# #2.2: Creating an RNA-seq analysis pipeline

SLURM scripting and processing
Using bioinformatics tools to process RNA-seq data
Transferring files between CHPC and personal computer
IGV

#### Access the slides and files here:

https://github.com/j-berg/bioinformatics\_bootcamp

#### Introduction to SLURM

- Schedule jobs to be run on high-performance compute nodes
- Keeps the usage fair for everyone
  - If you've run a bunch of jobs recently, you are put at the end of the queue
- Submit a SLURM script (a modified bash script)



### Scratch Directory

- Temporary file storage
- Unlimited space
- Be sure to clear out files you don't need!
- If using human/mouse samples, need to build your index here due to storage constraints

Location:

\$ mkdir /scratch/general/lustre/uNID

### SLURM job header

```
#!/bin/bash <-tell script where to find bash #SBATCH --time=72:00:00 <- script time-out <- how many nodes to use #SBATCH --o /uufs/chpc.utah.edu/common/home/uNID/slurmjob-%j <- Where to write the job log #SBATCH --partition=notchpeak <- Where to run the job
```

source /uufs/chpc.utah.edu/common/home/uNID/miniconda3/etc/profile.d/conda.sh

source activate class <- Access conda environment

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% is a magic character -- %j will auto-fill SLURM job ID

#### Create a workflow for genome indexing

```
#!/bin/bash
#SBATCH --time=72:00:00
#SBATCH --nodes=1
#SBATCH -o /uufs/chpc.utah.edu/common/home/u
                                                   /slurmjob-%j
#SBATCH --partition=notchpeak
source /uufs/chpc.utah.edu/common/home/u
                                                /miniconda3/etc/profile.d/conda.sh
source activate class
SCRUSER=/scratch/general/lustre/u
REF=/scratch/general/lustre/u
                                    /yeast_index
GTF=$REF/Saccharomyces_cerevisiae.R64-1-1.100.gtf
# initialize reference folder in scratch directory
mkdir -p $REF
mv ~/reference_yeast/* $REF
# Isolate FASTA files
mkdir -p $REF/fastas
mv $REF/*.fa $REF/fastas
# Generate STAR index
STAR --runMode genomeGenerate --genomeDir $REF/genome --genomeFastaFiles $REF/fastas --sjdbGTFfile $REF/$GTF --runThreadN
32 --sjdbOverhang 50
```

#### Run a workflow for genome indexing

```
Every 1.0s: squeue -u u

JOBID PARTITION NAME USER ST TIME NODES NODELIST(REASON)
1398079 notchpeak make_yea u R 3:04 1 notch012
```

### Creating an analysis workflow

```
source /uufs/chpc.utah.edu/common/home/u
                                               /miniconda3/etc/profile.d/conda.sh
source activate class
SCRDIR=/scratch/general/lustre/$USER/$SLURM_JOBID
mkdir -p $SCRDIR
INPUT=/uufs/chpc.utah.edu/common/home/u
                                              /seq_files
REF=/scratch/general/lustre/u
                                    /yeast_index
GTF=$REF/Saccharomyces_cerevisiae.R64-1-1.100.gtf
FILES=(SRR1166442 SRR1166443 SRR1166444 SRR1166445 SRR1166446 SRR1166447)
OUTPUT=/uufs/chpc.utah.edu/common/home/u
                                               /seq_output
 Move raw data
mkdir -p $SCRDIR/input
mkdir -p $SCRDIR/output
cp $INPUT/* $SCRDIR/input
mkdir -p $SCRDIR/output/preprocess
 nkdir -p $SCRDIR/output/alignment
 nkdir -p $SCRDIR/output/postprocess
mkdir -p $SCRDIR/output/count
mkdir -p $SCRDIR/output/qc
mkdir -p $OUTPUT
cd $SCRDIR/.
```

### Creating an analysis workflow

```
for FILE in ${FILES[@]}; do
      echo "Processing ${FILE}"
       fastp -- thread 20 -1 30 -q 28 \
              -i $SCRDIR/input/${FILE}.fastq \
              -o $SCRDIR/output/preprocess/${FILE}.fastq \
              -j $SCRDIR/output/preprocess/${FILE}.json \
              -h $SCRDIR/output/preprocess/${FILE}.html
      echo "Preprocessing QC..."
       fastqc -q $SCRDIR/output/preprocess/${FILE}.fastq
              -o $SCRDIR/output/qc/${FILE}
      STAR --runThreadN 20 --sjdbOverhang 50 \
              --outSAMunmapped Within --outSAMtype BAM Unsorted --quantMode TranscriptomeSAM \
              --genomeDir $REF/genome \
               --sjdbGTFfile $GTF \
              --readFilesIn $SCRDIR/output/preprocess/${FILE}.fastg \
              --outFileNamePrefix $SCRDIR/output/alignment/${FILE} #will end in _Aligned.bam
       samtools sort -- threads 20 \
              -o $SCRDIR/output/postprocess/${FILE}_sorted.bam \
               $SCRDIR/output/alignment/${FILE}_Aligned.bam
       samtools index -@ 20 \
               $SCRDIR/output/postprocess/${FILE}_sorted.bam
      echo "Counting..."
      htseq-count -q -f bam -m intersection-nonempty -t exon -i gene_id -r pos -s no \
              $SCRDIR/output/postprocess/${FILE}_sorted.bam \
               $GTF > $SCRDIR/output/count/${FILE}.tsv; done
```

## Creating an analysis workflow

```
# Summarize QC
multiqc $SCRDIR/output

# Clean-up
mv $SCRDIR/output/postprocess/*_sorted.bam $OUTPUT
mv $SCRDIR/output/postprocess/*_sorted.bam.bai $OUTPUT
mv $SCRDIR/output/count/*.tsv $OUTPUT
mv $SCRDIR/output/qc/*.html $OUTPUT
mv $SCRDIR/output/qc/*.html $OUTPUT
mv $SCRDIR/output/*.html $OUTPUT
```

### Helpful SLURM commands

```
Start a slurmjob:
$ sbatch script.sh
Cancel a slurmjob:
$ scancel jobid
See your jobs in queue:
$ squeue -u uNID
Live update of queue:
$ watch -n1 squeue -u uNID
```

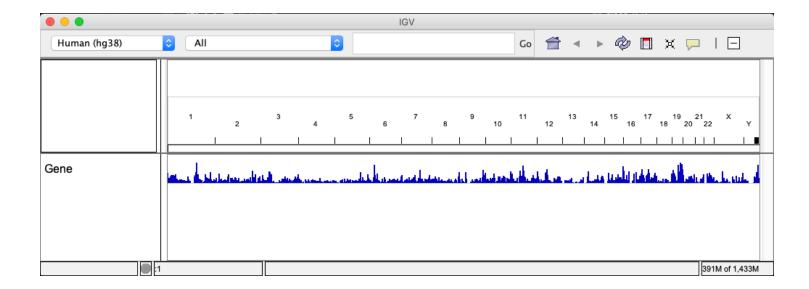
## Transferring files to your personal computer

From your personal computer/terminal

```
$ scp uNID@notchpeak.chpc.utah.edu:~/path/to/file.bam ./
$ scp uNID@notchpeak.chpc.utah.edu:~/path/to/file.bam.bai ./
```

#### Visualizing read pile-ups with IGV

- Download IGV:
  - https://software.broadinstitute.org/software/igv/download
- Drag and drop BAM file into viewer



#### Homework

- For the dataset you previously downloaded, download the appropriate reference files for that model organism
- Generate a genome index and store in the Scratch directory
- Create and run a script that processes each of the files you downloaded
- Transfer one of the alignment files to your personal computer and open in IGV. Find a gene whose transcripts (isoforms) seem to be differentially expressed.