

Complete genome sequence of *Solobacterium moorei* JCM 10645^T isolated from a human stool sample

Kota Oshibuchi,^{1,2,3} Jiayue Yang,^{1,2} Nozomu Obana,⁴ Shinji Fukuda,^{1,2,4,5,6} Kazuharu Arakawa^{1,2,7}

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT *Solobacterium moorei* JCM 10645^T is an obligately anaerobic Gram-positive bacterium that was isolated from a human stool sample, generally known as a bacterium associated with sepsis, bacteremia, halitosis, and periodontal disease. In this study, we report the complete genome sequence of this strain, which is 2.615 Mbp with a 37.2% GC content.

KEYWORDS *Solobacterium moorei*, complete genome, nanopore sequencing, genome assembly

Solobacterium moorei JCM 10645^T is a Gram-positive, bacilliform, non-spore-forming, and obligately anaerobic bacterium that has been isolated from a human stool sample by Kageyama et al. (1). This bacterium is the type strain of *Solobacterium moorei*. This species is generally known as a bacterium associated with sepsis, bacteremia, halitosis, and periodontal disease (2). Here, we analyzed the complete genome sequence of *Solobacterium moorei* JCM 10645^T.

Solobacterium moorei JCM 10645^T, obtained from the Japan Collection of Microorganisms, was cultured in GAM medium at 37°C in an anaerobic condition for 24 hours. The genomic DNA was extracted from the harvested cells using Genomic-tip 20/G (Qiagen) according to the manufacturer's protocol for both long- and short-read sequencing. A long-read sequencing library was prepared using the Rapid Barcoding Sequencing Kit (SQK-RBK004) (Oxford Nanopore Technologies) and sequenced using an R10.3 Flow-cell (FLO-MIN111) on a GridION X5 device (GridION software release 21.05.25; Oxford Nanopore Technologies). Base calling (Super Accurate Mode), demultiplexing, and adapter removal were performed using Guppy v.6.3.8 software. Assembly was performed using all sequenced long reads (641.7 Mb, 227,408 reads in total, N50: 4.643 kb) by Canu version 2.2 (3). Short-read sequencing was performed for error correction using the HyperPlus library preparation kit (Kapa Biosystems) and a MiSeq sequencer (Illumina) with Reagent Kit v3 600 cycles as 300 bp paired ends (Illumina). Using filtered reads by fastp tool (<https://github.com/OpenGene/fastp>; 4.6M out of 9.7M paired reads), error correction was then performed by aligning the reads using Burrows-Wheeler Aligner v.0.7.11 (4), and subsequently, the polished sequence was then generated using Pilon v.1.23 (5). Assembly quality was assessed using CheckM (6) on the DFAST server leading to a predicted completeness score of 100% (Erysipelotrichaceae marker lineage). The genome was annotated using the DDBJ Fast Annotation and Submission Tool (DFAST) version 1.2.18 (7). All software was used with default settings unless otherwise specified.

The assembled genome consists of one linear chromosome of 2,615,268 bp with a GC content of 37.2%, harboring 2,599 protein-coding genes, 6 rRNAs, and 43 tRNAs. Volatile sulfur compound production has been reported for this species (8), and BLASTP search for the hydrogen-sulfide-producing enzyme MegL (SwissProt: MEG_L_FUSNN) (9) identified a potential homolog by bi-directional identity ($e = 2e-103$). The availability

Editor André O. Hudson, Rochester Institute of Technology, Rochester, New York, USA

Address correspondence to Kazuharu Arakawa, gaou@sfc.keio.ac.jp.

Kota Oshibuchi is an employee of Shiseido Co. Ltd., but the company had no role in any part of the research.

See the funding table on p. 2.

Received 10 October 2023

Accepted 26 October 2023

Published 29 November 2023

Copyright © 2023 Oshibuchi et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

of the genome resource can potentially contribute to the molecular mechanisms of the association of *Solobacterium moorei* in halitosis.

ACKNOWLEDGMENTS

We thank Tomoki Takeda, Sora Ishikawa, and Yasuha Watanabe for technical support and suggestions. The sequencing and assembly were conducted in the Genome Engineering Workshop course of the Systems Biology Program, Graduate School of Media and Governance, Keio University.

This work was supported in part by research funds from the Yamagata Prefectural Government and Tsuruoka City, Japan.

AUTHOR AFFILIATIONS

¹Institute for Advanced Biosciences, Keio University, Tsuruoka, Yamagata, Japan

²Systems Biology Program, Graduate School of Media and Governance, Keio University, Fujisawa, Kanagawa, Japan

³MIRAI Technology Institute Yokohama, Yokohama, Japan

⁴Transborder Medical Research Center, Institute of Medicine, University of Tsukuba, Tsukuba, Ibaraki, Japan

⁵Gut Environmental Design Group, Kanagawa Institute of Industrial Science and Technology, Kawasaki, Kanagawa, Japan

⁶Laboratory for Regenerative Microbiology, Juntendo University Graduate School of Medicine, Bunkyo-ku, Tokyo, Japan

⁷Faculty of Environment and Information Studies, Keio University, Fujisawa, Kanagawa, Japan

AUTHOR ORCIDs

Nozomu Obana  <https://orcid.org/0000-0002-0637-7904>

Shinji Fukuda  <http://orcid.org/0000-0001-5161-9880>

Kazuharu Arakawa  <http://orcid.org/0000-0002-2893-4919>

FUNDING

Funder	Grant(s)	Author(s)
Yamagata Prefectural Government and Tsuruoka City	YPGTC2022	Shinji Fukuda Kazuharu Arakawa Jiyue Yang

AUTHOR CONTRIBUTIONS

Kota Oshibuchi, Formal analysis, Writing – original draft, Writing – review and editing, Investigation | Jiyue Yang, Resources, Writing – review and editing | Nozomu Obana, Resources, Writing – review and editing | Shinji Fukuda, Resources, Writing – review and editing | Kazuharu Arakawa, Conceptualization, Data curation, Formal analysis, Funding acquisition, Methodology, Project administration, Resources, Supervision, Validation, Writing – original draft, Writing – review and editing

DATA AVAILABILITY

The chromosome sequence reported here was deposited in DDBJ under the accession number [AP028934](#), and the raw reads were deposited in the Sequence Read Archive (SRA) under the BioProject accession number [PRJNA1013036](#) as [SRR25909846](#) and [SRR25909847](#) runs.

REFERENCES

1. Kageyama A, Benno Y. 2000. Phylogenetic and phenotypic characterization of some eubacterium-like isolates from human feces: description of *Solobacterium moorei* gen nov. Microbiol Immunol 44:223–227. <https://doi.org/10.1111/j.1348-0421.2000.tb02487.x>
2. Vancauwenberghe F, Dadamio J, Laleman I, Van Tornout M, Teughels W, Coucke W, Quirynen M. 2013. The role of *Solobacterium moorei* in oral malodour. J Breath Res 7:046006. <https://doi.org/10.1088/1752-7155/7/4/046006>
3. Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722–736. <https://doi.org/10.1101/gr.215087.116>
4. Li H, Durbin R. 2010. Fast and accurate long-read alignment with burrows-wheeler transform. Bioinformatics 26:589–595. <https://doi.org/10.1093/bioinformatics/btp698>
5. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>
6. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. Checkm: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. Genome Res 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>
7. Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34:1037–1039. <https://doi.org/10.1093/bioinformatics/btx713>
8. Tanabe S, Grenier D. 2012. Characterization of volatile sulfur compound production by *Solobacterium moorei*. Arch Oral Biol 57:1639–1643. <https://doi.org/10.1016/j.archoralbio.2012.09.011>
9. Chen Y-W, Camacho MI, Chen Y, Bhat AH, Chang C, Peluso EA, Wu C, Das A, Ton-That H, Biswas I. 2022. Genetic determinants of hydrogen sulfide biosynthesis in *Fusobacterium nucleatum* are required for bacterial fitness, antibiotic sensitivity, and virulence. mBio 13:e0193622. <https://doi.org/10.1128/mbio.01936-22>