# BPL_TEST2_Batch_design_space script with PyFMI ver 2.7.4

The key library PyFMI v2.7.4 is installed and downgrading is done Numpy v1.19.1. To simplify this we first install conda.

After the installation a small application BPL_TEST2_Batch_design_space is loaded and run. You can continue with this example if you like.

```
!lsb_release -a # Actual VM Ubuntu version used by Google
```

```
No LSB modules are available.
Distributor ID: Ubuntu
Description:    Ubuntu 18.04.6 LTS
Release:        18.04
Codename:       bionic
```

```
%env PYTHONPATH=
```

```
env: PYTHONPATH=
```

```
!wget https://repo.anaconda.com/miniconda/Miniconda3-py37_4.12.0-Linux-x86_64.sh
!chmod +x Miniconda3-py37_4.12.0-Linux-x86_64.sh
!bash ./Miniconda3-py37_4.12.0-Linux-x86_64.sh -b -f -p /usr/local
import sys
sys.path.append('/usr/local/lib/python3.7/site-packages/')
```

```
Package ruamel_yaml conflicts for:
ruamel_yaml==0.15.100=py37h27cfd23_0
conda==4.12.0=py37h06a4308_0 -> ruamel_yaml[version='>=0.11.14,<0.17']

Package yaml conflicts for:
conda==4.12.0=py37h06a4308_0 -> ruamel_yaml[version='>=0.11.14,<0.17'] -> yaml
ruamel_yaml==0.15.100=py37h27cfd23_0 -> yaml[version='>=0.2.5,<0.3.0a0']
yaml==0.2.5=h7b6447c_0

Package colorama conflicts for:
conda-package-handling==1.8.1=py37h7f8727e_0 -> tqdm -> colorama
tqdm==4.63.0=pyhd3eb1b0_0 -> colorama
colorama==0.4.4=pyhd3eb1b0_0

Package lxml conflicts for:
pyfmi -> lxml
lxml

Package fmilib conflicts for:
pyfmi -> fmilib[version='>=2.2,<2.3.0a0']
fmilib

Package requests conflicts for:
conda==4.12.0=py37h06a4308_0 -> requests[version='>=2.18.4,<3']
```

```
conda==4.12.0=py37h06a4308_0 -> requests[version='>=2.18.4,<3']
requests==2.27.1=pyhd3eb1b0_0

Package gmp conflicts for:
suitesparse -> mpfr[version='>=4.1.0,<5.0a0'] -> gmp[version='>=6.2.1,<7.0a0']
mpfr -> gmp[version='>=6.2.1,<7.0a0']
gmpThe following specifications were found to be incompatible with your system

  - feature:/linux-64::__glibc==2.27=0
  - feature:|@/linux-64::__glibc==2.27=0
  - brotlipy==0.7.0=py37h27cfd23_1003 -> libgcc-ng[version='>=7.3.0'] -> __gli
  - cffi==1.15.0=py37hd667e15_1 -> libgcc-ng[version='>=7.5.0'] -> __glibc[ver
  - conda-package-handling==1.8.1=py37h7f8727e_0 -> libgcc-ng[version='>=7.5.0
  - cryptography==36.0.0=py37h9ce1e76_0 -> libgcc-ng -> __glibc[version='>=2.1
  - fmilib -> libgcc-ng[version='>=7.5.0'] -> __glibc[version='>=2.17']
  - gmp -> libgcc-ng[version='>=7.5.0'] -> __glibc[version='>=2.17']
  - libffi==3.3=he6710b0_2 -> libgcc-ng[version='>=7.3.0'] -> __glibc[version=
  - libgcc-ng==9.3.0=h5101ec6_17 -> __glibc[version='>=2.17']
  - libstdcxx-ng==9.3.0=hd4cf53a_17 -> __glibc[version='>=2.17']
  - libxml2 -> libgcc-ng[version='>=9.3.0'] -> __glibc[version='>=2.17']
  - libxslt -> libgcc-ng[version='>=9.3.0'] -> __glibc[version='>=2.17']
  - metis -> libgcc-ng[version='>=7.5.0'] -> __glibc[version='>=2.17']
  - mpfr -> libgcc-ng[version='>=7.5.0'] -> __glibc[version='>=2.17']
  - ncurses==6.3=h7f8727e_2 -> libgcc-ng[version='>=7.5.0'] -> __glibc[version
  - numpy -> libgcc-ng[version='>=7.3.0'] -> __glibc[version='>=2.17']
  - openssl==1.1.1n=h7f8727e_0 -> libgcc-ng[version='>=7.5.0'] -> __glibc[vers
  - pycosat==0.6.3=py37h27cfd23_0 -> libgcc-ng[version='>=7.3.0'] -> __glibc[v
  - pyfmi -> libgcc-ng[version='>=7.5.0'] -> __glibc[version='>=2.17']
  - python==3.7.13=h12debd9_0 -> libgcc-ng[version='>=7.5.0'] -> __glibc[versi
  - readline==8.1.2=h7f8727e_1 -> libgcc-ng[version='>=7.5.0'] -> __glibc[vers
  - ruamel_yaml==0.15.100=py37h27cfd23_0 -> libgcc-ng[version='>=7.3.0'] -> __
  - sqlite==3.38.2=hc218d9a_0 -> libgcc-ng[version='>=7.5.0'] -> __glibc[versi
  - tk==8.6.11=h1ccaba5_0 -> libgcc-ng[version='>=7.5.0'] -> __glibc[version='
  - yaml==0.2.5=h7b6447c_0 -> libgcc-ng[version='>=7.3.0'] -> __glibc[version=
```

```
!conda update -n base -c defaults conda --yes
```

```
Collecting package metadata (current_repodata.json): done
Solving environment: done

## Package Plan ##

  environment location: /usr/local

  added / updated specs:
    - conda


The following packages will be UPDATED:

  openssl            conda-forge::openssl-1.1.1o-h166bdaf_0 --> pkgs/main::ope

The following packages will be SUPERSEDED by a higher-priority channel:

  ca-certificates    conda-forge::ca-certificates-2022.9.2~ --> pkgs/main::ca-
  certifi            conda-forge/noarch::certifi-2022.9.24~ --> pkgs/main/linu
  conda              conda-forge::conda-22.9.0-py37h89c186~ --> pkgs/main::con


Preparing transaction: done
```

```
     Verifying transaction: done
     Executing transaction: done
     Retrieving notices: ...working... done


!conda --version
!python --version

     conda 22.9.0
     Python 3.7.13


!conda install -c conda-forge pyfmi==2.7.4 --yes # Install the key package

     Collecting package metadata (current_repodata.json): done
     Solving environment: done

     ## Package Plan ##

       environment location: /usr/local

       added / updated specs:
         - pyfmi==2.7.4


     The following packages will be UPDATED:

       ca-certificates    pkgs/main::ca-certificates-2022.07.19~ --> conda-forge::c
       conda              pkgs/main::conda-22.9.0-py37h06a4308_0 --> conda-forge::c

     The following packages will be SUPERSEDED by a higher-priority channel:

       certifi            pkgs/main/linux-64::certifi-2022.9.24~ --> conda-forge/nc


     Preparing transaction: done
     Verifying transaction: done
     Executing transaction: done
     Retrieving notices: ...working... done


!conda install numpy=1.19.1 --yes # Need to downgrade numpy

     Collecting package metadata (current_repodata.json): done
     Solving environment: failed with initial frozen solve. Retrying with flexible
     Collecting package metadata (repodata.json): done
     Solving environment: done

     ## Package Plan ##

       environment location: /usr/local

       added / updated specs:
         - numpy=1.19.1


     The following packages will be downloaded:

         package                    |            build
         ---------------------------|-----------------
```

```
    blas-1.0                    |         openblas          46 KB
    ------------------------------------------------------------
                                           Total:          46 KB

The following NEW packages will be INSTALLED:

  blas               pkgs/main/linux-64::blas-1.0-openblas None
  numpy-base         pkgs/main/linux-64::numpy-base-1.19.1-py37h75fe3a5_0 None

The following packages will be SUPERSEDED by a higher-priority channel:

  ca-certificates    conda-forge::ca-certificates-2022.9.2~ --> pkgs/main::ca-
  certifi            conda-forge/noarch::certifi-2022.9.24~ --> pkgs/main/linu
  conda              conda-forge::conda-22.9.0-py37h89c186~ --> pkgs/main::con
  numpy              conda-forge::numpy-1.21.6-py37h976b52~ --> pkgs/main::num



Downloading and Extracting Packages
blas-1.0             | 46 KB     | : 100% 1.0/1 [00:00<00:00, 10.95it/s]

ChecksumMismatchError: Conda detected a mismatch between the expected content
for url 'https://repo.anaconda.com/pkgs/main/linux-64/blas-1.0-openblas.conda'
  download saved to: /usr/local/pkgs/blas-1.0-openblas.conda
  expected sha256: c85b5d0a336b5be0f415c71fd7fe2eca59e09f42221bfa684aafef5510b
  actual sha256: 5dc5483db0d9785b19e021cee418a8ee03e0ff0e5ebd0b75af4927746604e
```

Now specific installation and the run simulations. Start with connecting to Github. Then upload the four files:

- FMU - BPL_TEST2_Batch_design_space_no_noise_linux_jm_cs.fmu
- Setup-file - BPL_TEST2_Batch_design_space_no_noise_explore.py
- FMU - BPL_TEST2_Batch_design_space_with_noise_linux_jm_cs.fmu
- Setup-file - BPL_TEST2_Batch_design_space_with_noise_explore.py

```
# Filter out DepracationWarnings for 'np.float as alias' is needed - wish I could m
import warnings
warnings.filterwarnings("ignore")
```

```
%%bash
git clone https://github.com/janpeter19/BPL_TEST2_Batch_design_space
```

```
Cloning into 'BPL_TEST2_Batch_design_space'...
```

```
%cd BPL_TEST2_Batch_design_space
```

```
/content/BPL_TEST2_Batch_design_space/BPL_TEST2_Batch_design_space
```

# BPL_TEST2_Batch_design_space - demo

In this notebook the design space for a batch cultivation process is determined and visualized. The example is kept as simple as possible. The culture grow on a substrate S and the cell conentration X inrease until the substrate is consumed. We study the problem first without any measurement noise and then later with measurement noise and use one separate FMU for each.

The end criteria for a batch is here when the subdstrate level has decreased below a certain predefined level and that time is called time_final:

- S < Smin

The evaluation of the batch culture is just in terms of the obtained value of cell concentration at the end in combination with how long time the culture took. The batch is accepted provided the culture fullfil the two requirements:

- X_final > X_final_min
- Time_final < time_final_max

The question is what range of process parameters Y and qSmax that can be allowed to still get accepted batches.

Here we simply use brute force and sweep through a number combinations of process parameters and evaluate by simulation the result for each parameter setting. We get rather clear-cut corners in the process parameter space that result in acceptable batches.

In the later part we introduce substrate measurement error and in this way introduce some uncertainty in the determination of end of batch. The impact of this measurement noise is that the design space get more rounded corners.

The practical experimental approach is usually to just use a few parameter combinations and evaluate these and from that information calculate the design space. Usually "process linearity" assumption is used. The combination of this experimental approach with brute force simulation is discussed in reference [1].

# 1 Batch end detection - no measurement noise

Here we load a system model without noise. Thus detection of end of batch is an event in continuous time.

```
run -i BPL_TEST2_Batch_no_noise_explore.py
```

```
      Linux - run FMU pre-comiled JModelica 2.4

      Model for bioreactor has been setup. Key commands:
       - par()       - change of parameters and initial values
       - init()      - change initial values only
       - simu()      - simulate and plot
       - newplot()   - make a new plot
       - show()      - show plot from previous simulation
```

```
# Adjust the diagram size
%matplotlib inline
plt.rcParams['figure.figsize'] = [30/2.54, 24/2.54]
```

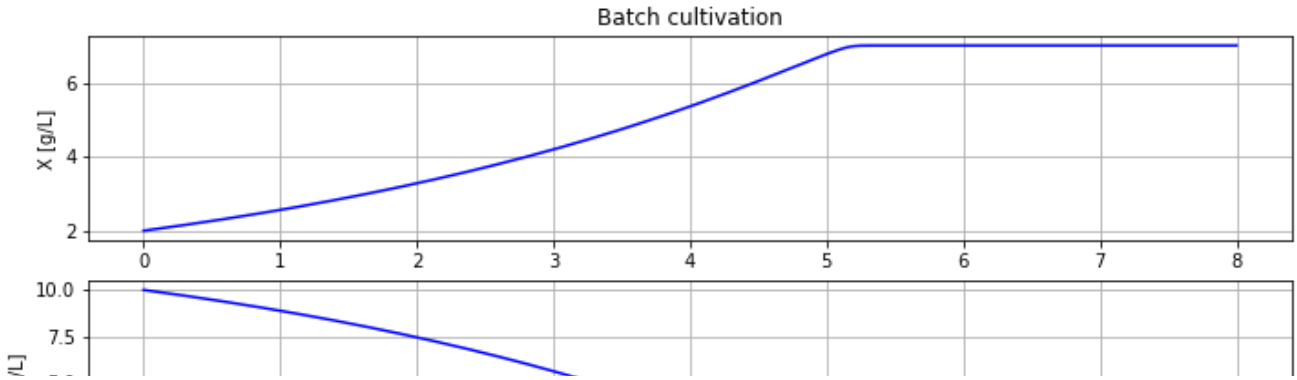
## 1.1 Batch evaluation

The first an example of batch that has an end of batch that fulfills the criteria for acceptance. In the following diagram we see examples of impact of variation on the criteria for acceptance.

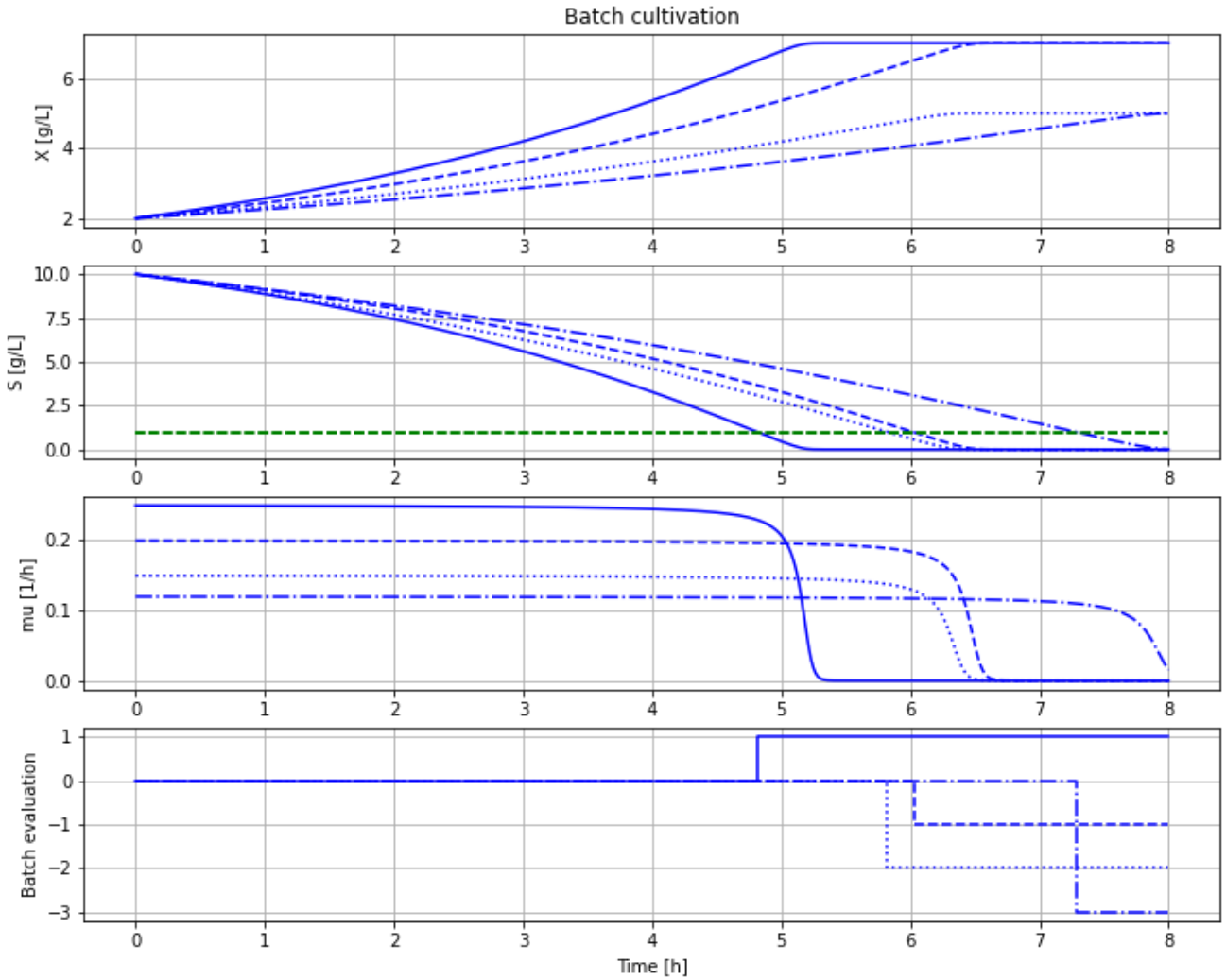
The variable batch_evaluation goes from 0 to either 1 or a negative value when end of batch is detected. A positive value 1 means that the acceptance criteria is fullfilled and a negative value -1, -2 or -3 is obtained if one or more criteria for acceptance is not fullfilled.

```
# Nominal parameters
par(S_min=1.0, time_final_max=6.0, X_final_min=5.0)
init(VX_0=2, VS_0=10)
par(Y=0.5, qSmax=0.5, Ks=0.1)
```

```
# Simulation of nominal parameters that gives a batch that meed the end criteria
newplot(plotType='TimeSeries_2')
simu(8)
```




```
# Exammple of process parameter changes and how they meet the end criteria
newplot(plotType='TimeSeries_2')
par(Y=0.50, qSmax=0.50); simu(8)  # - pass (solid line)
par(Y=0.50, qSmax=0.40); simu(8)  # - fail criteria time_final < 6.0 (dashed line)
par(Y=0.30, qSmax=0.50); simu(8)  # - fail criteria X_final > 5.0 (dotted line)
par(Y=0.30, qSmax=0.40); simu(8)  # - fail both criteria (dash dotted line)
```

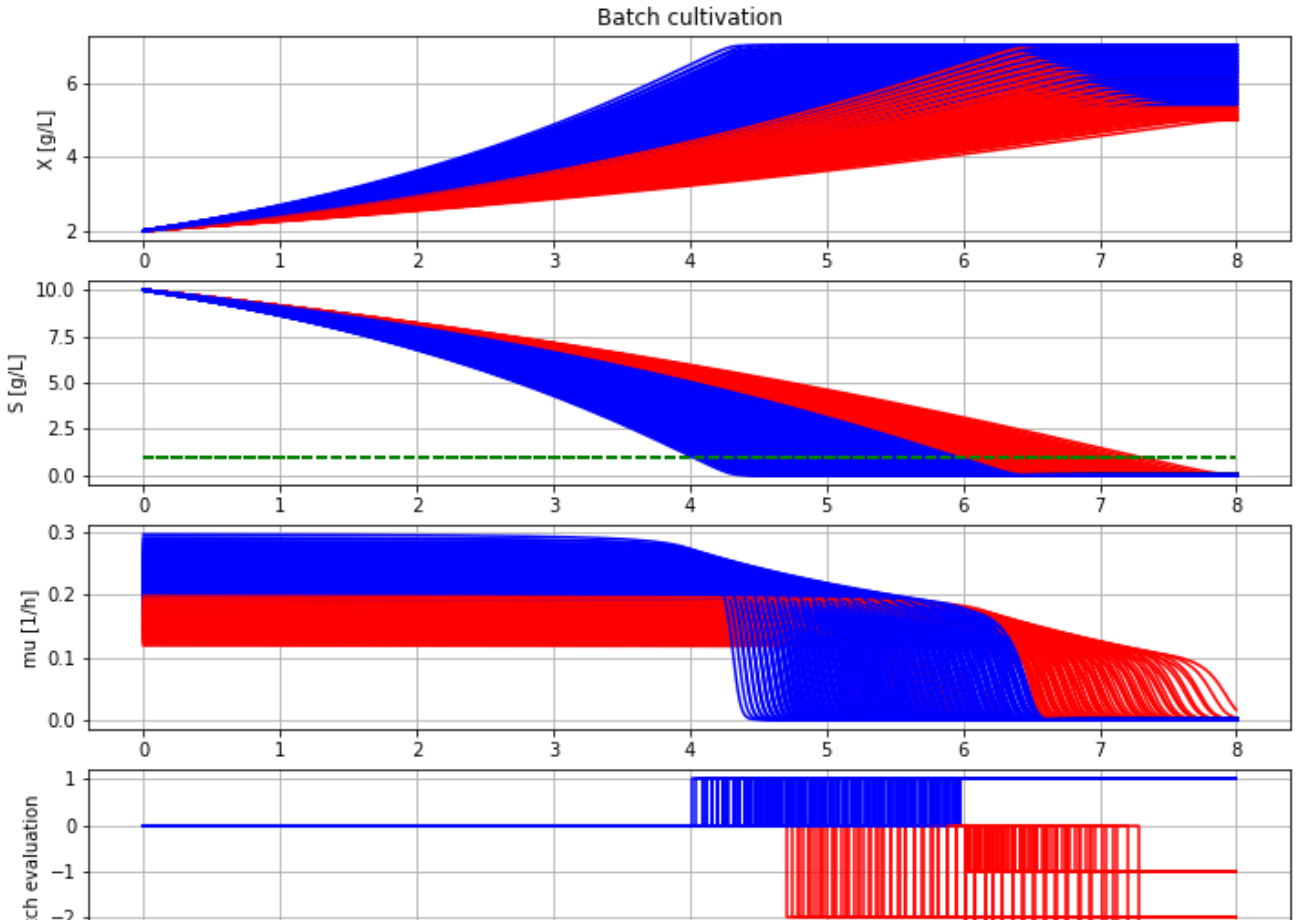



We see that the accepted batch (solid line) finish first. The batches that fail take longer time and two of them has also lower cell concentration at the end.

## 1.2 Batch evaluation under process variation - parameter sweep

Now let us systematically sweep through a number of combinations of process parameters Y and qSmax and evaluate the batches and visualise the result.

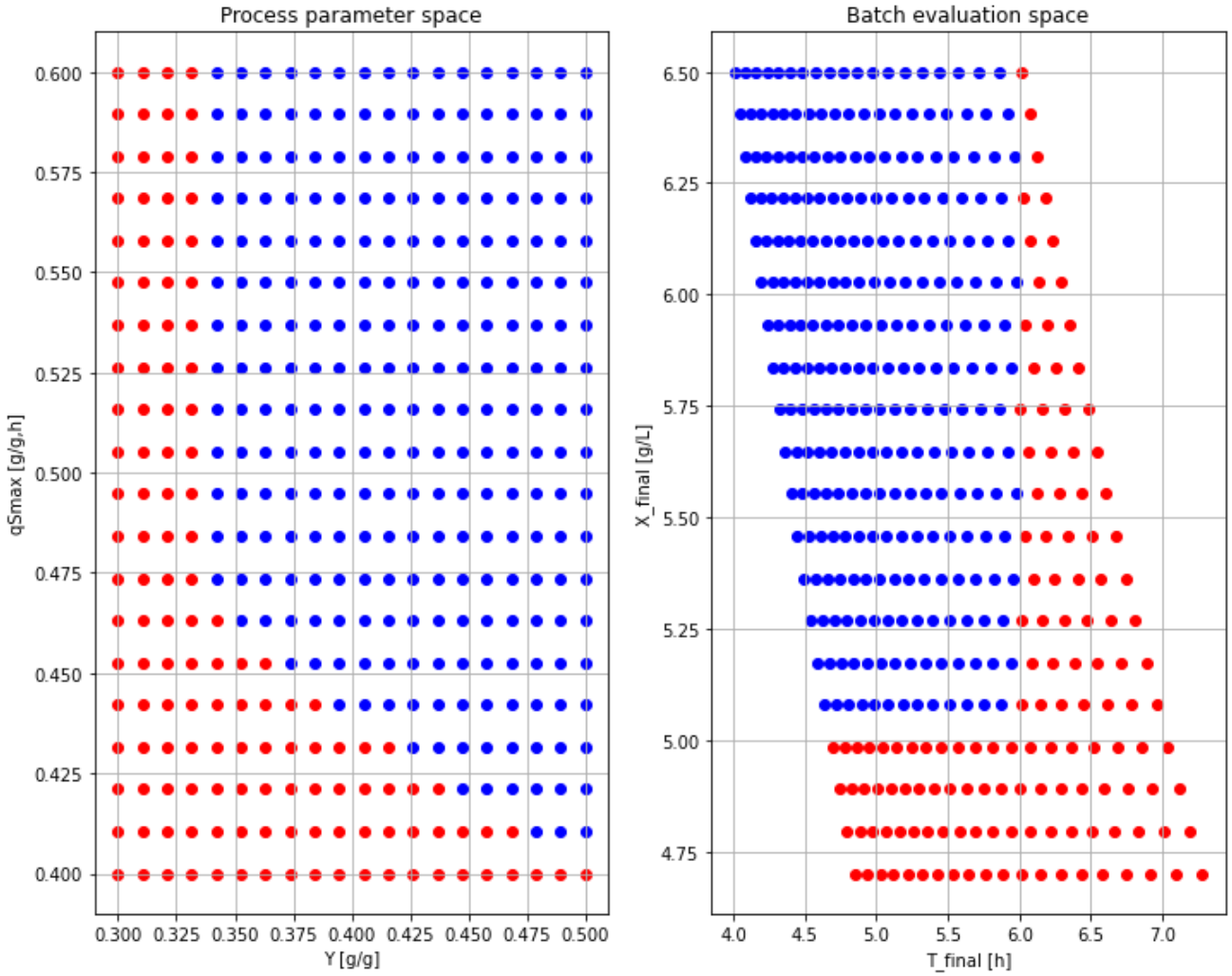
```
# Define sweep ranges and storage of final data
nY = 20
nqSmax = 20
Y_range = np.linspace(0.3,0.5,nY)
qSmax_range = np.linspace(0.4,0.6,nqSmax)
data = np.zeros([nY,nqSmax,5])


# Run parameter sweep - takes a few minutes
newplot(plotType='TimeSeries_2_diagrams')
init(VX_0=2, VS_0=10)

for j in range(nY):
    for k in range(nqSmax):
        par(Y=Y_range[j])
        par(qSmax=qSmax_range[k])
        simu(8)

        # Store final results
        data[j,k,0] = Y_range[j]
        data[j,k,1] = qSmax_range[k]
        data[j,k,2] = sim_res['monitor.time_final'][-1]
        data[j,k,3] = sim_res['monitor.X_final'][-1]
        data[j,k,4] = sim_res['monitor.batch_evaluation'][-1]

        # Plot simulation results
        if sim_res['monitor.batch_evaluation'][-1] > 0:
            ax1.plot(sim_res['time'], sim_res['bioreactor.c[1]'],'b-')
            ax2.plot(sim_res['time'], sim_res['bioreactor.c[2]'],'b-')
            ax2.plot([0, simulationTime], [model.get('monitor.S_min'), model.get('m
            ax3.plot(sim_res['time'], sim_res['bioreactor.culture.q[1]'],'b-')
            ax4.step(sim_res['time'],sim_res['monitor.batch_evaluation'],where='pos
        else:
            ax1.plot(sim_res['time'], sim_res['bioreactor.c[1]'],'r-')
            ax2.plot(sim_res['time'], sim_res['bioreactor.c[2]'],'r-')
            ax2.plot([0, simulationTime], [model.get('monitor.S_min'), model.get('m
            ax3.plot(sim_res['time'], sim_res['bioreactor.culture.q[1]'],'r-')
            ax4.step(sim_res['time'],sim_res['monitor.batch_evaluation'],where='pos

plt.show()
```




Batches represented by blue lines are those that in the end got accepted. The red ones failed.

```
# Show end results
plt.figure()
ax1 = plt.subplot(1,2,1)
ax2 = plt.subplot(1,2,2)

for j in range(nY):
    for k in range(nqSmax):
        if data[j,k,4] > 0:
            ax1.scatter(data[j,k,0],data[j,k,1],c='b')
        else:
            ax1.scatter(data[j,k,0],data[j,k,1],c='r')
ax1.grid()
#plt.axis([0, 0.8, 0, 0.8])
ax1.set_ylabel('qSmax [g/g,h]')
ax1.set_xlabel('Y [g/g]')
ax1.set_title('Process parameter space')

for j in range(nY):
    for k in range(nqSmax):
        if data[j,k,4] > 0:
            ax2.scatter(data[j,k,2],data[j,k,3],c='b')
        else:
            ax2.scatter(data[j,k,2],data[j,k,3],c='r')
ax2.grid()
#plt.axis([0, 8, 0, 8])
ax2.set_xlabel('T_final [h]')
```

```
ax2.set_ylabel('X_final [g/L]')
ax2.set_title('Batch evaluation space')
plt.show()
```




Here we visualize the previous simulations results in a different way with foucse on the end result. Each dot in the left diagram (process parameter space) represent a simulation that give a result in the rigiht diagam (batch evaluation space). The blue dots are those batches that were accepted and the red ones those that failed.

The blue dots in the process parameter space show the "design space" for the acceptance criteria we have.

## 2 Batch end detection - with measurement noise

Here we load a system model with normal noise added to the sampled value of substrate concentration. The measurement of substrate conentration usually has a higher variation than measuremetn of cell concentration and therefore we focus here on the impact on substrate conentrations.

Thus detection of end of batch is now in discrete time with a give samplePeriod (default 0.1 hour). This discreteization also introduce an error in detection of the end point. By changing this sample intervall to shorter values you can see the impact of this error but not done here.

```
run -i BPL_TEST2_Batch_with_noise_explore.py
```

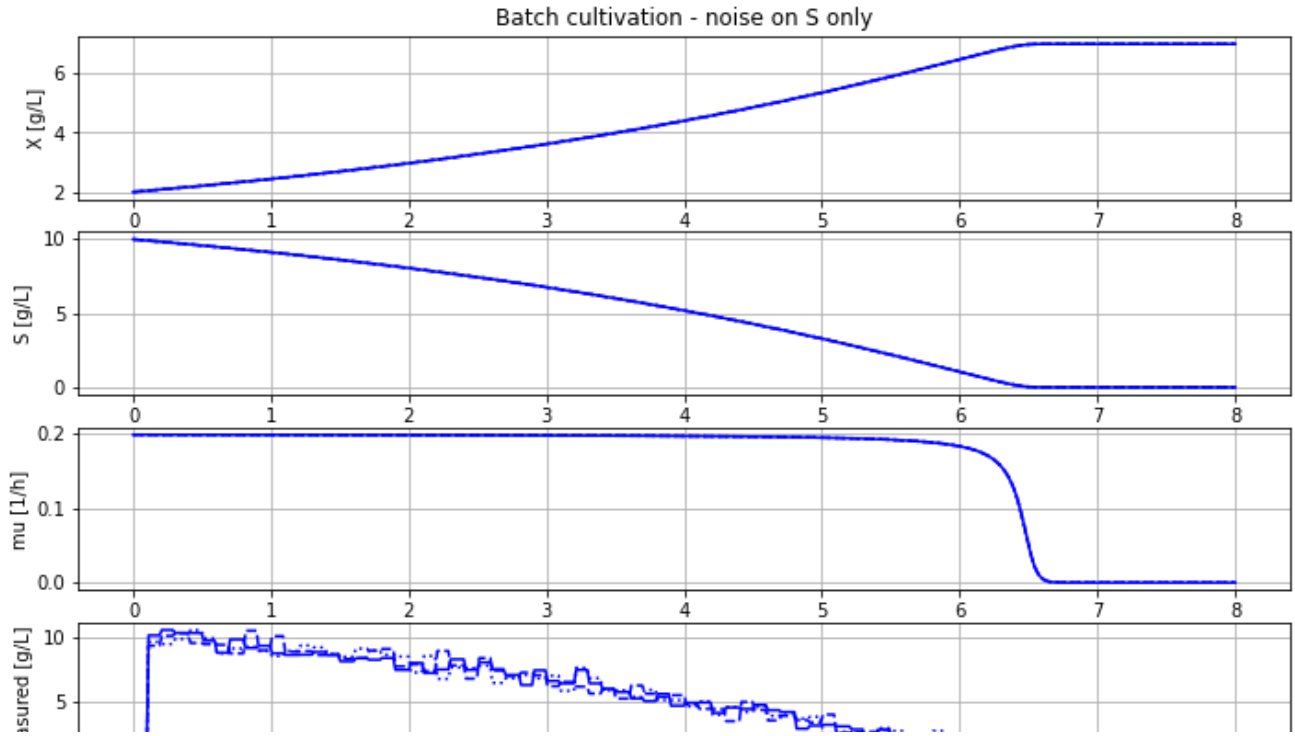
```
    Linux - run FMU pre-comiled JModelica 2.4

    Model for bioreactor has been setup. Key commands:
     - par()       - change of parameters and initial values
     - init()      - change initial values only
     - simu()      - simulate and plot
     - newplot()   - make a new plot
     - show()      - show plot from previous simulation
     - disp()      - display parameters and initial values from the last simulatic
     - describe()  - describe culture, broth, parameters, variables with values /

    Note that both disp() and describe() takes values from the last simulation

    Brief information about a command by help(), eg help(simu)
    Key system information is listed with the command system_info()
    <Figure size 850.394x680.315 with 0 Axes>
```

## 2.1 Batch evaluation under substrate measurement error

Here we see an example of how substrate measurement noise directly affect the evaluation of the batch from accetable to not acceptable.

```
# Nominal parameters
par(S_min=1.0, time_final_max=6.0, X_final_min=5.0)
init(VX_0=2, VS_0=10)
par(Y=0.5, qSmax=0.5, Ks=0.1)
par(sigma=0.48, samplePeriod=0.1)
```

```
# Simulation of nominal parameters that gives a batch that meed the end criteria
newplot(plotType='TimeSeries_2')
par(Y=0.5, qSmax=0.4);
for value in [2,3,5]: par(seed=value); simu(8)
```

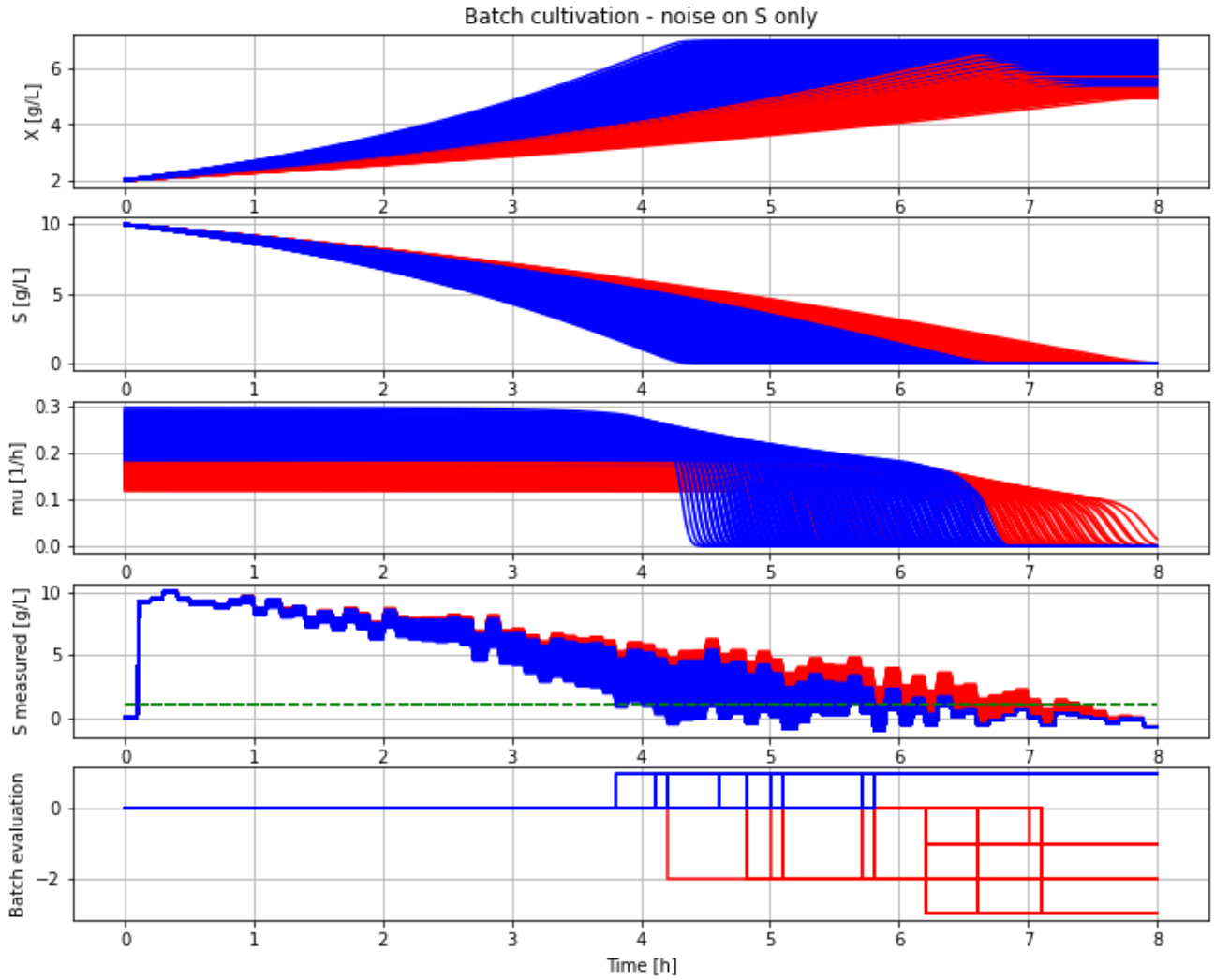



## 2.2 Batch evaluation under process variation and measurement error - parameter sweep

Now let us again systematically sweep through a number of combinations of process parameters Y and qSmax and evaluate the batches and visualise their result.

```
# Define sweep ranges and storage of final data
nY = 20
nqSmax = 20
Y_range = np.linspace(0.3,0.5,nY)
qSmax_range = np.linspace(0.4,0.6,nqSmax)
data = np.zeros([nY,nqSmax,5])


# Run parameter sweep - takes a few minuts
newplot(plotType='TimeSeries_2_diagrams')
par(sigma=0.48, seed=1, samplePeriod=0.1)

for j in range(nY):
    for k in range(nqSmax):
        par(Y=Y_range[j])
        par(qSmax=qSmax_range[k])
        simu(8)

        # Store final results
        data[j,k,0] = Y_range[j]
        data[j,k,1] = qSmax_range[k]
        data[j,k,2] = sim_res['monitor.time_final'][-1]
        data[j,k,3] = sim_res['monitor.X_final'][-1]
        data[j,k,4] = sim_res['monitor.batch_evaluation'][-1]

        # Plot simulation results
```

```
        if sim_res['monitor.batch_evaluation'][-1] > 0:
            ax1.plot(sim_res['time'], sim_res['bioreactor.c[1]'],'b-')
            ax2.plot(sim_res['time'], sim_res['bioreactor.c[2]'],'b-')
            ax3.plot(sim_res['time'], sim_res['bioreactor.culture.q[1]'],'b-')
            ax4.plot(sim_res['time'], sim_res['sensor.out.c[2]'],'b-')
            ax4.plot([0, simulationTime], [model.get('monitor.S_min'), model.get('m
            ax5.step(sim_res['time'],sim_res['monitor.batch_evaluation'],where='pos
        else:
            ax1.plot(sim_res['time'], sim_res['bioreactor.c[1]'],'r-')
            ax2.plot(sim_res['time'], sim_res['bioreactor.c[2]'],'r-')
            ax3.plot(sim_res['time'], sim_res['bioreactor.culture.q[1]'],'r-')
            ax4.plot(sim_res['time'], sim_res['sensor.out.c[2]'],'r-')
            ax4.plot([0, simulationTime], [model.get('monitor.S_min'), model.get('m
            ax5.step(sim_res['time'],sim_res['monitor.batch_evaluation'],where='pos

plt.show()
```




```
# Show end results
plt.figure()
ax1 = plt.subplot(1,2,1)
ax2 = plt.subplot(1,2,2)
```

```
for j in range(nY):
    for k in range(nqSmax):
        if data[j,k,4] > 0:
            ax1.scatter(data[j,k,0],data[j,k,1],c='b')
        else:
            ax1.scatter(data[j,k,0],data[j,k,1],c='r')
ax1.grid()
#plt.axis([0, 0.8, 0, 0.8])
ax1.set_ylabel('qSmax [g/g,h]')
ax1.set_xlabel('Y [g/g]')
ax1.set_title('Process parameter space')

for j in range(nY):
    for k in range(nqSmax):
        if data[j,k,4] > 0:
            ax2.scatter(data[j,k,2],data[j,k,3],c='b')
        else:
            ax2.scatter(data[j,k,2],data[j,k,3],c='r')
ax2.grid()
#plt.axis([0, 8, 0, 8])
ax2.set_xlabel('T_final [h]')
ax2.set_ylabel('X_final [g/L]')
ax2.set_title('Batch evaluation space')
plt.show()
```

We see that we get somwehat different results in the parameter space. The acceptable region with blue dots (design space) get a more rounded corner. The vertical left line is also more rugged.

With much more simulations we could get a better idea of the probablity that a batch is accepted and determine the design space in proabilistic sense.

## 3 Summary

We have worked through a simple example of evaluation of batch cuöture with given acceptance criteria and how that criteria can be translated to acceptable variation in process parameters, i.e. the design space.

In the deterministic case we get a rather clear cut design space.

In the more realistic case with subsrate measurment noise included we get a more complicated design space, but still similar.

The stochastic model introduce erorrs both due to the added normal noise in the substrate concentration, and due to the fact that we use time discrete system for the noise. The impact of the time discrete check when batch has ended can be made smaller by chosing a smaller sample intervall. This was not studied here and is left for the interested reader.

Note...

## References

[1] Axelsson J.P. and A. Elsheikh: “An example of sensitivity analysis of a bioprocess using Bioprocess Library for Modelica”, Proceedings MODPROD, Linköping, Sweden 2019, see presentation here.

## Appendix

```
disp('culture')
```

```
    Y : 0.5
    qSmax : 0.6
    Ks : 0.1
```

```
describe('mu')
```

```
    Cell specific growth rate variable : 0.0 [ 1/h ]
```

```
describe('parts')
```

```
['bioreactor', 'bioreactor.culture', 'liquidphase', 'monitor', 'MSL', 'sensor'
```

```
describe('MSL')
```

```
MSL: 3.2.2 build 3 - used components: Noise.NormalNoise
```

```
system_info()
```

```
System information
 -OS: Linux
 -Python: 3.7.15
 -Scipy: not installed in the notebook
 -PyFMI: 2.7.4
 -FMU by: JModelica.org
 -FMI: 2.0
 -Type: FMUModelCS2
 -Name: BPL_TEST2.BatchWithNoise
 -Generated: 2022-10-18T07:35:35
 -MSL: 3.2.2 build 3
 -Description: Bioprocess Library version 2.1.0
 -Interaction: FMU-explore ver 0.9.5
```