## **BPL\_TEST2\_Perfusion - demo**

In [1]: run -i BPL\_TEST2\_Perfusion\_fmpy\_explore.py

This notebook explore perfusion cultivation in comparison with ordinary continuous cultivation (chemostat) and use comparable settings to earlier notebook. Further you see here examples of interaction with the simplified commands par(), init(), simu() etc as well as direct interaction with the FMU which is called "model" here. The last simulation is always available in the workspace and called "sim\_res". Note that describe() brings mainly up from descriptive information from the Modelica code from the FMU but is complemented by some information given in the Python setup file.

```
Windows - run FMU pre-compiled JModelica 2.14
       Model for bioreactor has been setup. Key commands:
        - par() - change of parameters and initial values
        - init()

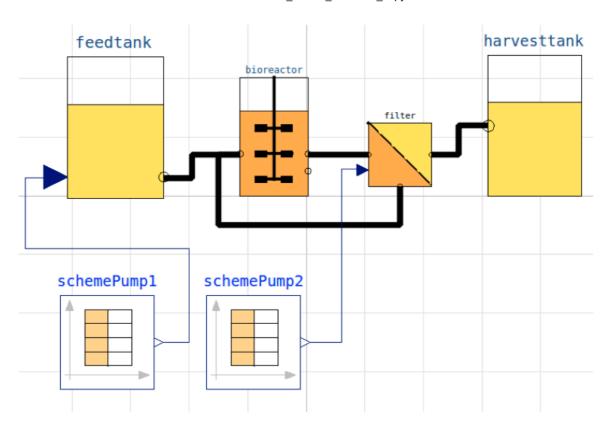
    change initial values only

       - simu()
                     - simulate and plot
       - newplot() - make a new plot
                   - show plot from previous simulation
        - show()
                    - display parameters and initial values from the last simulation
        - disp()
        - describe() - describe culture, broth, parameters, variables with values/units
       Note that both disp() and describe() takes values from the last simulation
       and the command process_diagram() brings up the main configuration
       Brief information about a command by help(), eg help(simu)
       Key system information is listed with the command system_info()
In [2]: %matplotlib inline
```

No processDiagram.png file in the FMU, but try the file on disk.

plt.rcParams['figure.figsize'] = [25/2.54, 20/2.54]

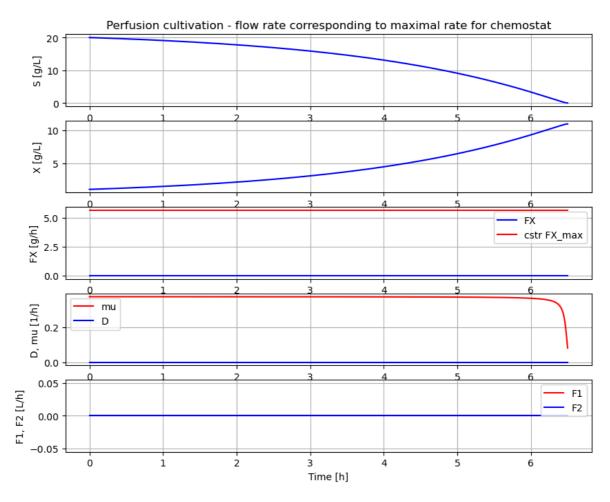
In [3]: process\_diagram()



```
In [4]: # Process parameters used throughout
    par(Y=0.5, qSmax=0.75, Ks=0.1)  # Culture
    par(filter_eps=0.10, filter_alpha_X=0.02, filter_alpha_S=0.10)  # Filter
    par(S_in=30.0)  # Inlet subs
    init(V_start=1.0, VX_start=1.0)  # Process in
    eps = parDict['filter_eps']  # Pump schea
```

```
In [5]: # Simulation of process with flow rate clot to wash-out for chemostat

init(VS_start=20)  # Process initial
 par(pump1_t1=10, pump2_t1=10)  # Pump schedule - recyc
 par(pump1_F1=2.5*0.155, pump2_F1=2.5*0.155/eps)
 par(pump1_t2=940, pump2_t2=940, pump1_t3=950, pump2_t3=950, pump1_t4=960, pump2_
 newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate f
 simu(6.5)
```



```
In [6]: # Concentration factor of the filter
    c=model_get('filter.retentate.c[1]')/model_get('filter.inlet.c[1]')
    print('Conc factor of perfusion filter =', np.round(c,3))
```

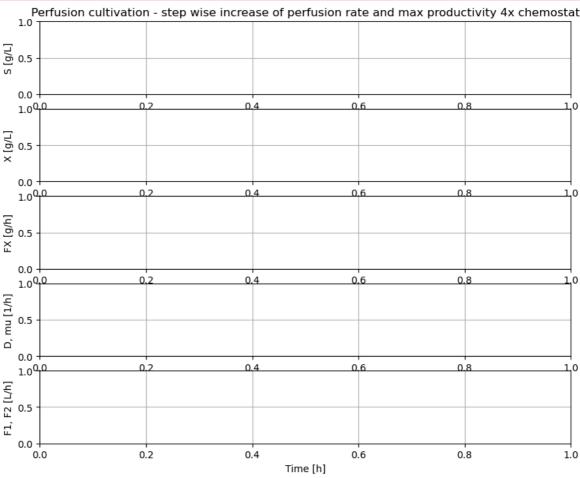
Conc factor of perfusion filter = 1.186

```
In [7]: c_data=sim_res['filter.retentate.c[1]']/sim_res['filter.inlet.c[1]']
    print('Conc factor variation', np.round(min(c_data[151:]), 3),'to', np.round(max
```

Conc factor variation 1.186 to 1.186

```
FMICallException
                                          Traceback (most recent call last)
Cell In[8], line 16
     13 par(pump1_F4=2.5*0.83, pump2_F4=2.5*0.83/eps)
     15 newplot(title='Perfusion cultivation - step wise increase of perfusion ra
te and max productivity 4x chemostat')
---> 16 simu(60)
File \\VBoxSvr\Modelica\GitHub\Colab\BPL_TEST2_Perfusion\BPL_TEST2_Perfusion_fmpy
_explore.py:565, in simu(simulationTime, mode, options, diagrams)
           start_values = {parLocation[k]:parDict[k] for k in parDict.keys()}
    564
           # Simulate
--> 565
           sim_res = simulate_fmu(
    566
             filename = fmu_model,
    567
             validate = False,
    568
             start time = 0,
    569
             stop time = simulationTime,
             output_interval = simulationTime/options['ncp'],
    570
             record_events = True,
   571
    572
             start_values = start_values,
             fmi_call_logger = None,
   573
             output = list(set(extract_variables(diagrams) + list(stateDict.keys
   574
()) + key_variables))
   575
    577
           simulationDone = True
    579 elif mode in ['Continued', 'continued', 'cont']:
File ~\miniconda3\envs\fmpy0320\Lib\site-packages\fmpy\simulation.py:761, in simu
late_fmu(filename, validate, start_time, stop_time, solver, step_size, relative_t
olerance, output_interval, record_events, fmi_type, start_values, apply_default_s
tart_values, input, output, timeout, debug_logging, visible, logger, fmi_call_log
ger, step_finished, model_description, fmu_instance, set_input_derivatives, remot
e_platform, early_return_allowed, use_event_mode, initialize, terminate, fmu_stat
e, set_stop_time)
   759
            result = simulateME(model description, fmu, start time, stop time, so
lver, step_size, relative_tolerance, start_values, apply_default_start_values, in
put, output, output_interval, record_events, timeout, step_finished, validate, se
t_stop_time)
    760 elif fmi type == 'CoSimulation':
--> 761 result = simulateCS(model description, fmu, start time, stop time, re
lative_tolerance, start_values, apply_default_start_values, input, output, output
interval, timeout, step finished, set input derivatives, use event mode, early r
eturn_allowed, validate, initialize, terminate, set_stop_time)
    763 if fmu instance is None:
   764
           fmu.freeInstance()
File ~\miniconda3\envs\fmpy0320\Lib\site-packages\fmpy\simulation.py:1272, in sim
ulateCS(model_description, fmu, start_time, stop_time, relative_tolerance, start_
values, apply_default_start_values, input_signals, output, output_interval, timeo
ut, step_finished, set_input_derivatives, use_event_mode, early_return_allowed, v
alidate, initialize, terminate, set stop time)
  1270
                        break
  1271
                else:
-> 1272
                   raise e
   1273 else:
   1275
           t_input_event = input.nextEvent(time)
File ~\miniconda3\envs\fmpy0320\Lib\site-packages\fmpy\simulation.py:1258, in sim
ulateCS(model_description, fmu, start_time, stop_time, relative_tolerance, start_
values, apply_default_start_values, input_signals, output, output_interval, timeo
```

```
ut, step_finished, set_input_derivatives, use_event_mode, early_return_allowed, v
alidate, initialize, terminate, set_stop_time)
   1256 try:
   1257
            if time + output_interval <= stop_time:</pre>
-> 1258
                fmu.doStep(currentCommunicationPoint=time, communicationStepSize=
output interval)
   1259
                n_steps += 1
   1260
                time = n_steps * output_interval
File ~\miniconda3\envs\fmpy0320\Lib\site-packages\fmpy\fmi2.py:580, in FMU2Slave.
doStep(self, currentCommunicationPoint, communicationStepSize, noSetFMUStatePrior
ToCurrentPoint)
    579 def doStep(self, currentCommunicationPoint, communicationStepSize, noSetF
MUStatePriorToCurrentPoint=fmi2True):
           self.fmi2DoStep(self.component, currentCommunicationPoint, communicat
ionStepSize, noSetFMUStatePriorToCurrentPoint)
File ~\miniconda3\envs\fmpy0320\Lib\site-packages\fmpy\fmi2.py:215, in _FMU2._fmi
2Function.<locals>.w(*args)
    212 if restype == fmi2Status: # status code
            # check the status code
    214
            if res > fmi2Warning:
                raise FMICallException(function=fname, status=res)
--> 215
    217 return res
FMICallException: fmi2DoStep failed with status 3 (error).
    Perfusion cultivation - step wise increase of perfusion rate and max productivity 4x chemostat
```



```
In []: #describe('cstrProdMax')
In []: # The maximal biomass productivity before washout is obtained aroudn 40 hours
    np.round(model_get('harvesttank.inlet.F')*model_get('harvesttank.inlet.c[1]'),1)
In []: # Thus perfusion (with this filter) brings a productivity improvement of about
    np.round(23.5/5.6,1)
In []: # Finally we check the filter flow rates at time 40 hour - note the negative sig
    model_get('filter.inlet.F')
In []: model_get('filter.filtrate.F')
In []: model_get('filter.retentate.F')
```

## **Summary**

- The perfusion filter had a concentration factor of cells around 1.08 and re-cycling flow was set to a factor 10 higher than the perfusion rate and changed when perfusion rate was change to keep the ratio factor 10.
- The first simulation showed that by cell retention using perfusion filter the process could be run at a perfusion flow rate at the maximal flow rate possible for corresponding chemostat culture and cell concetration increased steadily.
- The second simulation showed that with a proper startup cell concentration, the cell concentration remained constant when perfusion rate increased in a similar way as what we see in a chemostat.
- The second simulation also showed that biomass productivity in this case was increased by a factor 4.2 compared to chemostat.
- If the perfusion rate increased to higher levels washout started but the decrase of cell concentration was slow.

Some of you who read this may have your perfusion experience with CHO-cultures. For such cultures the cell concentration do increase with increase of perfusion rate and there are understood reasons for that. But for this simplified process as well as microbial processes they typically keep cell concentration constant when flow rate is chaged, and that under quite wide conditions. I will try come back to this phenomena in a later notebook.

## **Appendix**

```
In [11]: # List of components in the process setup and also a couple of other things like
         describe('parts')
        ['bioreactor', 'bioreactor.culture', 'D', 'feedtank', 'filter', 'harvesttank', 'l
        iquidphase', 'MSL', 'schemePump1', 'schemePump2']
In [12]: describe('MSL')
        MSL: RealInput, RealOutput, CombiTimeTable, Types
In [13]: system_info()
        System information
         -OS: Windows
         -Python: 3.12.3
         -Scipy: not installed in the notebook
         -FMPy: 0.3.20
         -FMU by: JModelica.org
         -FMI: 2.0
         -Type: CS
         -Name: BPL_TEST2.Perfusion
         -Generated: 2024-05-12T21:00:01
         -MSL: 3.2.2 build 3
         -Description: Bioprocess Library version 2.2.0
         -Interaction: FMU-explore for FMPy version 1.0.0
 In [ ]:
```