BPL_TEST2_Perfusion script with PyFMI

The key library PyFMI is installed.

After the installation a small application BPL_TEST2_Perfusion is loaded and run. You can continue with this example if you like.

!lsb_release -a # Actual VM Ubuntu version used by Google → No LSB modules are available. Distributor ID: Ubuntu Ubuntu 22.04.4 LTS Description: Release: 22.04 Codename: iammv %env PYTHONPATH= → env: PYTHONPATH= !python --version → Python 3.11.11 !wget https://repo.anaconda.com/miniconda/Miniconda3-py311_24.11.1-0-Linux-x86_64 !chmod +x Miniconda3-py311_24.11.1-0-Linux-x86_64.sh !bash ./Miniconda3-py311_24.11.1-0-Linux-x86_64.sh -b -f -p /usr/local import sys sys.path.append('/usr/local/lib/python3.11/site-packages/') → --2025-02-07 13:47:54-- https://repo.anaconda.com/miniconda/Miniconda3-py311 Resolving repo.anaconda.com (repo.anaconda.com)... 104.16.191.158, 104.16.32. Connecting to repo.anaconda.com (repo.anaconda.com)|104.16.191.158|:443... con HTTP request sent, awaiting response... 200 OK Length: 145900576 (139M) [application/octet-stream] Saving to: 'Miniconda3-py311 24.11.1-0-Linux-x86 64.sh' Miniconda3-py311_24 100%[==========] 139.14M 97.3MB/s in 1.4s 2025-02-07 13:47:56 (97.3 MB/s) - 'Miniconda3-py311_24.11.1-0-Linux-x86_64.sh PREFIX=/usr/local Unpacking payload ... Installing base environment... Preparing transaction: ...working... done Executing transaction: ...working... done installation finished.

!conda update -n base -c defaults conda --yes

→ Channels:

defaults

Platform: linux-64

Collecting package metadata (repodata.json): done

Solving environment: done

Package Plan

environment location: /usr/local

added / updated specs:

- conda

The following packages will be downloaded:

| package | build | |
|--|---------------------------------------|------------------|
| ca-certificates-2024.12.31 certifi-2025.1.31 | h06a4308_0 py311h06a4308_0 | 128 KB 163 KB |
| | Total: | 291 KB |

The following packages will be UPDATED:

ca-certificates
certifi

2024.11.26-h06a4308_0 --> 2024.12.31-h00 2024.8.30-py311h06a4308_0 --> 2025.1.31-py31

Downloading and Extracting Packages:

certifi-2025.1.31 | 163 KB | : 0% 0/1 [00:00<?, ?it/s] ca-certificates-2024 | 128 KB | : 0% 0/1 [00:00<?, ?it/s]

ca-certificates-2024 | 128 KB | : 100% 1.0/1 [00:00<00:00, 17.96it/s]

Preparing transaction: done Verifying transaction: done Executing transaction: done

!conda --version
!python --version

conda 24.11.1 Python 3.11.11

!conda config --set channel_priority strict

!conda install -c conda-forge pyfmi --yes # Install the key package

 $\overline{2}$

Preparing transaction: done Verifying transaction: done Executing transaction: done

Notes of BPL_TEST2_Perfusion

This notebook explore perfusion cultivation in comparison with ordinary continuous cultivation (chemostat) and use comparable settings to earlier notebook. Further you see here examples of

interaction with the simplified commands par(), init(), simu() etc as well as direct interaction with the FMU which is called "model" here. The last simulation is always available in the workspace and called "sim_res". Note that describe() brings mainly up from descriptive information from the Modelica code from the FMU but is complemented by some information given in the Python setup file.

Now specific installation run a simulation and notebook for that Start with connecting to Github. Then upload the two files:

- FMU-BPL TEST2 Perfusion linux om me.fmu
- Setup-file BPL_TEST2_Perfusion_explore.py

```
%%bash
git clone https://github.com/janpeter19/BPL_TEST2_Perfusion
→ Cloning into 'BPL TEST2 Perfusion'...
%cd BPL_TEST2_Perfusion
/content/BPL TEST2 Perfusion
run -i BPL_TEST2_Perfusion_explore.py
→ Linux - run FMU pre-comiled OpenModelica
    Model for bioreactor has been setup. Key commands:
     - par()

    change of parameters and initial values

     - init()

    change initial values only

     - simu()
                   simulate and plot
     - newplot() - make a new plot
     - show()

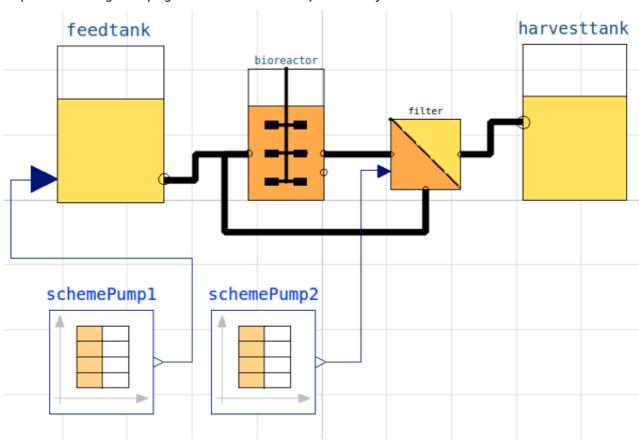
    show plot from previous simulation

     - disp()

    display parameters and initial values from the last simulation

     - describe() - describe culture, broth, parameters, variables with values/ur
    Note that both disp() and describe() takes values from the last simulation
    and the command process_diagram() brings up the main configuration
    Brief information about a command by help(), eg help(simu)
    Key system information is listed with the command system_info()
# Filter out DepracationWarnings for 'np.float as alias' is needed - wish
import warnings
warnings.filterwarnings("ignore")
%matplotlib inline
plt.rcParams['figure.figsize'] = [25/2.54, 20/2.54]
process_diagram()
```

No processDiagram.png file in the FMU, but try the file on disk.



```
# Process parameters used throughout
par(Y=0.5, qSmax=0.75, Ks=0.1)  # Culture
par(filter_eps=0.10, filter_alpha_X=0.02, filter_alpha_S=0.10)  # Filter
par(S_in=30.0)  # Inlet subst
init(V_start=1.0, VX_start=1.0)  # Process ini
eps = parDict['filter_eps']  # Pump schedu
```

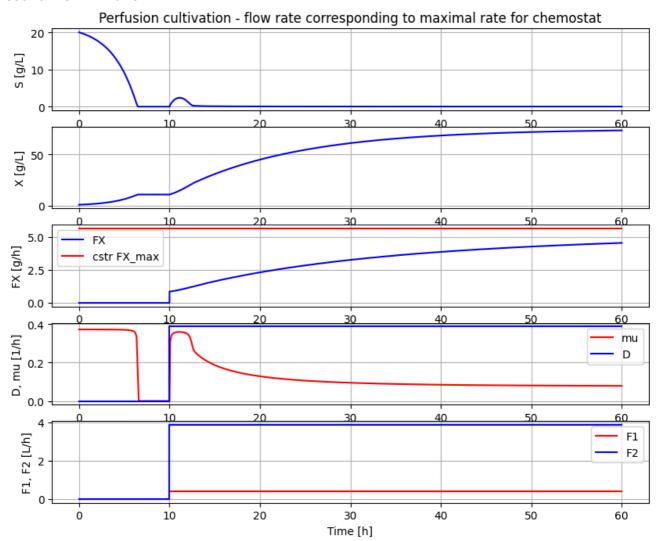
Simulation of process with flow rate close to wash-out for chemostat

newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate fo simu(60)

```
\overline{2}
```

Could not find cannot import name 'dopri5' from 'assimulo.lib' (/usr/local/lil Could not find cannot import name 'rodas' from 'assimulo.lib' (/usr/local/lib, Could not find cannot import name 'odassl' from 'assimulo.lib' (/usr/local/lil Could not find ODEPACK functions. Could not find RADAR5

Could not find GLIMDA.

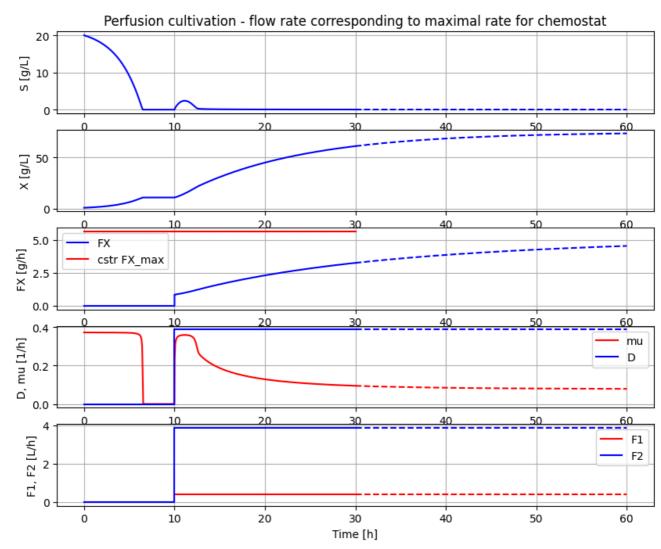


Simulation of process with flow rate close to wash-out for chemostat

```
init(VS_start=20)
                                                         # Process initial
                                                         # Pump schedule - recycl
par(pump1_t1=10, pump2_t1=10)
par(pump1_F1=2.5*0.155, pump2_F1=2.5*0.155/eps)
par(pump1_t2=940, pump2_t2=940, pump1_t3=950, pump2_t3=950, pump1_t4=960, pump2_t
newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate fo
simu(30)
```

simu(30,'cont')





Note the inability of the OpenModelica FMU to handle simu('cont') properly.

```
# Concentration factor of the filter
c=model.get('filter.retentate.c[1]')[0]/model.get('filter.inlet.c[1]')[0]
print('Conc factor of perfusion filter =', np.round(c,3))

→ Conc factor of perfusion filter = 1.179

c_data=sim_res['filter.retentate.c[1]']/sim_res['filter.inlet.c[1]']
print('Conc factor variation', np.round(min(c_data[151:]), 3), np.round(max(c_data[151:]), 3))
```

init(VS_start=150)

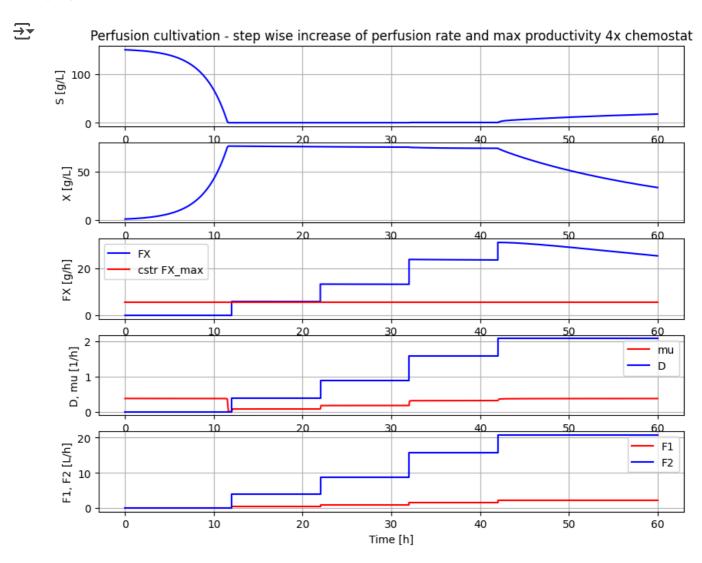
Simulation of process with step-wise increase of pefusion rate until wash-out.
This means that re-circulation rate change at the same time as the perfusion ra

```
par(pump1_t1=12, pump2_t1=12)
par(pump1_F1=2.5*0.155, pump2_F1=2.5*0.155/eps)
par(pump1_t2=22, pump2_t2=22)
par(pump1_F2=2.5*0.35, pump2_F2=2.5*0.35/eps)
par(pump1_t3=32, pump2_t3=32)
par(pump1_F3=2.5*0.63, pump2_F3=2.5*0.63/eps)
par(pump1_t4=42, pump2_t4=42)
par(pump1_F4=2.5*0.83, pump2_F4=2.5*0.83/eps)
```

Process initial varied

Pump schedule - recycl

newplot(title='Perfusion cultivation - step wise increase of perfusion rate and m simu(60)



```
# Simulation without a plot and just to check typical values at high production r
simu(40)
c data=sim res['filter.retentate.c[1]']/sim res['filter.inlet.c[1]']
print('Conc factor variation', np.round(min(c_data[190:]), 3), 'to', np.round(max
→ Conc factor variation 1.162 to 1.179
#describe('cstrProdMax')
# The maximal biomass productivity before washout is obtained aroudn 40 hours
np.round(model.get('harvesttank.inlet.F')[0]*model.get('harvesttank.inlet.c[1]')[
→ 23.5
# Thus perfusion (with this filter) brings a productivity improvement of about
np.round(23.5/5.6,1)
→ 4.2
# Finally we check the filter flow rates at time 40 hour - note the negative sign
model.get('filter.inlet.F')[0]
→ 15.7499999999998
model.get('filter.filtrate.F')[0]
<del>→</del> -1.575
model.get('filter.retentate.F')[0]
```

Summary

- The perfusion filter had a concentration factor of cells around 1.08 and re-cycling flow was set to a factor 10 higher than the perfusion rate and changed when perfusion rate was change to keep the ratio factor 10.
- The first simulation showed that by cell retention using perfusion filter the process could be run at a perfusion flow rate at the maximal flow rate possible for corresponding chemostat culture and cell concetration increased steadily.
- The second simulation showed that with a proper startup cell concentration, the cell concentration remained constant when perfusion rate increased in a similar way as what we see in a chemostat.
- The second simulation also showed that biomass productivity in this case was increased by a factor 4.2 compared to chemostat.

 If the perfusion rate increased to higher levels washout started but the decrase of cell concentration was slow.

Some of you who read this may have your perfusion experience with CHO-cultures. For such cultures the cell concentration do increase with increase of perfusion rate and there are understood reasons for that. But for this simplified process as well as microbial processes they typically keep cell concentration constant when flow rate is chaged, and that under quite wide conditions. I will try come back to this phenomena in a later notebook.

```
conditions. I will try come back to this phenomena in a later notebook.
# List of components in the process setup and also a couple of other things like l:
describe('parts')
['bioreactor', 'bioreactor.culture', 'D', 'feedtank', 'filter', 'harvesttank'
describe('MSL')
MSL: 3.2.3 - used components: RealInput, RealOutput, CombiTimeTable, Types
system_info()
\rightarrow
    System information
     -OS: Linux
     -Python: 3.11.11
     -Scipy: not installed in the notebook
     -PyFMI: 2.16.3
     -FMU by: OpenModelica Compiler OpenModelica 1.25.0~dev-133-ga5470be
     -FMI: 2.0
     -Type: FMUModelME2
     -Name: BPL.Examples TEST2.Perfusion
     -Generated: 2024-11-06T21:37:58Z
     -MSL: 3.2.3
     -Description: Bioprocess Library version 2.3.0
     -Interaction: FMU-explore version 1.0.0
```

Start coding or generate with AI.