# Simulation and inference of phylodynamic individual level models

Justin Angevaare<sup>1</sup>, Zeny Feng<sup>1</sup>, Rob Deardon<sup>2</sup>

<sup>1</sup>University of Guelph <sup>2</sup>University of Calgary

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#### Overview

Phylodynamics

Individual level models

Transmission pathway ILMs

Phylodynamic ILMs

Pathogen.jl

### **Phylodynamics**

- The joint or conditional investigation of the dynamics of disease spread and evolution
- Appropriate when epidemiological and evolutionary processes occur at similar time scales
- Requires dense genetic sampling of a pathogen during an epidemic

### Phylodynamics software I

- ► Typically a phylogenetic tree is inferred, and from that a transmission network generated
- ► MCMC; Metropolis-Hasting with specialized proposal methods for exploring phylogenetic tree space used
- ▶ Some challenges persist with low acceptance rates

#### Phylodynamics software II

- ▶ We leverage parameters from Individual Level Models (ILMs) (Deardon et al., 2010)
- Generate phylogenetic tree from transmission network proposals
  - ▶ "high acceptance" proposals

#### **ILMs**

- ► Time inhomogeneous Poisson process of disease spread through a heterogeneous population
- ▶ Inference typically conducted in Bayesian framework

#### Exposure in ILMs

- ▶ Individuals are in a single disease state during any period t
- Transition of i from susceptible to exposed occurs with rate

$$\lambda_{SE}(i,t) = \left[\Omega_{S}(i) \sum_{k \in I_{(t)}} \Omega_{T}(k) \kappa(i,k)\right] + \epsilon(i,t) \text{ for } i \in S_{(t)}$$
(1)

#### where.

- $ightharpoonup I_{(t)}$  is the set of infectious individuals during time period t,
- $ightharpoonup S_{(t)}$  is the set of susceptible individuals during time period t,
- $ightharpoonup \Omega_S(i)$  is an susceptibility function,
- $ightharpoonup \Omega_T(k)$  is a transmissability function,
- $\blacktriangleright \kappa(i,k)$  is an infection kernel, and,
- $\epsilon(i, t)$  is a sparks function.

#### **ILM** likelihood

$$L(D|\theta) = \prod_{t=1}^{T-1} \psi(t) v(t) \exp\left\{-v(t)\Delta_t\right\},\tag{2}$$

where,

$$\psi(t) = \begin{cases} \frac{\lambda_{SE}(i,t)}{v(t)} & \text{if } i \in (S_{(t)} \cap E_{(t+1)}) \\ \frac{\lambda_{EI}(j,t)}{v(t)} & \text{if } j \in (E_{(t)} \cap I_{(t+1)}) \\ \frac{\lambda_{IR}(k,t)}{v(t)} & \text{if } k \in (I_{(t)} \cap R_{(t+1)}) \end{cases}$$

$$v(t) = \sum_{i=0}^{\infty} \lambda_{SE}(i,t) + \sum_{i=0}^{\infty} \lambda_{EI}(j,t) + \sum_{i=1}^{\infty} \lambda_{IR}(k,t).$$
 (4)

where  $\Delta_t$  is the length of  $t^{th}$  inter-event period.

### Transmission pathway ILM extension

- Extension allowing simulation and inference of infection sources
- Necessary for phylodynamic extension

#### Exposure in transmission pathway ILM

 Separate exposure rates are defined for each susceptible-infectious combination as

$$\lambda_{SE}^*(i,k,t) = \Omega_S(i)\Omega_T(j)\kappa(i,j) \text{ if } i \in S_{(t)}, k \in I_{(t)},$$
 (5)

and for exposures from an exogenous source as

$$\lambda_{SE}^*(i,t) = \epsilon(i,t) \text{ if } i \in S_{(t)}.$$
 (6)

### Transmission pathway ILM likelihood

$$L(D|\theta) = \prod_{t=1}^{T-1} \zeta(t) v(t) \exp\left\{-v(t)\Delta_t\right\},\tag{7}$$

where,

$$\zeta(t) = \begin{cases} \frac{\lambda_{SE}^*(i,k,t)}{\upsilon(t)} & \text{if } i \in (S_{(t)} \cap E_{(t+1)}) \text{ by endogenous exposure by } k \\ \frac{\lambda_{SE}^*(i,t)}{\upsilon(t)} & \text{if } i \in (S_{(t)} \cap E_{(t+1)}) \text{ by exogenous exposure} \\ \frac{\lambda_{EI}(j,t)}{\upsilon(t)} & \text{if } j \in (E_{(t)} \cap I_{(t+1)}) \\ \frac{\lambda_{IR}(k,t)}{\upsilon(t)} & \text{if } k \in (I_{(t)} \cap R_{(t+1)}) \end{cases}$$

$$(8)$$

#### Phylodynamic ILM extension

- ► Joint model of disease spread and evolution through a heterogeneous population
- Combines transmission pathway ILM with a phylogenetic tree consistent with transmission network
- ▶ Exposure times are assumed to be pathogen divergence dates

#### Phylodynamic ILM simulation

- ► Gillespie (1977) stochastic simulation method can be utilized:
- Event time is generated from an exponential distribution based on sum of rates
- Event type is generated from discrete distribution
- Event rates are updated

### Phylodynamic ILM inference: likelihood

- ► Full likelihood is the product of the transmission pathway ILM likelihood (Eq. 7) with corresponding phylogenetic tree likelihood
- ▶ Phylogenetic tree likelihood can be calculated using the pruning algorithm of Felsenstein (1973, 1981)

#### Phylodynamic ILM inference: MCMC I

- Specialized MCMC algorithms necessary for phylodynamic II M
- ► Algorithm must efficiently explore full posterior distribution of:
  - ► Augmented data (transmission network & event timings)
  - ▶ ILM and substitution model parameters

#### Phylodynamic ILM inference: MCMC II

- ► We implement an MCMC algorithm with several different step types:
  - 1. Propose ILM or substitution model parameters sample
  - 2. Propose new set of event times
  - 3. Propose new set of exposure sources

#### MCMC III

Proposals to transmission network are generated with a discrete probability distribution. An individual i will be proposed to have been exposed by k with probability

$$\frac{\lambda_{SE}^{*}(i,k,t)}{\lambda_{SE}^{*}(i,t) + \sum_{k} \lambda_{SE}^{*}(i,k,t)},$$
(9)

and by an exogenous source with probability

$$\frac{\lambda_{SE}^*(i,t)}{\lambda_{SE}^*(i,t) + \sum_k \lambda_{SE}^*(i,k,t)}.$$
 (10)

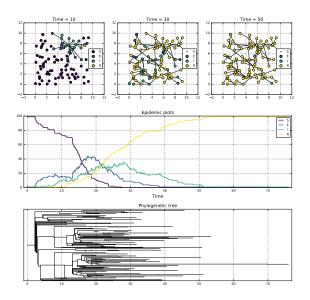
▶ This is accounted for in Metropolis-Hastings acceptance ratio

### Pathogen.jl

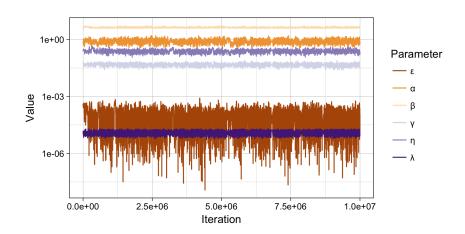
- jangevaare/Pathogen.jl is a flexible full featured package which is in development for Phylodynamic modelling in Julia
- Julia is a fast, high level language designed for scientific computing applications that is approaching it's 1.0 release
- ▶ SEIR, SEI, SIR, and SI ILMs with fully customizable  $\Omega_S$ ,  $\Omega_T$ ,  $\kappa$ ,  $\epsilon$ ,  $\Omega_L$ , and  $\Omega_R$ . Many substitution models.
- Initial work has included some simplifying assumptions regarding pathogen diversity.



# Phylodynamic simulations in Pathogen.jl



# Phylodynamic inference in Pathogen.jl



# Initial findings and ongoing work

- Inference of  $\lambda_{SE}^*(i, t)$ , and exogenous exposures sensitive to external diversity
- Incorporation of more realistic external pathogen source diversity
- Incorporation of within host diversity
- Development of guidelines for MCMC tuning
- Further speed and usability optimizations in Pathogen.jl

#### References

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#### Thank you!

□ jangevaa@uoguelph.ca | ○ jangevaare







#### Latency in ILMs

▶ Transition of *j* from exposed to infectious occurs with rate

$$\lambda_{EI}(j,t) = \Omega_L(j) \text{ for } j \in E_{(t)}$$
(11)

where,

- $\triangleright$   $E_{(t)}$  is the set of exposed individuals during time period t, and,
- $\triangleright \Omega_L(i)$  is a latency function.

#### Removal in ILMs

▶ Transition of *k* from infectious to removed occurs with rate

$$\lambda_{IR}(k,t) = \Omega_R(k) \text{ for } k \in I_{(t)}$$
 (12)

where,

•  $\Omega_R(k)$  is a removal function.

# Phylogenetic tree likelihood I

$$\begin{split} L(D|\theta) = & \pi \times S_N, \\ S_N = & [P_{N_L} \times S_{N_L}] \odot [P_{N_R} \times S_{N_R}] \\ S_{N-1} = & [P_{N-1_L} \times S_{N-1_L}] \odot [P_{N-1_R} \times S_{N-1_R}], \\ & \cdots \\ S_{\frac{N+1}{2}+1} = & \left[ P_{\frac{N+1}{2}+1_L} \times S_{\frac{N+1}{2}+1_L} \right] \odot \left[ P_{\frac{N+1}{2}+1_R} \times S_{\frac{N+1}{2}+1_R} \right], \\ P_j = & \exp(Q \times d_j), \end{split}$$

# Phylogenetic tree likelihood II

where,

- $\blacktriangleright$   $\pi$  is a vector of nucleotide frequencies according to a nucleotide substitution model, Q
- $\triangleright$   $S_i$  is a vector of nucleotide likelihoods at node j
- ▶ ⊙ is the component-wise product
- N is the total number of nodes
- $\triangleright$   $S_1, \ldots, S_{\frac{N+1}{2}}$  are observed
- $\rightarrow$   $j_L$ ,  $j_R$ , represent the left and right children nodes of node j, and.
- ▶ d<sub>j</sub> is the length of the branch connecting node j to its parent node.

Felsenstein's pruning algorithm requires that the nucleotide likelihoods at internal nodes are calculated following a postorder traversal of the phylogenetic tree. As each site is assumed to be independent, the full likelihood of any phylogenetic tree is the product of site-specific likelihoods.

#### Alternative continuous time ILM likelihood I

$$L(D|\theta) = \prod_{t=1}^{T-1} \left[ \prod_{i \in (S_{(t)} \cap E_{(t+1)})} P_{SE}(i,t) \prod_{j \in (E_{(t)} \cap I_{(t+1)})} P_{EI}(j,t) \right]$$

$$\prod_{k \in (I_{(t)} \cap R_{(t+1)})} P_{IR}(k,t) \prod_{i \in (S_{(t)} \cap S_{(t+1)})} P_{SS}(i,t)$$

$$\prod_{j \in (E_{(t)} \cap E_{(t+1)})} P_{EE}(j,t) \prod_{k \in (I_{(t)} \cap I_{(t+1)})} P_{II}(k,t) \right], \quad (13)$$

where,

- ▶ D are the observed data, which includes observations of infectiousness and removal times for each individual and any risk factor information required by  $\Omega_S$ ,  $\Omega_T$ ,  $\kappa$ ,  $\Omega_L$ , and  $\Omega_R$ ,
- $m{ heta}$  are the model parameters and augmented event timings for all observed individuals, and...

#### Alternative continuous time ILM likelihood II

$$P_{SE}(i,t) = \lambda_{SE}(i,t) \exp \{-\lambda_{SE}(i,t)\Delta_{t}\},$$

$$P_{EI}(j,t) = \lambda_{EI}(j,t) \exp \{-\lambda_{EI}(j,t)\Delta_{t}\},$$

$$P_{IR}(k,t) = \lambda_{IR}(k,t) \exp \{-\lambda_{IR}(k,t)\Delta_{t}\},$$

$$P_{SS}(i,t) = \exp \{-\lambda_{SE}(i,t)\Delta_{t}\},$$

$$P_{EE}(j,t) = \exp \{-\lambda_{EI}(j,t)\Delta_{t}\},$$

$$P_{II}(k,t) = \exp \{-\lambda_{IR}(k,t)\Delta_{t}\}.$$
(14)
(15)
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