

CTS505 Project 1

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The solutions for the tasks listed below should be saved in separate `.R` files, e.g. `task1.R`, `task2.R`. Please include loaded libraries, loading of files, and all other commands in the files.

Task 1.

- (a) Download a list of gene IDs with expression values and false discovery rate (FDR) columns from https://links.jakobilab.org/cts505_genes.
- (b) Identify the organism used for the experiment based on the provided gene IDs for the next tasks.

Task 2.

- (a) Read the provided input file into R with an appropriate function.
- (b) Use the `biomaRt` package to retrieve gene names and Uniprot IDs for the provided gene IDs using the identified organism.
- (c) Print the top 10 highest expressed genes based on the logFC value into a new CSV file, `top10.csv`. Include the gene names and Uniprot IDs.

Task 3.

- (a) Use the `ggplot2` package to create an X-Y graph that plots `log10(FDR value)` on the Y axis and the logFC value on the X axis.
- (b) Set a title and descriptive X and Y axis labels and save the plot to PDF file named `xy_plot.pdf`.

Task 4.

- Download the list of gene counts stored in the following file
https://links.jakobilab.org/p2_counts
 - Identify the organism used for the experiment based on the provided gene IDs for the next tasks.
- (a) Establish a reasonable grouping of the provided samples based on the sample names.
- (b) Read the provided input file into **R** with an appropriate function.
- (c) Use the **biomaRt** package or any other suitable ID conversion package to retrieve gene names for gene IDs.

Task 5.

- (a) Develop a suitable design formula as discussed in the class to perform differential gene expression analysis between your chosen groups.
- (b) Use the **edgeR** workflow to build an **DGElist** object.
- (c) Using **edgeR**, perform filtering and normalization of the imported count data.
- (d) Lastly, perform differential gene expression analysis and note the number of up- and down-regulated genes.

Task 6.

- (a) Save a result table with differentially expressed genes filtered by p-value and FDR (0.05) and sorted by logFC.

Please send your R scripts & plots as attachment via email to tjakobi@arizona.edu.

In case of questions please do not hesitate to contact me.