MAD – Data Analysis & Biostatistics in R Getting to Work

James R. Hunter, Ph.D.

DIPA, EPM, UNIFESP

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Administrative Notes

- Piazza is a bust. Not going to use it
 - Questions through all the traditional means
- \bullet Chapter 9 has had a major revision. If you downloaded before 5/9
 - Need a new download

Section 1

Homework from Week One

Question 1 - R as Calculator and PEMDAS

• Evaluate this expression in R; round to 1 decimal place

$$\frac{\log_{10}(67000)*92+36}{\sqrt{759}}$$

```
res <- ((log10(67000) * 92) + 36) / sqrt(759)
res

## [1] 17.42286
round(res, 1)
```

[1] 17.4

Question 2 – Indexing a Vector

Vector of 20 random numbers created with

```
set.seed(42) # so everyone produces the same answer
x <- round(rnorm(20, mean = 100, sd = 10), 2) # returns x</pre>
```

```
## [1] 113.71 94.35 103.63 106.33 104.04 98.94 115.12
## [11] 113.05 122.87 86.11 97.21 98.67 106.36 97.16
```

- What is the 2nd element of x?
 - x[2] = 94.35
- What is the class and type of x?
 - class(x) = numeric
 - typeof(x) = double
- What is the maximum value of x?
 - \rightarrow max(x) = 122.87

Question 3 - Information about Data Frame

- Based on data frame "einstein_soro.rds"
 - ▶ Binary stored form of data about COVID-19 sorological tests performed at Albert Einstein Hospital
 - From FAPESP Data Consortium files

200 obs. of 10 variables:

• 1st Step – Load Data Frame

'data.frame':

```
soro <- readRDS("C:/Users/james/OneDrive/Documents/MAD/MAD-Infecto-2020/einstein_so
str(soro)</pre>
```

```
##
   $ pacid
               : chr "b6d668e4f818f7b3643ed593b8fb902bf9d2501e" "a090625661c06e9c
   $ dt_collect: chr "28/05/2020" "11/05/2020" "16/06/2020" "10/06/2020" ...
##
##
   $ analysis : chr "IgM, COVID19" "IgG, COVID19" "IgG, COVID19" "COVID IgM Inte
   $ result
               : chr "0.74" "0.03" "0.02" "Não reagente" ...
##
## $ unit.
          : chr "AU/ml" "AU/ml" "AU/ml" "NULL" ...
##
   $ reference : chr "<=0.90" "<=0.90" "<=0.90" "...
##
   $ sex : Factor w/ 2 levels "female", "male": 2 1 1 2 1 1 1 2 1 2 ...
## $ birth_yr : num 1989 1975 1997 2006 1983 ...
##
   $ uf
               : Factor w/ 25 levels "AC", "AL", "AM", ...: 24 9 24 24 24 24 24 24 24
               : Factor w/ 21 levels "BARUERI", "CAMPINAS", ...: 19 NA 19 19 19 19 19
##
   $ city
```

Questions about soro

- How many cases are in this data base?
 - ▶ First line of str() tells you: 200 obs. of 10 variables
 - ► Can also calculate as nrow(soro) = 200
- How many variables are in this data base?
 - First line of str()
 - Can also calculate as length(soro) = 10
- What is the class and type of soro?
 - class(soro) = data.frame
 - typeof(soro) = list
- What is the date of collection (dt_collect) of the 3rd case?
 - soro\$dt_collect[3] = 16/06/2020

soro Questions e and f

Print the 5th to the 10th birth years (birth_yr).

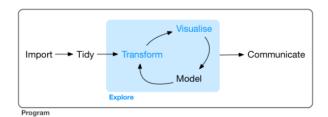
```
soro$birth_yr[5:10]
```

```
## [1] 1983 1963 1988 1971 1968 1976
```

- How many different cities are represented in this sample?
 - ▶ levels(soro\$city) = 21

Section 2

Data Analysis Projects



Project Workflow

 Tidyverse diagram shows principal steps in organizing computational tasks

However

- Much planning necessary before we even start with Import
- Right at start of project

Initial Issues in Project Planning – Biological/Medical

- What is my research topic? What is the large-scale question?
- What has science said on this topic previously?
 - Literature search
 - What sources to use (PubMed, Google Scholar, etc.)?
 - When will I have read enough?

Initial Issues in Project Planning – Technique

- What is the type of study I will conduct?
 - Experimental
 - Observational
 - Case-Control
 - Other type?
- What is my research hypothesis?
- What experiments/field work/data gathering support that?
- What will be my primary analytic technique?
 - What have others done?
 - ★ Were they effective?
 - Produce interesting results?

Data for My Project

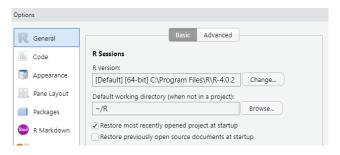
- What data do I need?
 - Categorical variables
 - Result variables
 - **★** Numeric
 - ⋆ Other Data Types
- How much data do I need?
 - Number of cases (n)
 - Statistical power to achieve significance

Section 3

Projects in RStudio

Active Directories

- When you start RStudio
 - Uses the default directory from your "Tools/Global Options" setting



• If you want a different directory for an analysis, need to use setwd() to direct R there

setwd() and getwd()

- To set a new working directory, need to use command setwd()
 - ▶ With full path name in quotes as argument
- Example for my thesis research in HIVAIDS
 - setwd(C:/Users/james/OneDrive/Documents/HIVAIDS/Drug_
- Complementary function getwd()
 - No argument
 - Shows what is current active directory
- ## [1] "C:/Users/james/OneDrive/Documents/MAD/MAD-Infecto

Why Do We Need *Projects*?

- You will work on different projects that have
 - Different databases
 - Different scripts
 - Different documents
- Throwing all your files from all projects in 1 R directory will make you crazy
 - Many projects could have upwards of 500 files
- Planning project workflow implies organization
- Build an R data analysis project around a reserved workspace

RStudio Projects Allow Group Members to Work Together Better

- Everyone has a different directory structure on their machines
 - I have projects in my R directory
 - All my HIV projects are in my HIVAIDS directory
 - All the versions of this course are in my MAD directory
- What is your directory structure?
- Projects create their own working directory (active directory)
 - Same for everyone who uses the same project structure and file
 - No more constantly having to type km-long file paths
 - ★ And getting them wrong

If the first line of any of your R scripts is

setwd("C:\Users\jenny\path\that\only\I\have")
Someone from the Tidyverse team will come to your office
and set your computer on fire

They have promised

The best way to avoid such a consequence is create a new RStudio project for each new activity

How to Create a RStudio Project

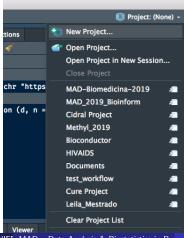
- Find "Project:(None)" text
 - ▶ Top right of screen above Environment, etc. tab



So far, no active project

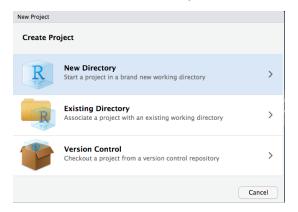
Create New Project

- Click on arrow to right of "(None)"
- Drop down menu will open
- Click "New Project ..." option



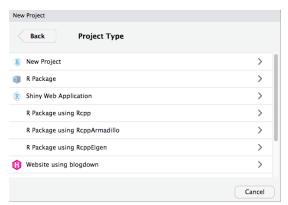
Create Project Window

- Options to choose a new or existing directory
- Version Control option more advanced
- For this case, choose "New Directory"



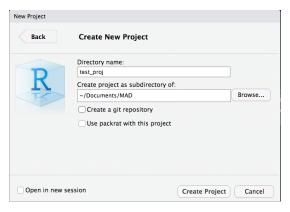
Types of New Projects

- RStudio has many types of new projects
- We want just a "New Project": click on that

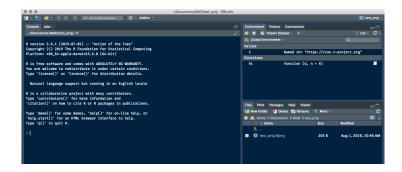


Final Creation Screen

- Give the project a name and a location
- Click Create Project button
- For now, ignore git and packrat boxes



Here Is Your Project



Subdirectories

- Projects can become large, VERY LARGE
 - Thesis project: 1,978 files, 383 sub-directories, 425 MB
- Sub-directories for files of different types.
 - Like setting up a project
 - Helps keep files organized
- My preferred subdirectory structure for a new project
 - Data
 - Raw Data
 - Docs
 - Graphics
 - Programs/Scripts
 - Slides
- Keep raw data separate from any other data files
 - Cleaned, subsets, etc.
 - ▶ You may need it again!

Section 4

We're HERE!

Problem

- Various projects and various directories could have very different paths
- Path to test_proj on my old Mac Air:
 - "/Users/jameshunter/Documents/MAD/test_proj"
- Path to test_proj on my current Windows laptop:
 - "C:/Users/james/OneDrive/Documents/MAD/test_proj"
- If I wanted to use my path to go to a graphics subdirectory that both had
 - It would be a mess!

Solution: here::here()

- here package does one thing
 - Lets you know what is full path to working directory
- Simple form: no arguments

here::here()

[1] "C:/Users/james/OneDrive/Documents/MAD/MAD-Infecto-2020"

Go to graphics Subdirectory of Current Project

- Give the function the argument "graphics"
- Thereafter, can use variable gr to refer to full path of location

```
gr <- here::here("graphics")
gr</pre>
```

- ## [1] "C:/Users/james/OneDrive/Documents/MAD/MAD-Infecto-2020/graphics"
 - Note new package::function notation
 - 2 colons call a function directly from a package without having to load package

Section 5

Scripts and Programming

Why Scripts? - A Review

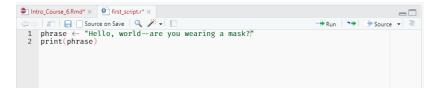
- Combine commands into coherent whole
- Store your commands
 - Reuse or recall at later date
- Facilitate correction of error and re-running program
- Scripts can be either
 - Separate documents (.r files)
 - Blocks of code in a larger .rmd document
 - ★ Like this one

Basic Scripts

- Open new script document (.r) with
 - CTRL-Shift-N
 - Plus icon in upper left of screen
- Editor is a basic text editor
- Example
 - Assign sentence: "Hello, world—wear a mask!" to the variable phrase
 - Print phrase to the screen

Program Text

```
phrase <- "Hello, world--wear a mask!"
print(phrase)</pre>
```



• Execute program with Run or Source buttons

Section 6

Loops

General

- Way to execute series of commands repeatedly
- Executes until encounters condition that ends it
- 2 Flavors
 - for loops
 - ▶ while loops

for Loops

Format

```
for(var in vector) {
  code
  code
  code
}
```

- for starts loop
- Number of repetitions of loop inside the parentheses
 - ► Repetitions:
 - ★ var variable name
 - ★ vector values variable can assume
- {} pair of curly braces
- Inside braces: lines of code you want to execute

Example for Loop

- Square whole numbers from 5 to 10
- Denote variable as i
- Vector

```
► c(5, 6, 7, 8, 9, 10)
```

```
▶ 5:10
```

```
for(i in 5:10){
    print(i^2)
}

## [1] 25
## [1] 36
## [1] 49
## [1] 64
## [1] 81
## [1] 100
```

What the Loop Did

- 1st Iteration
 - ▶ Assigned 5 to i
 - ▶ Squared i (i^2)
 - Wrote i to screen (25)
- 2nd Iteration
 - Assigned 6 to i
 - Squared i
 - Wrote i to screen (36)
- Same for 3rd to 6th
- At end of 6th iteration (i <- 10)
 - Could not find a new value to give to i
 - Stopped

More Realistic Example

- Take a DNA string and transform it to a vector of letters
- Length of the string unknown
- Can be calculated with nchar() function: gives number of characters

```
library(tidyverse)
seq <- "CCTCAAATCACTCTTTGGCAACGACCCTTAGTCACAATAAAAGTAGGGGA"
seq_length <- nchar(seq)
seq_vector <- character(seq_length) # create vector to hold result
for (i in 1:nchar(seq)) {
   seq_vector[i] <- tolower(str_sub(seq, i, i))
}
seq_vector</pre>
```

while Loops

- Similar structure to for loops but work differently
- Keeps repeating until condition you put in while statement turns FALSE
- Consult text
- Used much less than for

Loops vs. Vectorized Statements

- R is vectorized
 - Interpreter will act on all elements of a vector at once
 - Don't need to loop item by item
- Should we use loops at all?
- Computer speed today reduces the speed advantage of vectorized statements
 - Except for VERY large datasets, loops are practically equivalent
 - Millions of cases, hundreds of variables
- Don't give up on loops
 - ▶ Logic is usually easier to program than many vectorized functions
 - Can save much overall time in terms of programming

Section 7

Conditional Statements (if. . . then. . . else)

Thought Problem

- We are reading sequences of nucleotides
 - DNA alphabet: ACGT
 - RNA alphabet: ACGU
- We could say
 - If the sequence is DNA then the alphabet is ACGT.
- More complete version
 - ▶ If the sequence is DNA then the alphabet is ACGT else the alphabet is ACGU.

If Statements in R

Basic structure

```
if(condition) {
  code
  code
  code
} else {
     code
     code
     code
     code
}
```

Simple Example: No else Clause

```
x <- 0
if (x == 0) {
  print("x equals 0")
}
## [1] "x equals 0"</pre>
```

More Complete Logical Test

```
x <- 0

if (x == 6) {
   print("x equals 6")
} else {
    print("x does not equal 6")
}</pre>
```

[1] "x does not equal 6"

3 Nested Conditions – DNA/RNA Example

```
seq type <- "DNA"
if (seq_type == "DNA") {
  print("ACGT")
} else {
    if (seq type == "RNA"){
      print("ACGU")
    } else {
      print("sequence neither DNA nor RNA")
```

[1] "ACGT"

ifelse() Function

- For simple logical tests
- ifelse(test, true, false)
- 3 Arguments
 - test: logical test
 - true: result if TRUE
 - false: result if FALSE
- If result of test is TRUE, returns true value
- If result of test is FALSE, returns false value

ifelse() Example

```
X <- 2
res <- ifelse(X > 10, "greater than", "less than or equal
paste("X is", res, "10")
```

[1] "X is less than or equal to 10"

More Complicated if...then...else Logic

- Function in dplyr package: case_when()
- Can more comfortably handle large number of alternative cases
- More advanced

Section 8

Importing Files to R

Types We Will Work With

- Comma-Delimited Files (.csv)
- Excel Files (.xls or .xlsx)
- FASTA files

Section 9

Comma-Delimited Files (.csv)

Importing a .csv File

- A plain text file that is in rectangular form
- Commas separating the fields
- Interpretable by Excel
- Most common file type you will import
- Use readr::read_csv() to import it
 - Function reads file
 - Treats commas as Excel does: separators between columns
 - Imports it to a tibble

```
1 df ← readr::read_csv(file, col_names = TRUE, col_types = NULL, na = c("", "NA"), skip = 0)
```

read_csv() Arguments

- file = file path to reach the file
 - ▶ Use of here::here()
- col_names = Is the first row the names of the columns or data?
 - ► TRUE means 1st row are the column names; read_csv() assumes so
 - FALSE means it has data rather than names

col_types = Argument

- read_csv() will try to guess the correct data type based on the column content
- If you want to let R guess, enter NULL or omit the argument
- If you want to specify column types, use a string of characters from list:
 - ▶ c = character
 - ▶ i = integer
 - ightharpoonup n = number
 - ▶ d = double
 - ▶ I = logical
 - ▶ f = factor
 - ▶ D = date
 - ▶ T = date time
 - ▶ t = time
 - ▶ ? = guess
 - to skip the column

Notes on col_types =

- Column string must have exactly the same number of characters as the data has columns
 - If you have a data set with five columns, the string must have five characters
 - ★ Ex. col_types = "cfn-c"
 - ★ Means character-factor-numeric-skip-character

Two Other Key Arguments

- Specifying missing data
 - ► read_csv() assumes that missing data will be encoded with
 - ★ the string "NA" or a blank string
 - If data uses other codes, need to specify them
 - ★ "99" is a common indicator of missing data in social science
 - ★ Must specify the na = argument as na = c("", "NA", "99")
- Skip rows: skip =
 - Datasets with metadata, etc. in 1st few rows
 - ► To skip these, set a positive value for the skip = argument
 - ★ Ex. skip = 4 will start reading the database on the fifth row

Simple .csv Example

- classe_simples.csv records info on a small school class
 - ▶ Name, home state, age and scores on a standardized test (1 10)

	Α	В	C	D
1	nome	estado	idade	nota
2	Pedro	DF	34	4.12
3	Joana	PE	59	6.1
4	Isabel	SP	26	9.34
5	Pedro	GO	67	6.75
6	Bia	RJ	53	5.85
7	Pedro	RJ	51	7.49
8	Tomas	TO	46	5.78
9	Isabel	PE	66	4.2
10	Tomas	GO	21	8.78
11	Ricardo	TO	51	8.58
12				

Importing classe_simples.csv

- Assign the function to a name for the data frame classe1
- Run a simplified command with no column spec
 - ► R will guess

```
classe1 <- read_csv(here::here("classe_simples.csv"))
classe1</pre>
```

Result

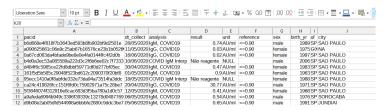
```
## Parsed with column specification:
## cols(
##
     nome = col character(),
##
     estado = col_character(),
##
     idade = col double(),
##
     nota = col double()
## )
## # A tibble: 10 x 4
##
              estado idade nota
      nome
##
      <chr> <chr>
                     <dbl> <dbl>
              DF
##
    1 Pedro
                        34
                            4.12
##
    2 Joana
             PΕ
                        59 6.1
##
    3 Isabel
             SP
                        26 9.34
    4 Pedro
              GO
                        67
                            6.75
##
##
    5 Bia
              RJ
                        53 5.85
    6 Pedro
              RJ
                        51 7.49
##
    7 Tomas
              TO
                        46 5.78
##
##
   8 Isabel
             PE
                        66 4.2
    9 Tomas
              GO
##
                        21 8.78
## 10 Ricardo TO
                        51
                            8.58
```

classe_simples.csv Results

- First line contained variable names; R used them
- If we had wanted to specify columns, we could have used
 - col_types = "ccnn"

More Complex Data Set

- Example we will use throughout data cleaning process
- 99 case version of Einstein Hospital COVID-19 sorological tests
 - einstein_soro_tests.csv
- First look at "messy", ie. real data



Constructing the Import Command

- Variable names in first line
 - col_names = can be omitted
- No need to use skip =
- NA argument
 - result and unit variables use "Não reagente" and "NULL"
 - birth_yr has 3 NA values
 - Unclear if they mean 0 or could not get a meaningful result
 - ► Create argument: na = c("Não reagente", "NULL", "NA")

col_types = Argument

- pacid: awkward, but obviously a string
- dt_collect: Date, but in non-standard format
 - R will parse as character type
 - We can reformat it in tidying phase
- analysis: character string
- result: should be numeric, but has text entries
 - R will parse as character type
- unit & reference fields: character strings
 - reference could be numeric, but leave to tidying
- sex: character, but better as factor: only 2 values
- birth yr: numeric
- uf & city: character
- Using codes, full argument: col_types = "cccccfncc"

Ready to Import

```
einstein soro <- read csv(here::here("einstein soro tests.csv").
                                                                                                                                      col types = "cccccfncc",
                                                                                                                                      na = c("Não reagente", "NULL", "NA"))
glimpse(einstein_soro)
## Rows: 99
## Columns: 10
## $ pacid
                                                                                  <chr> "b6d668e4f818f7b3643ed593b8fb902bf9d2501e", "a090625661c...
## $ dt collect <chr> "28/05/2020", "11/05/2020", "16/06/2020", "10/06/2020", ...
## $ analysis
                                                                                  <chr> "IgM, COVID19", "IgG, COVID19", "IgG, COVID19", "COVID I...
## $ result
                                                                                  <chr> "0.74", "0.03", "0.02", "Não reagente", "0.47", "0.9", "...
                                                                                  <chr> "AU/ml", "AU/ml", "AU/ml", NA, "AU/ml", "AU/ml", NA, "AU...
## $ unit
## $ reference
                                                                                 <chr> "<=0.90", "<=0.90", "<=0.90", "", "<=0.90", "<=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "=0.90", "
## $ sex
                                                                                  <fct> male, female, female, female, female, female, female, male...
## $ birth_yr
                                                                                  <dbl> 1989, 1975, 1997, 2006, 1983, 1963, 1988, 1971, 1968, 19...
## $ uf
                                                                                  <chr> "SP", "GO", "SP", "
## $ city
                                                                                  <chr> "SAO PAULO", NA, "SAO PAULO", "SAO PAULO", "SAO PAULO", ...
```

Section 10

Working with Excel Files

Advice on Working within Excel - 1

- If you are preparing .csv files,
 - Save different types of data in different files
- If you are working with .xlsx files,
 - Save different types of data in different sheets (tabs)
- Data should be placed in a block of data
 - Without any blank lines
- First line should have variable names, if you are using them
- Each column should contain only one class of data
 - Numeric, character, logical, . . .

Advice on Working within Excel - 2

- Zeros are always "0"
 - ▶ Never "-", " " (blank space) or other character
- Missing data should always be "NA"
 - ▶ Never "0", "99" or other text
- Start making the data conform to the tidy data rules here
 - Each column should be a variable
 - Each line should be a case
- Never use colors or design elements

Importing Excel Files

- Do not have to save files as ".csv" first
- readxl package
 - read_excel()
 - Works with both .xls and .xlsx formats
- Differs from read_csv() primarily in handling of column types
- Also has arguments for
 - Specifying tabs within workbook
 - Specifying specific region on a given sheet

```
df ← read_excel(path, sheet = NULL, range = NULL, col_names = TRUE, col_types = NULL, na = "", skip = 0)
```

Arguments for read_excel()

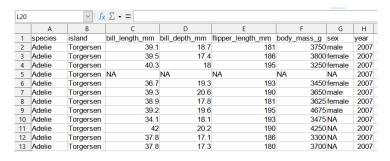
- Specifying what to download
 - Path for file
 - Sheet name
 - range in normal Excel format: "A1:B25"
- col_names = functions same as read_csv()
- na = and skip = function the same

Column Types in read_excel()

- Uses whole words instead of single letters
- If you want to make a factor, need to do so in tidying process
- Column types:
 - date
 - guess (trust R to make a good choice)
 - list
 - logical
 - numeric
 - skip
 - text

Spreadsheet Example

- R standard data set: Palmer Penguins
 - 8 characteristics of 344 penguins
 - ▶ penguins.xlsx



Example Arguments

- Column types
 - Argument is unnecessary as R will parse values correctly
 - ▶ If we want to specify it
 - * col_types = c("text", "text", "numeric", "numeric",
 "numeric", "numeric", "text", "numeric)
 - ▶ NA needed for measurement columns since missings exist
 - ★ na = "NA"

Import File

```
penguin <- readxl::read_excel(here::here("penguins.xlsx"), na = "NA")</pre>
glimpse(penguin)
## Rows: 344
## Columns: 8
## $ species
                       <chr> "Adelie", "Adelie", "Adelie", "Adelie", "Adelie",...
## $ island
                       <chr> "Torgersen", "Torgersen", "Torgersen", "Torgersen...
## $ bill_length_mm
                       <dbl> 39.1, 39.5, 40.3, NA, 36.7, 39.3, 38.9, 39.2, 34....
## $ bill depth mm
                       <dbl> 18.7, 17.4, 18.0, NA, 19.3, 20.6, 17.8, 19.6, 18....
## $ flipper_length_mm <dbl> 181, 186, 195, NA, 193, 190, 181, 195, 193, 190, ...
## $ body_mass_g
                       <dbl> 3750, 3800, 3250, NA, 3450, 3650, 3625, 4675, 347...
## $ sex
                       <chr> "male", "female", "female", NA, "female", "male",...
## $ vear
                       <dbl> 2007. 2007. 2007. 2007. 2007. 2007. 2007. 2007. 2007. 2...
```

Section 11

FASTA Files

Read a FASTA File

- Most common storage format for nucleotide (DNA or RNA) and amino acid sequences
- Text-based representation using single-letter codes (alphabets)
- Each sequence can be preceded by a line of sequence name and comments
- Like a .csv file: pure text (uses text editor)
- To prepare sequences for analysis
 - Use specialized programs for reading and parsing
- New package (bioseq) provides easy and complete functions
 - Not in original set of downloaded packages
 - Install on your machine with install.packages("bioseq") in Console
- bioseq::read fasta() takes 2 arguments
 - file name and path
 - Type of sequence: "DNA" (default), "RNA" or "AA"

FASTA File Example

- DNA sequences of the gag polyprotein of the HIV-1 virus
 - ▶ Reference genome HXB-2
- Sequence stored in file HIVHXB.fa
- Sequence is 1,503 base pairs long.



Import the File

```
seq <- bioseq::read_fasta(here::here("HIVHXB.fa"), "DNA")
stringr::str_sub(seq, 1, 60)</pre>
```

- # [1] "ATGGGTGCGAGAGCGTCAGTATTAAGCGGGGGAGAATTAGATCGATGGGAAAAAATTCGG"
 - Result a character string vector
 - First 60 nucleotides using the subsetting function of stringr package.

Section 12

Go to Presentation Class_2b