# BPL\_IEC\_validation script with FMPy

The key library FMPy is installed.

After the installation a small application BPL\_IEC\_validation is loaded and run. You can continue with this example if you like.

!lsb\_release -a # Actual VM Ubuntu version used by Google No LSB modules are available. Distributor ID: Ubuntu Description: Ubuntu 22.04.3 LTS Release: 22.04 Codename: jammy %env PYTH0NPATH= → env: PYTH0NPATH= !wget https://repo.anaconda.com/miniconda/Miniconda3-py312\_24.3.0-0-Linux-x86\_64. !chmod +x Miniconda3-py312\_24.3.0-0-Linux-x86\_64.sh !bash ./Miniconda3-py312 24.3.0-0-Linux-x86 64.sh -b -f -p /usr/local import sys sys.path.append('/usr/local/lib/python3.12/site-packages/') → --2024-05-21 07:02:13-- https://repo.anaconda.com/miniconda/Miniconda3-py312 Resolving repo.anaconda.com (repo.anaconda.com)... 104.16.191.158, 104.16.32. Connecting to repo.anaconda.com (repo.anaconda.com)|104.16.191.158|:443... com HTTP request sent, awaiting response... 200 OK Length: 143351488 (137M) [application/octet-stream] Saving to: 'Miniconda3-py312\_24.3.0-0-Linux-x86\_64.sh' Miniconda3-py312\_24 100%[===========] 136.71M 187MB/s in 0.7s 2024-05-21 07:02:14 (187 MB/s) - 'Miniconda3-py312\_24.3.0-0-Linux-x86\_64.sh' : PREFIX=/usr/local Unpacking payload ... Installing base environment... Preparing transaction: ...working... done Executing transaction: ...working... done installation finished. !conda update -n base -c defaults conda --yes → Channels: - defaults Platform: linux-64

Collecting package metadata (repodata.json): done

Solving environment: done

## Package Plan ##

environment location: /usr/local

added / updated specs:

- conda

The following packages will be downloaded:

package	build	
conda-24.5.0 frozendict-2.4.2 openssl-3.0.13	py312h06a4308_0 py312h06a4308_0 py312h06a4308_0 h7f8727e_1	1.2 MB 36 KB 5.2 MB
	 Total:	6.5 MB

The following NEW packages will be INSTALLED:

frozendict pkgs/main/linux-64::frozendict-2.4.2-py312h06a4308\_0

The following packages will be UPDATED:

conda 24.3.0-py312h06a4308\_0 --> 24.5.0-py312h0 openssl 3.0.13-h7f8727e\_0 --> 3.0.13-h7f872

Downloading and Extracting Packages:

openssl-3.0.13 | 5.2 MB | : 0% 0/1 [00:00<?, ?it/s] conda-24.5.0 | 1.2 MB | : 0% 0/1 [00:00<?, ?it/s]

openssl-3.0.13 | 5.2 MB | : 0% 0.002997347135570501/1 [00:00<01:27

frozendict-2.4.2 | 36 KB | : 44% 0.43853215920344746/1 [00:00<00:00, conda-24.5.0 | 1.2 MB | : 1% 0.01293349794914382/1 [00:00<00:20,

openssl-3.0.13 | 5.2 MB | : 100% 1.0/1 [00:00<00:00, 2.59it/s] conda-24.5.0 | 1.2 MB | : 100% 1.0/1 [00:00<00:00, 1.25it/s]

Preparing transaction: done Verifying transaction: done Executing transaction: done

!conda --version
!python --version

→ conda 24.5.0 Python 3.12.2

!conda install -c conda-forge fmpy --yes # Install the key package



Preparing transaction: done Verifying transaction: done Executing transaction: done #!conda install matplotlib --yes

Now specific installation and the run simulations. Start with connecting to Github. Then upload the two files:

- FMU BPL\_IEC\_operation\_linux\_om\_me.fmu
- Setup-file BPL\_IEC\_operation\_fmpy\_explore.py

```
%%bash
git clone https://github.com/janpeter19/BPL_IEC_validation

Cloning into 'BPL_IEC_validation'...

%cd BPL_IEC_validation

/content/BPL_IEC_validation
```

#### BPL IEC validation

Author: Jan Peter Axelsson

Here I try to reproduce somre results from Jonas Månssons master thesis report "Control of chromatography comlumn in production scale", TFRT-5599.

The interaction withe the model has changed slightly so that linear flow rate LF = F/area is used. The column volume is now specified in terms of (cross-section) area times height. Thus by changing area the scale up works fine and what you do in practice. Further the time unit changed from seconds to minutes.

The model has 5 states as listed below, 3 states in liquid mobile phase and 2 states for the gel for bound proteins. In a few of Jonas figures also bound ions of the buffer are plotted but I do not do that. Bound ions of the buffer can be calculated from the difference of total binding capaciety  $Q_{av}$  and bound protein and bound protein antagonist, see section 5.1 in the report.

The molecular weights listed below are not used in the simulations. They just give typical values and the molecular weight for bound protein and antagonist protein is just arbitrary here.

Parameters of the model in general as well as time scale are all arbitrary and focus is on qualitative aspects of the model.

The height of the column is 20 cm which is a common size in the industry.

```
run −i BPL_IEC_fmpy_explore.py

Linux − run FMU pre-comiled OpenModelica 1.23.0-dev

Model for bioreactor has been setup. Key commands:

- par() - change of parameters and initial values
```

```
- init() - change initial values only
```

- simu() simulate and plot
   newplot() make a new plot
- show() show plot from previous simulation
- disp()
   display parameters and initial values from the last simulation
   describe()
   describe culture, broth, parameters, variables with values/un

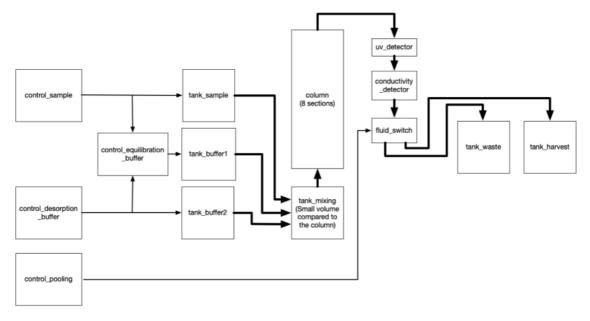
Note that both disp() and describe() takes values from the last simulation and the command process\_diagram() brings up the main configuration

Brief information about a command by help(), eg help(simu) Key system information is listed with the command system\_info()

```
plt.rcParams['figure.figsize'] = [30/2.54, 24/2.54]
```

# The process diagram is made outside Modelica to illustrate the configuration process\_diagram()

No processDiagram.png file in the FMU, but try the file on disk.



describe('chromatography'); print(); #describe('liquidphase')

Ion exchange chromatorgraphy controlled with varying salt-concentration. The

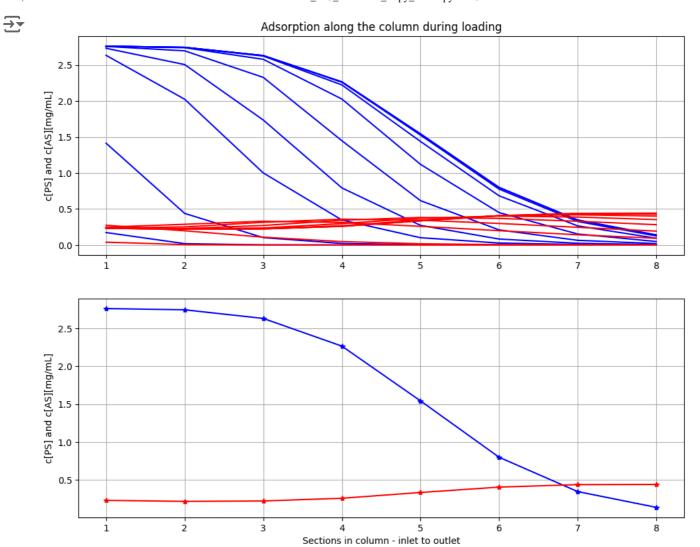
### Loading or adsorption

The parameter notation and values are the same as in the referred report. However the flow rate is here denoted F while q in the report. The column is diveded in n=8 sections and set at compilation time. The values are arbitrarly chosen in the report and the focus is on qualitative aspects of the model.

The simplified model describe only the column in terms of volume and does not distinguish a high column with a small diameter from a lower with larger diameter.

The parameters k1, k2, k3, k4 and Q\_av are given relative volume and with increased column volume a larger capacity is thus obtained.

```
# Loading of the column - try to reproduce Jonas figure 13.
newplot(title='Adsorption along the column during loading', plotType='Loading')
# Sample
par(P_in=1.0, A_in=1.0, E_in=0)
# Column properties
par(k1=0.3, k2=0.05, k3=0.05, k4=0.3, Q_av=3.0)
par(height=20, diameter=0.714)
par(x m=0.3)
# Operation
par(E_in_desorption_buffer=8)
par(LFR=12)
par(scale_volume=False)
par(start_adsorption=0, stop_adsorption=50)
par(start_desorption=150, stationary_desorption=450)
par(start_pooling=220, stop_pooling=450)
# Simulation
simu(100)
```

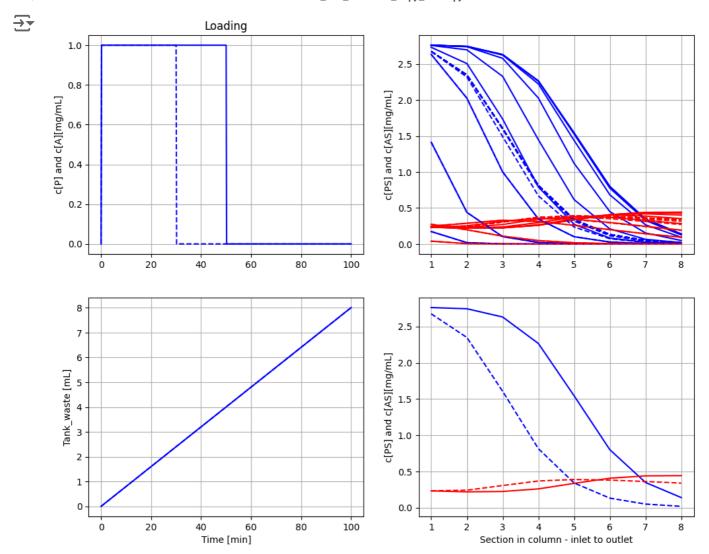


The results are the same af Figure 13 in [1].

# We just check that we had the same volume flow rate as Jonas describe('F')

→ Column ackumulated volume flow : 0.08 [ mL ]

describe('V')



To the left the inlet loading over time. To the right upper concentration along the column at different times and in steady states finally To the right lower concentrations along the column in steady state.

We see that a shorter time and then less material makes less of the column capacity used.

Note that the flow through the column is constant despite change from sample to just buffer 1, and shown in how the volume of the waste tank increase with time.

## Elution or desorption

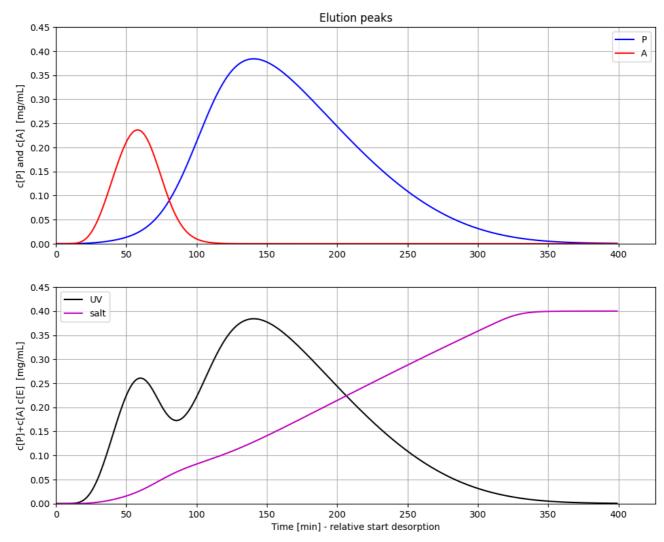
```
# Elution of the column
newplot(title='Elution peaks', plotType='Elution')

# Sample
par(P_in=1, A_in=1.0, E_in=0)

# Operation
par(E_in_desorption_buffer=8)
par(LFR=12.0, start_adsorption=0, stop_adsorption=50, start_desorption=150, stati

# Simulation
simu(550)
```





The results are the same af Figure 14 in [1].

The upper diagrams shows the column outlet concentrations of P and A over time.

The lower diagram shows the sum (or possibly the UV signal) at column outlet as well as the salt concentration. We have some separation between the two peaks.

Note that the salt concentration deviates slightly from the linear increse between time 50 to 100. This is due to ion interaction with P and A in the column. This is phenomenon can also be seen

in real data. The ion-salt concentration is scaled with factor 0.05 to get comparable concentrations to P and A.

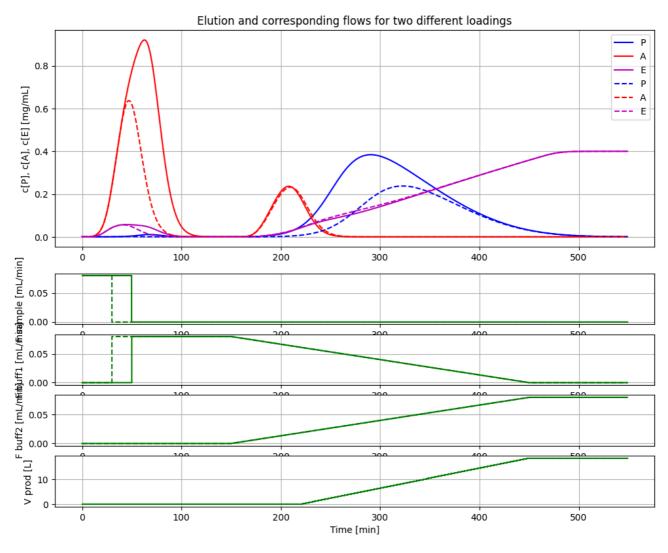
I have here simulated time 150 of adsorbtion and then started elution. Here is time counted as zero at time of start of elutions. Not sure how long Jonas simulated to get steady state before he did elution.

# More complete visualization of the elution phase and the different flows newplot(title='Elution and corresponding flows for two different loadings', plotT par(stop\_adsorption=50); simu(550)

# Simulation with changed parameter t2
par(stop\_adsorption=30); simu(550)

# Reset changed parameter
par(stop\_adsorption=50)





Here a diagram that shows the peaks at the outlet as shown in the previous diagram. Below the flow rates of the three different sorces. Here time is 0 at start of adsorbtion and elution starts at time 150.

Automatic pooling based on UV-mmeasurement is tested in another notebook.

### Change of resin properties

```
# Elution of the column
newplot(title='Elution peaks for different resin parameters k1 and k2', plotType=
# Sample
par(P_in=1, A_in=0.0, E_in=0)
# Operation
par(E_in_desorption_buffer=8)
par(LFR=12.0, start_adsorption=0, stop_adsorption=50, start_desorption=150, stati
# Simulations
par(k1=0.05, k2=0.50); simu(550)
par(k1=0.05, k2=0.25); simu(550)
par(k1=0.05, k2=0.05); simu(550)
par(k1=0.25, k2=0.05); simu(550)
par(k1=0.50, k2=0.05); simu(550)
# Adjust diagrams
ax1.set_ylim([0, 0.65])
ax2.set_ylim([0, 0.65])
plt.show()
→
```

Elution pople for different racin parameters 11 and 12