

## Protein Extraction and Calibration Curve

- If storing samples for more than 30 days, add a protease inhibitor to hemolymph extract.
- With the larva constrained and folded in half, exposing the dorsum, make an incision through the cuticle at the proleg to extract hemolymph
- Pipet the hemolymph into 100µL of PBS
- Place the test tube into ice to slow oxidation and add inhibitor(s).
- Store supernatant overnight at -80°C

## Protein Quantification using Bradford Assay kit See: Pierce Bradford Protein Assay Kit.pdf

Prepare the Diluted Albumin (BSA) Standards

Dilution Scheme for Standard Test Tube and Microplate Protocols (Working Range = 100-1500µg/mL)

Vial	Volume of diluent (µl) $\text{dH}_2\text{O}$	Volume (µl) and source of BSA	Final BSA concentration (mg/ml)
A	0	300 Stock	2
B	125	375 Stock	1.5
C	325	325 Stock	1
D	175	175 Vial B dilution	0.75
E	325	325 Vial C dilution	0.5
F	325	325 Vial E dilution	0.25
G	325	325 Vial F dilution	0.125
H	400	100 Vial G dilution	0.025
I	400	0	0 = Blank

## Standard Microplate Protocol (Working Range = 100-1500µg/mL)

- Upto 7 treatments can fit on a plate
  - 3 Reps of each dilution
- Prepare three (100µl) dilutions of each of your protein supernatant samples: 1X, 10X, 20X.
  - [diluting volume/#'x' volume] = diluted volume
  - Ex: 100µl/1x = 30µl of unknown need to be added → mark 100% stock
  - Ex: 100µl/10x = 10µl of unknown need to be added → mark 10% stock
  - Ex: 100µl/20x = 5µl of unknown need to be added → mark 5% stock
- Mix each sample for 5 secs and Pipette 5µL of each standard or unknown sample into the appropriate microplate wells.
- Prepare Coomassie reagent, allow it to equilibrate to room temperature (WR). 200µl needed per sample plus 30%
- ((#stds + #unk)\*(3rep)\*(250) + (0.3\*WR needed) = WR needed
- Add 200µL of the Coomassie Reagent to each well.
- Cover plate with *Press'n Seal*, mix with plate shaker for 30 seconds, then incubate at room temperature for 10 minutes.
- Measure the absorbance at or near 595 nm on a plate reader.