# BioSpecimen Exchange for Neurological Disorders (BioSEND)

PREDICT-HD 3.0

**Training Webinar- PHD3** 



## **BioSEND Training Webinar Overview**

- 1. Study Reminders
- 2. Site Equipment
- 3. Biospecimen Collection Protocol
- 4. Study Visit Protocol
- 5. Kits & Samples
  - Requesting Kits
  - Labels
  - Sample Collection & Processing
  - Sample Shipment
- BioSEND Website
- 6. Contact Information



#### **Study Reminders**

#### Please remember...

- Biospecimens are limited, valuable resources.
- Standardization and quality are key!
- Reference the BioSEND Manual of Procedures as needed.
- Do not replace or supplement any kit components without first receiving approval from BioSEND.



# Site Equipment

#### The following items are to be supplied by the site:

- Personal protective equipment
- Alcohol prep pads
- Butterfly needles and hubs
- Tourniquet
- Gauze pads
- Bandages
- Sharps bin and lid

- Microcentrifuge tube rack
- Test tube rack
- Crushed ice
- Pipettes and pipette tips
- 4°C Centrifuge
- -80°C Freezer
- Dry ice

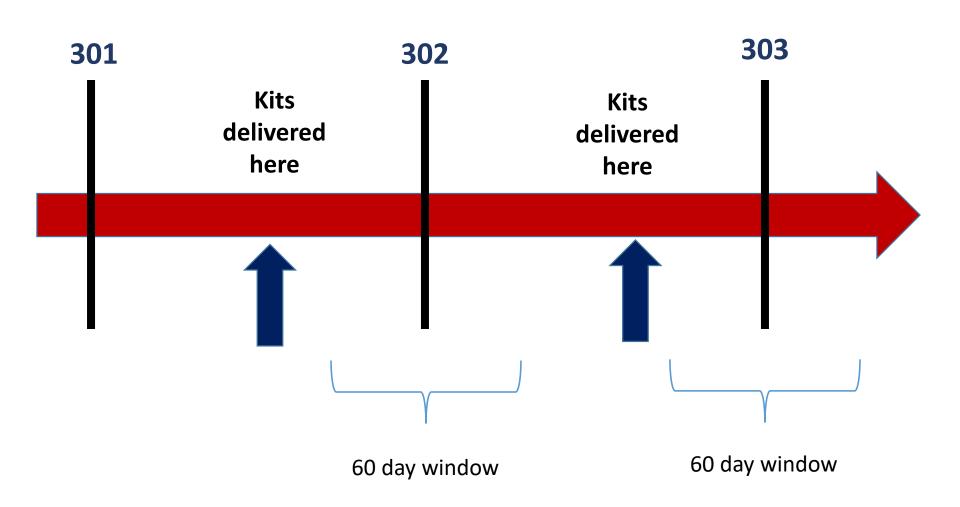


# Biospecimen Collection Protocol All specimen types FROZEN

	301 (BL)	302 (12M)	303 (24M)
Whole Blood (1 x 6ml)	X	X	X
Plasma (12 x 1.5ml)	X	X	X
Buffy Coat (4 aliquots)	X	X	X
<b>CSF</b> (13 x 1.5ml)	X	X	X



# **Study Visit Protocol**





# Study Visit Protocol

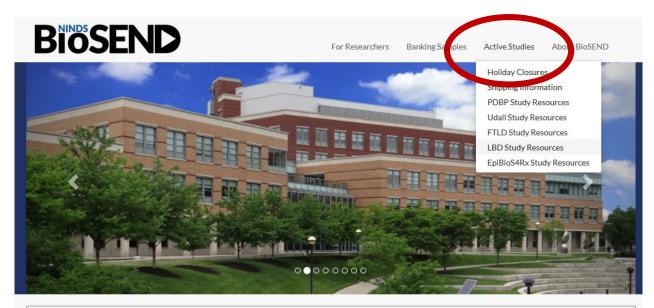
- After subject completes baseline visit, BioSEND sets up automated kit sending schedule for subject's subsequent visits
- Schedule gives 2 month window around the longitudinal study visit target (1 month on either side)
- BioSEND will send kits prior to start of study window
  - Reduces effort for study coordinators
  - Sites only need to order kits if visit will occur AHEAD of the study visit window
- All study visit target dates are determined from Baseline Visit (not from last study visit date)



# Requesting Kits



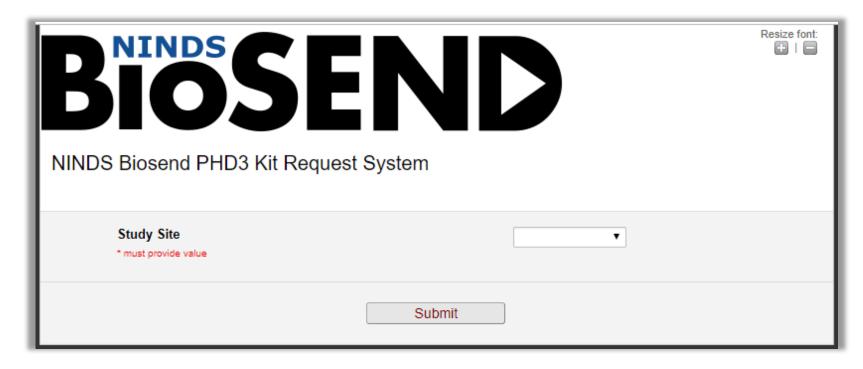
#### NINDS BioSEND Website







# BioSEND Kit Request Module

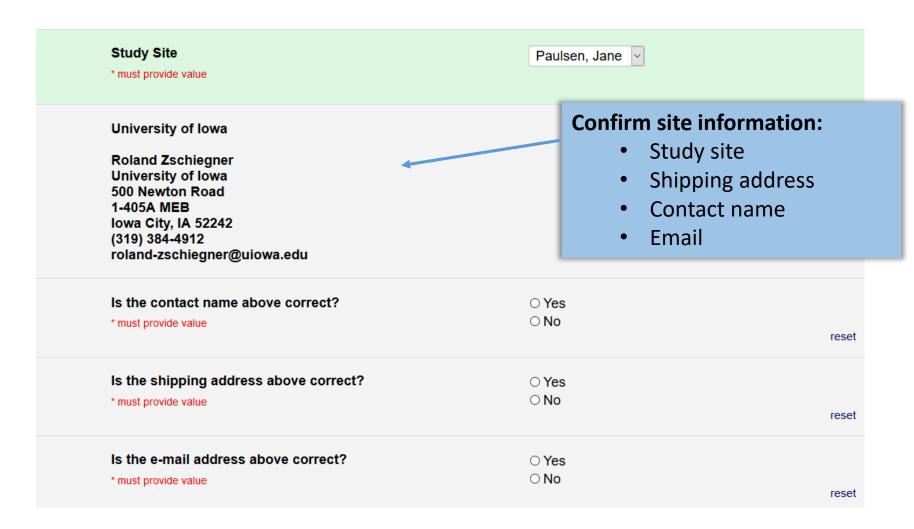


- http://kits.iu.edu/biosend/PHD3
- Choose your site PI from the drop-down list.



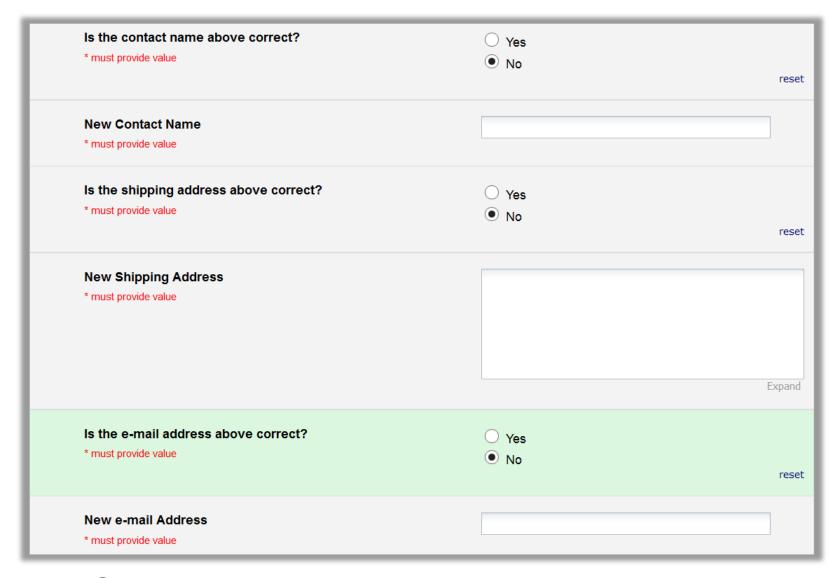
## BioSEND Kit Request Module

NINDS Biosend PHD3 Kit Request System





# BioSEND Kit Request Module





- Is the information correct?
- Provide the correct information if needed

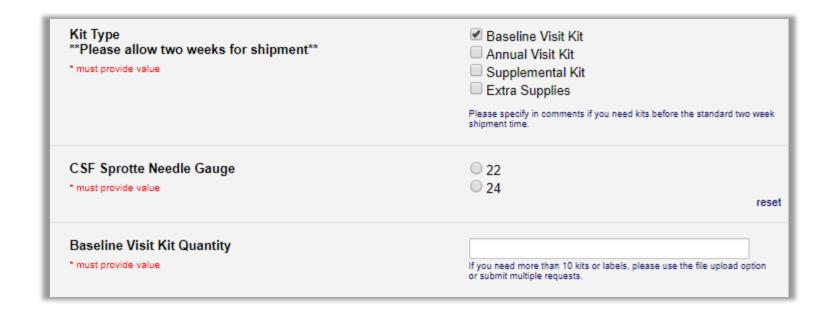
# BioSEND Kit Request Module: Kit Type



 Typically, you will only need to order kits for the Baseline visit



# BioSEND Kit Request Module: Baseline Kit

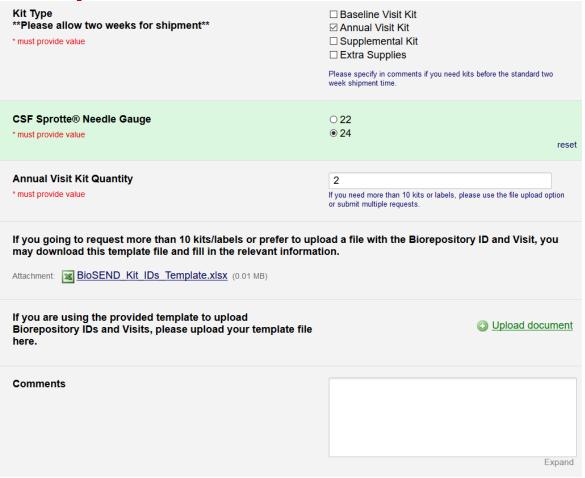


- BioSEND creates ST numbers for baseline kits
- Enter preferred CSF needle gauge
- Enter kit quantity



## BioSEND Kit Request Module: Annual Visits

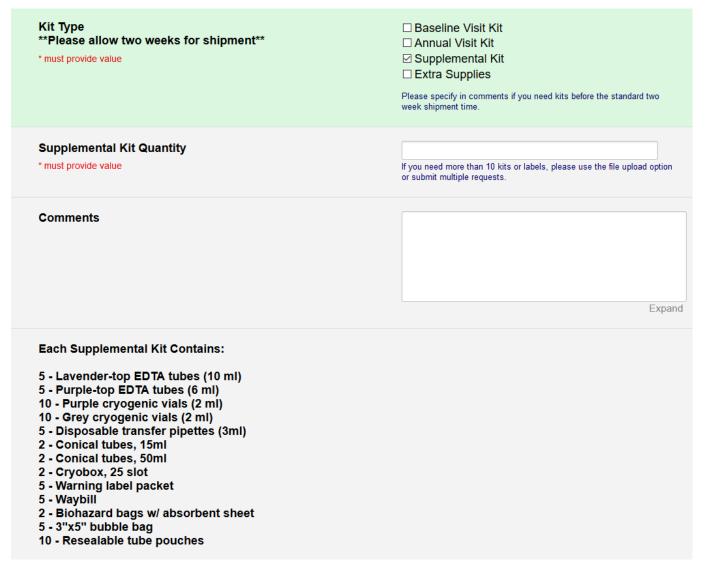
Only use this if the subject will be seen ahead of a study visit window



Input subject/visit needed in Comments so correct labels are provided



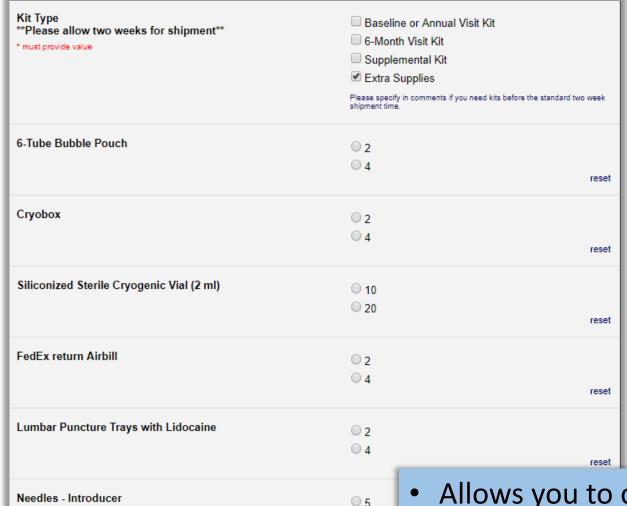
#### BioSEND Kit Request Module: Supplemental Kit





Contains a variety of extra kit pieces

#### BioSEND Kit Request Module: Extra Supplies





 Allows you to choose specific supplies and particular quantities

#### BioSEND Kit Request Module: Multiple Orders

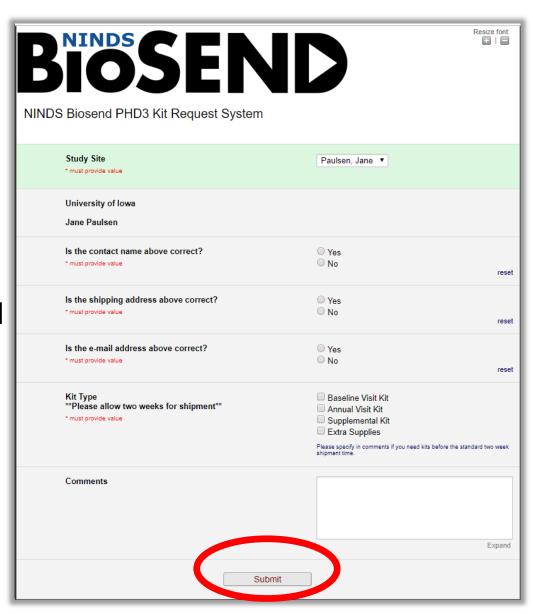


 You can order more than one type of kit in a single kit request



## BioSEND Kit Request Module: Submit

- Click "Submit" to turn in your request.
- The BioSEND staff will notify you that your request has been received and address any issues.





# Labels



# Types of Labels

Case Label

**Collection Tube Label** 

**Aliquot Label** 

ST-10012345: PHD3:301



**BioSend** 

**Identify study and Visit** 

0004468119



**BioSEND** 

ST-10001234

Baseline

Plasma

For collection tubes

BioSEND-PHD3

**PLASMA** 

ST-10001234

For cryovials; will come pre-labeled



## Case Label

ST-10012345: PHD3:301



**BioSend** 





**Biorepository Name** 



#### Case Labels

#### Case labels are placed:

- On the plastic biohazard bag of the cryovial transport box.
- On the plastic biohazard bag for the EDTA tube.
- On the lid of frozen shippers









#### **Collection Tube Label**

0004468119



**BioSEND** 

ST-10001234

Baseline

Plasma



**Biospecimen Number** 



**Subject Number** 



**Visit Type** 



**Specimen Type** 



## Aliquot Tube Label

BioSEND-PHD3

**PLASMA** 

ST-10001234

Study Abbreviation





\*Note: The tube itself with have a unique barcode printed in both 2D format (on bottom of tube) and human readable format (alongside of tube).

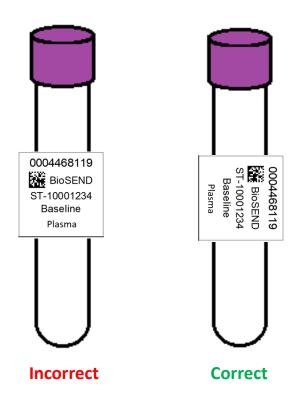


#### **Collection Tube Labels**

Collection Tube Labels are placed on:

All collection tubes

Be sure that the tube has the correct specimen type on the label





# **Aliquot Labels**

 Cryovials will be pre-labeled, but please verify that you are using the correct cryovial for the correct specimen type

Grey cryovials are used for Buffy Coat and CSF aliquots



BioSEND-PHD3 CSF ST-10001234

OR

BioSEND-PHD3 BUFFY COAT ST-10001234 Purple cryovials are used for plasma aliquots



BioSEND-PHD3 PLASMA ST-10001234



# Sample Collection & Processing

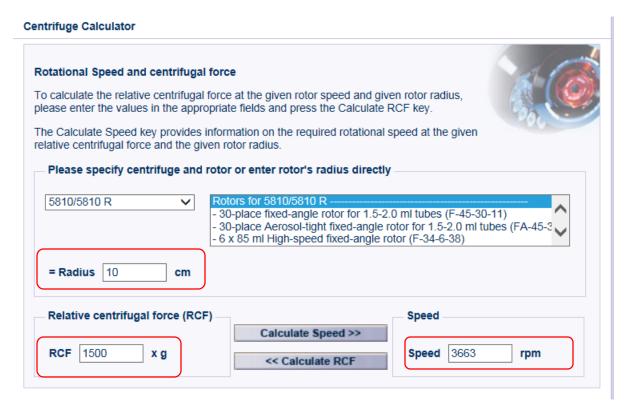
#### **Reminders:**

- Collection of biospecimen blood samples should be from subjects who have been <u>fasting for 8 hours</u> or more. If fasting is not feasible, follow suggested low-fat diet.
- G force ≠ RPM
- All specimens should be frozen and stored <u>UPRIGHT</u>
  - For RNA and whole blood specimens, please freeze samples upright in a non-styrofoam rack
  - For plasma, buffy coat, serum, and CSF aliquots, please freeze samples upright in the cryobox provided



## **Calculating Centrifugation Speed**

https://www.eppendorf.com/CA-en/centrifuge-speed-calculator/



\*The 3663 rpm speed was calculated using a hypothetical radius of 10 cm and a RCF of 1500 x g.



# Order of Specimen Collection

- 1. EDTA 6 ml (purple top) blood collection for DNA
- 2. EDTA **10 ml (lavender top)** blood collection for plasma and buffy coat

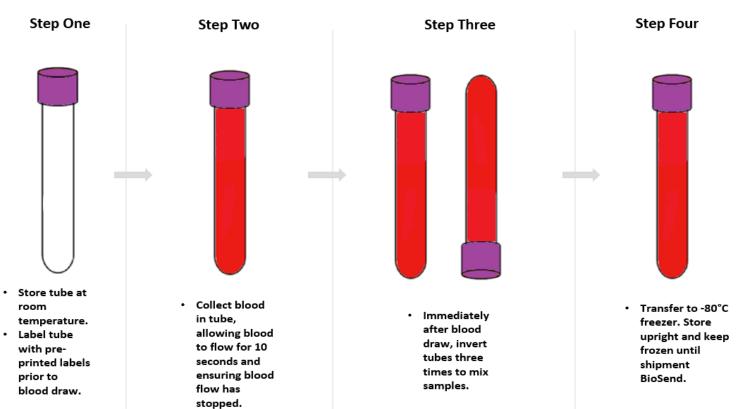






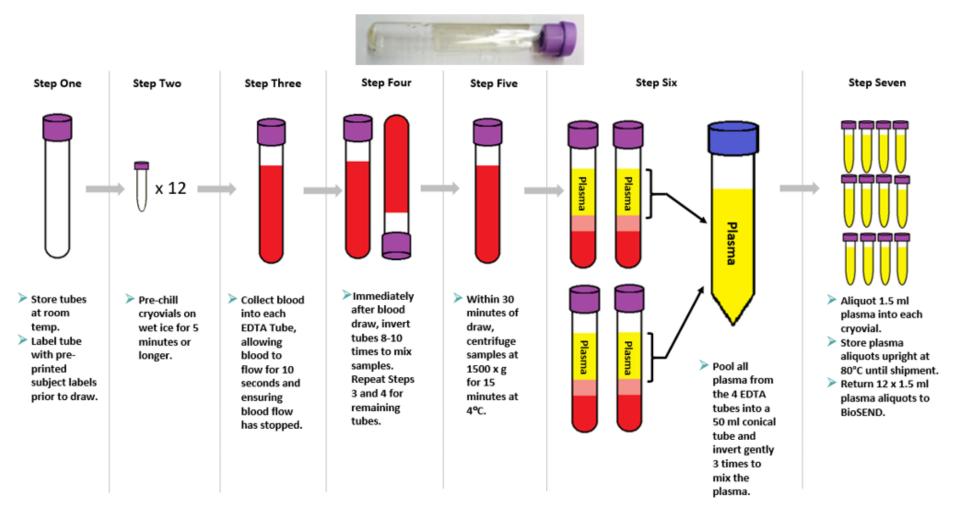
#### Whole Blood (6 ml Lavender Top Tube)





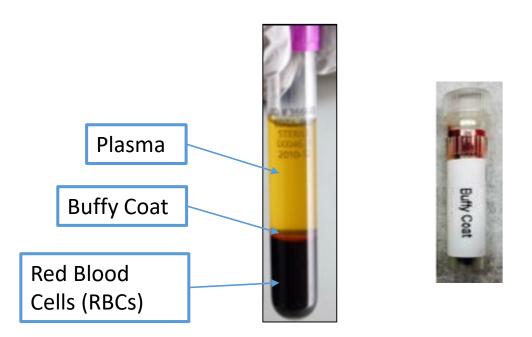


#### Plasma Preparation (10ml Purple EDTA Tubes)





#### **Buffy Coat Collection**



Collect the buffy coat layer using the transfer pipet provided. Residual plasma as well as some RBCs will be included in this collection. A buffy coat will be reddish in color due to RBCs. Freeze buffy coats upright on dry ice or -80 freezer. Store in -80 freezer until shipment to BioSEND.



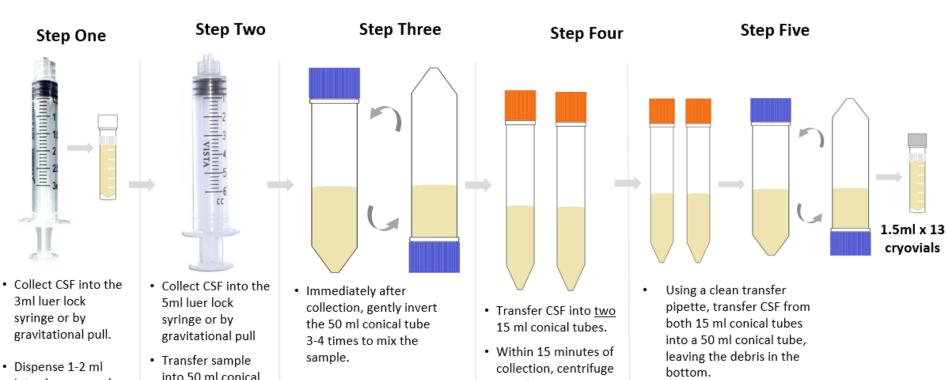
#### **Lumbar Puncture Procedure**

24g or 22g spinal needle provided in custom LP tray

- Prepare transfer and aliquot tubes (NOT in LP tray but in kit)
  - Do NOT pre-chill aliquot tubes



#### **CSF Collection and Preparation**



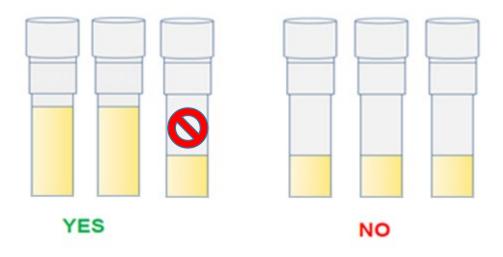
- into clear-capped cryovial.
- · Send to local lab for testing.
- into 50 ml conical tube.

- - samples at room temperature at 2000 x g for 10 minutes.
- Gently invert the 50 ml conical tube 3-4 times to mix the sample.
- Aliquot 1.5 ml into the grey capped cryovials.
- Store CSF aliquots upright at -80°C until shipment to BioSEND.



## Plasma, Buffy Coat and CSF Aliquots

- Fill cryovials to 1.5ml
- Over-filled vials may burst in freezer
- Ship material to BioSEND
  - 12 Plasma aliquots
  - 2 Buffy Coat
  - 13 CSF aliquots
- Do NOT send residual volumes to BioSEND





### **Blood Collection: Troubleshooting**

#### Issue #1: Collection tube with little/no vacuum

- Always check expiration dates before beginning blood draw and discard expired tubes
  - Tubes expire on last day of month printed on tube
- Store tubes at ambient temperature
  - Extreme temperatures can affect vacuum
- Keep extra collection tubes from supplemental kit nearby during blood draw to replace "bad" tubes. These can also be requested through the Kit Request Module.
- If frequent occurrence, report tube type and lot numbers to Indiana University



### **Blood Collection: Troubleshooting**

#### Issue #2: Hemolyzed (pink/red) plasma

Cause: Blood Collection Methods	Corrective Action
Improper venipuncture site	Draw from median cubital, basalic, and cephalic veins from antecubital region of arm
Prolonged tourniquet use	Tourniquet should be released after no more than 1 min, excessive fist clenching should be avoided
Not allowing alcohol to dry on skin before venipuncture	Without touching, allow the venipuncture site to air dry
Lumen of needle too close to inner wall of vein (indicated by slow blood flow)	
Use of too large/small bore needle resulting in excess force applied to blood	Avoid using too small/large needle. Needle size dependent on the subject's physical characteristics & amount of blood to be drawn. Most commonly used sizes are 19 – 23.
Pulling/pushing plunger too fast while drawing/transferring blood	Avoid drawing the syringe plunger too forcefully when collecting blood
	Ensure all blood collection assemblies are fitted securely, to avoid frothing



#### **Blood Collection: Troubleshooting**

#### Issue #2: Hemolyzed (pink/red) plasma

Cause: Sample Processing Methods	Corrective Actions
Vigorous mixing/shaking	Gently invert blood collection tube when mixing additive with specimen, follow guidelines in Biologics Manual regarding number of times to invert each type of tube
Not allowing serum to clot for recommended time	Serum tubes without clot activator should be allowed to clot for 60 min in a vertical position
Exposure to excessive heat or cold	Keep samples at ambient temp
Prolonged contact of serum/plasma with cells	Do not store uncentrifuged samples beyond recommended time



Reference: BD's "Tech Talk" newsletter, Vol. 2, No. 2, October 2003 (http://www.bd.com/vacutainer/pdfs/techtalk/TechTalk Jan2004 VS7167.pdf)

# Sample Shipment



### Frozen Samples

- All samples are shipped frozen
  - Plasma, buffy coat, whole blood and CSF
- Ship Monday-Wednesday Only via FedEx Priority Overnight
- Schedule FedEx® pickup
- Email Sample Record and Shipment Notification Form including FedEx® tracking number AHEAD OF SHIPMENT





### Packaging and Shipping Frozen Samples









Place cryobox in biohazard bag with absorbent sheet, seal, and label with case label.



## **Packaging and Shipping Frozen Samples**





Place Whole Blood and buffy coats in bubble wrap pouches and seal



Place in second biohazard bag with absorbent sheet, seal, and label with case label.



### Packaging and Shipping Frozen Samples



Place approx. 2-3 inches of dry in bottom of shipper



Place biohazard bags upright in box



**FILL** dry ice to top of box



# **Packaging & Shipping Troubleshooting**

#### **Issue: Broken/Damaged Tubes**

Cause	Preventative Action
Over filling tubes	Fill tubes to suggested volume. If any sample still remains, place in an additional tube
Improper packaging	Ensure the tubes are securely placed into the bubble wrap pouch and are placed in a separate bag from the boxed plasma, serum, and CSF.
Rough shipping conditions	Extra bubble wrap may be needed to pad blood tubes
Extreme changes in temperature (ambient→freezer; freezer→dry ice)	Wrapping the tubes in bubble wrap before freezing may help slow the cooling process



# Shipping Frozen Samples

- Hold packaged samples in a -80°C freezer until pickup.
- Samples should be received at BioSEND within 2 weeks of collection.



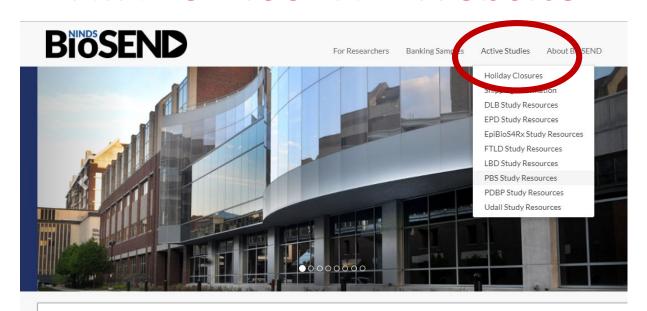


# Sample Shipment Notification Form

Site Name: Principal Investigator:	
Coordinator: Telephone: Email:	
Please list only ONE subject per Sample Record Summary and Shipment Notification	<u>Form</u>
Clinical Identifier: Subject ID (ST# from pre-printed labels):	
Gender: Visit Type:	
Age in Years: Plus Months:	
n the table below, please indicate the date of specimen collection and number of tubes/aliquots submitte	ed.
Completed by Submitter/Site	
Number of Tubes/ Aliquots sent to  Dates of Draw Specimen Type BioSEND Notation of Problems	
DNA	
RNA	
Buffy Coat	
Plasma	
Plasma Serum	



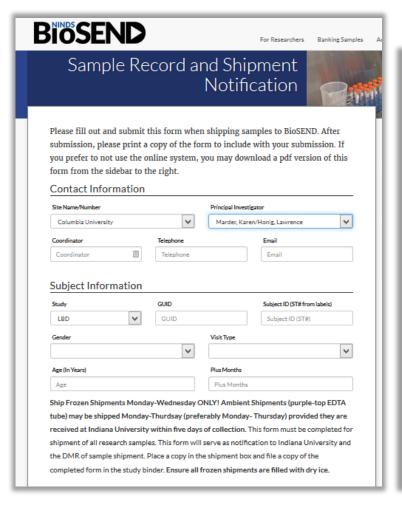
#### NINDS BioSEND Website

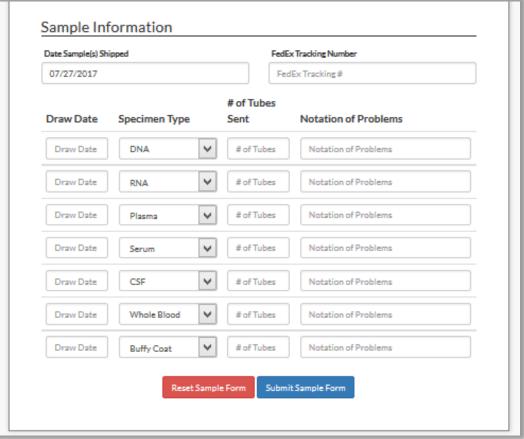






## Sample Shipment Notification Form, Online







# **Holiday Closures**

Date	Holiday
January 1	New Year's Day
3 <sup>rd</sup> Monday in January	Martin Luther King, Jr Day
4 <sup>th</sup> Monday in May	Memorial Day
July 4	Independence Day (observed)
1 <sup>st</sup> Monday in September	Labor Day
4 <sup>th</sup> Thursday in November	Thanksgiving
4 <sup>th</sup> Friday in November	Friday after Thanksgiving
December 25	Christmas Day



### **BioSEND Contact Information**

• Questions?

Please contact: Claire Wegel (cwegel@iu.edu)

Email: biosend@iu.edu



# **Questions?**

