BPL_TEST2_Perfusion script with FMPy

The key library FMPy is installed.

After the installation a small application BPL_TEST2_Perfusion is loaded and run. You can continue with this example if you like.

!lsb_release -a # Actual VM Ubuntu version used by Google

No LSB modules are available.

Distributor ID: Ubuntu

Description: Ubuntu 22.04.3 LTS

Release: 22.04 Codename: jammy

%env PYTHONPATH=

```
→ env: PYTHONPATH=
```

```
!wget https://repo.anaconda.com/miniconda/Miniconda3-py312_24.3.0-0-Linux-x86_64.
!chmod +x Miniconda3-py312_24.3.0-0-Linux-x86_64.sh
!bash ./Miniconda3-py312 24.3.0-0-Linux-x86 64.sh -b -f -p /usr/local
import sys
sys.path.append('/usr/local/lib/python3.12/site-packages/')
```

 \rightarrow --2024-10-03 06:17:51-- https://repo.anaconda.com/miniconda/Miniconda3-py312 Resolving repo.anaconda.com (repo.anaconda.com)... 104.16.191.158, 104.16.32. Connecting to repo.anaconda.com (repo.anaconda.com)|104.16.191.158|:443... con HTTP request sent, awaiting response... 200 OK Length: 143351488 (137M) [application/octet-stream] Saving to: 'Miniconda3-py312_24.3.0-0-Linux-x86_64.sh'

Miniconda3-py312_24 100%[===========] 136.71M 61.1MB/s in 2.2s

2024-10-03 06:17:54 (61.1 MB/s) - 'Miniconda3-py312_24.3.0-0-Linux-x86 64.sh'

PREFIX=/usr/local Unpacking payload ...

Installing base environment...

Preparing transaction: ...working... done Executing transaction: ...working... done installation finished.

!conda update -n base -c defaults conda --yes

 \rightarrow

```
Downloading and Extracting Packages:
openssl-3.0.15
                      | 5.2 MB
                                        0% 0/1 [00:00<?, ?it/s]
                                  | :
conda-24.9.1
                      | 1.1 MB
                                        0% 0/1 [00:00<?, ?it/s]
                                  | :
certifi-2024.8.30
                                        0% 0/1 [00:00<?, ?it/s]
                      | 163 KB
                                  1 :
ca-certificates-2024 | 130 KB
                                  | :
                                        0% 0/1 [00:00<?, ?it/s]
frozendict-2.4.2
                      | 36 KB
                                        0% 0/1 [00:00<?, ?it/s]
openssl-3.0.15
                      | 5.2 MB
                                        0% 0.003007460830410892/1 [00:00<02:03
ca-certificates-2024 | 130 KB
                                       12% 0.12323429860849944/1 [00:00<00:02,
                                  1 :
certifi-2024.8.30
                      | 163 KB
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                                  | : |
openssl-3.0.15
                      | 5.2 MB
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ca-certificates-2024 | 130 KB
                                  | : 100% 1.0/1 [00:00<00:00,
                                                                 3.08s/it]
frozendict-2.4.2
                      | 36 KB
                                  | : 100% 1.0/1 [00:00<00:00,
                                                                 2.22it/s]
frozendict-2.4.2
                      | 36 KB
                                  | : 100% 1.0/1 [00:00<00:00,
                                                                 2.22it/s]
certifi-2024.8.30
                      | 163 KB
                                  | : 100% 1.0/1 [00:00<00:00,
                                                                 2.21it/s]
                      | 5.2 MB
                                  | : 100% 1.0/1 [00:01<00:00,
                                                                 1.04s/it]
openssl-3.0.15
conda-24.9.1
                      | 1.1 MB
                                  | : 100% 1.0/1 [00:01<00:00,
                                                                 1.05s/it]
```

Preparing transaction: done Verifying transaction: done Executing transaction: done

!conda --version
!python --version

conda 24.9.1 Python 3.12.2 !conda install -c conda-forge fmpy --yes # Install the key package

₹

```
#!conda install -c conda-forge matplotlib --yes
#!conda install -c conda-forge scipy --yes
#!conda install -c conda-forge openpyxl --yes
#!conda install -c conda-forge xlrd --yes
```

Notes of BPL_TEST2_Perfusion

This notebook explore perfusion cultivation in comparison with ordinary continuous cultivation (chemostat) and use comparable settings to earlier notebook. Further you see here examples of interaction with the simplified commands par(), init(), simu() etc as well as direct interaction with the FMU which is called "model" here. The last simulation is always available in the workspace and called "sim_res". Note that describe() brings mainly up from descriptive information from the Modelica code from the FMU but is complemented by some information given in the Python setup file.

Now specific installation run a simulation and notebook for that Start with connecting to Github. Then upload the two files:

- FMU BPL_TEST2_Perfusion_linux_om_me.fmu
- Setup-file BPL_TEST2_Perfusion_fmpy_explore.py

```
%%bash
git clone https://github.com/janpeter19/BPL_TEST2_Perfusion
Cloning into 'BPL_TEST2_Perfusion'...
%cd BPL_TEST2_Perfusion
/content/BPL_TEST2_Perfusion
run -i BPL_TEST2_Perfusion_fmpy_explore.py
→ Linux – run FMU pre-comiled OpenModelica
    Model for bioreactor has been setup. Key commands:
     - par()

    change of parameters and initial values

     - init()

    change initial values only

                   simulate and plot
     - simu()
     - newplot()
                   - make a new plot
     - show()
                   show plot from previous simulation
     - disp()

    display parameters and initial values from the last simulation
```

- describe() - describe culture, broth, parameters, variables with values/ur

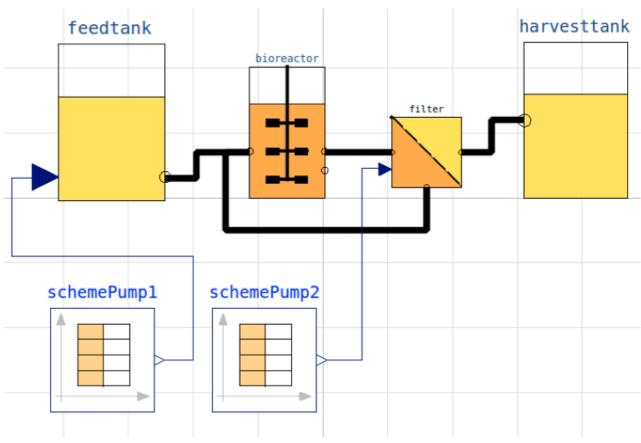
Note that both disp() and describe() takes values from the last simulation and the command process_diagram() brings up the main configuration

Brief information about a command by help(), eg help(simu) Key system information is listed with the command system_info()

%matplotlib inline
plt.rcParams['figure.figsize'] = [25/2.54, 20/2.54]

process_diagram()

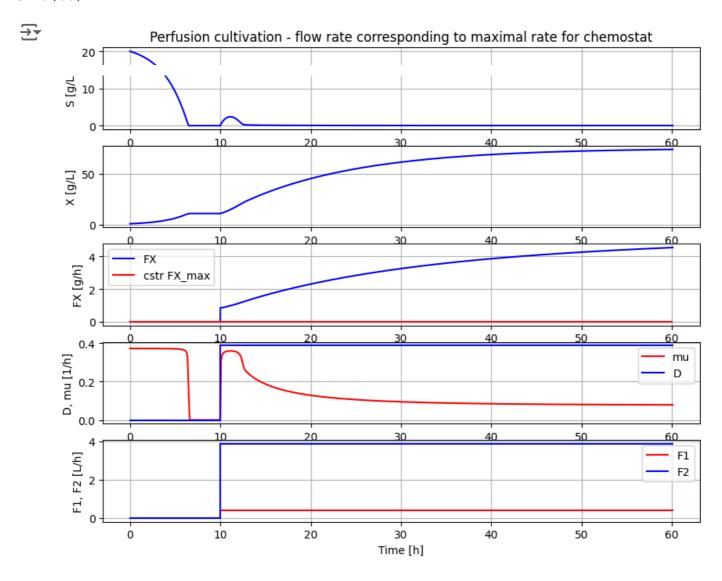
No processDiagram.png file in the FMU, but try the file on disk.



```
# Process parameters used throughout
par(Y=0.5, qSmax=0.75, Ks=0.1)  # Culture
par(filter_eps=0.10, filter_alpha_X=0.02, filter_alpha_S=0.10)  # Filter
par(S_in=30.0)  # Inlet subst
init(V_start=1.0, VX_start=1.0)  # Process ini
eps = parDict['filter_eps']  # Pump schedu
```

Simulation of process with flow rate clot to wash-out for chemostat

newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate fo simu(60)

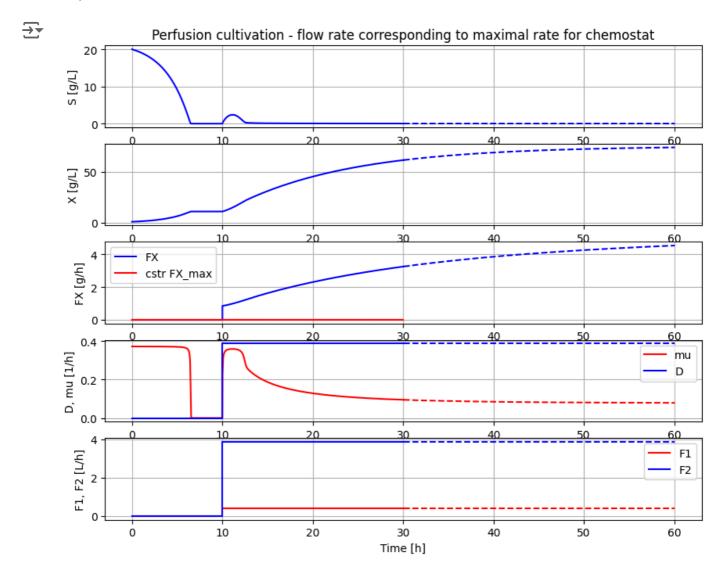


Simulation of process with flow rate close to wash-out for chemostat

```
init(VS_start=20)  # Process initial
par(pump1_t1=10, pump2_t1=10)  # Pump schedule - recycl
par(pump1_F1=2.5*0.155, pump2_F1=2.5*0.155/eps)
par(pump1_t2=940, pump2_t2=940, pump1_t3=950, pump2_t3=950, pump1_t4=960, pump2_t
newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate fo
```

newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate fo simu(30)

simu(30,'cont')



```
# Concentration factor of the filter
c=model_get('filter.retentate.c[1]')/model_get('filter.inlet.c[1]')
print('Conc factor of perfusion filter =', np.round(c,3))
```

Econc factor of perfusion filter = 1.179

```
c_data=sim_res['filter.retentate.c[1]']/sim_res['filter.inlet.c[1]']
print('Conc factor variation', np.round(min(c_data[151:]), 3), np.round(max(c_dat

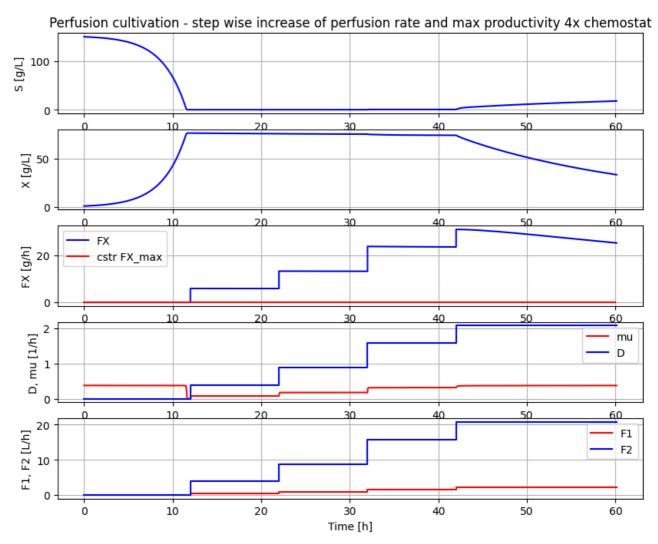
Conc factor variation 1.179 1.179

# Simulation of process with step-wise increase of pefusion rate until wash-out.
# This means that re-circulation rate change at the same time as the perfusion rate init(VS_start=150)

# Process initial varied
```

newplot(title='Perfusion cultivation - step wise increase of perfusion rate and m simu(60)





```
# Simulation without a plot and just to check typical values at high production r
#simu(40)
#c_data=sim_res['filter.retentate.c[1]']/sim_res['filter.inlet.c[1]']
#print('Conc factor variation', np.round(min(c_data[190:]), 3), 'to', np.round(ma
#describe('cstrProdMax')
```

The maximal biomass productivity before washout is obtained aroudn 40 hours
np.round(model_get('harvesttank.inlet.F')*model_get('harvesttank.inlet.c[1]'),1)

→ 25.2

Thus perfusion (with this filter) brings a productivity improvement of about np.round(23.5/5.6,1)

```
→ 4.2
```

```
# Finally we check the filter flow rates at time 40 hour - note the negative sign model_get('filter.inlet.F')

→ 20.74999999999996

model_get('filter.filtrate.F')

→ -2.07499999999997

model_get('filter.retentate.F')

→ -18.674999999999997
```

Summary

- The perfusion filter had a concentration factor of cells around 1.08 and re-cycling flow was set to a factor 10 higher than the perfusion rate and changed when perfusion rate was change to keep the ratio factor 10.
- The first simulation showed that by cell retention using perfusion filter the process could be run at a perfusion flow rate at the maximal flow rate possible for corresponding chemostat culture and cell concetration increased steadily.
- The second simulation showed that with a proper startup cell concentration, the cell
 concentration remained constant when perfusion rate increased in a similar way as what
 we see in a chemostat.
- The second simulation also showed that biomass productivity in this case was increased by a factor 4.2 compared to chemostat.
- If the perfusion rate increased to higher levels washout started but the decrase of cell concentration was slow.

Some of you who read this may have your perfusion experience with CHO-cultures. For such cultures the cell concentration do increase with increase of perfusion rate and there are understood reasons for that. But for this simplified process as well as microbial processes they typically keep cell concentration constant when flow rate is chaged, and that under quite wide conditions. I will try come back to this phenomena in a later notebook.

```
# List of components in the process setup and also a couple of other things like l: describe('parts')

['bioreactor', 'bioreactor.culture', 'D', 'feedtank', 'filter', 'harvesttank' describe('MSL')
```