BPL_TEST2_Perfusion - demo

This notebook explore perfusion cultivation in comparison with ordinary continuous cultivation (chemostat) and use comparable settings to earlier notebook. Further you see here examples of interaction with the simplified commands par(), init(), simu() etc as well as direct interaction with the FMU which is called "model" here. The last simulation is always available in the workspace and called "sim_res". Note that describe() brings mainly up from descriptive information from the Modelica code from the FMU but is complemented by some information given in the Python setup file.

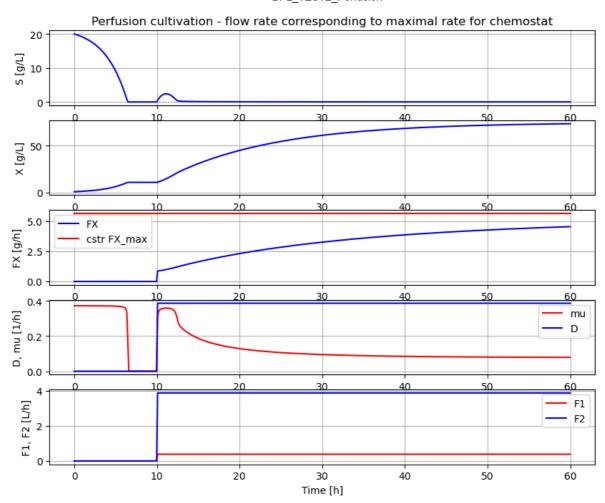
```
In [1]: run -i BPL_TEST2_Perfusion_explore.py
                   Windows - run FMU pre-compiled JModelica 2.14
                    Model for bioreactor has been setup. Key commands:
                      - par()
                                                   - change of parameters and initial values
                                                    - change initial values only
                      - init()

    simulate and plot

                      - simu()
                      - newplot() - make a new plot

    show plot from previous simulation

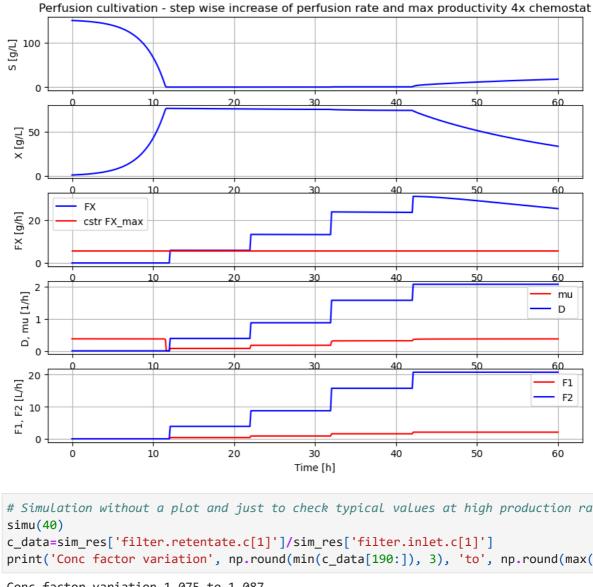
                      - show()
                      - disp() - display parameters and initial values from the last simulation
                      - describe() - describe culture, broth, parameters, variables with values / unit
                   Note that both disp() and describe() takes values from the last simulation
                    Brief information about a command by help(), eg help(simu)
                    Key system information is listed with the command system_info()
In [2]: %matplotlib inline
                    plt.rcParams['figure.figsize'] = [25/2.54, 20/2.54]
In [3]: # Process parameters used throughout
                    par(Y=0.5, qSmax=0.75, Ks=0.1)
                                                                                                                                                                                     # Culture
                    par(filter_eps=0.10, filter_alpha_X=0.02, filter_alpha_S=0.10)
                                                                                                                                                                                     # Filter
                                                                                                                                                                                     # Inlet substro
                    par(S in=30.0)
                    init(V_0=1.0, VX_0=1.0)
                                                                                                                                                                                     # Process initi
                    eps = parDict['filter_eps']
                                                                                                                                                                                      # Pump schedule
In [4]: # Simulation of process with flow rate clot to wash-out for chemostat
                    init(VS_0=20)
                                                                                                                                                            # Process initial
                    par(pump1_t1=10, pump2_t1=10)
                                                                                                                                                            # Pump schedule - recycle
                    par(pump1 F1=2.5*0.155, pump2 F1=2.5*0.155/eps)
                    par(pump1 t2=940, pump2 t2=940, pump1 t3=950, pump2 t3=950, pump1 t4=960, pump2 t4=9
                    newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate for
                    simu(60)
```



```
In [5]: # Concentration factor of the filter
    c=model.get('filter.retentate.c[1]')[0]/model.get('filter.inlet.c[1]')[0]
    print('Conc factor of perfusion filter =', np.round(c,3))

Conc factor of perfusion filter = 1.369
```

In [6]: c_data=sim_res['filter.retentate.c[1]']/sim_res['filter.inlet.c[1]']
print('Conc factor variation', np.round(min(c_data[151:]), 3), np.round(max(c_data
Conc factor variation 1.369 1.649



```
In [8]: # Simulation without a plot and just to check typical values at high production rat
         print('Conc factor variation', np.round(min(c_data[190:]), 3), 'to', np.round(max(
         Conc factor variation 1.075 to 1.087
In [9]: describe('cstrProdMax')
         Calculate from the model maximal chemostat productivity FX_max : 5.625 [ g/h ]
In [10]: # The maximal biomass productivity before washout is obtained aroudn 40 hours
         np.round(model.get('harvesttank.inlet.F')[0]*model.get('harvesttank.inlet.c[1]')[0]
Out[10]: 23.5
In [11]: # Thus perfusion (with this filter) brings a productivity improvement of about
         np.round(23.5/5.6,1)
Out[11]: 4.2
In [12]: # Finally we check the filter flow rates at time 40 hour - note the negative sign f
         model.get('filter.inlet.F')[0]
Out[12]: 15.74999999999998
In [13]: model.get('filter.filtrate.F')[0]
Out[13]: -1.575
        model.get('filter.retentate.F')[0]
```

Summary

- The perfusion filter had a concentration factor of cells around 1.08 and re-cycling flow was set to a factor 10 higher than the perfusion rate and changed when perfusion rate was change to keep the ratio factor 10.
- The first simulation showed that by cell retention using perfusion filter the process could be run at a perfusion flow rate at the maximal flow rate possible for corresponding chemostat culture and cell concetration increased steadily.
- The second simulation showed that with a proper startup cell concentration, the cell concentration remained constant when perfusion rate increased in a similar way as what we see in a chemostat.
- The second simulation also showed that biomass productivity in this case was increased by a factor 4.2 compared to chemostat.
- If the perfusion rate increased to higher levels washout started but the decrase of cell concentration was slow.

Some of you who read this may have your perfusion experience with CHO-cultures. For such cultures the cell concentration do increase with increase of perfusion rate and there are understood reasons for that. But for this simplified process as well as microbial processes they typically keep cell concentration constant when flow rate is chaged, and that under quite wide conditions. I will try come back to this phenomena in a later notebook.

Appendix

```
In [15]: disp('culture')
    Y : 0.5
    qSmax : 0.75
    Ks : 0.1

In [16]: describe('mu')
    Cell specific growth rate variable : 0.315 [ 1/h ]

In [17]: # List of components in the process setup and also a couple of other things like Lidescribe('parts')
    ['bioreactor', 'bioreactor.culture', 'D', 'feedtank', 'filter', 'harvesttank', 'lidescribe('MSL')
    MSL: RealInput, RealOutput, CombiTimeTable, Types

In [19]: system_info()
```

System information

-OS: Windows
-Python: 3.10.6

-Scipy: not installed in the notebook

-PyFMI: 2.9.8

-FMU by: JModelica.org

-FMI: 2.0

-Type: FMUModelCS2

-Name: BPL_TEST2.Perfusion
-Generated: 2022-10-06T08:40:14

-MSL: 3.2.2 build 3

-Description: Bioprocess Library version 2.1.0

-Interaction: FMU-explore version 0.9.6e

In []: