from 'package:Matrix':
##
##
       mean
## The following object is masked from 'package:ggplot2':
##
##
       stat
## The following objects are masked from 'package:stats':
##
       binom.test, cor, cor.test, cov, fivenum, IQR, median, prop.test,
##
##
       quantile, sd, t.test, var
## The following objects are masked from 'package:base':
##
##
       max, mean, min, prod, range, sample, sum
library(matrixStats)
## Attaching package: 'matrixStats'
## The following objects are masked from 'package:mosaic':
##
       count, iqr
##
```

```
## The following object is masked from 'package:dplyr':
##
## count
library(qvalue)
```

# Exercises for Week1

# Statistics for genomics

How to summarize collection of data points: The idea behind statistical distributions

1. Calculate the means and variances of the rows of the following simulated data set, and plot the distributions of means and variances using hist() and boxplot() functions. [Difficulty: Beginner/Intermediate]

```
#sample data matrix from normal distribution
gset=rnorm(600,mean=200,sd=70)
data=matrix(gset,ncol=6)

row_means=rowMeans(data)
row_vars= apply(data,1,var)

par(mfrow=c(2,2))
hist(row_means)
boxplot(row_means)
hist(row_vars)
boxplot(row_vars)
```

2. Using the data generated above, calculate the standard deviation of the distribution of the means using the sd() function. Compare that to the expected standard error obtained from the central limit theorem keeping in mind the population parameters were  $\sigma = 70$  and n = 6. How does the estimate from the random samples change if we simulate more data with data=matrix(rnorm(6000,mean=200,sd=70),ncol=6)? [Difficulty: Beginner/Intermediate]

```
gset=rnorm(600,mean=200,sd=70)
data=matrix(gset,ncol=6)
row_means=rowMeans(data)

means_sd = sd(row_means)
ctl_sd=70/sqrt(6)

#The standard deviation of the sample means is similar to the standard deviation from CLT.

data2=matrix(rnorm(6000,mean=200,sd=70),ncol=6)
row_means2=rowMeans(data2)
means_sd2 = sd(row_means2)

#Changing to 6000 did not make a difference, the estimate from the random sampling is still very simila
```

- 3. Simulate 30 random variables using the rpois() function. Do this 1000 times and calculate the mean of each sample. Plot the sampling distributions of the means using a histogram. Get the 2.5th and 97.5th percentiles of the distribution. [Difficulty: Beginner/Intermediate]
- 4. Use the t.test() function to calculate confidence intervals of the mean on the first random sample pois1 simulated from the rpois() function below. [Difficulty: Intermediate]

5. Use the bootstrap confidence interval for the mean on pois1. [Difficulty: Intermediate/Advanced]

```
#Bootstrapping is when we don't know the population parameter (Lambda in this example).So we take sampl
set.seed(100)
pois1= do(1000) * mean(rpois(30,lambda=5))
boot_samples = do(1000)*sample(pois1,30,replace=T)
quantile(boot_samples[,1],p=c(0.025,0.975))
### 2.5% 97.5%
```

#### How to test for differences in samples

## 4.166667 5.866667

1. Test the difference of means of the following simulated genes using the randomization, t-test(), and wilcox.test() functions. Plot the distributions using histograms and boxplots. [Difficulty: Intermediate/Advanced]

```
set.seed(101)
gene1=rnorm(30,mean=4,sd=3)
gene2=rnorm(30,mean=3,sd=3)

t.test(gene1,gene2)
wilcox.test(gene1,gene2)

#Randomization
#Save the real difference between the gene1 and geen2 groups
#Generate a new data frame using gene1 and gene2 values (in esp column) with 30 test and 30 control lab
org.diff=mean(gene1)-mean(gene2)
```

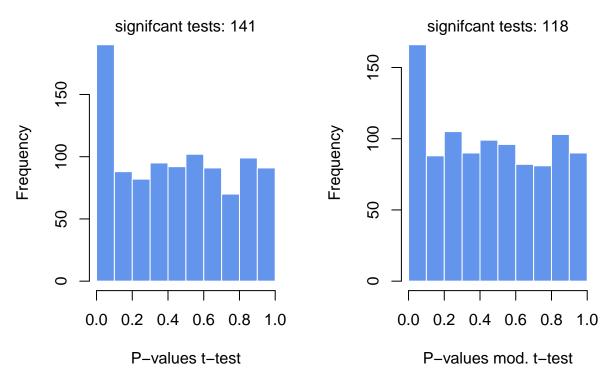
2. Test the difference of the means of the following simulated genes using the randomization, t-test() and wilcox.test() functions. Plot the distributions using histograms and boxplots. [Difficulty: Intermediate/Advanced]

```
set.seed(100)
gene1=rnorm(30,mean=4,sd=2)
gene2=rnorm(30,mean=2,sd=2)
## Same as above
t.test(gene1,gene2)
wilcox.test(gene1,gene2)
#Randomization
#Save the real difference between the gene1 and geen2 groups
#Generate a new data frame using gene1 and gene2 values (in esp column) with 30 test and 30 control lab
org.diff=mean(gene1)-mean(gene2)
gene.df=data.frame(exp=c(gene1,gene2),
                  group=c(rep("test",30),rep("control",30)))
#Shuffle by the group labels and get the means for control and test, get the difference, repeat 1000 ti
shuffled_means = do(1000) * diff(mosaic::mean(exp ~ shuffle(group), data=gene.df))
hist(shuffled_means[,1],
     xlab="null distribution | no difference in samples",
    main=expression(paste(H[0]," :no difference in means") ),
    xlim=c(-2,2),
     col="cornflowerblue",
     border="white")
abline(v=quantile(exp.null[,1],0.95),col="red")
abline(v=org.diff,col="blue" )
text(x=quantile(exp.null[,1],0.95),y=200,"0.05",adj=c(1,0),col="red")
text(x=org.diff,y=200,"org. diff.",adj=c(1,0),col="blue")
```

3. We need an extra data set for this exercise. Read the gene expression data set as fol-

lows: gexpFile=system.file("extdata", "geneExpMat.rds", package="compGenomRData") data=readRDS(gexpFile). The data has 100 differentially expressed genes. The first 3 columns are the test samples, and the last 3 are the control samples. Do a t-test for each gene (each row is a gene), and record the p-values. Then, do a moderated t-test, as shown in section "Moderated t-tests" in this chapter, and record the p-values. Make a p-value histogram and compare two approaches in terms of the number of significant tests with the 0.05 threshold. On the p-values use FDR (BH), Bonferroni and q-value adjustment methods. Calculate how many adjusted p-values are below 0.05 for each approach. [Difficulty: Intermediate/Advanced]

```
#Read the data
gexpFile=system.file("extdata", "geneExpMat.rds", package="compGenomRData")
data=readRDS(gexpFile)
#For the whole data
#Check if the variances are similar for the t.test:
col_vars= apply(data,2,var)
	t Welch Two Sample t-test since variances are different:
test=data[,1:3]
control=data[,4:6]
n1=3
n2 = 3
stats::t.test(test,control)
##
## Welch Two Sample t-test
## data: test and control
## t = -14.74, df = 4115, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -0.2222141 -0.1700416
## sample estimates:
       mean of x
                     mean of y
## -0.0004465826 0.1956812552
#Testing each gene:
#The formula for the Welch's t-test is: Differences of means/ variance
#Calculate the differences of the means of 2 groups:
diff_means=rowMeans(test)-rowMeans(control)
#Get the estimate of pooled variance
pooled_var = sqrt(
  (rowVars(test)*(n1-1) + rowVars(control)*(n2-1)) / (n1+n2-2) * (1/n1 + 1/n2)
  )
#Estimate t statistic without moderated variance and calculate P-value of rejecting null
t = diff_means / pooled_var
p = 2*pt(-abs(t), n1+n2-2)
#Same with the moderated variance
mod.pooled_var = (pooled_var + median(pooled_var)) / 2 # moderation in variation
t.mod <- diff_means / mod.pooled_var</pre>
```



```
#Pvalue correction for the t-test and the moderated t-test:
qvalues <- qvalue(p)
qvaluesmod <- qvalue(p.mod)
bonf.pval=p.adjust(p,method ="bonferroni")
bonf.pvalmod=p.adjust(p.mod,method ="bonferroni")
fdr.adj.pval=p.adjust(p,method ="fdr")
fdr.adj.pvalmod=p.adjust(p.mod,method ="fdr")

#For t-test

num_sig_q<- sum(qvalues$qvalues<0.05)
num_sig_bonf<- sum(fdr.adj.pval<0.05)
paste("Qvals:", num_sig_q)</pre>
```

## [1] "Qvals: 100"

```
paste("Bonferroni:", num_sig_bonf)

## [1] "Bonferroni: 25"

paste("FDR:", num_sig_fdr)

## [1] "FDR: 99"

#For the moderated t-test

num_sig_q<- sum(qvaluesmod$qvalues<0.05)
num_sig_bonf<- sum(bonf.pvalmod<0.05)
num_sig_fdr<- sum(fdr.adj.pvalmod<0.05)

paste("Qvals:", num_sig_q)

## [1] "Qvals: 100"

paste("Bonferroni:", num_sig_bonf)

## [1] "Bonferroni: 2"

paste("FDR:", num_sig_fdr)

## [1] "FDR: 100"</pre>
```

# Relationship between variables: Linear models and correlation

Below we are going to simulate X and Y values that are needed for the rest of the exercise.

```
# set random number seed, so that the random numbers from the text
# is the same when you run the code.
set.seed(32)

# get 50 X values between 1 and 100
x = runif(50,1,100)

# set b0,b1 and variance (sigma)
b0 = 10
b1 = 2
sigma = 20
# simulate error terms from normal distribution
eps = rnorm(50,0,sigma)
# get y values from the linear equation and addition of error terms
y = b0 + b1*x+ eps

# 1. Run the code then fit a line to predict Y based on X. [Difficulty:**Intermediate**]
```

```
model = lm(y \sim x)
summary(model)
# 2. Plot the scatter plot and the fitted line. [Difficulty:**Intermediate**]
plot(x,y)
abline(model,col="red")
# 3. Calculate correlation and R^2. [Difficulty:**Intermediate**]
corr = cor(x,y)
r_sqr=corr^2
# 4. Run the `summary()` function and
#try to extract P-values for the model from the object
#returned by `summary`. See `?summary.lm`. [Difficulty:**Intermediate/Advanced**]
model_summary = summary(model)
model_summary
# 5. Plot the residuals vs. the fitted values plot, by calling the `plot()`
#function with `which=1` as the second argument. First argument
#is the model returned by `lm()`. [Difficulty:**Advanced**]
plot(lm(y~x), which = 1)
# 6. For the next exercises, read the data set histone modification data set. Use the following to get
hmodFile=system.file("extdata",
                    "HistoneModeVSgeneExp.rds",
                     package="compGenomRData")
# There are 3 columns in the dataset. These are measured levels of H3K4me3,
# H3K27me3 and gene expression per gene. Once you read in the data, plot the scatter plot for H3K4me3 v
histone_data=readRDS(hmodFile)
plot(histone_data$H3k4me3, histone_data$measured_log2, xlab = "Measured levels of H3K4me3", ylab= "Gen
# 7. Plot the scatter plot for H3K27me3 vs. expression. [Difficulty:**Beginner**]
plot(histone_data$H3k27me3, histone_data$measured_log2, xlab = "Measured levels of H3k27me3", ylab= " G
# 8. Fit the model for prediction of expression data using: 1) Only H3K4me3 as explanatory variable, 2)
```

```
fit1 = lm(histone_data$measured_log2 ~ histone_data$H3k4me3)
fit2 = lm(histone_data$measured_log2 ~ histone_data$H3k27me3)
fit3 = lm(histone_data$measured_log2 ~ histone_data$H3k4me3 + histone_data$H3k27me3)
summary(fit1)
summary(fit2)
summary(fit3)
#All terms are significant
# 10. Is using H3K4me3 and H3K27me3 better than the model with only H3K4me3? [Difficulty:**Intermediate
#Comparing R-squared values, which is: R-squared = Explained variation / Total variation. So higher R-s
```