

Gviz visualization: Copy Number Variations

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Installation of BiocManager and Gviz (= Bioconductor package)

```
# Install BiocManager if not yet installed
# Install Gviz package if not yet installed
if (!requireNamespace("BiocManager", quietly = TRUE)){
  install.packages("BiocManager")}
if (! requireNamespace("Gviz", quietly = TRUE)) {
  BiocManager::install("Gviz")
}
# Load BiocManager
# library(BiocManager)
```

Loading Gviz package

Function to create a table with GRCh37 coordinates

```
## separate function requires tidyr package
library(tidyr)
Gviz_table <- function(filename) {
  X <- read.csv(file=filename, header = TRUE, sep=";")
  ##### GRCh37 coordinates in first assembly
  tablepart1 <- X[grep("^GRCh37", X$assembly1),]
  id1 <- tablepart1[1]
  if(any(names(tablepart1) == 'variant_region_id')){regionid1 <- tablepart1[2]}
  if(any(names(tablepart1) == 'study_ID')){studyid1 <- tablepart1[4]}
  Chr_1a <- tablepart1[["Chr_1"]]
  Chr_1a <- gsub("[A-Za-z]", "", Chr_1a)
  Chr_1a <- sub("", "chr", Chr_1a)
  assembly1 <- tablepart1[["assembly1"]]
  assembly1 <- gsub(".*?:", "", assembly1)
  assembly1 <- separate(data = as.data.frame(assembly1), col = assembly1,
    into = c("start", "end"), sep = "-")
  if(any(names(tablepart1) == 'variant_region_id') &
    any(names(tablepart1) == 'study_ID')){
    GRCh37_assembly1 <- data.frame(id1,Chr_1a,assembly1,regionid1,studyid1)
  } else {
    GRCh37_assembly1 <- data.frame(id1,Chr_1a,assembly1)
  }
  names(GRCh37_assembly1)[names(GRCh37_assembly1) == "assembly1"] <- "assembly"
```

```

names(GRCh37_assembly1)[names(GRCh37_assembly1) == "Chr_1a"] <- "chr"
#GRCh37_assembly1["genome"] <- "hg19"
##### GRCh37 coordinates in second assembly
tablepart2 <- X[grep("^GRCh37", X$assembly2),]
id2 <- tablepart2[1]
if(any(names(tablepart2) == 'variant_region_id')){regionid2 <- tablepart2[2]}
if(any(names(tablepart2) == 'study_ID')){studyid2 <- tablepart2[4]}
Chr_1b <- tablepart2[["Chr_2"]]
Chr_1b <- gsub("[A-Za-z]", "", Chr_1b)
Chr_1b <- sub("", "chr", Chr_1b)
assembly2 <- tablepart2[["assembly2"]]
assembly2 <- gsub(".*?:", "", assembly2)
assembly2 <- separate(data = as.data.frame(assembly2), col = assembly2,
                      into = c("start", "end"), sep = "-")
if(any(names(tablepart2) == 'variant_region_id') &
    any(names(tablepart2) == 'study_ID')){
  GRCh37_assembly2 <- data.frame(id2,Chr_1b,assembly2,regionid2,studyid2)
} else {
  GRCh37_assembly2 <- data.frame(id2,Chr_1b,assembly2)
}
names(GRCh37_assembly2)[names(GRCh37_assembly2) == "assembly2"] <- "assembly"
names(GRCh37_assembly2)[names(GRCh37_assembly2) == "Chr_1b"] <- "chr"
#GRCh37_assembly2["genome"] <- "hg19"
##### Gviz table
GRCh37_Gviz <- rbind(GRCh37_assembly1,GRCh37_assembly2)
GRCh37_Gviz <- GRCh37_Gviz[order(-GRCh37_Gviz[1]),]
}

```

Input tables and GRanges Object

```

### results-CNV-dbVar.csv
csvfile <- read.csv(file="results-CNV-dbVAR.csv", header = TRUE, sep=";")
csvfile

```

##	CNV_variant_id	variant_region_id	type	study_ID
## 1	49623411	nsv4457776	['copy number variation']	nstd102
## 2	49353191	nsv4358278	['copy number variation']	nstd102
## 3	49353005	nsv4358092	['copy number variation']	nstd102
## 4	49350830	nsv4355917	['copy number variation']	nstd102
## 5	49349701	nsv4354788	['copy number variation']	nstd102
## 6	49349293	nsv4354380	['copy number variation']	nstd102
## 7	49345450	nsv4350537	['copy number variation']	nstd102
## 8	49344315	nsv4349402	['copy number variation']	nstd102
## 9	48468240	nsv3904885	['copy number variation']	nstd102
## 10	48466558	nsv3903203	['copy number variation']	nstd102
## 11	48466447	nsv3903092	['copy number variation']	nstd102
## 12	48453939	nsv3890584	['copy number variation']	nstd102
## 13	45807136	nsv2779094	['copy number variation']	nstd37
## 14	17813982		[]	nstd102
## 15	17813734		[]	nstd101
## 16	3740775	nsv533414	['copy number variation']	nstd37

```

## 17      3739972      nsv532611 ['copy number variation'] nstd101
## 18      3738955      nsv531594 ['copy number variation'] nstd101
## 19      3738954      nsv531593 ['copy number variation'] nstd101
## 20      3738649      nsv531288 ['copy number variation'] nstd101
## 21      1212838      nsv497563 ['copy number variation'] nstd37
## 22      1137112      [] nstd37
##      clinical_assertion Chr_1      assembly1 Chr_2
## 1      ['Pathogenic'] 19 GRCh37:260911-4788357 19
## 2      ['Pathogenic'] 19 NCBI36:184565-4650484 19
## 3      ['Pathogenic'] 19 NCBI36:210395-6746622 19
## 4      ['Pathogenic'] 19 NCBI36:3959558-4714171 19
## 5      ['Pathogenic'] 19 NCBI36:3505633-4641977 19
## 6      ['Pathogenic'] 19 NCBI36:1923244-9620555 19
## 7      ['Pathogenic'] 19 GRCh37:3076808-4796782 19
## 8      ['Pathogenic'] 19 GRCh37:3338022-4833151 19
## 9      ['Pathogenic'] 19 GRCh37:68029-59110290 19
## 10     ['Pathogenic'] 19 GRCh37:260912-59097160 19
## 11     ['Pathogenic'] 19 GRCh37:260912-58956888 19
## 12     ['Pathogenic'] 19 GRCh37:3120160-9732820 19
## 13     ['Pathogenic'] 19 GRCh37:260912-58956888 19
## 14     [] 19 GRCh37:260912-58956888 19
## 15     [] 19 GRCh37:260912-58956888 19
## 16     ['Pathogenic'] 19 NCBI36:210395-6746622 19
## 17     ['Pathogenic'] 19 NCBI36:3505633-4641977 19
## 18     ['Pathogenic'] 19 NCBI36:3959558-4714171 19
## 19     ['Pathogenic'] 19 NCBI36:1923244-9620555 19
## 20     ['Pathogenic'] 19 NCBI36:184565-4650484 19
## 21     ['Pathogenic'] 19 GRCh37:3338022-4833151 19
## 22     [] 19 GRCh37:3338022-4833151 19
##      assembly2
## 1      GRCh38.p12:260911-4788345
## 2      GRCh37.p13:233565-4699484
## 3      GRCh37.p13:259395-6795622
## 4      GRCh37.p13:4008558-4763171
## 5      GRCh37.p13:3554633-4690977
## 6      GRCh37.p13:1972244-9759555
## 7      GRCh38.p12:3076810-4796770
## 8      GRCh38.p12:3338024-4833139
## 9      GRCh38.p12:68029-58598923
## 10     GRCh38.p12:260912-58585793
## 11     GRCh38.p12:260912-58445521
## 12     GRCh38.p12:3120162-9622144
## 13     GRCh38.p12:260912-58445521
## 14     GRCh38.p12:260912-58445521
## 15     GRCh38.p12:260912-58445521
## 16     GRCh37.p13:259395-6795622
## 17     GRCh37.p13:3554633-4690977
## 18     GRCh37.p13:4008558-4763171
## 19     GRCh37.p13:1972244-9759555
## 20     GRCh37.p13:233565-4699484
## 21     GRCh38.p12:3338024-4833139
## 22     GRCh38.p12:3338024-4833139

```

```
### dataframe to convert
```

```
CNVdbVar <- Gviz_table("results-CNV-dbVAR.csv")
```

```
CNVdbVar
```

```
##      CNV_variant_id  chr   start      end variant_region_id study_ID
## 1      49623411 chr19  260911  4788357      nsv4457776  nstd102
## 2      49353191 chr19  233565  4699484      nsv4358278  nstd102
## 3      49353005 chr19  259395  6795622      nsv4358092  nstd102
## 4      49350830 chr19  4008558  4763171      nsv4355917  nstd102
## 5      49349701 chr19  3554633  4690977      nsv4354788  nstd102
## 6      49349293 chr19  1972244  9759555      nsv4354380  nstd102
## 7      49345450 chr19  3076808  4796782      nsv4350537  nstd102
## 8      49344315 chr19  3338022  4833151      nsv4349402  nstd102
## 9      48468240 chr19    68029 59110290      nsv3904885  nstd102
## 10     48466558 chr19  260912 59097160      nsv3903203  nstd102
## 11     48466447 chr19  260912 58956888      nsv3903092  nstd102
## 12     48453939 chr19  3120160  9732820      nsv3890584  nstd102
## 13     45807136 chr19  260912 58956888      nsv2779094  nstd37
## 14     17813982 chr19  260912 58956888      nstd102
## 15     17813734 chr19  260912 58956888      nstd101
## 16       3740775 chr19  259395  6795622      nsv533414  nstd37
## 17       3739972 chr19  3554633  4690977      nsv532611  nstd101
## 18       3738955 chr19  4008558  4763171      nsv531594  nstd101
## 19       3738954 chr19  1972244  9759555      nsv531593  nstd101
## 20       3738649 chr19  233565  4699484      nsv531288  nstd101
## 21       1212838 chr19  3338022  4833151      nsv497563  nstd37
## 22       1137112 chr19  3338022  4833151      nstd37
```

```
### reduce lengths of CNVs, so features/ids can be visualized
```

```
CNVdbVar$start[as.numeric(CNVdbVar$start)<4669000] <- 4669000
```

```
CNVdbVar$end[as.numeric(CNVdbVar$end)>4724100] <- 4724100
```

```
### convert dataframe CNVdbVar to GRanges Object
```

```
CNVdbVar_GR <- makeGRangesFromDataFrame(CNVdbVar, keep.extra.columns=TRUE)
```

```
CNVdbVar_GR
```

```
## GRanges object with 22 ranges and 3 metadata columns:
```

```
##      seqnames      ranges strand | CNV_variant_id variant_region_id
##      <Rle>      <IRanges> <Rle> |      <integer>      <factor>
## [1] chr19 4669000-4724100      * |      49623411      nsv4457776
## [2] chr19 4669000-4699484      * |      49353191      nsv4358278
## [3] chr19 4669000-4724100      * |      49353005      nsv4358092
## [4] chr19 4669000-4724100      * |      49350830      nsv4355917
## [5] chr19 4669000-4690977      * |      49349701      nsv4354788
## ...      ...      ...      ...      ...
## [18] chr19 4669000-4724100      * |      3738955      nsv531594
## [19] chr19 4669000-4724100      * |      3738954      nsv531593
## [20] chr19 4669000-4699484      * |      3738649      nsv531288
## [21] chr19 4669000-4724100      * |      1212838      nsv497563
## [22] chr19 4669000-4724100      * |      1137112
##      study_ID
##      <factor>
## [1] nstd102
```

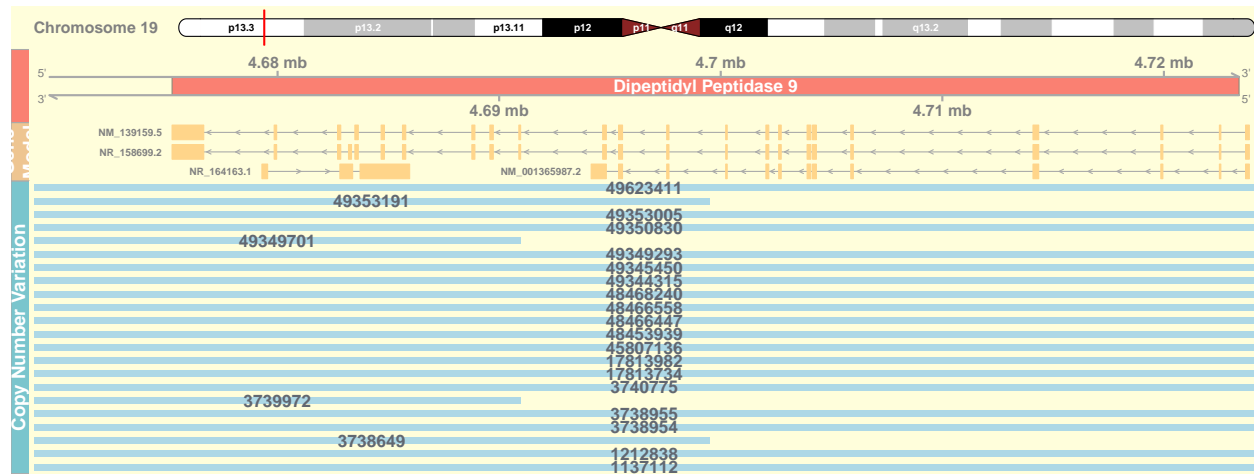
```
##      [2]  nstd102
##      [3]  nstd102
##      [4]  nstd102
##      [5]  nstd102
##      ...      ...
##     [18]  nstd101
##     [19]  nstd101
##     [20]  nstd101
##     [21]   nstd37
##     [22]   nstd37
##  -----
##  seqinfo: 1 sequence from an unspecified genome; no seqlengths
```

Gene Model with CNVs - DPP9

Show the figure in new window. Variant ids are shown.

```
chr1 <- as.character(unique(seqnames(CNVdbVar_GR)))
itrack1 <- IdeogramTrack(genome = "hg19", chromosome = chr1,
                        background.panel = "#FFFEDB",
                        background.title = "#FFFEDB")
GeneModel <- read.csv(file="results-transcripts-UCSC.csv", header = TRUE, sep=";")
grtrack1 <- GeneRegionTrack(GeneModel, genome = "hg19", chromosome = "chr19",
                           name = "Gene Model",
                           transcriptAnnotation = "symbol",
                           background.title = "#EEC591",
                           background.panel = "#FFFEDB",
                           col.border.title = "dark gray",
                           cex.title = 1.1, showId = TRUE)
gtrack1 <- GenomeAxisTrack(background.panel = "#FFFEDB",
                           background.title = "#FA8072",
                           range=IRanges(start = 4675239,
                                           end = 4723855,
                                           names = "Dipeptidyl Peptidase 9"),
                           col.border.title = "dark gray", showId = TRUE)
atrack1 <- AnnotationTrack(CNVdbVar_GR,
                           name = "Copy Number Variation",
                           background.title = "#7AC5CD",
                           background.panel = "#FFFEDB",
                           col.border.title = "dark gray",
                           cex.title = 1.1,
                           feature = CNVdbVar$CNV_variant_id,
                           showFeatureId = T, cex.feature = 1,
                           fontcolor.feature = "#616771")
plotTracks(list(itrack1,gtrack1,grtrack1,atrack1),
           from = 4669000, to = 4724100,
           col = NULL,
           add53 = TRUE, add35 = TRUE,
           cex = 1.1,
           cex.id = 1.2,
           col.id = "white",
           fill.range = "#FA8072",
           showBandId = TRUE, cex.bands = 0.7,
```

```
fontface = 2, title.width = 0.3, sizes = c(0.3,0.5,0.4,2),
margin = 0)
```



Documentation to create nice plots

Usage: <https://www.bioconductor.org/packages/devel/bioc/vignettes/Gviz/inst/doc/Gviz.html>

Color Chart: <https://github.com/EarlGlynn/colorchart/wiki/Color-Chart-in-R>