Korean J Parasitol Vol. 55, No. 5: 473-480, October 2017 https://doi.org/10.3347/kjp.2017.55.5.473

ORIGINAL ARTICLE

Genetic Diversity and Natural Selection in 42 kDa Region of *Plasmodium vivax* Merozoite Surface Protein-1 from China-Myanmar Endemic Border

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Abstract: *Plasmodium vivax* merozoite surface protein-1 (PvMSP1) gene codes for a major malaria vaccine candidate antigen. However, its polymorphic nature represents an obstacle to the design of a protective vaccine. In this study, we analyzed the genetic polymorphism and natural selection of the C-terminal 42 kDa fragment within PvMSP1 gene (Pv MSP1₄₂) from 77 *P. vivax* isolates, collected from imported cases of China-Myanmar border (CMB) areas in Yunnan province and the inland cases from Anhui, Yunnan, and Zhejiang province in China during 2009-2012. Totally, 41 haplotypes were identified and 30 of them were new haplotypes. The differences between the rates of non-synonymous and synonymous mutations suggest that PvMSP1₄₂ has evolved under natural selection, and a high selective pressure preferentially acted on regions identified of PvMSP1₃₃. Our results also demonstrated that PvMSP1₄₂ of *P. vivax* isolates collected on China-Myanmar border areas display higher genetic polymorphisms than those collected from inland of China. Such results have significant implications for understanding the dynamic of the *P. vivax* population and may be useful information towards China malaria elimination campaign strategies.

Key words: Plasmodium vivax, merozoite surface protein-1, genetic polymorphism, natural selection, Myanmar, China

INTRODUCTION

Malaria is a major infectious disease in the Greater Mekong Subregion (GMS) in Asia. Although there has been a considerable decrease in the incidence of malaria in China [1], Yunnan Province still has the highest transmission area of vivax malaria in China, particularly in the southern border areas adjacent to Myanmar. *Plasmodium vivax* is also the most widely distributed species of all 5 human malaria parasites in Southeast Asia and accounts for 65% of malaria cases in Asia and South America [2]. More attention is being focused on malaria today than any time since the world's last efforts to achieve eradica-

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tion over 40 years ago. The global community is now discussing strategies aimed at dramatically reducing malarial disease burden and the eventual eradication of all types of malaria everywhere. As a consequence, *P. vivax*, which has long been neglected and mistakenly considered benign, is now entering into the strategic debates taking place on malaria epidemiology and control, drug resistance, pathogenesis, and vaccines. Thus, contrary to the past, the malaria research community is becoming more aware and concerned about the widespread spectrum of illness and death caused by up to a couple of hundred million cases of vivax malaria each year [3].

Taking account of the facts above, availability of *P. vivax* malaria vaccine is highly desirable. Advanced studies on genetic diversity of the most variable domain of vaccine candidate *P. vivax* merozoite surface proteins (PvMSPs) in field isolates of different countries have been carried on and demonstrated that the diversity of MSPs in *P. vivax* is presumed be associated to parasite immune evasion and be important for the rationale

Received 19 March 2017, revised 16 September 2017, accepted 18 September 2017.
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of malaria vaccine designs [4,5]. Since the 42 kDa fragment of Plasmodium merozoite surface protein-1 (PvMSP1) contains known B- and T-cell cell epitopes, a PvMSP142 vaccine antigen may be capable of conferring protection mediated by providing antigen-specific T-cell help for B-cells and antibody production [6]. Several previous studies have reported the presence of acquired antibodies against the C-terminus part of the protein called PvMSP1₁₉ or PvMSP1₄₂ antigens among individuals during natural P. vivax infections [7,8]. Immunological studies performed on animal models have also proved that the PvMSP1₁₉ or PvMSP1₄₂ is one of the promising vaccine candidates against asexual stages of the malaria [9]. Although genetic polymorphisms in the central repeat region of PvMSP1 has been investigated among other countries in Southeast Asia on P. vivax isolates [10], the data is not available for the C-terminus region of this antigen from southern border areas adjacent to Myanmar and the inland cases in China.

The present study aimed to identify the genetic diversity and haplotypes of the gene fragment coding PvMSP1₄₂ in *P. vivax* isolates of malaria cases in China-Myanmar border (CMB) areas and Yunnan, Zhejiang, and Anhui province of inland of China. Moreover, the natural selection of the gene fragment coding PvMSP1₄₂ was tested in 4 *P. vivax* populations from CMB areas and inland of China.

MATERIALS AND METHODS

Ethics statement

This study was conducted according to the principles expressed in the Declaration of Helsinki. Blood collections were made with full informed consent of the patients and following institutional ethical guidelines that were reviewed and approved by the ethics committee at National Institute of Parasitic Diseases, Chinese Center for Disease Control and Prevention.

Blood samples, DNA extraction, and purification

Blood samples were obtained from 77 symptomatic and microscopically confirmed *P. vivax* malaria patients in China during 2009 to 2012. These examined samples included 59 imported cases of CMB areas based on their traveling history, including 14, 19, 17, and 9 cases each year from 2009 to 2012, respectively, and additional 18 patients from inland China. Among the 18 samples from inland China, 6 of them were from Anhui province collected in 2009, 6 from Zhejiang prov-

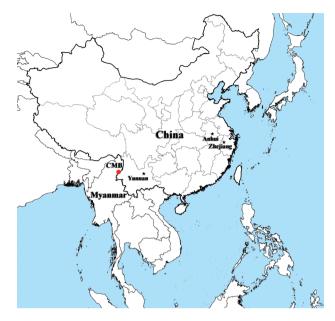


Fig. 1. The map of *P. vivax* samples collection. Sample collection areas in this study are indicated in black pentagrams (Anhui, Yunnan, and Zhejiang provinces from inland China) and red dot (China-Myanmar border area, CMB).

ince collected in 2009 and 6 from Yunnan province collected in 2010 (Fig. 1). Annual parasite incidence (API) for Anhui, Zhejiang and Yunnan province was 0.0210, 0.0352, and 0.2186 per 10,000 person-years in 2015 [11]. Because the transmission of malaria had been controlled in a very low level in local China, only limited sporadic cases of malaria inland were collected here. These samples collected from different villages in Yunnan province treated as the inland malaria cases from these febrile patients haven't been abroad within 1 month. All the patients' *P. vivax* infection were diagnosed by microscopic examination of thin and thick blood smears and further confirmed by nested PCR as described previously [12].

Genomic DNA was isolated from 200 µl of venous blood which collected in a sterile heparinized tube from the patients who were found positive for *P. vivax*. Approximately 100 µl of blood each patient was used and added 100 µl PBS to get the final volume of 200 µl. Then, DNA was extracted from the whole blood by using the QIAamp DNA mini kit (QIAGEN, Shanghai, China), according to the manufacturer's instructions. The purified DNA was dissolved in 150 µl TE buffer (10 mM Tris-HCl, 1 mM EDTA; pH 8.0) and stored at -20°C until use.

PCR amplification and analysis of *P. vivax* field isolates The *P. vivax* fragment (comprising PvMSP1₄₂ kDa amino ac-

ids) of PvMSP142 was amplified by polymerase chain reaction (PCR). In this study, the specific primer was designed according to the P. vivax MSP1 complete gene sequence of PlasmoDB (PVX_099980) [13]. The primers are: Pv1SF (5'-AGAAG AAAAC GTAGC AGCAA-3') and Pv1SR (5'-AAGCC CAGTT CAGTT CAGAA CTCA-3'). PCR reaction volumes were 50 µl. The cycling parameters for PCR amplification was performed under the following conditions: initial denaturation 5 min at 94°C, 35 cycles of denaturation at 94°C for 30 sec, annealing at 55°C for 30 sec, and extension at 72°C for 1 min 30 sec, followed by a final extension at 72°C for 5 min. PCR mixture reagents contained 1 µl of DNA, 0.5 units of ExTag or LATag DNA polymerase (Takara, Shiga, Japan), 0.2 mM of each primer, 0.1 mM dNTPs in a 25 µl of reaction mix with 1.5 mM MgCl₂. The PCR products were examined by electrophoresis in a 1% agarose gel, visualized with an ultraviolet trans-illuminator and purified with PCR purification kits (Qiagen). Then, the purified PCR products were sequenced using the forward primers on an ABI PRISM 3700 DNA capillary sequencer, by BGI Company (Shenzhen, Guangdong, China). All unique mutations were carefully checked, and ambiguous bases were confirmed by resequencing. We also carried out a BLAST search on PlasmoDB Genebank Database of P. vivax to compare these successfully sequenced isolates with those previously identified from China and the Asia-pacific subregion. Accurate alignment of the sequences was performed by in ClustalX version 2.0 [14]. The new sequences were deposited in Gen-Bank with accession nos. JX490129-JX490156, JX993754, and JX993755.

Nucleotide diversity and natural selection test of *P. vivax* field isolates

Nucleotide and amino acid sequences were aligned using Clustal W in MEGA 5.0 [15]. First, nucleotide diversity (π) was computed in 100 bp sliding window and 25 bp step size using DnaSP v.5.0 [16]. Then, to detect natural selection acting on these coding sequences, the rates of non-synonymous (dN) to synonymous (dS) substitution (dN/dS) was calculated with DnaSP v.5.0. If the amino acid change is deleterious, purifying selection, then dN/dS<1; only when the amino acid change offers a selective advantage, the dN/dS is>1. In addition, Tajima's D was used to test neutrality of this gene fragment in DnaSP v.5.0 [16,17]. A remarkable negative value of Tajima's D reveals an excess of rare variants as expected under positive and negative selection or population size expansion. Whereas,

a significant positive value demonstrates an excess of high-frequency variant as expected under balancing selection or under population structure. Finally, to describe the genetic similarities among PvMSP1₄₂ haplotypes, we constructed networks by the median joining method from 41 unique haplotypes on the basis of PvMSP1₄₂ sequences in Network 4.5 [18].

Phylogenetic analysis of PvMSP1 from *P. vivax* field isolates

The phylogenetic relationships were derived from the PvM-SP1₄₂ sequences. In case of individuals that carried an identical sequence (it is possible that individuals from different locations shared the same sequence), only 1 sequence was included for the tree reconstruction. The reference sequences were chosen from GenBank. Then, partitioned Neighbor-Joining method was performed in MEGA 5.0 [15,19] to construct the phylogenetic tree (with Kimura-2 parameter distance, branch support with 1000 bootstrap replicates, and complete deletion of gaps). MSP1₄₂ fragment gene of *P. cynomolgi* strain Berok was set as the out group.

RESULTS

Haplotype variations in *P. vivax* of different field isolates

We successfully amplified and sequenced the gene encoding PvMSP1₄₂ fragment (1,209 bp, corresponding to amino acid positions 1350-1752 in PvMSP1) shown in Supplementary Fig. S1. Of 77 isolates from 4 geographic locations, 41 haplotypes were detected on the entire PvMSP1₄₂ fragment (Table 1). From these 41 haplotypes, 30 were new haplotypes. Only 1 single haplotype was detected among 6 isolates from Anhui, all these samples were collected from Bengbu city and 2 haplotypes from 6 isolates in Zhejiang province. In contrast, other 6 isolates from Yunnan province distributed in different villages are detected in 6 different haplotypes and 35 haplotypes are detected from 59 imported cases of CMB areas.

Only 3 haplotypes were detected for PvMSP1₁₉ fragment in comparison with 38 haplotypes for PvMSP1₃₃ fragment in the all sequenced isolates. The consistent pattern was observed in the *P. vivax* populations from CMB areas and inland China (Table 1).

Nucleotide diversity and natural selection of different PvMSP1₄₂ fragment from different *P. vivax* isolates

The overall nucleotide diversity (π) of PvMSP1₄₂ for all of 77

Table 1. Halpotype diversity, nucleotide diversity, and natural selection of Plasmodium vivax MSP142

Fragment	Hª	π^{b}	dN°	dS ^d	dN/dS	Tajima's D	P-value
42 kDa							
All samples (n = 77)	41	0.01901	0.02188	0.01016	2.15354	2.44824	< 0.05
Border areas ^f (n = 59)	35	0.01836	0.02085	0.01082	1.92699	2.01030	>0.05
Inland China (n = 18)	8	0.01317	0.01615	-0.00328	4.92378	0.48590	>0.05
Anhui (n=6)	1	n.a ^e	n.a	n.a	n.a	n.a	n.a
Yunnan (n=6)	6	0.01803	0.02234	0.00366	6.10383	0.23699	>0.05
Zhejiang (n=6)	2	0.01235	0.01546	0.00209	7.39713	1.37681	>0.05
33 kDa							
All samples (n = 77)	38	0.02617	0.03026	0.01401	2.15989	2.57163	< 0.05
Border areas (n = 59)	33	0.02523	0.02879	0.01489	1.93351	2.11738	< 0.05
Inland China (n = 18)	8	0.01818	0.02237	0.00456	4.90570	0.54855	>0.05
Anhui (n=6)	1	n.a	n.a	n.a	n.a	n.a	n.a
Yunnan (n=6)	6	0.02467	0.03067	0.00509	6.02554	0.27447	>0.05
Zhejiang (n=6)	2	0.01716	0.02159	0.00290	7.44483	1.37681	>0.05
19 kDa							
All samples (n=77)	3	0.00063	0.00071	0.00037	1.91892	-0.76528	>0.05
Border areas (n = 59)	3	0.00073	0.00081	0.00048	1.68750	-0.73272	>0.05
Inland China (n=18)	2	0.00033	0.00042	n.a	n.a	-1.16467	>0.05
Anhui (n=6)	1	n.a	n.a	n.a	n.a	n.a	n.a
Yunnan (n=6)	2	0.00098	0.00126	n.a	n.a	-0.93302	>0.05
Zhejiang (n=6)	1	n.a	n.a	n.a	n.a	n.a	n.a

^aH: the number of haplotypes.

0.15 — Myanmar — Zhejiang — Yunnan 0.05 — Yunnan 0.05 — Myanmar — Zhejiang — Yunnan 0.05 — Myanmar — Yunnan 0.05 — Yunnan 0.05 — Myanmar — Yunnan 0.05 — Myanmar — Yunnan 0.05 — Yunnan 0.05 — Myanmar — Yunnan 0.05 — Myanmar — Yunnan 0.05 — Myanmar — Yunnan 0.05 — Yunnan 0.05 — Myanmar — Myanmar — Yunnan 0.05 — Myanmar — Myanmar — Myanmar — Myanmar — Yunnan 0.05 — Myanmar — Myanmar

Fig. 2. Nucleotide diversity per site (π) at PvMSP1₄₂ from *P. vivax* isolates collected along the China-Myanmar border areas, local regions of Yunnan and Zhejiang provinces in inland China.

isolates was 0.01901, and π values were 0.01803 and 0.01235 for the isolates from Yunnan and Zhejiang province of inland China, respectively, as well as π value was 0.01836 for the isolates from the CMB areas, with the peak on nucleotide positions from 476 to 525 bp (Fig. 2), which located at the C-terminal 33 kDa fragment within PvMSP1 gene (PvMSP1₃₃) (Supplementary Figs. S2, S3).

The rates of non-synonymous (dN) to synonymous (dS) substitution (dN/dS) of PvMSP1₄₂ for all of 77 isolates was

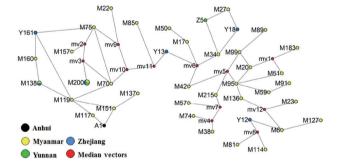


Fig. 3. The network of PvMSP142 from *P. vivax* isolates collected along the China-Myanmar border areas, local regions of Anhui, Yunnan, and Zhejiang provinces in China.

2.15354, and the rates of dN/dS were 6.10383, 7.39713, and 1.92699 for the isolates from Yunnan, Zhejiang province of inland China and CMB areas, respectively, suggesting a positive selection for PvMSP1₄₂ of *P. vivax* populations from inland China and CMB areas. The overall Tajima's D value of PvMSP1₄₂ was 2.44824 (P<0.05) for all of 77 isolates, and the Tajima's D values were 0.23699, 1.37681, and 2.01030 for the isolates from the Yunnan, Zhejiang province of inland China and CMB areas, respectively, which also indicated balancing selection for PvMSP1₄₂ of *P. vivax* populations from CMB areas.

 $^{{}^{\}mathrm{b}}\pi$: nucleotide diversity.

^cdN: the rates of nonsynonymous substitutions.

ddS: the rates of synonymous substitutions.

en.a.: not applicable.

^fBorder areas: China-Myanmar border areas.

The network of 41 haplotypes from 77 isolates of PvMSP1₄₂ showed that most prevalent haplotypes originated from Myanmar followed by Yunnan and Zhejiang provinces of China. Moreover, the haplotypes in the studied *P. vivax* populations were highly diverse (35/59) in the Myanmar population, and in extremely case, all isolates from Yunnan showed independent haplotypes. In contrast, Zhejiang and Anhui population in inland China showed the low haplotype diversity. The haplotypes were consistent with the distribution of allele frequencies as shown (Table 1; Fig. 3).

Nucleotide diversity and natural selection of different PvMSP1₃₃ and PvMSP1₁₉ fragments from different *P. vivax* isolates

The overall nucleotide diversity (π) of PvMSP1₃₃ for all of 77 isolates was 0.02617, and π values were 0.02467 and 0.01716 for the isolates from Yunnan and Zhejiang province of inland China, respectively, as well as π value was 0.02523 for the isolates from the CMB areas (Table 1; Fig. 2). The overall nucleotide diversity (π) of PvMSP1₁₉ for all of 77 isolates was 0.00063, and π values were 0.00098 and 0.00073 for the isolates from Yunnan province of inland China and the CMB areas, respectively (Table 1; Fig. 2).

The rates of dN/dS for PvMSP1₃₃ were 6.02554, 7.44483, and 1.93351 for the isolates from Yunnan, Zhejiang province of inland China and CMB areas, respectively, suggesting the positive selection located at PvMSP1₃₃ fragment of *P. vivax* populations from inland China and CMB areas. Furthermore, the rate of dN/dS for PvMSP1₁₉ was 1.68750 for the isolates from CMB areas.

Moreover, the overall Tajima's D value of PvMSP1₃₃ was 2.57163 (P < 0.05) for all of 77 isolates, and the Tajima's D values were 0.27447, 1.37681, and 2.11738 for the isolates from Yunnan, Zhejiang province of inland China and CMB areas, respectively, also suggesting a positive balancing selection of PvMSP1₃₃ in the CMB population (Tajima's D = 2.11738, P < 0.05) (Table 1). However, Tajima's D values were negative for PvMSP1₁₉ in the CMB areas (-0.73272) and Yunnan population (-0.93302), indicating the purifying selection (Table 1).

Phylogenetic diversity of PvMSP1₄₂ from *P. vivax* of different field isolates

The phylogenic tree showed that all these isolates of *P. vivax* clinical patients from different regions such as Myanmar, Thailand, Singapore, Bangladesh, South Korea, India, Vietnam, and

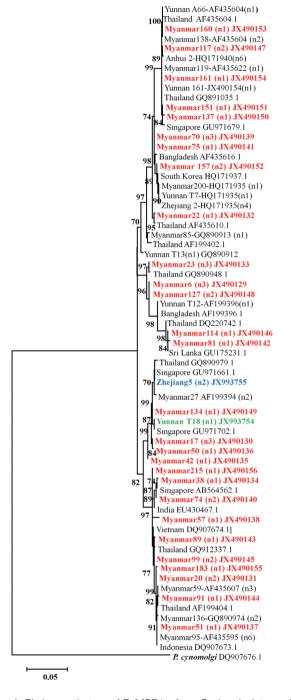


Fig. 4. Phylogenetic tree of PvMSP1₄₂ from *P. vivax* isolates collected along the China-Myanmar border areas, local regions of Anhui, Yunnan, and Zhejiang provinces in inland China. Novel sequences identified in this study are indicated in red, blue, and green. Scale bar indicates nucleotide substitutions per site.

Indonesia having high prevalence (Fig. 4). What's more, the *P. vivax* clinical samples showed distinct differences haplotypes among the isolates collected from different province in China

or different villages in Yunnan province. The phylogenetic analysis revealed that newly identified haplotypes from China were clustered differently. The gene coding PvMSP1₄₂ from field isolates collected in Anhui province has only 1 haplotype and close to these isolates from Myanmar. Two different haplotypes of PvMSP1₄₂ were detected from Zhejiang isolates. One sequence coding PvMSP1₄₂ from Zhejiang province is close to these from Yunnan province and Myanmar; while another one also detected in Zhejiang isolates has been demonstrated to be close to isolates from Singapore and Thailand. All these genes coding PvMSP1₄₂ from Yunnan are from those patients who haven't been abroad within 1 month.

DISCUSSION

The *P. vivax* parasite exhibits higher genetic diversity than *P. falciparum*, especially for the gene families associated with merozoite invasion or immune response modulation (e.g., the *msp3*, *vir*, and *msp7* gene families) [20-22]. The high genetic diversity and natural selection of *P. vivax* vaccine targets is common existed in isolates world-wide [23,24]. The PvMSP1 locus codes for a major asexual blood-stage antigen currently proposed as a malaria vaccine candidate antigen. Reports of extensive polymorphism of this protein from field isolates and clones from different geographical areas remain a major challenge. Numerous studies on the genetic diversity of PvMSP1 in *P. vivax* field isolates have been carried out in many different geographic areas [25,26]. However, there is no available data for PvMSP1₄₂ from southern border areas adjacent to Myanmar and the inland cases in China.

In this study, we present several sets of genetic information for PvMSP1₄₂ of populations from inland China and CMB areas at first time. We found 35 and 8 haplotypes of PvMSP1₄₂ for the isolates from Myanmar and China during 2009-2012, respectively. We also documented varied types of haplotypes characteristic of high genetic diversity in the studied region compared to other endemic regions. This high genetic diversity of PvMSP1₄₂ fragments were consistent with that of *P. vivax* field isolates collected in Cambodia and Thailand [27].

Of the 41 haplotypes, 30 were new haplotypes including 28 of them from Myanmar, characterizing of multiple clonality. The same single haplotype was documented in each of the inland isolates from Anhui compared to those of Myanmar, 2 different haplotypes from isolates from Zhejiang and diverse multiple haplotypes found in Yunnan similar to Myanmar.

This finding indicated that geographical proximity between Myanmar and Yunnan China border which showed that vector dynamic and/or human motility might have been important contributing factors in malaria parasite transmission and degree of endemicity [28]. In recent years, malaria transmission has been controlled in a very low level in Anhui province, China and these cases collected here from Bengbu city were localized sporadic malaria cases [29]. These results are consistent with previous studies that genetic diversity of the malaria parasites has been shown to be associated with the levels of endemicity and transmission intensity.

Genetic diversity analysis revealed that the majority of polymorphic sites were in the 33 kDa portion and significant proportion of the identified polymorphisms occurred probably as result of reported positive selection pressure on this region while 19 kDa regions remained highly conserved. The similar results for the positive selection of PvMSP1₃₃ were reported in the *P. vivax* isolates from India and Sri Lanka several years ago [30,31]. The frequent occurrences of non-synonymous substitutions relative to synonymous ones and high value of Tajima's D indicate the polymorphism of antigen enable parasites to avoid host immune pressure and host immune responses likely play a role in maintaining the polymorphism of *P. vivax* MSP1 alleles.

The halpotype network demonstrated that parasite populations are highly heterogenetic and dynamics of the disease transmission in these endemic areas [32]. PvMSP1 gene codes for a major malaria vaccine candidate antigen. But its polymorphic nature represents an obstacle to the design of a protective vaccine. Present study will be helpful for the development of PvMSP1 based vaccine against P. vivax malaria and provide evidence driven knowledge towards development of effective control interventions in Myanmar and appropriate measures in achieving China malaria elimination goals. Noteworthy, Myanmar is one of the major malaria endemic countries in the South-East Asia region, the genetic diversity of the malaria parasite circulating in CMB areas provides additional supportive information. In total, we documented 11 synonymous and 112 non-synonymous haplotypes of which 71.11% and 36.66% previously reported. Of the 11 synonymous polymorphisms, 7 were previously identified. The change might be contributed to evolutionary and/or environmental changes characterized by different patterns compared to natural and geographical studies in the Great Mekong region.

Interestingly, the network analysis of identified haplotypes

of PvMSP1₄₂ showed that most prevalent haplotypes originated from Myanmar followed by Yunnan and Zhejiang provinces of China. This information is vital and indicates that understanding the genetic diversity and network provides insights into parasite strains dynamics in the region, and design of most appropriate programmes and interventions in reducing or blocking the transmission, curving the spread of parasite as well as containment of increasing resistant strain in the Great Mekong Region.

Phylogenic tree also showed a substantial degree of variability of the origin of the parasites. Although, all P. vivax clinical isolates, originated from the same species but analysis of these isolates showed distinct differences with the high prevalence of isolates from different countries of Myanmar, Thailand, Singapore, Bangladesh, South Korea, India, Vietnam, and Indonesia and different regions in China. Our findings are consistent with high malaria endemicity in Myanmar, where with the long borders proximity, haplotype diversity has been high comparable to the endemicity in vivax population from inland areas of China such as Anhui and Zhejiang province were lower. However, further studies on a larger population from these endemic geographic areas are required not only to determine the nationwide parasite genetic mapping and detailed malaria molecular epidemiology in CMB areas to provide evidence based decision and effective interventions [33,34].

ACKNOWLEDGMENTS

This work was supported by the National Key Research and Development Program of China (nos. 2016YFC1202000, 2016YFC1202001, 2016YFC1202003 and 2016YFC1200500), the National Natural Science Foundation of China (no. 81101266), the WHA-WHO Demonstration Project (no. UN-OPS/ANDI/G/2016/01), the Foundation of National Science and Technology Major Program (no. 2012ZX10004-220), and the Fourth Round of Three-Year Public Health Action Plan (2015-2017) in Shanghai, P. R. China (no. GWIV-29).

CONFLICT OF INTEREST

We have no conflict of interest related to this work.

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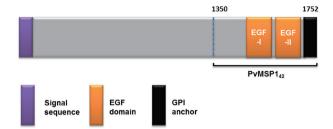


Fig. S1. Schematic diagram of *Plasmodium vivax* merozoite surface protein 1 (PvMSP1). The gene fragment encoding PvMSP1 $_{42}$ (including 2 EGF domains and a glycosylphosphatidylinositol anchor) was amplified and sequenced.

Fig. S2. The nucleotide sequence alignment of PvMSP1₄₂ from *P. vivax* isolates collected along the China-Myanmar border areas, local regions of Anhui, Yunnan and Zhejiang provinces in inland China.

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99.seq 14.seq 17.seq 18.seq 20.seq 22.seq 32.seq 33.seq 34.seq 36.seq 37.seq 38.seq 44.seq 45.seq 50.seq 51.seq				TG. TGA. TGA. TGA.	C			. AG	G.AG.AG.AG.ATG.AG.AG.AG.AG.AG.AG.AG.AG.A
99.seq 14.seq 17.seq 18.seq 19.seq 20.seq 27.seq 32.seq 33.seq 33.seq 34.seq 34.seq 35.seq 45.seq 50.seq 51.seq 53.seq			ACCACCACCACCACCACCACC	TG. TGA. TGA. TGA. TGA. TGA.	C			. AG	G.AG.AG.AG.ATG.AG.AG.AG.AG.AG.AG.AG.AG.AG.AG.AG.AG.A
9.seq 14.seq 17.seq 18.seq 19.seq 20.seq 27.seq 32.seq 33.seq 34.seq 36.seq 37.seq 38.seq 44.seq 45.seq 50.seq 51.seq			ACCACCACCACCACCACC	TG. TGA. TGA. TGA. TGA.	C			. AG	G.AG.AG.AG.ATG.AG.AG.AG.AG.AG.ATG.ATG.A

Fig. S2. Continued

157.seq		AC	CTGA	G			G	т	480
158.seq							A	G	480
215.seq							AG	G.A	480
160.seq		AC	CTGA	G	C .	G	G	т	480
162.seq									480
167.seq									
-									
172.seq									
173.seq									480
183.seq							AG	G.A	480
191.seq							T		480
199.seq							AG	G.A	480
200.seg		AC	CTGA	G			G	т	480
203.seq						7	AG	.G.A	480
Anhui 11.seq									
Anhui 2.seq									
Anhui 4.seq									480
Anhui_5.seq									
Anhui_9.seq									
Anhui_10.seq									480
									480
Yunnan 161.seq		AC	CTGA	G			G	т	480
Yunnan A66.seq		AC	CTGA	G			G	т	480
Yunnan T7.seg			CTGA	G		G	G	т	480
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Zhejiang_7.seq							T		480
	400	F00	F10	F20	520	F 4.0	550	F.C.0	`
	490	500	510	520	530	540	550	560	
6.seq	490 A							1	
6.seq 17.seq			.CA		AT.C				560
-	A	GAGT.	.CA		AT.C				560 560
17.seq 20.seq			.CA		AT.C GT				560 560 560
17.seq 20.seq 22.seq		GAGT.	.CA CA(	G.1	GT				560 560 560 560
17.seq 20.seq 22.seq 23.seq			.CA		GT CG.				560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq	A	GAGTAGGAGT.	.CA(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A(A	G.AAG.1	CAT.C A GT CG CAT.C				560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq	A	. G . AGT	.CA( 	G.1	C G T. C				560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq	A		C A	G.1 G.AA. G.AG.1 G.AA.	GTA				560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq	A		C A	G.1 G.A	GTAAA				560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq	A	G . AGT	C A	G.1 G.AG.1 G.AA. G.AA. G.AA.	CA T.C				560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq	A	G .AGTT.AG .AGTT.AG .AGTT.AG .AGTG .AGTG .AGT.	C A	G.1	CAT.CAGT.CAAGT.CAAGTAAGTAA				560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq	A	G .AGTT.AG .AGTT.AG .AGTT.AG .AGTG .AGTG .AGT.	C A	G.1	CAT.CAGT.CAAGT.CAAGTAAGTAA				560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq	A	G . AGT G . AGT G . AGT T . A . G . AGT T . A . G . AGT G . AGT G . AGT T . A	C A	G. T. G. G. T. G.	CA T.C				560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq	A	. G . AGT G . AGT T . A . T . A . T . A	C	G. A. A. G. A. A. A. A. A. G. A.	CA. T.C A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A A				560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq 63.seq	A		C A	G. A. A. G. A. G. A. G. A. G. A. A. G. A.	CA. T.C. A. G. T. C. A. G. T. C. A. G. T. G. T. G. T. G. T. G. T. G. T. C.				560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq 63.seq	A	. G . AGT G . AGT T . A . G . AGT T . A . G . AGT T . A . G . AGT G . AGT T . A . T . A	C A	G. A. A. G.	CA. T.C.  A. G. T. C.  A. T. C.  A. T. C.  A. G. T.  G. T.  G. T.  A. A.  G. T.  G. T.  A. C.  A. T.  C.  A. T.  C.  A. T.  C.  C.  C.  C.  C.  C.  C.  C.  C.				560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq	A	G .AGTT .AG .AGTT .AG .AGTT .AT .AG .AGTT .AT .AT .AT .A.	C. A	G. A. A. G. A. G. A. G. A. A. G. A. A. G.	CA. T.C. A. G. T. C. A. A. G. T. C. A. A. G. T. G. G. T. C. C. G. T. C. C. G. T. C. C. G. T. C. C. G. T. C.				560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq 74.seq	A	G .AGTT .AG .AGTT .AG .AGTT .AG .AGTT .AT .AT .AT .AT .AT .AT .AT .A.	C. A.	G. A. A. G. A. A. G. A. G. A. G. A. G. A. G. A. A. G. A.	CA. T.C. A. G. T. C. A. A. G. T. C. A. A. G. T. C. A. A. G. T. C. G. T. C. G. G. T. T. C. C. G. G. T. T. C. T. T. C. G. T. T. C. G. T. T. C. G. T. T. C. G. T. T. T. C. G. T. T. T. C. G. G. T.				560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 63.seq 63.seq 64.seq 70.seq 74.seq 75.seq	A	G .AGTT.AG.AGTT.AG.AGTT.AT.AT.AT.AT.AT.AT.AT.AT.AT.A.	C. A.	G. A. A. G.	CA. T.C. A. G. T. A. A. G. T. A.				560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 50.seq 51.seq 57.seq 59.seq 66.seq 70.seq 74.seq 75.seq 78.seq	A	G . AGT G . AGT T . A . G . AGT T . A . G . AGT T . A . T . A . T . A . T . A . T . A . T . A	C A	G. T.	CA T. C				560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq 74.seq 75.seq 78.seq 81.seq	T A C T A A A A A A A A A A A A A A A A	G . AGT G . AGT T . A . G . AGT T . A . G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A	C A	G. T.	CA. T.C. A. G. T. C. A. A. G. T. C. A. G. T. C. A. G. T. G. T. G. T. G. T. G. T. C. G. T. C. G. T. C. G. T. C. C. G.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 50.seq 51.seq 57.seq 59.seq 66.seq 70.seq 74.seq 75.seq	A	G . AGT G . AGT T . A . G . AGT T . A . G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A	C A	G. T.	CA. T.C. A. G. T. C. A. A. G. T. C. A. G. T. C. A. G. T. G. T. G. T. G. T. G. T. C. G. T. C. G. T. C. G. T. C. C. G.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq 74.seq 75.seq 78.seq 81.seq	T A C T A A A A A A A A A A A A A A A A	G . AGT G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A	C A	G. T.	CA. T.C. A. G. T. C. A. A. G. T. C. A. A. G. T. G. G. T. G. T. G. T. C. G. T. C. G. T. C. G. T. C. C. G. CA. T. C. C. G. CA. T. C. C. G. C. T.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq 74.seq 78.seq 78.seq 81.seq 82.seq	T A C T A A A A A A A A A A A A A A A A	G .AGTG .AGTT .A .G .AGTT .A .G .AGTT .A .G .AGTT .A .T .A	C A	G. A. A. G. T. G. A. G.	C. G. T. C. C. G. G. T. C. C. G. G. T. C. G. G. T. C. G. G. T. C. G. G. T. C. G. G. C. C. G. C. C. C. G. C. C. C. G. C.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq 74.seq 75.seq 78.seq 82.seq 85.seq 89.seq	A	G .AGTG .AGTT .A .G .AGTT .A .G .AGTG .AGTG .AGTG .AGTT .A .T .A	C A	G. A. A. G. T. G. A. G. A. G. A. G. A. G. A. G. A. G. T. G.	CA T.C. A. G. T. C. A. A. G. T. A. A. G. T. C. C. G. T. C. G. T. C. C. G. G. T. C. G. G. G. T. T. C. C. G. G. T. C. C. G. C. G. T. C. C. G. G. T. C. C. G. G. T. C. C. G. C. G. C. T. C. C. G. C. C. C. G. C. C. C. G. C. C. G. C.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 63.seq 66.seq 70.seq 74.seq 75.seq 78.seq 89.seq 81.seq 89.seq 91.seq	A	G .AGTT .AG .AGTT .AG .AGTT .AG .AGTT .AG .AGTT .AT .A.	C. A.	G. A. A. G. T. G.	CA T.C.  A. G. T. C.  A. T.C.  A. T.C.  A. A. G. T.  G. T.  G. T.  A. A. C.  C. G.  C. G.  C. G.  C. T.  C. G.  C. T.  C. G.  C. G.  C. T.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 63.seq 66.seq 70.seq 74.seq 75.seq 78.seq 89.seq 89.seq 91.seq 95.seq	A	G .AGTT .AG .AGTT .AG .AGTT .AT .A.	C. A. C. C.	G. T.	CA T.C.  A. G. T. C.  A. A. C. G. T.  G. T. C.  C. G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.  G. T. C.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 27.seq 38.seq 40.seq 50.seq 51.seq 57.seq 59.seq 66.seq 70.seq 74.seq 75.seq 78.seq 81.seq 82.seq 89.seq 89.seq 99.seq 99.seq	A	G . AGT T . A . G . AGT T . A . G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A	C A	G. T.	CA T.C		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 66.seq 70.seq 71.seq 75.seq 89.seq 81.seq 82.seq 89.seq 99.seq	T A C T A A A A A A A A A A A A A A A A	G . AGT T . A . G . AGT T . A . G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A	C A	G. T.	CA. T.C. A. G. T. C. A.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 59.seq 66.seq 70.seq 74.seq 75.seq 78.seq 81.seq 82.seq 89.seq 991.seq 998.seq 999.seq 114.seq	T A C T A A A A A A A A A A A A A A A A	G . AGT G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A	C A	G. T.	CA T.C. A. G. T. C. A.		T		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq 74.seq 75.seq 89.seq 81.seq 82.seq 89.seq 99.seq 99.seq 114.seq 117.seq	T A C T A A A A A A A A A A A A A A A A	G . AGT G . AGT T . A . G . AGT T . A . G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . AGC . AGC	C A	G. T.	CA T.C. A. G. T. C. A.		T.		560 560 560 560 560 560 560 560 560 560
17.seq 20.seq 22.seq 23.seq 23.seq 38.seq 40.seq 40.seq 50.seq 51.seq 57.seq 59.seq 66.seq 70.seq 74.seq 75.seq 78.seq 81.seq 82.seq 89.seq 991.seq 998.seq 999.seq 114.seq	T A C T A A A A A A A A A A A A A A A A	G . AGT G . AGT T . A . G . AGT T . A . G . AGT T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . T . A . AGC . AGC	C A	G. T.	CA T.C. A. G. T. C. A.		T.		560 560 560 560 560 560 560 560 560 560

Fig. S2. Continued

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119.seg	AAAGCAG	560
120.seg	TACAG.AA.GT	
127.seg	ACAG.TAT.C	
132.seq	TTACAG.AA.GT	
133.seq	CTGAGTCAAGA	
134.seq	CTGAGTCAAGAA	
136.seq	TACAG.AA.GT	560
137.seq	AAAGCAG	
138.seq	AAAGCAG	560
144.seq	TTACAG.AA.GT	560
145.seq	ACAG.TAT.C	560
150.seq	CTGAGTCAAGAA	560
151.seq	AAAGCAG	560
153.seq	TTACAG.AAGT	560
154.seq	TTACA.G.AA.GT	560
156.seq	A	560
157.seq	A	560
158.seq	A	560
215.seq	TA.G.AA.GT	
160.seq	AAAGCAG	
162.seq	A	
167.seq	TACAG.AAGT	
172.seq	A	
173.seq	A	
183.seq	T	
191.seq	CTGAGTCAAG	
199.seq	TACAG.AA.GT	
200.seq	ACG	
203.seq Anhui 11.seq	AAAGC.AGAG	
Anhui 2.seq	A	
Anhui 4.seq	AAAGCAG	
Anhui 5.seq	AAAGCAG	
Anhui 9.seq	AAAGCAGCG	
Anhui 10.seq	A	
	gCTGAGTCAAG	
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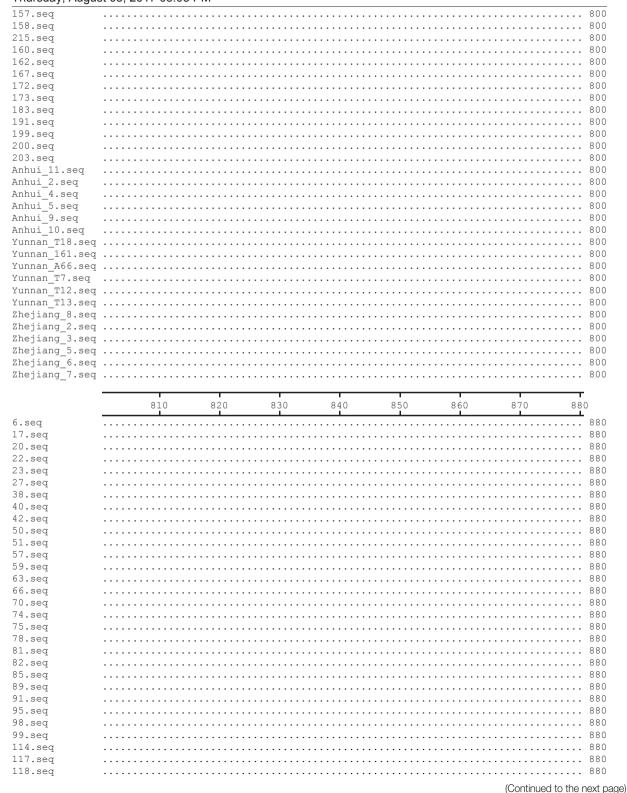


Fig. S2. Continued

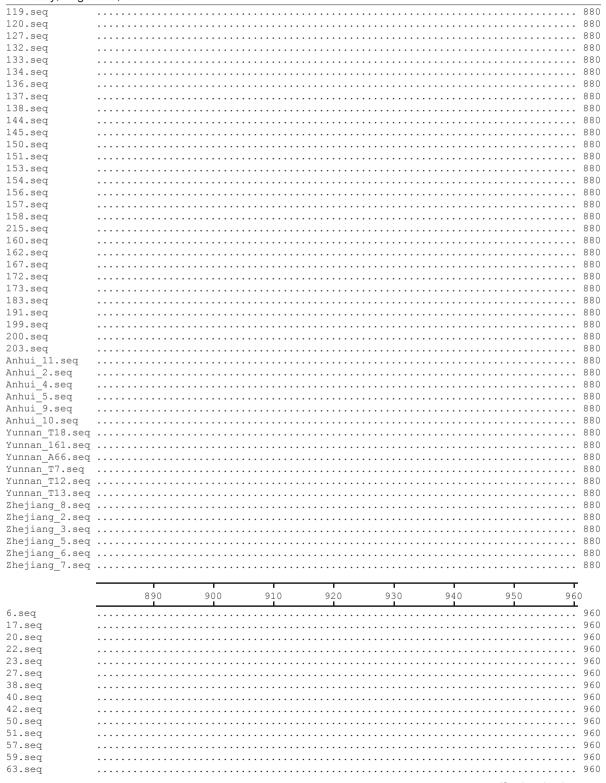


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Zhejiang_7.seq		120

Fig. S3. The amino acid sequence alignment of PvMSP1₄₂ from *P. vivax* isolates collected along the China-Myanmar border areas, local regions of Anhui, Yunnan, and Zhejiang provinces in inland China.

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17.seq 20.seq 22.seq	E.D.TTAK Q.L.S.V.QNLVN.QD.K.NE I.TSD.TT	.T	190		<u> </u>	220	230 2	40 . 718 . 718 . 718 . 718 . 718
17.seq 20.seq 22.seq 23.seq	E.D.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE	.T	190		NN	220	230 2	40 . 718 . 718 . 718 . 718 . 718 . 718 . 718
17.seq 20.seq 22.seq 23.seq 27.seq	E.D.TT. AK .Q.L.S.V.QNL.VN.QD.K.NE I.TSD.TT. AK .Q.L.S.V.QNL.VN.QD.K.NE	LN	190	l		220	230 2	40 . 718 . 718 . 718 . 718 . 718 . 718 . 718 . 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq	E.D.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE	LN	190	l		220	230 2	40 . 718 . 718 . 718 . 718 . 718 . 718 . 718 . 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq	E.D.TT. AK .Q.L.S.V.QNL.VN.QD.K.NE I.TSD.TT. AK .Q.L.S.V.QNL.VN.QD.K.NE	LN	190	1		220	230 2	40 . 718 . 718 . 718 . 718 . 718 . 718 . 718 . 718 . 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq	E.D.T.T.AK Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK Q.L.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NE	T. LN	190	l		220	230 2	40 . 718 . 718
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17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq	E.D.T.T.AK Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK Q.L.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NE	LN	190	l		220	230 2	40 . 718 . 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq	E.D.T.T.AK Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK Q.L.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.S.V.QN Q.L.S.V.QNL.VQN	T	190			220	230 2	40 . 718 . 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 50.seq 51.seq 57.seq 59.seq 63.seq	E.D.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE .L.VN.QD.K.NE .L.VN.QD.K.NE .L.VN.QD.K.NE .L.S.V.QNQ.L.S.V.QNL.VN.QD.K.NE .L.VN.QD.K.NE	T	190	l		220	230 2	40 • 718 • 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 40.seq 51.seq 57.seq 59.seq 63.seq 66.seq	L. VN. QD. K. NE . L. VV. QN L. VN. QD. K. NE I.T. SD. T. T. AK . Q. L. S. V. QN L. VN. QD. K. NE . L. VN. QD. K. NE	T	190			220	230 2	40 - 718 - 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq 63.seq 66.seq 70.seq	E.D.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK .Q.L.S.V.QNE.D.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.S.V.QNQ.L.S.V.QNL.VN.QD.K.NE .L.VN.QD.K.NE .L.VN.QD.K.NE .L.VN.QD.K.NE	LN	190	l	N N	220	230 2	40
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 59.seq 66.seq 70.seq 74.seq	E.D.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NE	T. LN	190		N N	220	230 2	40 - 718 - 718
17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 63.seq 66.seq 70.seq 74.seq 75.seq	E.D.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NE I.T.SD.T.T.AK .Q.L.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.S.V.QNL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NEL.VN.QD.K.NE	T. LN	190		N N	220	230 2	40 - 718 - 718
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17.seq 20.seq 22.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 63.seq 66.seq 70.seq 75.seq 75.seq 78.seq 78.seq 82.seq	L. VN.QD.K.NE L. VN.QD.K.NE LL. VN.QD.K.NE L. VN.QD.K.NE L. VN.QD.K.NE L. VN.QD.K.NE LL. VN.QD.K.NE	T	190		N N	220	230 2	40
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17.seq 20.seq 22.seq 23.seq 23.seq 27.seq 38.seq 40.seq 42.seq 50.seq 51.seq 57.seq 63.seq 66.seq 70.seq 74.seq 78.seq 81.seq 82.seq 89.seq		S	190			220	230 2	40
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Fig. S3. Continued

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119.seq	I. E. AN AA . S	8
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127.seg	E.D.TT. AK.T 71	
132.seq	LVN.OD.K.NE.I	
133.seg	Q.L.S.V.QN. LN. 71	
134.seg	O.L.S.V.ON. LN. 71	
136.seg	.L.DN.QD.K.NE.I. 71	
137.seq	I. E. AN AA. S	
138.seq	I. E. AN. AA. S	
144.seq	.L. VN.OD.K.NE.I. 71	
145.seq	E.D.T.T.AK.T. 71	
150.seq	O.L.S.V.ON. LN 71	
151.seq	I. E. AN. AA. S	
153.seq	L. VN.OD.K.NE.I. 71	
154.seq	L. VN.OD.K.NE.I. 71	
156.seq	E.D.TT. AK.T 71	
157.seq	I SD.T. T S	
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215.seq	.L. VN.OD.K.NE.I. 71	
160.seq	I. E. AN AA. S	
162.seq	I. SD.T. T. S. N. 71	
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173.seq	IE. AN.AAS	
183.seq	L. VN. OD. K. NE. I	
191.seq	O.L.S.V.ON LN N 71	
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200.seq	I. SD.T.T.S. N. 71	
203.seq	. L. VN.OD.K.NE.I	
Anhui 11.seq	I. E. AN AA. S	
Anhui 2.seq	I. E. AN. AA. S	
Anhui 4.seq	I. E. AN. AA. S	
Anhui 5.seq	IE. AN.AA S	
Anhui_9.seq	IE. AN.AA S	
Anhui 10.seg	IE.AN.AAS	
	Q.L.S.V.QN. LN. 71	
	I. E. AN AA S	
	I. E. AN AA. S	
	ISD.T.TS	
	E.D.TTAK.T	
	Q.L.SD.TTS	
	TSD.T.TS	
	ISD.TTS	
	ISD.TTS	
	Q.L.S.V.QNLN71	
	I SD.T.T S N	
	Q.L.S.V.QNLN	
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	250 260 270 280 290 300 310 320	
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42.seq	95	38
50.seq	95	38
51.seq	95	8
57.seq	95	8
59.seq	95	8
63.seq	95	38
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#### Fig. S3. Continued

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66.seq	 958
70.seq	 958
74.seq	 958
75.seq	 958
78.seq	 958
81.seq	 958
82.seq	 958
85.seq	 958
89.sea	 958
91.seq	
95.sea	
98.seq	
99.seq	
114.seq	
117.seq	
118.seq	
119.seq	
120.seq	
127.seq	
132.seq	
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151.seq	
153.seq	
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156.seq	
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203.seq	
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Anhui_2.seq	
Anhui_4.seq	
Anhui_5.seq	
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Anhui_10.seq	
Yunnan_T7.seq	 958
Yunnan_T12.seq	 958
Yunnan_T13.seq	 958
<pre>Zhejiang_8.seq</pre>	 958
Zheijang 7 seg	958

Fig. S3. Continued

	330	340	350	360	370	380	390	40
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				E				
q								
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9								
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a a								
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4 7								
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q P								
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11.seq								
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Fig. S3. Continued

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Anhui 5.seq		119
Anhui 9.seq		
Anhui 10.seq		
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	EE	
Yunnan_T12.seq		119
Yunnan_T13.seq		119
Zhejiang 8.seq		119
Zhejiang 2.seg		119
Zheiiang 3.seg		119
	·	
Zile Jiang_/.seq		113
	<del></del>	
	<u> </u>	
6.seq		120
17.seq		120
20.seq		120
22.seq		120
23.seq		120
27.seq		120
38.seq		120
40.seq	•••	120
42.seq	•••	120
50.seq		120
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51.seq	•••	120
57.seq	•••	120
59.seq	•••	120
63.seq	•••	120
66.seq		120
70.seq		120
74.seq		120
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85.seq		120
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91.seq		120
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95.seq	•••	
98.seq	•••	120
99.seq	•••	120
114.seq	···	120
117.seq		120
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119.seq		120
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145.seq		120
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151.seq		120
153.seq		120
154.seq		120
156.seq		120
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Fig. S3. Continued

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160.seq	•••	120
162.seq	•••	120
167.seq	•••	120
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203.seq	•••	120
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Anhui_2.seq	•••	120
Anhui_4.seq	•••	120
Anhui_5.seq	•••	120
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Anhui_10.seq	•••	120
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Yunnan_161.seq		120
Yunnan_A66.seq		120
Yunnan_T7.seq		120
Yunnan_T12.seq		120
Yunnan_T13.seq		120
Zhejiang_8.seq		120
<pre>Zhejiang_2.seq</pre>		120
<pre>Zhejiang_3.seq</pre>		120
Zhejiang_5.seq		120
<pre>Zhejiang_6.seq</pre>		120
Zhejiang_7.seq	•••	120

Decoration 'Decoration #1': Hide (as '.') residues that match the Consensus exactly.