

# BPL\_IEC\_operation script with PyFMI

The key library PyFMI is installed.

After the installation a small application BPL\_IEC\_operation is loaded and run. You can continue with this example if you like.

```
In [1]: !lsb_release -a # Actual VM Ubuntu version used by Google
```

```
No LSB modules are available.  
Distributor ID: Ubuntu  
Description:    Ubuntu 22.04.4 LTS  
Release:        22.04  
Codename:       jammy
```

```
In [2]: %env PYTHONPATH=
```

```
env: PYTHONPATH=
```

```
In [3]: !python --version
```

```
Python 3.11.11
```

```
In [4]: !wget https://repo.anaconda.com/miniconda/Miniconda3-py311_24.11.1-0-Linux-x86_64.s  
!chmod +x Miniconda3-py311_24.11.1-0-Linux-x86_64.sh  
!bash ./Miniconda3-py311_24.11.1-0-Linux-x86_64.sh -b -f -p /usr/local  
import sys  
sys.path.append('/usr/local/lib/python3.11/site-packages/')
```

```
--2025-03-27 08:21:43-- https://repo.anaconda.com/miniconda/Miniconda3-py311_24.11.1-0-Linux-x86_64.sh
Resolving repo.anaconda.com (repo.anaconda.com)... 104.16.191.158, 104.16.32.241, 2606:4700::6810:20f1, ...
Connecting to repo.anaconda.com (repo.anaconda.com)|104.16.191.158|:443... connected.
HTTP request sent, awaiting response... 200 OK
Length: 145900576 (139M) [application/octet-stream]
Saving to: 'Miniconda3-py311_24.11.1-0-Linux-x86_64.sh'

Miniconda3-py311_24 100%[=====>] 139.14M  83.4MB/s   in 1.7s

2025-03-27 08:21:45 (83.4 MB/s) - 'Miniconda3-py311_24.11.1-0-Linux-x86_64.sh' saved
[145900576/145900576]

PREFIX=/usr/local
Unpacking payload ...

Installing base environment...

Preparing transaction: ...working... done
Executing transaction: ...working... done
installation finished.
```

```
In [5]: !conda update -n base -c defaults conda --yes
```

Channels:  
- defaults  
Platform: linux-64  
Collecting package metadata (repodata.json): - 00\ 00| 00/ 00- 00\ 00| 00/ 00- 00\  
00| 00/ 00- 00done  
Solving environment: | 00/ 00done

## Package Plan ##

environment location: /usr/local

added / updated specs:  
- conda

The following packages will be downloaded:

package	build	
-----	-----	
ca-certificates-2025.2.25	h06a4308_0	129 KB
certifi-2025.1.31	py311h06a4308_0	163 KB
openssl-3.0.16	h5eee18b_0	5.2 MB
-----	-----	
Total:		5.5 MB

The following packages will be UPDATED:

ca-certificates	2024.11.26-h06a4308_0 --> 2025.2.25-h06a4308_0
certifi	2024.8.30-py311h06a4308_0 --> 2025.1.31-py311h06a4308_0
openssl	3.0.15-h5eee18b_0 --> 3.0.16-h5eee18b_0

Downloading and Extracting Packages:

openssl-3.0.16	5.2 MB	: 0% 0/1 [00:00<?, ?it/s]
certifi-2025.1.31	163 KB	: 0% 0/1 [00:00<?, ?it/s]
ca-certificates-2025	129 KB	: 0% 0/1 [00:00<?, ?it/s]
ca-certificates-2025	129 KB	: 100% 1.0/1 [00:00<00:00, 10.70it/s]
openssl-3.0.16	5.2 MB	: 3% 0.02982593950162064/1 [00:00<00:03, 3.43s/it]
certifi-2025.1.31	163 KB	: 100% 1.0/1 [00:00<00:00, 9.80it/s]
ca-certificates-2025	129 KB	: 100% 1.0/1 [00:00<00:00, 9.07it/s]
ca-certificates-2025	129 KB	: 100% 1.0/1 [00:00<00:00, 9.07it/s]

Preparing transaction: - 00done  
Verifying transaction: | 00/ 00- 00done  
Executing transaction: | 00done

```
In [6]: !conda --version  
!python --version
```

```
conda 24.11.1  
Python 3.11.11
```

```
In [7]: !conda config --set channel_priority strict
```

```
In [8]: !conda install -c conda-forge pyfmi --yes # Install the key package
```

```
Channels:
- conda-forge
- defaults
Platform: linux-64
Collecting package metadata (repodata.json): - 22\ 22| 22/ 22- 22\ 22| 22/ 22- 22\
22| 22/ 22- 22\ 22| 22/ 22- 22\ 22| 22/ 22- 22\ 22| 22/ 22- 22\ 22| 22/ 22- 22\ 22|
22/ 22- 22\ 22done
Solving environment: / 22- 22\ 22| 22done
```

## Package Plan ##

environment location: /usr/local

added / updated specs:

- pyfmi

The following packages will be downloaded:

package	build		
-----	-----		
_x86_64-microarch-level-3	2_broadwell	8 KB	conda-forge
assimulo-3.6.0	py311h083bc19_0	1.1 MB	conda-forge
certifi-2025.1.31	pyhd8ed1ab_0	159 KB	conda-forge
conda-25.3.0	py311h38be061_0	1.1 MB	conda-forge
fmilib-2.4.1	hac33072_1	383 KB	conda-forge
gmp-6.3.0	hac33072_2	449 KB	conda-forge
libamd-3.3.3	haaf9dc3_7100102	49 KB	conda-forge
libblas-3.9.0	31_h59b9bed_openblas	16 KB	conda-forge
libbtf-2.3.2	h32481e8_7100102	27 KB	conda-forge
libcamd-3.3.3	h32481e8_7100102	46 KB	conda-forge
libcbblas-3.9.0	31_he106b2a_openblas	16 KB	conda-forge
libccolamd-3.3.4	h32481e8_7100102	42 KB	conda-forge
libcholmod-5.3.1	h59ddab4_7100102	1.1 MB	conda-forge
libcolamd-3.3.4	h32481e8_7100102	33 KB	conda-forge
libcxsparse-4.4.1	h32481e8_7100102	118 KB	conda-forge
libgcc-14.2.0	h767d61c_2	828 KB	conda-forge
libgcc-ng-14.2.0	h69a702a_2	52 KB	conda-forge
libgfortran-14.2.0	h69a702a_2	52 KB	conda-forge
libgfortran-ng-14.2.0	h69a702a_2	53 KB	conda-forge
libgfortran5-14.2.0	hf1ad2bd_2	1.4 MB	conda-forge
libgomp-14.2.0	h767d61c_2	449 KB	conda-forge
libklu-2.3.5	hf24d653_7100102	142 KB	conda-forge
liblapack-3.9.0	31_h7ac8fdf_openblas	16 KB	conda-forge
libldl-3.3.2	h32481e8_7100102	24 KB	conda-forge
libopenblas-0.3.29	pthreads_h94d23a6_0	5.6 MB	conda-forge
libparu-1.0.0	h17147ab_7100102	91 KB	conda-forge
librbio-4.3.4	h32481e8_7100102	47 KB	conda-forge
libspex-3.2.3	had10066_7100102	79 KB	conda-forge
libspqr-4.3.4	h852d39f_7100102	213 KB	conda-forge
libstdcxx-14.2.0	h8f9b012_2	3.7 MB	conda-forge
libstdcxx-ng-14.2.0	h4852527_2	53 KB	conda-forge
libsuitesparseconfig-7.10.1	h92d6892_7100102	42 KB	conda-forge
libumfpack-6.3.5	heb53515_7100102	424 KB	conda-forge
metis-5.1.0	hd0bcaf9_1007	3.7 MB	conda-forge
mpfr-4.2.1	h90cbb55_3	620 KB	conda-forge

numpy-2.2.4	py311h5d046bc_0	8.6 MB	conda-forge
openssl-3.4.1	h7b32b05_0	2.8 MB	conda-forge
pyfmi-2.16.3	py311h9f3472d_0	5.2 MB	conda-forge
python_abi-3.11	2_cp311	5 KB	conda-forge
scipy-1.15.2	py311h8f841c2_0	16.4 MB	conda-forge
suitesparse-7.10.1	ha0f6916_7100102	12 KB	conda-forge
sundials-7.1.1	ha52427a_0	907 KB	conda-forge
-----			
Total:		56.1 MB	

The following NEW packages will be INSTALLED:

_x86_64-microarch~	conda-forge/noarch::_x86_64-microarch-level-3-2_broadwell
assimulo	conda-forge/linux-64::assimulo-3.6.0-py311h083bc19_0
fmilib	conda-forge/linux-64::fmilib-2.4.1-hac33072_1
gmp	conda-forge/linux-64::gmp-6.3.0-hac33072_2
libamd	conda-forge/linux-64::libamd-3.3.3-haaf9dc3_7100102
libblas	conda-forge/linux-64::libblas-3.9.0-31_h59b9bed_openblas
libbtf	conda-forge/linux-64::libbtf-2.3.2-h32481e8_7100102
libcamd	conda-forge/linux-64::libcamd-3.3.3-h32481e8_7100102
libcbblas	conda-forge/linux-64::libcbblas-3.9.0-31_he106b2a_openblas
libccolamd	conda-forge/linux-64::libccolamd-3.3.4-h32481e8_7100102
libcholmod	conda-forge/linux-64::libcholmod-5.3.1-h59ddab4_7100102
libcolamd	conda-forge/linux-64::libcolamd-3.3.4-h32481e8_7100102
libcxsparse	conda-forge/linux-64::libcxsparse-4.4.1-h32481e8_7100102
libgcc	conda-forge/linux-64::libgcc-14.2.0-h767d61c_2
libgfortran	conda-forge/linux-64::libgfortran-14.2.0-h69a702a_2
libgfortran-ng	conda-forge/linux-64::libgfortran-ng-14.2.0-h69a702a_2
libgfortran5	conda-forge/linux-64::libgfortran5-14.2.0-hf1ad2bd_2
libklu	conda-forge/linux-64::libklu-2.3.5-hf24d653_7100102
liblapack	conda-forge/linux-64::liblapack-3.9.0-31_h7ac8fdf_openblas
libldl	conda-forge/linux-64::libldl-3.3.2-h32481e8_7100102
libopenblas	conda-forge/linux-64::libopenblas-0.3.29-pthreads_h94d23a6_0
libparu	conda-forge/linux-64::libparu-1.0.0-h17147ab_7100102
librbio	conda-forge/linux-64::librbio-4.3.4-h32481e8_7100102
libspex	conda-forge/linux-64::libspex-3.2.3-had10066_7100102
libspqr	conda-forge/linux-64::libspqr-4.3.4-h852d39f_7100102
libstdcxx	conda-forge/linux-64::libstdcxx-14.2.0-h8f9b012_2
libsuitesparsecon~	conda-forge/linux-64::libsuitesparseconfig-7.10.1-h92d6892_7100102
libumfpack	conda-forge/linux-64::libumfpack-6.3.5-heb53515_7100102
metis	conda-forge/linux-64::metis-5.1.0-hd0bc9f9_1007
mpfr	conda-forge/linux-64::mpfr-4.2.1-h90cbb55_3
numpy	conda-forge/linux-64::numpy-2.2.4-py311h5d046bc_0
pyfmi	conda-forge/linux-64::pyfmi-2.16.3-py311h9f3472d_0
python_abi	conda-forge/linux-64::python_abi-3.11-2_cp311
scipy	conda-forge/linux-64::scipy-1.15.2-py311h8f841c2_0
suitesparse	conda-forge/linux-64::suitesparse-7.10.1-ha0f6916_7100102
sundials	conda-forge/linux-64::sundials-7.1.1-ha52427a_0

The following packages will be UPDATED:

conda	pkgs/main::conda-24.11.1-py311h06a430~ --> conda-forge::conda-25.3.0-py311h38be061_0
libgcc-ng	pkgs/main::libgcc-ng-11.2.0-h1234567_1 --> conda-forge::libgcc-ng-14.2.0-h69a702a_2

```
libgomp                pkgs/main::libgomp-11.2.0-h1234567_1 --> conda-forge::libgomp
-14.2.0-h767d61c_2
libstdcxx-ng           pkgs/main::libstdcxx-ng-11.2.0-h12345~ --> conda-forge::libstdc
xx-ng-14.2.0-h4852527_2
openssl                pkgs/main::openssl-3.0.16-h5eee18b_0 --> conda-forge::openssl
-3.4.1-h7b32b05_0
```

The following packages will be SUPERSEDED by a higher-priority channel:

```
certifi                pkgs/main/linux-64::certifi-2025.1.31~ --> conda-forge/noarch::
certifi-2025.1.31-pyhd8ed1ab_0
```

Downloading and Extracting Packages:

```
scipy-1.15.2           | 16.4 MB | : 0% 0/1 [00:00<?, ?it/s]
numpy-2.2.4            | 8.6 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

```
libopenblas-0.3.29    | 5.6 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

```
pyfmi-2.16.3          | 5.2 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

```
metis-5.1.0           | 3.7 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

```
libstdcxx-14.2.0      | 3.7 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

```
openssl-3.4.1         | 2.8 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

```
libgfortran5-14.2.0   | 1.4 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

```
conda-25.3.0          | 1.1 MB  | : 0% 0/1 [00:00<?, ?it/s]
```

assimulo-3.6.0 | 1.1 MB | : 0% 0/1 [00:00<?, ?it/s]

libcholmod-5.3.1 | 1.1 MB | : 0% 0/1 [00:00<?, ?it/s]

sundials-7.1.1 | 907 KB | : 0% 0/1 [00:00<?, ?it/s]

libgcc-14.2.0 | 828 KB | : 0% 0/1 [00:00<?, ?it/s]

mpfr-4.2.1 | 620 KB | : 0% 0/1 [00:00<?, ?it/s]



gmp-6.3.0 | 449 KB | : 0% 0/1 [00:00<?, ?it/s]

libgomp-14.2.0 | 449 KB | : 0% 0/1 [00:00<?, ?it/s]

libumfpack-6.3.5 | 424 KB | : 0% 0/1 [00:00<?, ?it/s]

fmilib-2.4.1	383 KB	:	0% 0/1 [00:00<?, ?it/s]
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libspqr-4.3.4	213 KB	:	0% 0/1 [00:00<?, ?it/s]
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scipy-1.15.2 0s/it]	16.4 MB	:	0% 0.0019058779654147827/1 [00:00<00:52, 53.0
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pyfmi-2.16.3 s/it]	5.2 MB	:	1% 0.005967906113297332/1 [00:00<00:17, 17.55
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metis-5.1.0 s/it]	3.7 MB	:	0% 0.004175799528999174/1 [00:00<00:26, 26.28
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numpy-2.2.4 5s/it]	8.6 MB	:	0% 0.0018194029373407579/1 [00:00<01:03, 63.7
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scipy-1.15.2 s/it]	16.4 MB	:	14% 0.13722321350986436/1 [00:00<00:01, 1.26
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pyfmi-2.16.3	5.2 MB	:	52% 0.5162238788002192/1 [00:00<00:00, 2.97i
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t/s] numpy-2.2.4 t/s]	8.6 MB	: 28% 0.28200745528781745/1 [00:00<00:00, 1.55i
metis-5.1.0 t/s]	3.7 MB	: 71% 0.7098859199298596/1 [00:00<00:00, 3.99i
scipy-1.15.2 t/s]	16.4 MB	: 35% 0.3525874236017348/1 [00:00<00:00, 1.41i
numpy-2.2.4 t/s]	8.6 MB	: 79% 0.7859820689312074/1 [00:00<00:00, 3.06i
metis-5.1.0	3.7 MB	: 100% 1.0/1 [00:00<00:00, 3.99it/s]
libstdcxx-14.2.0 2s/it]	3.7 MB	: 0% 0.0042177278432850495/1 [00:00<01:24, 84.8
pyfmi-2.16.3	5.2 MB	: 100% 1.0/1 [00:00<00:00, 2.70it/s]
scipy-1.15.2 t/s]	16.4 MB	: 57% 0.5736692675898496/1 [00:00<00:00, 1.71i
libopenblas-0.3.29	5.6 MB	: 100% 1.0/1 [00:00<00:00, 2.69it/s]
libopenblas-0.3.29	5.6 MB	: 100% 1.0/1 [00:00<00:00, 2.69it/s]
openssl-3.4.1 7s/it]	2.8 MB	: 1% 0.0055741049077571376/1 [00:00<01:20, 80.9
scipy-1.15.2 t/s]	16.4 MB	: 77% 0.7699746980275722/1 [00:00<00:00, 1.77i
libgfortran5-14.2.0	1.4 MB	: 100% 1.0/1 [00:00<00:00, 40.39s/it]

libstdcxx-14.2.0	3.7 MB	: 100% 1.0/1 [00:00<00:00, 2.33it/s]
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libstdcxx-14.2.0	3.7 MB	: 100% 1.0/1 [00:00<00:00, 2.33it/s]
numpy-2.2.4	8.6 MB	: 100% 1.0/1 [00:00<00:00, 3.06it/s]

conda-25.3.0	1.1 MB	: 1% 0.01395284272989957/1 [00:00<00:39, 40.28s/it]
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scipy-1.15.2	16.4 MB	: 95% 0.947221348811147/1 [00:00<00:00, 1.76it/s]
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libcholmod-5.3.1	1.1 MB	: 1% 0.014870549794649543/1 [00:00<00:39, 40.13s/it]
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assimulo-3.6.0	1.1 MB	: 100% 1.0/1 [00:00<00:00, 39.51s/it]
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openssl-3.4.1	2.8 MB	: 100% 1.0/1 [00:00<00:00, 2.00it/s]
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openssl-3.4.1 | 2.8 MB | : 100% 1.0/1 [00:00<00:00, 2.00it/s]

conda-25.3.0 | 1.1 MB | : 100% 1.0/1 [00:00<00:00, 40.28s/it]

libcholmod-5.3.1 | 1.1 MB | : 100% 1.0/1 [00:00<00:00, 40.13s/it]

sundials-7.1.1 | 907 KB | : 2% 0.01763373830085844/1 [00:00<00:37, 38.66  
s/it]

libgcc-14.2.0 | 828 KB | : 2% 0.01932337522187561/1 [00:00<00:35, 35.73  
s/it]

mpfr-4.2.1 | 620 KB | : 3% 0.025811696239942908/1 [00:00<00:26, 26.91  
s/it]

gmp-6.3.0 | 449 KB | : 4% 0.03561313321233331/1 [00:00<00:19, 19.73  
s/it]

gmp-6.3.0 | 449 KB | : 100% 1.0/1 [00:00<00:00, 19.73s/it]

mpfr-4.2.1 | 620 KB | : 100% 1.0/1 [00:00<00:00, 26.91s/it]

libgcc-14.2.0 | 828 KB | : 100% 1.0/1 [00:00<00:00, 35.73s/it]

sundials-7.1.1 | 907 KB | : 100% 1.0/1 [00:00<00:00, 38.66s/it]

libgomp-14.2.0 | 449 KB | : 4% 0.03562807972826631/1 [00:00<00:20, 21.21  
s/it]

libumfpack-6.3.5 | 424 KB | : 4% 0.037731330084655984/1 [00:00<00:19, 20.36  
s/it]

libgomp-14.2.0 | 449 KB | : 100% 1.0/1 [00:00<00:00, 21.21s/it]

libumfpack-6.3.5 | 424 KB | : 100% 1.0/1 [00:00<00:00, 20.36s/it]

libspqr-4.3.4 | 213 KB | : 8% 0.07503068271326775/1 [00:00<00:09, 10.45  
s/it]



fmilib-2.4.1 | 383 KB | : 4% 0.04180391656566945/1 [00:00<00:18, 18.84  
s/it]

libspqr-4.3.4 | 213 KB | : 100% 1.0/1 [00:00<00:00, 10.45s/it]

fmilib-2.4.1 | 383 KB | : 100% 1.0/1 [00:00<00:00, 18.84s/it]

metis-5.1.0 | 3.7 MB | : 100% 1.0/1 [00:00<00:00, 3.99it/s]

... (more hidden) ...

... (more hidden) ...

scipy-1.15.2	16.4 MB	: 100% 1.0/1 [00:01<00:00, 1.76it/s]
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libgfortran5-14.2.0	1.4 MB	: 100% 1.0/1 [00:01<00:00, 1.04it/s]
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libgfortran5-14.2.0	1.4 MB	: 100% 1.0/1 [00:01<00:00, 1.04it/s]
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libstdcxx-14.2.0	3.7 MB	: 100% 1.0/1 [00:01<00:00, 2.33it/s]
------------------	--------	--------------------------------------

libopenblas-0.3.29	5.6 MB	: 100% 1.0/1 [00:01<00:00, 2.69it/s]
--------------------	--------	--------------------------------------

assimulo-3.6.0 | 1.1 MB | : 100% 1.0/1 [00:01<00:00, 1.39s/it]

assimulo-3.6.0 | 1.1 MB | : 100% 1.0/1 [00:01<00:00, 1.39s/it]

openssl-3.4.1 | 2.8 MB | : 100% 1.0/1 [00:01<00:00, 2.00it/s]

conda-25.3.0 | 1.1 MB | : 100% 1.0/1 [00:02<00:00, 2.11s/it]

conda-25.3.0 | 1.1 MB | : 100% 1.0/1 [00:02<00:00, 2.11s/it]

libcholmod-5.3.1 | 1.1 MB | : 100% 1.0/1 [00:02<00:00, 2.17s/it]

libcholmod-5.3.1 | 1.1 MB | : 100% 1.0/1 [00:02<00:00, 2.17s/it]

gmp-6.3.0 | 449 KB | : 100% 1.0/1 [00:02<00:00, 2.21s/it]

gmp-6.3.0 | 449 KB | : 100% 1.0/1 [00:02<00:00, 2.21s/it]

mpfr-4.2.1 | 620 KB | : 100% 1.0/1 [00:02<00:00, 2.30s/it]

mpfr-4.2.1 | 620 KB | : 100% 1.0/1 [00:02<00:00, 2.30s/it]

libgcc-14.2.0 | 828 KB | : 100% 1.0/1 [00:02<00:00, 2.42s/it]

libgcc-14.2.0 | 828 KB | : 100% 1.0/1 [00:02<00:00, 2.42s/it]

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libgomp-14.2.0 | 449 KB | : 100% 1.0/1 [00:02<00:00, 2.72s/it]

libgomp-14.2.0 | 449 KB | : 100% 1.0/1 [00:02<00:00, 2.72s/it]

libumfpack-6.3.5 | 424 KB | : 100% 1.0/1 [00:02<00:00, 2.76s/it]

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libspqr-4.3.4	213 KB	: 100% 1.0/1 [00:02<00:00, 2.81s/it]
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libspqr-4.3.4	213 KB	: 100% 1.0/1 [00:02<00:00, 2.81s/it]
numpy-2.2.4	8.6 MB	: 100% 1.0/1 [00:03<00:00, 3.06it/s]

fmilib-2.4.1	383 KB	: 100% 1.0/1 [00:03<00:00, 2.87s/it]
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fmilib-2.4.1 | 383 KB | : 100% 1.0/1 [00:03<00:00, 2.87s/it]

... (more hidden) ...

scipy-1.15.2 | 16.4 MB | : 100% 1.0/1 [00:04<00:00, 1.76it/s]















```
Preparing transaction: - 00\ 00| 00done
Verifying transaction: - 00\ 00| 00/ 00- 00done
Executing transaction: | 00/ 00- 00\ 00| 00/ 00- 00\ 00| 00/ 00- 00\ 00| 00/ 00- 00\
00| 00/ 00- 00\ 00| 00/ 00done
```

## Preparation of BPL\_IEC\_operation

Now specific installation and the run simulations. Start with connecting to Github. Then upload the two files:

- FMU - BPL\_IEC\_Column\_system\_linux\_om\_me
- Setup-file - BPL\_IEC\_explore

```
In [9]: %%bash
git clone https://github.com/janpeter19/CONF_2023_08_NPCW24
```

```
Cloning into 'CONF_2023_08_NPCW24'...
```

```
In [10]: %cd CONF_2023_08_NPCW24
/content/CONF_2023_08_NPCW24
```

## BPL\_IEC\_operation

Authors: Karl Johan Brink and Jan Peter Axelsson

In this notebook we show operation of a typical ion-exchange chromatography step. The impact of pH is also illustrated.

The model is based on the simplified model [1].

```
In [11]: run -i BPL_IEC_explore.py
```

Linux - run FMU pre-compiled OpenModelica

Model for the process has been setup. Key commands:

- par() - change of parameters and initial values
- init() - change initial values only
- simu() - simulate and plot
- newplot() - make a new plot
- show() - show plot from previous simulation
- disp() - display parameters and initial values from the last simulation
- describe() - describe culture, broth, parameters, variables with values/units

Note that both disp() and describe() takes values from the last simulation and the command process\_diagram() brings up the main configuration

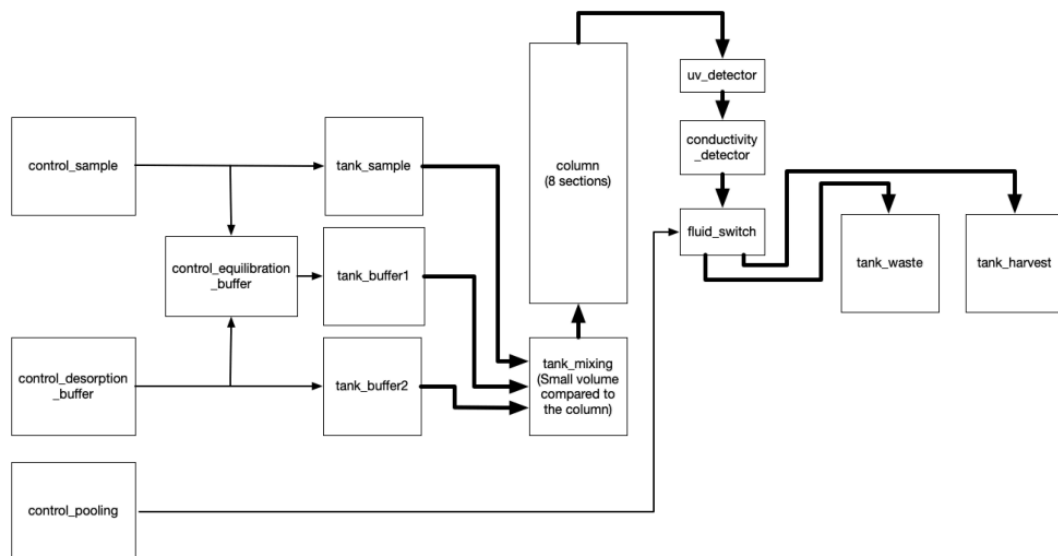
Brief information about a command by help(), eg help(simu)

Key system information is listed with the command system\_info()

```
In [12]: %matplotlib inline
plt.rcParams['figure.figsize'] = [36/2.54, 30/2.54]
```

```
In [13]: process_diagram()
```

No processDiagram.png file in the FMU, but try the file on disk.



## 1 Typical parameters for a pilot scale ion exchange chromatography column process setup

```
In [14]: # From given column height (h) diameter (d) and Linear flow rate (Lfr)
# actual column volume (V) and volume flow rate (VFR) are calculated below.

from numpy import pi
h = 20.0
d = 1.261
a = pi*(d/2)**2
```



```

V = h*a
print('V =', np.round(V,1), '[mL]')

lfr = 48
VFR = a*lfr/60
print('VFR =', np.round(VFR,1), '[mL/min]') # Pumps

```

V = 25.0 [mL]  
VFR = 1.0 [mL/min]

```

In [15]: # Sample concentration product P_in and antagonist A_in
par(P_in = 1.0)
par(A_in = 1.0)
par(E_in = 0.0)

# Column properties are described by the size and binding capacity of the resin Q_a
par(height = h)
par(diameter = d)
par(Q_av = 6.0)

# Remaining salt koncentration in the column from prvious batch and eliminated duri
init(E_0 = 50)

# Salt koncentration of the desorption buffer
par(E_in_desorption_buffer = 8.0)

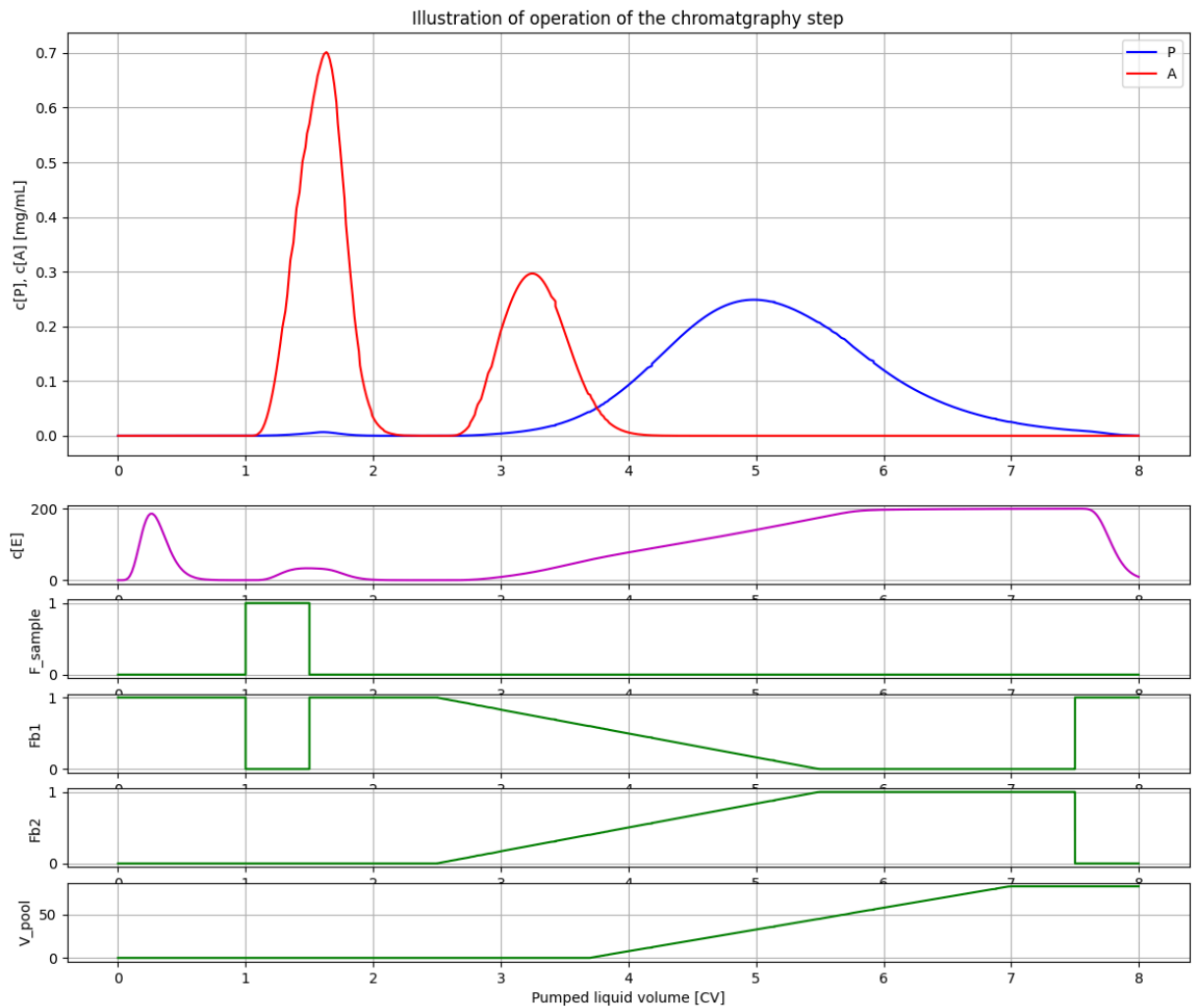
# Flow rate rate through the
par(LFR=lfr)

# Switching points during operation are conveniently described in terms of multiple
CV_ekv = 1.0
CV_ads = 0.5
CV_wash = 1.0
CV_desorb = 3.0
CV_start_pool = 1.2
CV_stop_pool = 4.5
CV_ekv2 = 2.5
par(scale_volume=True, start_adsorption=CV_ekv*V, stop_adsorption=(CV_ekv+CV_ads)*V
par(start_desorption=(CV_ekv+CV_ads+CV_wash)*V, stationary_desorption=(CV_ekv+CV_ad
par(stop_desorption=7.5*V)
par(start_pooling=(CV_ekv+CV_ads+CV_wash+CV_start_pool)*V, stop_pooling=(CV_ekv+CV_

# Simulation and plot of results
newplot(title='Illustration of operation of the chromatgraphy step', plotType='Elut
simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)

```

Could not find cannot import name 'dopri5' from 'assimulo.lib' (/usr/local/lib/python3.11/site-packages/assimulo/lib/\_\_init\_\_.py)  
 Could not find cannot import name 'rodas' from 'assimulo.lib' (/usr/local/lib/python3.11/site-packages/assimulo/lib/\_\_init\_\_.py)  
 Could not find cannot import name 'odassl' from 'assimulo.lib' (/usr/local/lib/python3.11/site-packages/assimulo/lib/\_\_init\_\_.py)  
 Could not find ODEPACK functions.  
 Could not find RADAR5  
 Could not find GLIMDA.



Comments of steps of operations:

1. Time: 0-1 hours - equilibration. Just to illustrate the equilibration process the first part of the column is given an initial value of salt concentration.
2. Time: 1-1.5 hours - sample is loaded on the column. The product P is adsorbed to the column and just a small amount passes through and goes to the waste. The antagonist A is much less adsorbed.
3. Time: 1.5-2.5 hours - washing 1. The column comes to equilibrium and both antagonist and product comes down to low levels.
4. Time: 2.5-5.5 hours - desorption. A linear gradient of increasing salt concentration is applied. First the antagonist and later the product comes out.
5. Time: 5.5-7.5 hours - washing 2. The column has constant salt concentration and stationary desorption.
6. Time: 3.7-7.0 hours - pooling of product. The start- and stop of pooling are chosen with trade-off between maximizing the product pooled and minimize the amount of antagonist in the pooling.

7. Time: 7.5-8.0 hours - desorption stopped and salt is washed out and preparation of the next batch to come.

Note that step 4 and 5 is parallel to step 6.

```
In [16]: # Check mass-balance of P and A
P_mass = model.get('tank_harvest.m[1]') + model.get('tank_waste.m[1]')
A_mass = model.get('tank_harvest.m[2]') + model.get('tank_waste.m[2]')
print('P_mass [mg] =', P_mass)
print('A_mass [mg] =', A_mass)
```

```
P_mass [mg] = [12.42212162]
A_mass [mg] = [12.48878113]
```

These values should be compared with the expected value 12.5 mg, i.e. half a column volume with sample concentration 1 mg/L. The difference is due to numerical errors during simulation.

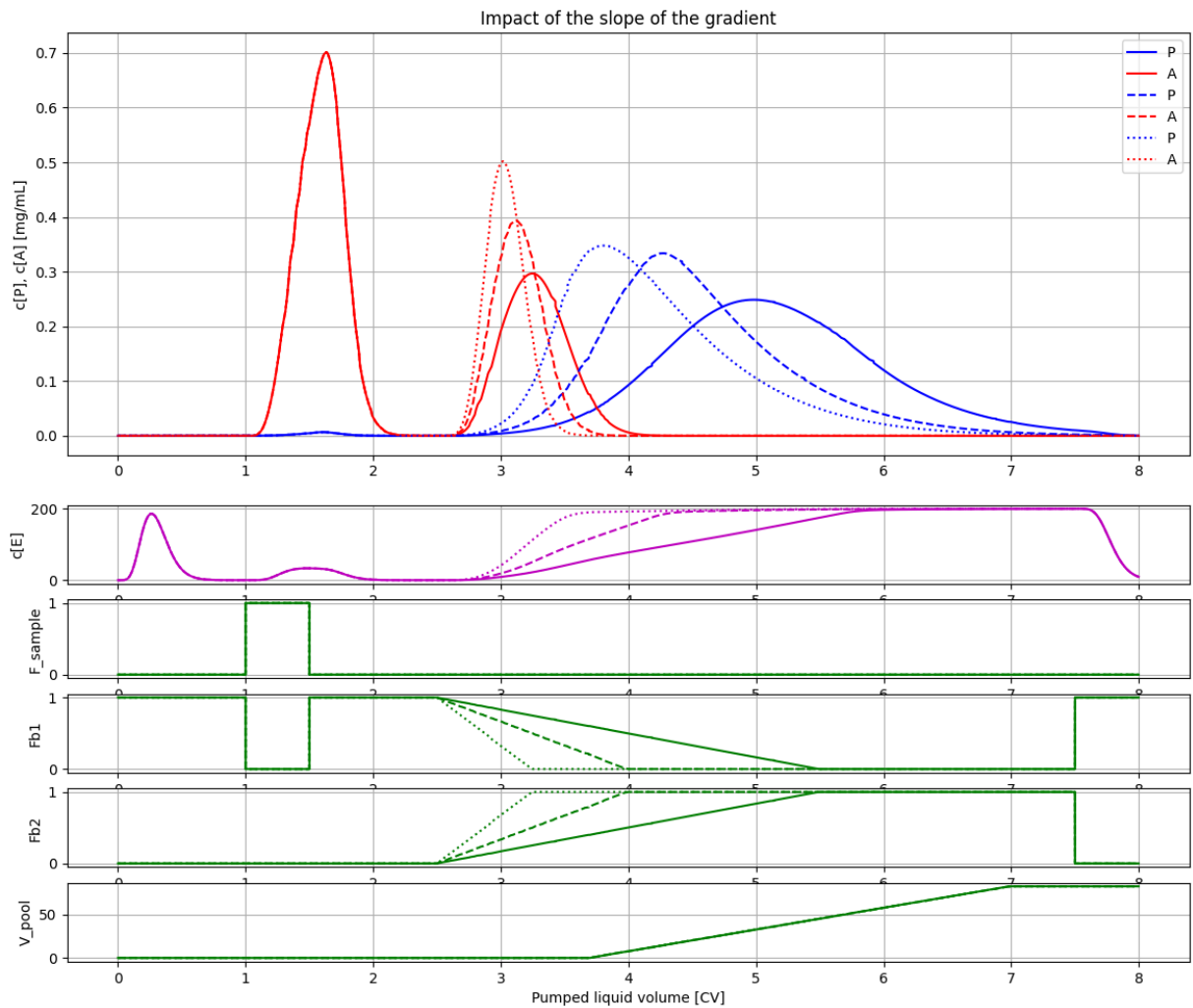
## 2 The impact of the slope of the desorption gradient

```
In [17]: # Simulations showing the impact of change of slope of the desorption gradient
newplot(title='Impact of the slope of the gradient', plotType='Elution-conductivity')

# Same gradient as before
par(start_desorption=(CV_ekv+CV_ads+CV_wash)*V, stationary_desorption=(CV_ekv+ CV_a
par(stop_desorption=7.5*V)
simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)

# Gradient finishes after 0.5 of the volume
par(stationary_desorption = (CV_ekv + CV_ads + CV_wash + 0.5*CV_desorb)*V )
simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)

# Gradient finishes after 0.25 of the volume
par(stationary_desorption = (CV_ekv + CV_ads + CV_wash + 0.25*CV_desorb)*V )
simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)
```



Note the pens shift style for each simulation in the order: solid, dashed, dotted, dash-dotted.  
The actual simulations done you see in the preceeding cell.

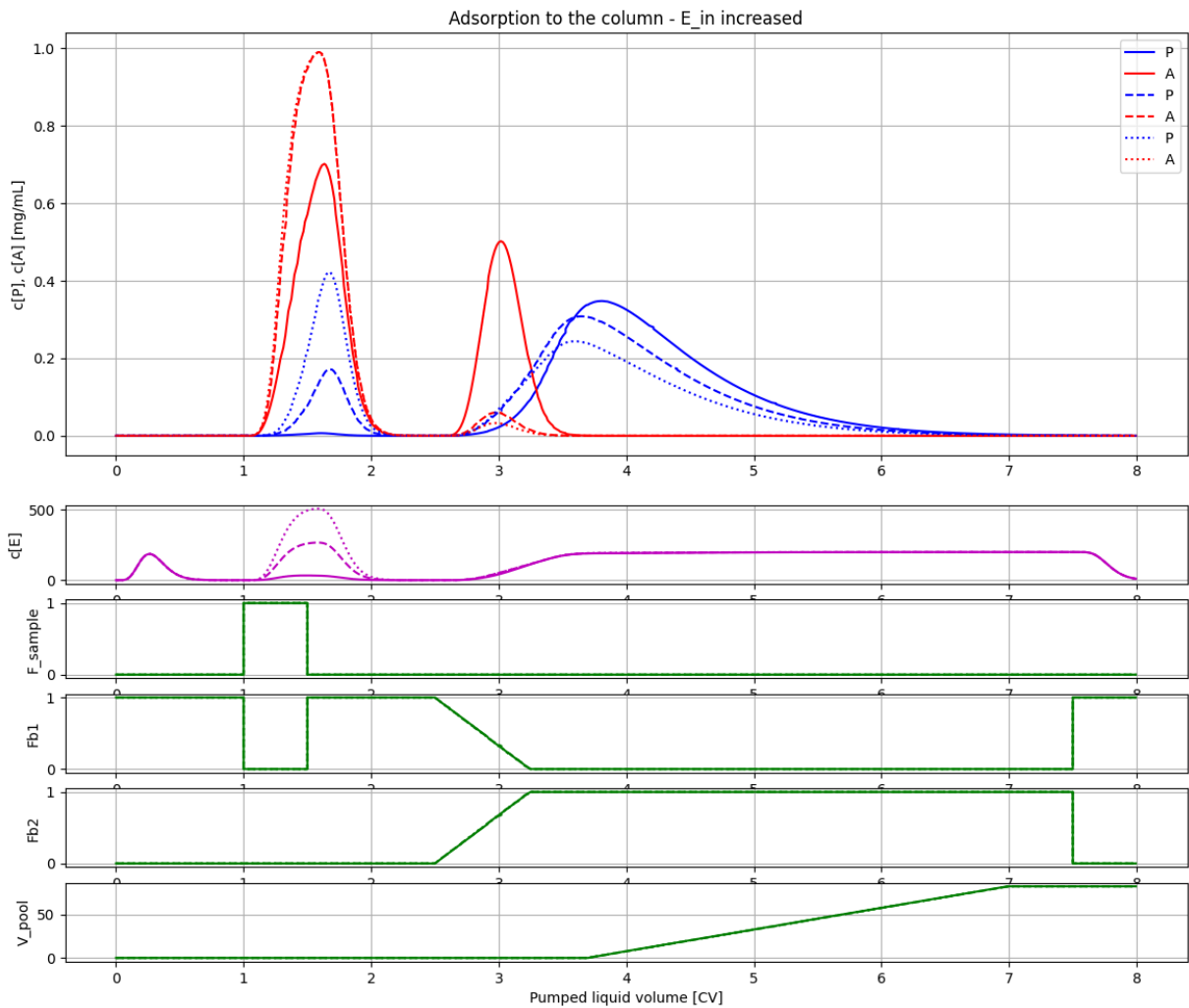
### 3 The impact of salt concentration in the sample

```
In [18]: # Let us investigate the impact of increasing salt concentration in the sample E_in

# Simulate and plot the results
newplot(title='Adsorption to the column - E_in increased', plotType='Elution-conduc

for value in [0, 10, 20]:
    par(E_in=value)
    simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)

# Restore default values
par(k2=0.05, k4=0.3, E_in=0)
```



Note, that increased salt concentration in the sample affect binding of both proteins. During adsorption less is bound. During desorption less product P can be harvested but the fraction of antagonist A may be lowered. Thus, some product is lost but the quality in terms of purity is improved.

## 4 The impact of change of binding strength due to pH

There are many factors that contribute to the binding strength. A most important factor is the pH-value of the resin and the characteristic iso-electric point of the protein. The binding strenght can be seen as proportional to the difference.

The binding strength of the resin is described by the quotient  $K_P = k_1/k_2$  for the protein P and similarly  $K_A = k_3/k_4$  for the protein A.

Below a few help-functions that describe this idea of the pH difference and its impact on binding strength in terms of the parameters  $k_1$ ,  $k_2$ ,  $k_3$ , and  $k_4$  of the protein-resin interaction.

```
In [19]: # Define function that describe the proportionality of binding strength of
# the pH difference of the iso-electric point and the resin

def KP_pH_sensitivity(pI_P=8.0, pH_resin=7.0):
    coeff_pH = 6.0
    return coeff_pH*(pI_P-pH_resin)

def KA_pH_sensitivity(pI_A=7.1667, pH_resin=7.0):
    coeff_pH = 1.0
    return coeff_pH*(pI_A-pH_resin)

def par_pH(pI_P=8.0, pI_A=7.1667, pH_resin=7.0, TP=3.33, TA=20.0):
    if (pI_P > pH_resin) & (pI_A > pH_resin):
        par(k2 = 1/(TP*KP_pH_sensitivity(pI_P=pI_P, pH_resin=pH_resin)))
        par(k4 = 1/(TA*KA_pH_sensitivity(pI_A=pI_A, pH_resin=pH_resin)))
    else:
        print('Both pI_P > pH_resin and pI_A > pH_resin must hold - no parameter ch
```

```
In [20]: # The default parameters of the column
disp('column')
```

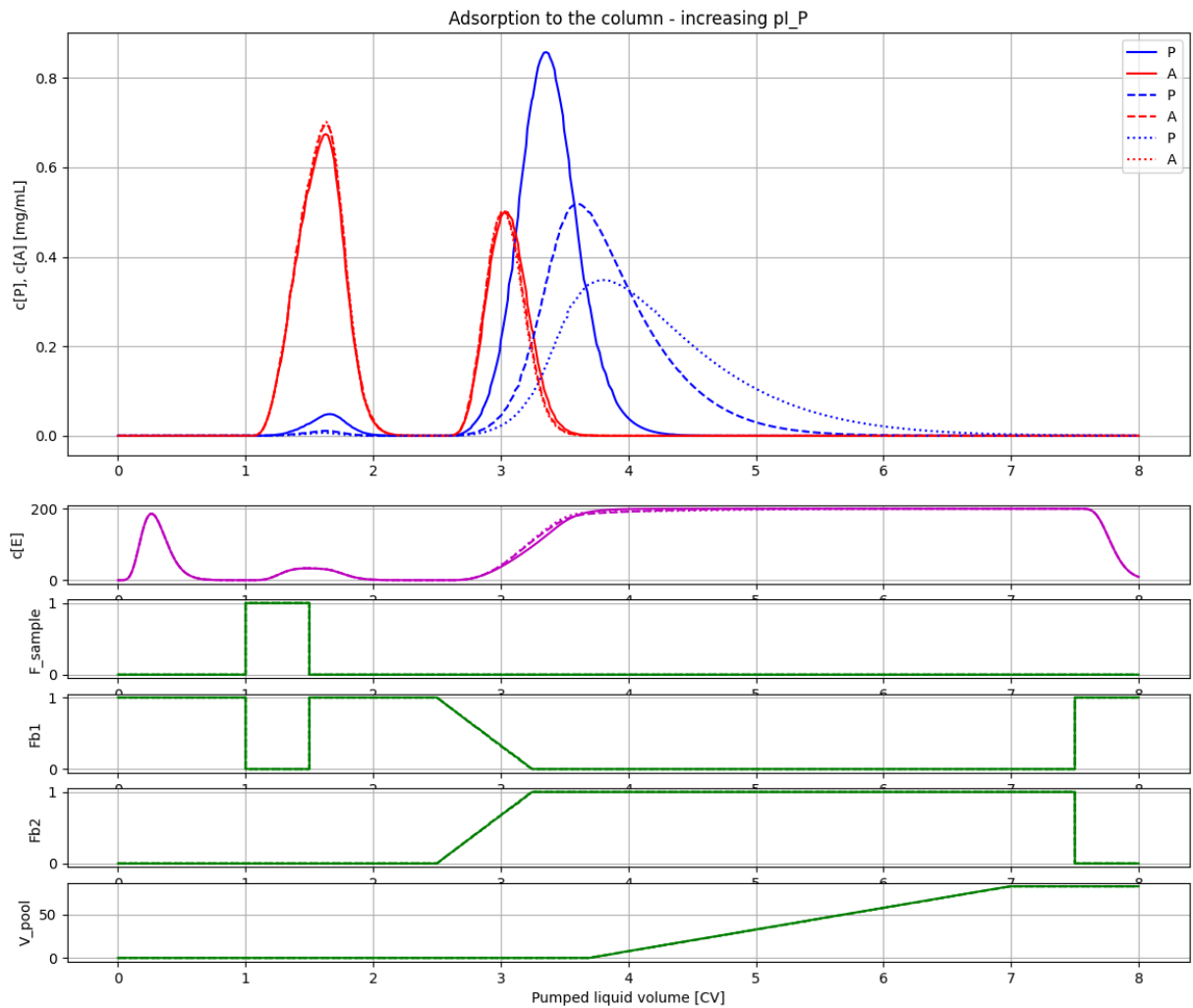
```
diameter : 1.261
height : 20.0
x_m : 0.3
k1 : 0.3
k2 : 0.05
k3 : 0.05
k4 : 0.3
Q_av : 6.0
E_0 : 50.0
```

```
In [21]: # Let us investigate the impact of change of the iso-electric pH for protein P

# Simulate and plot the results
newplot(title='Adsorption to the column - increasing pI_P', plotType='Elution-condu

for value in [7.2, 7.6, 8.0]:
    par_pH(pI_P=value, pI_A=7.1667, pH_resin=7.0)
    simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)

# Restore default values
par(k2 = 0.05, k4 = 0.3)
```



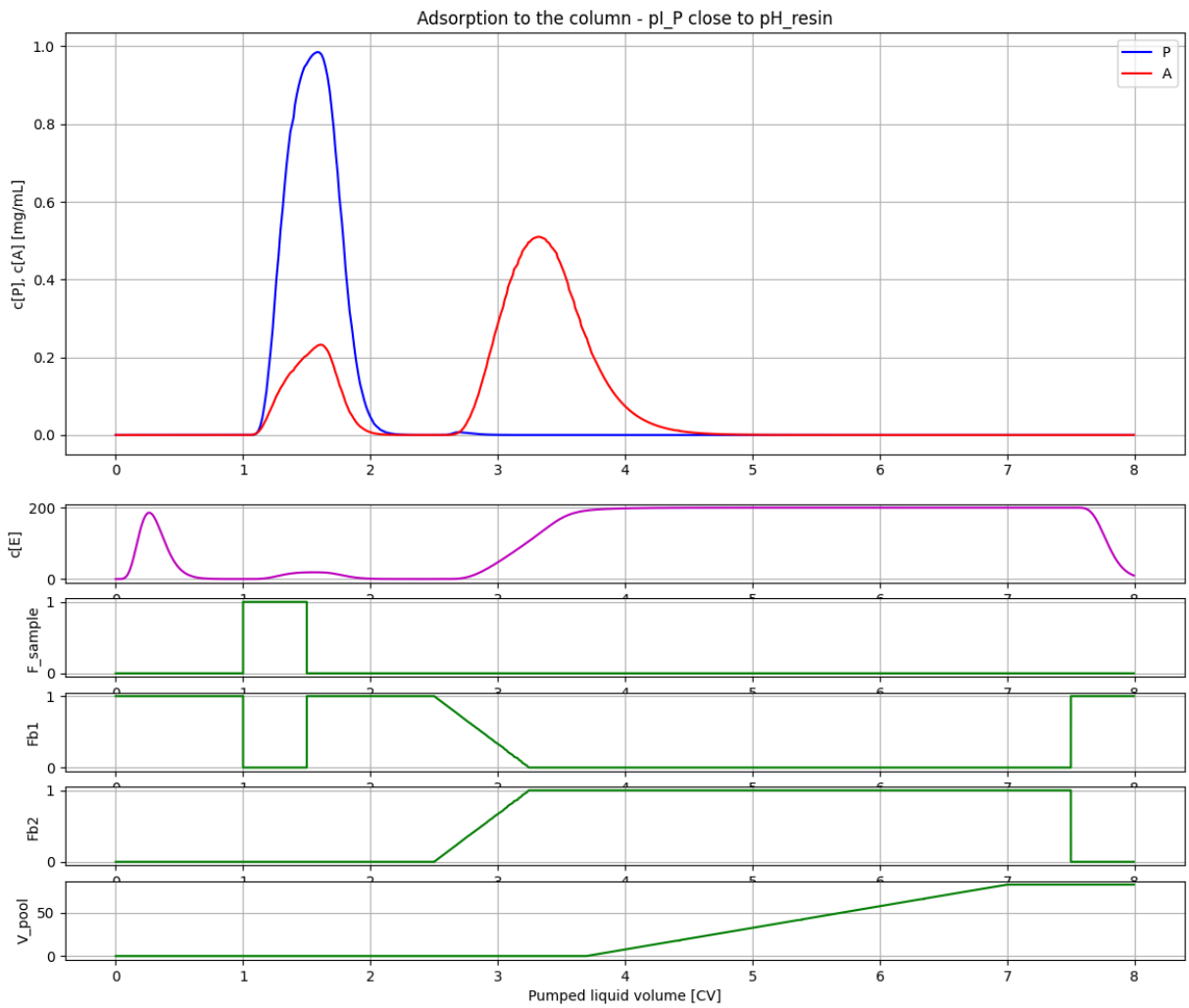
Note, with increasing  $pI_P$  the binding of P increase which leads less loss of product during adsorption. During desorption the peak height is lower with increasing binding strenght, but the total amoiunt of product P that can be harvested is higher, due to the smaller loss during adsorption.

```
In [22]: # Let us investigate the impact of pI_P close to pH_resin

# Simulate and plot the results
newplot(title='Adsorption to the column - pI_P close to pH_resin', plotType='Elutio

for value in [7.0001]:
    par_pH(pI_P=value, pI_A=8)
    simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)

# Restore default values
par(k2=0.05, k4=0.3)
```



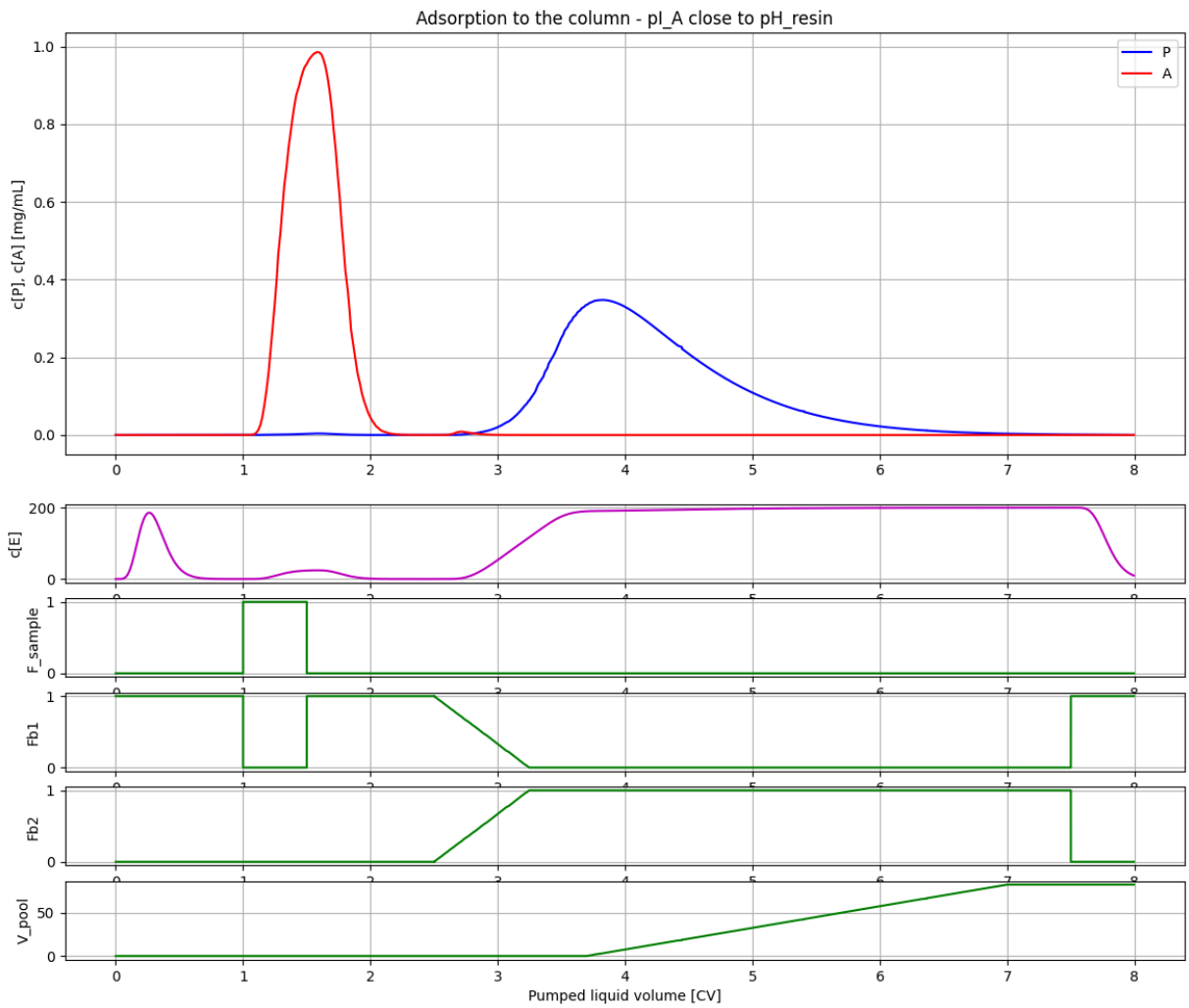
```
In [23]: # Let us investigate the impact of  $pI_A$  close to  $pH_{resin}$ 

# Simulate and plot the results
newplot(title='Adsorption to the column -  $pI_A$  close to  $pH_{resin}$ ', plotType='Elutio

for value in [7.001]:
    par_pH( $pI_P=8.0$ ,  $pI_A=value$ )
    simu(( $CV_{ekv}+CV_{ads}+CV_{wash}+CV_{desorb}+CV_{ekv2}$ )* $V/VFR$ )

# Restore default values
par( $k2=0.05$ ,  $k4=0.3$ )
```



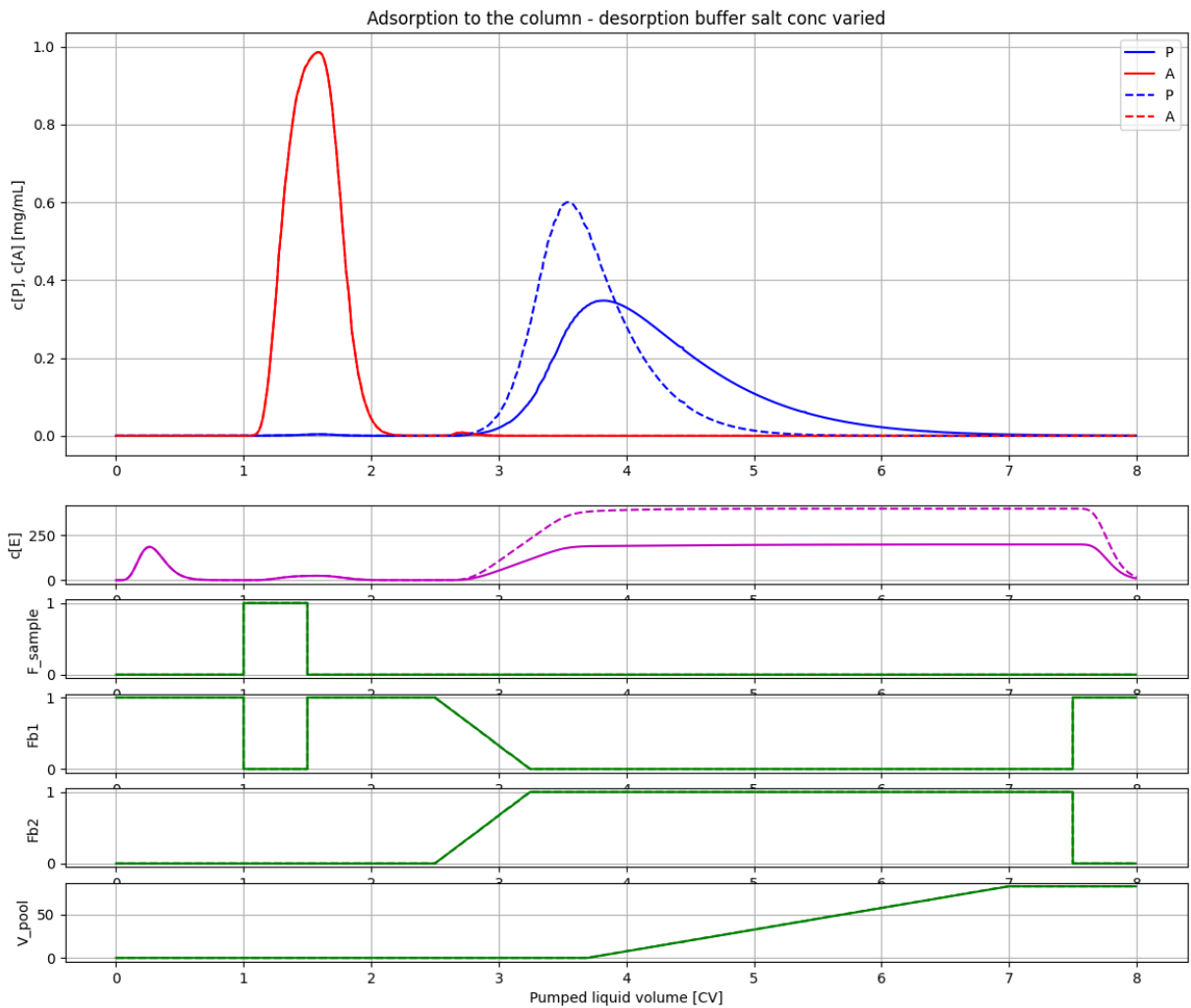


```
In [24]: # Let us also investigate the impact of salt concentration of the desorption buffer

# Simulate and plot the results
newplot(title='Adsorption to the column - desorption buffer salt conc varied', plot

for value in [8.0, 16.0]:
    par(E_in_desorption_buffer=value)
    par_pH(pI_P=8.0, pI_A=7.001, pH_resin=7.0)
    simu((CV_ekv+CV_ads+CV_wash+CV_desorb+CV_ekv2)*V/VFR)

# Restore default values
par(E_in_desorption_buffer=8.0)
par(k2=0.05, k4=0.3)
```



## 5 Breakthrough curve often used during process development

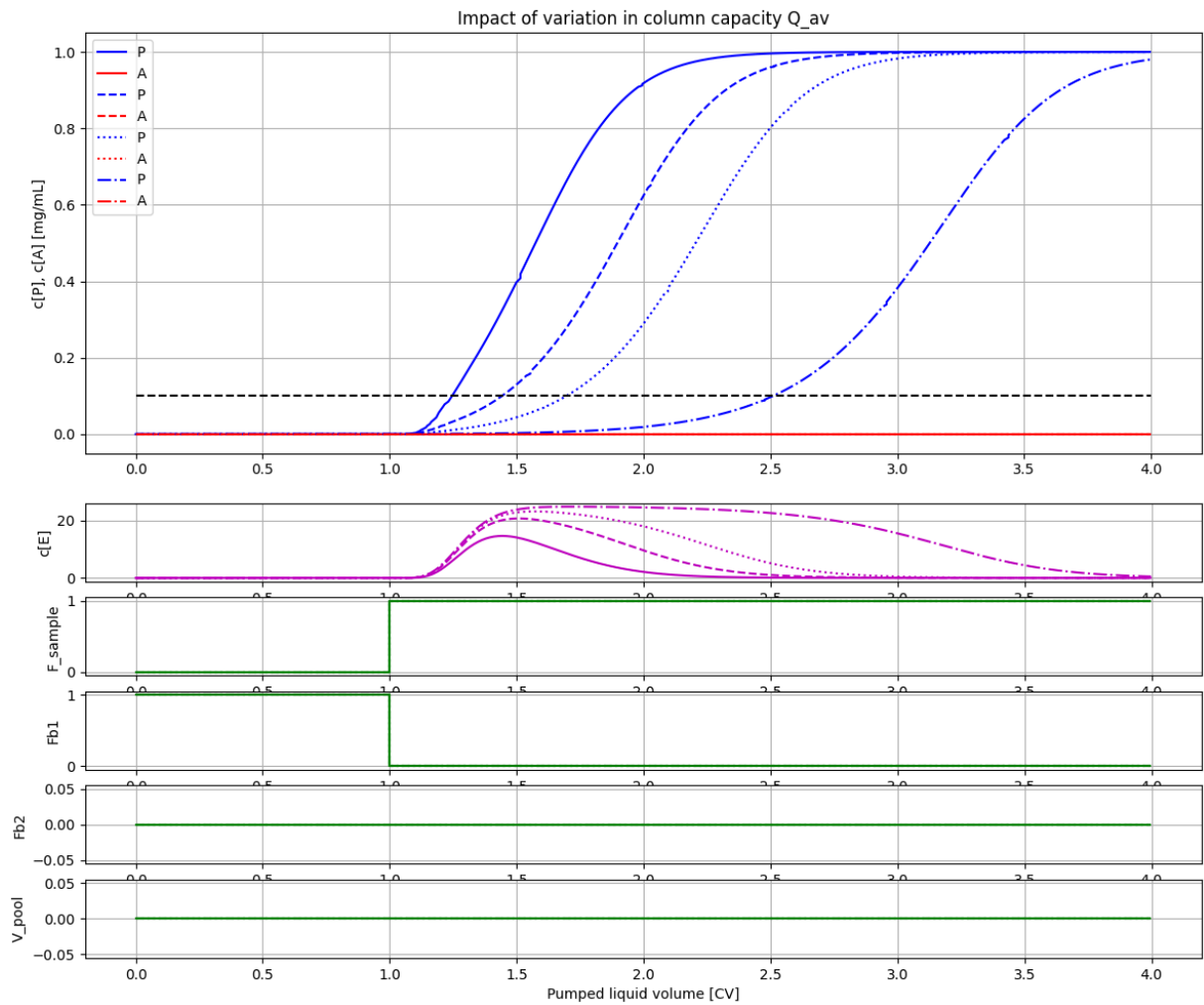
```
In [25]: # Experiment to check column capacity  $Q_{av}$  often called breakthrough curve
par(P_in=1, A_in=0, E_in=0)
init(E_0 = 0)
par( $Q_{av}$ =6.0)

par(scale_volume=True, start_adsorption=1*V, stop_adsorption=4.01*V)
par(start_desorption=10*V, stationary_desorption=10.5*V, stop_desorption=11*V)
par(start_pooling=11*V, stop_pooling=12*V)

newplot(title='Impact of variation in column capacity  $Q_{av}$ ', plotType='Elution-cond
for value in [1, 2, 3, 6]: par( $Q_{av}$ =value); simu(4.0*V/VFR)

# Linje för 10% UV
ax1.plot([0,4], [0.1,0.1], 'k--')

# Restore default parameters
par( $Q_{av}$ =6.0, A_in=1.0)
```



With greater column capacity  $Q_{av}$  the longer it takes before the concentration of protein start to increase. Note, that the salt concentration increase initially during adsorption but then go back to low levels. This phenomenon is also seen experimentally.

## 6 Summary

The simplified simulation model was found useful to describe operational aspects of ion exchange chromatography.

- The model describe qualitatively well the impact of typical operational changes in the flow rate.
- The model also describe qualitatively well the impact of changes in iso-electric point of the proteins relative the pH of the resin.
- The small deviations in salt concentration from linear increase during the gradient in the salt buffer is also what you see in reality.

## References

1. Månsson, Jonas, "Control of chromatography column in production scale", Master thesis TFRT-5599, Department of Automatic Control, LTH, Lund Sweden, 1998.
2. Pharmacia LKB Biotechnology. "Ion Exchange chromatography. Principles and Methods.", 3rd edition, 1991.
3. Jungbauer, Alois and Giorgio Carta, "Protein Chromatography: Process Development and Scale-Up", Wiley 2nd edition, 2020.

## Appendix

```
In [26]: describe('MSL')
```

```
MSL: 3.2.3 - used components: RealInput, RealOutput, CombiTimeTable, Types
```

```
In [27]: system_info()
```

```
System information
-OS: Linux
-Python: 3.11.11
-Scipy: not installed in the notebook
-PyFMI: 2.16.3
-FMU by: OpenModelica Compiler OpenModelica 1.21.0
-FMI: 2.0
-Type: FMUModelME2
-Name: BPL_IEC.Column_system
-Generated: 2023-04-21T10:58:26Z
-MSL: 3.2.3
-Description: Bioprocess Library version 2.1.1
-Interaction: FMU-explore version 0.9.8
```

```
In [27]:
```