

# Gviz visualization: Copy Number Variations

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## Installation of BiocManager and Gviz (= Bioconductor package)

```
# Install BiocManager if not yet installed
# Install Gviz package if not yet installed
if (!requireNamespace("BiocManager", quietly = TRUE)){
  install.packages("BiocManager")}
if (! requireNamespace("Gviz", quietly = TRUE)) {
  BiocManager::install("Gviz")
}
# Load BiocManager
# library(BiocManager)
```

## Loading Gviz package

```
# Load Gviz package
library(Gviz) # Version 1.30.3
# Loading required package: S4Vectors
# Loading required package: stats4
# Loading required package: BiocGenerics
# Loading required package: parallel
# Loading required package: IRanges
# Loading required package: GenomicRanges
# Loading required package: GenomeInfoDb
# Loading required package: grid

# Set working directory (path/to/folder)
setwd("C:/Users/irisr/Desktop/CSV")
```

## Function to create a table with GRCh37 coordinates

```
## separate function requires tidyr package
library(tidyr)
Gviz_table <- function(filename) {
  X <- read.csv(file=filename, header = TRUE, sep=";")
  ##### GRCh37 coordinates in first assembly
  tablepart1 <- X[grep("^GRCh37", X$assembly1),]
  id1 <- tablepart1[1]
```

```

if(any(names(tablepart1) == 'variant_region_id')){regionid1 <- tablepart1[2]}
if(any(names(tablepart1) == 'study_ID')){studyid1 <- tablepart1[4]}
Chr_1a <- tablepart1[["Chr_1"]]
Chr_1a <- gsub("[A-Za-z]", "", Chr_1a)
Chr_1a <- sub("", "chr", Chr_1a)
assembly1 <- tablepart1[["assembly1"]]
assembly1 <- gsub(".*?:", "", assembly1)
assembly1 <- separate(data = as.data.frame(assembly1), col = assembly1,
                      into = c("start", "end"), sep = "-")
if(any(names(tablepart1) == 'variant_region_id') &
    any(names(tablepart1) == 'study_ID')){
  GRCh37_assembly1 <- data.frame(id1,Chr_1a,assembly1,regionid1,studyid1)
} else {
  GRCh37_assembly1 <- data.frame(id1,Chr_1a,assembly1)
}
names(GRCh37_assembly1)[names(GRCh37_assembly1) == "assembly1"] <- "assembly"
names(GRCh37_assembly1)[names(GRCh37_assembly1) == "Chr_1a"] <- "chr"
#GRCh37_assembly1["genome"] <- "hg19"
##### GRCh37 coordinates in second assembly
tablepart2 <- X[grep("^GRCh37", X$assembly2),]
id2 <- tablepart2[1]
if(any(names(tablepart2) == 'variant_region_id')){regionid2 <- tablepart2[2]}
if(any(names(tablepart2) == 'study_ID')){studyid2 <- tablepart2[4]}
Chr_1b <- tablepart2[["Chr_2"]]
Chr_1b <- gsub("[A-Za-z]", "", Chr_1b)
Chr_1b <- sub("", "chr", Chr_1b)
assembly2 <- tablepart2[["assembly2"]]
assembly2 <- gsub(".*?:", "", assembly2)
assembly2 <- separate(data = as.data.frame(assembly2), col = assembly2,
                      into = c("start", "end"), sep = "-")
if(any(names(tablepart2) == 'variant_region_id') &
    any(names(tablepart2) == 'study_ID')){
  GRCh37_assembly2 <- data.frame(id2,Chr_1b,assembly2,regionid2,studyid2)
} else {
  GRCh37_assembly2 <- data.frame(id2,Chr_1b,assembly2)
}
names(GRCh37_assembly2)[names(GRCh37_assembly2) == "assembly2"] <- "assembly"
names(GRCh37_assembly2)[names(GRCh37_assembly2) == "Chr_1b"] <- "chr"
#GRCh37_assembly2["genome"] <- "hg19"
##### Gviz table
GRCh37_Gviz <- rbind(GRCh37_assembly1,GRCh37_assembly2)
GRCh37_Gviz <- GRCh37_Gviz[order(-GRCh37_Gviz[1]),]
}

```

## Input tables and GRanges Object

```

### results-CNV-dbVar.csv
csvfile <- read.csv(file="results-CNV-dbVAR.csv", header = TRUE, sep=";")
csvfile

```

```

##      CNV_variant_id variant_region_id                                type study_ID

```

```

## 1      49623411      nsv4457776 ['copy number variation'] nstd102
## 2      49353191      nsv4358278 ['copy number variation'] nstd102
## 3      49353005      nsv4358092 ['copy number variation'] nstd102
## 4      49350830      nsv4355917 ['copy number variation'] nstd102
## 5      49349701      nsv4354788 ['copy number variation'] nstd102
## 6      49349293      nsv4354380 ['copy number variation'] nstd102
## 7      49345450      nsv4350537 ['copy number variation'] nstd102
## 8      49344315      nsv4349402 ['copy number variation'] nstd102
## 9      48468240      nsv3904885 ['copy number variation'] nstd102
## 10     48466558      nsv3903203 ['copy number variation'] nstd102
## 11     48466447      nsv3903092 ['copy number variation'] nstd102
## 12     48453939      nsv3890584 ['copy number variation'] nstd102
## 13     45807136      nsv2779094 ['copy number variation'] nstd37
## 14     17813982      [] nstd102
## 15     17813734      [] nstd101
## 16      3740775      nsv533414 ['copy number variation'] nstd37
## 17      3739972      nsv532611 ['copy number variation'] nstd101
## 18      3738955      nsv531594 ['copy number variation'] nstd101
## 19      3738954      nsv531593 ['copy number variation'] nstd101
## 20      3738649      nsv531288 ['copy number variation'] nstd101
## 21      1212838      nsv497563 ['copy number variation'] nstd37
## 22      1137112      [] nstd37
##      clinical_assertion Chr_1      assembly1 Chr_2
## 1      ['Pathogenic'] 19 GRCh37:260911-4788357 19
## 2      ['Pathogenic'] 19 NCBI36:184565-4650484 19
## 3      ['Pathogenic'] 19 NCBI36:210395-6746622 19
## 4      ['Pathogenic'] 19 NCBI36:3959558-4714171 19
## 5      ['Pathogenic'] 19 NCBI36:3505633-4641977 19
## 6      ['Pathogenic'] 19 NCBI36:1923244-9620555 19
## 7      ['Pathogenic'] 19 GRCh37:3076808-4796782 19
## 8      ['Pathogenic'] 19 GRCh37:3338022-4833151 19
## 9      ['Pathogenic'] 19 GRCh37:68029-59110290 19
## 10     ['Pathogenic'] 19 GRCh37:260912-59097160 19
## 11     ['Pathogenic'] 19 GRCh37:260912-58956888 19
## 12     ['Pathogenic'] 19 GRCh37:3120160-9732820 19
## 13     ['Pathogenic'] 19 GRCh37:260912-58956888 19
## 14      [] 19 GRCh37:260912-58956888 19
## 15      [] 19 GRCh37:260912-58956888 19
## 16     ['Pathogenic'] 19 NCBI36:210395-6746622 19
## 17     ['Pathogenic'] 19 NCBI36:3505633-4641977 19
## 18     ['Pathogenic'] 19 NCBI36:3959558-4714171 19
## 19     ['Pathogenic'] 19 NCBI36:1923244-9620555 19
## 20     ['Pathogenic'] 19 NCBI36:184565-4650484 19
## 21     ['Pathogenic'] 19 GRCh37:3338022-4833151 19
## 22      [] 19 GRCh37:3338022-4833151 19
##      assembly2
## 1      GRCh38.p12:260911-4788345
## 2      GRCh37.p13:233565-4699484
## 3      GRCh37.p13:259395-6795622
## 4      GRCh37.p13:4008558-4763171
## 5      GRCh37.p13:3554633-4690977
## 6      GRCh37.p13:1972244-9759555
## 7      GRCh38.p12:3076810-4796770
## 8      GRCh38.p12:3338024-4833139

```

```
## 9   GRCh38.p12:68029-58598923
## 10  GRCh38.p12:260912-58585793
## 11  GRCh38.p12:260912-58445521
## 12  GRCh38.p12:3120162-9622144
## 13  GRCh38.p12:260912-58445521
## 14  GRCh38.p12:260912-58445521
## 15  GRCh38.p12:260912-58445521
## 16  GRCh37.p13:259395-6795622
## 17  GRCh37.p13:3554633-4690977
## 18  GRCh37.p13:4008558-4763171
## 19  GRCh37.p13:1972244-9759555
## 20  GRCh37.p13:233565-4699484
## 21  GRCh38.p12:3338024-4833139
## 22  GRCh38.p12:3338024-4833139
```

```
### dataframe to convert
```

```
CNVdbVar <- Gviz_table("results-CNV-dbVAR.csv")
CNVdbVar
```

##	CNV_variant_id	chr	start	end	variant_region_id	study_ID
## 1	49623411	chr19	260911	4788357	nsv4457776	nstd102
## 2	49353191	chr19	233565	4699484	nsv4358278	nstd102
## 3	49353005	chr19	259395	6795622	nsv4358092	nstd102
## 4	49350830	chr19	4008558	4763171	nsv4355917	nstd102
## 5	49349701	chr19	3554633	4690977	nsv4354788	nstd102
## 6	49349293	chr19	1972244	9759555	nsv4354380	nstd102
## 7	49345450	chr19	3076808	4796782	nsv4350537	nstd102
## 8	49344315	chr19	3338022	4833151	nsv4349402	nstd102
## 9	48468240	chr19	68029	59110290	nsv3904885	nstd102
## 10	48466558	chr19	260912	59097160	nsv3903203	nstd102
## 11	48466447	chr19	260912	58956888	nsv3903092	nstd102
## 12	48453939	chr19	3120160	9732820	nsv3890584	nstd102
## 13	45807136	chr19	260912	58956888	nsv2779094	nstd37
## 14	17813982	chr19	260912	58956888		nstd102
## 15	17813734	chr19	260912	58956888		nstd101
## 16	3740775	chr19	259395	6795622	nsv533414	nstd37
## 17	3739972	chr19	3554633	4690977	nsv532611	nstd101
## 18	3738955	chr19	4008558	4763171	nsv531594	nstd101
## 19	3738954	chr19	1972244	9759555	nsv531593	nstd101
## 20	3738649	chr19	233565	4699484	nsv531288	nstd101
## 21	1212838	chr19	3338022	4833151	nsv497563	nstd37
## 22	1137112	chr19	3338022	4833151		nstd37

```
### reduce lengths of CNVs, so features/ids can be visualized
```

```
CNVdbVar$start[as.numeric(CNVdbVar$start)<4669000] <- 4669000
CNVdbVar$end[as.numeric(CNVdbVar$end)>4724100] <- 4724100
```

```
### convert dataframe CNVdbVar to GRanges Object
```

```
CNVdbVar_GR <- makeGRangesFromDataFrame(CNVdbVar, keep.extra.columns=TRUE)
CNVdbVar_GR
```

```
## GRanges object with 22 ranges and 3 metadata columns:
```

```
##      seqnames      ranges strand | CNV_variant_id variant_region_id
```

```
##           <Rle>           <IRanges> <Rle> |           <integer>           <factor>
## [1] chr19 4669000-4724100 * | 49623411 nsv4457776
## [2] chr19 4669000-4699484 * | 49353191 nsv4358278
## [3] chr19 4669000-4724100 * | 49353005 nsv4358092
## [4] chr19 4669000-4724100 * | 49350830 nsv4355917
## [5] chr19 4669000-4690977 * | 49349701 nsv4354788
## ...      ...      ...      ...      ...
## [18] chr19 4669000-4724100 * | 3738955 nsv531594
## [19] chr19 4669000-4724100 * | 3738954 nsv531593
## [20] chr19 4669000-4699484 * | 3738649 nsv531288
## [21] chr19 4669000-4724100 * | 1212838 nsv497563
## [22] chr19 4669000-4724100 * | 1137112
##      study_ID
##      <factor>
## [1] nstd102
## [2] nstd102
## [3] nstd102
## [4] nstd102
## [5] nstd102
## ...      ...
## [18] nstd101
## [19] nstd101
## [20] nstd101
## [21] nstd37
## [22] nstd37
## -----
## seqinfo: 1 sequence from an unspecified genome; no seqlengths
```

## Gene Model with CNVs - DPP9

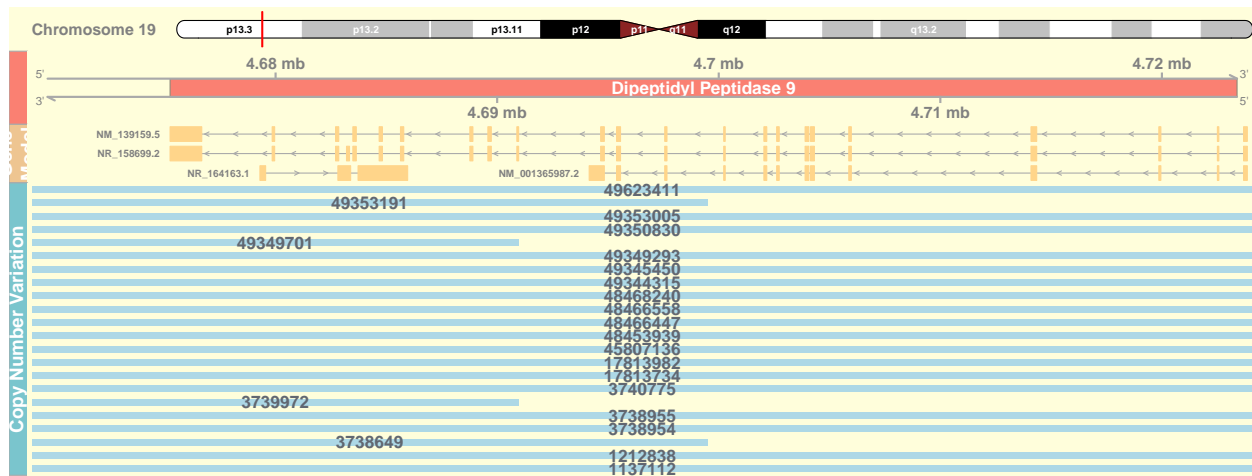
Show the figure in new window. Variant ids are shown.

```
chr1 <- as.character(unique(seqnames(CNVdbVar_GR)))
itrack1 <- IdeogramTrack(genome = "hg19", chromosome = chr1,
                        background.panel = "#FFFEDB",
                        background.title = "#FFFEDB")
GeneModel <- read.csv(file="results-transcripts-UCSC.csv", header = TRUE, sep=";")
grtrack1 <- GeneRegionTrack(GeneModel, genome = "hg19", chromosome = "chr19",
                           name = "Gene Model",
                           transcriptAnnotation = "symbol",
                           background.title = "#EEC591",
                           background.panel = "#FFFEDB",
                           col.border.title = "dark gray",
                           cex.title = 1.1, showId = TRUE)
gtrack1 <- GenomeAxisTrack(background.panel = "#FFFEDB",
                           background.title = "#FA8072",
                           range=IRanges(start = 4675239,
                                           end = 4723855,
                                           names = "Dipeptidyl Peptidase 9"),
                           col.border.title = "dark gray", showId = TRUE)
atrack1 <- AnnotationTrack(CNVdbVar_GR,
                           name = "Copy Number Variation",
                           background.title = "#7AC5CD",
```

```

background.panel = "#FFEDB",
col.border.title = "dark gray",
cex.title = 1.1,
feature = CNVdbVar$CNV_variant_id,
showFeatureId = T, cex.feature = 1,
fontcolor.feature = "#616771")
plotTracks(list(itrack1,gtrack1,grtrack1,atrack1),
  from = 4669000, to = 4724100,
  col = NULL,
  add53 = TRUE, add35 = TRUE,
  cex = 1.1,
  cex.id = 1.2,
  col.id = "white",
  fill.range = "#FA8072",
  showBandId = TRUE, cex.bands = 0.7,
  fontface = 2, title.width = 0.3, sizes = c(0.3,0.5,0.4,2),
  margin = 0)

```



## Documentation to create nice plots

Usage: <https://www.bioconductor.org/packages/devel/bioc/vignettes/Gviz/inst/doc/Gviz.html>

Color Chart: <https://github.com/EarlGlynn/colorchart/wiki/Color-Chart-in-R>