

✓ BPL_IEC_validation script with FMPy

The key library FMPy is installed.

After the installation a small application BPL_IEC_validation is loaded and run. You can continue with this example if you like.

```
!lsb_release -a # Actual VM Ubuntu version used by Google
```

```
⇒ No LSB modules are available.
   Distributor ID: Ubuntu
   Description:    Ubuntu 22.04.3 LTS
   Release:        22.04
   Codename:       jammy
```

```
%env PYTHONPATH=
```

```
⇒ env: PYTHONPATH=
```

```
!wget https://repo.anaconda.com/miniconda/Miniconda3-py312_24.3.0-0-Linux-x86_64.
!chmod +x Miniconda3-py312_24.3.0-0-Linux-x86_64.sh
!bash ./Miniconda3-py312_24.3.0-0-Linux-x86_64.sh -b -f -p /usr/local
import sys
sys.path.append('/usr/local/lib/python3.12/site-packages/')
```

```
⇒ --2024-11-07 10:43:07-- https://repo.anaconda.com/miniconda/Miniconda3-py312
Resolving repo.anaconda.com (repo.anaconda.com)... 104.16.191.158, 104.16.32.
Connecting to repo.anaconda.com (repo.anaconda.com)|104.16.191.158|:443... co
HTTP request sent, awaiting response... 200 OK
Length: 143351488 (137M) [application/octet-stream]
Saving to: 'Miniconda3-py312_24.3.0-0-Linux-x86_64.sh'
```

```
Miniconda3-py312_24 100%[=====>] 136.71M  105MB/s in 1.3s
```

```
2024-11-07 10:43:08 (105 MB/s) - 'Miniconda3-py312_24.3.0-0-Linux-x86_64.sh' :
```

```
PREFIX=/usr/local
Unpacking payload ...
```

```
Installing base environment...
```

```
Preparing transaction: ...working... done
Executing transaction: ...working... done
installation finished.
```

```
!conda update -n base -c defaults conda --yes
```

```
⇒
```

07/11/2024, 11:49

BPL_IEC_validation_fm


py_colab.ipynb - Colab

openssl-3.0.15	5.2 MB	:	0%	0/1	[00:00<?, ?it/s]
conda-24.9.2	1.1 MB	:	0%	0/1	[00:00<?, ?it/s]
certifi-2024.8.30	163 KB	:	0%	0/1	[00:00<?, ?it/s]
ca-certificates-2024	130 KB	:	0%	0/1	[00:00<?, ?it/s]
openssl-3.0.15	5.2 MB	:	0%	0.003007460830410892/1	[00:00<01:59
certifi-2024.8.30	163 KB	:	10%	0.09811307196196202/1	[00:00<00:03,
ca-certificates-2024	130 KB	:	12%	0.12323429860849944/1	[00:00<00:02,
conda-24.9.2	1.1 MB	:	1%	0.013754463022707792/1	[00:00<00:26
openssl-3.0.15	5.2 MB	:	35%	0.34886545632766347/1	[00:00<00:00,
certifi-2024.8.30	163 KB	:	100%	1.0/1	[00:00<00:00, 2.60it/s]
certifi-2024.8.30	163 KB	:	100%	1.0/1	[00:00<00:00, 2.60it/s]
ca-certificates-2024	130 KB	:	100%	1.0/1	[00:00<00:00, 2.57it/s]
ca-certificates-2024	130 KB	:	100%	1.0/1	[00:00<00:00, 2.57it/s]
frozendict-2.4.2	36 KB	:	100%	1.0/1	[00:00<00:00, 1.89it/s]
openssl-3.0.15	5.2 MB	:	100%	1.0/1	[00:01<00:00, 1.01it/s]
conda-24.9.2	1.1 MB	:	100%	1.0/1	[00:01<00:00, 1.00it/s]

Preparing transaction: done
Verifying transaction: done
Executing transaction: done

```
!conda --version
!python --version
```

 conda 24.9.2

 Python 3.12.2

```
!conda install -c conda-forge fmpy --yes # Install the key package
```



EXECUTING TRANSACTION: DONE

```
#!conda install matplotlib --yes
```

Now specific installation and the run simulations. Start with connecting to Github. Then upload the two files:

- FMU - BPL_IEC_operation_linux_om_me.fmu
- Setup-file - BPL_IEC_operation_fm.py_explore.py

```
%%bash
```

```
git clone https://github.com/janpeter19/BPL_IEC_validation
```

📁 Cloning into 'BPL_IEC_validation'...

```
%cd BPL_IEC_validation
```

📁 /content/BPL_IEC_validation

✓ BPL IEC validation

Author: Jan Peter Axelsson

Here I try to reproduce some results from Jonas Månssons master thesis report "Control of chromatography column in production scale", TFRT-5599.

The interaction with the model has changed slightly so that linear flow rate $LF = F/\text{area}$ is used. The column volume is now specified in terms of (cross-section) area times height. Thus by changing area the scale up works fine and what you do in practice. Further the time unit changed from seconds to minutes.

The model has 5 states as listed below, 3 states in liquid mobile phase and 2 states for the gel for bound proteins. In a few of Jonas figures also bound ions of the buffer are plotted but I do not do that. Bound ions of the buffer can be calculated from the difference of total binding capacity Q_{av} and bound protein and bound protein antagonist, see section 5.1 in the report.

The molecular weights listed below are not used in the simulations. They just give typical values and the molecular weight for bound protein and antagonist protein is just arbitrary here.

Parameters of the model in general as well as time scale are all arbitrary and focus is on qualitative aspects of the model.

The height of the column is 20 cm which is a common size in the industry.

```
run -i BPL_IEC_fm.py_explore.py
```

Linux - run FMU pre-comiled OpenModelica

Model for bioreactor has been setup. Key commands:

- par() - change of parameters and initial values
- init() - change initial values only
- simu() - simulate and plot
- newplot() - make a new plot
- show() - show plot from previous simulation
- disp() - display parameters and initial values from the last simulation
- describe() - describe culture, broth, parameters, variables with values/units

Note that both disp() and describe() takes values from the last simulation and the command process_diagram() brings up the main configuration

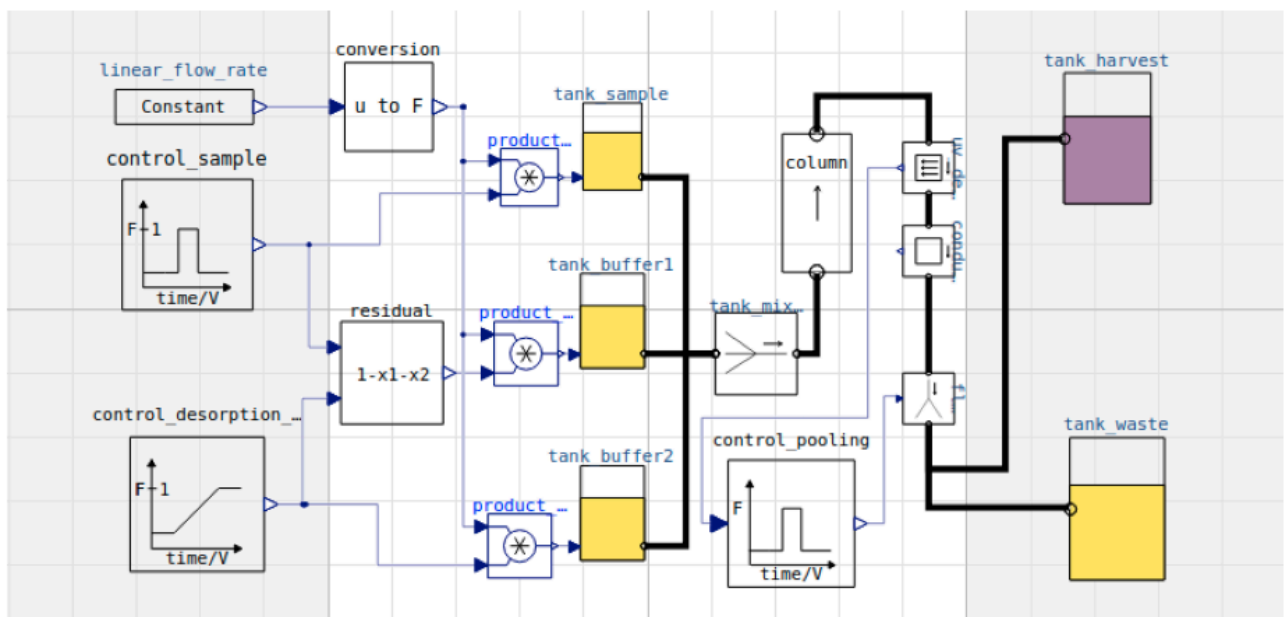
Brief information about a command by help(), eg help(simu)

Key system information is listed with the command system_info()

```
plt.rcParams['figure.figsize'] = [30/2.54, 24/2.54]
```

The process diagram is made outside Modelica to illustrate the configuration
process_diagram()

No processDiagram.png file in the FMU, but try the file on disk.



```
describe('chromatography'); print(); #describe('liquidphase')
```

Ion exchange chromatography controlled with varying salt-concentration. The

✓ Loading or adsorption

The parameter notation and values are the same as in the referred report. However the flow rate is here denoted F while q in the report. The column is divided in $n=8$ sections and set at compilation time. The values are arbitrarily chosen in the report and the focus is on qualitative aspects of the model.

The simplified model describe only the column in terms of volume and does not distinguish a high column with a small diameter from a lower with larger diameter.

The parameters k_1 , k_2 , k_3 , k_4 and Q_{av} are given relative volume and with increased column volume a larger capacity is thus obtained.

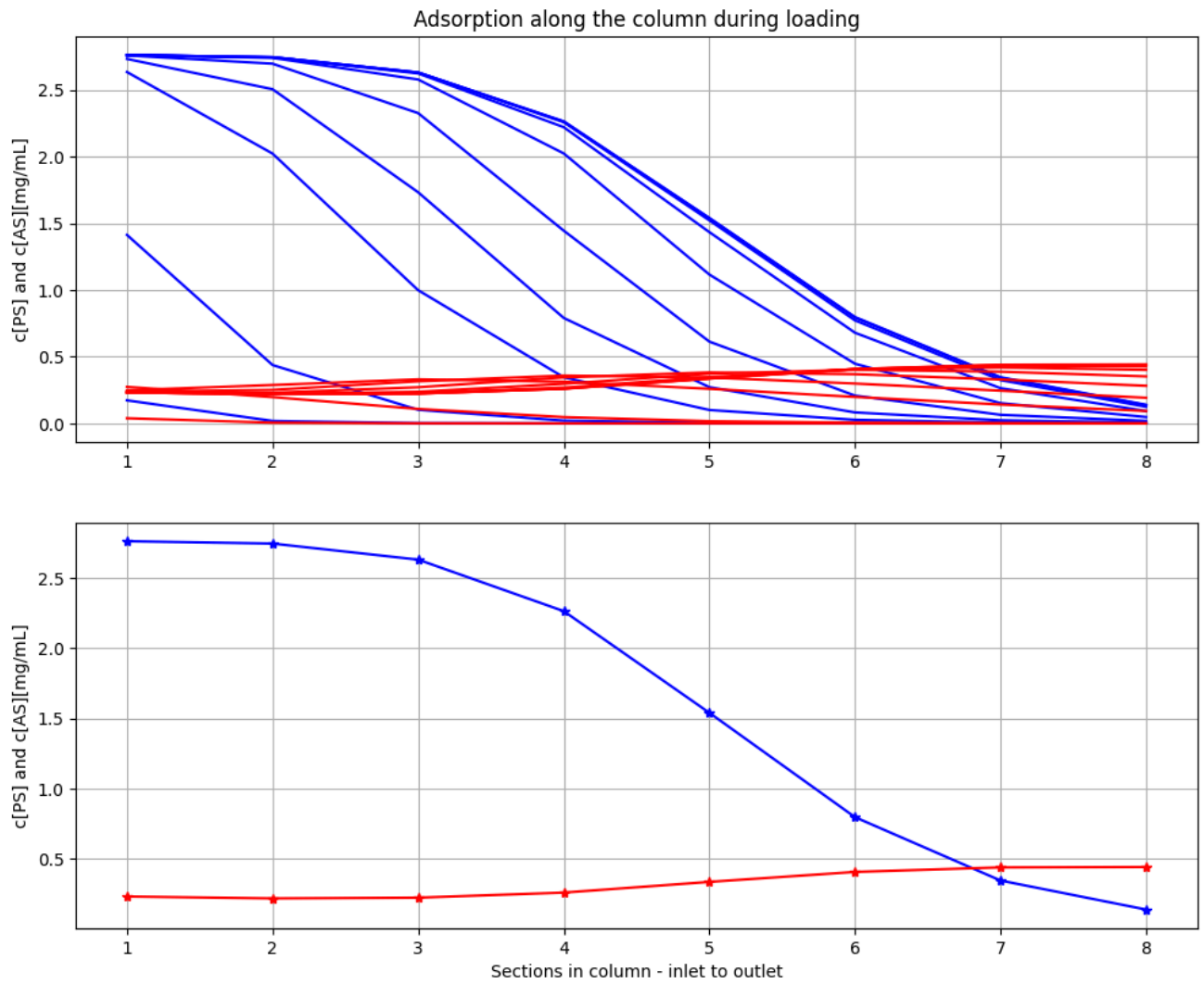
```
# Loading of the column – try to reproduce Jonas figure 13.
newplot(title='Adsorption along the column during loading', plotType='Loading')

# Sample
par(P_in=1.0, A_in=1.0, E_in=0)

# Column properties
par(k1=0.3, k2=0.05, k3=0.05, k4=0.3, Q_av=3.0)
par(height=20, diameter=0.714)
par(x_m=0.3)

# Operation
par(E_in_desorption_buffer=8)
par(LFR=12)
#par(scale_volume=False)
par(start_adsorption=0, stop_adsorption=50)
par(start_desorption=150, stationary_desorption=450)
par(start_pooling=220, stop_pooling=450)

# Simulation
simu(100)
```



The results are the same as Figure 13 in [1].

```
# We just check that we had the same volume flow rate as Jonas
#describe('F')
```

```
describe('V')
```



Column volume total - derived : 8.008 [mL]

```
model_get('column.x_m')
```

⇒ 0.3

```
model_get('column.V_m')
```

⇒ 2.4023570526441924

```
describe('column.n')
```

⇒ Number of sections of the column : 8.0 [None]

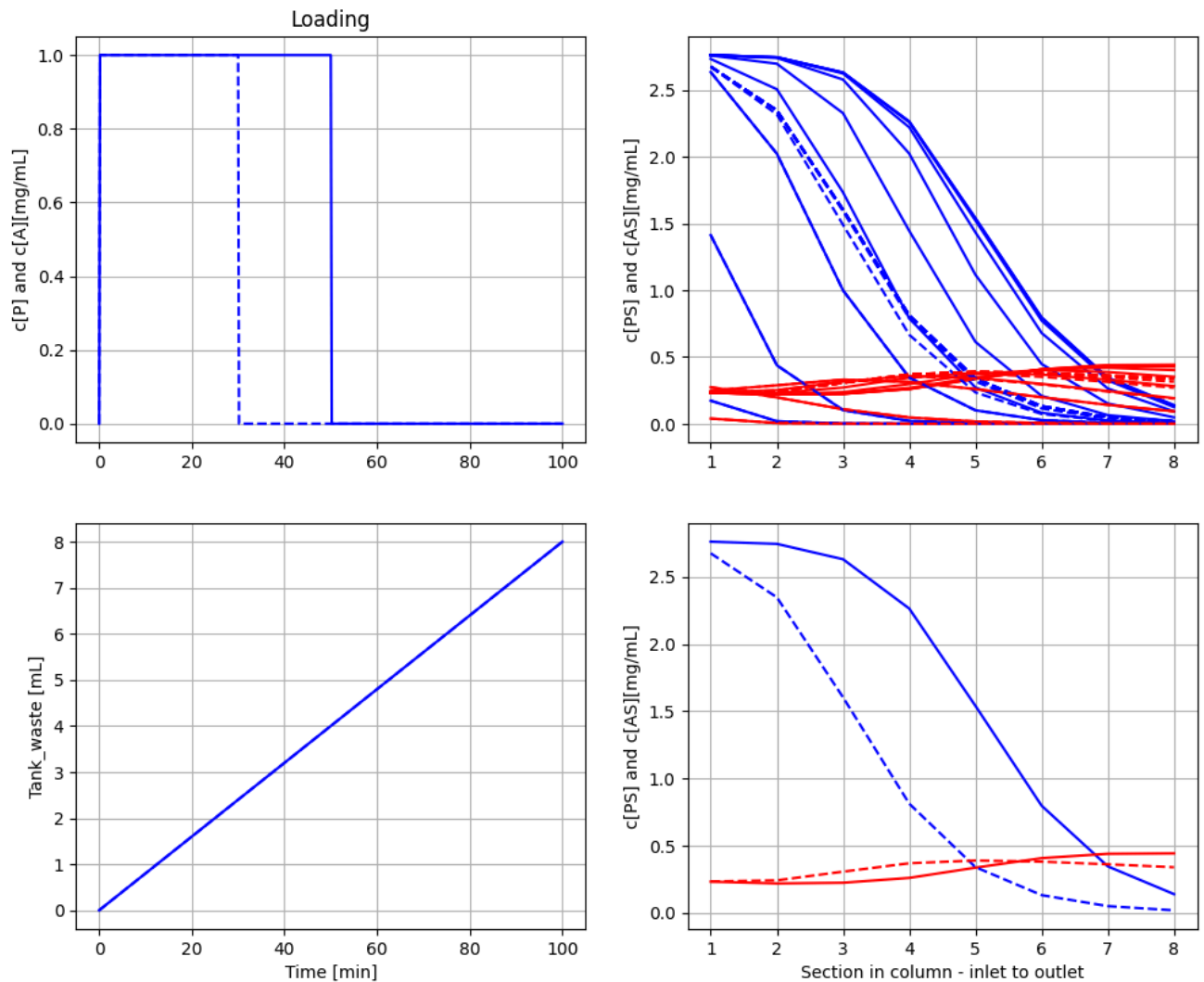
```
model_get('column.column_section[1].V_m')
```

⇒ 0.30029463158052405

```
# Impact of shorter time for loading and then less material  
newplot(title='Loading', plotType='Loading-combined')  
show()
```

```
# Simulation with changed parameter t2  
par(stop_adsorption=30); simu(100)
```

```
# Reset changed parameter  
par(stop_adsorption=50)
```

To the left the inlet loading over time. To the right upper concentration along the column at different times and in steady states finally To the right lower concentrations along the column in steady state.

We see that a shorter time and then less material makes less of the column capacity used.

Note that the flow through the column is constant despite change from sample to just buffer 1, and shown in how the volume of the waste tank increase with time.

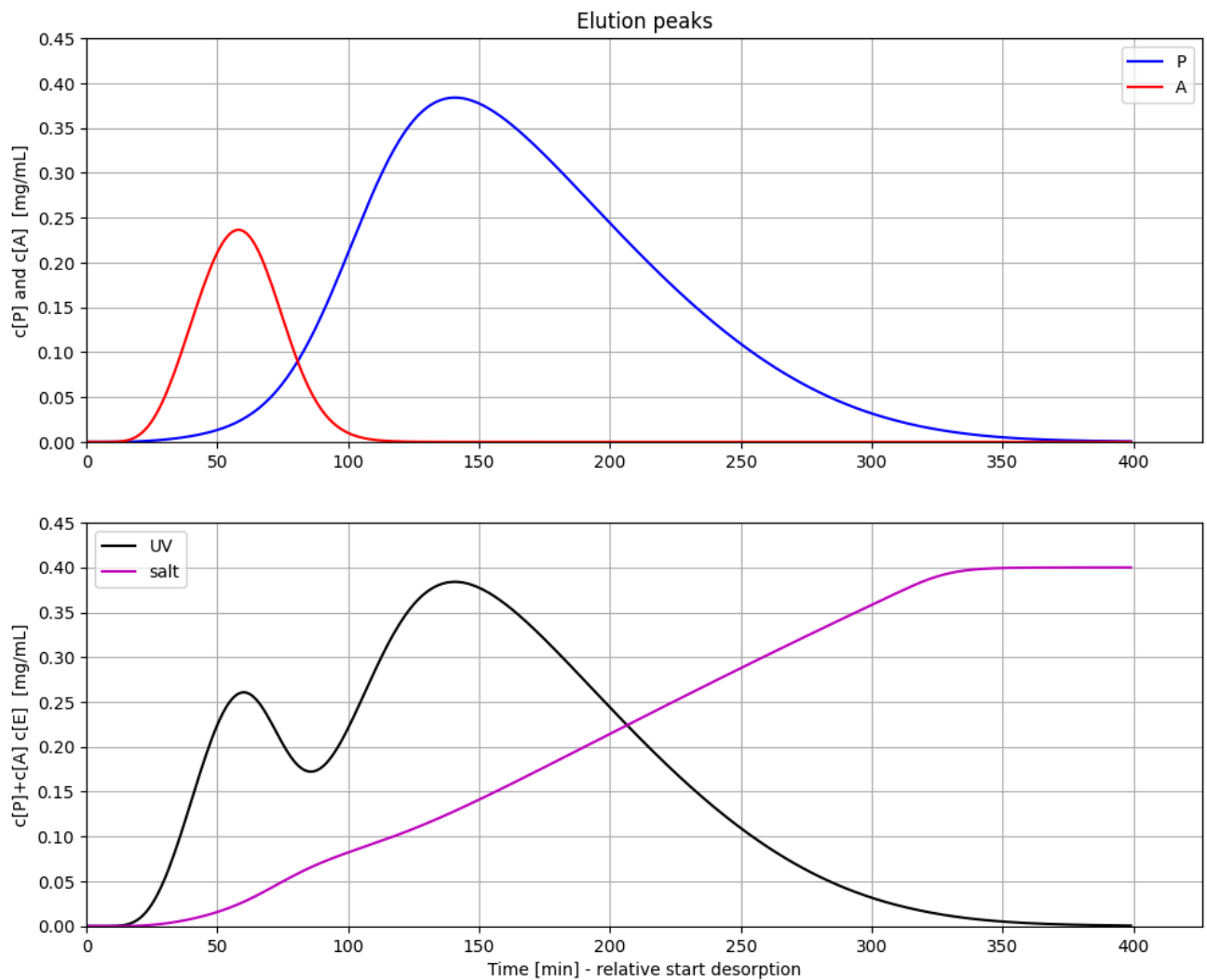
✓ Elution or desorption

```
# Elution of the column
newplot(title='Elution peaks', plotType='Elution')

# Sample
par(P_in=1, A_in=1.0, E_in=0)

# Operation
par(E_in_desorption_buffer=8)
par(LFR=12.0, start_adsorption=0, stop_adsorption=50, start_desorption=150, stati

# Simulation
simu(550)
```



The results are the same as Figure 14 in [1].

The upper diagram shows the column outlet concentrations of P and A over time.

The lower diagram shows the sum (or possibly the UV signal) at column outlet as well as the salt concentration. We have some separation between the two peaks.

Note that the salt concentration deviates slightly from the linear increase between time 50 to 100. This is due to ion interaction with P and A in the column. This phenomenon can also be seen

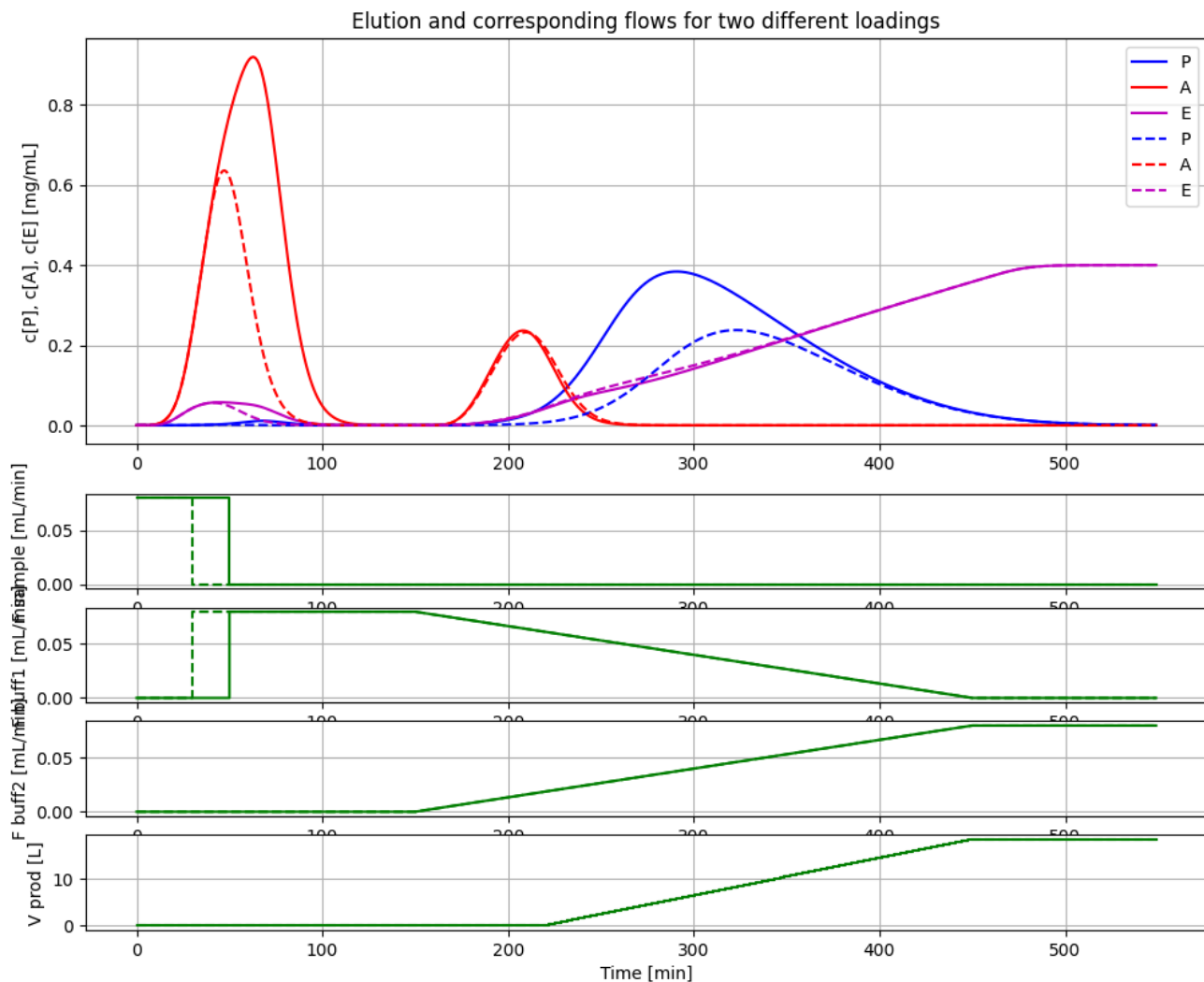
in real data. The ion-salt concentration is scaled with factor 0.05 to get comparable concentrations to P and A.

I have here simulated time 150 of adsorption and then started elution. Here is time counted as zero at time of start of elutions. Not sure how long Jonas simulated to get steady state before he did elution.

```
# More complete visualization of the elution phase and the different flows
newplot(title='Elution and corresponding flows for two different loadings', plotT
par(stop_adsorption=50); simu(550)

# Simulation with changed parameter t2
par(stop_adsorption=30); simu(550)

# Reset changed parameter
par(stop_adsorption=50)
```



Here a diagram that shows the peaks at the outlet as shown in the previous diagram. Below the flow rates of the three different sources. Here time is 0 at start of adsorption and elution starts at time 150.

Automatic pooling based on UV-measurement is tested in another notebook.

✓ Change of resin properties

```

# Elution of the column
newplot(title='Elution peaks for different resin parameters k1 and k2', plotType=

# Sample
par(P_in=1, A_in=0.0, E_in=0)

# Operation
par(E_in_desorption_buffer=8)
par(LFR=12.0, start_adsorption=0, stop_adsorption=50, start_desorption=150, stati

# Simulations
par(k1=0.05, k2=0.50); simu(550)
par(k1=0.05, k2=0.25); simu(550)
par(k1=0.05, k2=0.05); simu(550)
par(k1=0.25, k2=0.05); simu(550)
par(k1=0.50, k2=0.05); simu(550)

# Adjust diagrams
ax1.set_ylim([0, 0.65])
ax2.set_ylim([0, 0.65])
plt.show()

```

