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Economic Feasibility of Seaweed Aquaculture in Southern California

A Group Project submitted in partial satisfaction of the requirements for the degree of
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by

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Table of Contents

Abstract	1
Executive Summary	2
Candidate species and cultivation methods	2
Economic feasibility	3
Co-culture with shellfish aquaculture	3
Recommendations.....	3
Background & Project Objectives.....	5
Significance	5
Seaweed around the globe	5
Seaweed in the United States	6
Seaweed in California.....	6
Potential for seaweed in the Southern California Bight	7
Environmental benefits and challenges	8
Project objectives	9
1. Identification of candidate species and cultivation methods	10
Introduction	10
Methods	11
Finding suitable seaweeds for cultivation in the Southern California Bight	11
Determining optimal conditions and growth rates	12
Results and Analysis	13
Finding suitable seaweeds for cultivation in the Southern California Bight	13
Determining optimal conditions and growth rates	16
Discussion	21
Limitations to growth rate estimates	21
Conclusion	22
2. Feasibility of seaweed aquaculture in southern California.....	23
Introduction	23
Methods	24
Model inputs	24
Model outputs	27

Results and Analysis	28
Break-even price point at year five	28
Onshore versus offshore cultivation methods.....	29
Species variability in costs	33
Assessing the seaweed market opportunities	34
Model sensitivity to changes in growth rate, risk and market price	36
Discussion	37
Onshore vs offshore.....	37
Effect of permitting on feasibility.....	38
Opportunities to reduce capital and operating costs for seaweed aquaculture	39
Conclusion	40
3. Integrating seaweed into an existing mussel farm	41
Introduction	41
3.1 Economically optimal mussel-seaweed farm.....	42
Methods	42
Calculating profit per unit of production for <i>M. galloprovincialis</i>	43
Estimating demand curve for edible seaweed	43
Calculating profit per unit of production for <i>G. pacifica</i>	44
Results and Analysis	46
Discussion	48
3.2 Offshore field experiment.....	49
Methods	49
Study site	49
Mussel growth and condition index	49
Seaweed growth study.....	50
Statistical analysis.....	50
Results	50
Mussel growth (length).....	50
Mussel quality (condition index)	52
Seaweed Growth	53
Discussion	54
Conclusion	55

4. Concluding remarks and recommendations	56
Cultivation techniques	56
Marketing	57
Political Atmosphere	57
Outlook	58
References	59
Appendices	69
Appendix A: Biology and life-cycle of seaweed	69
Appendix B: Hydrographic conditions at our farm location in Santa Barbara, CA.....	72
Appendix C: Seaweed data	72
Appendix D: Seaweed yield summary	78
Appendix E. Cost information	79
Appendix F. Cash flow analyses (cont.).....	87

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Abstract

Global aquaculture production of seaweed is a staggering 23.8 million tonnes per year, with a net worth of \$6.4 billion USD. The majority of edible seaweeds consumed in California is imported from Asia and local seaweed product is overwhelmingly sourced from wild harvesters in northern California. Reliance on imports and wild harvesting can have large environmental consequences and put additional pressure on regional marine ecosystems. As the demand for local seaweed in California increases interest has grown in developing a seaweed aquaculture industry through both onshore and offshore systems. However, the technical and economic feasibility of growing native California seaweed species as local alternatives to the major imported species — nori, kombu, and wakame— has not been assessed. Our project investigated which seaweed species have the most potential based on market demand and suitability to local environmental conditions. We identified four promising candidates in the Southern California Bight and the appropriate methods for their cultivation. We then developed a bioeconomic model to highlight the key factors that determine the economic feasibility of establishing a seaweed aquaculture industry in the region. Finally, we undertook a preliminary assessment of seaweed integration into an existing local mussel farm. Our results show that regional seaweed aquaculture is economically feasible, that there are important considerations for siting production on or offshore, and that co-culturing seaweed with shellfish can provide a jumpstart to the industry as a whole. These findings provide stakeholders and policymakers a blueprint for prioritizing next steps in expanding sustainable seaweed aquaculture in southern California.

Executive Summary

Seaweed aquaculture is an emerging industry in the United States, but has been established in many Asian countries for centuries. Over one-fifth of global annual aquaculture production comes from seaweed, with the vast majority of that being used for direct human consumption (Moffit, 2014). Seaweed is produced for a wide variety of uses, including pharmaceuticals, biofuel, and agricultural products, but food and health are human necessities that may motivate the more rapid expansion of local, sustainable aquaculture. The rich nutritional profile and low environmental footprint of seaweed contribute to its promise as a sustainable and healthy food source. To date, increasing demand for edible seaweeds in California has been primarily filled by imported products (NOAA Fisheries, 2017). However, the environmental and oceanographic conditions of California's waters are an untapped resource for seaweed cultivation as evidenced by the dense stands of kelp forest along the coast. The lack of need for freshwater and pesticide inputs makes seaweed aquaculture a prime candidate to sustainably address food security issues in California.

In our project, we focused on seaweed species that satisfy a growing health-conscious community and have a high potential in the southern California food market. We assessed the economic feasibility of starting a seaweed farm in onshore (i.e. land-based) and offshore. Here, offshore is defined as mariculture occurring in ocean but does not necessarily refer to cultivation in deep and distant waters. The main barriers impeding the growth of the industry are high initial capital costs, permitting uncertainty, and a lack of established cultivation methods specific to the region. Our report aims to provide researchers, future farmers, and policy makers comprehensive scientific knowledge about ideal candidate species for edible seaweed cultivation, prospective economic feasibility, and mechanisms to circumvent some of those barriers in the short term.

Candidate species and cultivation methods

We utilized three criteria to identify edible seaweed species that are ideal for cultivation and sale in southern California: 1) The algae must occur naturally throughout the whole region, 2) have a pre-existing food market in California, and 3) must have established aquaculture techniques. The first condition helps promote a cohesive and collaborative local industry and the latter two are desirable by aquaculturists who want to buy into a system that has a proven history of success. Applying these criteria to the native species of the Bight, the following four species emerge as the most promising candidates:

- *Gracilaria pacifica* (ogo or limu) - a delicate red seaweed used as a topping for poke and rice bowls
- *Pyropia perforata* (nori) - a local variant of a highly marketable red genus that is commonly dried for sushi rolls
- *Ulva lactuca* (sea lettuce) - a productive green species found in soups and seaweed salads
- *Laminaria setchellii* (kombu) - a sturdy brown seaweed that is sought after for use in soup stock or consumed whole

Optimal harvest season and potential productivity for a locale can be estimated by using knowledge of species physiological tolerance ranges and a combination of ambient

environmental conditions, such as water temperature, nutrient availability, and photoperiod. The final decision facing a farmer is which grow-out structure to use for a particular species. All four aforementioned species can be grown in tanks onshore. For offshore cultivation, longline culture is most appropriate for *G. pacifica* and *L. setchellii*, whereas *P. perforata* and *U. lactuca* should be raised used nets. Since the latter two species are more susceptible to rough wave action, siting them in calmer or shallower waters will decrease the likelihood of fragmentation.

Economic feasibility

We synthesized projected farm yield, initial required capital, and annual operating costs through a bioeconomic model to investigate the key factors that will affect the economic feasibility of seaweed aquaculture in southern California. Seaweed growth rate has a major influence on the viability of a farm system and *L. setchellii* outperformed the other three candidate species largely due to its high biological productivity and low annual harvest costs. However, as a brown seaweed, it typically has a lower market value than a red species such as *P. perforata*. This suggests potential trade-offs in species selection between productivity and marketability. Our research also highlights the pros and cons of siting seaweed production on and offshore. Offshore production can be more economically viable due to its lower annual operational costs, but a farmer may be forced to make sacrifices in terms of product quality and increased risk. Additionally, securing a permit for offshore aquaculture has proven to be more difficult than for onshore systems. An aquaculture farmer can have more control over product quality onshore but this comes at the price of higher labor and energy costs. Our models predicts an average price of \$11.42/kg across the four species for an onshore farmer to recoup their costs over a five-year time period compared to \$6.84/kg offshore. Regardless of location, these “break-even” prices suggest that the cost of seaweed aquaculture in southern California prohibits successful competition on the global market, where seaweed is processed for use as a food additive. Instead, the seaweed industry must position itself to provide a value-added produce or specialty item to be viable.

Co-culture with shellfish aquaculture

In California and elsewhere around the globe, seaweed aquaculture has been increasingly associated with shellfish cultivation. These two extractive organisms have been traditionally paired with fed finfish aquaculture, a process known as integrated multitrophic aquaculture (IMTA). However, shellfish and seaweed can also be grown together in isolation. Shellfish farmers benefit from diversifying their product portfolio and potential seaweed farmers can leverage existing permits for shellfish aquaculture to avoid having to obtain a new one. Sharing required infrastructure and novel “3-D” cultivation methods allows for increased yield from a single farm unit. This project demonstrates that an existing mussel farm could increase its profit by substituting seaweed production on to the farm. It is also possible that there are ecological benefits of growing the two species in unison, such as localized buffering of ocean acidification by seaweeds, but further empirical research is needed to corroborate these theories.

Recommendations

Seaweed aquaculture can be economically feasible in southern California but barriers exist that prevent the development of a regional industry. We recommend stakeholders should focus their efforts on adapting current cultivation techniques for use in California waters, identifying the most effective marketing strategies, and creating a political framework in support of sustainable

best management practices. Future steps will include validating the growth rate estimates utilized in this report and quantifying the health and environmental benefits of seaweed aquaculture for consumers and decision makers. Further research should investigate co-culture with shellfish and the polyculture of multiple seaweed species to maximize economic and biological outcomes. Through increased collaboration between researchers, policy makers, and aquaculture farmers, sustainable seaweed aquaculture can be the next innovation in California's rich history of food production.

Background & Project Objectives

Significance

Aquaculture is poised to help address food security issues as the world's population continues to grow (FAO, 2014). A shift towards more domestic production in the United States would allow for the creation of many jobs and stricter control of the sustainability and environmental impact of the industry. While finfish and shellfish aquaculture have received significant attention in the academic and political realms, the cultivation of edible seaweeds is a relatively novel idea throughout much of the U.S. coastal zone. Seaweeds, or macroscopic algae, are nutrient rich, require very few inputs (including no freshwater), and thrive in the upwelling zone along the coast of the western United States. These facets make the production of seaweed for direct human food use appealing to consumers, policy makers, and aquaculturists alike.

Interest in seaweed aquaculture is growing here in southern California, but many questions remain as to the feasibility of the industry. To our knowledge, there has been no assessment of the economic feasibility of growing native California seaweed species for food use. Additionally, no review has summarized which native seaweed species to target, what cultivation methods are most appropriate, and the limitations and uncertainties regarding seaweed aquaculture in California. Our project seeks to address these questions and provide a framework for future studies to build upon when assessing the economic feasibility of seaweed in California and other regions. Additionally, identifying the most ideal seaweed candidates will help direct future research toward these species.

Seaweed around the globe

23.8 million tonnes of seaweed are produced globally every year, valued at \$6.4 billion USD (Moffitt, 2014). Over 95% of seaweed products are farmed, accounting for over a fifth of global annual aquaculture production (Moffitt, 2014). Out of the 33 countries that harvest seaweed, China is the largest producer by a significant margin with limited production in Western countries. Cultivation of seaweed grew 50% globally between 2004 and 2014 (Nayar & Bott, 2014). However, the demand for it is still outpacing supply as the global seaweed market doubled from 2000 to 2012 (West et al., 2016). Out of all groups cultivated, the market for red seaweeds is growing the most rapidly at 8% annually (West et al., 2016).

Seaweed produced for human consumption as fresh, dried, or processed products makes up 80% of the value of the global market while the remainder is mainly used in pharmaceuticals, agricultural fertilizer, and aquaculture feed (Buchholz et al., 2012; McHugh, 2003; Nayar & Bott, 2014; West et al., 2016). Algal hydrocolloids, such as carrageenan and agar, are used as emulsifying, gelling, or water retention agents. However, sustainable food production is a popular topic with which more people can engage compared to seaweed cultivation for alternative energy use or high-end cosmetics that are non-essential. In the international edible seaweed market, the most dominant species are the red seaweeds *Kappaphycus alvarezii* and *Eucheuma spp.*, which are used to produce carrageenan (Moffitt, 2014). These are followed by seaweeds consumed dried or raw, such as *Laminaria japonica* (kombu or Japanese kelp),

Gracilaria spp. (ogo, limu), *Undaria pinnatifida* (wakame) and *Pyropia spp.* (nori). *Pyropia* (formerly referred to as *Porphyra*) is by far the most valuable seaweed species with two species, *P. yezoensis* and *P. tenera*, constituting the bulk of global production (Jensen, 1996; Levine & Sahoo, 2009).

Numerous health benefits are attributed to seaweed including high levels of iodine, fiber, and trace minerals, although nutrient contents vary greatly across species (Fleurence, 1999; MacArtain et al., 2007). For example, the calcium concentration in some seaweeds exceeds that of whole milk, making it particularly attractive for use in health food or vegan products (MacArtain et al., 2007). Given the health benefits associated with consuming seaweed, tremendous growth potential exists for this product in health-conscious markets within the US, Canada, and Europe.

Seaweed in the United States

The supply of farmed, wild harvested, and imported seaweed has grown over the past decades in the United States as the American palate begins to accept and integrate seaweed. Seaweed imports increased 65% from 1999 to 2008 (Nayar & Bott, 2012). In 2016, the U.S. imported over 31 metric tons of seaweed, a 19.7% increase from 2015 (NOAA, 2017). This seaweed was valued at about \$146 million USD (NOAA, 2017). Most edible seaweed production occurs on the east coast and in Hawai'i, largely through the harvest of *Chondrus crispus* (irish moss) and *Palmaria palmata* (dulse). More recently, aquaculturists have begun cultivating *Saccharina latissima* (sugar kelp) and *Gracilaria spp* (ogo or limu) as well. There is also an increasing effort in the US to move towards an ecosystem-based approach known as integrated multi-trophic aquaculture (IMTA), a technique that harnesses the benefits of culturing two or more species from different trophic levels together.

Rapid increase in demand for seaweed products is expected to continue domestically due to the potential for a new, sustainable, and healthy protein source (Buck et al., 2017). This has already been seen in Maine where seaweed sales doubled every year for the first 6 years of seaweed aquaculture production, between 2009-2015 (Lem, 2016). The easiest market entry point for seaweed domestically is in high-end restaurants. Chefs are drawn by its rich nutrition profile and are experimenting with it at increasing rates (Mouritzen, 2012). Out of the most commonly consumed species, *Pyropia spp.* and *Palmaria palmata* have been identified as having the highest protein quantity and quality respectively (Holdt and Kraan, 2011). Despite varying substantially across species, the nutritional aspect of seaweed is crucial to its potential marketability (MacArtain et al., 2007; McHugh, 2003).

Seaweed in California

Despite limited seaweed aquaculture research and development in California, seaweed harvest is growing rapidly. In 2004, harvested seaweed amounted to over 33,000 metric tons, making it the 5th most landed taxon by weight on the west coast (Norman et al., 2007). As of 2014, harvesting of kelp (*Macrocystis pyrifera*) has declined rapidly, while edible seaweed harvest (e.g. species from *Pyropia*, *Laminaria*, and *Monostroma*) has increased (CDFAW, 2014).

The seaweed harvest occurs primarily in northern California, with many commercial harvesters operating in Mendocino County and near San Francisco (e.g. Strong Arm Farms in Sonoma,

Mendocino Sea Vegetable Company and Rising Tide Sea Vegetables in Mendocino). There are strict harvesting regulations including permitted areas, protected species, and limits on the annual harvest quantities. Commonly hand harvested seaweed species in California are *Macrocystis pyrifera* (giant kelp), *Nereocystis luetkeana* (bull kelp), *Mastocarpus papillatus* (turkish washcloth), *Laminaria setchellii* (kombu) and *Stephanocystis osmundacea* (bladder chain kelp) (CDFW, 2014).

While coastal seaweed aquaculture has developed to varying extents on the eastern seaboard and in Hawai'i, businesses and stakeholders are just starting to consider growing seaweed off the coast of California. Currently, no ocean farmed seaweed exists in California. Suitable local seaweed species, methods for culturing them in the eastern Pacific Ocean, and the economic feasibility of farm methods still need to be identified. Therefore, seaweed aquaculture remains limited in California, with the only existing production occurring in tanks on land.

The three current land based seaweed farms in California are: Monterey Bay Seaweeds in Moss Landing, Carlsbad Aquafarm in Carlsbad, and The Cultured Abalone Farm in Santa Barbara. The farms in Santa Barbara and Carlsbad primarily grow shellfish but augment their operations with production of *Gracilaria pacifica*. Meanwhile, Monterey Bay Seaweed grows *Palmaria palmata* (Northeast Atlantic dulse), *Gracilaria spp.*, and *Ulva spp* in recirculating tanks as its sole products. In Santa Barbara, *G. pacifica* is used for abalone and fish feed, and sold to restaurants and food processors. Monterey Bay Seaweeds sells to chefs and high-end restaurants in the wider Monterey Bay Area (e.g. San Francisco, Santa Cruz), and these seaweeds are processed live and transported in saline water to their destination. The existing seaweed market can provide useful insight into potential seaweed candidates, target markets, and ways to add value through processing.

Potential for seaweed in the Southern California Bight

Nutrient-rich, upwelled waters off the southern California coast provide a unique opportunity to cultivate seaweed with minimal inputs (Sautter & Thunell, 1991). These are the same waters that support the algae-dominated kelp forest ecosystems that line the region's coastline (Brzezinski et al., 2013). The environmental conditions of the Southern California Bight, the coastline between Point Conception and San Diego, also cater to the strict light and temperature requirements that many cultivated seaweeds have. Water temperatures and photoperiods in the region are relatively homogenous year-round (Appendix B). In addition to physical and chemical ocean properties, the Southern California Bight is also an epicenter for the health-conscious food market in the United States (Guthman, 2004). Having immediate access to the target market will likely lead to more economic success for the seaweed aquaculture industry (D. Bush, pers comm, 2018; M. Graham, pers comm, 2018).

Despite the numerous promising characteristics of the Southern California Bight for seaweed aquaculture, the industry remains underdeveloped. Part of the current dialogue in the industry focuses on the decision to site production onshore or offshore. Delicate species that are likely to fragment benefit from the less dynamic conditions when cultured on land (M. Graham, pers comm). Onshore farms are much easier to get permitted but are typically more energy intensive and expensive to operate (D. Bush, pers comm, B. Friedman, pers comm, 2017; Huguenin, 1976;). Open-ocean farms could be a good alternative to land-based systems since there are few inputs in the form of energy, seawater intake, and added nutrients (Nobre et al 2010).

However, open-ocean farms don't come without challenges in California. Spatial conflicts with other ocean users such as the Navy, commercial fishers, and tourism operations exist, but could be minimized with careful planning (Gentry et al., 2017). Understanding the tradeoffs between onshore and offshore may enable decision makers to make more informed choices regarding future seaweed aquaculture development along the Southern California Bight.

Environmental benefits and challenges

Encouraging local cultivation has significant environmental implications, as most seaweed consumed in southern California is imported (Figure 1). Imported produce items, such as seaweed, have a significantly higher carbon footprint due to their transport and electricity use for storage, packaging, and transport operations (Jones, 2002; Sim, 2007). Imported produce also contributes more to climate change, ocean acidification, and abiotic depletion than produce that is domestically grown (Sim, 2007). Furthermore, there are likely added benefits to growing seaweed in our marine environment. These range from absorbing excess terrestrial nutrients present in runoff to moderating ocean acidification (Hirata et al 1994, Grote 2016, Kang et al 2014, Kim et al 2015). The last characteristic is particularly relevant to shellfish aquaculture, as shellfish development can be retarded by shifts in pH and carbonate chemistry (Chung et al., 2013).

As climate shifts the regular range of local ocean conditions, it may be pertinent to consider integrating seaweed into an IMTA or co-culture system to help mitigate localized effects of ocean warming and acidification. Additionally, some species of seaweed have high nutrient uptake capacities, reincorporating nutrients like ammonium that is released from suspended mussel farms (Cohen & Neori, 1991; Grote, 2016; Kang et al., 2014; Kim et al., 2015). While shellfish could also provide environmental benefits as filter-feeders, expansion of the industry should be coupled with conservative practices that limit the net footprint in our oceans. Thus, integrating new species into an existing farm could be a more sustainable practice moving forward.

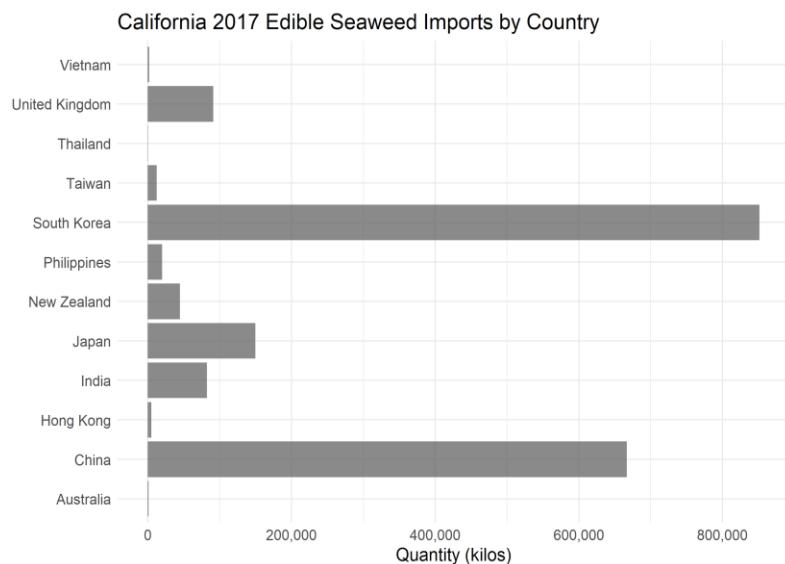


Figure 1. Edible seaweed imports to California in 2017 by country. Total imports by weight is 1,926,273 kg valued at an average of \$13.38/kg. Source: NOAA Fisheries, Fisheries Statistics Division: Commercial Fisheries Statistics, 2017 Cumulative Trade Data by U.S. Customs District.

In general, concerns still persist regarding the greater aquaculture industry like escape, benthic and aquatic pollution from waste and excess feed, and use of antibiotics that may create drug-resistant parasites and bacteria. However, many of these do not apply to properly managed seaweed aquaculture (Costa-Pierce, 2002). Cultivating native species greatly reduces the ecological impacts of escape. Seaweeds are net nutrient sinks which limits their environmental footprint and makes them valuable in IMTA systems (Troell et al., 2009). Finally, no pesticide or freshwater inputs are required to grow seaweed in the Southern California Bight. This is particularly important given the region's limited water resources and frequent bouts with drought (MacDonald, 2007).

There are still concerns about local seaweed aquaculture offshore in addition to the spatial issues outlined in the previous section. The most unrelenting concern is marine mammal entanglement (R. Lovell, pers comm, 2017). There have been 5 reported cases globally of turtle and cetacean entanglement in offshore mussel farm equipment, which is structurally similar to seaweed aquaculture equipment (Young, 2015). Although entanglement is suspected to be underreported, 5 occurrences in the last two decades suggest fairly small risk (Lindell and Bailey, 2015). Moreover, the risk of marine mammal aquaculture entanglement is significantly less than that of traditional fishing gear (Young, 2015).

Project objectives

Perhaps the greatest challenge to growing seaweed in the Southern California Bight is that interested parties simply don't have the framework for a successful seaweed aquaculture operation in the region. While more information is available for onshore farming, questions remain for both onshore and offshore systems about what species to grow, when and where to grow those species, what cultivation method to use, and how profitable these operations could be. Because seaweed aquaculture in the Southern California Bight promises less risk, fewer environmental threats, and more environmental benefits than traditional terrestrial agriculture and other forms of aquaculture, there is growing interest both onshore and offshore from groups like Ventura Shellfish Enterprise, the Port of San Diego, and Catalina Sea Ranch. Our project completed a comprehensive analysis of the ideal candidate species with regards to the requirements for their cultivation and the relative economic feasibility of doing so. To accomplish this, we set out the following project objectives:

- 1) Identify southern California candidate seaweed species and the appropriate cultivation methods
- 2) Determine key factors that influence economic feasibility in each species-farm system through the use of a bioeconomic model
- 3) Perform a preliminary investigation of incorporating seaweed into an existing mussel farm

Through the first objective, we selected the most promising species from those native to the Southern California Bight using a mixed criteria of technical ease and market suitability. In the next section, we projected the economic feasibility of the seaweed aquaculture industry in the region via a synthesis of estimated farm costs and annual biological yield. We also outlined the variables and processes that stakeholders should focus on to improve business sustainability. Finally, we examined the benefits to both industries of integrating seaweed cultivation into existing mussel aquaculture through our partnership with Santa Barbara Mariculture, an open-ocean mussel farm off the coast of Hope Ranch.

1. Identification of candidate species and cultivation methods

Introduction

A major challenge for seaweed aquaculture in the Southern California Bight is the lack of research and development done on native seaweeds for aquaculture in the region (see Background). Seaweed farming is a multi-stage process that is shaped by the life-history and ocean condition preferences of the target species. These two factors greatly influence the cultivation methods used and the respective costs (Pereira & Yarish, 2008). Before establishing a seaweed farm, a farmer must decide 1) where to place the farm (onshore or offshore), 2) which species to grow, 3) the best method and timing for cultivating the species, and 4) the economic feasibility of these decisions (Radulovich et al., 2015; Titlyanov & Titlyanova, 2010). In this chapter, we describe the decisions facing a potential seaweed farm, steps that must be taken at the various phases, and provide the best available data to inform the first three questions above.

Farm location and which species to produce are the first decisions that must be evaluated by an interested aquaculturist. For intensive seaweed farming, the two possible locations are onshore or in the ocean (i.e. offshore). Onshore pertains to seaweed cultivated on-land and includes pond and tank culture; offshore is used to describe any operation where seaweed is grown in the ocean, regardless of the proximity of the farm to the shore (McHugh, 2003). As discussed in the Background, onshore production offers benefits in the way of permitting ease and risk reduction for fragile species that may fragment (M. Graham, pers comm, 2017). Conversely, offshore farms are typically cheaper to operate and are less energy intensive, but the lack of control over environmental conditions often results in highly variable annual production (Huguenin, 1976).

The life history, ocean condition preferences, locality of species, cultivation potential, and marketability all have to be considered when selecting a seaweed species to cultivate (Radulovich et al., 2015). Interested farmers are unlikely to choose a species that isn't cultivated globally (B. Friedman, pers comm, 2017). Farms should only consider seaweeds native to the region to avoid the risk of introducing a foreign species. Selecting a native seaweed species also provides confirmation that the species can grow in that region for at least part of the year. It is important to compare the optimal environmental conditions for each species to those observed in the farms locale to ensure that the seaweed will have the appropriate levels of light,

nutrients, temperature, and other environmental variables (Santelices, 1999). Furthermore, the local oceanographic regime can influence the chosen seaweed's reproductive schedule which will have a significant impact on farm operations. The reproductive schedule directly limits the growth and harvest seasons of some species of seaweed (Titlyanov & Titlyanova, 2010). Some species have complicated life cycles that require land-based hatchery facilities that can carefully control water temperature, nutrients, and light (McHugh, 2003). Hatcheries are the sites at which reproduction is induced to seed the production units that will later be placed offshore.

Seaweeds are grouped into three taxonomic clades: red (*Rhodophyta*), green (*Chlorophyta*), and brown (*Phaeophyta*). Each group has unique differences in reproductive and life-history modes (Appendix A). The wide variety of reproductive, physiological, and morphological forms amongst seaweeds results in highly specialized cultivation strategies that must be determined for each species. Broadly, there are four major steps in the cultivation of macroalgae: 1) hatchery phase, 2) nursery phase, 3) grow-out phase, and 4) harvest phase but not all species require each of these phases.

Once a farmer has chosen a species of seaweed to grow, they must select a growing method. Broadly, several methods of offshore cultivation exist including: fixed pole, semi floating raft, floating raft, open water ropes, nearshore ropes, baskets, and nets (Kim et al., 2017). Some taxa have multiple applicable cultivation methods while others are typically only grown using one structure (Kim et al., 2017). The most common cultivation methods onshore are growing seaweed in tanks or ponds (Kim et al., 2017; McHugh, 2013). These various cultivation methods depend on farm location and incur vastly different costs depending on the species and locale.

After life history, cultivation potential, and ocean conditions are accounted for, marketability needs to be considered. The chosen species must have a proven market for increased chance of success. While many markets for seaweed exist and have been discussed throughout this paper, we have chosen to focus on the direct for human consumption market. The majority of the value global seaweed aquaculture is in the food market and local farmers can tap into its higher price point to increase their economic viability (D. Bush, pers comm, 2018; M. Graham, pers comm, 2017; McHugh, 2003). The entire evaluation process from site location to marketability is outlined in further detail in the methods section of this chapter.

Methods

To determine ideal candidate species, their optimal cultivation structures, and their theoretical growth rates in the Southern California Bight, we used the following six procedures: 1) identify species that occur throughout the Southern California Bight, have a food market, and are from a taxa widely cultivated in aquaculture systems, 2) determine the ideal grow-out structures both on and offshore, 3) establish oceanographic conditions at our model site, 4) match these conditions with the ambient environmental requirements for the seaweed species, 5) determine the best cultivation schedule regarding hatchery, out-planting, and harvesting, and 6) ascertain high and low growth rates for the seaweed species in offshore and onshore regions.

Finding suitable seaweeds for cultivation in the Southern California Bight

We first identified all seaweed taxa that grow throughout the full range of the Southern California Bight using the Jepson Herbarium at the University of California, Berkeley, which resulted in 191

genera of species (UC Berkeley, n.d.). We only used species that occur throughout the entire Southern California Bight since the growth rates for species reaching their thermal limits will be limited, and thus, not appropriate for mass cultivation. Furthermore, using seaweed that grows outside of its native range raises concerns with introducing a foreign species. Based on these criteria, we excluded six seaweed species that have been found at only at the very northern range of the Southern California Bight (Santa Barbara County) and in low densities. This included the following species: *Postelsia palmaeformus*, *Palmaria mollis*, *Alaria marginata*, *Gigartina papillata*, and *Fucus gardneri*. Next, we cross-referenced this list with the genera that have a direct human consumption/food market in Southern California (Abbott & Hollenberg, 1992; Kim et al., 2017; Titlyanov & Titlyanova, 2010). Finally, we eliminated any species which are not grown in aquaculture systems globally (McHugh, 2013; Titlyanov & Titlyanova, 2010). For the remaining candidate species, we conducted a literature review and spoke with industry experts to determine the appropriate grow-out structures (e.g. nets, tanks) needed for each species in offshore and onshore regions in the Southern California Bight.

Determining optimal conditions and growth rates

Seaweed species growth rates in offshore regions are highly dependent on oceanographic conditions. Thus, it is necessary to time ideal growth conditions with the offshore outplanting/cultivation period of the seaweed species. We reviewed the literature to identify what environmental settings our seaweed candidates prefer and compared those against historical ocean conditions recorded by NOAA, SCCOOS, UC Santa Barbara LTER, and CalCOFI for our chosen farm location at Santa Barbara Mariculture in Santa Barbara, CA (Appendix B). The exact coordinates for the study are as follows: 34°23'39.2"N 119°45'09.8"W. We chose this location because it is the only permitted and functioning offshore aquaculture farm of any type in the region. However, Santa Barbara Mariculture grows shellfish and does not currently grow seaweed.

After matching optimal seaweed growth conditions to the seasonality of those factors at our study site, we compared the potential growing season to how the life cycle of the seaweed affects harvest and may be affected by ocean conditions. We were then left with an optimal growing and harvest season. For onshore, the ocean temperature remained an important factor in determining seasonality but nutrient levels were not as important since farmers can easily manipulate these by adding fertilizer to their system.

Once we established what to grow, how to grow it, and when to grow it, we had to estimate the growth rate and annual yield for each candidate species. The following methodology was used to select growth rates:

1. Determined monthly (Jan-Dec) values of temperature, dissolved inorganic nitrogen, current speed, photoperiod, and irradiance in regions representative of our client's farm location. Sources: SCCOOS, CalCOFI Stations, UC Santa Barbara (UCSB) LTER, and NOAA reports of historic values.
2. Collected discrete daily relative growth rates for a specific set of environmental conditions from various seaweed culture experiments, culture handbooks, and publications.
3. Where species specific data were lacking, we used morphologically, genetically, or reproductively similar species as a proxy for the local Southern California Bight species.

4. Selected the stocking density most commonly used in literature or one that led to optimal growth in the absence of a ubiquitous value.
5. Chose the growth rates or growth rate range that best matches the environmental system in the Southern California Bight and our understanding of the biology of our local species. At times, an estimated or measured final yield at the end of each harvest period (e.g. 7 kg per meter of longline) as a more reliable indicator of potential productivity.
6. Selected a representative high and low growth rate if there were clear peak and slow periods/months during a single harvest season or a single year. We did not include a requirement that these two were sourced from the same study for every species.
7. When peak and slow periods for our local species were not available, we used algal abundance studies (e.g. UCSB LTER) to approximate which months out of the year we were likely to see the species grow its best.

Results and Analysis

Finding suitable seaweeds for cultivation in the Southern California Bight

We identified nine species that are extant throughout the entire Southern California Bight and are sold for direct human consumption (Table 1.1). From these nine species, we excluded those that do not have a food market in California or are not from genera with established aquaculture practices. This resulted in our four candidate species that we used for further analysis regarding their economic feasibility in southern California. Below, we present information on our candidate species and the other five species that were excluded from our study.

Table 1.1. The nine seaweed species that were considered for their potential in seaweed aquaculture in the Southern California Bight. The final four candidate species and five excluded species, along with the justification for exclusion, are shown.

Selection	Species
Candidate species	<i>Gracilaria pacifica</i> <i>Pyropia perforata</i> <i>Ulva lactuca</i> <i>Laminaria setchellii</i>
No food market	<i>Chondracanthus exasperatus</i> <i>Agardhiella coulteri</i> <i>Hypnea spp</i>
No aquaculture cultivation	<i>Eisenia arborea</i> <i>Gelidium robustum, Gelidium spp</i>

Candidate species

Gracilaria pacifica

Gracilaria pacifica, or “ogo”, is grown in aquaculture systems, occurs from Santa Barbara to San Diego (and beyond), and has a food market (Abbott & Hollenberg, 1992; D. Bush, pers comm, 2018; Kim et al., 2017). It is generally consumed fresh as a topping for poke bowls or dried and processed for use as a rice seasoning. This species is grown onshore in tank culture in southern California currently and can be cultivated offshore on ropes, cage-like structures, and on the nearshore benthos (Kim et al., 2017; Yarish et al., 2012).



Figure 1.1. Tank cultivation of *G. pacifica* (Mazza, 2017).



Figure 1.2. Fixed pole net cultivation of *Pyropia* (McHugh, 2013).

aquaculture system experimentally (Waaland, 1977). Commonly known as “nori”, *P. perforata* is often dried and processed into thin sheets for sushi rolls. We selected *P. perforata* as a candidate species due to its potential value and the wide cultivation of its genus. *Pyropia spp.* are commonly grown on nets via fixed pole, semi floating rafts or floating rafts offshore (Figure 1.2; Kim et al., 2017; Levine & Sahoo, 2009).

Ulva lactuca

Both species of *Ulva* common in California are found in our range (Abbott & Hollenberg, 1992). *Ulva lactuca* is grown in aquaculture systems and has a food market (M Friedlander, pers comm, 2018; Titlyanov & Titlyanova, 2010). “Sea lettuce” is frequently utilized fresh in seaweed salads and soups. *Ulva lactuca* are generally grown on nets offshore or in tank cultures onshore (Brandenburg, 2016; Cohen and Neori, 1991).



Figure 1.3. *U. lactuca* in the wild (Peters, 2006).



Figure 1.4. A kelp species grown on longlines as *Laminaria setchellii* would be (The Seaweed Site, 2018)

Laminaria setchellii

Laminaria spp. are grown commercially in aquaculture systems, occur in our range, and have a food market (Abbott & Hollenberg, 1992; Mendocino Sea Vegetables; Rising Tide Sea Vegetables; Titlyanov & Titlyanova, 2010). Therefore, we chose to consider *L. setchellii* as a native representative of this genus. While this species is not currently consumed by humans, other Laminariales, or “kombu”, are a staple product in Asian cuisines and are consumed raw, cooked, and dried. *Laminaria spp.* are grown on longlines or lines via floating rafts offshore and in tanks onshore (Figure 1.4; McHugh, 2013; Peteiro et al., 2006; Titlyanov & Titlyanova, 2010).

Species that do not meet all criteria

Chondracanthus exasperatus

Chondracanthus exasperatus grows in our target region and is being/has been grown on a small scale in Washington (Waaland, 2004). The market for this species exists solely in the cosmetic industry and thus did not fit our criteria for having a food market in the U.S. (Boratyn, 2000; Lewallen & Lewallen, 1996; Waaland, 2004).

Eisenia arborea

This species grows in Santa Barbara and has been identified as having potential for commercial cultivation (Abbott & Hollenberg, 1992; Zertuche-Gonzalez et al., 2016). However, the food market for this species is very small and primarily exists in Asia (Zertuche-Gonzalez et al., 2016). There is no evidence of a U.S. market. Furthermore, it is not currently cultivated in aquaculture systems and there is generally a lack of information on this species outside of wild harvesting it for abalone feed or experimenting with it in powder form to treat allergies (Zertuche-Gonzalez et al., 2016). The lack of a current food market and the lack of information for the commercial cultivation of this species caused us to eliminate it from consideration.

Gelidium robustum and *Gelidium spp.*

G. robustum grows throughout our target region and is most productive in nutrient rich, upwelling regions (Abbott & Hollenberg, 1992; Santelices, 1991). There are potentially small food markets in China and Japan for *Gelidium spp.* (Doty et al., 1987; FAO, 1994; Fotedar & Phillips 2011; Salinas, 1991). However, the cultivation of *Gelidium spp.* is still experimental and not currently seen as economically viable (Cremades, 2012; Doty et al., 1987; Fotedar & Phillips, 2011; Salinas, 1991). Given the lack of established wide-scale cultivation practices, we excluded *Gelidium spp.*

Agardhiella coulteri

This species occurs within the Southern California Bight range but there is no evidence of a direct human consumption market (Abbott & Hollenberg, 1992). Other seaweeds in the *Agardhiella* genus are cultivated for their phycocolloids and are starting to be used for shellfish feed (Garr et al., 2009; Chopin et al., 1989; Huang & Rorrer, 2002).

Hypnea spp.

Hypnea spp. grow in the northern end of the Southern California Bight (i.e. Santa Barbara County), but are extremely rare (Jepson Herbarium, 2018). The genus is commercially cultivated, but there is no evidence of a food market and is typically processed as a phycocolloid (Wallner et al., 1992; Ding et al., 2013; Ganeshan et al., 2005). Given that *Hypnea* does not grow throughout our entire study range, it was excluded.

Determining optimal conditions and growth rates

With our list of candidate species finalized, we identified ideal environmental conditions, growth rates, and the seasonality of the grow-out and harvest periods for each species in both an onshore and offshore system. When data for our candidate species in our region was not available, we tried to use data from studies done with synonymous species (in regard to life cycle and morphology) at similar latitudes with similar ocean current patterns. All final parameters discussed in this section are summarized in Appendix C.

Gracilaria pacifica

Most of the studies found in literature for offshore *Gracilaria* growth were performed on other species (i.e. *G. gracilis*) or had non-nutrient limiting environments (concentrations of NO₃- between 15.3 and 33.5 uM). Maximum growth rates reported were as high as 11% (Davison, 2015; Wakibia et al., 2011). However, these rates would most likely be limited by nutrient availability in the Santa Barbara Channel. To estimate growth at our client's farm off Hope Ranch, Santa Barbara, we used a two-week field growth study that was conducted by the Sustainable Aquaculture Research Center (SARC) onsite in March 2016 for *Gracilaria pacifica*. It is likely that March falls during the most productive time of year for *G. pacifica* due to the nutrient concentrations being the highest around this time, according to historic trends from UCSB LTER and CalCOFI monitoring data.

From our research, we predicted that *G. pacifica* growth is most influenced by light and nutrient availability. This high value taken from the SARC experiment for relative growth rate was 3.98% per day (Table 1.2). The low growth rate is taken from the Halling et al. (2005) study that grew *Gracilaria chilensis* offshore in Chile. While this is not a perfect replicate of Santa Barbara, both locations have Mediterranean climates with equatorial currents (pole to equator, colder waters). We set the lower end of our growth range at 1.44% (Table 1.2), a value that was one standard deviation lower than the mean reported in that study. The stocking density (0.4 kg/m) was taken from Dawes (1995), which produced similar growth rates to that in the SARC experiment.

Table 1.2. Low and high growth rates for candidate species both offshore and onshore. In some instances, the most reliable growth estimate found in the literature was a final annual production instead of daily relative growth rate. These are indicated as such below and the equivalent relative growth rate was calculated for use in our bioeconomic model.

Species	Offshore Growth	Onshore Growth	Data Sources
<i>Gracilaria pacifica</i>	1.4 - 3.98% RGR per day	5.48 – 6.9% RGR per day	SARC, Halling et al 2005; SARC
<i>Ulva spp.</i>	3.3 – 6.67 kg/m ² annual production	10 – 17.6% RGR per day	Titlyanov & Titlyanova 2010; Duke et al 1989; Hernandez et al 2005
<i>Pyropia perforata</i>	2.96 - 6.52 kg/m ² annual production	10 – 16% RGR per day	Parker 1974, Miura 1975; Kim et al 2007
<i>Laminaria spp.</i>	7 – 16 kg/m annual production	0.85 – 1.3 % RGR per day	Peteiro & Freire 2013, Watson & Dring 2013; Azevedo et al 2016

G. pacifica may have a slower growing period for months December - February and in June given the nutrient concentration in Santa Barbara. As a peak growing season was available in the literature for the Southern California Bight, we relied on a combination of expert opinion, oceanographic monitoring data, and abundance estimates of native *Gracilaria spp.* in the area (D. Reed, pers comm, 2018; UCSB LTER). *Gracilaria pacifica* naturally occurs on Mohawk Reef near the Santa Barbara Harbor between the months of March and October with a peak between May and July (SB LTER). There are fairly low amounts of DIN (NO₃, NO₂, NH₃) between July and November (< 1.2 uM), which start to drop in June to 2 uM (UCSB LTER). The longest photoperiod is from March to October (UCSB LTER). Therefore, it is likely that *G. pacifica* is nutrient limited in the summer and limited by light availability in the winter time (Santelices, 1999). We found one case in Chile where *Gracilaria* is cultivated every 7 months, but here we use the average cultivation period of 6 months (Winberg, 2011). We determined that *G. pacifica*'s ideal harvest period would likely be March through June at a minimum, but extended it through August to obtain a six month cultivation season (Figure 1.5).

Data for onshore *G. pacifica* growth rates was sourced directly from a local aquaculture farm in Santa Barbara. Stocking density and growth rate were both taken from a SARC pilot growth study conducted onshore in Goleta, California during late summer and early fall of 2017 (J. Couture, pers comm, 2018). We used 6.9% and 5.48% as the high and low daily relative growth rate from that study. Stocking density associated with the reported growth was 3.17 kg/m³. Based on prior experience, *G. pacifica* has the ability to grow year round January to December in an onshore tank system (D. Bush, pers comm, 2018). We set a weekly harvest frequency (D. Bush, pers comm, 2018; Davison & Piedrahita, 2015).

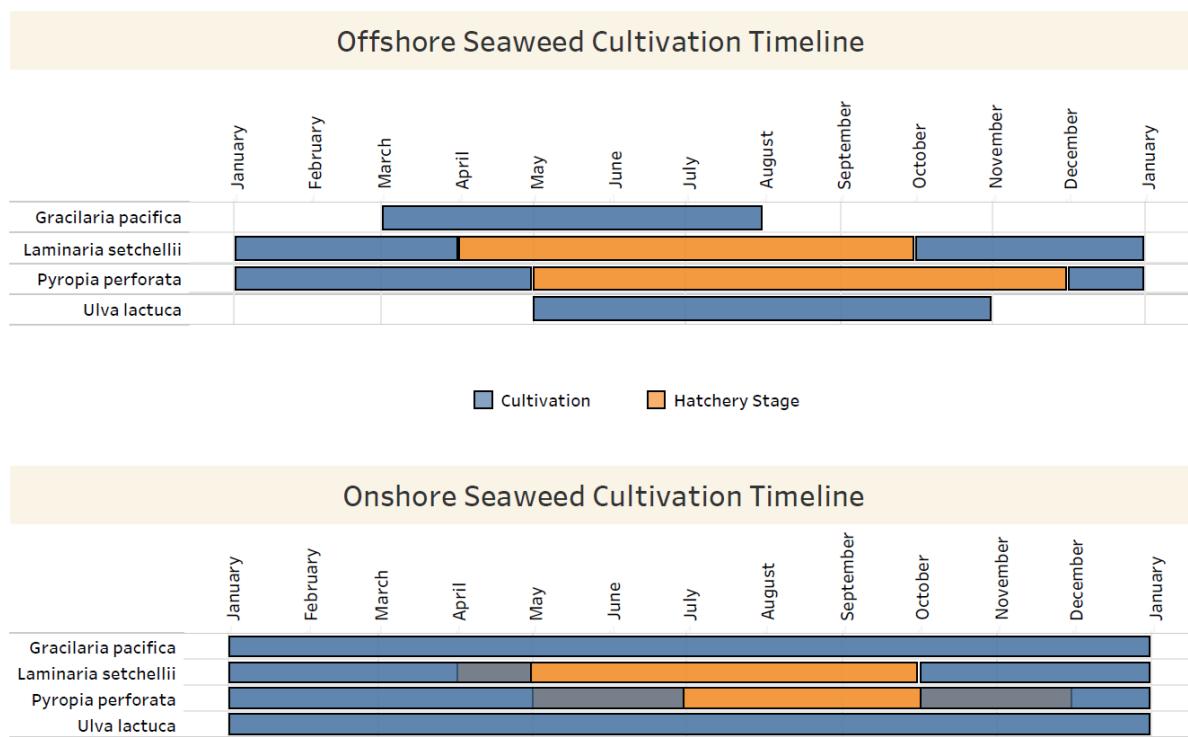


Figure 1.5. Onshore and offshore cultivation timelines for the four candidate species.

Ulva lactuca

Ulva lactuca is initially cultivated by allowing its spores to settle out onto nets, and we assumed an initial stocking density of 0.25 kg per m². Then we used a final reported yield of 0.5 – 1.0 kg dry weight per m² (Titlyanov & Titlyanova, 2010). Using an average conversion factor from literature of 6.67 kg wet weight to 1 kg dry weight, we determined the high and low wet weight yield to be 6.67 kg per m² and 3.3 kg per m² respectively (Azevedo et al., 2016; Hernandez et al., 2005; Roesijadi et al., 2008; Watson & Dring, 2007). From there, we back calculated the low and peak growth rates to be 2.6% and 4.96% per m² per day. While these growth rates from Titlyanov and Titlyanova (2010) were lower than other growth rates reported, the high growth rates observed in other studies were instantaneous and recorded only for a short period of time.

We determined the thermal optimum for *Ulva lactuca* to take place from May to October given the North Sea range of 15-20 °C (Figure 1.5). During this time, temperatures in Santa Barbara are over 15 °C and photoperiod is also the highest (between 13.97 and 11.33 hours). *U. lactuca* reproduces from June-August, but the reproductive phase occurs for only a short period during peak season and doesn't affect the harvest (M. Freidlander, pers comm, 2018). Natural abundances in Santa Barbara are generally highest around spring and summer (D. Reed, pers comm, 2018). *Ulva* can survive in low nutrient ranges as long as the temperature is below 20 °C, as is the case at our farm location (Duke et al., 1989). Therefore, we expect *U. lactuca* to still grow well during summer despite lower nutrient levels.

There were several onshore *Ulva* spp. studies available for *U. curvata* and *U. lactuca*, many of which reported extremely high growth rates. The estimates range from 10 and 65% relative

growth per day (Carl et al., 2014; Duke et al., 1989; Neori et al., 1991). In a constrained system, such as a tank or a densely packed offshore farm, density-dependent factors slow down growth or cause mortality (Neori et al., 1991). The low growth rate of 10% per day is a reasonable expectation, but we had to limit the high growth rate based on our physical farm constraints. First, harvest frequencies reported for *Ulva spp.* in onshore systems were in the range of 3-4 weeks, so we used an average of 3.5 weeks (M. Friedlander, pers comm, 2018; van den burg et al., 2012). Then, the initial (0.375 kg/m³) and maximum (27.8 kg/m³) stocking density were used to calculate the maximum growth rate over a single harvest period (Duke et al., 1989; Hernandez et al., 2005). The calculated high growth rate was 17.6% per day, which falls in the range of growth reported in literature. Similar to *G. pacifica*, *Ulva spp.* can persist vegetatively for at least a year from the same source, so stocking can occur yearly (Bolton et al., 2009; M. Friedlander, pers comm, 2018).

Pyropia perforata

We used a final yield value of 1100 sheets per net (18m x 1.5m net size) for our high growth estimate using a conversion factor of 25 sheets per 1kg of fresh *Pyropia perforata* (Levine & Sahoo, 2009; Parker, 1974). We assumed that four nets were seeded and deployed during the annual growing period (KMO, 1982). Yield values for *Pyropia spp.* vary greatly, but 1100 sheets per net is well within the lower production ranges found in literature. Given that *Pyropia spp.* has to be seeded on nets in a hatchery, stocking densities aren't relevant or reported. In order to calculate a growth rate, we needed a stocking density and assumed a value of 1kg/m². The average harvest period for *Pyropia spp.* is 37.5 days (Kim et al., 2017; Levine and Sahoo, 2009). We had to back-calculate a growth rate given these parameters to yield 1100 sheets per net. Our final value was 3.2% per day per m². For our low growth rate, we completed the same calculation but used a very conservative final yield of 500 sheets per net and calculated growth to be 1.05% per day per m² (Miura, 1975).

Through our literature review, we were not able to elucidate the nutrient requirements for *P. perforata*. *P. perforata* can withstand extremely high levels of nutrients, but there is no mention in any study of the minimum required nutrient input (Romero, 2009). At the genus level, *Pyropia spp.* prefer 100-200 mg/m³ of nitrogen for optimal growth, which is within the 43-200 mg/m³ available at the farm site in Santa Barbara (Chen & Xu, 2005). Light is overwhelmingly the most important factor for *P. perforata* growth and 8-16 hours of light per day is optimal for all *Pyropia spp.* (Pereira, 2006). Multiple studies suggest that 10-20 °C is the optimal temperature for *Pyropia spp.* (Kim et al., 2007; Pereira et al., 2006). There is no optimal temperature estimated/recoded specifically for *P. perforata*.

Wild populations of *P. perforata* are most abundant in mid spring to early fall, however this is not the best harvest season for the species (Romero, 2009). *Pyropia perforata* releases carpospores when photoperiods exceed 12 hours at low temperatures coinciding with upwelling periods (Pacheco-Ruiz et al., 2005). Photoperiods in Santa Barbara exceed 12 hours between April and September, upwelling occurs between March and May, and temperatures are lower in the spring (Brzezinski et al., 2013). Thus, sporing will likely begin in the spring and harvest should be completed around then to avoid allocation of metabolic energy to reproduction. Nutrients are lowest between July and November, so there is concern that *Pyropia* may not grow well during this period (Appendix A). We chose to set that harvest period as the harvest period of December to April as the best balance between all the environmental factors. This time period is colder in temperature, avoids photoperiods over 12 hours, and takes advantage of

the upwelling and higher nutrient availability. This is when *Pyropia* spp. are grown in Japan, but Japanese *Pyropia* spp. prefer cold water and ocean conditions are more variable in the Western Pacific (MacFarlane, 1966).

Onshore, we used the high and low relative growth rates of 16% and 10% per day per m³ from the only study to conduct research on four species of Pyropia in land-based tanks (Kim et al., 2007). Other studies have reported significantly higher relative growth rates than were stated in the Kim et al. (2007) study. However, this may be due to the fact that the other experiments used younger seaweed blades (< 10 cm) that uptake nutrients faster and grow more rapidly (Kim et al., 2007). The initial stocking density we used was from an onshore culture experiment in Israel (Israel et al., 2006).

The Israel et al. (2006) study found that peak growth for *Pyropia* spp. in tank culture was between December and March when temperatures ranged from 14 - 18 °C. Given that information, the most appropriate cultivation period in Santa Barbara is October to June where temperatures range from 14 – 17 °C on average (Appendix B). We used the harvest frequency of 2.5 weeks from that same study. However, they were only able to cultivate *Pyropia* for 20 weeks (4.7 months), which may be due to higher water temperatures (>20 °C) in Israel outside of that time frame that were unsuitable for *Pyropia* growth. Another concern was that a photoperiod of 12 or more hours could induce sporing. The use of shade structures could extend the cultivation period for an onshore farm (especially between April and June). Since water temperature in Santa Barbara reaches a max of 19.4 °C in July, cultivation should end in June when sporing in a hatchery can take place.

Laminaria setchellii

Similarly to *P. perforata*, the growth rate for *L. setchellii* had to be derived from final production quantities. We used an annual yield of 16 kg per meter of longline, taken from a study on *Saccharina latissima* (related species in the same family) in Spain (Peteiro & Freire, 2013). Unlike many other groups of edible seaweeds, it is common for Laminariales to be allowed to grow uninterrupted for an entire annual season before being harvested (Watson & Dring, 2013). We calculated a peak growth rate of 1.7% per day. We used a yield value of 7 kg per meter longline from a study on *Laminaria digitata* to calculate a low growth rate of 1.1% (Watson & Dring, 2013). Relative growth rates exist in the literature for *L. setchellii*, but they were observed in a controlled experimental setting and are likely less to be accurate than a final farm yield value. Given the large variation in reported growth rates across Laminariales, both *S. latissima* and *L. digitata* are likely to give us accurate first-order approximations for our local candidate species (Lane et al., 2006);

Wild abundances of *Laminaria* spp. occur between August to December at Mohawk Reef (SB LTER). *Laminaria digitata* cultivation was reported to be poor in the summertime after May, with lots of biofouling from epiphyte growth (Watson & Dring, 2013). The optimal cultivation period reported was from November to May. A recent SeaGrant report indicated that the best cultivation period for *Saccharina latissima* in the Southern California Bight is October to March, validating previous studies that have been done for *S. latissima* and *L. digitata* (Lester et al., 2014). This may be the best estimate for our local species since it is conducted near our study site.

Onshore, we selected high and low growth rates from Azevedo et al. (2016) in Portugal, which has a similar temperate latitude. Our high and low values were 0.85% and 1.3% per day per m³ respectively. We used a lower stocking density of 8 kg/m³ to help prevent excessive epiphyte development and photoinhibition, ensuring product suitability for high-value applications such as human food. Water temperatures during this growth period were between 15 – 16 °C, which was congruent with Santa Barbara ocean temperature (Azevedo et al., 2016).

We used similar cultivation period as offshore, leaving time in the summer to propagate seeded lines. Although the temperature ranges in Santa Barbara indicate we could cultivate *Laminaria* onshore year-round, it would be much slower in the summer between May and September given local water temperature (Azevedo et al., 2016). *Laminaria* grows best during April in the Azevedo et al. (2016) study, where ocean conditions were similar to what exists in Santa Barbara. We extended the cultivation period onshore to the end in April since it was still in the optimal temperature range established by Azevedo et al. (2016).

As the tanks were initially stocked with 8 kg/m³, harvest frequency depended on time it took to reach 12.6 kg/m³ (harvest weight). Therefore, harvest frequency varied with growth rate. We used the logistic equation provided in Hernandez et al. (2005) to get a harvest frequency of 15 days for the high growth rate (14 harvests annually over 7 months) amounting to 64 kg annually from each 1000 L tank. At the low growth rate, harvest occurred every 23 days (9 harvests annually), which produced 41.4 kg annually from each tank.

Discussion

Limitations to growth rate estimates

While going through the literature to identify suitable growth rate estimates for our candidate species, we encountered a number of challenges we believe, if addressed, will improve the transferability and interpretation of seaweed aquaculture research in the future. There is significant inconsistency in the way stocking density of seaweed is reported in literature, which makes it difficult to use in the context of aquaculture. The three main farm structures we used in our project are longline, nets, and tanks. Each of those has inherently different measurement dimensions. Preparing the seaweed in each of those units would be easiest if the stocking density matched the unit dimensions of those structures. For example, if a seaweed growth study is conducted on longlines, the stocking density in kg per meter should be reported. If a study is conducted in tanks, the stocking density in kg per meter cubed should be included.

The duration of many seaweed growth experiments only last a few days or a few weeks. Results often try to capture maximum growth rate, which is induced by creating specific environmental conditions in lab (e.g. high nutrient concentration). In nature, it is rare for these maximum reported daily growth rates to be sustained for a long period of time. Determining maximum potential of seaweed growth is important for identifying optimal conditions. However, when applied to aquaculture operations, the conditions are often limited to regional patterns or ability to cover costs required to replicate those optimal conditions. Maximum stocking density is

rarely reported, but very important to determining the constraints of a farm and how often a species should be harvested to prevent fouling or overcrowding.

As we continue to research local species for suitability in aquaculture systems in California, certain standards of reporting need to be established to prevent additional roadblocks. It is vital that applied seaweed aquaculture research heavily incorporate aspects from farmers, phycologists, and policy makers. Research in applied aquaculture can be made more adaptable to other regions by including detailed information about factors that influence seaweed growth such as temperature, nutrients, and light. Our project summarizes key building blocks in seaweed aquaculture research and identified available data for each. As a next step, it would be beneficial for future studies to confirm the reported data for species in our region.

Conclusion

The rich coastal waters off the coast of California house a grand biodiversity of seaweed species. We identified the four most promising aquaculture candidates (common name in parentheses) for direct human consumption. These species occur naturally throughout the entirety of the Southern California Bight, have an existing food market in California, and have documented aquaculture cultivation techniques: *Gracilaria pacifica* (ogo), *Ulva lactuca* (sea lettuce), *Pyropia perforata* (nori), and *Laminaria setchellii* (kombu). We found that tank culture was the most suitable grow out method for all four species onshore through consultation with literature and industry experts. Offshore, the choice of growth structure hinges on intrinsic species properties and prevailing oceanographic conditions. Our research supports the use of longline culture for *G. pacifica* and *L. setchellii* and net culture for *U. lactuca* and *P. perforata*. This abbreviated list can serve as a baseline for further seaweed aquaculture research and investigation in the region.

2. Feasibility of seaweed aquaculture in southern California

Introduction

At its core, any farming operation is a joint venture in biology and economics. In the first chapter, we outlined the relevant biological parameters that will determine the productive capacity of a seaweed aquaculture system. In this chapter, we turn our attention to the associated costs and potential profits to project the economic feasibility of seaweed aquaculture in the Southern California Bight. These depend on the respective seaweed species and their realized productivity, the capital and operating costs, and the sales prices that can be achieved. The combination of these factors will either enable or restrict a potential seaweed industry's ability to compete successfully within the existing market. We constructed a bioeconomic model to synthesize this information and provide stakeholders a mechanism to investigate patterns and draw conclusions about different potential farm systems.

Bioeconomic models are a tool by which complex natural systems can be simplified into the key relationships that relate their physical, ecological, and economic components (Allen et al. 1984). Furthermore, bioeconomic models can be paired with sensitivity analyses to allow researchers to identify how strongly certain input parameters affect model output. For seaweed aquaculture, this is valuable as it illustrates to resource managers and potential farmers where efforts should be focused to reduce uncertainty or alter inputs to maximize economic/biological outcomes.

Costs associated with seaweed aquaculture can be broken down into initial capital costs and annual operating costs. Choice of farm structure, type of materials, species cultivated, labor force, and farm location can all influence economic feasibility significantly. Keeping costs low is necessary to ensuring the profitability of a farm (Watson & Dring, 2011). The species grown and farm location have a huge impact on the total costs. Seaweeds that are cultivated through a hatchery stage require additional infrastructure. Onshore and offshore farm systems in particular have different fixed independent (e.g. environmental assessment), fixed dependent (e.g. structural costs), and marginal costs (e.g. labor). While some costs, such as annual registration, are fixed regardless of size ("independent"), many costs depend on the size of the farm ("dependent"). Identifying comparative, average farm sizes for an onshore and offshore system that have reasonable initial capital costs was crucial to our analysis. Fixed costs for onshore systems require materials that are more energy intensive, since the system aims to replicate ocean conditions, such as current flow and optimal photoperiod (Pereira & Yarish, 2008). Given that needed infrastructure for seaweed or other aquaculture species does not exist yet in California, overall costs will likely be greater than for those in countries and regions that have developed aquaculture industries (Watson & Dring, 2011).

Prices obtained for seaweed used for human consumption vary greatly across markets and seaweed species (McHugh, 2003; Walsh and Watson, 2011). In the global market, prices are typically low and local seaweed producers would be competing against low-value imports to

access this market (Kim et al., 2017). The average price of edible dried seaweed that is imported into California is \$13.38/kg, with the majority sourced from South Korea and China (NOAA Fisheries, Fisheries Statistics Division, 2017). With the difficulty of competing on the global scale, seaweed farmers are tapping into the growing food-to-table movement to access the value-added seaweed market that sells to high-end restaurants or the local produce market to achieve a price premium (Mouritsen, 2012). In Hawaii, fresh *Gracilaria* has been collected and sold as a salad vegetable for several decades, due to a unique cultural history (McHugh, 2003). Some species are significantly more valuable than others. The variation is largely due to nutritional content and food usage (Nayar & Bott, 2014). For example, *Pyropia* is by far the most valuable seaweed, while *Laminaria* usually fetches a lower price (Nayar & Bott, 2014). Identifying the markets that a seaweed industry in southern California can access will inform industry development, species selection, and cultivation methods.

We developed a bioeconomic model to identify the economic feasibility of establishing different types of seaweed farms in Southern California. The species-farm combinations we investigated include *Pyropia perforata*, *Gracilaria pacifica*, *Ulva lactuca*., and *Laminaria setchellii* in both onshore and offshore locations.

Methods

Model inputs

Biological parameters

The first section describes the way in which we collected the relevant growth information from primary literature and field experts. Relative daily growth rate was the main input into our model. It was represented as additional biomass accrued, expressed in percentages of the previous day's total. After using the collected values to calculate total annual yield, we evaluated them against a range of empirical farm yields from literature (Appendix D) and industry-established harvest frequencies (Appendix C). For any large discrepancies between annual yield predicted and the range of empirical yields from literature, we re-evaluated our inputs to see if there was anything biasing our results. We chose to examine the model's sensitivity to yield by running three deterministic scenarios with low, medium, and high growth rates respectively (Appendix C). The reproductive nature (vegetative or sexual) and the main abiotic factors that limit growth in each candidate species were used to calculate the productivity of our model farm systems.

Farm size

We set our offshore model system to be 25 acres (10.4 hectares), the same operating size as our client's offshore mussel farm. We then needed a way to equitably compare the offshore and onshore model systems. We first calculated the annual yield in an offshore system for each species using the mean growth rate, an average of our reported high and low values. That yield was used in conjunction with the onshore information (e.g. stocking density, growth rate) to determine number of tanks needed to match the productivity estimated offshore. We did this on a per-species basis using the mean growth rate specific to each location. This allowed us to

scale the costs of the onshore aquaculture system to an appropriate level for that amount of production.

Another factor we had to consider for comparison was the means of acquiring land or space in the ocean. As stated in § 15405 of the California Fish and Game code, there is a 25 year limit to leases of the marine benthos for projects in state waters (CA Dept. of Fish & Wildlife, 2007). This forces offshore aquaculture permitting to take on a structure akin to long-term renting. It is possible to purchase land for development onshore, but land prices vary highly with location and represent a substantial capital investment. Instead, it is more likely that a farmer who wanted to grow seaweed on land would try to rent space from a port, harbor, or other coastal organization.

For example, the Natural Energy Laboratory in Hawaii leases space on their Host Park property, which gives tenants access to the existing seawater pump, electricity, and an already largely permitted area (NEHLA, 2016). To become a tenant at the Host Park, interested parties must submit a business plan and apply for some additional development permits through the State of Hawaii, and if approved, will be leased the land for a 30-year period. Costs for Host Park include annual leasing fees, a fraction of annual revenue (i.e. 5% for commercial use), and initial construction costs such as installing electricity hook-ups and a smaller access pipe for seawater. Several port districts including San Diego, Los Angeles, and Ventura have shown interest in the development of infrastructure to support aquaculture ("AltaSea at the Port of Los Angeles", 2018; "Blue Economy Incubator", 2016; "Ventura Shellfish Enterprise", 2017), but are still in planning phases. Port of Hueneme leases space to a business that cultivates limpets for the pharmaceutical industry, but information for the system in place in Hawaii was more immediately available (Stellar Biotechnologies, Inc, 2011). Therefore, we used the latter to help structure the leasing costs for our onshore model farm.

Cost data

We conducted a literature review and consulted with experts to estimate the costs of on and offshore seaweed farms and logistical requirements for each of the cultivation methods (Appendix E). The most applicable and detailed cost information came from Petrell et al. (1993) which utilized significant cost information from a study of seaweed farming in Washington state in the 1980s (Druel, 1980). Due to the limited availability and considerable variation in relevant cost data, we also consulted with six aquaculture experts (farmers and researchers) to validate costs (D. Marquez, D. Bush, S. Lindell, D. Bailey, B. Friedman, N. Caruso). Earlier, we determined a net structure for *P. perforata* and *U. lactuca* and longlines for *G. pacifica* and *L. setchellii* were the ideal offshore grow-out methods (Kim et al., 2017, see Chapter 1; Levine & Sahoo, 2009; Peteiro et al., 2016; Redmond et al., 2014; Titlyanov & Titlyanova, 2010). Onshore cultivation for all species occurred in tanks, as this method is predominantly used in California and developed nations (Kim et al., 2017).

Since *G. pacifica* and *L. setchellii* are both longline species, we used costs from a *Laminaria digitata* longline seaweed farm in Ireland for both scenarios (Watson & Dring, 2011). Costs were converted to US\$ (exchange rate 1 euro = \$1.19) and adjusted for inflation (\$1 in 2011 equal to \$1.11 in 2017) (Bureau of Labor Statistics, 2017). Since more equipment (e.g. longlines,

anchors) was needed in our model farm, cost estimates were calculated at the \$/m of longline level and scaled up. Our model farm consisted of 125 longlines that were each 137m long, which mirrored the dimensions of the mussel longlines of our client. Therefore, we felt confident that they would be capable of existing in the offshore environment of the Santa Barbara Channel. Structural requirements were modeled using those currently in place at our client's farm, substituting Jeyco anchors for the concrete blocks used in this study. Fixed costs besides structural costs included hiring a consultant for the Environmental Impact Report, purchase of a boat, and construction costs for the hatchery, if applicable. Labor estimates for a similar model farm size were not provided in any available studies, and thus, expert advice for the effort needed to harvest and deploy lines was used. For 25-acres, the initial deployment (i.e. 'seeding') was estimated to cost \$3,000 (\$120 per acre) and harvesting costs were \$6,000 (\$240 per acre) with contribution from the owner operator and an additional employee (\$15/hr) (D. Marquez, pers comm, 2017). Other marginal costs include gas for boat, transportation of product, and annual permit fees.

Costs for *P. perforata* and *U. lactuca* were obtained from research conducted in Washington State assessing *Pyropia* culture (KMO, 1982). Despite an extensive literature search, no more recent source provided sufficiently detailed cost information. Costs were also adjusted for inflation (\$1 in 1980 = \$2.58 in 2017, Bureau of Labor Statistics). *Pyropia* culture in Washington was estimated for a protected intertidal region, which meant structure design was not appropriate for our exposed offshore region (Levine & Sahoo, 2009). Therefore, considerable modifications of the structure were required. Cost estimates for offshore net cultivation consisted of values from SB Mariculture's longline system and the fixed pole net structure available from KMO (1982). Under the new idealized structure design, nets were suspended from buoys used in the longline structure, and anchored into the muddy bottom using Jeyco anchors. Given that nets should occupy approximately 1/12th of the available cultivation area, 300 traditional saku nets (18m X 1.5m) were placed into the 25-acre farm and arranged parallel with the current (KMO, 1980; Levine & Sahoo, 2009). Regarding marginal costs, the cultivation of *Pyropia* is extremely labor intensive, and thus employee labor requirements for a 300-saku net farm were used from KMO (1982) while the same employee requirements for the longline cultivation were used for *U. lactuca*. Additional costs that varied between species included the operating and initial capital required for a hatchery and a drying machine for *P. perforata* and *L. setchelli*, since these species are typically sold as a dried, edible product (Watson & Dring, 2011; D. Marquez, pers comm, 2018; D. Bailey, pers comm, 2018).

Reliable and applicable estimates of onshore costs are extremely limited and typically confidential. We chose to use estimates for tank cultivation from a single, reliable study (Watson & Dring, 2011). Costs were validated and improved upon by two experts (D. Marquez & D. Bush, pers comm, 2018). Tank costs were estimated for operating a single 1000-L tank, which enabled easy scaling of costs with increasing seaweed production. Labor estimates were not provided in Watson & Dring (2011). We assumed one employee (\$45,000) could manage 60 tanks annually (D. Bush, pers comm, 2018). The lease structure, annual costs, and construction costs from NEHLA were used (NEHLA, n.d.). Finally, calculated prices were in units of wet weight to allow direct comparison between seaweed that is typically sold fresh (*Gracilaria* and

Ulva) and dried (*Laminaria* and *Pyropia*). However, farm systems growing *P. perforata* and *L. setchellii* include drying in their cost estimates.

Model outputs

Annual Production

We began by calculating the initial seaweed biomass (kg wet weight) of our different farm systems by multiplying the stocking density of the candidate species by the relevant structural unit (meter of longline, square meter of net, or tank). Seaweed growth studies commonly calculate a mean daily growth rate by comparing the final yield to the initial stocking density over the course of the experimental period (Duke et al., 1989; Santelices and Doty 1989; Yang et al, 2006; Azevedo et al., 2016). Rearranging that formula allowed us to project biomass yields with our estimated growth rates:

$$\text{biomass}_{\text{final}} = \text{biomass}_{\text{initial}} * e^{(\text{relative daily growth rate} * \text{numbers of days})} \quad (\text{one harvest period})$$

The number of days the seaweed was allowed to grow was dictated by the harvest frequency. To determine the final annual yield, we multiplied the production of one harvest by the number of harvest periods in one season. Species-farm scenarios that required reproductive cultivation by seeding didn't have initial stocking densities, since the standard is to report number of spores rather than a weight. Final yield was equally vital to our analysis and was used as our growth estimate value. Daily growth rate and initial stocking density for the species that spore were back-calculated from the final yield (See first chapter for further information on how estimates for growth were selected).

Net Present Value

Using a five year time horizon, the net present value of each of our farm systems was calculated by summing up the discounted revenues and expenditures over that time interval. Initial startup costs were accounted for in year 1. We assumed that all the seaweed produced could be sold at a single price point and sales were modeled in discrete annual time chunks. We did not investigate the potential effect of seasonality or flooding the market on sales. We chose to incorporate the costs of drying *P. perforata* and *L. setchellii* as they are more commonly sold in this manner. However, we converted the price point for the dried product as well as the final dried production back to their fresh weight equivalents for ease of comparison across species in our results and figures. We selected a dry weight to fresh weight conversion ratio of 1:6.67 (15%) as an intermediate of values found throughout literature (Azevedo et al., 2016; Hernandez et al., 2005; Roesijadi et al., 2008; Watson & Dring, 2007). A discount rate of five percent was used in this model. The five year time horizon we investigated was chosen to match the interval mandated by the California Coastal Act of 1976 during which local coastal programs must be reviewed by the California Coastal Commission (§ 30519.5). We decided that

this periodicity of the permitting and review process lent itself well to evaluating the feasibility of aquaculture operations.

Break Even Price

We determined the price point needed to have a net profit of \$0 at the end of five years. This “break even” price was proportional to the costs and amount of production for each model farm system. In examining these values, stakeholders are able to gauge the market that they would have to be able to enter to garner a profit in at least a five year time horizon. This translates to valuable information about the quality of product they would need to deliver and the necessary operational considerations to achieve such quality. A discount rate of five percent was also applied throughout these calculations.

Results and Analysis

The production predicted by our model within an offshore and onshore system varied across species. *L. setchelli* offshore had the greatest mean annual production (180 metric tons), followed by *G. pacifica* (52), *U. lactuca* (38), and *P. perforata* (36). The number of 1000-L tanks required to match offshore production for each species were *L. setchelli* (3389), *P. perforata* (764), *G. pacifica* (565), and *U. lactuca* (238). The productivity of the species was a considerable factor influencing economic feasibility.

Break-even price point at year five

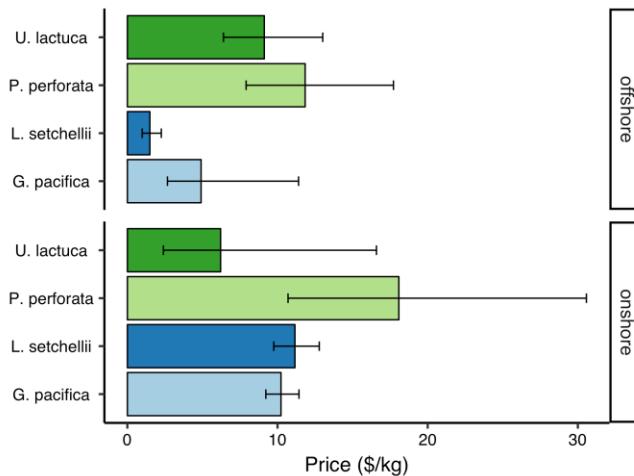


Figure 2.1: Break-even market price (\$/kg) at the end of 5-years for all candidate species both onshore and offshore. For direct comparison of fresh and typically processed seaweeds, kg units are all fresh weight. The colored bars represent the mean growth rate scenario and the bounding bars show the low and high growth rate scenario.

The market price needed to break-even after five-years for a species differed between the two growing locations. Offshore had a lower break-even price-point than onshore tank cultivation for

three out of four species (Figure 2.1). The mean values of the break-even price across all species were \$6.84/kg for offshore and \$11.42/kg onshore. *U. lactuca* was the only candidate species where the break-even price was lower onshore than offshore, at \$6.21/kg and \$9.12/kg, respectively. Additionally, it had the smallest price difference between cultivation sites across all four species. The largest difference in the break-even market price between on and offshore for the same species was *L. setchellii*, followed by *P. perforata*.

There was strong inter-species variation of the break-even within the onshore and offshore groups (Figure 2.1). Across all four species, *P. perforata* had the highest break-even price for on and offshore cultivation (\$18.08/kg and \$11.84/kg), and *L. setchellii* offshore (\$1.05/kg) and *U. lactuca* onshore (\$6.21/kg) were the least expensive to produce. The break-even price of *L. setchellii* offshore in our study is lower than offshore cultivation of *L. digitata* or *Saccharina latissima* reported in Ireland, which required a market value of \$2.97/kg (Dring et al., 2013). However, that study considered a three year time horizon. In offshore areas adjacent to wind-farms, the break-even price for *L. digitata* was estimated at \$1.74/kg dried (van den Burg et al., 2016). Reasons for the slight disparity between our calculated break-even price and those found in literature are related to the growth rates and scale of the farm operation.

The economic feasibility of the seaweed species depends upon the annual farm yield. Higher growth rates increase farm yields leading to more per acre productivity. This lowers the farm gate price, or price received by the seller, that farmers must obtain to begin profiting in five years (Figure 2.1). Considerable deviation between profitability under high and low growth rate scenarios for *P. perforata* and *U. lactuca* occurred, particularly onshore. Conversely, the growth rate ranges for *G. pacifica* and *L. setchellii* were narrower onshore. The offshore growth rate range for *G. pacifica* was large due to the difference between our relatively high growth from experimental results and lower growth values from experiments in Chile where water temperatures were cooler (Halling et al., 2005). Growth rates used in our model reflect the large variability of values obtained in literature under similar environmental conditions. This highlights the need for field-validated growth rates for our species under the environmental conditions found in the Southern California Bight.

Onshore versus offshore cultivation methods

The projected cash flow, or timeline of costs and revenues, over a five-year period differed between onshore and offshore for all species in our model (Figure 2.2; Appendix F). The strongest factors influencing break-even price for the candidate species for both on and offshore cultivation were initial production costs and crop productivity. Production cost includes the upfront structural costs of longlines/nets, buoys, tanks and size independent costs such as permits. Additionally, operating costs such as labor, annual permits, gas or electricity fees from pumping, and transportation of product contributed substantially to costs (Figure 2.2). Our costs for offshore regions fit within ranges reported in other regions in the world, such as nearshore areas in the Netherlands where initial costs for a 25-acre farm range from \$353,600 - \$1,076,400 (van den Burg et al., 2016). The paucity of cost information for onshore systems prevents us from comparing our cost information.

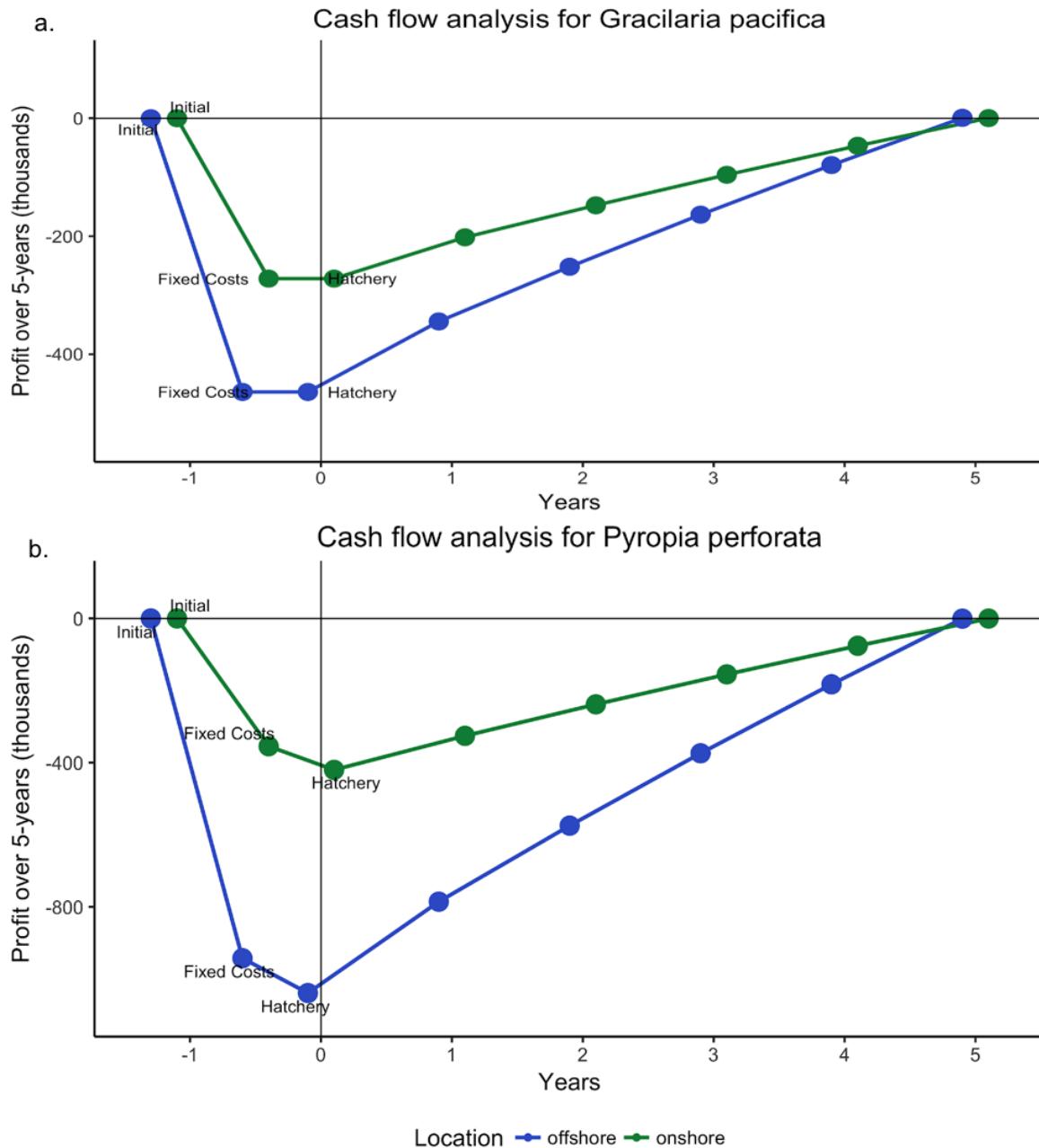


Figure 2.2. The cash flow (thousands of \$US) for the payback of a) *Gracilaria pacifica* and b) *Pyropia perforata* at the break-even price under mean growth scenarios at 5-years in onshore (green) and offshore (blue) locations (see Figure 2.1 for break-even information). The upfront capital costs are labelled on the graph, and include fixed costs such as permitting, structural costs, and costs associated with establishing a hatchery when needed (as for *P. perforata*).

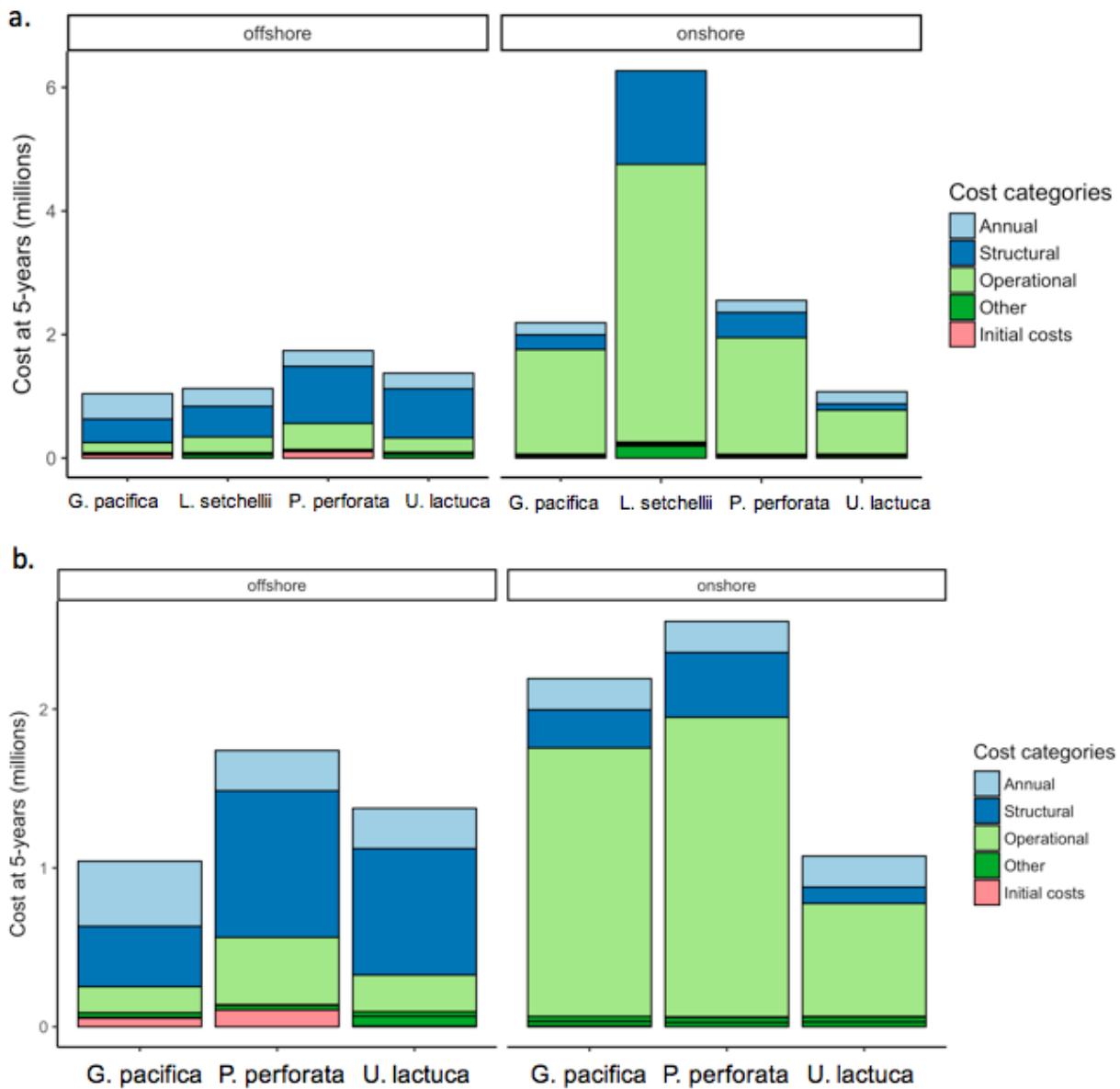


Figure 2.3. Allocation of the costs associated with each species-location combination over a 5-year period. The annual and marginal costs have been calculated over 5-years and discounted (0.05). The other category is for cost categories that comprised less than 5% of the total cost for each species. Figure (a) shows the cost including *Laminaria setchellii* onshore, and (b) excludes *L. setchellii* onshore for better visual analysis of cost composition.

The fixed costs for the onshore system were less expensive than fixed costs offshore between all species except *L. setchellii*. However, over the five-year time horizon, the larger marginal costs of onshore production resulted in lower break-even prices for offshore despite greater initial costs. For *G. pacifica*, the onshore scenario incurred startup costs that were \$200,000 less than offshore. The greater annual profitability offshore due to lower operating costs was sufficient to overcome the larger fixed costs and surpass the onshore scenario after 4.5 years.

Initial offshore costs were three times higher than onshore in another candidate species, *U. lactuca*. Initial costs for *L. setchellii* were uniquely five times higher onshore than offshore due to the requirement of a hatchery for the offshore system and the large number of tanks that were required for *L. setchellii* (Appendix F). Upfront costs associated with hatcheries increased the costs by \$90,000 for offshore species when a lease for land and hookup to water had to be purchased (e.g. NEHLA lease structure costs) or \$60,000 when the hatchery was adjacent to onshore tanks. Among the onshore systems, *G. pacifica* and *U. lactuca* did not require hatcheries. The driving factor for differences in profitability between on and offshore is the respective productivity of the species in the different environments and the high marginal costs associated with onshore production that reduced profitability over time for most species.

There was strong variability in composition of costs between onshore and offshore cultivation (Figure 2.3). As seen through the breakeven price, costs were considerably lower for offshore compared to onshore over a 5-year period. Onshore structural costs comprised on average 30% of the total costs, although this varied across species. Marginal costs (primarily labor) were the second largest remaining cost. Conversely, fixed dependent costs for offshore, which included nets, longlines, buoys and anchors were the dominating cost in this cultivation scheme. Marginal costs were substantially lower offshore, largely due to the lower labor requirements for crop maintenance offshore. We speculated that onshore systems require more monitoring to ensure all the pumps, aerators, and light fixtures are working properly. This is especially true for species such as *L. setchelli* that only require deployment and a single harvest of all crops offshore. Marginal costs are greater for *P. perforata* because of large labor requirements for harvests that are more frequent and the nursery phase where daily maintenance requires the raising and lowering nets.

Between the two offshore systems, the cost of the net structure was almost double that of the longline structure, which greatly increased the fixed dependent costs associated with *U. lactuca* and *P. perforata* (Figure 2.3). Adapting the net structure to an offshore environment increased buoy costs and doubled the number of anchors used compared to longline culture of *G. pacifica* and *L. setchelli* (Figures 2.3 and 2.4). This system may be more suitable for a nearshore, shallow environment. For *P. perforata* and *U. lactuca*, buoys, anchors, and anchor lines contributed to at least 5% to the total initial costs structures. Both anchors and buoys were major material costs for the longline grow-out system as well, but hatchery construction and the Environmental Impact Report were the next largest costs for *L. setchellii* and *G. pacifica* respectively (Figure 2.4). The bulk of the fixed cost for the onshore system was tanks and equipment installation. Costs associated with different species in the offshore cultivation system will vary considerably as structures must be adapted to the environmental conditions and particular species grown.

We were unable to estimate the cost for several aspects of on and offshore cultivation. Therefore, our costs are likely to underestimate the true value. For example, the cost of building and installing the tank system were not included for our model onshore farm. These construction costs include laying connecting pipes to the main seawater pump and any grading of the land or other construction requirements that would be required. Maintenance fees were

also not included, as they were unavailable. For offshore cultivation, the largest uncertainties came from missing estimates and assumptions made for *P. perforata* culture. We assumed that the estimated costs of the nursery frames for *Pyropia* culture in more inshore regions is representative of costs for an offshore nursery system that would enable the farmer to expose nets to the air. Additionally, the hatchery costs for all species are likely underestimates. The missing costs for the hatchery, particularly for *P. perforata*, include the cost of the oyster shells to seed the conchelis phase, a giant seeding wheel, and the freezer where nets will need to be stored until deployment (Levine & Sahoo, 2009; Blouin et al., 2011). Furthermore, costs around labor are uncertain, highly variable, and will be affected by the difficulty of harvesting and deploying material within the environment (van den Burg et al., 2016). Thus, our labor estimates will need to corroborated as the industry progresses. However, as Watson and Dring (2011) provide the most thorough cost information, our estimates are as accurate as is possible to obtain from literature. As with any economic feasibility assessment, changes in various parameters such as farm distance from harbor, wave exposure, or construction costs will drastically affect costs and therefore feasibility. Thus, we recommend interested parties use our research and framework, but tailor costs to their set up to improve the usefulness of the feasibility assessment.

Species variability in costs

L. setchellii had the greatest costs onshore, but the lowest costs offshore. This is due to the high productivity in offshore system (final yield per harvest: 7-11 kg/m longline), which required more than 3,000 tanks onshore to match. The productivity of *L. setchelli* onshore, based on the values from the only study available, was relatively low compared to other species cultivated onshore, such as *U. lactuca* (Neori et al., 1991, Azevedo et al., 2016). In many onshore and offshore aquaculture systems, *Ulva spp* are a major biofouling organism due to their high productivity (Fitridge et al., 2012). Controlling *U. lactuca* is a major problem for onshore cultivation of *G. pacifica* onshore (D. Bush, *pers comm*). This high productivity of *U. lactuca* onshore explains its low break-even price onshore and why it is more profitable onshore than other species. The poor profitability of *P. perforata* onshore and offshore is due to the high fixed costs and labor requirements offshore and low productivity per tank onshore. Across both onshore and offshore regions, *G. pacifica* was the second cheapest species to produce due to relatively high growth rates and not having any hatchery related costs (Oliveira et al., 2000; Abreu et al., 2011). However, having to purchase *G. pacifica* stock annually from a nearby producer for \$6/kg substantially increased annual costs (Figure 2.3). These costs could decrease if a bulk price rate for purchasing a constant supply of *G. pacifica* from the same supplier were arranged. Reducing capital and operating costs is one way to improve the economic feasibility of seaweed aquaculture in southern California.

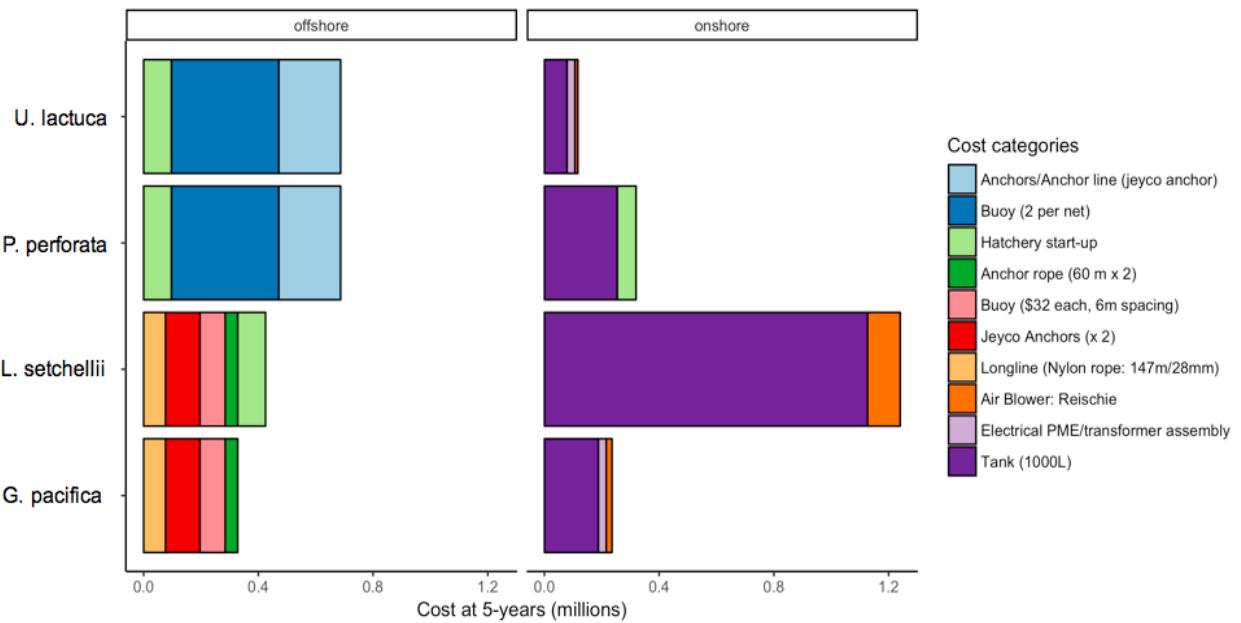


Figure 2.4. Fixed independent and dependent costs that contributed more than 5% to the initial cost for all species-location combinations.

Assessing the seaweed market opportunities

Market prices are a significant factor influencing seaweed aquaculture feasibility (van den burg et al., 2016). Break-even market-price values from Figure 2.1 can help indicate which market future seaweed farmers in the Southern California Bight must target in order to be viable. Under mean growth scenarios, the break-even price point ranged from \$1.50/kg for *L. setchellii* offshore to \$18.08/kg for *P. perforata* onshore (Table 2.1). Comparing the break-even price against global, US, and local market prices, seaweed will need to be sold into the high-end local market to restaurants or as a value-added produce item as in Hawai'i for the farm to begin to profit at 5-years (Tables 2.1 and 2.2). All species-location scenarios would need to access a local market except for *L. setchellii*, offshore, *G. pacifica* offshore, and *U. lactuca* onshore. These can be profitable within five-years at market prices similar to those in Hawai'i. None of the seaweed-location scenarios are able to produce seaweed at sufficiently low cost to access the global market. Therefore, local and regional consumer demand for seaweed will be a critical component in determining the economic feasibility of a seaweed industry in the Southern California Bight.

Table 2.1. Break-even market price for species at 5-years for mean annual production scenario.

Species	Location	Price (\$/kg)
Pyropia perforata	offshore	\$11.84
Pyropia perforata	onshore	\$18.08
Ulva lactuca	offshore	\$9.12
Ulva lactuca	onshore	\$6.21
Gracilaria pacifica	offshore	\$4.91
Gracilaria pacifica	onshore	\$10.23
Laminaria setchellii	offshore	\$1.50
Laminaria setchellii	onshore	\$11.16

Table 2.2. Market price (\$/kg) for seaweed species in the global, California, and US market. The costs associated with the different processing procedures (i.e. dried, raw) across species and markets are presented. Wet local values for *Pyropia perforata* and *Laminaria setchellii* were back-calculated from the dried price, assuming a 1:6.67 kg conversion from dried to wet biomass (Azevedo et al., 2016; Hernandez et al., 2005; Roesijadi et al., 2008; Watson & Dring, 2007).

Species	Location	Type	\$/kg	Reference
<i>Porphyra/Pyropia</i>	Global	Dried Wet	\$16 \$0.52 - \$2.40	McHugh, 2003; Nayar & Bott, 2014; Kim et al., 2017
<i>Laminaria spp</i>	Global	Dried Wet	\$2.80 \$0.14-\$0.42	McHugh, 2003; Nayar & Bott, 2014; Kim et al., 2017
<i>Gracilaria spp</i>	Global	Wet (agar)	\$0.27-\$1.20	Nayar & Bott, 2014; Kim et al., 2017
<i>Ulva lactuca</i>	Global	Dried Wet	\$0.40 \$0.07	Wholesale through Alibaba, 2017 van den burg, 2016
<i>Gracilaria spp</i>	Hawai'i	Wet	\$6.60	Paepae o He'eia, website
<i>Pyropia perforata</i>	Local	Dried (bulk) Wet	\$148.00 \$ 22.34	Strong Arm Farms, pers comm, 2017
<i>Laminaria setchellii</i>	Local	Dried (bulk) Wet	\$136.00 \$20.46	Strong Arm Farms, pers comm, 2017
<i>Gracilaria pacifica</i>	Local	Wet	\$15.40	Cultured Abalone Farm & Carlsbad Aquafarm (pers comm.)
<i>Ulva lactuca</i>	Local	Wet	\$15.40	Monterey Bay Seaweeds (pers comm). True price not shown for confidentiality reasons.

Our model found market price, in addition to cultivation costs and seaweed productivity, to be a crucial aspect impacting economic feasibility of seaweed cultivation. In Ireland, it was found that doubling the market price of *L. digitata* (from €1 to €2/kg) made a 100-ton seaweed farm economically feasible within 3-years (Dring et al., 2013). Additionally, in order for the offshore cultivation of *L. digitata* on a wind farm to be viable, market prices would need to increase 300% from current levels (van den burg et al., 2016). Research assessing the feasibility of seaweed farms in Europe and the US (Alaska and New England) concluded that seaweed must be sold into value-added markets to increase farm's profitability (Peteiro et al., 2016; Walsh & Watson, 2013; McDowell Group, 2017; Yarish et al., 2017). As studies in Europe and the US have highlighted, a viable seaweed industry in the Southern California will need to produce seaweed of sufficient quality to access a high-end value added market.

Under current demand, it is highly unlikely that 35 - 180 metric tons of seaweed would be absorbed annually by the high-value market in Southern California. Local production of fresh seaweed within the region is probably already above saturation for the edible market (D. Bush, pers comm). However, wild harvesters selling dried, processed seaweed in northern California have been able to sell significantly more to the local, high-end market (H. Herrmann, pers comm, 2018). There may be other reasons that the businesses here appear saturated, such as marketing strategy or food permitting issues. There is a lack of food safety regulations around raw seaweed that would normally enable producers to sell at retail outlets such as Whole Foods (D. Bush, pers comm). Another considerable hurdle is the relatively small demand for local seaweed products. While the importation of seaweed products into the US grew 65% between 2000-2008, the majority of this growth has come from processed seaweed products from Asia, such as nori packets (Nayar & Bott, 2014). The next step in assessing the feasibility of a seaweed industry in the region will require a market analysis to better understand current consumer demand in terms of quantity and desired product forms. Furthermore, steps to increase local demand and transfer demand from imported products to local value added seaweed products will be necessary. Transforming seaweed from a niche imported and irregularly consumed product to a value-added produce item that is able to capitalize on the farm-to-table movement could greatly improve the prospects of seaweed aquaculture in southern California and the United States more broadly.

Model sensitivity to changes in growth rate, risk and market price

Profitability of a seaweed farm will be strongly influenced by the productivity (growth rates and risk) and the market price that can be achieved. Our model was highly sensitive to changes in the growth rate, as seen in the break-even price graph (Figure 2.1). For example, the range for break-even market prices for *P. perforata* onshore was from \$10.70/kg to \$30.57/kg depending on realized productivity. Under mean growth rates, offshore profited at the lower price, however, onshore can be more profitable if high growth rates are achieved there. Growth rates for a species in the same environment can have significant temporal variation. Therefore, experiments to determine ideal site locations and species should occur over multiple years.

Other major factors that affect the per area yield include biofouling, extreme oceanographic events such as storms or temperature spikes, and suite suitability for each species (Pereira & Yarish, 2008; Fritridge et al., 2012; Radulovich et al., 2015). Both onshore and offshore aquaculture suffer from varying degrees of biofouling, which can have severe negative impacts on crop yield. During the nursery period of *Pyropia* aquaculture, nets are exposed to the air for 6 hours daily to prevent epiphytic organisms such as *Ulva spp* from overwhelming the nets and stunting juvenile *Pyropia* growth (Sahoo & Levine, 2009). Threat of biofouling by epiphytes often forces farmers to harvest crops even if environmental conditions are favorable for continued growth (Petiero et al., 2016). Therefore, understanding growth rates of the cultivated seaweed species and timing the outplanting and harvest to minimize biofouling is essential.

Risk is another concern for seaweed aquaculture, particularly in offshore environments (Petiero et al., 2016). Large winter storm events, such as those occurring in the Southern California Bight, can wipe-out the entire annual crop. This is of particular concern for species like *P. perforata* and *U. lactuca* that have thin foliose thalli and are less able to withstand wave action. Additionally, *P. perforata* grows best during the winter time period and would be most subjected to heavy storms. *L. setchellii* grows during winter as well, but kelp species are more hardy due to their strong holdfasts (Peteiro et al., 2016). Therefore, they may be the best option in areas along the Southern California Bight where wave action and storms are stronger or more frequent. Storm events, wave action, nutrient availability, biofouling, marine spatial conflicts, and sensitive habitats are several of the key important considerations for farm site selection (Gentry et al., 2017; Lester et al., 2014; Radulovich et al., 2015). When nutrients are not limiting, light levels and temperature account for over 75% of growth variability (Capo et al., 1999). However, each species has its own unique tolerances to environmental conditions (Appendix C). Thus, site selection to ensure nutrient requirements and environmental conditions are appropriate is of utmost importance and testing of the site for these parameters should be done before undertaking any intensive seaweed cultivation activity (Dawes et al., 1995; Santelices, 1999).

Discussion

Onshore vs offshore

Offshore cultivation performed better than onshore in our model under the mean growth scenarios. However, there are several additional aspects besides assumed profitability that need to be taken into account in an offshore system. These include risk, loss of control over farm conditions, variability in production, importance of site selection, and marketability and quality of the product (West, 2005; Levine & Sahoo, 2009). A major limitation of offshore aquaculture compared to onshore is the inability to control environmental parameters. For seaweed species, nutrient levels, light, and temperature are critical factors affecting growth (Santelices, 1999). Offshore farmers have a limited ability to regulate these elements. Controlling consistency of supply and quality of product is important in securing confidence from suppliers and buyers. Conversely, onshore cultivation provides greater control over environmental parameters. When ambient nutrient levels or light levels are low, onshore farmers can add fertilizer or increase light levels in tanks (Kim et al., 2017). This allows the farmer to

reduce some variability in production. To highlight the importance of nutrient availability on productivity, *P. yezoensis* offshore culture was attempted in Maine but failed due to insufficient nutrient availability to support growth (McVey et al., 2002). Therefore, having control over ambient environmental conditions is a benefit of onshore culture which may result in greater profitability.

Strong seasonality of the crops in offshore cultivation can result in a flooding of the market. This is particularly true for species such as *L. setchellii* where our model projects that 180 tons will become available over the period of a month. Flooding the market may also reduce the price point that the crop can be sold at, which is a weakness that negatively affects other products. One example of this is wild salmon wherein fishermen receive a lower value by flooding the market during harvest season (Valderrama & Anderson, 2010). They are forced to process and can the salmon instead of selling it fresh at a higher value. In juxtaposition, aquaculture can supply a fresh salmon product year-round. Therefore, producing a large quantity of product over such a short-term period means only very small quantities can be sold fresh, and the rest will need to be additionally processed either by drying or added into other products (e.g. ingredient in Blue Evolution seaweed pasta (Blue Evolution, n.d.)). The expense of this additional processing will raise the cost of production substantially but may be offset by receiving a higher market price (Yarish et al., 2017).

Another important caveat regarding offshore cultivation is the consistency of product value that can be achieved. A relative lack of control over ambient conditions offshore can hamper both the quantity and quality of the seaweed produced. For example, an ocean temperature spike resulting in damaged thalli or predation from other species such as fish or invertebrates will reduce the market value. While product quality is less of a concern if the seaweed is used for agricultural use or heavily processed for agar, it is crucial for the seaweed crop to obtain high-end market prices available through avenues such as restaurants. Therefore, when assessing the market that seaweed aquaculture should target, it is critical to ensure that the quality of the product can match that dictated by the market. For example, as wave action increases, the thalli of *Pyropia* become tougher, which reduces the grade of the crop (I. Levine, pers comm, 2018). Additionally, if the quality of the seaweed produced offshore is too low to access the local produce market, then economic feasibility might swing in favor of onshore production. Thus, the quality of product achieved is an essential component to the species and location decision.

Effect of permitting on feasibility

Permitting is a major factor influencing the feasibility of aquaculture in California. No permit has been granted in the offshore waters of California for over 20-years, although an established framework exists. Even in cases like seaweed aquaculture where documented environmental impacts are low, public opposition to ocean development can greatly slow the growth of the industry. Our model demonstrated that seaweed aquaculture can be economically viable in southern California with deference to a few key factors. However, given the political climate and strong opposition to aquaculture development in California and the U.S. more broadly, the economic viability of seaweed aquaculture may be irrelevant (Knapp, 2008). The permitting process is extremely fragmented and involves more than 10 federal, state, and local agencies

(CDFW, 2016). There is movement and strong interest in permitting seaweed aquaculture (e.g. Pharmasea, scientific trials by Salt Point Seaweed), but it is difficult to say if this will be sufficient to change the permitting landscape. The difficulty with permitting offshore has pushed aquaculturists onshore. However, onshore farms that cultivate seaweed typically grow other more valuable species (eg. abalone) to which they are able to feed unsold seaweed. Only one onshore seaweed farm exists in California.

While permitting was not an explicit factor included in our model, the history of aquaculture in California highlights the importance of developing permitting structures and resources for the cultivation of seaweed. Areas in California such as the Port of San Diego are working to establish a lease system and infrastructure to promote waterfront industries, including aquaculture. Our study employed the use of a lease-style permitting structure and associated costs for tenants that currently exist in Hawaii. Doing so enabled us to ignore the costs that onshore farmers would otherwise incur including purchasing land, installing a seawater pump (~\$30,000 dollars), and grading the land for an aquaculture facility. Given the high costs of ocean property land in southern California, it is difficult to determine if onshore seaweed aquaculture would be economically feasible given these high fixed capital costs. This highlights the importance of developing infrastructure, such as that proposed by the Port of San Diego, to assist with lowering initial fixed costs.

Opportunities to reduce capital and operating costs for seaweed aquaculture

A major barrier facing any new and existing US aquaculture industry is competing with cheaper imported products. These products are generally less expensive due to the lower costs of labor and cheap farm techniques enabled by lower environmental standards (e.g. spraying acid on *Pyropia* nets to control epiphytes in China; Levine & Sahoo, 2009). Therefore, keeping fixed and operating costs low is essential to give U.S. aquaculture a chance against imported products. There are several opportunities to reduce both capital and fixed costs for seaweed farming on and offshore, which include crop rotation offshore, reducing costs of net structure for *P. perforata* and *U. lactuca*, streamlining the permitting process, and reducing hatchery costs by creating a hatchery industry and university cooperative.

Cultivating different species offshore throughout the year will increase the annual productivity of the seaweed farm without drastically increasing starting capital costs. For example, growing *P. perforata* during the fall-winter months and *U. lactuca* in the spring-summer months enables faster payback of the expensive net structures and hatchery. Similarly, a farmer could grow *L. setchellii* during the winter-early spring, and *G. pacifica* in the spring-summer months using the same longline structure. This raises the societal benefit of permitting that section of the ocean for aquaculture by increasing the biomass per area of food produced annually. Another possible intervention is to reduce the high costs associated with installing the net structures offshore. If field studies determine that fewer buoys or less expensive anchors can be used, then *P. perforata* and *U. lactuca* farming becomes more profitable and may enable sale of this product at a produce market price.

As discussed above, securing a permit represents a major source of uncertainty in the viability of a seaweed farm, particularly the opportunity costs of the process (which can take up to 5 years or more). Therefore, state and federal regulatory agencies should work to streamline the permitting process to reduce both the time and costs required to obtain a permit. Finally, constructing and running a hatchery is a substantial cost for farmers. Developing a regional hatchery through university-industry collaboration could leverage the existing facilities at universities and reduce the costs to seaweed farmers. Additionally, this collaborative hatchery could also provide students or researchers an opportunity to address knowledge gaps regarding development of hatchery techniques and genetic selection for superior native seaweed strains.

Conclusion

Our study found strong variability in the costs and break-even prices among the candidate seaweed species. For the majority of species, the offshore model system had a lower break-even price than onshore. The fixed costs for offshore were more expensive than onshore for every species except for *L. setchellii*. However, the greater operational costs of onshore resulted in higher costs over the 5-year period. Our model results were highly sensitive to species growth rate and market prices. The importance of these factors necessitates further investigation regarding the validation of *in situ* growth rates for our native species and a market analysis to understand current consumer demand and potential for growth. Additionally, it will be important to develop collaborative partnerships between academia, resource agencies, and the aquaculture industry to overcome the considerable knowledge gaps and lack of infrastructure that exist in farming native seaweed species.

3. Integrating seaweed into an existing mussel farm

Introduction

In the first two chapters, we outlined potential candidate species, their appropriate cultivations techniques (see section 1), and considerations of their relative economic feasibility (see section 2). While we have identified some potential barriers and solutions to help mitigate the bottlenecks for developing seaweed aquaculture in California, it may take some time before these are realized. On the east coast and abroad, aquaculture is increasingly moving towards an ecosystem-based approach known as integrated multi-trophic aquaculture (IMTA) - a technique that harnesses the benefits of culturing two or more trophic level species together (Soto, 2009). For example, researchers in the U.S. Northeast identified three *Pyropia spp.* that could play an important bioremediation role in an integrated system with finfish (Carmona et al., 2005).

Current costs and environmental uncertainty of breeding finfish offshore in California have led to considerations of an IMTA model growing only shellfish and seaweed, which we will refer to as co-culture. Encouraging local production of seaweed could reduce our global footprint from importation and offset some of the environmental footprint of shellfish produced locally (Jones, 2002; Sim, 2007; Troell et al. 2009). The economic diversification of co-culture could provide significant benefits in an offshore system that has elements of high risk due to changes in weather (i.e. storms) and ocean conditions. One ocean condition that can be detrimental for shellfish development due to shifts in carbonate chemistry is ocean acidification, or the lowering of global ocean pH (Chung et al., 2013).

The ocean in the Southern California Bight tends be more acidic than usual in the spring due to upwelling. Furthermore, shellfish operations can be put on hold if elevated ocean toxin levels are detected. These toxins can be potent, such as domoic acid, which can cause severe illness or death after consumption (CA Dept. of Public Health, 2017). When domoic acid is at levels over 20 µg per gram of shellfish meat, commercial shellfish operations may be required to close (CA Ocean Science Trust, 2016). This period is often unpredictable, lasting between 3-6 months and incurring large economic losses to the farmer (B. Friedman, pers comm, 2017). Growing a second crop that is not susceptible to domoic acid or ocean acidification could allow the farmer to be more spatially, temporally, and economically efficient. Furthermore, co-culture of seaweed on existing shellfish farms can reduce start-up cost and ease the permitting process needed to register new aquaculture operations (Holdt & Edwards, 2014).

Aside from economic, spatial, and temporal benefits, co-culture can provide local ecological benefits. Shellfish farms have relatively low impact and can even improve water quality as the species filter feeds on nutrients in the water column (Lindahl et al., 2005). The benefits of this ecosystem service provided by shellfish are dependent upon ambient characteristics (i.e. nutrient availability and availability of biomass to absorb those nutrients). However, shellfish farms do release some nutrients into the water column and a healthy mussel bed could increase primary production in areas that have low nitrogen concentrations, which would otherwise limit phytoplankton productivity (Asmus & Asmus, 1991). Due to the net increase in ammonium around mussel farms, the farms tend to be sited in deeper water with good circulation (Jansen,

2012; Lindahl et al., 2005). However, this excess ammonium can be mitigated by primary producers in the ecosystem, like phytoplankton or macroalgae (i.e. seaweed).

Seaweeds have a large capacity to absorb dissolved inorganic nutrients (DIN) such as ammonium from intensive mariculture (Troell et al., 1999). Not only do they absorb excess DIN, but the seaweed themselves tend to grow at much faster rates in high-nutrient settings (Sanderson et al., 2012; Troell et al., 1999). There have been several experiments that reported notable DIN uptake by seaweed situated next to fish farms, urbanized coastal waters, and artificially-induced nutrient concentrations (Grote, 2016; Kang et al., 2014; Kim et al., 2015).

Of our candidate species, *Gracilaria pacifica* was an ideal species for initial field investigation at our client's mussel farm in Santa Barbara. It is relatively easy to grow vegetatively, doesn't require a hatchery stage, and is predicted to grow during the local domoic acid period. Furthermore, samples of *G. pacifica* were easy to obtain from our external advisor at the Cultured Abalone Farm for testing. Beyond those qualifications, it is the only seaweed species with an existing food market that has been cultivated in farm systems in both southern and northern California. Additionally, there is already a proof-of-concept for growing *G. pacifica* onshore and initial growth studies have been conducted for the species in the Santa Barbara Channel during peak upwelling season (D. Bush, pers comm, 2017; J. Couture, pers comm, 2018).

We undertook a two-step process to identify the economic and biological synergies of cultivating Mediterranean mussels (*Mytilus galloprovincialis*) and *G. pacifica* together. First, we performed an analysis of the market and demand for both products to determine the ideal production ratio of seaweed to mussels to maximize a farmer's profit. Then, we conducted an empirical study to identify growth and product quality benefits from adding seaweed culture into the framework of an existing mussel farm.

3.1 Economically optimal mussel-seaweed farm

We conducted a preliminary economic analysis of co-culture feasibility by 1) looking at our client's profits from *Mytilus galloprovincialis* (Mediterranean mussels) and 2) estimating the current demand of edible fresh seaweed from literature and industry.

Methods

We estimated profit per unit of production for both seaweed and mussels to determine the optimal mussel-to-seaweed production ratio on our client's 25 acre (10.4 hectares) offshore farm (same farm site used throughout this paper). We then calculated profit per unit of production (kg) by estimating the current seaweed demand in California, marginal cost for seaweed production, and our client's reported mussel profit per acre. The optimal ratio of production was where the profit per acre for both products converges.

We made several assumptions in our calculations, listed below:

- Aggregation of fresh and dried seaweed in the high-end, functional food market is a good estimate of the demand for *G. pacifica*

- The level of production our mussel farmer operates at has no impact on the overall mussel market, so the cost and revenue stay constant for all levels of production
- Marginal cost for seaweed is constant, since the costs for producing an additional unit of seaweed is mostly negligible until a certain farm size is exceeded
- Our client and any other shellfish farmer is a rational economic actor making profit-driven decisions

Controlling for farm size allowed us to account for the major costs in initial capital and required labor. We used farm size in units of acres as a measure of production, since there was a maximum amount of mussels and seaweed that could be feasibly produced in a single acre due to structural constraints of the farm equipment. The production capacity per acre also depended on the optimal stocking density of each species.

Calculating profit per unit of production for *M. galloprovincialis*

The data source we used to calculate profit per production was our client's reported marginal cost (C_M) and revenue (R_M) received for *M. galloprovincialis* during the years 2014-2016. In order to derive the profit produced by mussels alone, we removed any presumed shared costs to calculate the marginal cost per production and subtracted that from the average revenue per kg that our client has received over those three years. Shared costs are those that only need to be incurred once for the two species, such as annual registration and boat slip. We used the purchase of mussel seed, shipping, contract labor, and boat fuel for this aspect of the farm. Using the revenue, production quantity, and costs provided by our client, we calculated the average revenue and cost per kg to find profit:

$$\text{Profit/kg} = (R_M/\text{kg} - C_M/\text{kg}) \quad (\text{mussel profit})$$

Estimating demand curve for edible seaweed

While we were able to obtain total cost, revenue, and profit for the mussel portion of the farm, all the data for seaweed had to be estimated from literature and limited industry-provided information. We determined revenue per production (R_S) for seaweed in the hypothetical co-culture farm by first estimating the current demand and supply of local, edible seaweed in California. Then, we calculated the additional quantity that could be produced on top of current production. This gave us a new equilibrium price that converges with the current mussel price rate our client is receiving.

To estimate the demand curve for *G. pacifica*, we aggregated production (Q_S) and price (R_S) information from several literature sources and limited production data from five wild seaweed harvesters in northern California and three small-scale businesses growing seaweed throughout California. We calculated price and quantity produced for wild harvesters and farmers separately since the range of farm gate prices for harvested and dried seaweed is different from fresh, farmed seaweed. After calculating them separately, we converted dry weight to fresh weight using an intermediate ratio taken from literature (1:6.67 dry to fresh), and took their weighted average based on production (Azevedo et al., 2016; Hernandez et al., 2005; Roesijadi et al., 2008; Watson & Dring, 2007).

We took average farm gate prices of all 5 harvesters, but only had annual production values for a single harvester. Since it was not possible to retrieve the data from the other businesses, we

used an annual average of the single harvester's (Strong Arm Farm in Sonoma County) value for the other 4 harvesters. As they are all seasonally limited to summertime, we thought that it was fair to assume they would have similar annual production. Similarly, we obtained all three farm gate prices from the seaweed farmers, but only accessed annual production values for two of them. Since the Carlsbad Aquafarm is growing at a small, experimental scale, we used the Cultured Abalone Farm's production as a conservative estimate for the annual production of Monterey Bay Seaweeds due to limitations on data availability.

Once we estimated current production (Q_S) in kg, averaged price received (R_S) in \$/kg for edible seaweed in California, and gathered a price elasticity (ϵ) from a similar product (local lettuce) that could be used as a substitute for local seaweed, we back-calculated the slope and y-intercept of the demand curve. The elasticity value (ϵ) we used was -2.05, a value obtained from a study that examined demand for local produce (Xu et al., 2015).

$$R_S = m * Q_S + b \quad (\text{demand curve equation})$$

$$m = R_S/Q_S * 1/\epsilon; b = R_S(1-1/\epsilon) \quad (\text{solve for } m \text{ and } b)$$

Calculating profit per unit of production for *G. pacifica*

After creating the demand curve, which provided us the revenue per unit across various production levels, we calculated our marginal cost of seaweed production. One-time, capital costs were included with the regular annual costs estimated in Chapter 2 and any shared mussel-seaweed costs were subtracted from this value. The amount of seaweed production that can replace mussel lines in the 25 acre farm was calculated by weight (kg), and then converted to acres based on stocking densities as reported by CA Fish and Game Commission (2018). Marginal cost (C_S) for the hypothetical offshore seaweed farm was primarily comprised of stocking costs, seeding labor, harvest labor, and boat fuel. In our co-culture scenario, we also accounted for initial capital costs of adding in seaweed. The initial fixed costs (F_S) included materials such as longlines, anchors, and buoys (Table 3.1). Since initial costs (primarily farm materials) are usually quite high, we looked at two scenarios: 1) initial costs incurred in present day 2) initial capital costs incurred 5-years before present. Total costs per acre for seaweed was calculated with the following formula:

$$(\sum F_S + C_S) * (1/25\text{acres}) \quad (\text{present day})$$

$$[(\sum F_S * \frac{1}{(1+\delta)^5}) + C_S] * (1/25\text{acres}) \quad (5\text{-years before present})$$

Table 3-1. The initial capital costs included for seaweed cultivation

Materials	Cost
Longline	\$76,532
Anchor rope	\$43,633
Chain	\$16,363
Buoy	\$88,000
Trawl floats	\$4,418
Shackles	\$6,545
Tying rope	\$16,363
Jeyco anchor	\$120,000
Deployment	\$8,175
Total	\$380,028
Total (5-year discount)	\$297,762

An additional condition was that Q_{S_acre} , the amount of seaweed our client could feasibly produce (in units of acres) next to his mussels, had to be added to the current total production of local seaweed ($Q_{S_Current}$) so we could account for the change in equilibrium market price. The modified demand curve equation that we needed to use to optimize for Q_{S_acre} was:

$$R_S = m * (Q_{S_acre} * (\text{kg/acre}) + Q_{S_Current}) + b \quad (\text{modified demand curve})$$

For each of the two annual cost scenarios (\$380,028 and \$297,762), we optimized for the Q_{S_acre} in the modified demand curve equation that would give us an R_S that satisfied the following:

$$\text{Profits/acre} = (R_S/\text{kg} - C_S/\text{kg}) * (\text{kg/acre conversion}) \geq \text{Profit}_M/\text{acre} \quad (\text{optimization ratio})$$

Q_{S_acre} was rounded to a whole number that was equal to or greater than the profit per acre of mussel production.

Finally, we compared the absolute and percent change in total annual profit of the two cost scenarios to the original 25-acre mussel monoculture farm.

Results and Analysis

Since our client's production and profit data is confidential, we do not list the results of calculating profit per acre of mussel production. The summary of estimated current production and average price for seaweed is listed in Table 3.2. The market price for farmed seaweed was significantly higher than for wild harvested. This is likely due to the large range of prices (\$13-88/kg) that fresh seaweed is sold for among the three farms.

Table 3.2. Average market price and production for local California seaweed grouped by wild harvest and farmed. We use a 1:6.67 dry to fresh weight conversion to compare across products.

	Market Price	Production
Wild Harvested	\$19.87/kg*	6,136.36 kg
Farmed Seaweed	\$39.97/kg	1,518.18 kg
Aggregate	\$23.86/kg	7,654.55 kg

* The original average wild harvested market price was \$134.24 per dry kg of seaweed. We converted the units to fresh weight dollars for ease of comparison.

Since seaweed farming in California is still new or used as a supplementary and experimental crop, the prices they can be sold at may fluctuate for a while and converge towards an equilibrium price only after a steady market has been established. Conversely, the wild harvest seaweed businesses have been operating and expanding for at least a decade despite seasonal limitations (H. Herrmann, pers comm, 2018). Their market price is only \$6/kg more than the average price of edible seaweed imported into California.

The aggregate of the production and market prices for wild harvested and farmed seaweed is shown in Figure 3.1. There was little information available on price elasticity of edible, local seaweed product. However, we deemed that using the elasticity of local lettuce in Hawai'i was comparable in that both were competing against a large imported source. Here we assumed that supply estimated from harvester and farm data has matched the existing demand in California. In our next steps, we calculated the new equilibrium price (\$/kg) if we increased production at our client's farm, keeping marginal cost constant.

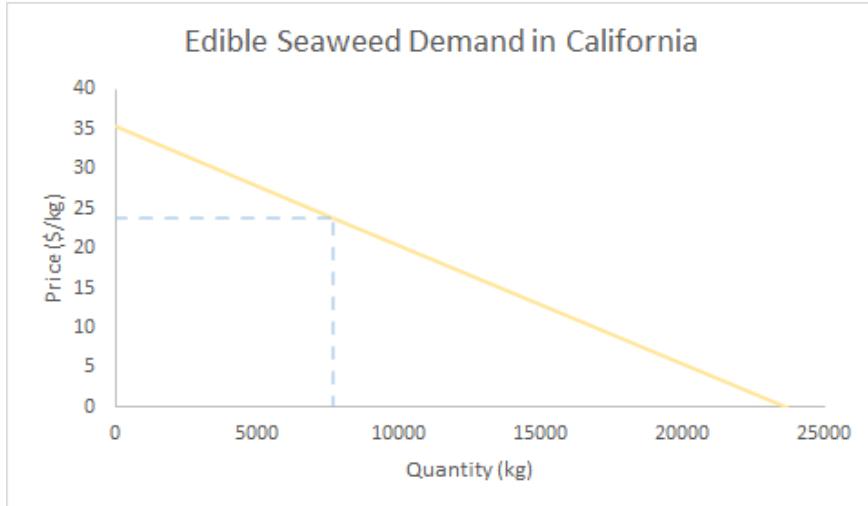


Figure 3.1. The edible seaweed demand curve ($y = -0.0015x + 35.50$) was constructed from production and price information from five northern California seaweed harvesters and three seaweed farmers. The blue dashed lines indicate the current aggregate quantity and market price.

When initial capital costs are incurred, the optimal ratio is 3 acres to 22 acres of seaweed and mussel production (Figure 3.2(a)). Given initial costs, we expect to have a low seaweed production compared to mussels. However, when the initial capital costs are discounted over a five-year period, the optimal amount of seaweed production only increases by a single acre (Table 3.3). In both co-culture scenarios, our client's profit per unit of acreage increased (Figure 3.2(b)).

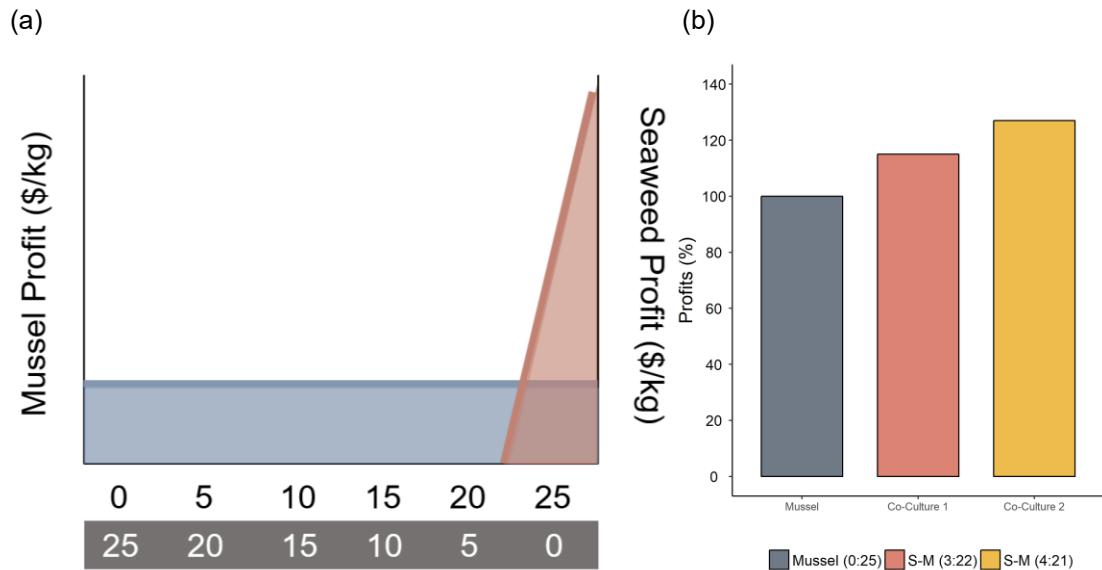


Figure 3.2. (a) The blue line is the constant profit per production of mussels and the red line is for seaweed. Shaded areas are total profit. The red shaded triangle is the added profit from co-culture. (b) In the first co-culture scenario 3:22 (pink), the increase in profits per year is about 20.1%. In the second co-culture scenario, optimal production ratio is 4:21 and the increase in profits is 31.5% (yellow).

Our preliminary analysis determined the optimal mussel to seaweed ratio by comparing the profit per unit of production for both products. The projected increase in profit from the co-culture scenarios would only occur with a shift in demand for local seaweed. The summary of our results are in Table 3.3.

Table 3-3. Prior to the addition of seaweed production from the hypothetical co-culture farm, the current production in California is 7,654 kg at an average farm gate price of \$23.86 ($\varepsilon = -2.05$). Marginal costs incorporate initial capital costs and annual costs, not listed here.

Scenario	Relative Profit	Added Profit	Added Production	Capital Costs	Marg. Cost	Equilib. Price
Co-Culture 1	115.3%	\$29,396.26	6,216 kg	\$380,027.50	\$8.74/kg	\$14.41/kg
Co-Culture 2*	127.2%	\$52,260.02	8,288 kg	\$297,761.50*	\$7.15/kg	\$11.25/kg

*In scenario Co-Culture 2, the initial capital costs were incurred 5-years prior so have been discounted ($\delta = 0.05$).

The new equilibrium price that we estimated from Co-Culture Scenario 1 was a lot closer to the current price per kg of imported edible seaweeds (\$13.38/kg). The additional production that would need to be demanded for market price to fall to \$14.41/kg is 6,216 kg. Given that current production is about 7,655 kg, this would require roughly a doubling of demand. However, this may not be unreasonable as interest in aquaculture has been growing in California and there is only one full-time seaweed farm currently.

Discussion

While the current market may not be able to absorb a 25-acre monoculture seaweed farm, growing seaweed in a co-culture system could be a conservative way to start expanding the industry. We assume that the dried and fresh seaweed market can share consumers. However, if the edible seaweed market grows, it is possible that these two markets diverge and demand will need to be calculated separately for a more accurate representation. While we assumed that the market is saturated in determining the optimal ratio, wild harvesters of seaweed seem to be able to sell their entire product fairly easily.

Even if we are underestimating production in California, there are significant hurdles to seaweed aquaculture production in the state. More research on where to market edible seaweed and where demand might be highest is a good next step. There are many challenges to selling local, fresh seaweed to the southern California market, but taking advantage of existing farms or infrastructure could be a good way to circumvent some of the bottlenecks. There are only three permitted offshore aquaculture plots in southern California currently, but working together to create co-culture or IMTA systems rather than building up individual monoculture sites will likely be most effective. Adding seaweed as an alternative crop could be beneficial to shellfish farmers during domoic acid closures or in the face of ocean acidification. Many opportunities exist for a farmer interested in co-culture. For example, it may be more efficient to partner with a processor to dry seaweed rather than absorbing the costs of creating that infrastructure on their own.

3.2 Offshore field experiment

Our client at Santa Barbara Mariculture approached us with a desire to investigate whether there were any synergistic benefits to culturing seaweed alongside the existing *M. galloprovincialis* crop (Mediterranean mussels). He had anecdotal evidence that his mussels seemed higher quality when his rope headlines were fouled with native *Gracilaria spp.* (B. Friedman, pers comm, 2017). Therefore, we tested whether there was an effect of growing *G. pacifica* nearby on mussel growth and meat quality. We also examined the technical ease of culturing *G. pacifica* and its potential productivity at an exposed open-ocean site. The following experiment took place over a five-month period from July through November 2017.

Methods

Study site

We conducted the field experiment at the Santa Barbara Mariculture farm - an exposed site in offshore waters of Santa Barbara ($34^{\circ}23'39.2''N$ $119^{\circ}45'09.8''W$). The site was located in 80 feet of water and the farm structure was typical of longline mussel culture with vertical droppers suspended by buoys over a 125 ft longline system. Dropper depth was between 15-35 ft below the surface. Depth was altered to increase growth depending on the season and other environmental factors (e.g. avoid duck predation). During the sampling period, water temperature likely ranged between 14.11 and 18.35 °C based on historical trends (CalCOFI Database, 2017; NOAA National Centers for Environmental Information, 2018). Note that this site is the same site used as the theoretical offshore location in the rest of our study.

Mussel growth and condition index

Mussel growth was tracked over a five-month period, with a mean initial shell length of 26.64 mm, determined by sampling one hundred individuals (Lander et al., 2012). The initial spat was seeded onto lines at a hatchery. We replicated the protocol from Lander et al. (2012) for sampling mussels. Growth for both the experimental ($n = 5$) and control ($n = 5$) sets of mussels were tracked monthly. The condition index was taken from 10 individuals grabbed at random per rope (i.e. $n = 100$ per sampling day). We transported the mussels on ice and then stored them in a freezer until further processing. We calculated the condition index as the ratio of dry meat weight to dry shell weight (%). Condition index was used because it is an indicator of high market value and the overall health of the mussel (Pauley et al., 1988). Prior to processing, we removed the byssal threads and measured the shell length, width, and height of each mussel using an electronic calipers to the nearest 0.01 mm. Next, we removed the mussel meat and measured the wet meat weight (WMW) in grams. We dried the mussel meat in an oven at 80°C and weighed it after 24 hours. We left shells out to air dry and then recorded their mass within 48-72 hours after initial processing.

Seaweed growth study

In contrast to the mussels, growth for *G. pacifica* was tracked bi-weekly for the first 3 months of the experiment, and monthly for the final 2 months. Rope culture is the most common cultivation method for *Gracilaria* globally, but netted bags are less labor intensive and thus more realistic for cultivation in the Southern California Bight (Oliveira et al., 2000). This was corroborated by a pilot study done performed by SARC at our client's site in which similar growth rates were observed for both rope and bag culture (J. Couture, pers comm, 2017). Therefore, we elected to use nylon mesh bags in our field experiment. The bags used were 2.74 m long with a diameter of 10.2 cm and cylindrical volume of 0.089 m³. Each bag consisted of 3 compartments (each ~91 cm long), which were created by tying nylon string across the diameter of the mesh.

Bags were stocked with 178 grams of *G. pacifica*. Each compartment contained approximately 59 grams of seaweed. Fresh *G. pacifica* was sourced from a local onshore farm where it is produced. We originally stocked the bags with more *G. pacifica* (277 g/bag section) but switched to a lower stocking density of 2 kg/m³ after a month of testing due to concerns of crowding (Nagler et al., 2003). The day before each experimental cycle, we transferred the bags to a nearby harbor where they remained submerged in seawater for approximately 12 hours. They were then placed in a cooler for transport to the field site the following morning. We secured the bags at 1m intervals between mussel droppers and suspended them from the main headrope using a polypropylene line. The top of each bag was approximately 5 m deep. As we deployed each set of bags, we removed the bags from the previous growth cycle (either two weeks or a month) and placed them in a cooler for return back to the lab and immediate processing. Prior to taking measurements, all *G. pacifica* was sorted by hand to avoid weighing epiphytes and other bio-fouling organisms. We recorded the wet weight of each compartment of the 5 bags using an electronic scale (0.01 g).

Statistical analysis

To test for effects of *G. pacifica* on final mussel length and mean condition index, we employed a pair of two-sided t-tests. We made an assumption of a normal distribution of sampling statistics due to the large number of total observations ($n_{mussel} = 500$, $n_{seaweed} = 90$) and an application of the central limit theorem. We used a multiple linear regression to examine the effect of *G. pacifica* on both condition index and length over time. We calculated the relative daily growth rate (%) of *G. pacifica* over the entire course of the field experiment as well as on a per-cycle basis. We used an alpha-level of significance of 0.05 unless otherwise stated.

Results

Mussel growth (length)

At the end of our five month field experiment, there was no discernible difference in final mean shell length between the control mussels and those grown alongside *Gracilaria pacifica*. The control mussels had an average length of 57.76 ± 4.11 mm ($n = 50$) whereas the experimental mussels had an average of 57.80 ± 6.47 mm ($n = 50$) (Table 3.4). A two-sided t-test concurred that there was no statistical difference in the mean shell length between the two groups at the end of month five ($t = -0.031$, $df = 81.1$, $p = 0.975$).

Table 3.4. The mean mussel shell length and standard deviation (mm) at the end of the experiment (month = 5) are shown (number of observations in italics). The mean and standard deviation of the initial mussels used to seed the experiment are included for comparison. A two-sided t-test showed no significant difference between mean shell length at month five for the control and experimental groups ($t = -0.031$, $df = 81.1$, $p = 0.975$).

Treatment	Time (Month)	Mean Length (mm)	Standard deviation (mm)
<i>None (Initial)</i>	0	26.64 (100)	5.68
<i>Control</i>	5	57.76 (50)	4.11
<i>Experimental</i>	5	57.80 (50)	6.47

The results of the multiple linear regression for shell length as a factor of time and treatment are shown in Table 3.5. The model significantly predicted shell length ($F(2,513) = 566.8$, $p < 0.001$, $R^2 = 0.687$). The regression coefficient for the experimental treatment was positive, but barely so ($\beta_{\text{experimental}} = 0.83 \pm 0.54$). However, treatment was not shown to be a significant predictor of final size ($p = 0.126$)

Table 3.5. Multiple linear regressions results for final shell length at the end of the experiment as a factor of time (months) and treatment (shown here in comparison to the control treatment). The model significantly predicted shell length ($F(2,513) = 566.8$, $p < 0.001$, $R^2 = 0.687$). Asterisks denote statistical significance at $\alpha = 0.05$

	Coefficient	Standard Error	t	p	Overall Adjusted R^2
<i>Intercept</i>	27.03	0.67	40.34	<0.001*	0.687
<i>Time (month)</i>	6.32	0.19	33.64	<0.001*	
<i>Experimental Treatment</i>	0.83	0.54	1.53	0.126	

Mussel quality (condition index)

Over the experimental time interval, condition indexes for all mussels ranged from 4.96 to 51.67 (Figure 3.3). We detected no statistically significant difference between the mean condition index of the control group (19.60 ± 6.32) and the mean (19.47 ± 7.02) of the experimental group co-cultured with seaweed ($t = 0.209$, $df = 509.7$, $p = 0.834$).

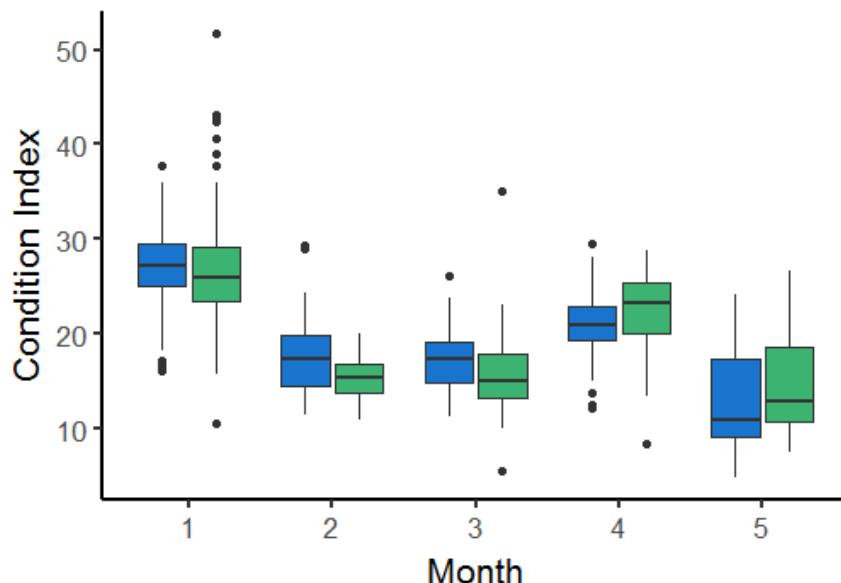


Figure 3.3. Monthly mean condition index ($n = 500$) of control (blue) and experimental mussels (green). A two-sided t-test showed no significant difference in condition index across the two treatments ($t = 0.209$, $df = 509.7$, $p = 0.834$).

Table 3.6 displays the results of the multiple linear regression of time and treatment on the condition index of mussels. The dual-factor regression model significantly predicted condition index ($F(2,514) = 77.25$, $p < 0.001$, $R^2 = 0.23$). Our model predicted that condition index was negatively correlated with time ($\beta_{\text{time}} = -2.23 \pm 0.18$) and the addition of seaweed ($\beta_{\text{experimental}} = -0.11 \pm 0.52$). However, the treatment factor was not demonstrated to be a significant predictor of mussel quality alone ($p = 0.826$).

Table 3.6. Multiple linear regressions results for condition index throughout the experiment as a factor of time (months) and treatment (shown here in comparison to the control treatment). The model significantly predicted condition index ($F(2,514) = 77.25$, $p < 0.001$, $R^2 = 0.23$). Asterisks denote statistical significance at $\alpha = 0.05$

	Coefficient	Standard Error	t	p	Overall Adjusted R^2
<i>Intercept</i>	26.11	0.64	40.89	<0.001*	0.228
<i>Time (month)</i>	-2.23	0.18	-12.43	<0.001*	
<i>Experimental Treatment</i>	-0.11	0.52	-0.22	0.826	

Seaweed Growth

The mean relative daily growth rate (%) of each of the eight experimental cycles was negative; indicating a loss of biomass once transplanted to the study site (Figure 3.4). We calculated the relative daily growth rate across the entire experiment to be -1.86 % ($n = 8$) when weights were applied for differential harvest period lengths. We also examined the subset of the cycles for which the stocking density (2 kg/m^3) referenced in Nagler et al. (2003) was applied. We calculated a growth rate of -1.82% ($n = 6$) when only these points are considered.

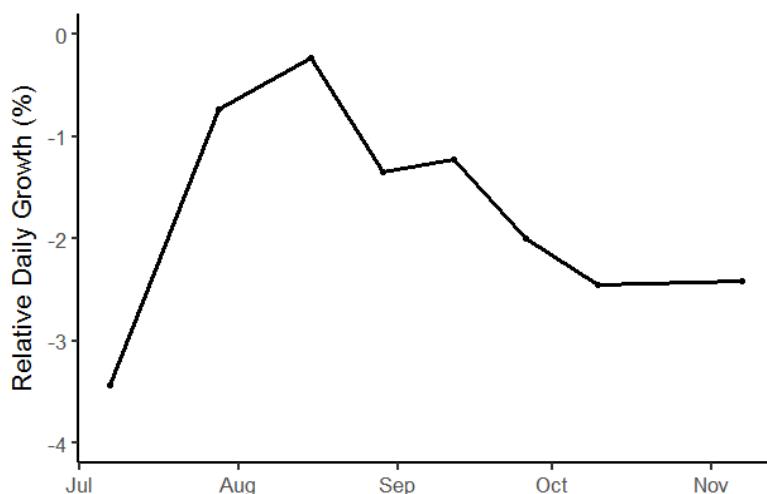


Figure 3.4. The mean relative daily growth rate of *G. pacifica* over eight growth cycles running from July through November of 2017. Cycles 3-8 (mid-August onwards) used a lower stocking density of 2 kg/m^3 (Nagler et al., 2003).

Discussion

The results of our field experiment indicate that there was no statistically significant effect of co-culturing *M. galloprovincialis* with *G. pacifica* on mussel length ($p = 0.975$) or condition index (0.834). However, we think it's possible that the results were confounded by the poor growth rates and health of the seaweed cultivated in our experimental trials. We only observed positive growth in 11% of all seaweed bag compartments over the course of our experiment and only 2.5% total achieved a cumulative growth of over 10% of its original stocking density ($n = 240$). Furthermore, we calculated the weighted mean relative daily growth rate (r) for our experimental seaweed trials to be -1.86%. For comparison, *G. pacifica* grown onshore in Santa Barbara County has been documented to reliably double in mass over a two-week period ($r \sim 0.05\%$) (D. Bush, pers comm, 2018). We believe this disparity in growth rates could have resulted from a variety of factors including cultivation method, seasonality, and factors inherent to offshore aquaculture.

We chose to cultivate *G. pacifica* in nylon mesh bags because of the relative ease of inserting them alongside the mussel droppers at our experimental site as well as promising results from previous growth studies in the area (J. Couture, pers comm, 2017; Oliveira et al., 2000). However, it is also commonly cultivated on ropes globally and in mesh cage-like structures (Yarish et al., 2012). Onshore, *G. pacifica* is grown loose in tanks as a tumble culture (D. Bush, pers comm). We think it is possible that our mesh bags constricted the seaweed and forced bunches of *G. pacifica* to clump together at times. This conceivably could have increased intraspecific competition for available nutrients and light. It may also have led to the build up of metabolic waste if the waste was not able to be flushed away by the current. We recommend that future studies investigate cultivating *G. pacifica* on ropes or in cage-like structures that are less constricting.

Due to temporal constraints on experimentation, our field experiment took place outside of the ideal cultivation period for *G. pacifica* from March to July (Figure 1.5). Therefore, we may have muted any potential seasonal benefits by growing *G. pacifica* outside of its optimal harvest period. Future studies should account for the preferred environmental conditions of both co-cultured species to increase the likelihood of observing any synergistic growth effects.

Our field experiment empirically highlighted several important tradeoffs when considering siting seaweed aquaculture onshore or offshore. We frequently observed partial to total bleaching of color in *G. pacifica*, an indication of poor health and overall market quality (D. Bush, pers comm). Bleaching is less common in onshore systems where farmers have more control over product quality. Farmers can modulate water temperature, current speed (flow rate), and nutrient availability with greater ease onshore. Due to this, many species cultured onshore can be raised year-round (Figure 1.5).

In addition to abiotic factors, Titlyanov and Titlyanova's (2010) review of seaweed cultivation methods identified biofouling as a major problem for the propagation of large-scale production systems. The most problematic fouling species are epiphytes and native seaweeds with rapid

growth rates, such as *Ulva spp*. Common methods of control include cultivating farmed species at greater depths, periodically exposing culture structures, and increasing the pH of the system through the application of acids (Titlyanov and Titlyanova, 2010). While we observed minimal epiphytic fouling offshore, our seaweed bags frequently returned from the farm site encrusted with an abundance of caprellids. Caprellids are omnivorous amphipods that have been considered for use in aquaculture systems as a means of epiphyte control (Fletcher, 1995). However, their presence greatly increased the processing time of the seaweed, which would lead to higher labor costs. Future studies should consider control mechanisms and the role of fouling organisms, both helpful and harmful, in determining the cultivation seasons of candidate species.

Conclusion

While we did not observe any biological synergies in our field experiment, we believe there are many promising facets of co-culturing seaweed with shellfish that are worthy of investigation. Our optimization model demonstrated that integrating *G. pacifica* cultivation could improve our client's profit under current market conditions. Our client could also diversify his product profile to decrease risk and be able to maintain income during domoic acid closures of mussel production. Furthermore, aquaculture producers can increase the productivity of ocean plots by implementing 3-D farm configurations as are currently being studied by researchers at the Woods Hole Oceanographic Institute (S. Lindell, pers comm, 2018). This could be valuable in the realm of marine spatial planning by reducing the need to obtain a new permit. Finally, seaweed aquaculture has been hypothesized to buffer decreases in pH (Chung et al., 2013) and eutrophication at a localized level (Newell, 1988). The former would help shellfish farmers increase the resiliency of their products to ocean acidification. The latter would offset nutrient loading from terrestrial systems and help stifle the development of harmful algal blooms. Further studies should investigate the environmental and ecological impacts of co-culturing at larger production and temporal scales.

4. Concluding remarks and recommendations

Seaweed aquaculture should have a place at the table in plotting the future production of local, healthy, and environmentally responsible food items here in California. Our research has identified candidate species with existing food markets and established production systems into which prospective farmers can buy. We demonstrated that in-state seaweed production can be economically feasible and that stakeholders should continue to evaluate potential productivity, control over product quality, and ease of permitting when deciding whether to site farms on or offshore. Finally, we outlined that shellfish aquaculture in southern California can help overcome some of the preliminary inertial barriers to seaweed aquaculture. However, it is crucial that actions are taken to promote the long-term viability of the industry as well. Efforts should be focused on improving cultivation techniques, marketing strategies, and the political atmosphere. Specifically, we recommend the following eight steps (Figure 4.1):

Cultivation Techniques

1. Validation of growth rates and farm structures
2. Solve bio-fouling issues
3. Polyculture of seaweeds
4. Co-culture of seaweeds with shellfish

Marketing

1. Further nutritional testing
2. Quantification of environmental benefits

Political Atmosphere

1. Increased collaboration between researchers, government, and farmers
2. Highlight benefits of seaweed aquaculture to push permitting

Cultivation techniques

Our model utilized the best available science to project the productivity of the *Gracilaria pacifica*, *Ulva lactuca*, *Pyropia perforata*, and *Laminaria setchellii* in the Southern California Bight. However, little to no research has been done on these candidate species in the region to corroborate these estimates. Therefore, an immediate need is for farmers and researchers to empirically **validate these growth rates and farm structures** on and offshore. This ground truthing will allow stakeholders to better calibrate our model findings and develop the most effective *in situ* cultivation techniques and management practices. For example, our field experiment highlighted the need to **solve bio-fouling issues** to prevent loss of product and the need for increased processing requirements. Finally, there is space for the development of creative grow-out methods to maximize the productivity of a plot and diversify farmers' economic portfolio to risk. The seasonal and spatial rotation of seaweed species (**polyculture**) and **co-culturing** them alongside shellfish can maximize economic efficiencies and ecological synergies.



Figure 4.1. A summary of our recommended next steps to encourage the long-term viability of seaweed aquaculture in southern California.

Marketing

A more detailed understanding of the market demand for fresh, local seaweed in southern California would allow farmers and policy makers to better scale the size of aquaculture production in state waters. In the short term, a market penetration study should be performed to quantify the demand and inform marketing strategies to promote the industry. Complementary research can be utilized to help augment these efforts. Further **quantification of the environmental footprint and benefits** of seaweed aquaculture can address concerns harbored by the public and policy makers about aquaculture more generally. This should include a life-cycle assessment of producing seaweed locally versus importing it from foreign markets. Additionally, quantifying and promoting the health benefits of consuming California seaweed through **further nutritional testing** will assist the acceleration of increasing demand in the region.

Political Atmosphere

Our work highlighted permitting difficulties, high capital costs, and cultivation knowledge gaps as the three main barriers to the development of a sustainable seaweed aquaculture industry in southern California. Researchers and stakeholders need to ensure that decision makers are armed with up-to-date and locally relevant information so that it is reflected in the regulatory atmosphere for aquaculture in California. Current research should be leveraged to **highlight the**

benefits of seaweed aquaculture and encourage the allocation of funds to support future studies. This will help foster political favor for the growth of environmentally conscious seaweed production. **Increased collaboration between research, government, and farmers** can help overcome the aforementioned barriers that cannot be addressed in isolation. For example, stakeholders should work together to develop a comprehensive food safety plan for the handling and sale of locally produced seaweeds. This will give distributors and consumers greater confidence to buy into domestic production. Additionally, pooling resources encourages the groundwork to be laid for necessary infrastructure. The establishment of commercial hatcheries would greatly reduce the initial cost of cultivation for farmers.

Outlook

Taken together, this suite of recommended next steps paves the way for seaweed aquaculture production in southern California. Stakeholders can use it to create a mutually beneficial business that is economically sound and ecologically friendly. Current and future Californians can enjoy a local food item with a rich nutritional profile and few resource inputs. Much like demand for the product itself, the seaweed industry will most likely not bloom in the span of a season. However, aquaculturists, scientists, and policy makers have the opportunity to make decisions in the short term to ensure the maturation of an environmentally responsible industry that will flourish into the future.

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Appendices

Appendix A: Biology and life-cycle of seaweed

For any agriculture or aquaculture production, understanding the biology and life-cycle of seaweed species is necessary to cultivate seaweed. The three major seaweed groups are red (Rhodophyta), green (Chlorophyta), and brown (Phaeophyta) algae; significant differences in reproductive and life-history modes exist between these groups. Asexual species, such as *Gracilaria pacifica*, are capable of producing multiple individuals from the same parent material through vegetative fragmentation or asexual spores (Wynne and Loiseaux 1976 in Lui et al., 2017). Conversely, sexual reproduction requires the release and fusion of gametophytes, forming a zygote that then undergoes cellular growth via mitosis.

The wide variety of reproductive, physiological, and morphological forms amongst seaweeds results in highly specialized cultivation strategies that must be determined for each species. Broadly, there are four major steps in the cultivation of macroalgae, which include 1) hatchery, 2) nursery phase, 3) grow-out phase, 4) harvest phase, but not all species require all phases. The life-cycles of these species drastically affect the cultivation methods and stages required during farming. Therefore it is crucial to understand the general differences between these groups and species.

Red seaweeds reproduction is characterized by a tri-phasic reproductive cycle and an 'alternation of generations,' which fluctuate between the adult gametophyte (male or female) and tetrasporophyte stage (Lobban and Harrison, 1994). In *Gracilaria*, the adult gametophyte and tetrasporophyte phases are indistinguishable (i.e. isomorphic), and the thalli of both phases are used for human consumption (Lobban and Harrison, 1994; Charrier et al., 2017).

Out of all cultivated seaweeds, *Pyropia spp.* has the most complicated life-history which necessitates a carefully controlled hatchery stage (Levine and Sahoo, 2009). Considerable work has gone into identifying and attempting to cultivate *Pyropia* species that can generate foliose thalli through asexual means and therefore are capable of seeding asexual spores onto the nets (e.g. Blouin et al., 2007). This bypasses the concelis phase, reducing the large costs and labor required for the hatchery phase. However, the majority of cultivated *Pyropia* species require a hatchery phase.

Within brown seaweeds, *Laminaria setchellii* gametophytes continue to develop until environmental cues (e.g. sufficient quantities of blue light) initiate reproduction, and the female gametophyte releases a pheromone that attracts motile male sperm. For large-scale cultivation, the environmental conditions that induce sporing and fertilization are carefully controlled for during the hatchery phase (see section below).

Ulva lactuca, a green species, can be propagated vegetatively in its unattached form (Titlyanov and Titlyanova, 2010). However, when grow-out occurs offshore spores are seeded onto the net or line in a hatchery or wild collection phase (Titlyanov and Titlyanova, 2010).

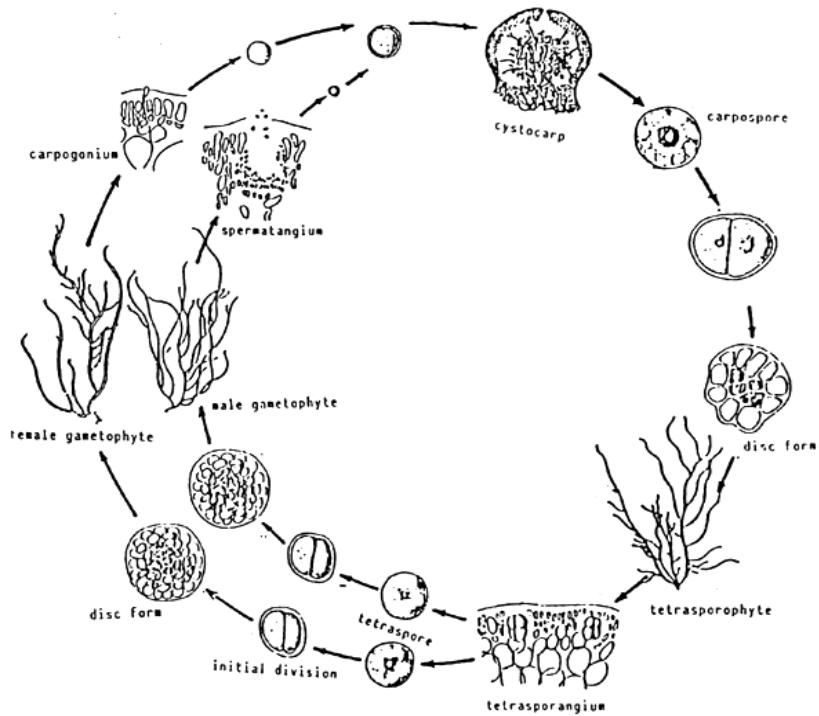


Figure A.1. Life-cycle of *Gracilaria* spp and representative of red seaweed, and a representative of a species that can propagate vegetatively (Figure from FAO, 1990).

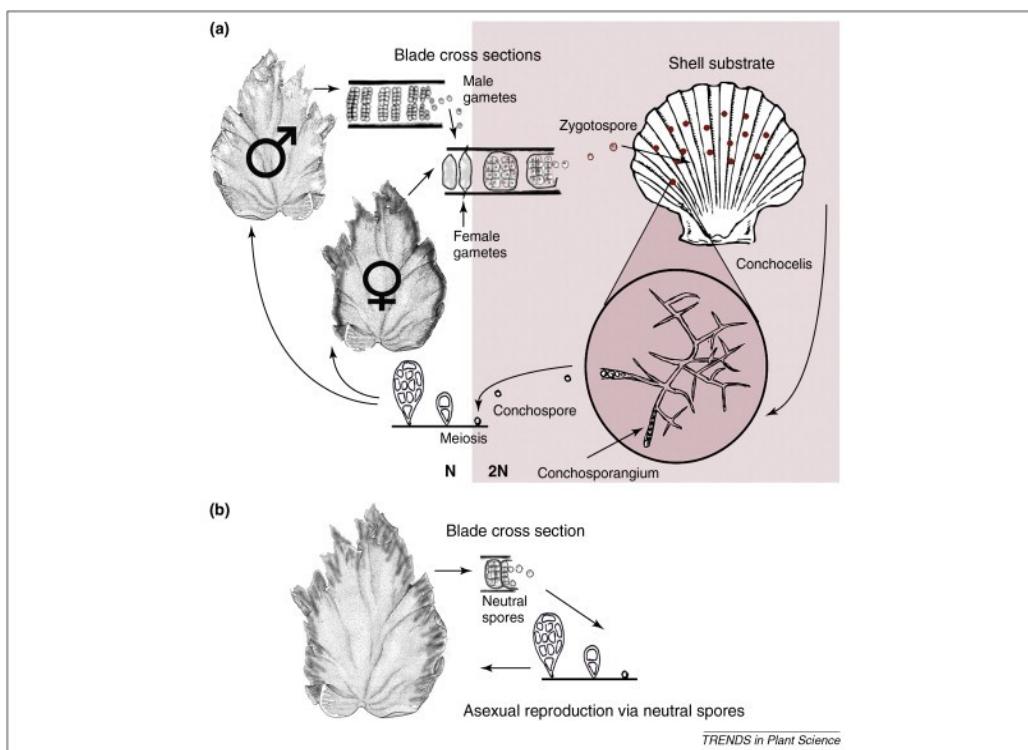


Figure A.2. Life-cycle of *Pyropia* spp, which is commonly a sexually reproducing species that requires a hatchery phase (Figure from Blouin et al 2010).

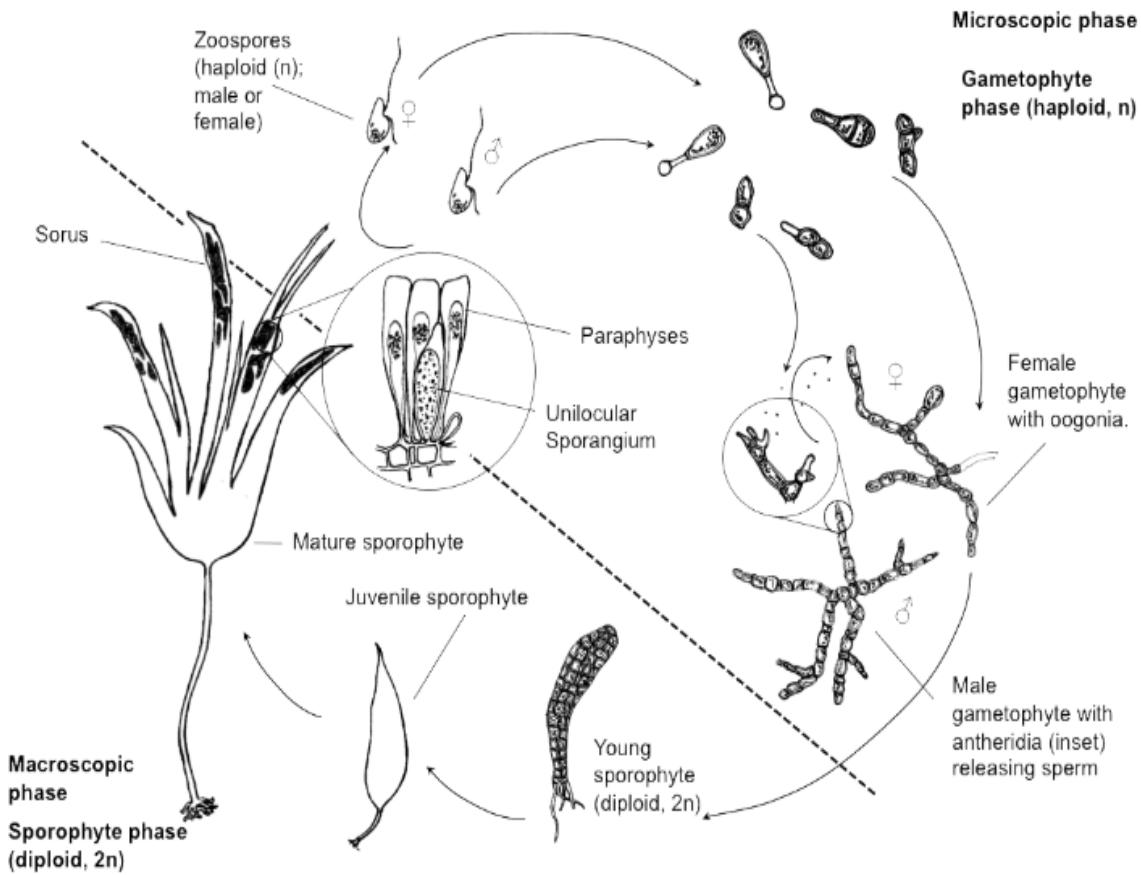


Figure A.3. Life-cycle of *Laminaria digitata* (Figure from Edwards and Watson, 2011).

Appendix B: Hydrographic conditions at our farm location in Santa Barbara, CA

Table B.1. Historic offshore ocean conditions at our farm site in Santa Barbara, CA. See graphs below for more information (Data sources: Brzezinski et al., 2013; Reed, 2017; CalCOFI, 2018; NOAA, 2018; Time and Date, 2018; Washburn, 2018).

	DIN (μM)	Photoperiod (hrs)	Surface Irradiance (mol photons / m^2d)	Temp ($^{\circ}\text{C}$)	Current (cm/s)	Salinity
January	3.9	10.12	38.63	14.27	7.31	33.33
February	3.8	10.98	48.25	13.96	7.68	33.38
March	6.5	11.95	58.20	13.80	7.63	33.5
April	7.3	13.05	65.95	14.02	7.69	33.68
May	4.70	13.97	71.18	15.30	7.77	33.68
June	2.00	14.45	66.95	16.40	8.05	33.58
July	1.1	14.23	68.46	17.57	7.18	33.53
August	1.2	13.43	64.96	14.47	6.93	33.48
September	1.20	12.38	56.39	17.80	7.88	33.45
October	1.10	11.33	50.37	17.17	8.44	33.41
November	1.00	10.37	44.16	16.55	8.79	33.44
December	2.50	9.87	43.49	13.90	7.97	33.36

Average Temperature near Bernard's Farm (2004-2017)

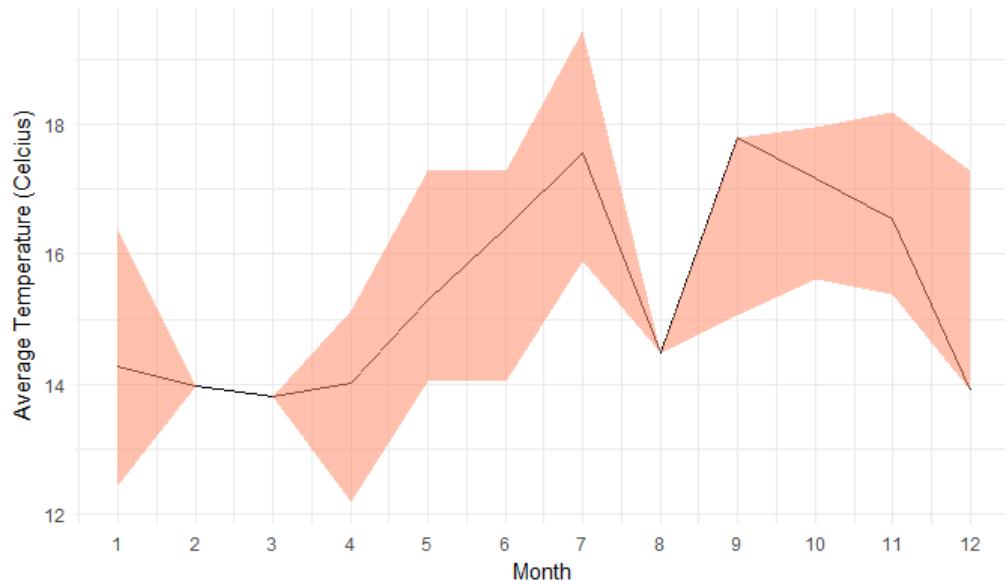


Figure B.1. Ocean temperature (°C) at our offshore farm location in Santa Barbara, CA was estimated from the monthly average over a 13-year time span (2004 to 2017) at CalCOFI station 81.7.43.5. Temperatures in May, June, September, and December were estimated using historic averages reported by NOAA for Santa Barbara or gapfilled using an average of adjacent months. Minimum and maximum temperatures are shown by the ribbon (Data sources: CalCOFI, 2018; NOAA, 2018).

Average DIN near Bernard's Farm (2002-2012)

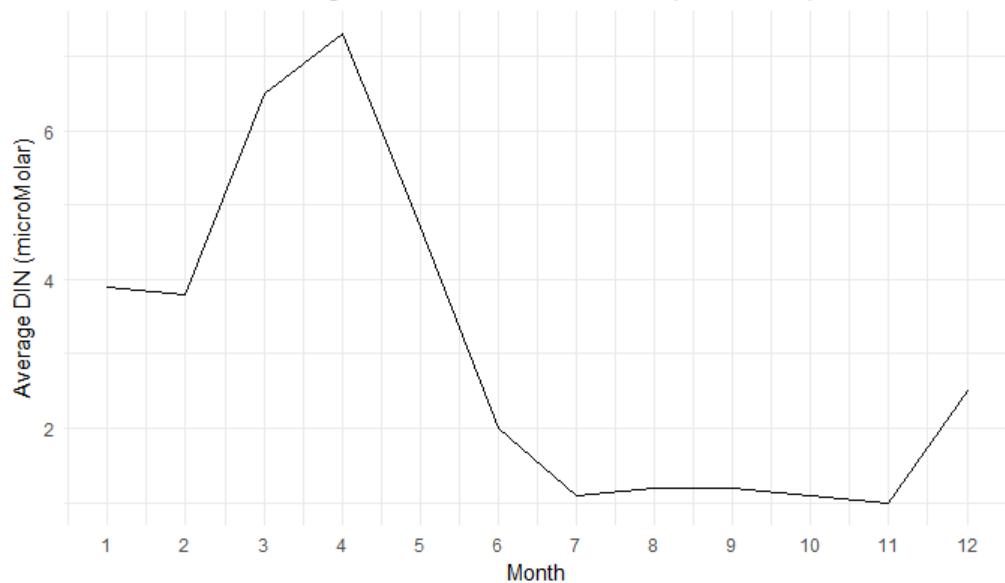


Figure B.2. Concentration of dissolved inorganic nutrients at our offshore farm location in Santa Barbara, CA was estimated from depth-averaged concentrations of nitrate, nitrite, and ammonium over the period 2002-2012. Original data collection was at sites Arroyo Burro, Arroyo Quemada, and Mohawk Reef. (Data source: Brzezinski et al. 2013).

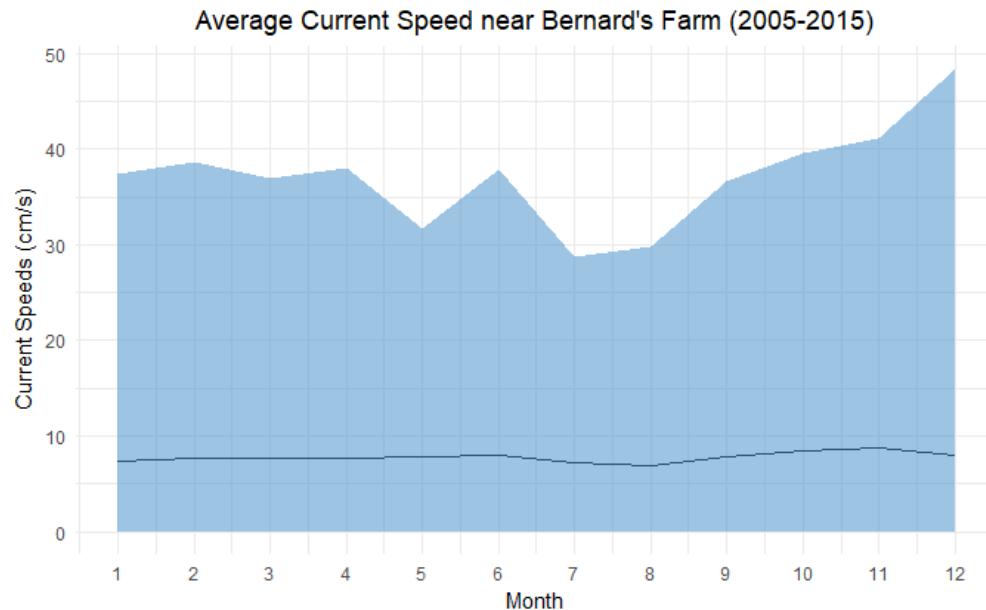


Figure B.3. Ocean current speed (cm/s) at the farm site was estimated from average and maximum moored CTD data at Mohawk Reef (2005-2015). Minimum current speed was assumed to be 0 cm/s for all months. (Data source: Washburn, 2018).

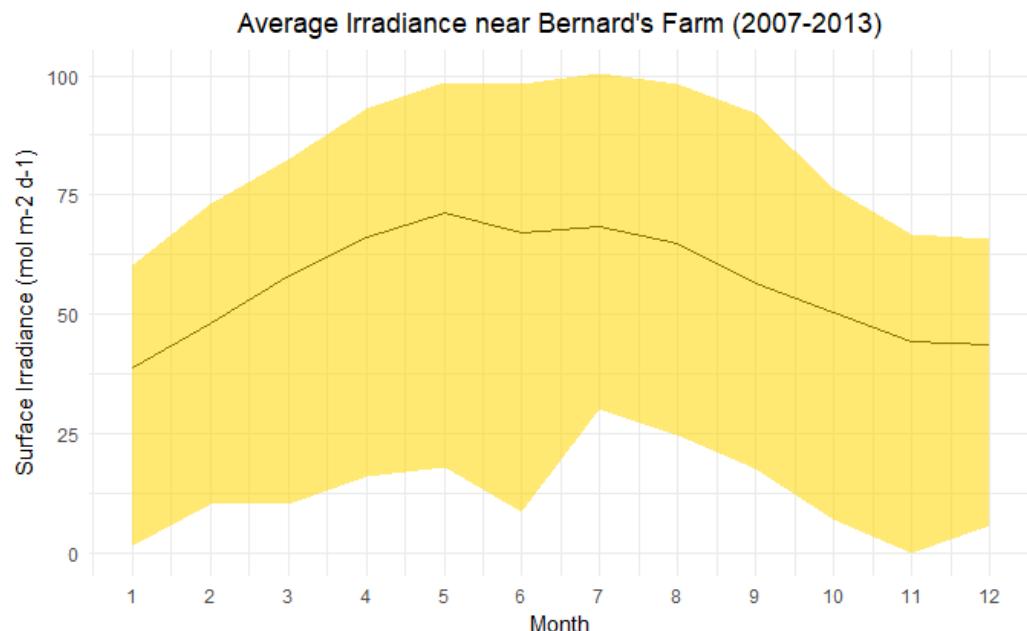


Figure B.4. Average monthly irradiance (2007-2014) in moles of photons per meter squared per day at our offshore farm location in Santa Barbara, CA. Minimum and maximum irradiance is shown in the ribbon. (Data source: Reed, 2017).

Data sources:

Brzezinski, M.A., D.C. Reed, S. Harrer, A. Rassweiler, J.M. Melack, B.M. Goodridge, and J.E. Dugan. (2013). Multiple sources and forms of nitrogen sustain year-round kelp growth on the inner continental shelf of the Santa Barbara Channel. *Oceanography* 26(3):114–123, <http://dx.doi.org/10.5670/oceanog.2013.53>.

CalCOFI. (2018). Hydrographic Cast Data March 1949 – Summer 2018 [Data file]. Retrieved from http://calcofi.org/downloads/database/CalCOFI_Database_194903-201708_csv_23Mar2018.zip.

NOAA National Oceanographic Data Center. (2018). Water Temperature Table of the Southern Pacific Coast. Available from https://www.nodc.noaa.gov/dsdt/cwtg/all_meanT.html.

Reed, D. (2017). SBC LTER: Kelp Removal Experiment: Daily photon irradiance at the surface and seafloor. Santa Barbara Coastal Long Term Ecological Research Project. doi:10.6073/pasta/e25f430bb589b4b16a3d97f6da2afe72

Time and Date. (2018). Yearly Sun Graph for Santa Barbara. Available from <https://www.timeanddate.com/sun/usa/santa-barbara>

Washburn, L., Gotschalk, C., & Salazar, D. (2018). Santa Barbara Coastal LTER Moored CTD and ADCP Data at Mohawk Reef (MKO), ongoing since 2005. Available from <http://sbc.lternet.edu/cgi-bin/showDataset.cgi?docid=knb-lter-sbc.2007>.

Washburn, L. (2018). SBCLTER: Ocean: Currents and Biogeochemistry: Moored CTD and ADCP Data at Mohawk Reef (MKO), ongoing since 2005. Santa Barbara Coastal LTER doi:10.6073/pasta/61ebe5fbec4663479ddae0eb5961d374.

Appendix C: Seaweed data

Table C.1. Low and high growth rates for candidate species both offshore and onshore. In some instances, the most reliable growth estimate found in the literature was a final annual production instead of daily relative growth rate. These are indicated as such below and the equivalent relative growth rate was calculated for use in our bioeconomic model.

Species	Offshore Growth	Onshore Growth	Initial Stocking Density	Max Stocking Density	Data Sources
<i>Gracilaria pacifica</i>	1.4 - 3.98% RGR per day	5.48 – 6.9% RGR per day	0.4 kg/m; 3.17 kg/m ³	11.9 kg/m ³	SARC, Halling et al 2005; SARC
<i>Ulva spp.</i>	3.3 – 6.67 kg/m ² annual production	10 – 17.6% RGR per day	0.375 kg/m ³	16.7 kg/m ² ; 27.8 kg/m ³	Titlyanov & Titlyanova 2010; Duke et al 1989; Hernandez et al 2005
<i>Pyropia perforata</i>	2.96 - 6.52 kg/m ² annual production	10 – 16% RGR per day	0.313 kg/m ³	NA	Parker 1974, Miura 1975; Kim et al 2007
<i>Laminaria spp.</i>	7 – 16 kg/m ² annual production	0.85 – 1.3 % RGR per day	8 kg/m ³	NA	Peteiro & Freire 2013, Watson & Dring 2013; Azevedo et al 2016

Table C.2. Notable ocean condition factors affecting candidate species growth and reproductive period, peak natural occurrence in Santa Barbara, harvest frequency, and the proposed cultivation season for each candidate species in offshore aquaculture systems.

Species	Factor(s) affecting growth	Reproductive Period	Natural Peak Occurrence (Bottom)	Harvest Frequency	Proposed Cultivation Season
<i>Gracilaria pacifica</i>	Nutrients (DIN), Light	May/July - October	March - October	Every 30 days	Feb-July
<i>Ulva lactuca, lobata, spp.</i>	Temperature, Light	June - August	August - October	Once a month	May-Oct
<i>Pyropia perforata</i>	Nutrients, Light, Temperature, Reproductive	Summer - Fall	March-Sep	Every 3 months	Dec-April
<i>Laminaria spp.</i>	Nutrients, Temperature, Reproductive	Summer - Fall	August - December	Once a year	October - March

Table C.3. Notable seawater condition factors affecting candidate species growth and reproductive period, peak natural occurrence in Santa Barbara, harvest frequency, and the proposed cultivation season for each candidate species in onshore aquaculture systems.

Species	Factor(s) affecting growth	Reprod. Period	Natural Peak Occurrence (Bottom)	Harvest Frequency	Proposed Cultivation Season
<i>Gracilaria pacifica</i>	Nutrients (DIN), Light	May/July - October	March - October	Weekly	Year round
<i>Ulva lactuca, lobata, spp.</i>	Temperature	June – August	August - October	3.5 weeks	Year round
<i>Pyropia perforata</i>	Nutrients, Light, Temperature, Reproductive	Summer - Fall (spores settle)	March-Sep	2.5 weeks	October to June
<i>Laminaria spp.</i>	Nutrients, Temperature, Reproductive	Summer - Fall	August (peak) - December	Every 2 months	October - April

Table C-4. Food market related information for California seaweeds.

Species	Geographic Range	Commercial Use	Nutritional Value	Farm Gate Price
<i>Gracilaria pacifica</i>	Alaska to Southern California	Agar/Food thickener Dried (human) Abalone feed	7-13% protein High fiber	Tank Grown: Carlsbad, \$15.4-22/kg; Goleta, \$8.8-17.6/kg; Monterey, \$88/kg
<i>Ulva spp.</i>	Alaska to Mexico	Popular edible: dried, toasted, or eaten fresh	13.6% protein	Tank Grown: Monterey \$88/kg
<i>Pyropia perforata</i>	Alaska to Baja (Intertidal and upper subtidal)	Dominant product in sushi, typically as dried sheets	36-47% dried protein	Commercial harvest: \$8 per oz, bulk \$110-143/kg
<i>Laminaria setchellii</i>	Alaska to Mexico (perennial)	Common in soups, often dried	8-15% protein	Commercial harvest: Mendocino \$97-105/lb (bulk); Sonoma \$171/kg

Appendix D: Seaweed yield summary

Table D.1. Model farm yield per unit of production

Species	Annual Yield per Unit	Farm Unit
Ogo	415 kg	1 longline (137 m)
Sea Lettuce	244 kg	1 net (27 m ²)
Nori	475 kg	1 net (27 m ²)
Kombu	1450 kg	1 longline (137 m)

Table D.2. Comparison of model calculated output and reported final yields taken from literature.

Species	On/off	Calculated Yield	Reported Final Yield
Ogo	Offshore	51,818 kg	>320,624 kg
Sea Lettuce	Offshore	4.68 kg/m ²	3.3 – 6.67 kg/m ²
Nori	Offshore	119.8 kg/net	80 - 160 kg/net
Kombu	Offshore	181,183 kg/25 acres	456,000 kg/25 acres
Ogo	Onshore	92 kg/1000-L tank	48 kg/1000-L tank
Sea Lettuce	Onshore	165 kg/1000-L tank	48 kg/1000-L tank
Nori	Onshore	47 kg/1000-L tank	48 kg/1000-L tank
Kombu	Onshore	53 kg/1000-L tank	48 kg/1000-L tank

Appendix E. Cost information

Table E.1. Costs for offshore cultivation of *Gracilaria pacifica* and *Laminaria setchellii* (Watson & Dring, 2011; D. Marquez, pers comm; B. Friedman, pers comm).

Materials	Single line (\$US 2017)	Total Farm Cost (US\$ 2017)
Fixed Dependent Costs		
Longline (Nylon rope: 147m/28mm)	\$612.26	\$76,531.88
Anchor rope (60 m x 2)	\$349.06	\$43,632.75
Chain (5m x 2)	\$130.90	\$16,362.50
Buoy (\$32 each, 6m spacing)	\$704.00	\$88,000.00
Trawl floats (x 2)	\$35.34	\$4,417.88
Shackles	\$52.36	\$6,545.00
Tying rope	\$130.90	\$16,362.50
Jeyco Anchors (x 2)	\$960.00	\$120,000.00
Deployment		\$8,175.00
Total cost (125 lines)		\$380,027.50
Fixed Independent Costs		
Boat (8mx2m Yamaha)		\$30,000.00

Motors (40hp)		\$3,870.00
Motors (25 hp)		\$3,354.00
Navigational/Radar buoy		\$3,711.00
Spar buoys (x4)		\$300.00
Truck used (transport)		\$13,000.00
Permits		
Permitting (FGC)		\$500.00
Permitting (Army Corp)		\$100.00
Permitting (CCC)		\$3,501.00
Broodstock collection (FGC)		\$525.00
Environmental Impact Report		\$25,000.00
Annual Costs		
Annual registration	\$545.00	\$545.00
Surcharge (\$25,000)	\$642.20	
Boat slip (avg. 5 yrs)	\$1,225.14	\$1,225.14

Boat maintenance	\$2,793.47	\$2,793.47
Rent (minimum: \$2 if over 10 acres)	\$250.00	\$250.00
Insurance (annual)	\$2,618.00	\$2,618.00
Telephone/postage (annual: harvest and deployment)	\$2,356.20	\$2,356.20
Diving (annual: harvest and deployment)	\$4,188.80	\$4,188.80
Protective clothing (annual)	\$1,309.00	\$1,309.00
Annual salary (owner)	\$50,000.00	\$50,000.00
Seeding labor (per)	\$3,000.00	\$3,000.00
Boat maintenance (annual)	\$2,793.47	\$2,793.47
Auto expenses (non-transport)	\$3,192.40	\$3,192.40
Marginal Costs (monthly)		
Cost of gas and oil for boat	\$483.63	\$2,898.00
Transport (produce and site)	\$244.67	\$244.67
Auto expenses (non-transport)	\$266.03	\$266.03
Harvest labor (each)	\$6,000.00	\$6,000.00

Gracilaria (stocking)	\$30,825.00	\$30,825.00
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Table E.2. Costs for offshore cultivation of *Pyropia perforata* and *Ulva lactuca* (Levine & Sahoo, 2009; KMO, 1982; Watson & Dring, 2011).

Material	Pyropia perforata (US\$ 2017)	Ulva lactuca (US\$ 2017)
Fixed Dependent Costs		
Seeding bags (Hamumbo Sheet)	\$3,204.36	NA
Nursery frames	\$55,902.92	NA
Giant wheel for net seeding	Missing	NA
Nets (\$14 each; 18m by 1.5 m)	\$69,660.00	\$19,902.86
Anchors/Anchor line (jeyco anchor)	\$215,550.00	\$215,550.00
Buoy (2 per net)	\$374,400.00	\$374,400.00
Rope-12mm	\$14,953.68	\$14,953.68
Rope-10mm	\$7,476.84	\$7,476.84
Rope-4mm	\$640.87	\$640.87
PVC each net (NA)	Missing	Missing
Baskets	\$1,548.00	\$1,548.00

Net bags	\$2,018.59	\$2,018.59
Equipment installation	\$61,920.00	\$61,920.00
Fixed Independent Costs		
Boat (8mx2m Yamaha)	\$60,000.00	\$30,000.00
Motors (40hp)	\$7,740.00	\$3,870.00
Motors (25 hp)	\$6,708.00	\$3,354.00
Harvester (net)	\$9,833.93	\$4,916.96
Motor (Honda 150-LPO)	\$4,295.70	\$2,147.85
Truck used (transport)	\$13,000.00	\$13,000.00
Navigational/Radar buoy	\$3,711.02	\$3,711.02
Spar buoys (x4)	\$300.00	\$300.00
Cold storage (nets)	Missing	NA
Permits		
Permitting (FGC)	\$500.00	\$500.00
Permitting (Army Corp)	\$100.00	\$100.00

Permitting (CCC)	\$3,501.00	\$3,501.00
Broodstock collection (FGC)	\$525.00	\$525.00
Environmental Impact Report	\$25,000.00	\$25,000.00
Annual Costs		
Annual registration	\$545.00	\$545.00
Surcharge (\$25,000)	\$642.20	\$642.20
Boat slip (avg. 5 yrs)	\$1,225.14	\$1,225.14
Rent (minimum: \$2 if over 10 acres)	\$250.00	\$250.00
Boat maintenance	\$2,793.47	\$2,793.47
Insurance (annual)	\$2,618.00	\$2,618.00
Telephone/postage (annual: harvest and deployment)	\$2,356.20	\$2,356.20
Diving (annual: harvest and deployment)	\$4,188.80	\$4,188.80
Protective clothing (annual)	\$1,309.00	\$1,309.00
Annual salary (owner)	\$50,000.00	\$50,000.00
Marginal		
Employee (2.5 Fulltime)	\$57,000.00	NA

Employee (deployment)	NA	\$6,000.00
Marginal costs (monthly)		
Cost of gas and oil for boat	\$1,260.00	\$483.63
Transport (produce and site)	\$244.67	\$244.67
Auto expenses (non-transport)	\$266.03	\$266.03
Harvest labor (each)	NA	\$6,000.00

Table E.3. Costs for onshore cultivation of seaweed species for a single tank (Watson and Dring, 2011; D. Marquez, pers comm; D. Bush, pers comm; N. Caruso, pers comm).

Materials	Per unit cost (US\$ 2017)
Tank (1000L)	\$332.49
Ball valves (2 cm)	\$23.56
Aerators	\$7.85
Air Blower: Reischie	\$1,898.05
Pipework	\$9.82
Joints, glue, sundries	\$6.55
Switch gear (installed)	\$392.70
Shading net (50m2)	\$3.93

Permitting (CCC)	\$3,501.00
Broodstock collection (FGC)	\$525.00
Seawater meter (hot and cold)	\$2,400.00
Plumber line	\$890.00
Electrical PME/transformer assembly	\$27,500.00
Telephone	\$250.00
One-acre lease	\$6,000.00
Annual registration	\$545.00
Surcharge (\$25,000)	\$642.20
Annual salary (owner)	\$50,000.00
Water pumping fee (inc. electricity)	\$12.31
Labor (1 FTE per 60 tanks)	\$3,750.00
Hatchery start-up	\$65,490.94
Hatchery annual	\$38,307.50
Dryer start-up	\$18,326.00
Dryer annual	\$3,921.06

Appendix F. Cash flow analyses (cont.)

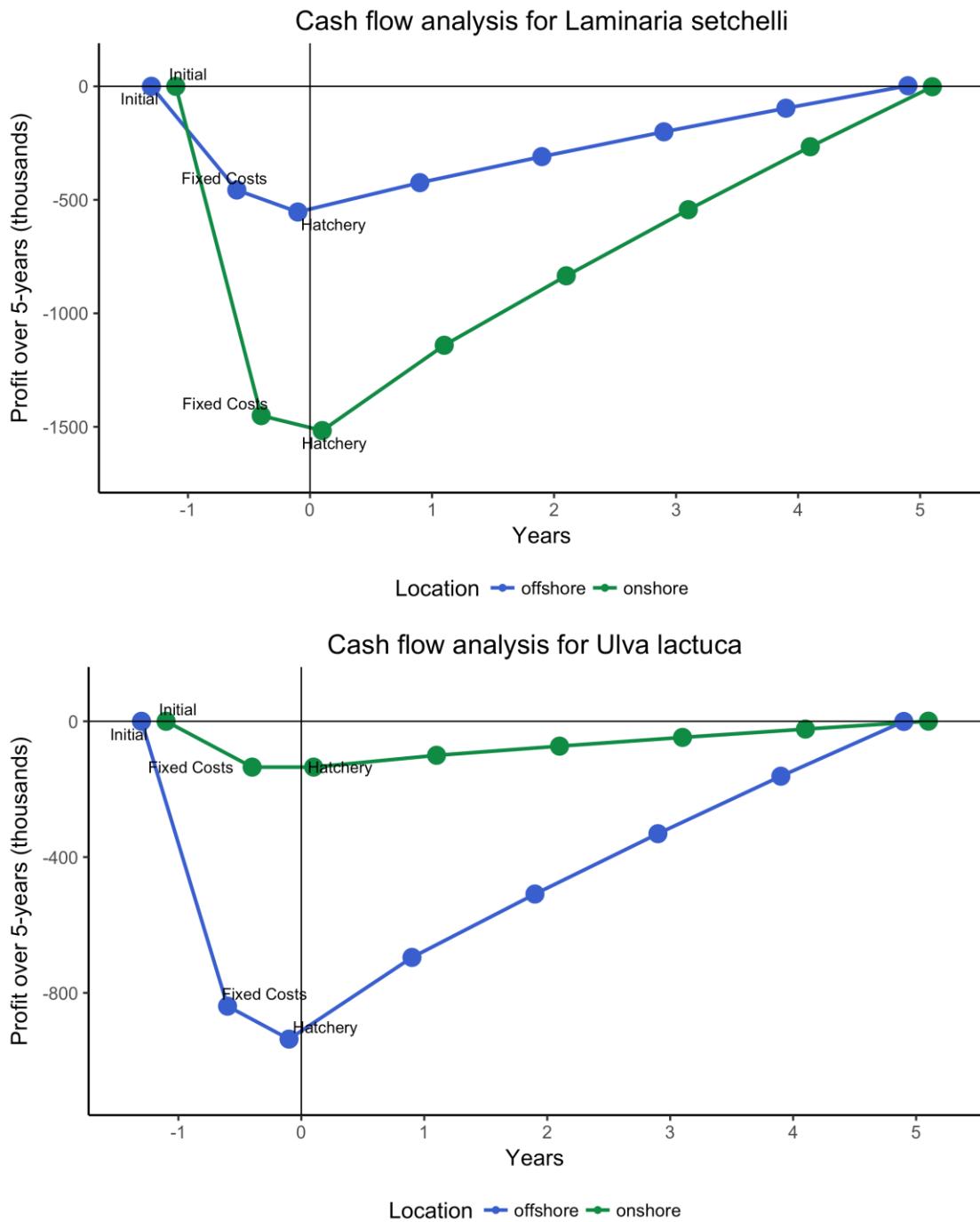


Figure F.1. The cash flow (thousands of \$US) for the payback of *Laminaria setchellii* and *Ulva lactuca* at the break-even price under mean growth scenarios at 5-years in onshore and offshore locations (see Figure 2.1 for break-even information). The upfront capital costs are labelled on the graph, and include fixed costs such as permitting and structural costs and costs associated with establishing a hatchery, for *Laminaria setchellii* and *Ulva lactuca*.