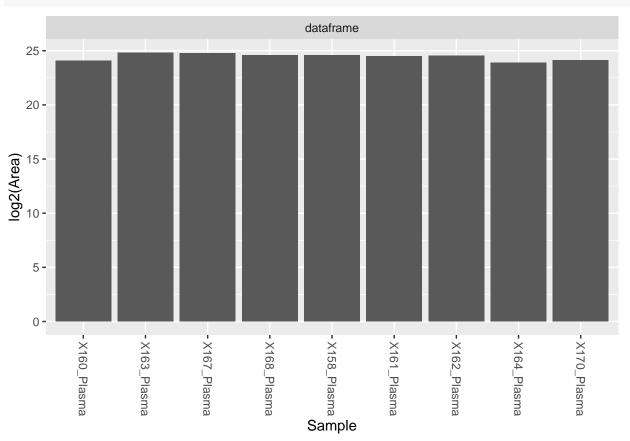
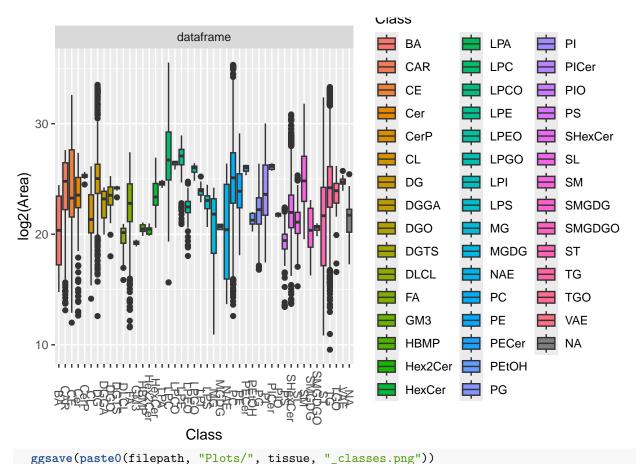
```
# Input Parameters
  tissue <- "Plasma"
  filepath <- "/Users/usa/Desktop/Zaganjor lab lipidomics results/lipidomics-analysis/"
# Load packages
  require(tidyverse)
  require(lipidr)
  require(viridis)
# Read in data
  tissue_data <- read.csv(pasteO(filepath, tissue, ".csv"))</pre>
# Remove extraneous col
  tissue_data$lipid_id <- NULL</pre>
# Remove "bad" entries (eg. 13-Docosenamide)
  tissue_data <- subset(tissue_data, grepl(":", lipids))</pre>
#remove NAs
  tissue_data <- na.omit(tissue_data)</pre>
# Convert Lipid Names to Systematic Nomenclature
  temp <- tissue data$lipids</pre>
  fix = sub(";(0)\d*)", "()\1)", temp)
  fix = sub(" 0-", "0 ", fix)
  fix = sub(" P-", "P ", fix)
  fix = sub("-FA", "/", fix)
  fix = sub(";(.*)$", "(\1)", fix)
  fix <- sub("([^\\)])\\|", "\\1(m)|", fix)
  fix \leftarrow sub("(.*\\mbox{(m}\))\\|(.*$)", "\\1|\\2(m2)", fix)
  fix <- sub("PE-Cer", "PECer", fix)</pre>
  fix <- sub("PI-Cer", "PICer", fix)</pre>
  fix <- sub("LPE-N ", "LPEN ", fix)</pre>
  fix <- sub("LPE-N ", "LPEN ", fix)</pre>
  tissue_data$converted_lipid_name <- fix</pre>
  all_match <- lipidr::annotate_lipids(tissue_data[[ncol(tissue_data)]])</pre>
  bad_match <- all_match %>% filter(not_matched)
  good_match <-subset(all_match, not_matched == "FALSE")</pre>
  tissue_data %>%
    select(lipids, converted_lipid_name) %>%
    write_csv(paste0(filepath, "Results/", tissue, "_name_converted.csv"))
# Begin lipidr analysis - setup lipidr object
  tissue_data$lipids <- tissue_data$converted_lipid_name</pre>
  tissue_data$converted_lipid_name <- NULL</pre>
  d <- as_lipidomics_experiment(tissue_data)</pre>
  meta <- read.csv(paste0(filepath, tissue, "_meta.csv"))</pre>
  d <- add_sample_annotation(d, meta)</pre>
# QC
 plot_samples(d, type = "tic", log = TRUE)
```

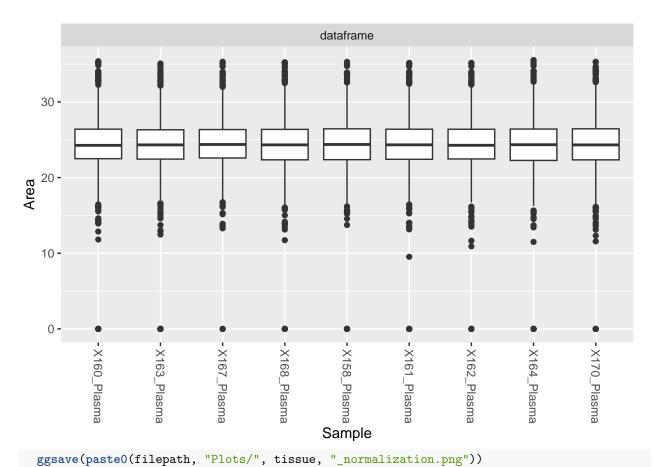


ggsave(paste0(filepath, "Plots/", tissue, "_samples.png"))
plot_lipidclass(d, "boxplot")

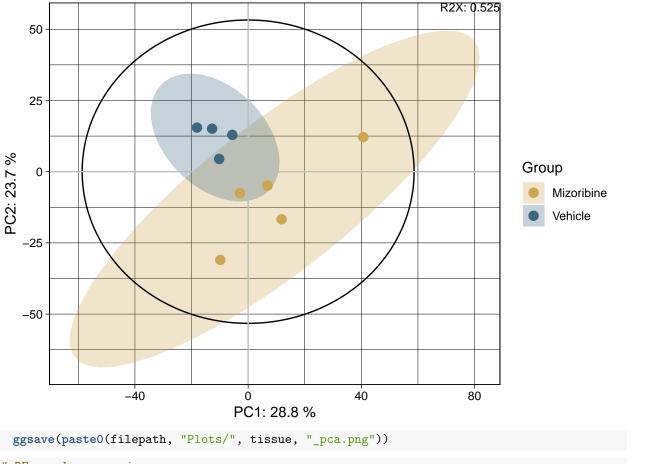


Resolve Duplicate Lipids (from multiple transitions) by selecting maximum, Normalize with PQN method,

d_summarized <- summarize_transitions(d, method = "max")
d_normalized <- normalize_pqn(d_summarized, measure = "Area", exclude = "blank", log = TRUE)
plot_samples(d_normalized, "boxplot")</pre>



```
# PCA
mvaresults <- mva(d_normalized, measure="Area", method="PCA")
pca_obj <- plot_mva(mvaresults, color_by="Treatment", components = c(1,2))
pca_obj + theme_linedraw() + scale_color_manual(values = c("Mizoribine" = "#CCA74E", "Vehicle" = "#3C</pre>
```



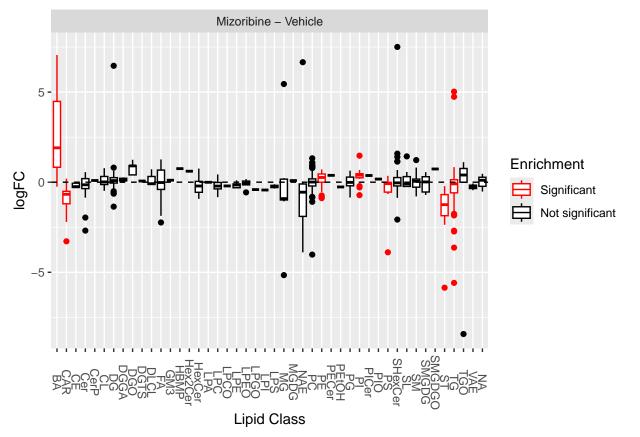
```
# DE - make comparisons
de_results <- de_analysis(data=d_normalized, group_col = "Treatment", Mizoribine - Vehicle, measure=
significant_molecules(de_results, p.cutoff = 0.05, logFC.cutoff = 0)

## named list()
write_csv(de_results, paste0(filepath, "Results/", tissue, "_de.csv"))

# LSEA
enrich_results <- lsea(de_results, rank.by = "logFC")
sig_lipidsets <- significant_lipidsets(enrich_results)
write_csv(enrich_results, paste0(filepath, "Results/", tissue, "_lsea.csv"))

# Visualizations

# Significant Lipid Classes (Boxplot)
plot_enrichment(de_results, sig_lipidsets, annotation="class")</pre>
```



```
ggsave(paste0(filepath, "Plots/", tissue, "_enrichment_boxplot.png"))
```

```
## Saving 6.5 \times 4.5 in image
```

```
# Volcano plot with TGs highlighted
de_results_volcano <- de_results %>%
    mutate(colorcode = ifelse(P.Value < 0.05, ifelse(Class=="TG", "Significant Triglycerides", "All Oth

p_volcano <- ggplot(de_results_volcano, aes(x = logFC, y = -log10(P.Value))) +
    geom_point(aes(color = colorcode)) +
    scale_color_manual(values = c("Not Significant" = "grey", "All Other Significant Lipids" = "#3C6780
    theme_linedraw() +
        xlab("Log2(Fold Change)") +
        ylab("-Log10(p value)") +
        labs(color = "Legend") +
        ggtitle(tissue) +
        theme(legend.position = "bottom") +
        geom_hline(yintercept = 1.3, linetype = "dashed", color = "darkgrey")

p_volcano</pre>
```



```
Legend • All Other Significant Lipids • Not Significant • Significant Triglycerides

ggsave(paste0(filepath, "Plots/", tissue, "_TG_volcano.png"))
```

```
## Saving 6.5 x 4.5 in image
```

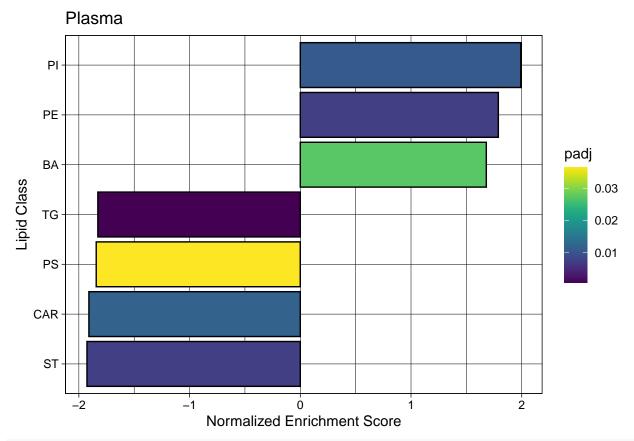
```
# Significant Lipid Classes (Barplot)
enrich_results$type <- sub("_.*", "", enrich_results$set)
enrich_results$class <- sub("^[^]*_", "", enrich_results$set)

lsea_tissue_data <- enrich_results %>%
    filter(type == "Class") %>%
    mutate(abs_NES = abs(NES))

df_sig <- lsea_tissue_data %>% filter(padj < 0.05) %>%
    arrange(desc(NES))

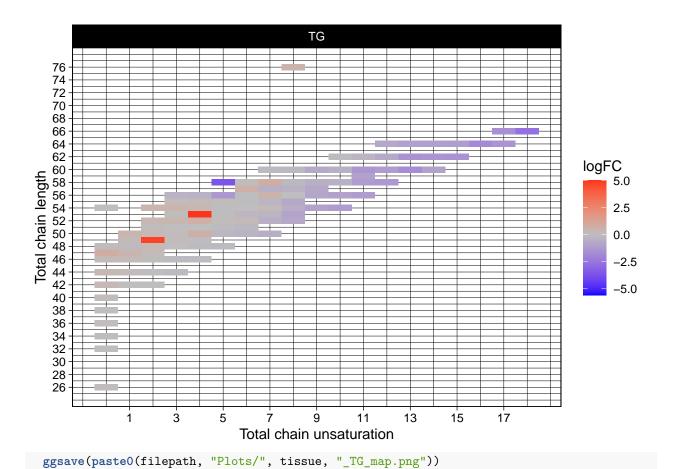
p_enrich <- ggplot(df_sig, aes(x = NES, y = reorder(class, NES), fill = padj)) +
    geom_bar(stat = "identity", color = "black") +
    scale_fill_continuous() + # Adjust the scale for continuous color
    labs(x = "Normalized Enrichment Score", y = "Lipid Class", title = tissue) +
    theme_linedraw() +
    scale_fill_viridis()</pre>
```

```
## Scale for fill is already present.
## Adding another scale for fill, which will replace the existing scale.
p_enrich
```



```
ggsave(paste0(filepath, "Plots/", tissue, "_enrichment_barplot_sig.png"))
```

Saving 6.5×4.5 in image # TG Heatmat de_results_tg <- de_results %>% filter(Class == "TG") p_grid <- ggplot(de_results_tg, aes(total_cs, total_cl, fill = logFC)) + geom_tile() +</pre> facet_wrap(~Class) + xlab("Total chain unsaturation") + ylab("Total chain length") + scale_fill_gradient2(midpoint = 0) + xlim(1, 18) + ylim(26, 76) + theme_linedraw() + scale_fill_gradient2(low = "blue", mid = "grey", high = "red") + scale_x_continuous(breaks = seq(1, 18, by = 2)) + scale_y_continuous(breaks = seq(26, 76, by = 2)) ## Scale for fill is already present. ## Adding another scale for fill, which will replace the existing scale. ## Scale for x is already present. ## Adding another scale for x, which will replace the existing scale. ## Scale for y is already present. ## Adding another scale for y, which will replace the existing scale. p_grid



Saving 6.5×4.5 in image