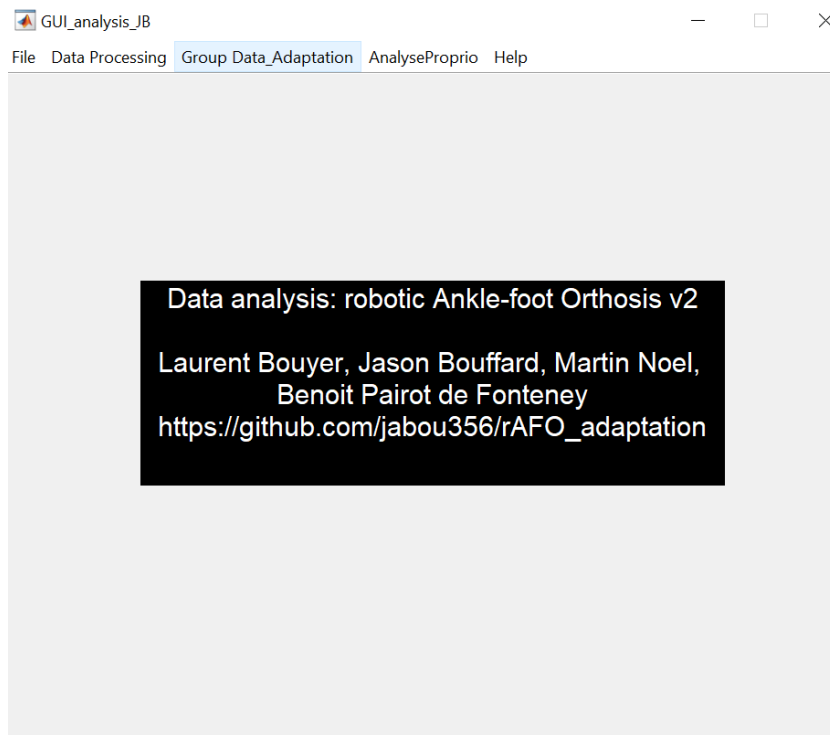


GUI_analysis_JB help file

GUI_analysis_JB has been developed by Jason Bouffard with the collaboration of Laurent Bouyer, Martin Noel (LOKOMAT) and Benoit Pairot de Fonteney between 2010 and 2018 to analyse data from the robotic ankle foot orthosis (Noel et al. 2008, IEEE Trans Neural Syst Rehabil Eng)



Here is a brief list of publications using the current or previous versions of GUI_analysis_JB, or embedded functions

Noel et al. 2008, IEEE Trans Neural Syst Rehabil Eng
Noel et al. 2009, J Neuroeng Rehabil
Blanchette et al. 2011, Gait Posture
Bouffard et al. 2014, J Neuroscience
Fournier-Belley et al. 2016, Gait Posture
Bouffard et al. 2016, Neural Plasticity
Bouffard et al. 2016, J Neurophysiol

Getting started

You can get the up-to-date version of the toolbox by downloading the latest release or cloning the master branch on: https://github.com/jabou356/rAFO_adaptation

To launch the program, just run the GUI_analysis_JB.m file. All necessary paths will be added to your Matlab Search paths. You will then be asked to open your project's parent folder. The

program will load the config.mat (see Annexe 1 for the content of config file) file or create and save a default one if it doesn't exist. If you want to modify the config.mat file, just do it manually. The program brings you back to the project's parent folder and save some files in it, so organize your data structure to navigate easily between subfolders during data analysis.

Step by step guide

Process individual data files

Transfer your .raw file from WinVision into a .mat

File->Convert *.raw vers *.mat

You will have to select the *.raw file you want to convert, then save it as a *.mat file. Repeat for each file.

Output: Inputfile.mat

data(1:number of channels,1:number of samples) matrix with unfiltered data.

Combine all .mat files for one subject into a single one

Data Processing -> Combine Files from WinVisio

This function will process data (apply filters and gains) and combine multiple files. The function will first ask you to go into your subject directory (i.e. the one with your .mat files). If you have a subject specific config.mat file in your subject directory, it will be loaded. If no such file exist, it will use the generic config.mat file loaded from the project parent directory. You should only add a subject specific config.mat file in the subject folder if it is necessary (different filter to use for this subject, different channel number, etc.).

Output: combined_data.mat:

data(1:number of channels,1:number of samples): matrix with unfiltered data

fdata(1:number of channels,1:number of samples): matrix with filtered data and appropriate gains

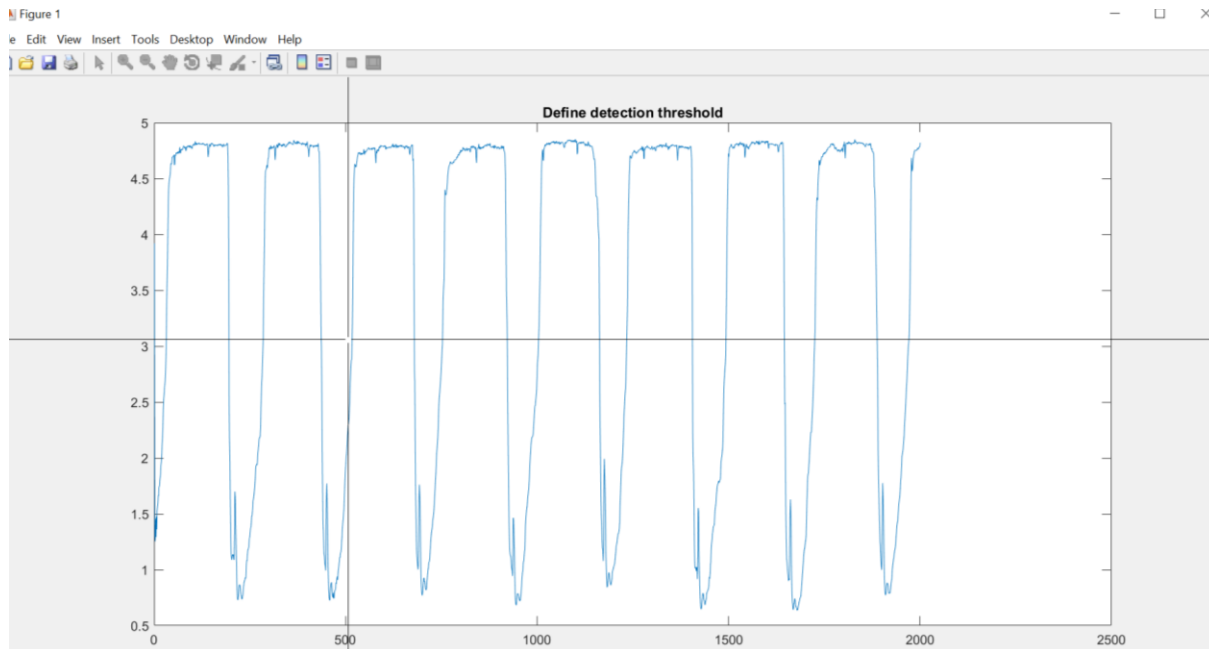
Cut file into individual strides

Data Processing -> Prepare Table -> Cut Tables Lokomat

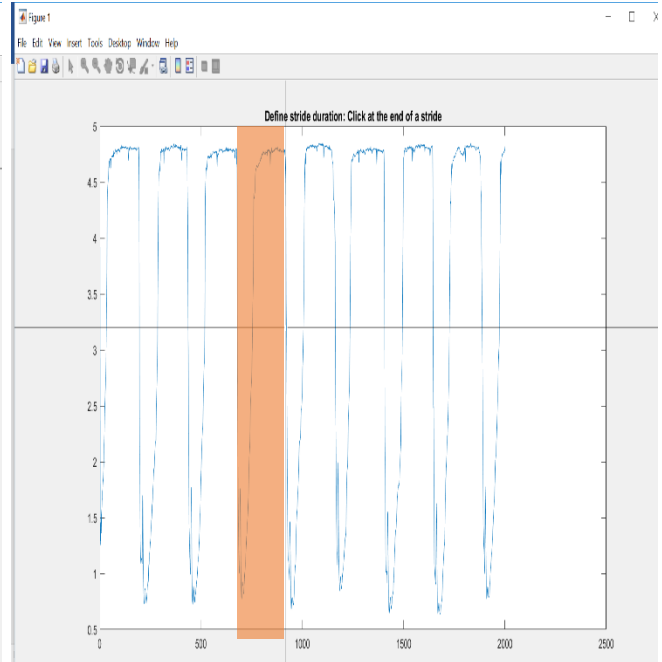
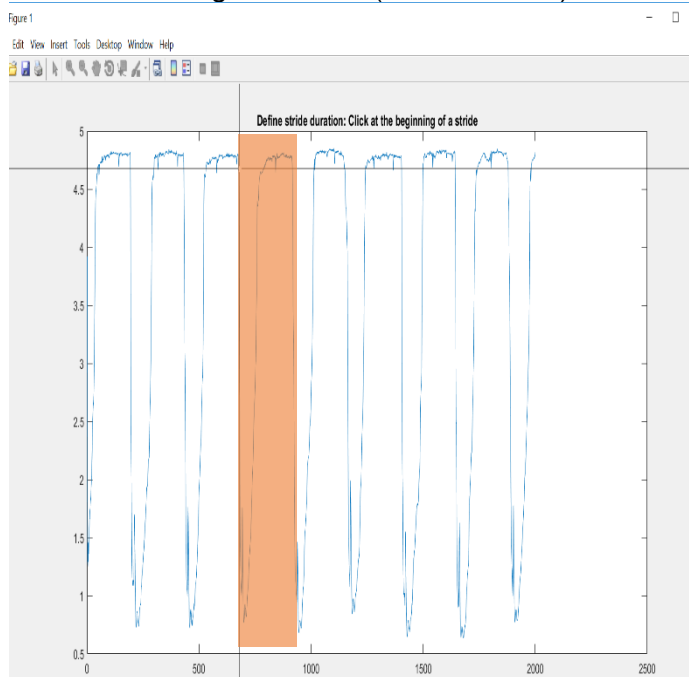
This function partition data in individual strides based on config.Sync_channel and other associated configuration parameters. It will also tag strides containing force field or reflexes (config.ISFF_channel, config.ISRFLX_channel).

Once you activate the menu, you will have to select your subject directory. Then the function will load the combined_data.mat file and will look for a subject specific config.mat (otherwise it will take the generic configuration file).

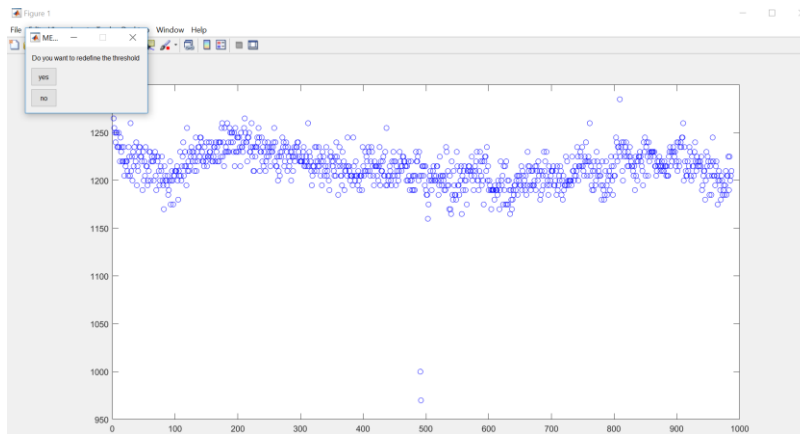
The config.Sync_channel's signal (usually heelstrike) will be plotted and you will have to select your synchronisation threshold. An optimal threshold should be crossed once by cycle (with in the config.trig_direction).



Then you will need to define an approximate stride duration by selecting the beginning and the end of a targeted stride (in the red box).

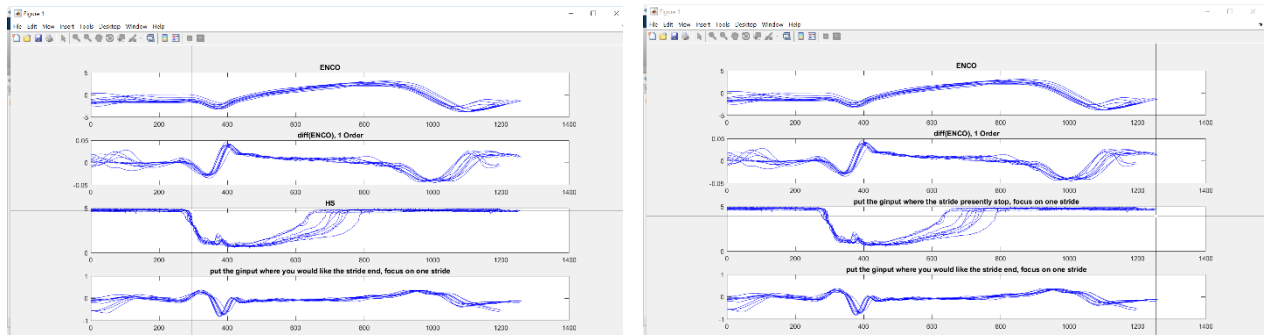


You will then be asked if you want to redefine the threshold based on a figure illustrating strides duration. If most of the strides fall within the range of expected duration, click No. If it is not the case, click on Yes and you will be asked to redefine your threshold and stride duration. This is an infinite loop. If you are not able to get a good synchronisation, just click No and modify your config.mat file to choose another Sync_channel or change other parameters (Sync_filter, Sync_diff).

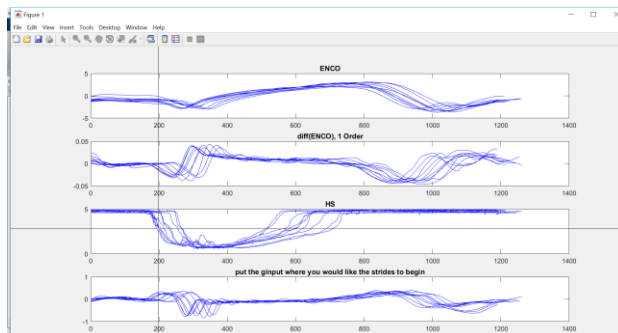


Once you accepted the data partitioning, another figure will be plotted with some signals you defined in `config.setOffsetChan` and `config.setOffsetdiff`. This step is to allow you to create an offset by choosing where you would like to see your strides ending or beginning.

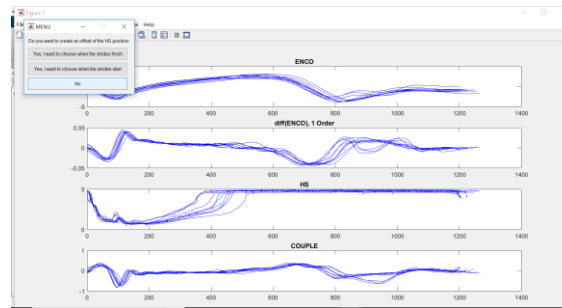
If you say that you would like to choose when the strides finish, you will have to focus on a single stride, define where you would like to see it finish and where it finished in the current figure. Then, the program will shift all strides by the amount of time separating your two clicks. In the example below, I indicated that I would like to see my stride finishing at the onset of heelstrike by focusing on the longest stride.



If you say that you would like to choose where the strides begin, you only need to click once, and the program will shift all strides from the selected xvalue (here around 200 ms).



In all case, you would like to get strides beginning at heelstrike (figure below. Those offsetting methods can be particularly useful when your config.Sync_channel is not a footswitch.



Output: Table_data.mat

Config : config file used to create Tables

Table: Contain partitioned data for all signals in the config.chan_name order.

Cycle_Table: Contains information about each stride.

(The structure of Table and Cycle_Table is presented in Annexe2.

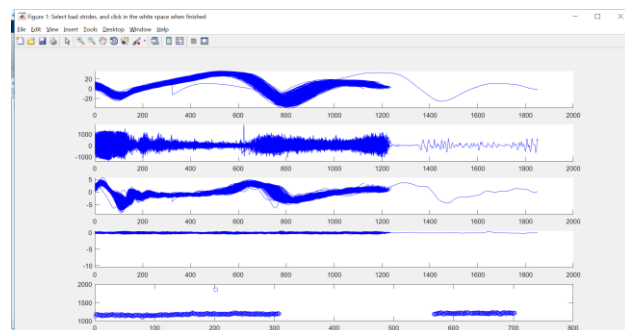
Remove bad trials

Data Processing -> validate cycles -> remove bad superpose

This function is used to visually scan the data and remove bad trials for further analyses. Bad trials are not deleted, but tagged in the 3rd row of Cycle_Table (1 = valid, 0 = non-valid).

When you call the remove bad trials function in the GUI, you will be asked to indicate which signals you would like to see to validate your strides. Enter each of the signal (as named in config.chan_name) to plot with quote and between {} (e.g. {'ENDO', 'TA', 'COUPLE', 'CONS_F'}).

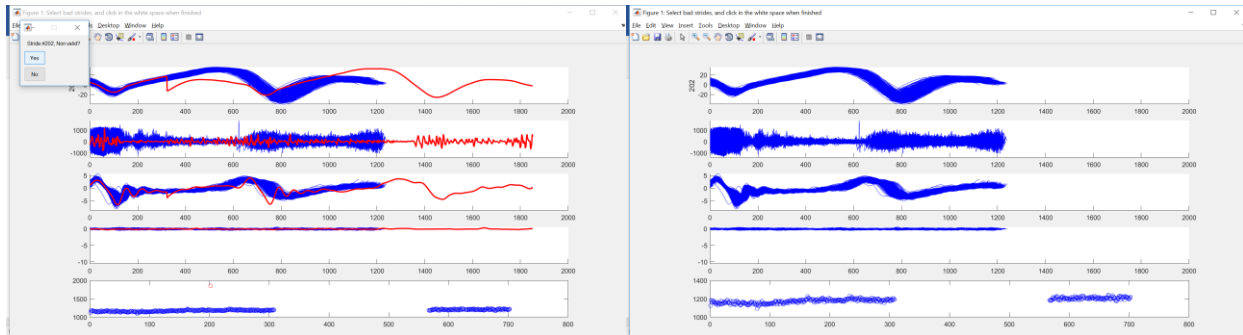
Then the selected channels will be plotted as well as each individual strides duration (lowest panel). First, strides without any event (Force fields or reflexes), will be presented.



From this figure, you can select a stride that appears not valid in any of the subplots (including the lowest one). It will then be highlighted in red and you will be asked if you want to remove this stride. If you click yes, the stride will be removed from the plot and marked as non-valid in Cycle_Table.

Output: Table_data.mat

Same as Cut_table, but with Cycle_Table updated.



When you consider that all the displayed strides are valid, you click in the white space in any subplot. Strides in the next condition (e.g. force field) will then be displayed.

Motor adaptation analysis (Group files)

Combine Group Data

GroupData Adaptation-> Combine Group Data

This function combines multiple Table_data.mat files into a Group structure for the selected signals.

When you call the function, you will first have to indicate if you already have a GroupData file. If you have one, you will load it to add new subjects at the end of the file. If you don't a new file will be initialized. Then you will have to indicate which signals you would like to combine into a Group Structure. . Enter each of the signal (as named in config.chan_name) to plot with quote and between {} (e.g. {'ENCO', 'TA', 'COUPLE','CONS_F'}). Note that if you want to generate built in ENCO and TA variables (will be presented late), you minimally need {'ENCO', 'TA', 'CONS_F'}.

When you are done adding subjects to the group structure, you will have to save it. Place the GroupData file in a strategic folder as new files will be saved there further in the analysis process.

Output: GroupData.mat

GroupData.Signal : Contains all data for a given Signal.

GroupData.Cycle_Table : Contains Cycle_Table information for all included subjects.

(The structure of GroupData file is presented in Annexe 3)

Identifications cycles

GroupData Adaptation-> Identification cycles

This function identify strides critical for the motor adaptation paradigm (Last baseline, Last force field, Last stride of the experiment).

You will just have to load your GroupData file.

Output: CyclesCritiques.mat

Annexe 1: config.mat

Channel identifier (all vectors, and cell array {chan names})

```
config.EMG_channels = [1,2,3,4,5]; %Original EMG chan number in WinVisio
```

```
config.Other_channels = [8,14,15,16]; % Original chan number (not EMG, not squared) in WinVisio
```

```
config.Square_channels = [10, 12]; % Original chan number (square waves) in WinVisio
```

```
config.chan_name = {'TA','SOL','GM','VL','RF','Knee','CONS_F',...  
    'ENCO','COUPLE','Response','HS'}; %chan_name:EMG goes first, then other_channels,  
then square channels
```

Identifier event channels with properties (all scalars)

```
config.ISFF_channel = 7; % Channel with FF command (e.g. CONS_F)
```

```
config.FFdetect_level = 0.6; % Threshold for FF detection (should be an positive scalar)
```

```
config.ISRFLX_channel = 0; %Channel to identify RFLX, catch, anticatch (e.g. Memory gate)
```

```
config.RFLXdetect_level = 0.4; % Threshold for event channel (should be a positive scalar)
```

Partition table into individual strides (all scalars or char)

```
config.Sync_channel = 11; % Channel # used to partition data
```

```
config.trig_direction = '<'; %Direction of the Sync_channel on which you want to trigger '<':  
descending, '>': ascending
```

```
config.trig_diff = 0; % set to diff order of sync chan (0 if no differentiation)
```

```
config.trig_lowpass = 0; %Set low pass filter frequency of sync chan (0 if no filter)
```

```
config.trig_Nlowpass = 0; % Set low pass order if to be used
```

```
config.pct_refractaire = 0.8; % Duration of refractory period (must be between 0 and 1)
```

Preprocessing properties

```
config.chan_gain = [250, 250, 250, 250, 250, 90, 4, 8, 10, 1, 1]; % vector with chan gains
```

```
config.EMG_filter = [20 450]; %:EMG :[low pass Fz, High pass Fz]
```

```
config.Other_filter = 15; % Other channles:[Low pass Fz]
```

```
config.Order = 2; %filter order (divide the wanted number by two as we use filtfilt)
```

```
config.sFz = 1000; %Sampling frequency (scalar)
```

Config for Adaptation Analyses

`config.useSync = 1; %useSync:1: Use sync data for analyses (e.g. middle pushoff), 0:start at first data point of each stride`

Config for Proprio Analysis

`config.AnkleName = 'ENCO'; % Name of Ankle signal for proprio`

`config.COUPLEName = 'COUPLE'; % Name of COUPLE signal for proprio`

`config.ResponseName = 'Response'; % Name of Response signal for proprio`

`config.ForcecommandName = 'CONS_F'; % Name of Force command signal for proprio`

`config.ResponseTh = 1.5; % Threshold for the detection of the button`