

## ▼ BPL\_TEST2\_Perfusion script with FMPy ver 0.3.15

The key library FMPy v0.3.15 is installed.

After the installation a small application BPL\_TEST2\_Perfusion is loaded and run. You can continue with this example if you like.

```
!lsb_release -a # Actual VM Ubuntu version used by Google

No LSB modules are available.
Distributor ID: Ubuntu
Description:   Ubuntu 22.04.2 LTS
Release:      22.04
Codename:     jammy

%env PYTHONPATH=

env: PYTHONPATH=

!wget https://repo.anaconda.com/miniconda/Miniconda3-py310_23.1.0-1-Linux-x86_64.sh
!chmod +x Miniconda3-py310_23.1.0-1-Linux-x86_64.sh
!bash ./Miniconda3-py310_23.1.0-1-Linux-x86_64.sh -b -f -p /usr/local
import sys
sys.path.append('/usr/local/lib/python3.10/site-packages/')

--2023-09-26 10:19:26--  https://repo.anaconda.com/miniconda/Miniconda3-py310_23.1.0-1-Linux-x86_64.sh
Resolving repo.anaconda.com (repo.anaconda.com)... 104.16.130.3, 104.16.131.3, 2606:4700::6810:8303, ...
Connecting to repo.anaconda.com (repo.anaconda.com)|104.16.130.3|:443... connected.
HTTP request sent, awaiting response... 200 OK
Length: 74403966 (71M) [application/x-sh]
Saving to: 'Miniconda3-py310_23.1.0-1-Linux-x86_64.sh.1'

Miniconda3-py310_23 100%[=====] 70.96M  147MB/s   in 0.5s

2023-09-26 10:19:27 (147 MB/s) - 'Miniconda3-py310_23.1.0-1-Linux-x86_64.sh.1' saved [74403966/74403966]

PREFIX=/usr/local
Unpacking payload ...

Installing base environment...

Downloading and Extracting Packages

Downloading and Extracting Packages

Preparing transaction: done
Executing transaction: done
installation finished.

!conda update -n base -c defaults conda --yes
```

```

Preparing transaction: done
Verifying transaction: done
Executing transaction: done

!conda --version
!python --version

conda 23.7.4
Python 3.10.13

!conda install -c conda-forge fmpy --yes # Install the key package

Collecting package metadata (current_repodata.json): done
Solving environment: \
The environment is inconsistent, please check the package plan carefully
The following packages are causing the inconsistency:

- conda-forge/noarch::typing-extensions==4.8.0=hd8ed1ab_done

## Package Plan ##

environment location: /usr/local

added / updated specs:
- fmpy

The following packages will be UPDATED:

typing_extensions  pkgs/main/linux-64::typing_extensions~ --> conda-forge/noarch::typing_extensions-4.8.0-pyha770c72_0

The following packages will be SUPERSEDED by a higher-priority channel:

ca-certificates    pkgs/main::ca-certificates-2023.08.22~ --> conda-forge::ca-certificates-2023.7.22-hbcca054_0
certifi            pkgs/main/linux-64::certifi-2023.7.22~ --> conda-forge/noarch::certifi-2023.7.22-pyhd8ed1ab_0
conda              pkgs/main::conda-23.7.4-py310h06a4308~ --> conda-forge::conda-23.7.4-py310hff52083_0

Downloading and Extracting Packages

Preparing transaction: done
Verifying transaction: done
Executing transaction: done

!conda install -c conda-forge matplotlib --yes

Collecting package metadata (current_repodata.json): done
Solving environment: -
CondaError: KeyboardInterrupt

#!conda install -c conda-forge scipy --yes

#!conda install -c conda-forge openpyxl --yes

#!conda install -c conda-forge xlrd --yes

```

## ▼ Notes of BPL\_TEST2\_Perfusion

This notebook explore perfusion cultivation in comparison with ordinary continuous cultivation (chemostat) and use comparable settings to earlier notebook. Further you see here examples of interaction with the simplified commands `par()`, `init()`, `simu()` etc as well as direct interaction with the FMU which is called "model" here. The last simulation is always available in the workspace and called "sim\_res". Note that `describe()` brings mainly up from descriptive information from the Modelica code from the FMU but is complemented by some information given in the Python setup file.

Now specific installation run a simulation and notebook for that Start with connecting to Github. Then upload the two files:

- FMU - BPL\_TEST2\_Perfusion\_linux\_om\_me.fmu
- Setup-file - BPL\_TEST2\_Perfusion\_fmipy\_explore.py

```
%%bash
```

```
git clone https://github.com/janpeter19/BPL_TEST2_Perfusion
```

```
fatal: destination path 'BPL_TEST2_Perfusion' already exists and is not an empty directory.
```

```
-----
CalledProcessError                                Traceback (most recent call last)
```

```
<ipython-input-8-f787d3948132> in <cell line: 1>()
```

```
----> 1 get_ipython().run_cell_magic('bash', '', 'git clone https://github.com/janpeter19/BPL_TEST2_Perfusion\n')
```

```
-----
  4 frames -----
```

```
<decorator-gen-103> in shebang(self, line, cell)
```

```
/usr/local/lib/python3.10/dist-packages/IPython/core/magics/script.py in shebang(self, line, cell)
```

```
243         sys.stderr.flush()
```

```
244         if args.raise_error and p.returncode!=0:
```

```
--> 245             raise CalledProcessError(p.returncode, cell, output=out, stderr=err)
```

```
246
```

```
247     def _run_script(self, p, cell, to_close):
```

```
CalledProcessError: Command 'b'git clone https://github.com/janpeter19/BPL_TEST2_Perfusion\n'' returned non-zero exit sta
```

SEARCH STACK OVERFLOW

```
%cd BPL_TEST2_Perfusion
```

```
/content/BPL_TEST2_Perfusion
```

```
run -i BPL_TEST2_Perfusion_fmipy_explore.py
```

```
Linux - run FMU pre-comiled OpenModelica 1.21.0
```

```
Model for bioreactor has been setup. Key commands:
```

- `par()` - change of parameters and initial values
- `init()` - change initial values only
- `simu()` - simulate and plot
- `newplot()` - make a new plot
- `show()` - show plot from previous simulation
- `disp()` - display parameters and initial values from the last simulation
- `describe()` - describe culture, broth, parameters, variables with values/units

Note that both `disp()` and `describe()` takes values from the last simulation and the command `process_diagram()` brings up the main configuration

Brief information about a command by `help()`, eg `help(simu)`

Key system information is listed with the command `system_info()`

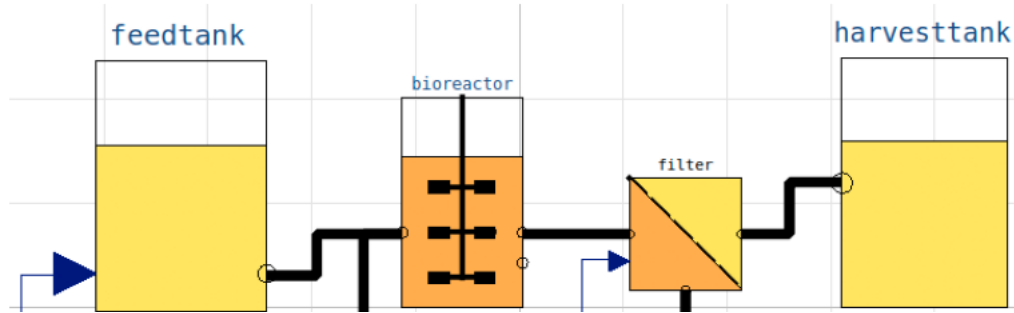
```
%matplotlib inline
```

```
plt.rcParams['figure.figsize'] = [25/2.54, 20/2.54]
```

```
process_diagram()
```



No processDiagram.png file in the FMU, but try the file on disk.

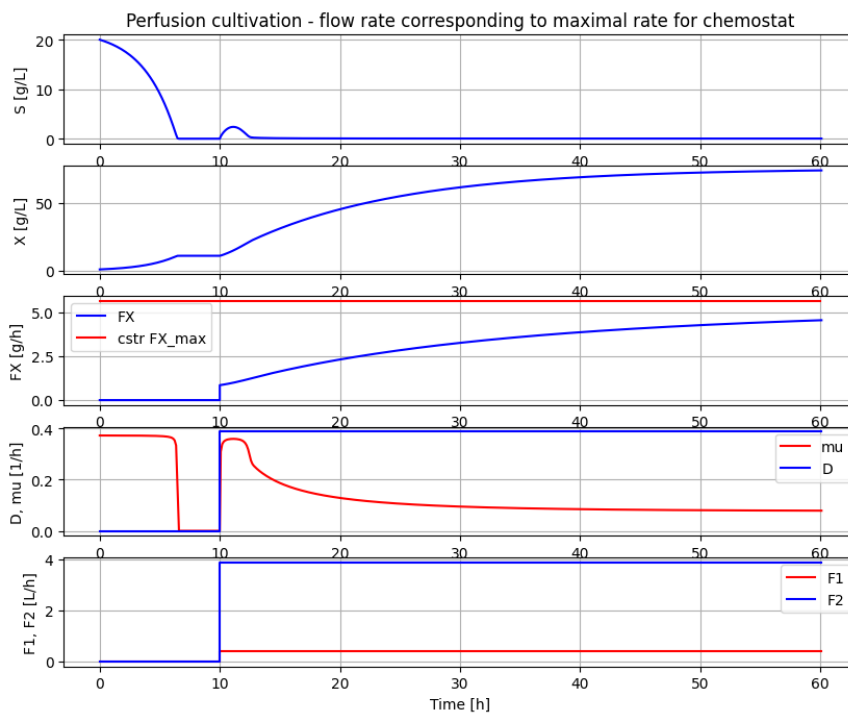


```
# Process parameters used throughout
par(Y=0.5, qSmax=0.75, Ks=0.1)
par(filter_eps=0.10, filter_alpha_X=0.02, filter_alpha_S=0.10)
par(S_in=30.0)
init(V_0=1.0, VX_0=1.0)
eps = parDict['filter_eps']
```

# Simulation of process with flow rate plot to wash-out for chemostat

```
init(VS_0=20)
par(pump1_t1=10, pump2_t1=10)
par(pump1_F1=2.5*0.155, pump2_F1=2.5*0.155/eps)
par(pump1_t2=940, pump2_t2=940, pump1_t3=950, pump2_t3=950, pump1_t4=960, pump2_t4=960)
```

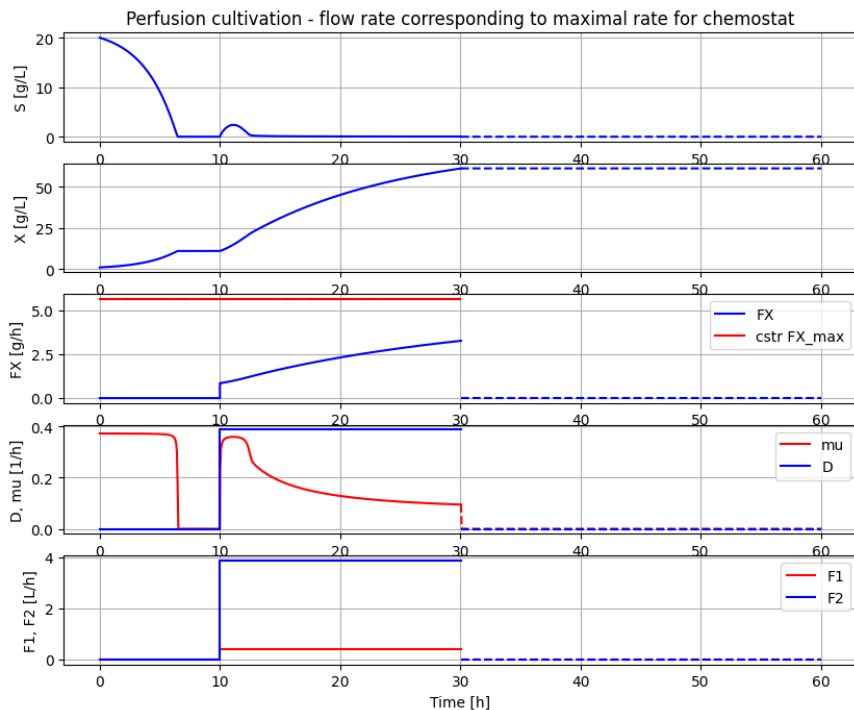
```
newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate for chemostat')
simu(60)
```



# Simulation of process with flow rate close to wash-out for chemostat

```
init(VS_0=20)
par(pump1_t1=10, pump2_t1=10)
par(pump1_F1=2.5*0.155, pump2_F1=2.5*0.155/eps)
par(pump1_t2=940, pump2_t2=940, pump1_t3=950, pump2_t3=950, pump1_t4=960, pump2_t4=960)

newplot(title='Perfusion cultivation - flow rate corresponding to maximal rate for chemostat')
simu(30)
simu(30, 'cont')
```



```
# Concentration factor of the filter
c=model_get('filter.retentate.c[1]')/model_get('filter.inlet.c[1]')
print('Conc factor of perfusion filter =', np.round(c,3))

Conc factor of perfusion filter = 1.186

c_data=sim_res['filter.retentate.c[1]']/sim_res['filter.inlet.c[1]']
print('Conc factor variation', np.round(min(c_data[151:]), 3), np.round(max(c_data[151:]),3))

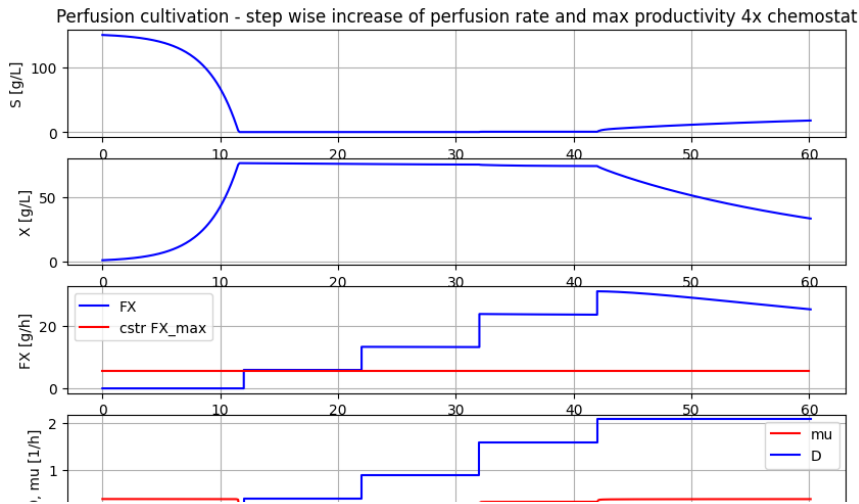
Conc factor variation 1.186 1.186

# Simulation of process with step-wise increase of perfusion rate until wash-out.
# This means that re-circulation rate change at the same time as the perfusion rate.

init(VS_0=150)                                # Process initial varied

par(pump1_t1=12, pump2_t1=12)                  # Pump schedule - recycle flow 10 times perfusion flow
par(pump1_F1=2.5*0.155, pump2_F1=2.5*0.155/eps)
par(pump1_t2=22, pump2_t2=22)
par(pump1_F2=2.5*0.35, pump2_F2=2.5*0.35/eps)
par(pump1_t3=32, pump2_t3=32)
par(pump1_F3=2.5*0.63, pump2_F3=2.5*0.63/eps)
par(pump1_t4=42, pump2_t4=42)
par(pump1_F4=2.5*0.83, pump2_F4=2.5*0.83/eps)

newplot(title='Perfusion cultivation - step wise increase of perfusion rate and max productivity 4x chemostat')
simu(60)
```



```
# Simulation without a plot and just to check typical values at high production rate
#simu(40)
#c_data=sim_res['filter.retentate.c[1]']/sim_res['filter.inlet.c[1]']
#print('Conc factor variation', np.round(min(c_data[190:]), 3), 'to', np.round(max(c_data[190:]),3))

#describe('cstrProdMax')
```

```
# The maximal biomass productivity before washout is obtained aroundn 40 hours
np.round(model_get('harvesttank.inlet.F')*model_get('harvesttank.inlet.c[1]'),1)
```

25.2

```
# Thus perfusion (with this filter) brings a productivity improvement of about
np.round(23.5/5.6,1)
```

4.2

```
# Finally we check the filter flow rates at time 40 hour - note the negative sign for outflow
model_get('filter.inlet.F')
```

```
model_get('filter.filtrate.F')
```

-2.0749999999999997

```
model_get('filter.retentate.F')
```

-18.674999999999997

## Summary

- The perfusion filter had a concentration factor of cells around 1.08 and re-cycling flow was set to a factor 10 higher than the perfusion rate and changed when perfusion rate was change to keep the ratio factor 10.
- The first simulation showed that by cell retention using perfusion filter the process could be run at a perfusion flow rate at the maximal flow rate possible for corresponding chemostat culture and cell concentration increased steadily.
- The second simulation showed that with a proper startup cell concentration, the cell concentration remained constant when perfusion rate increased in a similar way as what we see in a chemostat.
- The second simulation also showed that biomass productivity in this case was increased by a factor 4.2 compared to chemostat.
- If the perfusion rate increased to higher levels washout started but the decrease of cell concentration was slow.

Some of you who read this may have your perfusion experience with CHO-cultures. For such cultures the cell concentration do increase with increase of perfusion rate and there are understood reasons for that. But for this simplified process as well as microbial processes they typically keep cell concentration constant when flow rate is chaged, and that under quite wide conditions. I will try come back to this phenomena in a later notebook.

```
# List of components in the process setup and also a couple of other things like liquidphase and D
describe('parts')
```

```
['bioreactor', 'bioreactor.culture', 'D', 'feedtank', 'filter', 'harvesttank', 'schemePump1', 'schemePump2']
```

```
describe('MSL')
```

MSL: 3.2.3 - used components: RealInput, RealOutput, CombiTimeTable, Types

```
system_info()
```

System information

- OS: Linux
- Python: 3.10.12
- Scipy: not installed in the notebook
- FMPy: 0.3.15
- FMU by: OpenModelica Compiler OpenModelica 1.21.0
- FMI: 2.0
- Type: ME
- Name: BPL\_TEST2.Perfusion
- Generated: 2023-04-20T12:25:10Z
- MSL: 3.2.3
- Description: Bioprocess Library version 2.1.1
- Interaction: FMU-explore for FMPy version 0.9.8