

Building a sequence aligner capable of affine gap penalties.

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October 2018

Introduction

Protein-protein sequence alignment is the basis and first step of many bioinformatical methods. In 1970, the first draft of the Needleman-Wunsch algorithm for global sequence alignment was first introduced [1]. This was a simple algorithm, with no gap penalties (only rewards or penalties for matches or mismatches), and cubic complexity. Through the years, many great additions have come to this first basis for sequence alignment, including gap penalties, *affine gap penalties*, a simplification to quadratic complexity, and the option for finding the best local alignments with the Smith-Waterman algorithm [2].

In this short project, an aligner capable of finding both global and local alignments while using the affine gap penalty is implemented and briefly analyzed.

Theory

Presented below are the respective algorithms for global and local alignment using affine gap penalties.

Needleman-Wunsch with Affine Gap Penalty

Input:

Sequences A and B to be aligned.

Variables:

$$i \in \{1, 2, \dots, n\}$$

$$j \in \{1, 2, \dots, m\}$$

$$M(i, j)$$

$$G_x(i, j)$$

$$G_y(i, j)$$

where n and m are the lengths of sequences A and B , respectively.

Initialization:

$$\begin{aligned}
M(0, 0) &= 0 \\
G_x(i, 0) &= h + g \times i \\
G_y(0, j) &= h + g \times j \\
\text{all other cells: } &-\infty
\end{aligned}$$

where h is the penalty for opening a new gap and g is the penalty for extending a gap.

Calculations:

For all i and j , do:

$$M(i, j) = \max \begin{cases} M(i-1, j-1) + s(x_i, y_j), \\ G_x(i-1, j-1) + s(x_i, y_j), \\ G_y(i-1, j-1) + s(x_i, y_j) \end{cases} \quad (1)$$

$$G_x(i, j) = \max \begin{cases} M(i-1, j) + h + g, \\ G_x(i-1, j) + g \end{cases} \quad (2)$$

$$G_y(i, j) = \max \begin{cases} M(i, j-1) + h + g, \\ G_y(i, j-1) + g \end{cases} \quad (3)$$

where s is the scoring matrix used, x_i is the amino acid at position i in sequence A , and y_j is the amino acid at position j in sequence B .

Traceback

Start at $\max(M(n, m), G_x(n, m), G_y(n, m))$ and follow pointers back to $M(0, 0)$, $G_x(0, 0)$, or $G_y(0, 0)$, introducing gaps in A and B when moving in G_x or G_y respectively, and matches when moving in M .

Smith-Waterman with Affine Gap Penalty**Input:**

Sequences A and B to be aligned.

Variables:

$$\begin{aligned}
i &\in \{1, 2, \dots, n\} \\
j &\in \{1, 2, \dots, m\} \\
M(i, j) \\
G_x(i, j) \\
G_y(i, j)
\end{aligned}$$

where n and m are the lengths of sequences A and B , respectively.

Initialization:

$$\begin{aligned}
M(0, 0) &= 0 \\
G_x(i, 0) &= \max(h + g \times i, 0) \\
G_y(0, j) &= \max(h + g \times j, 0) \\
\text{all other cells: } &-\infty
\end{aligned}$$

where h is the penalty for opening a new gap and g is the penalty for extending a gap.

Calculations:

For all i and j , do:

$$M(i, j) = \max \begin{cases} M(i-1, j-1) + s(x_i, y_j), \\ G_x(i-1, j-1) + s(x_i, y_j), \\ G_y(i-1, j-1) + s(x_i, y_j), \\ 0 \end{cases} \quad (4)$$

$$G_x(i, j) = \max \begin{cases} M(i-1, j) + h + g, \\ G_x(i-1, j) + g \end{cases} \quad (5)$$

$$G_y(i, j) = \max \begin{cases} M(i, j-1) + h + g, \\ G_y(i, j-1) + g \end{cases} \quad (6)$$

where s is the scoring matrix used, x_i is the amino acid at position i in sequence A , and y_j is the amino acid at position j in sequence B .

Traceback

Start at largest $M(i, j)$ and follow pointers back to any of $M(0, 0) = 0$, introducing gaps in A and B when moving in G_x or G_y respectively, and matches when moving in M .

EMBOSS Implementation

Commonly available implementations of these algorithms are available via the EMBOSS software package [3]. The EMBOSS implementation follow slightly different rules however;

- When opening a gap, there is no gap extension penalty added to the first gap.
- In the cases that a sequence starts with or ends in a gap, no gap opening penalty is added to the starting and/or ending gap.

Method

Both Needleman-Wunsch and Smith-Waterman with affine gap penalties were included in the final script created. Additionally, for ease of comparison, an option was included to run the script with the EMBOSS scoring method. This script was called *aligner.py*.

Some light analysis of time efficiency and method complexity was performed, optimizing the script further. Details can be found in the *Project_supplementary.ipynb* jupyter notebook. The optimized script is called *aligner_optimized.py*. Both instances of the script was kept separately for ease of comparison.

Results

The resulting aligner is complexity $O(nm)$, as derived from tests in *Project_supplementary.ipynb*, and can manage many different kinds of alignments, as described by its help statement:

```
usage: aligner_optimized.py [-h] [-g GAP] [-o OPENGAP] [-e] [-l]
                             [-m MATRIX] [-c CHARBREAK]
A B
```

Aligns sequence A and sequence B using affine gap penalties. If you want to align with regular gap penalties, set the `--opengap` flag to 0.

positional arguments:

A	Sequence A
B	Sequence B

optional arguments:

-h, --help	show this help message and exit
-g GAP, --gap GAP	Gap extension penalty (should most often be a negative number).
-o OPENGAP, --opengap OPENGAP	Gap opening penalty (should most often be a negative number).
-e, --emboss	Use EMBOSS rules for alignment, meaning you don't receive gap extension when opening a gap, and having gaps at the end or beginning of a sequence does not count as opening a gap.
-l, --local	Make a local alignment rather than a global.
-m MATRIX, --matrix MATRIX	Which scoring matrix to use. The matrix should be saved as a python dictionary in plain-text. Default behaviour is using the inbuilt PAM250.
-c CHARBREAK, --charbreak CHARBREAK	

How many characters per row you want the output to have. Default is no linebreaks.

A quick analysis of the runtime over aligning randomized sequences of varying length support the complexity prediction, as seen in figure 1.

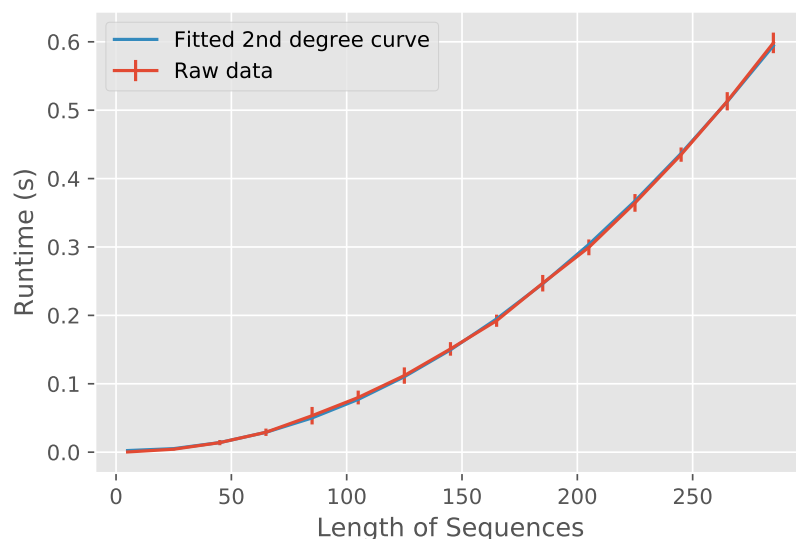


Figure 1: The time for the aligner to finish plotted against the length of the sequences it's tasked to align. All measurements were made with PAM250 matrix, global alignment, open gap penalty of -5, extend gap penalty of -1, and EMBOSS mode turned off. The times are means over 100 random sequences, with the bars showing standard deviation.

Test Cases

Some examples comparing to the official EMBOSS NEEDLE[3]:

Example 1

EMBOSS:

```
#=====
#
# Aligned_sequences: 2
# 1: HBA_HUMAN
# 2: HBA_MOUSE
# Matrix: EPAM250
```

```

# Gap_penalty: 5.0
# Extend_penalty: 1.0
#
# Length: 142
# Identity:      122/142 (85.9%)
# Similarity:    136/142 (95.8%)
# Gaps:          0/142 ( 0.0%)
# Score: 609.0
#
#
#=====

HBA_HUMAN      1 MVLSPADKTNVKAAWGKVGAGHAGEYGAEALERMFSLFPTTKTYFPHFDLS      50
      ||||..||:|:|||||:|:|:|:|||||||..|||||||:|
HBA_MOUSE      1 MVLSGEDKSNIAAWGKIGGHGAEYGAEALERMFASFPTTKTYFPHFDVS      50

HBA_HUMAN     51 HGSAQVKGHGKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFK      100
      |||||||||||||||:|:|:|:|:|:|:|:|:|:|:|:|
HBA_MOUSE     51 HGSAQVKGHGKKVADALASAAGHLDDLPGALSALSDLHAHKLRVDPVNFK      100

HBA_HUMAN     101 LLSHCLLVTLAAHLP AEFTPAVHASL DKFLASVSTVLTSKYR      142
      |||||||||||:|:|:|:|||||||||||||||||
HBA_MOUSE     101 LLSHCLLVTLASHHPADFTPAVHASL DKFLASVSTVLTSKYR      142

```

Output from my own aligner, run with the same penalties (and the EMBOSS flag):

```

SCORE: 609

MVLSPADKTNVKAAWGKVGAGHAGEYGAEALERMFSLFPTTKTYFPHFDLS
::::::::::::::::::::::::::::::::::::::::::::::::::::
MVLSGEDKSNIAAWGKIGGHGAEYGAEALERMFASFPTTKTYFPHFDVS

HGSAQVKGHGKKVADALTNAVAHVDDMPNALSALSDLHAHKLRVDPVNFK
::::::::::::::::::::::::::::::::::::::::::::::::::::
HGSAQVKGHGKKVADALASAAGHLDDLPGALSALSDLHAHKLRVDPVNFK

LLSHCLLVTLAAHLP AEFTPAVHASL DKFLASVSTVLTSKYR
::::::::::::::::::::::::::::::::::::::::::::::::::::
LLSHCLLVTLASHHPADFTPAVHASL DKFLASVSTVLTSKYR

```

Example 2

EMBOSS:

```

#####
# Program: needle

```

```

# Rundate: Fri 19 Oct 2018 10:41:02
# Commandline: needle
#   -auto
#   -stdout
#   -asequence emboss_needle-I20181019-104058-0584-90315017-p2m.asequence
#   -bsequence emboss_needle-I20181019-104058-0584-90315017-p2m.bsequence
#   -datafile EPAM250
#   -gapopen 5.0
#   -gapextend 1.0
#   -endopen 10.0
#   -endextend 0.5
#   -aformat3 pair
#   -sprotein1
#   -sprotein2
# Align_format: pair
# Report_file: stdout
#####

#=====
#
# Aligned_sequences: 2
# 1: FABPH_HUMAN
# 2: FABP5_MOUSE
# Matrix: EPAM250
# Gap_penalty: 5.0
# Extend_penalty: 1.0
#
# Length: 137
# Identity:      64/137 (46.7%)
# Similarity:    99/137 (72.3%)
# Gaps:          6/137 ( 4.4%)
# Score: 352.0
#
#
#=====

FABPH_HUMAN      1 M--VDAFLGTWKLVD SKNFDDYMKSLGVGFATRQVASMTKPTTIEKNGD      48
|  :...|.|:|::|.|:|::| |. | | | :|.|:|:|:| |. | | :|:
FABP5_MOUSE      1 MASLKDLEGWRLMES HGFE EYMKELGVGLALRKMAAMAKPDCIITCDGN      50

FABPH_HUMAN     49 ILTLKTHSTFKNTEISFKLGVFEFDETTADDRKVKSI VTL DGGKLVHLQKW      98
.:|:|:| |.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.
FABP5_MOUSE     51 NITVKTESTVKTTVFSCNLGEKFDETTADGRKTETVCTFQD GALVQHQQW      100

FABPH_HUMAN     99 DGQETTLVRELIDGKLIL--TLTHGTAVCTRTRYEKEA      133
| |:|:|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.|.

```

FABP5_MOUSE 101 DGKESTITRKLKDGKMIVECVMNNAT--CTRVYEKVQ 135

Output from my own aligner, run with the same penalties (and the EMBOSS flag):

SCORE: 352.0

```
M--VDAFLGTWKLVD SKNFDDYMKSLGVGFATRQVASMTKPTTIEKNGD
: .....
MASLKDLEGKWRLMESHGFEEYMKELGVGLALRKMAAMAKPDCIITCDGN
```

```
ILTLKTHSTFKNTEISF--KLGVEFDETTADDRKVKSIIVTL DGGKLVHLQ
..... : .....
NITVKTESTVKTT-V-FSCNLGEKFDETTADGRKTETVCTFQD GALVQHQ
```

```
KWDGQETTTLVRELIDGKLILT--LTHGTAVCTRTRYEKEA
..... : .....
QWDGKESTITRKLKDGKMIVECVMNNAT--CTRVYEKVQ
```

As you can see, these alignments are not identical, but do have identical scores, indicating either a fault within the aligner, or a non-unique solution to the alignment. Since the scores are identical, the latter seems more probable.

Future Work

The current implementation is in Python, and while this makes it easy to modify and read, it also makes it slow. If this implementation should have any real value, it would have to be reimplemented in a compiled language, such as C.

References

- [1] Saul B Needleman and Christian D Wunsch. A general method applicable to the search for similarities in the amino acid sequence of two proteins. *Journal of molecular biology*, 48(3):443–453, 1970.
- [2] Temple F Smith and Michael S Waterman. Comparison of biosequences. *Advances in applied mathematics*, 2(4):482–489, 1981.
- [3] I. Rice, P. Longden and A. Bleasby. Emboss: The european molecular biology open software suite. *Trends in Genetics*, 16(6):276–277, 2000.