Kernel Density Estimation Model (KDEM) Vignette Instructions

J.L. Tracy

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This vignette provides steps for using example monarch roosting (migratory) occurrence data and a human population grid from Tracy et al. (submitted) to generate a training set ensemble (TSE) of kernel density estimate models (KDEMs) for eastern monarch migration pathways.

1) Open a new empty ArcMap project and add the raster *pop10kmn3* (year 2000 human population density in a 3 km radius generalized to a 10 km resolution) (raster located in *KDMVignetteData* folder). This raster is in the North America Albers Equal Area Conic projection that is used for generating KDEs. Make sure the monarch roost data point shapefile *MonarchRoosts\_2002\_2006East.shp* is in the same folder as the raster.

2) Use Python script editor such as IDLE to open the ArcPython script *MonarchRoostDataProcessingBatch.py* (in the ArcPythonScript folder). Follow the instructions at the top of the script for setting the Geoprocessing Environments in the above ArcMap project so that the output coordinates, processing extent, snap raster and cell size set are set to match the raster *pop10kmn3*. Make sure the specified directories in the script match the above directory for data in step 1 on your computer. Open the ArcPython window in the ArcMap project, and paste in several lines at a time of commands to run from the ArcPython script *MonarchRoostDataProcessingBatch.py*. This script will project the roost data and use the human population density raster to upweight monarch roost data in low population areas, creating the shapefile *rstdenpopall.shp*.

3) Use an R script editor like R Studio or Tinn-R to open the R script *KDEModel\_MonarchRoostTSE.R* (located in the RScript folder). Make sure FunctDirect (line 25) matches the location of *KDEMSubset\_GridTrainTestEvalAIC\_Calib\_Function.R* on your computer. Also, make sure the InDirect directory in the script matches the desired location on your computer (line 56). Run the R script from lines 1 to 128 to establish various parameters and plot the shapefile *rstdenpopall.shp.*

4) Similar to step 2, use the ArcPython script *MonarchRoostBackgroundExtentRaster\_PseudoabsenceRasterGeneration.py* to generate a background raster and pseudoabsence raster for later deriving Background points (not used for KDEM, but used if running MaxEnt) and Pseudoabsence points in R script below.

5) Going back to R script of step 3, start running code from line 136 to 364. This step creates Pseudoabsence points and monarch roost data point training set shapefiles for the KDE training set ensembles in the next step.

6) Similar to step 2, use the ArcPython script *MonarchRoostKfoldDataProjection\_KDEProcessingBatch\_PopDenInd.py* to generate a training set ensemble of three KDE surface rasters.

7) Back in the R script, run lines 374 to 518 for generating the training set ensemble of three KDEM rasters and an output a csv file of evaluation statistics in the InDirect directory. A new output subdirectory will be generated called something like “MonRstKDEMCNAA” (unless you changed parts of name in R script) for producing the three training set KDEM rasters.

8) Similar to step 2, use the ArcPython script *MonarchRoostKfoldDataProjectionTSE\_KDEProcessingBatch.py* to combine the three above generated training set KDEMS into a Training Set Ensemble (TSE) that can be used to display a minimum consensus (value 0 is no color, 1-3 is same color) or 100% consensus (values 0-2 is no color, 3 is color) binary calibration model.

Tracy JL, Tracy JL, Kantola T, Baum KA, Coulson RN (submitted) Modeling fall migration pathways and spatially identifying potential migratory hazards for the eastern monarch butterfly. Submitted to Landscape Ecology