

CELLULAR SIGNALING MECHANISMS UNDERLYING THE
ANGIOGENIC RESPONSE TO MYCOBACTERIAL
INFECTION

by

William Jared Brewer

Department of Molecular Genetics and Microbiology
Duke University

Date: _____
Approved:

David M. Tobin, Supervisor

J. Andrew Alspaugh

John F. Rawls

Christopher Kontos

Dennis Ko

Dissertation submitted in partial fulfillment of the
requirements for the degree of Doctor of Philosophy
in the Department of Molecular Genetics and Microbiology
in the Graduate School of
Duke University

2022

ABSTRACT

CELLULAR SIGNALING MECHANISMS UNDERLYING THE ANGIOGENIC RESPONSE TO MYCOBACTERIAL INFECTION

by

William Jared Brewer

Department of Molecular Genetics and Microbiology
Duke University

Date: _____
Approved:

David M. Tobin, Supervisor

J. Andrew Alspaugh

John F. Rawls

Christopher Kontos

Dennis Ko

An abstract of a dissertation submitted in partial fulfillment of the
requirements for the degree of Doctor of Philosophy
in the Department of Molecular Genetics and Microbiology
in the Graduate School of
Duke University

2022

Copyright © 2022 by William Jared Brewer
All rights reserved

Abstract

Pathological angiogenesis is a widespread biological phenomenon that influences the progression of various diseases, including autoimmune conditions, cancers, and microbial infections. One infection in particular, tuberculosis, is associated with the induction of a potent pro-angiogenic signaling cascade that facilitates bacterial growth and accelerates disease progression. A synthesis of early studies on bacterial factors that drive host angiogenesis with modern genetic findings identified the mycobacterial glycolipid trehalose 6-6'-dimycolate (TDM) as a critical factor driving vascular endothelial growth factor (VEGFA) production and angiogenesis during mycobacterial infection. Despite these recent findings, many of the underlying host response mechanisms remain unknown.

The introductory chapter will serve to introduce the reader to the major concepts addressed in this work: *Mycobacterium tuberculosis* and the disease it causes, the role of macrophages in health and disease, the function of pattern recognition receptors in detecting microbial ligands, the specific downstream intracellular signaling pathway of interest for this work (mediated by the transcription factor, nuclear factor of activated T cells, NFAT), the contributions of angiogenesis to diverse contexts and pathologies, and the promise of host-directed therapies to overcome challenges associated with traditional treatment approaches in infectious disease.

Chapter 2 describes the new and existing methodological approaches that were

required to complete this work. This work utilizes the zebrafish-*Mycobacterium marinum* model of tuberculosis infection to facilitate in depth *in vivo* observation and quantitation of these phenomena. Using this model in tandem with human macrophage cell culture, I was able to model major aspects of the host-pathogen interface, enabling me to identify a critical role for a macrophage-C-type lectin receptor-NFATC2-VEGFA signaling axis required for the angiogenic response to mycobacterial infection and TDM, findings that comprise the core of this work and are detailed at length in Chapter 3.

The analysis of the large amounts of data generated in this work required creative approaches to data processing and analysis. To this end, I have developed a set of novel processing modalities in Python and R that are capable of the rapid and reproducible processing of images as well as certain aspects of automated data collection therefrom. These macros, many written for the FIJI/ImageJ programming environment, serve as the infrastructure on which the rest of this work has been built. These will be detailed in Chapter 4.

Finally, this body of work leaves many questions as yet unanswered. While it is clear that NFAT signaling is required for VEGFA production, the precise mechanism by which this may work is unclear and could be mediated by either direct DNA binding or indirect activation or cooperative binding with some other transcriptional activator. There also exist a variety of other potential NFAT- and angiogenesis-related phenotypes worthy of exploring using the tools and approaches I have developed. It is my hope that the findings herein stimulate further study on the contributions of NFAT signaling to the host immune response to mycobacterial infection and evaluation of the potential of NFAT inhibition as host-directed therapy to tuberculosis.

Dedication

For my constant motivation and inspiration
who taught me to read and write
who always believed I could do anything I set my mind to
who supported me doing anything, no matter how silly
who provided a rock of stability in a tumultuous world
who dreamt of this
– my Mamaw Barb

I hope I have made her proud.

List of Tables

3.1	Observed relationships between individual NFAT isoform activation and VEGFA production	276
3.2	Nearest gene neighbors for the ST sgRNAs (all are intergenic).	276
5.1	Selection criteria for MINCLE homologs	371
5.2	List of putative zebrafish MINCLE homologs	375
5.3	Constitutively active NFATC2 constructs	401
A.1	List of Antibodies Used	448
A.2	Bacterial Strains	449
A.3	Chemicals	450
A.4	Commercial Assays	452
A.5	Cell Lines	452
A.6	Model Organisms and Strains	453
A.7	Recombinant DNA and Plasmids	453
A.8	Software	455
A.9	Equipment	455
A.10	Oligonucleotides	456

List of Figures

1.1	Chemical structure of TDM	22
1.2	Granuloma structure schematic	50
1.3	Schematic of NFAT activation mechanism	82
1.4	Functional domains in NFAT family proteins	84
2.1	<i>flk1:eGFP^{s843}</i> larval zebrafish at 6 days post fertilization	166
3.1	Graphical abstract	237
3.2	Schematic of the possible pathways downstream of C-type lectin activation	242
3.3	Diagram of trunk injections in the larval zebrafish	243
3.4	Extracellular bacterial accumulation corresponds to <i>vegfaa</i> induction	244
3.5	Extracellular bacterial growth correlates with angiogenesis	245
3.6	<i>myd88</i> is dispensable for angiogenesis	247
3.7	Chemical inhibition of SYK prevents TDM-mediated Angiogenesis	248
3.8	<i>card9</i> is dispensable for angiogenesis	249
3.9	<i>asc</i> is dispensable for angiogenesis	251
3.10	FK506 treatment inhibits angiogenesis during infection	251

3.11	FK506 treatment inhibits TDM-mediated angiogenesis	253
3.12	<i>nfatc3a</i> is dispensable for angiogenesis	254
3.13	<i>nfatc3a</i> is dispensable for angiogenesis	255
3.14	Expression of NFAT isoforms in the granuloma and kidney macrophages	256
3.15	<i>nfatc2a</i> is necessary for angiogenesis during infection	257
3.16	Design of <i>irg1:VIVIT-tdTomato</i>	259
3.17	<i>lyz:VIVIT-tdTomato</i> does not impact angiogenesis	260
3.18	<i>irg1:VIVIT-tdTomato</i> inhibits angiogenesis during infection	261
3.19	<i>irg1:VIVIT-tdTomato</i> blocks TDM-mediated angiogenesis	262
3.20	Schematic of the CLARITY-clearing approach	263
3.21	<i>nfatc2a</i> is necessary for angiogenesis in adult granulomas	265
3.22	Granuloma heterogeneity	266
3.23	<i>irg1:VIVIT-tdTomato</i> inhibits granuloma angiogenesis in the adult zebrafish	267
3.24	NFAT inhibition decreases bacterial burden	269
3.25	Model of extracellular exposure of γ - <i>Mtb</i> to THP-1 macrophages	270
3.26	NFAT inhibition blocks <i>VEGFA</i> transcription in THP-1 macrophages	271
3.27	NFAT inhibition blocks <i>VEGFA</i> production and secretion in THP-1 macrophages	272
3.28	Immunofluorescence of γ - <i>Mtb</i> -exposed THP-1 macrophages	273
3.29	Quantitation of THP-1 macrophage immunofluorescence with NFAT inhibition	274
3.30	NFAT isoform immunofluorescence	275

3.31	NFAT isoform immunofluorescence quantitation	275
3.32	NFATC2 sgRNA targets	277
3.33	<i>NFATC2</i> targeting in THP-1 macrophages inhibits VEGFA production	278
3.34	Validation of <i>NFATC2</i> knockout cells	279
3.35	Quantitation of <i>NFATC2</i> knockout THP-1 macrophages	280
3.36	Summary figure	286
3.37	Putative NFATC2 and HIF-1 α Binding Sites in the VEGFA promoter	287
4.1	GUI for manual maximum intensity projections	300
4.2	GUI for maximum intensity projection with automatic image opening	304
4.3	First part of the GUI for metamorpher, which allows for the compilation of multidimensional Metamorph images	317
4.4	Second part of the GUI for metamorpher, allowing the user to select the false colors for the different image channels	318
4.5	GUI for the existing “nd stack builder” script	320
4.6	GUI for cell isolation in the surface plot analysis pipeline	324
4.7	GUI for measuring the output from “3D Surface Plot”	327
4.8	GUI for measuring bacterial burden from larval zebrafish	363
5.1	Major findings and lingering questions	368
5.2	Expression profiles of two putative MINCLE homologs in the zebrafish	373
5.3	Single-cell RNA-seq expression profile for 56379	373
5.4	Single-cell RNA-seq expression profile for 79903	374
5.5	Single-cell RNA-seq expression profile for 77975	374

5.6	Structural alignment of MINCLE and 56379	376
5.7	Structural alignment of MINCLE and 79903	377
5.8	Structural alignment of MINCLE and 77975	377
5.9	HIF-1 α regulatory mechanisms	381
5.10	scRNA-seq expression data for <i>hif1ab</i>	396
5.11	scRNA-seq expression data for <i>vegfaa</i>	396
5.12	scRNA-seq expression data for <i>flt1</i>	424

Acknowledgements

This document is the culmination of my entire twenty-five-year-long academic career to date and would have been impossible without all of the people along the way who have supported, inspired, and taught me all I needed to get here. The number of people who warrant thanks exceeds my capacity to thank them and the space allotted.

From the time of my birth, I had one member of my family who always knew I could be something amazing and labored to ensure I became all that and more – my Mamaw Barb. While she was lost to us far too soon, her contributions to my life echo into the present and, were her life's circumstances different, she could have been the one to become the first Brewer with a Ph.D. with her brilliant mind, dedication, and love of knowledge.

My journey into the world of learning would not have been possible without the manifold contributions of those primary and secondary school teachers who saw my potential and worked to help me achieve it. From kindergarten all the way to my senior year of high school I had beacons who guided me to overcome the difficult cultural and familial conditions into which I was born and raised. These contributions are what set me on the path to where I am today and I will forever appreciate those who strove to uplift students from the difficult educational environment of Barbourville.

The inflection point of my life was the four years I spent at Transylvania University. It was truly the place where everything changed. This was where I really found myself and was pushed to my intellectual limits and beyond and I discovered a true passion for teaching. Paul Duffin introduced me to the wonders of bench science and gave me a platform to learn, discover, and fail that provided me the precise mentality I needed to succeed in graduate school. Scott Whiddon has always been my #1 cheerleader and has been an invaluable source of support both throughout my time at Transylvania and into the present day. Michael Cairo sparked in me a love for political science and gave me the support, affirmation, and down-to-earth advising I needed to find my own academic identity. Lastly, Jeff Freyman gave me the most rigorous and exhilarating set of academic experiences of my life to date – the courses I took with him showed me the power to change someone’s life in the classroom because his courses, approach to teaching, and pursuit of the Truth changed my entire worldview. More than perhaps anybody else, Jeff showed me how academia can be a path to the *bonum vitae* and set me on this path I am now at yet another crossroads along.

Of course, college is not merely the classes you take and I had the privilege of meeting countless friends who taught me who I was and who I wanted to become, many of whom have been with me continuously for the past decade. Bobby Puckett is the first of my peers I had ever met who filled me with intellectual awe and his mentorship, especially in my first-year of college, set me on a path to further success. He believed in me, encouraged me, humbled me, and supported me all along this path I have taken and fostered my emotional and mental development in untold ways through my entire first-year and continuously into the present. I also had the privilege to surround myself with an amazing set of immediate peers – Chanslor Gallenstein, Drew Cunningham, Ryan Hanson, Susan Northcutt, and Quentin Becker – who have

been there from the day we moved into Clay Hall until today. And Rachel Johnson kept my ego in check as she outperformed and outclassed me at every turn and taught me so much about humility and grace (and what tuberculosis was). As my college career advanced, I had the opportunity to meet incredible people at every stage who made a difference. Jared Auton is an impeccable soul with a fervor for art who taught me about the beauty of the world around us. Daniel Martell and John Mays gave me invaluable opportunities in mentorship and taught me how *I* could make a real difference in the lives of others. And Daniel Cooper is shining light of positivity and joy who keeps me thinking with an optimistic mindset and with nothing but hope for the future. There are undoubtedly others I have missed, but the sum of those four years and the people who touched my life throughout them have been essential to my personal growth and success.

Graduate school itself, of course, has been a wonderful experience in growth and challenge. From the first days of this experience, there has been one person who I have been able to rely on for support – Andy Alspaugh. He has always believed in me and my ability to go on and has always encouraged me to pursue my dreams. His kindness is an immeasurable inspiration I can only vainly attempt to replicate for others. Graduate school has given me a body of peers who I have had the joy to meet and learn from at various stages, including Joe Saelens, Eric Walton, Ryan Finethy, Jessie Finethy, Molly Matty, Allison Roder, Shannon Esher, Hannah Brown, Al Harding, Matt Sacco, Stephen Walsh, Dulcemaria Hernandez, and Mollie Sweeney. These people have all impacted my life in critical ways and have helped me grow into the scientist (and adult) I am today. Lastly, I would like to thank Mark Cronan, the devil always on my shoulder encouraging me to try the risky experiment and who kept me in the lab way too late with riveting scientific, political, and personal conversation. He is my model of how to be a great scientist and I am fortunate to

have had the pleasure to know him. My time in Durham has also brought a lost connection back – Austin Farmer – one of the very few who also left our hometown to experience something different and pursue his dreams and we have picked right back up where we left off ten years ago.

Of course, my advisor, David Tobin, has been a constant and unconditional source of support and encouragement through the long experimentation that was required for much of the work presented in these coming pages. He provided me the academic and scientific freedom to find my path in science and to fail (repeatedly) with the unwavering faith that I would succeed. His personal kindness has also made the tribulations of these six years possible and he is truly a selfless soul with the unfettered desire to help others succeed. The trials of graduate school would have been impossible to overcome without his constant support. The rest of my thesis committee also warrants acknowledgement for being there for me since the very beginning and supporting, encouraging, and constructively criticizing me along the way – John Rawls, Chris Kontos, Dennis Ko, and Andy Alspaugh.

Lastly, I want to thank my loving partner in this life, Kristen Stockdale, who has believed in me and supported me throughout the past 4 years. As is often said of the long-suffering partner, she has tolerated the days, nights, and weeks that I was completely absent and, when I finally had a break, welcomed me home with no (visible) resentment and for that I will be forever thankful. She brings balance to my life and reminds me of all the other things that there are to do in the world beyond the lab and her presence reminds me I should probably just go home instead of running that 6 PM gel.

Contents

Abstract	iv
Dedication	vi
List of Tables	vii
List of Figures	viii
Acknowledgements	xii
1 Introduction	1
1.1 Tuberculosis	1
1.1.1 History of Tuberculosis	3
1.1.2 The Pathogenic Lifestyle of <i>Mycobacterium tuberculosis</i>	6
1.1.3 Treatments for Tuberculosis and their Mechanisms of Action .	12
1.1.4 The Mycobacterial Cell Wall	17
1.1.5 Trehalose 6-6'-Dimycolate (TDM)	21
1.2 Macrophages in Development and Disease	30
1.2.1 The M1-M2 (False) Dichotomy	31
1.2.2 Developmental Origins	32
1.2.3 Alveolar Macrophages in Tuberculosis	36
1.2.4 Recruited Macrophages in Tuberculosis	38

1.2.5	Host Resistance and Susceptibility to Infection Mediated by Macrophages	41
1.2.6	The Granuloma	46
1.3	Pattern Recognition in Innate Immunity	53
1.3.1	Major Families of Pattern Recognition Receptors	54
1.3.2	LPS as a Model Molecular Pattern	59
1.3.3	Signaling Mechanisms Downstream of C-Type Lectin Receptors	61
1.3.4	C-Type Lectins in Tuberculosis	71
1.3.5	The TDM Receptor	74
1.4	Nuclear Factor of Activated T Cells (NFAT)	80
1.4.1	Clinical Utility of NFAT Inhibitors	85
1.4.2	Differentiation of Individual Isoforms	88
1.4.3	Activity of NFAT in Myeloid Populations	117
1.4.4	The NFAT Renaissance	121
1.5	Angiogenesis	125
1.5.1	Angiogenesis in Cancer	127
1.5.2	The Relative Failure of Bevacizumab	129
1.5.3	Non-Angiogenic Functions of VEGFA	134
1.5.4	Macrophage-Endothelial Crosstalk in Angiogenesis	136
1.5.5	Historical Observations of Angiogenesis in Tuberculosis	141
1.5.6	Modern Studies on Granuloma Angiogenesis	142
1.6	Host-Microbe Interactions to Study Cell Biological Processes	147
1.6.1	Host-Directed Therapies: History and Promise	148
2	Scientific Methodology	153
2.1	Transparency in Science and Data Availability	153
2.2	Zebrafish as a Model Organism	154

2.2.1	Zebrafish and the History of Developmental Biology	157
2.2.2	Modern Applications of Laboratory Zebrafish	160
2.2.3	Zebrafish as a Model System for the Study of Host-Microbe Interactions	162
2.2.4	Zebrafish a Model System for Vascular Biology	166
2.2.5	Challenges in the Use of the Zebrafish Model	169
2.3	<i>Mycobacterium marinum</i> -Zebrafish Model of Tuberculosis	172
2.3.1	History and Merits of the Zebrafish- <i>M. marinum</i> Model . . .	172
2.3.2	Zebrafish Husbandry	175
2.3.3	Deficits of Mouse Models of Tuberculosis	180
2.3.4	Comparison to Other Models	184
2.4	Transgenic Tools in the Zebrafish	185
2.4.1	Technical Description of New Transgenic Lines	187
2.5	Use of CRISPR/Cas9 in Zebrafish	190
2.5.1	Crispant Assays	196
2.5.2	Prospectus on Cutting Edge and Future Tools in the Zebrafish	198
2.6	CLARITY and Confocal Microscopy	200
2.6.1	Technical Description	201
2.7	<i>Mycobacterium marinum</i>	203
2.7.1	Culture Conditions	204
2.7.2	CFU Assays	205
2.8	THP-1 as a Macrophage Model	207
2.8.1	Culture Conditions	208
2.8.2	qRT-PCR	209
2.8.3	ELISA	215
2.8.4	Immunofluorescence	219

2.8.5	Lentivirus Construction	230
2.8.6	Transduction	231
2.9	Quantification and Statistical Analysis	231
2.10	R	232
2.11	FIJI/ImageJ	233
3	Macrophage NFATC2 Mediates Angiogenic Signaling During Mycobacterial Infection	236
3.1	Abstract	236
3.2	Introduction	238
3.3	Results	243
3.3.1	Macrophage Induction of <i>vegfaa</i> and Angiogenesis during Mycobacterial Infection	243
3.3.2	Genetic <i>card9</i> Deficiency Does Not Compromise Angiogenesis During Mycobacterial Infection	246
3.3.3	Pharmacological Inhibition of NFAT Activation Limits Angiogenesis in Response to Mycobacteria and TDM	252
3.3.4	The Isoform <i>nfatc2a</i> is Specifically Required for Angiogenesis During Infection	254
3.3.5	Macrophage-NFAT is Essential for Angiogenesis Induction <i>in vivo</i>	258
3.3.6	NFAT Activation is Essential for Angiogenesis in Adult Granulomas	263
3.3.7	Macrophage-specific NFAT Inhibition in Mature Granulomas Reduces Angiogenesis	264
3.3.8	Inhibition of NFAT Signaling Results in Decreased Bacterial Burden	268
3.3.9	Pharmacological Inhibition of NFAT in Human THP-1 Macrophages Limits VEGFA Induction by <i>Mycobacterium tuberculosis</i>	269
3.3.10	Requirement of human NFATC2 for VEGFA induction	271

3.4	Discussion	277
3.4.1	Limitations of the Study	288
3.5	Acknowledgements	290
3.6	Author Contributions	290
4	Novel Computational Approaches to Image Processing and Quantitation	292
4.1	FIJI/ImageJ	293
4.2	Python	295
4.2.1	Napari	296
4.3	R	296
4.4	Maximum-Intensity Projection and Composite Image Generation . .	297
4.5	Building Multi-Dimensional Images from Metamorph File-Saving Logic	310
4.6	Surface Plot Analysis for Cellular Distribution of Labeled Proteins .	319
4.7	py-LaRoMe	330
4.8	Experimental Blinding via a Single-Click Command Line Interface .	338
4.9	User-Friendly Analysis of RNA Sequencing Data using Kallisto/Sleuth in a Python Environment	343
4.10	Bacterial Burden Analysis by Fluorescence Intensity in a Semi-Automated Manner with a User-Friendly Graphical Interface	355
5	Conclusions and Future Directions	365
5.1	The Zebrafish MINCLE	367
5.2	Integration of Hypoxia Signaling	378
5.2.1	HIF for HIF's Sake	380
5.2.2	Macrophage Metabolism in Immunity	388
5.2.3	HIF-NFAT Interactions	398
5.3	Transcriptional Coregulatory Mechanisms	400

5.3.1	NFAT:AP-1 Interactions	403
5.3.2	NFAT:NFκB	404
5.4	New Genetic Approaches to the Study of NFAT Signaling	407
5.5	Mycobacterial Interactions with Other Aspects of Vascular Biology .	410
5.5.1	Vascular Permeability	411
5.5.2	Pericytes	414
5.5.3	Extracellular Matrix	417
5.5.4	Other Angiogenic Stimuli	420
5.6	Precise Contributions of VEGFA Signaling to Mycobacterial Burden .	422
5.7	VEGFR2-Dependent Signaling Pathways in the Endothelium	427
5.8	Lymphangiogenesis	428
5.9	Generalizability	434
5.10	A Pressing Need for More Objective Approaches to Image Analysis .	440
5.10.1	Approaches to Measuring Angiogenesis	441
5.10.2	High-Throughput Quantitation of Cell Features	442
5.11	NFAT Targeting as a Host-Directed Therapy?	444
5.12	Closing Remarks	446
A	Reagents Used	448
B	Response to Reviewers for Brewer et al., 2022b	459
Bibliography		483
Biography		705

Chapter 1

Introduction

This work integrates a detailed interrogation of host-pathogen interactions in the specific context of C-type lectin signaling responses and angiogenesis. In this section, I will introduce the major concepts needed to interpret the data-driven findings in later chapters. These topics will include: the disease itself, the fundamental biology of macrophages, the principles of C-type lectin signaling, the NFAT signaling pathway, and angiogenesis. Each section will incorporate historical background, foundational findings in the field of interest, and contextualization for how this information educates later approaches and experimental decisions.

1.1 Tuberculosis

Of all the infectious agents to have ever afflicted humankind, *Mycobacterium tuberculosis* is perhaps the most imminently successful (Barberis et al., 2017). The primary cause of potentially greater than one billion human deaths since 1800 alone (Murray, 2004; Paulson, 2013), “the White Plague”¹ has had profound impact on the cultural

¹This nickname is in juxtaposition to the Black Plague, the historical disease caused by *Yersinia pestis*, which decimated much of the global human population in the 1300s (Perry and Fetherston, 1997).

and political development of the modern world and continues to impact the lives of most people around the world today², with a disproportionate impact on those who live in economically developing countries. Fallaciously considered a disease of antiquity, this disease co-evolved with humankind (Hershkovitz et al., 2015) and today manifests in active disease in greater than 10 million people³ each year and has killed greater than one million people per year each year since records or estimates have been available (Saleem and Azher, 2013; World Health Organization, 2021) with the case and death burden rising due to health system neglect exposed by the COVID-19 pandemic ongoing at the time of this writing (Pai, Kasaeva, and Swaminathan, 2022).

Mycobacterium tuberculosis has long been of basic scientific interest on account of the myriad ways in which it undermines host immune responses to establish a replicative niche within the human lung (Stanley et al., 2003; Monack, Mueller, and Falkow, 2004; Nguyen and Pieters, 2009; Baxt, Garza-Mayers, and Goldberg, 2013; Hmama et al., 2015; Yu et al., 2019). *M. tuberculosis* infection results in the formation of caseating granulomas⁴ encapsulated by a complex network of immune cells within which the bacteria replicate (Pagan and Ramakrishnan, 2018). Over evolutionary time, these bacteria have innovated novel ways of subverting nearly every host-protective immune response while exacerbating maladaptive ones in pursuit of survival and transmission (Ernst, 2012; Rahman et al., 2020; Guan et al., 2021; Chandra, Grigsby, and Philips, 2022). This makes the study of tuberculosis not only the study of microbiology and immunology, but a fascinating study in the basic

²For additional reading on this subject of how tuberculosis has impacted the development of human society, see (Chalke, 1962; Dubos and Dubos, 1987; Day, 2017).

³Greater than 30 million people are at constant risk of infection by cohabiting with active tuberculosis patients (Ross et al., 2021).

⁴The biology of these granulomas is a fascinating microcosm of developmental and cell biology which will be discussed at length in Subsection 1.2.6.

principles of cell biology informed by evolution and ecology, interwoven with history.

1.1.1 History of Tuberculosis

The overwhelming prevalence of tuberculosis in the 18th and 19th centuries led to a profound degree of cultural salience for this disease in the daily lives of the people of those times. Responsible for the deaths of many preeminent public figures of these eras⁵ and approximately 25% of all deaths by any cause (Centers for Disease Control, 2016), it became a ubiquitous dimension of the literary *Zeitgeist* as well. Perhaps most famously, tuberculosis is depicted as the disease that afflicts the Lowood School in Charlotte Brontë's *Jane Eyre*, among other novels depicting the disease then known as *consumption* for the way in which it leads to cachexia⁶, pallor, hemoptysis⁷, and ultimately death (Brontë, 1847; Loddenkemper, Lipman, and Zumla, 2015).

This cachexia is a defining feature of tuberculosis across the diverse organisms afflicted by it; such progressive, consumptive wasting unable to be ameliorated by improved nutrition is an unusual presentation strongly reminiscent of many cancers and rather dissimilar from most infectious diseases (Silva and Faccioli, 1988; Tisdale, 2002; Martignoni, Kunze, and Friess, 2003; Chang et al., 2013). Indeed, as medi-

⁵The number of such public figures is far too great to list. From the 1840s and 1850s alone, tuberculosis was responsible for the deaths of Andrew Jackson (seventh president of the United States), Henry Clay (Secretary of State, Speaker of the House, three-time presidential candidate for the Whig Party), John C. Calhoun (Vice President, Secretary of State), Alexis de Tocqueville (famed French observer of American culture and author of the classic of political theory, *Democracy in America*), Henry David Thoreau (naturalist author of *Walden*), and Emily Brontë (author of *Wuthering Heights*). Additionally, it is thought that George Washington may have been afflicted with intestinal tuberculosis, explaining a lifetime of gastrointestinal troubles (Chernow, 2010) and, in the modern day, Ringo Starr of *The Beatles* fame had tuberculosis and was confined to a sanitorium for some time (Starr, 2016).

⁶To be described more extensively shortly, cachexia is a progressive wasting that results in unmitigable weight loss.

⁷The racking cough that accompanies tuberculosis is often depicted by women on chaise lounges coughing blood into a white handkerchief; the reality, of course, is far more dire (Giles, 1802–1881; Bouvier, 1842–1865; Robinson, 1858; Alexander, 1895).

cal understanding of diseases progressed beyond concepts of humoral imbalance⁸, a prevailing theory was that tuberculosis was a hereditary form of cancer due to the way it spread within families (Frith, 2014). The functional and consequential similarities between tuberculosis and cancer are replete and will be a theme returned to throughout this document. However, as Louis Pasteur's germ theory (Smith, 2012) came to be more widely accepted in the same period in the Industrial Revolution as Joseph Lister's antiseptic practices (Toledo-Pereyra, 2010), it became a subject of scientific inquiry to identify the potentially infectious bases for seemingly transmissible diseases⁹.

Robert Koch was the first to document the tubercle bacillus as the infectious etiology of tuberculosis in 1882 (Koch, 1882; Cambau and Drancourt, 2014). *Mycobacterium tuberculosis* proved to be a foundational instrument in the broader development of the discipline of microbiology and provided Koch with the rationale for the development of what we know as the Koch's postulates¹⁰¹¹, a procedural set of

⁸This theory of humoral imbalance was prevalent from the time of the ancient Greeks until the 1800s, when more scientific approaches to the understanding of human disease were developed (Lagay, 2002). This theory posited that disease arose from an imbalance of the four major humors: blood, yellow bile, black bile, and phlegm. These constituted the human corpus and their balance dictates human health. We now know, of course, that diseases generally have specific causes that are able to be diagnosed by modern medicine.

⁹This having the caveat that tuberculosis does not always appear to be transparently transmissible. The disease can take months to years to manifest in infected persons, leading to long time delays between putative exposure and active disease (Behr, Edelstein, and Ramakrishnan, 2018).

¹⁰The postulates are as follows: the organism should be present in afflicted individuals, but not in healthy individuals; the organism should be able to be grown in pure culture; inoculation of a healthy host should recapitulate the disease; the organism should be able to be harvested back from the new host. Of course, each and every one of these postulates have been broken in pursuit of identifying disease-causing agents, but this remains the basis of identifying new infectious etiologies (Segre, 2013).

¹¹There is a parallel set of postulates known as the molecular Koch's postulates which are used to define the contribution of a particular gene to the virulence of a pathogen, which can be used to systematically and functionally dissect the genetic underpinnings of pathogenesis. These postulates, first proposed by Stanley Falkow are a functional adaptation of the original postulates for the molecular era (Falkow, 1988; Falkow, 2004).

criteria for determining the infectious etiology of any disease. This, along with the identification of the anthrax bacillus (*Bacillus anthracis*), sparked the beginning of the modern era of microbiology (Koch, 1876). It was only once the causative agent of tuberculosis had been identified that it became possible to earnestly pursue curative therapies and vaccines¹², which came swiftly thereafter to varying degrees of efficacy. By 1921, Albert Calmette and Camille Guérin had attenuated the closely related and conspecific *Mycobacterium bovis* for use as a vaccine, creating the BCG live attenuated vaccine still in use today¹³ (Hawgood, 2007).

Once thought to have been functionally banished to the annals of history, tuberculosis, after a steady decline in cases throughout the middle of the 20th century¹⁴, came roaring back later in the century with the introduction of HIV into the human population in the 1980s. HIV/AIDS causes an immunodeficiency that increases susceptibility to infection, disease, and death from tuberculosis, which has become an AIDS-defining illness (Selwyn et al., 1992; Drobniowski, Pozniak, and Uttley, 1995). Compounding these troubles, particular strains of *M. tuberculosis* have developed resistance to many of the commonly used antibiotic therapies available, leading to the emergence of multi-drug resistant strains that increase cost of treatment, acute

¹²This is an activity that Koch himself pursued, to some controversy. He publicly promised a vaccine made of homogenized bacilli; while this application failed, it provided the foundation of the tuberculin now used in the diagnostic skin test taken by millions of Americans annually (Goetz, 2014).

¹³The BCG vaccine is annually given to millions of newborns around the world to prevent tuberculosis, but demonstrates, at best, variable efficacy (Rodrigues, Diwan, and Wheeler, 1993; Andersen and Doherty, 2005; Schrager et al., 2020; Trauer et al., 2021) and a short period of protection against tuberculosis, despite inducing a lifetime's worth of immunomodulation and protection from lung cancer and all-cause death (Higgins et al., 2016).

¹⁴This was coincident with, but likely unrelated to, the development of effective antibiotic therapies. Indeed, the modern disparity between tuberculosis rates between the United States and Western Europe and much of the rest of the world is thought to have more to do with improved living conditions, growing herd immunity, and improved nutrition rather than the use of antibiotics as downward trends actually began 100 years prior to the discovery of streptomycin in 1944 (Lowbury, 1958).

patient burden, and duration of therapy (Matteelli, Roggi, and Carvalho, 2014).

1.1.2 The Pathogenic Lifestyle of *Mycobacterium tuberculosis*¹⁵

The study of *M. tuberculosis* has long focused on factors that drive its virulence; that is, its ability to cause disease, evade immune responses, overcome antibiotic therapy, and transmit to new hosts (Glickman and Jacobs, 2001). These studies have revealed innumerable ways by which this pathogen accomplishes these tasks and many potential targets for new antitubercular therapies both via bacterial factors and host-protective and -detrimental immune responses. In many regards, the study of tuberculosis is the study of so-called “anti-immunology” given the vast spectrum of mechanisms used to undermine the host and the frequent failure of the host to mount a sufficient immune response (Finlay and McFadden, 2006; Diacovich and Gorvel, 2010).

In addition to the clear relevance of the study of tuberculosis to human health, the unique biological features of this acid-fast, genetically-insular¹⁶, non-motile, slow-growing¹⁷ mycobacterial species make it a fertile ground for basic scientific studies into the way that both saprophytic and pathogenic species of bacteria adapt to adverse environments and ultimately establish a productive niche (Pieters and Gatfield,

¹⁵For excellent reviews on this topic, see J. A. Philips and J. D. Ernst (2012). “Tuberculosis pathogenesis and immunity”. In: *Annu Rev Pathol* 7, pp. 353–84 and A. O’Garra et al. (2013). “The immune response in tuberculosis”. In: *Annu Rev Immunol* 31, pp. 475–527.

¹⁶Some, but not all *Mycobacterium* spp. can undergo horizontal gene transfer, but the mechanisms are poorly understood. Indeed, the challenge of shuttling DNA across the cell wall was a major impediment in the development of genetic tools to study mycobacterial genetics in the lab. However, *M. tuberculosis sensu stricto* is not thought to be competent for natural gene transfer (Krzywinska, Krzywinski, and Schorey, 2004; Galagan, 2014; Derbyshire et al., 2014; Reva, Korotetskiy, and Ilin, 2015; Merker et al., 2015; Boritsch et al., 2016; Madacki et al., 2021).

¹⁷*M. tuberculosis* doubles approximately once every 24 hours, compared to *Escherichia coli* which can double once every 20 minutes in log-phase.

2002; Houben, Nguyen, and Pieters, 2006; Falkinham III, 2009; Prasanthi and Murty, 2014; Ghodbane et al., 2014; Delafont et al., 2014). The physiological features of the bacillus – a thick, hydrophobic cell wall (Jarlier and Nikaido, 1994; Chatterjee, 1997; Jankute et al., 2015), unique export and import systems (Braibant, Gilot, and Content, 2000; Pandey and Sassetti, 2008; Tullius et al., 2011; Houben, Korotkov, and Bitter, 2014; Wong, 2017; Palmer, 2017; Famelis et al., 2019; Bunduc, Bitter, and Houben, 2020), and novel mechanisms for cell division (Dziadek et al., 2003; Hett and Rubin, 2008; Kieser and Rubin, 2014; Odermatt et al., 2020) and stress tolerance (Garg et al., 2015; Peddireddy, Doddam, and Ahmed, 2017) – make this a fascinating case study in the evolutionary processes that drive niche adaptation and, indeed, niche creation (Chapman, 1971; Warner and Mizrahi, 2007; de Chastellier, 2009; Gengenbacher and Kaufmann, 2012; Lerner et al., 2016; Honda, Virdi, and Chan, 2018; Gagneux, 2018; Sarathy and Dartois, 2020; Pereira et al., 2020; Lovewell et al., 2021).

That related members of the same genus of bacteria occupy such diverse infectious niches across a wide spectrum of organisms (from fish and amphibians and reptiles to birds and mammals and in every major organ system), with many also possessing stages of growth in the environment is a testament to the extent to which these species have evolved structures and responses that can accommodate a wide range of physical and chemical stressors (Thoen, Karlson, and Himes, 1981; Palmer, Welsh, and Hostetter, 2011; Hershberg, 2016; Saelens, Viswanathan, and Tobin, 2019; Larsen et al., 2020). By contrast, some species, notably *M. tuberculosis* and *M. leprae*¹⁸, are tightly adapted to a more limited range of hosts and are thought to have lost the capacity for long-term survival outside of a mammalian host, although this is now

¹⁸Leprosy, the classical Biblical affliction, is another prominent human disease caused by a mycobacteria. The biology of leprosy is fascinating and will be touched on sporadically throughout this dissertation (Schamberg, 1899).

under some contention with the identification of infection-competent *M. tuberculosis* from soil samples (Martinez et al., 2019; Ploemacher et al., 2020; Borham et al., 2022; Mtetwa et al., 2022). This diversity within the genus offers abundant opportunity for gene-structure-function discovery to uncover factors both required for maintaining an environmental niche as well as those specifically required for either commensal or pathogenic association with the host, an approach that has long been fruitful in the discovery of novel virulence factors (Sassetti and Rubin, 2003; Ehrt, Rhee, and Schnappinger, 2015) but comparatively neglected in the basic bacteriological study of environmental mycobacteria.

When a pathogenic *Mycobacterium* infects a naïve host, an intricate set of cellular and signaling responses occur at the interface of the host and the bacterium that facilitate either successful clearance or establishment of a productive infection (Davies and Grange, 2001b; Gagneux et al., 2006a; Turner et al., 2017; Bohrer et al., 2021). The summative effect of these immune responses determines the outcome of the infection and careful balancing on the part of the host must take place to both kill the pathogen and minimize host pathology (Casadevall and Pirofski, 1999; Casadevall and Pirofski, 2003). Taking human tuberculosis infection as the model, productive infection will result in patients coughing aerosolized droplets that often contain a single bacillus (Churchyard et al., 2017; Mathema et al., 2017). These individual bacilli can then be inhaled by a naïve person nearby, establishing a new cycle of infection. Once that person has inhaled this bacterium¹⁹, lung-resident macrophages²⁰

¹⁹Unlike many other diseases, increasing the infectious inoculum paradoxically reduces the risk of disease – large clumps of bacteria are unable to reach the distal lung where they are better able to establish infection (Cambier et al., 2014).

²⁰Also known as alveolar macrophages, these are one of many, many different types of tissue-resident macrophages. While it is well beyond the scope of this section to explore fine distinctions, but suffice it to say that macrophages in each tissue niche are functionally distinct from one another and exhibit distinct responses to stimuli. Tissue-resident macrophages are also established from a non-hematopoietic origin from the fetal yolk and are replication-competent, allowing them to

phagocytose the bacteria. In many instances (approximately 90% of the time), the macrophage will successfully kill the bacteria, but in the other 10%, the bacteria are able to survive, replicate, and progress to either latent or active disease (North and Jung, 2004; Verrall et al., 2014; Pai et al., 2016; Kroidl et al., 2022).

If initial clearance or containment fails, a complex cascade of events proceeds. After phagocytosis, the macrophage sets in motion signaling processes that should result in fusion between the phagosome and extant lysosomes within the cytosol (Abramovitch et al., 2011). However, the bacteria, through a combination of structural features in the cell wall (more on this in Subsection 1.1.5) and secreted effector proteins, blocks phagosomal-lysosomal fusion to establish a replicative niche in the macrophage within a parasitophorous vacuole (McDonough, Kress, and Bloom, 1993; Pieters, 2008; Casonato et al., 2014). Subsequently, the outpouring of secreted effectors from the bacteria across the phagosomal membrane into the host cytosol results in profound reprogramming that blocks apoptosis (Keane et al., 1997; Keane, Remold, and Kornfeld, 2000; Yokobori et al., 2012; Maueroder et al., 2016; Ahluwalia et al., 2017; Beckham et al., 2017; Wong, 2017), downregulates production of select cytokines and chemokines (while enhancing the expression of others) (Fortune et al., 2004; Madan-Lala et al., 2011; Cambier, Falkow, and Ramakrishnan, 2014), directs the macrophage to recruit additional macrophages to further the replicative cycle (Cambier et al., 2014; Appelberg et al., 2015; Pagan et al., 2015), and guides the macrophage out of the lung proper and into the interstitial space surrounding the lungs, the actual site of primary infection (Peters and Ernst, 2003; Ramakrish-

self-renew *in situ* (Davies et al., 2013a; Davies et al., 2013b; Epelman, Lavine, and Randolph, 2014; Ginhoux and Jung, 2014; Ginhoux and Guilliams, 2016). The study of macrophage biology is one of immense challenge and opportunity to uncover novel roles for these multifunctional cells, which are now known to play important roles in processes as diverse as germ cell maturation, metabolism, sleep, cardiovascular disease, and many more (Davies et al., 2013a; Hussell and Bell, 2014). Some of these will be addressed in greater detail in Section 1.2.

nan, 2012; Upadhyay, Mittal, and Philips, 2018; Cohen et al., 2018). Due to the slow growth of *M. tuberculosis*, it can take several days before the macrophage has become filled with such a large number of bacteria that it necroses²¹, allowing the bystander macrophages and neutrophils to be infected anew (Banos-Mateos et al., 2017). This cycle continues as more immune cells are recruited by the escalating infection and more extracellular bacteria accumulate (Lin et al., 2006; Orme and Basaraba, 2014; Cadena, Flynn, and Fortune, 2016; Corleis and Dorhoi, 2020). These stages of infection, cell lysis, aggregation, and further recruitment eventually result in the formation of what we know as a granuloma (Ramakrishnan, 2012; Pagan and Ramakrishnan, 2018).

Tuberculous granulomas are complex aggregates of (primarily) macrophages that have differentiated into a less inflammatory²², epithelioid state that encapsulate a central focus of extracellular bacteria within a necrotic core (Boros, 2003; Rubin, 2009; Cronan et al., 2016). These epithelioid macrophages are augmented by the full spectrum of other immune cell types – inflammatory macrophages, neutrophils, basophils, eosinophils, natural killer cells, T cells, B cells, and a range of stromal cells (Russell, 2007; Ramakrishnan, 2012). As a discrete structure, these provide a full immunological nexus integrating essentially every identifiable immune cell population and may provide a pathology-minimizing approach to infection containment despite relative costs and benefits that will be explored more so later on (Casadevall and Pirofski, 2003). They also display a great deal of heterogeneity from granuloma to

²¹There is some interesting biology behind this, where *M. tuberculosis* actively inhibits apoptosis to allow itself to grow to high burden within the macrophage and then lyse it to be released into the extracellular milieu (Keane, Remold, and Kornfeld, 2000).

²²Some might say M2-like state, but the M1-M2 binary axis is, rightfully, under attack in the era of single cell -omics. This false dichotomy neglects the full spectrum of activation states possible for macrophages and the plasticity with which a given macrophage can change its activation state based on the particular set of signals being received at a granular point in time (Martinez and Gordon, 2014; Ley, 2017). See also Subsection 1.2.1.

granuloma and person to person, reflecting key variations in the immune and bacterial responses over time (Cadena, Fortune, and Flynn, 2017; Gideon et al., 2022). These granulomas, although extractable intact from their environment, must exist in a tissue environment not of their own immediate design (Kaplan et al., 2003; Datta et al., 2015; Cronan et al., 2018; McCaffrey et al., 2022). These microenvironmental factors, while of long-standing interest in tumor biology, have yet to be properly explored in the context of tuberculous granulomas. Although immune cells can be readily recruited, the extrapulmonary space is a pre-existing physical location that can be remodeled to some degree but is inherent in the course of the infection. Mycobacteria are tasked with manipulating these tissues, which they do not directly infect²³, to further their own lifestyle. One of the ways they do this, and which is the focus of much of this work, is by inducing the pathological growth of blood vessels toward the site of infection in a process known as angiogenesis.

The clinical description of hemoptysis is a clear manifestation of this vascular involvement in the disease (Souders and Smith, 1952; Middleton et al., 1977; Turner, Basaraba, and Orme, 2003; Turner et al., 2003). As the granuloma cavitates and releases its contents into the airway, the encapsulating vasculature is destroyed. While such damage could have occurred incidentally by damaging the vessels that service the alveoli, it is now clear that much of this hemorrhage is the product of the encapsulating vascular web that develops around the granuloma over the course of the infection (Cudkowicz, 1952; Oehlers et al., 2015; Datta et al., 2015). These clinical observations have, however, left unaddressed the role for this vasculature in the de-

²³This is not true *sensu stricto* as Cohen et al. (2018) observed transient infection of pulmonary epithelial cells, but this fades within a few days – the consequences of these infections are not known. Lasting memory of these events may influence outcomes of other infections or superinfecting mycobacteria, similar to findings in influenza viruses, which leave behind a pool of surviving epithelial cells after infection (Heaton et al., 2014; Hamilton et al., 2016; Dumm et al., 2019; Fiege et al., 2019; Dumm et al., 2020).

velopment, progression, transmission, and treatment of the disease. The former two questions are the subject of the present work while the latter two remain promising avenues of future discovery.

1.1.3 Treatments for Tuberculosis and their Mechanisms of Action

Mycobacterial infections are uniquely integrative biological phenomena that require a delicate balance between the host and the pathogen. The host, in order to eradicate the bacteria, seeks a potent but highly specific sterilizing immune response while the bacteria, seeking to establish a replicative niche, must evade these host defenses. Both must avoid inducing excessive tissue damage, which may reduce the fitness of both organisms – the host through impaired survival and reproduction and the pathogen by early termination of a route of transmission. Chemotherapeutic treatment of infection seeks to tip the scales back in favor of the host by targeting processes essential for bacterial survival. Historically, treatment for bacterial infections has been through the application of these bacteria-targeting antibiotics²⁴, despite their mechanism of action rarely being understood at the time of clinical introduction (Osborne, 2013; D'Ambrosio et al., 2015). Modern tuberculosis infections are treated with a four-drug cocktail of antibiotics over the course of six to nine months: isoniazid, ethambutol, pyrazinamide, and rifampin (Grace et al., 2019; Dorman et al., 2021). Should the bacteria exhibit resistance to one or more of these, a state known as multi- or extensive-drug resistance (MDR/XDR), additional drugs with further host

²⁴The most classic example is penicillin, which was discovered in 1941 by Alexander Fleming and sparked a revolution in the treatment of countless battlefield diseases during World War II, saving countless lives (Fleming, 1941; Henderson, 1997). While many bacteria have developed resistance to penicillin through expression of penicillin-binding proteins or beta-lactamase, other antibiotics have filled in some of these gaps to treat these resistant strains (Eagle, 1954; Sabath et al., 1977; Murray and Mederski-Samaroj, 1983; Klare et al., 1992).

toxicity are used: kanamycin, ciprofloxacin, and cycloserine are common choices, although new drugs are slowly coming onto the market (Quenard et al., 2017; Nahid et al., 2019; Jang and Chung, 2020). Of these, bedaquiline appears to have the most promise in improving the overall treatment of tuberculosis, but long-term impact remains to be seen (Mahajan, 2013; Pym et al., 2016; Furin, Lessem, and Cox, 2017).

The first modern, clinically effective treatment for tuberculosis was pioneered by the discovery of streptomycin from *Streptomyces griseus* in 1943 (Hinshaw, Feldman, and Pfuetze, 1946; Hinshaw, Pyle, and Feldman, 1947; Comroe, 1978). Unlike its antibiotic predecessor, penicillin, streptomycin was effective in killing *Mycobacterium tuberculosis* bacilli *in vitro*. However, due to its lack of oral bioavailability, the use of this drug was limited to hospitals and clinics able to deliver the drug intravenously. Additionally, like many of the attempts at drug development for tuberculosis that had preceded it²⁵, it was not particularly effective at eliminating disease when used alone and resistance quickly emerged (Pfuetze and Pyle, 1949; Ruiz et al., 2003). Streptomycin is an aminoglycoside antibiotic that interferes with protein biosynthesis by poisoning the 30S subunit of the ribosome (Sharma et al., 2007). This mechanism is common to all of the diverse bacteria against which streptomycin is effective, making it a good general purpose antibiotic, if somewhat limited in the face of the unique features of mycobacterial anatomy.

Thus, the introduction of a mycobacteria-specific antibiotic in the form of isoniazid in 1952 was a major breakthrough in the treatment of this disease (Goldman and Braman, 1972). Orally bioavailable and highly effective at killing mycobacteria, it comes with the dose limiting side effects of peripheral neuropathy and occasionally fatal hepatitis that make it a less than perfect therapeutic option (Maddrey and

²⁵One of these, para-aminosalicylic acid (PAS) is an interesting, if distracting tale in the history of microbiology. For more information, see (Dubovsky, 1988; Donald and Diacon, 2015)

Boitnott, 1973; Black et al., 1975; Metushi et al., 2011; Metushi, Utrecht, and Phillips, 2016; Diallo et al., 2018). Nevertheless, it is still in use today on account of its synergy with other antimycobacterials and independent efficacy. Isoniazid works by targeting mycobacterial cell wall synthesis via InhA inhibition to block earlier stages of fatty acid biosynthesis (Timmins and Deretic, 2006; Vilchezze and Jacobs, 2007). This prevents the synthesis of the mycolic acids that comprise the outermost layer of the cell wall and which are essential for mycobacterial survival, growth, and immune evasion (Rozwarski et al., 1998; Lei, Wei, and Tu, 2000; Vilchezze et al., 2006; Timmins and Deretic, 2006).

Recognizing the inherent limitations of isoniazid monotherapy, additional drugs came into use over the next twenty years. Next on the list of drugs was ethambutol, which entered into use in 1961 (Thomas et al., 1961; Chakraborty and Rhee, 2015). Ethambutol, like isoniazid, targets the synthesis of the cell wall, this time by inhibiting the enzymatic ligation of arabinogalactan, which bridges the lower peptido-glycan layer and the outer mycolic acid layer, destabilizing the cell wall and increasing bacterial susceptibility to immune killing (Safi et al., 2013; Zhang et al., 2020b). Interestingly, the precise mechanism of action of ethambutol remains unknown despite over 60 years of extensive study, although cross resistance with isoniazid is common, suggesting some overlapping target pathways (Gupta et al., 2006; Zhang et al., 2020b).

Rifampin (1965) was the next addition and has an entirely distinct mechanism of action compared to the previous entrants. Targeting multiple, simultaneous, essential biological pathways is an excellent and repeatedly proven way of killing pathogens and preventing the emergence of resistance to all of the drugs simultaneously as such coordinate genetic mutation is exceedingly unlikely and even more so given the slow mu-

tation rate of *Mycobacterium* species²⁶ (Pletz, Hagel, and Forstner, 2017). Rifampin targets RpoB, the major subunit of the bacterial DNA-dependent RNA polymerase, which is essential for transcription (Wehrli, 1983; Telenti et al., 1993). Although mutations have arisen that confer resistance to rifampin, these have particular fitness costs on the bacteria under conditions lacking antibiotic pressure (Billington, McHugh, and Gillespie, 1999; Mariam et al., 2004; Gagneux et al., 2006b; Gagneux, 2009; Xu et al., 2021a). Rifampin has proven to be an excellent antimycobacterial drug with a comparatively favorable side effect profile compared to the other commonly used drugs and is universally used for treatment of both latent and active tuberculosis²⁷.

To round out the four drug cocktail generally recommended for the first-line treatment of tuberculosis today, pyrazinamide (1972) is the most mechanistically interesting of this antibiotic panel (Yeager, Munroe, and Dessau, 1952; Steele and Des Prez, 1988; Millard et al., 2019). It appears to work by diffusion into the acidic necrotic center of the granuloma where protons activate the prodrug by nucleophilic attack of the amide bond, allowing it to be enzymatically converted by the bacteria into pyrazinoic acid, the active antimicrobial (Kalinda and Aldrich, 2012; Zhang et al., 2013; Zhang et al., 2019; Lamont and Baughn, 2019; Villagracia et al., 2020; Lamont, Dillon, and Baughn, 2020). The low pH maintains the stoichiometry in favor of the protonated pyrazinoic acid form over the conjugate base pyrazinoate, facilitat-

²⁶At a rate of approximately two base pair changes per 10,000 replications, it can take weeks or months for this slow growing bacteria to happen upon a resistance-conferring mutation (McGrath et al., 2014).

²⁷Rifampin treatment is the origin of the oft-repeated but foundationally untrue warning given to women on hormonal contraceptives to beware antibiotics. While rifampin has been shown to reduce blood estrogen concentrations, there is no evidence of other drugs having these effects. Common antibiotics are vanishingly unlikely to interfere with oral contraceptives and these warning labels are both unnecessary and misleading (Skolnick et al., 1976; Zhanel et al., 1999; Barditch-Crovo et al., 1999; Archer and Archer, 2002; Simmons et al., 2018a; Simmons et al., 2018b).

ing diffusion into the cytosol of the bacteria across the hydrophobic cell membrane. Despite the knowledge that has been ascertained about the precise conditions under which pyrazinamide is active, the mechanism of action remains under hot contention with a variety of different mechanisms proposed and the most recent – that it inhibits the synthesis of the essential fatty acid and metabolic carrier coenzyme A – still under dispute as the mechanism of resistance is typically through mutations in *pncA*, the pyrazinamidase (Zhang et al., 2003; Shi et al., 2011; Kalinda and Aldrich, 2012; Zhang et al., 2014; Dillon et al., 2017; Lamont, Dillon, and Baughn, 2020). Pyrazinamide, entirely by accident, takes advantage of the particular biological environment of the infecting bacteria to specifically target the pathogen. As a relatively innocuous prodrug that is activated at the precise site of infection, it is able to ameliorate some of the toxic effects that would be associated with direct use of pyrazinoic acid while concentrating active drug where bacteria are actively growing with passive diffusion moving additional prodrug into the granuloma to be activated and trapped inside the necrotic caseum (Wade and Zhang, 2004; Gopal et al., 2016). Replicating such effects with other prodrugs seems like an effective means of improving safety and efficacy profiles, although the rational design of such drugs is certainly far from trivial.

Modern antibiotic development generally has been stymied by a lack of incentive for the expenditure of research and development funding on costly, low profit drugs (Joshi, 2011). As new antibiotics are likely to be reserved for cases with extensive antibiotic resistance and are likely to be cost-prohibitive for broad use, few have been developed despite pressing need. One of the success stories is that of bedaquiline, which was first approved in 2012 and represents an entirely new class of antibiotics. The development of bedaquiline required \$500 million in public investment compared to \$100 million in investment by the profiteering corporation, Janssen Biotech (Os-

borne, 2013; Leibert, Danckers, and Rom, 2014; Gotham et al., 2020). Bedaquiline is a potent and highly effective drug reserved for use in multidrug resistant (MDR) and extensively drug resistant (XDR) cases of tuberculosis and which acts to block ATP synthase to shut down bacterial metabolism and directly leads to bacterial death (Nguyen et al., 2018; Sarathy, Gruber, and Dick, 2019).

1.1.4 The Mycobacterial Cell Wall

Given that inhibition of cell wall biosynthesis is a common and highly effective mechanism of action for many antimycobacterial drugs²⁸, this structure is of clear importance to the survival and pathogenic success of these bacteria. *In vitro*, mycobacteria are unique microbes that grow in intricate serpentine cords along agar plates (Koch, 1882; Glickman, Cox, and Jacobs, 2000). These cords were among the first observations that helped to classify diverse mycobacterial species together and defining the ontogeny of these cords was of immense concern to early mycobacteriologists. By the 1950s these scientists had identified what they called the cord factor – a discrete molecule required for the cording effect seen in mycobacteria and that is, indeed, able to replicate key features of cording in isolation, even in the absence of bacteria, demonstrating that cording is a biophysical property of this underlying factor (Sorkin, Erlenmeyer, and Bloch, 1952; Bloch, Sorkin, and Erlenmeyer, 1953; Bloch and Noll, 1955; Behling et al., 1993; Hunter, Venkataprasad, and Olsen, 2006). This cording phenotype can play important roles during infection, where cords are difficult or impossible for macrophages to phagocytose and allow for immune evasion (Bernut et al., 2014; Ufimtseva et al., 2018).

The chemical composition of this cord factor was soon determined and this allowed

²⁸Isoniazid, ethambutol, ethionamide, pyrazinamide (indirectly), cycloserine, and vancomycin, among others (Nataraj et al., 2015).

it to be given a name – trehalose 6-6'-dimycolate or TDM (Noll et al., 1956). TDM features a trehalose head group and two long mycolate tails that can extend up to C₁₀₀ in length, creating an incorrigibly hydrophobic molecule that forms a thick amphipathic bilayer at the surface of the mycobacteria with the trehalose moieties facing the outside world and attached to the arabinogalactan layer below with a dimensionally thick²⁹ interior of interleaved mycolic acid chains (Glickman, 2008; Welsh, Hunter, and Actor, 2013; Bansal-Mutalik and Nikaido, 2014; Alderwick et al., 2015; Adhyapak et al., 2020; Batt, Minnikin, and Besra, 2020). TDM is the predominant mycolic acid species in this cell wall layer³⁰ and has been studied since its discovery for the ways in which it contributes to mycobacterial fitness in a diverse range of environments, including and especially the human host with even the earliest observations noting its toxicity (Bloch, Sorkin, and Erlenmeyer, 1953; Noll, 1956).

The bacterial surface is encapsulated by concentric layers of distinct biomolecules that serve distinct structural and immunomodulatory functions (Neyrolles and Guihot, 2011). Peptidoglycan is a ubiquitous feature of bacteria and the roles of peptidoglycan in modulating host immune responses have been reviewed extensively elsewhere (Kang et al., 1998; Guan and Mariuzza, 2007; Wolf and Underhill, 2018). Beyond and interspersed with the peptidoglycan cell wall, arabinogalactans serve as critical supports for the outer mycomembrane. However, these arabinogalactans also serve important immunomodulatory functions in yet another demonstration of the economizing nature of mycobacterial host subversion. For instance, lipoglycans can be detected by TLR2 and stimulate host inflammatory responses important for macrophage recruitment and granuloma formation (Gilleron et al., 2006).

²⁹Up to 40 nm in thickness.

³⁰40% of the total dry mass of mycobacteria is comprised of lipids, making these a critical part of the lifestyle of mycobacteria and exceptional among other bacterial genera (Jackson, 2014).

In addition to TDM, the cell wall or outer membrane contains many other biologically active compounds that play important roles in both bacterial homeostasis and host-pathogen interactions (Yu et al., 2012; Alderwick et al., 2015; Rens et al., 2021). One of these, phthiocerol dimycoceroserate (PDIM) appears to play a specific role in mycobacterial growth in the lung but not in other organs, perhaps implying a role for this lipid in adaptation to the primary site of infection in modern human tuberculosis (Cox et al., 1999). This molecule is able to mask underlying immunostimulatory molecules from detection by the host immune system; in the absence of PDIM, TLR-dependent recruitment of macrophages enhances bacterial clearance (Cambier et al., 2014; Day et al., 2014). It has also been suggested that PDIM facilitates mycobacterial escape from the phagosome of both host macrophages and lymphatic endothelial cells and, eventually, release into the extracellular space (Quigley et al., 2017; Lerner et al., 2018). Finally, in an entirely novel mechanism of action, PDIM dissociates from the *Mycobacterium* and integrates into the host cell membranes, where it suppresses immune activation; this depends on host cholesterol, providing an opportunity for host-directed intervention to overcome PDIM-dependent immunomodulation via use of widely prescribed statins (Stancu and Sima, 2001; Cambier et al., 2020).

A second class of modulatory lipids are the phenolic glycolipids (PGLs). PGL has been found to be essential for the bacterial transition from survival within tissue-resident macrophages, which have some microbicidal activity, into patrolling monocytes, which are thought to be even more permissive for bacterial growth (Sinsimer et al., 2008; Barnes et al., 2017). PGL stimulates the tissue-resident macrophages to produce CCL2, a monocyte-attractive chemokine, which enables the transfer of the bacteria into a more permissive growth environment. Acute blockade of CCL2 signaling improved bacterial killing by trapping them within the tissue-resident macrophages (Cambier et al., 2014; Cambier et al., 2017). In leprosy, PGL has been found to

stimulate macrophages to produce reactive nitrogen species that are able to damage nerve fibers, a major contributor to the demyelination effect seen in leprosy (Madigan et al., 2017). In tuberculosis, these reactive nitrogen species are thought to act as a microbial mechanism, but these radical species can also act to damage host tissues as well (Ohno et al., 2003; Voskuil et al., 2011; Idh et al., 2017). A final interesting cell wall lipid is sulfolipid-1, which had been thought to paradoxically act to restrict bacterial growth in macrophages (Gilmore et al., 2012; Seeliger et al., 2012), but has since been identified to specifically activate nociceptive neurons in the lung to induce coughing as a means to facilitate transmission to new hosts (Ruhl et al., 2020).

Due to this complex outer membrane structure, mycobacteria do not fit into standard binary classifications of bacteria within the Gram staining system³¹. While Gram-negative bacteria feature both an inner and outer phospholipid membrane, Gram-positive bacteria have only a single plasma membrane but are encased in a thick layer of peptidoglycan (Salton, 1953; Carbone, Valadez, and Pickett, 1982). Although evolutionarily derived from the Gram-positive bacterial phylum *Actinomycetota*³², mycobacteria possess features of both Gram-positive and Gram-negative bacteria: they have a single phospholipid bilayer and a thick peptidoglycan layer,

³¹The Gram stain utilizes a series of stains that bind to the peptidoglycan layer of Gram positive bacteria and render them visibly violet while Gram negative bacteria are counterstained magenta (Bartholomew and Mittwer, 1952).

³²This large phylum of bacteria includes incredible diversity and a number of other important human pathogens with varying degrees of relatedness to *Mycobacterium*. A notable example is *Corynebacterium diphtheriae*, the causative agent of diphtheria, which also produces mycolic acids, albeit shorter in length (van der Peet et al., 2015). The existence of a highly effective vaccine to diphtheria while no effective vaccine exists against tuberculosis is emblematic of the divergent strategies these species use to undermine their hosts. *C. diphtheriae* produces a classical toxin, diphtheria toxin, that is responsible for much of the pathology of disease while *M. tuberculosis* was thought to lack toxins until the discovery of the tuberculosis necrotizing toxin (TNT), although this is only selectively expressed and not thought to be absolutely essential for disease (Danilchanka et al., 2014; Sun et al., 2015; Pajuelo et al., 2018; Tak et al., 2019; Pajuelo, Gonzalez-Juarbe, and Niederweis, 2020; Pajuelo et al., 2021; Tak, Dokland, and Niederweis, 2021; Izquierdo Lafuente et al., 2021).

but also have an additional membrane comprised of mycolic acids which is occasionally referred to as the *mycomembrane* (Brennan and Nikaido, 1995; Chatterjee, 1997; Brennan, 2003; Jankute et al., 2015). This mycomembrane is the defining characteristic of mycobacteria and, as we have seen, plays critical roles in the lifestyle of the bacteria. Many of these roles are exercised by a single component – trehalose 6-6'-dimycolate, or TDM.

1.1.5 Trehalose 6-6'-Dimycolate (TDM)

The mycomembrane and its primary constituent, TDM, have many well-defined roles in providing tolerance to environmental stress, detoxifying reactive oxygen species (Chan et al., 1989), providing dehydration resistance (Harland et al., 2008), and modulating host immune responses (Lima et al., 2001; Ryall, Kumazawa, and Yano, 2001; Harland et al., 2008; Das, Murray, and Cross, 2013; Asmar et al., 2016; Hunter et al., 2016; Patin et al., 2017). TDM, for instance, is able to block a key step in phagosomal maturation, which would normally be able to kill the bacteria after uptake into phagocytic immune cells, including macrophages and neutrophils (Spargo et al., 1991; Indrigo, Hunter, and Actor, 2003; Axelrod et al., 2008). TDM also stimulates the production of a range of cytokines by macrophages and can autonomously induce granuloma formation (Korf et al., 2005). The broad ability of TDM to mediate mycobacterial interactions with the environment is one of the critical dimensions in the evolution of mycobacteria and the ability to then utilize novel modifications on this same molecular framework to undermine host immune responses appears to have been essential for their transition to a pathogenic or commensalistic³³ lifestyle

³³This notion of commensal mycobacteria warrants a vast degree of additional study. Although the laboratory model of non-pathogenic mycobacteria, *Mycobacterium smegmatis*, was isolated from syphilitic chancres and, later, smegma, very little is known about the niche of these commensal mycobacteria, how they maintain a neutral or neutral-positive relationship with their (often transient) hosts, and how their presence impacts host immunity to future encounters with pathogenic

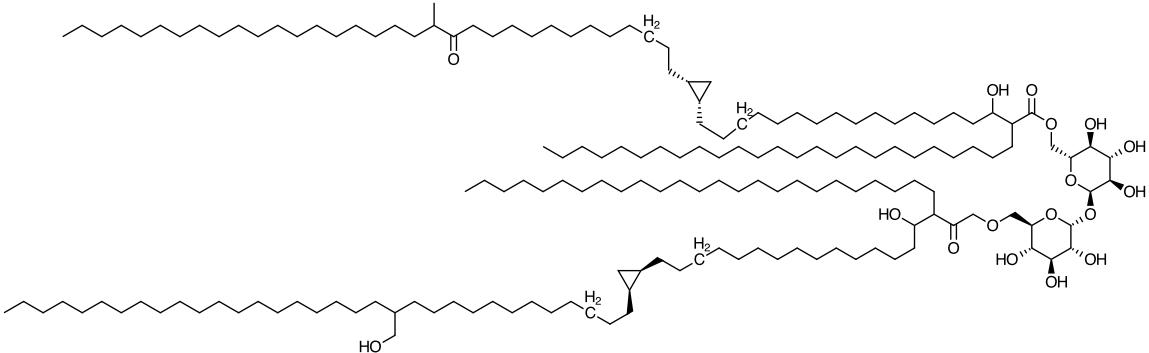


Figure 1.1: TDM is the primary component of the mycobacterial cell wall and can be seen here. At defined locations along the length of the mycolate chains, modifications can be added by a set of defined enzymes. The most notable of these modifications are the unusual and biologically rare cyclopropane modifications, which provide TDM a number of immunomodulatory characteristics.

in association with eukaryotic hosts ranging from amoeba³⁴ to fish to humans (Hagedorn et al., 2009; Salah, Ghigo, and Drancourt, 2009; Falkinham III, 2009; Delafont et al., 2014; Honda, Virdi, and Chan, 2018).

TDM, in many ways, defines the lifestyle of mycobacteria. As mentioned previously, this remarkably hydrophobic (indeed, wax-like) structure provides bacteria a potent tool for surviving not only harsh and fickle environmental conditions but also the conditions likely to be encountered in a host. This structure has been thoroughly dissected over the past decades of research, and a range of modifications are known that influence both the biochemical properties of the cell wall and the ways in which host organisms respond to this structure. Along the length of the mycolic acid tails, there are four main classes of modifications that may be present in two major locations. These modifications include methoxy, methyl, keto, and cyclopropyl groups,

mycobacteria (Robinson and Huppler, 2017).

³⁴As with many common human pathogens, it is theorized that mycobacteria developed the strategies necessary to overcome human macrophage responses on the amoebic training ground; the strategies utilized by amoeba to digest their prey are similar to those utilized by macrophages to eradicate invaders and virulence strategies that target these pathways are likely to be translatable across diverse organisms (German, Doyscher, and Rensing, 2013; Escoll et al., 2013; Price and Vance, 2014; Abukhalid et al., 2021; Fu et al., 2021).

which can be located at either proximal or distal locations (Sugawara et al., 2002; Minnikin et al., 2002; Takayama, Wang, and Besra, 2005; Rao et al., 2005; Rao et al., 2006; Bhatt et al., 2008; Barkan et al., 2009; Walton et al., 2018). Some of these modifications are represented along the structure shown in Figure 1.1. Of these, the most research interest has centered on the very unusual cyclopropane modifications, which add a great deal of energetic ring strain to the molecule and are, generally, unusual biological modifications due to their inherent instability and energy investment required to create (Wessjohann, Brandt, and Thiemann, 2003; Bach and Dmitrenko, 2004).

Cyclopropane modification of the proximal modification site (at approximately C₁₆) has been identified in both *cis* and *trans* isomers, each with distinct immunological properties and with both present within individual mycobacteria (Glickman, Cox, and Jacobs, 2000; Glickman, Cahill, and Jacobs, 2001; Rao et al., 2005; Rao et al., 2006). The *cis* modification was described first and is added to TDM by the protein product of the bacterial gene *pcaA*. *M. tuberculosis* deficient in *pcaA* is hypoinflammatory in a mouse model of infection, suggesting that *cis*-cyclopropane modified TDM is pro-inflammatory (Rao et al., 2005). Loss of this gene results in an overall reduction in bacterial survival. This somewhat contradictory result indicates that aspects of the host inflammatory response are important for bacterial survival and replication, findings that have since been replicated in a variety of other contexts in respect to tuberculosis disease (Koul et al., 2004; Flynn and Chan, 2005; Huynh, Joshi, and Brown, 2011; Sasindran and Torrelles, 2011; Tobin et al., 2012). Alternately, *trans*-cyclopropane modification of TDM is catalyzed by CmaA2³⁵ and this orientation was found to be hypoinflammatory (Rao et al., 2006).

³⁵Some redundancy is also seen with $\Delta mmaA2$, but these must have important different roles; MmaA2 also modifies the distal site of the mycolate chains (Glickman, 2003; Barkan et al., 2010).

Similar to $\Delta pcaA$ *M. tuberculosis*, loss of *cmaA2* resulted in a bacterial growth defect and prompt clearance of the bacteria, but by an alternative mechanism. Instead of a muted inflammatory response, $\Delta cmaA2$ *M. tuberculosis* induced hyperinflammation. Loss of both ($\Delta pcaA\Delta cmaA2$) results in hyperinflammation as well, suggesting that *trans*-cyclopropanation is a mechanism to dampen detection and this modification predominates in the absence of *pcaA* (Barkan et al., 2012). This body of work, largely from the Glickman lab, established a variety of important immunological roles for enantiomerically distinct versions of the same biomolecule that differ at only a single chemical site. This specificity is evocative of the high degree to which mycobacterial species have adapted to their hosts by developing novel modifications and mechanisms to perturb the immune response in their favor. By contrast, little has been described for roles of these modifications outside of modulating host immunity, suggesting that these may be innovations specific to the host-pathogen interaction, especially as these enzymes are only present in pathogenic species and are absent from *M. smegmatis*.

Models of TDM-host cell interactions are often lacking by virtue of the underlying chemistry of TDM. The profound hydrophobicity of TDM limits the avenues by which it can be experimentally presented to cells³⁶. On the surface of mycobacteria, TDM is (a) mixed with a range of other co-stimulatory molecules that may be important for the function of TDM (Rhoades et al., 2003; Torrelles and Schlesinger, 2010; Mazurek et al., 2012), (b) presented along the curved surface of a roughly-cylindrical bacillus (McCarter and Hastings, 1935; Krokowski et al., 2018), (c) constantly subject to remodeling as the chemically reactive components are oxidized (Chan et al., 1989; Hett, Chao, and Rubin, 2010; Meniche et al., 2014; Shaku, Ealand, and

³⁶For instance, direct addition of TDM to tissue culture media would result in a film at the surface, or at best, seemingly inert micelles.

Kana, 2020), and (d) exists in different physical conformations based on the degree of bacterial cording and abundance of dead mycobacteria³⁷. *In vitro*, these are difficult aspects to model and two major methods have emerged to agonize cultured cells with TDM: on the surface of polystyrene microbeads (Bloch, 1950; Retzinger et al., 1982; Behling et al., 1993; Indrigo, Hunter, and Actor, 2003; Geisel et al., 2005; Bowdish et al., 2009) and through evaporative monolayers on the surface of tissue culture plastic (Schabbing, Garcia, and Hunter, 1994; Hunter, Venkataprasad, and Olsen, 2006; Harland et al., 2008; Ishikawa et al., 2009; Miyake et al., 2013; Zhao et al., 2014). On beads of a small ($<5\text{ }\mu\text{m}$) diameter or in aqueous micelles, it adopts a bilayer configuration somewhat similar to that seen on live individual bacilli; on larger beads or on a plane, it acts as a monolayer. The monolayer configuration is more inflammatory but was also thought unlikely to exist *in vivo*, although this is subject to debate and cording may be a mechanism of mimicking this presentation based on the stage of the infection. Additionally, both interpretations rely on pure TDM in the absence of the complete milieu present on live mycobacteria; the inverse (removal of TDM from otherwise complete lipids) may be yet more informative as to the role of this unique compound.

While no comprehensive comparison between these exposure routes has been done, vague historical descriptions assign them two different behaviors. Dense surface monolayers of TDM are cytotoxic to cells and trigger a highly inflammatory response (Retzinger et al., 1982; Schabbing, Garcia, and Hunter, 1994; Hunter, Venkataprasad, and Olsen, 2006; Welsh, Hunter, and Actor, 2013); on the other hand, TDM on the

³⁷I hypothesize that TDM exerts distinct functions in early and late infection – early on, manipulation of the phagocytic macrophage is likely to be essential while in late infection it may be more important to modulate granuloma macrophages in the context of larger bacterial burden. Thus, it is sensible to think that the presentation on single bacilli and that on large clumps of bacilli may activate different receptors to different degrees. This is supported by analogy to Goodridge et al. (2011) but strict demonstration is likely to be challenging with existing tools and models.

surface of beads (although with some variation based on the diameter) tends to drive a more regulated response that still differs in some regards from that induced by whole, metabolically inactive mycobacteria (Bowdish et al., 2009; Welsh, Hunter, and Actor, 2013). While whole mycobacteria undoubtedly have other molecular patterns that augment the overall immune response, it is likely that the full breadth of the immune response to TDM has yet to be fully uncovered on account of deficient models to do so³⁸. The physiological relevance of these monolayer-like configurations of TDM is up to some debate, but there is some evidence that planes of TDM from dead mycobacteria can form *in vivo* (Schabbing, Garcia, and Hunter, 1994; Hunter et al., 2006; Glickman, 2008). Additionally, the clumps of bacteria that form within the necrotic core may grow in larger macrostructures resembling these artificial planes. Despite the relative opacity of these historical observations, what is clear is that TDM must be presented to cells in particular arrangements to have particular effects, which is seen in head-to-head comparisons between heat-killed *M. tuberculosis* and gamma-irradiated *M. tuberculosis* (Datta et al., 2006; Cha et al., 2015; Gleeson et al., 2016; Yang et al., 2018; Krokowski et al., 2018; Mosavari et al., 2021). While gamma-irradiated *M. tuberculosis* maintain their shape and structure, heat-killed *M. tuberculosis* are broken down, dampening their ability to stimulate the immune system (Carpenter et al., 1959; Secanella-Fandos et al., 2014).

Moonlighting

Pathogenic microorganisms are often constrained by genomic size – too small and too few essential functions can be encoded; too large and the risk of duplication errors

³⁸Some model of total lipid reconstitution on freely associating bacillus-sized beads within an extracellular matrix may get us closer to this – isolating lipid activity in a native context but in the absence of confounding bacterial physiology (Retzinger et al., 1982; Rhoades et al., 2003; Sakamoto et al., 2010).

and energetic cost of maintenance becomes prohibitory (Ranea et al., 2005; Huberts and Klei, 2010; Bobay and Ochman, 2017). There is therefore a great deal of evolutionary pressure to economize and multitask – why have two proteins to do two functions if one can do both? That is the precise logic underlying many bacterial toxins, secreted effectors, and structural features (Gupta et al., 2019). One of the most famous of these multifunctional proteins, often dubbed moonlighting proteins³⁹, is the alpha-enolase from *Streptococcus pneumoniae* (Bergmann et al., 2001). Enolase is an enzyme critical to glycolysis and converts 2-phosphoglycerate to phosphoenolpyruvate, which is an essential intermediate in the breakdown of glucose into pyruvate to generate ATP and NADH. However, *S. pneumoniae* also secretes this normally cytosolic enzyme onto the surface of the outer membrane, which allows it to interact with host plasminogen and catalyze its conversion into active plasmin (Bergmann, Schoenen, and Hammerschmidt, 2013). Plasmin degrades host fibrin clots, leading to enhanced tissue invasion and pathogenicity through avoidance of pathogen containment by host fibrin and increased dissemination (Whiting et al., 2002; Weiser, Ferreira, and Paton, 2018). By evolutionary addition of new properties, fusion of two unrelated proteins into a single protein, alterations of protein localization, or novel layers of regulation, bacteria can, in a very efficient way, exert multiple essential functions from single biological products.

Within mycobacteria, there are many notorious examples of “moonlighting” proteins, including the enolase of *M. tuberculosis*, which also binds plasminogen⁴⁰ (Rahi

³⁹Conceptually, of course, moonlighting is purely orientational. While the given example is one instance where a historically well-defined enzyme has additional functions based on localization, other multifunctional enzymes that can target both bacterial and host substrates or that have distinct functions when cytosolic or periplasmic or secreted are unlikely to be given this title unless they bear high homology to universally conserved proteins.

⁴⁰As will become relevant later in this document (Subsubsection 5.5.3), plasmin also has important regulatory function during angiogenesis. How mycobacterial engagement of plasmin(ogen) alters the overall angiogenic response is an interesting question that warrants future study.

et al., 2017). Indeed, enolase appears to be a highly conserved moonlighting protein, with plasminogen binding having been demonstrated for numerous other species as well (Seweryn et al., 2007; Vanegas et al., 2007; Agarwal et al., 2008; Candela et al., 2009; Floden, Watt, and Brissette, 2011; Figueiredo et al., 2015). *M. tuberculosis* has other metabolic enzymes that are used to subvert the host immune response. For instance, mycobacterial MenJ is a vitamin K oxidoreductase that confers intracellular survival within macrophages independent of its catalytic activity (Kumar et al., 2020b). Similarly, ArgD is an acetylornithine aminotransferase that also acts as a potent TLR4 agonist (Nehvi et al., 2022). This repurposing of metabolic enzymes is one of many notable ways of subverting host immunity while optimizing genetic coding space (Banerjee et al., 2004; Banerjee et al., 2007; Henderson, Lund, and Coates, 2010). Given that *M. tuberculosis* has lost approximately 30% of its genomic content relative to environmental mycobacteria, streamlining has played an important role in adaptation to an exclusively pathogenic lifestyle (Stinear et al., 2008).

Similar to protein examples, which tend to be more obvious, the structural features of the bacteria can also serve important moonlighting functions in the sense that single compounds can play key roles in seemingly unrelated phenomena. TDM is an excellent example of this – it is a conserved feature of non-pathogenic mycobacterial species, suggesting that this feature likely emerged to address environmental needs that preceded the need to engage with host immunity (Kremer et al., 2002; Pacheco et al., 2013). However, the adaptation to a pathogenic lifestyle has resulted in TDM being repurposed with additional modifications to suit these new demands. Indeed, TDM serves such a wide array of important functions in the physiology of (especially pathogenic) mycobacteria that to assign it a “major” function would be rather fallacious. As a major structural component of the cell wall, defense from the environment, broadly speaking, is clearly the overarching theme of this sophisticated

glycolipid.

Strictly in the context of host immunity, TDM had been generally ascribed a few major roles: (a) blockade of lysosomal-phagosomal fusion (Indrigo, Hunter, and Actor, 2003; Axelrod et al., 2008; Patin et al., 2017), (b) alteration in expression of major immunoregulatory cytokines (Perez et al., 2000; Indrigo, Hunter, and Actor, 2002; Bowdish et al., 2009; Sakamoto et al., 2013), (c) induction of humoral immunity (Fujiwara et al., 1999; Ryll, Kumazawa, and Yano, 2001; Fujita et al., 2005), (d) stimulation of hypercoagulation (Retzinger et al., 1982; Retzinger, 1987; Donnachie, Fedotova, and Hwang, 2016), (e) interaction with the stromal milieu (Sakamoto et al., 2010), (f) mediation of granuloma formation (Bekerkunst, 1968; Hunter et al., 2006; Lee et al., 2012), and more. Delipidation of mycobacteria results in a profound alteration of the overall inflammatory response *in vitro* and results in efficient bacterial killing by macrophages but perturbed expression of IL-1 β , TNF- α ⁴¹, IL-6, and IL-12. It is now thought that many of these functions are mediated by recognition of TDM by surface host receptors⁴², a topic that will be returned to shortly. However, the expression of these critical cytokines (among many others) regulated by TDM results in profound changes in the overall tone and tempo of the inflammatory response that, in aggregate, contribute to granuloma formation, a process we now know to be dependent on both pro- and anti-inflammatory signaling molecules, including IL-4, IL-13, IFN- γ , TNF- α , and nitric oxide (Flynn et al., 1993; Cooper et al., 1993; Bergeron et al., 1997; MacMicking et al., 1997; Kaneko et al., 1999; Akdis et al., 2011; Cavalcanti et al., 2012; Cronan et al., 2021). These processes are intimately

⁴¹TNF- α was once commonly called “cachectin” for its ability to induce cachexia (Tracey, Lowry, and Cerami, 1988). TNF- α is probably the defining cytokine of tuberculosis infection and is critical for bacterial control (Orme, 1998), although others may, with equal validity, argue that this honor goes to IFN- γ .

⁴²Whether or not these receptors detect mycobacteria within the phagosome is unknown – this may be able to explain some, but not all, of the influence of TDM on infected macrophages.

linked with the phenotype that will be further explored throughout this work: the TDM-dependent induction of VEGFA within granuloma macrophages and resultant neighboring angiogenesis.

1.2 Macrophages in Development and Disease

Macrophages are developmentally essential for the survival of vertebrate organisms. These phagocytic cells of the hematopoietic lineage perform vital roles in organ development, immune maturation, vascular remodeling, scavenging for cellular debris, and defense against pathogens (Wynn, Chawla, and Pollard, 2013; Wattrus et al., 2022). They are both continually replenished by hematopoiesis in the bone marrow and migration into destination tissues as well as self-renewing within essentially every tissue in specialized form – microglia in the brain, alveolar macrophages in the lung, Kupffer cells in the liver, osteoclasts in the bone, Langerhans cells in the skin, histiocytes in connective tissue, and varying generically-named macrophages in the heart, blood vessels, adipose tissue, lymphatic tissue, gonads, and the intestinal tract (Davies et al., 2013a; Bain and Schridde, 2018; Na et al., 2019). Under normal conditions, these macrophages circulate through the blood as precursors known as monocytes which, themselves, have important roles in immunity (Sampath et al., 2018; Mass, 2018). Upon activation by an outside stimulus, these monocytes will extravasate and undergo tissue-educated differentiation into macrophages. Macrophages are the central orchestrators of immunity to tuberculosis, acting as the first responders and comprising the host-pathogen interface, being the primary cellular constituent of the granuloma (Pagan and Ramakrishnan, 2018). These macrophages must be capable of both initiating and resolving inflammation, finely tuning their responses dynamically over the course of an insult in order to simultaneously clear

invading pathogens, limit the pathology of inflammation, and ultimately repair damaged tissues (Unanue and Allen, 1987; Murray and Wynn, 2011; Watanabe et al., 2019).

1.2.1 The M1-M2 (False) Dichotomy

For the past two decades, macrophage research has been dominated by the concept of the M1-M2 binary of polarization states (Italiani and Boraschi, 2014; Mills, 2015). This concept, introduced by Mills et al. (2000) is a useful paradigm to understand the activity of macrophages *in vitro* and mirrors certain aspects of *in vivo* biology, but proves inadequate to capture the diversity and ironies of macrophage behaviors. M1 macrophages are described as “classically” activated and are produced in response to TLR agonists (see Subsection 1.3.1 for more details) and IFN- γ while M2(a) macrophages differentiate in response to IL-4/IL-13 (Wynn, Chawla, and Pollard, 2013; Mills and Ley, 2014). This neatly classifies these macrophages into inflammatory and anti-inflammatory in a deliberate parallel to the T_H1/T_H2 T cell paradigm. Further research has identified a number of subtypes of M2 macrophages, including M2b (immune complexes, IL-1, TLR agonists), M2c (IL-10, TGF- β , glucocorticoids), and M2d (IL-6, TLR agonists, adenosine) (Huang et al., 2018; Viola et al., 2019). What this research has revealed is that macrophages are fascinatingly diverse cells, able to respond intelligently to different stimuli and dynamically alter their responses through integration of new information. In the era of single cell RNA sequencing and an ever-greater number of single cell -omics approaches, it has become apparent that *in vivo*, this binary does not exist. These subtypes are defined based on their treatment *in vitro* with a limited and defined subset of stimuli (Wynn, Chawla, and Pollard, 2013; Gosselin et al., 2014); in a tissue context, cells are continually bombarded with often conflicting signals that they must synthesize to create a coherent set

of actions (Murray et al., 2014) and macrophages are imprinted by their phagocytic history, generating further layers of heterogeneity (A-Gonzalez et al., 2017). What has been proposed instead, by Nahrendorf and Swirski (2016) in a beautiful opinion piece, is a network of macrophage activation possibilities that realistically acknowledges the limitations of the current model. Macrophages are capable of expressing both M1 and M2 signatures simultaneously in ways that defy typical classification, but if only a limited subset of markers are assessed, then a macrophage population may be erroneously deemed “M1” or “M2”. This limits the degree to which the dominant M1-M2 model is able to reflect any real biology *in vivo*.

While this M1-M2 nomenclature will inevitably come up throughout the course of this thesis, it is suggested that any in-text use be taken as a pure reflection of the historical context of the publication being cited and lack of additional information to call them otherwise⁴³. I will not use these terms to describe any of the macrophages in the experiments presented in Chapter 3 and this choice is deliberate in pursuit of understanding the biology of the macrophage, not simply what it might be called.

1.2.2 Developmental Origins

Two cell types are a ubiquitous presence in every tissue in the body: endothelial cells and macrophages. These two tissues serve as the support system for all other bodily functions. The endothelium does so by providing oxygen and nutrients to these tissues, as well as providing a signaling infrastructure to alter tissue development and

⁴³The distinctions between different T cell differentiation states appear to be more in line with proper differentiation and not simply activation. Type I responses are those that induce inflammation while type II responses are responsible for tolerance and resolution. There are additional T cell activation states (many of them) that reveal some of the discrete loci along the gradient which immune responses can exist. This is beyond the scope of the present work, but I am not denying the existence of immune polarization and type responses, merely the binary classification of the highly plastic macrophages based on highly controlled *in vitro* exposures to defined cytokines.

homeostasis⁴⁴. Macrophages, on the other hand, serve to actively maintain tissue structure and health through the production of a range of cytokines, chemokines, and signaling molecules and patrol tissues to clear cell debris (Wynn, Chawla, and Pollard, 2013). During infection or other tissue damage, these tissue-resident macrophages are able to quickly respond to eradicate most invaders and return the tissue to homeostasis. Macrophages are also intimately involved in the development of most or all tissues and emerge early in embryonic development to serve in this role.

Macrophages first emerge from the yolk sac via primitive hematopoiesis at approximately six weeks post fertilization in humans (Geissmann, 2010; Yona et al., 2013; Feyaerts et al., 2022). Each of these in its own tissue is essential for remodeling and homeostasis both in early embryonic development and in developmental processes throughout life as they self-renew in addition to contributions from circulating monocytes (Jappinen et al., 2019). Microglia, the first of these to emerge, are important for response to neurological damage and scavenge for cell debris; developmentally, they are important for synaptic pruning and maturation for the solidification of knowledge and behavior (Wynn, Chawla, and Pollard, 2013; Lavin et al., 2015). In the absence of microglia, synapses grow unrestricted and form aberrant connections in a way that mirrors the immature brain and greater numbers of neurons develop and survive but lack mature synapses indicative of cohesive pruning (Hammond, Robinton, and Stevens, 2018). And, in a theme that will be present throughout different contexts, these macrophages are important for the elaboration

⁴⁴The relative contributions of these two cell types to development have largely been revealed by the differential lethality of mutations affecting these individual systems, which both arise from a common hemangioblast precursor that was implied by the existence of the *cloche* mutation (Stainier et al., 1995; Vogeli et al., 2006). Mutation in *myb* result in a total loss of blood cells, but this is viable into adult stages of development in the zebrafish (Lipsick, 2010; Soza-Ried et al., 2010). By contrast, loss of the two homologs of *VEGFA* in the zebrafish (*vegfaa* and *vegfab*) or the *kdrl* receptor results in embryonic lethality (Carmeliet et al., 1996). These will be discussed slightly more in Subsection 2.2.1.

of the brain vasculature. Microglia secrete VEGFA to guide the production of blood vessels throughout the brain; given the sizable demands of the brain for oxygen and glucose, this is a patently important role for these cells from the earliest stages of life (Dudiki et al., 2020).

Subsequently, other types of specialized macrophages emerge and populate other organs, including Langerhans cells, Kupffer cells, and alveolar macrophages (Davies et al., 2013a; Wynn, Chawla, and Pollard, 2013; Lavin et al., 2015; Gordon and Pluddemann, 2017). These macrophages must all deal with the environment at large either through direct contact in the airway or the skin or indirectly via toxin processing in the liver. Langerhans cells are important for immediate host defense against invading pathogens at the skin and are additionally required at homeostasis for important self-renewal processes, including hair growth and skin shedding (Merad, Ginhoux, and Collin, 2008; Theret, Mounier, and Rossi, 2019). Kupffer cells are responsible for modulating liver filtering functions, including by modulation of liver uptake of cholesterol; they are also able to combat pathogens and facilitate the removal of dying cells and are important for liver regeneration (Theret, Mounier, and Rossi, 2019; Demetz et al., 2020).

Alveolar macrophages – the primary tissue-specific macrophage subtype of interest in tuberculosis – are extremely plastic cells that are essential for the maintenance of lung health both from natural cellular turnover in the lung and resorption of pulmonary mucus and due to the abundance of potential respiratory pathogens faced in the daily life of any lunged animal (Hussell and Bell, 2014). Alveolar macrophages are self-renewing and, after establishment from the yolk sac during development, are capable of perpetuating themselves throughout life with only modest contributions from circulating monocytes (Hashimoto et al., 2013; Yona et al., 2013; Varol, Mild-

ner, and Jung, 2015). Inflammation in the lungs is clearly harmful for normal bodily functions and hyper-inflammation is a death knell (Kemp and Lockey, 2002). Thus, alveolar macrophages must finely tune their responses to pathogens and lung damage to facilitate pathogen clearance and tissue repair. In general, alveolar macrophages have a muted inflammatory immune response *in vitro*⁴⁵, evocative of their delicate role in maintaining lung function (Joshi, Walter, and Misharin, 2018; Svedberg et al., 2019). Like most other tissue-resident macrophages, these cells evade clear classification on the M1-M2 polarization axis and can span the full range of these phenotypes depending on the circumstance and in response to cytokines generated by other cells (Hussell and Bell, 2014; Svedberg et al., 2019).

In their particular tissues, all of these macrophage subsets are responsible for both inducing inflammation and resolving it. These cells must plastically respond to ever-changing conditions to exact the proper type of response at the appropriate time (Pollard, 2009). They must also change over the course of the life of the organism, transitioning from early roles in aiding the development of tissues to maintaining homeostasis and, eventually, managing the aging process (Linehan and Fitzgerald, 2015; Duong et al., 2021; Guimaraes et al., 2021; De Maeyer and Chambers, 2021). An interesting additional role for macrophages that is just beginning to be studied is their capacity to physically transmit signals between tissues. A recent study found that pigment cells in the zebrafish were remodeled based on interactions with macrophages and, in their absence, melanocyte patterning is disrupted. This argues that macrophages, via their intrinsic motility, may be able to augment long-range communications to facilitate tissue remodeling and repair (Eom and Parichy, 2017). Macrophages also exist at the very foundation of immune system development, being

⁴⁵Although it is relatively easy to harvest alveolar macrophages by bronchoalveolar lavage, providing them a stable environment *ex vivo* seems to be rather challenging. While this is the closest that can be gotten to *in vivo*, this too comes with some caveats.

the master regulators of hematopoietic stem cell survival, thus determining the clonal populations that contribute to all hematopoietic lineages throughout life (Watrus et al., 2022).

1.2.3 Alveolar Macrophages in Tuberculosis

Alveolar macrophages are the first responders to tuberculosis infection, being already present at the site (Flynn and Chan, 2001). Their biology is fundamentally a reflection of the niche in which they reside – they utilize oxidative phosphorylation as a metabolic modus to take advantage of the abundant oxygen present and respond modestly and slowly to insult as a means to minimize pathology (Joshi, Walter, and Misharin, 2018). These macrophages are thus of great interest for the roles they play in the overall pathogenesis of *M. tuberculosis*. Alveolar macrophages have been shown to be more susceptible to infection by mycobacteria, in general, than are the interstitial macrophages to soon arrive at the site, in part due to their mixed activation state and delayed responses (Kahnert et al., 2006; Madden et al., 2022), although circulating monocytes appear to be more susceptible still (Cambier et al., 2014; Cambier et al., 2017). Early studies found that *M. tuberculosis* induces apoptosis of alveolar macrophages to facilitate their escape into the interstitial space (Keane et al., 1997); further study actually uncovered that, although virulent *M. tuberculosis* was able to induce apoptosis, it actually did so *less* than avirulent strains, suggesting that some virulence factor limited apoptosis to provide the bacteria a temporary replicative niche (Keane, Remold, and Kornfeld, 2000; Cohen et al., 2018). Subsequent findings on apoptosis as a host-protective response align well with these data and necrosis is *M. tuberculosis*'s preferred means of cellular exit from macrophages (Behar et al., 2011), much of which is interconnected with the process and consequences of efferocytosis by other innate immune cell types (Martin et al., 2012). Some of the

foundational studies on *M. tuberculosis* macrophage subversion were in the specific context of alveolar macrophages; Mwandumba et al. (2004) found that the bacteria are able to reside in pH-neutral phagosomes within alveolar macrophages, allowing them to survive and replicate. Consistent with these findings, it was uncovered that depleting alveolar macrophages actually results in improved bacterial control, implicating alveolar macrophages as a weak link in the overall response to tuberculosis – shielding the bacteria until their population had expanded beyond the reasonable limits of control able to be exerted by recruited macrophages and neutrophils (Leemans et al., 2001).

These infected macrophages are also able to shuttle the bacteria out of the lung and into the interstitium, an event that long precedes bacterial infection of other cell types and the establishment of a productive infection (Cohen et al., 2018). However, alveolar macrophages from patient contacts have increased capacity to restrict bacterial growth, perhaps in an instance of “trained immunity”⁴⁶ exerting protective effects after sub-clinical exposure (Carranza et al., 2006; Divangahi et al., 2021).

If alveolar macrophages are such poor responders to tuberculosis, are other cell types better able to control the infection? This has been a critical question in the development of new therapeutic options. While a thorough discussion of what is known about neutrophils is beyond the scope of this work, the general conclusion on the role of neutrophils in the pathogenesis of tuberculosis infection has been equivocal, with different groups using different models coming to disparate and difficult-to-

⁴⁶The concept of trained immunity or innate immunological priming is an interesting concept that has been put into use (largely by accident) for decades. While it is beyond the scope of the present work to give it its proper discussion, further reading can be found at (Arts et al., 2018; Moorlag et al., 2020; Khan et al., 2020; Netea et al., 2020; Katzmarski et al., 2021; Trauer et al., 2021; Kaufmann et al., 2022; Katzmarski et al., 2022). The mechanism of BCG in the treatment of bladder cancer is largely through trained immunity (Redelman-Sidi, Glickman, and Bochner, 2014) and BCG is being evaluated as a trained immunity vaccine against COVID-19 and potentially future pandemics for which no specific vaccine yet exists (Redelman-Sidi, 2020).

reconcile conclusions (Fortune and Rubin, 2007; Yang et al., 2012; Srivastava, Ernst, and Desvignes, 2014). On the other hand, the critical importance of hematopoietic-derived macrophages is clear: these cells are absolutely required for the effective control of tuberculosis infection and, in their absence, bacteria grow uncontrolled in the host (Clay et al., 2007; Pagan et al., 2015; Matty et al., 2019).

1.2.4 Recruited Macrophages in Tuberculosis⁴⁷

At homeostasis, non-tissue resident macrophages circulate through the blood stream as monocytes. While these comprise a minority of the total macrophages in the body, they are key respondents to sites of insult to mediate pathogen control and wound healing. In the classical model, these then differentiate into more M1-like macrophages, expressing IFN- γ , TNF- α , and IL-1 β (Arango Duque and Descoteaux, 2014). Despite their critical importance, relatively little is actually known about the biology of macrophages derived from circulating monocytes in the initial events of the immune response to tuberculosis. For the first fifteen days of infection (in mice), the entire course of infection is dictated by alveolar macrophages, which are the near-exclusive hosts of mycobacteria until that time point, at which time monocyte-derived macrophages and neutrophils become the major host cells (Cohen et al., 2018). Recruited monocytes sense a number of chemoattractants to direct them to the site of infection and then differentiate into macrophages, including type I interferon and CCL2 (Peters et al., 2001; Peters et al., 2004; Antonelli et al., 2010; Desvignes, Wolf, and Ernst, 2012). Antonelli et al. (2010) concluded that the recruitment of these macrophages exacerbated infection as these were pathogen-permissive cells; however, this paints a picture of *all* macrophages as being detrimental for disease when synthesized with the

⁴⁷While books could be written on the body of *in vitro* macrophage responses during infection, I have deliberately attempted, with varying success, to narrow the focus onto known *in vivo* biology to aid in differentiating macrophages by their ontogeny.

findings from Leemans et al. (2001), who found that alveolar macrophage depletion ameliorated disease. What to make of this then? It appears that macrophage-derived type I interferons (IFN- α/β) are, indeed, detrimental during tuberculosis and engage in discrete functions from the classically protective IFN- γ . Additionally, CCR2 is an important protective chemokine through its ability to recruit interstitial macrophages to the lung (Peters et al., 2001), although the net effect of this appears to result overall in worse disease outcomes (Cambier et al., 2014). Many of the ultimate effects are ascribed to a failure to recruit T cells after infection, but clearly there are potentially essential roles for these recruited macrophages in directly protecting against tuberculosis.

Much of what we know about the interactions between macrophages and pathogenic mycobacteria come from *in vivo* observations in the zebrafish model⁴⁸⁴⁹. Genetic depletion of all of the macrophages in the developing zebrafish larva results in unrestricted growth of bacteria, suggesting that patrolling macrophages are important mediators of host protection during the earliest events of infection (Clay et al., 2007). Similar to the findings from Cohen et al., it was found that these macrophages are important for the dissemination of the bacteria into other tissues; it is possible that macrophages are broadly able to fulfill this role depending on the particular disease context and alveolar macrophages happen to be the population of interest in pulmonary tuberculosis. These findings contrasted with those found in previous models, where wholesale depletion of all macrophages in the mouse lung was

⁴⁸For now, this is the extent of required detail. More will be provided in Section 2.2, but suffice to say that, given all that is known about the differences in *in vitro* and *in vivo* macrophage biology, this serves as a surrogate, optically accessible host for studying the earliest events of infection in a native tissue context.

⁴⁹There are many studies of the *in vitro* interactions between macrophages and mycobacteria. These are extremely valuable studies that model important aspects of these interactions that have later been demonstrated to be critical *in vivo*. I will focus on those findings that are directly relevant to the *in vivo* context for these purposes.

able to decrease bacterial burden (Leemans et al., 2005). Further, it is clear from human population studies that macrophage deficiencies increase susceptibility to tuberculosis (Hambleton et al., 2011), although functional differences in macrophage “deficiency” and macrophage eradication are obviously possible, as are unintended consequences of genetic mutants that are unrelated to their effects on macrophage number. Nevertheless, the aggregate of these data demonstrates that patrolling macrophages recruited to the site of infection can exert protective activity against tuberculosis while resident alveolar macrophages are unable to do so on account of their tissue-specific niche and more limited microbicidal activity.

These data evoke the roles of macrophages in “undisturbed” tuberculosis infection, but a comprehensive analysis must integrate the existing therapies used to treat tuberculosis, especially in the context of drug-resistant strains. Much ink has been spilled on the role of the granuloma as a physical barrier to therapeutic access during infection (Driver et al., 2012; Ekins, 2014; Cicchese et al., 2020; Cronan, 2022), but Adams et al. (2011) identified a role for macrophage drug efflux in inducing drug tolerance in mycobacteria. By allowing the bacteria to grow undisturbed while being exposed to bactericidal concentrations of antitubercular drugs, the macrophages can act as a protective niche against treatment. While these roles clearly did not evolve on account of the modern treatment regimen, they reveal an interesting dimension of the cell biology of the bacteria, wherein macrophages actively efflux drugs that ultimately protects the pathogen; notably, treatment with efflux pump inhibitors lowers intra-macrophage bacterial burden by increasing the effective drug concentration. Experimentation to identify whether this process is similarly protective against metallotoxicity or other endogenous factors would provide further context to the evolutionary origins of this phenomenon.

1.2.5 Host Resistance and Susceptibility to Infection

Mediated by Macrophages

Macrophages play extremely complex and often contradictory roles in the control of infection based on their responses at particular times along a very fine gradient, where seemingly subtle shifts can result in radically different functional outcomes. While there is a great deal known about outright susceptibility to infection, including the Mendelian susceptibility to mycobacterial disease, which is caused by errors in the response to IFN- γ (Bustamante et al., 2014), many or most of these levels of resistance are likely to be mediated by macrophage behaviors, which are certain to be the major player for any response that does not stimulate an adaptive response detectable by skin test. Particular genetic disorders are known to lead to increased susceptibility to mycobacterial infections, including normally opportunistic mycobacteria. For instance, lysosomal storage disorders alter macrophage behavior and migration to increase patient susceptibility to tuberculosis (Berg et al., 2016). More general approaches have also been taken to identify macrophage genes that mediate susceptibility to tuberculosis (Thuong et al., 2008), but this remains a field worth additional exploration.

There is an enormous body of literature on the contributions of macrophages in the general execution of control in tuberculosis infection. Although literature descriptions tend to focus on the many ways that the bacterium can subvert the capacity of the macrophage to kill it, the previously detailed evidence on the consequences of macrophage depletion suggest that macrophages are capable of some level of effective control (Pagan et al., 2015). Macrophages natively express a spectrum of pattern recognition receptors (PRRs), genetically encoded receptors that bind discrete ligands from bacteria, viruses, fungi, parasites, and damaged-self which allows cells of the

innate immune system to discriminate between self and non-self. These receptors are not only capable of activating the macrophage to execute more effective antimicrobial functions, they also serve as a critical step in macrophage antigen presentation for the purpose of stimulating a potentially protective adaptive response.

The earliest studies on this topic identified a set of cytokines that were clearly important for the control of tuberculosis: TNF- α , IFN- γ , and IL-12 among them (Flynn et al., 1993; Cooper et al., 1993; Flynn et al., 1995; Cooper et al., 1997). Further studies on human genetics found that variations in the NRAMP1 gene conferred resistance to tuberculosis infection (Bellamy et al., 1998), perhaps through increased metal ion transport into the phagosome (Davies and Grange, 2001a). Similarly, polymorphisms in the promoter for hepcidin, which regulates iron transport, alter susceptibility to disseminated tuberculosis (Liang et al., 2017). Indeed, one of the major functions of IFN- γ and TNF- α is to further activate macrophages to execute bactericidal control (Kaufmann, 2002). TNF- α is also essential for the development and stability of granulomas in the mouse, presenting a multifaceted role for this critical cytokine (Chakravarty et al., 2008). Despite the classical model of mycobacterial blockade of phagosomal-lysosomal fusion, some cells manage to complete this fusion and kill the contained mycobacteria (Kaufmann, 2002).

Subsequent studies have revealed additional macrophage gene products that undermine the host immune response and serve important bacterially beneficial roles. Two of these – CCL2 and MMP9 – are important for recruiting additional macrophages and restructuring the interstitial microenvironment (Volkman et al., 2010; Cambier et al., 2014). Due to the liabilities impose by these genes on the progression of disease, it seems plausible that targeting them may offer an alternative path for disease treatment. Additionally, unlike previous studies, these offer unequivocal host

susceptibility responses; knockout of IFN- γ or TNF- α result in increased bacterial burden and defective granuloma formation whereas abolition of MMP9 and CCL2 resulted in reductions in both granuloma formation *and* bacterial burden.

Other signaling events can also play important roles in host defense against tuberculosis infection, especially other aspects of the cytokine environment that can exact control of mycobacteria. One of the most recently described of these is the capacity for GM-CSF to activate macrophages to control bacterial replication, a process that is inhibited by HIV-1 coinfection (Bryson et al., 2019). Interestingly, addition of GM-CSF to the macrophages facilitates renewed control, suggesting that cytokine re-regulation may be an avenue for new therapies as it is in many other chronic infections; by returning the milieu to a proper tone, it may be possible to encourage macrophages to kill the bacteria more effectively despite the measures taken by the bacteria or comorbidities to prevent this outcome. The nature of macrophages during HIV infection is an interesting question given that these cells can be directly infected by the virus and that tuberculosis is such a common and severe disease among AIDS patients. Alveolar macrophages display altered activation status and serve as a reservoir for HIV, which compromises the earliest events of the response to infection (Evans and Wansbrough-Jones, 1996; Herbein et al., 2002; Cassol et al., 2010; Cribbs et al., 2015; Clayton et al., 2017; Kruize and Kootstra, 2019). These alterations make the macrophages resistant to killing by cytotoxic T lymphocytes as well as impaired for the induction of key anti-inflammatory responses (Porcheray et al., 2006; Herbein and Varin, 2010; Clayton et al., 2018; Neff et al., 2020). Together, the increased susceptibility to tuberculosis infection during AIDS results in combinatorial innate and adaptive immune suppression that leads to rapid disease progression due to the effects of HIV in killing CD4 $^{+}$ T cells and modulating macrophages responses with mycobacterial killing of macrophages within the lung (Mariani et al., 2001).

Non-proteinic signals can also intersect with these events, including nitric oxide, hydrogen sulfide, and superoxide, which are widely produced metabolites that have many functions in homeostasis and disease. Nitric oxide is a potent vasodilator⁵⁰ and oxidant that is able to kill mycobacteria, but also prevents the infiltration of pathogen-permissive neutrophils into the granuloma, reducing bacterial burden (MacMicking et al., 1995; MacMicking, Xie, and Nathan, 1997; MacMicking et al., 1997; Szabo, 2007; Mishra et al., 2017b). Hydrogen sulfide also acts in an immunosuppressive fashion and prevents macrophage-mediated inflammation and reduces production of protective cytokines, but this suppressive phenotype results in increased bacterial burden (Rahman et al., 2020). Superoxide production promotes tolerance to infection to sustain host survival even at the same bacterial burden, a new and more complex role for these simple compounds on determining host outcomes (Olive et al., 2018).

Macrophage cell death pathways can play important roles in priming later steps of protection. Through the process of necrosis or necroptosis, macrophages release bacteria into the tissue to infect new macrophages, a process that the bacteria actively exploits through production of TNT⁵¹ (Guirado, Schlesinger, and Kaplan, 2013; Pa-

⁵⁰Nitric oxide release is the primary mechanism of action for nitroglycerin treatment during infarction.

⁵¹For many years, tuberculosis was thought to lack traditional toxins that define the pathogenicity of many other bacterial pathogens, such as *Shigella* and *Vibrio cholerae* (Holmgren, 1981; Tesh and O'Brien, 1991; Gyles, 2007; Guichard et al., 2013). However, recent findings have identified the existence of the tuberculosis necrotizing toxin, or TNT, a subunit of the outer membrane protein CpnT that can be cleaved off and exported into the host macrophage where it induces an immunologically blunted necrosis, allowing the bacteria to escape and infect new host cells (Danilchanka et al., 2014; Pajuelo et al., 2021; Tak, Dokland, and Niederweis, 2021; Izquierdo Lafuente et al., 2021). TNT acts as an NAD⁺ glycohydrolase, depleting host cell stores of NAD⁺ and inducing cell death (Sun et al., 2015; Tak et al., 2019; Pajuelo, Gonzalez-Juarbe, and Niederweis, 2020). *M. tuberculosis* itself encodes an anti-toxin that prevents bactericidal activity while maintaining effective control of host cells (Sun et al., 2015). Although only recently more appreciated, this toxin serves as a major mechanism of host cell killing by *M. tuberculosis* that is far more “traditional” than the major mechanisms of immune subversion that will be discussed throughout much of the remainder of this thesis.

juelo et al., 2018). Part of this necrosis is mediated by macrophage responses to TNF- α , which results in cell death through overload of mitochondrial-dependent reactive oxygen species production (Roca et al., 2019; Roca et al., 2022). However, not all macrophages succumb to death by this mechanism and many will undergo apoptosis, a protective form of cell death associated with enhanced bacterial clearance and nitric oxide production (Herbst, Schaible, and Schneider, 2011; Divangahi, Behar, and Remold, 2013). Trapping the bacteria within apoptotic bodies allows other macrophages and neutrophils to undergo unimpeded phagosomal-lysosomal fusion to destroy the contents (Molloy, Laochumroonvorapong, and Kaplan, 1994; Behar et al., 2011; Dallenga et al., 2017; Mahamed et al., 2017).

Additionally, host resistance genes may be an alternative direction for these sorts of studies – alteration in the function of genes that decrease likelihood of infection or increase clearance may be more difficult to isolate from human population studies, but may be addressable with enough statistical power or with *in vitro* approaches (Bourgeois, Smith, and Ko, 2021). This model has been proposed as a means of heterozygous advantage⁵² for diseases like Tay Sachs without rigorous experimental validation (Spyropoulos et al., 1981). Another disorder, Gaucher disease, has additional recent data supporting a heterozygous advantage against tuberculosis, although further data is required to dissect the impact of this on human populations over time (Fan et al., 2022). One notable finding from Das et al. (2013) is that human genotype at the *MIF* locus is a strong determinant of susceptibility to infection, with high-expressing genotypes conferring protection through the upregulation of DECTIN-1 expression on macrophages⁵³. Another example explored the contri-

⁵²Such analogies rely on the widely known model of heterozygous advantage against malaria infection among sickle cell trait carriers. It is tempting to think that such a phenomenon could exist for tuberculosis, but it is unclear whether it does.

⁵³As will be discussed more later on, DECTIN-1 has known roles in the immune response to

butions of mycobacteria-induced cell death pathways on disease outcomes and found that, while mycobacteria secrete a potent mitotoxin (ESAT6) able to induce necrotic cell death in infected macrophages, the macrophages can utilize mTOR and oxidative phosphorylation as a mechanism to limit necrotic cell death through mitochondrial damage and resist infection via increased macrophage longevity. This demonstrates yet another role for macrophages in host defense as in the absence of mTOR mycobacteria grow unrestricted (Pagan et al., 2022). This also add nuance to the previously discussed results from Roca et al. (2019) as, while mycobacteria are able to subvert critical cellular pathways to induce necrosis, this is able to be somewhat mitigated by the macrophages themselves in a potent defense mechanism. More subtle gene-gene interactions or gene-environment interactions might underly relative resistance to tuberculosis infection, which can be epidemiologically observed among many people who are chronically exposed but never tuberculin skin test convert and among people who become latently infected but never develop active disease (Flynn, Chan, and Lin, 2011; Orme, Robinson, and Cooper, 2015).

1.2.6 The Granuloma

After many rounds of bacterial replication and reinfection of macrophages, the nodus of infection begins to coalesce into a structure known as a granuloma, the true epicenter of tuberculosis infection. These granulomas are sophisticated immunological foci that contain the entire spectrum of immune cells within this confined structure, all seeking to eradicate the invading mycobacteria but finding themselves incapable of doing so. As alluded to previously (Subsection 1.1.2), these structures are comprised of uniquely activated macrophages that span the traditional M1-M2 axis as

tuberculosis (Rothfuchs et al., 2007; Marakalala, Graham, and Brown, 2010; Wagener et al., 2018), although the overall effect is rather muted and may not be essential for effective mycobacterial control.

well as granulocytes of all varieties, lymphoid cells, and novel cell types including foamy macrophages⁵⁴ and multinucleated giant cells. However, as has been reviewed comprehensively by (Pagan and Ramakrishnan, 2018)⁵⁵, the macrophage is the central driver of the structure and function of the granuloma – they arrive at the site of infection and initiate the cascade of events that ultimately defines the course of tuberculosis disease.

The granuloma has been long proposed to have both host-protective and host-detrimental effects and the precise balance between them likely varies from granuloma to granuloma. However, the invading mycobacteria have extensively evolved to survive within the granuloma and it has thus become a known quantity in their evolution and they respond extensively to the unique physiology of that environment (Ramakrishnan, Federspiel, and Falkow, 2000; Gagneux, 2018); indeed, they appear to home to existing granulomas and integrate into them unscathed when introduced as a superinfection (Cosma, Humbert, and Ramakrishnan, 2004). Pathogenic mycobacteria also secrete effectors into the macrophage that encourage granuloma formation, providing further evidence of the bacterial-beneficial roles of this structure (Volkman et al., 2004; Volkman et al., 2010).

As the nexus of immunological activity in tuberculosis, it is also the source of a dizzying array of cytokines and chemokines that comprise the overall response to infection, many of which are produced by macrophages. Macrophage-derived TNF- α and IFN- γ ⁵⁶ are important for control of infection and, in their absence, the bacteria

⁵⁴As will be more interesting later, it has been demonstrated that NFAT signaling is important for the development of foam cells (Du et al., 2021); whether the macrophage-specific VIVIT approach used later (Chapter 3) might alter foamy macrophage formation is unknown and difficult to test with present tools.

⁵⁵A. J. Pagan and L. Ramakrishnan (2018). “The Formation and Function of Granulomas”. In: *Annu Rev Immunol* 36, pp. 639–665

⁵⁶Although T cells produce most of the total IFN- γ , macrophages remain a potentially physiologi-

grow unrestricted (Flynn et al., 1993; Flynn et al., 1995; Fenton et al., 1997; Algood, Lin, and Flynn, 2005). However, the bacteria also modulate the expression and downstream responses of these cytokines to further their own lifestyle in what has often been referred to as an evolutionary arms race (Ting et al., 1999). Others have roles that are just beginning to be uncovered. All of these signals are largely induced by engagement between bacterial- or host-derived ligands with granuloma macrophages and serve to modulate critical aspects of the granuloma microenvironment. Much like the tumor microenvironment (see Subsection 1.5.1), the granuloma microenvironment is a key aspect of the progression of the disease, but these features are only now beginning to be dissected properly to understand how neighboring stromal cells impact the immune response, in the broadest sense.

Granuloma formation is a process that remains poorly understood. While initial findings in the 1990s found that critical host-protective cytokines were required for granuloma development, these did not necessarily explain the full expanse of the phenotype and were unable to differentiate granuloma formation from gross differences in mycobacterial burden (Flynn et al., 1993; Flynn et al., 1995). TNF- α especially has important roles in inducing macrophage microbicidal activity, so it is difficult to disentangle these roles from any contribution to granuloma formation *per se* (Ramakrishnan, 2013a). More recent efforts have focused on other signaling events that might be more limited in scope but have more granuloma-specific effects. Torraca et al. (2017) found that CXCR4 signaling⁵⁷ is an important aspect of granuloma for-

cally relevant source of this critical cytokine (which is regulated by NFAT) (Darwich et al., 2009; Robinson et al., 2010). As a caveat, no known studies have utilized a *LysM*-Cre; *Ifng*^{f/f} approach to begin to assess the functional contributions of macrophage IFN- γ even in the presence of T cell-derived interferon.

⁵⁷The primary ligand for CXCR4 is CXCL12. Although the primary cellular source of CXCL12 in the granuloma remains poorly defined, a more functional interrogation of this chemokine in the response to infection may reveal interesting new factors that contribute to the formation and maintenance of the granuloma.

mation and did so by sustaining granuloma angiogenesis. In the absence of CXCR4, granulomas developed a lessened degree of vasculature and correspondingly lower bacterial burden, potentially through signaling to vascular pericytes (Pollard, 2009). This left other questions unaddressed, but set the stage for future studies on particular host determinants. Disrupting the structure of the granuloma via inhibition of protective host immunity leads to increased bacterial burden (Flynn et al., 1993; Flynn et al., 1995; Juffermans et al., 2000; McElvania Tekippe et al., 2010), but disruption through inhibition of the epithelialization event itself results in prolonged survival and improved bacterial killing (Cronan et al., 2016). This work disentangles aspects of the unending argument over the host-beneficial and host-detrimental nature of the granuloma, which resolves into what it was always going to be: a structure that under some circumstances and in some regards is host-beneficial but in others prevents the execution of an effective immune response. The underlying distinctions depend on the immune status of the host and, likely, bacterial genetic background and environmental factors, such as smoking and personal history (Glickman and Schluger, 2016).

Heterogeneity

Granulomas, like the cells that comprise them, are incredibly heterogeneous structures, varying between individual granulomas within the same person and from person to person (Matty et al., 2015; Lenaerts, Barry, and Dartois, 2015). This heterogeneity is a reflection of the complex states in which macrophages can exist and their ontologies (Kiss et al., 2018). Within individual humans, the dynamics of individual granulomas showcase the diverse possible outcomes for a given infectious focus, a pattern reflected in some infection models (Lin et al., 2014; Lenaerts, Barry, and Dartois, 2015). The granuloma by the grossest histological observation contains ep-

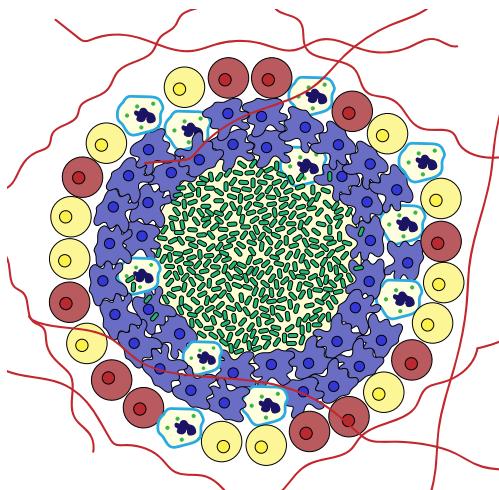


Figure 1.2: Schematic of the basic structure of a granuloma. The innermost layers are comprised of epithelioid macrophages that encase the necrotic core while outer layers are comprised of a mixture of granulocytes and lymphocytes. The whole structure is encased in a web of vasculature.

ithelioid macrophages, motile inflammatory macrophages, multinucleated giant cells, foamy macrophages, and macrophages in various stages of various types of cell death in addition to all the other immune cell populations in the granuloma. However, even among neighboring granulomas, these relative proportions can vary widely and one can achieve effective sterilization while the other cavitates.

This diversity within individuals in rate of progression and outcome provides a full spectrum of possibilities for each granuloma that has largely escaped our language to describe them except in sweeping intervals. Such language, including “centroacinar⁵⁸, perifocal, tree-in-bud⁵⁹, metabolically active, exsudative⁶⁰, suppurative⁶¹,

⁵⁸Within the alveolar space.

⁵⁹Bronchiolar spread leading to fluid filling the cavity, which radiologically resembles a budding tree branch (Gosset, Bankier, and Eisenberg, 2009).

⁶⁰filled with circulatory fluid, but not necessarily red blood cells.

⁶¹purulent, pus-filled.

miliary⁶², caseous, circumscribed, and fibro-calcified⁶³” is but a sampling of the possible terms that pathologists can use to describe these lesions (Ehlers and Schaible, 2012). Such striking variation can be readily seen in the images that accompany Subsection 3.3.6 and Figure 3.22, where the size and shape of these structures can vary widely even within an individual.

One of the many sources of this heterogeneity lies not in the genetics but in the life history of the person. Alveolar macrophages are long-lived cells that must handle a vast diversity of insults on a daily basis and it has been shown that these insults alter their biology in a permanent fashion and likely are propagated to progeny of these mitotic cells (Woodruff et al., 2005; Hodge et al., 2007; Berg et al., 2016; Glickman and Schluger, 2016; A-Gonzalez et al., 2017; Duan et al., 2017). Behaviors such as smoking are clearly inhibitory of alveolar macrophage responses and are known to exacerbate tuberculosis (Hodge et al., 1996; Woodruff et al., 2005; Glickman and Schluger, 2016). What is not as well known is how these environmental factors alter the structure of the granuloma over the long term, especially once the granuloma has established in the interstitium and has a larger proportion of the cell mass comprised of recruited macrophages.

Even macrophages within otherwise unperturbed organs can display remarkable morphological and expressive heterogeneity (Gordon and Taylor, 2005; Gordon and Pluddemann, 2017). This intrinsic variation from macrophage to macrophage would suggest that the outcome of tuberculosis exposure *could* plausibly be determined based on which individual in a population of macrophages is first infected (Verrall et al., 2014). While this would be challenging or impossible to test experimentally, hopefully some future approach will allow this level of micro-scale dissection of indi-

⁶²Literally, millet seed-like lesions along the lung.

⁶³Encased in fibrotic tissue that has undergone calcification and partial ossification.

vidual macrophage heterogeneity in their interactions with *M. tuberculosis* *in vivo* or *ex vivo*. For instance, flow-cell based approaches may allow for single macrophage-*M. tuberculosis* interactions to be modeled and subsequent single cell ATAC-seq or RNA-seq may identify subtle differences between macrophages that predispose them to infection or resistance within a single host. Studies from the zebrafish had displayed such within cell-type heterogeneity clearly – regardless of what the initial phagocytic cell is, the bacteria then subvert host responses to drive the recruitment of more permissive monocytes and macrophages, demonstrating variations in the resting capacity of these phagocytes for bacterial killing (Cambier et al., 2014). This manipulation allows the bacteria to “select” their future host cells by specifically recruiting those least likely to have bactericidal capacity. This baseline heterogeneity within the host presents an exploitation schema for the bacteria that may begin at the very first encounter (Ramakrishnan, 2013b).

The initial events of infection remain difficult to study in most models, but insights from the zebrafish have allowed direct observation of the heterogeneous outcomes of these early events and, more importantly, potential sources of this heterogeneity which can be genetic, environmental (including the microbiome), or without clearly identifiable cause. The experimenter is burdened by entropy – there is a great deal of intractable noise in every system and one of these sources of noise is the microbiota of the host. While no comprehensive study has yet been done on particular commensal species⁶⁴ and their potentially protective or deleterious roles during tuberculosis infection, it is known that commensal species can undermine mycobacterial immune subversion by triggering TLR-dependent sensing mechanisms that negate the shield-

⁶⁴A more thorough study of bacteria-bacteria interactions during tuberculosis infection would be an intriguing new dimension to the understanding of infection. As bacteria have evolved sophisticated mechanisms of intra-kingdom competition, it is likely the case that existing colonization with particular strains of bacteria or fungi imparts resistance to tuberculosis but these have not yet been studied in any depth.

ing exercised by the infecting mycobacteria (Ramakrishnan, 2013b; Cambier et al., 2014).

In the granuloma, there exists an essentially random (Orme and Basaraba, 2014) distribution of lymphocytes around the periphery of the structure. While it is clear that the innermost macrophage layers have some reasonable degree of “organization” as a product of their epithelial junctions with one another, other portions of the granuloma can vary substantially (Cronan et al., 2016). Even within the epithelioid layers, the macrophages remain respectably motile and are able to move throughout the granuloma to continuously remodel this structure (Cronan et al., 2018). These features likely contribute to the diverse outcomes seen within individuals in as-yet unappreciated ways.

1.3 Pattern Recognition in Innate Immunity

For over 100 years, the cells of the innate immune system were thought to be essentially blind scavengers that existed solely to pick up debris for presentation to and activation of adaptive immune responses (Iwasaki and Medzhitov, 2010). How could cells saddled strictly with their somatic genotype go about “intelligently” identifying pathogens? The first clues came from gene homology between the cloned human IL-1 receptor and a developmental protein from *Drosophila*, Toll (Lehmann and Nusslein-Volhard, 1986; Heguy et al., 1992). These similarities between IL-1R and Toll first informed the mechanism by which IL-1 α / β act on cells and then led to the identification of a range of proteins that shared these homologous domains required to activate NF- κ B (Rock et al., 1998; O’Neill, Golenbock, and Bowie, 2013). These proteins, dubbed the Toll-like receptors (TLRs) proved to be the foundation of the modern understanding of a very sophisticated innate immune system that, in the vast

majority of cases, is solely responsible for the prevention of disease (Janeway, 2005). Additionally, these discoveries opened doors into the study of evolutionarily conserved mechanisms of pathogen defense, as TLRs are conserved across both *Animalia* and *Plantae*⁶⁵ (Staskawicz et al., 1995; Armant and Fenton, 2002; Ausubel, 2005). Since that time, numerous other families of receptors, known as pattern recognition receptors (PRRs), have been identified that are able to detect and respond to various pathogen-derived ligands, commonly referred to pathogen⁶⁶-associated molecular patterns. These PAMPs serve as the foundation of the innate immune system and give it an intrinsic ability to detect and respond to threats, both from pathogens and from internal signals indicative of inflammation or damage. To varying degrees, all of these pathways have been implicated in the host response to tuberculosis, although TLR- and CLR-mediated responses stand out as the most well-studied in this regard and seemingly the most prominent during infection (Stamm, Collins, and Shiloh, 2015).

1.3.1 Major Families of Pattern Recognition Receptors

Decades of research have revealed a number of families of pattern recognition receptors that are responsible for detecting diverse sets of widely conserved ligands from bacteria, viruses, fungi, and parasites. All of these have members that can respond, in some degree, to mycobacterial infection, making all of them relevant to the study of tuberculosis (Ishikawa, Mori, and Yamasaki, 2017). As alluded to previously, the Toll-like receptor family was the first to be identified. There are 13 known

⁶⁵This is mostly true – plants have structurally similar proteins that recognize similar ligands, but this is thought to have arisen by convergent evolution rather than a single shared origin (Johnson et al., 2003b).

⁶⁶These are occasionally referred to as microbe-associated molecular patterns; this nomenclature successfully captures the reality that these ligands are not restricted to pathogenic species, but has not become widely used.

TLRs in the human genome, each of which detects a number of ligands ranging from lipopolysaccharide from Gram-negative bacteria (TLR4) and flagellin⁶⁷ (TLR5) to peptidoglycan⁶⁸ (TLR2) and RNA (TLR7) (Kawai and Akira, 2007). In addition to the TLR family, there are nucleotide oligomerization domain (NOD)-like receptors (NLRs), retinoic acid-inducible gene I (RIG-I)-like receptors (RLRs), absent in melanoma 2-like receptors (ALRs), and C-type lectin receptors (CLRs). All of these families can activate NF-κB signaling, highlighting the central importance of this pathway in activating immunoresponsive genetic programs and executing host defense against pathogens. However, most of these receptors also engage more specific immune pathways downstream that provide them with unique responses tailored to the classes of pathogens to which they are responding.

TLR activation has long been the model for understanding the host innate immune response to pathogens. TLR activation induces a somewhat monotone response dominated by NF-κB (Arbibe et al., 2000; Kawai and Akira, 2007; Kawai and Akira, 2009; Brandt et al., 2013). Most of the TLRs utilize a downstream adaptor protein, MYD88, while others (TLR3 and some TLR4 responses) utilize a different adaptor, TRIF (Yamamoto et al., 2003), to transduce signals which ultimately lead to the phosphorylation of inhibitor of nuclear factor kappa B (I-κB) by the IκB kinase (IKK), which releases NF-κB for translocation into the nucleus (Wright et al., 1990; Kawai et al., 1999; Triantafilou and Triantafilou, 2002; Yamamoto et al., 2003; Akira and Takeda, 2004; Takeda and Akira, 2004; Kawasaki and Kawai, 2014). This transcription factor family is often viewed in a somewhat monolithic manner, but is comprised of five independent transcription factors (NF-κB1, NF-κB2, RelA, RelB,

⁶⁷Flagellin is the monomeric subunit used to generate bacterial flagella in motile strains that use flagella for movement.

⁶⁸Peptidoglycan is a major component of the bacterial cell wall and is present in essentially all bacteria.

and c-Rel) that generally act as heterodimers with one another; this raises the potential for subtle and as yet undescribed levels of regulatory complexity⁶⁹ based on the active heterodimers in different contexts (Rice, MacKichan, and Israel, 1992; Baeuerle and Henkel, 1994; Finco and Baldwin, 1995; Oeckinghaus and Ghosh, 2009; Ghosh et al., 2012; Liu et al., 2017; Albensi, 2019). To date, little has been done to fully characterize these distinctions, although some work has found different transcription factor bindings sites to be preferentially bound by some dimers and not others (Siggers et al., 2011; Ramsey et al., 2019; Florio et al., 2022). Nonetheless, NF-κB activation writ large is able to activate transcription of a full spectrum of cytokines and chemokines, including IL-1 β , IL-6, IL-12, IL-17, IFN- α/β , TNF- α , and IFN- γ (Pahl, 1999; Gilmore, 2006; Liu et al., 2017). This robust pro-inflammatory response is essential for effective clearance of many pathogens and also increases the resistance of surrounding tissue to further infection by intracellular pathogens. As the TLRs can recognize such a diverse array of ligands, this makes them a critical inducer of potent host-protective responses through the induction of microbicidal pathways and recruitment of additional immune cells (Kawai and Akira, 2007; Kawasaki and Kawai, 2014).

NLRs are conserved across metazoans and detect cytosolic PAMPs to defend against intracellular pathogens (Martinon and Tschopp, 2005; Creagh and O'Neill, 2006; Clarke, 2014; Motta et al., 2015). NLRs, like TLRs, activate NF-κB, but can also activate mitogen-activated protein kinase (MAPK) signaling, including p38 and JNK (Shaw et al., 2008; Franchi et al., 2009; Zhong, Kinio, and Saleh, 2013; Saxena and Yeretssian, 2014; Platnich and Muruve, 2019; Velloso et al., 2019). NLRs detect a range of primarily bacterial ligands, including gamma-D-glutamyl-meso-di-

⁶⁹Principally, there are 20 different permutations of these proteins just on their own and not accounting for the complex additional layers of possible regulation with additional binding partners.

aminopimelic acid (iE-DAP) from Gram-negative bacteria, muramyl dipeptide from pan-bacterial peptidoglycan, *Legionella* flagellin, and bacterial RNA (Franchi et al., 2009; Zhong, Kinio, and Saleh, 2013; Saxena and Yeretssian, 2014). NLRs initiate the formation of inflammasomes: large, cytosolic, punctate structures that process pro-IL-1 β and pro-IL-18 for secretion and paracrine signaling. Within the responding cell itself, however, the combination of MAPK and NF- κ B signaling provides a different signaling tone to that induced by TLRs. MAPKs are known to induce the expression of IL-1 α/β , IL-10, IL-12, and TNF- α (Dong, Davis, and Flavell, 2002; Arthur and Ley, 2013; Soares-Silva et al., 2016), which are potent immunomodulators, with TNF- α inducing cell death and inflammation while IL-10 is a potent anti-inflammatory chemokine (Couper, Blount, and Riley, 2008; Ouyang et al., 2011; Soares-Silva et al., 2016). Work remains to be done to more thoroughly define the specific MAPK-dependent responses downstream of NLR activation, which may inform more targeted approaches to immunotherapy in the context of infection.

RLRs are an additional class of cytosolic sensors, most commonly implicated in the antiviral immune response as they detect aberrant nucleic acids, usually those with abnormal features that would not typically be found in eukaryotic cells, including double-stranded RNA and uncapped single-stranded RNA of varying lengths (Kawai and Akira, 2010; Loo and Gale, 2011; Rehwinkel and Gack, 2020; Jia, Fu, and Tang, 2021). The RLR family is composed of RIG-I itself as well as MDA5; these have distinct affinity for different ligands, but both ultimately activate IRF3/7 to induce transcription of IFN- α/β , classic type I interferons that activate autocrine and paracrine JAK/STAT signaling to promote antiviral responses in neighboring tissues (Loo and Gale, 2011). These responses are notable in the history of hepatitis C treatment, as the previous standard of care treatment for hepatitis C was treatment with IFN- α , which facilitated viral clearance in many cases (Rong and

Perelson, 2010). Interestingly, RIG-I signaling is also able to mediate some immune responses against *M. tuberculosis* and drives the production of IFN- β through MAVS to exacerbate pathology; *Mavs* mutant mice had dramatically improved survival after challenge (Cheng and Schorey, 2018). The precise ligands for MAVS produced by *M. tuberculosis* remain unknown, but may involve outer membrane vesicles, which can carry RNA cargo (Prados-Rosales et al., 2011; Brown et al., 2015; Tsatsaronis et al., 2018; Dauros-Singorenko et al., 2018; Gupta and Rodriguez, 2018; Chiplunkar et al., 2019).

ALRs are the newest addition to the class of PRRs and serve to detect cytosolic DNA – a potent signal of some sort of pathogenic invasion, either by DNA viruses or some bacteria, or cellular damage. These direct sensors of cytosolic DNA appear to be largely redundant with the existing cGAS pathway as both of these pathways activate STING, which induces the interferon response cascade (Gray et al., 2016). However, an alternative hypothesis suggested that ALRs *antagonize* the interferon response by inhibiting cGAS-mediated activation of STING (Nakaya et al., 2017). These genes are important for promoting the pathogenesis of tuberculosis, as in the absence of IFI16 and the effector IRF3, *M. tuberculosis* is attenuated for growth in a mouse model (Manzanillo et al., 2012). It is hypothesized, although seemingly untested, that each of the 13 ALR genes (in mice) have distinct families of targets and exerts these modulating effects in distinct contexts in cooperation and antagonism with STING (Nakaya et al., 2017). As this pathway is relatively new to study, there is still a great deal to be learned about how it mediates type I interferon responses and how those interact with engagement of other PRR signaling pathways. However, early studies have already begun to identify consequences of misregulation of these genes, including in the development of autoimmunity (Caneparo et al., 2018).

1.3.2 LPS as a Model Molecular Pattern

Lipopolysaccharide (LPS) from Gram-negative bacteria has long been the model for understanding ligand-receptor interactions and signal transduction in the context of PAMP-PRR pairs. On the cell surface, LPS is detected by a complex of CD14, MD2, and TLR4, making it a rather unusual mechanism of detection in comparison to other TLRs, which can either independently detect ligands or act in heterodimers with other members of the TLR family. This complex coordinates the activation of MYD88 and TRIF, which activate NF- κ B signaling as previously described (Beutler, 2000; Miller, Ernst, and Bader, 2005). For intracellular LPS, which might be encountered in the context of a cytosolic bacterial infection, detection by Caspase-11 mediates the formation of inflammasomes to process IL-1 β and IL-18 to defend against these intracellular Gram-negative bacterial pathogens (Wright et al., 1990; Kawai et al., 1999; Triantafilou and Triantafilou, 2002; Kayagaki et al., 2013; Shi et al., 2014; Rathinam, Zhao, and Shao, 2019; Vasudevan et al., 2022). LPS-TLR4 interactions mediate key aspects of sepsis – a systemic inflammatory disorder set on by infection – as well as normal immune responses (Freudenberg et al., 2008).

TLR4 was the first of the TLR family to be characterized and remains the most thoroughly studied (Lu, Yeh, and Ohashi, 2008). The ostensible ligand (albeit indirectly via CD14) for TLR4 was identified as LPS, a highly conserved pattern on the outer leaflet of the outer membrane of Gram-negative bacteria (Wright et al., 1990; Guha and Mackman, 2001; Kayagaki et al., 2013; Shi et al., 2014). LPS, while once again often stated as a monolithic entity, is in fact a whole family of diverse lipoglycans that vary widely in saccharide antigen and lipid composition, which has become an active area of study (Raetz et al., 2007; Maldonado, Sa-Correia, and Vallvano, 2016; Steimle, Autenrieth, and Frick, 2016; SenGupta et al., 2016; Kutsch et

al., 2020; Dickinson et al., 2022). LPS, as the name suggests, is comprised of a lipid moiety that docks into the outer leaflet of the outer membrane of Gram negative bacteria and is connected to a series of well-defined and highly diverse glycans in different physical locations along the molecule; the outermost of these is the O-antigen, which in some species can antagonize adaptive immune responses (Dominguez-Medina et al., 2020). In addition to the highly variable O-antigen, other regions within the molecule can vary within a single bacterium, over time, and across species.

The precise composition of the lipid tails of LPS alters its ability to bind TLR4 and induce inflammatory responses. Pathogenic species of Gram-negative bacteria tend to have six (6) lipid tails on LPS that activate TLR4 while commensal or environmental species have five (5) or fewer lipid tails that do not activate TLR4⁷⁰ (Dixon and Darveau, 2005; Ueda et al., 2010; Lebeer, Vanderleyden, and De Keersmaecker, 2010; Gyorfy, Duda, and Vizler, 2013; Tan et al., 2015; Steimle, Autenrieth, and Frick, 2016; Simpson and Trent, 2019). Precisely why and how these differences have emerged and evolutionary rationales for the failure of pathogenic species to adopt immune evading tetra- or penta-acyl LPS and instead adopt hepta-acyl LPS during infection (SenGupta et al., 2016) is the subject of ongoing work, but it seems undoubted that some aspects of the TLR-dependent response pathway must offer benefit to the bacteria and provide an avenue of bacterial subversion against the host immune response, although this remains understudied in non-tuberculous bacterial infections (Park et al., 2013; McGuire and Arthur, 2015).

The ability for variations of a single structure to modulate such diverse host immune responses establishes a model for pathogens to intricately manipulate the fea-

⁷⁰For instance, the oral opportunistic pathogen *Porphyromonas gingivalis* produces a tetraacylated LPS that actually inhibits TLR4 activation by hexaacylated LPS from *Escherichia coli* (Darveau et al., 2004; Zhang et al., 2008b; Herath et al., 2013).

tures of these widely shared and physiologically essential patterns to alter immune response capacity and, potentially, drive bacterially-beneficial responses through perversion of the “intended” role of these host receptors. The molecule of interest for my purposes, TDM, is comparatively less studied in the context of host immunity, driving some uncertainty around the direct mediators of this response. Additionally, the specific presentation and conformation of TDM seems to impact the immune response generated to this ligand. *In vitro*, TDM has been demonstrated to adopt different conformational states based on surface composition (for instance, at the air-water interface) and geometry, although the *in vivo* relevance of these different states is contentious (Behling et al., 1993; Hunter et al., 2006). While this will be detailed in Subsection 1.3.5, the major receptors known to detect TDM are within the C-type lectin receptor family and are known as MINCLE and MCL.

1.3.3 Signaling Mechanisms Downstream of C-Type Lectin Receptors

While TLR activation *per se* is a rather monotonous response that is predominantly driven by NF- κ B, CLRs terminate in at least two known downstream signaling pathways. In addition to NF- κ B (via a pathway to be discussed shortly), they are capable of activating the nuclear factor of activated T cells (NF-AT or NFAT) pathway (Goodridge, Simmons, and Underhill, 2007; Dambuza and Brown, 2015). This ability to activate multiple layers of transcriptional regulation either at the same time or under different contexts (length of time, strength of agonism, particular ligand) offers CLRs a powerful additional mechanism of modulating the tone of the immunological response in response to particular insults (Brown, Willment, and Whitehead, 2018).

CLRs are a diverse class of pattern recognition receptors that are defined by their

use of divalent calcium (Ca^{2+}) to coordinate the binding of carbohydrate patterns (Hosoi, Imai, and Irimura, 1998; Dodd and Drickamer, 2001), generally segregated into two major classes: QPD (glutamine-proline-aspartate) motif lectins, which bind galactose-containing sugars, and EPN (glutamate-proline-asparagine) motif lectins, which bind mannose- or glucose-containing sugars (Holtet et al., 1997; Zelensky and Gready, 2005; Furukawa et al., 2013; Alenton et al., 2017). QPD-containing C-type lectins are, in general soluble or secreted proteins and include the likes of human tetranectin (CLEC3B), an extracellular matrix-interacting protein, and herring antifreeze protein, which mediates the breakdown of ice crystals in the blood of cold-water fish (Ewart and Fletcher, 1993; Nielsen et al., 1997; Ewart et al., 1998; Graversen et al., 1998; Liu et al., 2007).

These receptors are critical mediators of antibacterial, antiviral, and antifungal immunity and have evolved to encompass a vast range of different receptors with unique ligand specificities and expression patterns (Hoving, Wilson, and Brown, 2014; Tang et al., 2018). CLRs are known to respond primarily to these carbohydrate-linked ligands via their lectin domains (Dodd and Drickamer, 2001; McGreal, Miller, and Gordon, 2005). Many biomolecules are sugar-modified, including those from bacteria, fungi, viruses, archaea, and eukaryotes (both self and pathogens) (Rudd et al., 2001; Ohtsubo and Marth, 2006). This allows CLRs to be a major pathway for the response to host-derived damage-associated molecular patterns (DAMPs) as well as PAMPs (Garcia-Vallejo and Kooyk, 2009). By contrast, EPN C-type lectins play a diverse set of roles and many are the classical members of the CLR family, with many being transmembrane receptors (Sancho and Sousa, 2012). Most notable among these CLRs is DECTIN-1, the archetypal member of the family which has long been studied for its roles in antifungal immunity, but has now been discovered to have a diverse set of roles in other conditions, including responses to bacterial pathogens (including

mycobacteria) and in autoimmunity (Brown et al., 2002; Brown et al., 2003; Brown, 2006; Yadav and Schorey, 2006; Schorey and Lawrence, 2008; Reid, Gow, and Brown, 2009; Drummond and Brown, 2011; Wagener et al., 2018; Deerhake et al., 2021).

DECTIN-1 has provided the scientific foundation of much of the knowledge we have about the mechanisms of signaling downstream of CLR activation. DECTIN-1-mediated signaling responses are known to be protective against fungal infections, making this a centerpiece of defense against common fungal pathogens, including *Malassezia* and *Candida*⁷¹ (Shiokawa, Yamasaki, and Saijo, 2017). Although not the focus of the present work, DECTIN-1 also has a number of important roles in antimycobacterial defense, which established the importance for this class of C-type lectin receptors in tuberculosis (Yadav and Schorey, 2006). DECTIN-1 is a single-pass transmembrane receptor that uses a large C-type lectin domain to engage with various ligands, most notably β -glucans, to stimulate responses in myeloid cells (Brown et al., 2007). DECTIN-1 itself possesses an intracellular YxxL/I (hemITAM) motif that is then phosphorylated by an adaptor kinase, spleen tyrosine kinase⁷² (SYK) (Kerrigan and Brown, 2011; Getahun and Cambier, 2015; Bauer and Steinle, 2017). It has been shown that stabilization of SYK facilitates stronger immunological responses and fosters protection against fungal infections; cells actively degrade this kinase to limit inflammation, sometimes to the detriment of effective immune clearance (Sohn, Gu, and Pierce, 2003; Wirnsberger et al., 2016; Zhu et al., 2016). The ITAM motif

⁷¹Interestingly, this role for C-type lectin receptors in defending against fungi is not ubiquitous and it seems that *Cryptococcus* has evolved mechanisms to avoid detection by most or all of those present in the human host (Walsh et al., 2017). These masking mechanisms can be perturbed by changes in the cell wall composition of the fungi, suggesting that this is an adaptive mechanism of protection for the pathogen (Esher et al., 2018).

⁷²SYK itself has many important roles in macrophage and B cell biology, but to spare the distraction, see Mocsai, Ruland, and Tybulewicz (2010) for a nice review of these roles. SYK is critical for phagosomal-lysosomal fusion and other roles within macrophages (Tabata et al., 2020). This also makes SYK a poor genetic target as disruption is embryonic lethal in mice (Yanagi et al., 2001).

is critical for the binding and activation of downstream processes from a range of tyrosine kinase-dependent receptors, including B cell receptor activation (Monroe, 2006; Bauer and Steinle, 2017). This sets off a complex series of signaling events that activate CARD9, ASC, and/or PLC γ 2, eventually resulting in NF- κ B activation in the former two instances and NFAT activation in the latter (Geijtenbeek and Gringhuis, 2009; Drummond and Brown, 2013). For DECTIN-1 specifically notable roles have been defined for both of these branches in this signaling pathway but much less is known about these pathways downstream of other, related receptors.

CARD9

CARD9-mediated signaling has long been considered to be the dominant modality of signaling downstream of C-type lectin receptors and is important for transcriptional responses to and defense against many intracellular pathogens (Hsu et al., 2007; Hara et al., 2007). This pathway was identified as a critical mediator of TLR-independent antifungal immunity through DECTIN-1, establishing the prototype for C-type lectin signaling responses (Gross et al., 2006). Human CARD9 deficiencies have a very specific susceptibility to fungal infections, but interestingly not other types of infections (Drummond and Lionakis, 2016; Drummond, Franco, and Lionakis, 2018).

One of the major focuses has been on the importance of CARD9-BCL10-MALT1 (CBM) signalosomes as a unique consequence of CLR agonism (Marakalala, Graham, and Brown, 2010; Drummond et al., 2011; Drummond and Lionakis, 2016; Marakalala and Ndlovu, 2017; Drummond, Franco, and Lionakis, 2018). Despite utilizing a method of activation that has more in common with B cell receptor activation than TLRs (Monroe, 2006), the functional downstream consequence is the same: nuclear translocation of NF- κ B through IKK and associated induction of im-

mune response genes including IL-2⁷³, IL-10, TNF- α , and more (Sancho and Sousa, 2012). Furthermore, the evidence is extremely strong that CBM-dependent signaling is critical for the response to a variety of fungal pathogens and that these generally type I responses are a potent defense against infection (Willment and Brown, 2008; Hardison and Brown, 2012; Drummond, Franco, and Lionakis, 2018). Despite this, early studies found that unprimed macrophages do not utilize the CBM signalosome and require priming in order to active CARD9-dependent transcriptional responses (Goodridge et al., 2009). These variations in the ability for DECTIN-1 to induce CARD9 activation may be useful in discriminating between different types of downstream responses; that study specifically found that bone-marrow derived macrophages exhibited DECTIN-1 responses that were essentially independent of CARD9 while dendritic cells promoted CARD9-dependent signaling. The responses downstream of C-type lectin activation through CARD9 tend to activate T_H1 and T_H17 responses, which are important for effective fungal control and defense against mycobacterial infections (Zanoni et al., 2005; Zenaro, Donini, and Dusi, 2009; Drummond et al., 2011; Lyadova and Panteleev, 2015). However, newer datasets have provided evidence of a range of genes that depend on CLR activation but are CARD9-independent (Deerhake et al., 2021). Some of these genes are likely to be NFAT-dependent while others may be activated by as-yet unidentified pathway or through more indirect mechanisms.

Specifically in tuberculosis immunity, CARD9 is important for limiting bacterial growth; in its absence, mice develop severe pneumonia and rapidly succumb to infection. Several different C-type lectin receptors can activate CARD9, so mutation

⁷³While all of these genes are confusing from most perspectives, this one is extremely confusing and requires a bit of gymnastics to come to – why would it be assumed that NF- κ B signaling is mediating IL-2 expression in the context of the body of literature stating that NFAT is the almost exclusive regulator of IL-2 with no affirmative evidence of that being the case?

of this adaptor protein compromises signaling through several distinct receptors that recognize a range of ligands (Wagener et al., 2018). Some of these effects may be mediated by loss of DECTIN-1-dependent responses, but much is likely to be mediated by other receptors, as will be discussed in Subsection 1.3.5 (Marakalala, Graham, and Brown, 2010; Marakalala and Ndlovu, 2017). CARD9 is not only essential for transcriptional responses downstream of C-type lectin receptors, it is also required for the transcriptional induction of MINCLE downstream of MCL. This provides a priming step that depends on CARD9 and then feeds-forward with additional CARD9-dependent MINCLE expression for the duration of agonism (Zhao et al., 2014).

This pathway also has known roles in exacerbating autoimmunity. A definitive study on the matter by Deerhake et al. (2021) demonstrated that Card9⁷⁴ activation during experimental autoimmune encephalomyelitis drove pathology despite the overall protective effects of Dectin-1 activity. This study generated a comprehensive dataset of the Card9-dependent signaling events in myeloid cells after exposure to the Dectin-1 agonist curdlan. This will provide a great deal of value to future experimenters attempting to isolate the contributions of distinct signaling pathways to an observed C-type lectin-dependent phenotype.

As will be discussed more exhaustively in Subsubsection 1.5.4, macrophages associated with tumors exert particular levels of regulation over the biology of the tumor itself. In this context, CARD9 is able to be activated by tumors to drive increased tumor metastasis (Yang et al., 2014c). In the context of wound healing, CARD9 is required for effective closure; this offers a different side to CARD9 signaling, which is

⁷⁴Throughout this document, whenever I am synthesizing data from different sources or speaking of a pathway in the abstract, I will typically use human gene nomenclature (Tweedie et al., 2021), however, when discussing particular studies in depth, I will adopt the nomenclature of the model organism used (in this case, the mouse). I hope to minimize any confusion on this topic by using species-specific nomenclature only sparingly and when relevant to the topic at hand.

often viewed as strictly inflammatory, but is evidently important for wound healing processes as well although the mechanisms remain largely unknown (Kanno et al., 2017).

Inflammasomes

CLR activation also results in the activation of ASC-dependent canonical inflammasomes, which process pro-IL-1 β and pro-IL-18 for secretion and paracrine and autocrine signaling (Gross et al., 2011) and play critical roles during both development and pathogen responses (Tyrkalska et al., 2019). This occurs through the formation of an intracellular signaling complex with many distinct subunits that, together, mediate the activation of inflammatory programs and pyroptosis (Pandey et al., 2021). These IL-1 β - and IL-18-dependent responses depend on the previously mentioned IL-1R which utilizes MYD88 to drive signaling, linking it into common sets of signals that are induced by TLR activation. This single pathway thus plays a critical and somewhat circular role in various facets of the host response downstream of TLR activation, which unifies the response tone while potentially restricting response diversity; while TLRs are somewhat broad in their expression pattern, the induction of IL-1 β secretion activates all neighboring cells that express IL-1R, which is practically ubiquitous in environment-facing tissues (Deyerle et al., 1992; Malik and Kanneganti, 2018). This makes this pathway extremely powerful for increasing the local inflammatory tone to block the replication and spread of (especially intracellular) pathogens, but subject to a complex set of subversive mechanisms utilized by bacteria, fungi, and viruses (Poeck and Ruland, 2010; MacMicking, 2012; Tavares, Burgel, and Bocca, 2015; Wein and Sorek, 2022). Comparatively less work has been done to characterize the activating mechanisms induced by CLRs that result in inflammasome formation, but this could serve as an exacerbatory mechanism to heighten

inflammation in response to particular classes of signal. Notably, LPS (the canonical TLR4 ligand) is also capable of activating inflammasomes, albeit by an entirely distinct Caspase-11-dependent mechanism (Hagar et al., 2013; Pilla et al., 2014; Vanaja et al., 2016; Finethy et al., 2020).

Inflammasome activation has many described roles in the control of mycobacterial infections and is actively manipulated by the bacteria, suggesting that C-type lectin mediated activation of this pathway may play a role in host defense against mycobacteria as it does against fungi (Hardison and Brown, 2012; Wassermann et al., 2015). Knockout of the mediator caspases results in increased bacterial burden in a strictly innate immune context (Kenyon et al., 2017). However, it has been demonstrated that *M. tuberculosis* actively blocks activation of the inflammasome, suggesting that this pathway may be of variable efficacy in defense against mycobacteria, or that inflammasome-mediated activation of gasdermin D is a more important mediator of these effects than is pro-IL-1 β processing (Master et al., 2008; Qu et al., 2020). Yet another plausible alternative incorporates non-canonical or inflammasome-independent functions of ASC, as ASC knockout mice demonstrate increased susceptibility to infection while NLRP3 and Caspase-1 are both dispensable for protection (McElvania Tekippe et al., 2010). There are also a number of tools that can be utilized to study this process, including a fluorescent ASC knock-in line that report inflammasome activation through visible formation of the characteristic cytosolic specks (Kuri et al., 2017). Such controversy within the field makes this a prime area for future clarifying study although it may be a technical challenge to disentangle some of these complex factors. Despite known role for CLR signaling in activating the inflammasome, the primary activation mechanism in mycobacterial infection is presumed to be via plasma membrane damage, although this does not rule out contributions from other pathways (Beckwith et al., 2020).

NFAT

Lastly, CLRs can activate NFAT signaling, which is a signaling pathway common across developmental processes but is thought to be a unique consequence of CLR agonism in respect to PRR activation and, as such, is often used as a reporter for CLR activity (Aramburu et al., 1998; Chow, Rincon, and Davis, 1999; Hooijberg et al., 2000; Jauliac et al., 2002; Wilkins et al., 2004; Goodridge, Simmons, and Underhill, 2007; Fuller et al., 2007; Goodridge and Underhill, 2008; Zhao et al., 2014; Hannanta-Anan and Chow, 2016; Bendickova et al., 2020). Despite the relative uniqueness of this pathway to CLRs, comparatively less work has been done to characterize it vis-à-vis the CARD9-NF- κ B pathway (Goodridge, Simmons, and Underhill, 2007; Goodridge et al., 2009; Deerhake et al., 2021). This pathway can induce the expression of EGR2, EGR3, COX2, IL-2, IL-10, and others, which is a more anti-inflammatory or immunoregulatory set of signals than the heavily type I signals induced by NF- κ B (IL-1, IFN- γ , etc.) (Saraiva and O'Garra, 2010). The interplay between these divergent signaling consequences is poorly understood, but is likely important in determining the overall tone and kinetics of the response to infection. This pathway will be given a much more extensive dissection in Section 1.4, but suffice to say that the consequences of this pathway's activity downstream of CLR activation are critical but poorly understood mediators of the overall response in tandem with NF- κ B activation.

Phagocytosis

One of the key integration mechanisms between C-type lectin receptors and NFAT pathway activation is the contribution that C-type lectins make to myeloid cell phagocytosis. These cells are widespread phagocytic receptors and are critical for the up-

take of fungi (Robinson et al., 2006). Could this be the case during mycobacterial infections? Seemingly so. The mannose receptor (CD206) is thought to be a major uptake receptor for mycobacteria, as ectopic expression drives the internalization of mycobacteria in other cell types (Schlesinger, 1993). While this is somewhat redundant with other mechanisms of uptake, this suggests a fundamental role for C-type lectins in driving mycobacterial phagocytosis (Ernst, 1998). One of the central consequences of the phagocytosis of any particle is the activation of the NFAT signaling pathway – tying this pathway intimately to the response to diverse infections and activation of adaptive immunity (Khameneh et al., 2017). Given that NFAT signaling is at the root of these earliest host interactions with mycobacteria, it is bizarre that this pathways has been comparatively neglected for so long. It is my hope that the discussion provided here and the resulted presented later on will spur further investigation into the contributions of calcineurin-NFAT signaling to the earliest events of innate immunity (Jayachandran et al., 2007; Jayachandran et al., 2008).

An excellent study by Goodridge et al. (2011) found that the activation of DECTIN-1 was dependent upon binding immobilized ligand. Soluble β -glucan particles were unable to activate DECTIN-1 while those same particles immobilized on plastic were sufficient to do so, providing a model wherein C-type lectin receptors can discriminate between happenstance binding of soluble ligands and direct microbial recognition to only induce phagocytosis when bound by a sufficiently large particle (Elder et al., 2017). This prevents microbes from shedding small decoy ligands to distract the innate immune system and induce aberrant and energetically costly phagocytosis. This phagocytosis is intimately linked to the activation of NFAT, where internalization of particles through phagocytic receptors (primarily C-type lectins) facilitates increased signaling activity (Fric, Zelante, and Ricciardi-Castagnoli, 2014). Whether this is the case for every conceivable ligand is unknown

and unlikely, but it, at minimum, offers yet another mechanism for host detection and potential microbial subversion.

These phagocytic activities require an intricate (and often physical) association between the inducing receptor with calcium ion channels, calmodulin, calcineurin, and mTOR. It has been found that STIM1 physically links phagosomes to the endoplasmic reticulum to enhance phagocytosis; this release of calcium via STIM1 is then able to activate NFAT to drive further transcriptional responses meant to address the contents of the phagocytic vesicle, to varying degrees of success (Nunes et al., 2012). The contributions of NFAT in controlling intracellular mycobacterial growth is wholly unknown but is presumably subject to extensive modulation by the infecting bacteria. Such limitation of NFAT responses within the macrophage while it is infected may be a compelling explanation for the extracellular exposure-specific responses observed in Subsection 3.3.1.

1.3.4 C-Type Lectins in Tuberculosis

In addition to DECTIN-1, there are a number of other C-type lectin receptors that have come under study for their contributions to host immunity. DECTIN-1 itself is thought to bind to mycobacteria, although the ligand remains unknown and DECTIN-1 is dispensable for effective antimycobacterial responses (Yadav and Schorey, 2006; Rothfuchs et al., 2007; Schorey and Lawrence, 2008; Marakalala et al., 2011). One of these other receptors largely predates acknowledgement of C-type lectins as a major pattern recognition signaling pathway but is critical for bacterial uptake. As previously mentioned, the mannose receptor is a major contributor to mycobacterial phagocytosis in a manner partially redundant with complement opsonization and uptake (Schlesinger, 1993; Kang and Schlesinger, 1998). However,

this mode of uptake does not induce lysosomal fusion and is thus a pro-bacterial mode of cell entry, allowing access to permissive host environments and evading bactericidal immune cells (Astarie-Dequeker et al., 1999; Goyal, Klassert, and Slevogt, 2016; Rajaram et al., 2017).

DC-SIGN was the first C-type lectin receptor to have described roles in host protection against *M. tuberculosis*. It was initially described as the major receptor for mycobacteria on the surface of dendritic cells, although it is now known to also be expressed by macrophages, especially alveolar macrophages (Geijtenbeek, Engering, and Van Kooyk, 2002; Tailleux et al., 2003; Tailleux et al., 2005). The general consensus was that DC-SIGN expression was a mechanism for host subversion by mycobacteria, but that does not adequately explain the function of DC-SIGN *in primo loco* (van Kooyk and Geijtenbeek, 2003), especially as the mycobacteria specifically target DC-SIGN to inhibit host immunity (Geijtenbeek et al., 2003). In this context, binding of *M. tuberculosis* mannose-capped lipoarabinomannan (ManLAM) (Maeda et al., 2003; Koppel et al., 2004; Pitarque et al., 2005) to DC-SIGN induced IL-6, IL-10 and IL-12 expression by dendritic cells; similar effects were seen with binding of HIV-1, *Helicobacter pylori*, and *Aspergillus*, making DC-SIGN a rather promiscuous receptor of pan-kingdom ligands ranging from bacteria to fungi to viruses (Serrano-Gomez, Leal, and Corbi, 2005; Gringhuis et al., 2009; den Dunnen, Gringhuis, and Geijtenbeek, 2009). Interestingly, this gene is all but impossible to study in the murine model because mice have no fewer than eight DC-SIGN duplicates, evidence of the evolutionary pressures likely to have necessitated such diversification of function, which may be common to C-type lectin receptors more generally (Tanne et al., 2009; Garcia-Vallejo and Kooyk, 2013).

There are other C-type lectin receptors that engage distinct aspects of host im-

munity to tuberculosis (Mishra et al., 2017a). DECTIN-2 has been shown to detect mannose-capped lipoarabinomannans and induce the expression of a by now familiar cast of cytokines – IL-2, IL-6, IL-10, and TNF- α . This ligand is another component of the mycobacterial cell wall and is likely to be exposed after bacterial killing or other loss of cell wall integrity, so is unlikely to be a phagocytic receptor for mycobacteria, but may be an important mediator of bystander cell activation (Marakalala and Ndlovu, 2017). All of these receptors have major downstream pathways that run through CARD9-NF- κ B, which has been shown to be important for mycobacterial control and limitation of inflammatory responses through IL-10 (Dorhoi et al., 2010).

Two additional members of the EPN-containing superfamily of CLRs are MCL and MINCLE⁷⁵. MCL, originally dubbed DECTIN-3⁷⁶, is expressed by myeloid cells at baseline and is a comparatively desensitized receptor with low affinity for its primary known ligand, TDM (Zhu et al., 2013; Miyake et al., 2013; Zhao et al., 2014). MINCLE, on the other hand, is tightly regulated and only induced after cellular priming by some other stimulus, including MCL activation (Wells et al., 2008; Patin, Orr, and Schaible, 2017). MINCLE has much higher affinity for TDM and, seemingly, a broader range of agonizing ligands, although the latter discrepancy may be a result of historical scientific focus rather than authentic biological difference (Richardson and Williams, 2014; Feinberg et al., 2016; Hansen et al., 2019).

⁷⁵Interestingly, engagement of MINCLE by fungi suppresses the protective DECTIN-1-mediated responses. How these receptors can be manipulated by pathogens to be either active or repressive is an interesting biochemical and biophysical question that has yet to be fully addressed (Wevers et al., 2014).

⁷⁶And for historical reasons, is still occasionally called this in the modern literature.

1.3.5 The TDM Receptor

TDM exerts similarly diverse functions as LPS and is also detected by host pattern recognitions receptors (PRRs) including TLR2 – another member of the Toll-like receptor family – and two C-type lectin receptors (CLRs), MINCLE and MCL. As discussed in Subsection 1.1.4 and Subsection 1.1.5, TDM is a structurally essential component of mycobacteria; the absence of TDM renders the bacteria susceptible to killing by immunological, chemical, and environmental stressors (Rao et al., 2005; Kan-Sutton, Jagannath, and Hunter, 2009; Moliva et al., 2019). In addition to the important structural aspects of TDM, it also possesses a number of chemical and biological functions in interactions between pathogenic mycobacteria and their hosts.

Chemically, TDM is radically different from nearly any other biomolecule that an organism is likely to encounter. Comprised of a trehalose head group – an unusual di-glucose that is never synthesized by animals (Elbein et al., 2003) – attached to profoundly hydrophobic, extremely long, and diversely modified branched fatty acid tails, TDM is directly cytotoxic to cells through disruption of plasma membrane integrity (Figure 1.1) (Retzinger et al., 1982; Sakamoto et al., 2010). For over 50 years after its initial characterization as an important structural feature of mycobacteria, the host receptor, if any, remained unknown. It was only in 2009 that a trio of publications began to define two distinct signaling pathway families that could be activated by TDM – a TLR2-dependent pathway and a Macrophage inducible C-type lectin (MINCLE)/Fc γ R-dependent pathway (Werninghaus et al., 2009; Ishikawa et al., 2009; Bowdish et al., 2009). The TLR2/MARCO/CD14⁷⁷-dependent response

⁷⁷Interestingly, CD14 has a variety of MYD88-independent roles and can induce NFAT activation (see Subsection 1.4.3) which may be one mechanic by which these conflicting pathways can be resolved (Jiang et al., 2005; Nakata et al., 2006). Alternately, the pathways could play complimentary or antagonistic roles in particular contexts over the course of infection; all of these seem like reasonable hypotheses, but research on the TLR2-pathway has largely stagnated in fa-

axis was first described by David Russell's group in 2009 and established an important role for this complex in regulating part of the expression of TNF α , IL-6, and IL-1 β (Manukyan et al., 2005; Bowdish et al., 2009). While this was a very thorough and comprehensive project that elucidated many cell biological aspects of macrophage exposure to TDM, it failed to account for the complete response to TDM and left open the possibility that other pathways may play critical roles in mediating this response.

MYD88

MYD88 is a critically important adaptor protein downstream of most TLR-dependent responses⁷⁸. In addition, MYD88 is essential for transcriptional responses downstream of IL-1R activation, making it essential for both the generation and transduction of important pro-inflammatory signals. MYD88 executes these functions by forming what is known as the myddosome, which is a complex oligomeric structure comprised of MYD88, which directly binds TLRs, and IRAK4 (Latty et al., 2018). This complex, through a series of intermediate steps, activates IKK to induce NF- κ B. This is then essential for transcription of many pro-inflammatory cytokines including TNF- α , IL-1 β , and IL-6. These responses aid in the eradication of various pathogens and prime an effective adaptive immune response (Balka and De Nardo, 2019), although it is functionally dispensable for adaptive immunity to tuberculosis despite a failure to restrict bacterial growth through innate mechanisms (Shi et al., 2003; Scanga et al., 2004; Berod et al., 2014).

As would be expected, MYD88 is critically important for restricting mycobacte-

vor of the MCL/MINCLE pathway. Additionally, these MYD88-independent signaling functions predominate in previously unstimulated monocytes and macrophages (Bjorkbacka et al., 2004)

⁷⁸Aside from TLR3 and some signals from TLR4, MYD88 is essential for TLR-mediated responses (Takeda and Akira, 2004; Kawasaki and Kawai, 2014).

rial growth during infection, although all the receptor-mediated contributions to this control have not been fully addressed (Underhill et al., 1999; Shi et al., 2003; Sugawara et al., 2003; Scanga et al., 2004; Holscher et al., 2008; Berod et al., 2014; Cervantes, 2017; Hosseini et al., 2021). Much of this response may be mediated by IL-1R-dependent responses rather than TLRs, suggesting a relatively minor role for TLRs in host defense to mycobacteria (Fremond et al., 2007; Holscher et al., 2008; Cambier et al., 2014). Adaptive responses are able to develop even in the absence of MYD88, implicating innate response failure in the overall increase in mortality (Fremond et al., 2004). Despite these important roles in host defense, it was previously unknown whether MYD88 played a role in important granuloma processes, such as angiogenesis. MYD88 has known roles in contributing to angiogenesis in tumors and wound healing (Macedo et al., 2007; Zhang et al., 2020a), but our work has found that it is dispensable for angiogenesis in response to TDM, suggesting the existence of an alternative, TLR- and IL-1R-independent signaling pathway for inducing VEGFA expression (see Figure 3.6) (Walton et al., 2018).

MINCLE/MCL

Preliminary results had indicated that a Fc γ R-SYK-CARD9 signaling axis was important for innate immune activation by TDM (Werninghaus et al., 2009). Knockout of *Card9* resulted in nearly abolished expression of a far more expansive range of cytokines and chemokines and led to the hypothesis that some co-receptor was actually responsible for directly binding TDM and that this co-receptor likely lacked its own ITAM motif, as it required Fc γ R to provide one in *trans*. The intersection of the data on Fc γ R in TDM detection (Werninghaus et al., 2009) and MINCLE's known dependence on Fc γ R to provide an ITAM (YxxL/I_{x(6-8)}YxxL/I) motif in *trans* for signaling (Yamasaki et al., 2008) led to a logical hypothesis: that perhaps MINCLE

was the sought-after TDM receptor.

MINCLE, itself, was viewed with some interest due to its curious regulatory pattern after having been identified as a macrophage-specific gene induced by C-EBP β ⁷⁹, but until 2008, no ligands had been identified (Akira et al., 1990; Balch et al., 1998). The first identified ligand, SAP130, is a nuclear protein that is exposed to the extracellular milieu after necrotic cell death, which is then able to activate macrophages to scavenge cellular debris (Yamasaki et al., 2008). These early observations were, themselves, clues to the pleiotropic nature of MINCLE activation as a strictly inflammatory response to cell death would be inappropriate in tone for the majority of innocuous programmed and incidental cell death events that occur almost constantly in the day-to-day lives of organisms comprised of billions of cells. This assigned an initial role to MINCLE but broke from the standard pattern of CLRs detecting PAMPs; however, many of these receptors had already been identified to bind seemingly unrelated ligands. Ultimately, a seminal paper in 2009 established a direct ligand-receptor binding interaction for TDM and MINCLE, establishing this as the dominant pathway for TDM detection and host signal transduction (Ishikawa et al., 2009).

Following this, it remained somewhat unclear what pathways were capable of inducing *CLEC4E* expression in response to pure TDM. Over evolutionary time, an ancestral CLR had undergone a tandem duplication to form the modern *CLEC4E* (MINCLE) and *CLEC4D* (MCL) genes. Based on conserved sequence and differential expression patterns, MCL became a compelling candidate to be the basal receptor for TDM. Indeed, although it took four years, MCL was identified as a low-affinity

⁷⁹In the original literature, this protein was commonly referred to as the nuclear factor of interleukin-6, or NF-IL6, although this name has since fallen into disuse (Akira et al., 1990; Matsumoto et al., 1999).

TDM binding receptor that could mediate the upregulation of MINCLE⁸⁰ (Miyake et al., 2013). Further studies elaborated the latter pathway to incorporate a biphasic and ultimately heterodimeric model of MCL/MINCLE⁸¹ activation, with MCL being activated first, inducing *MINCLE* expression and then the two acting as a heterodimer over longer courses of signaling (Lobato-Pascual et al., 2013; Yamasaki, 2013; Furukawa et al., 2013; Kerscher et al., 2016b). Additionally, it has been found that this heterodimer is able to mediate phagocytosis, offering yet another potential route of entry into phagocytes for mycobacteria (Yamasaki, 2013; Lobato-Pascual et al., 2013).

While the mechanisms of detecting TDM are, themselves, biochemically interesting as an organism evolves to detect this rather unusual compound, the functional consequences of MINCLE-TDM interactions remained unknown and, to this day, remain conflictual. Lee et al. (2012) found that MINCLE expression on neutrophils was protective against infection with *M. tuberculosis* Erdman while Behler et al. (2012) found that MINCLE expression in alveolar macrophages was important for responses to BCG and in the absence of MINCLE, the mice exhibited increased bacillary load. However, Heitmann et al. (2013) found that *Clec4e*^{-/-} mice had no defect in bacterial control after infection with either *M. tuberculosis* H37Rv or BCG. These results are difficult to reconcile, but each uses different strains, routes of infection, and infectious doses, leading to further difficulty in analysis⁸². A subsequent study found that MIN-

⁸⁰Many inflammatory stimuli are able to induce the expression of MINCLE. It was originally identified as an NF-IL6 target gene, which can also be induced by MYD88 signaling and other danger signals (Kerscher et al., 2016a).

⁸¹In the literature, these protein products are often listed using mouse-specific nomenclature as Mincl and Mcl for the sake of being more word-legible. MINCLE and Mincl are the protein products of the genes CLEC4E and Clec4e; MCL and Mcl are the protein products of CLEC4D) and Clec4d, from humans and mice respectively.

⁸²As suggested by Lang (2013), culture conditions and mouse colony hygiene may also underly these differences. In retrospect it is impossible to know, but this set of papers has stymied future research designed to clarify the importance of this ligand-receptor interaction during infection.

CLE on dendritic cells was also important for control of disseminated tuberculosis of the spleen (Behler et al., 2015) and complementary studies identified a critical role for MCL in more general defense against tuberculosis (Wilson et al., 2015). Human genetic studies have investigated polymorphisms in the human MINCLE gene and have not yet found any association with tuberculosis, although none of the investigated mutations were loss of function (Bowker et al., 2016). By contrast, the inducing receptor MCL seems to have a stronger body of evidence supporting its role in mediating protective responses, with knockout mice having higher bacterial burden and mortality despite having functional MINCLE⁸³ (Wilson et al., 2015). While the picture as a whole comes out somewhat murky, it may be the case, similar to responses downstream of DECTIN-1 in experimental autoimmune encephalomyelitis, that the net effect of MINCLE is protective, but other factors can change the relative benefit by antagonizing other immunological pathways in ways that depend on bacterial or mouse strain, microbiome, or other as-yet unidentified factors.

In addition to binding to mycobacterial TDM, MINCLE has a number of other ligands, including fungal PAMPs, such as those from *Malassezia* and, perhaps, *Candida* (Wells et al., 2008; Yamasaki et al., 2009; Ishikawa et al., 2013). In response to *Malassezia*, MINCLE induces IL-6 and TNF- α , aiding in host defense against this opportunistic fungal pathogen (Lu, Nagata, and Yamasaki, 2018). MINCLE can also bind related mycolic acids from other species, including *Corynebacterium* mycolic acids, which are much shorter than the comparable species in *Mycobacterium* (van der Peet et al., 2015). This suggests that MINCLE is a versatile receptor with a broad set of ligands, the specificity of which is probably determined by higher orders of regulation, including potential heterodimers with other receptors and conforma-

⁸³As will be discussed later, result of full-locus deletion mutants across both MCL and MINCLE would be an interesting addition to our knowledge of the role of TDM detection *per se* to antimycobacterial immunity rather than this gene-centric model.

tional changes (Jegouzo et al., 2014). MINCLE, like similar C-type lectin receptors, is able to induce a variety of downstream signaling cascades, including the one which became the focus of this work, NFAT.

1.4 Nuclear Factor of Activated T Cells (NFAT)

NFAT signaling activation results in diverse cell- and context-dependent outcomes. This gene family was first described as a transcription factor that regulated the production of IL-2 in T cells (Shaw et al., 1988; Jain et al., 1993; Northrop et al., 1994). This pathway could be inhibited by blocking the phosphatase activity of calcineurin, suggesting that NFAT was regulated by calcineurin (Jain et al., 1993; Loh et al., 1996). While it was initially studied for its diverse roles in regulating lymphoid biology, it has since had widely diverse roles ascribed to it in nearly all cell types including cardiomyocytes, endothelial cells, skeletal muscle, pancreatic β islets, oligodendrocytes, keratinocytes, and myeloid cells (Kegley et al., 2001; Stevenson et al., 2001; Horsley and Pavlath, 2002; Crabtree and Olson, 2002; Al-Daraji et al., 2002; Muller and Rao, 2010; Fric et al., 2012a; Weider et al., 2018).

This pathway has had a vast range of roles ascribed in the context of infectious disease (as already briefly discussed in Subsubsection 1.3.3), cancer (Muller and Rao, 2010), autoimmunity (Park et al., 2020), and development (Horsley and Pavlath, 2002; Crabtree and Olson, 2002). In this section, I will review some of these roles and provide a more comprehensive review of the unique roles of the different isoforms as a means of informing the later results particular to NFATC2.

NFAT has many features that make it a transcription factor family of broad basic as well as translational interest. The NFAT family is comprised of five members: NFATC1 (also known as NFAT2), NFATC2 (NFAT1), NFATC3 (NFAT4), NFATC4

(NFAT3), and NFAT5 (Rao, Luo, and Hogan, 1997). Historical reasons have resulted in a convoluted nomenclature⁸⁴, so for the sake of consistency, the NFATCx⁸⁵ naming scheme will be used throughout this document. NFAT5 is a special, and distantly related, member of this family that appears to be important for the transcriptional response to osmotic stress, but unlike all of the other members, is constitutively nuclear and not regulated by changes in cytosolic calcium concentration via calcineurin (Go et al., 2004; Kumar et al., 2020a).

The four calcium responsive members have long been passively assumed to be functionally redundant, with their roles defined by their patterns of tissue expression (Chow, Rincon, and Davis, 1999; Macian, 2005; Klein et al., 2006; Mancini and Toker, 2009). All of them seem to be derived from an ancestral single isoform that was duplicated over the course of evolution (although intermediate representatives with greater than one but fewer than four isoforms seem to be unknown among modern species) (Muller and Rao, 2010). However, evolution has provided each of these isoforms distinctive biophysical properties that allow them to have non-redundant roles even in cell types where more than one is expressed simultaneously. Most notable is their alterations in sensitivity to changes in calcium: while NFATC2 has a persistent response after strong activation near the plasma membrane and endomembranes, NFATC3 rapidly traffics in and out of the nucleus in response to

⁸⁴As often happens in science when multiple independent lab groups discover proteins at the same time, the naming can become a challenge as the field as whole reconciles two or more distinct naming schema. In this case, no resolution has ever come about. The NFATc subnomenclature was meant to designate that they are calcium-responsive and calcineurin-dependent and distinct from NFAT5, the modern homolog of the ancestral protein with high sequence similarity from humans to sponge. In choosing to maintain the NFATc nomenclature, I take no position on the relative merits of the two systems. Additional, now largely outdated, naming schemes had an additional name for each of the isoforms that I will address only as needed throughout this document. Typically, the text will obfuscate the use of these alternative names and translate them into the desired nomenclature.

⁸⁵Classically, these are referred to as "NFATcX" (with a lowercase "c"), but this is inconsistent with human gene nomenclature and has been discarded for this document.

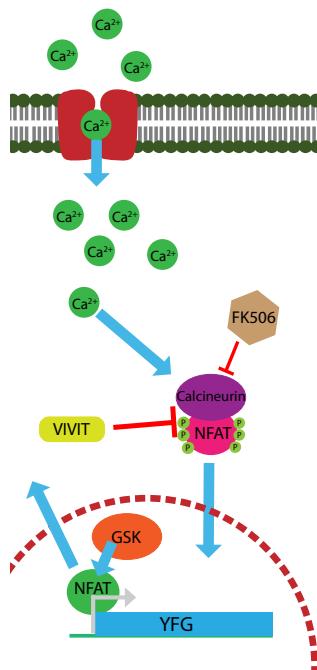


Figure 1.3: This dramatically simplified schematic shows the mechanism of NFAT activation: calcium influx activates calcineurin to dephosphorylate NFAT to allow for nuclear translocation; this process can be reversed through GSK-mediated phosphorylation that exposes a nuclear export sequence. This induction can be inhibited by a number of reagents, including FK506 and VIVIT.

small magnitude changes in calcium and requires high levels of nuclear calcium to maintain activity (Rinne, Banach, and Blatter, 2009; Ulrich et al., 2012; Yissachar et al., 2013; Kar and Parekh, 2015; Kar et al., 2016).

NFAT proteins require the phosphatase calcineurin for their activation. Upon an increase in calcium, calcineurin dephosphorylates NFAT to expose a nuclear localization sequence (NLS); once in the nucleus, kinases (including GSK3 proteins and protein kinase A) are able to phosphorylate NFAT to drive it back into the cytosol in inactive form; the balance of these effects determine the overall transcriptional response (Figure 1.3) (Crabtree and Olson, 2002). This shuttling behavior allows existing pools of NFAT to rapidly modulate host responses, including developmental, immunological, and pathological responses. This also allows for rapid tuning of the

longevity of the response, presumably allowing for the induction of different genes and to different degrees based on the length of activation. Although no work has ever been done to define such distinctions, the principles of biochemical affinity dictate that more accessible chromatin with more NFAT binding sites would be activated prior to those in less accessible configurations or with fewer sites more distal from the transcriptional start site, which may require long periods of strong activation to be induced. Defining these different classes of genes in different cell types would provide a far greater depth of understanding for the consequences of NFAT activation and timing of intervention for maximum medical benefit.

NFAT proteins are by and large structurally similar and are comprised of three broad domains: the N-terminal domain, the DNA binding domain, and the C-terminal domain. The DNA binding domain shares strong homology with other REL-like proteins, placing NFAT on a branch of the NF- κ B evolutionary tree. While the C-terminal domains are relatively understudied, much is known about the N-terminal domain, which contains a nine amino acid transactivation domain (9aaTAD) and relatively unstructured regions important for interacting with a range of binding partners, including AP-1 transcription factors (Boise et al., 1993; Martinez et al., 2015). The N-terminal domain also contains the calcineurin binding motif (SPRIEIT) and the serine-proline domains that determine the phosphorylation status of the protein and exposure of the sequestered NLS for importin trafficking (Rao, Luo, and Hogan, 1997). A simplified diagram of this is shown in Figure 1.4.

The foundational work on NFAT was done almost entirely in T cells, where it was found to be important not only for intra-T cell differentiation into T_H2 cells, but also in dendritic cells for the initial production of IL-2, suggesting that NFAT is an inducer of anti-inflammatory signaling cascades that drive type II responses to facil-



Figure 1.4: This simplified depiction of some of the major domains of interest that are shared among all of the NFAT proteins. The SP domains are dephosphorylated by calcineurin to induce nuclear translocation and rephosphorylated by GSK to induce nuclear export by either revealing or concealing the embedded NLS. The other domains regulate protein-protein interactions and DNA binding.

itate tolerance (Granucci et al., 2001; Granucci et al., 2003; Malek and Bayer, 2004). NFAT is also essential for induction of IL-4/IL-13, the archetypal anti-inflammatory (or inflammation-resolving) cytokines (Agarwal, Avni, and Rao, 2000; Wierenga and Messer, 2000; Monticelli and Rao, 2002; Monticelli, Solymar, and Rao, 2004) which are essential for granuloma formation (Cronan et al., 2021). NFAT activity regulates many aspects of the T cell lifecycle from proliferation to exhaustion, making it central to the lives of these cells (Ranger et al., 1998b; Crabtree and Olson, 2002; Malek and Bayer, 2004; Martinez et al., 2015; Yahia-Cherbal et al., 2019). However, a binary type I/II classification for this pathway is ultimately elusive, as it is also critical for regulating the expression of TNF- α and IFN- γ , critically important pro-inflammatory cytokines and is required for T_H1 specification (Nathan et al., 1983; Sica et al., 1997; Kiani et al., 2001; Porter and Clipstone, 2002; Ke et al., 2006; Kaminuma et al., 2008; Fric et al., 2012b). The pleiotropic nature of this pathway makes it of especial interest in the context of host-pathogen interactions where a robust inflammatory response is needed to kill the invading pathogen, but moderation is required to prevent excessive tissue damage (Casadevall and Pirofski, 2003). These distinct effects of NFAT depend on both the cell type in which it is acting and the particular

isoform activated, as these proteins have diverse but differentiable roles depending on such variables, among others.

1.4.1 Clinical Utility of NFAT Inhibitors

The central role of NFAT in the immune system has long been appreciated, albeit in a rather focused context, via the widespread application of NFAT inhibitory drugs in the clinic. Two drugs are widely used to block calcineurin activation and suppress immune responses: cyclosporine A and tacrolimus (Mihatsch et al., 1998). These drugs were discovered and developed for clinical use to target the T cell response and prevent organ transplant rejection by blocking the affinity maturation and proliferation of anti-graft T cells. The profound and global immune suppression that accompanies the use of these drugs has prevented their use in other contexts for fear of increased susceptibility to infectious diseases (van Sandwijk, Bemelman, and Ten Berge, 2013). The weakness of these drugs is that they block all calcineurin activity in all cell types, leading to a vast range of collateral targets that manifests in renal toxicity (likely through endothelial damage) and neurological side effects – a better approach would be to find a way to locally target only the disease-relevant interacting partner of calcineurin (in this case, NFAT) (Randhawa, Starzl, and Demetris, 1997; Nankivell et al., 2016). Halfway approaches have emerged using tacrolimus (and derivatives) through its use as a topical ointment for atopic dermatitis, but this is inherently limited to skin conditions (Cheer and Plosker, 2001; Al-Daraji et al., 2002; Gupta, Adamiak, and Chow, 2002; Gutfreund et al., 2013).

Topical tacrolimus is site-localized with minimal skin absorption and exhibits far fewer side effects than comparable use of corticosteroids (Gutfreund et al., 2013). Approaches focused on targeted inhibition of misregulated pathways have clear promise

in improving overall disease outcomes, including in the context of NFAT inhibition, but what is needed is a generalizable mechanism to deliver potent and localized cellular inhibition of NFAT. Future efforts toward this end may apply adeno-associated virus (AAV) vectors, liposomes, or other delivery mechanisms to drive the expression of VIVIT⁸⁶ or CRISPR/Cas9 in specific tissues at particular times (Colombo et al., 2022).

The imminent importance of NFAT became obvious in the organ transplant era. The recipient immune system will wage immunological war against non-self organs, which requires recipients to undergo life-long immunosuppressive therapy. One of the most successful approaches to preventing organ transplant rejection has been the use of calcineurin inhibitors, including cyclosporine and tacrolimus (Ellis, 1995; Mihatsch et al., 1998; Scott et al., 2003; Lerut et al., 2008). These calcineurin inhibitors block the NFAT-mediated transcription of IL-2, IL-4, TNF- α , and IFN- γ , which dampens the adaptive immune response (especially T cell mediated responses) and dramatically extends the useful lifespan of the transplanted organ, most notably in the case of liver transplants (McCaffrey et al., 1993; Moench et al., 2007).

Although many immunosuppressive therapies have markedly increased risks for various opportunistic infections, calcineurin inhibitors are comparatively spared from this disadvantage. For instance, alemtuzumab, which depletes B and T cells, increases the risk of a number of bacterial, viral, and fungal infections, including *Staphylococcus*, Hepatitis B, and *Cryptococcus* (Fishman, 2007; Harris et al., 2021). By contrast, tacrolimus has potent antifungal activity and few infectious disease risks have been noted with tacrolimus monotherapy (McAlister et al., 2006; Singh et al., 2016b; Emal

⁸⁶VIVIT will be discussed more extensively in later sections, but is a peptide-based inhibitor of the calcineurin-NFAT interface that acts in a functionally dominant-negative fashion by utilizing a higher-affinity version of the SPRIEIT (or PxIxIT) sequence natively found in the NFAT proteins.

et al., 2019; Gong et al., 2021; Papon et al., 2021). This evidence would suggest compensatory mechanisms are available to fend off many pathogens while maintaining enough targeted immunosuppression to prevent organ rejection. This provides evidence of the potential for the use of NFAT inhibitors in an infectious disease context without overtly inhibiting the overall immune response to the infection.

New approaches have begun to be developed for NFAT inhibition. One notable example is the development of a comparatively more selective calcineurin-NFAT inhibitor, INCA-6 (Roehrl, Wang, and Wagner, 2004; Roehrl et al., 2004). While tacrolimus acts through binding of FK506-binding proteins that then complex with and inactivate calcineurin, INCA-6 selectively blocks the interaction between calcineurin and NFAT, sparing some or most of the other functions of calcineurin while maintaining potent immunosuppression. While this inhibitor has not been exhaustively trialed, this general theme offers some promise for more targeted therapies that can overcome some of the side effects of traditional calcineurin inhibitors while maintaining most of the benefits.

Another approach that has more recently begun to gain traction is the development of isoform-selective inhibitors. The four NFAT isoforms have selective expression and activation profiles that make them conceivably differentiable biochemically. Select recent work has begun to do exactly that, by taking a structure-guided approach to identifying regions of the proteins unique to particular isoforms and targeting them with small molecules (Kitamura and Kaminuma, 2021). While no isoform-specific inhibitors are yet available, this will likely change in the coming years as distinct roles for these proteins continue to be unveiled.

Beyond small molecule based approaches, the age of personalized medicine opens possibilities for gene therapy approaches to ameliorate pathology caused by one or a

combination of NFAT isoforms in particular tissues. For instance, a T cell-targeted NFATC2 mutation may result in comparable graft-sparing immunosuppression to tacrolimus while limiting deleterious consequences to a single cellular compartment. With the advent of chimeric antigen receptor T cells (CAR T cells), the use of autologous donation for genetic modification has both clinical approval and potential additional practical applications (June, 2007). Alternately, new delivery mechanisms may make it possible to selectively deliver traditional small molecules to particular cell types, perhaps through molecular caging approaches or liposomal delivery to discrete tissues (Hu et al., 2019; Mukhtar et al., 2020). These possibilities and others foreshadow a future of greater specificity in targeting NFAT signaling, on both the isoform and tissue fronts.

1.4.2 Differentiation of Individual Isoforms

As alluded to previously, the different NFAT isoforms have begun to be assigned discrete functions in different contexts. As part of the present work sought to define isoform-specific roles for NFATC2, it may be of utility to dissect the known differentiable functions of these four isoforms, which may further inform future studies. There is a great deal of interesting biology that derives from the different expression patterns and properties of these proteins, which will be briefly reviewed here. While an early publication (Masuda et al., 1995) evaluated the expression of the then-three known isoforms of NFAT across a range of tissues via Northern blot, little immediate work was done to evaluate if this expression carried any functional consequences but this set off a flurry of study on the roles of these isoforms in many different tissues, which is the focus of this section.

NFATC1

Like most⁸⁷ of the NFAT isoforms, NFATC1 was first investigated in the context of lymphocyte biology, having been isolated from a bovine thymus and from human Jurkat T cells (Northrop et al., 1994). This protein was initially found to be important for T cell activation and relatively restricted in expression pattern based on the cDNA library hybridization approaches used at the time. NFATC1 was found to bind and activate the IL-2 promoter, suggesting an important role in the differentiation of T cells into CD4⁺ effector T cells (including both T_H1 and T_H2 cells) and, later, memory T cells (Monticelli and Rao, 2002; Oestreich et al., 2008; Torgerson et al., 2009; Martinez et al., 2015; Klein-Hessling et al., 2017; Yahia-Cherbal et al., 2019). Regulation of this essential pathway in T cell biology made this isoform of especial biological interest going forward given the importance of T cells in infection, autoimmunity, and cancer. It was subsequently found that NFATC1 is an important host factor for HIV-1 replication; this is tied to the importance of NFATC1 in T cell activation as HIV-1 requires T cell activation for efficient replication (Kinoshita et al., 1997) and NFATC1 binds the HIV-1 long-terminal repeats (Romanchikova et al., 2003). Further evidence of this conflicted nature of NFATC1 was evinced by findings that mice doubly deficient in NFATC1 and NFATC2 displayed both effector CD4⁺ T cell differentiation defects and B cell hyperinflammation and hyperproliferation, ascribing these two NFAT isoforms key roles in not only lymphocyte activation, but in the regulation of tonally appropriate response to both pathogens and at homeostasis (Peng et al., 2001). In addition to its roles in regulating IL-2, further studies found that NFATC1 (again in collaboration with NFATC2) regulates IL-4 transcription. Specifically, NFATC1 deficiency led to diminished production of IL-4, demonstrating

⁸⁷Really, all but NFATC4.

that NFATC1 is a positive regulator of IL-4 production and T_H2 responses (Monticelli and Rao, 2002). A further study found that, conversely to the effect of NFATC1 and NFATC2 on the IL-4 promoter, NFATC1 was incapable of ectopically inducing *TNF* transcription in a manner that depended upon the C-terminal activation domain of NFATC2, providing compelling evidence at one level at which NFAT isoforms can be differentially regulated (Kaminuma et al., 2008).

While work has largely slowed on this pathway in the context of T cells, additional work built on this foundation to further dissect specific aspects of T cell biology regulated by NFATC1. In the modern era of cancer immunotherapy, factors regulating PD-1 expression became of interest and it was found that NFATC1 is required for PD-1 expression after T cell activation, which gives this pathway an important role in regulating anti-cancer responses, especially in the context of PD-L1/PD-1 checkpoint inhibitor treatment (Oestreich et al., 2008). NFATC1 was also found to control the cytotoxicity of CD8⁺ T cells, giving it key roles on both arms of T cell differentiation. In the absence of NFATC1, T cells have reductions in cytotoxicity and fail to control infection by bacterial pathogens (Klein-Hessling et al., 2017). NFATC1 activity in some ways defines this differentiation process as FOXP3, a major regulator of T_{reg} development, downregulates the transcription-level expression of *NFATC1* to block the expression of T_{eff} cytokines (Torgerson et al., 2009). This was an early indicator of the ways in which the NFAT isoforms can be dynamically regulated at the transcription level, a phenomenon worthy of further discussion later on.

Recently, far more work has been done to define the role of NFATC1 in other realms of biology; most notably, important roles for NFATC1 have been uncovered in osteoclast differentiation, muscle cells, (lymphatic) endothelial cell biology, cardiac development, podocytes, other immune cell types, and various aspects of stem cell

and cancer biology.

Osteoclasts are complex bone-degrading cells ontologically derived from macrophages after M-CSF and RANKL stimulation⁸⁸. In this realm, NFATC1 has been found to be the master regulator of osteoclast differentiation, constituting perhaps the single most important biological role for this isoform. While individual isoform mutants tend to have mild effects in T cells and combinatorial mutation (as we will see) is necessary for complete disruption of function, in osteoclasts NFATC1 is the sole necessary member in this differentiation process from macrophages into osteoclasts (Kim and Kim, 2014). The regulatory mechanisms of the NFAT family logically fit into the overall kinetics of bone biology; the high calcium environment primes the calcineurin-NFAT axis and this environment is required for osteoclastogenesis (Negishi-Koga and Takayanagi, 2009). Study of osteoclast biology has also revealed interesting and potentially informative roles in the interplay of NF-κB and NFATC1 transcriptional activation, with NF-κB having been shown to bind the *NFATC1* promoter (Muhammad et al., 2014). A further study found that the NF-κB protein RelB was able to inhibit the transcription of *NFATC1*, adding yet more nuance to the interaction between these proteins (Zhao et al., 2015). Whether or not NF-κB can bind other NFAT promoters to drive increased transcription remains unknown. These studies also identified auto regulatory functions of NFATC1, which binds its own promoter to drive transcriptional upregulation during differentiation (Asagiri et al., 2005). NFATC1 is initially activated through the combinatorial effects of TRAF6, c-Fos, STAT3, and p38 signaling after RANKL binding to RANK; ectopic expression of NFATC1 in development facilitates osteoclast differentiation in the absence of RANKL (Takayanagi et al., 2002; Matsumoto et al., 2004; Huang et al., 2006; Yang

⁸⁸These cells work in tandem with osteoblasts to regulate bone mass. Interestingly, NFAT is important on both sides, playing important regulatory roles in bone deposition as well (Winslow et al., 2006).

et al., 2019; Huang et al., 2020)⁸⁹. Additional signals from ITAM-linked receptors through NFATC1 are also important for inducing these responses in a non-redundant fashion with RANK signaling (Koga et al., 2004). The absence of functional osteoclasts leads to progressive osteopetrosis in the mouse (Takayanagi et al., 2002; Kim and Kim, 2014). This paper led to a series of follow-ups defining some of the previously mentioned regulatory mechanisms. Particular inflammatory stimuli, including activation of DECTIN-1 is able to inhibit osteoclastogenesis through inhibition of NFATC1 via *MAFF* promoter occupancy (Zhu et al., 2017). These studies have revealed in remarkable detail the absolutely indispensable role for NFATC1 signaling in the process of osteoclastogenesis and homeostasis by integrating environmental factors with a series of intersecting pathways. One notable feature of osteoclasts is that they are multinucleated cells⁹⁰ and their fusion is dependent on NFATC1 activity, a process that appears to be conserved across various tissue types (Kim et al., 2008); another major multinucleated cell type with important roles for NFATC1 is that of muscle cells.

Muscle cells allowed for a more thorough exploration of NFATC1 as an electrical and mechanical sensor. Skeletal muscle is another context in which multiple NFAT isoforms are playing important roles, with NFATC1, NFATC2, and NFATC3 all having described roles (Delling et al., 2000; Stevenson et al., 2001; McCullagh et al., 2004; Amberg et al., 2004; Tothova et al., 2006; Layne et al., 2008; Ehlers,

⁸⁹The text of (Huang et al., 2020) (presumably) erroneously refers to NFATC2 and, given the abundance of evidence against NFATC2 playing a role in osteoclastogenesis, I will assume that NFATC1 was intended here.

⁹⁰One of the interesting phenomena within the tuberculous granuloma is the formation of what are known as multinucleated giant cells, which are an undercharacterized component of the granuloma structure. A potential role for NFAT in inducing the formation of these multinucleated giant cells is compelling by analogy to osteoclastogenesis, but this remains purely speculative (Forkner, 1930; Zhu and Friedland, 2006; Lay et al., 2007; Zhu et al., 2007; Shrivastava and Bagchi, 2013; Mezouar et al., 2019; Losslein et al., 2021).

Celona, and Black, 2014). For NFATC1 specifically, others have found that NFATC1 can detect nerve activity as exercise stimulation induces the nuclear translocation of NFATC1 while extensive rest results in shuttling back to the cytoplasm, likely through mechanosensitive calcium channels (Tothova et al., 2006). NFATC1 also plays a key role in muscle regeneration; after muscle damage, NFATC1 activity is correlated with increased *MYOD* expression (Sakuma et al., 2003), but during muscle fiber type differentiation, NFATC1 negatively regulates MYOD protein by binding and blocking MYOD interaction with p300. The net effect of this is that NFATC1^{hi} myocytes become slow-twitch muscle fibers while NFATC1^{low} myocytes become fast-twitch fibers (Ehlers, Celona, and Black, 2014).

An early histological observation of *Nfatc1*^{-/-} mice was that they possessed a heart defect leading to pulmonary and aortic valve abnormalities that resulted in congenital death by E15 (Pompa et al., 1998; Ranger et al., 1998a). These defects gave an important role to NFATC1 in regulating endothelial development and lead to speculation that NFATC1 may regulate some aspect of VEGFR signaling, as VEGFR is the master regulator of endothelial biology. Indeed, VEGFA stimulation of human pulmonary valve endothelial cells (HPVECs) led to NFATC1 activation (and not that of any other isoforms) through VEGFR2, which induces proliferation of the HPVECs (Johnson et al., 2003a). NFATC1 was also activated in the endocardium by RANKL, the same ligand that drives myeloid differentiation into osteoclasts; this shared pathway suggests that RANKL may broadly activate NFATC1 in tissues where the receptor is present (Combs and Yutzey, 2009). This was among the first mechanistic descriptions of a role for NFAT in the endothelium, where it is now known to be an important transcription factor mediating angiogenesis, but in a manner that is somewhat limited by the particular endothelial subtype and microenvironment (specifically in the endocardium) (Courtwright et al., 2009; Jang et al., 2010; Wu

et al., 2012; Wu, Baldwin, and Zhou, 2013; Gunawan et al., 2020). This work *in toto* establishes a developmentally essential role for endocardial NFATC1 expression in the formation of pulmonary and aortic valves and a potentially broader role of NFATC1 in the regulation of particular aspects of endothelial cell biology. A slightly broader study looked at this phenomenon from a pan-NFAT perspective and found similar effects, where interactions between endocardial and myocardial NFAT responses determined the development of the heart valve in a VEGFA-dependent manner (Chang et al., 2004).

In contrast to the rather specific role for NFATC1 in endocardial development, NFATC1 is broadly essential to lymphatic endothelial specification and elaboration, a process known as lymphangiogenesis and which will be discussed at length in Section 5.8. NFATC1 plays an essential and non-redundant role in regulating the formation of lymph sacs and valves, for which it also serves as a definitive genetic marker (Shin et al., 2019). *Nfatc1*^{-/-} mice have a normal vascular system but a reduction in lymphatic proliferation and morphogenesis (Kulkarni, Greenberg, and Akeson, 2009; Norrmen et al., 2009). Similar to previous findings in the endocardium, the activation of NFATC1 depended on VEGFR2⁹¹ activity, providing further evidence of the unique relationship between this receptor and transcription factor in mediating major processes in endothelial biology.

The activity of NFATC1 in immune cell types other than lymphocytes is relatively unclear. There is evidence that TNF-TNFR signaling on macrophages activates macrophage NFATC1, driving increased osteoclastogenesis. However, this was studied primarily in the context of bone homeostasis (as discussed previously) and not in the context of infection. Whether macrophage TNFR signaling through NFATC1

⁹¹This is, in fact, the finding, that VEGFR2 activity on venous endothelial cells drives the proliferation of the lymphatic endothelium, not VEGFR3 as might be assumed.

contributes to the biology of infection in any way remains, as yet, unknown⁹² (Yarilina et al., 2011). TNF- α also contributes to podocyte⁹³ damage through an NFATC1-dependent mechanism. TNF induces alterations in cholesterol metabolism that allows NFATC1 to induce apoptosis, which in turn compromises kidney function⁹⁴ (Zhang et al., 2013; Pedigo et al., 2016).

Lastly, NFATC1 has been found to have variable, but influential roles in the biology of stem cells and during particular cancers, notably hepatocellular carcinoma (Daniel et al., 2014). However, the general theme of NFATC1 activation in the context of cancer is the induction of an epithelial-mesenchymal transition that promotes dissemination and invasion. This was first described in 2006 in a publication that found that NFATC1 is over expressed in pancreatic carcinoma and drives increased expression of c-myc, a classical oncogene (Buchholz et al., 2006; Eerola et al., 2019). As is common with c-myc activation, this drives increased proliferation and reduced contact inhibition. This set a foundation for exploration of this pathway in other cell types and suggested that inhibition of NFATC1 may be a promising therapeutic target in carcinoma. This pathway was then translated to hepatocellular carcinoma, in which c-myc, once again, increases proliferation of the cancer (Wang et al., 2012). Notably, ectopic expression of NFATC1 in non-expressing cancers increases cell invasion and motility through repression of *CDH1*⁹⁵ expression (Oikawa et al., 2013). This model

⁹²This becomes more compelling in the context of new roles for TNF in the formation of multinucleated giant cells within the granuloma (Mezouar et al., 2019). Study of whether this is strictly dependent on NFATC1 signaling would be an very consistent insight and compelling new tools to perturb this cell type in the granuloma.

⁹³Podocytes are epithelial cells that reside within the glomerulus of the kidney and aid in filtering blood.

⁹⁴This would logically suggest that tacrolimus therapy could be beneficial to kidney health after transplant if the effects could spare the deleterious effects on the endothelium (Randhawa, Starzl, and Demetris, 1997; Nankivell et al., 2016).

⁹⁵Epithelial cadherin

was later integrated with potential cancer initiating stimuli through evaluation of NFATC1-STAT3 transcriptional complexes⁹⁶ that can be induced by chronic inflammation to promote KRAS^{G12D}-driven carcinogenesis using a pancreatitis model. This raises an obvious question: can pharmacological inhibition of NFATC1 improve cancer prognosis, given that we have two highly efficacious calcineurin-NFAT inhibitors with FDA approval – tacrolimus and cyclosporine A. Indeed, treatment of mice bearing prostate carcinomas with either drug improved disease outcome by reducing proliferation and metastasis⁹⁷ (Kawahara et al., 2015a). Identical outcomes were found by the same group in the context of bladder cancer⁹⁸ (Kawahara et al., 2015b). From these results, it is reasonable to conclude that NFATC1 can act as an oncogene across a range of different oncological contexts through driving increased c-myc expression (Buchholz et al., 2006; Flockhart et al., 2009; Seifert et al., 2009; Oikawa et al., 2013; Kawahara et al., 2015a; Kawahara et al., 2015b; Liu et al., 2021a). Other oncogenic targets of NFATC1 have been proposed including BMI1 (Wu et al., 2019) and ITGA5 (Eerola et al., 2019), although more exhaustive characterization is in order.

However, the literature is not unidimensional on this topic. A more recent study reevaluated NFATC1 in hepatocellular carcinoma and found that *low* NFATC1 expression was correlated with increased tumor progression and size and that NFATC1 activates *FASL* to drive apoptosis (Xu et al., 2018a). Conversely, NFATC1 overex-

⁹⁶This same NFATC1-STAT3 transcriptional complex has been shown to be important for osteoclastogenesis, continuing on the theme that context is essential for interpreting consequences of pathway activation (Baumgart et al., 2014)

⁹⁷The potential for a targeted delivery to the prostate seems compelling given the abundance of surface markers for prostate cancer and slow disease progression in many cases (Bradford et al., 2006).

⁹⁸The bladder cancer finding is interesting and little has been done to study the intersectional benefit of the standard bladder cancer treatment – BCG inoculation – and some of these other drugs. It would be intriguing if NFAT inhibition altered BCG-dependent cytotoxicity against bladder cancer, as this could reveal further mechanisms of NFAT in mycobacterial responses more generally.

pression suppressed proliferation via an EGR2-dependent response pathway (Wang et al., 2020a). This would suggest that NFATC1 is a tumor suppressor in hepatocellular carcinoma. While this is an outlier in the overall schema, it is probable that NFATC1 can act as either pro- or anti-tumor depending on additional, as-yet-unknown factors or epistatic interactions.

A variety of minor roles have been uncovered for NFATC1 in regulating particular populations of stem cells. In the hair follicle, NFATC1 activity is related to quiescence while downregulation of NFATC1 spurs proliferation (Horsley et al., 2008; Keyes et al., 2013). This finding explained a known clinical phenomenon wherein patients receiving cyclosporine A display accelerated hair growth and established an important regulatory role for this protein in cycles of growth and quiescence in this tissue. In contrast to its roles in cancer, here NFATC1 activity correlates with inhibition of growth. Similar effects are seen in lung stem cells, where these NFATC1-expressing cells are induced toward terminal differentiation during regeneration and replacement of alveolar epithelia (Lee et al., 2014).

While comprehensive profiling of the presence of NFAT isoforms in particular immune cell subtypes and under different conditions is broadly lacking, one cell type in which NFATC1 is known to be active is neutrophils. Exposure of neutrophils to allergic antigens⁹⁹ or immunoglobulin E activated them to express *COX2* and secrete prostaglandin E2¹⁰⁰ in a NFAT-dependent manner (Vega et al., 2007). Subsequent work investigated this pathway in the context of mast cell biology and, although the

⁹⁹These neutrophils were purified from patients with known allergies and compared to “normal” neutrophils from patients who lacked the allergy.

¹⁰⁰There is much that can be said about the role of prostaglandin E2 (PGE₂) in tuberculosis immunity (Serhan and Chiang, 2008). One of the major roles that has been uncovered is as a protective lipid able to modulate the balance between IL-1 and IFN- α/β to improve disease outcomes (Mayer-Barber et al., 2014). The study of host lipid signaling molecules is an interesting field of tuberculosis biology but is beyond the present scope of this work.

approaches are somewhat discordant¹⁰¹, found an important role for NFATC1 in regulating the transcript-level expression of HIF-1 α under hypoxia, potentially linking NFAT activity to the induction of hypoxic responses and, importantly for this work, VEGFA expression and angiogenesis (Walczak-Drzewiecka et al., 2008). No further work has been done to evaluate these results on the NFAT-HIF-1 α interaction, but this is one potential pathway by which the results given in Chapter 3 might operate, although with a different isoform – NFATC2.

NFATC2

NFATC2 was actually the first of the NFAT proteins to be identified, in 1988 by Gerald Crabtree. This protein was found to be required for the T cell activation cascade and bound the IL-2 promoter as well as the LTR of HIV-1 and was assumed to be a single gene product and not a member of a larger family of proteins, akin to the situation in *Drosophila* (Shaw et al., 1988). Anjana Rao subsequently found that NFATC2 was activated by calcineurin and could be inhibited by clinical calcineurin inhibitors and that NFATC2 engaged in cooperative binding with members of the AP-1 transcription factor family to induce transcription of IL-2 (Jain et al., 1993). These two studies provided us the general model of NFAT activity that has been repeatedly validated into the present. Subsequent results added some subtlety to this described transcriptional effect through the discovery that NFATC2 acted in an immunoinhibitory fashion, with *Nfatc2*^{-/-} mice demonstrating increased B cell proliferation in response to *Leishmania* and exacerbated allergic responses to ovalbumin (Xanthoudakis et al., 1996).

¹⁰¹For instance, they precipitated the HIF-1 α promoter with NFATC1 but utilized NFATC4 over-expression to demonstrate increased HIF-1 α expression. Under a model where all of the NFAT isoforms are equivalent, this is a reasonable enough approach, but lacks the subtlety that both more modern and more classical approaches might take.

As might be expected, this founding member of the NFAT family was also the first to have described roles beyond the adaptive immune system with Aramburu et al. (1995) finding that NFATC2 was activated in natural killer cells¹⁰² after stimulation of Fc γ RIII. Intriguingly and in an observation rarely noted in the literature, this activation of NFATC2 drove the transcription of *NFATC1*, suggesting NFAT cross-family regulatory patterns and the potential for NFAT transcriptional induction to be an additional layer of regulation within this pathway.

But by 2000, it was found that NFATC2 was an important repressive determinant of chondrogenesis as mice lacking NFATC2 exhibited increased chondrocyte¹⁰³ development and ossification and appeared to act as a tumor suppressor (Ranger et al., 2000). This tumor suppressor hypothesis introduced by the authors would be more thoroughly explored some time later by comparison of the effect of activation of NFATC1 and NFATC2 in fibroblasts. A constitutively active NFATC2 induced apoptosis in these cells while a similarly modified NFATC1 induced proliferation and transformation; conversely, *Nfatc2*^{-/-} mice were more susceptible to tumor formation (Robbs et al., 2008). However, together, these two proteins repress the development of osteoarthritis (Greenblatt et al., 2013). This mechanistic differentiation between these closely related isoforms was a strong indication that, despite having a common DNA binding motif and overlapping gene expression, that these isoforms were differentiable in function and, likely, act in a balancing manner in normal cells. Other studies found that NFATC2 inhibits the expression of *CDK4*, a kinase important for cell division and often misregulated in cancer (Baksh et al., 2002; O'Leary, Finn, and

¹⁰²While this is not a dramatic deviation from the theme, as natural killer cells are derived from the lymphoid compartment, it was an early indication that T cell activation was not the whole tale of NFAT.

¹⁰³Chondrocytes are cells that produce and maintain cartilage between bones and are essential for healthy skeletogenesis.

Turner, 2016) and CCNA2, a key cell cycle protein (Carvalho et al., 2007).

Similar to the previous description for NFATC1, NFATC2 is a critical regulator of T and B cell activation, but primarily in concert with NFATC1 (Peng et al., 2001). Individual roles for NFATC2 came later, with a notable publication finding that IL-6 detection by T cells drives increased NFATC2 expression and subsequent T_H2 differentiation and colitis, as this signaling pathway drives NFATC2-dependent transcription of IL-4 (Diehl et al., 2002; Weigmann et al., 2008). Unlike NFATC1, which appears to more broadly induce the expression of both pro- and anti-inflammatory cytokines, NFATC2 generally regulates the expression of anti-inflammatory signaling cascades; NFATC2-deficient mice have reduced expression of IL-10, one of the classical, anti-inflammatory, type II cytokines¹⁰⁴ (Lee et al., 2009). Further contributing to this is that while constitutively active versions of both NFATC1 and NFATC2 are capable of activating transcription of the IL-4 promoter, knockout mice for NFATC2 demonstrate dramatically reduced IL-4 production while NFATC1 knockout mice show *increased* IL-4 production (Monticelli and Rao, 2002) and delays in IL-4 resolution, conferring a resistance defect to *Leishmania* (Kiani et al., 1997). Conversely, NFATC2 alone appears to regulate TNF- α and IFN- γ expression in T cells as expression of NFATC2 was able to induce TNF- α while expression of NFATC1 was unable to do so, an ability conferred by the C-terminal activation domain (Teixeira et al., 2005; Kaminuma et al., 2008). TNF- α , although considered a classical pyrogenic cytokine, is highly pleiotropic and induces context dependent protection from bacterial infections and cancers but can induce autoimmunity and allergy. Signaling from this single molecule can induce a spectrum of phenotypes from proliferation to necrosis (Gough and Myles, 2020). Notable is the essential role of TNF- α in host defense to

¹⁰⁴Along with IL-4 and IL-13, IL-10 is a primary stimulant of inflammation-resolution after insult.

tuberculosis, implicating by proxy a potential role for NFATC2 in this process¹⁰⁵.

For the purposes of this document, one of the most impactful publications on NFATC2 came through an investigation on the contribution of NFATC2 to tuberculosis pathogenesis in mice (Via et al., 2012). While a more comprehensive analysis of how these findings integrate with the findings I will present in Chapter 3 is well warranted and will be addressed in Section 3.4, this was one of the, if not the, first studies on NFAT signaling as a contributor to tuberculosis pathogenesis and/or host defense and remains the primary study on the topic prior to my work presented in Chapter 3 and in Brewer et al. (2022b). *Nfatc2*^{-/-} mice have impaired adaptive immune responses to *M. tuberculosis* infection, with CD4⁺ T cells having especially notable defects. This study used the tuberculosis-resistant BALB/c background and infected with *M. tuberculosis* HN878, a more recent clinical isolate¹⁰⁶. *Nfatc2* null mice had 10³ greater bacterial burden and greatly enhanced dissemination at the terminal time point, although they had equivalent burden up to approximately 6 weeks post infection. Previous studies had identified that NFATC2 was required for production of both IFN- γ and TNF- α by T cells, but the findings here displayed an unusual mispatterning, with greater levels of TNF- α at 4 weeks post infections, a phenotype specific to CD4⁺ T cells. This collection of data suggested that NFATC2 was important for tuberculosis disease control over the entire course of infection, although the precise immunological mechanism remained unknown, with the majority of the phenotype attributed to these misregulated CD4⁺ cells.

¹⁰⁵Given the positive and negative roles that TNF- α has in the pathology of tuberculosis, NFAT may offer some mechanism for modulating expression without complete blockade, which may be one way that TNF- α -targeted therapies could go about shifting the balance of host immunity and host damage (Mootoo et al., 2009).

¹⁰⁶Both are somewhat unconventional choices, especially for the time. BALB/c mice are more T_H2 skewed than C57BL/6 mice, which are very T_H1 skewed. Mutation of NFATC2 on the BALB/c background should make the overall immune response more inflammatory, based on all of the findings already detailed.

NFATC2 has also been demonstrated to have important roles in antifungal responses with mesenchymal stromal cells within the adipose tissue, an unusual finding that suggests an important role for this pathway in various aspects of cell-autonomous defense as well as innate immunity (Tidu et al., 2021). Such findings may inform the use of calcineurin inhibitors in graft-versus-host disease, as the immune responses of stromal cells may be important in mediating antifungal control both from the intestine as well as invasive skin infections.

One of the innovations of the new millennium in the study of NFAT was more thorough characterization of double knockout phenotypes. We have already seen studies conducted with NFATC1 and NFATC2 double knockout animals¹⁰⁷, but none with other isoform combinations. Later studies investigated NFATC2 and NFATC3 combinatorial effects and uncovered evidence that double knockout of these isoforms renders CD4⁺ T cells unresponsive to suppression by T_{regs}, causing systemic hyperinflammation (Bopp et al., 2005) while NFATC2 mutation alone has no effect on T cell development (Cante-Barrett, Winslow, and Crabtree, 2007). This effect also causes intrinsic differentiation into hyperproliferative T_H2 cells, which may explain aspects of the inflammation seen in this context (Ranger et al., 1998a; Rengarajan, Tang, and Glimcher, 2002). Both of these had been previously seen to be immunosuppressive isoforms, so their combinatorial knockout appears to exacerbate the inflammatory phenotypes of the single knockout (Hodge et al., 1996; Oukka et al., 1998) while potentially allowing NFATC1 unrestricted access to promoter elements rather than the wild-type counterbalancing that likely occurs (Ranger et al., 1998a).

The capacity for NFATC2 to regulate growth became important for the develop-

¹⁰⁷These have high overlapping expression in T cells, were the first to be discovered, and are most similar to one another, providing a solid basis to begin study with these.

ment of muscle cells, which must undergo syncytiation¹⁰⁸ to become multinucleated. It was noted that *Nfatc2*^{-/-} mice had reduced muscle size compared to wild-type mice. These mice are capable of forming normal numbers of muscle fibers but are incapable of muscle hypertrophy, endomitosis, and myoblast fusion required for standard musculature (Horsley et al., 2001; Pavlath and Horsley, 2003; Schulze et al., 2005). Keeping with this theme, NFATC2 is also required for cardiac hypertrophy. NFATC2 is the dominant isoform in the heart and myocardial-specific constitutively active calcineurin-induced hypertrophy was abrogated in the *Nfatc2* null background. In this context, NFATC2 is clearly detrimental as knockout of NFATC2 protected mice from various pathologies with no impact on exercise-induced cardiac enlargement (Bourajjaj et al., 2008). In β islets in the pancreas, NFATC2 activation is required to induce proliferation, which depends on the transcriptional target *NR4A1* (Simonett et al., 2021). This develops an important theme for NFATC2, which generally appears to regulate important growth and cell division phenomena in contrast to the role of the other isoforms.

NFATC2 was also found to contribute to tumor-associated macrophage biology by promoting further macrophage infiltration into melanomas, which corresponded to increased growth and metastasis (Daniel et al., 2014). Notably, this phenomenon was present in response to overexpression of NFATC2 in either the melanoma itself or the tumor-associated macrophages (Liu et al., 2018). It had previously been found that NFATC2 transcriptionally inhibits *MITF* expression, inducing dedifferentiation which facilitates immune evasion (Perotti et al., 2016). Further publications on melanoma found that, through the induction of IL-8 and MMP3, NFATC2 in-

¹⁰⁸The formation of a syncytium can occur through either endomitosis, where cells divide their genome and nuclei but fail to undergo cytokinesis, or through fusion, where two cells fuse to form a larger cell comprised of the cellular contents of both cells. Many viruses induce the formation of cellular syncytia through cell-cell fusion, which seems to enhance their replication and spread (Jessie and Dobrovolsky, 2021).

creases tumor growth and metastasis (Shoshan et al., 2016). Similar effects were seen in lung adenocarcinoma, with more exhaustive work investigating the influence of NFATC2 on recurrence and survival, where high expression increased the likelihood of recurrence (Xiao et al., 2017); depletion of NFATC2 conversely decreased invasion and migration (Liu, Zhao, and Wu, 2013). While this phenotype is more reminiscent of those seen with NFATC1, these isoforms can clearly display some elements of redundancy and inappropriate activity, especially in a context like cancer. An interesting paper found an NFATC2 translocation event fusing EWSR1 (the classic Ewing sarcoma RNA binding protein) with NFATC2 through a chromosome 20:chromosome 22 fusion event that drove Ewing sarcoma. In these translocations, the N-terminal domains of NFATC2 were replaced by the N-terminal region of EWSR1, driving a constitutively active NFATC2 linked to the transactivating segment of EWSR1 (Szuhai et al., 2009).

Relevant for this work, roles for NFATC2 in myeloid cells are relatively understudied and much of what is known is derived from studies in dendritic cells¹⁰⁹. As previously mentioned, important pro-tumor roles for NFATC2 in tumor-associated macrophages have been identified (Liu et al., 2018). An earlier study examined NFATC2 in the context of myeloid development and found that myeloid progenitors expressed high levels of NFATC2 that diminished over the course of development, however this did not address any changes that might happen with further differentiation or functional stimulation in the context of disease (Kiani et al., 2004). A study on the underlying ontogeny of inflammatory bowel disease found that NFATC2 nuclear translocation in macrophages was essential for the development of the disease

¹⁰⁹Dendritic cells can derive from either the myeloid or lymphocytic compartments. Despite this caveat, for most purposes, most of the circulating dendritic cells are “conventional” dendritic cells from a myeloid origin while a minority are plasmacytoid (Shortman and Liu, 2002; Collin and Bigley, 2018).

and could be regulated by the NFAT kinase, LRRK2 (Liu et al., 2011). A follow-up study (which did not look at the isoform-specific level) used the VIVIT peptide to inhibit NFAT activation in macrophages and found that this provided a benefit in the context of ulcerative colitis in a mouse model (Elloumi et al., 2012). This inhibition blocked the expression of several major cytokines implicated in this disease, including TNF- α , IFN- γ , and IL-12. NFATC2 is also essential for the formation of foam cells through negative regulation of PPAR γ (Du et al., 2021), which are a specific lipid-laden subset of macrophages known to be present within the granuloma and to provide a replicative niche for mycobacteria (Russell et al., 2009; Kim et al., 2010b; Johansen et al., 2018; Guerrini and Gennaro, 2019; Agarwal et al., 2020; Agarwal, Gordon, and Martinez, 2021). Unlike NFATC1, NFATC2 is thought to be absent from neutrophils, the other major cell type in the initial-early response to tuberculosis infection, suggesting that any primary role for NFATC2 in the immune response to tuberculosis is most likely to be restricted to the responding macrophages (Greenblatt et al., 2010). NFATC2 in mast cells has recently been identified to be important for the mast cell-mediated production of IL-3 and, indirectly, IL-9, which are key cytokines that regulate macrophage and T cell biology (Sabbaghi et al., 2021). While not a crystalline picture, this establishes some generally known roles for NFATC2 in myeloid cell biology, an important foundation for my work presented in Chapter 3.

NFATC3

One of the major roles that have been described for NFATC3 is in vascular biology, especially as it pertains to pulmonary hypertension (Nieves-Cintron et al., 2007; Nieves-Cintron et al., 2008). The first indication in this direction was that NFATC3 in tandem with NFATC4 is important for vascular patterning during mouse development, where mutation of either both of these isoforms or of calcineurin causes a lethal

failure of angiogenesis, rendering the fetus unable to develop properly organized vasculature (Graef et al., 2001). Additional characterization of these double mutants revealed profound defects in the cardiac tissue itself, exhibiting mitochondrial dysfunction (Kegley et al., 2001; Bushdid et al., 2003). Later, it was discovered that NFATC3 alone was responsible for cardiac hypertrophy in response to hypertension, giving this isoform a key role in cardiovascular responses more broadly (Wilkins et al., 2002). Under a condition described as “Ca²⁺ sparklets,” it was uncovered that these calcium signals were able to stimulate cardiac calcineurin to drive the activation of NFATC3, which induces a variety of hypertensive phenotypes, including increases in cytosolic calcium and increased muscle tone (Nieves-Cintron et al., 2007). A pair of papers from the same group found that a set of micro RNAs were able to regulate the activity of NFATC3 in the context of cardiac hypertrophy, adding a novel layer of regulation to the NFAT family of proteins and suggesting that other isoforms may be regulated by miRNAs under specific circumstances (Lin et al., 2009; Wang et al., 2010).

Shortly thereafter, a new player in the regulation of NFATC3 was introduced – hypoxia. Under conditions of hypertrophy, the cardiac muscle experiences increased hypoxia and it was a question of whether or not these previously described NFATC3 phenotypes may be integrated with, independent of, or in opposition to that hypoxia. A study found that hypoxia increased the expression of endothelin-1 (a vasoconstrictive signaling peptide), which activated arterial NFATC3 in a manner facilitated by changes in actin polymerization (Frutos et al., 2011). These somewhat discordant observations set the stage for further studies on the interactions of hypoxia, a classic stimulus of angiogenesis, and the NFAT signaling pathway. Similar to other hypoxia-responsive pathways, this one is also responsive to superoxide and other reactive oxygen species, suggesting that there may be some connection between the

activation of NFATC3 and HIF-1 α either directly or indirectly (Ramiro-Diaz et al., 2014). Outside of the heart proper, NFATC3 is required for muscle development; knockout results in decreased muscle size in adult mice via alterations in myosin heavy chain expression (Delling et al., 2000; Kegley et al., 2001). Other alterations are seen other arterial smooth muscle, reducing the expression of potassium channels after angiotensin treatment (Stevenson et al., 2001; Amberg et al., 2004); a similar effect is seen in bladder smooth muscle cells (Layne et al., 2008). This argues for a broader and multifaceted role for NFATC3 in regulating the biology of smooth muscle cells and suggests that this may be an interesting opportunity for future study of the contributions of NFATC3 to various muscle conditions.

More minor roles for NFATC3 have been identified in astrocytes, which are support cells within the brain. NFATC3 is activated by glutamate receptors, which induce influxes of calcium. Despite uncovering this activity, no particular role was ascribed to NFATC3 in these cells (Jones et al., 2003). The role for NFATC3 in liver regeneration has also been assessed and this isoform encourages hepatocyte proliferation (Pierre et al., 2009). These anecdotal roles for NFATC3 suggest broader roles in other tissues that have not been fully explored. In the days of single cell transcriptomics, it is likely worthwhile to explore the role of this isoform and others in tissues with clearly enriched expression of single isoforms.

By intriguing contrast to the previously discussed findings on NFATC2, NFATC3 was found to be an important *positive* determinant of chondrogenesis (Ranger et al., 2000; Tomita et al., 2002). NFATC3 activity induces *BMP2*, which drives differentiation of chondrocytes from mesenchymal cells. This suggests an interesting potential interplay between NFAT isoforms where they may be able to competitively inhabit particular binding motifs and repress or activate transcription based on the activ-

ity of other transcriptional coregulators exclusive to one or a subset of the isoforms. A more comprehensive assessment of these sort of interactions would certainly provide an invaluable resource in interpreting some of these polarized results found with distinct isoforms.

NFATC3, more than any of the other isoforms, has been found to have a number of roles in macrophages, courtesy of the observation that NFATC3 nuclear localization was required for TLR-mediated inflammatory responses (Minematsu et al., 2011) but that NFATC3 nuclear localization was inhibited by NF-κB as well (Conboy et al., 1999). As has been seen previously, NFAT protein can have antagonistic roles with NF-κB and this is the explanation provided here. These unusual results (both necessary and inhibitory) are somewhat at conflict with the remainder of the literature, however, as TLR agonists were incapable of directly activating NFAT and this effect was mediated by existing pools of nuclear NFATC3 in the cells they were using that appeared to mediate this phenotype. This suggested a new level of transcriptional regulation that depended on some layer of interaction between NFATC3 and NF-κB that prevented excessive NF-κB activity. A complementary study evaluated the contributions of NFATC3 in Gram-negative sepsis and found that NFATC3 was important for the release of pro-inflammatory cytokines and compounds, specifically nitric oxide¹¹⁰. Again, these results argue against a direct role for TLRs in inducing NFAT, but suggest that existing nuclear NFATC3 or some other stimulus facilitate transcriptional responses in cooperation with TLR-induced signals (Ranjan et al., 2014). During lung injury, NFATC3 was found to exacerbate lung pathology due to

¹¹⁰Nitric oxide has many described roles in both modulating vascular dilation and in host defense against mycobacteria. Early studies found an essential role for the inducible nitric oxide synthase in protection against *M. tuberculosis*, adding yet another key pathway that is regulated, at least in part, by NFAT (MacMicking et al., 1995; MacMicking, Xie, and Nathan, 1997; MacMicking et al., 1997).

regulation of CCR2¹¹¹ and TNF- α production by macrophages¹¹² (Karpurapu et al., 2018). These NFATC3-dependent macrophage activities drove neutrophil recruitment and vascular damage; genetic removal of NFATC3 accelerated lung repair and diminished inflammatory responses. Finally, in 2019, an answer was uncovered as to what, if any stimulus, was causing NFAT activation in macrophages during sepsis. TRPV4, a calcium channel responsive to vanilloid activation, was found to increase Ca²⁺ influx in macrophages to exacerbate the production of reactive oxygen species and cytokines; inhibition of this channel damped these effects and improved mouse survival after injury (Li et al., 2019). Other studies have identified an important role for NFATC3 in inhibiting the formation of foam cells¹¹³ in atherosclerosis. Transgenic overexpression of NFATC3 was protective against atherosclerosis and prevented the formation of foam cells via miR-204, making NFAT *activation* potentially of therapeutic use in this disease (Liu et al., 2021b).

Additional studies have identified important roles for NFATC3 in concert with NFATC1 in antifungal responses in neutrophils. Greenblatt et al. (2010) found that cyclosporine treatment of mice increased their susceptibility to *Candida* infection through blockade of calcineurin activity. Through a genetic approach, they found that NFATC1 and NFATC3 were redundantly required for the induction of IL-10, but were dispensable for fungal killing and overall murine survival, suggesting that calcineurin

¹¹¹CCR2 is a chemokine of two minds: essential for the recruitment of macrophages in response to mycobacterial infection and ultimately detrimental to the outcome of disease by recruiting the wrong kinds of monocytes and macrophages. This will be detailed in Subsection 1.6.1, but for now, CCR2 induction can be considered an incidental pro-bacterial response during tuberculosis although it is likely protective in most other disease contexts (Cambier et al., 2014).

¹¹²The intimate role of these cytokines in particular may suggest as-yet unknown roles for this protein in mediating host susceptibility to tuberculosis. The development of a macrophage-specific knockout model for NFATC3 would be an interesting approach to understanding the pathways responsible for inducing expression of these host-deleterious factors.

¹¹³These lipid-laden cells also develop during granuloma formation; it would be interesting to see if the NFAT approaches in Chapter 3 alter the formation of these in the granuloma and, if so, the consequences of those changes.

is executing NFAT-dependent and -independent functions within neutrophils during fungal infection – dissection of these contributions and any alterations resulting from NFATC1/NFATC3 knockout in a more granular way would certainly add to our understanding of this pathway more generally.

None of these sections would be complete without a description of the role of NFATC3 T cell biology, where, once again, it plays an important role in thymocyte positive selection and T_{reg} activity (Ho et al., 1995; Bopp et al., 2005). In the absence of NFATC2 and NFATC3, T_{regs} are unable to inhibit CD4 $^{+}$ T cells, assigning these isoforms inhibitory roles toward the helper activity of CD4 $^{+}$ T cells. NFATC3 is also specifically required for positive selection and null mice are unable to generate CD4 $^{+}$ /CD8 $^{+}$ double-positive thymocytes; indeed, this is an effect specific to NFATC3 as rescue experiments with other isoforms failed to restore the generation of double positive cells (Cante-Barrett, Winslow, and Crabtree, 2007). The motifs present on NFATC3 that grant this specific activity were not identified but understanding of these would greatly enhance our model for isoform-specific protein features able to differentiate the activity of proteins within this family. NFATC3 is also capable of inducing expression of IFN- γ and TNF- α while downregulating IL-4 and IL-5 expression; these findings are gain-of-function experiments as no role was able to be identified with mutation of NFATC3 alone (Chen et al., 2003). Aside from these roles, relatively little else is known about this isoform in T cells, unlike the extensive roles already described for NFATC1 and NFATC2.

NFATC4

As mentioned in the previous section, NFATC3 and NFATC4 have a number of cooperative roles, including in the developing vasculature and in cardiac development (Graef et al., 2001; Bushdid et al., 2003). However, NFATC4 has additional roles that

are somewhat unusual for the NFAT family of proteins as it is minimally expressed in the immune system and has found a variety of roles in diverse somatic tissues, including adipocytes, hepatocytes, and neurons. However, this is likely the least studied of this family, so further roles are likely to be discovered in the coming years.

NFATC4 has been of some interest in the pathology of diabetes due to its ability to regulate the expression of genes in adipocytes, including by inhibition of the transcription of adiponectin, an important biomarker for diabetes and obesity. While the ability for NFATC4 to inhibit expression of adiponectin was demonstrated *in vitro*, whether this pathway was important for the expression *in vivo* is unknown (Kim et al., 2006). Conversely, active NFATC4-mediated transcription in adipocytes is associated with the expression of a variety of pro-inflammatory cytokines, including TNF- α (Kim et al., 2008). Adipocyte LPIN1 is able to repress NFATC4 activation to mute these inflammatory responses, making LPIN1 a compelling anti-inflammatory target in obesity (Kim et al., 2010a). This provides a somewhat unified model, as chronic inflammation and obesity are known to be correlated and adipocyte NFATC4 activity may contribute to obesity-induced inflammation.

NFATC4 has also been ascribed important roles in mediating liver damage; this is in contrast to the role NFATC3 plays in facilitating liver repair and furthers the arguments for more targeted, isoform-level approaches to understanding the functions of these proteins. In non-alcoholic steatohepatitis (NASH), knockdown of *NFATC4* liberated PPAR α , which enabled renewed fatty acid metabolism to reduce the burden of lipids in hepatocytes; the activity of NFATC4 also increased macrophage recruitment via inflammatory signaling from the hepatocytes, further contributing to the inflammation seen in NASH (Du et al., 2020). These sorts of protein:protein interactions are a prelude for the later findings presented from Poli et al. (2022), which

investigated the cytosolic activity of NFAT in more specific detail. Later studies also revealed a role for NFATC4 in ethanol-induced cell death via inhibition of PPAR γ (Wu et al., 2021). In both cases, inhibition of NFATC4 resulted in improvements in disease and argues that this pathway is an key regulator of liver damage that is serving a maladaptive role in this context.

NFATC4 also has a number of described roles within neurons and neural progenitors. The earliest study found that NFATC4 was required for regulating the expression of the vasoactive intestinal peptide, an important neurotransmitter, but did not look at this activity in neuronal cells *per se* (Symes, Gearan, and Fink, 1998). The first description of NFATC4 specifically in neurons found it to be responsive to L-type calcium-channels and to induce ITPR1 expression, implicating it in hippocampal neurons and memory formation (Graef et al., 1999). Further studies have expanded upon this initial finding and found important roles for NFATC4 in regulating apoptosis in the cochlea (Benedito et al., 2005; Luoma and Zirpel, 2008), cortical neurons (Vashishta et al., 2009), and adult-born neurons (Quadrato et al., 2012). Moreno et al. (2015) found that hypoxia in neural stem cells activated NFATC4 and that this pathway is a major contributor to stem cell proliferation, differentiation, and self-renewal through regulation of ID2. Another study found that NFATC4 inhibits β -catenin signaling to drive neuronal proliferation and induce cell cycle exit (Huang et al., 2011). Together, these studies identify an important role for NFATC4 in regulating neuronal development and behavior; further studies in a knockout mouse should explore the behavioral and intellectual consequences of this disruption.

An older study also found that NFATC4 *inhibits* breast cancer motility (Fougere et al., 2010). This study found that inhibition of NFATC4 increased motility by

releasing inhibition of *LCN2* gene expression. This is in converse to a previous study that had found that NFATC4 was an interacting partner for the estrogen receptor and facilitated the expression of estrogen-responsive genes; knockdown of NFATC4 diminished breast cancer cell growth and was proposed as a potential therapeutic target (Zhang et al., 2005). This is a further demonstration of the pleiotropic nature of the NFAT pathway, but may also reflect differences in the biology of breast cancer given the dizzying array of subtypes and subtype-specific treatments now available.

NFATC4 has other miscellaneous described roles that warrant further investigation. During cardiac hypertrophy, overexpression of SIRT6 is able to inhibit the transcription and activation of NFATC4, which prevents excessive hypertrophy. This study was done primarily in the context of overexpression, so it would have been interesting to see if any physiological stimuli could activate SIRT6 to exert these effects as well (Li et al., 2018b). In the endothelium, NFATC4 was described as an important mediator of vascular inflammation; activated ORAI1 calcium channels drove NFATC4 activation and inhibition of these channels resulted in reduced inflammatory signatures in the vasculature (Yu et al., 2018). Both of these suggest that NFATC4 acts as a pro-inflammatory transcription factor during diverse cardiovascular responses.

Distinct Regulatory Mechanisms

As has become apparent in the previous section, one of the notable roles of NFAT is not only its ability to induce transcription, but the many contexts in which it inhibits transcriptional responses, especially those induced by NF-κB. While transcription factors are generally thought of as *inducers* of transcriptional responses, the complex interactions between this family and other families of transcription factors as well as between NFAT isoforms is an interesting subject for future discovery. For instance, in the context of angiogenesis, it is thought that NF-κB induction of the Fas lig-

and increased endothelial apoptosis and inhibits angiogenesis while NFAT-mediated activation of cFLIP, a pro-survival gene. Thus, the functional antagonism between these two transcription factors determines the overall degree and timing of angiogenesis within the endothelium (Aurora et al., 2010). Previously, Conboy et al. (1999) had found antagonism between NFAT and NF- κ B in the process of macrophage activation, where NF- κ B induces TNF- α while NFATC3 opposes that transcriptional induction. In the opposite direction, in osteoclasts, NF- κ B induces transcription of NFATC1, a key aspect of the differentiation of osteoclasts from macrophages (Asagiri et al., 2005). Future models of NFAT activation need to reconcile the tension between this and the NF- κ B signaling pathway in different contexts given their ability to mutually inhibit one another; it is even likely that these tensions have some level of isoform-specific regulation that is, to date, completely unknown.

Of course, one of the major mechanisms of differentiating the members of the NFAT family is in their expression profile. Isoforms that are exclusive to particular tissues are likely to exert specific roles in those tissues. However, in the presence of overlapping expression, there must be additional signals or distinctions that allow them to exert distinct phenotypes rather than functioning strictly redundantly. While the breadth of these differences are not yet known, one of the major aspects that have been studied is in the dynamics of nuclear import and export across the different isoforms (Chow et al., 1997; Rinne, Banach, and Blatter, 2009; Ulrich et al., 2012; Yissachar et al., 2013; Kar and Parekh, 2015; Kar et al., 2016). Each isoform also has a range of different spliceoforms that can modulate activity (Vihma, Pruunsild, and Timmusk, 2008; Mancini and Toker, 2009). For instance, NFATC2 can be spliced into at least three functionally distinct isoforms with different abilities to activate transcription with one being inhibitory toward the others (Chuvpilo et al., 1999). These levels of regulation are sure to be more complex than we presently understand

and offer another avenue of fruitful future study.

The NFAT isoforms share a common DNA binding motif (5'-GGAAA-3'), which enables them to exhibit the redundancies previously described (Rao, Luo, and Hogan, 1997; Chen et al., 1998). However, these isoforms may also have distinct DNA binding motifs to which they bind more efficiency than others, which can offer an additional layer of regulation. For instance, NFATC2 can bind with greater affinity to the sequence 5'-C^mGGAA-3', which will comprise some, but not all of the consensus sequences (Ray et al., 2021). Differential binding affinity for the other isoforms has not yet been exhaustively detailed, but future studies are likely to uncover preferences among them for neighboring nucleotides around the consensus sequence. An additional layer of regulation is via posttranscriptional modifications, including SUMOylation, which has been noted for NFATC1 and ubiquitination, which has been described for NFATC4 (Fan et al., 2008; Xiao et al., 2021). These modifications, among others, are likely to be present on other isoforms and to influence the overall NFAT transcriptional response although they have not been well characterized in general.

A further layer of relatively understudied biology is the aggregation of NFAT around RNA scaffolds in the cytosol, making them RNA binding proteins. This description is consistent with our immunofluorescence observations of perispeckled NFAT foci in the cytosol of THP-1 macrophages (in Subsection 3.3.10) and makes for an interesting additional layer of potentially variable regulation (Sharma et al., 2011). Knowledge as to whether or not different NFAT isoforms bind to different RNA scaffolds or are differentially regulated by that binding would add to our understanding of the complex mechanisms that allow some to be activated while other remain inactive within the same cells.

A quartet of foundational studies set the stage for some of the mechanisms by which the different NFAT isoforms can be regulated by distinct mechanisms. Ulrich et al. (2012) observed that NFATC3 and NFATC4 displayed distinct activation kinetics in hippocampal neurons, with NFATC3 rapidly activating while NFATC4 slowly activated over a longer period of time. This interestingly made NFATC3 the functionally dominant isoform and knockdown of this single isoform abrogated most of the responses even in the presence of NFATC4. This effect was thought to be mediated by different sensitivity of the isoforms to rephosphorylation by GSK3 β . Yissachar et al. (2013) compared the kinetics and dynamics of NFATC2 and NFATC3 and found that, while NFATC3 exhibited rapid nuclear localization and rapidly shuttles in and out of the nucleus over time, NFATC2 exhibits a slower response that steadily increases over time with increasing signal duration. While these distinctions had been grossly noted for some time (Chow et al., 1997; Rinne, Banach, and Blatter, 2009), the precise mechanisms were unclear. A later study by Kar and Parekh (2015) found that the precise cellular location of the Ca²⁺ flux resulted in different effects; NFATC2 depended on a close association with ORAI1 channels while NFATC3 also required high nuclear calcium for sustained activity; this rise in nuclear calcium depends on inositol triphosphate receptors on the nucleus (Kar et al., 2016). These interesting layers of regulation are thought to be mediated by differences in the SP3¹¹⁴ motif that governs nuclear import and export and which is the target of both calcineurin and NFAT kinases. As a layer of additional subtlety, it appears that these differences in kinetics can vary between different cell types, as NFATC2 translocates quickly in neurons while NFATC4 exhibits relatively slow translocation (Vihma et al., 2016). These effects may, in part, be regulated by different cytosolic compartments utilized

¹¹⁴NFAT proteins have a series of serine-proline (SP) phosphorylation sites in the N-terminal domain that regulate its nuclear localization. One of these, the SP3 site, seems to be of additional importance in this process. See Figure 1.4.

by the resting NFAT isoforms and their proximity to various calcium channels and calcineurin (Ulengin-Talkish and Cyert, 2022). The degree to which findings in particular tissues can be generalized to others is not yet known and may be difficult to assess until further studies have done more exhaustive comparisons (perhaps in T cells or uterine epithelia, where all of them are expressed to some degree).

A last area of potential future study is on the descriptively named NFATC2 interacting protein (*NFATC2IP*). This protein has few described roles (Sun et al., 2018; Xu et al., 2021b; Huang et al., 2022) despite having been found to interact with NFATC2 and uncovering the specific roles that it is playing in modulating the activity of NFATC2 or other NFAT isoforms would offer a further layer of regulation to NFATC2 that is likely to influence its ability to induce specific responses and may play some role in the kinetics of NFATC2 observed previously (Yissachar et al., 2013; Kar and Parekh, 2015; Kar et al., 2016). Investigating *nfatc2ip* knockout zebrafish is likely to identify important roles for this protein in mediating the overall course of infection and should offer an opportunity to expand our knowledge of this pathway in the pathogenesis of tuberculosis.

1.4.3 Activity of NFAT in Myeloid Populations¹¹⁵

NFAT was discovered relatively early on to be one of the major and defining responses to CLR activation. Early studies had identified a novel SYK-dependent pathway that was required for transcriptional responses to zymosan through DECTIN-1 (Rogers et al., 2005). This work set the stage for the discovery of the signaling mechanisms downstream of SYK activation that were required to induce immune responses. CARD9

¹¹⁵For additional reviews on this subject, see J. Fric et al. (2012b). “NFAT control of innate immunity”. In: *Blood* 120.7, pp. 1380–9 and K. Bendickova, F. Tidu, and J. Fric (2017). “Calcineurin-NFAT signalling in myeloid leucocytes: new prospects and pitfalls in immunosuppressive therapy”. In: *EMBO Mol Med* 9.8, pp. 990–999

was discovered first as a master regulator of CLR-dependent anti-fungal signaling (Subsubsection 1.3.3) (Gross et al., 2006; Hara et al., 2007), but this was not long to be the only pathway engaged by DECTIN-1. Defined by Goodridge, Simmons, and Underhill in 2007 as an important response mechanism, NFAT activation has been co-opted over the years as an experimental tool to measure CLR activation because TLRs do not¹¹⁶ activate NFAT (Yamasaki et al., 2009; Ishikawa et al., 2013; Furukawa et al., 2013; Richardson and Williams, 2014; Hattori et al., 2014).

By using either NFAT proteins fused to fluorescent proteins to monitor nuclear localization or the DNA regulatory elements for NFAT to drive luciferase or GFP from a minimal promoter, it is possible to capture a report of NFAT activation with high sensitivity or with rapid response times (Aramburu et al., 1998; Chow, Rincon, and Davis, 1999; Jauliac et al., 2002; Wilkins et al., 2004; Gwack et al., 2006; Colella et al., 2008; Rinne, Banach, and Blatter, 2009; Kar and Parekh, 2015; Kar et al., 2016). This has been used dozens of times in the literature to define the specificity of a response for a particular receptor and ligand. Despite the ironic ubiquity of this approach as experimental tool, very little subsequent work has been done to define the functional consequences of NFAT activation downstream of CLR activation, especially in the specific context of MINCLE or MCL agonism. Given the specificity of the NFAT response, there must be important biological consequences of this pathway being activated during infection, but these have been broadly neglected.

¹¹⁶This is presented as the dogma in the literature although it warrants some degree of elaboration. Zanoni et al. found that CD14, the TLR4 co-receptor for LPS, induced the activation of NFAT in dendritic cells that was independent of TLR4, MYD88, and TRIF, suggesting TLR-independent signaling mechanisms govern NFAT activation in this context while Minematsu et al. found that NFATC3 and NFATC4 were required for TLR responses in macrophages. Notably, both CD14 and DECTIN-1 localize to lipid rafts and recruit PLC- γ 2, offering a potential unifying model on the mechanisms of action at play here (Xu et al., 2009b). On the other hand Kang et al. found that NFAT activation was a negative regulator of TLR responses. As previously summarized, the bulk of the evidence is that TLRs themselves cannot directly activate NFAT although co-receptors and other effects may be able to engage this pathway (Zanoni and Granucci, 2012).

An early study found that NFAT was required for the induction of EGR2¹¹⁷, EGR3, M-CSF, COX2, IL-2, IL-10, IL-12, and myeloperoxidase transcriptional responses and prostaglandin E2 production in response to zymosan through DECTIN-1 (Goodridge, Simmons, and Underhill, 2007). Conversely, TNF- α and IL-6 were independent of NFAT in this context and instead dependent on TLR2 and MYD88. These functional distinctions between the different pathways that could be engaged by a single ligand are important for determining these relative contributions and more selective therapeutic targeting of particular pathways. Activation of NFAT is critical for control of various fungal infections through both direct myeloid activation as well as important priming of the adaptive immune system through induction of TNF- α , IL-2, IL-10, and IL-12. Interestingly, it seems that genetic inhibition of NFAT is sensitizing to a greater degree than pharmacological treatment, likely due to the antifungal activity of calcineurin inhibitors and lack of NFAT among fungi, which possess a distinct transcription factor, *Crz1*, downstream of calcineurin (Onyewu et al., 2004; Sugita et al., 2005; Xu et al., 2009c; Lev et al., 2012; Thewes, 2014; Herbst et al., 2015; Zelante et al., 2017; Yang et al., 2022).

Classically, the TNF-TNFR signaling pathway induces transcriptional responses primarily through NF- κ B, although it is understood that other transcription factors can mediate downstream responses through TNFR. One of these, intriguingly, is the NFAT signaling pathway, which has been shown to be induced in macrophages through NFATC1 in a manner reminiscent of that seen in osteoclastogenesis (Yarilina et al., 2011). This offers the opportunity for TNF- α to feed-forward by driving its own transcription through NFAT. Given that these experiments were conducted under chronic TNF- α stimulation, this may be uniquely relevant to the biology of the

¹¹⁷EGR2 is required for the development of natural killer T cells, so this is another mechanism whereby myeloid NFAT responses can drive adaptive immunity (Lazarevic et al., 2009).

granuloma where TNF- α is a major component of the cytokine milieu and is critical to host defense against mycobacteria.

A recent review has highlighted some of the known roles for NFAT activity in neutrophils in mediating important protective immune responses (Vymazal et al., 2021). This review posits that neutrophil NFAT signaling is a master regulator of neutrophil responses through the detection of various microbial ligands and subsequent induction of protective responses, most notably during fungal infection and after exposure to LPS through CD14. This review focused on the consequences of NFAT inhibition via calcineurin inhibitors on neutrophil function and concluded that suppression of neutrophil functions may increase patient susceptibility to various opportunistic infections, although these models struggle to completely differentiate the impact on different cell populations (Herbst et al., 2013). This is nonetheless an important consideration and makes NFAT targeting perhaps most appealing in the context of excessive inflammation. The study of these pathways within opportunistic fungal pathogens also in some ways limits the scope of these studies; while these are important and relevant pathogens in people receiving these immunosuppressive therapies after solid organ transplant, this may not reflect the behavior that would occur at lower doses or in otherwise healthy people.

Furthermore, there is somewhat of an NFAT renaissance occurring in the literature at the time of writing. Several new papers have emerged in the past several months identifying novel new roles for NFAT signaling in a variety of (predominantly hematopoietic) tissues, giving new emphasis to this long-neglected pathway (Deerhake et al., 2021; Poli et al., 2022; Peuker et al., 2022). This work will be more thoroughly reviewed in the next section, Subsection 1.4.4. My work discussed in later chapters adds to this body of NFAT-dependent responses and, hopefully, encourages

additional future work to define the roles of this important but understudied pathway in the response to not only tuberculosis but the full range of human diseases that engage CLR signaling, especially fungal diseases and additional autoimmune disorders (Brewer et al., 2022b). This pathway has been neglected for some time due to a focus on CARD9-dependent signaling, but has now come back into the spotlight for its important roles in mediating diverse aspects of immunity.

1.4.4 The NFAT Renaissance

As previously alluded to, a number of major papers have been published in the past few months assigning new roles to NFAT, with many of them specifically in cells of the hematopoietic lineage – neutrophils, macrophages, and platelets. The last major review of the role of NFAT in these cells was published by Fric et al. in 2012 and, compared to now, the picture was rather broad in the roles that NFAT plays in mediating response to various conditions. At the time it was understood that NFAT plays an important role in the development of myeloid cells, where it inhibits their differentiation; inhibition of NFAT signaling increased the percentage of cells that differentiate into the monocyte/dendritic cell lineage vis-à-vis neutrophils. While these roles for NFAT in development have important implications for the clinical use of NFAT inhibitors, they do not address the role of NFAT in these cells after development. Seminal early findings connected the NFAT signaling pathway to the signaling response to different PAMPs, including β -glucan and zymosan via DECTIN-1 in macrophages and dendritic cells and LPS via CD14 exclusively in dendritic cells (Goodridge, Simmons, and Underhill, 2007; Zanoni et al., 2009). NFAT also has a number of role in the microbicidal activities of neutrophils and phagocytosis in macrophages. NFAT inhibition in the absence of adaptive immunity was known to increase susceptibility to fungal infections via blockade of the expression of anti-

inflammatory cytokines.

Probably the first truly modern, comprehensive investigation of the role of NFAT in the innate immune response was published in late 2021 by Deerhake et al. This study, concurrent with the present work, evaluated the role of DECTIN-1 signaling in the pathogenesis of experimental autoimmune encephalomyelitis (Deerhake et al., 2021) here at Duke. DECTIN-1 expression was protective from the disease, but CARD9-dependent signaling exacerbated disease progression, demonstrating important roles for NFAT signaling in protection. Use of both genetic and pharmacological methods in a range of cell types found that myeloid cells stimulated with DECTIN-1 agonists drive activation of NFAT to produce neuroprotective cytokines, including oncostatin M and, importantly for this work, VEGFA. This modern, dataset-driven analysis is a key resource for understanding the spectrum of NFAT-dependent phenotypes induced by C-type lectin receptors and, while focused on one particular ligand, remains evocative of the potential for C-type lectin receptors to induce pro-angiogenic signaling. In their context, primary neutrophils were far more responsive to curdlan stimulation than were bone-marrow derived macrophages, although as has been discussed in Subsubsection 1.3.3, unprimed macrophages have overall dampened responses and different priming approaches may have provided differing results.

The study by Poli et al. (2022) is exceptionally interesting as it studied the role of NFAT in platelets which, in mammals, are anucleated. Thus, this work studies the influence of NFAT on mediating cellular responses in the absence of transcription. Inhibition of NFAT in platelets using transgenic VIVIT expression exacerbated Gram-negative sepsis by inducing hypercoagulation, in part through interactions with neutrophils to induce neutrophil extracellular trap release. NFATC2 is dephosphorylated by activation of the thrombin receptor, which then inhibits activation of

integrin α IIb β 3. Inhibition of NFAT leads to hyperactivation of integrin α IIb β 3 and increased coagulation of platelets. This increased integrin activity also increases platelet binding to circulating neutrophils; NFAT inhibition also leads to increased platelet degranulation, a marker of coagulative activity and inflammation and drives the recruitment of additional leukocytes. Use of a platelet-specific VIVIT mouse demonstrated a localized effect specific to platelets that drove increased inflammation in a model of Gram-negative sepsis. This work is interesting in the context of previous findings on the role of ITAM-linked receptors on platelets, which activate SYK, a known upstream activator of NFAT (Manne et al., 2015). The precise mechanism here remains unknown – what interacting partners could an NFAT protein have in an anuclear cell to mediate these activities? Such potential was alluded to by Du et al. (2020) and Du et al. (2021) although differentiation of these roles is challenging in nucleated cells, making the specific application of platelets both a biological and technical innovation. An exploration of the protein-protein interaction network of NFAT proteins could reveal interesting new biology applicable to nucleated cells as well; the function of this ostensible transcription factor in other realms of biology offers interesting opportunities to more specifically target those aspects of NFAT to modulate its activity. Clearly, new tools and deeper understandings of NFAT protein topology will be required to differentiate these classes of functions in these cells. This work highlights the potentially *pro*-inflammatory nature of NFAT inhibition (a potential benefit for those taking NFAT-inhibitory drugs for organ transplants) and a potential means to increase blood clotting responses for those with clotting disorders.

Peuker et al. (2022) found that deletion of calcineurin in myeloid cells was able to reduce the size of colorectal tumors in a mouse model in a manner dependent on microbiota-mediated activation of NFAT. NFATC1-dependent expression of IL-6 from macrophages induced the tumoral expression of two genes that negatively regulate

T cell activation, B7H3 and B7H4; treatment of antibiotic-treated mice with LPS or myeloid-VIVIT expressing mice with IL-6¹¹⁸ is able to restore the expression of these proteins within the tumor and promote tumor growth. Through selective ablation of particular immune cell populations, this effect was isolated to the response to neutrophils. However, this publication utilized some evidence of a role for MYD88 in contributing to this phenotype to attempt to link TLR4-MYD88 activation to NFAT activation, despite no other publication having ever made this argument and repeated historical demonstrations that TLRs could not activate NFAT and actually antagonized NFAT activity¹¹⁹. The reality in this situation is likely cooperative roles for NFAT and MYD88-mediated NF-κB pathways rather than a common signaling origin; this is especially notable for the studies testing various TLR agonists on the effect on IL-6 expression, which may be additionally sensitive to these sorts of combinatorial effects where both LPS (TLR4 ligand) and Pam3CSK4 (TLR1/2 ligand) induce IL-6 in an NFAT-dependent manner. Both LPS and Pam3CSK4 could be mediating these effects directly through CD14-NFAT signaling, but this hypothesis was not tested in this instance; it would certainly be of some interest to investigate whether these described phenotypes are dependent specifically on CD14 or if there are yet further mechanisms at play.

¹¹⁸The role of IL-6 in this context is intriguing given the described role for IL-6 in the induction of M2d macrophages, which prolifically produce VEGFA. Study of this polarization state during these sorts of infections is one way to get a more complete profile of macrophage responses within the traditional M1-M2 schema (Huang et al., 2018).

¹¹⁹One study, Liu et al. (2008a) argued for an NFAT-dependent cardiac response downstream of LPS, but did not explore a precise mechanism; CD14-mediated signaling seems likely but further study would need to be done.

1.5 Angiogenesis

Angiogenesis is a fundamental process in the development of vertebrate animals, which require the elaboration of blood vessels into all of the corporeal tissues in order to supply them with oxygen and structurally essential signaling cues (Adams and Alitalo, 2007; Carmeliet and Jain, 2011). This process is an elaborate coordination between the endothelium itself, stimulating cells (fibroblasts, somites, macrophages, and cancer cells among them), the surrounding connective tissue, and mural cells that maintain the vasculature (Armulik, Genove, and Betsholtz, 2011; Stratman et al., 2017). This process also requires an intricate series of intracellular signaling events within endothelial cells to direct these processes of growth (but not too much) and directionality (Simons, Gordon, and Claesson-Welsh, 2016).

Much of what is known about angiogenesis was first established in the optically transparent and genetically tractable zebrafish model, which allows for very fine observation of the very earliest processes of blood vessel formation (Chan et al., 2002; Chavez et al., 2016). The initial stages of this are known as vasculogenesis, in which endothelial cells as such differentiate from hemangioblasts early in development (Vogeli et al., 2006; Bertrand and Traver, 2009). Once a basic vasculature has been established, it can be remodeled and extended upon by the process of angiogenesis (Koch and Claesson-Welsh, 2012). Much of this process is regulated by a single pathway – the vascular endothelial growth factor A/vascular endothelial growth factor receptor 2 (VEGFA/VEGFR2) signaling axis (Olsson et al., 2006; Chung and Ferrara, 2011; Villanueva, 2016). Despite the seeming simplicity in this, there are additional layers of regulation at every conceivable level. The detection of VEGFA by an endothelial cell sets off signals that drive proliferation and growth toward the

source of the chemokine, which can be a variety of cell types, including macrophages (Olsson et al., 2006). These tip cells sprout along the VEGFA gradient and regulate the cells behind them as a mechanism to prevent excessive branching¹²⁰ (Tammela et al., 2011; Sakabe et al., 2017). In the absence of both of the *VEGFA* homologs in the fish, *vegfaa* and *vegfab*, the somitic vasculature is completely absent and only the dorsal aorta, the caudal vein, and other vasculogenic vessels are able to form through VEGFA-independent processes (Liang et al., 2001; Covassin et al., 2006; Bahary et al., 2007). Notably, these phenomena are profoundly difficult to study in the mouse, which must overcome tissue hypoxia as an environmental stressor from very early stages of embryonic development while the externally developing larval zebrafish are perfused with oxygen until many days post fertilization. To complete the growth of normal blood vessels, the stalk of the angiogenic vessel must open a lumen, a process that is still not well understood but is known to be mediated, at least in part, by macrophages (Gerri et al., 2017).

This entire process must be finely regulated to maintain vascular homeostasis. Too much angiogenesis is clearly bad, as seen in cancer, but even in normal tissues, vascular hyperproliferation can result in retinal disorders and psoriasis (Gupta and Zhang, 2005; Bisht, Dhasmana, and Bist, 2010; Malecic and Young, 2017). Endothelial cells utilize a variety of downregulatory mechanisms to block excessive angiogenesis, including apoptosis and transcriptional repressors (Stefanec, 2000; Chavakis and Dimmeler, 2002; Dimmeler, Haendeler, and Zeiher, 2002; Garcia-Barros et al., 2003; Duval et al., 2003; Ben Shoham et al., 2012). Thus, the weight of pro-angiogenic factors must exceed that of the inhibitory factors in order to induce a productive net

¹²⁰This is highly analogous to the ways in which plants regulate their growth. In plants, the growth tip secretes hormones along a gradient that prevents the activation of new growth points until the hormone concentration drops sufficiently low. This facilitates directional growth until the plant is large enough to warrant further branching.

angiogenic response.

These same fundamental processes are at play during pathological angiogenesis as these processes are similarly regulated by VEGFA/VEGFR2, but depending on the cause of the pathology, are notably disordered. While developmental angiogenesis is orderly and ultimately results in the formation of “normal” blood vessels, pathological angiogenesis often lacks the fine-tuning needed to form stable vessels with high integrity and are thus leaky, highly branched, and lack lumen formation. During these events, a much more complex and chaotic set of signals are at play which influence the endothelial biology of the region; ideally, the course of the response to the perturbation would restore homeostasis and facilitate vascular normalization and, eventually, regression. Additionally, the endothelium itself is a major source of signaling that supports tissue homeostasis and responses, a role only recently appreciated (Amersfoort, Eelen, and Carmeliet, 2022) but long understood by proxy of the *cloche* mutation (Stainier et al., 1995; Vogeli et al., 2006). The vast degree of specification which the endothelium undergoes based on its surroundings is a key aspect of endothelial cell biology which can be difficult to study, much like modeling tissue-resident macrophages can be nearly impossible *in vitro*. Nevertheless, these roles have been increasingly dissected through the use of single cell transcriptomics to evaluate the diversity among inter- and intra-tissue endothelium, revealing so-called unexpected levels of heterogeneity in these tissues (Chavkin and Hirschi, 2020).

1.5.1 Angiogenesis in Cancer

Tissue perturbations, such as those caused by wounds, often drive the invasion of blood vessels toward the site of insult as a mechanism to facilitate tissue repair. However, angiogenesis can serve as a maladaptive response in many contexts. Most

famous of these is in tumor biology, where these vessels serve as a supply of oxygen and glucose, a route of dissemination to distal sites, and a paradoxical barrier to the effective delivery of curative chemotherapeutics (Park et al., 2016; Yoncu et al., 2017; Fujita and Akita, 2017). One of the foundational observation in solid tumors was that they were often encased in vascular webs that differed substantially from those seen in neighboring, unaffected tissues (Chung and Ferrara, 2011). The size and cellular density of these tumors created a hypoxic environment at the center of the tumor that drove the activation of HIF-1 α and subsequent transcriptional responses geared toward the alleviation of this hypoxia; this pathway is also key in the metabolic switch in cancer that drives the Warburg effect (Mesange et al., 2014; Masoud and Li, 2015; Courtney et al., 2015). One of the most efficient means of alleviating cellular hypoxia is to shorten the distance between the cell and arteries, which are able to supply the cell with oxygenated blood. Under homeostatic conditions, no cell is more than 1 millimeter from the nearest blood vessel (Weber, Ju, and Borner, 2020). Tumors are thus under profound selective pressure to increase the availability of oxygen or perish and do so via upregulation of VEGFA signaling both endogenously and via tumor-associated macrophages (Riabov et al., 2014; Napione, Alvaro, and Bussolino, 2017). However, the deregulated transcriptional responses of these tumors drive the production of new blood vessels that fail to properly mature and remain highly permeable to blood solutes while adapting to and influencing the tumor microenvironment (Qian et al., 2009; Carlson et al., 2021). This vascular permeability is advantageous to the tumor by allowing glucose and other nutrients to diffuse out of the vessels and into the tumor microenvironment (Park et al., 2016). Such leakiness would logically appear to be a boon to chemotherapeutic delivery, but actually serves as a monumental barrier to achieving sufficiently high drug concentrations at the site of the tumor to reach an effective dose (Goel, Wong, and Jain,

2012; Datta et al., 2015; Yonucu et al., 2017).

In the transition toward chemotherapeutic options with lessened toxicity, a number of kinase inhibitors and monoclonal antibodies were developed that target a specific receptor on those blood vessels required for their growth and maintenance: the vascular endothelial growth factor receptor 2 or VEGFR2 (Potente, Gerhardt, and Carmeliet, 2011; Shibuya, 2011; Welti et al., 2013; Ranieri et al., 2014). This tyrosine kinase receptor triggers a downstream transcriptional response cascade that results in endothelial proliferation and directed growth toward the source of the ligand, VEGFA¹²¹. By inhibiting either the enzymatic activity of the receptor using kinase inhibitors or blocking the interaction between the receptor and the ligand using monoclonal antibodies, effective regression and normalization of the vascular webs around tumors can be achieved.

1.5.2 The Relative Failure of Bevacizumab

These factors and others led to active development of vascular targeting therapies throughout the 1990s and early 2000s, which culminated in the approval of bevacizumab to treat metastatic colon cancer in 2004, only 15 years after the discovery of VEGFA (Leung et al., 1989; Ferrara et al., 2004; Muhsin, Graham, and Kirkpatrick, 2004; Gordon and Cunningham, 2005; Kabbinavar et al., 2005; McCormack and Keam, 2008). Bevacizumab is a humanized monoclonal antibody targeting VEGFA, blocking ligand-receptor binding, VEGFR2 activation, and resultant angiogenesis (Keating, 2014). The promise of bevacizumab was to adjunctively enhance the effi-

¹²¹VEGF, or VEGFA, is the “canonical” VEGFR2 ligand, but there are four independent VEGF genes (VEGFA-D), each with different affinity for different VEGFRs (VEGFR1-3) and with different properties. VEGFB modulates VEGFA signaling in angiogenesis, while VEGFC and VEGFD are lymphangiogenic chemokines, which stimulate the growth of lymphatic vessels. This process remains relatively unstudied in the context of mycobacterial infection (Harding et al., 2015), but seems likely to play important roles in regulating the course of infection.

cacy of other drugs while starving the tumor of needed oxygen – a theoretical one-two punch at the cell biology of these tumors. However, the story ultimately proved more complicated and bevacizumab has been only a limited success and other vascular-targeting therapies have fared little better. Bevacizumab is a humanized monoclonal antibody that very potently ($K_D=1.1$ nM) blocks the interaction between VEGFR2 and VEGFA and induces vascular regression by binding and sequestering VEGFA (Papadopoulos et al., 2012; Yang et al., 2014b). This therapy has become standard of care for a subset of tumor types¹²² and physiological locations, but the mystery remains why this therapeutic strategy targeting a highly conserved, indeed nearly ubiquitous (Donnem et al., 2018), feature of tumors is not more broadly applicable and generally successful (Van Meter and Kim, 2010). Such a therapy could, in theory, have been a universally beneficial adjunct through metabolic attack on the tumor coupled with improved drug delivery but proved to be comparatively limited. Initially approved only for metastatic colorectal cancer, it began to accumulate other indications for some time, until further trials in certain cancers found limited efficacy over the long term and relapsed cancers often demonstrating resistance and increased aggressiveness through poorly studied reprogramming mechanisms (Rivera and Bergers, 2015; Itatani et al., 2018).

It seems that the physiological stress of hypoxia appears to drive upregulation of compensatory pathways within the tumor itself – the escalating hypoxia in the local region drives rapid amplification of VEGFA and PIGF production to alleviate such detrimental hypoxia (Alidzanovic et al., 2016; Gardner, Madu, and Lu, 2017). By this mechanism it is proposed that tumors increase the local concentration of VEGFA beyond the stoichiometry and binding affinity between bevacizumab and VEGFA or

¹²²Colorectal cancer, non-small cell lung cancer, breast cancer, ovarian cancer, and cervical seem to be susceptible to inhibition (Kazazi-Hyseni, Beijnen, and Schellens, 2010; Garcia and Singh, 2013; Tewari et al., 2014; Botrel et al., 2016; Gridelli et al., 2018; Baraniskin et al., 2019).

induce other growth factors (FGF- β and VEGFC among them) to promote vascular relapse and renewed angiogenesis toward the site in a return to *status quo ante* (Zhao et al., 2017; Michaelsen et al., 2018; Haibe et al., 2020; Montemagno and Pages, 2020; Zahra, Sajib, and Mikelis, 2021). These therapies can also induce extracellular matrix remodeling in ways that actually block effective drug delivery (Rahbari et al., 2016). Tumors, through the secretion of such a range of different factors, also coopts the vasculature itself and turns the vasculature into a “malignant” state where it will grow even in the absence of stimulus due to the environment of the tumor itself (Frentzas et al., 2016). Another facet of VEGFA signaling in cancer that has become more recently appreciated is the autocrine role of VEGFA production in stimulating tumor cell proliferation; just as endothelial cells proliferate in response to VEGFA, so too do some cancer cells (Goel and Mercurio, 2013; Ntellias et al., 2020). While this should add yet another putatively beneficial aspect to angiogenesis-targeting therapies, the reality is much more complex.

Perhaps the most interesting of the stories of attempted use of bevacizumab is in glioblastoma. Despite having been given FDA approval for use in recurrent glioblastoma and initial studies having shown some promise (Segerstrom et al., 2006), there was little evidence of long-term efficacy until further meta-analyses were conducted enabling greater understanding – it was found that treatment was only mildly successful and that withdrawal led to an increase in tumor vasculature and may alter certain aspects of the biology of the tumor to host detriment (Iwamoto et al., 2009). Later cell biological studies found that treatment of tumors with bevacizumab decreased tumor-associated vasculature but actually increased tumor invasiveness into the parenchyma, much of which is mediated by increased HIF-1 α activity (Keunen et al., 2011). A truly proper double-blind clinical assessment of the potential use of bevacizumab in glioblastoma was finally conducted in 2014 found either no improvement

(Gilbert et al., 2014) or a mild benefit (Chinot et al., 2014). The general conclusion is that tumors are initially responsive to bevacizumab but most will progress within months with potential increases in invasiveness and dissemination at that point – whether bevacizumab *per se* alters the tumor in such a way to make it more aggressive is unclear (Li et al., 2017). One potential explanation is through induction of an epithelial-to-mesenchymal transition that encourages proliferation and cellular motility to other sites. Additionally, the tumors express other pro-angiogenic factors able to compensate for the loss of VEGFA signaling, including PIGF, angiopoietins, CXCR4 (Xu et al., 2009a), FGF (Tamura et al., 2017), and VEGFC (Villefranc et al., 2013; Zhao et al., 2017; Michaelsen et al., 2018). Overall, bevacizumab treatment of glioblastoma seems to offer a slight advantage to progression-free survival but no advantage whatsoever to long-term survival; the progression-free survival is likely through the immediate improvements seen radiologically after initiation but recurrence then compensates for any temporary advantage (Wick et al., 2017; Zhan et al., 2019).

New approaches to targeting tumor-associated vasculature are clearly needed (Cesca et al., 2013; Hara et al., 2016). Vascular targeting as a theme has clear potential as an adjunct to other therapies, but the current approaches are limited in their utility (Neri and Bicknell, 2005). As mentioned previously, one approach would be to target VEGFA production at the root – by blocking transcription factors needed to induce VEGFA. Additional approaches would be to target axillary pathways also important for vascular endothelial cell biology in tumors – the interaction between angiopoietins and the TIE receptors is one primary possibility (Hato, Tabata, and Oike, 2008; Fujita and Akita, 2017). Other options include somehow artificially mimicking normoxia (with a chemical hydroxyl donor?) for the tumor to block HIF-1 α activation, if HIF-1 α is truly the dominant means of inducing VEGFA in this context.

Lastly, one of the major hazards of this tumor vasculature is the leakiness, which strongly contributes to tumor growth. By inducing vascular normalization, rather than regression, through a combination of VEGFR- and TIE2-targeting therapies, it may be possible to simultaneously improve chemotherapeutic delivery without inducing compensatory responses in the tumor. Alternatively, inducing vascular perfusion may also offer another anti-tumor strategy that improves chemotherapeutic delivery although the utility of this approach remains unclear (Rivera and Bergers, 2015).

Given the challenges associated with targeting the vasculature directly and compensatory pathways, many of which are induced by HIF-1 α , alternative approaches are needed to more effectively target vascular growth in tumors. HIF-1 α -directed therapeutic options remain functionally non-existent in 2022, with none having been approved for broad use and none directly targeting the activity of the protein itself (Sharma, Sinha, and Srivastava, 2022). All existing approaches impact HIF-1 α activity indirectly, by altering the activity of upstream regulatory factors or major downstream response pathways. HIF-1 α , aside from its role in inducing angiogenesis, exerts a panoply of pro-tumor effects, including promoting the epithelial-mesenchymal transition, which enhances the metastatic capacity of the cells and driving glycolysis in the Warburg effect (Courtney et al., 2015; Sharma, Sinha, and Srivastava, 2022). The most functionally promising drug candidates are actually those that *agonize* HIF-1 α ¹²³ and drive *increased* local angiogenesis, which is rather beneficial for a number of disorders, including major burns and diabetes (Dor et al., 2002; Atluri and Woo, 2008; De Rosa, Di Stasi, and D'Andrea, 2018). However, existing inhibitors through either direct or indirect mechanisms remain either impotent or excessively

¹²³This is typically by inhibiting the upstream prolyl hydroxylase enzymes that hydroxylate HIF-1 α in the oxygen-dependent degradation domain, resulting in its proteosomal degradation. In the absence of effective activity of these oxygen-dependent hydroxylases, HIF-1 α is able to remain active.

toxic *in vivo*. It has long been established that other transcriptional pathways¹²⁴ are important for the production of VEGFA and these may prove to be a more fertile ground for discovery of anti-angiogenic therapies that target required upstream factors.

A final point worthy of consideration on this topic is the nature of the vasculature itself in the periphery of the tumor. While much has been done to *physically* characterize these vessels, a comprehensive look at the biological ways in which these vessels differ from normal vasculature and how that might impact the spectrum of therapeutic options is lacking (Jambusaria et al., 2020). To my knowledge, there are no resources available that have characterized the vascular endothelium of solid tumors from humans and certainly nothing on the scale of the many genomic and transcriptomic atlases of various tumors and cell lines (Pepin et al., 2012; Kahn et al., 2021; Carlson et al., 2021). The lessons learned from such an effort would be of immense benefit the study of these tumors, developing more targeted therapies, and likely inform our understanding of the granuloma-associated vasculature as well.

1.5.3 Non-Angiogenic Functions of VEGFA

VEGFA is also able to exert a number of roles in regulating non-endothelial biology, a set of roles somewhat unappreciated in the literature at large. These early roles were first identified in hematopoietic stem cells, where VEGFA signaling is important for inducing proliferation and preventing apoptosis among this population (Gerber et al., 2002). Other roles for VEGFA have been identified in regulating cancer growth, as briefly discussed in Subsection 1.5.1 (Goel and Mercurio, 2013).

¹²⁴Including SMAD3/4, TGF- β , p300, and a series of AP-1 transcription factors including c-Fos and Jun-D (Marconcini et al., 1999; Gray et al., 2005; Jeon et al., 2007; Schmidt et al., 2007; Thangarajah et al., 2009; Nam, Park, and Kim, 2010; Kwon et al., 2012; Catar et al., 2013; Yoshitomi et al., 2021).

This phenomenon of VEGFA-VEGFR signaling inducing cancer cell survival and proliferation is widespread and may represent a general mechanism of cell survival by coopting the pro-growth, pro-survival signals normally used by the endothelium (Mercurio, Lipscomb, and Bachelder, 2005; Wiszniak and Schwarz, 2021).

A key role of VEGFA is in the recruitment of myeloid cells, which has been demonstrated in a number of contexts. Acting through these myeloid cells, further angiogenic effects can be exacted on the vasculature that are independent of vascular sensing of VEGFA directly (Cursiefen et al., 2004). There are an number of pro-angiogenic therapies in the works for disorders like myocardial infarction, burns, and diabetes, and these have been seen to induce the recruitment of inflammatory types of macrophages whereas anti-inflammatory macrophages can be recruited by FGF and produce VEGFA *in situ* (Lucerna et al., 2007). A model wherein VEGFA produced by “M2” macrophages recruits “M1” macrophages could present a feedback cycle that limits excessive angiogenesis through macrophage-macrophage interactions (Barbay et al., 2015). Macrophages have also been noted to enhance VEGFA-driven angiogenesis by tumor xenografts, suggesting potential roles for either VEGFA-mediated induction of VEGFA within macrophages in a feed-forward loop or direct macrophage-endothelial interactions (Britto et al., 2018).

VEGFA can also act directly on the macrophages to modulate their polarization state and influence control of tumor growth (Dineen et al., 2008; Incio et al., 2016; Zhang et al., 2021a). One study in particular found that the VEGFR2⁺ macrophages induces an immunosuppressive effect within the tumor microenvironment and inhibits macrophage-mediated killing of tumor cells. Given the levels of VEGFA found in the granuloma, similar effects may be present although expression of *kdr* and *kdrl*

are limited¹²⁵ (Cronan et al., 2021). By contrast, granuloma macrophages express high levels of *flt1* (VEGFR1), so the previously described effects of VEGFR1 on macrophages may be more relevant (Incio et al., 2016). These roles are comparatively underexplored in the context of infection, where the influence of VEGFR1 and VEGFR2 signaling within macrophages is likely to be a critical determinant of their bactericidal capacity.

Lastly, in the context of mycobacterial infections, VEGFA has been found to have important roles in regulating granuloma inflammation independent of its ability to induce vascularization. Harding et al. (2019) found that VEGFA is induced in granulomas in mice and that blockade reduces inflammation without compromising bacterial containment, even in this model system that lacks robust angiogenic responses to so-called granulomas. This is mediated by reduced macrophage recruitment to the site of infection potentially through P2RX7 activation (Matty et al., 2019). This study provides evidence of the importance of VEGFA in recruiting immune cells to the site of infection, although this recruitment does not manifest as alterations in bacterial burden. It is possible or probable that these effects may interact with alterations in vasculature that are able to be observed in other models of mycobacterial infection and should be accounted for in future studies.

1.5.4 Macrophage-Endothelial Crosstalk in Angiogenesis

As previously alluded to, macrophages are essential mediators of angiogenesis during development and in the context of cancer and various vascular disorders (Sunderkotter et al., 1994; Ribatti et al., 2007; Chung and Ferrara, 2011; Weis and Cheresh, 2011). This role has been widely appreciated for over fifty years and early studies

¹²⁵The nomenclature for zebrafish VEGFRs is somewhat confused due to a genome duplication and results in two copies of VEGFR2. For a nomenclature guide, see Bussmann et al. (2008).

demonstrated that tumor angiogenesis was inhibited by monocyte depletion, which also resulted in a decrease in tumor size (Polverini et al., 1977; Sunderkotter et al., 1994).

While much has been studied regarding the contributions of macrophages to angiogenesis during cancer, less has been established in the realm of macrophage contributions to developmental angiogenesis. So-called “M2” macrophages are known to contribute to angiogenesis as these reparative macrophages produce VEGFA during wound healing and in response to other insults (Jetten et al., 2014). This macrophage-mediated angiogenesis also appears to depend on IL-10 and TNF- α signaling, although the contributions of different cytokines is unclear (Leibovich et al., 1987; Nakamura et al., 2015). One area of interest is the ability for macrophages to physically engage the endothelium to mediate angiogenesis, similar to some of the previous descriptions of microglia stimulating and pruning neuronal synapses. In the developmental absence of macrophages, fewer vessels are able to sprout and connect in the brain (Fantin et al., 2010). Macrophages physically bring together sprouting tips of endothelial cells to facilitate new connections between tip cells and create new vascular connections¹²⁶ (Liu et al., 2016).

These processes can actually be mediated by macrophages of many different phenotypes, including so-called M1 macrophages. However, these different states of these macrophages induce distinct transcriptional changes in the endothelium itself, which would likely alter further macrophage behavior in the context of a native tissue. However, in engineered tissues, M1 macrophages induce early neovascularization while the other cell types were unable to do so (Graney et al., 2020). Unfortunately, this study was focused on the biology of the endothelium and did not profile the re-

¹²⁶This process, called anastomosis, is interesting in development, but beyond the scope of the present discussion.

sponses of the macrophages themselves, leading to important questions about what macrophage-derived factors were modulating these responses. In any case, this study suggests that it is not simply M2d macrophages that are able to induce angiogenesis within tissues.

The endothelium is a critical moderator of macrophage behavior and does so through the expression of important integrins and chemokines that constrain and activate macrophages. During insult (pathogenic, mechanical, thermal, or otherwise), the endothelium becomes an adherent surface for myeloid cells that drives their recruitment (Kalucka et al., 2017). Some of these junction proteins can be specifically induced by inflammatory signals, which allow circulating monocytes to bind and extravasate into the tissue. These proteins include a family of selectins as well as CDH5, PCAM, and ICAM (Imhof and Aurrand-Lions, 2004). The endothelium is also a source of CCL2, a potent myeloattractive chemokine, which, as we have seen, serves a detrimental role in host defense against tuberculosis (Martin et al., 2007; Cambier et al., 2014).

One of the ways that the endothelium and macrophages are known to physically associate is through TIE2-expressing macrophages remaining in contact with angiopoietin-2-expressing endothelial cells (De Palma et al., 2007; De Palma and Naldini, 2011; Baer et al., 2013). TIE2-expressing macrophages are pro-angiogenic cells that interact with the endothelium directly to provide paracrine signaling to the endothelium to facilitate angiogenesis. While these cells have been extensively explored in the context of tumors, where, as a subset of tumor-associated macrophages, they contribute to both angiogenesis and dissemination¹²⁷, they likely exist at homeosta-

¹²⁷This is now, interestingly, under dispute as it has been observed that in the absence of TIE2-expressing macrophages, angiogenesis and dissemination can both still occur. Thus, the role for these interesting macrophage subsets remains more obscure than ever; further studies will be required to reconcile these observations from the body of literature with these newer findings

sis and in more transient conditions as well and play important roles in regulating endothelial remodeling (Lewis, De Palma, and Naldini, 2007; Duran et al., 2021).

A study from Britto et al. (2018) found that during cancer, macrophages were responsible for physically guiding endothelial cells along the VEGFA gradient toward the tumor. This was independent of the macrophages' ability to produce VEGFA and abolition of macrophages even in VEGFA-expressing tumors resulted in impeded angiogenesis. This provides macrophages a direct role in physically guiding blood vessels toward the source of VEGFA, potentially through some regulation of the activity of the tip cell. These sorts of physical associations between macrophages and the endothelium are uniquely able to be dissected within the zebrafish model.

These physical associations are now known to be mediated by the activation of HIF-1 α in macrophages. HIF-1 α is essential for vascular development, but the underlying rationale was unknown. Gerri et al. (2017) found that macrophage expression of HIF-1 α mediated physical interactions between macrophages and the endothelium that enabled critical aspects of developmental angiogenesis and angiogenesis during wound repair; in the absence of HIF-1 α , macrophages are unable to interact with the endothelium and repair breaks in the vasculature. There is much fertile ground left to be explored on the HIF-1 α signaling targets that enable macrophages to engage with the vasculature, but this sets the foundation for these activities to exist during both developmental and pathological angiogenesis, a pattern long repeated in the history of the field.

(Jakab et al., 2022).

Tumor-Associated Macrophages

The most notable of these pro-angiogenic effectors is, of course, VEGFA. The process of tumor growth eventually requires the neovascularization of the tumor in order to continue growing and this process can be mediated by tumor-associated macrophages. However, it is not simply that tumor-associated macrophages express VEGFA, they also express a whole panoply of cytokines that, in a feed-forward loop, drive increased VEGFA expression and angiogenesis. These cytokines include IL-1 β , TGF- β , and M-CSF, all of which can induce further HIF-1 α activation in additional macrophages (Carmi et al., 2009; Nicholas, Oniku, and Sumbayev, 2010; Guo et al., 2016).

Macrophages, both monocyte-derived and tissue-resident, initially respond to growing tumors and act as antigen-presenting cells to present tumor antigens to the adaptive immune system that stimulate B cells to produce antibodies and T cells to develop into cytotoxic CD8 $^{+}$ T cells (Laviron and Boissonnas, 2019). While macrophages are initially protective against tumors, they transition into a more immunosuppressive, pro-tumor state after the initial failure to induce effective adaptive immunity and manipulation by the tumor microenvironment (Noy and Pollard, 2014). Tumor-associated macrophages are classically thought to be “M2-like” macrophages on account of their seeming indisposition toward killing the cancerous cells; they migrate into hypoxic regions of the tumor and, upon activation of HIF-1 α ¹²⁸, begin to secrete VEGFA and drive angiogenesis toward the tumor. While tumors can directly produce these pro-angiogenic factors, they are greatly aided by the complementary activity of these macrophages.

¹²⁸The ironic relationship between M1 polarization and HIF-1 α activity belies the notion that VEGFA is an M2-associated cytokine. Clearly there are more factors at play in this paradigm than HIF-1 α directly regulating VEGFA if VEGFA levels within so-called M1 macrophages are actually limited relative to M2 macrophages, where HIF-1 α activity is more limited (Sunderkotter et al., 1994; Carmi et al., 2009; Harding et al., 2019; Yu et al., 2021).

One of the means by which tumor-associated macrophages can facilitate changes in the tissues surrounding the tumor to facilitate invasion is by the production of matrix metalloproteases, which have well-known roles in degrading the extracellular matrix and facilitating angiogenesis. This loosening of the extracellular matrix enables tumor growth, increased macrophage recruitment, and the growth of endothelial cells (Lamagna, Aurrand-Lions, and Imhof, 2006). These matrix metalloproteases also play important roles in promoting mycobacterial growth, suggesting further parallels between these disease contexts (Volkman et al., 2010). This sets the stage for further pro-angiogenic factors to stimulate the extensive vascularization seen in many tumors and in tuberculosis.

1.5.5 Historical Observations of Angiogenesis in Tuberculosis

Histopathological characterization of tuberculous granulomas has served as the foundation of our knowledge of the structural features of tuberculosis disease. All of the major features were described on the basis of this histological analysis, including the necrotic core, the epithelioid macrophage layer¹²⁹, the lymphocytic cuff, and – important for this work – a surrounding web of vasculature.

In contrast to the normal lung, tuberculous lungs demonstrated dramatic aberrant vascularization in the proximity of the tubercle, arguing for some role of the disease in inducing this process; these vessels had even been observed to derive from the aorta, which is a remarkable subversion of the central circulation of the whole body (Cudkowicz, 1952). Such vascular re- and dis-organization was a notable radiological

¹²⁹This layer was known as such for decades prior to a proper characterization in Cronan et al. (2016). These cells pseudo- (or properly) differentiate into macrophages possessing notable epithelial characteristics.

and histological feature of the disease, but what role it may play was, as yet, unknown.

There was rife speculation on the role of this vasculature in the pathology of the disease. Bloody sputum had long been recognized as a primary differentiating factor between tuberculosis and other potential sources of lung disease, suggesting that vascular damage was a major consequence of cavitation and transition to active disease. While some viewed the angiogenesis as circumstantial – granulomas at sterile wounds surrounding foreign objects also become vascularized – others speculated that this may be a host-protective effect, allowing for the recruitment of additional immune cells to the site of the infection (Miller, 1938; Cudkowicz, 1952; Gale, 1957; Oehlers et al., 2015). There lingered the third possibility that rather than being either incidental or host-protective, that this effect may be a maladaptive response, serving to benefit the bacteria to the detriment of the host (Cudkowicz, 1952). Answers to this question would have to wait for quite some time as the tools to properly dissect the contributions of pathological angiogenesis to tuberculosis disease outcome remained many years in the future.

1.5.6 Modern Studies on Granuloma Angiogenesis

After a decades-long period of relative inactivity, a series of papers in the early-2000s began to explore mechanisms underlying the granuloma angiogenesis, although these early studies lacked effective tools to dissect this biology in the context of infection *sensu stricto*. A pair of early papers investigated the contribution of TDM to the induction of angiogenesis. The chronological first to be published, Sakaguchi et al. (2000) did extremely thorough work to dissect the chemokines required for angiogenesis and the cellular source of these factors using an air pouch model in the mouse along with complementary work *in vitro*. This study was interested primarily in the

potential for these purified mycolic acids to be used for pro-angiogenic effects, potentially for treatment of burns or diabetes, although little discussion is provided on this front. Saita et al. (2000) developed a rat corneal model of angiogenesis and applied purified TDM to induce an effective angiogenic response that depended on VEGFA and there was some speculation that TDM may contribute to the neovascularization effect seen in the proximity of tuberculosis granulomas in human patient samples, but little follow-up work was able to be done at that time. These efforts combined a targeted characterization of the cytokine response and piecewise inhibition of these factors to identify both VEGFA and IL-8 (CXCL8) as important pro-angiogenic factors caused by TDM. With relatively limited tools and the lack of an effective model, no work was able to be done to thoroughly characterize this angiogenesis phenotype in the context of infection or the consequences of blocking angiogenesis on disease progression.

Intermittent subsequent work had found important roles for angiogenesis-relevant host genes in tuberculosis pathogenesis, but the connection had not yet been drawn in full. For instance, a study found that endothelin signaling was important for host control of tuberculosis infection, pointing to a specific role of the vascular responses in the control of infection, even in the background of a C57BL/6 mouse (Correa et al., 2014). Because there is no obvious angiogenic effect in this background, these effects are likely mediated by physiological changes in the lung vasculature. Their contributions to vasoconstriction may, in some way, limit nutrient access to the site of infection although these mechanisms may be worthy of future exploration.

The need for a tractable model required innovative new approaches to studying this phenomenon. Classical mouse models fail to form epitheloid granulomas with associated vasculature and many other mammalian models lack the exhaustive set of

tools required for a thorough mechanistic dissection of this process. While a more comprehensive discussion of the zebrafish-*Mycobacterium marinum* model will wait for Section 2.2, this model offered a set of unique advantages – most notably in regards to genetic tractability and optical transparency – that enabled pioneering studies on the mechanisms underlying the angiogenic response and the functional consequences of inhibiting this process. This study from Oehlers et al. (2015) identified that growth of vasculature toward the site of infection increased bacterial burden and that inhibition of this process may serve as a useful host-directed therapeutic approach. Concurrently, Datta et al. (2015) developed a rabbit model of infection, which is a model in which granulomas form along with associated vasculature. This work sought to comprehensively characterize the vasculature and identified notable commonalities between the heterogeneous and structurally abnormal vessels seen in tuberculosis and those seen in solid tumor cancers, a historical expertise of Rakesh Jain’s group. The structural deficits of these vessels serve as a barrier to the effective delivery of small molecules because they leak out of the vessels and form a hazy corona around the granuloma without effectively penetrating it. Normalization and regression of these vessels with bevacizumab was able to improve effective small molecule delivery to the inner layers of the granuloma, suggesting that this may be an option for adjunctive therapy to improve drug delivery.

Shortly thereafter, Polena et al. (2016) interrogated granuloma angiogenesis as a potential mechanism of dissemination and, using Matrigel plugs impregnated with *M. tuberculosis*, identified an angiogenic response to infection in SCID¹³⁰ mice that depended on macrophage-derived VEGFA and which facilitated spread from the plug to distal organs. One of the challenges with these latter studies is that they only observe the functional consequences of angiogenesis inhibition for a limited period of

¹³⁰Severe combined immunodeficiency mice lack functional lymphocytes.

time (three or eight days in the Datta et al. (2015) study and two weeks in Polena et al. (2016)) and Polena et al. (2016) utilize a severely immunocompromised mouse model, where granulomas fail to mimic the human pathology and any protective role for the adaptive immune system is neutralized. More recently, Torraca et al. (2017) found a critical role for CXCR4, an important chemokine receptor, in driving angiogenesis in a manner that was independent of VEGFA production. While VEGFA is clearly a major determinant of angiogenesis, it is not the only possible determinant and its interaction with other signaling molecules may be important for the overall phenotype.

These studies also raised several important questions:

- How does VEGFA inhibition interact with the use of existing therapies? Can an additive or synergistic effect be achieved by combining the bacterial burden reduction effects seen in Oehlers et al. (2015) and Polena et al. (2016) with the improved drug delivery identified by Datta et al. (2015) in the context of standard-of-care anti-tuberculosis therapy?
- Does this vascularization effect extend to other granulomatous diseases? For instance, it is known that *Schistosoma* interacts directly with the endothelium, but does the granuloma response itself facilitate growth or transmission via modulation of the surrounding vasculature (Chaves, 1966; Shariati et al., 2011; Pereira et al., 2013)? Similarly, the fungal pathogens *Cryptococcus* and *Histoplasma* induce granuloma formation in the course of disease and the role for angiogenesis in these contexts remains unknown¹³¹ (Martidis et al., 1999; Coenjaerts et al., 2004). Conversely, sarcoidosis granulomas are avascular; what

¹³¹Given that both of these pathogens can disseminate hematogenously, there is reason to believe that blood vessel biology plays some role in the development of these diseases, although how it might be doing so is largely unknown (Kauffman, 2007).

functional distinctions might exist that discriminate between whether or not a granuloma sub-type is heavily vascularized or avascular given that macrophages play the defining role in all cases (Kambouchner et al., 2011).

- Pathogens must actively subvert host immunity in order to survive and transmit and have evolved sophisticated mechanisms for doing so. Given our knowledge that mycobacteria benefit from host angiogenesis, could it be the case that they have developed specific mechanisms for inducing this process?

This last point is notable given the previously discussed foundational findings on the ability of TDM to induce angiogenesis. Could the bacteria be actively modifying TDM in order to enhance the degree of vascularization? Observations from Sakaguchi et al. attempted to dissect some elements of this using TDM from *Rhodococcus* spp., but was unable to functionally differentiate these configurations. Additionally, Polena et al. utilized heat killed *M. tuberculosis* and, while their thesis was that VEGFA upregulation was dependent on ESX-1, found a substantial upregulation of VEGFA in the absence of active metabolism or structural integrity. These findings suggested a heat-stable factor independent of secreted effectors could mediate at least some percentage of the overall angiogenesis phenotype and that it might depend on TDM.

To this end, Walton et al. genetically and biochemically interrogated the mechanisms underlying the TDM-dependent angiogenesis phenotype. This work identified TDM as sufficient to induce angiogenesis using a novel application of the zebrafish model, wherein TDM emulsified in incomplete Freund's adjuvant is injected into the trunk of the larval zebrafish, mirroring the approach used in Oehlers et al. for live mycobacterial infection. Furthermore, this work identified *cis*-cyclopropyl modification of TDM as necessary for the induction of this phenotype, a process mediated by the bacterial enzyme PcaA. $\Delta pcaA$ *M. marinum* is attenuated for *in vivo* growth

due to deficient angiogenesis, a process that can be complemented in *trans* by co-infection with wild-type *M. marinum*. This work serves as the foundation of the subsequent work presented here as it left a major question as yet unaddressed: **what signal transduction pathways within responding macrophages are capable of detecting and responding to TDM to drive VEGFA production and downstream angiogenesis?**

1.6 Host-Microbe Interactions to Study Cell Biological Processes

Pathogens are nature's born cell biologists. For their survival, they depend on the coordinated subversion of host responses to infection, having to overcome the restrictive force of 1,000 interferon-stimulated genes, a fully equipped innate and adaptive immune system, and the physical architecture of the infectious focus (Byrne, Lehmann, and Landry, 1986; MacMicking, 2014). The literature is replete with examples how the study of the mechanisms that pathogens use to undermine the host sheds light on normal, homeostatic host processes including DNA replication, RNA splicing and modification, protein biosynthesis, and organelle regulation to cell-cell communication, critical cell receptors, secretion pathways, and post-translational modifications (Compans and Roberts, 1994). While much of the early work was conducted using viruses, modern understanding of cell biology synthesizes findings from across pathogens, including bacteria and fungi (Welch, 2015). This deepened understanding of cell biology can lead to the development of more efficacious treatments of infection through modulation of host responses, an approach known as host-directed therapy.

1.6.1 Host-Directed Therapies: History and Promise

One of the defining characteristics of tuberculosis infection is the formation of caseating granulomas. These granulomas, formerly known as tubercles¹³², are the most notable and ubiquitous pathology of human tuberculosis. These granulomas are a highly conserved immunological response to any object, pathogen or otherwise, that the immune system is unable to clear and are an imminently visible and clinically definitive manifestation of tuberculosis¹³³. For reasons that remain poorly understood, but likely related to the inflammatory biases of the C57BL/6 and other mouse models, these mice do not form granulomas¹³⁴ after being infected with *Mycobacterium tuberculosis* and mice do not harbor a species of *Mycobacterium* that infects them in the wild (Orme, 1998; Apt, 2011; Harper et al., 2012). This has set the mouse on an evolutionary trajectory where potentially adaptive (or maladaptive) responses to mycobacterial infection fail to occur. No matter the relative costs or benefits to the host of granuloma formation, the inability of any as yet known mouse model (with the partial exception of the C3H/FeJ model) to form granulomas compromises their ability to serve as a physiologically relevant model of some, but not all, aspects of human tuberculosis.

A major challenge has been the specific identification of diseases, stimuli, and biological consequences that drive angiogenic effects. While the angiogenic response

¹³²Hence, *tubercul*-osis.

¹³³A large body of work exists on the mechanisms that *Schistosoma* eggs use to induce parasite-beneficial granuloma formation. However, even in the absence of active biological induction of granulomas, sterile but indigestible objects will induce granuloma formation, albeit with some distinguishing characteristics (Boros, 2003).

¹³⁴Strangely, these mice do form granulomas in response to *Schistosoma* and other stimuli, suggesting something distinguishing about mycobacterial infection or the production of central necrosis and perhaps offering clues as to the unique characteristics of the tuberculous granuloma (Fu et al., 2012).

to tumors is thought to be mediated primarily through a hypoxia-dependent mechanism, the angiogenic response to other stimuli is far less homogeneous. For instance, in the context of the tuberculous granuloma, these structures initially form in the oxygenated environment of the human lung, which encounters 21% oxygen in air approximately 16 times per minute – not an environment that would generally facilitate a hypoxia response. While it is certainly possible in occluded or fluid-filled sites to create acute hypoxia, the angiogenic response within the lung would be assumed to rapidly and efficiently alleviate this stressor. No systematic comparison has been done to truly measure the precise oxygen tension in these granulomas from either humans or non-human primates, so it remains difficult to make sweeping assertions¹³⁵. Coarse assessments with tools like pimonidazole have been performed, but these are non-quantitative by their nature and impossible to effectively calibrate *in vivo* (Cousins et al., 2016). Regardless, the experimental identification of particular mycobacterial components able to induce angiogenesis suggests more sophisticated immunological mechanisms at play than simple hypoxia.

A bacteria-centric approach to treatment of tuberculosis seems logical, as bacteria possess many functions that humans lack entirely that are necessary for their pathogenicity, making these appealing targets for drugs. However, this opens the door to the emergence of resistance when treatment is unable to clear the infecting bacteria and a tolerant or resistant population then expands anew. This creates a compelling niche for a new approach to the treatment of chronic bacterial (and fungal and viral) infections: host-directed therapy. Host-directed therapies have long been used in cancer; indeed, anti-angiogenic therapy is one of the earlier examples

¹³⁵This is partially due to lack of tools to do so. HIF-1 α itself is often used as an endogenous measure of oxygen tension, but this is unreliable as HIF can be regulated at multiple other levels and could only be conceivably used at known identical transcription and translation rates, which is impractical.

of a host-directed therapy to cancer. But translating such therapies to infectious disease has, thus far, proven difficult or impractical. One of the reasons is a lack of understanding of the underlying mechanisms that could be targeted to benefit the host to bacterial detriment; another is the difficulty in interfering with host processes in ways that are specific to the site of infection while minimizing overall toxicity. While host toxicity is generally acceptable collateral damage in life-or-death cancer treatment, this is often viewed less favorably when treating infectious diseases for which pathogen-targeting therapies are thought superior.

Despite these challenges, mycobacterial infections, as a product of the unique intersectionality of host and bacterial biology in the granuloma, offer a spectacular opportunity to develop host-directed therapies that shorten time to cure, abbreviate the current drug regimen, prevent the emergence of antibiotic resistance, and, ultimately, fulfill the World Health Organization's goal of eradicating tuberculosis by 2050¹³⁶ (Schwegmann and Brombacher, 2008).

Host-directed therapeutic approaches can take many forms: antibodies, repurposed drugs, cell therapies, and more (Zumla et al., 2016). Many have been suggested, some of which have been reviewed by Wallis and Hafner (2015). They work by a number of different mechanisms, but can be categorized into a few main classes: enhancing antimicrobial responses (Khan et al., 2016), modulating inflammation, and altering stromal cell behavior. Of these, the most attention has been given to enhancing

¹³⁶Disease eradication has long been a stated goal of many public health campaigns, but has thus far been successful precisely twice: against the scourge of smallpox (in 1977) and against rinderpest (a disease of cattle, in 2011) (Mariner et al., 2012; Hopkins, 2013). Current campaigns show promise in the eradication of dracunculiasis (or guinea worm) in the immediate future, with cases down to 14 in 2021 (Kreier, 2022). Others, including polio, yaws, and rabies, remain elusive despite all having effective vaccines or treatments, are human-exclusive (or have a known, discrete, treatable reservoir), and declining case counts. In the eyes of many, polio is an exceptional disappointment given how close we have come, but the continued need for the use of the oral polio vaccine makes eradication all but impossible in the immediate term.

antimycobacterial responses as there are a number of useful autophagy activators that may be able to more effectively kill intracellular bacteria (Wallis and Hafner, 2015). Modulation of the expression of known deleterious responses, like the production of CCL2 and MMP9 offer additional avenues of drug discovery (Volkman et al., 2010; Cambier et al., 2014). However, these all come with important limitations in that they may only be effective at particular stages of disease – how prominent is the role of autophagy in later stages of disease? On the other hand, by attempting to modify the stromal microenvironment, it may be possible to address the granuloma *in medias res* and enhance responses through these mechanisms. This is distinct from but analogous to the approach taken checkpoint inhibitors in cancer: nothing about the tumor itself has changed, but the way the immune system is engaging with it changes fundamentally.

The most successful currently utilized host-directed therapy against tuberculosis is the use of corticosteroids to inhibit excessive inflammation during disease, a phenomenon that is able to be genetically predicted based on the individuals genotype at the *LTA4H* locus (Tobin et al., 2012; Tobin, 2015). Additional therapies may be as trivial as the addition of excess vitamin D₃, which may act to enhance antimicrobial responses in granuloma macrophages (Tobin, 2015). Another proposed approach is through the modulation of IL-1 and IFN- α/β signaling. While IL-1 mediated responses can be protective against tuberculosis, type I interferon responses are thought to be host-deleterious; both pathways can be address simultaneously through the administration of prostaglandin E2 and zileuton to improve host responses through the eicosanoid pathway (Mayer-Barber et al., 2014; Kaufmann et al., 2014; Ji et al., 2021).

This work and the foundational work that preceded it present angiogenesis as a

potential target of host-directed therapy against tuberculosis (Oehlers et al., 2015; Datta et al., 2015; Polena et al., 2016; Oehlers et al., 2017; Walton et al., 2018). This approach in broad strokes has already demonstrated clinical value through the use of thalidomide to treat juvenile tuberculous meningitis as thalidomide¹³⁷ is both a VEGFA and TNF- α inhibitor (Kaufmann et al., 2014; van Toorn et al., 2021). I propose that this strategy can be improved or augmented through the use of more targeted approaches that undermine upstream events required for the production of VEGFA in such a manner that the granuloma is left few options for developing resistance to anti-angiogenic therapy (Kiran et al., 2016). Additional work will need to identify the comparative benefits of anti-angiogenic versus normalization therapy in directing host-beneficial responses that are favorable for drug delivery, which should be a major focus of near-future work.

The work presented here focuses on a few critical themes: the diversity of factors that can contribute to the net effect of a particular insult on the immune response, the networked nature of immune responses beyond simple binaries, and the vast and poorly understood complexity beyond the simple dogmas we accept today. In the context of mycobacterial infection, the balance of inflammatory and anti-inflammatory responses determines the ability of the host to survive infection. Beyond infection, this balance of signals creates human predisposition to allergies, autoimmunity, cancer, heart disease, and many other disorders. A deeper understanding of the ways that individual signal transduction cascades can drive both pro- and anti-inflammatory responses is essential for the development of better therapeutics disorders of the immune system in the broadest sense.

¹³⁷Thalidomide is famous for the tragedy of severe congenital birth defects among mothers who took the drug during pregnancy, which left a severe stigma on the drug, despite more recent findings that it is a safe and effective treatment for a variety of disorders, including cancer and infectious diseases (McBride, 1961; D'Amato et al., 1994; Franks, Macpherson, and Figg, 2004).

Chapter 2

Scientific Methodology

Detailed explanation of scientific methodology is the foundation of scientific reproducibility. In this chapter, I will introduce the major model organism used for this work – *Danio rerio*, the zebrafish – along with the aquatic pathogen used to model human tuberculosis, *Mycobacterium marinum*. Additional descriptions of tissue culture, molecular biology, genetic, and biochemical approaches are also provided¹.

2.1 Transparency in Science and Data Availability

In the absence of reproducibility and open access to both the results and the raw data from which the results are derived, science is a fruitless venture. A fundamentally human enterprise, science is subjected to the limits of human understanding and human perfectability. The only way to hedge against our base failings as scientists is to make both the product and the process as transparent as possible. To that end, all of the raw images and quantitation used in this manuscript have been made publicly available at Brewer et al. (2022a) along with all of the R scripts used to analyze the data and any processing macros or other analysis scripts written in Python.

¹Portions of this methodology section are extracted from Brewer et al. 2022 and are incorporated into this chapter *in lieu* of direct inclusion in Chapter 3, where the results can be found.

Presentation of major uses of these languages can be found in Chapter 4. Any raw images that were unable to be made available due to size limitations can be requested from the author.

Additionally, the raw data to generate this dissertation is available on Github at <https://github.com/jaredbrewer/dissertation>, including source bibliography files (without PDF attachments) and an extended bibliography with additional links and information (generated automatically by EndNote and presented unedited; not all references will be included in the final manuscript).

2.2 Zebrafish as a Model Organism

Laboratory model organisms have been a staple of research since the dawn of the scientific endeavor, but only in the past century has model standardization allowed for improvements in reproducibility and reliability among experiments. In the 1970s and into the 1980s George Streisinger in Oregon sought a model that would allow for the full visual access to development only possible in oviparous organisms (Streisinger et al., 1981). Although *Xenopus* frogs had been in use for some time, their long time to sexual maturity (up to 2 years for the tragically tetraploid *Xenopus laevis*, the dominant model at the time) and other challenges led researchers to a fish model, at the root of the land-adapted branch of the tree of life. A purchase at a local pet store led to the establishment of the imminently powerful zebrafish model, enabling seminal and otherwise impossible findings in developmental biology (Bradford et al., 2022). This model has since found applications in nearly every field of biology for many of the same reasons: optical transparency, extremely rapid development, high fecundity, and genetic tractability (Grunwald and Eisen, 2002; Eisen, 2020). These features make the zebrafish a potent and robust tool for the study of many different

biological processes and, thanks to their intolerance for inbreeding, have remained a genetically diverse outbred² model for research that allows for more sophisticated modeling of complex processes with the caveat that it also fuels a need for high n-values due to inherent variation between individuals. Conversely, detectable effects in a high-noise environment may be more robust associations than those found in a more monoclonal environment representative of only a single instance in a vast range of genomic possibilities.

Danio rerio is a small (1-2 cm in length) freshwater fish natively found in the Ganges River and tributaries in India (Engeszer et al., 2007; Arunachalam et al., 2013; Parichy, 2015). A fetching and robust fish in the pet trade, they possess reflective stripes that play a role in predator evasion, as well as contributing to their mating cycle. They spawn freely, releasing individual eggs into the water where they can be fertilized by an engaged male³. The larvae consume waterborne microorganisms for food as they undergo metamorphosis, changing key elements of the body plan to develop the adult anatomy. The fish develop into sexual maturity over the course of approximately 5-8 weeks and are typically fully grown within 3 months, allowing for rapid generation times. Their small size facilitates rearing large numbers of them at once, making it possible to do experiments with large numbers of replicates to overcome some of the heterogeneity present in the model.

Only in the past twenty years has an earnest effort been put forth to develop the

²The scale of zebrafish outbreeding is difficult to define, even among strains that are used in research laboratories. For instance, the majority of the work in subsequent chapters is done in the *AB background, a classic wild-type reference strain used around the world. This strain, similar to other strains, has upwards of 6000 copy number variations between individuals (~15% of the genome) in addition to approximately 1 single nucleotide polymorphism (SNP) for every 500 bases of genome sequence (Guryev et al., 2006; Brown et al., 2012; Balik-Meisner et al., 2018). Experimentalist anecdotes of the intolerance of the zebrafish for inbreeding are ubiquitous, as this widespread genome-level heterozygosity appears to confer some important advantages to individuals, especially in regard to fertility.

³Or some bystander male.

zebrafish as a model for immunological studies (Davis et al., 2002; Langenau et al., 2003; Hsu et al., 2004). Although it has long been known that zebrafish, like all vertebrates, possess the full repertoire of immune cells and responses (Liongue et al., 2009; Lugo-Villarino et al., 2010), little was done with that knowledge until more recently, given the perceived benefits of other models⁴. Now zebrafish have become a widely used model in the study of diseases of the immune system ranging from cancer to neurological disorders to tuberculosis, thanks to pioneering work by Leonard Zon, Lalita Ramakrishnan, David Langenau, and others. The features that made the zebrafish beloved for studies of developmental and cell biology also translate into this context, facilitating optical, genetic, and biochemical dissection of the immune system during disease.

Coinciding with their divergence from the other clades of fish, the teleost fish experienced a whole genome duplication that has left them with redundant copies of many proteins with substantive consequences for their evolution (Meyer and Schartl, 1999; Amores et al., 2011; Howe et al., 2013; Glasauer and Neuhauss, 2014). Such a duplication enables diversification of function, increased mutational tolerance, and transcriptional inactivation of one or the other copies with little ability to *a priori* predict which is expressed (Opazo et al., 2013; Voldoire et al., 2017). This duplication has led to much headache for zebrafish researchers, but has opened up interesting avenues of discovery in the realm of genetic compensation and protein evolution (Moleri et al., 2011; Boudinot et al., 2011; Rossi et al., 2015; Stainier, Kontarakis, and Rossi, 2015; El-Brolosy and Stainier, 2017; Stainier et al., 2017; El-Brolosy et al., 2019; Szatal and Stainier, 2020; Kontarakis and Stainier, 2020). This is one of the features of the model that must be considered in its application as it can increase the difficulty

⁴More on this will be provided in Subsection 2.3.3, but for now we will allow the zebrafish to stand on its own merits.

in determining genetic mechanisms at play in a given phenotype; new approaches using F₀ crisprants have allowed for more rapid phenotypic assessment. Notable for this work, the zebrafish possess 6 NFATc isoforms: *nfatc1*, *nfatc2a*, *nfatc2b*, *nfatc3a*, *nfatc3b*, and *nfatc4*.

2.2.1 Zebrafish and the History of Developmental Biology

Based on the known features of the zebrafish, it was clear that the zebrafish had potential to be a powerful model to study the development of many major organ systems and tissues in a small, amenable vertebrate, but the tools to do so were lacking (Bakkers, 2011). After the proper establishment of the zebrafish model by Streisinger et al., work was done to finely map the earliest events of development and compare these to those known from other models, including *Xenopus* and the mouse (Kimmel, Sepich, and Trevarrow, 1988; Kimmel, 1989; Kimmel et al., 1995). One of the major needs in the field was the development of genetic tools with which to assess the contribution of different genes to distinct aspect of vertebrate development (Driever et al., 1994; Mullins et al., 1994), an initiative spearheaded by Wolfgang Driever and Mary Mullins that resulted in the famous “Zebrafish Issue” of Development in December 1996, featuring 37 articles all related to the results of these enormous genome-wide mutagenesis screens (Haffter et al., 1996; Driever et al., 1996; Knapik et al., 1996; Nusslein-Volhard, 2012; Mullins et al., 2021). These parallel screens uncovered factors impacting every conceivable aspect of vertebrate development, including brain (Schier et al., 1996; Heisenberg et al., 1996; Jiang et al., 1996; Brand et al., 1996a; Stemple et al., 1996; Odenthal et al., 1996a), craniofacial (Whitfield et al., 1996; Malicki et al., 1996b; Schilling et al., 1996; Piotrowski et al., 1996; Neuhauss et al., 1996), neurological (Malicki et al., 1996a; Furutani-Seiki et al., 1996; Abdelilah et al., 1996; Baier et al., 1996; Karlstrom et al., 1996; Trowe et al., 1996), early events

(Kane et al., 1996a; Kane et al., 1996b), body morphogenesis (Brand et al., 1996b; van Eeden et al., 1996b; van Eeden et al., 1996a; Hammerschmidt et al., 1996b; Mullins et al., 1996; Hammerschmidt et al., 1996a), cardiovascular (Stainier et al., 1996; Chen et al., 1996), hematopoiesis (Weinstein et al., 1996; Ransom et al., 1996), pigmentation (Kelsh et al., 1996; Odenthal et al., 1996b), and more (Granato et al., 1996; Pack et al., 1996). These tools set the foundation for decades of research in developmental biology as the precise genes underlying these effects remained largely mysterious even after their chromosomal location had been, in many cases, mapped – such was the nature of the pre-genomics era.

Although not a part of this monumental screen, a single example of the power of early zebrafish embryonic development studies lies in the *cloche*^{m39} mutation. This spontaneous mutant displayed total failure of vasculogenesis and hematopoiesis, providing evidence of the common developmental origin of these two processes and implying the existence of the so-called hemangioblast⁵ (Stainier et al., 1995). While further studies were conducted on this gene to characterize its activity in determining endothelial and hematopoietic fate; its first described function was as an upstream regulator of VEGFR2 (*flk1* or *kdr*)⁶ expression and that this effect was mediated cell-autonomously (Liao et al., 1997; Parker and Stainier, 1999). Some additional insight on regulated gene networks was revealed with the gene underlying *cloche* having cross-regulatory functions with two other genes, *scl* and *hhex*, where *cloche* mutants have disrupted expression of *hhex* and *scl* (Liao et al., 2000). While sporadic subsequent studies were conducted, without a genetic identity, it became more difficult to

⁵This since-confirmed multipotent cell type that can become either hematopoietic stem cells or endothelium was long hypothesized (Sabin, 1920; Murray, 1932) but took until the early 2000s to confirm their existence (Xiong, 2008).

⁶Due to a genome duplication, zebrafish have two copies of genes similar to VEGFR2 in humans, *kdr* and *kdr*, resulting in a confused nomenclature. For the standard references for these genes, see Bussmann et al. (2008)

generate novel insights on this gene (Qian et al., 2005). Finally, in 2016, Reischauer et al. identified *npas4l* as the gene responsible for *cloche*, which has resulted in renewed interest in this gene, where it has been found to be important for regulation of a range of critical hematopoietic genes (Marass et al., 2019) as well as part of a gene network that determines endothelial and pronephron fate (Mattonet et al., 2022). *npas4l*, while absent from mammalian genomes, provides insight into the evolution of hemangioblasts and shares many functional overlaps with ETV2, a transcription factor in mammals and fish that is absent in birds and reptiles (Vogeli et al., 2006; Weng et al., 2020).

While the precise story of *cloche* is not essential to the work presented here, the general theme is key to an appreciation for the zebrafish as a model organism – general findings at the foundation of developmental biology can feed forward into ever more sophisticated understandings as the technologies develop that enable these clearer perspectives. The zebrafish also enables study of developmentally lethal genes in ways that other vertebrate models cannot; the *cloche* mutant fish dies by 7 days post fertilization, which would make study within the living mouse possible only (partially) by vivisection or at terminal time points⁷. *cloche*, in juxtaposition to other lines, including the *myb* mutant and *vegfaa/vegfab* mutants, reveals a fundamental role for the endothelium in vertebrate development independent of tissue oxygenation and blood supply. What roles the endothelium plays in signaling to support tissue development remain an active area of fascinating study. Additionally, the range of available tools in the zebrafish that developed over the course of the study on *cloche* serves as a microcosm for progress in the field at large, where finer tools allowed for more detailed examinations of particular phenotypes – first through transgenic tools

⁷The analogous mutation in mammals, of *ETV2*, results in completely non-viable embryos and the precise moment of developmental failure is difficult to define; all embryos perish by E9.0 and lack both hematopoietic and endothelial cell lineages (Garry, 2016).

and *in situ* hybridizations, then through microarrays and RNA-seq, and ultimately through CRISPR/Cas9 and whole-genome sequencing. As more tools become available, including knock-in technologies and more affordable single cell -omics, more and more of these gross phenotype-driven historical observations will be able to be studied in deep molecular detail.

2.2.2 Modern Applications of Laboratory Zebrafish

Zebrafish are a widely applicable model to many fields of biology, including developmental biology, cell biology, cancer biology, immunology, neurology, animal behavior, toxicology, pharmacology, and more. As mentioned above, zebrafish are a classical model in developmental biology, but their utility persists into the present, where they are widely used to study many of the important early events in vertebrate development at high optical and temporal resolution. Beyond this, zebrafish are among the most amenable models in use for the study of toxicology and pharmacology on account of their facile breeding and ease of manipulation. They have also become a powerful model in animal behavior, where work by Randall Peterson and others has established the zebrafish as a useful model for drug discovery and profiling – zebrafish can be exposed to various pharmaceutical agents and then, using known responses, quantitatively profiled for behavioral changes (MacRae and Peterson, 2003; Zon and Peterson, 2005; Rihel et al., 2010; Kokel et al., 2010; Kokel and Peterson, 2011; Peterson and Macrae, 2012; MacRae and Peterson, 2015; Bruni et al., 2016; Patton, Zon, and Langenau, 2021). Like all models, the zebrafish had benefitted in recent years from the development of a variety of new technologies that have enabled ever-finer examination of interesting biology.

Leonard Zon has also served as a major force in the advancement of the zebrafish

model, this time in the realm of cancer immunology. Zon and colleagues were at the forefront of developing transgenic tools that would facilitate study of the interactions between the immune system and xenograft tumors and the utilizing these newly possible findings to advance the field's knowledge of host-cancer interactions (Amatruda et al., 2002; Trede et al., 2004; Cagan, Zon, and White, 2019; McConnell and Zon, 2021). Langenau et al. (2003) developed a novel zebrafish model that drove the expression of the myc oncogene within the lymphoid compartment, which was able to rapidly generate leukemia in the fish. Such a platform is useful for being able to generate genetically-defined cancer-bearing animals that can then be subjected to medium- to high-throughput screening approaches for drugs, drug targets, and other genes that contribute to the biology of these cancers in humans.

Through integrated study of the immune system with these tumors in the context of new adult zebrafish models (including the optically transparent *casper* and *crystal* models), interesting new approaches have been developed, enabling new genetic, optical, and therapeutic approaches to the study of cancer (Stern and Zon, 2003; White, Rose, and Zon, 2013; Yan et al., 2019; Gomez-Abenza et al., 2019; Hason and Bartunek, 2019; Yan et al., 2021) and hematopoietic disorders (Jong et al., 2011; Tang et al., 2014). Through novel genetic barcoding, it is now possible to analyze these cell on the level of individual cell dynamics, an unprecedented look at the biology of these tumors (Sankaran, Weissman, and Zon, 2022). These approaches have been especially powerful in the study of melanoma, for which the zebrafish provide an excellent genetic model (Kaufman et al., 2016). This work has led to clinical trials for new cancer treatments, streamlining the bench-to-bedside pipeline (Hanna et al., 2021). Zebrafish are now being proposed as “patient avatars,” enabling laboratory screening of therapeutics against the patient's particular tumor xenografted into zebrafish to predict patient responses to chemotherapy (Li et al., 2012; Sertori et al.,

2016; Yan et al., 2019; Fazio et al., 2020; Sertori et al., 2022). This is but a limited sampling of the diverse roles that zebrafish have found in the laboratory, but sets a model for how linear developments over time can shape it into a powerful model even in unexpected contexts.

2.2.3 Zebrafish as a Model System for the Study of Host-Microbe Interactions

In the same time frame as the Zon lab’s foundational discoveries in the study of cancer, the Ramakrishnan lab was developing the zebrafish to study host-pathogen interactions. While much of the understanding of tuberculosis had derived from either *in vitro* models or from the mouse, both lacked some important features that would enable a clearer understanding of these interactions (Davis et al., 2002). While this set the stage for two decades of intensive study of mycobacteria (see Section 2.3), it also set off a cascade of study on various pathogens – including other bacteria, fungi, and viruses – utilizing the zebrafish as a surrogate host and taking advantage of what makes the zebrafish scientifically compelling in other fields: gene and response conservation with humans, optical transparency, and genetic amenability (Kanther and Rawls, 2010; Angosto and Mulero, 2014; Levraud et al., 2014).

While the precise details often vary somewhat⁸, the sweep of zebrafish immunology broadly parallels that of mammalian immunology (Renshaw and Trede, 2012; Zou and Secombes, 2016). Additionally, the high degree of genetic conservation between zebrafish and humans allows for functional interrogation of conserved processes using the zebrafish as an amenable and optically accessible host (Gomes and Mostowy,

⁸While zebrafish and mammals share many cytokines and receptors, others are difficult to divine. For instance, zebrafish lack GM-CSF and its receptor, as well as IL-2, IL-3 and IL-5, among other genes (Stachura et al., 2013; Pazhakh and Lieschke, 2018; Lawir et al., 2019). The diversity in the C-type lectin receptor lineage is of especial trouble for this work.

2020). They share all of the same major subtypes of immune cells across both the innate and adaptive compartments, allowing for one-to-one comparisons across systems (Thisse and Zon, 2002; van der Sar et al., 2004). More specifically, the major subtypes of macrophages are also conserved, with microglia and Kupffer cells being present through development (Oosterhof, Boddeke, and Ham, 2015; Shwartz, Goessling, and Yin, 2019). Importantly, both zebrafish and mammals have the major cytokines and cytokine receptors required for an effective immune response against pathogens, allowing findings from the fish to translate into a human context. And useful for the experimenter, the developmental timeline of the zebrafish allows for the temporal segregation of innate and adaptive responses in an otherwise immunocompetent host (Sullivan et al., 2017; Masud, Torraca, and Meijer, 2017).

The zebrafish has become a useful model for the study of a range of pathogens beyond the one used in this work, *Mycobacterium marinum* (Brannon et al., 2009; Benard et al., 2012; Briolat et al., 2014) (discussed further in Section 2.3). Other bacterial pathogens have been used in the zebrafish to study the immune response in detail, including *Salmonella* (van der Sar et al., 2003), *Pseudomonas aeruginosa* (Pont and Blanc-Potard, 2021), *Staphylococcus aureus* (Prajsnar et al., 2008), and *Vibrio cholerae* (Runft et al., 2014). Zebrafish have even found use as useful model of sepsis, allowing for rapid screening of relevant host determinants through live visualization of the cascade of inflammatory events that drive this complex immune system hyperreaction (Ruyra et al., 2014; Barber, Fleming, and Mulvey, 2016; Philip et al., 2017).

Probably the most extensively studied zebrafish-pathogen interaction outside of *M. marinum*, various elements of *Pseudomonas* biology have been dissected using the larval zebrafish. *Pseudomonas* is a prevalent infection among cystic fibrosis patients and

the particular virulence factors and host determinants of infection remain under active study. First used in 2009 by Clatworthy et al., they were readily able to validate the *in vivo* attenuation of known virulence mutants, allowing further study on novel determinants of virulence and their mechanism of action in the interaction with both host macrophages and neutrophils in the larval model. Indeed, the zebrafish allowed for the study of the contributions of the gene underlying cystic fibrosis (CFTR) in the innate immune response and found that *cftr* morphant zebrafish neutrophils were deficient in their ability to kill *Pseudomonas*, implicating altered immune responses in part of cystic fibrosis susceptibility to infection (Phennicie et al., 2010). Subsequent work sought to address the known biofilm interactions between *Candida* and *Pseudomonas*. While *in vitro* studies had assumed that these organisms were antagonistic, this *in vivo* study using the larval zebrafish found that they synergistically enhanced one another's growth and drove increased zebrafish mortality. Such understandings of microbe-microbe interactions in the context of a host expand our ability to model the human disease condition, where cystic fibrosis patients are often afflicted with polymicrobial infections (Bergeron et al., 2017).

Recently the Huttenlocher and Perfect labs have utilized the zebrafish as a model host to study host-fungal interactions, both in *Cryptococcus* and *Candida* (Gratacap and Wheeler, 2014; Johnson et al., 2018). Use of this model revealed new dimensions of the hematogenous dissemination of *Cryptococcus* and the role of macrophages in both controlling the infection and providing a replicative niche (Tenor et al., 2015; Davis et al., 2016). This model has the potential to expand for the study of other fungal pathogens and partially supplant the *Galleria mellonella* model with one that has more advanced genetic tools, more straightforward husbandry, and vertebrate immunity (Rosowski et al., 2018). In the standard mouse model, the hematogenous dissemination effect seen in human cryptococcosis patients is difficult to observe

in real-time, so this approach allows for much deeper analysis of macrophage-yeast interactions in a native tissue context.

Zebrafish are also useful in other aspects of host-microbe interactions, including in the microbiome. Pioneering work from John Rawls and Jeff Gordon established the use of gnotobiotic zebrafish to study conserved and unique host responses to microbial colonization and composition (Rawls, Samuel, and Gordon, 2004; Rawls et al., 2006). This work has expanded dramatically over time and has also contributed valuable tools back into the zebrafish toolkit that allow for the study of immune responses in diverse contexts (Kanther et al., 2011). The zebrafish has become a powerful host for studying the otherwise impossible-to-directly-observe immune-microbe interactions in the gut, shedding light on the role of these interactions in health and disease with great precision (Park et al., 2019; Murdoch and Rawls, 2019; Murdoch et al., 2019; Espenschied et al., 2019).

In all instances, the presence of a rich *in situ* innate immune system coupled with live observation has allowed researchers to rapidly screen for novel virulence factors involved in manipulating the host immune response, through bacterial subversion of both macrophages and neutrophils (Torraca and Mostowy, 2018). The tractability and genetic conservation between zebrafish and humans also enables the study of host determinants and the use of the zebrafish as a heterologous host will likely expand as the need for ever-higher-throughput screen grows to find novel dimensions of the host-pathogen response. As the zebrafish develops the initiation of the adaptive immune system also allows the zebrafish to function as a platform to evaluate vaccine candidates, perhaps cutting costs and increasing the rapidity with which new approaches can be tested (Myllymaki et al., 2017).

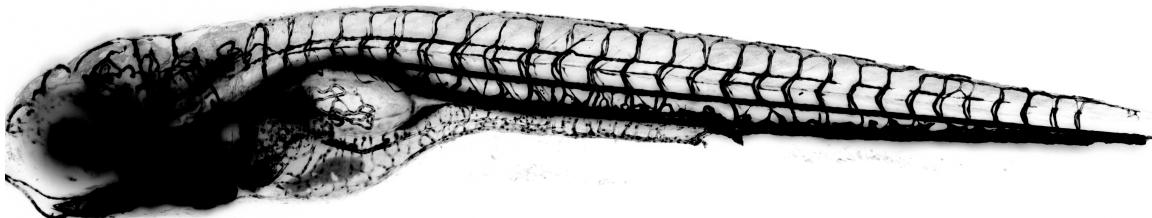


Figure 2.1: Black and white image of the *flk1:eGFP^{s843}* larval zebrafish, showing the regularly patterned intersomitic vasculature. This transgenic line allows for facile visualization and quantification of various angiogenic processes due to the highly stereotyped developmental patterns of the larval zebrafish endothelium.

2.2.4 Zebrafish a Model System for Vascular Biology

The zebrafish is a powerful model for studying various aspects of vascular development and endothelial cell biology (Wilkinson and van Eeden, 2014). As we have already seen with the tale of the *cloche* mutation, the zebrafish allows for sophisticated genetic and optical analysis of the fine details of vascular development. The zebrafish offers an array of benefits for the study of vascular processes including strong conservation of underlying genetic pathways, optical accessibility from the earliest events of development, oxygen perfusion by passive diffusion for most of the larval stages (Ellertsdottir et al., 2010), a superabundance of genetic tools (Figure 2.1), and a highly stereotyped pattern of vascularization, from which even minor perturbations can be detected (Jin et al., 2005). These features and more make the zebrafish perhaps the gold-standard model for studies of vascular patterning.

The biology of the zebrafish opens the possibility of studying processes in vascular development that are impossible to do in a viviparous animal. External fertilization and development makes it possible to precisely determine the stage at which vascular malformations induce developmental failure in a highly oxygenated environment. These things are possible to varying degrees in the mouse, for instance,

but the number of replicates and the resolution of observation are unmatched in the zebrafish.

Even the earliest events of vascular specification can be modeled in the zebrafish and translated essentially one-to-one into humans and zebrafish have been instrumental in identifying these cellular differentiation events that give rise to the earliest hemangioblasts and the consequences thereof. These hemangioblasts function as a bipotent lineage able to differentiate into both angioblasts and hematopoietic stem cells, generating the blood and lymphatic vasculature as well as all of the hematopoietic lineages (Vogeli et al., 2006). Angioblasts migrate individually toward the midline of the zebrafish and coalesce to form the dorsal aorta, a process that can be watched in real time with fluorescent imaging (Lawson and Weinstein, 2002). This early process is mediated by a short peptide and functions independently of VEGFA despite VEGFA being expressed by this point in development (Nasevicius, Larson, and Ekker, 2000; Liang et al., 2001; Hogan and Schulte-Merker, 2017). At later points in development, cells along the dorsal aorta of the zebrafish larvae dedifferentiate and become the hemogenic endothelium, from which hematopoietic stem cells arise to see the development of myeloid and erythroid populations by approximately 24 hours post fertilization (Gore et al., 2012).

Later steps depend on angiogenesis to form new blood vessels, a process intimately dependent on VEGFR signaling. In the absence of VEGFR, vascular sprouting is profoundly reduced and subsequent vasculature is malformed; this also suggests that there are some non-VEGF cues that initiate angiogenesis but are unable to properly regulate it to generate a mature endothelial system (Hogan and Schulte-Merker, 2017). These may be aberrant vasculogenesis efforts or angiogenesis induction by other mechanisms that will be discussed at greater length in further sections.

These developmental angiogenic processes have a great deal in common with the mechanisms underlying pathological angiogenesis, as highlighted in Section 1.5. During development, the somites of the zebrafish express Vegfaa to induce the formation of the stereotypic intersomitic vasculature that comprises the vascular patterning along the trunk; such developmental *vegfaa* expression can be seen clearly in transgenic approaches to visualizing this pathway (Karra et al., 2018; Walton et al., 2018). Despite the central role of VEGFA in angiogenesis, there are other auxiliary factors that are also capable of inducing angiogenesis, including BMPs, plexins, Notch, and CXCR4. The role of CXCR4 has been discussed in Subsection 1.2.6, but in development this chemokine can guide endothelial migration to ensure proper vascular connections and proper physical differentiation of the blood and lymphatic vascular systems. This role for CXCR4 is thought to be in mediating migration of already active angiogenesis and not in the initiation stage (Schuermann, Helker, and Herzog, 2014).

An interesting aspect of vascular development that remains somewhat mysterious while also being a critical dimension of vascular-targeting therapeutic strategies is the process of lumen formation within the growing vasculature. While the events of tip cell initiation, migration, and guidance have been detailed in some depth, the process of opening a lumen through which blood can flow is less well detailed, partially for lack of effective *ex vivo* complementary strategies for study. One known mechanism for this is through vacuolar fusion between neighboring cells, making the entire vessel one cell in thickness with a hollow center running along the length of the structure (Kamei et al., 2006). Such a process might be expected to predominate in the long, thin vascular spindles seen in our model of zebrafish granuloma formation, although detailed analysis of the structural features of these vessels remains lacking.

The regularly patterned intersomitic vasculature of the larval zebrafish makes it an excellent platform to study the angiogenic potential of a vast range of possible stimuli. In my work, I have used this patterned region to measure perturbations in a highly sensitive manner, as has been done by others previously (Oehlers et al., 2015; Walton et al., 2018). The developmental regularity of this location offers many advantages for studying these processes and the area generally has only sparse macrophage residency at early points in development, allowing for some degree of macrophage recruitment and then angiogenesis to be experimentally studied. This location seems like a prime potentiator of future studies on pro- and anti-angiogenic drug screening given the ease with which stimuli can be implanted into the trunk of the larval zebrafish.

2.2.5 Challenges in the Use of the Zebrafish Model

While extremely powerful, the zebrafish is still not a perfect model; the model chosen to address any question must be the one that most clearly mirrors the process being examined (and, for the zebrafish in particular, the tools to address the question must exist within the model). Challenges with zebrafish can be summarized in four major issues: fish-specific differences in biology, the aforementioned whole genome duplication, lack of true inbred models, and a lack of biochemical reagents to study zebrafish proteins.

The focus of this work is the study of tuberculosis disease, which in humans is a predominantly pulmonary infection that spreads from person to person via aerosolized droplets from the respiratory tract. Zebrafish, being fish, do not have lungs and are thus rendered impotent when it comes to the study of mycobacteria-lung interactions, including those with alveolar macrophages. While the zebrafish faithfully recapitulates many aspects of tuberculosis disease and the shared genetic programs

that drive macrophage immune responses, granuloma formation, angiogenesis, and host protection, any aspects that are specific to a given tissue may not be able to be captured with this model and other models should be employed in a complementary fashion to improve overall biological understanding.

The ancestral whole genome duplication has resulted in both a great deal of redundancy within the zebrafish genome that requires much more exhaustive genetic work than the often one-to-one gene pairs in humans and mice. While this does not inhibit the potential for discovery, it increases experimenter labor and can slow progress as the “functional” ortholog is sought out of a potential set of partially homologous genes. On the other hand, the inverse can often occur – unexpected roles for zebrafish proteins then translate into a shared function in the mammalian counterpart, which can typically only be discovered either incidentally or through unbiased screening approaches. This is not to suggest that such issues cannot occur in the mouse, as they have very famously with studies on DC-SIGN (Tanne et al., 2009; Garcia-Vallejo and Kooyk, 2013) and IRGM (Singh et al., 2006; Henry et al., 2009; Bekpen et al., 2009; Singh et al., 2010; Bekpen, Xavier, and Eichler, 2010; Dockterman and Coers, 2022), but that such issues tend to be more frequent and troublesome in the zebrafish model.

While the outbred nature of common laboratory zebrafish lines is an advantage in many respects, the lack of inbreeding fuels two issues: a poor quality reference genome and high degrees of heterogeneity within the population regarding particular responses, especially immune responses. The reference genome was a monumental effort and resulted in reasonable coverage across 1.4 gigabases (Howe et al., 2013). However, long regions of repeats and ambiguous calls along many introns have left it inferior to comparable mouse and human genomes, which were constructed from

extremely laborious but precise bacterial artificial chromosome cloning and Sanger sequencing (Osoegawa et al., 2000). While the reference strains were derived from doubled haploid founders, there remains substantial genetic diversity within these so-called strains⁹

Lastly, the lack of antibodies available for the fish is a ubiquitous complaint among zebrafish researchers. While mice have >5000 antibodies available from Abcam and humans have nearly 20,000, zebrafish have 105 at the time of this writing in December 2022. While obviously not the full breadth of all available antibodies, it demonstrates the relative lack of these tools in the zebrafish, which inhibits protein localization studies in fixed tissue as well as a more trial-and-error approach to studying genes of interest. While the situation is steadily improving, what is needed is a community-wide initiative to generate and share (ideally sequence-defined, monoclonal) antibodies from a variety of host species against a range of proteins, ideally at highly conserved locations for use in other models. Such a resource would greatly accelerate discovery in the fish, but seems to be many years off, by which time, perhaps, new computational methods to antibody-target binding will make the laborious process of host immunization and then affinity purification a moot point and enable a genetics-first approach to antibody discovery (Wilman et al., 2022; Hummer, Abanades, and Deane, 2022; Akbar et al., 2022b; Akbar et al., 2022a; Shan et al., 2022). Libraries of camelid single domain antibodies are pushing ever-closer to this elusive goal (Moutel et al., 2016; Valdes-Tresanco et al., 2022). A program comparable to AlphaFold able to predicting high-affinity binding to particular proteins would

⁹As detailed at <https://zfin.org/ZDB-GENO-960809-7>, the *AB zebrafish lineage is derived from an isolated but widely intercrossed set of lines without a dramatic series of bottlenecks to force true homozygosity (likely because this is impossible). The large numbers of offsprings and parents for each new generation ensures a relative degree of outbreeding that becomes obvious with further genomic studies (Holden and Brown, 2018; Suurvali et al., 2020; Deng et al., 2022; Bradford et al., 2022).

revolutionize all of biomedical science in a way that nothing else (including CRISPR and RNA-seq) have.

2.3 *Mycobacterium marinum*-Zebrafish Model of Tuberculosis

Work by the Ramakrishnan group, began in part in Stanley Falkow's lab, led to the development of a heterologous model system for the study of tuberculosis. *M. tuberculosis* is an extremely slow-growing pathogen¹⁰ that must be handled under biosafety level 3 (BSL-3) conditions. These requirements make working with *M. tuberculosis* challenging; deficiencies in the mouse model compound the issue and lead to difficulty in studying specific aspects of the host-pathogen interface. Thus, the development of a complementary model able to facilitate visual access and that reproduces key aspects of the human disease had the potential to unlock new insights into these interactions and, over the past twenty years, have done precisely that (Myllymaki, Bauerlein, and Ramet, 2016).

2.3.1 History and Merits of the Zebrafish-*M. marinum* Model

Studies began in the mid-1990s set the stage for the emergence of a new model of tuberculosis infection. Ramakrishnan and Falkow utilized a closely related pathogenic species of mycobacteria, *Mycobacterium marinum* to demonstrate a temperature-dependent persistence in cultured cells. A subsequent study found that *M. marinum* was able to infect and cause consumptive disease in *Rana pipiens* frogs as well as

¹⁰The doubling time is approximately 24 hours and the time to visualizable colonies is on the scale of three or four weeks.

infect human extremities (Ramakrishnan et al., 1997; Ramakrishnan, 1997; Cosma et al., 2006). A new model was also developed using carp leukocytes, allowing macrophage-mycobacterial interactions to be studied at BSL-2 conditions (El-Etr, Yan, and Cirillo, 2001). A foundational study in 2002 set the tone for the next two decades of research into host-microbe interactions in the zebrafish. Davis et al. took advantage of the optical transparency and manipulative amenability of the zebrafish larvae to infect them with *M. marinum*. *M. marinum* is a globally dispersed pathogen of fish and amphibians that causes tuberculosis in fish, which tends to manifest in superficial lesions, spinal deformities, and wasting and which can occasionally infect humans, typically on the superficial extremities where the body temperature is within the growth range of the bacterium (Aronson, 1926; Parisot, 1958; Gray et al., 1990; Hashish et al., 2018). The use of this heterologous host-pathogen system allowed for the first ever *in vivo* visualization of the early processes of granuloma formation through the interactions between the invading bacteria and the responding host macrophages, which serve as the first responding innate immune cells to mycobacterial infections (Davis et al., 2002; Davis and Ramakrishnan, 2009). This ability to dissect the relative contributions of the innate immune system in an unmodified organism has enabled many studies on the specific roles of macrophages and neutrophils in host immune control and has highlighted the imminent importance of these early responses in infection control that had been ignored by the IFN- γ and T cell-biased control seen in C57BL/6 mouse models (Lesley and Ramakrishnan, 2008).

Further developments over the following years, most notably by Swaim et al. in 2006, established the zebrafish as a sophisticated and multifaceted model that allows for both comprehensive live imaging of the early processes of infection and dissection of the later stages of infection using adult zebrafish that form granulomas morphologically similar to those formed by humans in response to both *M. tuberculosis* and

during opportunistic infections by *M. marinum*. These findings set the stage for the continued development of the zebrafish-*M. marinum* model of tuberculosis and has enabled the study of processes of human disease that have been long described but previously unable to be evaluated.

While a great deal was done to develop the model over time, there remained important questions about the ability for these findings to be translated into humans; mice are the preeminent preclinical model and to translate findings in the fish directly into humans could, in certain instances, dramatically cut the bench-to-bedside timeline. A pair of seminal papers (Tobin et al., 2010; Tobin et al., 2012) established the *LTA4H* locus as a major determinant of the course of tuberculous meningitis – individuals with either homozygous allele were at increased risk of poor outcomes and treatment of one of the homozygous populations with dexamethasone was proposed as a host-directed therapy to improve outcomes in this deadliest of tuberculosis manifestations. Indeed, subsequent studies found that this patient stratification strategy was able to determine patient response to dexamethasone in tuberculous meningitis (Thwaites, Toorn, and Schoeman, 2013; Prasad, Singh, and Ryan, 2016; Thuong et al., 2017; Wilkinson et al., 2017; Davis, Meintjes, and Wilkinson, 2018), although there may be additional factors worth considering beyond just *LTA4H* genotype (Siddiqi et al., 2021).

Today, the zebrafish-*M. marinum* model continues to proliferate in utility and acceptance. Indeed, as the field of zebrafish research as a whole advances technologically, it allows the advantages of the fish to shine through even more so in the study of host-pathogen interactions. New technologies for spatial transcriptomics will be greatly enhanced by our knowledge base in zebrafish development and the existing optical transparency of the larval fish; the day is likely not far distant when a com-

prehensive, spatially-aware transcriptional atlas of the developing zebrafish larva will be available analogous to that of *Caenorhabditis elegans*, which will only accelerate our knowledge of host-tuberculosis dynamics through added dimensional awareness (Packer et al., 2019).

As alluded to, one of the most useful incidental features of the fish is the temporal segregation of innate and adaptive immune responses (Myllymaki, Bauerlein, and Ramet, 2016). While the adult zebrafish possesses the complete vertebrate immune system, including both myeloid and lymphoid compartments, the larval zebrafish is limited to innate immune responses (Cronan and Tobin, 2014). This allows the researcher to differentiate the contributions of these two distinct systems to the overall phenotype and to, in an unmodified host, ascribe discrete roles to the innate immune system. This is one of the major reasons why so much has been learned about macrophage-mycobacterial interactions using the larval zebrafish – it allows for analysis of these interactions in the context of a native, immunocompetent host but without confounding intervention from adaptive immunity.

2.3.2 Zebrafish Husbandry

Zebrafish husbandry serves as one of the major advantages of the model. Social fish, they can be maintained at a reasonably high density per unit volume as adults¹¹, which can then spawn ~200 eggs per week per female. Our fish are kept on a constantly recirculating rack system from Aquaneering, which replaces 90%+ of the volume of each of ~1,000 tanks once per hour, 24 hours a day. The fish are diurnal animals and seasonally-indifferent photoperiodic breeders, which greatly eases experimenter access, as they will reproduce reliably only in the morning, which can

¹¹Approximately 250 mL of water per fish is more than sufficient for rapid growth to adulthood and breeding success.

be artificially set to the convenience of the laboratory. The fish are kept at 28.5°C on a 14hr-10hr light-dark cycle and kept in either 3 or 6 L tanks. Reverse osmosis water is maintained at 600-700 µS conductivity by addition of Instant Ocean Sea Salt (#SS15-10) and a pH between 7.0 and 7.4 (buffered by automated addition of sodium bicarbonate; Arm & Hammer Pure Baking Soda [#426292]). All of this work was performed in accordance and compliance with policies approved by the Duke University Institutional Animal Care and Use Committee (protocol #A091-20-04).

Infected adult zebrafish are maintained at an identical 14hr-10hr light cycle at 28.5°C in an isolated incubator (ThermoFisher #PR505755L) physically separated from the primary fish system. Fish are kept at no greater than 1 fish/100 mL of water in Aquaneering crossing cages (Aquaneering #ZHCT100) and are fed daily with the standard fish diet in our lab (Skretting #GEMMA Micro 500). Water is changed daily using water taken from the primary fish system. While sex is not a standard factor we account for in the analysis of our experiments, approximately equal numbers of fish of each sex are used for infection experiments. Any adult fish exhibiting signs of imminent distress or morbidity¹² are euthanized. Standard adult experiments are conducted for a minimum of 14 days, although infection will continue to progress over the course of approximately four to six weeks if needed. Fish are humanely euthanized at the conclusion of experimentation by tricaine overdose and decapitation.

For infection, fish were anesthetized in 120 µg/mL tricaine. Single-use aliquots of single cell suspensions of *M. marinum* were thawed and diluted in sterile PBS and zebrafish were injected with 10 µL of a solution containing 200-1000¹³ fluorescent

¹²inability to right, flared scales, labored breathing, obvious open wounds

¹³While the precise number is kept consistent within each experiment and subsequent replicates, different initial inocula are chosen based on the desired kinetics of disease. Lower initial doses

bacteria using a back-loaded insulin syringe (BD #08290-3284-38). Injection is done into the peritoneal cavity of the fish, achieving a systemic infection within the cavity and which spreads to all major organ systems¹⁴.

The larval zebrafish, on account of the high fecundity of the model, is a key tool in these studies. We can readily collect >1,000 embryos at once and can reasonably infect ~200 per hour, allowing for high n-value screening approaches. This, in addition to their remarkable ability to recover and highly stereotyped developmental patterning, facilitates reproducible studies in many fields of biology including the work described herein. Embryos are collected in the early- to mid-afternoon and allowed to develop in the presence of 0.001% methylene blue in E3 medium (5 mM NaCl (Fisher Scientific #S271), 178 µM KCl (VWR #BDH9258), 328 µM CaCl₂ (VWR #BDH9224), 400 µM MgCl₂ (Ward's Scientific #470301)) at no more than 150 larvae per dish. At 24 hours post fertilization, they are transferred into E3 supplemented with 1-phenyl-2-thiourea¹⁵ (PTU, Sigma-Aldrich #P7629) at a final concentration of 45 µg/mL to prevent melanization if they are to be used for imaging studies. Otherwise, they are allowed to develop in unmodified E3 medium until 3-4 days post fertilization, when they are put into the nursery system to be raised to adulthood. Infected larval zebrafish are euthanized prior to 8 days post fertilization in all instances. Zebrafish are of indeterminate sex¹⁶ until they reach the juvenile

are likely to be superior for identifying factors regulating bacterial control while higher doses are better if the desired outcome is to study aspects of granuloma biology, as high starting doses tend to result in a greater number of granulomas forming earlier in disease. This often appears to balance out as the disease progresses and some functional upper limit on bacillary load is reached.

¹⁴Although brain involvement is only rarely seen. It is not a standard tissue for us to analyze in the lab, but previous analysis has generally failed to identify meningitis as a manifestation of *M. marinum* infection in the zebrafish.

¹⁵The ability to taste phenylthiourea (also known as phenylthiocarbamide) is an interesting genetic trait in humans that can predispose individuals to enjoying the taste of various cruciferous vegetables (Kim et al., 2003).

¹⁶This is only strictly true for laboratory strains of zebrafish (Kochakpour and Moens, 2008). Wild

stage of development, so no distinctions are or can be made on the basis of sex in larval zebrafish studies.

For ease of manipulation and the minimization of distress, larval zebrafish are anesthetized in approximately 160 µg/mL of tricaine (MS-222 or Tricaine-S¹⁷, Syndel #ANADA 200-226) prior to injection. Injection of *M. marinum* is done by injecting larvae into a developmentally undefined peri-notochordal space between the somitic muscle layers along the trunk at 2 days post fertilization. Approximately 50-150 fluorescent bacteria are injected, spreading along the anterior-posterior length of the fish and establishing a largely localized infection along the avascular trunk. This infection location allows for facile quantitation of vascular aberrations, an essential prerequisite for this study. Injection of TDM is conducted similarly. Trehalose 6-6'-dimycolate from *Mycobacterium bovis* (TDM, Sigma-Aldrich #T3034) was re-suspended in 2:1 v/v chloroform:methanol at 1 mg/mL and stored at -80°C. Prior to use, the liquid is evaporated under vacuum and emulsified in incomplete Freund's adjuvant (IFA, Sigma-Aldrich #F5506) at 2 mg/mL. Larvae are then anesthetized and injected with approximately 10-20 nL of TDM/IFA or IFA alone along the trunk in this same undefined peri-notochordal space. The droplets coalesce into spheres within 10-15 minutes and remain in place for the entire experimental duration. Larvae are allowed to recover in E3 medium supplemented with PTU and raised in a 28.5°C incubator.

Images were captured on a Zeiss Z1 compound epifluorescent microscope and pro-

zebrafish have a sex determining region on chromosome 4 that has been repeatedly lost under laboratory culture conditions, for reasons that remain mostly unknown (Howe et al., 2013; Wilson et al., 2014; Parichy, 2015). In the lab, a variety of factors seem to influence the eventual sex determination of the fish, including rearing density and caloric allotment, in addition to various genetic factors (Kossack and Draper, 2019). Despite the lack of strict genetic determinants of sex, the standard 1:1 female:male ratio is generally maintained under normal conditions.

¹⁷Now known as “Syncaleine”

cessed in FIJI/ImageJ. The Z stacks were panned across to identify aberrant vasculature, which was then traced using the "Segmented Line" tool. This was used to capture the length to a Results Table, which was then used for subsequent analysis and plotting.

Source Code 2.1: This is a reference model for the analysis used to evaluate differences in angiogenesis across different genetic and treatment groups. In all instances, raw vascular measurements from FIJI/ImageJ are imported, paired with a key file (either genotype or file name depending on blinding strategy), and then plotted with ggplot.

```

library(ggplot2)
library(ggbeeswarm)
library(scales)
library(extrafont)
library(reshape)
library(plyr)
library(ggsignif)
library(RColorBrewer)
library(FSA)
library(gghighlight)
library(mdthemes)

setwd("~/Documents")
today <- format(Sys.time(), "%m%d%y")

ttl <- expression(paste(bolditalic("nfatc2a")^"xt69"))

nfatc2a <- read.csv("./Nfatc2a_Infection/JB292/nfatc2a/JB292_quant.csv",
  header = T)
nfatc2a$exp <- "JB292"
nfatc2a <- nfatc2a[, c("Label", "Length")]
key <- read.csv("./Nfatc2a_Infection/JB292/JB292_genotypes.csv", header =
  T)
key$rep <- paste(key$rep, ".czi", sep = "")

nfatc2a.vasc <- aggregate(Length ~ Label, nfatc2a, sum)
colnames(nfatc2a.vasc) <- c("rep", "vasc")

merged <- merge(nfatc2a.vasc, key, by="rep", all=TRUE)
nfatc2a.merged <- na.omit(merged)

results <- dunnTest(nfatc2a.merged$vasc ~ as.factor(nfatc2a.merged$gen),
  two.sided = F, method = "bh")

```

```

results

nfatc2a.plot <- ggplot(nfatc2a.merged, aes(x = gen, y = vasc, color, fill
    ↵ = gen)) + # scale_y_continuous(limits = c(0,1100)) +
  geom_beeswarm(aes(shape = gen, size = 5), dodge.width = 0.9) +
  scale_shape_manual(values = c(21, 22, 24)) +
  geom_boxplot(aes(fill = gen), alpha = 0.25, outlier.shape = NA,
    ↵ position = position_dodge(width = 0.9)) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
    ↵ position_dodge(width = 0.9)) +
  xlab("Genotype") + ylab("Length of Abnormal Vasculature ( mm)") +
  geom_signif(y_position = c(450, 525, 600), xmin = c(1, 1, 2), xmax =
    ↵ c(2, 3, 3), annotations = c("p < 0.001", "p < 0.001", "p < 0.001"),
    ↵ textsize = 5.5, color = "black") +
  scale_x_discrete(limits = c("ctrl", "het", "mut"), labels =
    ↵ c("Wild-type", "Heterozygous", "Mutant")) +
  scale_fill_manual(name = "Genotype", labels = c("ctrl", "het", "mut"),
    ↵ values = c("firebrick3", "springgreen3", "deepskyblue")) +
  theme(legend.position = "none") +
  guides(color = "none") + theme_minimal() +
  ggtitle(ttl) +
  theme(text = element_text(size = 20, face = "bold"), plot.title =
    ↵ element_text(hjust = 0.5)) +
  theme(legend.position = "none")

fn <- paste("JB292_nfatc2a_", today, ".png", sep = "")
ggsave(fn, nfatc2a.plot, width = 6.625, height = 5.875, units = "in", dpi
    ↵ = 300)

```

2.3.3 Deficits of Mouse Models of Tuberculosis

One model in particular, the C57BL/6 *Mus musculus* mouse model, has become a ubiquitous feature of every major research institution all over the world due to their clonal nature¹⁸, relative ease of use, and minimal expense¹⁹. However, their genetic homogeneity fails to reproduce many phenotypes seen in human disease, making them

¹⁸Genetic diversity between individual C57BL/6 mice is in the range of 10-20 single nucleotide polymorphisms per individual in a genome of 2.5 gigabases – a remarkable degree of isogenicity (Bryant, 2011; Sarsani et al., 2019).

¹⁹A single C57BL/6 mouse from Jackson Laboratories (jax.org) at the time of writing is \$24 USD.

an excellent model for some disorders and an insufficient one for others. Such loss of heterozygosity far from models the human condition, where there are an estimated 20 million base pairs of difference from person to person (The 1000 Genomes Project Consortium et al., 2015). The difficulties of the mouse model are nowhere more apparent than in developmental biology. Although mouse viviparous development is extremely well defined and stereotyped over the course of gestation, that is precisely the challenge. Gestation is an internal and ongoing process of physiological and anatomical development and while it is possible to catalog the process of development in snapshots in time through vivisection, it is impossible to understand the kinetics and processes of development using a model that does not allow for immediate visual accessibility. While mice faithfully recapitulate certain aspects of human development, it is far from perfect and other approaches are able to access different types of knowledge in greater detail.

The mouse has served as the model for immunology for the past 50 years. It has enabled monumental discoveries that have resulted in new medications and therapies to treat nearly every conceivable human disease and is the foundation of every single chemotherapeutic medicine on the market today. The diminutive mouse is an outstanding model for a vast array of human diseases and continues to be the go-to model for many processes and disorders. However, classical inbred mouse models, including C57BL/6 and other popular lines, including BALB/c, A/J, and 129S1, fail to replicate defining characteristics of tuberculosis in ways that compromise our ability to apply findings from these models to the kinetics and pathology of human disease. For instance, the C57BL/6 mouse is highly resistant to acute tuberculosis disease; these mice can be infected with standard laboratory strains of *M. tuberculosis* and succumb approximately 300 days later²⁰.

²⁰Bacterial strain variations and dosage can alter this somewhat, but infection with the reference

Historically, the C57BL/6 mouse model of tuberculosis has served as the gold standard for studies on host-pathogen interactions in tuberculosis and has been used to identify major host-protective factors as well as bacterial virulence factors. However, even among other mouse models, the C57BL/6 model has clear challenges in the field of tuberculosis biology. For one, C57BL/6 does not form necrotic granulomas under standard laboratory doses of laboratory strains of *M. tuberculosis* (Orme, 1998). This discordance between the observed human phenotype and the mouse model leaves an abundance of room for misinterpretation of data that may or may not be translatable to the human disease context. The loci of infection in this mice becomes a granulocytic infiltrate, with abundant cellular involvement and engagement with the bacteria, including the presence of lymphocytes (Ulrichs and Kaufmann, 2006; Hunter, Jagannath, and Actor, 2007). One of the standard arguments that the granuloma has host-detrimental aspects is that the physical structure blocks T cell-mycobacteria interactions, which may be able to serve an important host-protective role. That this is incapable of being modeled in the C57BL/6 model leaves room for competing models to model these elements of granuloma biology.

More recently, the C3H/FeJ “Kramnik” mouse model has come into widespread use. This model is able to model granuloma biology, as ~50% of infected mice will form granulomas (Harper et al., 2012; Lenaerts, Barry, and Dartois, 2015). However, this is also a hypersusceptible model of infection, with the average mouse succumbing to infection within approximately 3 months. In humans, disease progression can be of either very rapid progression over the course of weeks or develop in severity over the course of many years (Timmersma et al., 2011). An alternative model has been developed through knockout of *Sst1*, which results in increased granuloma-like

strain H37Rv will result in a highly reproducible and sudden series of death almost a year after initial infection.

morphology in the mouse, but this has only been investigated in *Mycobacterium avium* models of infection (Rosenbloom et al., 2021). Thus, there remains an unmet need for a model that (a) forms granulomas and mirrors other important aspects of human disease and (b) exhibits a spectrum of disease presentation similar to that seen in humans.

Other mouse models could potentially fulfill some of these demands, if appropriately chosen. One approach has been to more precisely mimic the infectious trajectory of human disease by instilling mice with ultra-low doses of *M. tuberculosis* by aerosol, which seems to facilitate a more necrotic granuloma phenotype, perhaps suggesting that the previously used larger doses (~300 CFU) were inducing a distinct immune response that led to more granulocytic infiltration (Plumlee et al., 2021). Recently, work by Clare Smith from the Sassetti lab has established the collaborative cross mouse collection as a fruitful tool for the discovery of novel aspects of host immunity to tuberculosis, including IFN- γ -independent defense, the role of important T cell integrins, and more across approximately 50 inbred-to-homozygosity mouse strains (Smith et al., 2016; Smith et al., 2022). Further and ongoing dissection of these strains will not only reveal further determinants of host immunity and bacterial susceptibility, but may provide the elusive granuloma mouse model, although what factors render most mice incapable of forming human-like caseating granulomas remains unknown; these host determinants of granuloma formation are also intriguing aspects of this axis that may reveal new contributions from genes of previously unknown function. However, the process of granuloma formation is one of synthetic tone – the overall response itself may be more important than the contribution of any one factor, aside from major basal determinants, like STAT6 signaling (Cronan et al., 2021) and some modicum of host defense through major immunostimulatory cytokines (Flynn et al., 1993; Flynn et al., 1995). In the long term, as understandings

of the host determinants of granuloma formation are better understood, knock-in and humanized models may provide an excellent means of modeling granulomas in the mouse but this seems like a far distant development that will require a great deal of work in the meanwhile to clarify the contributions of these diverse factors.

2.3.4 Comparison to Other Models

Other popular laboratory models of tuberculosis are able to form granulomas, including rabbits and guinea pigs; the former is relatively resistant to tuberculosis while the latter is highly susceptible (Heppleston, 1949; Dorman et al., 2004; Clark, Hall, and Williams, 2014). However, these tend to require maintenance via relative outbreeding, are larger mammals with associated higher husbandry costs, and are devoid of most useful genetic tools. This left a clear gap in our ability to understand some of the aspects of this important human disease that required innovative new approaches and a whole new paradigm. Additionally, these models do not escape the challenges of working in a BSL-3 environment with *M. tuberculosis*. While the cynomolgous macaque model of tuberculosis is an excellent representation of human tuberculosis, the costs associated with the use of macaques is enormous (Pena and Ho, 2015). A cost-efficient, genetically amenable small vertebrate model of infection was a clear gap in the resources available in the late 1990s and the zebrafish was able to fill some of these gaps in our ability to model these infectious processes. As time has progressed, our ability to use the zebrafish to model different aspects of mycobacterial infection has expanded, with the zebrafish now able to be used for studies in *M. leprae*, an incorrigibly difficult organism to study due to its peculiar lifestyle and agonizing doubling time (\sim 12 days) (Madigan et al., 2017) and *M. abscessus*²¹,

²¹The treatment of these non-tuberculous mycobacteria is a fascinating tale in the innovation of novel therapeutic approaches in patients with extensively drug resistant strains of these species. Recent studies have applied mycobacteriophages derived from the SEA-PHAGE project, an un-

a close relative of *M. marinum* (Stinear et al., 2008; Halloum et al., 2016; Bryant et al., 2016).

2.4 Transgenic Tools in the Zebrafish

Since that first monumental forward genetic screen in 1996, the arsenal of tools available to the zebrafish researcher has expanded dramatically. This expansion began rather soon thereafter, with a major *insertional* mutagenesis screen being published, which allows for more rapid barcoding and identification of the disrupted gene than earlier ethylnitrosourea mutagenesis approaches (Amsterdam et al., 1999). Additionally, the ability to inject plasmid DNA and express exogenous constructs in the fish was discovered relatively early on (Stuart, McMurray, and Westerfield, 1988; Lele and Krone, 1996), but was extremely laborious in both time investment and effort required. The race was on for a technology that would be able to introduce DNA into the zebrafish embryo that would efficiently transmit across generations. The results of these labors were two complementary technologies: I-SceI meganuclease-mediated transgenesis (Thermes et al., 2002) and *tol2*-mediated transgenesis (Kwan et al., 2007). I-SceI transgenesis utilizes a “meganuclease” that recognizes and cleaves a specific, 18 base pair sequence only known to exist in the *Saccharomyces* mitochondria. This, coupled with some useful biophysical properties, allows it to protect foreign DNA within cells until it can interact and integrate with the genome (Grabher, Joly, and Wittbrodt, 2004; Soroldoni, Hogan, and Oates, 2009). While other approaches were attempted, including the use of retroviruses (Kurita, Burgess, and

dergraduate-driven research project collecting environmental phages able to infect *M. smegmatis*, to selectively infect and kill *M. abscessus* (Jordan et al., 2014; Dedrick et al., 2019; Dedrick et al., 2021; Dedrick et al., 2022). The ability to leverage a science education effort in SEA-PHAGES to uncover therapeutic phages able to save lives is an incredible testament to the power of basic science to lead to unexpected clinical breakthroughs and sets forth a model for the discovery of additional mycobacteriophages that might be able to similarly treat drug-resistant tuberculosis.

Sakai, 2004), nothing else matched this system for ease of use and efficacy. However, research in medaka had begun utilizing transposons to mediate transgenesis to great effect and this was viewed as the future. Sleeping Beauty was an early attempt, but has largely fallen into disuse (Davidson et al., 2003), while *tol2*, with its large cargo capacity (Balciunas et al., 2006), has become the dominant mode, largely thanks to the tol2kit, which is a Gateway-cloning compatible kit of promoter, gene, and 3' UTR elements that can be used to rapidly clone components to generate transgenic zebrafish – conceptualization to injection can be streamlined into ten days or less if needed (Kwan et al., 2007). This method was used in this work to generate the major new transgenic lines that enabled the work in Chapter 3²².

Newer technologies are making the zebrafish ever-more amenable to genetic manipulations and optical control. For a more in-depth analysis of the state and future of genetic tools in the zebrafish, see Subsection 2.5.2. The current range of tools available in the zebrafish is vast and able to specifically dissect key aspects of the host-pathogen interface and new tools are released all the time. For instance, it is possible to drive the expression of multiple individual proteins off of a single polycistronic transcript through the use of 2A peptides (Kim et al., 2011). Relevant to the studies to come is the ability to visualize changes in calcium, which can be seen using calmodulin-modified green and red fluorescent proteins known as genetically encoded calcium indicators, the most notable of which being GCaMP and its many

²²I have, however, adapted the *tol2* destination vector used throughout this work, pDEST tol2 Ubb pA, to an I-SceI-compatible destination vector for use with that system. The cloning steps are identical to the *tol2* vector. This vector was created by two step PCR amplification of the ccdB/CmR cassette and the backbone. The ccdB/CmR cassette was amplified with 5'-ACCGGT GGGAGGCGTTCGGGCCACAGCGGGACCATGATTACGCCAACGC-3' and 5'-GACGTCTA GGGATAAACAGGGTAATTGGTACCGTAAACGACGG-3' while the backbone was amplified with 5'-GACGTCCCGCTGTGGCCCCAACGCCTCCGCATCAGCGCAATTCAATTGG-3' and 5'-ACCGGTATTACCCTGTTATCCCTAGCAGGATAAACCTTGATGC-3'. These fragments were then digested with AgeI-HF (NEB #R3552L), AatII (NEB #R0117L), and DpnI (NEB #R0176L), heat-inactivated, and ligated together with T4 DNA ligase (NEB #M0202S) to produce pDEST I-SceI Ubb pA.

variants (Nakai, Ohkura, and Imoto, 2001; Dana et al., 2016; Zhang et al., 2021b). These have been used to both measure and manipulate calcium flux in cells to alter their movement and immune responses (Beerman et al., 2015). The zebrafish offers the possibility to do calcium imaging across the entire larva at high spatial resolution, especially through use of newer microscopy techniques like LightSheet (Reynaud et al., 2008; Kim et al., 2017).

An additional tool is the use of gene trapping techniques, that insert either transcriptional repressors or polyadenylation sequences into genes and allow for these to be rescued through use of Cre or other methods. These have allowed for various tagging approaches and have generated libraries of strains with fluorescence driven off endogenous promoters. Conversely, by placing native genes under the control of inducible promoters, it is possible to determine temporal effects of particular genes during development (Ma et al., 2017). These tools and others to be discussed later offer the opportunity to study the mechanisms of NFAT activation and the biology of these proteins in their native context. An *nfatc2a* gene trap line would be ideal, but none has yet been identified (Clark et al., 2012; Ichino et al., 2020).

2.4.1 Technical Description of New Transgenic Lines

The p5e *irg1* construct was generated by restriction digestion of *irg1*-pTol2linkerswitch (Sanderson et al., 2015) (a gift from Christopher Hall) with FseI and XmaI and then blunted using T4 DNA polymerase (NEB #M0203S) per the manufacturer's instructions. Simultaneously, p5e MCS (Kwan et al., 2007) was PCR linearized using inverted T3 and T7 promoter primers (5'-CCCTATACTGAGTCGTATTAC-3', 5'-TCCCTTAGTGAGGGTTAAT-3'), digested with DpnI and PCR purified. These fragments were then ligated using T4 DNA ligase (NEB #M0202S) to generate p5e

irg1. This plasmid was then recombined with pME tdTomato (Addgene #135202), p3e *ubb* pA (Addgene #188702), and pDEST tol2 *ubb* pA (Addgene #188701) by Gateway cloning (ThermoFisher #12538120) to generate the pTol2 *irg1*:tdTomato construct that was then injected into single cell embryos alongside 15 ng/µL *tol2* mRNA (Balciunas et al., 2006) in 1x Tango buffer (ThermoScientific #BY5). Candidate founders were selected based on fluorescence at 3 dpf, raised to adulthood, and outcrossed to *AB to establish the line, which transmits at ~50% frequency, suggesting a single insertion locus and has exhibited stable expression over ~6 generations.

Tg(*irg1*:VIVIT-*tdTomato*^{xt38}), in which the NFAT inhibitory peptide VIVIT conjugated to the fluorescent protein tdTomato is expressed strictly in macrophages, was constructed by recombination of p5E *irg1* (Addgene #188698), pME VIVIT NS (Addgene #188699), p3E tdTomato (Addgene #188700), and pDEST tol2 *ubb* pA (Addgene #188701). Reactions were incubated at equimolar ratios overnight in a 25°C thermocycler with heated lid, with volumes calculated using the “LR Ratios Calculator” Excel document (Brewer et al., 2022a). The *irg1* promoter was first described by Sanderson et al. as a macrophage-specific inducible promoter, but our lab has found that this element often drives basal expression in macrophages as well, likely in an insertion-site-dependent manner²³. Tg(*irg1*:*tdTomato*^{xt40}) was similarly generated by recombination of p5E *irg1*, pME tdTomato, p3E Ubb pA, and pDEST tol2 Ubb pA. Both of these lines express tdTomato at baseline within macrophages by 48 hours post fertilization.

The development of Tg(*lyz*:VIVIT-*tdTomato*^{xt39}) was conducted similarly, replac-

²³*irg1*, or *acod1* is an aconitate dehydrogenase that is specifically induced within macrophages by interferon responses. While we generally use this as a tool to mark macrophage populations, there is interesting biology related to the product of aconitate dehydrogenase, itaconate. The roles of itaconate in infection are many and will be explored further in Subsection 5.2.2 (ONeill2019; Nair et al., 2018; Wu et al., 2020; Lin et al., 2021; Coelho, 2022; Peace and O'Neill, 2022; Chen et al., 2022).

ing p5e *irg1* with p5e *lyz* (Hall et al., 2007). These fish were injected at the single-cell stage and screened for tdTomato expression at 2 days post fertilization. Positive founders were raised to adulthood and screened for transmission. A single founder was identified that transmitted at ~50% frequency and these were then used to further establish this transgenic line, which has sustained stable expression over approximately 4 generations.

The middle element, pME VIVIT NS was constructed by a synthetic templated PCR after annealing. Two oligonucleotides from Integrated DNA Technologies (IDT) were annealed by heating to 95°C and then slowly cooled to room temperature (sense: 5'-GCCATCATGGCAGGACCACACCCGGTGATTGTTATCACTGGACCACA TGAGGAG-3', anti-sense: 5'-CTCCTCATGTGGTCCAGTGATAACAATCAC CGGGTGTGGTCCTGCCATGATGGC-3'). This was then used as a template for PCR using two primers to add the *attB1* and *attB2* sites required for Gateway recombination into pDONR 221 (forward: 5'-GGGGACAAGTTGTACAAAAAA GCAGGCTGCCATGGCAGGACC-3', reverse: 5'-GGGGACCACTTGTAC AAGAAAGCTGGTACTCCTCATGTGGTCCAGTG-3'). This PCR product was then column purified and recombined into pDONR 221 (ThermoFisher #12536017) using BP Clonase II (ThermoFisher #11789020) to generate pME VIVIT NS (no stop) (Addgene #188699). Constructs were verified by either Sanger sequencing or whole plasmid sequencing from Plasmidsaurus and have been submitted to Addgene, which provides additional whole plasmid sequencing verification.

Genotyping to differentiate the *irg1:tdTomato^{xt40}* and *irg1:VIVIT-tdTomato^{xt38}* lines can be performed where necessary (either for intentional experimental blinding or due to incidental mixing of fish during husbandry or experimentation) by PCR and gel electrophoresis. Primers (5'-GATTAGGTGACACTATAGATTCA

GAGCTCGCACAGG-3', 5'-ATCTCGAACTCGTGGCC-3') amplify across the 3' end of the *irg1* promoter and into the 5' end of the tdTomato insert. VIVIT+ fish display a 236 bp band while tdTomato-only fish display a 163 bp band. No band is seen in sibling fish lacking an *irg1* transgene.

2.5 Use of CRISPR/Cas9 in Zebrafish

One of the major advantages of the zebrafish model is that, unique among vertebrate model organisms, it is trivial to generate targeted knockouts of essentially any gene using CRISPR/Cas9, which can be performed by even new researchers in the course of a couple of hours. Compare this to the mouse where it takes highly technical single cell injection and implantation to screen for new lines, which can often cost thousands of dollars to outsource. Extensive work has been done to establish different applications of this technology to generate F₀ individuals for high-throughput screening, knock-in technology to use endogenous promoters and tag genes, whole-segment deletions for total gene removal, and more.

One of the huge advantages of CRISPR/Cas9 in zebrafish is the ability to use “crispants” to assess phenotypes, which is very rapid and allows for efficient screening of different genes than the standard injection, rearing, and isolating single heritable mutations pipeline. This approach has been extensively studied and described elsewhere, but with the correct approach, these results often faithfully recapitulate phenotypes seen in stable mutants (Zhang et al., 2017; Wu et al., 2018). This approach transforms six months of waiting into as little as a week from injection to preliminary data – a huge time savings that allows for more informed experimentation.

Generation of mutants in *myd88*, *card9*, *nfatc2a*, *nfatc3a*, ENSDARG00000079903,

ENSDARG00000077975, and ENSDARG00000056379 was performed as described previously (Moreno-Mateos et al., 2015). Briefly, the oligonucleotides produced by CRISPRscan were utilized as a PCR template paired with the common sgRNA tail oligo (5'-AAAAGCACCGACTCGGTGCCACTTTCAAGTTGATAACGG ACTAGCCTTATTTAACTTGCTATTCTAGCTCTAAAAC-3'). These were mixed at equimolar ratios (5 µL each from 10 µM stocks) into a standard Q5 (NEB #M0491S) reaction mixture containing 2x concentration of dNTPs (NEB #N0447S) and thermocycled using the following parameters: 98°C – 30 sec, [98°C – 5 sec, 45°C – 30 sec, 72°C – 15 sec] x 24, 72°C – 5 min, 4°C – ∞. This product was then PCR purified using a commercial kit by the manufacturer's instruction (Macherey-Nagel #740609). This product was then used in an *in vitro* transcription reaction using the NEB T7 HiScribe kit (NEB #E2040S) with the following adjustments: 17 µL template, 2 µL GTP, 2 µL CTP, 2 µL ATP, 2 µL UTP, 2 µL enzyme, 3 µL buffer and left to react overnight at 37°C. This was then purified using the Monarch Total RNA Miniprep kit (NEB #T2010S). RNA was diluted to 500 ng/µL in TE and stored at -80°C until use. On the morning of injection, 1 µL of RNA was added to 1 µL of 63 µM recombinant Cas9 protein (IDT DNA #1081059) in 1x Tango buffer (ThermoFisher #BY5). This mixture was then injected into single cell embryos and these were then either used directly for experiments or raised to adulthood to be screened as potential founders. Alleles were identified by outcrossing of mosaic adults to wild-type *AB and Sanger sequencing of F₁ adults. DNA extraction was conducted by cellular lysis in 50 mM sodium hydroxide as described previously (Meeker et al., 2007). Briefly, either adult zebrafish tail fins or whole larvae were collected in 50 mM NaOH in H₂O and lysed at 98°C for 12 minutes in a thermocycler and then neutralized by 1:10 addition of a solution of 1M Tris-HCl (pH 8) in 10x TE (100 mM Tris, 10 mM EDTA). This solution was then directly used as the template for downstream PCR

reactions.

Once a mutation has been established, efficient and cost-effective genotyping is essential for differentiating the different genotypes. High-throughput and researcher blinding are also important dimensions in this and one major method of blinded genotyping at scale is via high-resolution melt analysis, or HRMA. HRMA utilizes a qPCR-based amplification with a super-saturating concentration of dsDNA-binding dyes to sensitively detect changes in the thermal melting profile of DNA (Reed, Kent, and Wittwer, 2007; Thomas et al., 2014); changes in the base-pair composition will alter the melting profile of the DNA. While larger mutations are generally easier to differentiate, this assay can theoretically detect changes as small as a A/G or C/T transition, but is often limited by the detection precision of the instrument. We have thus employed a streamlined combination of the described rapid and efficient DNA extraction via alkaline lysis (Meeker et al., 2007) with HRMA for the purposes of genotyping at every possibility. Other methods are only utilized if HRMA has failed, thus the use of classical PCR and restriction digestion to genotype *nfatc2a* mutants.

The allele *myd88^{xt2924}* was generated by injection of a single guide RNA into single-cell embryos (guide sequence: 5'-TAATACGACTCACTATAGGCAGACTGGAGGACAGGTTTAGAGCTAGAA-3'). Adult zebrafish derived from the injected embryos were crossed to wild-type fish, and the progeny were assessed via HRMA as described previously. F primer: 5'-CCGAAAGAACTGGTCTGTTCC-3'; R primer: 5'-ACGAGTTCCCAGTCCGTCA-3'. Larvae exhibiting lesions at the *myd88* target locus were further analyzed by sequencing, and an allele exhibiting a 22 bp deletion was identified. This deletion begins at amino acid 39 (of 284) and results

²⁴This allele was originally published in E. M. Walton et al. (2018). “Cyclopropane Modification of Trehalose Dimycolate Drives Granuloma Angiogenesis and Mycobacterial Growth through Vegf Signaling”. In: *Cell Host Microbe* 24.4, 514–525 e6 and was generated by Rebecca Beerman.

in a frameshift and premature translation termination after 59 amino acids. F₁ adult fish heterozygous for this allele were identified and pooled, and larvae used for the experiments requiring myd88 knockouts were generated by crossing these F₁ adults in the *flk1:eGFP* background. The resulting progeny exhibited Mendelian ratios of homozygous wild-type, heterozygous mutant, and homozygous mutant alleles via HRMA.

The allele *card9^{xt31}* was generated by injection of a single guide RNA into single-cell embryos (guide sequence: 5'-TAATACGACTCACTATAGGGCAAGGTGCTGAGCAGCGTTTAGAGCTAGAA-3'). We identified an allele containing a 28 bp insertion, resulting in an immediate downstream frameshift leading to a premature termination codon at amino acid 59 (with missense mutations beginning at amino acid 47). Genotyping was performed using high-resolution melt analysis (HRMA) using the MeltDoctor Master Mix (Applied Biosystems #4415450) with primers flanking the sgRNA site (5'-CCTTATCTGAGACAGTGCAAGGTGC-3', 5'-TTACCAACTTGCAGCGTCTG-3'). Amplification for Sanger sequencing was performed using primers (5'-GTTTCCCAGTCACGACCGAATGCTTCTCATCAAGACC-3', 5'-CGAATGCTTCTCATCAAGACC-3'. Sequencing was performed with the "M13F(-40)" primer from Eton Biosciences (5'-GTTTCCCAGTCACGAC-3').

The allele *nfatc2a^{xt69}* was generated by simultaneous injection of two neighboring guide RNAs to increase odds of a larger intervening deletion (guide sequences: 5'-TAATACGACTCACTATAGGGCTGCGAGAACGGGCCACGTTTAGAGCTAGAA-3', 5'-TAATACGACTCACTATAGGCAGCCCGTGCACGGGTTAAGCTAGAA-3'). While this strategy failed in its initial purpose of generating a guide-spanning deletion, we identified a mutation consisting of a complex, bipartite insertion/deletion resulting from independent sgRNA activity leading to a net 4 bp

insertion and frameshift leading to a premature termination codon at amino acid 272 (of 894, prior to the DNA binding domain)²⁵. Genotyping can be performed by one of two distinct restriction digest-based methods. The original method was performed by restriction digest of the ~500 bp PCR product produced by the listed sequencing primers (5'-TAGAAGGCACAGTCGAGGCTCGAGGCTTCTGGAGACCTCTG TCC-3', 5'-TGACACACATTCCACAGGGTCTCTAGAGGTTGCCCTTCATA TCCTGC-3', underlined portion base pairs with the genomic sequence); digestion was with PflMI (NEB #R0509) directly in the PCR reaction mixture. PCR was performed using LongAmp Taq (NEB #M0323) strictly for reasons of buffer compatibility with the restriction enzyme. Digestion was carried out for ~3 hours at 37° in the presence of rSAP (NEB #M0371) to minimize background. Sanger sequencing was conducted on undigested PCR products using the vendor (Eton Biosciences) supplied “BGH Reverse” primer (5'-TAGAAGGCACAGTCGAGG-3') corresponding to the appended 5' tail of the forward PCR primer.

The second method utilizes a separate set of primers (5'-CCTCTATGCAAACGCA CCTACG-3', 5'-GTGATGCTCCTTGTGGCAC-3') to generate a 102-106 bp PCR product spanning the mutation site. This PCR is performed in 20 μL reaction volumes using Taq polymerase (NEB #M0285L) (again, for reasons of buffer compatibility) and 1 μL MwoI (NEB #R0573L) is added directly to the reaction mixture after thermocycling, which is then incubated at 60°C for 1 hour. The reaction is then visualized on a 2-3% agarose gel impregnated with SYBR Safe dye. In our hands, this second method is faster, easier, more robust, and more cost-effective. In both cases, the wild-type product is unable to be cut (single larger band) while the mutant

²⁵One of the caveats to this approach is the remnant protein could act in a dominant negative fashion on the NFAT pathway. Future production of a whole locus deletion would be excellent confirmation of the phenotypes seen here, but in the context of the other results presented in Subsection 3.3.10 it seems fair to assume that the phenotype is not due to this purely speculative effect.

is cleaved into two similarly sized smaller bands (a slightly hazy single lower band); the heterozygotes are differentiated by the presence of both bands. Confirmatory Sanger sequencing was performed as needed.

The allele *nfatc3a^{xt59}* was generated using an individual sgRNA (5'-TAATAC GACTCACTATAAGGCAGTTGCAGTAGTCATGTTTAGAGCTAGAA-3') and a mutation was identified containing a 22 bp deletion leading to a premature termination codon at the 8th amino acid (of 1074). The allele was identified by PCR amplification and Sanger sequencing using F: 5'-GTTTCCCAGTCACGA CCAGAAGGTCGAGCAGTTGG-3' and R: 5'-AACGTGTTCGCCTTGC-3'. Sequencing used the "M13F(-40)" primer supplied by the vendor (Eton Biosciences) (5'-GTTTCCCAGTCACGAC-3'). Genotyping was routinely conducted by high-resolution melt analysis (HRMA) using the MeltDoctor Master Mix (ThermoFisher #4415450) with primers flanking the sgRNA site (5'-AAAGAGTCGGTGTACATA GACGGG-3', 5'-CGAAGATCAGTCTGAAGTCCAGC-3').

The allele *xt41* in ENSDARG00000079903 was generated by injection of two single guide RNAs (5'-TAATACGACTCACTATAAGGAGGCAATAAGTGGAAAGTGT TTTAGAGCTAGAA-3' and 5'-TAATACGACTCACTATAAGGCAATAAGTG GAAGTGGGGTTTAGAGCTAGAA-3') into single-cell embryos and a mutation was identified containing a 13 bp insertion in exon 4. Sequencing was conducted with F: 5'-GTTTCCCAGTCACGACTCATTATTAAGAGTGAAGAGAAGCA GG-3' and R: 5'-TGTTTTGTAGGAATCCGATGC-3' using the "M13F(-40)" primer supplied by the vendor (Eton Biosciences) (5'-GTTTCCCAGTCACGAC-3'). Genotyping was conducted by HRMA using 5'-ACAGAGACTGGAGGCAATA AGTGG-3' and 5'-CCTTGATTCACTGGTGAGTTATCCACC-3'.

The allele *xt42* in ENSDARG00000077975 was generated by injection of two single

guide RNAs (5'-TAATACGACTCACTATAAGCGAGGACTTCTGTGGATGTT TTAGAGCTAGAA-3' and 5'-TAATACGACTCACTATAAGTCATATCTCCAT TTGTCGGTTTAGAGCTAGAA-3') into single-cell embryos and a mutation was identified containing an 8 bp deletion in exon 4. Sequencing was conducted with F: 5'-GTTTCCCAGTCACGACATGAGCTGGTCTGAGAGC-3' and R: 5'-CATGAA CGTTTACCACTTACCC-3' using the "M13F(-40)" primer supplied by the vendor (Eton Biosciences) (5'-GTTTCCCAGTCACGAC-3'). Genotyping was conducted by HRMA using 5'-CAGAGGTTCATATCTCCATTGTCGAGG-3' and 5'-TTGC CCTCGATCTCTCGTCAG-3'.

The allele *xt43* in ENSDARG00000056379 was generated by injection of a single guide RNA (5'-TAATACGACTCACTATAAGTGAAAGTGTGAGAGGTCGTT TTAGAGCTAGAA-3') into single-cell embryos and a mutation was identified containing a 13 bp insertion in exon 2. Sequencing was conducted with F: 5'-GTTT TCCCAGTCACGACGACCTATACTCTCATCACAGAGC-3' and R: 5'-GTCAG ACACAGATGCATTGC-3' using the "M13F(-40)" primer supplied by the vendor (Eton Biosciences) (5'-GTTTCCCAGTCACGAC-3'). Genotyping was conducted by HRMA using 5'-CACAAAGGAGTGAAGTGTGAGAGG-3' and 5'-GAGCA ATAAGCAGGACAAGAGAAACC-3'.

2.5.1 Crispant Assays

To generate mosaic knockouts in genes of interest, we synthesized sgRNAs targeting the first exon of the respective genes. For *nfatc2a* we used 5'-TAATACGACTCACT ATAGGTCACTCAGGTGAAGTGTGTTTAGAGCTAGAA-3' and for *nfatc3a* we used 5'-TAATACGACTCACTATAAGTAGAGGCACTGACCTGCGGTTT AGAGCTAGAA-3'. For prospective genotyping of these alleles, we used HRMA

to assess approximate editing efficiency; this can only act as a rough proxy due to limitations and feasibility of exhausting genetic analysis of these mosaic larvae. For *nfatc2a*, we used the following primers: 5'-CTCTTTACGGCGAAAAAGCTGC-3', 5'-GAAACAAACCTTGAAGTCCTGTTGG-3'. For *nfatc3a* we used: 5'-AAAGAGTCGGTGTACATAGACGGG-3', 5'-CGAAGATCAGTCTGAAGTCCAGC-3'. We had already begun generating the future stable alleles *nfatc2a*^{xt69} and *nfatc3a*^{xt59} and used these listed sgRNAs to increase our likelihood of introducing a functional mutation in these genes and to normalize target location and sgRNA number.

For my contributions to Cronan et al. (2021), I designed and generated crisprants in the IL-4R receptor homologs in the zebrafish using the strategy put forth by Wu et al. (2018). The two orthologs are sufficiently similar at the nucleotide level to allow for simultaneous targeting by the same guide RNAs, which may imply a misannotation or some level of total functional redundancy – more targeted deep sequencing approaches would be required to determine which may be the case. These guide RNAs were synthesized as previously described and injected into embryos at the one-cell stage. Four pooled guide RNAs were used with the following sequences: 5'-TAATACGACTCACTATAAGGGCTTGGCAGACGAGTGTGGTTTAGAGCTAGAAATAGC-3', 5'-TAATACGACTCACTATAAGGTGATCGGATGTCTTGCACGTTTAGAGCTAGAAATAGC-3', 5'-TAATACGACTCACTATAAGGGAAACTTTCATGTTACCTGTTAGAGCTAGAAATAGC-3', and 5'-TAATACGACTCACTATAAGGCCAGGCCGTCTGTGATTGTTAGAGCTAGAAATAGC-3'.

2.5.2 Prospectus on Cutting Edge and Future Tools in the Zebrafish

Targeted gene editing with CRISPR/Cas9 has unlocked boundless opportunities to develop new genetic tools in the zebrafish. A large need in the community is greater access to tools that allow for cell-type specific gene ablation on the model of Cre-*loxP* in the mouse (Housden et al., 2017). Indeed, Cre drivers for the zebrafish exist, but incorporating *loxP* sites into native genomic sequence is a particular challenge due to technical limitations. This has led to some alternative approaches being developed, although the ultimate use of directed Cre-*loxP* gene disruption likely remains the goal. For instance, it is possible to specifically express sgRNAs in cell-types of interest using ribozyme-mediated cleavage, allowing for tissue-restricted gene ablation (Yin et al., 2015; Wang et al., 2021). While the effects are more challenging to validate, this offers a transgenic-based approach to gene ablation that is readily incorporated into existing workflows and is able to be flexibly used through use of the Tg(*ubb:Cas9^{xt48}*) fish that I founded during my rotation.

Site-specific addition of new genetic elements is a pressing need and it may be that tools are finally arriving to accomplish these goals. The new GeneWeld approach allows for microhomology mediated end-joining to specifically integrate exogenous genetic elements with reasonable efficiency, which should add new approaches to endogenous tagging and *loxP* incorporation, although this system has yet to come into widespread use (Wierson et al., 2020). This strategy has been used to knock-in fluorescent proteins, tags, Cre recombinase, and more, with utility sure to expand further (Almeida et al., 2021; Liu et al., 2022a). Others have taken slightly older approaches through gene traps to use Cre recombination to turn genes on and off in particular cell types. Gene traps, fundamentally, required luck and perseverance

to establish as their integration loci are basically random (Sugimoto et al., 2017). The ability to target particular genes with CRISPR/Cas9 to mediate the specific integration of *loxP* sites remains somewhat elusive. Part of the challenge is the extremely rapid cell generation time of zebrafish embryos, which makes technologies developed for the much slower mouse embryo difficult or impossible to use thanks to cell cycle stage-specific DNA repair strategies (Hustedt and Durocher, 2016; Prill and Dawson, 2020).

In the future, these knock-in strategies need to be exhaustively optimized to allow for trivial generation by researchers in the same way that *tol2*-mediated transgenesis has been optimized. It is difficult to know what variables warrant the most optimization, but pharmacological treatment of zebrafish embryos to alter the balance of DNA repair outcomes seems to be promising (Nakade et al., 2014; Luo et al., 2018). Relatedly, the ability to efficiently generate precise edits in the zebrafish genome will allow for more effective zebrafish disease modeling that mirror human mutations; base editors have shown some promise but are limited in their scope and precise knock-in strategies are likely to be more extensively utilized.

Optogenetics have also been used in the zebrafish in the past to exercise fine control over the timing of gene induction, but these have been limited by unusual toxicity. Recent developments have allowed for a more robust system of optogenetics, which in the background of tissue-specific expression, has the opportunity to allow intra-granuloma manipulation of gene expression to study the specific effects of gene induction or disruption within a localized structure (Deisseroth, 2015). These will certainly complement existing transgenic approaches and can likely be integrated into these more advanced knock-in based strategies (Reade et al., 2017).

Lastly, the future of knockouts in the zebrafish is clearly through locus-spanning

deletions even if these approaches come with some downsides. The abundance of evidence on the topic of genetic compensation has led to fears that conclusions drawn about the roles of particular genes are the product of transcriptional changes in related genes through an as-yet unknown RNA-dependent mechanism (Rossi et al., 2015; El-Brolosy et al., 2019). While this has the potential to disrupt other genes via removal of trans-acting elements within gene introns, the risks of genetic compensation via mutant mRNA degradation seem too high to simply ignore. An alternate and perhaps easier strategy would be to selectively mutate critical domains in one's protein of interest to generate in-frame mutations that disrupt protein function without generating early stop codons. This is, however, not always possible and may not reveal sufficient information about the function of diverse domains along the length of a gene. Ultimately, this seems to require additional labor on the part of the researcher to effectively characterize their gene of interest through multiple complementary strategies – not necessarily an enticing prospect, but one that will improve the reliability of the literature going forward.

2.6 CLARITY and Confocal Microscopy

One of the central advantages of the larval zebrafish is its optical transparency, which allows for intravital imaging through every tissue. This advantage is lost in the adult due to the development of skin pigmentation and other factors. In other animals, this lack of optical access is a severe impediment to understanding the fundamental biological processes. Karl Deisseroth thus created the CLARITY technique, which uses a custom fixation method to immobilize proteins and nucleic acids within a tissue hydrogel and then wash away birefringent lipids, generating an optically clear tissue that can be imaged at high depth and resolution (Chung and Deisseroth, 2013;

Yang et al., 2014a). This method was adapted for use in the zebrafish by Cronan et al. (2015) and is used to study the biology of granulomas in the adult zebrafish in this work.

2.6.1 Technical Description

CLARITY fixation and clearing was conducted as previously described (Cronan et al., 2015). In brief, adult zebrafish were euthanized in tricaine, decapitated, and disemboweled. Visceral organs were immersed in an ice-cold A₁P₄ CLARITY solution (4% paraformaldehyde (EMS #15710), 1% acrylamide (Bio-Rad #1610140), 0.05% bisacrylamide (Bio-Rad #1610142), 0.0025 g/ml radical initiator (Wako Chemical #VA-044) in 1x final concentration PBS (Corning #46013CM) and nutated at 4°C for 2-3 days prior to overlay with mineral oil (Fisher Scientific #BP2629) and polymerized at 37°C for 3 hours. Hydrogel samples were collected, washed in 1x PBS, and then immersed in clearing solution at 37°C (8% sodium dodecyl sulfate (Bio-Basic #SD8119) in 200 mM boric acid (Sigma-Aldrich #B0394), pH 8.5), which was changed every 2-3 days until samples were optically clear. These samples were washed in 1x PBS supplemented to 0.1% Triton-X (Fisher Scientific #BP151) for two days at 37°C with daily solution changes to remove excess SDS from the tissue. These tissues were then individually placed into black, opaque microcentrifuge tubes (Argos Technologies #EW-06333-80) and immersed in refractive index matching solution (RIMS) (40 g, Histodenz (Sigma-Aldrich #D2158), 30 mL 20 mM phosphate buffer (4.043 g Na₂HPO₄ (VWR #BDH9296), 678.7 mg NaH₂PO₄ (Sigma-Aldrich #S9638), 1 L diH₂O), 0.01% sodium azide (NaN₃) (Sigma-Aldrich #71290)) with rotation for at least 24 hours prior to imaging (Yang et al., 2014a).

Imaging was conducted on a spinning disk microscope (Zeiss AxioObserver Z1 con-

nected to an XCite 120 LED Boost with an XLight 2TP, 89North LDI, Hamamatsu C13440 and captured on a Dell Precision Tower 5810 running Windows 10 Enterprise with Metamorph 7.10.5.476) in a MatTek dish (#P35G-1.5-14-C) with optical bottom. Additional RIMS was added to the dish to cover the sample and minimize refraction during imaging. We panned across the proximal surface of the organ bundles to identify granulomas in each individual sample and captured Z-stack images of each of the identifiable granulomas at the maximum possible optical depth in the fish. This is able to capture the majority (but perhaps not all) of the granulomas present in a given fish due to inherent limitations in lens working distance.

All image processing was conducted in FIJI/ImageJ (Schneider et al., 2012). In-focus Z planes were identified and processed with the Maximum Intensity Projection function using a Jython macro (see Section 4.4). These files were saved and then subjected to cropping where the frame was cropped to the vasculature immediately surrounding each granuloma. This distance was unable to be precisely normalized across granulomas due to the heterogeneity in size and shape of the granulomas themselves as well as the nature of their varying physiological locations. Cropped images were then blinded using the blindrename.pl script (Salter, 2016). Images were opened in ImageJ and vessels were traced using the segmented line tool, added to the Region of Interest (ROI) Manager tool and then measured for distance in pixels. The ROI files were saved and have also been provided in the Zenodo archive. Total length was then converted to microns based on the conversion factor provided by the microscope ($1 \text{ px} = 0.6552 \mu\text{m}$). Resulting .csv files were processed in Excel to remove unnecessary tag information from file names and then all subsequent analysis was performed in R using RStudio (R Core Team, 2022; RStudio Team, 2022).

2.7 *Mycobacterium marinum*

As previously discussed, *Mycobacterium marinum* is a powerful tool to study host-mycobacterial interactions in the zebrafish, but there are several critical elements of its biology that make it so useful. It was originally discovered in a salt water fish and later described as the cause of an outbreak in humans from a public swimming pool (Aronson, 1926; Linell and Norden, 1954). This bacterial species has a super-sized genome of 6.6 megabases, which is fully 50% larger than the genome of *M. tuberculosis*, reflecting adaptation of *M. tuberculosis* to a human-restricted niche via gene loss. *M. marinum* possesses orthologs of >80% of *M. tuberculosis* genes with extremely high amino acid identity (Stinear et al., 2008). This allows *M. marinum* to model many aspects of *M. tuberculosis* pathogenesis, which is my primary utility for this species in my work. This extensive genetic conservation allows us to investigate conserved pathogenic mechanisms using the heterologous zebrafish-*M. marinum* system, which offers the benefits of the zebrafish with a natural pathogen in a BSL-2 environment (Tobin and Ramakrishnan, 2008).

However, this excess coding space offers the opportunity to uncover novel aspects of mycobacterial biology through an interrogation of the genes unique to *M. marinum* that may allow it to either survive in the environment for extended periods or facilitate infection of its natural hosts, which are mostly fish, amphibians, and reptiles. These extra genes, of which there are approximately 2,500, have largely unassigned functions and an ecology-centered approach to the autotelic study of *M. marinum* would certainly reveal interesting adaptations to these diverse environments which can range from hot to cold, wet to dry, and transitory stages through various hosts spanning multiple different phylogenetic classes (Stinear et al., 2000; Stamm and

Brown, 2004). Further studies on the genetic diversity that exists within *M. marinum* may offer further hints if comparative genomics/pathogenicity approaches were taken to the study of virulence within *M. marinum* in diverse model hosts (Das et al., 2018).

2.7.1 Culture Conditions

All strains are derived from *M. marinum* strain M (ATCC #BAA-535) (Ramakrishnan and Falkow, 1994). Hygromycin-resistant fluorescent strains expressing the tdTomato (Oehlers et al., 2015), mCerulean, or EBFP2 fluorescent proteins have been described previously (Takaki et al., 2013). Bacterial culture was carried out on either 7H10 agar (Difco #262710) plates supplemented with Middlebrook OADC growth supplement (10% v/v; Sigma-Aldrich #M0678) and 50 µg/mL Hygromycin B (ThermoFisher #10687010) or liquid 7H9 media (Difco #271310) supplemented with Middlebrook OADC growth supplement (10% v/v), 0.05% Tween 80 (Sigma-Aldrich #P1754), and 50 µg/mL Hygromycin B.

Single cell preparations of these bacteria were prepared and stored as single-use aliquots at -80°C. Briefly, bacteria were grown at 33°C in 50 mL 7H9 supplemented with 10% OADC (Sigma-Aldrich #M0678), 0.05% Tween-80 (Sigma-Aldrich #P1754), and 50 µg/ml hygromycin B (Invitrogen #10687010) (7H9 Complete). Once cultures reach OD₆₀₀ 0.55-0.8, they are spun down at 4600 rcf for 15 minutes and resuspended in 5 mL PBS-T (1x PBS with 0.05% tyloxapol (Sigma-Aldrich #T8761)) and bring to 25 mL total in PBS-T. They are spun and washed 2x in 25 mL PBS-T each time and then resuspend in 2 mL of 7H9 with 10% OADC (Freezing 7H9) and split into 250 µL aliquots and homogenized 10x using a 1 mL syringe and 27G needle (BD #309623). Next, a soft spin at 770 rcf for 1 minute is done to pellet

larger clumps and the supernatants are collected and then push the pooled supernatants through a 5 μm filter (Millipore #SLSV025LS) using a 10 mL syringe. The suspension is collected in 1.5 mL microfuge tubes and spun at 10000 rcf for 5 minutes. Final resuspension of pellet is done in freezing 7H9 and aliquoted into single use aliquots and concentration is calculated by fluorescent bacteria on a hemocytometer and by colony forming units on selective media.

2.7.2 CFU Assays

Colony forming unit assays were conducted by complete homogenization of whole adult zebrafish after euthanasia by tricaine overdose and external cleansing of the skin using 70% ethanol. A single 6.5 mm ceramic bead (Omni #19-682) was added to in a pre-filled bead mill tube containing 2.8 mm stainless steel beads (Sigma-Aldrich #Z763829-50EA) and then the fish was homogenized on a bead mill (MP Bio #116004500) for a single 25 second interval at 5 meters/second. Lysate was plated on 7H10 plates supplemented with 10% OADC, hygromycin B (50 $\mu\text{g}/\text{mL}$), amphotericin B (Gibco #15290-026) (10 $\mu\text{g}/\text{mL}$), and polymyxin B (Cayman Chemical #14157) (25 $\mu\text{g}/\text{mL}$). Lysate was plated in serial 1:10 dilutions up to 10^{-5} . Cultures were grown for 10-14 days prior to counting visible colonies. Where possible (due to contamination inherent to the assay), confirmatory counting was performed at 21 days after plating to capture slow-growing colonies, which are quite rare in our hands. Plates displaying overt contamination that occluded colony growth were excluded from further analysis.

Source Code 2.2: This code displays an example of the code used to generate the plots for CFU quantitation. The CFUs are normalized to the median value of the control group in their respective experiments and then statistically compared by Student's t test and plotted using ggplot.

```

library(ggplot2)
library(scales)
library(extrafont)
library(reshape)
library(plyr)
library(ggsignif)
library(RColorBrewer)
library(FSA)
library(gghighlight)

setwd("~/Documents")
today <- format(Sys.time(), "%m%d%y")

cfu <- read.csv("./tdTomato_VIVIT_CFU_median_021622.csv", header = T)

t.test(cfu$rel_count~cfu$gen)

labs <- c(expression(paste(bolditalic("irg1:tdTomato"))),
          expression(paste(bolditalic("irg1:VIVIT"))))

ttl.1 <- expression(atop(paste(bolditalic("irg1:tdTomato ")), bold("v.")),
                     paste(bolditalic("irg1:VIVIT ")), bold("CFU")))

cfu.plot <- ggplot(cfu, aes(x = gen, y = rel_count, color = gen, fill =
  gen)) + scale_y_continuous(limits = c(0,1.25)) +
  geom_dotplot(position = position_jitterdodge(jitter.width = 0.35,
  dodge.width = 0.9), binaxis = "y", binwidth = 0.05, method =
  "histodot", stackdir = "centerwhole", stackratio = 0.1, dotsize =
  1, binpositions = "all") +
  geom_boxplot(aes(fill = gen), alpha = 0.25, outlier.shape = NA,
  position = position_dodge(width = 0.9)) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
  position_dodge(width = 0.9)) +
  xlab("Genotype") + ylab("Relative CFU") +
  geom_signif(y_position = c(1.1), xmin = c(1), xmax = c(2), annotations
  = c("p = 0.018"), textsize = 5.5, color = "black") +
  scale_x_discrete(limits = c("tdTomato", "VIVIT"), labels = labs) +
  scale_fill_manual(name = "Genotype", labels = labs, values =
  c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
  guides(color = "none") + theme_minimal() +
  labs(title = ttl.1) +
  theme(text = element_text(size = 20, face = "bold"), plot.title =
  element_text(size = 24, face = "bold", hjust = 0.5), plot.subtitle
  = element_text(size = 24, face = "bold", hjust = 0.5)) +

```

```
theme(legend.position = "none")  
  
cfu.plot  
  
fn <- paste("JB30[234]_cfu_", today, ".png", sep = "")  
ggsave(fn, cfu.plot, width = 4, height = 5.875, units = "in", dpi = 300)
```

2.8 THP-1 as a Macrophage Model

THP-1 (ATCC #TIB-202) cells are a human monocytic cell culture line derived from a 1-year-old 46XY patient suffering from acute myelogenous leukemia in 1980 (Tsuchiya et al., 1980). They have become a widespread and useful model to study certain aspects of macrophage biology *in vitro* as they can be differentiated into macrophage-like cells by addition of phorbol 12-myristate-13-acetate (PMA) to the media. This makes them a facile model – replicating in suspension at baseline, but becoming adherent, non-replicating, and terminally differentiated after treatment with PMA. While they are widely used, they are an imperfect model and cannot model certain aspects of tissue macrophage biology and are only exposed to the defined set of stimuli provided in the media. Additionally, they are variably aneuploid, with reports providing evidence of pseudo-diploid cells up to tetraploidy and beyond (Odero et al., 2000; Adati et al., 2009; Kasai et al., 2022). Despite these disadvantages, they remain among the most faithful models of general macrophage biology and macrophage responses to tuberculosis and typically align well with studies conducted on mouse bone marrow-derived macrophages and human peripheral blood monocyte-derived macrophages, arguably the two best models for macrophage biology *in vitro* (Mendoza-Coronel and Castanon-Arreola, 2016).

Here, we use them in order to model particular aspects of the host interaction with *M. tuberculosis* not only to complement our findings in the fish but also to enable new

avenues of discovery that are impossible in the complex *in vivo* setting. The relative ease of genetic and pharmacological manipulation of these cells allows for screening of macrophage phenotypes with relative ease that can then inform later approaches to be taken in the zebrafish. Going forward, using these model systems together is likely to be a potent means of honing in on interesting phenotypes that can then be dissected in parallel across both systems.

2.8.1 Culture Conditions

The cells used for these experiments were sourced from the Duke Cell Culture Facility and tested for mycoplasma prior to receipt. Cells are cultured in RPMI-1640 (Sigma-Aldrich #R8758) supplemented with glucose (Sigma-Aldrich #G8769), HEPES (Gibco #15630), sodium pyruvate (Gibco #11360) and 10% non-heat inactivated FBS (Sigma-Aldrich #F2442) in T-75 flasks (CellStar #658170) in a 37°C incubator with 5% CO₂. Cells were cultured for no greater than 10-12 passages prior to use and were then disposed of. Extended passage generated strange aberrations in the responsiveness of the cells and seemed to, generally, result in a loss of responsiveness and increase in baseline VEGFA production. I found this effect rather inexplicable and unusual, but has been occasionally commented on in the literature (Stokes and Doxsee, 1999; An et al., 2009; Chanput, Peters, and Wickers, 2015; Lund et al., 2016; Gazova et al., 2020); I have no other rational basis for these observed effects, but it is certainly one challenge of working with these cells. Several hypotheses are possible, but none strike one as especially compelling – freezing may offer some sort of “reset” or rejuvenation mechanism or different culture conditions may alter their phenotype in a semi-permanent manner. If this is to be the case, then a more robust system may be required going forward or single clone expansion and testing for sensitivity may be useful as we found clones of safe-targeting lentivirus-transduced

cells that were able to respond to gamma-irradiated *M. tuberculosis* after weeks of continuous culture under puromycin selection, suggesting that imprinted cell-to-cell heterogeneity and population drift or selection under culture may result in the effects described. Founding new lineages of cells known to respond vigorously to *M. tuberculosis* may be a useful way of making these assays more robust and reproducible going forward.

2.8.2 qRT-PCR

qRT-PCR is a powerful and highly precise method for quantitating the amount of a transcript present in a sample. This utilizes RNA extraction followed by quantitative reverse transcription to turn the RNA into complementary DNA, which can then be measured by standard PCR amplification in the presence of a dsDNA-binding dye. Under optimal conditions, this reaction proceeds by doubling the amount of product present each cycle, which can then be directly translated into a relative quantity by base 2 transformation relative to a control transcript. These data are then analyzed by the $2^{-\Delta\Delta C_t}$ method, which makes some important but not always valid assumptions about the raw data (Livak and Schmittgen, 2001; Cikos, Bukovska, and Koppel, 2007). For instance, this method assumes that the PCR efficiency for all of the primer sets is 100%, which is not always true. However, this is the gold standard for comparing the relative quantities of a transcript across two samples and also allows for comparisons of the quantities of two or more different transcripts. This method is ubiquitously used for its relative ease and resource economy while remaining highly accurate (Dvinge and Bertone, 2009; Perkins et al., 2012; Pabinger et al., 2014).

Source Code 2.3: This is a qRT-PCR-specific package-free approach to the analysis of C_t values from a 7500 Fast qPCR machine. The advantage is that the existing qPCR analysis packages are all, to varying degrees, difficult to use all to perform a relatively simple set of operations. This is also the first example in my code of using for loops to processively create all of the plots needed – this is a pattern that should be replicated in the future for ease and consistency.

```

# Jared Brewer
# Created: 05 November 2020
# Last Edited: 10 November 2022
# RT-qPCR Analysis

library(plyr)
library(dplyr)
library(ggplot2)
library(data.table)
library(reshape2)

# Read in a file formatted in a wide form - you're best off doing this
#   manually in Excel or in a text editor.
# Row names: Samples
# Col names: Gene Targets

today <- format(Sys.time(), "%m%d%y")
thp1 <- read.csv("./gmtb_inca6_qpcr_all.csv", header = T)

# Do some simple math - it appends them to the end (very important).
thp1$dVEGFA <- thp1$VEGFA - thp1$GAPDH
thp1.agg <- aggregate(dVEGFA ~ cond + rep + exp, thp1, mean)

for (exp in thp1.agg$exp[!duplicated(thp1.agg$exp)]) {
  ref <- mean(thp1.agg[thp1.agg$exp == exp & thp1.agg$cond ==
    "dmso_ctrl"]$dVEGFA)
  thp1.agg$ref[thp1.agg$exp == exp] <- ref
}

thp1.agg$ddCt <- thp1.agg$dVEGFA - thp1.agg$ref
thp1.agg$rq <- 2**-(thp1.agg$ddCt)

thp1.agg <- separate(thp1.agg, cond, into = c("drug", "bacteria"), sep =
  "_", remove = T)

labs <- c(expression(paste(bold("Control"))),
  expression(paste(bolditalic(" Mtb"))))

for (exp in thp1.agg$exp[!duplicated(thp1.agg$exp)]) {

```

```

subs <- thp1.agg[thp1.agg$exp == exp,]
thp1.aov <- aov(subs$rq ~ subs$drug*subs$bacteria)
print(TukeyHSD(thp1.aov))
if (exp == 1) {
  lims <- c(13, 14.5)
}
if (exp == 2) {
  lims <- c(11, 12.5)
}
if (exp == 3) {
  lims <- c(13, 14.5)
}
thp1.plot <- ggplot(subs, aes(x = drug, y = rq, fill = bacteria)) +
  scale_y_continuous(limits = c(0, 15)) +
  geom_beeswarm(aes(shape = drug, size = 5), dodge.width = 0.9,
    show.legend = F) +
  scale_shape_manual(values = c(21, 22, 24)) +
  geom_boxplot(aes(fill = bacteria), alpha = 0.25, outlier.shape = NA,
    position = position_dodge(width = 0.9)) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
    position_dodge(width = 0.9)) +
  xlab("Drug Treatment") + ylab("RQ VEGFA") +
  geom_signif(y_position = lims, xmin = c(0.8, 1.2), xmax = c(1.2,
    2.2), annotations = c("p < 0.001", "p < 0.001"), textsize = 5.5,
    color = "black") +
  scale_x_discrete(limits = c("dmso", "inca"), labels = c("DMSO",
    "INCA-6")) +
  scale_fill_manual(name = "Bacteria", labels = labs, values =
    c("firebrick3", "deepskyblue")) + theme(legend.position = "none")
  +
  guides(color = "none") + theme_minimal() +
  theme(text = element_text(size = 20, face = "bold"), plot.title =
    element_text(hjust = 0.5)) +
  theme(legend.position = "bottom")

ggsave(paste("THP-1_VEGFA_qPCR_exp", exp, "_", today, ".png", sep =
  ""), thp1.plot, width = 5.15, height = 6.2, dpi = 300)

# This code generates bar plots -- you need a single point to plot that
# represents all of the various images and then data to populate the
# error bars.

sem <- ddply(subs, c("drug", "bacteria"), summarize, mean = mean(rq),
  sem = sd(rq)/sqrt(length(rq)))
sem <- transform(sem, lower = mean-sem, upper = mean+sem)

```

```

colnames(sem) <- c("drug", "bacteria", "rq", "sem", "lower", "upper")

thp1.col <- ggplot(sem, aes(x = drug, y = rq, fill = bacteria)) +
  scale_y_continuous(limits = c(0, 15)) +
  geom_col(position = position_dodge(1)) +
  geom_errorbar(data = sem, aes(ymin = lower, ymax = upper), width =
    0.25, position = position_dodge(1), size = 1.5, color = "black") +
  xlab("Treatment") + ylab("RQ VEGFA") + #
  scale_y_continuous(trans='log10') +
  scale_x_discrete(limits = c("dmso", "inca"), labels = c("DMSO",
    "INCA-6")) +
# c("-INCA-6 \n" mmtb, "+INCA-6, \n -Mtb", "-INCA-6, \n +Mtb",
#   "+INCA-6, \n +Mtb")
  scale_fill_manual(name = "Bacteria", labels = labs, values =
    c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
  geom_signif(y_position = lims, xmin = c(0.8, 1.2), xmax = c(1.2,
    2.2), annotations = c("p < 0.001", "p < 0.001"), textsize = 5.5,
    color = "black") +
# guides(color = "none") +
# dark_mode() +
  theme_minimal() +
  theme(text = element_text(size = 20, face = "bold")) +
  theme(legend.position="bottom")

ggsave(paste("THP-1_VEGFA_qPCR_bar_exp", exp, "_", today, ".png", sep =
  ""), thp1.col, width = 5.15, height = 6.2, dpi = 300)
}

thp1.dagg <- aggregate(rq ~ drug + bacteria + exp, thp1.agg, mean)

thp1.aov <- aov(rq ~ drug * bacteria, data = thp1.dagg)
TukeyHSD(thp1.aov)

lims = c(12, 14, 10)

thp1.plot <- ggplot(thp1.dagg, aes(x = drug, y = rq, fill = bacteria)) +
  # scale_y_continuous(limits = c(0,15)) +
  geom_beeswarm(aes(shape = drug, size = 5), dodge.width = 0.9,
    show.legend = F) +
  scale_shape_manual(values = c(21, 22, 24)) +
  geom_boxplot(aes(fill = bacteria), alpha = 0.25, outlier.shape = NA,
    position = position_dodge(width = 0.9)) +

```

```

stat_boxplot(geom='errorbar', width = 0.25, position =
  ↵  position_dodge(width = 0.9)) +
xlab("Drug Treatment") + ylab("RQ VEGFA") +
geom_signif(y_position = lims, xmin = c(0.8, 1.2, 1.8), xmax = c(1.2,
  ↵  2.2, 2.2), annotations = c("p < 0.001", "p < 0.001", "p = 0.04"),
  ↵  textsize = 5.5, color = "black") +
scale_x_discrete(limits = c("dms0", "inca"), labels = c("DMS0",
  ↵  "INCA-6")) +
scale_fill_manual(name = "Bacteria", labels = labs, values =
  ↵  c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
guides(color = "none") + theme_minimal() +
theme(text = element_text(size = 20, face = "bold"), plot.title =
  ↵  element_text(hjust = 0.5)) +
theme(legend.position = "bottom")

ggsave(paste("THP-1_VEGFA_qPCR_agg_all", "_", today, ".png", sep = ""),
  ↵  thp1.plot, width = 5.15, height = 6.2, dpi = 300)

sem <- ddply(thp1.dagg, c("drug", "bacteria"), summarize, mean =
  ↵  mean(rq), sem = sd(rq)/sqrt(length(rq)))
sem <- transform(sem, lower = mean-sem, upper = mean+sem)
colnames(sem) <- c("drug", "bacteria", "rq", "sem", "lower", "upper")

thp1.col <- ggplot(sem, aes(x = drug, y = rq, fill = bacteria)) +
  geom_col(position = position_dodge(1)) +
  geom_errorbar(data = sem, aes(ymin = lower, ymax = upper), width =
    ↵  0.25, position = position_dodge(1), size = 1.5, color = "black") +
  xlab("Treatment") + ylab("RQ VEGFA") + #
  ↵  scale_y_continuous(trans='log10') +
  scale_x_discrete(limits = c("dms0", "inca"), labels = c("DMS0",
    ↵  "INCA-6")) +
# c("-INCA-6 \n" mmtb, "+INCA-6, \n -Mtb", "-INCA-6, \n +Mtb",
#   ↵  "+INCA-6, \n +Mtb")
  scale_fill_manual(name = "Bacteria", labels = labs, values =
    ↵  c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
  geom_signif(y_position = c(12, 15, 8), xmin = c(0.8, 1.2, 1.8), xmax =
    ↵  c(1.2, 2.2, 2.2), annotations = c("p < 0.001", "p < 0.001", "p =
      ↵  0.04"), textsize = 5.5, color = "black") +
# guides(color = "none") +
# dark_mode() +
  theme_minimal() +
  theme(text = element_text(size = 20, face = "bold")) +
  ↵  theme(legend.position="bottom") #, plot.background =
  ↵  element_rect(fill = "grey15"),

```

```

# legend.background = element_rect(fill = "grey15"), panel.background =
  element_rect(fill = "grey10"))

ggssave(paste("THP-1_VEGFA_qPCR_agg_bar", "_", today, ".png", sep = ""),
  thp1.col, width = 5.15, height = 6.2, dpi = 300)

```

Technical Details

THP-1 cells were transdifferentiated into macrophage-like cells using 50 ng/mL PMA (phorbol 12-myristate-13-acetate) (Sigma-Aldrich #P148), seeded in 24 well cell culture treated plates at a concentration of 5×10^5 cells/ml and incubated at 37°C/5%CO₂ for 48 hours. After that the PMA media was changed using complete RPMI 1640 media and incubated at 37°C/5%CO₂ for 24 hours (rest day). Then the cells were exposed to 0.5 mL of gamma-irradiated *M. tuberculosis* (BEI #NR-49098) in 25% glycerol (Sigma-Aldrich #G7757) diluted in RPMI-1640 at a final concentration of 1 mg/mL. Cells were spun at 100 rcf for 5 min and incubated at 37°C/5%CO₂ for 8 hours.

Cells then had media removed and 300 μL of Trizol was added and cells were vigorously resuspended and moved into 1.5 mL microfuge tubes. RNA extraction was conducted by addition of 0.7 volumes of 1x TE (Sigma-Aldrich #T9285) and 100 μL of chloroform (EMD Millipore #CX1055). After spinning at 17,000 rcf for 30 minutes at 4°C, the upper aqueous layer was transferred to another tube, and 100 μL of 24:1 chloroform:isoamyl alcohol (Sigma-Aldrich #25666) was added. The tubes were then shaken by hand and spun for another 30 minutes at 17,000 rcf at 4°C. The top aqueous layer was removed and final cleanup was done using the RNA Cleanup Kit (NEB #T2040L) per the manufacturer's instructions.

cDNA synthesis was performed using the LunaScript RT SuperMix Kit (NEB #E3010L) by the manufacturer's instructions. RT-PCR was performed using the

Luna Universal qPCR Master Mix (NEB #M3003L) in an Applied Biosystems 7500 Fast (ThermoFisher #4351106) per the manufacturer's instructions. Final calculations were conducted in R.

2.8.3 ELISA

An enzyme-linked immunosorbent assay (ELISA) is an effective means of quantifying the amount of protein present in a supernatant and is often used to quantify cytokine production in a variety of contexts. For my purposes, I am most interested in the production of VEGFA by THP-1 cells. The sandwich ELISA we use coats the plate with one antibody, which then fixes the protein of interest in place and is then detected by binding of another antibody at another location on the same protein. While these kits are widely manufactured, it is principally possible to use any two well-defined antibodies to measure any protein whose expression might change due to experimentation. These are somewhat targeted by nature, but reward the experimenter with highly quantitative and standardized data able to be converted into the concentration of the protein in the media.

Source Code 2.4: This older code is used to analyze ELISA data from a plate reader. It includes the internal logic to generate a standard curve and then use that standard curve to convert optical density values into a quantitative measurement of the concentration of VEGFA in the supernatant.

```
# Jared Brewer
# Created: 25 September 2021
# Last Edited: 10 November 2022
# ELISA Analysis

library(drc)
library(ggplot2)
library(ggbeeswarm)
library(scales)
library(extrafont)
library(reshape)
```

```

library(plyr)
library(ggsignif)
library(RColorBrewer)
library(FSA)
library(gghighlight)

stdcrv <- read.csv("./THP1_ELISA_StdCrv_092721.csv", header = T)

model1 <- drm(OD~Conc,
               fct=LL.4(names=c("Slope", "Lower", "Upper", "ED50")),
               data=stdcrv)
plot(model1)

##

exp3 <- read.csv("./THP1_ELISA_Rep3-0924_092721.csv", header = T)

exp3 <- aggregate(exp3$od,
                   by = list(rep = exp3$dup, treat = exp3$treat, cond =
                     ↪ exp3$cond),
                   FUN = function(x) c(mean = mean(x)))

DOSEEx <- ED(model1, exp3$x, type = "relative", display = F)
exp3.data <- as.data.frame(DOSEEx)
exp3.data$adjust <- exp3.data$Estimate*3

exp3$conc <- exp3.data$adjust
exp3[is.na(exp3)] <- 0
names(exp3)[names(exp3) == "x"] <- "raw"

exp3.aov <- aov(conc ~ treat*cond, data = exp3)

##

exp2 <- read.csv("./THP1_ELISA_Rep2-0923_092721.csv", header = T)

DOSEEx.2 <- ED(model1, exp2$od, type = "relative", display = F)
exp2.data <- as.data.frame(DOSEEx.2)
exp2.data$adjust <- exp2.data$Estimate*3

exp2$conc <- exp2.data$adjust
exp2[is.na(exp2)] <- 0

exp2.aov <- aov(conc ~ treat*cond, data = exp2)

```

```

##

# The standard curve on this one was broken for some reason, probably
# because it was the first try. But it is fine for confirmation given
# the other results and the consistence within the experiment.

exp1 <- read.csv("./R_VEGF_ELISA_gMtb_INCA-6_8+24hr_091721.csv", header =
#<-- T)
exp1.aov <- aov(blanked_od ~ treat*cond, data = exp1)

labs <- c(expression(paste(bold("Control"))),
          expression(paste(bolditalic(" Mtb"))))

# 

elisa.plot <- ggplot(exp3, aes(x = cond, y = conc, color, fill = treat))
#<-- + scale_y_continuous(limits = c(0,22)) +
  geom_quasirandom(aes(shape = cond, size = 5), dodge.width = 0.9,
#<-- show.legend = F) +
  scale_shape_manual(values = c(21, 22)) +
  geom_boxplot(aes(fill = treat), alpha = 0.25, outlier.shape = NA,
#<-- position = position_dodge(width = 0.9)) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
#<-- position_dodge(width = 0.9)) +
  xlab("Exposure Condition") + ylab("[VEGFA] pg/mL") +
  geom_signif(y_position = c(15, 17, 19), xmin = c(0.8, 0.8, 1.8), xmax =
#<-- c(1.8, 2.2, 2.2), annotations = c("p < 0.0001", "p = 0.101", "p <
#<-- 0.0001"), textsize = 8, color = "black", vjust = -0.35) +
  scale_x_discrete(limits = c("control", "mtb"), labels = labs) +
  scale_fill_manual(name = "Treatment", labels = c("DMSO", "40 M INCA-6
#<-- "), values = c("firebrick3", "deepskyblue")) +
  theme_minimal() +
  theme(text = element_text(size = 20, face = "bold")) +
  theme(legend.position="bottom")

fn <- paste("exp3_ELISA_", today, ".png", sep = "")
ggsave(fn, elisa.plot, width = 5, height = 7, units = "in", dpi = 300)

# 

elisa.plot <- ggplot(exp2, aes(x = cond, y = conc, color, fill = treat))
#<-- + scale_y_continuous(limits = c(0,40)) +
  geom_quasirandom(aes(shape = cond, size = 5), dodge.width = 0.9,
#<-- show.legend = F) +

```

```

scale_shape_manual(values = c(21, 22)) +
geom_boxplot(aes(fill = treat), alpha = 0.25, outlier.shape = NA,
← position = position_dodge(width = 0.9)) +
stat_boxplot(geom='errorbar', width = 0.25, position =
← position_dodge(width = 0.9)) +
xlab("Exposure Condition") + ylab("[VEGFA] pg/mL") +
geom_signif(y_position = c(30, 34, 38), xmin = c(0.8, 0.8, 1.8), xmax =
← c(1.8, 2.2, 2.2), annotations = c("p < 0.0001", "p = 0.193", "p =
← 0.0001"), textsize = 8, color = "black", vjust = -0.35) +
scale_x_discrete(limits = c("control", "mtb"), labels = labs) +
scale_fill_manual(name = "Treatment", labels = c("DMSO", "40 M INCA-6
← "), values = c("firebrick3", "deepskyblue")) +
theme_minimal() +
theme(text = element_text(size = 20, face = "bold")) +
theme(legend.position="bottom")

fn <- paste("exp2_ELISA_", today, ".png", sep = "")
ggsave(fn, elisa.plot, width = 5, height = 7, units = "in", dpi = 300)

# 

elisa.plot <- ggplot(exp1, aes(x = cond, y = blanked_od, color, fill =
← treat)) + scale_y_continuous(limits = c(0,1.5)) +
geom_quasirandom(aes(shape = cond, size = 5), dodge.width = 0.9,
← show.legend = F) +
scale_shape_manual(values = c(21, 22)) +
geom_boxplot(aes(fill = treat), alpha = 0.25, outlier.shape = NA,
← position = position_dodge(width = 0.9)) +
stat_boxplot(geom='errorbar', width = 0.25, position =
← position_dodge(width = 0.9)) +
xlab("Exposure Condition") + ylab("Blanked OD") +
geom_signif(y_position = c(0.75, 1, 1.25), xmin = c(0.8, 0.8, 1.8),
← xmax = c(1.8, 2.2, 2.2), annotations = c("p < 0.0001", "p = 0.205",
← "p < 0.0001"), textsize = 8, color = "black", vjust = -0.35) +
scale_x_discrete(limits = c("control", "mtb"), labels = labs) +
scale_fill_manual(name = "Treatment", labels = c("DMSO", "40 M INCA-6
← "), values = c("firebrick3", "deepskyblue")) +
theme_minimal() +
theme(text = element_text(size = 20, face = "bold")) +
theme(legend.position="bottom")

fn <- paste("exp1_ELISA_", today, ".png", sep = "")
ggsave(fn, elisa.plot, width = 5, height = 7, units = "in", dpi = 300)

```

Technical Details

Cells were cultured identically to previous, except they were plated in 96 well cell culture treated plates and exposed to gamma-irradiated *M. tuberculosis* for a total of 24 hours to facilitate VEGFA production and secretion. Supernatants were collected and spun down to remove cell debris and then the upper layer was collected for further analysis. ELISA was performed according to the manufacturer's instructions (R&D Systems #DY293B). Absorbance was read on an Agilent Synergy LX plate reader. For one of the replicates shown in Chapter 3, the standard curve failed and so the normalized OD values are shown instead.

2.8.4 Immunofluorescence

As extensively detailed for the zebrafish, the ability to see something happen is extremely impactful in one's ability to understand it. While live cell imaging is possible in tissue culture, we have utilized fixed cells and immunostaining to capture a static point in time so that we can build a relationship between NFAT activation and VEGFA production on the cellular level.

THP-1 cells were plated on 4-well chamber slides (MatTek #CCS-4) and differentiated with PMA at 50 ng/mL for 48 hours. Media was then replaced with fresh RPMI-1640 and cells were allowed to rest for 24 hours prior to further stimulation. Cells were then treated by addition of 1 mg/mL final concentration gamma-irradiated *M. tuberculosis*, 40 µM INCA-6 (Cayman Chemicals #21812), and/or vehicle controls (25% glycerol in PBS or DMSO (Fisher Scientific #BP337), respectively). Cells were then incubated at 37°C, 5% CO₂ for 8 hours and then fixed in 4% PFA in 1x PBS for 20 minutes. Cells were then washed twice in 0.25% NH₄Cl (Sigma-Aldrich #254134) or 0.15M glycine (to neutralize), rinsed in PBS, blocked in 2.5%

donkey serum (Fisher Scientific #50413253) in 1x PBS and 0.01% NaN₃ for at least 20 minutes, and then incubated in primary antibody overnight at 4°C. Cells were then rinsed, secondary antibody was added and cells were again incubated overnight at 4°C. After 5x rinses in PBS, cells were dipped in distilled H₂O and mounted in DAPI Fluoromount-G (SouthernBiotech #0100-20), which was allowed to set overnight at RT in the dark. Slides were either stored at 4°C in the dark prior to visualization or visualized immediately.

Images shown in the figures were digitally adjusted for brightness and contrast in FIJI/ImageJ (Schindelin et al., 2012) and all adjustments were applied uniformly across the images within an experiment. All quantitation was performed based on the unadjusted brightness and contrast values and thresholded to better capture positive signal and eliminate the background fluorescence ubiquitous in these images.

To capture differences in VEGFA expression across different experimental conditions, we programmatically blinded a subset of images from each experimental condition using blindrename.pl (Salter, 2016) and, using the *Cell Counter* plugin in FIJI/ImageJ, we marked each nucleus (as a proxy for cell number), each cell that visually expressed VEGFA at a minimum/maximum bit value of 100/1500, cells that had nuclear translocation of NFATC2, cells at the intersection of these two factors, and, when applicable, Cas9 expression. These values were exported and subsequently processed in R.

Source Code 2.5: This script is used to analyze all of the various immunofluorescence quantitation performed in these studies and includes distinct blocks of code for the inhibitor, isoforms, and lentivirus assays and the data processing used to generate the plots in ggplot.

```
# Jared Brewer
# Created: 19 June 2022
# Last Edited: 15 November 2022
```

```

# Cell Culture Analysis Pipeline

library(DataCombine)
library(tidyr)
library(ggplot2)

today <- format(Sys.time(), "%m%d%y")

# The key is a master key with all of the included experiments in one
# file.
key <- read.csv("./THP1_quant/keyfile.csv")
colnames(key) <- c("original", "file.path")

replacer <- function(x){
  str_replace(x, ".tif", "")
}

key <- key |> mutate_all(funs(replacer))

treat.labs <- c(expression(paste(bold("Control"))),
                  expression(paste(bolditalic(" Mtb"))))

# Inhibitor Analysis

counts.dir <- "./THP1_quant/inhibitor"
counts <- list.files(counts.dir, full.names = T)

inhibitor <- data.frame()

for (file in counts) {
  if (endsWith(file, ".csv")) {
    csv <- read.csv(file)
    base <- basename(file)
    fn.col <- mutate(csv, file.path = paste0(base))
    inhibitor <- bind_rows(inhibitor, fn.col)
  }
}

inhibitor = subset(inhibitor, Slice %in% "Total")
inhibitor <- dplyr::select(inhibitor, c("Type.1", "Type.2", "Type.3",
                                         "Type.4", "file.path"))
colnames(inhibitor) <- c("num_cells", "vegf_pos", "nfat_nuc",
                        "intersect", "file.path")
inhibitor <- data.frame(lapply(inhibitor, function(x) {gsub(".csv", "", x)}))

```

```

inhibitor <- merge(inhibitor, key, by = "file.path", all = F)

inhibitor <- data.frame(lapply(inhibitor, function(x) {gsub("mtb_inca",
  ↵ "mtbinca", x)}))
inhibitor <- separate(inhibitor, original, into = c("slide", "condition",
  ↵ "rep"), sep = "_", remove = T)

# Not sure at what point in this pipeline it got confused and thought
  ↵ there were characters?
inhibitor[, c(2:5)] <- sapply(inhibitor[, c(2:5)], as.numeric)

inhibitor$pct_vegf <- inhibitor$vegf_pos/inhibitor$num_cells
inhibitor$pct_int <- inhibitor$intersect/inhibitor$vegf_pos
inhibitor$pct_nuc <- inhibitor$nfat_nuc/inhibitor$num_cells
inhibitor$vegf_nuc <- inhibitor$nfat_nuc/inhibitor$vegf_pos
inhibitor$vegf_int <- inhibitor$intersect/inhibitor$vegf_pos
inhibitor$int_tot <- inhibitor$intersect/inhibitor$num_cells
inhibitor$nuc_vegf <- inhibitor$intersect/inhibitor$nfat_nuc
inhibitor$vegf_nuc_norm <-
  ↵ (inhibitor$nfat_nuc/inhibitor$vegf_pos)/inhibitor$num_cells

inhibitor[is.na(inhibitor)] <- 0

replaces = data.frame(from = c("ctrl", "inca", "mtb", "mtbinca"), to =
  ↵ c("dmso_ctrl", "inca_ctrl", "dmso_mtb", "inca_mtb"))
inhibitor <- FindReplace(data = inhibitor, Var = "condition",
  replaceData = replaces, from = "from", to =
    ↵ "to", exact = T)
inhibitor <- separate(inhibitor, condition, into = c("drug", "bacteria"),
  ↵ sep = "_", remove = T)

main.labs <- c(expression(paste(bold("DMSO"))),
  expression(paste(bold("INCA-6"))))

inh.aov <- aov(inhibitor$pct_vegf ~
  ↵ inhibitor$treatment*inhibitor$isoform)
TukeyHSD(inh.aov)

inhibitor.plot <- ggplot(inhibitor, aes(x = drug, y = pct_vegf*100, color
  ↵ = bacteria, fill = bacteria)) +
  scale_y_continuous(limits = c(0,50)) +
  geom_beeswarm(aes(shape = drug, size = 5), dodge.width = 0.9,
    ↵ show.legend = F) +
  scale_shape_manual(values = c(21, 22, 23, 24)) +

```

```

geom_boxplot(aes(fill = bacteria), alpha = 0.25, outlier.shape = NA,
  ↵ position = position_dodge(width = 0.9), fatten = 5) +
stat_boxplot(geom='errorbar', width = 0.25, position =
  ↵ position_dodge(width = 0.9)) +
xlab("Drug Treatment") + ylab("% VEGFA+ Cells") +
geom_signif(y_position = c(40, 45, 30), xmin = c(0.8, 1.2, 1.8), xmax =
  ↵ c(1.2, 2.2, 2.2), annotations = c("p < 0.001", "p < 0.001", "p =
  ↵ 0.885"), textsize = 5.5, color = "black") +
scale_x_discrete(limits = c("dmso", "inca"), labels = main.labs) +
scale_fill_manual(name = "Treatment", labels = treat.labs, values =
  ↵ c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
guides(color = "none") + theme_minimal() +
theme(text = element_text(size = 20, face = "bold"), plot.title =
  ↵ element_text(hjust = 0.5)) +
theme(legend.position="bottom")

fn <- paste("VEGFA_inhibitor_", today, ".png", sep = "")
ggsave(fn, inhibitor.plot, height = 6, width = 5, units = "in", dpi =
  ↵ 300)

inh.aov <- aov(inhibitor$pct_nuc ~ inhibitor$treatment*inhibitor$isoform)
TukeyHSD(inh.aov)

inhibitor.plot <- ggplot(inhibitor, aes(x = drug, y = pct_nuc*100, color
  ↵ = bacteria, fill = bacteria)) +
  scale_y_continuous(limits = c(0,75)) +
  geom_beeswarm(aes(shape = drug, size = 5), dodge.width = 0.9,
  ↵ show.legend = F) +
  scale_shape_manual(values = c(21, 22, 23, 24)) +
  geom_boxplot(aes(fill = bacteria), alpha = 0.25, outlier.shape = NA,
  ↵ position = position_dodge(width = 0.9), fatten = 5) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
  ↵ position_dodge(width = 0.9)) +
  xlab("Drug Treatment") + ylab("% Nuclear NFAT") +
  geom_signif(y_position = c(65, 70, 40), xmin = c(0.8, 1.2, 1.8), xmax =
  ↵ c(1.2, 2.2, 2.2), annotations = c("p < 0.001", "p < 0.001", "p =
  ↵ 0.998"), textsize = 5.5, color = "black") +
  scale_x_discrete(limits = c("dmso", "inca"), labels = main.labs) +
  scale_fill_manual(name = "Treatment", labels = treat.labs, values =
  ↵ c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
guides(color = "none") + theme_minimal() +
theme(text = element_text(size = 20, face = "bold"), plot.title =
  ↵ element_text(hjust = 0.5)) +
theme(legend.position="bottom")

```

```

fn <- paste("NFAT_nuc_inhibitor_", today, ".png", sep = "")
ggsave(fn, inhibitor.plot, height = 6, width = 5, units = "in", dpi =
  300)

inh.aov <- aov(inhibitor$int_tot ~ inhibitor$treatment*inhibitor$isoform)
TukeyHSD(inh.aov)

inhibitor.plot <- ggplot(inhibitor, aes(x = drug, y = int_tot*100, color
  = bacteria, fill = bacteria)) +
  scale_y_continuous(limits = c(0,50)) +
  geom_beeswarm(aes(shape = drug, size = 5), dodge.width = 0.9,
  show.legend = F) +
  scale_shape_manual(values = c(21, 22, 23, 24)) +
  geom_boxplot(aes(fill = bacteria), alpha = 0.25, outlier.shape = NA,
  position = position_dodge(width = 0.9), fatten = 5) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
  position_dodge(width = 0.9)) +
  xlab("Drug Treatment") + ylab("% VEGFA+ and NFAT+") +
  geom_signif(y_position = c(35, 40, 25), xmin = c(0.8, 1.2, 1.8), xmax =
  c(1.2, 2.2, 2.2), annotations = c("p < 0.001", "p < 0.001", "p =
  1.00"), textsize = 5.5, color = "black") +
  scale_x_discrete(limits = c("dmso", "inca"), labels = main.labs) +
  scale_fill_manual(name = "Treatment", labels = treat.labs, values =
  c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
  guides(color = "none") + theme_minimal() +
  theme(text = element_text(size = 20, face = "bold"), plot.title =
  element_text(hjust = 0.5)) +
  theme(legend.position="bottom")

fn <- paste("NFAT_VEGFA_inhibitor_", today, ".png", sep = "")
ggsave(fn, inhibitor.plot, height = 6, width = 5, units = "in", dpi =
  300)

# Isoforms Analysis

counts.dir <- "./THP1_quant/isoforms"
counts <- list.files(counts.dir, full.names = T)

isoforms <- data.frame()

for (file in counts) {
  if (endsWith(file, ".csv")) {
    csv <- read.csv(file)
    base <- basename(file)
    fn.col <- mutate(csv, file.path = paste0(base))
  }
}

```

```

    isoforms <- bind_rows(isoforms, fn.col)
}
}

isoforms = subset(isoforms, Slice %in% "Total")
isoforms <- dplyr::select(isoforms, c("Type.1", "Type.2", "Type.3",
  ↵ "Type.4", "file.path"))
colnames(isoforms) <- c("num_cells", "vegf_pos", "nfat_nuc", "intersect",
  ↵ "file.path")
isoforms <- data.frame(lapply(isoforms, function(x) {gsub(".csv", "", x)
  ↵ }))
isoforms <- merge(isoforms, key, by = "file.path", all = F)
isoforms <- separate(isoforms, original, into = c("treatment", "MAX",
  ↵ "isoform", "rep"), sep = "_", remove = T)
isoforms <- data.frame(lapply(isoforms, function(x) {gsub("uninfected",
  ↵ "control", x)}))

isoforms[, c(2:5)] <- sapply(isoforms[, c(2:5)], as.numeric)

isoforms$pct_vegf <- isoforms$vegf_pos/isoforms$num_cells
isoforms$pct_int <- isoforms$intersect/isoforms$vegf_pos
isoforms$pct_nuc <- isoforms$nfat_nuc/isoforms$num_cells
isoforms$vegf_nuc <- isoforms$nfat_nuc/isoforms$vegf_pos
isoforms$vegf_int <- isoforms$intersect/isoforms$vegf_pos
isoforms$int_tot <- isoforms$intersect/isoforms$num_cells
isoforms$nuc_vegf <- isoforms$intersect/isoforms$nfat_nuc
isoforms$vegf_nuc_norm <-
  ↵ (isoforms$nfat_nuc/isoforms$vegf_pos)/isoforms$num_cells

isoforms[is.na(isoforms)] <- 0

iso.aov <- aov(isoforms$pct_int ~ isoforms$treatment*isoforms$isoform)
TukeyHSD(iso.aov)

isoforms.plot <- ggplot(isoforms, aes(x = isoform, y = pct_int*100, color
  ↵ = treatment, fill = treatment)) +
  scale_y_continuous(limits = c(0,120)) +
  geom_beeswarm(aes(shape = isoform, size = 5), dodge.width = 0.9,
  ↵ show.legend = F) +
  scale_shape_manual(values = c(21, 22, 23, 24)) +
  geom_boxplot(aes(fill = treatment), alpha = 0.25, outlier.shape = NA,
  ↵ position = position_dodge(width = 0.9), fatten = 5) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
  ↵ position_dodge(width = 0.9)) +
  xlab("Isoform") + ylab("% Double Positive Cells / VEGFA+ Cells") +

```

```

geom_signif(y_position = c(70, 115, 105, 60), xmin = c(0.8, 1.2, 1.8,
← 2.8), xmax = c(1.2, 2.2, 2.2, 4.2), annotations = c("p < 0.001", "p
← < 0.001", "p < 0.001", "p > 0.5"), textsize = 5.5, color = "black")
← +
scale_x_discrete(limits = c("nfatc1", "nfatc2", "nfatc3", "nfatc4"),
← labels = c("NFATC1", "NFATC2", "NFATC3", "NFATC4")) +
scale_fill_manual(name = "Treatment", labels = treat.labs, values =
← c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
guides(color = "none") + theme_minimal() +
theme(text = element_text(size = 20, face = "bold"), plot.title =
← element_text(hjust = 0.5)) +
theme(legend.position="bottom")

fn <- paste("NFAT_isoforms_", today, ".png", sep = "")
ggsave(fn, inhibitor.plot, height = 7, width = 7, units = "in", dpi =
← 300)

# Lentivirus Analysis

counts.dir <- "./THP1_quant/lentivirus"
counts <- list.files(counts.dir, full.names = T)

lentivirus <- data.frame()

for (file in counts) {
  if (endsWith(file, ".csv")) {
    csv <- read.csv(file)
    base <- basename(file)
    fn.col <- mutate(csv, file.path = paste0(base))
    lentivirus <- rbind(lentivirus, fn.col)
  }
}

lentivirus <- subset(lentivirus, Slice %in% "Total")
lentivirus <- dplyr::select(lentivirus, c("Type.1", "Type.2", "Type.3",
← "Type.4", "Type.5", "Type.6", "file.path"))
colnames(lentivirus) <- c("num_cells", "vegf_pos", "nfat_nuc",
← "cas9_pos", "cas9_vegf", "intersect", "file.path")

lentivirus <- data.frame(lapply(lentivirus, function(x) {gsub(".csv", "", 
← x) }))
lentivirus <- merge(lentivirus, key, by = "file.path", all = F)
lentivirus <- separate(lentivirus, original, into = c("slide",
← "genotype", "treatment", "rep"), sep = "_", remove = T)

```

```

lentivirus <- data.frame(lapply(lentivirus, function(x) {gsub("ST", "CT",
  ↵  x) }))

lentivirus[, c(2:7)] <- sapply(lentivirus[, c(2:7)], as.numeric)

lentivirus$pct_vegf <- lentivirus$vegf_pos/lentivirus$num_cells
lentivirus$pct_int <- lentivirus$intersect/lentivirus$vegf_pos
lentivirus$pct_nuc <- lentivirus$nfat_nuc/lentivirus$num_cells
lentivirus$nuc_vegf <- lentivirus$intersect/lentivirus$nfat_nuc
lentivirus$int_tot <- lentivirus$intersect/lentivirus$num_cells
lentivirus$cas9_pct <- lentivirus$cas9_vegf/lentivirus$vegf_pos

lentivirus[is.na(lentivirus)] <- 0

main.labs <- c(expression(paste(bold("Safe Targeting"))),
  expression(paste(bolditalic("NFATC2"))))

lenti.aov <- aov(lentivirus$pct_vegf ~
  ↵  lentivirus$treatment*lentivirus$genotype)
TukeyHSD(lenti.aov)

lenti.plot <- ggplot(lentivirus, aes(x = genotype, y = pct_vegf*100,
  ↵  color = treatment, fill = treatment)) +
  # scale_y_continuous(limits = c(0,50)) +
  geom_beeswarm(aes(shape = genotype, size = 5), dodge.width = 0.9,
  ↵  show.legend = F) +
  scale_shape_manual(values = c(21, 22)) +
  geom_boxplot(aes(fill = treatment), alpha = 0.25, outlier.shape = NA,
  ↵  position = position_dodge(width = 0.9), fatten = 5) +
  stat_boxplot(geom='errorbar', width = 0.25, position =
  ↵  position_dodge(width = 0.9)) +
  xlab("Genotype") + ylab("% VEGF+ Cells") +
  geom_signif(y_position = c(40, 45), xmin = c(0.8, 1.2), xmax = c(1.2,
  ↵  2.2), annotations = c("p = 0.002", "p = 0.026"), textsize = 5.5,
  ↵  color = "black") +
  scale_x_discrete(limits = c("CT", "NFATC2"), labels = main.labs) +
  scale_fill_manual(name = "Treatment", labels = treat.labs, values =
  ↵  c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
  guides(color = "none") + theme_minimal() +
  theme(text = element_text(size = 20, face = "bold"), plot.title =
  ↵  element_text(hjust = 0.5)) +
  theme(legend.position = "bottom")

fn <- paste("lenti_VEGF_", today, ".png", sep = "")

```

```

ggsave(fn, lenti.plot, width = 6.375, height = 5.875, units = "in", dpi =
    ↵ 300)

lenti.aov <- aov(lentivirus$int_tot ~
    ↵  lentivirus$treatment*lentivirus$genotype)
TukeyHSD(lenti.aov)

lenti.plot <- ggplot(lentivirus, aes(x = genotype, y = int_tot*100, color
    ↵  = treatment, fill = treatment)) +
    # scale_y_continuous(limits = c(0,50)) +
    geom_beeswarm(aes(shape = genotype, size = 5), dodge.width = 0.9,
    ↵  show.legend = F) +
    scale_shape_manual(values = c(21, 22)) +
    geom_boxplot(aes(fill = treatment), alpha = 0.25, outlier.shape = NA,
    ↵  position = position_dodge(width = 0.9), fatten = 5) +
    stat_boxplot(geom='errorbar', width = 0.25, position =
    ↵  position_dodge(width = 0.9)) +
    xlab("Genotype") + ylab("% VEGFA+ and NFAT+ of Total") +
    geom_signif(y_position = c(35, 40), xmin = c(0.8, 1.2), xmax = c(1.2,
    ↵  2.2), annotations = c("p = 0.002", "p = 0.003"), textsize = 5.5,
    ↵  color = "black") +
    scale_x_discrete(limits = c("CT", "NFATC2"), labels = main.labs) +
    scale_fill_manual(name = "Treatment", labels = treat.labs, values =
    ↵  c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
    guides(color = "none") + theme_minimal() +
    theme(text = element_text(size = 20, face = "bold"), plot.title =
    ↵  element_text(hjust = 0.5)) +
    theme(legend.position = "bottom")

fn <- paste("lenti_int_", today, ".png", sep = "")
ggsave(fn, lenti.plot, width = 6.375, height = 5.875, units = "in", dpi =
    ↵ 300)

lenti.aov <- aov(lentivirus$nuc_vegf ~
    ↵  lentivirus$treatment*lentivirus$genotype)
TukeyHSD(lenti.aov)

lenti.plot <- ggplot(lentivirus, aes(x = genotype, y = nuc_vegf*100,
    ↵  color = treatment, fill = treatment)) +
    # scale_y_continuous(limits = c(0,50)) +
    geom_beeswarm(aes(shape = genotype, size = 5), dodge.width = 0.9,
    ↵  show.legend = F) +
    scale_shape_manual(values = c(21, 22)) +
    geom_boxplot(aes(fill = treatment), alpha = 0.25, outlier.shape = NA,
    ↵  position = position_dodge(width = 0.9), fatten = 5) +

```

```

stat_boxplot(geom='errorbar', width = 0.25, position =
  ↵  position_dodge(width = 0.9)) +
xlab("Genotype") + ylab("% VEGF+ and NFAT+ \n of NFAT+ Cells") +
geom_signif(y_position = c(70, 80), xmin = c(0.8, 1.2), xmax = c(1.2,
  ↵  2.2), annotations = c("p = 0.003", "p = 0.021"), textsize = 5.5,
  ↵  color = "black") +
scale_x_discrete(limits = c("CT", "NFATC2"), labels = main.labs) +
scale_fill_manual(name = "Treatment", labels = treat.labs, values =
  ↵  c("firebrick3", "deepskyblue")) + theme(legend.position = "none") +
guides(color = "none") + theme_minimal() +
theme(text = element_text(size = 20, face = "bold"), plot.title =
  ↵  element_text(hjust = 0.5)) +
theme(legend.position = "bottom")

fn <- paste("lenti_VEGFnuc_", today, ".png", sep = "")
ggsave(fn, lenti.plot, width = 6.375, height = 5.875, units = "in", dpi =
  ↵  300)

```

Many of the antibodies used in this study were an extremely kind gift from Aviva Symes at the Uniformed Services University. She had kept them in cold storage for greater than 20 years before passing them on to me and they have proven indispensable to many of these studies, allowing us to profile all of the different NFAT isoforms as well as all of them at once using the pan-NFAT antibody, which desperately needs to be recreated, preferably as a monoclonal antibody. This would certainly make studies on NFAT more approachable to many other groups around the world.

Zeiss filter sets used were:

- Filter Set 50 (Cy5, Alexa Fluor 647)
- Filter Set 47 (CFP)
- Filter Set 38 (GFP, Alexa Fluor 488)
- Filter Set 43HE (tdTomato, Alexa Fluor 555)
- Filter Set 46 (YFP)
- Filter Set 49 (DAPI)

2.8.5 Lentivirus Construction

We sought to generate lentiviruses able to target multiple single guide RNAs to the same gene to maximize overall mutation rate and allow us to conduct experiments in mixed pools of heterogeneous cells, to minimize functional passage number. We therefore adopted a hybrid approach, inserting the sgRNA targeting array and hUbC promoter from Kabadi et al. (2014) (Addgene #53190, a kind gift from Charles Gersbach) into the NotI/XbaI site of the lentiCRISPRv2 plasmid from Sanjana, Shalem, and Zhang (2014) (Addgene #52961, a kind gift from Feng Zhang), creating a hybrid plasmid that simultaneously expressed Cas9, the puromycin resistance marker, and up to 4 single guide RNAs from a single plasmid.

This resulting transfer empty vector (pLV hUbC-Cas9-P2A-Puro_BsmBI-sgRNA-BsmBI, Addgene #188703) was digested with Esp3I FastDigest (ThermoFisher #FD0454) precisely as previously described (Kabadi et al., 2014) in the presence of equal masses (~200 ng each) of constituent sgRNA expression plasmids driven from mU6, hU6, 7SK, or hH1 RNA pol III promoters, ligated with T4 ligase (NEB #M0202S), and cloned into NEB Stable (NEB #C3040H) cells. Resulting plasmids were screened by restriction digestion and full plasmid sequencing.

Single guide RNA expression plasmids were cloned from pHU6-gRNA, pmU6-gRNA, ph7SK-gRNA, and pH1-gRNA as described previous (Kabadi et al., 2014). The guide sequences for both NFATC2 and the safe targeting loci were chosen from a database of available guides and safe loci in the human genome to model the DNA damage response from sgRNA targeting without overt toxicity or phenotypic changes (Morgens et al., 2017). These plasmids were purified and used in subsequent steps.

The appropriate lentivirus transfer plasmid was transfected into HEK-293T cells

alongside pMD2.G (Addgene #12259) and psPAX2 (Addgene #12260) (both kind gifts from Didier Trono) (plus sfGFP-C1 to mark transfected cells, Addgene #54579, a kind gift of Michael Davidson & Geoffrey Waldo) in a 4:3:1(:0.5) mass ratio using TransIT-Lenti reagents (Mirus Bio #MIR-6603) (Pedelacq et al., 2006). Supernatants were collected 48 hours post transfection and immediately used to transduce THP-1 cells in the presence of 8 µg/mL polybrene (Sigma-Aldrich #TR-1003-G). Approximate titer was determined by infecting additional HEK-293T cells with varying dilutions of the supernatant although this was largely superfluous after many days of selection under puromycin and passage.

2.8.6 Transduction

THP-1 cells were seeded in complete RPMI-1640 media supplemented with 8 µg/mL polybrene (Sigma-Aldrich #TR-1003-G) in two non-treated six-well plates at a concentration of 1×10^6 cells/ml in each well. One six-well plate was infected with 1mL of pLV-ST and the other with pLV-NFATC2. The lentivirus infected THP-1 cells were spun at 1500 rcf/2 hours/22°C, gently resuspended and incubated at 37°C/5%CO₂ for 72 hours. Transduced cells were selected with 2 µg/mL puromycin (Sigma-Aldrich #P4512) for 48 hours and then kept in complete RPMI-1640 with 1 µg/mL puromycin until time of assay, typically 10 to 20 days in total.

2.9 Quantification and Statistical Analysis

All assays were performed under experimental blinding. For all assays where the genotype or experimental condition of the fish was apparent to the experimenter during data gathering (for instance, experiments in adults and the VIVIT assays in larvae), the resulting images were computationally blinded prior to analysis with

either blindrename.pl (Salter, 2016) or an in-house Python translation (available at doi://10.5281/zenodo.6816429 or Section 4.8). For assays where the genotype is unknown (in-cross of heterozygotes experiments for *myd88*, *card9*, *nfatc2a*, *nfatc3a* in the larvae), blinding was inherent in the design of the experiment and genotypes were matched to the individual fish *post hoc*.

Statistical analysis was performed using R 4.2.2 *Innocent and Trusting* within the latest version of RStudio IDE, 2022.12 *Elsbeth Geranium* on macOS Monterey 12.6.1 (R Core Team, 2022; RStudio Team, 2022). Graphing was performed using ggplot2 (Wickham, 2016; Wickham et al., 2022a). All statistical tests performed and the resulting significance values are indicated in figures and figure legends.

2.10 R

The R statistical environment began as a competing/complementary/intercompatible alternative to the existing S statistical programming language, but has become nearly ubiquitous in the life sciences over the past fifteen years or so. Offering new users an accessible environment in which to learn programming and with a large community, this language allows for reproducible, open-source analysis and visualization of nearly any scientific data. R has become immensely flexible and libraries have been developed to accommodate any and every kind of numerical data. This also allow for community reanalysis and reinterpretation of existing data by facilitating the direct sharing of analysis methods and raw data.

R would not be where it is today without RStudio (soon to be renamed “Posit”), which offers a feature-complete integrated development environment complete with in-line output and a user-friendly interface with various easy-to-navigate panels. This interface has allowed much of the work I have completed to occur when I was less

knowledgeable about the underlying mechanics of R as a language. New users almost uniformly use RStudio as their entrance to R and it is powerful enough for more advanced users as well.

These features allowed me to develop a comprehensive set of infinitely reproducible analysis scripts to analyze all of the data presented here. These scripts, along with the raw data, are available on Zenodo at doi://10.5281/zenodo.6816429. R (version 4.2.2, *Innocent and Trusting*) was accessed via RStudio (*Elsbeth Geranium* version 2022.12.0) on macOS Monterey 12.6.1 (R Core Team, 2022; RStudio Team, 2022). The libraries used to complete this work were dplyr (Wickham et al., 2022b), reshape (Wickham, 2022), and FSA (Ogle et al., 2022). Graphs utilized ggplot2 (Wickham, 2009; Wickham, 2016; Wickham et al., 2022a), gghighlight (Yutani, 2022), ggbeeswarm (Clarke and Sherrill-Mix, 2017), ggsignif (Ahlmann-Eltze and Patil, 2021), scales (Wickham and Seidel, 2022), extrafont (Chang, 2022), and RColorBrewer (Neuwirth, 2022).

2.11 FIJI/ImageJ

FIJI/ImageJ is an extremely powerful image processing application created initially by Wayne Rasband and now largely maintained by Curtis Reuden with support from the National Institutes of Health (Schneider, Rasband, and Eliceiri, 2012). Developed primarily in Java, this portable application has been instrumental in image analysis pipelines for over 20 years. Completely free and open-source, ImageJ as well as its successor ImageJ2 and the bundled plugins included with FIJI, as well as the broader community of plugins, are the standard against which any competing applications are compared. This imminently accessible and, obviously, cost-effective means of analyzing images opens this process to labs all over the world (Girish and

Vijayalakshmi, 2004). Through the interpretation of images as numerical pixel values, ImageJ is able to perform nearly every conceivable mathematical operation on images to facilitate their revisualization and interpretation. Due to the numerical infrastructure of scientific imaging data, various aspects of these images can be distilled into various quantitative measurements, which is used throughout this work to analysis the angiogenesis phenotype. While the built-in capabilities of ImageJ have evolved over time, the primary advantage of using ImageJ is access to a robust programming environment based on Java in which to create plugins and macros in a range of Java-compatible programming languages, including Groovy, R, and Python as well as a beginner-friendly IJ1 macro language that, while limited, is able to be written trivially by new users, thanks in part to the excellent macro recorder that will translate GUI-based interactions into a set of executable macro commands. For instance, a basic script to measure thresholded pixel intensity within a user-defined area and record it to a ResultsTable is a simple as:

```
setThreshold(600, 4096);
// setTool("freehand");
run("Measure");
close();
```

This work has necessitated the development of a range of these scripts and plugins, which have serviced the analysis of these images in the background both through automating repetitive processing tasks (Z projections, channel splitting, channel merging, file saving, lookup table application, etc.) and extracting quantitative data from these images in a high-throughput fashion. These can all be found in Chapter 4. In a number of instances, I was forced to translate several Java plugins into Python

to allow for streamlined access to the underlying API; these are also described in Chapter 4.

New, user-friendly RNA-seq analysis pipelines were developed in Python around the combination of Kallisto (Bray et al., 2016) and Sleuth (Pimentel et al., 2017) to facilitate analysis and visualization of the data in Saelens et al. (2022). This analysis pipeline is presented in Section 4.9.

Image analysis was conducted using the FIJI (Schindelin et al., 2012; Rueden et al., 2017) expansion of ImageJ (Schneider, Rasband, and Eliceiri, 2012). Analysis pipelines were written in Jython (v.2.7.2) (van Rossum, 1995) and executed within the ImageJ Jython interpreter. All scripts are provided via Zenodo at the doi listed above. Additional description of the use of FIJI/ImageJ can be found in Section 4.1.

Chapter 3

Macrophage NFATC2 Mediates Angiogenic Signaling During Mycobacterial Infection¹

3.1 Abstract

As introduced in the previous chapters, pathogenic mycobacteria manipulate the host immune response to generate a productive infection that enables them to transmit to new hosts. To do so, they must both subvert protective immune responses and agonize the pathways that provide them benefit in the way of nutrients, susceptible host cells, or routes of transmission. This necessitates the engagement of both the immune cells themselves and the surrounding stromal cells, all of which much be subverted to generate the optimal pro-bacterial response within the limits of bacterial control of host physiology. One of the mechanisms by which this has been found to occur is through a specific modification of the specialized cell wall lipid, trehalose 6-6'-dimycolate (TDM). This mycobacterial glycolipid has been found to direct angiogenesis

¹Most of the data in this chapter is from W. J. Brewer et al. (2022b). “Macrophage NFATC2 Mediates Angiogenic Signaling During Mycobacterial Infection”. In: *Cell Rep* 41.111817, text extensively modified.

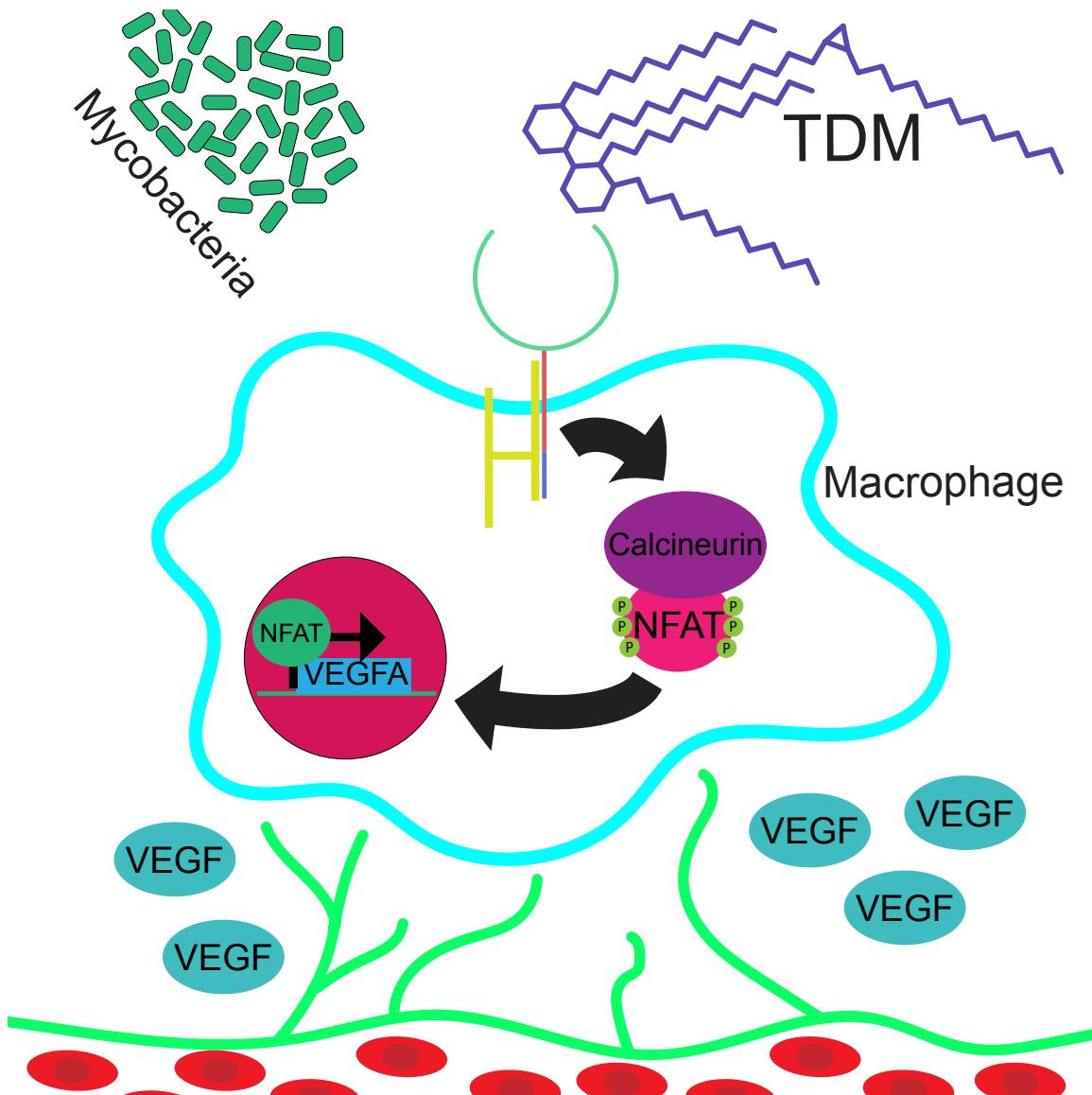


Figure 3.1: This is a graphical representation of the work completed here, which found a novel role for NFAT activation downstream of macrophage-mycobacterial surface interactions that facilitated the production and secretion of VEGFA to drive angiogenesis.

toward nascent granulomas to enhance overall bacterial burden. The present study utilizes the zebrafish-*Mycobacterium marinum* infection model to define the signaling basis of the host angiogenic response. Through intravital imaging and targeted, cell-specific peptide-based inhibition, I have identified macrophage-specific activation of NFAT signaling as essential to TDM-mediated angiogenesis *in vivo*. Exposure of human cells to *Mycobacterium tuberculosis* results in robust induction of VEGFA that correlates with and is dependent on a signaling pathway downstream of host TDM detection and culminates in NFATC2 activation. As granuloma-associated angiogenesis is known to serve bacterial-beneficial roles, these findings identify potential host targets to improve tuberculosis disease outcomes.

3.2 Introduction

The host rejoinder to infection is driven by an intricately regulated, but occasionally discordant or maladaptive, immune response to pathogenic stimuli at the cell-intrinsic, innate, and adaptive levels (MacMicking, 2004; Finlay and McFadden, 2006; Iwasaki and Medzhitov, 2010; MacMicking, 2012; Kim et al., 2012; Haldar et al., 2015; Wilburn et al., 2022). Although an inflammatory immune response is essential to host survival and pathogen killing, an overly robust response is clearly deleterious to the host, as seen in sepsis (Casadevall and Pirofski, 2003; Finethy et al., 2020). However, as we have seen in previous chapters, the inflammatory response can also serve pro-bacterial purposes, rendering the immune response impotent. The contributions of immune cells to host defense have been widely studied, but there is growing appreciation for the contributions non-immune populations, including stromal cells and the endothelium (Honan and Chen, 2021; Worrell and MacLeod, 2021; Honan and Chen, 2021; Amersfoort, Eelen, and Carmeliet, 2022) make in shaping

the host response to both acute and chronic infections (Mueller and Germain, 2009; Rando, MacMicking, and James, 2013; Krishnamurty and Turley, 2020). Pathogens have long been known to possess sophisticated mechanisms to undermine signaling pathways in immune cells and, more recently, have been shown to manipulate development and homeostatic tissue processes to force them toward pathogen-beneficial states (Menzies and Kourteva, 1998; Guichard et al., 2013).

Mycobacterium tuberculosis is among history's most widespread and successful pathogens. It has evolved an array of sophisticated mechanisms that manipulate its human host to enable bacterial survival, replication, and transmission. Upon infection, *M. tuberculosis* sets in motion an intricate immune response wherein innate immune cells, consisting initially of macrophages, congregate at the bacterial focus and then undergo an epithelioid transformation and interdigitate to form an encased granuloma, the hallmark feature of tuberculosis (Subsection 1.2.6), which provides both the replicative niche and the major host-pathogen interface of tuberculosis disease (Cronan et al., 2016; Pagan and Ramakrishnan, 2018; Cronan et al., 2021). Granuloma-associated vasculature has long been noted in human and animal models of tuberculosis (Cudkowicz, 1952; Russell, Barry, and Flynn, 2010) but the mechanisms of induction and precise contributions to infection are not yet fully understood (Subsection 1.5.6).

Many of the major pathological features of mycobacterial granulomas, including associated vascularization, are conserved from zebrafish to humans (Swaim et al., 2006; Bohrer et al., 2021). Zebrafish can be infected with a natural pathogen, *Mycobacterium marinum*, which induces a robust angiogenic response during granuloma formation in both the larva and adult (Section 2.3). This process, much like that in humans, non-human primates, and rabbits, is associated with production of a pro-

angiogenic chemokine, Vegfaa, at the site of infection (Oehlers et al., 2015). This chemokine has long been known to be a critical regulator of angiogenesis in both developmental and pathological contexts (Leung et al., 1989; Adams and Alitalo, 2007; Chung and Ferrara, 2011). Similarly, human granulomas have been shown to express VEGFA and are physically associated with blood vessels that penetrate the outer granulomatous layers (Guirado and Schlesinger, 2013; Datta et al., 2015). Subsequent work has demonstrated a role for these vessels in supporting bacterial growth and in dissemination of the bacilli from their primary site of infection (Polena et al., 2016). Additional roles for VEGFA in non-angiogenic processes have also been noted, suggesting that angiogenic signaling cascades can also alter the biology of granuloma macrophages to exacerbate disease (Harding et al., 2019). Additional roles have been proposed for the related lymphangiogenesis (Alitalo, Tammela, and Petrova, 2005; Duong, Koopman, and Francois, 2012; Lerner et al., 2020), where pharmacological blockade of these vessels of the lymphatic system also offer host-protective benefits (Harding et al., 2015). Recent profiling of human and non-human primate granulomas have confirmed the presence of aberrant vasculature associated with *M. tuberculosis* granulomas (Cronan et al., 2021; Gideon et al., 2022; McCaffrey et al., 2022), although more complete characterization of the endothelium itself is lacking.

Pathogenic mycobacteria have evolved specialized mechanisms to promote and accelerate angiogenesis and modulate other aspects of endothelial biology (Oehlers et al., 2017). Notably, the extensively modified and essential outer cell envelope component trehalose 6-6'-dimycolate (TDM) is cis-cyclopropanated by the enzyme PcaA (Glickman, Cox, and Jacobs, 2000; Rao et al., 2005). Mutation of *pcaA* results in a reduction in granuloma angiogenesis and concomitant reduction in bacterial burden; correspondingly, cyclopropanated TDM alone is sufficient to induce host angiogenesis (Saita et al., 2000; Sakaguchi et al., 2000; Walton et al., 2018) (Subsection 1.5.6). As

pcaA-dependent vascularization supports bacterial growth, factors driving angiogenesis represent potential sites of therapeutic intervention yet the signals that mediate this host process remain unclear.

TDM is an extraordinarily long-chain, hydrophobic (C_{60} - C_{100}) glycolipid (Noll et al., 1956; Noll, 1956; Behling et al., 1993; Hunter, Venkataprasad, and Olsen, 2006) that has been shown to be detected in cell culture and murine models by host C-type lectin receptors, most notably MCL (CLEC4D) and MINCLE (CLEC4E), as well as by Toll-like receptor 2 (TLR2), CD14, and MARCO (Bowdish et al., 2009; Matsunaga and Moody, 2009; Ishikawa et al., 2009; Miyake et al., 2013) (see also Subsection 1.3.5). Canonically, C-type lectin signaling is transmitted through a CARD9-NF- κ B signaling pathway that results in the transcription and production of TNF- α , IL-1 β , IL-6 and other cytokines (Yamasaki et al., 2008; Goodridge et al., 2009; Lobato-Pascual et al., 2013; Zhao et al., 2014; Deerhake et al., 2021) (Subsubsection 1.3.3, Section 1.4). However, beyond CARD9, a number of other downstream signaling pathways are engaged by C-type lectin activation and likely control discrete aspects of signaling in largely unappreciated ways (Goodridge, Simmons, and Underhill, 2007; Deerhake et al., 2021).

Here, I synthesize findings from zebrafish and cell culture models to define the *in vivo* angiogenic response induced by pathogenic mycobacteria. Contrary to classical models of C-type lectin signaling, I find that cis-cyclopropanated TDM exerts its pro-angiogenic effects through an alternative NFAT-driven pathway rather than canonical CARD9-NF- κ B signaling. I use peptide-mediated, cell-specific inhibition of NFAT to demonstrate that both early and mature granuloma angiogenesis are dependent upon macrophage-NFAT signaling. I identify *NFATC2* as the predominant isoform mediating *VEGFA* induction and angiogenesis *in vitro* and *in vivo*. These findings

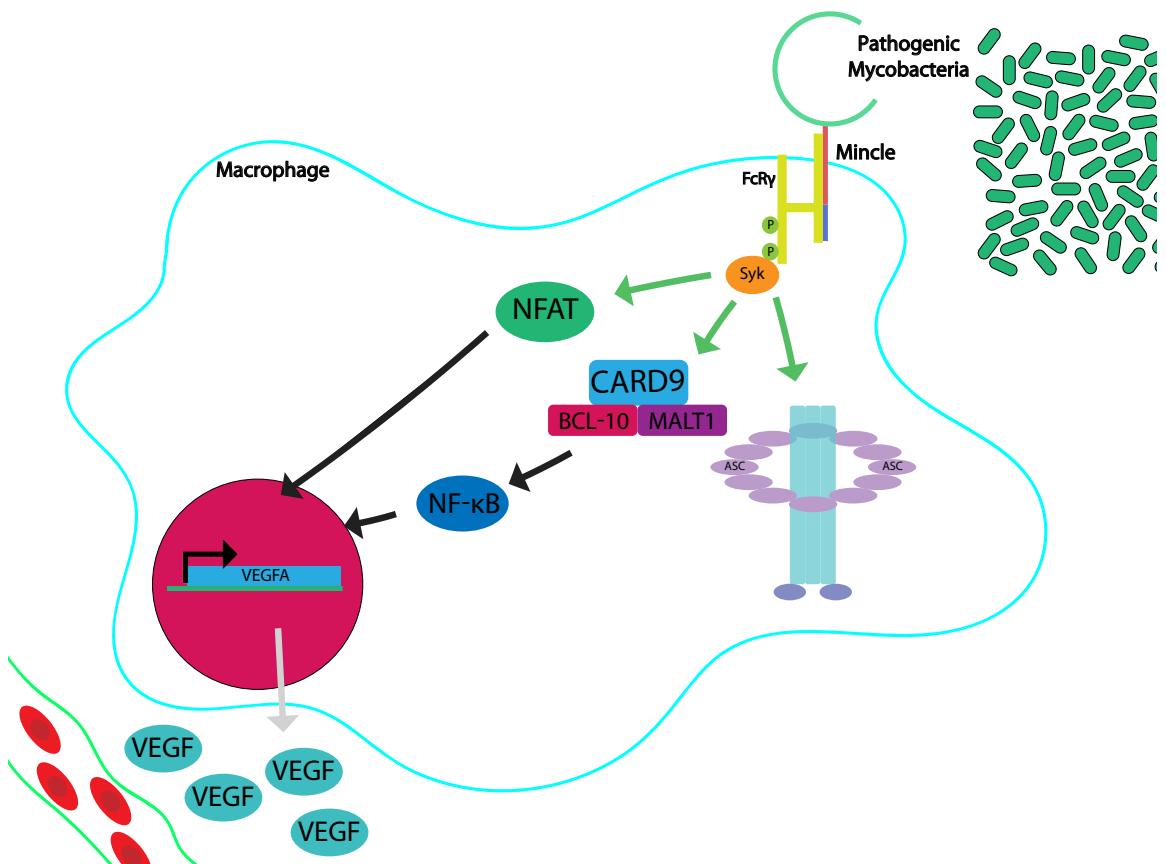


Figure 3.2: The central hypothesis of this work is that a discrete pathway downstream of C-type lectin receptor activation is mediating the induction of VEGFA within macrophages to drive angiogenesis during mycobacterial infections. This schematic displays the hypothesized pathway options that could be playing these roles: CARD9, NFAT, or inflammasome activation. This work sought to dissect these possibilities to better understand the mechanism of angiogenesis during tuberculosis infection.

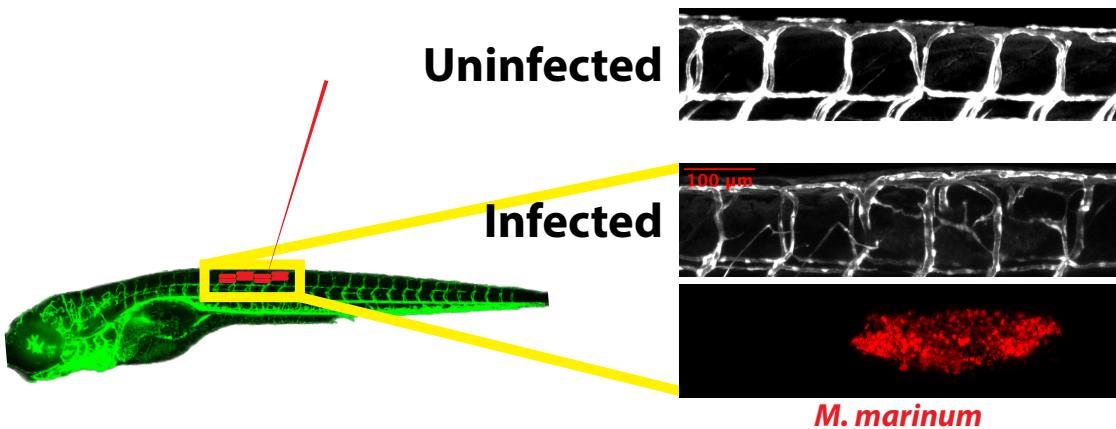


Figure 3.3: Pictorial representation of the site of infection and angiogenic outcomes from infection. After injection into a peri-notochordal space along the trunk of the fish, the bacteria replicate and blood vessels grow toward the site of infection, as compared to the uninfected reference.

define the basis of granuloma-associated angiogenesis during pathogenic mycobacterial infections and suggest new targets for host-directed therapeutic interventions during tuberculosis.

3.3 Results

3.3.1 Macrophage Induction of *vegfaa* and Angiogenesis during Mycobacterial Infection

Injection of live *Mycobacterium marinum* into the dorsal trunk of the zebrafish larva is sufficient to induce a robust angiogenic response adjacent to nascent granulomas in a macrophage-dependent manner (Oehlers et al., 2015) (Figure 3.3). The stereotyped vasculature along this region of the larva allows facile quantitation of neovascularization during and after granuloma formation or other insult (Lawson and Weinstein, 2002; Jin et al., 2005; Gore et al., 2012; Matsuoka and Stainier, 2018) (Subsec-

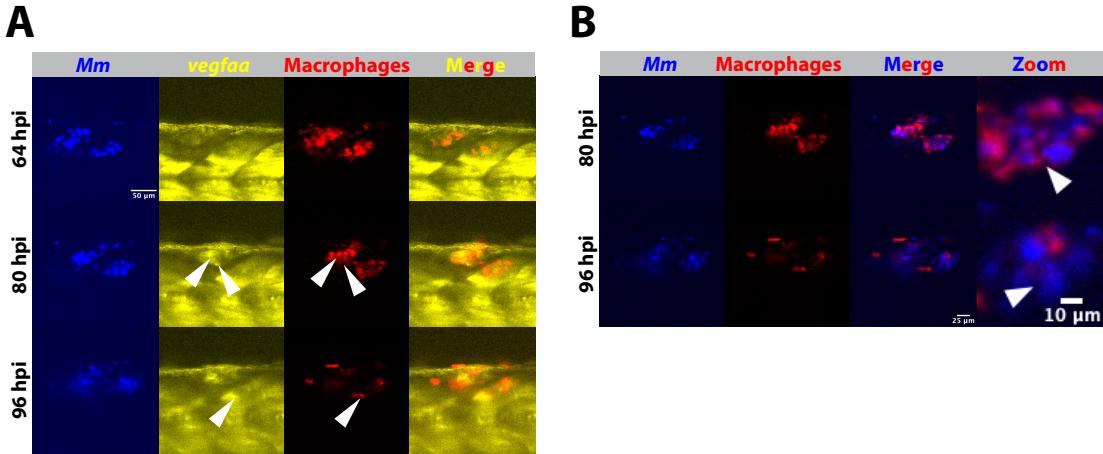


Figure 3.4: Around 60 hours post infection, macrophages interacting with the extracellular bacteria at the site of infection begin to express *vegfaa*, as reported by the *vegfaa:eGFP^{pd260}* transgene. In (A), a time series can be seen showing *vegfaa*-expressing macrophages and in (B), macrophages can be seen engaging with what appears to be extracellular bacteria that has accumulated over the course of the infection.

tion 2.2.4). Our lab has previously demonstrated that cis-cyclopropanated trehalose 6-6'-dimycolate (TDM) is required for the induction of *vegfaa* and angiogenesis at the site of infection. Furthermore, we found that genetic blockade of Vegfaa signaling was sufficient to abolish angiogenesis during infection with wild-type mycobacteria (Walton et al., 2018). Taken together, these findings suggest that the failure to induce *vegfaa* is a primary contributor to the loss of angiogenesis in *pcaA*-deficient granulomas.

To study this phenomenon further, I began by examining the kinetics of *vegfaa* induction to identify the cellular source of Vegfaa during granuloma formation. To test whether macrophages were playing this role, Eric Walton developed a macrophage-specific reporter using the previously described *acod1* promoter (also known as *irg1*), Tg(*irg1:tdTomato^{xt40}*) (from here, *irg1:tdTomato*). *irg1* has been found to be expressed specifically in zebrafish macrophages and is upregulated during infec-

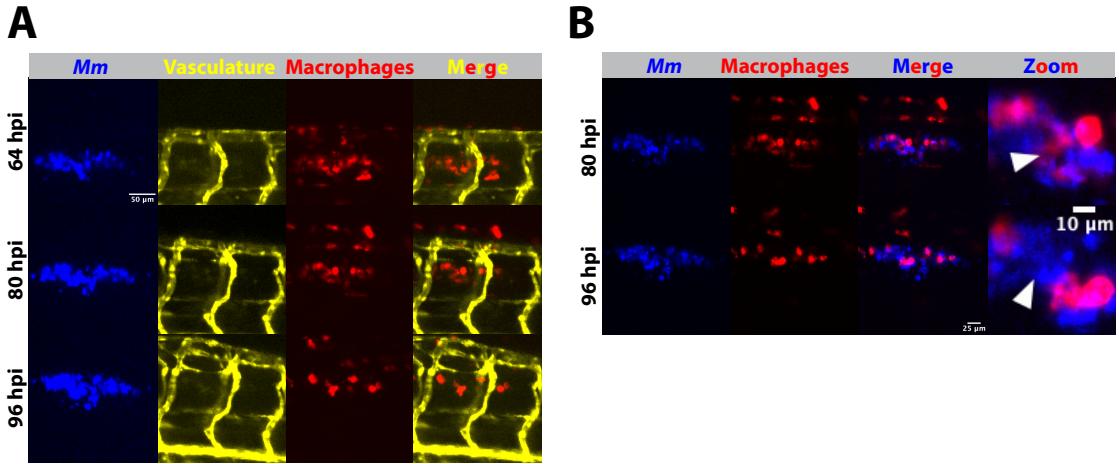


Figure 3.5: At later time points, after *vegfaa* induction, blood vessels proliferate and begin to grow toward the site of infection in association with nearby macrophages. (A) shows this process occurring over the time period studied and culminating in a new vessel nearly spanning the intersomitic vasculature. (B) displays the accumulation of extracellular bacteria and macrophage killing over this time period.

tion (Sanderson et al., 2015; Kwon et al., 2022). I then crossed this line with the *vegfaa* reporter line TgBAC(*vegfaa:eGFP^{pd260}*) (*vegfaa:eGFP* throughout) (Karra et al., 2018) and infected double transgenic *irg1:tdTomato; vegfaa:eGFP* progeny with *M. marinum* expressing eBFP2 (*Mm-eBFP2*) to simultaneously visualize bacteria, macrophage localization, and *vegfaa* production *in vivo* (Takaki et al., 2013).

Imaging was commenced at a time point that preceded robust induction of *vegfaa:eGFP* but would allow us to capture the maximum time span of these events due to difficulty in keeping larvae alive for longer courses of embedded imaging. I observed an increase in *vegfaa* reporter signal over time that appeared largely localized to macrophages (Figure 3.4A). I observed that bacteria initially grew primarily intracellularly within individual macrophages at 36 hours post infection but began to grow in characteristic extracellular cords by approximately 84 hours post infection with little to no intracellular containment at this site by 96 hours post infection (Figure 3.4B). This cording phenomenon is comparatively accelerated in the trunk

compared to traditional caudal vein infection, perhaps due to the lack of vascularity in this muscular region. The increase in extracellular growth coincided with the induction of eGFP signal in macrophages at 64 hours (Figure 3.4A), suggesting that, at low overall burden, intracellular detection is unable to induce *vegfaa* expression while extracellular engagement correlates with *vegfaa* expression during early stages of granuloma formation (Figure 3.4).

I next visualized the production of angiogenic vessels throughout infection in parallel to our characterization of *vegfaa* induction. Due to an inability to separate discrete emission wavelengths using two GFP reporter lines, I was unable to examine all four components (bacteria, *vegfaa* induction, macrophages, and vasculature) simultaneously. To relate this process directly to the angiogenesis observed in mycobacterial granulomas, I crossed the *irg1:tdTomato* macrophage reporter to the Tg(*kdr1:eGFP^{s843}*) (from here, *flk1:eGFP*) line, which labels vascular endothelium (*flk1:eGFP; irg1:tdTomato*) (Jin et al., 2005). Under the same conditions and burden at which I infected the *vegfaa* and macrophage dual reporter line, I observed robust vascularization at approximately 96 hours post-infection, subsequent to initial granuloma formation and *vegfaa* induction (Figure 3.5A, B).

3.3.2 Genetic *card9* Deficiency Does Not Compromise Angiogenesis During Mycobacterial Infection

Given these observations suggesting that macrophages engaging extracellular bacteria are an important source of *vegfaa* expression, I identified pattern recognition receptor (PRR) signaling pathways that had been implicated in the literature in host responses to TDM, a major external component of the mycobacterial cell envelope. We had previously found that *myd88* was dispensable for the induction of angiogenesis in

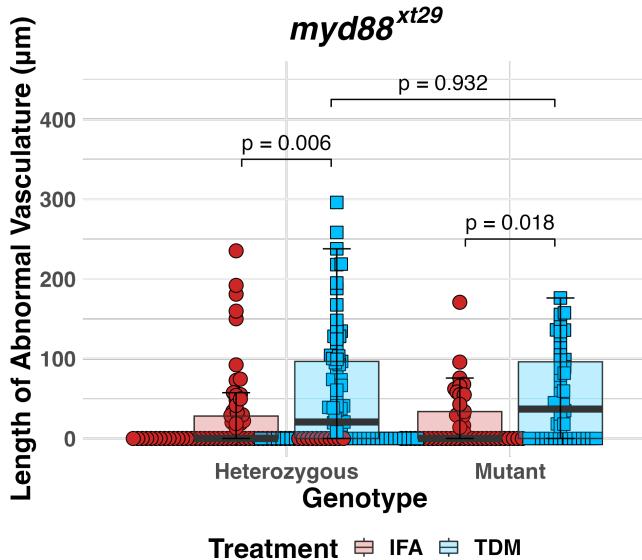


Figure 3.6: *myd88* is dispensable for angiogenesis during infection, suggesting a TLR- and IL-1R-independent signaling pathway is necessary for inducing angiogenesis downstream of TDM detection. Larval zebrafish were injected with IFA or TDM, imaged and quantitated, and then matched to genotype. Adapted from personal contributions to Walton et al. (2018).

response to TDM *in vivo* (Figure 3.6) (Bowdish et al., 2009; Walton et al., 2018). This suggested that the described TLR2- or IL-1R-mediated responses that function downstream of TDM detection in some contexts were unlikely to be required for this process². Rather, we found that the Fc γ R homologs in zebrafish, *fcer1g* and *fcer1gl*, are required for the full angiogenic response to TDM (Walton et al., 2018), implicating C-type lectin receptors signaling in mediating this response (Richardson and Williams, 2014; Zhao et al., 2014). I then utilized a well-characterized inhibitor of a downstream kinase in this pathway to investigate the role of SYK in mediating these responses. As expected, treatment of larval zebrafish with BAY 61-3606 resulted in an abolition of TDM-dependent angiogenesis and providing pharmacological evidence of the importance of C-type lectin signaling responses in mediating the angiogenic

²This could not, however, rule out any potential role for CD14-mediated signaling independent of TLR2. One issue is that the zebrafish do not have an identified CD14 homolog, but additional data clarifies things somewhat.

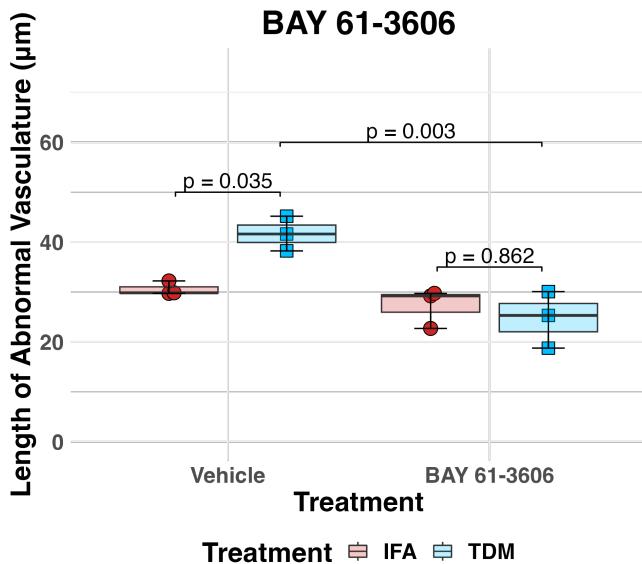


Figure 3.7: BAY 61-3606, a pre-clinical inhibitor of spleen tyrosine kinase (SYK), a necessary adaptor protein downstream of C-type lectin signaling, prevents angiogenesis in response to TDM. Each data point represents the mean of an individual experiment.

effects of TDM (Figure 3.7) (Ghotra et al., 2015; Xu et al., 2018b).

As many of the downstream activities of C-type lectin receptors have been ascribed to the activation of CARD9-NF- κ B signaling (Goodridge et al., 2009; Lobato-Pascual et al., 2013; Zhao et al., 2014; Williams, 2017; Deerhake et al., 2021), I assessed what role this pathway might play in angiogenesis during mycobacterial infection. Eric Walton developed a *card9* knockout zebrafish line using CRISPR/Cas9 that carries a 28 bp insertion, resulting in an early stop after 59 amino acids (*card9^{xt31}*) (Figure 3.8E). I then assayed these animals in the *flk1:eGFP* transgenic background by incrossing *flk1:eGFP*; *card9^{xt31/+}* animals and infecting the resulting offspring with tdTomato-fluorescent *M. marinum* (*Mm*-tdTomato) at 2 days post fertilization (Figure 3.8A, B) (Jin et al., 2005; Oehlers et al., 2015). I quantitated the resulting aberrant vasculature at 4 days post-infection under genotypic blinding and post hoc matched these measurements to genotype. To my surprise, there were no significant

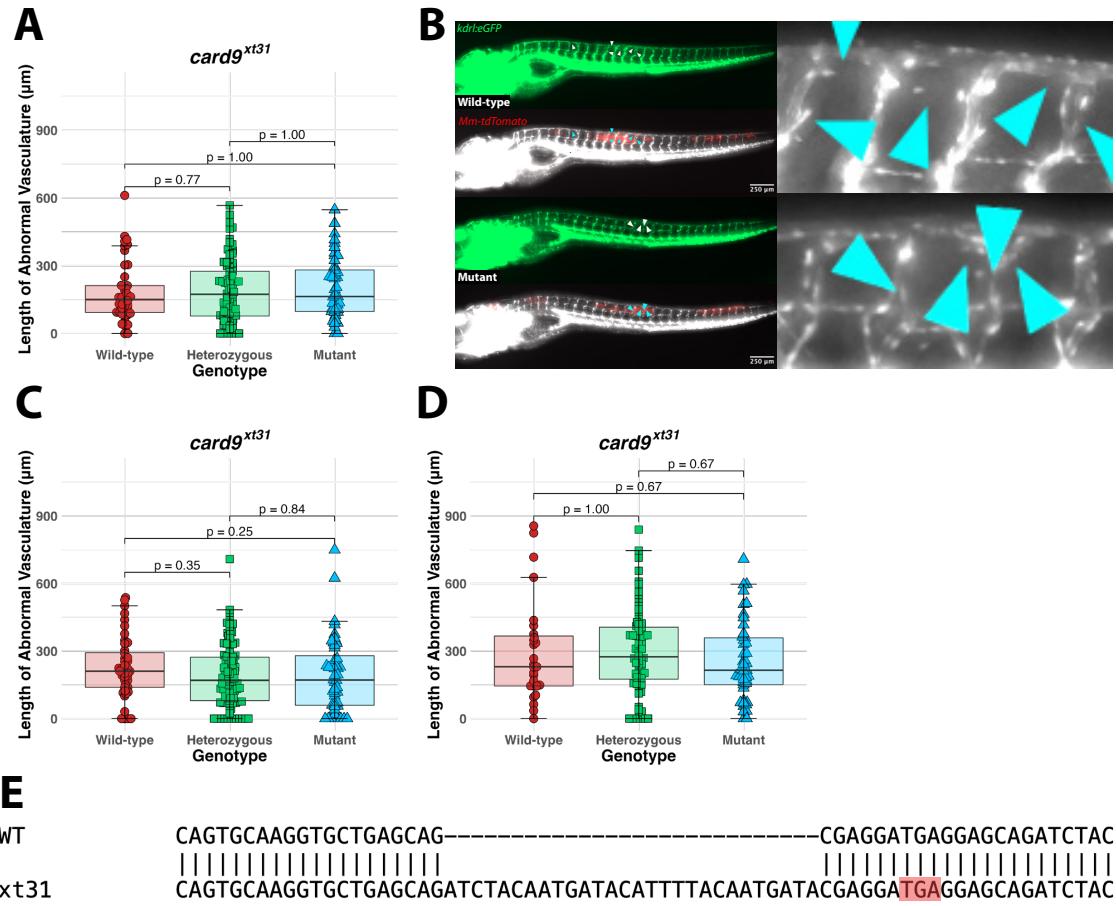


Figure 3.8: *card9*, the canonical downstream adaptor from C-type lectin signaling, is not required for angiogenesis during mycobacterial infection. (A), (C), and (D) show three replicates of this experiment from genotype-blind experiments. (B) shows a representative example of the effect seen, where no differences can be ascertained between genotypes. (E) displays the 28 base pair insertion generated, with the new early stop codon highlighted in red.

differences between the three genotypes across three independent replicates (Figure 3.8A, C, D), suggesting either redundancy between multiple established pathways or the existence of an alternative pathway downstream of TDM detection that was *fcer1g/fcer1gl*-dependent, but independent of both *myd88* and *card9*. Future work should more comprehensively assess the contributions of *card9* to other aspects of mycobacterial control.

As C- type lectin receptors are also known to activate inflammasome formation, I sought to assess whether non-canonical³ functions of the inflammasome may be required for the angiogenic response. We thus utilized an *asc* knockout line (*asc*^{w216}, which abolishes inflammasome nucleation and subsequent processing of IL-1 β , IL-18⁴, and Gasdermin D⁵). Similar to previous assays, I incrossed heterozygotes of *asc*^{w216} in the *flk1:eGFP* background (*flk1:eGFP; asc*^{w216}) and injected the resulting offspring with TDM or the IFA vehicle. I found that *asc*^{w216/w216} zebrafish were competent for angiogenesis in response to TDM (Figure 3.9), suggesting that this pathway was also dispensable for angiogenesis and narrowing our focus onto the NFAT signaling pathway.

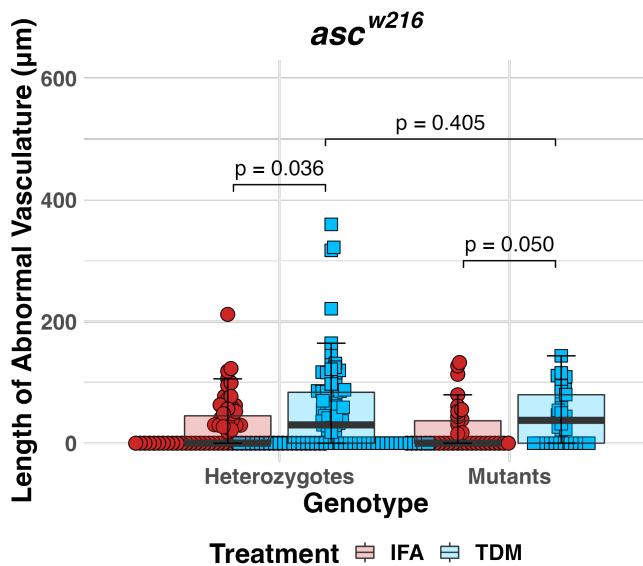


Figure 3.9: *asc*, the critical downstream adaptor protein for the formation of the inflammasome, is not required for angiogenesis in response to purified mycobacterial TDM.

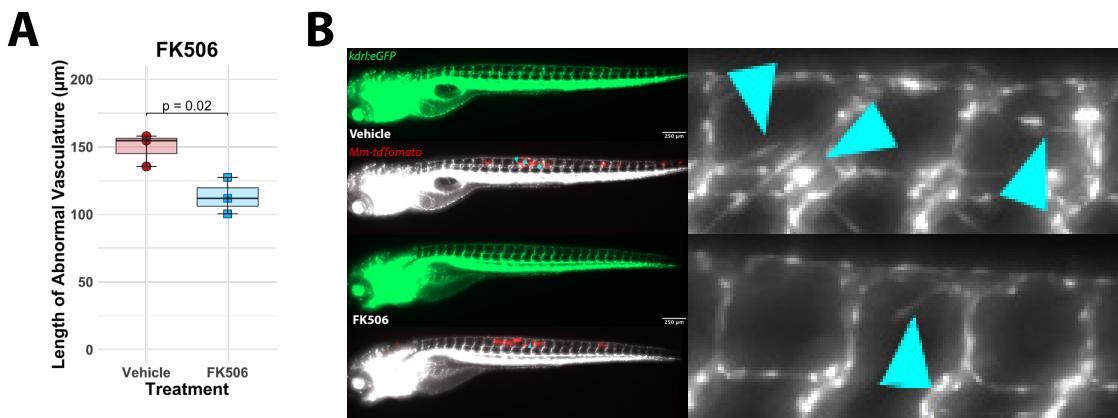


Figure 3.10: Treatment of infected larval zebrafish with the calcineurin-NFAT inhibitor FK506 at 125nM results in a reduction in infection-induced angiogenesis, suggesting a role for this pathway in inducing angiogenesis during infection. (A) Quantitation of this effect, where each data point represents the mean value from a single independent experiment. (B) Representative images of the degree of angiogenesis seen within and without FK506 treatment.

3.3.3 Pharmacological Inhibition of NFAT Activation Limits Angiogenesis in Response to Mycobacteria and TDM

Although many of the physiological consequences of C-type lectin receptor induction are often ascribed to CARD9-NF- κ B signaling, this PRR class is also known to activate a distinct transcription factor family with known roles in immunity – the nuclear factor of activated T cells, or NFAT (Goodridge, Simmons, and Underhill, 2007; Deerhake et al., 2021). This calcium-responsive transcription factor pathway is best described in its role regulating T cell biology, but there are numerous reports describing various roles for the members of this pathway in other cell types, including macrophages (see Section 1.4) (Symes, Gearan, and Fink, 1998; Jones, Conover, and Symes, 2000; Crabtree and Olson, 2002; Horsley and Pavlath, 2002; Elloumi et al., 2012). Given that there are four mammalian members of this pathway and six zebrafish homologs with potentially overlapping functions, I began with a pharmacological approach to globally inhibit NFAT signaling through all six zebrafish isoforms. Although this comes with caveats with specificity – as described in Subsubsection 1.4.2, NFATC1 has important roles in angiogenesis within the endothelium – it offers an opportunity to assess the general roles of this pathway as a first-pass approach.

³Based on the *myd88* data, we already knew at this point that IL-1 β signaling was dispensable for angiogenesis as IL-1R-dependent responses rely on MYD88 for intracellular signal transduction.

⁴Zebrafish do not have an annotated IL-18, although they may have some analogous that has not yet been identified.

⁵Given the known roles for adenosine receptor signaling in inducing angiogenesis, it was possible that inflammasome-mediated cell death pathways could be indirectly responsible for inducing angiogenesis, although this does not appear to be the case in this instance (Dusseau, Hutchins, and Malbasa, 1986; Montesinos et al., 2002; Clark et al., 2007; Auchampach, 2007).

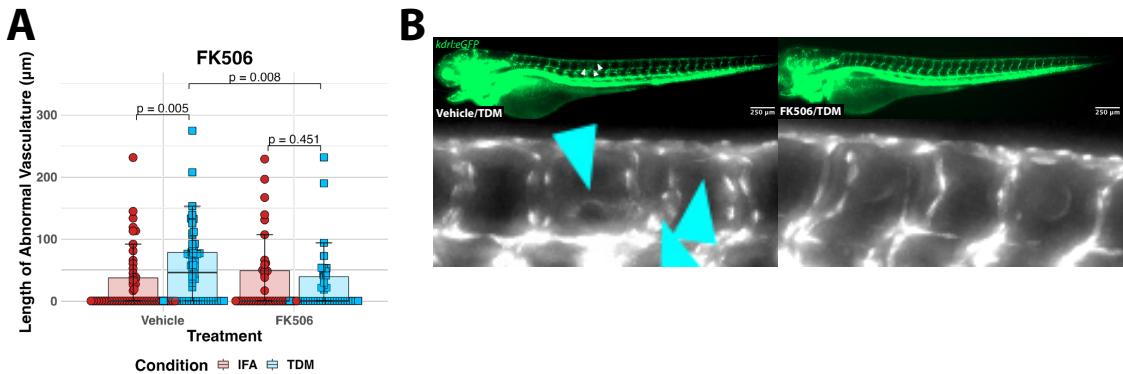


Figure 3.11: Treatment of infected larval zebrafish with the calcineurin-NFAT inhibitor FK506 at 125nM results in a reduction in TDM-induced angiogenesis, suggesting a role for NFAT signaling in inducing TDM-dependent pro-angiogenic signaling. (A) Quantitation of the angiogenesis seen in these assays. A subset of the total data points are shown for clarity and statistics are from the subset displayed. (B) Representative images from the TDM-injected groups showing a reduction in angiogenesis after FK506 treatment.

I first infected 2 days post fertilization *fkl1:eGFP* larval zebrafish with *Mm*-td-Tomato in the trunk and treated them with 125 nM FK506, a clinically utilized calcineurin inhibitor that blocks NFAT activation, for the duration of the experiment (Ellis, 1995) (Subsection 1.4.1). This minimal dose of FK506 was chosen due to developmental toxicities observed at higher doses in my hands and is in line with the dosage used by others (Kujawski et al., 2014). I imaged the fish at 4 days post infection and quantitated the degree of vasculature induced in the presence and absence of inhibitor under computational blinding. Even with at this low dose of FK506, I noted a small, but statistically significant reduction in the mean degree of neovascularization at this time point, consistent with a role for NFAT in controlling angiogenesis in response to *M. marinum* infection (Figure 3.10A, B) (Kujawski et al., 2014). To ask whether this effect was specific to recognition of TDM, I injected purified TDM or vehicle (incomplete Freund's adjuvant; IFA) alone into the trunks of 2 days post fertilization larvae. Treatment with FK506 resulted in a statistically significant reduction in the

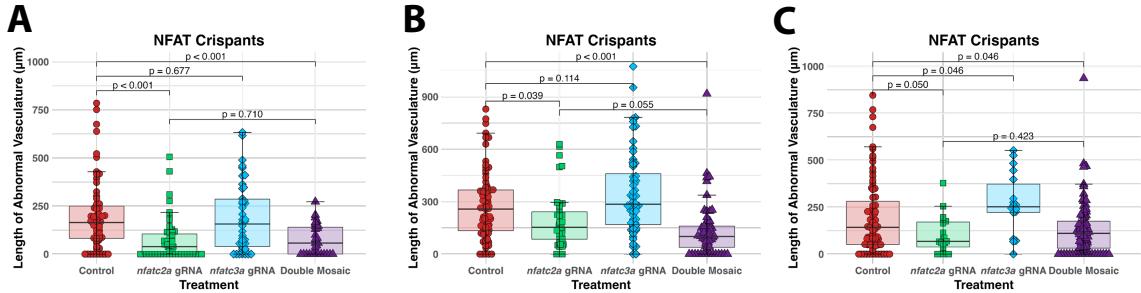


Figure 3.12: Assays in mosaic, CRISPR/Cas9 RNP-injected larval zebrafish suggested that *nfatc3a* was dispensable for the induction of angiogenesis during infection. Panels (A), (B), (C) show the results of three replicates of these mosaic injections.

degree of angiogenesis induced at 2 days post injection (Figure 3.11A, B), suggesting that this pathway was specifically relevant to TDM-dependent angiogenesis.

3.3.4 The Isoform *nfatc2a* is Specifically Required for Angiogenesis During Infection

Combining our observations on the correspondence of granuloma formation and the induction of *vegfaa* with our data implicating the NFAT pathway, I sought to identify NFAT isoforms that were enriched in granuloma macrophage populations. Aside from investigations made into *nfatc1*, which is restricted to the endocardium, lymphatic vessels, and the notochord during much of zebrafish development (see Subsubsection 1.4.2) (Pestel et al., 2016; Shin et al., 2019; Bagwell et al., 2020), little is known of the expression patterns of these genes in zebrafish⁶, especially in the context of infection. I first made use of published scRNA-seq datasets from mycobacterial granulomas in zebrafish and non-human primates for *nfat* transcripts that were robustly expressed in granuloma macrophages at the population level and identified both zebrafish *nfatc2a* and *nfatc3a* as plausible candidates (Cronan et al., 2021; Gideon et

⁶As mentioned previously in Subsubsection 1.4.2, the NFATC4 isoform is thought to not be expressed in the hematopoietic compartment, an observation backed up by some more recent data as well (Peuker et al., 2022).

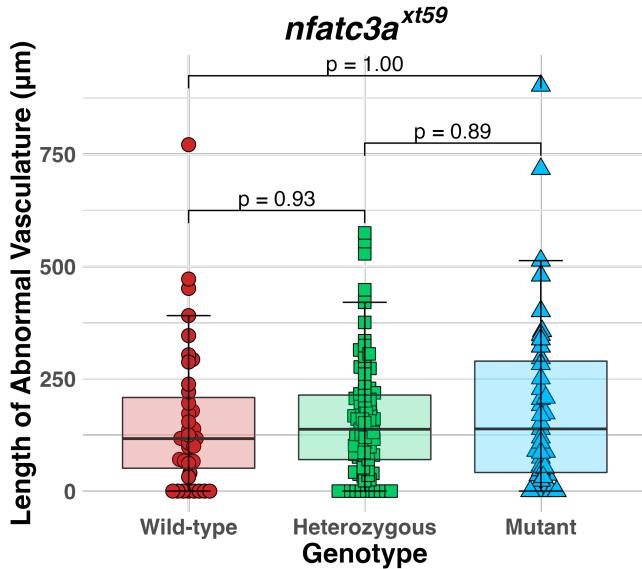


Figure 3.13: (A) *nfatc3a*, which is prominently expressed in granuloma macrophages, is not required for angiogenesis during mycobacterial infection. (B) Sequence alignment of the mutation present in *nfatc3a^{xt59}* compared to the reference sequence.

al., 2022). I also leveraged other existing datasets to identify *nfatc2a* and *nfatc3a* as the most highly expressed isoforms in adult zebrafish macrophages and in the granuloma based on the RNA-seq dataset in Cronan et al. (2016). This demonstrated substantially higher expression of these two isoforms in resting macrophages and these remained the most highly expressed within the granuloma, consistent with the expression patterns seen in Cronan et al. (2021) (??).

To examine potential roles for *nfatc2a* and *nfatc3a* in granuloma-associated angiogenesis *in vivo*, I first screened F₀ CRISPR-injected mosaic knockouts (“crispants”) to rapidly evaluate these candidate genes (Subsection 2.5.1). Using this approach, similar to that used previously by other groups, I assessed the relative roles of these two isoforms individually and in tandem, measuring the angiogenic response to mycobacterial infection in the *flk1:eGFP* background (Jao, Wente, and Chen, 2013; Hoshijima, Juryneac, and Grunwald, 2016; Wu et al., 2018; Hoshijima et al., 2019;

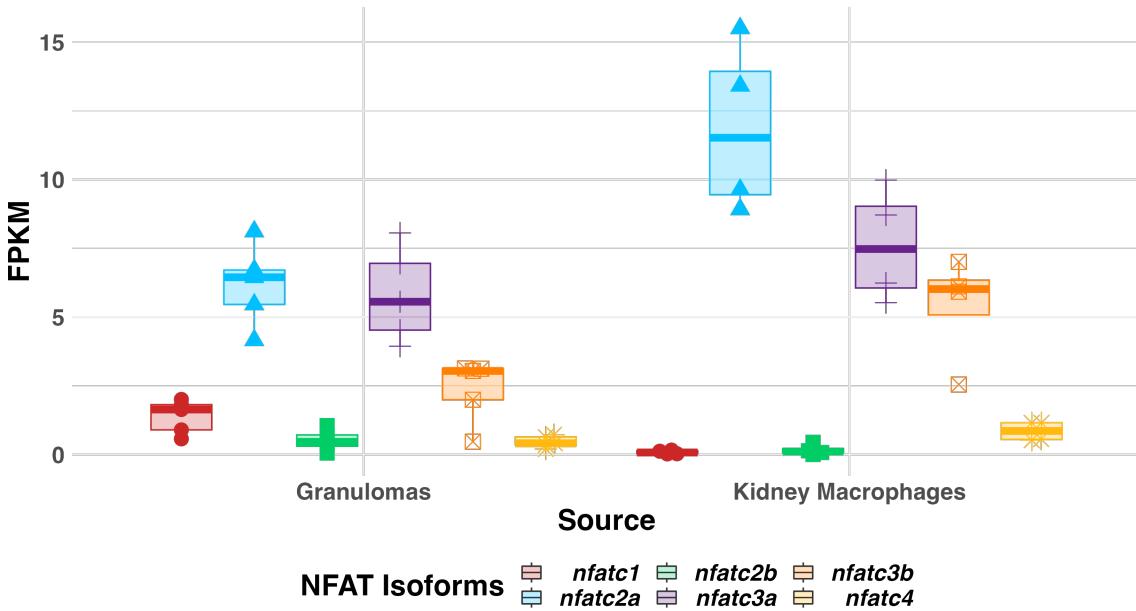


Figure 3.14: Each of the *nfat* proteins in the zebrafish is plotted by FPKM, demonstrating comparatively higher expression of *nfatc2a* and *nfatc3a* than the other isoforms, with *nfatc1*, *nfatc2b*, and *nfatc4* being notably lowly expressed in these populations, helping to narrow on particular candidates for genetic study.

Kroll et al., 2021). I found that *nfatc2a* inhibition resulted in a ~50-80% reduction in angiogenesis (Figure 3.14A-C). In contrast, *nfatc3a* had no effect on the length of ectopic blood vessels present (Figure 3.14A-C). The dual targeted double mosaics were statistically indistinguishable⁷ from the *nfatc2a* injected fish alone (Figure 3.14A-C). This allowed us to prospectively identify *nfatc2a* as an NFAT isoform required for full angiogenic response to mycobacteria while *nfatc3a*, despite expression in overlapping cell populations, appeared to be entirely dispensable for this process at this point in time (Figure 3.14A-C).

I then established stable, germ-line transmitting indel mutant alleles for both genes to validate our results from mosaic animals. Recapitulating our results in the F₀ gen-

⁷In one replicate, this group was near significance as being lower than the *nfatc2a* alone group and this is certainly a plausible effect, but *nfatc2a* was certainly the most prominent effect in these assays.

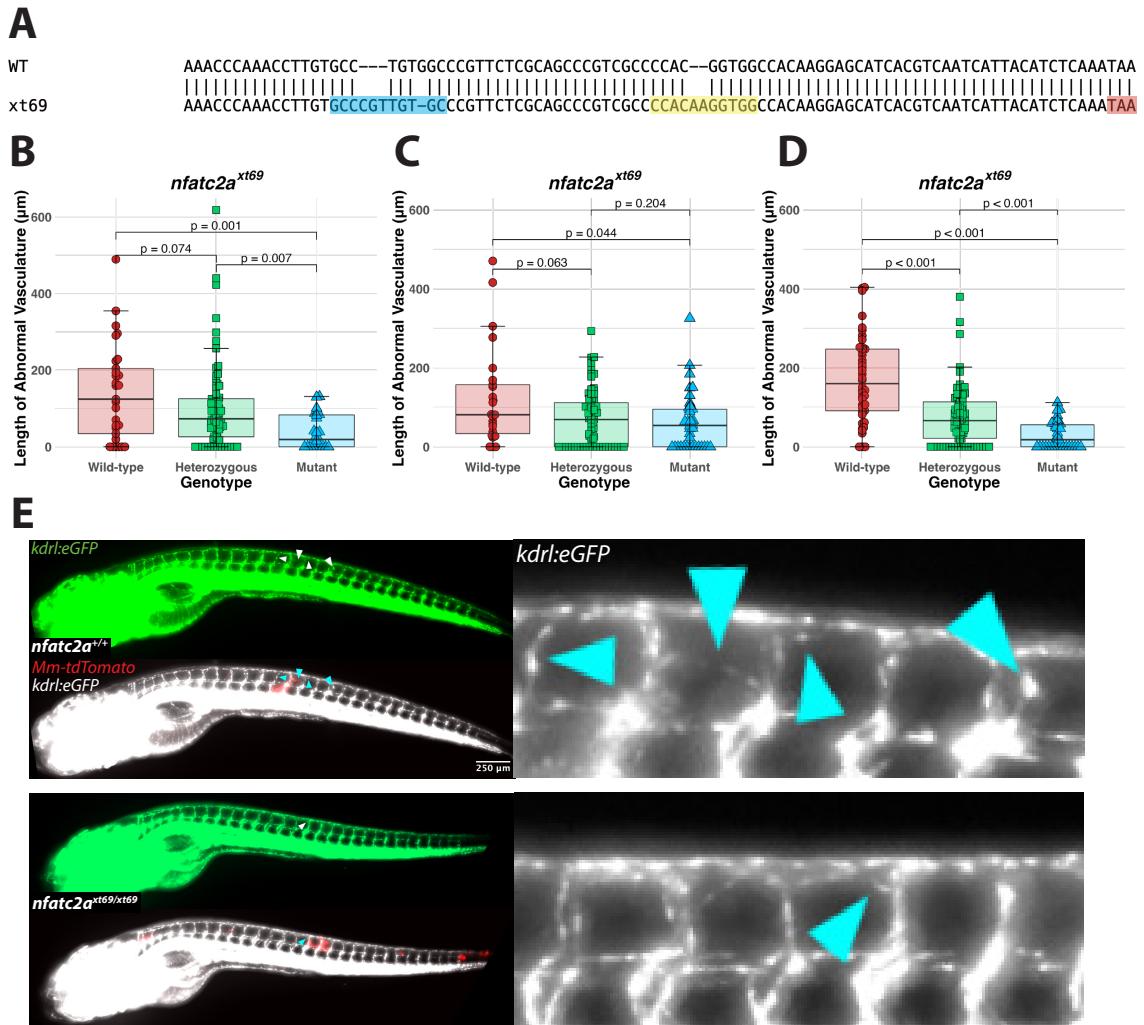


Figure 3.15: *nfatc2a* is required for the angiogenic response to infection. Use of the stable mutant line *nfatc2a^{xt69}* demonstrated an important role for this pathway in inducing angiogenesis. (A) diagrams the mutation I established. (B), (C) and (D) show three replicates of the infection experiment, demonstrating robust and reproducible reduction in angiogenesis in both the heterozygous and mutant conditions relative to wild-type. (E) shows a representative example of the effect seen, with robust angiogenesis under wild-type conditions and a significant reduction with *nfatc2a^{xt69/xt69}*.

eration, the *nfatc3a*^{xt59} mutation carrying a 22 bp deletion (leading to an early stop codon at amino acid 9 in exon 1) had no effect on angiogenesis at 4 days post infection (Figure 3.13A, B). I then developed a knockout line of *nfatc2a* bearing a net 4 bp insertion leading to an early stop codon in the second exon (at amino acid 273, frameshifted after amino acid 247), prior to the DNA-binding domain⁸ (*nfatc2a*^{xt69}) (Figure 3.15A). I repeated our angiogenesis assay using larvae from in-crosses of *flk1:eGFP*; *nfatc2a*^{xt69/+} animals that produced expected Mendelian ratios of wild-type, heterozygous, and homozygous mutant offspring. Consistent with the results from mosaic animals, homozygous knockout of *nfatc2a* was sufficient to reduce the degree of angiogenesis present in larval zebrafish at 4 days post infection (Figure 3.15B-E). Importantly, given the known role of NFAT isoforms in T cell function, these defects emerged prior to the developmental emergence of functional T cells, which does not occur until approximately 6 days post fertilization as the thymus develops (Trede et al., 2004). However, whole animal knockouts could not address potential roles for this gene in cell types other than macrophages in mediating these angiogenic processes.

3.3.5 Macrophage-NFAT is Essential for Angiogenesis

Induction *in vivo*

Given our observations on *vegfaa* induction in macrophages at the granuloma, I tested whether NFAT signaling was required specifically in macrophages for granuloma-associated angiogenesis. For *in vivo* inhibition of macrophage NFAT signaling during infection, I developed an approach that takes advantage of the NFAT-inhibitory pep-

⁸The nature of this mutation makes it plausible that it could be acting in a dominant negative manner against NFAT signaling more broadly, but this is argued against by the results from the mosaic assays, where the sgRNA targeted the first exon, prior to the calcineurin-interacting domain.

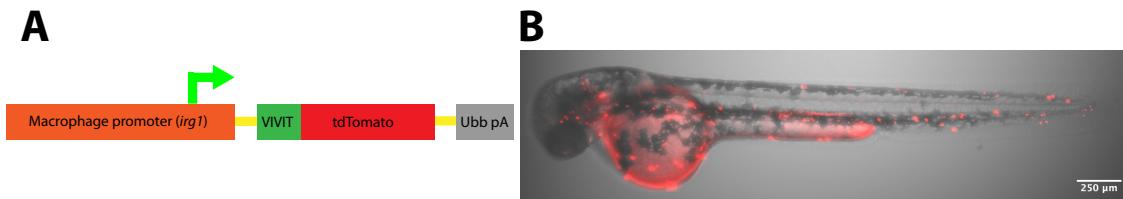


Figure 3.16: Design of the *irg1*:*VIVIT-tdTomato* construct. (A) shows the overall layout of the construct, with *irg1* driving the expression of the VIVIT peptide conjugated to tdTomato. (B) shows a representative *irg1*:*VIVIT-tdTomato*^{xt38} larva.

tide VIVIT which competitively inhibits calcineurin-dependent activation of all the NFATc isoforms (Aramburu et al., 1999). This approach has been successfully used as an exogenous treatment in cell culture (Deerhake et al., 2021) and mice (Noguchi et al., 2004; Elloumi et al., 2012; Rojanathammanee et al., 2015), through ectopic over-expression in cell culture (McCullagh et al., 2004), and, more recently, in transgenic mice (Poli et al., 2022; Peuker et al., 2022). I developed a transgenic zebrafish line in which VIVIT is expressed specifically in macrophages, Tg(*irg1*:*VIVIT-tdTomato*^{xt38}) (from here, simply *irg1*:*VIVIT*) (Figure 3.16A, B) (Sanderson et al., 2015). I assessed whether the macrophage-specific expression of VIVIT would be sufficient to reduce the degree of angiogenesis during infection in the trunk with wild-type *M. marinum* expressing mCerulean (*Mm-mCerulean*). I found that macrophage-specific VIVIT expression significantly reduced angiogenesis in response to infection (Figure 3.18A-D). This suggested a macrophage-specific role for NFAT signaling downstream of mycobacterial detection that was necessary to induce angiogenesis, presumably through the *nfatc2a* isoform.

To ask more directly whether the decreased angiogenesis observed in the NFAT-deficient macrophages was via the TDM-mediated pathway, I applied the TDM injection assay we had developed previously. I injected TDM or the IFA vehicle into the trunk of 2 days post fertilization larval zebrafish and measured the resulting

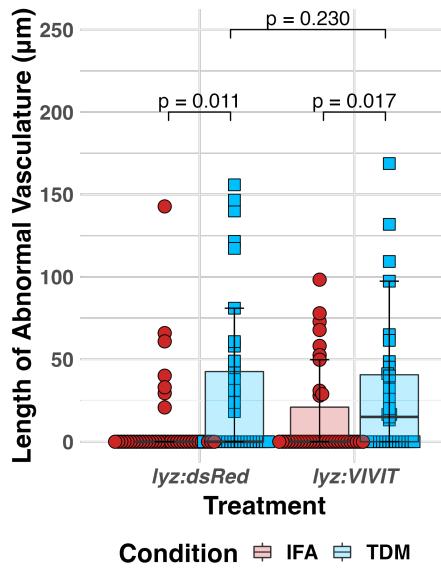


Figure 3.17: Neutrophil-specific VIVIT expression in larval zebrafish has no obvious impact on TDM-induced angiogenesis. Each data point represents a single fish from a subset of 150 larval zebrafish from a single experiment. The TDM assays can be a bit finicky, so these results are likely worth repeating in the context of infection, which would potentially reveal interesting neutrophil-specific activities for NFAT including in systemic infections and within the granuloma.

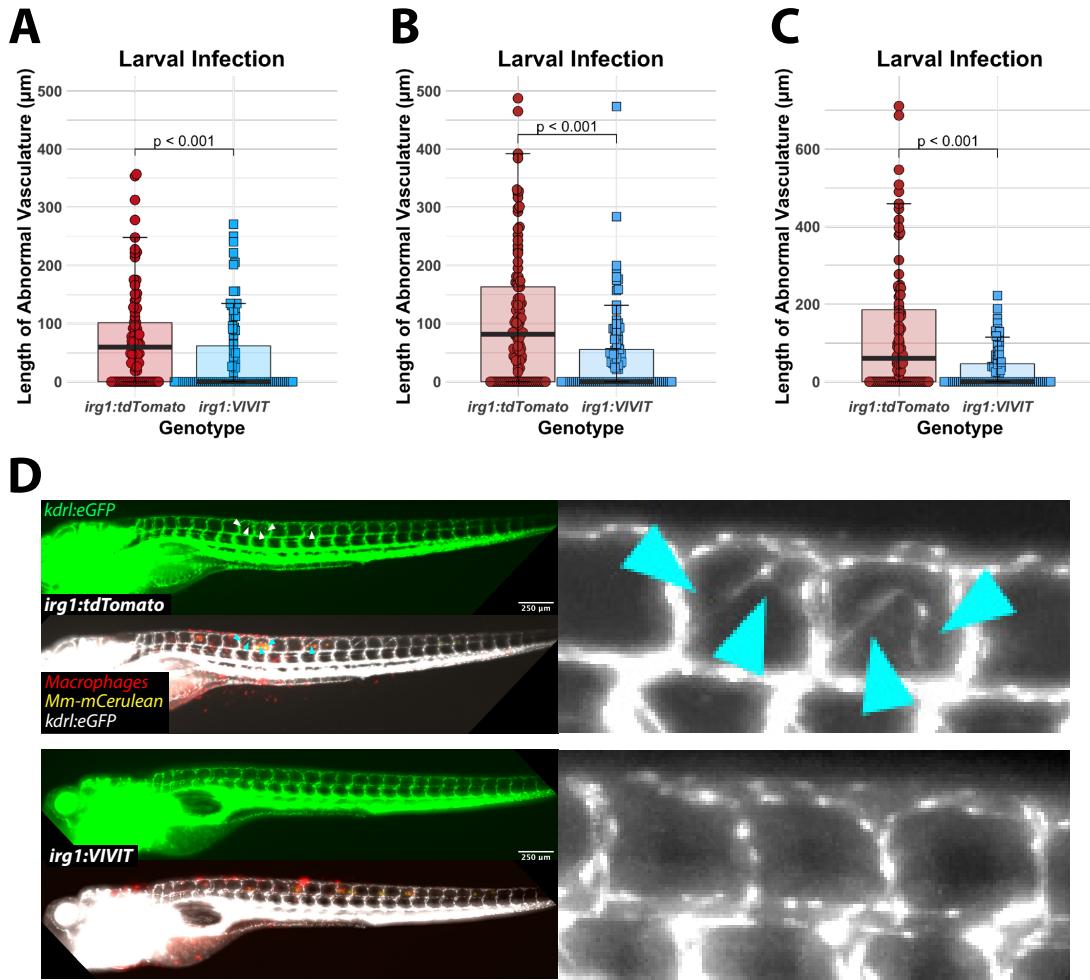


Figure 3.18: Macrophage-specific VIVIT expression in larval zebrafish inhibits infection-induced angiogenesis. (A), (B), and (C) show three replicates of this infection experiment and a robust reduction in the amount of angiogenesis with expression of the VIVIT peptide compared to *irg1:tdTomato*-only controls. (D) shows representative images of the effect observed, with reduced angiogenesis seen in the VIVIT condition.

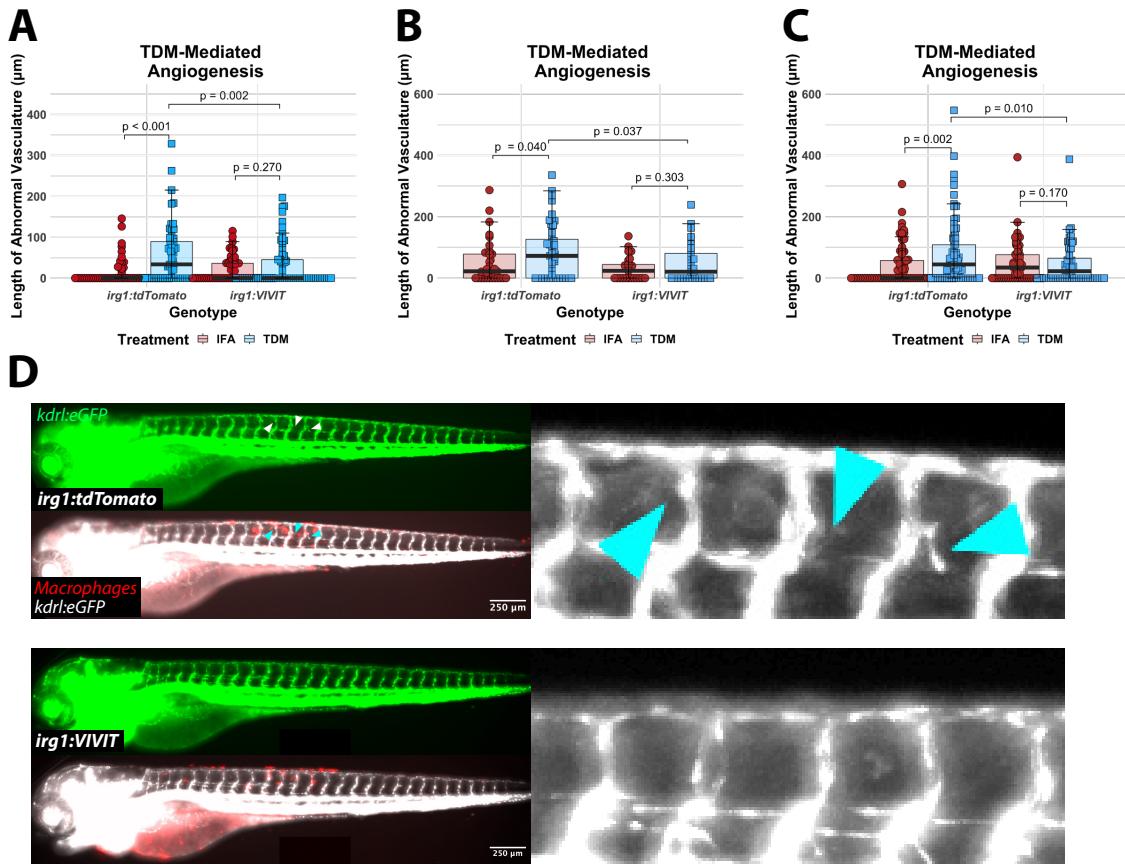


Figure 3.19: Macrophage-specific VIVIT expression reduces TDM-induced angiogenesis in the larval zebrafish. (A), (B), and (C) display three independent replicates of this experiment; (D) demonstrates the effect by representative images showing the reduction seen in the VIVIT-expressing condition.

angiogenesis at 2 days post injection (Walton et al., 2018). TDM was sufficient to induce angiogenesis *in vivo* and this effect was dependent upon functional macrophage NFAT signaling, with the degree of TDM-induced angiogenesis reduced to the level of the vehicle alone in *irg1:VIVIT-tdTomato* animals compared to *irg1:tdTomato* controls (Figure 3.19A-D). This is in contrast to the results I found using a *lyz:VIVIT-tdTomato^{xt39}* transgenic compared to *lyz:dsRed2^{nz50}*, where both conditions induced angiogenesis in response to TDM (Figure 3.17) (Hall et al., 2007).

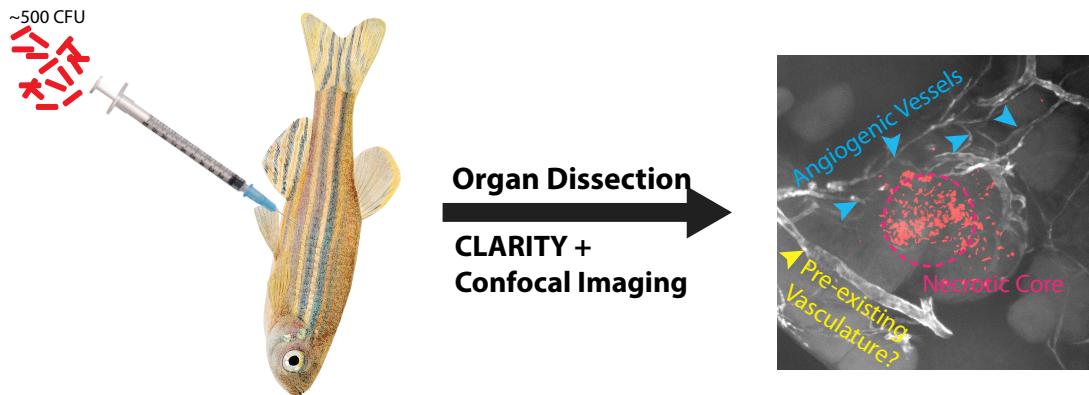


Figure 3.20: Schematic of the infection and CLARITY-clearing approach used for adult infection experiments. Adult zebrafish (>12 weeks post fertilization) are infected interperitoneally with 500 CFU of fluorescent *M. marinum* and then organs are harvested and fixed approximately 14 days later, cleared with detergent, and then imaged.

3.3.6 NFAT Activation is Essential for Angiogenesis in Adult Granulomas

Adult zebrafish are equipped with both innate and adaptive immunity and form mycobacterial granulomas that histologically mirror epithelioid human tuberculosis granulomas (Swaim et al., 2006), including induction of a surrounding vascular network (Cronan et al., 2015). To assess whether our findings in the larvae translated to a longer-term context in the presence of adaptive immunity, I infected adult *flk1:eGFP; nfatc2a^{xt69/xt69}* zebrafish and *flk1:eGFP; nfatc2a^{+/+}* siblings with *Mm*-tdTomato and examined their peritoneal organs at 18 days post infection after CLARITY-based clearing (Chung and Deisseroth, 2013; Cronan et al., 2015). Cleared organs were then imaged by spinning disk confocal microscopy at 100x total magnification with a 10x long working distance lens (Figure 3.20). I measured the total vascular network surrounding the granulomas in a programmatically blinded fashion (Salter (2016) and

Section 4.8) and found that *nfatc2a^{xt69/xt69}* fish had a significant reduction (~50%) in the length of the vascular network compared to wild-type siblings, further validating this gene as important for the angiogenic response *in vivo* (Figure 3.21A-D). These putatively neovascular vessels tend to be highly branched and to be comprised of a limited number of cells with small or non-existent luminal volume, indicating that they are still in the sprouting stage of angiogenesis and suggesting a potential failure to mature over time, perhaps indicative of leakiness or other vascular defects, although further physiological characterization would be required to be certain. I observed robust effects that are likely understated in our quantitation, as I could not make any formal distinction between thicker, existing vasculature present at baseline that happens to fall nearby the granuloma and the characteristic neovascularization more intimately associated with the granuloma and present in wild-type but reduced in *nfatc2a* mutants⁹ (Figure 3.22).

3.3.7 Macrophage-specific NFAT Inhibition in Mature Granulomas Reduces Angiogenesis

I next evaluated whether macrophage-specific NFAT inhibition had similar effects on vascularization in adult zebrafish. I infected adult *flk1:eGFP; irg1:tdTomato* and *flk1:eGFP; irg1:VIVIT-tdTomato* double transgenic zebrafish with *Mm-mCerulean* and examined visceral organs at 14 days post infection. I used confocal imaging to visualize individual CLARITY-cleared organs and measured the total length of granuloma-proximal vasculature under blinding as above (Salter, 2016). I found that the degree of vascularization was significantly reduced around granulomas from

⁹However, such a type of reanalysis is possible, as the images with the corresponding regions of interest are included in the data release, so these differences are possible to be measured by interested parties in the future.

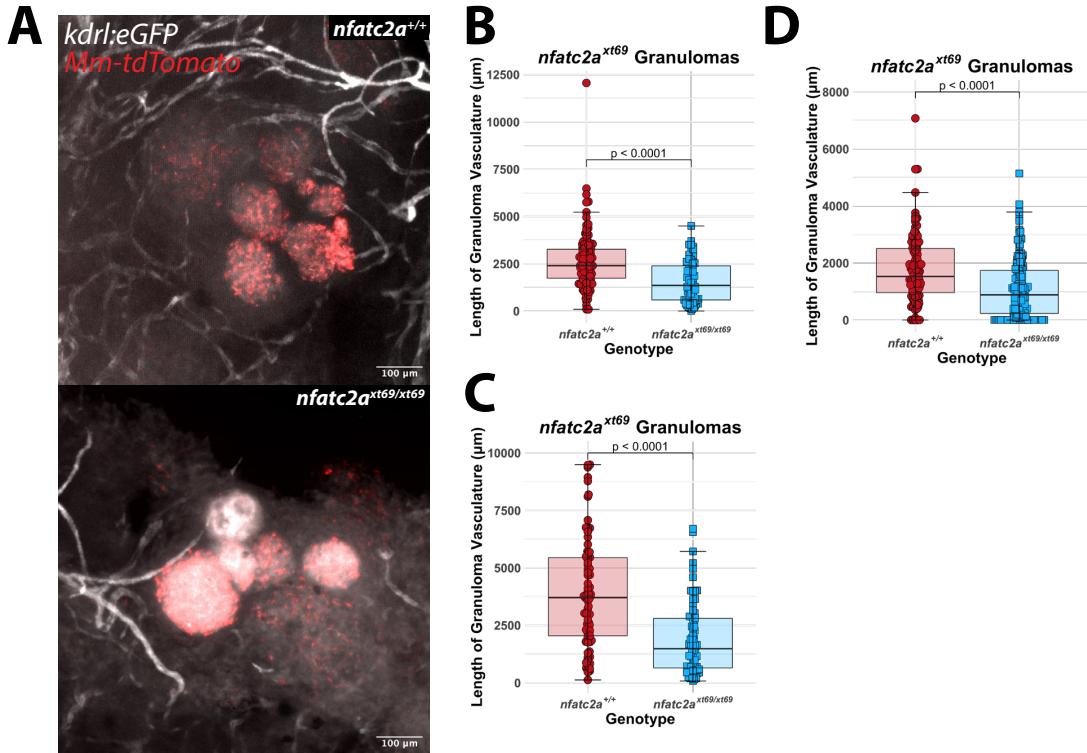


Figure 3.21: *nfatc2a* is required for robust angiogenesis in established granulomas. (A) shows images comparing wild-type and *nfatc2a^{xt69/xt69}* granulomas and a substantial reduction in the overall angiogenic effect can be seen. This is quantitated in (B), (C), and (D), with a 50% reduction in the total vasculature visible in the proximity of each granuloma.

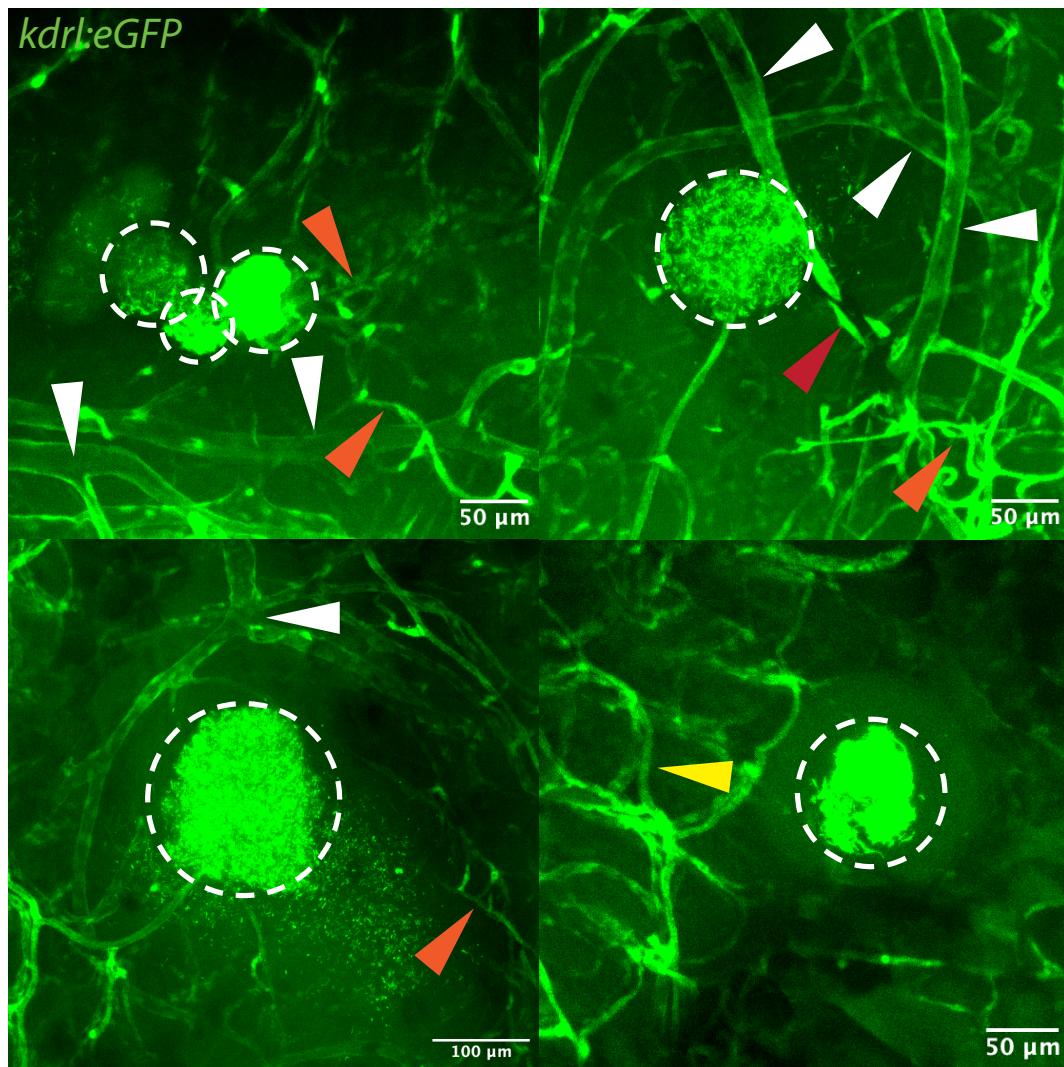


Figure 3.22: Mycobacterial granulomas display significant heterogeneity in vascularization and many are in physical proximity to mature, likely pre-existing vessels. One of the drawbacks of the adult angiogenesis measurements is significant background seen in the proximity of these granulomas, as seen in this figure.

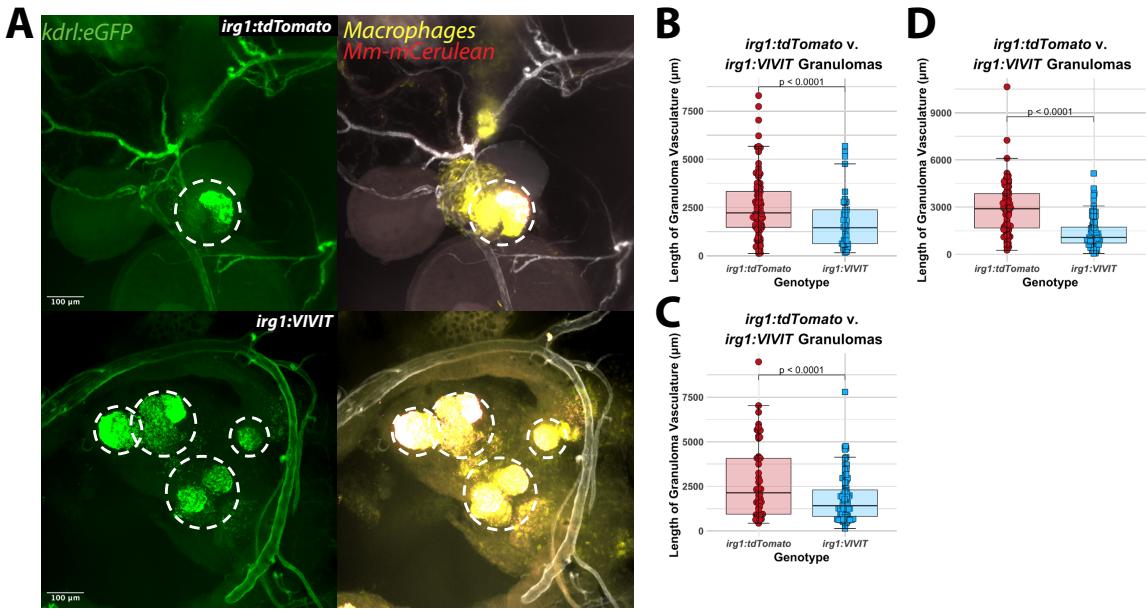


Figure 3.23: Macrophage-VIVIT expression inhibits granuloma angiogenesis in the adult zebrafish infection model. (A) shows images comparing *irg1:tdTomato* and *irg1:VIVIT-tdTomato* granulomas and a substantial reduction in the overall angiogenic effect can be seen. This is quantitated in (B), (C), and (D), with a 50% reduction in the total vasculature visible in the proximity of each granuloma.

irg1:VIVIT-tdTomato fish as compared to *irg1:tdTomato* fish (Figure 3.23A-D). The extent of the vascular network in the *irg1:VIVIT-tdTomato* condition was notably restricted in most cases or solely comprised of more mature, luminal vessels, suggesting a total failure to induce an angiogenic response (Figure 3.23A). These findings, consistent with our previous data from both larval zebrafish infections in the *irg1:VIVIT-tdTomato* background and in the *nfatc2a* mutant adult fish, point to a critical role for macrophage-specific NFAT activation in inducing the angiogenic response at mycobacterial granulomas. Furthermore, this establishes that NFAT function is broadly conserved from early larval infection through to the mature necrotic granulomas that characterize adult infection.

3.3.8 Inhibition of NFAT Signaling Results in Decreased Bacterial Burden

We had previously shown that inhibition of granuloma-associated vascularization is associated with decreased bacterial burden (Oehlers et al., 2015). Mycobacterial mutants unable to induce vascularization ($\Delta pcaA$), and pharmacogenetic inhibition of VEGFA or CXCR4 signaling result in lower bacterial burden, presumably due to functions of the aberrant vasculature promoting bacterial growth and/or inhibiting bacterial killing (Glickman, Cox, and Jacobs, 2000; Rao et al., 2005; Oehlers et al., 2015; Walton et al., 2018) or through more direct pro-bacterial activities of VEGFA on responding macrophages (see also Subsection 1.5.3) (Harding et al., 2019). To examine the effect on burden of inhibition of NFAT signaling, I performed colony forming unit (CFU) assays at timepoints after the induction of angiogenesis and granuloma maturation. I infected *nfatc2a^{+/+}* and *nfatc2a^{xt69/xt69}* adult zebrafish with *Mm-tdTomato* and plated them for CFU at 24 days post infection. I found that knockout of *nfatc2a* resulted in a ~50% decrease in median colony number compared to wild-type after extended infection (Figure 3.24A).

Finally, I evaluated the impact of macrophage-specific NFAT inhibition on whole organism bacterial burden. I infected adult zebrafish possessing either the *irg1:VIVIT-tdTomato* or *irg1:tdTomato* transgenes with *Mm-tdTomato* and then homogenized and plated these fish at 18 days post infection. I found that macrophage expression of the VIVIT peptide resulted in a median reduction of ~60% of the bacterial burden in these fish at this time point relatively soon after the formation of necrotic granulomas and robust induction of angiogenesis (Figure 3.24B).

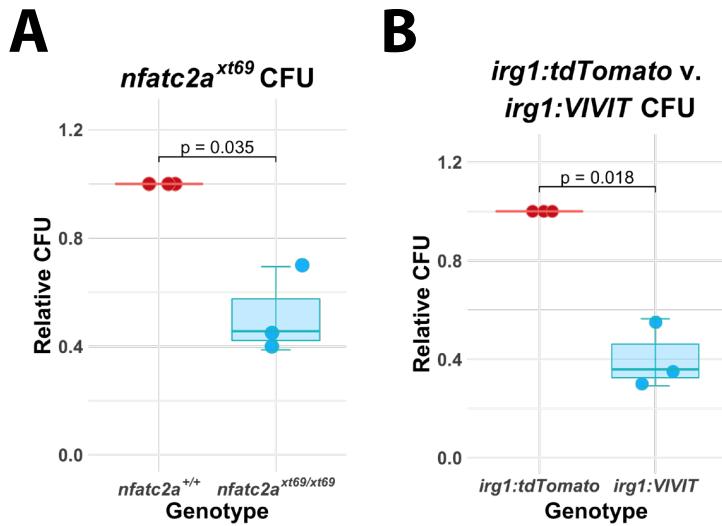


Figure 3.24: Inhibition of NFAT signaling results in a reduction in mycobacterial CFU at time points after granuloma formation. (A) shows a reduction in bacterial burden in *nfatc2a* mutant fish while (B) shows a similar reduction in burden in VIVIT-expressing fish.

3.3.9 Pharmacological Inhibition of NFAT in Human

THP-1 Macrophages Limits VEGFA Induction by *Mycobacterium tuberculosis*

The zebrafish mycobacterial infection model shares important conserved features with *M. tuberculosis* infection of humans, host response and granuloma angiogenesis (Swaim et al., 2006; Datta et al., 2015; Oehlers et al., 2015; Cronan et al., 2016; Cronan et al., 2021). In addition, important aspects of the response to cyclopropanated TDM appears to be largely maintained between zebrafish and humans (Rao et al., 2005; Walton et al., 2018). I next asked whether our findings discovered *in vivo* with the zebrafish-*M. marinum* model were conserved in human cells exposed to *M. tuberculosis*. Ana María Xet-Mull and I developed a cell culture model of macrophage-*M. tuberculosis* interactions using differentiated THP-1 monocytic cells

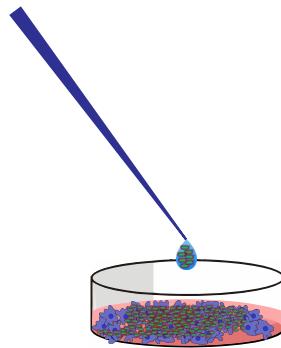


Figure 3.25: In our extracellular exposure model, clumps of gamma-irradiated *M. tuberculosis* is added to monolayers of THP-1 macrophages to stimulate them via primarily extracellular exposure rather than by phagocytosis.

exposed to γ -irradiated *Mycobacterium tuberculosis* H37Rv (γMtb), which produces the full spectrum of TDM species, presented to the cell in their native configuration (as compared to heat-killed *M. tuberculosis*, which disrupts cell envelope structure and organization) (Romero et al., 2014; Secanella-Fandos et al., 2014) (Figure 3.25) (Subsection 1.1.5). I found that exposure of differentiated THP-1 macrophages to γMtb was sufficient to induce VEGFA transcription as well as VEGFA secretion (Figure 3.26A-C, Figure 3.27A-C). To examine whether NFAT signaling is required for production and secretion of VEGFA we treated THP-1 macrophages with the small molecule inhibitor INCA-6, which specifically disrupts the interaction between the NFAT family members and their activating phosphatase, calcineurin by a mechanism similar to that of the VIVIT peptide (Roehrl, Wang, and Wagner, 2004; Roehrl et al., 2004). Strikingly, treatment of THP1 cells with INCA-6 during γMtb exposure significantly inhibited transcriptional induction of *VEGFA* (Figure 3.26A-C), as well as VEGFA secretion (Figure 3.27A-C). Immunofluorescence revealed robust translocation of NFAT (using an NFATC2 antibody) that was broadly correlated to VEGFA signal (Figure 3.28A-D, Figure 3.29A-C). Taken together these experiments suggest that human NFAT signaling is required for VEGFA production in response to *M.*

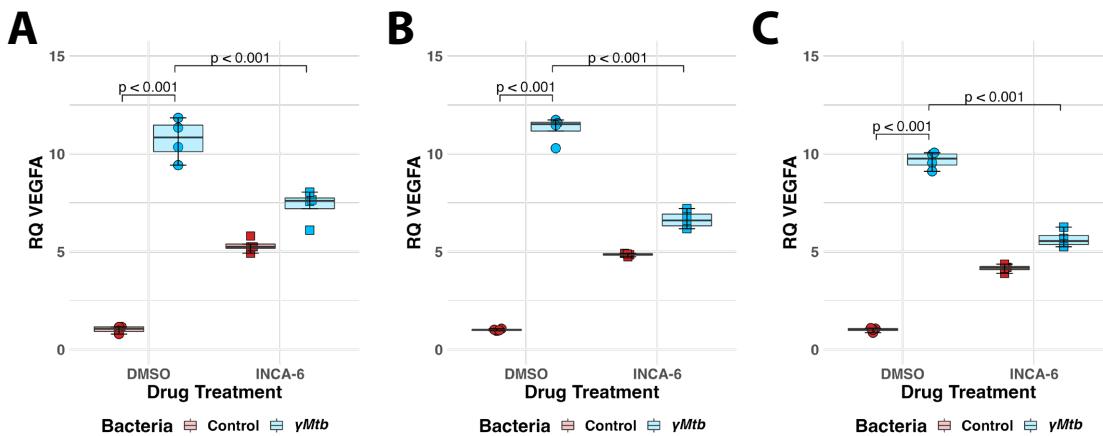


Figure 3.26: Treatment of THP-1 macrophages with gamma-irradiated *M. tuberculosis* results in the induction of *VEGFA* that is sensitive to inhibition by INCA-6, a calcineurin-NFAT inhibitor. Three independent replicates are shown in (A), (B), and (C).

tuberculosis exposure.

3.3.10 Requirement of human NFATC2 for VEGFA induction

To identify functionally important NFAT human isoforms, we exposed THP-1 macrophages to γMtb and subsequently used the secretion inhibitor brefeldin A to lock VEGFA within secreting cells. Simultaneous staining for each of the four human NFATc proteins along with VEGFA allowed us to identify NFAT isoforms that underwent changes in expression and localization and correlate this with VEGFA production (Figure 3.30A). While THP-1 macrophages express all of the isoforms to varying degrees, the most intense co-staining with VEGFA was found with NFATC2 (Figure 3.30B). Additionally, while each of the isoforms displayed structural and localization alterations (albeit not always nuclear) after γMtb exposure¹⁰, only NFATC2

¹⁰These localization patterns were rather remarkable and it is striking that they are not further commented on in the literature, if they are present in other cell types. These bodies of NFAT proteins residing in the cytosol seemed to dynamically reconfigure based on the activation of

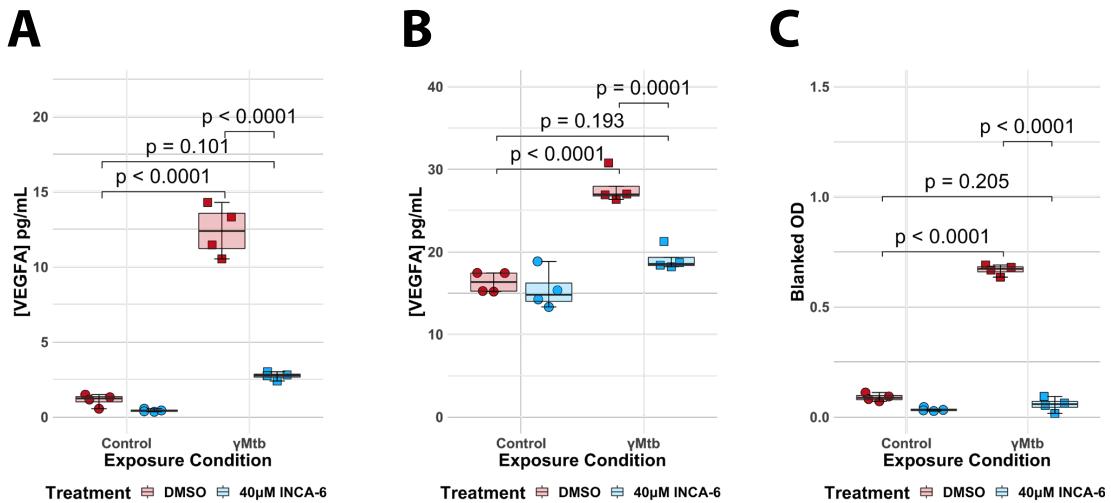


Figure 3.27: Treatment of THP-1 macrophages with gamma-irradiated *M. tuberculosis* results in the secretion of VEGFA and is sensitive to inhibition by INCA-6, a calcineurin-NFAT inhibitor. Three independent replicates are shown in (A), (B), and (C). The standard curve for the replicate in C was generated improperly and was unable to be used, so raw blanked OD values are provided instead.

showed robust nuclear localization that appeared to correspond to VEGFA induction in individual cells (Table 3.1). While some NFAT isoform translocation was observable with at least NFATC1 and, minorly, NFATC3, this generally had, at best, modest correspondence to the degree or presence of VEGFA production (Figure 3.31). While NFATC1 was correlated with VEGFA induction, the effect was much weaker than that seen with NFATC2. I quantified this effect by counting the number of VEGFA-expressing, NFAT nuclear localized cells and normalized to the number of VEGFA-expressing cells in total and found that NFATC2 most tightly corresponded to the induction of VEGFA in γMtb -exposed cells Figure 3.31. Given the strong correlation for NFATC2 with nuclear localization and VEGFA production after γMtb exposure, expression data from zebrafish and non-human primate granulomas, as well

the macrophage and to increase in size and quantity after stimulation. While there was plenty of nuclear translocation obvious, the cytosolic reconfigurations are also interesting and indicate these some of the isoforms might be playing cytosol-specific roles during immune responses and this dissection would be a fascinating addition to the body of NFAT literature.

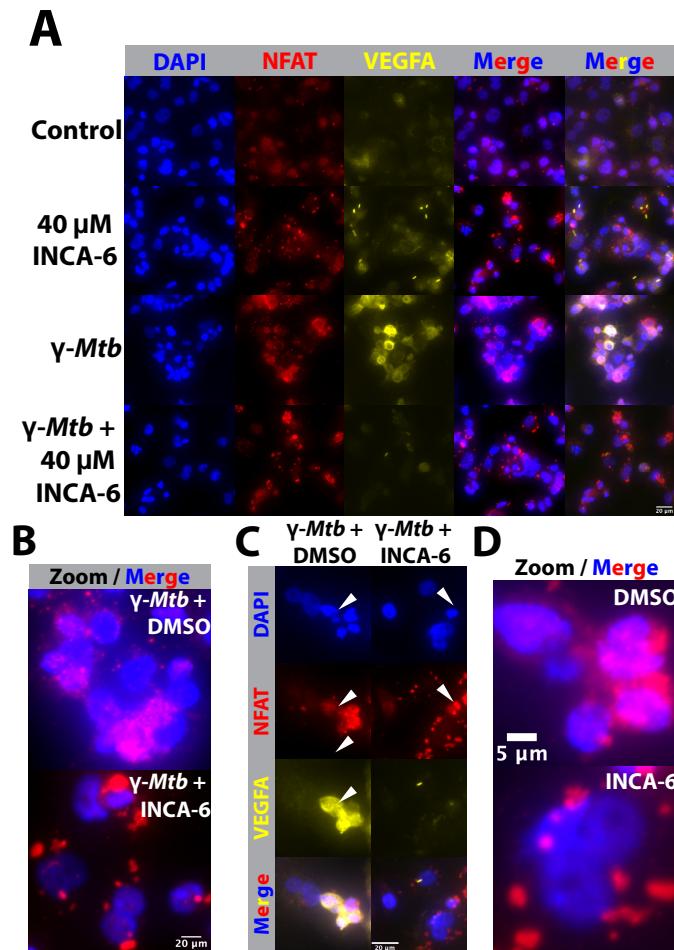


Figure 3.28: Immunofluorescence imaging of gamma-irradiated *M. tuberculosis* exposed THP-1 macrophages reveals robust NFAT protein translocation into the nucleus after exposure that corresponds to the induction of VEGFA. (A) shows robust VEGFA induction that can be inhibited by addition of INCA-6. (B) shows a magnified view of the NFAT nuclear translocation seen in (A). (C) shows further images and the correspondence between NFAT activation and VEGFA production. (D) shows the reduction in nuclear localization from the images seen in (C).

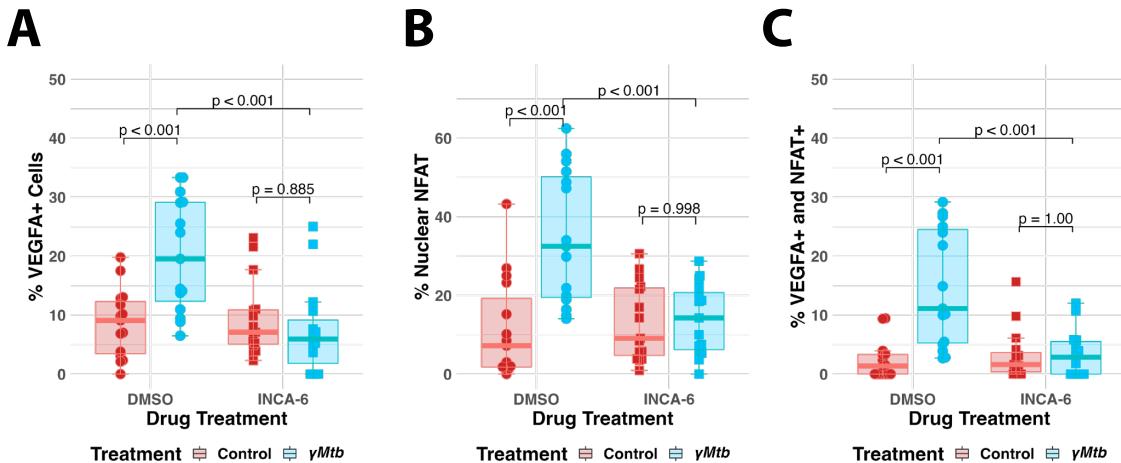


Figure 3.29: Quantitation of various aspects of the response seen by immunofluorescence after gamma-irradiated *M. tuberculosis*-exposure and INCA-6 treatment. (A) shows the overall percentage of VEGFA+ cells seen in each condition, with a significant increase with γMtb exposure that is inhibited by INCA-6. (B) shows a parallel phenotype, where γMtb induces NFAT nuclear localization that is sensitive to INCA-6 addition. (C) shows the percentage of the total cells in the field of view that demonstrate both NFAT nuclear localization and VEGFA expression.

as the *in vivo* zebrafish results implicating macrophage *nfatc2a* in *vegfaa* production and angiogenesis, I focused on human NFATC2 as the key isoform.

To test a functional role for human NFATC2 in macrophage induction of VEGFA during γMtb exposure, we used a lentivirus-mediated CRISPR/Cas9 approach to introduce high-efficiency disruption of NFATC2. Using techniques inspired by the zebrafish and backported to cell culture, we simultaneously expressed four distinct guide RNAs targeting NFATC2 or safe-targeting controls, to maximize the percentage of puromycin-resistant cells possessing complete null mutations (Wu et al., 2018). I compared these cells to those transduced with lentiviruses expressing safe-targeting control sgRNAs. (Figure 3.32, Table 3.2, Subsection 2.8.5) (Kabadi et al., 2014;

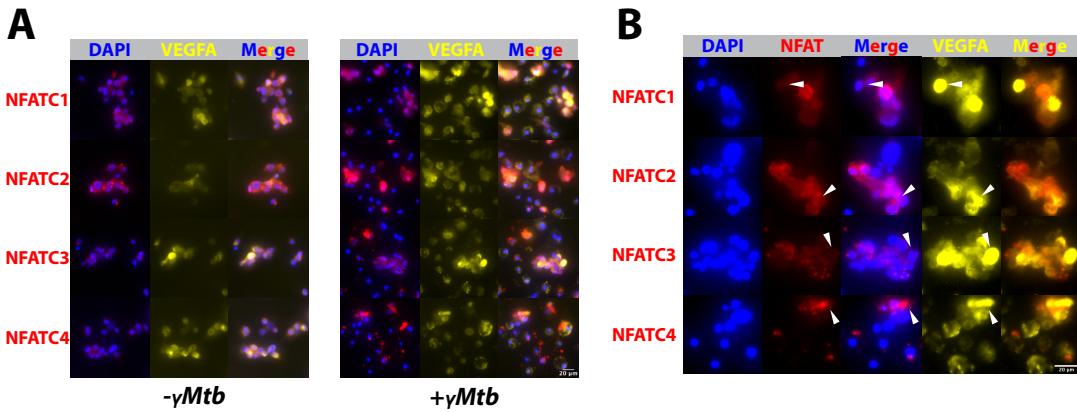


Figure 3.30: To identify particular NFAT isoforms that may be important for VEGFA production downstream of *M. tuberculosis* detection, immunofluorescence was performed against each of the isoforms. NFATC2 showed the most consistent induction of all of the isoforms. (A) shows alterations in the protein production and localization with and without *M. tuberculosis*. (B) displays a representative field where multiple VEGFA-expressing cells can be seen, but only nuclear localization of NFATC2 seems to correspond with this expression.

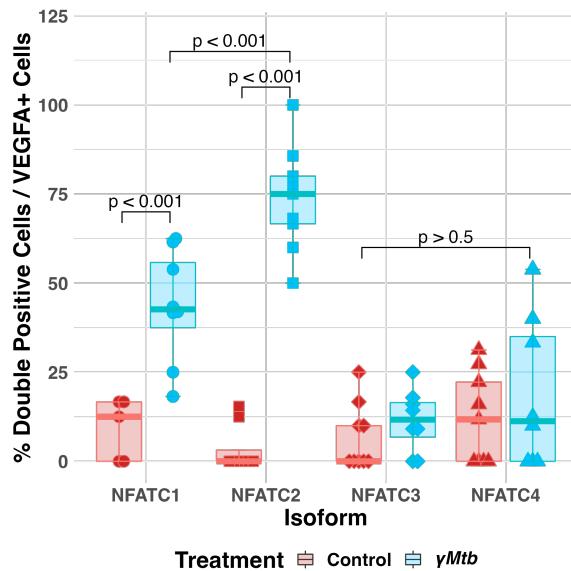


Figure 3.31: I quantitated the immunofluorescence images from THP-1 macrophages stained with antibodies against the four NFAT isoforms to assess the contribution of each protein to the VEGFA expression phenotype. Indeed, I found that NFATC2 most tightly corresponded to VEGFA induction by measuring double positive cells (those with both VEGFA expression and NFAT nuclear localization) and normalized to the total number of VEGFA positive cells.

Table 3.1: Relative correspondence between the nuclear localization and VEGFA induction across all four NFAT isoforms. I observed robust correspondence between NFATC2 and VEGFA and a lesser association between NFATC1 and VEGFA while the other two isoforms displayed no notable relationship between their nuclear localization and VEGFA induction.

	$-\gamma Mtb$	$+\gamma Mtb$	Relationship to VEGFA?
NFATC1	+	++	Modest relationship between nuclear localization and VEGFA expression.
NFATC2	++	+++	Nuclear localization generally corresponds to VEGFA expression.
NFATC3	+	++	Weak relationship between nuclear localization and VEGFA expression.
NFATC4	+/-	+	Expression increased but not obviously nuclear in most cells.

Table 3.2: Nearest gene neighbors for the ST sgRNAs (all are intergenic).

sgRNA	Nearest Gene Neighbor (Chromosome)
hU6 ST	CNTN1 (12)
mU6 ST	MTDH (8)
7SK ST	OPN5 (6)
hH1 ST	ENSG00000249941 (5)

Sanjana, Shalem, and Zhang, 2014; Morgens et al., 2017; Kitamura and Kaminuma, 2021). Due to technical challenges associated with long-term culture of THP-1 cells and to address heterogeneity among cellular responses, I focused these assays on VEGFA induction in these cells by immunofluorescence after γMtb exposure, as the effects by qRT-PCR and ELISA were difficult to interpret due to this underlying heterogeneity and the extended culture required for selection. Because the N-terminal epitope recognized by our NFATC2 antibody was upstream of the targeted sites, I was unable to examine functional protein levels directly and simultaneously in the immunofluorescence images (Figure 3.34A) and this resulted in a high frequency of protein mislocalization to the nucleus despite anticipated loss-of-function. However, I

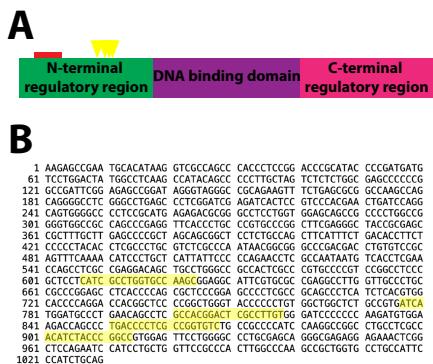


Figure 3.32: Based on our findings in THP-1 macrophages and the larval zebrafish, NFATC2 was selected for genetic targeting in the THP-1 macrophages. (A) displays the location of the four sgRNAs selected (in yellow arrows) and the location of the antibody epitope (red bar). (B) shows, on the genomic sequence itself, the location of the four sgRNAs.

found that transduced cells targeted by NFATC2 lentivirus generally failed to induce VEGFA while safe-targeting control lentivirus-transduced cells responded normally (Figure 3.33A-B), an effect that can be quantitated by percentage of VEGFA+ cells by multiple metrics (Figure 3.35A-C). These cells also demonstrated evidence of non-sense-mediated decay by NFATC2 transcript levels in suspension THP-1 monocytes (Figure 3.34B). Thus, macrophage NFATC2-mediated induction of VEGFA downstream of mycobacterial TDM exposure is conserved from zebrafish to human cells exposed to *M. tuberculosis*.

3.4 Discussion

This work uncovers an unexpected and novel role for macrophage NFAT activation in immune responses to pathogenic mycobacteria and the maladaptive angiogenic responses that occur during infection. This activation of NFAT is driven through recognition of bacterial cis-cyclopropanated trehalose 6-6'-dimycolate, a major constituent of the cell envelope in pathogenic mycobacteria, that we have previously

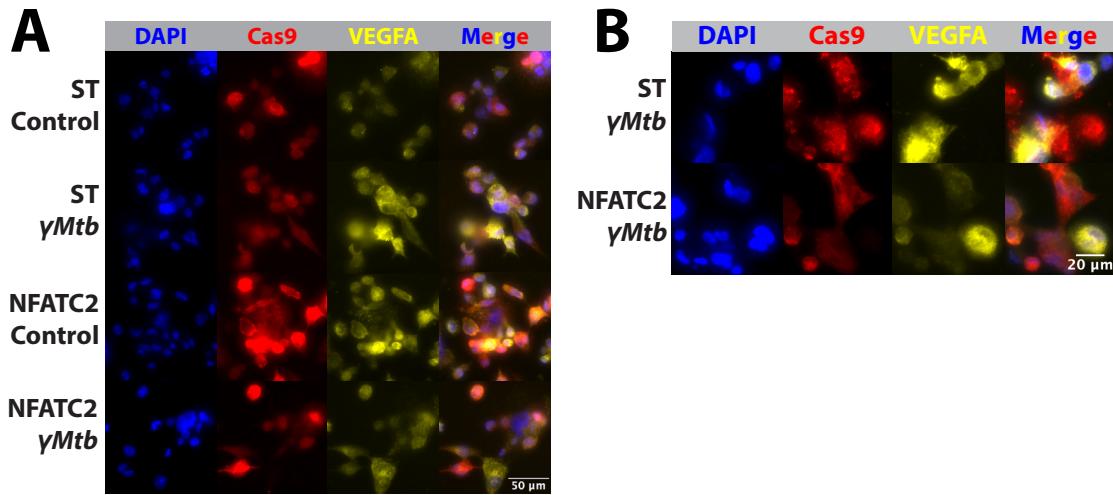


Figure 3.33: Lentivirus-mediated CRISPR/Cas9 targeting of NFATC2 results in reduced VEGFA expression after gamma-irradiated *M. tuberculosis* exposure. (A) shows that Cas9-expressing NFATC2-targeted cells have a reduction in VEGFA production while similar Cas9-expressing cells with safe targeting control sgRNAs robustly induce VEGFA. (B) shows this magnified. Note the higher-expressing NFATC2-targeted cell has minimal Cas9 expression, suggesting that these effects may depend on the efficiency of targeting and other factors. (C) shows quantitation of the percentage of VEGFA+ cells in a given field of view, with a statistically significant reduction in the NFATC2-targeted condition.

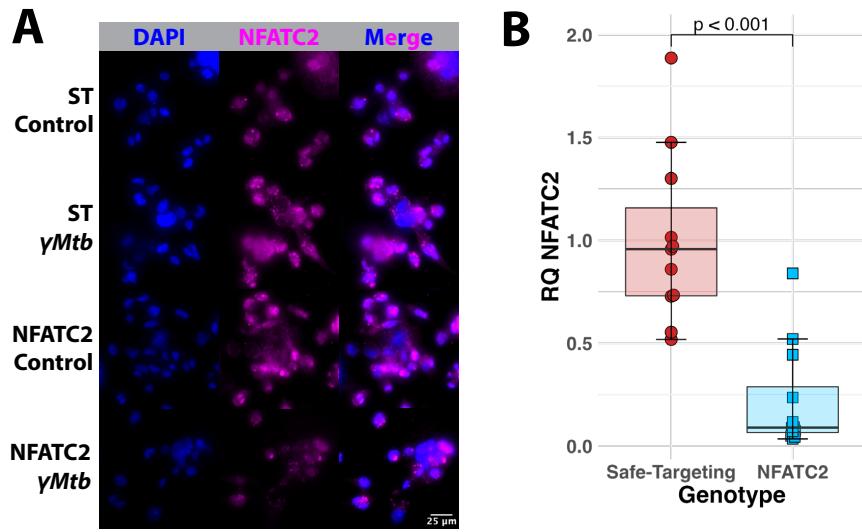


Figure 3.34: Validation of the lentivirus-mediated knockout approach. (A) shows aberrant localization of NFATC2, suggesting that the remaining protein has been functionally disrupted. Note that the antibody binding site is N-terminal to the sgRNA sites, a technical oversight in this design that made it difficult to quantify reductions in NFATC2 protein levels after targeting. (B) Nonsense-mediated mRNA decay in suspension THP-1 monocytes is reduced by NFATC2 targeting relative to safe targeting controls.

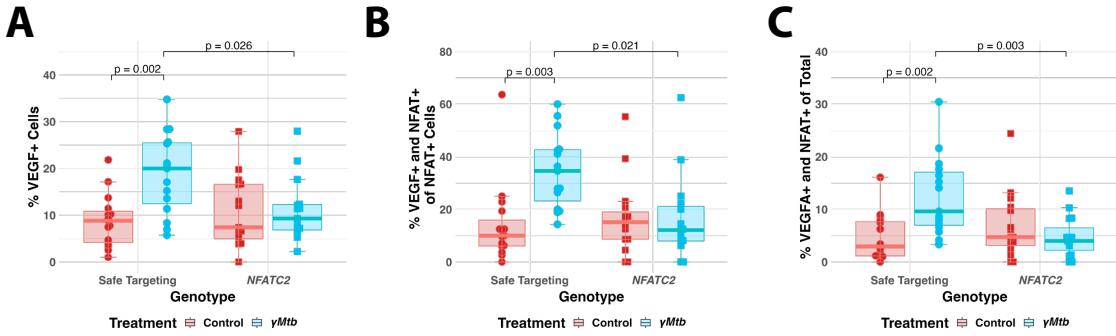


Figure 3.35: Blinded quantification of our lentivirus-transduced CRISPR/Cas9 THP-1 macrophages demonstrating a reduction in the degree of VEGFA expression that corresponds to reduced NFATC2 nuclear responsiveness to γMtb . (A) shows the total percentage of VEGFA+ cells out of the total cells present. (B) is the percentage of cells with both VEGFA expression and NFATC2 nuclear translocation out of the total number of cells with NFATC2 nuclear translocation. (C) shows the percentage of the total number of cells that have both VEGFA expression and NFATC2 nuclear translocation. Together these data implicate NFATC2 in mediating the VEGFA response to γMtb exposure.

found is necessary and sufficient to drive pathological angiogenesis (Walton et al., 2018). Identifying this unexpected role for NFAT in angiogenesis expands our understanding of the mechanisms governing mycobacterial pathogenesis and offers targets for potential host directed therapeutics. Traditionally, work on TDM-mediated C-type lectin activation has focused on CARD9 and NF- κ B signaling. Here, by contrast, I describe a specific role for alternative C-type lectin signaling responses through the NFAT pathway to drive VEGFA production and granuloma-associated angiogenesis.

VEGFA induction is a prominent feature of tuberculosis in human disease as well as in a number of animal models, including non-human primates, rabbits, mice, and zebrafish (Datta et al., 2015; Oehlers et al., 2015; Polena et al., 2016; Harding et al., 2019; Cronan et al., 2021; Gideon et al., 2022). I found that VEGFA was produced specifically within newly arrived macrophages at nascent granulomas. Macrophage populations are critical to VEGFA induction, as macrophage-specific inhibition of

NFAT signaling and depletion of macrophages generally result in reductions in granuloma-associated angiogenesis (Oehlers et al., 2015). We find that this extends to the macrophage-enriched NFAT isoform, *nfatc2a*, which is also required for angiogenesis. Using a human cell culture model, I found that NFATC2 was similarly engaged in human cells as amongst all NFAT isoforms, only NFATC2 underwent robust nuclear translocation in response to *M. tuberculosis* stimulation. Correspondingly, pharmacological inhibition of NFAT signaling in human cell culture as well as genetic inhibition of *NFATC2* resulted in reduced VEGFA production.

Although animal models of tuberculosis generally report high VEGFA expression in the granuloma, there are few studies that center on VEGFA induction in cell culture infection models (Polena et al., 2016). Through high-resolution time-lapses and reporter lines, I found that *vegfaa* induction generally does not occur until the formation of initial granulomas and is generally correlated with the appearance of extracellular bacteria that could be recognized by incoming, likely uninfected macrophages (Figure 3.4 and Figure 3.5). This concentration-dependent effect on signaling may reflect key aspects of the disease itself, wherein large masses of extracellular bacteria accumulate in the necrotic core of the granuloma, potentially triggering relatively insensitive and/or chronic C-type lectin signaling in this context. Given what is already known about the need for chronic stimulus to produce effective NFATC2 activation, this would coherently fit with existing models and would offer a mechanism wherein other NFAT isoforms could play other, potentially protective roles in early infection responses but NFATC2 is specific to later stages at high intensity and duration of activation (Yissachar et al., 2013; Kar and Parekh, 2015).

Consistent with the recognition of extracellular bacteria, exposure of human macrophage-like cells to γ -irradiated *M. tuberculosis* rapidly induced NFATC2-dependent

VEGFA signaling in a dose dependent manner. Standard cell culture infection models generally eliminate extracellular bacteria using gentamicin treatment and media changes, and so it is possible that engagement of this pathway by extracellular bacteria or TDM stimulation is a key component of this response. A survey of the literature and a variety (Lee et al., 2019; Pisu et al., 2020; Hall et al., 2021; Looney et al., 2021; Pu et al., 2021) of RNA-seq datasets from macrophage-*M. tuberculosis* infection experiments reveal modest or nonexistent induction of VEGFA, further supporting the notion that extracellular exposure to *M. tuberculosis* may be an important element of the angiogenic response and may reflect some aspects of the macrophage-*M. tuberculosis* interface within granulomas (Orme, 2014).

As its name suggests, the NFAT pathway plays an indispensable role in normal T cell biology. Accordingly, whole animal knockouts of NFAT in standard mouse models of *M. tuberculosis* infection – where granuloma formation itself may be limited – may have obscured a role for myeloid-specific effects of NFAT signaling (Via et al., 2012). This study investigated a role for NFATC2 in control of *M. tuberculosis* infection and found that NFATC2 was required for effective bacterial control and production of TNF- α and IFN- γ in CD4 $^{+}$ T cells despite no perturbation of the expression of these genes in dendritic cells, suggesting an adaptive immune response-specific defect in bacterial control. This may be of little surprise, given the extensive contacts that T cells have with mycobacteria within the lesions that develop in standard mouse models. Such a defect in T cell responses may be important for bacterial control when T cells are able to access the bacteria, but this condition is not reflective of the biology of the granuloma in other models or humans. It would be interesting to explore the phenotype of a *LysM-Cre; Nfatc2^{f/f}* mouse infected with tuberculosis to isolate the macrophage-dependent phenotypes. However, given our findings on macrophage NFAT contributions to angiogenesis, which is poorly modeled in the

mouse, either entirely new roles for NFAT may be uncovered or it may have little or no effect on bacterial growth and disease. In any case, the present results offer some tension with these historical findings and it may take additional work to definitively identify any role for NFATC2 in the overall pathogenesis of tuberculosis infection.

The zebrafish model, by looking at early timepoints, uncovered a role both in angiogenesis and, presumably as a consequence, bacterial control. Wholesale, longer-term inactivation of NFAT, which also plays important roles in T cells, would potentially compromise important aspects of a productive adaptive immune response during mycobacterial infection. While genetically manipulable animal models allow for cell-specific separation, any host-based therapeutic approaches might require cell-specific macrophage delivery methods (Hu et al., 2019; Mukhtar et al., 2020; Colombo et al., 2022), NFATC2-specific targeting (Kitamura and Kaminuma, 2021), and/or contend with the adaptive immune response¹¹, an important aspect of host resistance during mycobacterial infection.

While little else has been studied in the context of NFAT-mycobacterial interactions, a previous publication assessed the contributions calcineurin activation on the mycobactericidal activity of macrophages and found that mycobacteria manipulate an actin-binding protein called CORO1A (coronin-1A or TACO) to block phagosomal-lysosomal fusion and do so by activating calcineurin (Jayachandran et al., 2007). In the absence of CORO1A or after treatment with calcineurin inhibitors, lysosomes efficiently fused with the mycobacteria-containing phagosomes, mediating bacterial killing. This effect was found to be independent of transcription, implicating calcineurin in somehow modulating the process of phagosomal-lysosomal fusion,

¹¹Although NFATC2 appears to be largely redundant with both NFATC1 and NFATC2 for many, but not all adaptive responses, it is still important to, as specifically as possible, target the desired pathway for minimal off-target side effects.

potentially through some interaction with CORO1A and actin. However, a subsequent publication found that this process was independent of actin (Jayachandran et al., 2008), leaving it a yet-to-be resolved mystery how directly calcineurin-dependent activity blocks phagosomal-lysosomal fusion to facilitate mycobacterial survival and replication (Zimmerli et al., 1996). This is also intriguing in the context of the known activation of NFAT downstream of phagocytosis, which would be expected to be active in this context unless specifically blocked by the mycobacteria (Fric, Zelante, and Ricciardi-Castagnoli, 2014). If nothing else, this adds to the body of evidence suggesting that calcineurin/NFAT-based host directed therapies may have multiple modes of host-beneficial activity. Differentiating these diverse roles for calcineurin in host immunity will expand our understanding of the multitude of activities of the pathway and allow finer pharmacological approaches to inhibiting calcineurin-dependent processes.

It remains unclear why NFATC2, but not any of the other isoforms, is specifically required in macrophages for the induction of VEGFA, given evidence that the others are present in resting macrophages (Figure 3.30). The functional distinctions between the isoforms have long been of basic interest, but relatively few specific differences between them have been identified beyond basal regulation to provide tissue-specificity and more recent findings describing layers of kinetic regulation with isoform-specific stimulation thresholds, nuclear retention, and more (Lyakh, Ghosh, and Rice, 1997; Rao, Luo, and Hogan, 1997; Yissachar et al., 2013; Kar et al., 2014; Kar and Parekh, 2015; Kar et al., 2016). Anecdotally, I observed substantially higher expression of NFATC2 by immunofluorescence after γ Mtb exposure even with INCA-6 treatment, suggesting that there may be interesting upstream regulatory mechanisms, potentially dependent on HIF-1 α or NF- κ B that prime further NFAT-dependent responses. These novel levels of regulation offer opportunities for uncovering new features of the

cell biology of NFAT.

NFAT is also likely to be involved in modulating the expression of other important pro-angiogenic chemokines, including CXCL12, the primary ligand for CXCR4. CXCR4 has been previously demonstrated to be critical for granuloma formation and angiogenesis (Torraca et al., 2017) and NFAT has been shown to be able to regulate CXCL12 expression, at least in osteoblasts (Sesler and Zayzafoon, 2013). It may be reasonable to think that NFATC2 is also mediating CXCL12 expression during infection and is acting as a more central regulator of angiogenic responses. This hypothesis warrants further study; in any case, the stimulus for and transcriptional mediators of the observed CXCR4-dependent granuloma angiogenesis should be identified to add to our understanding of how these processes occur.

Here, I have identified the unique requirement for this single isoform in macrophages to induce angiogenesis in response to mycobacterial infection (Figure 3.36). One hypothesis is that NFATC2 has binding partner(s) unique among NFAT isoforms required for its effect on the VEGFA promoter. Whether this is HIF-1 α (the canonical regulator of VEGFA) or one of the many previously described interacting partners is, as yet, unknown, but could be tested either *in vitro* or *in vivo* with genetic or chemical approaches (Subsection 5.2.3). However, higher order regulatory mechanisms that result in the production of VEGFA in the absence of overt hypoxia have been understudied and this work proposes at least one potentially generalizable mechanism whereby NFATC2 activation results in VEGFA transcriptional upregulation, a process that can be inhibited with chemical and genetic intervention. Despite the widespread presence of the putative NFAT binding motif 5' -GGAAA-3' (Figure 3.37) in the proximal VEGFA promoter (Gearing et al., 2019), their influence on VEGFA transcription has been relatively unexplored as this specific effect is generally

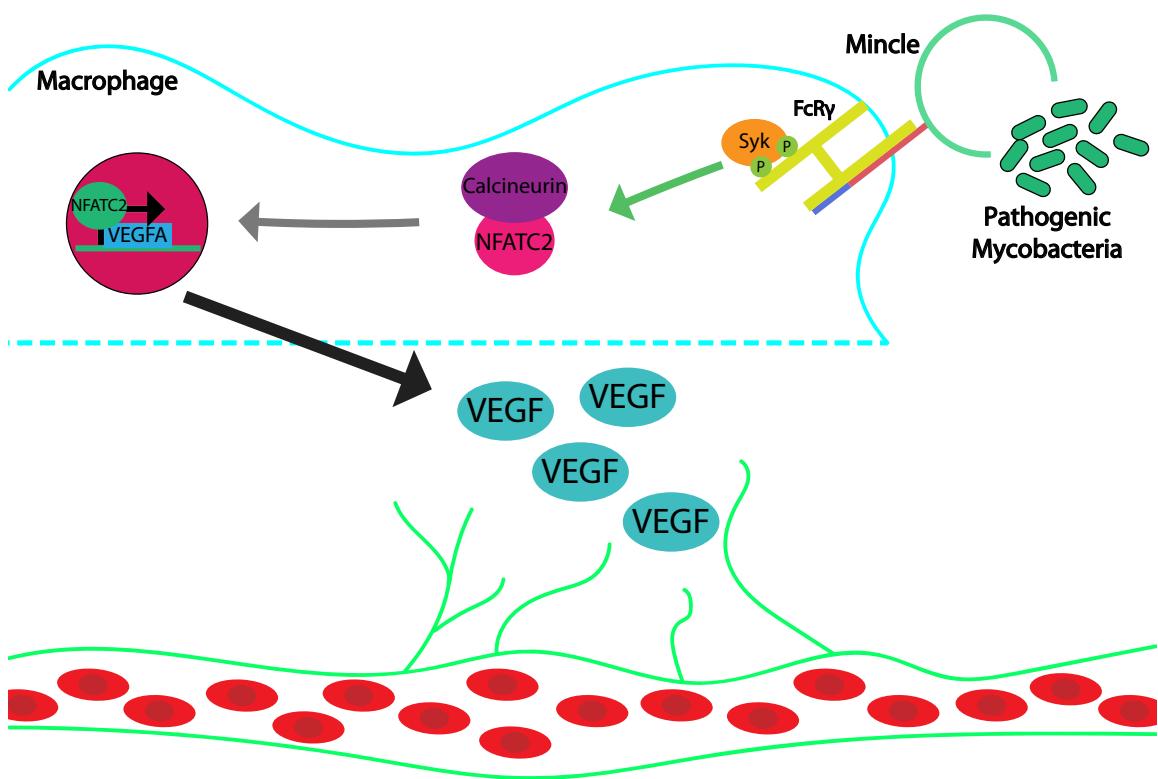


Figure 3.36: Upon completion of this work, I have identified a critical role for NFATC2 activation downstream of TDM detection in mediating the production of VEGFA by macrophages, which induces angiogenesis. This clarifies a major question in the field as to the underlying mechanisms of angiogenesis and opens new paths for investigation into the contributions of the NFAT signaling pathway to mycobacterial pathogenesis.

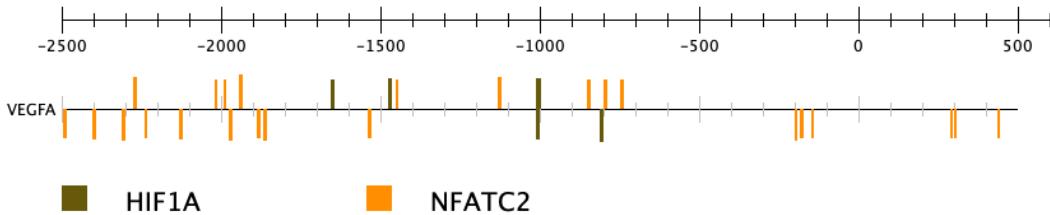


Figure 3.37: NFATC2 and HIF-1 α putative binding sites in the VEGFA promoter as determined by Ciiider. There are several NFAT binding sites in the promoter, the function of which has largely yet to be determined.

not seen in T cells or other cell types (Chang et al., 2004). This single publication from Chang et al. (2004) found that the -2400 site in the *VEGFA* promoter was specifically bound by NFATC proteins and acted to *repress* transcription. This is an intriguing hypothesis that might suggest that NFAT is able to repress *VEGFA* transcription at baseline but may act to induce it after stimulation via distinct binding sites or in concert with other transcription factors. Such results would be consistent with the mild but reproducible increase in VEGFA I observe with INCA-6 treatment in our experiments in THP-1 cells (Figure 3.26).

As described in Section 1.4, NFAT is involved in the induction of a variety of cytokines including IFN- γ and TNF- α , but whether NFAT-dependent transcriptional induction of these within macrophages has a role in the overall course of disease is unknown. This surely offers promise for the future as an expansion of the present work to incorporate a more complete view of the NFAT-dependent signals that influence mycobacterial disease progression.

A more comprehensive characterization of NFAT-dependent innate immune responses has begun in recent years (Deerhake et al., 2021; Peuker et al., 2022; Poli et al., 2022) (for a more detailed description, see Subsection 1.4.4), but this pathway

has remained unstudied in the context of macrophage signaling during mycobacterial infection. Furthermore, this work draws a connection between the induction of calcium fluctuations – which can occur in response to many different developmental, homeostatic, and pathological stimuli, including to mycobacterial infection (Malik, Denning, and Kusner, 2000; Kusner and Barton, 2001; Jayachandran et al., 2007; Jayachandran et al., 2008; Matty et al., 2019) – to the angiogenic response to that stimulation. Our identification of NFAT regulation of VEGFA offers a novel approach to both pro- and anti-angiogenic intervention in various pathological contexts.

3.4.1 Limitations of the Study

The limitations of this study, like all studies, are innumerable. While I am proud of what I have accomplished in pursuit of this work, there are many opportunities for the future as well as further work to clarify the results I have found here. I have identified interesting macrophage biology mediating an important disease-relevant phenotype during mycobacterial infection, but I do not see the same magnitude of bacterial burden change as with VEGFA-inhibition alone (Oehlers et al., 2015), suggesting pleiotropy in the NFAT signaling pathway with inhibition resulting in both pro- and anti-bacterial effects. What these antimycobacterial aspects of NFAT signaling are is worth investigation to better understand the full corpus of possible consequences to NFAT inhibition strategies for the treatment of infectious disease and the potential risks for patients already taking calcineurin inhibitors. To do so, RNA-seq or similar should be employed to better delineate these alterations in gene expression profile. It would also be beneficial to work toward disentangling some of the other effects of NFAT inhibition by simultaneous treatment of the zebrafish with VEGFR2 inhibitors in the *irg1:VIVIT* background to directly test non-angiogenic features of NFAT in mycobacterial infection.

I have thus far been unable to identify the particular macrophage receptor responsible for detecting TDM in the zebrafish. While work in THP-1 macrophages may be able to more affirmatively link MINCLE or MCL to the activation of NFAT, identification of the zebrafish receptor would make a substantive contribution to model development as well as the evolution of C-type lectin receptors more broadly. Cell-type specific knockout of *nfatc2a* would also aid greatly in definitively linking this gene to VEGFA induction and angiogenesis, but the tools to do so are not yet operational; perhaps future studies can begin with such intent and focus more stringently on the role of this isoform strictly in macrophages.

Some technical limitations are also interfering with interpretation of some results. Generation of stable knockouts of NFATC2 in THP-1 macrophages would make for more robust and cleaner assays, although it has the limitation of odd clonal behavior and loses the heterogeneity that can be advantageous in dissecting some phenotypes. The variable ploidy of these cells is also a challenge, so an alternative macrophage model or primary cells may be required to more effectively model some of these responses.

This macrophage-NFAT signaling pathway likely has important roles in other disease contexts that are not addressed here. Tuberculosis is the most common cause of granuloma formation, but it is far from the only cause, and any contributions of this pathway to granuloma biology in *Schistosoma*, *Cryptococcus*, or *Histoplasma* infections is not yet known. However, with the tools now available, these roles are likely to be easier to study in the future and new insights may be gleaned from such investigation. Angiogenesis is a common pathology in some autoimmune diseases, such as arthritis, and is prevalent among cancers; whether macrophage NFAT signaling might be mediating angiogenesis in these contexts is surely of future interest and

may reveal novel aspects of the host-tumor interface.

I have validated important aspects of our observations in the zebrafish with a mammalian cell culture model, but subsequent studies may warrant further integration of mammalian models of tuberculosis infection where angiogenesis is present (rabbits, guinea pigs, macaques) or human patient samples to better understand certain aspects of the underlying biology and validate NFAT induction during tuberculosis infection and potential correspondence to VEGFA production.

3.5 Acknowledgements

I would like to thank all the members of the Tobin and Smith labs for thoughtful contributions throughout the development of this work, Mari Shinohara and Elizabeth Deerhake for help and advice with the cell culture assays, Aviva Symes for sharing long-stored antibodies that would have otherwise been lost to science, Mark Cronan for comments on the manuscript, Emily Hunt and Iswarya Padmanaban for fish care and husbandry, Mirko Rosenau for adult zebrafish stock image, and BEI Resources (NIAID, NIH) for provision of the γ -irradiated *Mycobacterium tuberculosis* H37Rv. This work was funded by NIH grants AI130236, AI125517, and AI166304 (David M. Tobin) and HL160239 (W. Jared Brewer).

3.6 Author Contributions¹²

W.J.B. and D.M.T. designed the project and conceived experiments; W.J.B. performed all zebrafish experiments, A.Y. assisted with some aspects of zebrafish exper-

¹²This list of contributions is adapted from Brewer et al. (2022b), but the text of this chapter has been extensively modified from the source, making precise attribution of the writing somewhat difficult.

iments (especially microinjection of constructs to develop ultimately failed NFAT reporters) and provided assistance with zebrafish husbandry, M.I.S. assisted with CFU assays, and E.M.W. developed the *card9^{xt31}* knockout line and the *irg1:tdTomato^{xt40}* transgenic line. W.J.B. and A.M.X.-M. performed all cell culture experiments; W.J.B. analyzed all experimental data; and W.J.B. wrote the original published manuscript with substantial edits from D.M.T. and additional contributions to the methods sections by A.M.X.-M. and input from all authors.

Chapter 4

Novel Computational Approaches to Image Processing and Quantitation

High-resolution, high-throughput microscopy has opened possibilities for biological analysis that were inconceivable only a few short years ago, but the methods by which to analyze these data remain largely lacking. While heroic efforts have been made to use both standard thresholding methods as well as newer machine learning-powered methods to simplify and automate image processing, these approaches are often somewhat limited by the range of purposes for which they were designed and tested. While the open source nature of many of these approaches facilitates adaptation, this is both technically beyond the skillset of many scientists and impeded by, at best, variable quality documentation. Additionally, many of the tasks that experimenters seek to do are both relatively simple and highly repetitive but they are often unaware of the means by which to cut down on manual processing time in order to economize their time and energies. The scripts presented in this chapter are a combination of automation scripts that will process particular image types into smaller and more informative images (compression of Z-stacks, time-series, etc.) and

those meant for analysis of image data (pixel intensity distribution across an image, bacterial burden within larval zebrafish). While none of these are of the caliber to open entirely new methods of analysis, I hope they are a catalyst for others in the zebrafish community at large to explore the potential for computational automation to save time and frustration in the process of analyzing often thousands of very large, data-rich images.

All of the scripts in their latest versions will be able to be found in perpetuity at <http://github.com/jaredbrewer/image-analysis>. A static version of these has been created at Zenodo (<https://doi.org/10.5281/zenodo.7036029>). Scripts at the end for RNA sequencing analysis are available at Zenodo (<https://doi.org/10.5281/zenodo.6981721>).

4.1 FIJI/ImageJ

ImageJ was first developed in embryo in the 1980s as a means of viewing scientific images on the Mac, but matured into the program we know today in the 1990s when it was ported to Java. The original ImageJ1 is still developed and maintained today and serves as the foundation of its functional successor, FIJI. *Fiji Is Just ImageJ* is essentially ImageJ2 bundled with a set of useful plugins that aid in visualizing and analyzing scientific imaging data. Preeminent among these bundled plugins is a library of programming languages that can be used to interface with the underlying Java-based APIs that make FIJI function. Among the options, Python is likely the most widely known and written programming language among biologists and this opens up a great deal of potential for object-oriented programming centered on the analysis of images.

The default language in ImageJ, the ImageJ1 Macro language (.ijm), is approxi-

mately based around Java but lacks defined sets of methods (for most purposes) and objectification, has difficulty with interfacing with file systems, and exposes a great deal of underlying computer logic to the user. Take, for instance, the procedure for writing a *for* loop:

```
for (i = 0; i < 10; i++) {  
    print("Hello");  
}  
  
for i in range(1:10):  
    print("Hello")
```

The former example, written in the IJ1 macro language, exposes the user to looping over a set of indices from 0 to 10 and incrementing with each loop, uses curly braces to define the boundaries of the loop block, and requires each line to end with a semicolon. These are all standards features of many popular programming languages, but for FIJI, the key is to be as legible of a programming language as possible since the writers of most of these macros are *not* software engineers. Python, by contrast, increments for the user and will increment through an entire set of objects without needing to know how many objects are present. Blocks of code are delimited by spaces or tabs, so the relative location of the code on the line is informative as to what it relates to. Additionally, Python has well-studied and well-defined object types with specific associated methods that translate between all versions of Python. The benefits of the Python community only add to the value proposition of learning and using Python both generally and within ImageJ.

Not only this, but to loop through directories of files, the ImageJ macro language struggles even further, with little awareness of file structures or regular expressions whereas Python can trivially fetch lists of files by user-defined patterns from mixed directories and even search through subdirectories recursively. These features make

operating system-aware and object-oriented programming languages like Python a more convenient option for scaling up programs and defining subfunctions within them to build more complex applications.

4.2 Python

As alluded to, Python is a dynamically typed high-level programming language that is widely used for generating automation scripts of all varieties in addition to web server backend applications, data science, and even game creation (van Rossum, 1995). The advantages of Python are derived from the enormous community of developers who use Python and who continue to develop it as a language. In data science, the utility of pandas and numpy are unmatched and these alone make Python a language worth learning for any molecular biologist in the analysis of their quantitative data. Throughout this work, I have used Python both within the FIJI/ImageJ interpreter (which is based on Jython version 2.7.2) and via CPython 3.11, the standard distribution of Python, in the Terminal. While Jython is nominally different from standard Python, when written appropriately, it is simple enough to write version-universal code within the relatively narrow confines of what I have sought to accomplish thus far with it.

One of the key developments in the field of FIJI and Python is the release of PyImageJ, a complete API interface between CPython and ImageJ2, written in Python (Rueden et al., 2022). Going forward, it will be useful to reimagine some of these scripts and approaches to utilize the PyImageJ-defined API, as this is clearly the future of scripting in FIJI. Use of the image viewer in an *ad hoc* fashion is likely superior to an image-first approach for many tasks and PyImageJ cuts a lot of the unfortunate overhead from the graphical elements of FIJI. Unfortunately, such a translation is

nontrivial as the command structure and imports have changed quite dramatically and in ways that are not as well documented as the base API. However, such an implementation allows FIJI/ImageJ to make use of pandas, numpy, and OpenCV in Python, making for a more complete data science package and streamlining data collection and processing in a single language.

4.2.1 Napari

While FIJI/ImageJ have served as the key to the analyses presented both here and in Chapter 3, it is not the only image analysis software available. An alternative, written in pure Python, is napari, which is in the middle stages of alpha testing as of this writing (Sofroniew et al., 2022). While not yet feature complete, it is a lightweight alternative to FIJI and, with proper optimization, can likely outperform FIJI in the long run. By shedding dependence on Java, memory management and performance are able to be improved while the lowest-level image processing can be done in C invisible to the end user. At the present time, however, FIJI seems to be the most robust and widely accepted option and will likely remain so for the conceivable future. A bridging option, *napari-imagej* is in the works and promises to dramatically expand the napari feature set, but does not aid in napari independence from Java or older modalities (<https://github.com/imagej/napari-imagej>).

4.3 R

As previously detailed (Section 2.10), R is a convenient and easy-to-learn object-oriented statistical programming language. This language has made a monumental contribution to data science and is used extensively throughout this work to analyze and visualize the resulting datasets. While less extensively used in this segment than

Python, it is key to a lot of further analyses and is exhaustively used for all of the visualization seen in Chapter 3. Many of these same functions can be executed in Python, but I attempt to utilize the best tool for a given task at a given time and R is often the best, most flexible way to work with arrays and data frames.

4.4 Maximum-Intensity Projection and Composite Image Generation

The fundamental premise of the ImageJ macro system is to simplify repetitive tasks to free up user time for higher forms of analysis and the default language (the ImageJ1 macro language) makes it easy to write procedural operations to be done on single images, but is difficult to scale to whole directories of images or accept various types of user input. I have thus developed a set of scripts that allow the user to rapidly generate maximum-intensity projections from sets of images and then generate composite images. This often condenses hundreds of megabytes of data into <30 MB, makes for more flexible image viewing and understanding, and allows the images to be opened in essentially any number (many hundreds to thousands) on any modern personal computer. These procedures are also internally, if naïvely, memory managed, allowing them to run on most personal computers indefinitely to process the sometimes thousands or tens of thousands of images that can be generated over the course of an experiment with multiple wavelengths, Z stacks, XY positions, and times¹. In our lab, we primarily use output files from epifluorescent Zeiss microscopes, which generate .czi files and from Metamorph connected to a custom spinning disk confocal system, which generates .tiff files.

¹If you have 4 channels across 48 stage positions, capturing every 30 minutes for 48 hours, you would have 18,432 individual images based on Metamorph's file saving structure; this can clearly get out of hand very quickly.

The maximum intensity projection is an extremely common way of condensing multidimensional images into a single two-dimension representation by finding the brightest pixel at every XY position and accepting it as the most “in focus” pixel. The success of this depends on images being properly exposed, but in most instances will generate a reasonably sharp image ready to be quantitated or presented. In any case, data spanning multiple Z stacks requires some degree of integration to be usefully presented to others in the static format of a publication or manual scanning through libraries of hundreds of images and these pipelines will facilitate that type of visualization at scale.

Source Code 4.1: This script allows the user to open as many files as their memory allotment will allow and then to Z project them one at a time with custom start and end positions. This ability often generates cleaner, sharper images by individually selecting the lowest and highest in-focus frames, but necessarily takes more time than a more automated approach.

```
from ij import IJ, ImagePlus
from ij.plugin import ZProjector
from ij.gui import NonBlockingGenericDialog
import os, sys, random
from os import path

gui = NonBlockingGenericDialog("MIP Ranges")

gui.addNumericField("First Slice: ", 0, 0)
gui.addNumericField("Last Slice: ", 0, 0)

gui.showDialog()

if gui.wasCanceled():
    IJ.error("Dialog cancelled!")
    exit()
# Everything below this ~should~ be in an else statement, but if it isn't
→ broke, don't fix it.

bot = gui.getNextNumber()
top = gui.getNextNumber()

imp = IJ.getImage()
```

```

if bot is 0 and top is 0:
    bot = 1
    top = int(imp.getImageStackSize())
if int(bot) > int(top):
    IJ.error("Bottom is greater than top, invalid combination.")
    exit()
if int(top) > int(imp.getImageStackSize()):
    IJ.error("Top is greater than the dimensions of the image - try again")
    exit()
if int(bot) < 1:
    IJ.error("Out of range - bottom number must be greater than or equal to
    ↵ 1")
    exit()

outputdir = str(imp.getOriginalFileInfo().directory)

proj = ZProjector.run(imp, "max", int(bot), int(top))
# This line is to show your MIP - if you want to see it, remove the #
# proj.show()

title = proj.getTitle()
out = path.join(outputdir, title)
IJ.saveAs(proj, "Tiff", out)

#Comment out these lines if you'd like to see your images before you
    ↵ close them.
imp.close()
proj.close()

rand = random.randint(1, 100)

if rand > 85:
    IJ.run("Collect Garbage", "")
else:
    pass

```

The use of Source Code 4.1 is to process a set of already opened images and generate maximum intensity projections from these and, optionally, save them back into the directory that they came from. This relatively simple set of GUI-guided processes allows the user to, in two clicks, accomplish a task that previously would

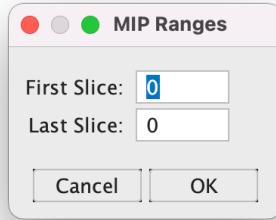


Figure 4.1: This is the entire graphical user interface for the manual maximum intensity projection and inputs required to run this script: the script asks for the top and bottom slices from the image of interest and then runs the rest of the script.

have required a great deal of menuing to accomplish. The goal is narrow, but this execution is extremely useful when working through large numbers of images. This is readily adaptable to any workflow as it has no features specific to a given image type or contents, so could easily be reused by many different individuals for their own purposes, like the subsequent scripts to process maximum intensity projections by other modes.

This script will first open a GUI with a simple set of inputs: the minimum and maximum slices that the user would like to use in creating the Z projection. This GUI is a “non-blocking” GUI, and can thus be opened in advance of deciding which slices to use – a convenience that frees the user from having to remember which slices were the best (Figure 4.1).

The internal logic of this macro seeks to minimize opportunities for user-derived error by checking whether the values provided are logical. The top slice needs to have a value greater than the bottom slice², but if both are left as the default (0),

²This is not a technical requirement – indeed, the ZProjector function itself can take these argument in either order, but it seemed to me that if the numbers are put in the wrong order that this suggested a user error in input and was not intended.

then it will create a maximum intensity projection across the whole image. This allows it to be a single-click convenience button for performing this operation on any number of images, although other options presented later are likely to be better choices for this. Additionally, if the top value is set to a number greater than the total number of slices, then it provides an error to the user. This adds some safety to the operations and prevents unintended operations, although the risk of data corruption is essentially non-existent as it saves the output with a new filename.

This, like many of the scripts further on, incorporates memory management to improve FIJI stability and prevent runaway memory leakage from Java, which very inefficiently clears out unused sectors even when the image has been closed. Thus, approximately one in every eight runs, the macro will run garbage collection to free up memory which can allow for more extended uptime for FIJI in the process of doing numbers of images far in excess of nominal memory space; however, for this, I would recommend looking to Source Code 4.2, which is written for essentially unlimited images in any reasonable memory buffer capacity.

A future version will incorporate a one-time checkbox that will allow the user to choose whether they would like to automatically reopen the dialog box for each image that they have open currently. If it is selected on the first run, it will then open for every subsequent image until cancelled. This should streamline these processes and further minimize clicks-per-operation, which should be the primary goal of these sorts of automation macros.

Source Code 4.2: A low overhead version of the manual maximum intensity projection script described above. Instead of opening all of the images first and then running the script, the script will processively open unanalyzed images one at a time and periodically garbage collect, allowing for entire directories to be processed at once on most reasonably modern computers.

```
from ij import IJ, ImagePlus
```

```

from ij.plugin import ZProjector
from ij.gui import NonBlockingGenericDialog, GenericDialog
import os, sys, random
from os import path

# This is a low-overhead version of the manMIPper that loads and displays
# images one at a time and only if needed.

gui = GenericDialog("File Directory: ")
gui.addDirectoryField("Z-Stacks Location", "~/Documents")
gui.addChoice("File Extension: ", [".czi", ".lif", ".lsm", ".tif",
    ".tiff", "Other"], ".tif")
gui.addStringField("Custom File Extension: ", "")
gui.showDialog()

basedir = str(gui.getNextString())

inputdir = basedir
outputdir = basedir + "MIPs/"
if not path.isdir(outputdir):
    os.makedirs(outputdir)

filenames = os.listdir(inputdir)

ext = str(gui.getNextChoice())
cusext = str(gui.getNextString())

extension = ""

if ext != "Other":
    extension = ext.lower()
elif ext == "Other":
    extension = cusext.lower()

rep_files = []

for files in filenames:
    if files.lower().endswith(extension) and "_thumb_" not in
        files.lower():
        file = os.path.join(inputdir, files)
        rep_files.append(file)

for f in rep_files:
    mips = [mip.lower() for mip in os.listdir(outputdir)]
    mip_title = str("MAX_" + os.path.basename(f)).lower()

```

```

if mip_title.replace(extension, ".tif") not in mips:
    imp = IJ.openImage(f)
    imp.show()

gui = NonBlockingGenericDialog("MIP Ranges")
gui.addNumericField("First Slice: ", 0, 0)
gui.addNumericField("Last Slice: ", 0, 0)
gui.showDialog()

if gui.wasCanceled():
    IJ.error("Dialog cancelled!")
    exit()

bot = gui.getNextNumber()
top = gui.getNextNumber()

if bot is 0 and top is 0:
    bot = 1
    top = int(imp.getImageStackSize())
if int(bot) > int(top):
    IJ.error("Bottom is greater than top, invalid combination.")
    exit()
if int(top) > int(imp.getImageStackSize()):
    IJ.error("Top is greater than the dimensions of the image - try
        ↵ again")
    exit()
if int(bot) < 1:
    IJ.error("Out of range - bottom number must be greater than or
        ↵ equal to 1")
    exit()

proj = ZProjector.run(imp, "max", int(bot), int(top))
title = proj.getTitle()
out = path.join(outputdir, title)
IJ.saveAs(proj, "Tiff", out)
imp.close()
proj.close()
else:
    pass
rand = random.randint(1, 100)
if rand > 85:
    IJ.run("Collect Garbage", "")
else:
    pass

```

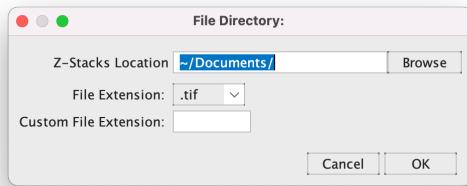


Figure 4.2: This GUI pop-up asks the user for the file directory that they would like to process and will process only the images directly in that folder; it also asks for the file extension so that it will only process the desired images in case the user has additional data in that folder.

Source Code 4.2, while not always the correct choice depending on user preferences and system capabilities, is much less memory hungry than the original version above, but typically results in slightly slower overall operations due to the delay in opening images. If the images are on fast internal storage, then this is certain to be faster than Source Code 4.1, but when reading from external storage, the I/O limitations will likely make it faster to open all of the images prior to processing unless system memory is severely limited. Nevertheless, the nature of this makes it very generally useful on older systems.

The user interface prompts the user to provide a directory that contains all of the desired images and provide the file extension from a pre-populated list or provide their own (Figure 4.2). These are the only user-inputs required to get this processing started; from there, a dialog box will open after each image has been opened requesting the first and last slices for maximum intensity projection, which can vary from image to image.

The internal logic of this script prevents the user from accidentally processing the same image more than once; this allows the processing to be done in sessions over time rather than all at once. The script looks for a file with the expected naming

structure for the output and, if it is found, removes the original file from the list of inputs. This also hedges against FIJI crashing or other errors. To do an image again due to some user input error, the user can either use Source Code 4.1 or remove the projection from the directory and the script will then allow it to be processed again.

This script also utilizes an identical memory management approach to the previous, which has allowed this to function over thousands of images at once. This offers a distinct advantage over manual memory management approaches that have to be used during manual processing and the requirement to open all of the images beforehand, a process inherently limited by available system memory.

Source Code 4.3: This script can be used in instances where the first and last stacks of a desired Z projection span the entire set of stacks provided. It will process an entire directory of images together and output the result into a subdirectory of the original.

```
from ij import IJ, ImagePlus
from ij.plugin import ZProjector
from ij.gui import GenericDialog
import os, sys, random
from os import path

gui = GenericDialog("File Directory: ")
gui.addDirectoryField("Z-Stacks Location", "~/Documents")
gui.addChoice("File Extension: ", [".czi", ".lif", ".lsm", ".tif",
    ".tiff", "Other"], ".tif")
gui.addStringField("Custom File Extension: ", "")
gui.showDialog()

basedir = str(gui.getNextString())

inputdir = basedir
outputdir = basedir + "MIPs/"
if not path.isdir(outputdir):
    os.makedirs(outputdir)

filenames = os.listdir(inputdir)

ext = str(gui.getNextChoice())
```

```

cusext = str(gui.getNextString())

extension = ""

if ext != "Other":
    extension = ext
elif ext == "Other":
    extension = cusext

rep_files = []

for files in filenames:
    if files.endswith(extension):
        file = os.path.join(inputdir, files)
        rep_files.append(file)

for f in rep_files:
    imp = IJ.openImage(f)
    if "MAX_" + imp.getTitle().rstrip(extension) + ".tif" not in
       os.listdir(outputdir):
        proj = ZProjector.run(imp, "max all")
        title = proj.getTitle()
        out = path.join(outputdir, title)
        IJ.saveAs(proj, "Tiff", out)
        imp.close()
        proj.close()
    rand = random.randint(1, 100)
    if rand > 85:
        IJ.run("Collect Garbage", "")
    else:
        pass

```

While the previous scripts have expected user input for each image, the skilled microscopist can select top and bottom slices that will suffice for generating Z projections during imaging itself. This means that the first and last slices of the projection are typically the first and last slices of the images *in toto*. This script takes a directory of images as an input and will perform total maximum intensity projections on all of them and save in a new subdirectory. After observation, any that seem incorrect can then be processed with one of the preceding scripts, especially Source Code 4.1.

The only prompting from the user is to provide a directory and file extension, just like Source Code 4.2. As a result of the lack of user intervention during processing, the internal logic of this script is much simpler and will simply perform top-to-bottom projections of each of the files in the directory and then save them in a subdirectory of the original directory provided. This operation can easily process hundreds of images per minute and cuts processing time down dramatically – whereas it can take 10-15 seconds per image doing it strictly by GUI, it can now do dozens to hundreds of images in that same time period. Not having to physically show the image also improves performance, although the user can opt to have it show the projections if desired by simple modifications to the code. A future version should incorporate an option to turn this on or off within the GUI; it should be reasonable for most computers to be able to show the maximum intensity projections for at least a few hundred images, although selecting such an option will come with some performance penalty.

Source Code 4.4: An interface to functions allowing slices in a Z-stack to be kept or removed as desired through function calls. This can integrate into other workflows and be connected to the previous scripts through higher-order wrappers.

```
from ij import IJ, ImagePlus, ImageStack, WindowManager
from ij.gui import GenericDialog
from ij.measure import Calibration
from ij.plugin import Plugin
from ij.process import ImageProcessor
from os import path

# Defaults:
first = 1
last = 9999
inc = 2
imp = IJ.getImage()

def sliceKeeper(imp, first, last, inc):
    imp = IJ.openImage(imp)
```

```

if not imp:
    IJ.noImage()
stack = imp.getStack()
if stack.getSize() is 1:
    IJ.error("Stack Required")
title = imp.getTitle().split(".")[0]
cal = imp.getCalibration()
impdir = str(imp.getOriginalFileInfo().directory)

def keepSlices(stack, first, last, inc):
    if last > stack.getSize():
        last = stack.getSize()
    newstack = ImageStack(stack.getWidth(), stack.getHeight())
    for slice in range(first, last + 1, inc):
        if slice > stack.getSize():
            break
        ip = stack.getProcessor(slice)
        newstack.addSlice(ip)
    imp = ImagePlus(title + "_kept", newstack)
    imp.setCalibration(cal)
    outputdir = path.join(impdir, title)
    IJ.saveAs(imp, "Tiff", outputdir)

keepSlices(stack, first, last, inc)

def sliceRemover(imp, first, last, inc):

    imp = IJ.openImage(imp)
    if not imp:
        IJ.noImage()
    stack = imp.getStack()
    if stack.getSize() is 1:
        IJ.error("Stack Required")
    title = imp.getTitle().split(".")[0]
    cal = imp.getCalibration()
    impdir = str(imp.getOriginalFileInfo().directory)

    def removeSlices(stack, first, last, inc):
        if last > stack.getSize():
            last = stack.getSize()
        newstack = stack.duplicate()
        for slice in range(first, last + 1, inc):
            if slice > stack.getSize():
                break
            newstack.deleteSlice(slice)

    removeSlices(stack, first, last, inc)

```

```
imp = ImagePlus(title + "_cut", newstack)
imp.setCalibration(cal)
outputdir = path.join(imkdir, title)
IJ.saveAs(imp, "Tiff", outputdir)

removeSlices(stack, first, last, inc)
```

FIJI/ImageJ comes with a built-in option to keep and remove particular slices (Image > Stacks > Tools > Slice Keeper / Slice Remover), but the native plugin does not readily fit into object-oriented programming pipelines like those used by Python and Java; it is very operational toward the currently open image and does not have a well-defined API. Thus, I have adapted the underlying logic of these plugins to be wrapped in various other scripts through calls to the defined functions, *sliceKeeper* and *sliceRemover*. For instance, this allows calls to the maximum intensity projection plugins to be funneled into this plugin to simultaneously generate the kept slices as well as the maximum intensity projection of those kept slices for some useful improvements to record keeping.

Integration of this as in import into the other scripts previously described with an additional option would be an intelligent way of making use of this plugin; it also reveals the many opportunities to expand upon and add functionality to the previously detailed plugins based on user needs. The addition of saving the exact slices used to generate maximum intensity projections along with some sort of metadata seems like it would improve data integrity in a meaningful way, although at the cost of increased storage requirements.

4.5 Building Multi-Dimensional Images from Metamorph File-Saving Logic

Metamorph is a commercial image capture and processing software that is used to gather images from a variety of different microscopy systems. It saves these images as a series of TIFF files separated based on channel, time, and scene but incorporates all of the Z positions for a given channel/time/scene in a single image and saves some useful metadata within .nd files. These ND files can then be utilized by a FIJI/ImageJ plugin developed by Cordelières (2005) to process these images into user-interpretable files, but this process, at least in our hands, is often slow and cumbersome, taking many hours or days to process approximately 0.5 to 1.5 terabytes of imaging data. Admittedly, this expansive scale of imaging is beyond the scope of what could possibly have been expected when this plugin was developed in 2005 (for Java version 5), but it has created difficulties in efficiently processing the images we have into a useful format.

To address these challenges, I have developed a novel processing modality in Python that can be used within FIJI that demonstrates substantial improvements in performance over the previous approach, with tasks generally reduced from hours to days to minutes to hours, all limited by input/output speeds from the disk, as these are often stored on USB 3.0 external storage drives. While the original ND stack builder offers a number of potentially useful features that are currently lacking from this implementation, this should cover most common use cases and, with a modular functional design, can be readily expanded upon in the future. While bugs are likely in these early versions, this should be a useful platform for the analysis of high-resolution

confocal images with high resolution across X, Y, Z, C, T, and S.

Source Code 4.5: This code allows for the efficient processing of images captured from Metamorph by leveraging the unified naming structure used by Metamorph to piece together corresponding images across all available dimensions. The code assumes as little as possible and explicitly requests necessary pieces of information from the user within a simple GUI. The simplicity of the underlying code and internal memory management allows for dramatic improvements in performance relative to existing approaches and should facilitate much more rapid image processing and data analysis while also offering a flexible platform the for the addition of further functionality.

```
from ij import IJ, ImagePlus
from ij.plugin import ZProjector, RGBStackMerge, Concatenator, Duplicator
from ij.gui import GenericDialog, NonBlockingGenericDialog
import os, sys, random, re
from os import path

gui = GenericDialog("Metamorph File Compiler")
gui.addMessage("Version 0.1a, 17 November 2022")
gui.addDirectoryField("Files Directory: ", "~/Documents")
gui.addCheckbox("Multiple Scenes?", True)
gui.addCheckbox("Multiple Time Points?", True)
gui.addCheckbox("Show Merged Image?", False)
gui.addCheckbox("Use Metadata from .nd File?", False)

gui.showDialog()

basedir = str(gui.getNextString())
multiscene = gui.getNextBoolean()
timelapse = gui.getNextBoolean()
opener = gui.getNextBoolean()
nder = gui.getNextBoolean()

outputdir = path.join(basedir, "comps")
if not path.isdir(outputdir):
    os.makedirs(outputdir)

inputs = os.listdir(basedir)
filenames = []
ext = []

for input in inputs:
    if "thumb" not in input and input.lower().endswith(".tif") and not
       input.startswith(".") and not input.startswith("_"):
        filenames.append(input)
```

```

    ext.append("." + input.rsplit(".", 1)[1])

ext = set(ext)

parts = []

for files in filenames:
    part = files.rsplit("_", 3)
    parts.append(part)

names = []
chans = []
scenes = []
times = []

for p in parts:
    names.append(p[0])
    if multiscene and timelapse:
        chans.append(p[1])
        scenes.append(int(p[2].replace("s", "")))
        times.append(int(p[3].split(".")[-1].replace("t", "")))
    elif multiscene:
        chans.append(p[1])
        scenes.append(int(p[2].split(".")[-1].replace("s", "")))
    elif timelapse:
        chans.append(p[1])
        times.append(int(p[2].split(".")[-1].replace("t", "")))
    else:
        chans.append(p[1]).split(".")[0]

names = list(set(names))
chans = list(set(chans))
scenes = list(sorted(set(scenes)))
times = list(sorted(set(times)))

chan_dict = {}
chan_pairs = ""
for chan in chans:
    chan_dict["C" + str(int(chans.index(chan) + 1))] = chan
    chan_pairs += "C" + str(int(chans.index(chan) + 1)) + ": " + chan +
    "\n"

chan_list = ["C" + str(int(chans.index(chan) + 1)) for chan in chans]
chan_list.append("None")

```

```

gui = NonBlockingGenericDialog("Color Matcher")
gui.addMessage(chan_pairs)
gui.addChoice("Red: ", chan_list, "None")
gui.addChoice("Green: ", chan_list, "None")
gui.addChoice("Blue: ", chan_list, "None")
gui.addChoice("Grey: ", chan_list, "None")
gui.addChoice("Cyan: ", chan_list, "None")
gui.addChoice("Magenta: ", chan_list, "None")
gui.addChoice("Yellow: ", chan_list, "None")

gui.showDialog()

red_pick = str(gui.getNextChoice())
green_pick = str(gui.getNextChoice())
blue_pick = str(gui.getNextChoice())
grey_pick = str(gui.getNextChoice())
cyan_pick = str(gui.getNextChoice())
magenta_pick = str(gui.getNextChoice())
yellow_pick = str(gui.getNextChoice())

colors = [red_pick, green_pick, blue_pick, grey_pick, cyan_pick,
          magenta_pick, yellow_pick]
for key, value in chan_dict.items():
    if key not in colors:
        del chan_dict[key]

chans = chan_dict

def metamorpher(names, chans, scenes, times, ext, colors, timelapse =
               True, multiscene = True, opener = False, nnder = False):

    def titler(nnder, name, scene):
        if nnder:
            if path.isfile(path.join(basedir, name + ".nd")):
                pos_dict = {}
                with open(path.join(basedir, name + ".nd"), "r") as nd_file:
                    for line in nd_file.readlines():
                        if line.startswith("\\"Stage"):
                            pos_dict[line.split(",")[0].strip("\\"\\n").replace("Stage",
                                "")] = line.split(",")[1].strip("\\"\\n\\r ")
                if str(scene) in pos_dict:
                    scene_name = pos_dict.get(str(scene))
                    title = "_" .join([name, scene_name])
            else:
                title = "_" .join([name, str(scene)])

```

```

    return title

def stiller(timelapse, combo_dict, name, chan, times, scene = "scene"):
    stills = []
    if timelapse:
        for time in times:
            still = path.join(basedir, "_" .join([name, chan, "s" +
                ↵ str(scene), "t" + str(time) + "" .join(ext)]))
            if path.isfile(still):
                still = ImagePlus(still)
                if still.getNSlices() > 1:
                    still = ZProjector.run(still, "max")
                    stills.append(still)
                else:
                    stills.append(still)
            if stills:
                combo_dict[key] = Concatenator.run(stills)
    else:
        still = path.join(basedir, "_" .join([name, chan, "s" + str(scene) +
            ↵ "" .join(ext)]))
        if path.isfile(still):
            still = ImagePlus(still)
            if still.getNSlices() > 1:
                still = ZProjector.run(still, "max")
                stills.append(still)
            else:
                stills.append(still)
        if stills:
            combo_dict[key] = ImagePlus(still)

def merge_and_save(combo_dict, colors, title, outputdir, opener):
    color_picker = []
    for color in colors:
        if color != "None":
            color_picker.append(combo_dict[color])
        elif color == "None":
            color_picker.append(None)
    merge = RGBStackMerge.mergeChannels(color_picker, False)
    if opener:
        merge.show()
    out = path.join(outputdir, title)
    del title
    IJ.saveAs(merge, "Tiff", out)
    rand = random.randint(1, 100)
    if rand > 85:

```

```

IJ.run("Collect Garbage", "")
else:
    pass

for name in names:
    if timelapse and multiscene:
        for scene in scenes:
            title = titler(nder, name, scene)
            print(title)
            combo_dict = {}
            for key, chan in chans.items():
                stiller(timelapse, combo_dict, name, chan, times, scene)
            if combo_dict:
                merge_and_save(combo_dict, colors, title, outputdir, opener)
    elif timelapse:
        title = name
        combo_dict = {}
        for key, chan in chans.items():
            stiller(timelapse, combo_dict, name, chan, times, scene)
        if combo_dict:
            merge_and_save(combo_dict, colors, title, outputdir, opener)
    elif multiscene:
        # If you have a multiple scene, non-timelapse image (I'm not sure
        # ↵ how or why it would have been set up like that?), there are
        # ↵ definitely better options out there, but this should work?
        for scene in scenes:
            title = titler(nder, name, scene)
            combo_dict = {}
            for key, chan in chans.items():
                stiller(timelapse, combo_dict, name, chan, times, scene)
            if combo_dict:
                merge_and_save(combo_dict, colors, title, outputdir, opener)
    else:
        title = name
        combo_dict = {}
        num_stacks = []
        for key, chan in chans.items():
            stiller(timelapse, combo_dict, name, chan, times, scene)
        if combo_dict:
            merge_and_save(combo_dict, colors, title, outputdir, opener)

metamorpher(names, chans, scenes, times, ext, colors, timelapse,
    ↵ multiscene, opener, nder)

gui = GenericDialog("All Done!")

```

```
gui.showDialog()
```

The main body of code simply serves to process the files within the user-provided directory into useful bits of information that can ultimately be used to regenerate the file names of the desired images. These pieces (names, channels, scenes, and times) are then fed into the `metamorpher()` function, which contains a number of child functions that unify the processing pipeline into a set of logical statements based on user-provided booleans concerning the nature of their images. Some of these pieces of information could be extracted from the `.nd` file in future versions, which would further reduce user error or enable further levels of user customization (for instance, the precise range of the Z projection or the number of time points that should be processed). By focusing on text-level processing, the advantages of Python are enhanced while limiting the need for extraneous image processing, as only the images that will actually be used are loaded into memory and, once loaded, are rapidly condensed (by Z projection) and cleared. This results in very rapid image concatenation and merging that can condense terabyte-scale data into more readily interpretable data on the gigabyte-scale that can be readily read into the memory of most standard consumer-grade computers. Indeed, one of the advantages of this processing modality is that, while performance on workstation-class computers with expansive memory will always be superior, many of these processing tasks are now possible on consumer-grade hardware. For instance, this script has been demonstrated to be able to process a time lapse file set containing 12 stage positions of 1024 X, 1024 Y, and 10 Z across three channels and 321 time points into an easily manipulable set of 12 images (one for each stage position) in less than an hour on a 2021 14" M1 Max MacBook Pro with 64 GB of unified memory. And thanks to the internal memory management of this script and the memory management of macOS, it should be

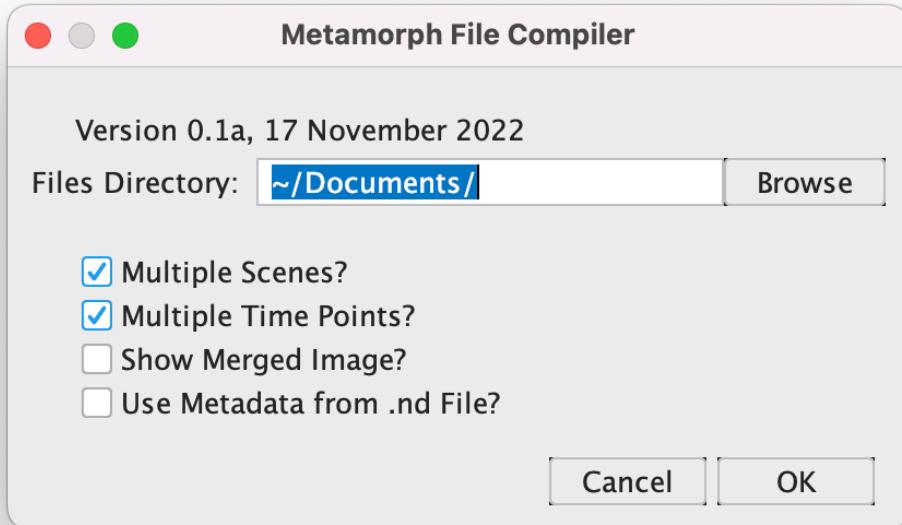


Figure 4.3: When the metamorpher script is run, this is the first GUI to pop up, which requests the file directory where the images are saved as well as some basic details about the experiment: was it a time lapse, where there multiple stage positions, would the user like to show the images once they are generated, and would they like to use information from the .nd file, which is currently limited to the names of the stage positions provided at the time of experimental set up. Future versions should more exhaustively use what metadata is included in the .nd file to emulate the existing behavior of “nd stack builder.”

reasonable to expect that such processing is possible (albeit potentially a bit slower) on less capable systems. In this instance, it was able to take >200 GB of files and condense them into 24 GB (~2 GB for each image set), which is relatively trivial to process.

Future versions of this script should seek partial feature parity with the original “nd stack builder” as a means of more fully displacing this functionality. In its current version, this script is unable to take a subset of Z positions for projection,

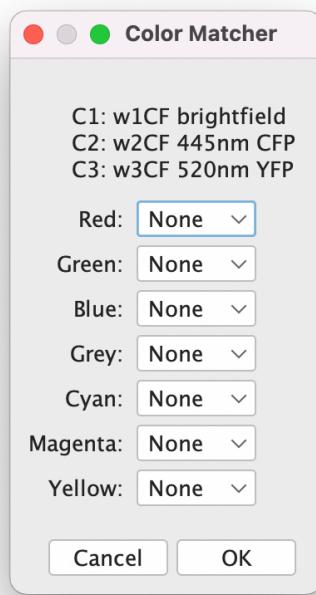


Figure 4.4: Once the directory has been selected, this will offer the use the option to match their channels with particular lookup tables based on the file name patterns. It helpfully provides which channel is which, which also offers flexibility across different types of experiments with different channels in different orders.

which should be functionality easily ported from previous scripts (Source Code 4.1). However, across such extensive time, the sample often moves and the full scope of the Z stack is often necessary to make for an approximately appropriate image across these time periods. Nevertheless, expanding this functionality seems useful for various edge-case scenarios. Advantageous to this is that the underlying processes are designed to be modular and have been consolidated into a number of defined functions that should allow for flexible expansion and addition of further logic. Additionally, the entire procedure is written in either base Python or using well-defined ImageJ functions, which should allow for relative code stability and, if Jython 3.x ever arrives, all of the code is written to be version independent and should function for 3.x versions of Python.

Lastly, this should be ported to PyImageJ to further future-proof this process. This should be a comparatively straight-forward translation task now that the underlying logic is defined, but the subtle incompatibilities between these two approaches to using Python for ImageJ are a meaningful hurdle.

4.6 Surface Plot Analysis for Cellular Distribution of Labeled Proteins³

The distribution of a protein across the cell body can indicate changes in function of the protein and alterations in cell behavior, but analysis of such changes is not quite trivial. There are methods for drawing lines and plotting fluorescence intensity, but this fails to integrate the entire cell in three dimensions in to the analysis as it proves to be only one dimensional along that line. To analyze the distribution of ARPC2 in

³Taken from personal contributions to J. W. Saelens et al. (2022). “An ancestral mycobacterial effector promotes dissemination of infection”. In: *Cell* 185.24, P4507–4525.E18.

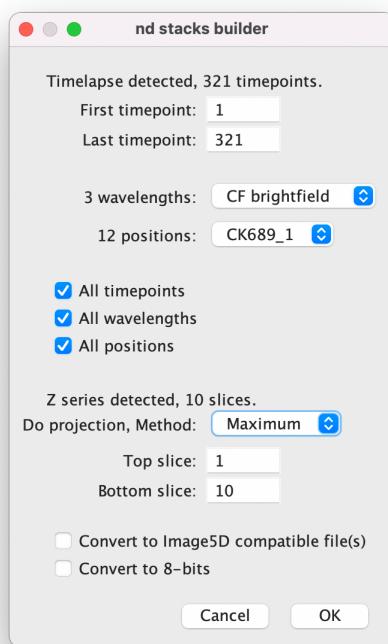


Figure 4.5: This screenshot of the GUI used for “nd stack builder” shows the options provided, which integrates with the .ND files included by Metamorph in the file directory. These contain simple string-based metadata that the “nd stack builder” uses to process these images which is in contrast to my patterning-based approach.

BLaER1 macrophages during *M. tuberculosis* infection, I developed a novel analysis pipeline that integrated the 3D Surface Plot plugin of ImageJ. This pipeline takes a raw image, isolates the desired channel, opens it in 3D Surface Plot for visualization, and then takes the user-generated output of 3D Surface Plot to find the maximum fluorescence along the longest axis of the cell. As this analysis is designed for macrophages, which can develop long spindles, this allows for quantitative proximal/distal differences in the localization of a protein to be determined.

Source Code 4.6: A script to isolate a single cell within a frame.

```

from ij import IJ, ImagePlus
from ij.io import DirectoryChooser, FileSaver
from ij.plugin import ChannelSplitter, ZProjector
from ij.plugin.frame import RoiManager
from ij.gui import GenericDialog
import os, sys
from os import path

gui = GenericDialog("File Directory: ")
gui.addDirectoryField("Cell Images", "~/Documents")
gui.addChoice("File Extension: ", [".czi", ".lif", ".lsm", ".tif",
    " .tiff", "Other"], ".tif")
gui.addStringField("Custom File Extension: ", "")
gui.addNumericField("Reference Channel: ", 2, 0)
gui.addNumericField("Measurement Channel: ", 1, 0)
gui.showDialog()

basedir = str(gui.getNextString())

inputdir = basedir
outputdir = basedir + "/Splits/"
if not path.isdir(outputdir):
    os.makedirs(outputdir)

filenames = os.listdir(inputdir)

ext = str(gui.getNextChoice())
cusext = str(gui.getNextString())

extension = ""

```

```

if ext != "Other":
    extension = ext
elif ext == "Other":
    extension = cusext

rep_files = []

for files in filenames:
    if files.endswith(extension):
        if not files.startswith("."):
            file = os.path.join(inputdir, files)
            rep_files.append(file)

for f in rep_files:
    imp = IJ.openImage(f)
    proj = ZProjector.run(imp, "max all")
    imps = ChannelSplitter.split(proj)
    for i in imps:
        title = i.getTitle()
        out = path.join(outputdir, title)
        IJ.saveAs(i, "Tiff", out)
        ImagePlus.close(i)

###

splits = os.listdir(outputdir)

tifs = []

for files in splits:
    if files.endswith(".tif"):
        file = os.path.join(outputdir, files)
        tifs.append(file)

names = []

for tif in tifs:
    base = str(tif.split("-", 1)[1])
    names.append(base)

names = set(names)

rm = RoiManager.getRoiManager()

tifs.sort(reverse = True)

```

```

ref = str(int(gui.getNextNumber()))
meas = str(int(gui.getNextNumber()))

for name in names:
    for tif in tifs:
        if tif.startswith(outputdir + "C" + ref) and tif.endswith(name):
            a = IJ.openImage(tif)
            IJ.setAutoThreshold(a, "MinError dark")
            IJ.run(a, "Analyze Particles...", "size=100.00-Infinity clear
                include add")
            ra = rm.getRoisAsArray()
            rm.reset()
            ImagePlus.close(a)
        if tif.startswith(outputdir + "C" + meas) and tif.endswith(name):
            b = IJ.openImage(tif)
            title = b.getTitle().lstrip(outputdir + "C" + meas + "-")
            out = path.join(outputdir, title)
            b.show()
            for r in ra:
                rm.addRoi(r)
            rm.select(b, 0)
            IJ.run(b, "Clear Outside", "")
            rm.reset()
            IJ.saveAs(b, "Tiff", out)

fin = GenericDialog("All Done!")
fin.showDialog()

```

The first step of this pipeline is to isolate the cell using some reference channel, which can also be the channel you are measuring if necessary, although I would recommend using some sort of pan-cytosolic or plasma membrane marker to more thoroughly mark the outline of the cell and to use that instead. The script presents a simple GUI (Figure 4.6) and allows the user to select the needed parameters and will then process entire folders of images into isolated cells in a single channel. A missing feature is that it needs to be able to isolate each cell from images that contain more than one cell, to save the user time on isolating cells in their own separate files by hand. This could conceivably be done by integrating logic around copying ROI

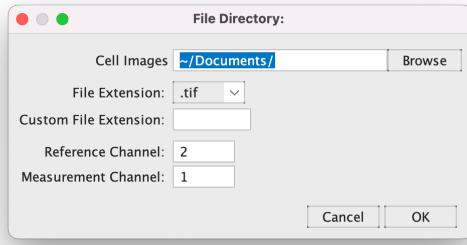


Figure 4.6: The user needs to provide a number of factors to allow for this function to operate properly: a file directory with the images that should be processed, the file extension, and a reference channel which should encompass approximately the entire cell, and the measurement channel, which is what the user wishes to actually measure.

contents into new images, but that is a work-in-progress at this time.

The output is a folder of single-channel files ready to be input into 3D Surface Plot, which can be launched with the following:

```
from ij import IJ, ImagePlus

imp = IJ.getImage()

IJ.run(imp, "3D Surface Plot",
      '''plotType=1
colorType=3
drawAxes=0
drawLines=0
drawText=0
grid=256
drawLegend=0
smooth=8.5
backgroundColor=000000
windowHeight=600
windowWidth=720'''')
```

The image then needs to be rotated until it is along its longest axis in XY and then oriented precisely perpendicular in XZ (this can be done by turning the axes

back on temporarily, although they must be turned off before the next step). This will give the longest two dimensional axis of the image and show a series of peaks and valleys corresponding to a smoothed-over rendition of the image. This can then be exported and, if desired, saved using:

```
from ij import IJ, ImagePlus
from os import path

outputdir = "" # Insert save directory as a string.
imp = IJ.getImage()
title = imp.getTitle()

out = path.join(outputdir, title)
IJ.saveAs(imp, "Tiff", out)
```

I recommend saving them for reference later, as the quantitation can occasionally be bugged by strange smoothing, etc. Any outliers will need to be inspected by hand and saving the intermediates can help a great deal with that. Additionally, saving allows further automation in the next step (although, as we will see, automating across all open images is also now possible and will be built into these options in the future).

Source Code 4.7: A script to automatically capture the signal at each point along an image and save it to a CSV file.

```
from ij import IJ, ImagePlus
from ij.gui import GenericDialog
from csv import writer
from os import path

gui = GenericDialog("Surface Plot Measure")
gui.addDirectoryField("File Location: ", "~/Documents")
gui.showDialog()

basedir = str(gui.getNextString())

inputdir = basedir
outputdir = basedir + "csvs/"
```

```

if not path.isdir(outputdir):
    os.makedirs(outputdir)

filenames = os.listdir(inputdir)

plot_files = []

for files in filenames:
    if files.startswith("Surface_Plot") and files.endswith(".tif"):
        file = os.path.join(inputdir, files)
        plot_files.append(file)

for f in plot_files:
    imp = IJ.openImage(f)
    title = imp.getTitle()

    IJ.run(imp, "8-bit", "")
    IJ.setRawThreshold(imp, 1, 255)
    IJ.run(imp, "Make Binary", "method=Default")

    width = imp.getWidth()
    height = imp.getHeight()

    pixel_width = []

    for num in range(0, width-1):
        pixel_width.append(num)

    pixel_height = []

    for num in range(0, height-1):
        pixel_height.append(num)

    max_coord = []

    for x_coord in pixel_width:
        for y_coord in pixel_height:
            if imp.getPixel(x_coord, y_coord)[0] == 255:
                max_coord.append([imp.getTitle(), x_coord, y_coord])

csv_file = path.join(outputdir, title.split(".", 1)[0] + ".csv")

with open(csv_file, "a") as csv:
    writer = writer(csv)
    for coord in max_coord:

```

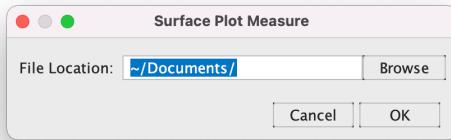


Figure 4.7: This simple GUI, reminiscent of previous examples, which asks only for the file directory that should be analyzed and will process all of the images matching the parameters in that directory. It selects specifically files that match the file naming pattern of surface plots and should be used directly after the previous script.

```
writer.writerow(coord)
```

This script is key to capturing the points along the abstracted profile of the cell. It is relatively simple in that it just writes to a CSV each XY coordinate where there is a signal, which can then be condensed by finding the maximum point at each X position in R, which is the final step in the analysis. This script presents a GUI to request the directory where these files are stored as the only necessary user input (Figure 4.7).

Source Code 4.8: An R script to capture the maximum point along the profile generated by the previous scripts and then calculate the area under the curve to compare different biological groups to one another.

```
# Jared Brewer
# Created: 21 July 2022
# Last Edited: 21 July 2022
# 3D Surface Profile Plot Analysis

library(ggplot2)
library(ggbeeswarm)
library(ggsignif)
library(dplyr)
library(DescTools)
library(stringr)

# Plug in the path were the .csv files are - make sure they are the only
# CSVs in the directory -
```

```

# unless you want to get more creative with some regex.

counts.dir <- ""
counts <- list.files(counts.dir, full.names = T)

cells <- data.frame()

# Our objective here is to combine all the individual CSV files into a
# single file, but to do a series of operations on it first.
# We read in the file, then aggregate into the minimum value
# (the data is inverted - need min, not max.)
# We then do some basic math operations to normalize the data across the
# length of the cell.

for (file in counts) {
  if (endsWith(file, ".csv")) {
    cells <- read.csv(file, header = F) |>
      aggregate(V3 ~ V1 + V2, data = _, min) |>
      mutate(pct_x = (V2 - min(V2))/(max(V2) - min(V2)),
             pct_y = (V3 - min(V3))/(max(V3) - min(V3))) |>
      mutate(x_pct_bin = ntile(pct_x, n=100),
             x_pct_corr = pct_x*100,
             y_pct_corr = -pct_y*100+100,
             treat = str_extract(V1, "WTCAFA|WTMAFA"),
             rep = as.numeric(str_extract(V1, "\\\d+$"))) |>
      rbind(cells)
  }
}

aucs <- data.frame()

# Now we want to measure the area under the curve for each replicate.
# We use nested for loops to read each treatment and each replicate for
# each treatment and filter.
# Then, we normalize across the total Y range, calculate the AUCs, and
# select only the columns we need.

for (t in unique(cells$treat)) {
  for (r in unique(cells$rep)) {
    aucs <- filter(cells, treat == t) |>
      filter(rep == r) |>
      mutate(y_pct_corr = y_pct_corr/sum(y_pct_corr)) |>
      mutate(per = AUC(x_pct_corr, y_pct_corr, from = min(0), to =
        max(25))/
```

```

        AUC(x_pct_corr, y_pct_corr, from = min(25), to =
          ↵ max(75)),
      mid = AUC(x_pct_corr, y_pct_corr, from = min(37.5), to =
          ↵ max(62.5))) |>
    select(per, mid, treat, rep) |>
  unique() |>
  na.omit() |>
  rbind(aucs)
}
}

t.test(aucs$per ~ aucs$treat)

# Plot the AUCs - must add your own labels.

auc.plot <- ggplot(aucs, aes(x = treat, y = per)) +
  geom_beeswarm(aes(shape = treat, size = 3)) +
  geom_boxplot(aes(fill = treat), alpha = 0.25, outlier.shape = NA,
    ↵ fatten = 5) +
  stat_boxplot(geom='errorbar', width = 0.25) +
  xlab("Genotype") + ylab("AUC") +
  geom_signif(y_position = c(0.6), xmin = c(1), xmax = c(2), annotations
    ↵ = c("p = 0.002"), textsize = 5.5, color = "black") +
  scale_x_discrete(limits = c("", ""), labels = c("", "")) +
  scale_fill_manual(name = "", labels = c("", ""), values = c("black",
    ↵ "black")) + theme(legend.position = "none") +
  guides(color = "none") + theme_minimal() +
  theme(text = element_text(size = 20, face = "bold"), plot.title =
    ↵ element_text(hjust = 0.5)) +
  theme_prism() +
  theme(legend.position = "none")

```

This R script takes the CSV format output from FIJI/ImageJ and processes it by finding the maximum value along the curve and using that as the peak relative fluorescence intensity for that X position and then calculates the area under the curve across a defined interval. For instance, the middle 50% of the cell can be defined as "central" and the outer 25% can be defined is "distal" and these values normalized to one another and then compared across different experimental groups. It was found that BLAER1 cells infected with *M. tuberculosis* expressing ancestral *esxM* had less

distal localization of ARPC2, suggesting that this bacterial effector was altering actin dynamics in these cells via modulation of ARPC2, a finding consistent with previous observations of macrophage from *Arpc2^{-/-}* mice (Rotty et al., 2017).

4.7 py-LaRoMe

The original purpose of LaRoMe (<https://github.com/BIOP/ijp-LaRoMe>) is to extract various features from images and is distributed as an ImageJ plugin. However, like many FIJI/ImageJ plugins, the design of LaRoMe is purely operational. In my attempts to utilize CellProfiler (Carpenter et al., 2006; Kamentsky et al., 2011; McQuin et al., 2018; Stirling et al., 2021) to automate capture of various details about the THP-1 macrophages from Subsection 3.3.9 and Subsection 3.3.10, I wished to incorporate LaRoMe into Python pipelines to increase my throughput of analysis. This, however proved challenging based on the original design of the plugins, as they did not accept an image as an argument and expected to use the currently active image window.

I therefore completely rewrote these plugins explicitly so that they would accept an image as the primary argument, which allows for easy integration into looping structures. In the process, the entire plugin was translated from Java to Python. It is hoped that in the future, these code bases can be reconciled so that the existing distribution of LaRoMe, managed by BIOP, can incorporate both the excellent GUI options already existing and allow for interfacing through scripts for high-throughput applications.

Source Code 4.9: A Python translation of the FIJI function “Label image to ROIs” from LaRoMe. This function allows the user to take images generated from CellProfiler and convert them into a set of regions of interest in the ROI Manager.

```

from ij import IJ, ImagePlus
from ij.gui import Wand, PolygonRoi, Roi
from ij.plugin import Duplicator
from ij.plugin.frame import RoiManager
from ij.process import ImageProcessor
import os

def labelsToROIs(imp):

    rm = RoiManager.getRoiManager()

    def L2R(imp):
        ip = imp.getProcessor()
        wand = Wand(ip)

        width = imp.getWidth()
        height = imp.getHeight()

        pixel_width = []

        for num in range(0, width - 1):
            pixel_width.append(num)

        pixel_height = []

        for num in range(0, height - 1):
            pixel_height.append(num)

        ip.setColor(0)

        for y_coord in pixel_height:
            for x_coord in pixel_width:
                if ip.getPixel(x_coord, y_coord) > 0:
                    wand.autoOutline(x_coord, y_coord)
                    if wand.npoints > 0:
                        roi = PolygonRoi(wand.xpoints, wand.ypoints, wand.npoints,
                                         Roi.TRACED_ROI)
                        roi.setPosition(imp.getCurrentSlice())
                        ip.fill(roi)
                        rm.addRoi(roi)
        rx = rm.getRoisAsArray()
        rm.reset()
        rois = []
        for r in rx:

```

```

if r not in rois:
    rois.append(r)
    rm.addRoi(r)

rm.runCommand(imp, "Show All")

copy_imp = imp.duplicate()
rm.reset()

dimensions = copy_imp.getDimensions()
nChannels = dimensions[2]
nSlices = dimensions[3]
nFrames = dimensions[4]

if (nChannels > 1 and nSlices > 1) or (nChannels > 1 and nFrames > 1)
    or (nSlices > 1 and nFrames > 1):
    print(imp.getTitle() + " is a hyperstack (multi c , z or t), please
        prepare a stack (single c, either z-stack or t-stack) from it.")
    pass
elif nChannels > 1 or nSlices > 1 or nFrames > 1:
    for stack in range(1, copy_imp.getImageStackSize() + 1):
        copy_imp.setPosition(stack)
        L2R(copy_imp)
else:
    L2R(copy_imp)

```

Source Code 4.9 allows the user to convert a labeled set of images, for instance from CellProfiler (Carpenter et al., 2006; Kamentsky et al., 2011; McQuin et al., 2018; Stirling et al., 2021), into a set of discrete regions of interest in the ROI Manager within FIJI. This allows for interesting layers of quantitation and segmentation that are otherwise difficult to perform within FIJI due to its difficulty in understanding the RGB layering in the TIFF files produced by CellProfiler. While this work ultimately did not bear much fruit, these approaches seem like they could be broadly useful for others in the future who are trying to do high-throughput, high-content data extraction from labeled images within a larger pipeline. The original implementation

is more user-friendly in many ways, but this implementation is, in my opinion, more flexible and with an easier to understand API.

Source Code 4.10: A Python translation of the FIJI function “ROIs to label image” from LaRoMe. This allows the user to use a set of ROIs to regenerate a label image, useful for creating masks on existing images and comparing areas between different channels.

```

from ij import IJ, ImagePlus
from ij.gui import Wand, PolygonRoi, Roi
from ij.plugin import Duplicator, Concatenator
from ij.plugin.frame import RoiManager
from ij.process import ImageProcessor
import os, re

def ROIsToLabels(rois, imp_path = None):

    rm = RoiManager.getRoiManager()
    rm.reset()
    rm.runCommand("Open", rois)
    rc = rm.getRoisAsArray()

    if not imp_path:
        pxm = []
        pym = []
        lrs = []
        for r in rc:
            lr = list(r)
            for l in lr:
                lrs.append(str(l).lstrip("java.awt.Point"))
        for lr in lrs:
            px = re.findall("x=\d+", lr)
            for p in px:
                pxm.append(int(p.lstrip("x=")))
            py = re.findall("y=\d+", lr)
            for p in py:
                pym.append(int(p.lstrip("y=")))

        imp_title = os.path.splitext(os.path.basename(rois))[0]
        imp = IJ.createImage(imp_title, "8-bit black", max(pxm), max(pym), 1)
    elif imp_path:
        imp = IJ.openImage(imp_path)

    def R2L(imp):

```

```

label_imp = imp.duplicate()
IJ.run(label_imp, "Grays", "")
IJ.run(label_imp, "16-bit", "")
rc = rm.getRoisAsArray()

stackN = label_imp.getImageStackSize()
isStack = bool()
if stackN > 1:
    isStack = True

if int(len(rc)) in range(1, 255):
    IJ.run(label_imp, "8-bit", "")
elif int(len(rc)) in range(256, 65535):
    IJ.run(label_imp, "16-bit", "")
else:
    IJ.run(label_imp, "32-bit", "")

for stack in range(1, stackN + 1):
    label_imp.getStack().getProcessor(stack).setValue(0)
    label_imp.getStack().getProcessor(stack).fill()

for i in range(1, len(rc)):
    if isStack:
        label_imp.setPosition(rc[i].getPosition())
    label_imp.getProcessor().setValue(i + 1)
    label_imp.getProcessor().fill(rc[i])

label_imp.setTitle("ROIsToLabels_" + imp.getTitle())
imp_dir = os.path.dirname(rois)
limp_title = label_imp.getTitle()
IJ.saveAs(label_imp, "Tiff", os.path.join(imp_dir, limp_title))

rm.reset()

dimensions = imp.getDimensions()
nChannels = dimensions[2]
nSlices = dimensions[3]
nFrames = dimensions[4]

if (nChannels > 1 and nSlices > 1) or (nChannels > 1 and nFrames > 1)
    or (nSlices > 1 and nFrames > 1):
    print(imp.getTitle() + " is a hyperstack (multi c, z or t), please
          prepare a stack (single c, either z-stack or t-stack) from it.")
    pass
elif nChannels > 1 or nSlices > 1 or nFrames > 1:

```

```

    for stack in range(1, imp.getImageStackSize() + 1):
        imp.setPosition(stack)
        R2L(imp)
    else:
        R2L(imp)

```

Source Code 4.10 essentially performs the opposite operation to that performed by Source Code 4.9. It can use a list of ROIs to generate a new image containing all of the uniquely labeled points and determines the bit-depth of the image based on the number of unique points required. This can be useful in the process of translating multiple labeled images onto one another as well as in integrating labels from multiple processing modalities.

Source Code 4.11: A Python translation of the FIJI function “ROIs to Measurement Image”. This combines the a defined set of ROIs (probably from labelsToROIs.py) and a raw image and generates an image that graphically represents measurements such as area or circularity.

```

from ij import IJ, ImagePlus
from ij.gui import Roi
from ij.plugin import Duplicator
from ij.plugin.frame import RoiManager
from ij.process import ImageProcessor, ImageStatistics
import math, os

def ROIsToMap(imp_path, rois, column_name):

    rm = RoiManager.getRoiManager()
    rm.reset()
    rm.runCommand("Open", rois)

    label = "Measure"
    choices = ["area", "angle", "anglevert", "ar", "circumference",
    ↵ "major", "minor", "mean",
    "median", "mode", "min", "max", "perimeter", "xcenterofmass",
    ↵ "ycenterofmass"]

    if str(column_name).lower() not in choices:
        print("Invalid column_name, try again")

```

```

    exit()

def R2M(im):
    imp2 = im.duplicate()
    saveloc = str(im.getOriginalFileInfo().directory)
    title = im.getTitle()

    if im.getBitDepth() == 32:
        IJ.run(imp2, "16-bit", "")

    stackN = imp2.getImageStackSize()
    isStack = bool()

    if stackN > 1:
        isStack = True

    for stack in range(1, stackN + 1):
        imp2.getStack().getProcessor(stack).setValue(0)
        imp2.getStack().getProcessor(stack).fill()

    IJ.run(imp2, "32-bit", "")

    rc = rm.getRoisAsArray()

    filling_value = 0

    for i in range(0, len(rc)):
        if isStack:
            imp.setPosition(rc[i].getPosition())
            imp2.setPosition(rc[i].getPosition())
            ip = imp.getProcessor()
            ip.setRoi(rc[i])

            ip2 = imp2.getProcessor()
            ip2.setRoi(rc[i])

            ip_stats = ip.getStatistics()

            if column_name.lower() == "area":
                filling_value = ip_stats.area
            elif column_name.lower() == "angle":
                filling_value = ip_stats.angle
            elif column_name.lower() == "anglevert":
                filling_value = ip_stats.angle - 90

```

```

    elif column_name.lower() == "ar":
        filling_value = ip_stats.major / ip_stats.minor
    elif column_name.lower() == "circumference":
        filling_value = 4 * math.pi * ip_stats.area /
            math.pow(rx[i].getLength(), 2)
    elif column_name.lower() == "major":
        filling_value = ip_stats.major
    elif column_name.lower() == "minor":
        filling_value = ip_stats.minor
    elif column_name.lower() == "mean":
        filling_value = ip_stats.mean
    elif column_name.lower() == "median":
        filling_value = ip_stats.median
    elif column_name.lower() == "mode":
        filling_value = ip_stats.mode
    elif column_name.lower() == "min":
        filling_value = ip_stats.min
    elif column_name.lower() == "max":
        filling_value = ip_stats.max
    elif column_name.lower() == "perimeter":
        filling_value = rc[i].getLength()
#
#        elif column_name.lower() == "pattern":
#            roi_name = rc[i].getName()
#            r = Pattern.compile(pattern)
#            m = r.matcher(roi_name)
#            if m.find():
#                group = m.group(1)
#                try:
#                    filling_value = Float.parseFloat(group)
#
elif column_name.lower() == "xcenterofmass":
    filling_value = ip_stats.xCenterOfMass
elif column_name.lower() == "ycenterofmass":
    filling_value = ip_stats.yCenterOfMass

ip2.setValue(filling_value)
ip2.fill(rc[i])
imp2.setProcessor(ip2)

imp2.setTitle(column_name.lower() + "_" + title)
imp2.show()
imp2.setRoi(0, 0, imp2.getWidth(), imp2.getHeight())
IJ.saveAs(imp2, "Tiff", os.path.join(saveloc, imp2.getTitle()))

R2M(imp)

```

The last function in the original LaRoMe transforms a set of ROIs into an image that graphically represents various qualities of the source image. This allows the user to transform a more basic segmented image into ROIs and then use those to measure various aspects of the image from which the ROIs were sourced; this recombines the data from labeled images with the primary data from which it was sourced to generate useful measurements, including the area of fluorescence within the ROI, the relative circularity, the minimum or maximum fluorescence intensity, and more. This utility warrants further exploration going forward in attempting to quantify major aspects of tissue culture immunofluorescence imaging; a more optimized pipeline could likely extract more useful data from the images I already have of various correspondences between NFAT nuclear localization and VEGFA production.

The rewrite allows all of these commands to be strung together to process arbitrary numbers of images all at once and, with intelligent file naming, it should be possible to go from CellProfiler-derived label images to meaningful quantitation of various aspects of the raw images across entire directories of image sets. “ROIs to Measurement Image” should probably have an optional GUI similar to the original wrapped around its commands, but I would refer those interested to simply use the original LaRoMe as this implementation and the original are at feature parity.

4.8 Experimental Blinding via a Single-Click Command Line Interface⁴

⁴Implementation from W. J. Brewer et al. (2022b). “Macrophage NFATC2 Mediates Angiogenic Signaling During Mycobacterial Infection”. In: *Cell Rep* 41.111817, original conception from Salter, 2016.

Source Code 4.12: A script to conduct computational filename blinding from the command line written in Python.

```
#!/usr/bin/python3

import os, csv, string, random, sys, argparse
from os import path

def blindrename(folder, r = False):

    # Root = bad for so many reasons.
    if os.geteuid() == 0:
        exit(f"For safety reasons, you must NOT be root when running
            ↳ renamer.py. Please become a non-root user, make sure that user
            ↳ has permissions to write to all files in {folder}, and try
            ↳ again.")

    # Needs a folder, if none is provided, exit.
    if not folder:
        exit("You must supply a folder name. I will randomly rename all files
            ↳ in that folder, and create a key file in .CSV format in that
            ↳ folder.")

    # The folder needs to already exist.
    if not path.isdir(folder):
        exit(f"{folder} is not a valid, existing folder. Try again - typo
            ↳ maybe?")

    # And it presumably needs to have some number of non-hidden files in
    # it. Seems harmless if it is empty, but not expected behavior.

    contents = os.listdir(folder)
    if len(contents) == 0:
        exit("Looks like that folder is empty!")

    # Avoid double renaming.
    if path.exists(os.path.join(folder, "keyfile.csv")):
        exit("Keyfile already exists. Have you already randomized that
            ↳ folder?")

    csvfile = open(path.join(folder, "keyfile.csv"), "w")
    writer = csv.writer(csvfile)
    writer.writerow(["original", "file.path"])

    chars = str(string.ascii_letters + string.digits)
```

```

print("Renaming: ")

for dirname, subdirs, files in os.walk(folder):
    subdirs[:] = [d for d in subdirs if not d.startswith(".")]
    files = [f for f in files if not f.startswith(".")]
    if r:
        old_names = [path.join(dirname, f) for f in files]
    else:
        old_names = [path.join(folder, f) for f in files if dirname is
                     folder]
    for old_name in old_names:
        if path.isfile(old_name):
            if path.basename(old_name) == "keyfile.csv":
                pass
            else:
                base = path.basename(old_name).rsplit(os.extsep, 1)
                # This ensures that there is a 1 index for files with no
                # extension
                base.append("")
                cloaked_name = "".join(random.choices(chars, k = 5))
                if base[1]:
                    new_name = path.join(dirname, cloaked_name + "." + base[1])
                if not base[1]:
                    new_name = path.join(dirname, cloaked_name)
                print(old_name, " to ", new_name)
                writer.writerow([old_name, new_name])
                os.rename(old_name, new_name)

print("Finished!")

def unblind(folder):

    if os.geteuid() == 0:
        exit(f"For safety reasons, you must NOT be root when running
             renamer.py. Please become a non-root user, make sure that user
             has permissions to write to all files in {folder}, and try
             again.")

    # Needs a folder, if none is provided, die
    if not folder:
        exit("You must supply a folder name. I will randomly rename all files
             in that folder, and create a key file in .CSV format in that
             folder.")

    # The folder needs to already exist.

```

```

if not path.isdir(folder):
    exit(f"{folder} is not a valid, existing folder. Try again - typo
        ↵ maybe?")

# And it presumably needs to have some number of non-hidden files in
# it. Seems harmless if it is empty, but not expected behavior.

contents = os.listdir(folder)
if ".DS_Store" in contents:
    contents.remove(".DS_Store")
if len(contents) == 0:
    exit("Looks like that folder is empty!")

# Needs a keyfile to function.
if not path.exists(os.path.join(folder, "keyfile.csv")):
    exit("No keyfile?")

csvfile = open(path.join(folder, "keyfile.csv"), "r")
reader = csv.reader(csvfile)

print("Renaming: ")

for file in reader:
    masked = file[1]
    original = file[0]
    if path.exists(masked):
        print(masked + " to " + original)
        os.rename(masked, original)

print("Finished!")

if __name__ == '__main__':
    parser = argparse.ArgumentParser(description='Blind rename/unblind a
        ↵ folder of files')
    parser.add_argument("function", nargs = 1, action = "store", help =
        ↵ "Which command would you like to run? [blindrename] or [unblind]")
    parser.add_argument("folder", nargs = 1, action = "store", help =
        ↵ "Provide the folder you wish to rename. Drag and drop is usually
        ↵ fine.")
    parser.add_argument("--r", default = False, required = False, action =
        ↵ "store_true", help = "Do you want to rename files in the
        ↵ subdirectories as well? If so, provide [--r]")

args = parser.parse_args()
print(args)

```

```
if "blindrename" in args.function:  
    blindrename(args.folder[0], args.r)  
if "unblind" in args.function:  
    unblind(args.folder[0])
```

In the process of data collection (especially image acquisition), the experimenter will typically assign each file a logical name indicative of what it contains. However, these file names are also a critical breakdown in analytical blinding and ways to avoid this are essential to prevent the introduction of excess experimenter bias into the process of analysis. This problem is clearly widespread, but solutions are difficult to come by. Individuals can have labmates or others to rename folders or files to obfuscate their contents, but this has the danger of the human element – it would not be that difficult to lose track of any alterations and make subsequent quantitation worthless. Thus, a computationally robust method is required.

Salter (2016) saw this issue and elegantly addressed it through a Perl script (Wall, Christiansen, and Orwant, 2000). The author generated a script that would allow you to provide a single argument – a folder – and have the contents renamed and output a keyfile. This implementation was excellent and used throughout my work, but as time passed, I found it lacking in two major areas: recursion and the ability to undo the renaming. I thus generated a Python implementation that was able to (optionally) recursively move through subdirectories, would provide the exact file path as the original name of the file (useful for feeding into R or FIJI), and came with the ability to provide a folder and keyfile and name all the files back to the original name. In my opinion, these features make for a more complete blinding solution and will hopefully be adopted more widely by users looking to reduce their experimental bias.

This script can be used by first making it executable:

```
chmod +x ./renamer.py
```

And then calling it by:

```
./renamer.py [function] [file directory to rename] [--r]
```

Where the “--r” flag is either left absent (for no recursion) or is provided “--r” for folder recursion through all non-hidden subdirectories. False is the default for safety. This is a reasonably user-friendly option via the command line is should make experimental blinding much easier. The function is either “blindrename” or “unblind” and the directory can be provided as a naked string (that is, no “” required).

Being written in Python, a GUI could probably be created that would allow this to be even more friendly to the technologically naïve user. This would be a relatively modest undertaking and is probably worth considering in the future; this would also allow for more effective cross-platform distribution as compiled binaries.

4.9 User-Friendly Analysis of RNA Sequencing

Data using Kallisto/Sleuth in a Python Environment⁵

While an old technology today, the analysis of RNA sequencing data still unfairly remains a challenge for the technologically naïve researcher. To ameliorate part of this problem, in the course of the work in Saelens et al. (2022), I developed a set of pipelines for the analysis of RNA-seq data using a combination of Kallisto and Sleuth, a pair of analysis and visualization applications that utilize pseudoalignment to calculate read counts and then display them in a Shiny application via R (Bray

⁵Taken and expanded from personal contributions to J. W. Saelens et al. (2022). “An ancestral mycobacterial effector promotes dissemination of infection”. In: *Cell* 185.24, P4507–4525.E18.

et al., 2016; Pimentel et al., 2017).

Conducting this portion of the work required acquainting myself with a number of commonly used computational tools, including cmake and a deeper knowledge of Python and how that can translate into generating a broadly useful cross-platform tool to analyze complex sequencing data. Doing so also required interfacing with FTP and other networking functions and navigating server directories to fetch reference cDNA from Ensembl.

While these were originally implemented as two parallel scripts, one for bacteria and the other for eukaryotes, they have since been consolidated into a single all-purpose script that requests different input based on what is available. The issue with bacterial analysis is that Ensembl has discontinued generating bioMarts for bacterial genomes and has adopted an unpredictable folder structure for fetching reference transcriptome files via FTP. Thus, the user will have to provide the reference transcriptome of their bacterial strain of choice, which can generally be acquired from species-specific databases (Mycobrowser being the notable one here) or from the set of available strains on NCBI or Ensembl. Nonetheless, this should offer a guided experience in the command line to the analysis of RNA sequencing data and provide output that can then be analyzed using either DESeq2 or Sleuth (Love, Huber, and Anders, 2014; Pimentel et al., 2017).

Source Code 4.13: A guided command line application for the analysis of bulk RNA-seq data using Kallisto.

```
#!/usr/bin/env python3

# macOS or Linux/WSL ONLY

# People need to have installed Anaconda for this to work. Presumably
# that can be done easily enough. Alt: execute another script first.
```

```

import re, os, ftplib, subprocess, glob, sys, shutil, platform

# This gives the script some self awareness. It finds itself and changes
# the working directory to that path (temporarily).
# This is important for executing the brew_installer.sh script.
script_path = os.path.dirname(os.path.realpath(__file__))
os.chdir(script_path)

# Welcome to the era of Apple Silicon.

if "macOS" in platform.platform():
    # This will check to see if several important system programs are
    # installed in sequential order.
    # If they are not, then it executes a script to install them, may
    # require user password.
    if not shutil.which('xcode-select'):
        subprocess.run(['./brew_installer.sh'])
    if not shutil.which('brew'):
        subprocess.run(['./brew_installer.sh'])
    else:
        subprocess.run(["brew", "install", "hdf5"])
    if "arm64" in platform.platform():
        kallisto = "./kallisto_as"
    elif "x86_64" in platform.platform():
        kallisto = "./kallisto_intel"
    elif "Linux" in platform.platform():
        kallisto = "./kallisto_linux"
        subprocess.run(["sudo", "apt-get", "install", "libhdf5-dev"])

from termcolor import colored, cprint
import Bio

fastq_dir = input("Enter the directory of your FASTQ files (drag and drop
# is fine): ")
fastq_dir = fastq_dir.strip()
try:
    os.chdir(fastq_dir)
except FileNotFoundError:
    text = colored("Looks like that directory does not exist - restart the
# script and try dragging and dropping the folder directly into
# Terminal.", "red")
    print(text)
    sys.exit(1)
except PermissionError:
    print(text)

```

```

    sys.exit(1)

phylum = input(colored('Is your organism an [animal], [plant], [fungus],\n'
    '↳ [protist], or [metazoan]? Input one of the options within the\n'
    '↳ brackets: ', 'cyan'))
phylum = phylum.lower()
organism_name = input(colored("Input the 'Genus species' for your\n"
    '↳ reference organism: ', 'magenta'))
org_split = organism_name.lower().split()
org_dir = "_" .join(org_split)

# This is where the user-friendliness comes in: use Ensembl's highly
↳ regular patterning (for eukaryotes) to fetch the needed file for
↳ supported organisms.

# Vertebrates/Model Animals:
if 'ani' in phylum:
    try:
        with ftplib.FTP('ftp.ensembl.org') as ftp:
            ftp.login('anonymous')
            ftp.cwd('/pub/current_fasta/{}/cdna/'.format(org_dir))
            for filename in ftp.nlst(pattern):
                fhandle = open(filename, 'wb')
                ftp.retrbinary('RETR ' + filename, fhandle.write)
                fhandle.close()
    except ftplib.error_perm:
        text = colored("It appears that your organism is not supported. Try\n"
            '↳ running again with a closely related species or check\n'
            '↳ spelling.", 'red')
        print(text)
        sys.exit(1)

# Fungi:
if 'fu' in phylum:
    try:
        with ftplib.FTP('ftp.ensemblgenomes.org') as ftp:
            ftp.login('anonymous')
            ftp.cwd('/pub/current/fungi/fasta/{}/cdna/'.format(org_dir))
            for filename in ftp.nlst(pattern):
                fhandle = open(filename, 'wb')
                ftp.retrbinary('RETR ' + filename, fhandle.write)
                fhandle.close()
    except ftplib.error_perm:

```

```

text = colored("It appears that your organism is not supported. Try
    ↵ running again with a closely related species or check
    ↵ spelling.", 'red')
print(text)
sys.exit(1)

# Plants:
if 'pl' in phylum:
    try:
        with ftplib.FTP('ftp.ensemblgenomes.org') as ftp:
            ftp.login('anonymous')
            ftp.cwd('/pub/current/plants/fasta/{}/cdna/'.format(org_dir))
            for filename in ftp.nlst(pattern):
                fhandle = open(filename, 'wb')
                ftp.retrbinary('RETR ' + filename, fhandle.write)
                fhandle.close()
    except ftplib.error_perm:
        text = colored("It appears that your organism is not supported. Try
            ↵ running again with a closely related species or check
            ↵ spelling.", 'red')
        print(text)
        sys.exit(1)

# Protists:
if 'pr' in phylum:
    try:
        with ftplib.FTP('ftp.ensemblgenomes.org') as ftp:
            ftp.login('anonymous')
            ftp.cwd('/pub/current/protists/fasta/{}/cdna/'.format(org_dir))
            for filename in ftp.nlst(pattern):
                fhandle = open(filename, 'wb')
                ftp.retrbinary('RETR ' + filename, fhandle.write)
                fhandle.close()
    except ftplib.error_perm:
        text = colored("It appears that your organism is not supported. Try
            ↵ running again with a closely related species or check
            ↵ spelling.", 'red')
        print(text)
        sys.exit(1)

# Metazoans:
if 'met' in phylum:
    try:
        with ftplib.FTP('ftp.ensemblgenomes.org') as ftp:
            ftp.login('anonymous')

```

```

        ftp.cwd('/pub/current/metazoa/fasta/{}/cdna/'.format(org_dir))
        for filename in ftp.nlst(pattern):
            fhandle = open(filename, 'wb')
            ftp.retrbinary('RETR ' + filename, fhandle.write)
            fhandle.close()
    except ftplib.error_perm:
        text = colored("It appears that your organism is not supported. Try
        ↵ running again with a closely related species or check
        ↵ spelling.", 'red')
        print(text)
        sys.exit(1)

# Error Handling:
valid_responses = ('ani', 'fu', 'pl')
if not any(s in phylum for s in valid_responses):
    text = colored("Looks like you've entered an improper phylum. Try
    ↵ again!", "red")
    print(text)
    sys.exit(1)

# Looks like everything works up to here now.

# Define some potentially useful variables that I can plug into
# subprocess.
kallisto = './kallisto'
index = 'index'
quant = 'quant'

# Fetch the index file (name independent, it just needs to end in the
# pattern defined above)
# All Ensembl cDNA files should have that precise formatting.

pattern = '*.cdna.all.fa.gz'

ref_cdna = glob.glob(pattern)
ref_cdna = ref_cdna[0]

exts = ('*.fastq.gz', '*.fq.gz', "*.fastq", "*fq")

threads = int(os.cpu_count())

# This checks whether you already have an index and, if so, whether you
# want to build a new one.
index_checker = os.path.isfile('kallisto_index')

```

```

if index_checker:
    indexed = input(colored("Would you like to build a fresh transcriptome
    ↵ index? This is optional [Y/N] ", 'green'))
    if indexed == "Y":
        if "bac" in phylum:
            ref_cdna = input(colored("Provide a path to the reference cDNA for
            ↵ your organism: ", "green")).rstrip()
            subprocess.call([kallisto, index, '-i', 'kallisto_index', ref_cdna])
    else:
        pass
else:
    if "bac" in phylum:
        ref_cdna = input(colored("Provide a path to the reference cDNA for
        ↵ your organism: ", "green")).rstrip()
        subprocess.call([kallisto, index, '-i', 'kallisto_index', ref_cdna])

while True:
    valid = ('pe', 'se')
    read_type = input(colored("Are your reads paired-end [PE] or
    ↵ single-end [SE]? Paired-end reads have two files for each
    ↵ condition - one with '_1' and one with '_2' [PE/SE] ", "yellow"))
    if read_type.lower() not in valid:
        text = colored("Looks like you had a typo in the last prompt!",
        ↵ "red")
        print(text)
        continue
    else:
        break

# This script runs for PE reads - it can estimate fragment length from
    ↵ this and doesn't need you to provide the information.

cnt = 0

if 'PE' in read_type.upper():
    fastqs = glob.glob("./*_1.f*q*")
    try:
        for forward in fastqs:
            reverse = forward.replace("_1.f", "_2.f")
            matcher = re.search("_1.f.*q*", forward).group(0)
            dir = forward.replace(matcher, "")
            subprocess.run([kallisto, quant,
            '-i', 'kallisto_index',
            '-o', dir + '_quant',

```

```

        '--bias',
        '-b', '200',
        '-t', threads,
        forward, reverse])
    cnt += 1
    print(cnt)
except:
    text = colored("Looks like something went wrong.", "red")
    print(text)
    sys.exit(1)
else:
    pass

# This needs a little bit more user-engagement - average fragment length
↳ and the SD are required, but can be substituted by guesses if it is
↳ not known (it rarely is).

cnt = 0

if 'SE' in read_type.upper():
    fastqs = glob.glob("./.*.f*q*")
    frag_len = 150
    standard_dev = 10
    frags = input("If known, input estimated average fragment length (from
      ↳ FastQC). If not known, hit enter: ")
    if not frags:
        frags = frag_len
    sd = input("If known, input the standard deviation of the fragment
      ↳ length (from FastQC). If not known, hit enter: ")
    if not sd:
        sd = standard_dev
try:
    for read in fastqs:
        matcher = re.search(".f.*q*", read).group(0)
        dir = read.replace(matcher, "")
        subprocess.run([kallisto, quant,
                      '-i', 'kallisto_index',
                      '-o', dir + '_quant',
                      '--bias',
                      '-b', '200',
                      '-t', threads,
                      '--single',
                      '-l', frags,
                      '-s', sd,
                      read])

```

```

        cnt += 1
        print(cnt)
    except:
        text = colored("Looks like something went wrong.", "red")
        print(text)
        sys.exit(1)
    else:
        pass

input(colored("All finished! You are ready to proceed with further
← analysis and visualization in RStudio.", 'green', 'on_white'))

```

This script automates many of the most confusing steps for end users by providing an FTP backend to, whenever possible, fetch the needed files aside from the .FASTQ files from the sequencer. Additionally, the use of Kallisto over more traditional alignment methods like STAR or Bowtie allows this to be trivially run on any modern computer without requiring the user to interface with a computing cluster, which is a relatively steep learning curve for many. Kallisto has other advantages as it is much faster and also more accurate than other alignment methods thanks to its pseudoalignment strategy versus more traditional, computationally intensive find-and-match approaches. Kallisto is also robust to unprocessed .FASTQ files, allowing the user to, in many cases, skip adapter trimming and other preprocessing steps. While there is still a place for STAR and HISAT alignments for *de novo* transcript discovery, in most instances, Kallisto will suffice for the task.

Source Code 4.14: Pipeline for the visualization of Kallisto-aligned RNA seq data using Sleuth. This version supports both eukaryotes and bacteria, albeit through two distinct methods of gathering gene lists.

```

# Sleuth Analysis of Kallisto Output
# Created: 29 September 2017
# Last Edited: 5 August 2022
# Jared Brewer

# You have to provide exactly two things:

```

```

# The directory where you did everything and set the working directory
# to that location using setwd()
# A sample table formatted with two columns: sample and condition and
# named sample_table.txt
# After running, you'll get an interactive window where you can play with
# your data and look for effects.
# There is merit in breaking out comparisons if you have a bunch of
# conditions...

setwd("~/esxM")

if (!require("BiocManager", quietly = TRUE))
  install.packages("BiocManager")
BiocManager::install(version = "3.15")

BiocManager::install(c("rhdf5", "biomaRt"))

install.packages("devtools")
devtools::install_github("pachterlab/sleuth")

library(sleuth)

library(tidyverse)
library(biomaRt)
library(data.table)
library(pheatmap)
library(tidyr)

# This analysis was optimized for animals - biomaRt access to bacterial
# genomes has been discontinued and is a mess.
mart <- read_tsv("./gene_list.txt")
t2g <- as.data.frame(mart>Name)
names(t2g)[1] <- "target_id"
t2g.2 <- t2g[!duplicated(t2g$target_id),]
t2g.2 <- as.data.frame(t2g.2)
names(t2g.2)[1] <- "target_id"
t2g.2$gene_id <- t2g.2$target_id

# Eukaryotes only:
mart <- useMart(biomart = "ENSEMBL_MART_ENSEMBL", dataset =
  "hsapiens_gene_ensembl")
t2g <- getBM(attributes = c("ensembl_transcript_id_version",
  "ensembl_gene_id", "external_gene_name"), mart = mart)
t2g <- rename(t2g, target_id = ensembl_transcript_id_version, ens_gene =
  ensembl_gene_id, ext_gene = external_gene_name)

```

```

# 

files.kal <- list.files(pattern = "./*_quant")
files.dirs <- sapply(files.kal, function(id) file.path(".", id))
sample.table <- read.table("./sample_table.txt", header = T)
sample.table <- dplyr::mutate(sample.table, path = files.dirs)

so <- sleuth_prep(sample.table,
                   full_model = ~condition,
                   target_mapping = t2g.2,
                   extra_bootstrap_summary = T,
                   aggregation_column = "target_id")
so <- sleuth_fit(so, ~condition, "full")
so <- sleuth_fit(so, ~1, 'reduced')
so <- sleuth_lrt(so, 'reduced', 'full')
# so <- sleuth_wt(so, which_beta = 'condition', which_model = 'full')

sleuth_table <- sleuth_results(so, 'reduced:full', 'lrt', show_all =
  FALSE)
sleuth_significant <- dplyr::filter(sleuth_table, pval <= 0.05)

sleuth_live(so)

de <- subset(sleuth_table, pval < 0.05)
paths <- enrichPathway(gene = trimws(de$entrez), pvalueCutoff = 0.2,
  readable = T, organism = "Mycobacterium_tuberculosis_h37rv")

tpm.mat <- sleuth_to_matrix(so, which_df = "obs_raw", which_units =
  "tpm")
tpm <- as.data.frame(tpm.mat)

# Read straight from Sleuth matrix:
setDT(tpm, keep.rownames = "genes")
esx <- tpm |> filter(endsWith(genes, "_5")) |> column_to_rownames(var =
  "genes")
esx <- as.matrix(esx)
write.csv(x = tpm.mat, file = "esx_5_tpm.csv")
pheatmap(esx, show_colnames = F, annotation_legend = T, filename =
  "rnaseq_heatmap_070622.png", width = 6, height = 2)

# Read from saved .csv file (this is the method used for the actual
figure):
esx <- read.csv("./esx_5_tpm.csv", header = T)
row.names(esx) <- esx$target_id
esx <- esx |> filter(endsWith(target_id, "_5"))

```

```
esx <- esx[-c(1:2)]
esx <- as.matrix(esx)
pheatmap(esx, show_colnames = F, annotation_legend = T, filename =
  "rnaseq_heatmap_070622.png", width = 6, height = 2)
```

Due to policy changes by Ensembl, retrieving pre-packaged marts for bacterial genomes is now impossible on the grounds that the number of bacterial strains has outgrown Ensembl's ability to provide these marts. Despite the difficulties this creates for finding reference gene lists for even the most common bacterial species (including even *Escherichia coli*), this can be subverted by converting gene lists available from species-specific repositories into a stripped-down mart-like object that can then be utilized for the downstream steps. No user wants to look at a list of Ensembl IDs after quantitation, and so provision of common gene names is an important component of any analysis. This analysis pipeline was generated specifically for *Mycobacterium tuberculosis*, but could easily be modified to support almost any species.

The logic for fetching and processing bioMarts remains in the script for use with eukaryotic species. As many of the later steps converge, the user can choose to run particular sets of lines based on their organism. Future versions might wrap some GUI elements or starting booleans to only execute the appropriate commands when needed, but in my experience, it is possible to get a new user working with this script with minimal overall effort.

4.10 Bacterial Burden Analysis by Fluorescence Intensity in a Semi-Automated Manner with a User-Friendly Graphical Interface

One of the major routine tasks in the field of zebrafish-*M. marinum* host-pathogen interactions is the quantitation of the total bacterial burden per larva. While it has been well established that the integrated fluorescence intensity of the image corresponds well to the colony forming units of bacteria present, the larval zebrafish has particular challenges. Many of these are attributable to autofluorescence from the yolk and pigment cells or the physical background of the imaging surface, but it is important to avoid catching these in the quantitation as these can vary greatly from fish to fish and are difficult to subtract *post hoc*. However, through clever approaches to image pre-processing it is possible to eliminate these sources of misquantitation and streamline analysis to minimize user intervention.

Source Code 4.15: This graphical user interface allows for automatic background subtraction from images of *M. marinum*-infected larval zebrafish and then quantitation of the remaining signal above a manually set threshold that captures as much of the true signal as possible.

```
from ij import IJ, ImagePlus
from ij.measure import ResultsTable, Measurements
from ij.plugin.frame import RoiManager
from ij.process import AutoThresholder
from ij.plugin import ZProjector
from ij.plugin.filter import ParticleAnalyzer
from ij.gui import GenericDialog, NonBlockingGenericDialog
import os, random, csv, sys, re
from os import path
```

```

def burden(directory, chan, min_threshold, ext, screen_threshold = "Otsu
    ↵ dark", proj_save = False, proj_show = False, imp_show = False,
    ↵ fish_channel = None, outline_threshold = "Triangle dark", brightfield
    ↵ = False, subset = 0, man_psize = 100000):

# directory is a string with path to the files you wish to analyze.
# chan is an integer of the channel # with the bacteria.
# min_threshold is an integer of the minimum threshold value (empirically
    ↵ determined by checking a few images)
# ext is the file extension (".czi", ".lsm", ".tif" are some examples) as
    ↵ a string
# screen_threshold is the threshold algorithm to find the yolk as a
    ↵ string. I found that "Otsu dark" works well, but others may be better
    ↵ in other circumstances.
# proj_save is a boolean that determines whether or not to save the MIPs
    ↵ as you go.
# proj_show is a boolean to determine whether to show the MIPs as they
    ↵ are processed. A bit memory intensive, but valuable for sanity
    ↵ checking.
# imp_show is a boolean for whether or not to show the source image (not
    ↵ recommended except with small datasets)
# fish_channel is an integer for the channel containing something we
    ↵ could use as the basis for an outline of the fish.
# outline_threshold is a string with the thresholding algorithm to find
    ↵ the whole fish (you want something generous, but probably not
    ↵ MinError or similar).
# brightfield is a boolean telling whether or not the selected
    ↵ fish_channel is the brightfield channel. This can be useful and very
    ↵ convenient, but requires some additional processing to be useful.
# subset is to run the function over a subset of the total number of
    ↵ images. Provide an integer value less than the total number of images
    ↵ in the directory. If subset is selected, the projections will
    ↵ automatically show.
# man_psize is a manual particle size value for the "Analyze
    ↵ Particles..." function. Default has worked well for me, but can be
    ↵ changed if needed.

filenames = os.walk(str(directory))
bfiles = []

if ext.startswith("."):
    # Rigid binding of extension per se.
    ext = ext.replace(".", "")

```

```

# This logic is a little over-complex for the task, but essentially I
# want to pull out the extension and then use it to match the end of
# the line. Why do it in two steps? It makes the code more flexible
# for ".tif" vs ".TIFF" and could allow future use of the ext in
# downstream applications if needed.
for dirpath, subdir, files in filenames:
    for file in files:
        mext = re.search("\." + ext + "\S*$", file, re.IGNORECASE)
        if mext:
            if file.endswith(mext.group()):
                bfiles.append(path.join(dirpath, file))

# Make sure all our measurements are set properly.
IJ.run("Set Measurements...",
"area mean min limit display redirect=None decimal=3")

rm = RoiManager.getRoiManager()
rt = ResultsTable.getResultsTable()
data = []

if subset > len(bfiles):
    wrong_subset = NonBlockingGenericDialog("Subset Error")
    wrong_subset.addMessage("The subset is greater than the number of
    files - running on whole directory.")
    wrong_subset.showDialog()
    subset = 0
if subset:
    bfiles = bfiles[0:int(subset)]
    proj_show = True
else:
    subset = 0

valid_thresholds = [str(i + " dark") for i in
    AutoThresholder.getMethods()]
valid_thresholds.append("Try all")
valid_thresholds.append("None")

if screen_threshold == "Try all" or outline_threshold == "Try all":
    if screen_threshold == "Try all":
        for f in bfiles:
            imp = IJ.openImage(f)
            proj = ZProjector.run(imp, "max all")
            proj.setC(int(chan))
            IJ.run(proj, "Auto Threshold", "method=[Try all] dark")
    if outline_threshold == "Try all":

```

```

for f in bfiles:
    imp = IJ.openImage(f)
    proj = ZProjector.run(imp, "max all")
    proj.setC(int(fish_channel))
    if brightfield:
        IJ.run(proj, "Find Edges", "slice");
    IJ.run(proj, "Auto Threshold", "method=[Try all] dark")
else:
    for i, f in enumerate(bfiles):
        imp = IJ.openImage(f)
        if imp_show:
            imp.show()
        if imp.getDimensions()[3] > 1:
            proj = ZProjector.run(imp, "max all")
        else:
            proj = imp.duplicate()
        if proj_show:
            proj.show()
        proj.setC(int(chan))
    if screen_threshold not in valid_thresholds:
        screen_threshold = "Otsu dark"

    if screen_threshold != "None":
        IJ.setAutoThreshold(proj, str(screen_threshold))

    IJ.run(proj, "Analyze Particles...",
           "size="+str(man_psize)+"-Infinity clear include add slice")

    if rm.getCount() > 1: # This is the dumbest source of a
                           # downstream error I have ever encountered. rm.getCount is an
                           # attribute and rm.getCount() is a method = two different
                           # outcomes, but no evaluation error.
        rm.runCommand(proj, "Select All")
        rm.runCommand(proj, "Combine")
        rm.runCommand(proj, "Delete")
        rm.runCommand(proj, "Add")

    rm.select(proj, 0)

    IJ.run(proj, "Clear", "slice")
    IJ.run(proj, "Select None", "")
    IJ.run(proj, "Remove Overlay", "")
    rm.reset()

```

```

# Build some logic for using the outline of the fish if provided.
# Brightfield images probably need even more work, but this will
# *probably* work for fluorescent channels.
if fish_channel:
    proj.setC(int(fish_channel))
    if brightfield:
        IJ.run(proj, "Find Edges", "slice");
    if outline_threshold not in valid_thresholds:
        outline_threshold = "Triangle dark"
    IJ.setAutoThreshold(proj, outline_threshold)
    IJ.run(proj, "Analyze Particles...",,
          "size="+str(man_psize)+"-Infinity clear include add slice")
    proj.setC(int(chan))
    ra = rm.getRoisAsArray()
    for r in ra:
        rm.addRoi(r)
    if rm.getCount() > 1:
        rm.runCommand(proj, "Select All")
        rm.runCommand(proj, "Combine")
        rm.runCommand(proj, "Delete")
        rm.runCommand(proj, "Add")

    rm.select(proj, 0)

else:
    IJ.run(proj, "Select All", "")

# We want this to work for any bit depth image, but due to
# limitations of the underlying plugin, may only work for 16-bit
# and lower images. Something weird could happen with 24-bit RGB
# images, but I can't stop people from doing weird things.

if proj.getBitDepth() == 24: # This should only be color images -
# seems a little risky.
    IJ.run(proj, "8-bit", "")
elif proj.getBitDepth() > 16: # The thresholds don't work for
# 32-bit images and FIJI doesn't really understand 10 or 12 bit
# images.
    IJ.run(proj, "16-bit", "")

IJ.setRawThreshold(proj, int(min_threshold),
                   2**int(proj.getBitDepth()) - 1)
IJ.run(proj, "Measure", "")
row = rt.getRowAsString(0).split("\t")

```

```

row.insert(0, path.join(imp.getOriginalFileInfo().directory,
    ↵ imp.getOriginalFileInfo().fileName))
if i == 0:
    headings = rt.getColumnHeadings().split("\t")
    headings.insert(0, "file_path")
data.append(row)

if not imp_show:
    imp.close()
if not proj_show:
    proj.close()

if proj_save:
    out = path.join(directory, proj.getTitle())
    IJ.saveAs(proj, "Tiff", out)

rand = random.randint(1, 100)
if rand > 85:
    IJ.run("Collect Garbage", "")
else:
    pass

# Write out the results to a CSV file for further processing
↳ (probably in R, but also fine for other options.)

with open(path.join(directory, "burden.csv"), "w") as csvfile:
    writer = csv.writer(csvfile)
    writer.writerow(headings)
    for datum in data:
        writer.writerow(datum)

gui = GenericDialog("Finished")
gui.addMessage("All Done! Output is located in " + directory)
gui.showDialog()

def burden_gui():

    values = [str(x + " dark") for x in AutoThreshold.getMethods()]
    values.append("Try all")
    values.append("None")

# Build the GUI elements.
gui = NonBlockingGenericDialog("Bacterial Burden Measurement")
gui.addDirectoryField("Image Folder: ", " ~/Documents")

```

```

gui.addChoice("File Extension: ", [".czi", ".lif", ".lsm", ".tif",
    ↵ "Other"], ".tif")
gui.addStringField("Custom File Extension: ", ".jpeg")
gui.addNumericField("Bacterial Fluorescence Channel #: ", 1)
gui.addNumericField("Minimum Threshold: ", 200)
gui.addMessage("'Try all' requires a subset to be selected - the fewer
    ↵ the better. Default = 5")
gui.addChoice("Background Removal Threshold: ", values, "Otsu dark")
gui.addCheckbox("Save MIPs?", False)
gui.addCheckbox("Show MIPs?", False)
gui.addCheckbox("Show Images?", False)
gui.addCheckbox("Use Fish Outline?", False)
gui.addToSameRow()
gui.addNumericField("Channel # for Fish Outline: ", 2)
gui.addChoice("Fish Threshold: ", values, "Triangle dark")
gui.addCheckbox("Brightfield Image?", False)
gui.addCheckbox("Only do a subset?", False)
gui.addToSameRow()
gui.addNumericField("If yes, how many images to try: ", 0)
gui.addNumericField("Manual Particle Size: ", 100000)
gui.showDialog()
if gui.wasCanceled():
    canceled = GenericDialog("Canceled")
    canceled.showDialog()
    sys.exit(0)

# Parse all of the GUI elements into variables that can be plugged into
    ↵ the burden function.
directory = str(gui.getNextString())
ext = str(gui.getNextChoice())
if ext == "Other":
    ext = str(gui.getNextString()).lower()
else:
    dump = gui.getNextString()

chan = int(gui.getNextNumber())
min_threshold = int(gui.getNextNumber())
screen_threshold = str(gui.getNextChoice())
proj_save = gui.getNextBoolean()
proj_show = gui.getNextBoolean()
imp_show = gui.getNextBoolean()
do_fish = gui.getNextBoolean()
if do_fish:
    fish_channel = int(gui.getNextNumber())
    outline_threshold = str(gui.getNextChoice())

```

```

brightfield = gui.getNextBoolean()
else:
    dump = gui.getNextNumber()
    dump = gui.getNextChoice()
    dump = gui.getNextBoolean()
    fish_channel = None
    outline_threshold = "Triangle dark"
    brightfield = False
subsetter = gui.getNextBoolean()
if subsetter:
    subset = int(gui.getNextNumber())
else:
    dump = gui.getNextNumber() # Another extraordinarily stupid source of
    ↵ error - every entry /must/ be accessed or you're going to
    ↵ frameshift - was feeding 0 into man_psize when subset = False and
    ↵ getting a correspondingly bad result (all points included).
    subset = 0
if screen_threshold == "Try all":
    subsetter = True
    if not subset:
        subset = 5
if outline_threshold == "Try all":
    subsetter = True
    if not subset:
        subset = 5
man_psize = int(gui.getNextNumber())

burden(directory = directory,
        chan = chan,
        min_threshold = min_threshold,
        ext = ext,
        screen_threshold = screen_threshold,
        proj_save = proj_save,
        proj_show = proj_show,
        imp_show = imp_show,
        fish_channel = fish_channel,
        outline_threshold = outline_threshold,
        brightfield = brightfield,
        subset = subset,
        man_psize = man_psize)

burden_gui()

```

This graphical user interface facilitates automatic processing of arbitrary numbers

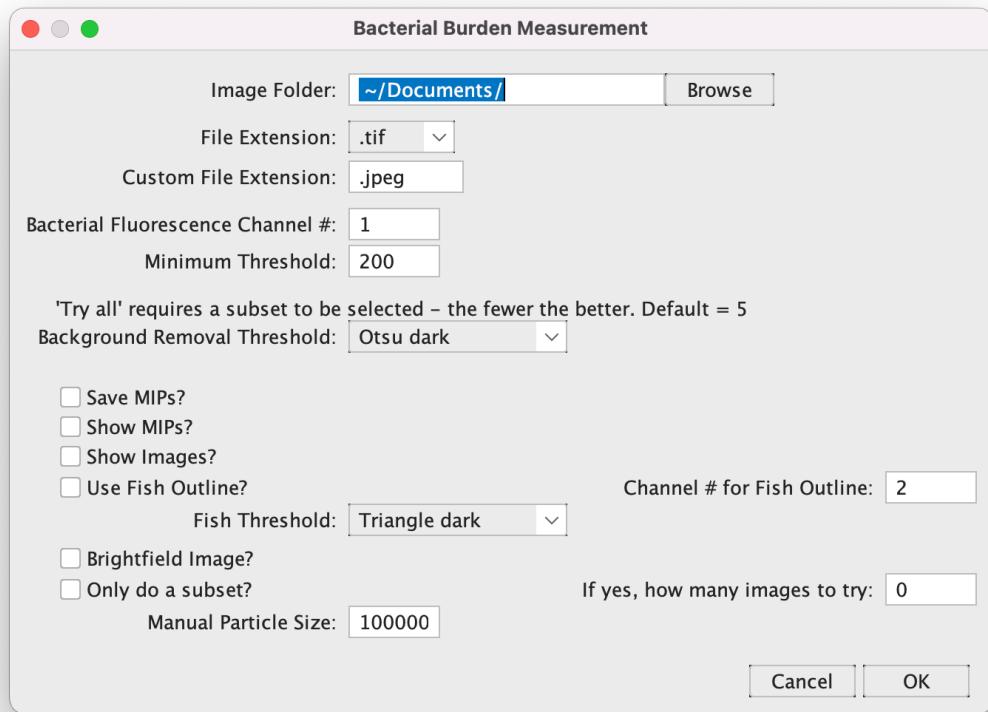


Figure 4.8: This graphical user interface guides the user through providing all of the parameters needed to measure burden from a set of larval zebrafish images. All of these parameters require some tweaking by the user, but allows for subsetting to try different combinations to optimize burden measurements within a single experiment.

of images at once by allowing users to select various parameters to test for appropriateness in their particular experiment (Figure 4.8). The underlying logic will automatically create Z projections if applicable and then use those for subsequent analysis. Users are encouraged to select a subset of images to start and the computational thresholds are used to capture any objects over a certain size for background removal, which will typically only capture the yolk and any background fluorescence. This approach also allows for more generous manual thresholds to be selected for quantitation, an issue that often arises in manual approaches to fluorescence quantitation due to the need to accommodate this autofluorescence.

It is my hope that this application, after further beta testing and refinement can supplant these manual approaches and replace them with something that free researcher time to conduct more experiments rather than spend many hours drawing circles around zebrafish in order to measure the bacterial burden of these fish. Additionally, this could in principle be adapted to measuring other aspects of zebrafish biology, from macrophage clustering to transcriptional reporter signals. The elegance of this approach is that it utilizes open-source and well-defined mechanisms for measuring signal over noise through the implementation of the automatic thresholds and wraps a set of utility functions within an interface that avoids the need for the end user to write burdensome macros to accomplish the same goal. This approach, after a minutes-long period of optimizing is able to measure the burden of thousands of larvae in mere minutes. The output can then be spot-checked for accuracy, as in a small subset of the larvae parts of the background may not be perfectly removed and these can then be reprocessed either manually or with altered parameters.

Chapter 5

Conclusions and Future Directions

This work has accomplished two primary goals: defining the intracellular signaling pathway within macrophages that is responsible for inducing angiogenesis during mycobacterial infection (the NFAT pathway) – a long-standing question in the field of tuberculosis pathogenesis – and setting the stage for future work to simplify and automate common procedures commonly used in the analysis of imaging data relevant to both zebrafish and tissue culture research. This dramatically expands our understanding of the host-pathogen interface in tuberculosis and debunks a widespread assumption, which was that hypoxia signaling is primarily responsible for the angiogenic response surrounding the granuloma. Instead, there is a discrete and specific immunological pathway that drives the expression of VEGFA to induce angiogenesis that seems to be functionally independent of hypoxia *sensu stricto*¹. This sets the stage for the subversion of other assumptive dogmas within the field, including what functional roles angiogenesis contributes to the growth or survival of mycobacteria. At the conclusion of the present work, many new questions have been generated while

¹This argument is supported by the fact that the larval zebrafish is oxygen perfuse, so hypoxia was clearly not the driving trigger in this context. Hypoxia may, however, synergize or otherwise interact with these immunological pathways within more mature granulomas in the adult, a question that will be addressed further in Section 5.2.

others remain unanswered and these offer fertile ground for new discoveries in the field of tuberculosis pathogenesis and host immunity.

Some of these lingering questions will be addressed in the coming weeks and months while others will stretch over the course of many years or decades as we delve into deeper and more sophisticated understandings of the fundamental processes governing the nature of the angiogenic response to and the role of VEGFA signaling in tuberculosis infection and how and when this can be a fruitful target for therapeutic intervention. This leaves a set of important questions (Figure 5.1) pertinent to model development, nuanced understanding of the biology of NFAT within (granuloma) macrophages, the intersections between this pathway and other, known pathways involved in angiogenic responses, and the future of imaging analysis in the context of ever-growing computational power.

- What is the TDM receptor in zebrafish and do they have an as-yet unannotated MINCLE homolog functionally equivalent to the human gene?
- How or why is NFATC2 special and is it sufficient to induce VEGFA or are there other factors also required for VEGFA expression?
- How does the NFAT pathway alter other aspects of macrophage behavior potentially relevant to tuberculosis biology and does this pathway intersect with HIF-1 α signaling? For instance, does this interface with dimensions of immunometabolism potentially mediated by hypoxia or other alterations in macrophage physiology?
- Do mycobacteria have other mechanisms for manipulating host vasculature and, if so, what are they and how can that enhance our overall understanding of the contributions of the vasculature to tuberculosis pathogenesis?

- Are our findings on the nature of NFATC2 in inducing tuberculous angiogenesis relevant to other disease contexts where VEGFA signaling plays an important role?
- Could NFAT offer a meaningful mechanism for inhibiting angiogenesis in the context of disease as a host-directed therapy?

These questions, among many others, are the subject of this concluding chapter and are summarized in Figure 5.1; it is hoped that a comprehensive presentation of these questions will stimulate future generations to pursue answers and that these will further inform our understanding of the pathogenesis of tuberculosis toward the goal of eradicating this contagion.

5.1 The Zebrafish MINCLE

As discussed in Subsection 1.3.5, data from human cell culture and mice has implicated MCL and MINCLE as the primary C-type lectin receptors for TDM, which induces a variety of downstream responses, seemingly including the upregulation of VEGFA and downstream angiogenesis (see Chapter 3). However, the precise identity of the homolog of MCL or MINCLE in the zebrafish remains unknown, making it difficult to precisely model these process or make one-to-one comparisons between humans and the zebrafish². In human, these two proteins arose from a tandem duplication and inversion at an unknown point in evolutionary history, although the two are ubiquitous across reptiles, birds, and mammals (Miyake et al., 2013; Richardson and Williams, 2014). Given the strong, bidirectional selective pressure on both

²There is some irony in the idea that this thesis is built around the concept of C-type lectin signaling but does not directly address the nature of the receptor itself in our model organism of choice. This is an undeniable gap in this work but one that can be patched over using the approaches to be outlined in this section.

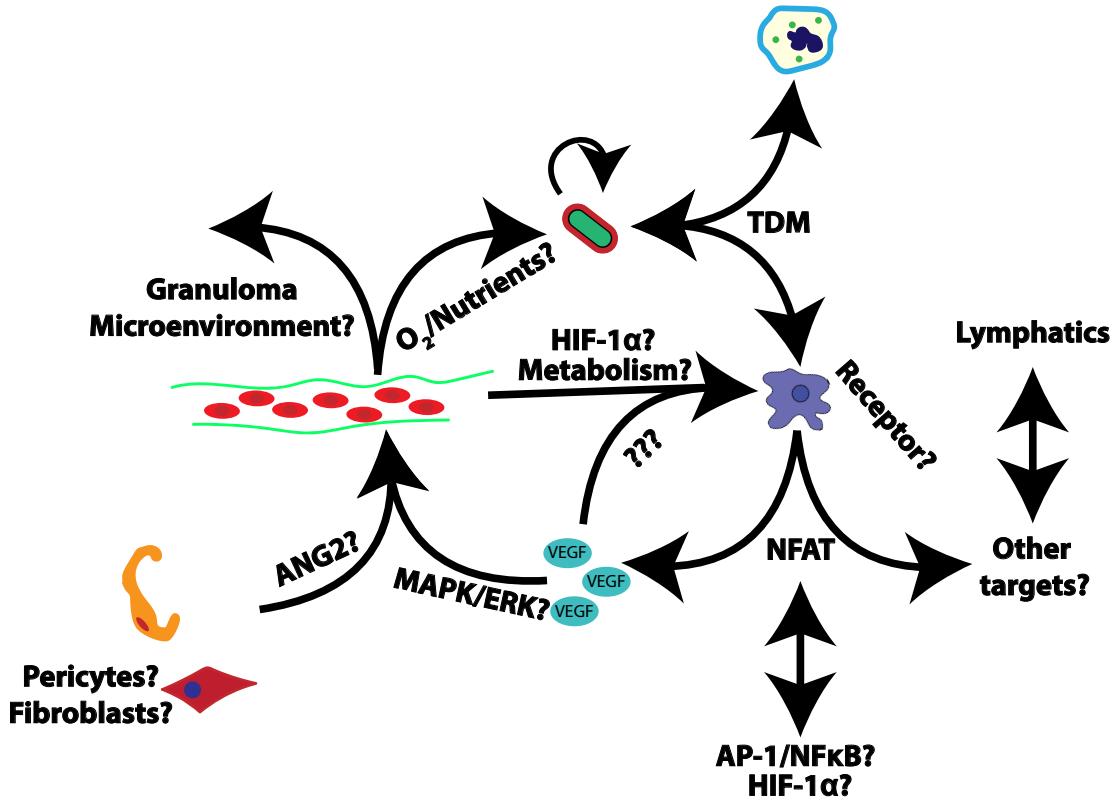


Figure 5.1: A graphical representation of the major findings and questions from this work. While we have now added a critical piece of the equation, there are many questions remaining and many directions for this project to go that effectively interrogate each of the different players, from the bacteria to macrophages to the endothelium and interesting aspects of the interactions between them which will be outlined in the coming sections.

host and pathogen to modulate host PRR activity, these divergences are expected even between closely related species (Rambaruth et al., 2015). This diversification is especially notable among CLRs: mice have no fewer than eight putative DC-SIGN homologs and a great deal of work had to be done to narrow down the functional ones in order to model human disease (Garcia-Vallejo and Kooyk, 2013); on the other hand, the bovine homolog of MINCLE was readily identifiable but had diverged in non-critical domains from the human MINCLE (Furukawa et al., 2013; Feinberg et al., 2013; Feinberg et al., 2016). Indeed, the murine Mincle is only 67% identical to the human MINCLE, despite an overlapping (but not completely identical) set of known ligands (Matsumoto et al., 1999). These aspects of structural diversity add unique complexities to the identification of any putative functional homolog in the fish, which may have substantially diverged from the ancestral protein as well as the mammalian versions. Despite these challenges, such identification would both substantially advance the zebrafish-*M. marinum* model and deepen our understanding of shared mechanisms of detection and response to C-type lectin receptor ligands.

Despite these challenges, there is an abundance of evidence that zebrafish possess an as-yet unidentified TDM receptor including, but not limited to: a long evolutionary history alongside pathogenic mycobacteria, the clear, *myd88*-independent immune response to purified TDM, and the *in vivo* attenuation of mutants lacking fully mature TDM³ (Rao et al., 2005; Walton et al., 2018). Together, these factors suggest that zebrafish have a receptor criteriologically similar to the human MINCLE/MCL that has yet to be identified by traditional sequence homology or synteny methods. Zebrafish have long been known to have functional homologs of other C-type lectin

³The attenuation phenotype could conceptually have been through TDM mediating direct cytotoxicity and inducing a damage response, but this was never as compelling as the notion of a specific immune response pathway given the obviously living, motile macrophages at the TDM interface (Bloch, Sorkin, and Erlenmeyer, 1953; Noll, 1956; Walton et al., 2018).

receptors despite difficulty in their identification (Yoder et al., 2004; Panagos et al., 2006; Petit et al., 2019). MINCLE has not been previously linked to angiogenesis, but the identification of this receptor in zebrafish would allow us to better understand the relevant pathway in humans; indeed, the role of MINCLE during infection is unclear given that some groups have demonstrated that it contributes to bacterial control (Behler et al., 2012; Lee et al., 2012; Behler et al., 2015; Wilson et al., 2015) while others have seen no effect (Heitmann et al., 2013). This has translational implications for modulating the activity of the human MINCLE to enrich for host-beneficial responses and also basic science implications in revealing the diversification of a receptor that maintains the ability to detect a common ligand.

Unlike MINCLE, the basal receptor MCL has clear roles in mediating protection against mycobacteria (Wilson et al., 2015). This might imply that the basal recognition capacities of MCL and potentially later downstream events that depend on MCL/MINCLE interactions are more important than the contributions of MINCLE alone. What is needed is a full locus-deletion mouse that removes both MCL and MINCLE to enable studies on the function of TDM detection *per se in vivo*. While the development of a mouse model to accomplish this would be an excellent contribution, we may be able to fill in some or most of these gaps using the zebrafish if we are able to identify the correct homolog(s) in the zebrafish genome.

Although large amino acid segments of CLRs are able to undergo radical changes in primary sequence with few deleterious effects, there are several domains that have been identified as absolutely essential for binding to TDM. A specific set of criteria for this selection are listed in Table 5.1 (Zelensky and Gready, 2005; Feinberg et al., 2013; Furukawa et al., 2013; Feinberg et al., 2016; Alenton et al., 2017; Bird et al., 2018). Based on these criteria, we have identified three putative homologs with

>50% amino acid similarity to the human CLEC4E in the carbohydrate binding domain (Table 5.1) and have identified transmitting nonsense mutations in each of them (Table 5.2) (Xu and Zhang, 2010). As further evidence, two of these homologs (77975 and 79903) are organized in tandem, mirroring the genomic organization of MINCLE and MCL in mammals.

Table 5.1: Criteria used to select putative zebrafish homologs of the human MINCLE.

Criteria	Rationale
Possesses a gEPNn motif	Of the two major carbohydrate recognition domain motifs, the EPN motif is known to bind glucose-derived sugars while QPD motifs are known to bind galactose-derived sugars. As trehalose is a di-glucose and MINCLE and MCL both possess this EPN motif, this is an important first-pass selection criterion.
Lacks an intracellular ITAM motif	In humans, both MINCLE and MCL use Fc γ R to signal as they lack their own ITAM motif. While not an essential quality to detect and respond to TDM, this would strengthen the similarities between the two; we have also published data implicating Fc γ R in the zebrafish, which argues in favor of this shared layer of similarity as well.
Induced by infection	Using existing RNA-seq datasets, expression of these genes under inflammatory stimulus is an important indicator that they may be acting similarly to MINCLE, which is an inducible gene responsive to various inflammatory stimuli.
Transmembrane helix	These surface receptors use a single-pass transmembrane helix to remain bound to the plasma membrane and transduce signals.
Hydrophobic amino acids in the CRD	One of the defining biochemical features of MINCLE is a small hydrophobic pocket that appears to be useful for binding to the mycolate tails of TDM; the presence of such a pocket would be evocative of further similarity to MINCLE.

Multiple approaches can be taken to identify the capacity of these proteins to bind TDM and generate meaningful biological responses. Going forward, I propose to take a biochemistry-first approach to this question as it enables greater flexibility in responding to new data and starts with a foundation of known interactions. Thus, I will utilize established methods of TDM blotting (Jegouzo et al., 2014) and wash recombinant carbohydrate recognition domain-streptavidin fusion protein lysate across the blots and then detect these interactions using standard biotinylated horseradish peroxidase detection. This will provide a quantifiable readout for both presence/absence of an interaction but also some indication of the strength of the interaction. Should none of these proteins efficiently bind TDM, it is relatively trivial to generate new chimeric proteins and test a range of others present in the zebrafish genome. This data can then be used to go back into the zebrafish to assess the *in vivo* consequences of this interaction and also allow for more flexibility in approach – rather than seeking the receptor, we can explore phenotypes that may be altered in this context across both angiogenic responses and more general immune responses which may defy easy phenotypic characterization given what we know from these previous mouse studies (Behler et al., 2012; Lee et al., 2012; Heitmann et al., 2013; Behler et al., 2015; Wilson et al., 2015).

In the zebrafish itself, I would actually propose to take a few steps backward and return to a mosaic-based screening approach. While the long-term goal should be to generate a full locus-spanning deletion of 77975 and 79903, it may be worthwhile to use the zebrafish as a screening platform to determine the potential contributions of these two genes separately and together in the angiogenesis phenotype and CRISPR/Cas9 ribonucleoprotein injections allow for such rapid screening and planning for whether it makes sense to invest the time and energy in generating a full deletion mutant. Given the unlikelihood of being able to ever generate a crossover

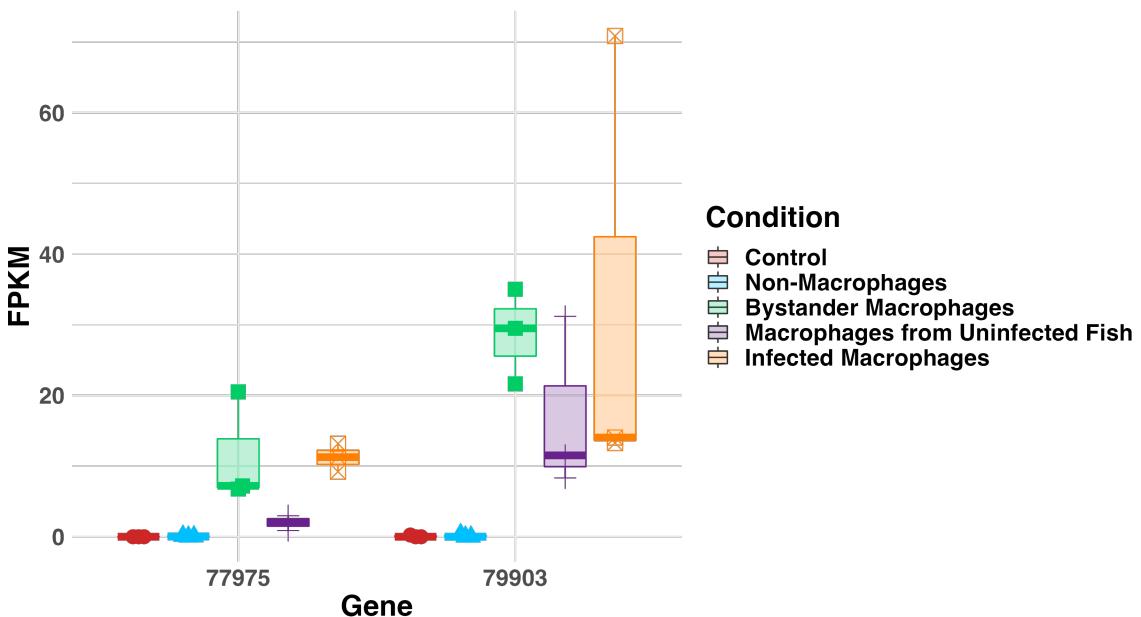


Figure 5.2: Comparison of the expression profiles of two of the putative MINCLE homologs, ENSDARG00000077975 and ENSDARG00000079903 based on existing RNA-seq datasets in the Tobin lab. Both of these genes are generally restricted to macrophages and are induced by the presence of mycobacterial infection, an effect exceptionally notable for 77975. This provides some evidence that these genes are regulated by mycobacterial infection and may be playing some role in this context.

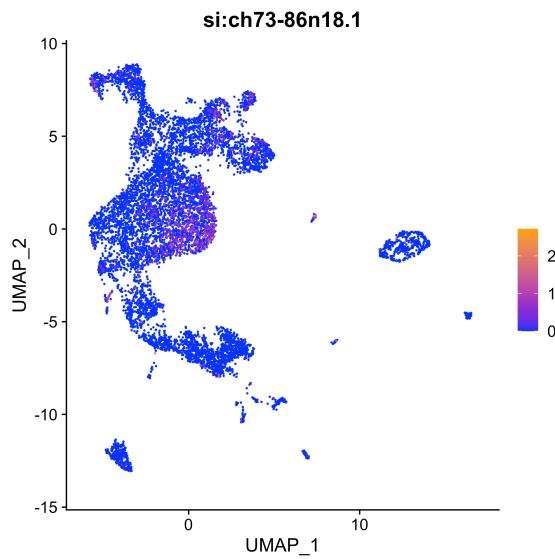


Figure 5.3: Based on the scRNA-seq from Cronan et al. (2021), ENSDARG00000056379 is reasonably expressed within subset of these macrophage populations, which include epithelioid macrophages and those that express *vegfaa*.

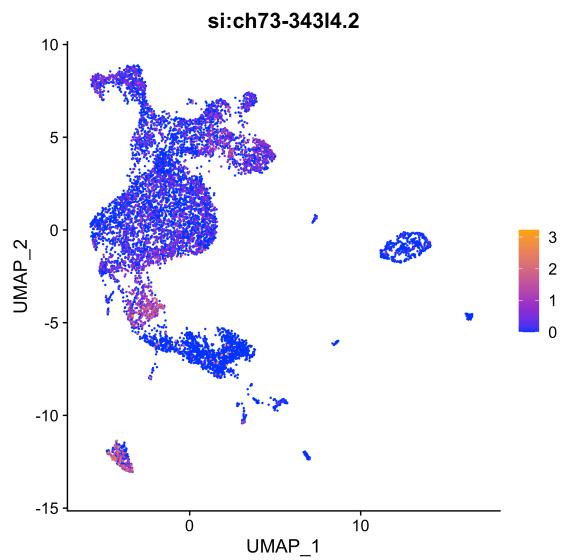


Figure 5.4: Based on the scRNA-seq from Cronan et al. (2021), ENSDARG00000079903 is reasonably expressed within subset of these macrophage populations, which include epithelioid macrophages and those that express *vegfaa*.

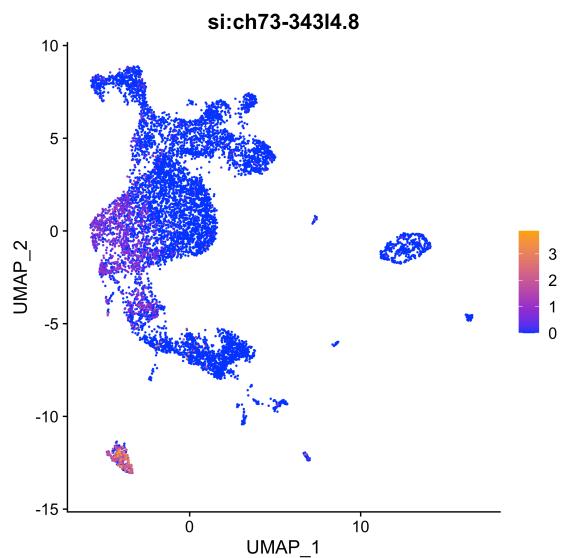


Figure 5.5: Based on the scRNA-seq from Cronan et al. (2021), ENSDARG00000077975 is reasonably expressed within subset of these macrophage populations, which include epithelioid macrophages and those that express *vegfaa*.

Table 5.2: Putative zebrafish MINCLE homologs with key details about their native structure and mutants that have been generated thus far.

Gene ID	56379	79903	77975
Length	263	263	170
CRD	kEPNn	gEPNn	gEPNn
Identity	41%	42%	42%
Similarity	50.7%	53.3%	60.3%
Mutation	::13	::13	Δ8
Site (a.a.)	43	207	108
RMSD	1.34	1.17	0.87
TM-score	0.86	0.85	0.86
Reference Coverage	95%	91%	92%
Target Coverage	91%	96%	100%

event to combine the two existing mutations, some alternative approach is clearly going to be required and this seems like a suitable place to start in pursuit of the zebrafish TDM receptor(s). These two genes make for especially compelling candidates as they are macrophage-specific and are induced by infection, even in bystander macrophages (Figure 5.2). This is further supported by expression data from the single cell RNA-seq experiment done for Cronan et al. (2021), which shows that these proteins are all widely expressed within different clusters of macrophages. Interestingly, they appear to show somewhat of a banding pattern across the spectrum of macrophages, perhaps indicating distinct roles in different subsets. Whether this is a true phenomenon or an artifact will require further investigation, but these appear to be promising candidates both as the TDM receptor and in other aspects of the innate immune response (Figure 5.3, Figure 5.4, Figure 5.5).

Additionally, it may be of some use to study the specific human MINCLE-mycobacteria interactions in the context of a whole immune system. Thus, going forward, it would be logical for future researchers to develop transgenic zebrafish that express

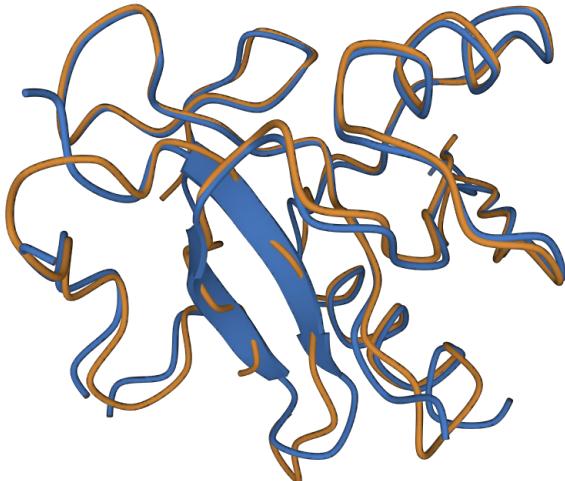


Figure 5.6: Pairwise structural alignment of the carbohydrate recognition domains of the human MINCLE and zebrafish 56379 showing strong structural overlap indicative of a common structure in excess of the amino acid level similarity.

human versions of MINCLE and MCL in macrophages and, perhaps, neutrophils, to assess the contributions of the human protein to conserved responses. This may also help to clarify some of the conflicting data in the literature around the role of MINCLE by using an overexpression model to capture the effects of excess MINCLE signaling. In the long term, gene replacement of the native MINCLE-like homolog with the human MINCLE would allow for a more authentically humanized model of macrophage biology within the zebrafish. Use of this as a complementation strategy if we can identify the zebrafish MINCLE would also serve as a more comprehensive humanized model to study the MINCLE-specific contributions to angiogenesis and bacterial control.

While these proteins were originally chosen based on the criteria listed in Table 5.1, the emergence of AlphaFold has allowed for highly accurate structure prediction for any sequence of amino acids, which has allowed for subsequent structural comparisons

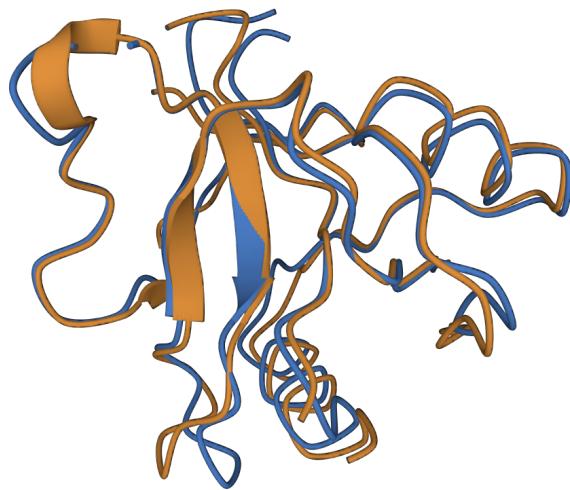


Figure 5.7: Pairwise structural alignment of the carbohydrate recognition domains of the human MINCLE and zebrafish 79903 showing strong structural overlap indicative of a common structure in excess of the amino acid level similarity.

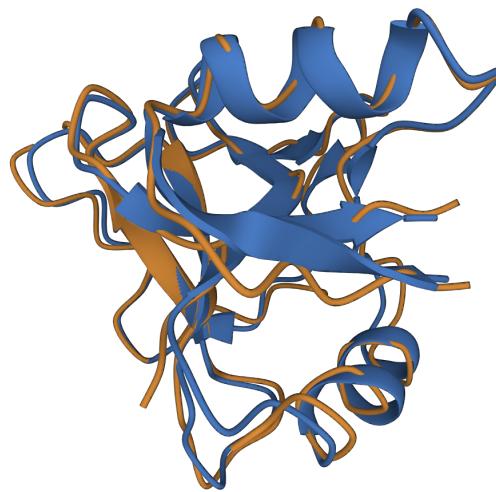


Figure 5.8: Pairwise structural alignment of the carbohydrate recognition domains of the human MINCLE and zebrafish 77975 showing strong structural overlap indicative of a common structure in excess of the amino acid level similarity.

between these genes and the human MINCLE extracellular domain that indicates a strong similarity between them based on TM-score and overall structural alignment (Table 5.2) (Jumper et al., 2021). I have conducted these pairwise alignments for each of the three genes of interest and this analysis indicates very high degrees of structural similarity among them, suggesting that they are strong candidates to bind similar ligands (Figure 5.6, Figure 5.7, Figure 5.8). However, this is disadvantaged by the fact that many carbohydrate recognition domains from diverse C-type lectins share major structural features, so these data must be taken into consideration with the primary sequence information to draw any meaningful conclusions (Table 5.2). Nevertheless, the access to structural information is immensely useful in characterizing some of these candidates and may allow for a better understanding of the features required for binding to TDM.

5.2 Integration of Hypoxia Signaling

The literature is replete with descriptions of HIF-1 α regulation of VEGFA production and signaling; the logical means by which to alleviate local hypoxia is through the recruitment of vasculature carrying oxygenated blood. This allows angiogenesis to occur when necessary and for the vasculature to remain quiescent under homeostatic conditions, where intravital oxygen concentrations are maintained at a high level within blood vessels and in neighboring tissues. However, in areas of pathogen invasion, tumor growth, or tissue damage, the local oxygen concentration can fall, triggering the activation of the HIF-1 α signaling pathway in neighboring cells. HIF-1 α is an oxygen-sensing protein that is expressed and rapidly degraded under normoxic conditions but stabilized under hypoxia. Under standard oxygen concentrations, two classes of regulatory proteins mediate prolylhydroxylation and proteosomal degrada-

tion of HIF-1 α in a process dependent on molecular oxygen (Hong, Lee, and Kim, 2004; Masoud and Li, 2015; Corcoran and O'Neill, 2016).

Alternatively, HIF-1 α can be induced through transcriptional alterations in the homeostatic stoichiometry of HIF-1 α itself and the two families of regulatory proteins, PHD and FIH. This allows HIF-1 α to be induced under normoxic conditions, the presumed whole-body state of the zebrafish larva (Ellertsdottir et al., 2010). This normoxic activation has been found to be important for myeloid immune responses, including those downstream of MINCLE activation, and may play an important role especially in the early signaling events of mycobacterial infection (Nishi et al., 2008; Schoenen et al., 2014; Schatz et al., 2016; Thompson et al., 2017).

In the environment of the granuloma, both the host and pathogen must adapt to reduced oxygen tension (Prosser et al., 2017). Mycobacteria can temporarily revert into a non-replicating state known as persistence, but this is not a viable strategy for long-term evolutionary success (Manabe and Bishai, 2000; Bentrup and Russell, 2001; Stewart, Robertson, and Young, 2003; Pandey and Sassetti, 2008; Ehrt, Schnappinger, and Rhee, 2018). However, during persistence, the bacteria are extremely difficult to kill as most antitubercular drugs are only effective on replicating bacteria⁴ (Veatch and Kaushal, 2018). Even during active growth, the bacteria and associated host cells must alter their metabolism to accommodate for reduced oxygen availability, with important consequences for host immunity (Tsai et al., 2006; Rustad et al., 2009; Harper et al., 2012; Galagan et al., 2013; Prosser et al., 2017). Despite our superficial knowledge about the importance and contributions of hy-

⁴This is true of antibiotics in general – by targeting pathways like cell wall and protein biosynthesis and transcription, these pathways are only absolutely essential for bacteria in the replicative phase. The ability to sporulate, like many *Bacillus* species, or to fall into periods of dormancy can allow for escape from bactericidal antibiotic therapy in the absence of overt resistance, a phenomenon often called tolerance (Higgins and Dworkin, 2012; Meredith et al., 2015; Levin-Reisman et al., 2017; Westblade, Errington, and Dorr, 2020).

poxia in the lifestyle of *M. tuberculosis*, we do not have a clear means to genetically manipulate these responses or to differentiate the roles of hypoxia *per se* from the activity of HIF-1 α signaling. Thus, going forward, new tools are going to be required to study not only the contributions of HIF-1 α in mycobacterial infections, but also the broader consequences of this pathway in macrophage physiology and how those contributions might intersect with the role of NFAT in inducing VEGFA production and angiogenesis in this environment.

5.2.1 HIF for HIF's Sake

HIF-1 α has only recently been intensively studied in the context of tuberculosis, with many of the foundational studies coming from Philip Elks's lab in the zebrafish⁵. These early studies utilized both dominant-negative and dominant-active versions of the zebrafish *hif1ab*⁶ to modulate the activity of this pathway, particularly in neutrophils, and have found that activation of HIF-1 α prolongs inflammation and improves mycobacterial clearance, suggesting that at early time points, HIF- α plays a protective role in infection (Elks et al., 2011; Elks et al., 2013; Hammond, Lewis, and Elks, 2020). These protective effects also extend to macrophages, where HIF-1 α stabilization enhances bacterial killing, presumably related to alterations in metabolism (Knight et al., 2018; Zenk et al., 2021). However, it has also been shown that HIF-1 α prolongs alveolar macrophage survival after injury, an effect that, if present during tuberculosis, may actually exacerbate disease due to the maladaptive

⁵There are also described roles for HIF-2 α in regulating certain aspects of neutrophil biology during mycobacterial infection, but this is beyond the scope of the presently proposed work (Thompson et al., 2014; Elks et al., 2015).

⁶As is all too common, but rarely quite so penetrant, zebrafish have two copies of all of the HIF-1 α subunits, making for an expansive coding space for these genes in the zebrafish genome and each copy appears to have some distinct effects, making for both opportunity and frustration (Elks et al., 2015).

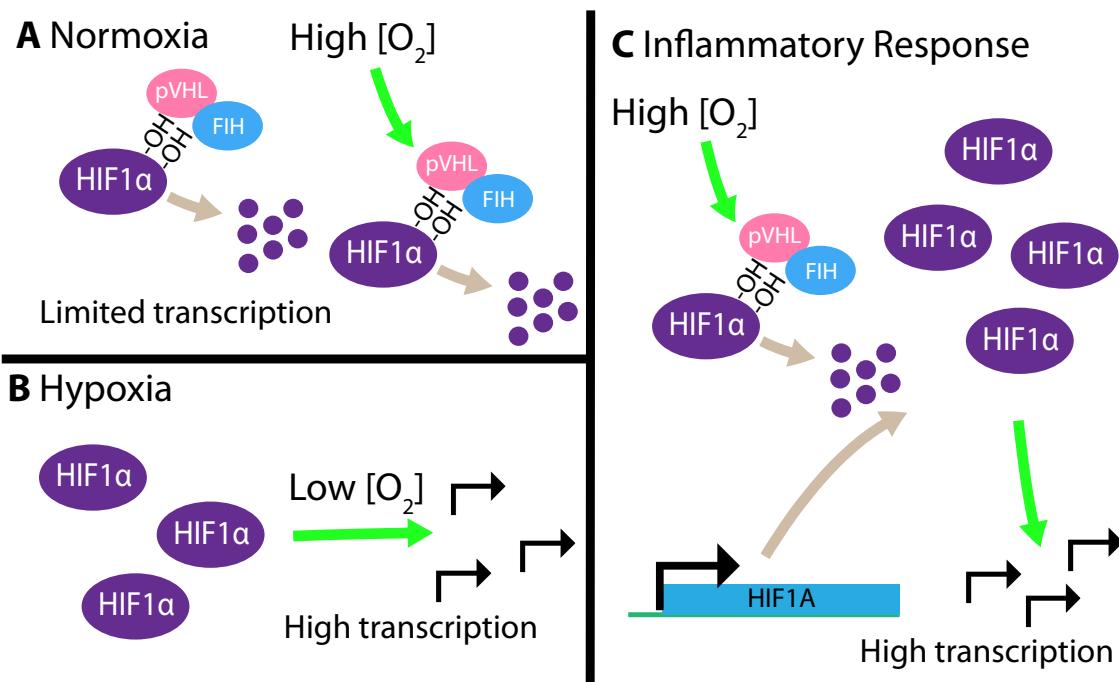


Figure 5.9: Schematic demonstrating the different states of HIF-1 α . (A) shows the standard normoxic state of HIF-1 α , where it is rapidly degraded by oxygen-dependent enzymes; (B) these enzymes are inhibited in hypoxia, leading to HIF-1 α -dependent transcription. (C) Immunological stimuli can result in HIF-1 α transcription, leading to functional activation even in the absence of overt hypoxia.

nature of alveolar macrophage responses; prompt necrosis of these cells would likely be more beneficial than their continued survival as a protective niche for the bacteria (Leemans et al., 2001; Woods et al., 2022). Other studies have also identified an essential role for HIF-1 α in both IFN- γ responses and TNF- α expression, both of which are essential for bacterial control (Flynn et al., 1993; Flynn et al., 1995; Braverman et al., 2016; Lewis and Elks, 2019). However, unlike the responses in macrophages, HIF-1 α stabilization in T cells inhibits effective cytotoxic responses, impairs host immunity, and induces disease tolerance, so it is clear that the balance of effects is critical to the determination of the overall role of HIF-1 α during infection (Tzelepis et al., 2018; Liu et al., 2022b).

More generally, HIF-1 α mediates both priming and responding to diverse insults.

Hypoxia is able to induce the expression of *TLR2* and *TLR6*, which are important for broad responses to various infections, including *M. tuberculosis* (Kuhlicke et al., 2007). However, once induced by the pathogen itself, HIF-1 α is a critical inducer of *IL1B* and *IL-1 β* which, as we have seen, is an important dimension of the host defense against mycobacterial infections (Ogryzko et al., 2019). Indeed, HIF-1 α is able to be activated indirectly by C-type lectin signaling, suggesting it may play a role in major antifungal and mycobacterial responses although the precise consequences of this mechanism of activation are still being worked out (Friedrich et al., 2017; Elder et al., 2019). As might be expected, genetic stabilization of HIF fosters more robust protective responses against multiple pathogens, paralleling findings from mycobacteria and making this a potential target for agonism as a means of fostering broad immune defense as well as trained immunity (Schild et al., 2020). This extends to macrophages specifically, which can exert improved killing of *M. tuberculosis* after HIF-1 α activation (Li et al., 2021). Different physiological conditions within the host can impair HIF activation and HIF-dependent responses, including high blood glucose, adding a potential dimension to the immunosuppression experienced by many diabetics (Teran et al., 2022). On the other hand, glucose influx is important for initial glycolysis initiation, so there is likely to be some degree of Goldilocks effect where the right amount of glucose can facilitate glycolytic metabolism but too much overwhelms the system and too little starves the cells (Stunault et al., 2018). Other studies have implicated HIF-1 α signaling in inhibiting macrophage necrosis in mycobacterial granulomas, indicating that this signaling pathway may exert some protective effects in particular contexts, although this study focused on *M. avium* granulomas which differ in some respects from *M. tuberculosis* granulomas (Cardoso et al., 2015).

Alterations in metabolism through HIF-1 α also underly so-called “trained immu-

nity⁷,.” The induction of aerobic glycolysis within macrophages primes them to respond more vigorously to future insults, an effect that appears to be able to last weeks, months, or years (Cheng et al., 2014). As HIF-1 α is a known mediator of IL-1 β induction, it appears that increased transcription in the absence of post-translational processing to increase the pool of latent pro-IL-1 β is one of the mechanisms by which this process is able to operate (Arts et al., 2018). Trained immunity may even serve as a transgenerational mechanism of protection against insults (Katzmarski et al., 2021), although this is controversial (Kaufmann et al., 2022; Katzmarski et al., 2022). The synthesis of the effects on metabolism with effects on macrophage biology and interactions with the surrounding stroma in mediating some of the effects of trained immunity still require a great deal of investigation, although this is certainly yet another realm where major families of immunologically relevant transcription factors surely play some role. It is known that the BCG vaccine is able to induce broad trained immunity, which is a potential mechanism by which BCG is thought to be able to protect against both bladder cancer, and, perhaps, COVID-19 and other respiratory viruses (Arts et al., 2016; Arts et al., 2018; Covian et al., 2019; van Puffelen et al., 2020; O’Neill and Netea, 2020).

This work nicely complements work from Didier Stainier’s lab, which used a combination of *hif1aa* and *hif1ab* mutant zebrafish and new macrophage-specific transgenic tools to manipulate the HIF-1 α signaling pathway and found that this pathway, specifically in macrophages, was critical for mediating developmental angiogenesis. Unlike the behaviors seen in neutrophils, specific expression of even a wild-type *hif1ab*

⁷Trained immunity is a process in which the innate immune system is primed to respond to future insults more effectively. This akin to the way that vaccinations prime the adaptive immune system, but through a completely different set of mechanisms and is non-specific, allowing for broad protection against diverse insults.

in macrophages was toxic to the cells and rendered them impotent⁸ (Gerri et al., 2017). This toxicity is likely due to sequestering of important binding partners, but may be due to metabolic changes that leave the cells unable to produce sufficient ATP, albeit by two divergent mechanisms – wild-type *hif1ab* may produce a strict reliance on glycolysis while dominant-negative *hif1ab* may force oxidative phosphorylation at rates exceeding the ability to produce pyruvate. A clearer understanding of the precise mechanism of this transgene-specific macrophage toxicity would likely reveal interesting new dimensions to the HIF-1 α pathway in macrophages⁹. This evokes an important function of this pathway in these cells that current tools remain unable to address.

HIF-1 α signaling is also implicated in various aspects of tumor biology and, as mentioned in Subsection 1.5.1, is important in mediating the angiogenic response in cancer. It also plays important functions in other aspects of cancer biology, including modulating tumor pH and the extracellular matrix (Dayan et al., 2008). These studies within tumors have also revealed interesting layers of transcriptional regulation within the HIF-1 α regulon and the functional differences between FIH and PHD, where PHD proteins are very strictly dependent on environmental oxygen while FIH has a much higher affinity for oxygen and can operate even at relative hypoxia. Given the different domains targeted by these proteins, HIF-1 α can exert biphasic effects at varying

⁸It is difficult to contextualize all of these results; mRNA injection of *hif1ab* results in a discrete set of less-severe phenotypes than transgenic expression of this cDNA strictly within macrophages? Additionally, this expression is clearly an impediment to immune responses but the Elks lab has repeatedly seen protection from whole-organism HIF-1 α stabilization and overexpression in neutrophils, arguing that there are confusing layers of additional regulation or compensation that we do not presently understand that stand in the way of effective transgenic control of this pathway.

⁹Other hypotheses include the overexpressed HIF-1 α sequestering essential cofactors from other proteins, breaking the stoichiometry of essential protein:protein interactions, or some other mechanism. That does not explain why mRNA overexpression and genetic knockout are both better tolerated, however.

oxygen tension, with certain genes upregulated at modest hypoxia and additional genes induced under severe hypoxia (Dayan et al., 2008). Such gradients of response may be critical to understanding this pathway during mycobacterial infection, where the timing of these changes in relative oxygen concentration may reflect distinct stages of disease.

To further the study of the HIF pathway in the context of mycobacterial infection in the zebrafish, a set of new tools should be made: one is a set of reporter constructs to better identify both hypoxic and transcriptional induction of *hif1ab* in macrophages and the other is a conditional approach to the expression of dominant negative and dominant active versions of HIF-1 α in macrophages, potentially enabling new cell-autonomous understanding of the role of this pathway in mycobacterial pathogenesis and angiogenesis.

Previous work in our lab using *in situ* hybridization for *phd3* mRNA revealed the upregulation of this gene surrounding the mycobacteria, indicating hypoxia (Oehlers et al., 2015). The Elks lab generated a transgenic line using a bacterial artificial chromosome containing the promoter for *phd3* that expresses GFP (Santhakumar et al., 2012), which serves as a useful spatial and temporal readout for HIF-1 α transcription factor activity across different tissues. However, this tool is unable to distinguish between normoxic and hypoxic activation or different cell types, so new tools are required to better address these questions.

During conditions of hypoxia, HIFs are degraded through hydroxylation in the oxygen dependent degradation domain (ODD) that contains two proline residues that are hydroxylated by PHD proteins, leading to proteasomal degradation. This ODD has been shown to be both necessary and sufficient to direct oxygen-dependent degradation, so it seems reasonable to use a macrophage-specific promoter to

drive expression of ODD linked to a fluorescent protein as a reporter for granuloma hypoxia. This would be stabilized at low oxygen concentration while being constitutively degraded under normoxia. This would allow for a clearer report of the degree of present hypoxia and complement existing tools, including pimonidazole (Huang et al., 1998; Cousins et al., 2016). If this tool alone were to be sufficient to induce macrophage toxicity, then an alternative approach could be taken to induce this transgene only when needed using degradation domains or inducible promoters, such as the *hsp70l* heat shock-inducible promoter¹⁰. Further discussion on degradation domain regulation will be provided shortly.

In parallel, a reporter is needed to provide a readout of normoxic activation of HIF-1 α , which is predominantly thought to be regulated at the transcriptional level and through accumulation of the citric acid cycle intermediates α -ketoglutarate and succinate (Tannahill et al., 2013; Palazon et al., 2014; Meng et al., 2018; Williams and O'Neill, 2018). Therefore, either ectopic expression or direct protein fusion strategies would be appropriate to the study of this pathway. If some promoter could be identified that responded comparably to the native *hif1ab* promoter or a CRISPR-mediated knockin could be generated, this would be a useful reporter of transcription-level induction of HIF-1 α , a process likely relevant to inflammatory responses during infection. Alternatively, a bacterial artificial chromosome-based transgenic could be constructed, which might be easier than screening for a knockin while more faithfully recapitulating the expression pattern than a promoter-based transgenic. This, in tandem with the previously mentioned ODD transgenes would allow for a more thorough dissection of the relative contributions of hypoxia *per se* and the ac-

¹⁰If this were to mediate macrophage toxicity, that would be informative as to the mechanism of the toxicity observed with expression of the full-length protein and I would hypothesize would indicate other, macrophage-specific essential targets of PHD and FIH that have yet to be identified. Expanding the regulatory landscape of these oxygen-dependent enzymes would shed a great deal of light on the total, integrated hypoxia response *in vivo*.

tivity of HIF-1 α . An additional approach would be to develop biosensors that can detect either succinate or α -ketoglutarate, examples of which already exist in the literature (Liu, Liu, and Wang, 2017; Luddecke et al., 2017; Suzuki et al., 2021). Developing macrophage-specific sensors to detect these HIF-1 α -inducing metabolites would offer another means to assess these effects while also offering a secondary approach to understand the mechanisms of HIF-1 α induction during immune responses to mycobacterial infection.

As previous transgenic attempts have failed, the expression of dominant negative-*hif1ab* and dominant active-*hif1ab* in macrophages will require new approaches. It seems that misregulation of HIF-1 α signaling results in some sort of developmental toxicity in the macrophage, so it is critical that HIF is only modulated in the time and place where it is most relevant. Thus, using established estradiol-responsive constructs, a set of fish should be made expressing dominant negative-*hif1ab* and dominant active-*hif1ab* covalently linked to ER50 degradation domains, which sequester proteins in the cytosol and target them for degradation except when in the presence of tamoxifen (Miyazaki et al., 2012). This would allow for the creation of HIF-modulating transgenes within macrophages that would be expected to have reduced toxicity and allow for time-targeted modulation of this pathway. Like the previous tools, this seems especially relevant in the context of mycobacterial granulomas from adult zebrafish, which are known to be hypoxic and more closely resemble human granulomas (Rustad et al., 2009; Harper et al., 2012; Oehlers et al., 2015). Alternatively, and perhaps even more elegantly, dominant negative-*hif1ab* could be created endogenously, through knock-in of a Cre-flippable trap cassette into intron 7 or 8; if placed in the “off” orientation, this would allow for normal HIF-1 α activity while induction of macrophage-specific Cre^{ERT2} would then allow for macrophage-specific activation of a dominant negative version of the protein through introduction

of a splice acceptor-stop codon-polyadenylation sequence (Ni et al., 2012). Even further creative approaches could enable tissue-specific induction of dominant active HIF-1 α through a similar approach by knocking in the remaining cDNA with the hydroxylated proline residues mutated and placing this, too, under control of an invertible *loxP* pair. Such an endogenous approach would also allow flexible pairing with other Cre drivers and simultaneous fluorescent tagging of cells in which the recombination has occurred, allowing these cells to be traced through the course of infection. These are likely to reveal not only new aspects of HIF modulation of angiogenesis, but broader impacts on the overall response to infection, especially given the central role of HIF-1 α in altering macrophage metabolism. These constitute hemizygous approaches able to be quickly paired with other tissue-specific Cre drivers, allowing for flexible application of these reagents in diverse scientific inquiries.

5.2.2 Macrophage Metabolism in Immunity

Upon activation, macrophages undergo a HIF-1 α dependent metabolic switch that corresponds, in part to their longevity and function (Kolliniati et al., 2022). While immediate/early-responding macrophages¹¹ undergo an inflammatory metabolic switch and begin to utilize aerobic glycolysis for metabolism as a means of rapidly (albeit inefficiently) generating energy, later arrivals and macrophages that survive over longer periods must utilize oxidative phosphorylation to optimize energy consumption over a longer course of immunological response (Kiran et al., 2016; Langston, Shibata, and Horng, 2017; Shi et al., 2019; Viola et al., 2019; Taylor and Scholz, 2022). Thus, while macrophages may initially utilize bursts of glycolysis to fuel rapid, oxidizing

¹¹In the literature, these are so-called “M1” macrophages and while useful to acknowledge the inflammatory and activated state of these cells, it is not clear to me nor does it seem likely that macrophages strictly use only one metabolic modus at a time, although many of them are relatively depleted of mitochondria (Biswas and Mantovani, 2012).

responses, this is an unsustainable mode of metabolism over the long term and these macrophages must either transition to the use of oxidative phosphorylation, disperse, or perish (Odegaard and Chawla, 2011; Howard and Khader, 2020). These metabolic switches are thought to be predominantly regulated by a combination of HIF-1 α , AKT, and mTOR, although this does not exclude the possibility of other factors influencing these changes, including, perhaps, the NFAT signaling pathway¹² (Karar and Maity, 2011; Covarrubias, Aksoylar, and Horng, 2015). The reason for the present interest in macrophage metabolism is many fold: there is comparatively little known about the immunometabolism of the granuloma itself (at least compared to that of macrophages *sensu stricto*), this process intimately involves HIF-1 α signaling (a classical mediator of angiogenesis), and is clearly an important dimension of a variety of protective immune responses and we currently lack a model that integrates what is likely the most important second messenger of eukaryotes (divalent calcium) with macrophage metabolism, which may allow for a more comprehensive integration of NFAT signaling with these broader changes in macrophage biology.

The immediate burst of glycolysis additionally generates important metabolic byproducts that can be repurposed for other cellular processes and which might have otherwise been consumed by the citric acid cycle (Kelly and O'Neill, 2015; Viola et al., 2019). One of the earliest observations on this point evaluated the consequences of loss of function of HIF-1 α on the ATP concentrations within myeloid cells and found dramatically reduced levels during inflammation (Cramer and Johnson, 2003). Notable among these are succinate and α -ketoglutarate, both of which are able to activate HIF-1 α and whose production is agonized by increased HIF-1 α activation

¹²Perhaps more likely is that the metabolic state of the cell influences the ability of NFAT to respond – this will require a long time to fully dissect, appreciate, and synthesize in order to develop the relationships that might exist within the world of possible macrophage responses in the context of their temporal metabolic state.

in a feed-forward loop. Wound sites and other sites of insult tend to have reduced oxygen tension, which drives the hypoxic activation of HIF-1 α as well. Together these inflammatory and chemical processes induce the production of VEGFA (as well as other cytokines and chemokines) to effectively respond to these stimuli. The macrophage-specific induction of HIF-1 α can act to mediate critical aspects of both endothelial cell biology as well as further macrophage responses in an autocrine and paracrine manner.

The glycolytic switch that occurs in these first responders is dependent upon and triggered by the inducible nitric oxide synthase (iNOS), which is induced by HIF-1 α and generates nitric oxide (NO). Nitric oxide is able to inhibit mitochondrial respiration, reducing the oxidative phosphorylation capacity of the cell and driving it toward alternative means of ATP production, the most biochemically compelling of which is aerobic glycolysis. The consumption of glucose by glycolysis without the ability to complete the citric acid cycle results in the accumulation of succinate and α -ketoglutarate, which further activate HIF-1 α . This activation of HIF-1 α drives the transcription of glucose transporters and lactate dehydrogenases to block mitochondrial utilization of the pyruvate end-product and results in mitochondrial wasting (Galvan-Pena and O'Neill, 2014; Russell, Huang, and VanderVen, 2019). This increase in glucose import has been shown to be important in the pathogenesis of leprosy, although specific experiments with other mycobacterial species would be required to validate the assumed generalizability of this process (Medeiros et al., 2016; Montoya-Rosales et al., 2016; Vance et al., 2019). HIF-1 α , as we have seen, also induces the expression of numerous cytokines and chemokines, making it an integral part of the overall response to the detection of diverse PAMPs, the model of which is LPS (see Subsection 1.3.2). As a major mediator of inflammatory responses, the HIF-1 α -dependent glycolytic phenotype of macrophages facilitates rapid, ener-

getically expensive responses to these acute threats but is unsustainable over the long term, forcing a net shift back to oxidative phosphorylation due to both cellular turnover and *in situ* metabolic reprogramming.

Recent findings in the field of tuberculosis pathogenesis indicate that the mechanistic target of rapamycin (mTOR) is a critical host protective factor against early mycobacterial infection through the regulation of oxidative phosphorylation (Pagan et al., 2022). Mycobacteria secrete ESAT6 to damage mitochondria, which results in a necrotic signaling cascade that leads to cell death (Roca et al., 2019), but the activity of mTOR limits this cytotoxicity. In the absence of mTOR, these macrophages are hypersensitive to killing by mycobacteria, resulting in a dramatic loss of bacterial restriction. This role for oxidative phosphorylation within macrophages in limiting mycobacterial growth argues against the classic model of glycolytic macrophages being more bactericidal and, rather, that increased macrophage longevity, even at the expense of “inflammation” is more productive for limiting infection. This role for macrophage metabolism in restricting bacterial growth is a further mechanism by which host metabolism can limit bacterial growth despite the efforts of the bacteria (Pernas, 2021). This adds yet another host-protective factor to the genetic arsenal of the host and vividly demonstrates the impact that loss of discrete pathways can have on the progression of disease.

In order to switch back to oxidative phosphorylation within a single cell, HIF-1 α must be degraded and alternative pathways induced. The net balance of inducible nitric oxide synthase-dependent production of nitric oxide and citrulline and arginase-dependent production of urea and ornithine appears to be critical for this shift (Qualls and Murray, 2016; Palmieri et al., 2020). This places arginine at the center of tuberculosis immunometabolism and its byproducts – through nitric oxide and itaconate –

determine key aspects of host immunity. Despite the traditional assignment of *NOS2* as an M1 marker and *ARG1* as an M2 marker, it is known that LPS – the most classical of M1 stimuli – can paradoxically induce arginase, which would argue that our current knowledge of the factors that contribute of these different states (if they are to be states as such) is lacking¹³ (El Kasmi et al., 2008). A further metabolite in this process, itaconate¹⁴, has a range of important immunomodulatory and directly antimicrobial properties that have only more recently been explored in the context of tuberculosis immunity (He et al., 2021; Jinich et al., 2022). As these processes reflect directly on amino acid metabolism, it is logical that other pathways related to amino acids would contribute to immune processes and indeed, the biosynthesis of other amino acids within macrophages appears to be important for granuloma formation, but the precise roles of these are relatively unclear.

A great deal of work has been done to study discrete genetic mutants in major genes along these pathways, but the interpretation of these data is challenging. One issue is that much has been done *in vitro* under standard LPS/IFN- γ or IL-4/IL-13 polarization schema or in the mouse, where more comprehensive profiling is possible but is hindered by attempts at placing these changes within dogmatic M1-M2 binaries; the knockout of *Arg1* or *Nos2* definitionally changes the cell's location within this dichotomy even if it does not fundamentally change the cellular identity of these populations. A more open-minded approach to the functional consequences of mutating critical metabolic genes within macrophages is needed to develop a more lucid

¹³In the presence of both iNOS and arginine, relative affinity and expression may play as-yet unexplored roles in shaping the overall tone of the immune response on the single cell level. The full categorization of this will require further technology development in single cell proteomics that is far more sensitive than current approaches.

¹⁴The role of itaconate in infection is interesting as this is a product of aconitate dehydrogenase, also known as *acod1* or *irg1*, which we use to mark macrophages. A more thorough investigation of the role of *acod1* in the granuloma is needed to understand the contributions of this definitive macrophage marker in the actual biology of infection (Nair et al., 2018).

understanding of the consequences of shifting macrophage metabolism along a fine gradient. All of these shifts are interwoven with HIF-1 α and depend on both inflammatory signals as well as the existence of formal hypoxia and the intersection between these signals has, thus far, been difficult to tease apart.

These metabolic changes are an important aspect of the capacity for macrophages to engage with mycobacteria, although these changes are not necessarily for the betterment of the host. Mycobacteria benefit from phagocytosis by macrophages undergoing glycolysis, even though these are traditionally thought to be more bactericidal phagocytes (Mehrotra et al., 2014; Escoll and Buchrieser, 2019; Shi et al., 2019). This is on account of the induction of fatty acid biosynthesis; mycobacteria utilize lipids as a primary carbon source and the production and uptake of additional lipids by the macrophage, including cholesterol, enhances bacterial growth (Gatfield and Pieters, 2000; Nazarova et al., 2019; Yan and Horng, 2020). Given the known roles for NFAT in regulating PPAR-dependent transcriptional responses and the essential role of NFATC2 in the induction of foamy macrophages, this indicates that NFAT-dependent responses may be critical determinants of macrophage lipid metabolism via their interactions with PPAR family transcription factors (Du et al., 2020; Du et al., 2021; Laval, Chaumont, and Demangel, 2021). This is evidenced by the ways in which *M. tuberculosis* subverts cellular metabolism through the use of the tuberculosis necrotizing toxin (TNT), which depletes the macrophage of NAD $^{+}$, which induces metabolic adaptation within the macrophage to compensate this loss of NAD $^{+}$ and transition back to oxidative phosphorylation in those instances where the macrophage does not simply die (Howard and Khader, 2020).

The products and intermediates in glycolysis have known antimycobacterial activity, including nitric oxide and prostaglandins (MacMicking et al., 1997; Mayer-Barber

et al., 2014), making macrophage metabolism a delicate balance between the bacteria's ability to manipulate the macrophage into a permissive state while minimizing the risks from these potentially antimicrobial responses (Osada-Oka et al., 2019). However, these metabolic changes also induce the production of IL-1 β , which has known protective activity against mycobacteria, suggesting that macrophages are able to utilize strategies to overcome these layers of bacterial subversion (Juffermans et al., 2000; Fremond et al., 2007; Corcoran and O'Neill, 2016; Ogryzko et al., 2019; Shi et al., 2019). Another aspect of this is wherein macrophage accumulate lipids for their own utility while barring bacterial access to these resources by the sequestration of them within cytosolic lipid droplets (Knight et al., 2018; Laval, Chaumont, and Demangel, 2021). The limitation of macrophage access to cholesterol through various means has the potential to induce forms of nutritional immunity against the bacteria and shift macrophages into more bactericidal states (Pandey and Sasse, 2008; Yang et al., 2014d; Babunovic et al., 2022). A more comprehensive evaluation of the overall metabolic drivers in macrophages during these immune responses is still very much a work in progress with many questions remaining about how mycobacteria are able to alter macrophage metabolic status for their benefit and compensatory mechanisms within the macrophage to overcome such subversion (Pagan et al., 2022). Many of these processes may be mediated by the induction of PPAR- γ , which is able to induce the import of cholesterol and other lipids and is known to be induced in granuloma macrophages, offering a complementary metabolic pathway to further define in the context of mycobacterial infection (Chawla et al., 2001).

Under sustained immunological stimulation during hypoxia, such as that found in the tuberculous granuloma, metabolic reprogramming toward oxidative phosphorylation occurs within macrophages that corresponds to downregulation of HIF-1 α (Mehrotra et al., 2014; Wilson, Mayr, and Weichhart, 2019). This raises a po-

tent conflict between the production of VEGFA, the M2-like identity of many of the VEGFA-expressing granuloma macrophages, and the downregulation of the presumptive master regulator of VEGFA, HIF-1 α . Some aspects of this conflict between the paradoxical decrease in HIF-1 α signaling and the increase in VEGFA production can now be resolved based on the present work, which presents an alternative model of a C-type lectin receptor-NFAT transduction pathway able to induce the transcription of VEGFA independent of hypoxia¹⁵. It may be that under this model the metabolic status of the macrophages is ultimately unimportant for VEGFA production or there are additional layers of intersectional regulation that depend on metabolism in ways as yet unknown. This also raises questions about why VEGFA is considered an M2 cytokine when its transcription was thought to be dependent upon a factor that is downregulated in the transition to M2-like states¹⁶, HIF-1 α ; additionally, VEGFA is known to be induced under some M1-like conditions, arguing that our general understanding misses some critical aspects of the ways in which angiogenesis can be induced. A more complete integration of the findings of this work with the general metabolic status of granuloma macrophages that produce VEGFA will be important for resolving some of these quandaries on the contribution of HIF-1 α to granuloma macrophage metabolism, antibacterial responses, and angiogenesis.

It is already known that altering macrophage metabolism by further upregulation of glycolysis is able to normalize surrounding vasculature through greedy consumption of glucose and resulting starvation of the vasculature (Wenes et al., 2016). These

¹⁵This is evidenced by the induction of angiogenesis in the larval zebrafish, which is perfused with oxygen, and the induction of VEGFA in THP-1 macrophages, which are cultured under 21% O₂ in air.

¹⁶Given the near ubiquitous expression of *hif1ab* in the granuloma and recent findings on the role of type II responses in inducing granuloma formation, this model is clearly lacking important details on the roles of these phenomena in mediating angiogenic signaling, but these more detailed approaches will hopefully spur new investigation in these realms.

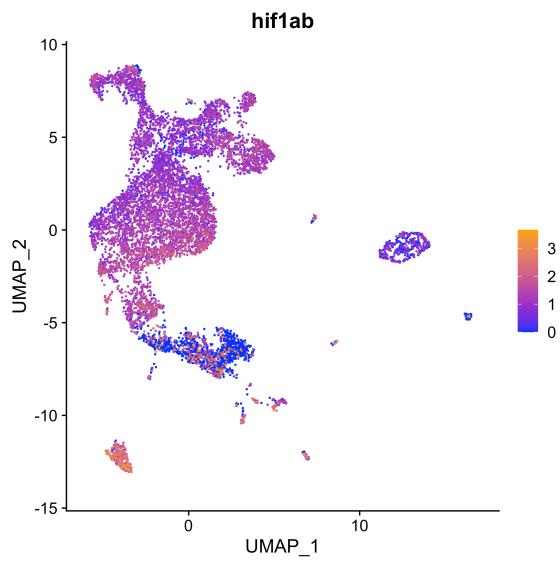


Figure 5.10: The primary zebrafish homolog of *HIF1A*, *hif1ab*, is highly expressed across diverse macrophage populations within the zebrafish, belying its supposed role as an M1 macrophage marker and suggesting that it may be playing important roles in mediating critical responses during mycobacterial infection.

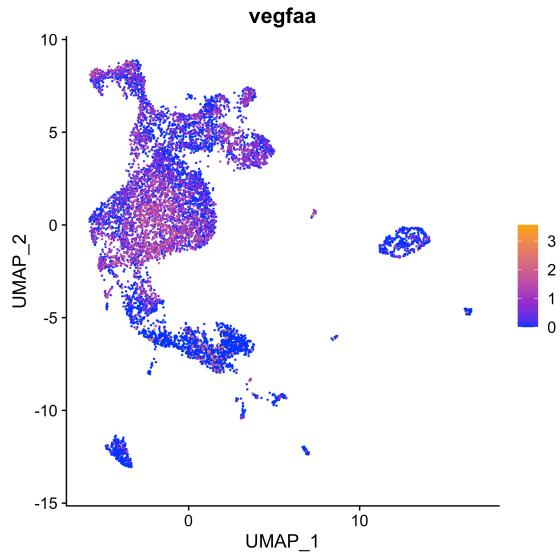


Figure 5.11: As we have seen previously, *vegfaa* is expressed in macrophages at the site of infection in larval zebrafish and is also highly expressed within certain granuloma macrophage populations, including both epithelioid and inflammatory macrophage subclasses.

effects are especially notable in tumors, where tumor-associated macrophages are heavily dependent on glycolysis and inhibition of glycolysis in these macrophages actually results in reduction of tumor burden through a shift back to more M1-like phenotypes (Rabold et al., 2017; Mehla and Singh, 2019). This raises some conflicts about the relative merits of modulating glycolysis within these tumors, but such relative contributions (as well as alterations in the growth of primary tumors and rates of metastasis) are worthy of further exploration (Mojsilovic et al., 2021). While glycolysis, of course, lies at the base of many different metabolic pathways, it may be the case that in the absence of glycolysis that other pathways are able to compensate within M1 macrophages to exercise these phenotypes while the increased metabolic demands of M2 macrophages result in them polarizing into more inflammatory states under such starvation conditions. Most (but not all) of the macrophages within the tumor microenvironment engage the citric acid cycle and oxidative phosphorylation for energy production as a more efficient means of producing energy, so subversion of the root of these processes (glycolysis) may offer a means of more broadly starving these cells with variable consequences (Yu et al., 2020; Puthenveetil and Dubey, 2020; Geeraerts et al., 2021). The net effect of the blockade of HIF-1 α in the tumor microenvironment results in diminished tumor growth and may be a useful target to metabolically reprogram tumors and tumor-associated macrophages (Hong, Lee, and Kim, 2004). However, more targeted approaches may allow for more thorough dissection of different cell types in different cancers and upregulation of glycolysis within tumor-associated macrophages concurrent with downregulation within the tumor itself may offer a potent combination able to even more effectively restrict tumor growth. These concepts may be translatable to the granuloma context, where increasing macrophage glycolysis may be beneficial in inducing vascular regression and potentiating bacterial killing while inhibition of glycolysis in T cells may be

beneficial to improve both disease tolerance and CD8⁺ responses (Phan, Goldrath, and Glass, 2017; Tzelepis et al., 2018; Russell et al., 2019; Wik and Skalhegg, 2022). Interestingly, it is known that NFAT signaling can control certain aspects of T cell metabolism, suggesting possible generalizability of this effect to other cell types at the intersection of diverse signals (Vaeth and Feske, 2018).

To better understand some of these effects within the granuloma, it is critical to expand the utility of single cell -omics approaches to capture the evolution of metabolism across the course of infection by metabolomics to gauge the contributions of various major metabolic pathways to the activities of discrete populations of cells (Somashekhar et al., 2011). This should also include a more comprehensive profile of the metabolites present within the necrotic core of the granuloma over time, which will reflect certain critical aspects of the immune response that can be inferred from these changes over time. Integration of these findings with the previously proposed new tools to study HIF-1 α may allow for a more complete picture of the contributions of HIF-1 α -dependent metabolic changes to bacterial control and open new avenues of host-directed therapeutic intervention able to shift different metabolic programs at different stages of disease.

5.2.3 HIF-NFAT Interactions

A tempting hypothesis is that HIF-1 α and NFATC2 are regulating or physically associating with one another to induce the expression of VEGFA. The model most consistent with the existing literature is that NFATC2 is upregulating the transcriptional expression of HIF-1 α under circumstances where HIF-1 α is already being stabilized; these interactions could also be reciprocal, where HIF-1 α activation induces the expression of further NFATC2, consistent with our previous observations (Section 3.4).

This would feed-forward the induction of VEGFA and may be essential for regulating these responses in normoxic environments or very early in infection.

A piece of standalone support for this model is a study conducted in mast cells that demonstrated a calcineurin-NFAT dependent upregulation of HIF-1 α at the transcriptional and protein levels. Treatment of these mast cells with ionomycin, which increases cytosolic calcium concentrations, induced these effects in a way that was able to be inhibited by tacrolimus treatment (Walczak-Drzewiecka et al., 2008). Additionally, this effect was exacerbated by culture under 1% O₂, a description that corresponds well to the behavior of NFATC3 in the myocardium, where hypoxia increased expression of endothelin-1 in a way that is sensitive to superoxide and the activity of NFATC4, which can also be activated by hypoxia (Frutos et al., 2011; Ramiro-Diaz et al., 2014; Moreno et al., 2015). Additional studies found that hypoxia activated NFATC3 to promote pulmonary smooth muscle proliferation in a way that might promote pulmonary hypertension (Hou et al., 2013). Hypoxia can also increase proliferation of human fibroblasts through NFATC2 independently of HIF-1 α but dependent on HIF-2 α (Senavirathna et al., 2018). None of these studies but the last investigated a mechanism to integrate this activation of NFAT under hypoxia with the roles of HIF-1 α despite these phenotypes having been previously described to correspond with HIF-1 α activity (Thackaberry et al., 2002; Li et al., 2014; Sonanez-Organis et al., 2016; Qi et al., 2017; Cui et al., 2021). Even the study by Senavirathna et al. only investigated this effect within a relatively limited scope (proliferation) and not in respect to the diversity of phenotypes that are stimulated by hypoxia.

To study these interactions, it seems that combinatorial chemical inhibition strategies could be utilized, although HIF-1 α inhibitors are relatively sparse in number and understudied (Viziteu et al., 2016). However, this would only scratch the surface in

pursuit of epistatic interactions between these proteins and would neglect the potential for protein-protein interactions in mediating part of the effect. A structure based approach targeting known interacting domains on HIF-1 α and its DNA binding domain may reveal important transactivation role for HIF-1 α as well as resolve some questions about the direct versus indirect nature of NFAT regulation of hypoxia responses. Much of this work could be done in a heterologous model (HEK-293T cells, for instance) to more efficiently assess these types of binary interactions (Graham et al., 1977). Larger scale screening approaches by two-hybrid would be another potential avenue for such a project to go as the interacting partners of various domains of NFATC2 remain only partially resolved. Proximity ligation approaches, which will be discussed more so later on, offer another possibility for dissecting the HIF-1 α interactome. Unfortunately, the state of reagents to work with toward these ends in the zebrafish are somewhat limited and additional groundwork would be required to study some of these processes in an *in vivo* context, although this seems like an excellent direction for a future Ph.D. project building on the current work as HIF-1 α seems to be the most likely complementary pathway for the induction of VEGFA during mycobacterial infections; there are, however, a number of potential alternative factors that could influence NFAT-dependent transcriptional induction of VEGFA.

5.3 Transcriptional Coregulatory Mechanisms

To further dissect the contributions of other transcription factors to the NFAT-dependent angiogenesis response, we can make use of our THP-1 macrophage platform to study the role of NFAT interacting partners in the overall response to *M. tuberculosis* exposure. While wild-type NFATC2 is able to interact with a panoply of other transcription factors through both C- and N-terminal domains, the domains

Table 5.3: Lentiviral expression constructs to assess the role of NFATC2 domains for induction of VEGFA signaling. All express the mPapaya fluorescent protein on the C-terminus to allow for direct visualization.

Plasmid	Utility
pLEX:mPapaya	Empty expression vector driving expression of only the conjugated fluorescent protein, for background comparison.
pLEX:CA-NFAT1	Expression of a constitutively active NFATC2 that drives transcriptional responses independent of calcineurin.
pLEX:CA-NFAT1-ΔDBD	Expression of a constitutively nuclear NFATC2 that is unable to bind DNA, to assess the contributions of NFAT binding on the induction of VEGFA.
pLEX:CA-NFAT1-ΔRIT	Expression of a constitutively active NFATC2 unable to interact with AP-1 transcription factors, to determine the contribution of this family to the VEGFA response.
pLEX:CA-NFAT1-ΔDBD-ΔRIT	Expression of a constitutively nuclear DNA binding domain mutant also unable to interact with AP-1 transcription factors.

important for particular binary interactions have been teased out over time, largely by Anjana Rao's lab. Thus, to simultaneously test the sufficiency of NFATC2 in inducing transcriptional responses and the importance of AP-1 transcription factor binding along with the ability for NFATC2 itself to bind DNA, I have developed a set of expression plasmids that drive expression of a range of different modified NFATC2 proteins (Table 5.3).

These plasmids will enable a genetics-first dissection of the activity of NFATC2 and its potential sufficiency to induce transcription of VEGFA. While, as noted in Section 3.4 and seen in Figure 3.37, the VEGFA promoter has a number of putative NFAT binding sites, the activity of these remain unknown. Using these new genetic

tools in the defined environment of THP-1 macrophages it should be possible to dissect the function of these sites¹⁷. Distant future approaches may utilize deeper genetic probing to identify particular sites of especial importance, as has been done in Chang et al. (2004). While that publication found one site that was bound by NFAT in the myocardium, the particular binding site of importance may vary by cell type and circumstance.

It is also possible that non-AP-1 transcription factors are important for NFAT activity and this is a trickier proposition to address, but could be studied through either immunoprecipitation-mass spectrometry or biotin tagging approaches via BioID (Roux et al., 2012). The advantage of the former is the identification of more stable interactions and could be done in tandem with DNase-seq, ChIP-seq, or ATAC-seq to identify promoter occupancy of this isoform. The latter may be a technically less challenging approach, however, and may provide a useful readout of the total set of interactions experienced by NFATC2 over the course of γ Mtb exposure. The former can also be done in genetically unmodified cells while the latter would require some (minimal) additional cloning and generation of yet another lentivirus for the purpose of tagging NFATC2 with one of the several biotin ligases now available (Cho et al., 2020). Both seem useful and complementary and should be considered going forward to more comprehensively understand the NFAT interactome during mycobacterial infections. I hypothesize that there are as-yet unobserved protein-protein interactions between NFATC2 and HIF-1 α that would potentially shed light on the transcriptional regulation of VEGFA in diverse contexts including in cancer and autoimmunity.

¹⁷This could also be translated to a more tractable model, like the previously described HEK-293T cells or even RAW 264.7 murine macrophages, which are more amenable transfection hosts that would allow for more rapid screening.

5.3.1 NFAT:AP-1 Interactions

The most exhaustively studied and first identified transcriptional partner for NFAT is the AP-1 transcription factor superfamily, which includes proteins in the JUN, FOS, ATF, and MAF families (Boise et al., 1993). While some of these may strike the reader as being critical regulators of cell growth and tumorigenesis, they also play critical roles in other aspects of normal biology including in immune responses (Macian, Lopez-Rodriguez, and Rao, 2001; Eferl and Wagner, 2003). Historical studies on the role of AP-1 in inducing VEGFA expression are equivocal, with one study finding that HIF-1 α -dependent induction of *VEGFA* is independent of AP-1 and a later study finding a cooperative role for HIF-1 α and AP-1 in inducing *VEGFA* (Finkenzeller, Technau, and Marne, 1995; Shih and Claffey, 2001). However, these studies on HIF-1 α -dependent VEGFA responses may or may not be directly relevant to the interactions between NFAT and AP-1. AP-1 binding sites are often adjacent to NFAT binding sites and these two transcription factors¹⁸ may cooperate to drive the transcription of VEGFA. These cooperative interactions may be close in proximity or over a long distance and may be direct protein:protein interactions or more indirect. Regardless, given the imminent importance placed on NFAT:AP-1 interactions, this seems to be an excellent place to start on a targeted list of potential interacting partners.

A few approaches can be taken. The first, utilizing the previously described pLEX:CA-NFAT- Δ RIT constructs, would give an indication about the importance of

¹⁸To avoid providing unnecessary amounts of detail, I will refer to AP-1 as a monolithic “transcription factor” much of the time. While each of the proteins in this family serve different functions, the subtleties are perhaps beyond the scope of this prospective overview. The guiding question of this section -- is *an* AP-1 transcription factor cooperating with NFAT to drive VEGFA transcription – is sufficiently broad that future experimentation would almost certainly eventually seek to identify, at the gene level, which is responsible. The canonical AP-1 transcription factor dimer is comprised of c-Fos and c-Jun, which may serve as a starting point for future investigation.

direct interactions with NFATC2, as these mutate the critical residues for complexing with AP-1 proteins. Additionally, there are a number of relatively broad inhibitors against AP-1 that could be used to test whether inhibition of AP-1 alone alters the VEGFA transcriptional response to mycobacteria *in vitro* or the overall angiogenic response *in vivo* (Huang et al., 1997; Makino et al., 2017). It has been previously demonstrated that AP-1 can activate VEGFA transcription as well as VEGFD, suggesting that this pathway may have as-yet unappreciated roles in inducing VEGFA during immune responses and may act in concert with NFAT to execute these functions¹⁹ (Shih and Claffey, 2001; Debinski et al., 2001; Josko and Mazurek, 2004; Wang et al., 2016; Guo et al., 2022).

5.3.2 NFAT:NFκB

As has been discussed previously (see Section 1.4), the interplay between NFAT and NF-κB is widely discussed but the precise biological consequences of these two transcription factor families in inducing VEGFA remain unknown. It has previously been shown that NF-κB is able to induce the transcription of VEGFA in various contexts (Lukiw et al., 2003; Xie et al., 2010; Greenberger et al., 2010) and is important for VEGFA expression in macrophages (Kiriakidis et al., 2003). Lukiw et al. (2003) even uncovered important interactions between HIF-1 α and NF-κB in the induction of VEGFA during hypoxia, suggesting the potential for there to be multiple layers of signaling required to induce VEGFA during particular context and which may vary based on the stimulus. Disentangling the complex interactions within this transcriptional triumvirate would be certain to reveal key new aspects of the biology of

¹⁹More classical data suggests that HIF-1 α -dependent VEGFA upregulation is independent of AP-1, although this data may be limited by the scope of the time and the particular experimental conditions. Or hypoxia-induced VEGFA may be independent of AP-1, but pathogen-induced VEGFA is dependent on it – these all remain to be explored (Finkenzeller, Technau, and Marme, 1995).

inflammatory VEGFA induction, although this would certainly be a major challenge requiring careful integration of genetic and biochemical approaches to understand promoter occupancy dynamics, protein:protein interactions, and regulatory networks underlying these phenomena.

The overarching theme in studies of NFAT-NF- κ B interactions is that they can either be antagonistic or cooperative and *a priori* prediction of which is likely to occur has been elusive (Fisher et al., 2006; Khalaf, Jass, and Olsson, 2013). Foundational studies in this field found cooperative roles for NF- κ B and NFAT in regulating the expression of IFN- γ (Sica et al., 1997) and that they were differentially induced by alterations in cellular calcium (Dolmetsch et al., 1997). Direct dimerization was first suggested with NFAT5, which established a model for the study of these interactions in the NFATc isoforms (Lopez-Rodriguez et al., 2001). NF- κ B and NFAT also directly interact to synergistically promote cardiac hypertrophy, roles consistent with those described for NFATC2, NFATC3, and NFATC4 (Subsubsection 1.4.2, Subsubsection 1.4.2, and Subsubsection 1.4.2) (Liu et al., 2012). And as we have seen in the context of osteoclast biology, NF- κ B can bind to the promoter of NFATC1²⁰ to drive increased expression to allow for osteoclast differentiation (Asagiri et al., 2005). However, activation of the specific NF- κ B member RELB has been shown to inhibit NFATC1 expression, so there may be important regulatory layers based on the particular NF- κ B family member engaged – a critical consideration that is difficult to account for with present knowledge²¹ (Zhao et al., 2015). And in the context of TLR agonism, NFATC3 nuclear occupancy was important for NF- κ B responses

²⁰The relevance to other NFAT proteins remains unknown, but at least NFATC2 has putative NF- κ B binding sites in the proximal promoter.

²¹One of the most important things to keep in mind is the arbitrary complex that is being introduced through this hypothesis. If NFAT/NF- κ B interactions are being mediated through either different dimers or trimers, there are suddenly >500 possible combinations of proteins that could offer subtle differences in response, but this becomes an insurmountable experimental hurdle.

but was not induced by TLRs and could either enhance or inhibit NF- κ B responses (Conboy et al., 1999; Minematsu et al., 2011). In the endothelium, the net effect of angiogenesis is determined by the balance of competitive promoter occupancy between NF- κ B and NFAT through the expression of FASL and cFLIP, respectively (Aurora et al., 2010). NF- κ B-dependent expression of FASL drives endothelial apoptosis while NFAT-dependent expression of cFLIP promotes cellular survival, despite both proteins binding both promoters. This pairwise antagonism between these two transcription factors offers an intriguing model that should be further explored in other contexts. Yet other regulatory factors may be involved in regulating these roles, but this serves as an interesting potential regulatory mechanism worthy of investigation in the context of macrophage VEGFA signaling given the composite evidence of NF- κ B importance for macrophage expression of VEGFA and the known layers of regulation surrounding NFAT and NF- κ B signaling and the data presented here on the role of NFAT in mediating this process during mycobacterial infection.

One of the interesting observations that came out of the work studying the NFATC2-dependent transcriptional responses to *M. tuberculosis* was the observation that γ *Mtb*-exposed THP-1 macrophages seemed to have greater production of NFATC2 protein by immunofluorescence, an unusual potential regulatory mechanism only rarely described in historical literature (Aramburu et al., 1995; Asagiri et al., 2005) and then only for NFATC1. The concept that there may be some sort of upstream priming event that stimulates NFATC2 production and/or downregulation of NFATC2 protein degradation offers the potential for a new layer of regulation of NFAT activity. Further experimentation will need to be done to confirm this and I would propose RNA-seq of THP-1 macrophages as an excellent first-attempt to categorize some of the macrophage-specific responses to extracellular exposure to *M. tuberculosis*, which is an as-yet unexplored dimension to *in vitro* studies of

macrophage-*M. tuberculosis* interactions and may reveal new mechanisms of intracellular subversion of macrophages by contrast with the numerous existing datasets on macrophage *infection*.

5.4 New Genetic Approaches to the Study of NFAT Signaling

One of the dominant tools in the field for the study of intracellular calcium flux is the use of GCaMP, a modified green fluorescent protein that fluoresces in response to calcium binding (Nakai, Ohkura, and Imoto, 2001; Zhang et al., 2021b). Since its initial development, many interactions have developed allowing for ever-finer detection of various aspects of cellular calcium concentrations and localization. These tools have been used in the fish to detect and manipulate cellular behavior in both immobilized and freely moving fish (Beerman et al., 2015; Kim et al., 2017). Calcium sensors have clear promise in better understanding the biology of NFAT activation, but likely need tethering to either the calcium channels or NFAT proteins themselves or a specific cellular compartment to increase spatial resolution; a whole-cell approach is no longer sufficient for the proper understanding of NFAT activity in this context and finer resolution would greatly aid in the identification of future mechanisms. Given findings from Kar and Parekh (2015) on the importance of nuclear calcium in regulating the activity of at least NFATC3, it would be beneficial to have new tools to monitor these alterations in real time.

An initial approach would take advantage of some of the new technologies available in the zebrafish (reviewed in Subsection 2.5.2) by generating an endogenously GCaMP-tagged *nfatc2a* in the zebrafish to monitor the timing of calcium flashes and

how those correspond to the activation state of the protein, able to be visualized by intercompartmental trafficking. By using high temporal and spatial resolution imaging offered by LightSheet (Reynaud et al., 2008), it would be possible to monitor on the seconds resolution the activation status of Nfatc2a within macrophages in response to mycobacterial infection. This finely tuned reporting behavior would reveal key details about the timing and kinetics of NFAT activation during infection and how that activation corresponds to the induction of *vegfaa* and angiogenesis.

In the long-term, it is likely to be worth exploring the possibility of generating a floxed or gene-trapped *nfatc2a* allele to allow study of this protein in a particular cellular context. This, paired with an *irg1:Cre^{ERT2}* line would provide the ability to tissue-specifically ablate or reconstitute Nfatc2a activity to more directly study this pathway's contributions to infection. While the use of VIVIT allows for gross examination of NFAT-dependent phenotypes, it would be beneficial to be able to study individual isoforms' contributions to given phenotypes. This is technically already possible but is rather laborious with current technologies. Should newer approaches to the generation of knock-in zebrafish lines emerge or these studies become a greater priority, this should be a foremost consideration in the continuation of this work.

Such a tool either on its own or by contrast with the *irg1:VIVIT* line should be analyzed by RNA-seq during mycobacterial infection to capture a comprehensive profile of the transcriptional changes resulting from inhibition of NFAT within macrophages. Bulk RNA-seq approaches are likely appropriate for this task and could be sourced from either macrophages in the granuloma from adult zebrafish or from macrophages in the early stages of granuloma development in the larva. This would provide an essentially comprehensive profile of the NFAT-dependent responses in these cells *in vivo* likely to also be a useful resource for the community at large in dissecting criti-

cal pathways for defense against mycobacteria and designing therapeutic options that properly balance the need to inhibit deleterious responses while enhancing protective ones.

It will also be interesting to start teasing out non-macrophage cellular contributions to angiogenesis. While we have already seen that depletion of macrophages reduces the angiogenic phenotype in response to mycobacterial infection (Oehlers et al., 2015), it is not known if other cells types may be able to compensate at other stages of infection. I have developed a *lyz:VIVIT-tdTomato* transgenic line that would allow for the functional dissection of these roles within neutrophils, in which NFAT may be playing distinct roles during infection, as has been described in other contexts (Vega et al., 2007; Herbst et al., 2015; Vymazal et al., 2021). Preliminary experiments suggest that this line does not alter angiogenesis after TDM injection, so other roles in the immune response are likely to be more relevant in this line (Figure 3.17). This would offer a new horizon of NFAT contributions during infection that could serve as an entire new Ph.D. project.

To more directly study the contributions of macrophages alone, however, we could apply the *irf8^{st95}* fish line, which lacks macrophages due to loss of a transcription factor essential for macrophage development, to study these processes in the larva and the adult²² (Tamura et al., 2005; Xu et al., 2012; Shiau et al., 2015). Ambitious approaches would incorporate reconstitution of the macrophage population in an otherwise wild-type *irf8^{st95/st95}* fish with *nfatc2a^{-/-}* macrophages to more precisely identify the requirement for these cells in angiogenesis. A slightly simpler approach would adopt the strategy used by Cronan et al. (2021) to conduct whole kidney mar-

²²Although it was originally assumed that the Mpeg1⁺ cells in the adult zebrafish were macrophages that developed independently of Irf8, it is now thought that these are actually B cells, indicating that survival without macrophages is possible in the zebrafish, although the fish are rather sickly and are generally sub-Mendelian in adulthood (Ferrero et al., 2020).

row transplantation into *myb* mutant larvae, reconstituting the entire hematopoietic system with *nfatc2a* mutant cells. Any of these approaches should allow for deeper study of macrophage NFATC2 biology in the zebrafish model through studies in both the larva and the adult, which could reveal both expected and unexpected tissue-specific roles for this gene in the process of pathological angiogenesis and host immunity.

5.5 Mycobacterial Interactions with Other Aspects of Vascular Biology

The focus of the present work has been on a rather coarse readout of vascular biology: angiogenesis or lack thereof. However, as was discussed briefly in Section 1.5, the biology of the endothelium is far more complex than just sprouting angiogenesis and the interaction of the granuloma with the vasculature is more complex as well. This makes it imperative to develop a more nuanced understanding of how these other factors may contribute to the pathology of the disease and how NFAT signaling may modulate these factors as well.

A radical approach that may be of some utility in addressing some of these more challenging questions would require the development of a new model wherein *ex vivo* cultured granulomas are co-cultured with endothelial cell lines to start addressing some of the contributions of different genetic factors to the ability of the granuloma to alter endothelial cell biology more directly. While it is known that human umbilical vascular endothelial cells (HUVECs) can respond to zebrafish Vegfaa, it may be worthwhile to explore a more native model. Endothelial cell lines from other fish species have been established and it could be of great utility to explore the possibility

of establishing something similar for the zebrafish (Luque et al., 2014; Pham et al., 2017). A genetically tractable system for generating these lines would allow for both granuloma-derived signals and endothelial responses to be measured in a more tightly controlled and readily quantified environment. Given many of these objectives involve more cell biology than simply the growth of vessels, the ability to observe them in greater isolation may prove necessary if these are to be effectively addressed.

5.5.1 Vascular Permeability

In addition to the overall growth of blood vessels, VEGFA has important roles in maintaining the integrity of blood vessels both through weakening integrin-mediated connections between neighboring cells and through further signaling to the neighboring mural cells, including pericytes, to modulate the access of vascular luminal contents to neighboring tissue in the direction of the source of VEGFA (Claesson-Welsh, 2015; Park-Windhol and D'Amore, 2016). However, VEGFA is not the only chemokine capable of modulating the integrity of the endothelium; another, related pathway known as the angiopoietin system is also known to play a major role in maintaining the structure of the endothelium and preventing (or encouraging) vascular leakage which may be critical for certain developmental events as well as effective immune responses.

A previous study from the Tobin lab found that angiopoietin signaling was a critical mediator of vascular permeability during mycobacterial infection, a finding consistent with that of tumors, where ANG2 signaling induces alterations in the biology of the endothelium through the Tyrosine kinase with Ig and EGF homology domain 2 (TIE2) pathway (Goel, Wong, and Jain, 2012; Thurston and Daly, 2012; Oehlers et al., 2017; Duran et al., 2021). Inhibition of angiopoietin signaling resulted in a

reduction in bacterial burden and improved vessel normalization, likely in part by blocking the function of VEGFA signaling. However, more recent findings have noted that granuloma macrophages do not seem to be a major source of angiopoietins²³, which creates a bit of a conundrum: if we know that these proteins are produced during infection and play a role in the overall course of disease, where are they coming from (Cronan et al., 2021)?

Angiopoietin signaling is a complex affair, with multiple ligands and receptors, but the focus is centered on the bifunctional ligand, angiopoietin-2²⁴ (ANGPT2 or ANG2) and the receptor TIE2²⁵. While at homeostasis and at some dosages ANG2 can *activate* TIE2 to promote vascular stability, in the context of inflammation, it becomes antagonistic through combinatorial effects with various cytokines and induces vascular permeability (Augustin et al., 2009). ANG1 is the standard agonist of this pathway and the activity of ANG1 maintains vascular integrity and prevents the expression of inflammatory integrins (Augustin et al., 2009). Leakiness induced by ANG2 results in a deregulated vascular environment that promotes tumor and, seemingly, bacterial growth; normalization with antibodies induces vascular regression and exerts anti-tumor effects. This also implicates ANG2 as an important mediator of angiogenic processes in concert with VEGFA. It is known that endothelial cells can produce their own ANG2 from Weibel-Palade bodies to signal in an autocrine and paracrine manner as part of normal vascular biology (Fiedler et al., 2004). Addi-

²³This is at least true of the 14 days post infection time point at which the samples were taken and similar findings can be found from Gideon et al. (2022) at four and ten weeks post infection.

²⁴Angiopoietin-1 is the main ligand during development and homeostasis, where knockout of ANG1 recapitulates the phenotypes of TIE2 knockout, but during pathology, alterations in ANG2 levels are the primary focus (Akwii and Mikellis, 2021). There are also a range of angiopoietin-like proteins that serve diverse functions that may be of interest but are target for far future investigation in this direction (Hato, Tabata, and Oike, 2008).

²⁵TIE1 primarily plays a complementary role with TIE2 as a heterodimer – this is a bit beyond the present topic but for further reading, Savant et al. (2015) and La Porta et al. (2018) have excellent studies on how TIE1 contributes to angiogenesis in development and cancer.

tionally, stromal cells in nearby tissues can respond to a cytokine environment and induce the expression of ANG2 (Hato, Tabata, and Oike, 2008; Huang et al., 2010; Thurston and Daly, 2012). This provides neighboring cells and the endothelium itself an important role in regulating endothelial biology and which may be at play in the context of the granuloma.

If nearby fibroblasts, pericytes, or the endothelium itself are producing ANG2, as would be suggested by Gideon et al. (2022) and paralleling tumor biology, then we are missing out on crucial aspects of stromal-granuloma interactions in the dataset from Cronan et al. (2021). What gene networks might be regulating these processes and are macrophages a major source of (a) key inducing cytokine(s) (Augustin et al., 2009)? These are difficult questions to address with existing tools and the gene duplication of the zebrafish presents fresh difficulties. Others in the lab have developed CRISPR-Cas9 mutations in zebrafish *angpt2a*, *angpt2b*, and *tie2*, which may be useful in dissecting some of these effects. Studies using these lines and, potentially, the development of novel *angpt2* transgenic reporters or application of broader microdissection approaches and further scRNA-seq would help to narrow on the cellular identity of ANG2-producing cells in the granuloma and how modulation of those cells could offer an alternative pathway toward vascular normalization in the granuloma either independent of or in concert with VEGFA-targeting therapies. As we have seen with bevacizumab (Subsection 1.5.2), it will likely be necessary to develop a multi-pronged approach to vascular targeting and a more thorough study of this pathway during mycobacterial infection would constitute a major contribution in that direction (Huang et al., 2010; Saharinen, Eklund, and Alitalo, 2017). An unlikely, but plausible, hypothesis would be that NFAT in macrophages is regulating the production of a cytokine or primary messenger that is stimulating ANG2 production by fibroblasts, pericytes, or the endothelium. Such an intercellular signaling network

would be a fascinating new aspect of granuloma angiogenesis biology worth further exploration. Despite the great deal that is known about the roles of ANG2-TIE2 signaling during cancer, it is not clear if there are discrete stimuli that are able to upregulate ANG2 expression or secretion by the endothelium and identification of such may serve as a further upstream mechanism of host-directed intervention in various diseases (Huang et al., 2010). Interestingly, the angiopoietin-TIE2 system also functions in and is important for lymphatic development, a topic that will be returned to in Section 5.8 (Eklund, Kangas, and Saharinen, 2017).

5.5.2 Pericytes

Pericytes are critical stromal cells that support various aspects of vascular cell biology. These cells are derived from myeloid and mesenchymal precursors and express a number of different pattern recognition receptors, making them important in regulating vascular inflammation and, potentially, vascular responses to infections (Stark et al., 2013; Yamazaki and Mukouyama, 2018). Despite these important functions in maintaining the vascular barrier, limiting vascular permeability, regulating angiogenesis, and more, essentially nothing is known about their contributions to tuberculosis (Bergers and Song, 2005). In addition, there are unusual subpopulations, especially within the brain, that have macrophage-like functions while remaining affiliated with the vascular endothelium; these cells likely conduct interesting immune processes about which we know essentially nothing (Balabanov et al., 1996; Venero Galanternik et al., 2017). Slightly more is known about their function in cancer, which may establish a model for our understanding in infectious disease.

Within tumors, pericyte coverage of vessels is highly variable and almost universally perturbed and pericytes are thought to play a bimodal role where high peri-

cyte coverage fosters primary tumor growth while low pericyte coverage facilitates metastasis (Ribeiro and Okamoto, 2015; Saharinen, Eklund, and Alitalo, 2017). Additionally, low pericyte coverage of blood vessels is associated with increased ANG2 expression, suggesting that pericyte-endothelial contacts or paracrine signaling may be important for limiting vascular permeability in addition to any physical role these cells may have in preventing vascular leakage. Because pericytes do not depend on VEGFA signaling for their maintenance, they are often left behind in the aftermath of anti-angiogenic therapy, where they are able to serve as a path for vascular regrowth after withdrawal of therapy (Keskin et al., 2015; Ribeiro and Okamoto, 2015). This suggests that a combination of anti-angiogenic therapy and pericyte-targeting therapy might be able to achieve more complete and sustained vascular regression.

The targeting of pericytes also offers another approach to understanding the function of vascular leakiness in influencing granuloma biology as others have demonstrated that pericytes are critical mediators of vascular integrity and that abolition of these cells can actually improve peri-tumor vascular normalization and restrict tumor growth (Keskin et al., 2015). Pericytes and the endothelium also communicate through a variety of immunological pathways, including inflammasome activation and IL-1 β signaling, making their interactions able to be perturbed by true inflammatory signals during disease, likely as an additional means of effectively responding to these signals (Kozma et al., 2021).

Other types of mural cells also exist that serve to support the vasculature and these may be of interest to study as well. For instance, Bower et al. (2017a) found that lymphatic vessels that run along blood vessels regulate their elaboration during vascular development, suggesting that crosstalk between these two critical circulatory systems may be important for the overall progression of angiogenesis in disease as well.

These networks, in addition to populations of macrophage-like pericytes, demonstrate that there is much left unknown about the vascular system and a more comprehensive spatial and transcriptional profiling of the endothelium and its supporting tissues in various disease states is needed to fully understand how the mural cells enveloping the vasculature respond to various stimuli and the consequences of these responses (Balabanov et al., 1996; Stark et al., 2013; Yamazaki and Mukouyama, 2018; Kozma et al., 2021).

Luckily, there already exist tools within the zebrafish that can allow for preliminary dissection of some of the roles of pericytes during mycobacterial infection. The TgBAC(*pdgfrb:mCitrine^{s1010}*) zebrafish line labels pericytes throughout development and may allow us to visualize some aspects of their biology using CLARITY techniques in the adult zebrafish granuloma as described in Section 2.6 (Vanhollebeke et al., 2015). Alterations in pericyte localization, vascular engagement, or even interactions with and within the granuloma itself may suggest an important role for these cells. Additionally, little is known about the contributions of the signaling molecules that these cells utilize for their normal biology and how those may influence disease. Use of genetic mutations in *pdgfrb* (the major signaling molecule that induces pericyte differentiation and proliferation) or tissue-targeted ablation approaches (*pdgfrb:nitroreductase*²⁶ or similar) may offer a look at the impact these cells have on the overall development of tuberculosis disease and their contributions to angiogenesis. Of all the as-yet unstudied stromal cells likely to influence the progression of disease, these strike me as the most interesting population given their heterogeneity and known functions in cancer.

²⁶Nitroreductase is a bacterial enzyme that reduces metronidazole into a toxic byproduct that kills cells and can be used to ablate specific cell populations, although this clearly has the potential to induce some immune responses to all of the dead cells (Sharrock et al., 2021).

5.5.3 Extracellular Matrix

Although this dissertation has not particularly addressed the biochemistry of the focal chemokine – VEGFA – it is an intriguing protein with multiple layers of regulation, including the existence of multiple splice variants. The two major ones, VEGFA¹²¹ and VEGFA¹⁶⁵, have distinct capacities for binding to the extracellular matrix (ECM), with VEGFA¹⁶⁵ binding to heparin while VEGFA¹²¹ freely diffuses through ECM (Shibuya, 2011; Fearnley et al., 2016). The combination helps to maintain gradients of chemokine able to be followed by both the endothelium and immune populations toward the productive source. These differences may determine different aspects of the overall angiogenic response and may offer an additional mechanism for subversion by the bacteria through engagement with the extracellular matrix.

Tetranectin

An interesting happenstance in my early graduate school career was a few months spent on the study of the zebrafish gene *clec3ba*, which is one of the two zebrafish homologs of the human *CLEC3B*, which encodes for tetranectin. Tetranectin is a plasminogen²⁷-binding protein produced by macrophages with very few described roles (Nielsen, Clemmensen, and Kharazmi, 1993). As implied by the gene name, tetranectin is a C-type lectin which we had naïvely thought might be a candidate for the MINCLE homolog based on the expression pattern by RNA-seq. This protein is able to block the binding of plasmin to the extracellular matrix, which reduces the rate of fibrin degradation (Mogues et al., 2004). Because so little is known about the

²⁷Plasminogen is an essential component of the fibrin-degradation pathway that clears fibrin clots. The balance of fibrin deposition and plasmin degradation determines the hemodynamics of clot formation. Fibrinogen is one of the many targets of aspirin that allows it to effectively thin the blood to prevent heart attacks (Bjornsson, Schneider, and Berger, 1989; Keragala and Medcalf, 2021).

biology of tetranectin aside from its structure (it exists as a trimer), some basic details about its role in plasminogen binding, and its potential as a biomarker for various cancers, this could offer a really novel opportunity to study something unexpected in the pathogenesis and angiogenic response to mycobacterial infection (Holtet et al., 1997; Nielsen et al., 1997; Graversen et al., 1998). One of the only modern papers I could find on this gene found that expression of tetranectin is protective during sepsis but that excessive endocytosis of tetranectin by macrophages resulted in increased pyroptosis and exacerbation of inflammation (Chen et al., 2020). This suggests that tetranectin can exert anti-inflammatory functions during this disease and could be a promising target for agonism during sepsis.

In my work on this gene, I found that disruption of *clec3ba* by CRISPR/Cas9 reduced the amount of angiogenesis in response to TDM injection in the larval zebrafish in a manner nearing statistical significance. While we realized that we were barking up the incorrect metaphorical tree in pursuit of the zebrafish MINCLE, these results are suggestive evidence that this gene may be playing a role in mediating angiogenesis, potentially in a broader sense but perhaps restricted to TDM or mycobacteria. In the search for non-VEGFA-dependent mechanisms to inhibit pathological angiogenesis, more active pursuit of these sorts of observations may provide new drug targets to either block or enhance angiogenesis.

Plasminogen

Given tetranectin's role in binding plasminogen, it begs the question of whether plasminogen itself may have some important role in mediating mycobacteria-induced angiogenesis. Previous studies had found that plasminogen knockout mice had reduced angiogenic responses to VEGFA, implicating plasmin degradation of fibrin in facilitating vessel growth (Oh, Hoover-Plow, and Plow, 2003). However, loss of plasminogen

results in fibrosis as there is no longer an effective mediator to degrade these fibrin deposits. Given that mycobacteria bind and activate plasminogen using alpha-enolase, it seems logical to expect that this protein serves pro-bacterial functions (Monroy et al., 2000). Indeed, interaction with plasminogen inhibits phagocytosis, but this does not properly address the role of this pathway in the granuloma (Echeverria-Valencia et al., 2019). To address this, it will be useful to generate knockout zebrafish able to model loss of function in plasminogen and fibrinogen. The role of fibrin and fibrin degradation in the granuloma is generally poorly understood so uncovering new roles for these hemodynamic processes is likely to reveal novel mediators of angiogenesis and host defense against mycobacteria.

The target of plasminogen, fibrin, also plays an important role in mediating the cellular effects of TDM. TDM bound to fibrinogen inhibits cellular necrosis and promotes the formulation of the granuloma whereas in the absence of fibrinogen, extensive cellular necrosis is able to occur (Sakamoto et al., 2010). This would suggest an immunosuppressive or immunomodulatory effect of fibrin on TDM²⁸; in the absence of plasmin to degrade fibrin, these effects may be magnified and angiogenesis further reduced. If that is the case, then local blockade of plasmin in the granuloma could be an interesting experimental approach to altering the granuloma microenvironment to be more hostile to bacterial growth. Some of these interactions have begun to be studied in recent years with a study from Hortle et al. (2019) demonstrating that fibrinogen mutant zebrafish had improved bacterial control due to mycobacterial induction of coagulation, a process known to be induced by TDM (Retzinger et al., 1982; Retzinger, 1987; Donnachie, Fedotova, and Hwang, 2016). It is possi-

²⁸These known interactions between TDM and fibrin are an additional confounding factor in *in vitro* studies of TDM responses; the goal should be to most accurately model the *in vivo* condition and the lack of these cofactors in pure TDM stimulation makes it difficult to accurately assess the relevance of types of responses to actual infection.

ble that, depending on the degree of fibrin deposition, various effects can be exerted where insufficient fibrionogenesis may be neutral or neutral-positive while insufficient fibrinolysis may offer benefits as well (Venkatasubramanian et al., 2016). These interactions, especially those between platelets and macrophages, warrant further study and it will be interesting to dissect plasmin/fibrin interactions as aspects of granuloma formation, the granuloma microenvironment, and angiogenesis (Feng et al., 2014; Lugo-Villarino and Neyrolles, 2014).

5.5.4 Other Angiogenic Stimuli

Vascular biology is vastly complex and there are many potentially influential angiogenic stimuli. As has already been discussed, Torraca et al. (2017) found a critical role for CXCR4 signaling in inducing angiogenesis in response to mycobacterial infection, but determining the signaling underlying CXCL12 expression from macrophages would certainly be of some interest. If NFAT is mediating the production of CXCL12, then this may offer an opportunity to intervene in a broadly pro-angiogenic pathway to exert host benefit by multiple mechanisms against which the pathogen may have little recourse.

Another critical pro-angiogenic signaling pathway is driven by the expression of TGF- β . Transforming growth factor beta is produced by macrophages and other immune cells, but also exerts autocrine functions that facilitate the upregulation of angiogenic signals, including VEGFA, as well as directly influencing angiogenesis by priming endothelial cells (Jeon et al., 2007; Ferrari et al., 2009; Goumans, Liu, and Dijke, 2009). Given the known roles for NFAT signaling in mediating the downstream responses to TGF- β , it is of interest to explore the role of this somewhat mysterious cytokine in regulating macrophage-endothelial crosstalk and potentially

important roles upstream of VEGFA in priming endothelial cells for proliferation (Cobbs and Gooch, 2007). Engoglin is a part of the TGF- β receptor complex and is a transmembrane protein that is induced in endothelial cells during inflammation to mediate the responses of the endothelium to TGF- β and other related cytokines and is thought to mediate certain aspects of blood vessel diameter and maturation after VEGFA-mediated angiogenesis (Liu et al., 2014; Sugden et al., 2017). This pathway thus makes a compelling case as being potentially dysfunctional during granuloma angiogenesis and may be an additional target that can facilitate vascular perfusion and normalization to improve drug delivery.

Endothelins are short peptides that regulate the vasculature that have previously been shown to play a role in tuberculosis. *M. tuberculosis* secretes an enzyme that liberates endothelin from the pro-peptide to allow for active signaling and this enhances bacterial growth (Correa et al., 2014). While this was originally proposed to be mediated by macrophage recognition of active endothelin, an alternative hypothesis is that this peptide is being activated by *M. tuberculosis* to modulate aspects of vascular biology, either through endothelin's roles in vasoconstriction or promoting endothelial survival. Such functions are comparatively understudied relative to VEGFA and may play important roles in mycobacterial subversion of the endothelial system independently of or in synthesis with TDM.

A final point is the potential role for novel modes of cellular differentiation to play a role in modulating host angiogenesis. Studies in development and cancer have identified mesenchymal to endothelial transitions that allow for the direct incorporation of mesenchymal stem cells and macrophages into the structure of the endothelium itself whereby these cells adopt defining characteristics of the endothelium, including expression of vascular endothelial cadherin (Wu et al., 2007; Zhang et al., 2008a;

Ubil et al., 2014). The contribution of the diverse types of macrophages within the granuloma, including those expressing both VEGFR1 and TIE2, to the vasculature is largely unknown, but the incorporation of mycobacterially subverted macrophages into the vasculature may be a further mechanism of coopting this system for the benefit of the invading bacteria (Hall et al., 2012; Cortie et al., 2014). Many further studies would be required to address this, but longer-term fate mapping of *flt1*⁺, *irg1*⁺ cells may open some possibilities for evaluating this process.

5.6 Precise Contributions of VEGFA Signaling to Mycobacterial Burden

Despite our knowledge that angiogenesis through VEGFA appears to increase bacterial burden, the precise mechanism by which this happens is unknown. Conceptually, these alterations could be through promotion of bacterial growth or inhibition of bacterial killing and our current knowledge is unable to definitively identify one, the other, or both of these mechanisms as contributing to the overall effect. Our oft-stated assumption is that this is mediated by the provision of additional oxygen and nutrients to the bacteria, but the identity of these “nutrients” is not clear. This is not the only possible hypothesis for what is happening, however, and dissecting other possibilities is important for clarifying what, precisely, is occurring at the vasculature-granuloma interface.

If oxygenation is truly the mechanism of enhanced bacterial growth, one would hypothesize that the induction of angiogenesis under artificial hypoxia in the larvae would result in no change in bacterial burden. This can be tested by use of a hypoxic chamber, in which the oxygen can be artificially lowered to 5% while sustaining larval

zebrafish survival (Rombough and Drader, 2009; Long et al., 2015). If oxygen is the primary driver of increased bacterial burden after angiogenesis, this effect should be nullified by culture under hypoxia. Conversely, if oxygen is not the primary driver, increased bacterial burden should be sustained under hypoxia. These results would be informative as to the nature of the growth of these blood vessels and their effects on mycobacterial growth.

If, on the other hand, these effects are mediated by the delivery of some nutrient, it may be necessary to devise some narrow list of potential solutes that are being delivered and devise methods to sequester them. Various vitamins and minerals are likely candidates and depleting the larval zebrafish of these either genetically or pharmacologically may reveal their contributions to mycobacterial growth in an angiogenesis-dependent fashion. If, however, none of these reveal an alteration in bacterial burden, an alternate hypothesis might be that either (a) the activity of VEGFA on the vasculature is incidental to the overall effect, which is actually mediated by changes in macrophage biology or (b) the increased angiogenesis facilitates recruitment of pathogen-permissive immune cells that ultimately increased bacterial burden through either inhibition of bacterial killing or facilitation of bacterial growth. These hypotheses are not mutually exclusive and teasing them apart will require careful experimental design.

To begin, macrophage-specific genetic approaches are required that inhibit the activity of VEGFR1 and VEGFR2 on macrophages, which are known to express these receptors. Genetic inhibition of these pathways through the use of dominant-negative approaches in the macrophage would allow for continued angiogenesis by the endothelium while inhibiting the paracrine effects of VEGFA production on the macrophages themselves (Stratmann et al., 2001; Tsou et al., 2002). If macrophage-

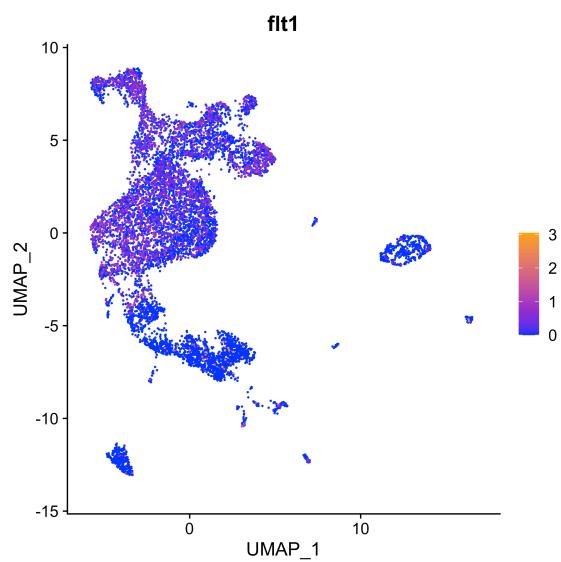


Figure 5.12: While there is minimal expression of *krd* and *kdrl* within granuloma macrophages, there is prominent and widespread expression of *flt1*, which is the gene encoding VEGFR1. This suggests that this VEGFR signaling pathway may be active within these macrophages, but the precise contributions of this pathway to the response to infection remains unknown. Development of new tools for the study of this pathway is likely to reveal important roles for VEGFR1-dependent signaling in shaping the immune response to mycobacterial infection.

specific VEGFR blockade results in decreased bacterial growth, this would suggest that the effect seen through inhibition of VEGFA is actually mediated by macrophages while the angiogenesis effect seen is purely circumstantial and a convenient readout of VEGFA signaling activity in the proximity of the granuloma. If this is the case, then *in vitro* approaches can also be taken by treating *VEGFA* knockout THP-1 or murine bone marrow-derived macrophages with VEGFA and assessing their capacity for bacterial killing and measuring bacterial replication within these macrophages. Reduction in bacterial killing after VEGFA treatment would indicate that VEGFA-dependent signaling on macrophages polarizes them into a more permissive state (Murakami et al., 2006; Murakami et al., 2008; Dineen et al., 2008; Lai et al., 2019; Okikawa et al., 2022); additionally, the rate of bacterial growth within each macrophage can be measured by live imaging to glean some insights as to the locus of activity – early killing would indicate more efficient phagosomal-lysosomal fusion whereas later effects might indicate some influence on the ability for macrophages to undergo apoptosis or efferocytose other dead macrophages containing mycobacteria. Further development of tools in bacterial genetics to study the particular effects on the bacteria – through lysosomal acidification, increased bacterial stress, cell death, or other effects – could offer additional tools for the study of these processes as a result of VEGFA-dependent macrophage reprogramming (McCann et al., 2007; Carroll et al., 2010; Abramovitch et al., 2011; Simeone et al., 2012; Tan et al., 2013; Sukumar et al., 2014; Abramovitch, 2018; Hayashi et al., 2018). In particular, use of mycobacterial cell death reporters could reveal alterations in rates of bacterial killing within the macrophage, providing a direct and quantifiable readout of this particular effect (Nkwouano et al., 2017; MacGilvary and Tan, 2018).

Testing the effects of macrophage VEGFR signaling on macrophage recruitment and behavior could be done either *in vivo* or *in vitro*, with a transwell assay per-

haps being the most compelling option. Forcing macrophages to travel along a VEGFA gradient toward a Matrigel plug of mycobacteria would reveal alterations in bactericidal capacity after stimulation with VEGFA along a gradient and mimic certain aspects of macrophage recruitment to the nascent granuloma. This could also be done *in vivo* by injecting $\Delta pcaA$ mycobacteria into the trunk along with recombinant Vegfaa and assessing effects on mycobacterial growth and macrophage behavior. Increased intracellular replication within Vegfaa-stimulated early macrophages would indicate that they have reduced bacterial killing capacity; increased recruitment might indicate that this is a mechanism by which the bacteria can drive recruitment of more permissive cells, similar to CCL2 (Cambier et al., 2014; Cattin et al., 2015). While this will require a great deal of further experimentation to effectively delineate, defining the actual role of VEGFA-VEGFR signaling in enhancing bacterial growth is essential to understanding what aspects of this axis are most fertile for host-directed intervention. Notably, granuloma macrophages generally lack expression of the VEGFR2 homologs in the zebrafish, but display prominent expression of VEGFR1 (Figure 5.12) (Cronan et al., 2021). This suggests that it may be possible to interfere with macrophage VEGFR signaling by targeting this single gene, which is known to be heterozygous viable in the mouse and has several available reagents in the zebrafish (Habeck et al., 2002; Thirunavukkarasu et al., 2007; Krueger et al., 2011; Ho and Fong, 2015; Rossi et al., 2016; Wild et al., 2017). Even reporter-based approaches through existing reporters would be of interest to understand the behavior of *flt1*-expressing macrophages in the granuloma (Krueger et al., 2011). It is certainly possible that the endothelium itself is somewhat incidental in this process and these effects are predominantly mediated by macrophages but, to date, the relative contributions are unknown.

5.7 VEGFR2-Dependent Signaling Pathways in the Endothelium

Despite the exhaustively detailed role for VEGFA-VEGFR2-dependent signaling in contributing to pathological angiogenesis during mycobacterial infections, the endothelial response pathways remain ambiguous. Given the diverse pathways that are induced after VEGFR2 agonism that mediate critical aspects of the overall angiogenic response, it is of some interest to develop a more lucid understanding of which pathway(s) within the endothelium are transducing these responses and their contributions to the overall angiogenic effect (Abhinand et al., 2016). These more discrete pathways may offer additional targets to block angiogenesis that are more difficult to compensate for than blunt VEGFR2 inhibition and will also shed further light on the specific pathways required during pathological angiogenesis, which may differ in meaningful ways from those pathways required for developmental angiogenesis.

A premier candidate for mediating the angiogenic effects during infection is the ERK pathway, which is one instance of the broader MAPK family (Shibuya and Claesson-Welsh, 2006; Wang et al., 2020b). There are a diverse set of tools available to study the activity of this pathway in the angiogenic sprouts, including sensitive ERK biosensors that have recently been published for use in the larval zebrafish (De Simone et al., 2021). Driving the expression of an ERK kinase translocation reporter from a vasculature-specific promoter, such as *kdr1* (also known as *flk1*) would allow for the fine visualization of the induction of ERK in these angiogenic processes, which can then be inhibited either pharmacologically with MEK inhibitors or with a vasculature-specific inducible dominant-negative ERK (Izumi et al., 2001). This would reveal any potential role for ERK signaling in the induction of angiogenesis during mycobacterial

infection. While this role is not inherently specific to pathological angiogenesis, it would offer a starting point for further exploration.

If it is the case that ERK signaling is dispensable for the induction of pathological angiogenesis, this should lead to the exploration of other pathways in a less biased fashion. To do so, I would propose microdissection of granulomas along with the granuloma-associated vasculature from adult zebrafish at earlier time points in infection (ten to twelve days post infection), to allow for the maximum pro-angiogenic signals to be captured. These cells can then be sorted for *kdr1:eGFP* signal and processed for proteomics, to compare to the nuclear and cytoplasmic compartments for the relative expression of various transcription factors in these compartments and compared to quiescent vasculature. The relative enrichment of particular transcription factors in the nuclear compartment might indicate that these factors are important for the angiogenic response and should be validated genetically through either knockout or dominant negative approaches. This has the potential to reveal pathological- or mycobacterial-specific pathways that might be more targeted to the vasculature and minimize side effects from broad angiogenic inhibition; it is also likely to reveal additional layers of mycobacterial subversion of host stromal responses important for the overall pathogenesis of tuberculosis.

5.8 Lymphangiogenesis

This project has focused strictly on the proliferation and growth of vascular endothelial cells during mycobacterial infections, a process known as (hem)angiogenesis. However, a parallel process exists concerning the lesser studied lymphatic vascular system and this is known as lymphangiogenesis. Lymphangiogenesis is known to play important roles in the progression of other infectious diseases, but its roles in

mycobacterial infection are comparatively unknown (Bowlin et al., 2021). The evolutionary segregation of blood and lymph circulation has led to distinct but very closely related mechanisms of regulation during development and pathology, which suggests a potential role for this pathway during tuberculosis infection as well. While this process is less extensively studied, it is a critical component of the vascular development of vertebrate organisms and is essential for proper fluid homeostasis and immune system function, as lymphatic vessels are the route along which antigen presenting cells “drain” into the lymph nodes to prime the adaptive immune system. Failures of lymphangiogenesis result in lymphoedema and general problems with fluid balance (Makinen et al., 2001). In the context of disease, the lymphatic system is essential for proper immune behavior, but during chronic conditions like cancer or tuberculosis, lymphangiogenesis can be subverted by the insult for their own benefit. In cancer, lymphangiogenesis is utilized to provide additional routes of metastasis away from the primary tumor; the role in tuberculosis is comparatively less well studied (Augustin et al., 2009; Huang et al., 2010; Duong, Koopman, and Francois, 2012; Stacker et al., 2014) and, indeed, dissemination in tuberculosis is still a growing field with many of the host-microbe interactions that facilitate this phenomenon still under extensive study (Saelens et al., 2022).

Lymphangiogenesis is primarily mediated through an analogous set of ligand-receptor pairs to that seen in hemangiogenesis. VEGFC and VEGFD²⁹ are the major chemokines that bind VEGFR3 on the lymphatic endothelium to stimulate lymphangiogenesis (Alitalo, Tammela, and Petrova, 2005; Kuchler et al., 2006; Haiko et al., 2008), a process which depends on functional ERK signaling, much like hemangiogenesis (Shin et al., 2016; Shin et al., 2017). During development, a portion of the

²⁹At least VEGFD acts non-redundantly and is also able to stimulate hemangiogenesis in the zebrafish and, given the findings presented in Subsection 1.5.1, both of these likely to play roles in other angiogenic processes (Bower et al., 2017b).

cardinal vein differentiates into lymphatic progenitors (Yaniv et al., 2006; Nicenboim et al., 2015) and begins to elaborate into a full lymphatic system by approximately 3 days post fertilization in the larval zebrafish; these budding vessels are guided by arterial cues, which allows much of this system to parallel the blood endothelial system (Bussmann et al., 2010; Jung et al., 2017). This process ultimately leads to a complete lymphatic system able to drain excess fluid and remove insults from within tissues, although zebrafish are not known to possess lymph nodes as mammals do and the primary sites of antigen presentation lie elsewhere (Korbut et al., 2016; van Lessen et al., 2017; Onder et al., 2017).

During cancer growth, the tumor itself and the tumor associated macrophages that surround it both contribute to lymphangiogenesis, which provides a means of dissemination for the tumor to distal body sites. Lymphangiogenesis and VEGFC/D can facilitate metastasis in the tumor although it is not clear whether this mechanism operates solely through induction of lymphangiogenesis or if VEGFC/D can act directly on the tumor itself, analogously to VEGFA autocrine activity (Stacker et al., 2014). The primary site of initial metastasis – to the lymph node – is a symptom of the lymphatic involvement in this disease and paralleling the dissemination patterns seen in tuberculosis (Ganchua et al., 2020). Of course, it was conceptually possible that the lymphatic endothelium was a bystander but it is clear that the lymphatic system actively remodels in response to tumor signals. It has since been demonstrated that VEGFC overexpression by cancer cells in the tumor drives increases lymphangiogenesis and early lymph node metastasis, implicating active manipulation on the part of the tumor in this process (Duong, Koopman, and Francois, 2012). Several different tumors also overexpress the lymphangiogenic effector CCBE1, which aids in processing VEGFC into its active form to potentiate lymphatic growth (Hogan et al., 2009; Le Guen et al., 2014; Li et al., 2018a; Zhao et al., 2018; Song et al., 2020).

Tumor-associated macrophages have been linked to this process through tumor-directed activation of the inflammasome that drives IL-1 β processing to increase lymphangiogenesis (Weichand et al., 2017). This interesting mechanism provides the tumor multiple means of inducing a full spectrum of different signals that facilitate its spread. Macrophages more broadly also play key roles in the induction of inflammatory lymphangiogenesis, also partially mediated by IL-1 β as well the induced expression of VEGFC and VEGFD during insult. Some sort of systematic promoter comparison, either computationally or *in vivo*, would help to decipher the regulatory mechanisms for each isoform. After tissue damage, it is critical to direct the growth of new lymphatic vessels toward the injury in order to facilitate drainage and immune access, but excessive lymphatic inflammation is harmful. An additional mechanism that macrophages can use to induce lymphangiogenesis is through IL-33 production, which induces a complex intracellular cascade that include nitric oxide signaling to drive inflammation (Han et al., 2017), which is protective against tuberculosis (Villarreal, Siefert, and Weiner, 2015; Pineros et al., 2017). Similarly, TNF- α has been shown to induce lymphangiogenesis by stimulating production of VEGFD³⁰ (Hong et al., 2016). Conveniently, the zebrafish is able to be utilized to model many of these processes in inflammatory lymphangiogenesis, allowing us to adapt it to the study of these processes in mycobacterial infection (Okuda et al., 2015).

To date, there have been two major sets of studies on the role of the lymphatic system in mycobacterial infection, which is a relative indictment on the attention that has been paid to this potentially significant contributor to disease. The first, from Harding et al. (2015), examined this pathway through chemical inhibition to block VEGFR3 activity. This study found that reduction of lymphangiogenesis re-

³⁰Could it be through NFAT? Perhaps and if so that would add another putatively positive aspect to targeting this pathway.

sulted in decreased proliferation of *M. tuberculosis*-specific T cells, suggesting that lymphangiogenesis blockade reduced access of antigen-presenting cells to the lymph nodes to stimulate T cell proliferation. However, this was done in the context of BCG “infection”, so the effects on bacterial burden were unable to be ascertained as BCG is attenuated and lacks critical virulence factors. Future studies on this pathway during infection with virulent *M. tuberculosis* seems to be in order as well as a more thorough examination of the consequences of lymphangiogenesis on disease progression over time. While blocking lymphangiogenesis may alter the ability of the immune system to generate a robust adaptive response, it may also alter cellular dynamics earlier in infection in host-beneficial ways, but these things are currently unknown. A synthesis of the larval and adult models of zebrafish infection seem to be promising methods for assessing some of these impacts.

The second set of studies interrogated the function of lymphatic endothelium as a replicative niche for *M. tuberculosis* (Lerner et al., 2016; Lerner et al., 2020). Although tuberculosis is classically a pulmonary infection and the pulmonary niche is the only effective route of transmission, the most common extrapulmonary presentation of tuberculosis is in the lymph nodes, a disease known from antiquity as scrofula (Bloch, 1973). The infection of the lymph nodes implied that the bacteria had a productive replicative niche in the lymph nodes, which could have simply been macrophages, but this study found that mycobacteria could directly infect the lymphatic endothelium (Lerner et al., 2016). Indeed, it appears that lymphatic endothelial cells are a relatively permissive host cell type, as the bacteria will cord within these cells as a mechanism of host immune evasion that can then block later macrophage phagocytosis (Lerner et al., 2020). Pathological examination of human pulmonary lymphatic vessels would be an interesting adjunct to understand how lymphatic endothelial cell disruption modulates the adaptive immune response and physically reshapes the

lung environment. This implies a binary role for lymphangiogenesis during infection, where it may be important for priming certain adaptive immune responses, but may also offer a convenient replicative niche; the net effect remains in need of study and the zebrafish may offer an ideal model in which to do so.

There are an abundance of tools in the zebrafish to study lymphangiogenic processes. The *lyve1* transgenic lines allow for visualization of lymphatic vessels throughout life, where they appear at approximately 3 days post fertilization (Okuda et al., 2012). This allows both larval and adult zebrafish to be used to study these processes, unlocking the full spectrum of disease. Additionally, a number of genetic tools exist to allow for study of this process. Notably, the *hsp70l:sFlt4^{bns82}* line is able to potently inhibit the generation of lymphatics through overexpression of a soluble decoy VEGFR3 after heat shock (Matsuoka et al., 2016). Together, these lines can offer both a means of visually assessing the state of the lymphatic endothelium as well as block lymphangiogenesis at various time points. Pioneering studies should evaluate the degree of lymphangiogenesis induced by *M. marinum* infection and, thorough genetic inhibition, study the role of this at both early and late stages of infection in the larva and the adult. It is my hypothesis that, especially in the adult, there is extensive lymphatic involvement that contributes to infection and, like previous studies, may provide an extragranulomatous replicative niche during earlier stages of infection (Lerner et al., 2016). The ability to study the contributions of lymphatics within a model that presents with necrotic, epithelioid granulomas offers the opportunity to also evaluate them as a potential route of dissemination complementary to hematogenous dissemination.

There are several proposed strategies to block lymphangiogenesis in these diverse contexts. One is, of course, through pharmacological inhibition of VEGFR3 signaling

in a manner analogous to that of bevacizumab, but this may be subject to subversion through compensatory VEGFR2 signaling. Another is to block aspects of the lymphatics that are required for growth, such as integrin α 5 (Dietrich et al., 2007) or fatty acid oxidation (Wong et al., 2017). The future of this remains uncertain, although these drugs may act as excellent adjuvants to existing antihemangiogenic therapies. A more comprehensive synthesis of both angiogenic families is needed to better understand their individual and combinatorial contributions to the overall pathology of tuberculosis; while antihemangiogenic therapies appear to be uniformly beneficial, the lymphatic system may be important for certain types of protective immune responses and specific targeting at certain points during the course of infection may alter the utility of antilymphangiogenic therapy.

5.9 Generalizability

One of the major questions is whether or not the findings I have presented in Chapter 3 are able to be generalized to other disease contexts. The most notable possibility is in cancer, where tumor-associated macrophages are well known to produce VEGFA to induce tumor angiogenesis (Subsubsection 1.5.4). Are these tumor associated macrophages detecting some tumor-derived ligand to induce these responses or is the production of VEGFA a product of a complex set of secreted signals that create the proper microenvironment? It may be that such a distinction is one without an underlying need for discrimination as the key lies in whether these signals – whatever they are – are inducing NFAT nuclear translocation. To my knowledge, no study has been done to look at the localization of NFAT in tumor associated macrophages from either cancer models or patient biopsies and this would make for one avenue to begin exploring potential contributions of this pathway to cancer progression.

The zebrafish and the tools I have made would also provide an excellent means to beginning to study these phenomena. For instance, the zebrafish *mitfa:BRAF^{V600E}* model could be used to induce melanomas in the zebrafish and then further histological profiling can be done to further examine the effects on tumor mass, vascularization, and other macrophage responses to the tumor in the *irg1:VIVIT* background (Patton et al., 2005; Brewer et al., 2022b). Complementary use of similar mouse models with *LysM-Cre*; *Nfatc2^{f/f}* or *ROSA26:lox-STOP-lox-VIVIT* lines already developed may offer further insight on this process in a mammalian context. The precise biology of tumor associated macrophages, while extensively described, is not particularly well understood and genetic approaches like these may offer further insight into some master regulators of tumor associated macrophage responses and present opportunities for reprogramming toward more effective tumor clearance. Such an approach can build upon work from the Zon lab to develop an optically accessible model to study tumor-macrophage interactions and the development of tumor-associated macrophages.

Other infectious diseases are known to induce the formation of granulomas or manipulate the vasculature in various ways. A great many diseases induce the formation of new blood vessels for their own purposes while others actively suppress the host angiogenic response (Osherov and Ben-Ami, 2016). For instance, *Cryptococcus neoformans* is a well-known granulomatous infection, where it can either conceal itself in quiescent or sterilizing lung granulomas or active, fulminant granulomas within the brain. Infection begins within macrophages where the yeast has to adapt to a new, hostile environment and subvert these responses for its own benefit, a process which has been described in some detail over the course of scientific investigation (Coelho, Bocca, and Casadevall, 2014). However, little is known or described about the associations between cryptococcal granulomas and the vasculature, with the subject

left unmentioned even in the most recent review on the subject³¹ (Ristow and Davis, 2021). More generally, the biology of the cryptococcal granuloma remains understudied with contributions of the surrounding stromal cells largely unknown and with few known host determinants despite many studies having been conducted on the contributions of C-type lectin receptors to host defense (Campuzano et al., 2017). However, C-type lectin signaling seems to have a mostly neutral effect on *Cryptococcus* infection despite CARD9 playing a critical role in host defense (Nakamura et al., 2007; Campuzano et al., 2017; Walsh et al., 2017), an effect reminiscent of the Janus-faced roles uncovered for DECTIN-1 signaling by Deerhake et al. (2021). Studying whether VEGFA and angiogenesis play a role in this context would certainly be interesting and, given the facility with which larval zebrafish can be infected with *Cryptococcus*, this may be a convenient platform to begin studying some of these interactions, as well as the macrophage-specific consequences of NFAT inhibition, as NFAT is thought to play important roles in protection against various fungal diseases (Goodridge, Simmons, and Underhill, 2007; Greenblatt et al., 2010; Tenor et al., 2015; Davis et al., 2016; Bojarczuk et al., 2016). The ability to adapt an existing model in the zebrafish to capture elements of this disease offers a compelling means of studying the contributions of NFAT signaling and angiogenesis in the innate responses to this prevalent disease of the immunocompromised (Lin and Heitman, 2006). These responses are comparatively understudied in *Cryptococcus* immunity and direct comparisons with what is known from *Candida* is challenging due to evolutionary distinctions and radically different disease presentation. Indeed, given that the existing antifungal drug itraconazole has known anti-angiogenic activity, it may be of interest to attempt to

³¹While absence of evidence is not evidence of absence, this is clearly a subject worthy of further exploration. Historical reviews note the extensive capillary involvement of *Cryptococcus*, suggesting that they are actively engaging the vasculature although with unknown consequence (Shibuya et al., 2005).

repurpose this drug as an anti-angiogenic therapeutic option in other contexts (Nacev et al., 2011).

Histoplasma is a further fungal infection that is known to induce granuloma formation and for which there seem to be additional descriptions of the vascular involvement of this infection. Similar to other infections, enhanced stabilization of HIF-1 α in this context resulted in improved bacterial killing (Friedrich et al., 2017; Friedrich et al., 2019). Despite these potentially beneficial roles for known pro-angiogenic pathways in the innate defense against *Histoplasma*, this does not resolve any role for the angiogenesis itself in this process. However, during later stages of granulomatous infection, extensive vasculitis in the periphery of these foci has been noted by histological examination (Mann, Fogarty, and Kincaid, 2000; Kauffman, 2007; Mukhopadhyay and Katzenstein, 2010). Interestingly, itraconazole is the primary therapy used for histoplasmosis and the anti-angiogenic contributions of this therapy to the overall course of disease resolution would certainly be of some interest in further clarifying the mechanisms of action (Kauffman, 2007). Models exist in the mouse for the study of this infection and have been well-described, but further work to characterize the host and pathogen factors underlying the granulomatous response in this disease seems to be in order (Heninger et al., 2006). Not only would this potentially reveal alternative treatment options, but would highlight further aspects of potentially generalizable granuloma biology.

Schistosomiasis is a classical model of what are called fibroid granulomas and this disease of the urogenital tract is known to induce VEGFA-dependent angiogenesis that benefits the growth of the parasitic eggs (Loeffler et al., 2002; Shariati et al., 2011; Pereira et al., 2013; Chabon et al., 2014; Figueiredo et al., 2015). However, the precise underlying mechanisms of this angiogenesis induction are unknown and ap-

pear to be mediated by specific macrophage-mediated detection of parasite ligands, but these have not been identified (Loeffler et al., 2002). It is tempting to think that this could be through the activation of a C-type lectin receptor, paralleling the findings presented in Brewer et al. (2022b). Identification of the underlying mechanism of VEGFA-induction by *Schistosoma* has implications in not only improving disease outcomes, but managing the hypertensive pathology of schistosomiasis, which is linked to parasite-induced VEGFA production (Chabon et al., 2014). The potential that this pathway is mediated by NFAT should be further explored, which is now possible in the zebrafish and might offer a compelling model for identifying both parasite ligands and host pathways in this disease (Takaki et al., 2021).

The role of angiogenesis in mold infections is the inverse, where *Aspergillus* is thought to inhibit the host VEGFA response in order to maintain itself within a vascular niche (Kontoyiannis, 2010). While the host responds by producing abundant VEGFA³², the fungus produces gliotoxin to destroy newly formed vessels and may also produce factors that suppress HIF-1 α (Ben-Ami, 2013). It thus becomes a compelling option that there may be non-targeted pathways by the fungus that could be repurposed to induce angiogenesis; NFAT activators may serve such a purpose by driving VEGFA expression, which has been suggested to be a potential host-directed therapeutic target in aspergillosis.

Rheumatoid arthritis is an autoimmune condition characterized by aberrant inflammation in the joints, which is often mediated by macrophages and T cells and for which anti-TNF- α therapy has proven remarkably effective (Feldmann, 2002). The macrophage and vascular involvement suggests a potential role for some intra-macrophage signaling pathway that induces the production of VEGFA, but the identity of

³²Could this be downstream of C-type lectin receptors?

this pathway is unknown, although assumed to be through the HIF-1 α (MacDonald et al., 2018). As anti-angiogenesis therapy has been shown in animal models to improve symptoms, this may offer another alternative to treatment for arthritis where other treatments have been unsuccessful (Paleolog, 2002; Elshabrawy et al., 2015). Indeed, treatment of patients with rheumatoid arthritis with tacrolimus ameliorates symptoms and has been shown to aid in treatment of otherwise non-responsive arthritis (Dutta and Ahmad, 2011). This mechanism could be through activity of tacrolimus on T cells, but may play equally important roles in inhibiting macrophage TNF- α and VEGFA production to improve symptoms. Alternatively, parts of the angiogenic response in arthritis are known to be mediated by aberrant TLR activation, which drives the production of ANG2 and, although we do not yet know if NFAT signalling mediates angiopoietin expression, this could be another potential mechanism of action here (Saber et al., 2011).

It may be the case that activation of C-type lectin receptors more broadly is able to induce the production of VEGFA, which would tie this pathway into a vast array of diseases given the vast variety of ligands able to agonize this pathway. Alternatively, it may be the case that pathogen-specific subversion of these receptors induces different relative proportions of the downstream pathways; if this were the case, the underlying mechanisms would be extremely revealing as to the nature of C-type lectin signaling and selective downstream pathway induction.

5.10 A Pressing Need for More Objective Approaches to Image Analysis

While microscopy serves as the basis of much experimentation and is ubiquitous in the zebrafish field, these sorts of data remain extremely difficult to analyze in an objective manner. While blinding approaches (Section 4.8) can help, what is truly needed is the development of more truly objective analytical approaches to glean quantitation from images. These features should, often, be discrete features that can be differentiated from the background but in practice this is not always the case. As often as not, human interpretation of these images is required to differentiate apparent from true effects, a problem that is magnified by heterogeneity within the samples. As we saw throughout Chapter 4 the manual aspects of image processing and analysis can be creatively subverted with existing tools, but end-to-end image analysis and parameter extraction would both ease the labor of the microscopist and would likely unlock new aspects of data analysis that are currently impractical or impossible.

Classically, image features are extracted through the utilization of image thresholding, which can be either static and manual or dynamic and automated. There are several of these automated thresholding methods built into FIJI/ImageJ, which allows for the intelligent selection of regions of interest, typically based on the intensity of the pixels that exceed a certain percentile of the total image. This is excellent for adapting to the image itself, but is challenging for comparing across images; static methods where a single minimum and maximum value is selected are superior although may not be appropriate for every image in a set. These approaches can be combined by using automated thresholding to calculate the static value to be used.

However, these neglect the possibility that a uniform measurement is required but the threshold itself can vary subtly from image to image. With existing methods, the best of both worlds is elusive.

The answer, so it seems, lies in intelligent application of machine learning algorithms. While these approaches are functionally in their infancy, it is already possible to attempt to train a model to analyze diverse aspects of images and classify them into distinct categories; it is more difficult to use those models to do things like precisely measure subtle changes. In the future, these will be important improvements to pursue with the goal of making scientific image analysis more reproducible, robust, and informative and simultaneously less laborious and subjective.

5.10.1 Approaches to Measuring Angiogenesis

Many have attempted and few have succeeded at creating approaches that are able to automate the measurement of angiogenesis in the zebrafish. While there seem to be a number of FIJI plugins that can measure blood vessel growth *in vitro*, the *in vivo* context presents some unique challenges even within such a developmentally defined model. Although there had been earlier utilities (Heath, Bicknell, and Simms, 2017), probably the first earnest attempt (from only this year) focused on developmental angiogenesis in the brain of the larval zebrafish and was able to develop a machine learning methodology to monitor quantitative features of the zebrafish cranial vasculature over time – an unquestionably impressive feat (Kugler et al., 2022). However, this is somewhat narrowly focused on its intended vascular context and I had substantial difficulty in translating this approach for use in intersegmental vasculature. While tailored approaches for each discrete problem is how we have gotten into this situation of being unable to utilize other's tools for our purpose, it may be that ei-

ther the approach taken by Kugler et al. needs to be accommodated somewhat to other regions of vasculature or bespoke tools will have to be made for each possible application.

To do so, I would propose a few key steps. First, it is important to develop a landmarked model of angiogenesis over the course of development that is able to provide a robust reference with acceptable built-in tolerances. While developmental angiogenesis in the zebrafish is highly stereotyped, there is unquestionably a degree of fluctuation from fish to fish in the exact trajectory of vascular patterning. Once such a model has been established, the model must be able to align new images to this reference and then detect any differences between the spectrum of possible references and the image presented to capture measurements about the aberrations – total length and volume, intersegmental location, correspondence to bacterial burden (if present), etc. It may be that this can be accomplished with traditional thresholding-based approaches, but training a machine learning model in PyTorch or TensorFlow and integrating with the new PyImageJ may be more feasible at scale and more generally applicable. This is a project worth substantial effort if the use of the larval zebrafish's intersomitic vasculature is to be used in future endeavors to measure vascular perturbations and would likely advance this model in and of itself by enabling more consistent measurements across individuals and labs.

5.10.2 High-Throughput Quantitation of Cell Features

Unlike the zebrafish, which is a complex multicellular organism, extracting useful information from cells in tissue culture *should* be comparatively easier, but as I have experienced, this is not always the case. Subtle variations in fluorescence intensity and patterning and algorithmic preference for within-image automated thresholding

versus pan-image uniform thresholding makes it difficult to accurately threshold cellular features at low magnification, which allows for more complete capture of fields of cells versus the biased selection of individual cells at high magnification. While it is relatively easy to segment cells at 63x or 100x with the use of proper markers, none of my attempts at automated quantitation of the images for the THP-1 experiments came to fruition. My desire was an objective capture of the nuclear localization of NFAT in these cells and both a binary call of VEGFA expression and some ability to, in a granular way, measure the amount of VEGFA being produced per cell out of approximately 100 cells per field of view. CellProfiler (Carpenter et al., 2006; Kamentsky et al., 2011; McQuin et al., 2018; Stirling et al., 2021) did an acceptable job at segregating nuclei and capturing large punctae of NFAT, but struggled to measure the diffuse staining patterns seen with VEGFA and would often omit cells that, by eye, I called as positive no matter the thresholding pattern used. Additionally, it was challenging to develop overlapping sets of measurements that allowed me to isolate individual cells with nuclear NFAT and VEGFA expression – a cytoplasmic marker likely would have helped, as would have sparser plating of the cells themselves. However, these workarounds in many ways defeat the aspirations of high-content imaging. While I have no delusion about the imaging done for this work being “high content,” as it becomes possible to automate the collection of terabytes of imaging data, it must be similarly possible to analyze these gigapixels of images in an efficient manner while also extracting the maximum amount of biologically relevant data from them.

While I do not think that machine learning is the answer to all that ails, it does seem likely to me that a properly trained model could allow for precisely these sorts of analyses. By developing a model that comprehends the correspondence between particular cellular features, it should be reasonable to allow the model to intelligently make judgements based on the proximity and intensity of nearby pixels of relevance.

Again, I suspect that one of the sources of issue in my previous attempts was the lack of a pan-cytosolic or plasma membrane marker able to effectively delineate neighboring cells. Further study on the factors that best facilitate automated image analyses in cell culture is evidently needed.

5.11 NFAT Targeting as a Host-Directed Therapy?

The elephant in the room at the conclusion of this work is whether or not pharmacological targeting of NFAT could be a viable option to use as a host-directed therapeutic against mycobacterial infections, including tuberculosis. While we do not yet know, based on our preliminary experiments restricted to targeting either the individual isoform *NFATC2* broadly or pan-NFAT signaling in macrophages, it may be a promising avenue for clinical study once more specific approaches have been developed to the targeting of this pathway. As mentioned in Section 3.4, there are a number of ways to overcome the traditional limitations of calcineurin inhibition either through the development of more targeted inhibitors (like cell-permeable VIVIT or INCA-6), cell-specific targeting with liposomes or AAVs, isoform-specific targeting to minimize the scope of the inhibition, location-specific delivery via nebulization, or some combination of all of the above (Aramburu et al., 1999; Roehrl, Wang, and Wagner, 2004; Roehrl et al., 2004; Kitamura and Kaminuma, 2021; Colombo et al., 2022). An alternative would be to add further layers of therapeutic intervention, potentially through the addition of exogenous IL-2 to overcome the T cell-mediated immunosuppression of NFAT-targeted therapy, an approach that has shown promise (Whitehouse et al., 2017). At present, it would require more exhaustive study on the disease outcomes of broad NFAT inhibition in humans to begin to develop a model

for targeting this pathway during disease.

It has been established that tacrolimus treatment is able to inhibit angiogenesis, although we are now able to propose a model where, at least in some instances, part of this angiogenesis inhibition is mediated by a failure of macrophages to produce VEGFA after inhibition of NFAT (Turgut et al., 2011; Shen et al., 2022). However, the synthesis of various factors that contribute to tacrolimus's ability to prevent angiogenesis may prove to be a strength. By targeting macrophages and the endothelium simultaneously, it may be possible to circumvent cellular resistance mechanisms within the granuloma that may attempt to induce further VEGFA to alleviate hypoxia. A more thorough study on the role of NFAT in the endothelium would certainly clarify some of these opportunities.

One of the major questions is whether NFAT targeting improves the efficacy of existing antimycobacterial therapies based on time to clearance. This too can be modeled reasonably effectively in the zebrafish model and is an approach that could have important ramifications for this as a host-directed therapy. If NFAT inhibition synergizes with existing therapies to accelerate bacterial killing, then that would certainly present a compelling argument for such a strategy; conversely, if NFAT inhibition reduces bacterial burden but hampers drug delivery, such would be the death knell for this approach. Even in the latter instance, this should stimulate further study to understand what endothelial features make for the most effective antimicrobial delivery and seek pathways that facilitate those specific alterations.

As an additional and perhaps more readily translated possibility, topical tacrolimus is already widely used clinically to treat atopic dermatitis and may be useful as an adjunctive therapy to treat either non-tuberculous mycobacterial infections in humans, including *M. marinum*, *M. ulcerans*, and *M. abscessus* or leprosy. The potential util-

ity of existing topical treatment to improve drug delivery to these largely topical infections is a compelling possibility that could likely be tested quickly in pre-clinical models of the former list of pathogens. Indeed, confirmation of the utility of such an approach in these other diseases may encourage further study on their utility in the context of *M. tuberculosis*.

5.12 Closing Remarks

The work presented here resolves a critical question in the world of tuberculosis pathogenesis and opens new doors in the fields of macrophage biology and angiogenesis by uncovering an unexpected and surprising role for the nuclear factor of activated T cells transcription factor family in mediating vascular endothelial growth factor production by macrophages and resulting angiogenesis during mycobacterial infection. Such a work has necessarily synthesized aspects of diverse disciplines within the biological sciences to address the standing question of what transcriptional regulation may be influencing the outgrowth of blood vessels during tuberculosis. While there is undoubtedly much more to learn about various factors influencing the activation of the angiogenic response, I believe this opens new doors to the study of the NFAT signaling pathway in the broader pathogenesis of a variety of mycobacterial infections after years of neglect. While other transcription factors have been deeply characterized in the innate immune response to infection, this critical player has comparatively languished. With this work adding to the body of studies newly emphasizing the critical importance of this pathway in various conditions, further studies are likely to reveal previously unknown levels of import in infection, cancer, autoimmunity, and more.

New technologies have greatly facilitated the study of this classical immune path-

way and embracing these as an opportunity to more exhaustively understand the contributions of myeloid NFAT signaling to diverse pathologies is critical for a more comprehensive view on both the development of infectious pathology and the mechanisms underlying the (side) effects of prominent immunosuppressive therapies. Integration of the diverse impacts of NFAT signaling across the entire organism would allow for a more comprehensive understanding of the roles of these proteins individually and in tandem in development, homeostasis, and immunity and inform new therapeutic options for conditions in which NFAT inhibition is beneficial.

The conduct of this work has required creativity and perseverance through a years-long process of personal and scientific growth. It has been an intensely human process of experimentation and discovery, success and failure, open-mindedness and a willingness to follow the data where it leads. Science succeeds whenever it is moving us, individually and collectively, toward a more complete understanding of the nuance and chaos of world around us.

Appendix A

Reagents Used

Table A.1: List of Antibodies Used

Reagent or Resource	Source	Identifier
polyclonal goat anti-human VEGFA antibody	R&D Systems	#AF-293
monoclonal mouse anti-Cas9 antibody	Cell Signaling	#7A9-3A3
Normal Goat IgG Control	R&D Systems	#AB-108-C
rabbit anti-human NFATC1 serum (against NH ₂ -CVSPKTTDPEEGFPRGLGA, residues 210 to 227)	Lyakh, Ghosh, and Rice, 1997; Symes, Gearan, and Fink, 1998	#801
rabbit anti-human NFATC2 serum (against NH ₂ -CSPPSGPAYPDDVLDYGLK, residues 53 to 70)	Lyakh, Ghosh, and Rice, 1997; Symes, Gearan, and Fink, 1998	#1777
rabbit anti-human NFATC3 serum (against NH ₂ -DLQINDPEREFLERPSRDHL, residues 130 to 149)	Lyakh, Ghosh, and Rice, 1997; Symes, Gearan, and Fink, 1998	#1689
rabbit anti-human NFATC4 serum (against NH ₂ -GRDLSGFPAPPGEPPA, residues 886 to 902)	Lyakh, Ghosh, and Rice, 1997; Symes, Gearan, and Fink, 1998	#889

rabbit anti-human NFATC4 serum (against NH ₂ -CDSKVVFIERGPDGKLQWEE, residues 614 to 632)	Lyakh, Ghosh, and Rice, 1997; Symes, Gearan, and Fink, 1998	#890
rabbit anti-human pan-NFAT serum (against NH ₂ -SDIELRKGETDIGRKNTRC)	Lyakh, Ghosh, and Rice, 1997; Symes, Gearan, and Fink, 1998	#796
donkey anti-goat IgG Alexa Fluor 647	ThermoFisher	#A-21447
donkey anti-goat IgG Alexa Fluor 555	ThermoFisher	#A-21432
donkey anti-rabbit IgG Alexa Fluor 647	ThermoFisher	#A-31573
donkey anti-rabbit IgG Alexa Fluor 555	ThermoFisher	#A-31572
donkey anti-mouse IgG Alexa Fluor 555	ThermoFisher	#A-31570
donkey anti-mouse IgG Alexa Fluor 488	ThermoFisher	#A-21202

Table A.2: Bacterial Strains

Reagent or Resource	Source	Identifier
<i>Mycobacterium marinum</i> M	ATCC	#BAA-535
<i>Mycobacterium marinum</i> M / pMSP12:mCerulean	Oehlers et al., 2015	N/A
<i>Mycobacterium marinum</i> M / pMSP12:tdTomato	Cambier et al., 2014	N/A
<i>Mycobacterium marinum</i> M / pMSP12:eBFP2	Takaki et al., 2013	N/A
Gamma-irradiated <i>Mycobacterium tuberculosis</i> H37Rv	BEI	#NR-49098
NEB 5-alpha Competent <i>Escherichia coli</i> (High Efficiency)	NEB	#C2987H
NEB 10-beta Competent <i>Escherichia coli</i> (High Efficiency)	NEB	#C3019H
NEB Stable Competent <i>Escherichia coli</i> (High Efficiency)	NEB	#C3040H

Table A.3: Chemicals

Reagent or Resource	Source	Identifier
Trizol	Ambion	#15596026
MicroAmp Fast Optical 96-Well Reaction Plate with Barcode, 0.1 mL	Applied Biosystems	#4346906
Spawning Tanks	Aquaneering	#ZHCT100
Baking soda (sodium bicarbonate)	Arm & Hammer	#426292
Insulin Syringes	BD	#08290-3284-38
Tuberculin Syringe (27G)	BD	#309623
SDS, 20%(w/v) solution, 1L	Bio-Basic	#SD8119
40% acrylamide	Bio-Rad	#1610140
2% bis-acrylamide	Bio-Rad	#1610142
Artemia	Brine Shrimp Direct	#BSEP6LB
Polymyxin B sulfate	Cayman Chemical	#14157
INCA-6	Cayman Chemical Roehrl et al., 2004	#21812
T-75 Flasks	CellStart	#658170
Molecular Biology Grade Water	Corning	#46000CI
10x PBS	Corning	#46013CM
7H10	Difco	#262710
7H9	Difco	#271310
Chloroform	EMD Millipore	#CX1055
16% Methanol-free Paraformaldehyde	EMS	#15710
Triton X-100	Fisher Scientific	#BP151
Dimethyl sulfoxide (DMSO)	Fisher Scientific	#BP231
Mineral oil	Fisher Scientific	#BP2629
Tween-80	Fisher Scientific	#BP337
Sodium chloride	Fisher Scientific	#S271
1x PBS	Gibco	#10010-023
Sodium pyruvate	Gibco	#11360
Amphotericin B	Gibco	#15290-026
HEPES	Gibco	#15630
Instant Ocean Sea Salt	Instant Ocean	#SS15-10
Hygromycin B solution	Invitrogen	#10687010
4-well Cell Culture Slides	MatTek	#CCS-4
35 mm Dish, No. 1.5 Coverslip, 14 mm Glass Diameter, Uncoated	MatTek	#P35G-1.5-14-C
Tris (base)	Millipore	#648311
Millex-SV 5.0 µm	Millipore	#SLSV025LS

T4 DNA Ligase		NEB	#M0202S
Taq 5x Master Mix		NEB	#M0285L
LongAmp Taq		NEB	#M0323L
rSAP		NEB	#M0371L
Q5 High-Fidelity DNA Polymerase		NEB	#M0491L
Q5 High-Fidelity 2X Master Mix		NEB	#M0492L
Deoxynucleotide (dNTPs) Solution Mix		NEB	#N0447L
XbaI		NEB	#R0145L
DpnI		NEB	#R0176L
XmaI		NEB	#R0180L
PflMI		NEB	#R0509L
MwoI		NEB	#R0573L
FseI		NEB	#R0588L
NotI		NEB	#R3189L
RNA Cleanup Kit (50 µg)		NEB	#T2040L
6.5mm ceramic beads		Omni	#19-682
Petri dishes for embryonic zebrafish		Sarstedt	#83-3902-500
BAY 61-3606	Selleck Chemicals		#S7006
FK506 (tacrolimus)	Selleck Chemicals		#S5003
Methanol	Sigma-Aldrich		#179337
Ammonium chloride	Sigma-Aldrich		#254134
24:1 chloroform:isoamyl alcohol	Sigma-Aldrich		#25666
Sodium azide	Sigma-Aldrich		#71290
Boric acid	Sigma-Aldrich		#B0394
Sodium phosphate monobasic monohydrate	Sigma-Aldrich		#D2158
Fetal bovine serum	Sigma-Aldrich		#F2442
Incomplete Freund's adjuvant (IFA)	Sigma-Aldrich		#F5506
Glycerol	Sigma-Aldrich		#G7757
Glucose solution	Sigma-Aldrich		#G8769
OADC	Sigma-Aldrich		#M0678
Phorbol-12-myristate-13-acetate (PMA)	Sigma-Aldrich		#P148
Tween-20	Sigma-Aldrich		#P1754
1-phenyl-2-thiourea	Sigma-Aldrich		#P7629
RPMI-1640	Sigma-Aldrich		#R8758
trehalose 6-6'-dimycolate (TDM) from M. bovis	Sigma-Aldrich		#T3034
Tyloxapol	Sigma-Aldrich		#T8761
100x Tris-EDTA (TE)	Sigma-Aldrich		#T9285
Polybrene	Sigma-Aldrich		#TR-1003-G

BeadBug homogenizer tubes with 2.8mm stainless steel beads	Sigma-Aldrich	#Z763829-50EA
Dry fish food	Skretting	#GEMMA Micro 500
DAPI Fluoromount-G	SouthernBiotech	#0100-20
Tricaine-S (MS-222)	Syndel	#ANADA 200-226
Brefeldin A Solution (1000X)	ThermoFisher	#00-4506-51
BP Clonase II	ThermoFisher	#11789020
LR Clonase II Plus	ThermoFisher	#12538120
FastDigest Esp3I (II class)	ThermoFisher	#FD0454
Calcium chloride	VWR	#BDH9224
Potassium chloride	VWR	#BDH9258
Sodium phosphate dibasic heptahydrate	VWR	#BDH9296
2,2'-Azobis[2-(2-imidazolin-2-yl)propane]dihydrochloride	Wako Chemicals	#VA-044
Magnesium chloride	Ward's Scientific	#470301

Table A.4: Commercial Assays

Reagent or Resource	Source	Identifier
MeltDoctor HRM Master Mix	Applied Biosystems	#4415450
Luna Universal qPCR Master Mix	NEB	#M3003X
Human VEGF DuoSet ELISA	R&D Systems	#DY293B-05
LunaScript RT SuperMix Kit	NEB	#E3010L
HiScribe T7 High Yield RNA Synthesis Kit	NEB	#E2040S

Table A.5: Cell Lines

Reagent or Resource	Source	Identifier
THP-1 monocytic cells	ATCC	#TIB-202
HEK-293T	ATCC	#CRL-2316

Table A.6: Model Organisms and Strains

Reagent or Resource	Source	Identifier
<i>Danio rerio</i> strain *AB	ZIRC	#ZDB-GENO-960809-7
Tg(<i>irg1:tdTomato</i> ^{xt40})	Brewer et al., 2022b	N/A
Tg(<i>irg1:VIVIT-tdTomato</i> ^{xt38})	Brewer et al., 2022b	N/A
Tg(<i>kdr1:eGFP</i> ^{s843})	Jin et al., 2005	N/A
TgBAC(<i>vegfaa:eGFP</i> ^{pd260})	Karra et al., 2018	N/A
Tg(<i>lyz:DsRed2</i> ^{nz50})	Hall et al., 2007	N/A
<i>nfatc2a</i> ^{xt69}	Brewer et al., 2022b	N/A
<i>nfatc3a</i> ^{xt59}	Brewer et al., 2022b	N/A
<i>card9</i> ^{xt31}	Brewer et al., 2022b	N/A
<i>myd88</i> ^{xt29}	Walton et al., 2018	N/A

Table A.7: Recombinant DNA and Plasmids

Reagent or Resource	Source	Identifier
p5E <i>irg1</i>	Addgene Sanderson et al., 2015	#188698
pME VIVIT NS	Brewer et al., 2022b; Addgene	#188699
p3E tdTomato	Walton et al., 2015; Addgene	#188700
pDEST tol2 Ubb pA	Walton et al., 2015; Addgene	#188701
pME tdTomato	Oehlers et al., 2015; Addgene	#135202
p3e Ubb pA	Walton et al., 2015; Addgene	#188702
pTol2 <i>irg1:VIVIT-tdTomato</i>	Brewer et al., 2022b	

pTol2 irg1:tdTomato	Brewer et al., 2022b	
pLV hUbC-Cas9-P2A-Puro_BsmBI-sgRNA	Brewer et al., 2022b; derived from Kabadi et al., 2014; Sanjana, Shalem, and Zhang, 2014; Addgene	#188703
pLV hUbC-Cas9-P2A-Puro sgRNA α NFATC2	Brewer et al., 2022b; Addgene	#188704
pLV hUbC-Cas9-P2A-Puro sgRNA α Safe Targeting Loci	Brewer et al., 2022b; Addgene	#188705
phU6 NFATC2	Brewer et al., 2022b; Addgene	#188708
pmU6 NFATC2	Brewer et al., 2022b; Addgene	#188709
p7SK NFATC2	Brewer et al., 2022b; Addgene	#188710
phH1 NFATC2	Brewer et al., 2022b; Addgene	#188711
phU6 ST	Brewer et al., 2022b; Addgene	#188712
pmU6 ST	Brewer et al., 2022b; Addgene	#188713
p7SK ST	Brewer et al., 2022b; Addgene	#188714
phH1 ST	Brewer et al., 2022b; Addgene	#188715
psPAX2	Addgene	#12260
pMD2.G	Addgene	#12259
sfGFP-C1	Pedelacq et al., 2006; Addgene	#54579
pLEX:ALFA-mPapaya	This work	N/A
pLEX:CA-NFAT1-ALFA-mPapaya	This work	N/A
pLEX:CA-NFAT1-ΔDBD-ALFA- mPapaya	This work	N/A
pLEX:CA-NFAT1-ΔRIT-ALFA-mPapaya	This work	N/A
pLEX:CA-NFAT1-ΔDBD-ΔRIT-ALFA- mPapaya	This work	N/A

Table A.8: Software

Application	Source/Citation
R, 4.2.2	R Core Team, 2022
RStudio, 2022.12 "Elsbeth Geranium"	RStudio Team, 2022
FIJI/ImageJ2, 2.9.0	Schindelin et al., 2012; Rueden et al., 2017
ImageJ, 1.53u	Schneider, Rasband, and Eliceiri, 2012
Python/Jython, 2.7.2	van Rossum, 1995
ggplot2, 3.3.5	Wickham, 2009; Wickham, 2016; Wickham et al., 2022a
dplyr, 1.0.9	Wickham et al., 2022b
gghighlight, 0.3.3	Yutani, 2022
ggbeeswarm, 0.6.1	Clarke and Sherrill-Mix, 2017
ggsignif, 0.6.3	Ahlmann-Eltze and Patil, 2021
blindrename.pl, 1.0	Salter, 2016
scales, 1.2.0	Wickham and Seidel, 2022
extrafont, 0.18	Chang, 2022
reshape, 0.8.9	Wickham, 2022
RColorBrewer, 1.1-3	Neuwirth, 2022
FSA, 0.9.3	Ogle et al., 2022
HRM Software, 3.0.1	ThermoFisher

Table A.9: Equipment

Equipment	Source	Identifier
MP Bio FastPrep 24 Classic (Bead Mill)	MP Bio	#116004500
Applied Biosystems 7500 Fast Real-Time PCR System	ThermoFisher	#4351106
Nikon Stereomicroscope	Nikon	#SMZ745
Nikon High Intensity Illuminator	Nikon	#NI-150
Eppendorf Femtojet 4x	Eppendorf	#5253000025
Precision Plant Growth Chamber, 504 L	ThermoFisher	#PR505755L
Zeiss AxioObserver Z1	Zeiss	N/A
X-Cite 120Q	Excelitas	#12-63000
Cryostat	Leica	#CM1860

Table A.10: Oligonucleotides

Description	Sequence (5' to 3')
VIVIT sense oligo	GCCATCATGGCAGGACCACACCCGGT GATTGTTATCACTGGACCACATGAGG AG
VIVIT anti-sense oligo	CTCCTCATGTGGTCCAGTGATAACAA TCACCGGGTGTGGCCTGCCATGATG GC
VIVIT attB1 primer F	GGGGACAAGTTGTACAAAAAAGCAG GCTGCCATCATGGCAGGACC
VIVIT attB2 primer R	GGGGACCACTTGTACAAGAAAGCTG GGTACTCCTCATGTGGTCCAGTG
irg1 promoter cloning primer F	CCCTATAGTGAGTCGTATTAC
irg1 promoter cloning primer R	TCCCTTAGTGAGGGTTAAT
nfatc2a gRNA 1	TAATACGACTCACTATAGGGCTGCGA GAACGGGCCACGTTTAGAGCTAGAA
nfatc2a gRNA 2	TAATACGACTCACTATAGGCAGCCG TCGCCCCACGGGTTTAGAGCTAGAA
nfatc3a gRNA 1	TAATACGACTCACTATAGGGCAGTT GCAGTAGTCATGTTAGAGCTAGAA
nfatc2a crispant gRNA	TAATACGACTCACTATAGGTCA GGTGAACCTGTCGTTTAGAGCTAGAA
nfatc3a crispant gRNA	TAATACGACTCACTATAGGTAGAGGC ACTGACCTGCGGTTTAGAGCTAGAA
myd88 gRNA	TAATACGACTCACTATAGGCCAGA CTGGAGGACAGGTTTAGAGCTAGAA
card9 gRNA	TAATACGACTCACTATAGGGCAAGGT GCTGAGCAGCGGTTTAGAGCTAGAA
ENSDARG00000056379 gRNA	TAATACGACTCACTATAGGTGAAGTG TTTGAGAGGTCGTTTAGAGCTAGAA
ENSDARG00000077975 gRNA 1	TAATACGACTCACTATAGGCAGGGAC TTTCTGTGGATGTTTAGAGCTAGAA
ENSDARG00000077975 gRNA 2	TAATACGACTCACTATAGGTCA TCCATTGTCGGTTTAGAGCTAGAA
ENSDARG00000079903 gRNA 1	TAATACGACTCACTATAGGGAGGCAA TAAGTGGAAAGTGTAGAGCTAGAA

ENSDARG00000079903 gRNA 2	TAATACGACTCACTATAAGGCATAAA GTGGAAGTGGGGTTTAGAGCTAGAA
nfatc2a PfM1 sequencing/genotyping primer F (+ BGH Reverse)	TAGAAGGCACAGTCGAGGCTCGAGGC TTTCTGGAGACCTCTGTCC
nfatc2a PfMI sequencing/genotyping primer R (+ GAG Reverse)	TGACACACATTCCACAGGGCTCTAG AGGTTGCCCTCATATCCTGC
nfatc2a MwoI genotyping primer F	CCTCTATGCAAACGCACCTACG
nfatc2a MwoI genotyping primer R	GTGATGCTCCTGTGGCAC
nfatc2a crispant HRMA primer F	CTCTTTTACGGCGAAAAAGCTGC
nfatc2a crispant HRMA primer R	GAAACAAACCTTGAAGTCCTGTTGG
nfatc3a crispant HRMA primer F	CCCGGGAATGAAGAGCTGG
nfatc3a crispant HRMA primer R	GTGTTCGCCTTGCGATCC
nfatc3a sequencing primer F (+ M13F-40)	GTTTCCCAGTCACGACCAGAACGGTC GAGCAGTTGG
nfatc3a sequencing primer R	AACGTGTTCGCCTTGC
nfatc3a HRMA primer F	AAAGAGTCGGTGTACATAGACGGG
nfatc3a HRMA primer R	CGAAGATCAGTCTGAAGTCCAGC
card9 sequencing primer F (+ M13F-40)	GTTTCCCAGTCACGACCAGAACGGTC CTCATCAAGACC
card9 sequencing primer R	CTTCAGATTGTCTTCAGAACTCTTAC C
card9 HRMA primer F	CCTTATCTGAGACAGTGCAAGGTGC
card9 HRMA primer R	TTACCAACTTGCAGGCGTCTG
myd88 HRMA primer F	CCGAAAGAAACTGGGTCTGTTCC
myd88 HRMA primer R	ACGAGTTCCCAGTCCGTCA
ENSDARG00000056379 sequencing primer F	GTGTTCCCAGTCACGACGACCTATACT TCTCATCACAGAGC
ENSDARG00000056379 sequencing primer R	GTCAGACACAGATGCATTGC
ENSDARG00000056379 HRMA primer F	CACAAAGGAGTGAAGTGTGAGAGG
ENSDARG00000056379 HRMA primer R	GAGCAATAAGCAGGACAAGAGAAACC

ENSDARG00000077975 sequencing primer F	GTTTCCCAGTCACGACATGAGCTGG TCTGAGAGC
ENSDARG00000077975 sequencing primer R	CATGAACGTTTACCACTTACCC
ENSDARG00000077975 HRMA primer F	CAGAGGTTCATATCTCCATTGTCGA GG
ENSDARG00000077975 HRMA primer R	TTGCCCTCGATCTCTCGTCAG
ENSDARG00000079903 sequencing primer F	GTTTCCCAGTCACGACTCATTATTAA AGAGTGAAGAGAAGCAGG
ENSDARG00000079903 sequencing primer R	TGTTTTGTAGGAATCCGATGC
ENSDARG00000079903 HRMA primer F	ACAGAGACTGGAGGCAATAAGTGG
ENSDARG00000079903 HRMA primer R	CCTTGATTCACTGGTGAGTTATCCAC C
VIVIT genotyping primer F	ATTCAGAGCTCGCACAGG
VIVIT genotyping primer R	ATCTCGAACTCGTGGCC
human VEGFA qPCR primer F	GAGGAGGGCAGAACATCACG
human VEGFA qPCR primer R	ACAGGATGGCTTGAAGATGTACTCG
human TNF qPCR primer F	GAGGCCAAGCCCTGGTATG
human TNF qPCR primer R	CGGGCCGATTGATCTCAGC
human GAPDH qPCR primer F	CTGGGCTACACTGAGCACC
human GAPDH qPCR primer R	AAGTGGTCGTTGAGGGCAATG
hU6 ST sgRNA Sequence	GATGGTGACAGTTGTCGA
mU6 ST sgRNA Sequence	GCTAAGTACTCTAACAGG
7SK ST sgRNA Sequence	GTGGATAACTCCTGAGT
hH1 ST sgRNA Sequence	GTGCAGTTCTCCGGGTTG
hU6 NFATC2 sgRNA Sequence	GACACCGGCGAGGGGTCA
mU6 NFATC2 sgRNA Sequence	GCTTGGCACCAGGCGATG
7SK NFATC2 sgRNA Sequence	GCCACGGACTCGCCTTGT
hH1 NFATC2 sgRNA Sequence	GGCCGGGTAGATGTGGCG
Common Tail Oligo	AAAAGCACCGACTCGGTGCCACTTT TCAAGTTGATAACGGACTAGCCTTAT TTAACTTGCTATTCTAGCTCTAAA AC

Appendix B

Response to Reviewers for Brewer et al., 2022b¹

The following is the cover letter that was provided along with the resubmission to *Cell Reports* for Brewer et al. (2022b). Written together with Dr. David Tobin.

5 November 2022

Dear Editors,

At the advice of Faby Rivas, Deputy Editor of *Immunity*, we are transferring our manuscript “Macrophage NFATC2 Drives [*sic*] Angiogenic Signaling During Mycobacterial Infection” to *Cell Reports*. Dr. Rivas wrote with our initial *Immunity* decision: “However, given a recent change in editorial policy that enables me to speak for *Cell Reports*, I can say that *Cell Reports* would be interested in taking your study to publication upon clarification of the quantification concerns raised by referees (minor revisions).”

As detailed in the Response to Reviewers document, we have both clarified and expanded on the quantification that we had previously performed. We have made

¹Lightly edited for structure and grammar, all content and critiques are left unmodified.

clear that our initial analyses were all performed blinded as well as performing additional extensive blinded and automated quantification as new Figure panels. These clarification as well as additional analyses have made our conclusions stronger. In addition, we performed one additional set of experiments to address variability in effect size that had concerned one of the reviewers. Our Response to Reviewers document describes this in more detail.

We think that this is an exciting story that would be of broad interest to *Cell Reports*' readers, as it touches on new or emerging concepts in mycobacterial pathogenesis, macrophage biology, engagement of endothelial cells during host immunity, immune cell/(non)-immune cell crosstalk, and NFAT signaling downstream of C-type lectin recognition of a specific bacterial lipid (a non-canonical pathway).

We are excited about multiple aspects of this work, which defines a completely unknown role for macrophage-specific NFAT signaling in mycobacterial pathogenesis, both in the zebrafish model and upon stimulation of a human cell line with *M. tuberculosis*. We had previously identified the impact of mycobacterium-induced angiogenesis in infection (Oehlers et al., 2015) as well as identifying an important bacterial modification of the mycobacterial cell envelope lipid trehalose dimycolate (TDM) that we showed is required for mycobacterium-induced angiogenesis (Walton et al., 2018). In the new work, we establish *in vivo* how TDM generates this host response, first ruling out *in vivo* the key canonical signaling molecules like CARD9. Through intravital imaging and targeted, cell-specific peptide-mediated inhibition, we identify macrophage-specific activation of NFAT signaling as essential to granuloma-associated angiogenesis. NFAT signaling has been extensively studied in the context of T cell biology, but has not, to our knowledge, been implicated in the macrophage response to mycobacterial infection.

We then combined single cell transcriptional data that we had generated from zebrafish granulomas and genetic approaches to identify the specific NFAT isoform required *in vivo*. In an *in vivo* genetic screen of multiple isoforms via CRISPR/Cas9 targeting, we found that *nfatc2a* in zebrafish was required for the response, while other NFATc isoforms were not. Finally, we extended our zebrafish *in vivo* findings to human cell lines and show that human *NFATC2* (the same isoform we identified in the zebrafish studies) mediates this response via macrophage production of the key pro-angiogenic molecule VEGFA downstream of *M. tuberculosis* TDM activation.

This conservation from fish to humans is particularly exciting and our results are consistent with single-cell analysis by ourselves and others of TB granulomas in zebrafish, mammalian models, and humans. Indeed one prominent theme of the more recent single cell profiling (scRNA-seq of zebrafish granulomas and non-human primate granulomas and MIBI-TOF analysis of human granulomas) has been a strong pro-angiogenic signature. Using the zebrafish model, we now provide new insight into the molecular basis for and consequences of that angiogenesis. Delineating this pathway also provides new therapeutic targets for host-directed approaches to treating TB.

Throughout, we use the tools available in the zebrafish to visualize the *in vivo* kinetics and dynamics of *Mycobacterium*-driven angiogenesis and host response, including multi-day timelapses using specific reporter lines, and CLARITY-cleared organs to analyze the intricate neovasculature that is driven by mycobacterial granulomas and which leads to accelerated bacterial growth and dissemination.

We are excited about these findings and hope that this work will be of broad interest to TB biologists and clinicians as well as those developing potential host-directed therapies for mycobacterial infections in pre-clinical models. The identification of a

specific NFAT isoform acting in macrophages suggests a new potential host target for therapies. In addition, because studies of NFAT have historically focused on T cells, uncovering an important role for NFAT signaling in macrophages should be particularly interesting to those studying macrophages and other myeloid cells in health and disease.

Thanks again for your consideration.

The following is the Response to Reviewers that accompanied the resubmission of Brewer et al. (2022b) to *Cell Reports*. Reviewer critiques from anonymous reviewers.

Response: We are grateful to both reviewers for their thoughtful review of our manuscript and have made a number of changes and improvements, including new experiments and analyses as well as text changes to strengthen our conclusions and highlight specific places where this work contributes novel findings likely to impact our understanding of NFAT signaling in macrophages, the engagement of NFAT signaling pathways downstream of C-type lectin recognition of microbial patterns, and specifically the role of these pathways in an important component of the host immune response to infection with pathogenic mycobacteria.

In addition to our textual changes highlighting the novelty and potential impact of the work, we present new, clearer and more magnified representative images throughout to make clearer both the robust angiogenic response we observe *in vivo* and our ability to quantitate *in vivo* at baseline and upon perturbation of this response. Importantly these changes can be uniquely accessed *in vivo* using the zebrafish model of mycobacterial infection.

In response to both reviewers, we have made much clearer that all our quantitative analysis was performed blinded (further details are now provided in the STAR Methods section), and in the new version we perform a number of automated and blinded image analyses to further support our conclusions. In particular, we now better quantitate the mammalian cell culture/*M. tuberculosis* studies, relating them in a blinded, quantitative manner to NFAT nuclear localization and VEGFA induction. We have added multiple new panels to both Figures 6 and 7 to include this quantitation.

In response to one reviewer's concern about variability in effect size in mammalian culture, we have better optimized our assay and present more consistent results: three new independent biological replicates that are now presented as primary data instead. The findings are completely consistent with our previous findings, but with less variability in effect size.

Based on reviewer comments, we have also added further discussion of the significance of our findings, including some discussion of the importance of extracellular bacteria engaging this signaling pathway and the reported unique signaling kinetics of NFATC2, the NFAT paralog that we identify here – both *in vivo* in the zebrafish and in mammalian cell culture experiments. We think that overall this work moves across two systems, one *in vivo* animal model in both larvae and adults, and another in human cell culture with *M. tuberculosis*, to identify a previously unknown role for NFAT signaling during mycobacterial infection and granuloma angiogenesis, which we know to be an important host

process manipulated by pathogenic mycobacteria to promote and sustain infection. Thus, this work also identifies a new host target for potential therapeutic interventions in TB, which, even in the age of COVID-19, has now again become the leading infectious cause of death worldwide.

Specific responses to each reviewer's comments are included below.

Reviewer #1: This study addresses the mechanism used by pathogenic mycobacteria and the mycobacterial lipoglycan trehalose dimycolate (TDM), to stimulate angiogenesis in the region of granulomas. This group has previously reported that induction of angiogenesis is pathogen-beneficial in their model of *M. marinum* infection in zebrafish, and has now addressed the signaling pathway involved in regulating macrophage production of proangiogenic VEGFA. The key finding in the manuscript is that macrophage NFATc, especially NFATc2a, signaling is essential, while CARD9 signaling is less important, for the effects observed. Certain of the results generated in the zebrafish models are confirmed using a human macrophage-like leukemia cell line exposed to irradiated *M. tuberculosis*. The major innovative finding is that NFAT signaling can be involved downstream of C-type lectin receptor signaling; a finding that may be generalizable.

Response: Thank you for the thoughtful review and for this assessment of the work, which we have now strengthened, including new experiments and analysis. We agree with the reviewer that the question of NFAT signaling downstream of C-type lectin receptor signaling and particularly in myeloid cells is almost entirely unstudied across fields, and that there

is particular importance and impact to understanding the architecture of this response generally and, moreover, the details for the immune response to a major human pathogen. Our work in zebrafish and work from multiple others with human clinical samples and in non-human primate models have identified the importance of VEGFA induction at the mycobacterial granuloma during pathogenesis, and, for some TB disease presentations, there are active clinical efforts to harness this pathway therapeutically. Moreover, as the reviewer points out, there is considerable novelty and generalizability in understanding myeloid engagement of NFAT signaling downstream of C-type lectin receptor activation.

Reviewer #1: There are two ways to consider the significance. With regard to promoting the understanding of mycobacterial/TB pathogenesis, the findings might be considered incremental, since the investigators have provided ample evidence of angiogenesis and its significance in this model. On the other hand, the finding of NFAT signaling in C-type lectin induction of VEGFA by macrophages has the potential to be generalizable to other C-type lectin signaling contexts.

There are no major concerns with the experimental, analytical, or data [*sic*], in the study. One concern is whether the finding that NFAT signaling downstream of C-type lectin engagement in macrophages contributes to angiogenesis is unique to this system, or if it is generalizable. More information on the possibility that the observations relate to signaling that is only engaged by a high concentration of agonist (i.e. numerous extracellular bacteria) could shed light on

this possibility².

Response: Thanks for this assessment and we agree that the finding of an important physiological role for NFAT signaling in macrophages, downstream of C-type lectin signaling is a largely undescribed and important aspect of NFAT and macrophage biology. As detailed above, we respectfully disagree with the characterization that this work in the context of TB is only a limited advance. While it fits together well with previous work we have published 1) describing the impact of angiogenesis on mycobacterial granulomas (Oehlers et al., 2015) and 2) identifying a specific bacterial enzymatic modification to a predominant cell envelope lipid that underlies this response (Walton et al., 2018) – the downstream host steps that mediate this important aspect of TB pathogenesis had not been studied. Here we found – both in an *in vivo* model that forms true granulomas and translating these findings into mammalian cell culture – that a previously undescribed engagement of NFAT signaling in macrophages drives granuloma-associated angiogenesis.

In this work, we provide the first real insight into how macrophage detection of the mycobacterial glycolipid TDM specifically manipulates the mycobacterium-infected host, leading to VEGFA induction and angiogenesis, a response that enables the bacteria to survive, replicate and disseminate and is therefore of active clinical interest therapeutically for TB. Notably, standard models of TB disease (C57BL/6

²In isolation, the findings reported here do not solve a major question in mycobacterial-host interactions, but may provide a step toward addressing the larger question of how angiogenesis is pathogen-beneficial in mycobacterial infections.

mice, for example) do not recapitulate the granulomas seen in humans and non-human primates, and so the zebrafish is one of the few genetically manipulable animal models where this important process can be dissected experimentally. This work 1) identifies activation of a previously unconsidered pathway that culminates in activation of NFATC2 as the key downstream transducer of VEGFA production during mycobacterial infection *in vivo* in the zebrafish models and in a human cell line; and 2) assesses the consequences of perturbing NFAT signalling, both using loss-of-function mutants as well as macrophage-specific inhibitory approaches in whole animals *in vivo*. Generally, NFAT induction and transduction of pathogen-associated signals have not been explored in myeloid cell biology, and so the link in both humans and zebrafish established here may also spur others to examine myeloid NFAT activation in different contexts during infection and disease.

Reviewer #1: The observation that induction of angiogenesis is delayed for several days post infection, when extracellular mycobacteria have accumulated, may be interesting and provide additional clues to the biological role of angiogenesis and mycobacterial infection. The observation suggests that the Clec-NFAT pathway might be due to low sensitivity and only activated by a high concentration of Clec ligand. It would be beneficial to address this possibility and consider its impact on mycobacteria-host interactions. The Discussion addresses the observation, but not its potential unique significance.

Response: We agree with this suggestion and have highlighted the potential unique significance further in the Discussion. We have tried to ad-

dress these observations throughout – first in Figure 1, where we put emphasis in the Results section on the fact that the induction of angiogenesis coincides with the emergence of extracellular bacteria, and, later in the paper, in the design of our mammalian cell culture assay. As we describe in the Discussion, it is notable that standard published THP-1 infection assays (which aim to eliminate extracellular bacteria by antibiotic treatment) have not detected VEGF production at later timepoints. We have included discussion of this point and further emphasized this point throughout.

Reviewer #1: The manuscript notes the involvement of calcium signaling in NFAT activation in classical systems, but doesn't address it experimentally. It would be interesting to know if intracellular calcium responses to mycobacteria and/or TDM through C-type lectins require high ligand/agonist concentrations, or if the noncanonical role of NFAT is calcium-independent. Inhibition by calcineurin blockade provides indirect evidence against the latter possibility, but doesn't prove it.

Response: Thanks for this suggestion and we agree that inhibition with multiple compounds and with the VIVIT peptide, while highly suggestive, does not fully distinguish between these possibilities. In the scope of other revisions we were not able to address this question experimentally, but have added a new sentence into the Discussion to mention the possibility of a calcium-independent potential role of NFAT and future research directions. We have also added additional quantitation in Figures 6 and 7 to make clear the dynamics and extent of the NFAT/INCA-6 effects and the effects of inhibiting NFATC2 in cell

culture.

Interestingly, in considering the reviewer's comments about the kinetics of NFATC2 activation, calcium dynamics, and exposure to extracellular bacteria, we came across literature implicating NFATC2 in a slower and more sustained process of activation relative to the other NFATc paralogs. This known property of NFATC2 and the associated references are now further highlighted in the discussion (Kar and Parekh, 2015; Kar et al., 2016) and may also relate to the reviewer's idea of persistent or higher extracellular ligand concentrations.

Reviewer #1: As presented, it is difficult to relate the images and the quantitation of angiogenesis. The images shown are low power, and it seems unlikely that such low power images were used to generate the quantitative data used to interpret the results.

Response: In order to assess angiogenesis comprehensively, we did image across the entirety of the animals for the larvae, but of course can magnify specific regions of angiogenesis as well as examining multiple planes. (What is shown are maximum intensity projections). For the larvae, we have now used higher magnification and clearer examples and images of the neovascularization in the figures (still representative of median data points). We also describe in additional detail our blinded quantitation for the larvae and automated (therefore also blinded) quantitation for the cell culture experiments, which revealed robust and significant differences in granuloma-associated angiogenesis as well as NFAT-dependence for VEGFA expression and angiogenesis in the cell culture experiments. The additional images and, in the case of the

cell culture experiments, new automated quantitation, are now incorporated into the main figures, and the STAR Methods now describes more extensively the methods we used for quantitation.

Reviewer #1: Very minor point: the legend for Fig3A is, “NFAT CRISPR Screen”, but results of only two NFATs are shown.

Response: We apologize for using the “screen” terminology, since this evokes large-scale CRISPR screens. We have removed the term and had only meant to imply that, based on the VIVIT and inhibitor results, we considered multiple NFAT paralogs. By combining expression analysis from published datasets with functional analysis of zebrafish CRISPR mutants that we made, we were able to identify *nfatc2a* as the key paralog required for the response.

Reviewer #1: Overall, the manuscript is very well written and organized. The methods, sample numbers, and analyses are all appropriate. As noted above, the link between images demonstrating angiogenesis and the quantitative data could be improved.

The videos are a minor addition without additional labeling on the videos themselves, to highlight the points made.

Response: Thanks for the suggestions, and we have reworked our magnification of the zebrafish images throughout the first few figures so that they are more apparent to the reader. We were trying to include the entirety of the animal to be transparent that we were not selectively showing only a part of each animal, but have now included more zoomed-in images of the angiogenesis that we score. In addition, we have added

detail regarding how we performed blinded quantitation in the STAR Methods section.

Reviewer #2: In this manuscript by Brewer et al. the authors examine the link between mycobacterial infection, host macrophages, and induction of host angiogenesis using zebrafish and human cell culture models. The authors cite a previously published report suggesting that mycobacterium-promoted increased vessel growth may facilitate bacterial growth and dissemination. They also note their own previously published study showing that a mycobacterium-derived cell wall component (TDM) promotes vegfaa production and angiogenesis at the site of mycobacterial infection and granuloma formation. Here, they seek to extend on their previous findings on TDM with new results showing that mycobacteria/TDM lead to activation of NFAT signaling, and NFAT-dependent induction of VEGF, in host macrophages. Although these findings are interesting and potentially highlight new targets for disrupting mycobacterial infection, the findings seem somewhat incremental in significance and a number of additional concerns detract from enthusiasm for this study.

Response: We appreciate the thoughtful feedback and perspective throughout. Although of course this is a judgement call, we disagree with the idea that these findings are incremental. NFAT signaling has not previously been implicated in granuloma-mediated angiogenesis, a physiologically important response to mycobacterial infection. Our initial work in the zebrafish identifying the biological relevance of angiogenesis during pathogenesis (Oehlers et al., 2015) came out just before

concurrent work establishing the relevance of angiogenesis in TB disease in humans and non-human primates, and there is active clinical interest in pursuing these pathways. Using the zebrafish model, we then established the specific mycobacterial lipid required for inciting angiogenesis (Walton et al., 2018), but how that lipid transduced the signal for VEGF production was not understood.

In this work, we provide the first real insight into how macrophage detection of that lipid specifically manipulates the mycobacterium-infected host, leading to VEGFA induction and angiogenesis, a response that enables the bacteria to survive, replicate and disseminate and is therefore of active clinical interest therapeutically for TB. Notably, standard models of TB disease (C57BL/6 mice, for example) do not recapitulate the granulomas seen in humans and non-human primates, and so the zebrafish is one of the few genetically manipulable animal models where this important process can be dissected experimentally. This work 1) identifies activation of a previously unconsidered pathway via NFATC2 as the key downstream transducer of VEGFA production during mycobacterial infection *in vivo* in the zebrafish models and in a human cell line; and 2) assesses the consequences of perturbing NFAT signaling, using both loss-of-function mutants as well as macrophage-specific inhibitory approaches in whole animals *in vivo*. Generally NFAT induction and transduction of pathogen-associated signals have not been explored in myeloid cell biology, and so the link in both humans and zebrafish established here may also spur others to examine myeloid NFAT activation in different contexts during infection and disease.

We detail below both experimental and analytical additions to address the reviewer's additional concerns. In particular, we had not previously explained fully the rigorous blinding methodology that we used throughout, and we have now made this much more explicit up front and in the STAR Methods. In addition, we include new automated computational analysis for the human cell culture work that confirms and strengthens our findings, presenting new panels featuring automated blinded quantitation in Figures 6 and 7.

Reviewer #2: Many of the zebrafish *in vivo* findings presented in Figs 1-5 are based on image data whose resolution and/or magnification do not always allow clear interpretation or clear visualization of the authors conclusions. Despite statistical significance in the authors measurements, the vessel effects appear modest, particularly in the larvae, and it is sometimes difficult to see how they were quantitated from the image examples presented, or how these sorts of limited vessel sprouts and segments, which are unlikely to be carrying much if any additional flow from the authors images, could be significantly influencing mycobacterial growth or dissemination. Since these measurements appear to rely on visual inspection and measurement of vessel/sprout length and are potentially more subjective than quantitative molecular measurement methods, it is also important that they are done fully blinded and that the blinding methods are fully described in the methods section.

Response: We appreciate raising these important points. We had not previously fully explained the rigorous blinding methodology that we had used throughout, and we have now made this much more explicit up

front and in the STAR Methods. In addition, we include new automated computational analysis for the human cell culture work that confirms and strengthens our findings, presenting new panels featuring automated blinded quantitation in Figures 6 and 7.

For the larvae, we have now used more highly magnified and clearer examples and images of the neovascularization in the figures (still representative of median data points). Regardless, our blinded quantitation reveals robust and significant differences in granuloma-associated angiogenesis in blinded analysis. We also note that, because of the timing (Oehlers et al., 2015; Walton et al., 2018), there is a small window of about 2 days (from 3 dpi to 5 dpi) as the response begins only once the initial granuloma has assembled. Unfortunately, the larval system does not permit the longer-term analysis that is available in adults. Although the larval effect sizes are reasonable and consistent, the adult effect size is more dramatic, as we can look over a two-week time-frame; use of the adults also allowed for traditional analysis of bacterial burden. The (blinded) quantitation in adults also certainly underestimates the effect size on angiogenesis, as it includes pre-existing vessels throughout that are included in all analyses. We have no automated way of excluding these pre-existing vessels in the adult and so have used the most conservative approach of including them, which necessarily drives up the background, but we also note that we do see qualitative differences in the *nfatc2* mutants and the macrophage-specific VIVIT line. Again, we had done all the quantitation previously in a completely blinded manner, and now have better described the process and blinding in the STAR Methods section as well

as increased the magnification of some of the images to make the effect more apparent.

Additional text has been added to the STAR Methods to reflect the blinding performed for the experiments and we apologize for any issues with clarity in the existing manuscript. We fully agree that the subjective nature of these types of measurements require blinding to be robust and reproducible. The blinding for these assays were conducted as follows: for assays where experimental blinding was difficult or infeasible (drug treatment assays, transgenic *irg1* fish, adult infections), computational blinding was conducted using either the blindrename.pl script (Salter, 2016) or an in-house developed Python translation of the same (included in the Zenodo submission accompanying this manuscript). This script renames all the files to a random string of characters and generates a keyfile to allow for post hoc matching of blinded images to treatment groups or genotypes. For all assays necessitating post hoc genotyping (larval infection of host mutants), genotypic blinding was used. Measurements were collected on each of the images prior to matching them to the determined genotype.

Reviewer #2: Besides vessel length and a few measurements of mycobacterial CFU in figure 5, no other quantitative data are provided for the zebrafish portion of this study (e.g., rigorous molecular measurement of NFAT activation or VEGF induction in host macrophages), despite the technical feasibility of such measurements, using the transgenic lines available to the authors. Furthermore, if the authors wish to confirm the requirement for macrophages in induction of host angiogenesis in ze-

brafish larvae *in vivo*, they could directly examine this by genetically ablating these cells, which can readily be accomplished in fish. The authors could also perform macrophage-specific targeting of *nfatc2a* and *nfatc3a* using *irg1:Cas9* and guide RNAs targeting each of these genes, to confirm the specific requirement for *nfatc2* in these cells *in vivo*.

Response: These are good suggestions, and we had tried a number of these approaches previously but, in the context of the larva, we are particularly limited by some of the tool availability and development. In our hands the larvae are too small to dissect spatially, and, at this particular developmental stage, there are significant levels of vegfaa production throughout, making whole-animal molecular measurements in this context uninformative. We have tried multiple approaches to generate NFAT reporter transgenic lines, including in-frame fusions and lines based on NFAT binding sites, but have thus far been unable to generate such a line. There were apparent toxic effects from whole-organism reporter transgenes and we were unable to identify transmitting founders from our attempts at macrophage-specific reporters. We are also limited by the lack of antibodies that exist for fish.

We also agree that the macrophage-specific targeting of *nfatc2a* would be a great experiment but do not currently have the technology to do so. We have tried for a number of years now to get a macrophage-specific Cas9 to express reliably, including using the *irg1* promoter, but without success. In collaboration with Qing Deng's laboratory we have published on functional neutrophil-specific Cas9 (Wang et al., 2021),

but macrophages have proven more challenging. There is one report of a macrophage-specific Cas9 from Graham Lieschke using a GAL4-UAS system (Isiaku et al., 2021). The requisite two lines are in Australia and would be an additional number of months to formally import, get through quarantine in our facility and validate, and we (and the zebrafish community generally) have often had issues with transgene silencing using the GAL4-UAS system. So while we recognize that this particular approach would add to the evidence presented in the paper, we don't think it is reasonably within the scope of a revision.

The suggested macrophage depletion experiment is one that we have done previously and published (Oehlers et al., 2015) showing dependence on macrophages for angiogenesis. In the current work, instead of depleting all macrophages generally, we have used the VIVIT peptide, specifically expressed in macrophages, to show that *in vivo*, macrophage-specific expression of VIVIT abrogates granuloma-associated angiogenesis. The conserved nature of the specific *nfatc2* gene in the response to pathogenic mycobacteria in the human cell culture experiments with macrophage-like THP-1 cells also contributes to further validation of the findings.

Reviewer #2: There are some related concerns about the *in vitro* human macrophage quantitative data in Figures 6 and 7. The different replicates of the VEGFA qPCR and ELISA results shown in fig. 6 and supplemental Fig. 4 have strongly differing results, suggesting there may be other variables that are not being adequately controlled in these assays that may be having substantially larger effects than the MTB

and INCA-6 effects, and making selection of a “representative” result somewhat unclear.

Response: We have tried to be complete and transparent throughout, showing each of the experiments in the initial submission (even the outlier that showed much higher induction) and we consistently see the INCA-6 effect. We agree that there is variation in the level of VEGFA induction. We have been completely transparent about the extent of the effect in our original assays (from 5-fold to 50-fold induction, with our main figure representation showing 12 fold induction). We believe that some of the heterogeneity in response may have come from the inherent “clumpiness” of the mycobacteria (they are lipid rich and tend to clump, providing uneven exposure) or properties in how the macrophages themselves were plated prior to infection. To confirm that we could improve on the control of other variables, we ran three additional independent experiments for the revision in which we now coated with poly-D-lysine before plating, as we had had to do for the immunofluorescence to see if this increased overall reproducibility. In the additional experiments, which we now present in Figure 6B and Supplemental Figures 4A and 4B, we see more consistency in the effect size, although the mean and median effect sizes are approximately the same as before. Thus, under two slightly different experimental protocols (one without poly-D-lysine and one with) we observe a consistent induction of VEGFA and consistent inhibition of VEGFA production upon INCA-6 treatment. For simplicity, we have incorporated the second set of experiments as the primary data. We have also included quantitation by three different methods: qRT-PCR, ELISA, and im-

munofluorescence. To make this clearer, we have included new Figures 6G and 6H to better relate the nuclear localization of NFAT to VEGF production and quantitate (in a blinded fashion) the results of INCA-6 exposure. To address the reviewer's concern, we have included data from three independent biological replicates across >5000 cells.

Reviewer #2: Is macrophage number equivalent in all these experiments? How are the values normalized? Despite activation of NFAT being a key point in this manuscript, the authors do not perform rigorous quantitative measurement of NFAT activation by careful measurement of nuclear/-cytoplasmic compartmentalization or other methods (this should be done in conjunction with simultaneous quantitative measurement of VEGFA induction). The data in Fig. 7 are notably deficient in this regard. The VIVIT experiments need validation of the efficacy and specificity of its effects on NFATC activation. And if the authors want assess the role of NFATC2 in their *in vitro* model, it would perhaps be better to select germline CRISPR knockouts of this gene in their cells, and perhaps perform siRNA knockdown to confirm.

Response: For the cell culture experiments, we plate the same number of macrophages for each experiment (2.5×10^5 per well in a 24-well plate, normalized to 5×10^5 cells per mL across all experiments regardless of plate size) and use aliquots of the gamma-irradiated *M. tuberculosis* and INCA-6. As mentioned above, some of the variability between replicates likely arises from the heterogeneity of *M. tuberculosis* exposure, with the extracellular bacteria tending to clump due to their incredibly lipid-rich cell envelope, but we also have gained some consistency via

coating the wells with poly-D-lysine. Notably, we think that the extracellular nature of the exposure may reflect conditions *in vivo*. We now mention in the Discussion that, as we observe in the zebrafish model, engagement of this pathway may be driven by extracellular bacteria.

For VIVIT validation, the peptide itself has been exhaustively validated over the last two decades and the zebrafish target site is 100% conserved for zebrafish *nfatc2a* as well as the other five *nfatc* paralogs. We also have a reliable readout for macrophage-specific VIVIT expression with the fluorescence construct. We agree that, in the future, a more thorough characterization of the effects that result from macrophage-specific VIVIT expression would be of interest and may provide further validation of this tool. The fact that *nfatc2a* mutants recapitulate the VIVIT phenotype is also notable in terms of the success of the VIVIT approach. As described above, we invested a lot of effort in trying to generate reporter lines that would give us an *in vivo* readout of NFAT activation, using multiple approaches, but were unable to generate such lines, and there are no available antibodies that appear to cross-react specifically with zebrafish NFAT (we have tried at least 4 different antibodies). So, while the zebrafish model provides important advantages in dissecting this process and the effects of mutants, drugs, and cell-specific genetic inhibition on the process *in vivo*, NFAT measurements that would be accessible in mammalian cell culture are thus far limited. But we think that the subsequent validation that we perform in mammalian cell culture highlights the relevance of the *in vivo* findings and the conservation of this response during mycobacterial infection.

In terms of the siRNA knockdown idea, we purchased and used the four commercially available *NFATC2* siRNAs from Qiagen. In contrast to the lentiviral approach that we ended up using (new Supplementary Figure 4J), we did not see efficient knockdown of *NFATC2* with these siRNAs. The purchased siRNAs, although supposedly guaranteed by Qiagen, were manufactured in 2011 and so, discouraged from this approach, we pursued the lentiviral constructs to directly assess NFATC2 in mammalian experiments with gamma-irradiated *M. tuberculosis*, to assess whether our *in vivo* findings in the zebrafish extended to human macrophages and to *M. tuberculosis*.

In both Figures 6 and 7, we have now used rigorous, blinded, quantitative computational analysis of immunofluorescence images of NFAT and VEGFA to precisely address the reviewer's concern. The automated quantitation is now described further in the STAR Methods section, the Results section and the figure legends.

In the new Figure 6, panels E, G, and H now in an unbiased, blinded way present quantitative assessment of VEGFA production in conjunction with NFAT nuclear localization in response to gamma-irradiated *M. tuberculosis* in the presence and absence of INCA-6 drug treatment, with analysis of >5000 cells.

In the new Figure 7, panels C, F, G, and H now use the same blinded quantitative approach to simultaneously assess VEGFA induction and NFAT nuclear localization in the context of the specific genetic inhibition of *NFATC2*. This blinded, quantitative analysis further confirms our initial findings and addresses concerns about the robustness of the

effects and the dataset.

Reviewer #2: (In re: clarity of reporting) Mostly yes, but more information on blinding methods for quantitative measurements (most of which appear to involve visual scoring of image data) in this manuscript would be helpful.

Response: As described in more detail above, we have better described our blinding methods for quantitative measurement (STAR Methods and description in Results) as well as applied new extensive and rigorous blinded and automated quantitation for Figures 6 and 7 in additional figure panels as requested by the Reviewer.

Bibliography

In this bibliography, a subset of the entries are in **bold** to indicate their relative importance and their potential interest for the reader. Other entries are provided annotation below the citation to provide additional context. These are distributed throughout this section and I hope they are of use to the reader in finding additional resources. Some of these will be publications that are directly relevant to the work presented in this document, others will be those publications that, through their methodology or findings, inspired me in the conduct of my own work.

A-Gonzalez, N., J. A. Quintana, S. Garcia-Silva, M. Mazariegos, A. Gonzalez de la Aleja, J. A. Nicolas-Avila, W. Walter, J. M. Adrover, G. Crainiciuc, V. K. Kuchroo, C. V. Rothlin, H. Peinado, A. Castrillo, M. Ricote, and A. Hidalgo (2017). “Phagocytosis imprints heterogeneity in tissue-resident macrophages”. In: *J Exp Med* 214.5, pp. 1281–1296.

– This study uncovers an interesting role for the phagocytic history of the macrophage in mediating its capacity for future responses. Specifically, this phagocytosis blunted the ability for macrophages to induce future inflammatory responses, rendering them comparatively less able to control pathogens than macrophages that lacked this phagocytic history. This would imply that “new” macrophages from the bone marrow are pre-disposed toward inflammatory responses until they have been educated by the tissue environment and phagocytosis to be more anti-inflammatory.

Abdelilah, S., E. Mountcastle-Shah, M. Harvey, L. Solnica-Krezel, A. F. Schier, D. L. Stemple, J. Malicki, S. C. Neuhauss, F. Zwartkruis, D. Y. Stainier, Z. Rangini, and W. Driever (1996). “Mutations affecting neural survival in the zebrafish *Danio rerio*”. In: *Development* 123, pp. 217–27.

Abhinand, C. S., R. Raju, S. J. Soumya, P. S. Arya, and P. R. Sudhakaran (2016). “VEGF-A/VEGFR2 signaling network in endothelial cells relevant to angiogenesis”. In: *J Cell Commun Signal* 10.4, pp. 347–354.

Abramovitch, R. B. (2018). “*Mycobacterium tuberculosis* Reporter Strains as Tools for Drug Discovery and Development”. In: *IUBMB Life* 70.9, pp. 818–825.

Abramovitch, R. B., K. H. Rohde, F. F. Hsu, and D. G. Russell (2011). “aprABC: a *Mycobacterium tuberculosis* complex-specific locus that modulates pH-driven adaptation to the macrophage phagosome”. In: *Mol Microbiol* 80.3, pp. 678–94.

Abukhalid, N., S. Islam, R. Ndzeidze, and L. E. Bermudez (2021). “*Mycobacterium avium* subsp. *hominissuis* Interactions with Macrophage Killing Mechanisms”. In: *Pathogens* 10.11.

Adams, K. N., K. Takaki, L. E. Connolly, H. Wiedenhoft, K. Winglee, O. Humbert, P. H. Edelstein, C. L. Cosma, and L. Ramakrishnan (2011). “Drug tolerance in replicating mycobacteria mediated by a macrophage-induced efflux mechanism”. In: *Cell* 145.1, pp. 39–53.

Adams, R. H. and K. Alitalo (2007). “Molecular regulation of angiogenesis and lymphangiogenesis”. In: *Nat Rev Mol Cell Biol* 8.6, pp. 464–78.

Adati, N., M. C. Huang, T. Suzuki, H. Suzuki, and T. Kojima (2009). “High-resolution analysis of aberrant regions in autosomal chromosomes in human leukemia THP-1 cell line”. In: *BMC Res Notes* 2, p. 153.

Adhyapak, P., A. T. Srivatsav, M. Mishra, A. Singh, R. Narayan, and S. Kapoor (2020). “Dynamical Organization of Compositionally Distinct Inner and Outer Membrane Lipids of Mycobacteria”. In: *Biophys J* 118.6, pp. 1279–1291.

Agarwal, P., T. W. Combes, F. Shojaee-Moradie, B. Fielding, S. Gordon, V. Mizrahi, and F. O. Martinez (2020). “Foam Cells Control *Mycobacterium tuberculosis* Infection”. In: *Front Microbiol* 11, p. 1394.

Agarwal, P., S. Gordon, and F. O. Martinez (2021). “Foam Cell Macrophages in Tuberculosis”. In: *Front Immunol* 12, p. 775326.

Agarwal, S., O. Avni, and A. Rao (2000). “Cell-type-restricted binding of the transcription factor NFAT to a distal IL-4 enhancer *in vivo*”. In: *Immunity* 12.6, pp. 643–52.

– This pioneering study identified the cell-type restricted roles for NFATC2 and found that this protein is only able to bind the IL-4 and IFN- γ promoters in T_H2 and T_H1 cells, respectively. This was the first major effort at identifying the specific

factors that allow NFAT proteins to exert cell-type specific activity beyond simply their expression. This study was very technically advanced for the time and skillfully defined the ways in which a single protein can regulate opposing transcriptional responses based on higher-order chromatin structural mechanisms.

Agarwal, S., P. Kulshreshtha, D. Bambah Mukku, and R. Bhatnagar (2008). “alpha-Enolase binds to human plasminogen on the surface of *Bacillus anthracis*”. In: *Biochim Biophys Acta* 1784.7-8, pp. 986–94.

Ahlmann-Eltze, C. and I. Patil (2021). “ggsignif: Significance Brackets for ggplot2”. In.

Ahluwalia, P. K., R. K. Pandey, P. K. Sehajpal, and V. K. Prajapati (2017). “Perturbed microRNA Expression by *Mycobacterium tuberculosis* Promotes Macrophage Polarization Leading to Pro-survival Foam Cell”. In: *Front Immunol* 8, p. 107.

Akbar, R., H. Bashour, P. Rawat, P. A. Robert, E. Smorodina, T. S. Cotet, K. Flem-Karlsen, R. Frank, B. B. Mehta, M. H. Vu, T. Zengin, J. Gutierrez-Marcos, F. Lund-Johansen, J. T. Andersen, and V. Greiff (2022a). “Progress and challenges for the machine learning-based design of fit-for-purpose monoclonal antibodies”. In: *MAbs* 14.1, p. 2008790.

Akbar, R., P. A. Robert, C. R. Weber, M. Widrich, R. Frank, M. Pavlovic, L. Scheffer, M. Chernigovskaya, I. Snapkov, A. Slabodkin, B. B. Mehta, E. Miho, F. Lund-Johansen, J. T. Andersen, S. Hochreiter, I. Hobæk Haff, G. Klambauer, G. K. Sandve, and V. Greiff (2022b). “In silico proof of principle of machine learning-based antibody design at unconstrained scale”. In: *MAbs* 14.1, p. 2031482.

Akdis, M., S. Burgler, R. Crameri, T. Eiwegger, H. Fujita, E. Gomez, S. Klunker, N. Meyer, L. O’Mahony, O. Palomares, C. Rhyner, N. Ouaked, A. Schaffartzik, W. Van De Veen, S. Zeller, M. Zimmermann, and C. A. Akdis (2011). “Interleukins, from 1 to 37, and interferon-gamma: receptors, functions, and roles in diseases”. In: *J Allergy Clin Immunol* 127.3, 701–21 e1–70.

Akira, S., H. Isshiki, T. Sugita, O. Tanabe, S. Kinoshita, Y. Nishio, T. Nakajima, T. Hirano, and T. Kishimoto (1990). “A nuclear factor for IL-6 expression (NF-IL6) is a member of a C/EBP family”. In: *EMBO J* 9.6, pp. 1897–906.

Akira, S. and K. Takeda (2004). “Toll-like receptor signalling”. In: *Nat Rev Immunol* 4.7, pp. 499–511.

Akwii, R. G. and C. M. Mikelis (2021). “Targeting the Angiopoietin/Tie Pathway: Prospects for Treatment of Retinal and Respiratory Disorders”. In: *Drugs* 81.15, pp. 1731–1749.

Al-Daraji, W. I., K. R. Grant, K. Ryan, A. Saxton, and N. J. Reynolds (2002). “Localization of calcineurin/NFAT in human skin and psoriasis and inhibition of calcineurin/NFAT activation in human keratinocytes by cyclosporin A”. In: *J Invest Dermatol* 118.5, pp. 779–88.

Albensi, B. C. (2019). “What Is Nuclear Factor Kappa B (NF-kappaB) Doing in and to the Mitochondrion?” In: *Front Cell Dev Biol* 7, p. 154.

Alderwick, L. J., J. Harrison, G. S. Lloyd, and H. L. Birch (2015). “The Mycobacterial Cell Wall–Peptidoglycan and Arabinogalactan”. In: *Cold Spring Harb Perspect Med* 5.8, a021113.

Alenton, R. R., K. Koiwai, K. Miyaguchi, H. Kondo, and I. Hirono (2017). “Pathogen recognition of a novel C-type lectin from *Marsupenaeus japonicus* reveals the divergent sugar-binding specificity of QAP motif”. In: *Sci Rep* 7, p. 45818.

Alexander, John White (1895). *Repose*.

Algood, H. M., P. L. Lin, and J. L. Flynn (2005). “Tumor necrosis factor and chemokine interactions in the formation and maintenance of granulomas in tuberculosis”. In: *Clin Infect Dis* 41 Suppl 3, S189–93.

Alidzanovic, L., P. Starlinger, D. Schauer, T. Maier, A. Feldman, E. Buchberger, J. Stift, U. Koeck, L. Pop, B. Gruenberger, T. Gruenberger, and C. Brostjan (2016). “The VEGF rise in blood of bevacizumab patients is not based on tumor escape but a host-blockade of VEGF clearance”. In: *Oncotarget* 7.35, pp. 57197–57212.

Alitalo, K., T. Tammela, and T. V. Petrova (2005). “Lymphangiogenesis in development and human disease”. In: *Nature* 438.7070, pp. 946–53.

Almeida, M. P., J. M. Welker, S. Siddiqui, J. Luiken, S. C. Ekker, K. J. Clark, J. J. Essner, and M. McGrail (2021). “Endogenous zebrafish proneural Cre drivers generated by CRISPR/Cas9 short homology directed targeted integration”. In: *Sci Rep* 11.1, p. 1732.

Amatruda, J. F., J. L. Shepard, H. M. Stern, and L. I. Zon (2002). “Zebrafish as a cancer model system”. In: *Cancer Cell* 1.3, pp. 229–31.

Amberg, G. C., C. F. Rossow, M. F. Navedo, and L. F. Santana (2004). “NFATc3 regulates Kv2.1 expression in arterial smooth muscle”. In: *J Biol Chem* 279.45, pp. 47326–34.

Amersfoort, J., G. Eelen, and P. Carmeliet (2022). “Immunomodulation by endothelial cells - partnering up with the immune system?” In: *Nat Rev Immunol* 22.9, pp. 576–588.

Amores, A., J. Catchen, A. Ferrara, Q. Fontenot, and J. H. Postlethwait (2011). “Genome evolution and meiotic maps by massively parallel DNA sequencing: spotted gar, an outgroup for the teleost genome duplication”. In: *Genetics* 188.4, pp. 799–808.

Amsterdam, A., S. Burgess, G. Golling, W. Chen, Z. Sun, K. Townsend, S. Farrington, M. Haldi, and N. Hopkins (1999). “A large-scale insertional mutagenesis screen in zebrafish”. In: *Genes Dev* 13.20, pp. 2713–24.

An, S., S. Kim, Y. Huh, T. R. Lee, H. K. Kim, K. L. Park, and H. C. Eun (2009). “Expression of surface markers on the human monocytic leukaemia cell line, THP-1, as indicators for the sensitizing potential of chemicals”. In: *Contact Dermatitis* 60.4, pp. 185–92.

Andersen, P. and T. M. Doherty (2005). “The success and failure of BCG - implications for a novel tuberculosis vaccine”. In: *Nat Rev Microbiol* 3.8, pp. 656–62.

Angosto, D. and V. Mulero (2014). “The zebrafish as a model to study the inflammasome”. In: *Inflammasome* 1.1.

Antonelli, L. R., A. Gigliotti Rothfuchs, R. Goncalves, E. Roffe, A. W. Cheever, A. Bafica, A. M. Salazar, C. G. Feng, and A. Sher (2010). “Intranasal Poly-IC treatment exacerbates tuberculosis in mice through the pulmonary recruitment of a pathogen-permissive monocyte/macrophage population”. In: *J Clin Invest* 120.5, pp. 1674–82.

Appelberg, R., D. Moreira, P. Barreira-Silva, M. Borges, L. Silva, R. J. Dinis-Oliveira, M. Resende, M. Correia-Neves, M. B. Jordan, N. C. Ferreira, A. J. Abrunhosa, and R. Silvestre (2015). “The Warburg effect in mycobacterial granulomas is depen-

dent on the recruitment and activation of macrophages by interferon-gamma". In: *Immunology* 145.4, pp. 498–507.

Apt, A. S. (2011). "Are mouse models of human mycobacterial diseases relevant? Genetics says: 'yes!'" In: *Immunology* 134.2, pp. 109–15.

Aramburu, J., L. Azzoni, A. Rao, and B. Perussia (1995). "Activation and expression of the nuclear factors of activated T cells, NFATp and NFATc, in human natural killer cells: regulation upon CD16 ligand binding". In: *J Exp Med* 182.3, pp. 801–10.

Aramburu, J., F. Garcia-Cozar, A. Raghavan, H. Okamura, A. Rao, and P. G. Hogan (1998). "Selective inhibition of NFAT activation by a peptide spanning the calcineurin targeting site of NFAT". In: *Mol Cell* 1.5, pp. 627–37.

Aramburu, J., M. B. Yaffe, C. Lopez-Rodriguez, L. C. Cantley, P. G. Hogan, and A. Rao (1999). "Affinity-driven peptide selection of an NFAT inhibitor more selective than cyclosporin A". In: *Science* 285.5436, pp. 2129–33.

– *Another study from Anjana Rao's lab, this time identifying a peptide, VIVIT, that is able to interfere with the interaction between calcineurin and NFAT. This is a critical tool in the execution of my project and, without these findings, none of the macrophage-specific phenotypes would have been possible to address in vivo.*

Arango Duque, G. and A. Descoteaux (2014). "Macrophage cytokines: involvement in immunity and infectious diseases". In: *Front Immunol* 5, p. 491.

Arbibe, L., J. P. Mira, N. Teusch, L. Kline, M. Guha, N. Mackman, P. J. Godowski, R. J. Ulevitch, and U. G. Knaus (2000). "Toll-like receptor 2-mediated NF-kappa B activation requires a Rac1-dependent pathway". In: *Nat Immunol* 1.6, pp. 533–40.

Archer, J. S. and D. F. Archer (2002). "Oral contraceptive efficacy and antibiotic interaction: a myth debunked". In: *J Am Acad Dermatol* 46.6, pp. 917–23.

Armant, M. A. and M. J. Fenton (2002). "Toll-like receptors: a family of pattern-recognition receptors in mammals". In: *Genome Biol* 3.8, REVIEWS3011.

Armulik, A., G. Genove, and C. Betsholtz (2011). "Pericytes: developmental, physiological, and pathological perspectives, problems, and promises". In: *Dev Cell* 21.2, pp. 193–215.

Aronson, J. D. (1926). "Spontaneous Tuberculosis in Salt Water Fish". In: *The Journal of Infectious Diseases* 39.4, pp. 315–320.

– *This paper was the first description of *Mycobacterium marinum* and established the histological findings on the granuloma that have served the field well ever since and defined the etiology of fish tuberculosis.*

Arthur, J. S. and S. C. Ley (2013). "Mitogen-activated protein kinases in innate immunity". In: *Nat Rev Immunol* 13.9, pp. 679–92.

Arts, R. J. W., A. Carvalho, C. La Rocca, C. Palma, F. Rodrigues, R. Silvestre, J. Kleinnijenhuis, E. Lachmandas, L. G. Goncalves, A. Belinha, C. Cunha, M. Oosting, L. A. B. Joosten, G. Matarese, R. van Crevel, and M. G. Netea (2016). "Immunometabolic Pathways in BCG-Induced Trained Immunity". In: *Cell Rep* 17.10, pp. 2562–2571.

Arts, R. J. W., Sjcfm Moorlag, B. Novakovic, Y. Li, S. Y. Wang, M. Oosting, V. Kumar, R. J. Xavier, C. Wijmenga, L. A. B. Joosten, Cbem Reusken, C. S. Benn, P. Aaby, M. P. Koopmans, H. G. Stunnenberg, R. van Crevel, and M. G. Netea (2018). "BCG Vaccination Protects against Experimental Viral Infection in Humans through the Induction of Cytokines Associated with Trained Immunity". In: *Cell Host Microbe* 23.1, 89–100 e5.

Arunachalam, M., M. Raja, C. Vijayakumar, P. Malaiammal, and R. L. Mayden (2013). "Natural history of zebrafish (*Danio rerio*) in India". In: *Zebrafish* 10.1, pp. 1–14.

Asagiri, M., K. Sato, T. Usami, S. Ochi, H. Nishina, H. Yoshida, I. Morita, E. F. Wagner, T. W. Mak, E. Serfling, and H. Takayanagi (2005). "Autoamplification of NFATc1 expression determines its essential role in bone homeostasis". In: *J Exp Med* 202.9, pp. 1261–9.

– *This was the first paper, to my knowledge, to investigate NFAT regulation of NFAT expression and found that NFATC1 acts in a feed-forward loop to drive its own expression to facilitate osteoclast differentiation. This principally opens the possibility that other NFAT isoforms may regulate either their own expression or that of other NFATs; this also begs the question whether other transcription factors can dynamically induce NFAT expression after some stimulus, although this mechanism of NFAT activation is comparatively understudied.*

Asmar, S., M. Sassi, M. Phelipeau, and M. Drancourt (2016). "Inverse correlation

between salt tolerance and host-adaptation in mycobacteria". In: *BMC Res Notes* 9, p. 249.

– *A very interesting study on the evolutionary rationale behind the divergent ability for mycobacteria to survive in the environment and within hosts. The ability to robustly tolerate salt is inversely correlated with success within hosts, so non-professionally pathogenic mycobacteria face severe fitness costs from host residence.*

Astarie-Dequeker, C., E. N. N'Diaye, V. Le Cabec, M. G. Rittig, J. Prandi, and I. Maridonneau-Parini (1999). "The mannose receptor mediates uptake of pathogenic and nonpathogenic mycobacteria and bypasses bactericidal responses in human macrophages". In: *Infect Immun* 67.2, pp. 469–77.

Atluri, P. and Y. J. Woo (2008). "Pro-angiogenic cytokines as cardiovascular therapeutics: assessing the potential". In: *BioDrugs* 22.4, pp. 209–22.

Auchampach, J. A. (2007). "Adenosine receptors and angiogenesis". In: *Circ Res* 101.11, pp. 1075–7.

Augustin, H. G., G. Y. Koh, G. Thurston, and K. Alitalo (2009). "Control of vascular morphogenesis and homeostasis through the angiopoietin-Tie system". In: *Nat Rev Mol Cell Biol* 10.3, pp. 165–77.

Aurora, A. B., D. Biyashev, Y. Mirochnik, T. A. Zaichuk, C. Sanchez-Martinez, M. A. Renault, D. Losordo, and O. V. Volpert (2010). "NF-kappaB balances vascular regression and angiogenesis via chromatin remodeling and NFAT displacement". In: *Blood* 116.3, pp. 475–84.

– *The question of how multiple immunologically relevant transcription factors interact with one another in various cell types is an important question in order to understand the actual role of these pathways in native tissues. In this study, the interactions between NF-κB and NFAT were evaluated and it was found that NF-κB induced vascular endothelial cell death via induction of FASL while NFAT promoted cell survival through cFLIP. The relative balance of these effect determined the overall angiogenic effect and evokes a model wherein the balance between these transcription factors can regulate major aspects of diverse responses across many tissues.*

Ausubel, F. M. (2005). "Are innate immune signaling pathways in plants and animals conserved?" In: *Nat Immunol* 6.10, pp. 973–9.

- Axelrod, S., H. Oschkinat, J. Enders, B. Schlegel, V. Brinkmann, S. H. Kaufmann, A. Haas, and U. E. Schaible (2008). “Delay of phagosome maturation by a mycobacterial lipid is reversed by nitric oxide”. In: *Cell Microbiol* 10.7, pp. 1530–45.
- Babunovic, G. H., M. A. DeJesus, B. Bosch, M. R. Chase, T. Barbier, A. K. Dickey, B. D. Bryson, J. M. Rock, and S. M. Fortune (2022). “CRISPR Interference Reveals That All-Trans-Retinoic Acid Promotes Macrophage Control of *Mycobacterium tuberculosis* by Limiting Bacterial Access to Cholesterol and Propionyl Coenzyme A”. In: *mBio*, e0368321.
- Bach, R. D. and O. Dmitrenko (2004). “Strain energy of small ring hydrocarbons. Influence of C-h bond dissociation energies”. In: *J Am Chem Soc* 126.13, pp. 4444–52.
- Baer, C., M. L. Squadrito, M. L. Iruela-Arispe, and M. De Palma (2013). “Reciprocal interactions between endothelial cells and macrophages in angiogenic vascular niches”. In: *Exp Cell Res* 319.11, pp. 1626–34.
- Baeuerle, P. A. and T. Henkel (1994). “Function and activation of NF-kappa B in the immune system”. In: *Annu Rev Immunol* 12, pp. 141–79.
- Bagwell, J., J. Norman, K. Ellis, B. Peskin, J. Hwang, X. Ge, S. V. Nguyen, S. K. McMenamin, D. Y. Stainier, and M. Bagnat (2020). “Notochord vacuoles absorb compressive bone growth during zebrafish spine formation”. In: *Elife* 9.
- Bahary, N., K. Goishi, C. Stuckenholz, G. Weber, J. Leblanc, C. A. Schafer, S. S. Berman, M. Klagsbrun, and L. I. Zon (2007). “Duplicate VegfA genes and orthologues of the KDR receptor tyrosine kinase family mediate vascular development in the zebrafish”.** In: *Blood* 110.10, pp. 3627–36.
- Baier, H., S. Klostermann, T. Trowe, R. O. Karlstrom, C. Nusslein-Volhard, and F. Bonhoeffer (1996). “Genetic dissection of the retinotectal projection”. In: *Development* 123, pp. 415–25.
- Bain, C. C. and A. Schridde (2018). “Origin, Differentiation, and Function of Intestinal Macrophages”. In: *Front Immunol* 9, p. 2733.
- Bakkers, J. (2011). “Zebrafish as a model to study cardiac development and human cardiac disease”. In: *Cardiovasc Res* 91.2, pp. 279–88.

- Baksh, S., H. R. Widlund, A. A. Frazer-Abel, J. Du, S. Fosmire, D. E. Fisher, J. A. DeCaprio, J. F. Modiano, and S. J. Burakoff (2002). “NFATc2-mediated repression of cyclin-dependent kinase 4 expression”. In: *Mol Cell* 10.5, pp. 1071–81.
- Balabanov, R., R. Washington, J. Wagnerova, and P. Dore-Duffy (1996). “CNS microvascular pericytes express macrophage-like function, cell surface integrin alpha M, and macrophage marker ED-2”. In: *Microvasc Res* 52.2, pp. 127–42.
- Balch, S. G., A. J. McKnight, M. F. Seldin, and S. Gordon (1998). “Cloning of a novel C-type lectin expressed by murine macrophages”. In: *J Biol Chem* 273.29, pp. 18656–64.
- Balciunas, D., K. J. Wangensteen, A. Wilber, J. Bell, A. Geurts, S. Sivasubbu, X. Wang, P. B. Hackett, D. A. Largaespada, R. S. McIvor, and S. C. Ekker (2006). “Harnessing a high cargo-capacity transposon for genetic applications in vertebrates”. In: *PLoS Genet* 2.11, e169.
- Balik-Meisner, M., L. Truong, E. H. Scholl, R. L. Tanguay, and D. M. Reif (2018). “Population genetic diversity in zebrafish lines”. In: *Mamm Genome* 29.1-2, pp. 90–100.
- Balka, K. R. and D. De Nardo (2019). “Understanding early TLR signaling through the Myddosome”. In: *J Leukoc Biol* 105.2, pp. 339–351.
- Banerjee, S., A. Nandyala, R. Podili, V. M. Katoch, K. J. Murthy, and S. E. Hasnain (2004). “*Mycobacterium tuberculosis* (Mtb) isocitrate dehydrogenases show strong B cell response and distinguish vaccinated controls from TB patients”. In: *Proc Natl Acad Sci U S A* 101.34, pp. 12652–7.
- Banerjee, S., A. K. Nandyala, P. Raviprasad, N. Ahmed, and S. E. Hasnain (2007). “Iron-dependent RNA-binding activity of *Mycobacterium tuberculosis* aconitase”. In: *J Bacteriol* 189.11, pp. 4046–52.
- Banos-Mateos, S., A. M. van Roon, U. F. Lang, S. L. Maslen, J. M. Skehel, and M. H. Lamers (2017). “High-fidelity DNA replication in *Mycobacterium tuberculosis* relies on a trinuclear zinc center”. In: *Nat Commun* 8.1, p. 855.
- Bansal-Mutalik, R. and H. Nikaido (2014). “Mycobacterial outer membrane is a lipid bilayer and the inner membrane is unusually rich in diacyl phosphatidylinositol dimannosides”. In: *Proc Natl Acad Sci U S A* 111.13, pp. 4958–63.

Baraniskin, A., B. Buchberger, C. Pox, U. Graeven, J. W. Holch, W. Schmiegel, and V. Heinemann (2019). “Efficacy of bevacizumab in first-line treatment of metastatic colorectal cancer: A systematic review and meta-analysis”. In: *Eur J Cancer* 106, pp. 37–44.

Barbay, V., M. Houssari, M. Mekki, S. Banquet, F. Edwards-Levy, J. P. Henry, A. Dumesnil, S. Adriouch, C. Thuillez, V. Richard, and E. Brakenhielm (2015). “Role of M2-like macrophage recruitment during angiogenic growth factor therapy”. In: *Angiogenesis* 18.2, pp. 191–200.

– *Pro-angiogenic therapy is likely to have many different unintended effects through enhancement of global VEGFA signaling and this study identifies a major effect, which is that the implantation of VEGFA in ischemia results in increased recruitment of M1-like macrophages, arguing for a chemoattractive role for VEGFA toward inflammatory macrophage subsets.*

Barber, A. E., B. A. Fleming, and M. A. Mulvey (2016). “Similarly Lethal Strains of Extraintestinal Pathogenic *Escherichia coli* Trigger Markedly Diverse Host Responses in a Zebrafish Model of Sepsis”. In: *mSphere* 1.2.

Barberis, I., N. L. Bragazzi, L. Galluzzo, and M. Martini (2017). “The history of tuberculosis: from the first historical records to the isolation of Koch’s bacillus”. In: *J Prev Med Hyg* 58.1, E9–E12.

Barditch-Crovo, P., C. B. Trapnell, E. Ette, H. A. Zatur, J. Coresh, L. E. Rocco, C. W. Hendrix, and C. Flexner (1999). “The effects of rifampin and rifabutin on the pharmacokinetics and pharmacodynamics of a combination oral contraceptive”. In: *Clin Pharmacol Ther* 65.4, pp. 428–38.

Barkan, D., D. Hedhli, H. G. Yan, K. Huygen, and M. S. Glickman (2012). “*Mycobacterium tuberculosis* lacking all mycolic acid cyclopropanation is viable but highly attenuated and hyperinflammatory in mice”. In: *Infect Immun* 80.6, pp. 1958–68.

Barkan, D., Z. Liu, J. C. Sacchettini, and M. S. Glickman (2009). “Mycolic acid cyclopropanation is essential for viability, drug resistance, and cell wall integrity of *Mycobacterium tuberculosis*”. In: *Chem Biol* 16.5, pp. 499–509.

Barkan, D., V. Rao, G. D. Sukenick, and M. S. Glickman (2010). “Redundant function of cmaA2 and mmaA2 in *Mycobacterium tuberculosis* cis cyclopropanation of oxygenated mycolates”. In: *J Bacteriol* 192.14, pp. 3661–8.

- Barnes, D. D., M. L. E. Lundahl, E. C. Lavelle, and E. M. Scanlan (2017). “The Emergence of Phenolic Glycans as Virulence Factors in *Mycobacterium tuberculosis*”. In: *ACS Chem Biol* 12.8, pp. 1969–1979.
- Bartholomew, J. W. and T. Mittwer (1952). “The Gram stain”. In: *Bacteriol Rev* 16.1, pp. 1–29.
- Batt, S. M., D. E. Minnikin, and G. S. Besra (2020). “The thick waxy coat of mycobacteria, a protective layer against antibiotics and the host’s immune system”. In: *Biochem J* 477.10, pp. 1983–2006.
- Bauer, B. and A. Steinle (2017). “HemITAM: A single tyrosine motif that packs a punch”. In: *Sci Signal* 10.508.
- Baumgart, S., N. M. Chen, J. T. Siveke, A. Konig, J. S. Zhang, S. K. Singh, E. Wolf, M. Bartkuhn, I. Esposito, E. Hessmann, J. Reinecke, J. Nikorowitsch, M. Brunner, G. Singh, M. E. Fernandez-Zapico, T. Smyrk, W. R. Bamlet, M. Eilers, A. Neesse, T. M. Gress, D. D. Billadeau, D. Tuveson, R. Urrutia, and V. Ellenrieder (2014). “Inflammation-induced NFATc1-STAT3 transcription complex promotes pancreatic cancer initiation by KrasG12D”. In: *Cancer Discov* 4.6, pp. 688–701.
- Baxt, L. A., A. C. Garza-Mayers, and M. B. Goldberg (2013). “Bacterial subversion of host innate immune pathways”. In: *Science* 340.6133, pp. 697–701.
- Beckham, K. S., L. Ciccarelli, C. M. Bunduc, H. D. Mertens, R. Ummels, W. Lugmayr, J. Mayr, M. Rettel, M. M. Savitski, D. I. Svergun, W. Bitter, M. Wilmanns, T. C. Marlovits, A. H. Parret, and E. N. Houben (2017). “Structure of the mycobacterial ESX-5 type VII secretion system membrane complex by single-particle analysis”. In: *Nat Microbiol* 2, p. 17047.
- Beckwith, K. S., M. S. Beckwith, S. Ullmann, R. S. Saetra, H. Kim, A. Marstad, S. E. Asberg, T. A. Strand, M. Haug, M. Niederweis, H. A. Stenmark, and T. H. Flo (2020). “Plasma membrane damage causes NLRP3 activation and pyroptosis during *Mycobacterium tuberculosis* infection”. In: *Nat Commun* 11.1, p. 2270.
- Beerman, R. W., M. A. Matty, G. G. Au, L. L. Looger, K. R. Choudhury, P. J. Keller, and D. M. Tobin (2015). “Direct *in vivo* Manipulation and Imaging of Calcium Transients in Neutrophils Identify a Critical Role for Leading-Edge Calcium Flux”. In: *Cell Rep* 13.10, pp. 2107–17.

Behar, S. M., C. J. Martin, M. G. Booty, T. Nishimura, X. Zhao, H. X. Gan, M. Divangahi, and H. G. Remold (2011). “Apoptosis is an innate defense function of macrophages against *Mycobacterium tuberculosis*”. In: *Mucosal Immunol* 4.3, pp. 279–87.

Behler, F., R. Maus, J. Bohling, S. Knippenberg, G. Kirchhof, M. Nagata, D. Jonigk, N. Izykowski, L. Magel, T. Welte, S. Yamasaki, and U. A. Maus (2015). “Macrophage-inducible C-type lectin MinCLE-expressing dendritic cells contribute to control of splenic *Mycobacterium bovis* BCG infection in mice”. In: *Infect Immun* 83.1, pp. 184–96.

– This study identified an unequivocal role for MCL in regulating host immunity to mycobacteria, unlike the confused results from Behler, Steinwede, Balboa, Ueberberg, Maus, Kirchhof, Yamasaki, Welte, and Maus (2012), Lee, Kang, Yan, Lee, Jeon, Cho, and Kim (2012), and Heitmann, Schoenen, Ehlers, Lang, and Holscher (2013). This indicates the basal responses through MCL are, potentially, more important for host control of infection than is MINCLE per se, which is consistent with an important role for MCL-MINCLE dimers in transducing important signals, which MCL may be able to do alone in the absence of MINCLE and in the presence of high concentrations of ligand.

Behler, F., K. Steinwede, L. Balboa, B. Ueberberg, R. Maus, G. Kirchhof, S. Yamasaki, T. Welte, and U. A. Maus (2012). “Role of MinCLE in alveolar macrophage-dependent innate immunity against mycobacterial infections in mice”. In: *J Immunol* 189.6, pp. 3121–9.

Behling, C. A., B. Bennett, K. Takayama, and R. L. Hunter (1993). “Development of a trehalose 6,6'-dimycolate model which explains cord formation by *Mycobacterium tuberculosis*”. In: *Infect Immun* 61.6, pp. 2296–303.

Behr, M. A., P. H. Edelstein, and L. Ramakrishnan (2018). “Revisiting the timetable of tuberculosis”. In: *BMJ* 362, k2738.

– Does latency occur in tuberculosis and, if it does, does it matter? In a controversial study, the authors dissect the historical data on this question and conclude that the vast majority of tuberculosis cases are incident to a primary exposure and transition into active disease quickly thereafter. This argues that latency, despite the specter it casts over the field at large, is only a minor contributor to the burden of tuberculosis disease and argues for deprioritizing research on latency to focus on the effects that drive primary infection after exposure. Given that the statistics are often cast that up to one-third of the global population is infected with tuberculosis, the contribution of this pool of potentially latently infected people to the actual disease burden appears

to be nominal and could conceivably be ignored in future discussions on how to best manage and eradicate tuberculosis.

Bekerkunst, A. (1968). "Acute granulomatous response produced in mice by trehalose-6,6-dimycolate". In: *J Bacteriol* 96.4, pp. 958–61.

Bekpen, C., T. Marques-Bonet, C. Alkan, F. Antonacci, M. B. Leogrande, M. Ventura, J. M. Kidd, P. Siswara, J. C. Howard, and E. E. Eichler (2009). "Death and resurrection of the human IRGM gene". In: *PLoS Genet* 5.3, e1000403.

Bekpen, C., R. J. Xavier, and E. E. Eichler (2010). "Human IRGM gene "to be or not to be"". In: *Semin Immunopathol* 32.4, pp. 437–44.

Bellamy, R., C. Ruwende, T. Corrah, K. P. McAdam, H. C. Whittle, and A. V. Hill (1998). "Variations in the NRAMP1 gene and susceptibility to tuberculosis in West Africans". In: *N Engl J Med* 338.10, pp. 640–4.

Ben-Ami, R. (2013). "Angiogenesis at the mold-host interface: a potential key to understanding and treating invasive aspergillosis". In: *Future Microbiol* 8.11, pp. 1453–62.

Ben Shoham, A., G. Malkinson, S. Krief, Y. Shwartz, Y. Ely, N. Ferrara, K. Yaniv, and E. Zelzer (2012). "S1P1 inhibits sprouting angiogenesis during vascular development". In: *Development* 139.20, pp. 3859–69.

Benard, E. L., A. M. van der Sar, F. Ellett, G. J. Lieschke, H. P. Spaink, and A. H. Meijer (2012). "Infection of zebrafish embryos with intracellular bacterial pathogens". In: *J Vis Exp* 61.

Bendickova, K., F. Tidu, M. De Zuani, M. H. Kohoutkova, I. Andrejcinova, A. Pompeiano, S. Belaskova, G. Forte, T. Zelante, and J. Fric (2020). "Calcineurin inhibitors reduce NFAT-dependent expression of antifungal pentraxin-3 by human monocytes". In: *J Leukoc Biol* 107.3, pp. 497–508.

Bendickova, K., F. Tidu, and J. Fric (2017). "Calcineurin-NFAT signalling in myeloid leucocytes: new prospects and pitfalls in immunosuppressive therapy". In: *EMBO Mol Med* 9.8, pp. 990–999.

– This is probably the most recent review on the subject of myeloid NFAT signaling, although the thesis of this is that clinical calcineurin inhibitors may impede myeloid

responses rather than the opportunities for inhibition of myeloid NFAT signaling in modulating responses for host benefit.

Benedito, A. B., M. Lehtinen, R. Massol, U. G. Lopes, T. Kirchhausen, A. Rao, and A. Bonni (2005). “The transcription factor NFAT3 mediates neuronal survival”. In: *J Biol Chem* 280.4, pp. 2818–25.

Bentrup, K. Honer zu and D. G. Russell (2001). “Mycobacterial persistence: adaptation to a changing environment”. In: *Trends Microbiol* 9.12, pp. 597–605.

Berg, R. D., S. Levitte, M. P. O’Sullivan, S. M. O’Leary, C. J. Cambier, J. Cameron, K. K. Takaki, C. B. Moens, D. M. Tobin, J. Keane, and L. Ramakrishnan (2016). “Lysosomal Disorders Drive Susceptibility to Tuberculosis by Compromising Macrophage Migration”. In: *Cell* 165.1, pp. 139–152.

Bergeron, A., M. Bonay, M. Kambouchner, D. Lecossier, M. Riquet, P. Soler, A. Hance, and A. Tazi (1997). “Cytokine patterns in tuberculous and sarcoid granulomas: correlations with histopathologic features of the granulomatous response”. In: *J Immunol* 159.6, pp. 3034–43.

Bergeron, A. C., B. G. Seman, J. H. Hammond, L. S. Archambault, D. A. Hogan, and R. T. Wheeler (2017). “*Candida albicans* and *Pseudomonas aeruginosa* Interact To Enhance Virulence of Mucosal Infection in Transparent Zebrafish”. In: *Infect Immun* 85.11.

*– This is a really nice study that investigates tri-kingdom interactions between fungi, bacteria, and animals to better understand the role of polymicrobial infections in determining disease outcomes and utilizes the zebrafish model to do so. Taking this as a model, the application of this pairwise strategy to understand how, for instance, airway commensal species interact with *Mycobacterium tuberculosis*, seems like a compelling way to understand the influence of the microbiome on tuberculosis disease outcomes and could likely be modeled in the zebrafish.*

Bergers, G. and S. Song (2005). “The role of pericytes in blood-vessel formation and maintenance”. In: *Neuro Oncol* 7.4, pp. 452–64.

Bergmann, S., M. Rohde, G. S. Chhatwal, and S. Hammerschmidt (2001). “alpha-Enolase of *Streptococcus pneumoniae* is a plasmin(ogen)-binding protein displayed on the bacterial cell surface”. In: *Mol Microbiol* 40.6, pp. 1273–87.

- Bergmann, S., H. Schoenen, and S. Hammerschmidt (2013). “The interaction between bacterial enolase and plasminogen promotes adherence of *Streptococcus pneumoniae* to epithelial and endothelial cells”. In: *Int J Med Microbiol* 303.8, pp. 452–62.
- Bernut, A., J. L. Herrmann, K. Kiss, J. F. Dubremetz, J. L. Gaillard, G. Lutfalla, and L. Kremer (2014). “*Mycobacterium abscessus* cording prevents phagocytosis and promotes abscess formation”. In: *Proc Natl Acad Sci U S A* 111.10, E943–52.
- Berod, L., P. Stuve, M. Swallow, C. Arnold-Schrauf, F. Kruse, M. V. Gentilini, J. Freitag, B. Holzmann, and T. Sparwasser (2014). “MyD88 signalling in myeloid cells is sufficient to prevent chronic mycobacterial infection”. In: *Eur J Immunol* 44.5, pp. 1399–409.
- Bertrand, J. Y. and D. Traver (2009). “Hematopoietic cell development in the zebrafish embryo”. In: *Curr Opin Hematol* 16.4, pp. 243–8.
- Beutler, B. (2000). “Tlr4: central component of the sole mammalian LPS sensor”. In: *Curr Opin Immunol* 12.1, pp. 20–6.
- Bhatt, A., A. K. Brown, A. Singh, D. E. Minnikin, and G. S. Besra (2008). “Loss of a mycobacterial gene encoding a reductase leads to an altered cell wall containing beta-oxo-mycolic acid analogs and accumulation of ketones”. In: *Chem Biol* 15.9, pp. 930–9.
- Billington, O. J., T. D. McHugh, and S. H. Gillespie (1999). “Physiological cost of rifampin resistance induced *in vitro* in *Mycobacterium tuberculosis*”. In: *Antimicrob Agents Chemother* 43.8, pp. 1866–9.
- Bird, J. H., A. A. Khan, N. Nishimura, S. Yamasaki, M. S. M. Timmer, and B. L. Stocker (2018). “Synthesis of Branched Trehalose Glycolipids and Their Mincle Agonist Activity”. In: *J Org Chem* 83.15, pp. 7593–7605.
- Bisht, M., D. C. Dhasmana, and S. S. Bist (2010). “Angiogenesis: Future of pharmacological modulation”. In: *Indian J Pharmacol* 42.1, pp. 2–8.
- Biswas, S. K. and A. Mantovani (2012). “Orchestration of metabolism by macrophages”. In: *Cell Metab* 15.4, pp. 432–7.

- Bjorkbacka, H., K. A. Fitzgerald, F. Huet, X. Li, J. A. Gregory, M. A. Lee, C. M. Ordija, N. E. Dowley, D. T. Golenbock, and M. W. Freeman (2004). “The induction of macrophage gene expression by LPS predominantly utilizes Myd88-independent signaling cascades”. In: *Physiol Genomics* 19.3, pp. 319–30.
- Bjornsson, T. D., D. E. Schneider, and Jr. Berger H. (1989). “Aspirin acetylates fibrinogen and enhances fibrinolysis. Fibrinolytic effect is independent of changes in plasminogen activator levels”. In: *J Pharmacol Exp Ther* 250.1, pp. 154–61.
- Black, M., J. R. Mitchell, H. J. Zimmerman, K. G. Ishak, and G. R. Epler (1975). “Isoniazid-associated hepatitis in 114 patients”. In: *Gastroenterology* 69.2, pp. 289–302.
- Bloch, H. (1950). “Studies on the virulence of tubercle bacilli; isolation and biological properties of a constituent of virulent organisms”. In: *J Exp Med* 91.2, 197–218, pl.
- Bloch, H. and H. Noll (1955). “Studies on the virulence of Tubercle bacilli; the effect of cord factor on murine tuberculosis”. In: *Br J Exp Pathol* 36.1, pp. 8–17.
- Bloch, H., E. Sorkin, and H. Erlenmeyer (1953). “A toxic lipid component of the tubercle bacillus (cord factor). I. Isolation from petroleum ether extracts of young bacterial cultures”. In: *Am Rev Tuberc* 67.5, pp. 629–43.
- Bloch, M. (1973). *The Royal Touch (Routledge Revivals): Sacred Monarchy and Scrofula in England and France*. Routledge Revivals: Selected Works of Marc Bloch. Taylor & Francis.
- Bobay, L. M. and H. Ochman (2017). “The Evolution of Bacterial Genome Architecture”. In: *Front Genet* 8, p. 72.
- Bohrer, A. C., E. Castro, Z. Hu, A. T. L. Queiroz, C. E. Tocheny, M. Assmann, S. Sakai, C. Nelson, P. J. Baker, H. Ma, L. Wang, W. Zilu, E. du Bruyn, C. Riou, K. D. Kauffman, Program Tuberculosis Imaging, I. N. Moore, F. Del Nonno, L. Petrone, D. Goletti, A. R. Martineau, D. M. Lowe, M. R. Cronan, R. J. Wilkinson, C. E. Barry, L. E. Via, D. L. Barber, A. D. Klion, B. B. Andrade, Y. Song, K. W. Wong, and K. D. Mayer-Barber (2021).** “Eosinophils are part of the granulocyte response in tuberculosis and promote host resistance in mice”. In: *J Exp Med* 218.10.

Boise, L. H., B. Petryniak, X. Mao, C. H. June, C. Y. Wang, T. Lindsten, R. Bravo, K. Kovary, J. M. Leiden, and C. B. Thompson (1993). “The NFAT-1 DNA binding complex in activated T cells contains Fra-1 and JunB”. In: *Mol Cell Biol* 13.3, pp. 1911–9.

Bojarczuk, A., K. A. Miller, R. Hotham, A. Lewis, N. V. Ogryzko, A. A. Kamuyango, H. Frost, R. H. Gibson, E. Stillman, R. C. May, S. A. Renshaw, and S. A. Johnston (2016). “*Cryptococcus neoformans* Intracellular Proliferation and Capsule Size Determines Early Macrophage Control of Infection”. In: *Sci Rep* 6, p. 21489.

Bopp, T., A. Palmetshofer, E. Serfling, V. Heib, S. Schmitt, C. Richter, M. Klein, H. Schild, E. Schmitt, and M. Stassen (2005). “NFATc2 and NFATc3 transcription factors play a crucial role in suppression of CD4+ T lymphocytes by CD4+ CD25+ regulatory T cells”. In: *J Exp Med* 201.2, pp. 181–7.

Borham, M., A. Oreiby, A. El-Gedawy, Y. Hegazy, H. O. Khalifa, M. Al-Gaabary, and T. Matsumoto (2022). “Review on Bovine Tuberculosis: An Emerging Disease Associated with Multidrug-Resistant *Mycobacterium* Species”. In: *Pathogens* 11.7.

Boritsch, E. C., V. Khanna, A. Pawlik, N. Honore, V. H. Navas, L. Ma, C. Bouchier, T. Seemann, P. Supply, T. P. Stinear, and R. Brosch (2016). “Key experimental evidence of chromosomal DNA transfer among selected tuberculosis-causing mycobacteria”. In: *Proc Natl Acad Sci U S A* 113.35, pp. 9876–81.

Boros, Dov L. (2003). *Granulomatous infections and inflammations : cellular and molecular mechanisms*. Washington, D.C.: ASM Press.

Botrel, T. E. A., L. G. O. Clark, L. Paladini, and O. A. C. Clark (2016). “Efficacy and safety of bevacizumab plus chemotherapy compared to chemotherapy alone in previously untreated advanced or metastatic colorectal cancer: a systematic review and meta-analysis”. In: *BMC Cancer* 16, p. 677.

Boudinot, P., L. M. van der Aa, L. Jouneau, L. Du Pasquier, P. Pontarotti, V. Briolat, A. Benmansour, and J. P. Levraud (2011). “Origin and evolution of TRIM proteins: new insights from the complete TRIM repertoire of zebrafish and pufferfish”. In: *PLoS One* 6.7, e22022.

Bourajjaj, M., A. S. Armand, P. A. da Costa Martins, B. Weijts, R. van der Nagel, S. Heeneman, X. H. Wehrens, and L. J. De Windt (2008). “NFATc2 is a necessary mediator of calcineurin-dependent cardiac hypertrophy and heart failure”. In: *J Biol Chem* 283.32, pp. 22295–303.

Bourgeois, J. S., C. M. Smith, and D. C. Ko (2021). “These Are the Genes You’re Looking For: Finding Host Resistance Genes”. In: *Trends Microbiol* 29.4, pp. 346–362.

Bouvier, Jules (1842–1865). *A mother cries in grief while comforting her dying daughter, who holds a rosebud with falling petals.*

Bowdish, D. M., K. Sakamoto, M. J. Kim, M. Kroos, S. Mukhopadhyay, C. A. Leifer, K. Tryggvason, S. Gordon, and D. G. Russell (2009). “MARCO, TLR2, and CD14 are required for macrophage cytokine responses to mycobacterial trehalose dimycolate and *Mycobacterium tuberculosis*”. In: *PLoS Pathog* 5.6, e1000474.

— This was the first truly earnest attempt to delineate a signaling pathway responsive to TDM detection and succeeded at identifying TLR2, MARCO, and CD14 as essential factors in the response to TDM. Despite these comprehensive findings, this model has largely been supplanted by the MINCLE/MCL pathway; nonetheless, all studies on TDM-dependent responses should consider the potential involvement of TLR2-MYD88 responses and how those might intersect with MINCLE/MCL-dependent responses.

Bower, N. I., K. Koltowska, C. Pichol-Thievend, I. Virshup, S. Paterson, A. K. Lagedijk, W. Wang, B. W. Lindsey, S. J. Bent, S. Baek, M. Rondon-Galeano, D. G. Hurley, N. Mochizuki, C. Simons, M. Francois, C. A. Wells, J. Kaslin, and B. M. Hogan (2017a). “Mural lymphatic endothelial cells regulate meningeal angiogenesis in the zebrafish”. In: *Nat Neurosci* 20.6, pp. 774–783.

Bower, N. I., A. J. Vogrin, L. Le Guen, H. Chen, S. A. Stacker, M. G. Achen, and B. M. Hogan (2017b). “Vegfd modulates both angiogenesis and lymphangiogenesis during zebrafish embryonic development”. In: *Development* 144.3, pp. 507–518.

Bowker, N., M. Salie, H. Schurz, P. D. van Helden, C. J. Kinnear, E. G. Hoal, and M. Moller (2016). “Polymorphisms in the Pattern Recognition Receptor Mincl Gene (CLEC4E) and Association with Tuberculosis”. In: *Lung* 194.5, pp. 763–7.

Bowlin, A., H. Roys, H. Wanjala, M. Bettadapura, G. Venugopal, J. Surma, M. C. Simon, and T. Weinkopff (2021). “Hypoxia-Inducible Factor Signaling in Macrophages Promotes Lymphangiogenesis in *Leishmania major* Infection”. In: *Infect Immun* 89.8, e0012421.

- Bradford, T. J., S. A. Tomlins, X. Wang, and A. M. Chinnaiyan (2006). “Molecular markers of prostate cancer”. In: *Urol Oncol* 24.6, pp. 538–51.
- Bradford, Y. M., C. E. Van Slyke, L. Ruzicka, A. Singer, A. Eagle, D. Fashena, D. G. Howe, K. Frazer, R. Martin, H. Paddock, C. Pich, S. Ramachandran, and M. Westerfield (2022). “Zebrafish information network, the knowledgebase for *Danio rerio* research”. In: *Genetics* 220.4.
- Braibant, M., P. Gilot, and J. Content (2000). “The ATP binding cassette (ABC) transport systems of *Mycobacterium tuberculosis*”. In: *FEMS Microbiol Rev* 24.4, pp. 449–67.
- Brand, M., C. P. Heisenberg, Y. J. Jiang, D. Beuchle, K. Lun, M. Furutani-Seiki, M. Granato, P. Haffter, M. Hammerschmidt, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, F. J. van Eeden, and C. Nusslein-Volhard (1996a). “Mutations in zebrafish genes affecting the formation of the boundary between midbrain and hindbrain”. In: *Development* 123, pp. 179–90.
- Brand, M., C. P. Heisenberg, R. M. Warga, F. Pelegri, R. O. Karlstrom, D. Beuchle, A. Picker, Y. J. Jiang, M. Furutani-Seiki, F. J. van Eeden, M. Granato, P. Haffter, M. Hammerschmidt, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, and C. Nusslein-Volhard (1996b). “Mutations affecting development of the midline and general body shape during zebrafish embryogenesis”. In: *Development* 123, pp. 129–42.
- Brandt, K. J., C. Fickentscher, E. K. Kruithof, and P. de Moerloose (2013). “TLR2 ligands induce NF-kappaB activation from endosomal compartments of human monocytes”. In: *PLoS One* 8.12, e80743.
- Brannon, M. K., J. M. Davis, J. R. Mathias, C. J. Hall, J. C. Emerson, P. S. Crosier, A. Huttenlocher, L. Ramakrishnan, and S. M. Moskowitz (2009). “*Pseudomonas aeruginosa* Type III secretion system interacts with phagocytes to modulate systemic infection of zebrafish embryos”. In: *Cell Microbiol* 11.5, pp. 755–68.
- Braverman, J., K. M. Sogi, D. Benjamin, D. K. Nomura, and S. A. Stanley (2016). “HIF-1alpha Is an Essential Mediator of IFN-gamma-Dependent Immunity to *Mycobacterium tuberculosis*”. In: *J Immunol* 197.4, pp. 1287–97.
- Bray, N. L., H. Pimentel, P. Melsted, and L. Pachter (2016). “Near-optimal probabilistic RNA-seq quantification”. In: *Nat Biotechnol* 34.5, pp. 525–7.

Brennan, P. J. (2003). "Structure, function, and biogenesis of the cell wall of *Mycobacterium tuberculosis*". In: *Tuberculosis (Edinb)* 83.1-3, pp. 91–7.

Brennan, P. J. and H. Nikaido (1995). "The envelope of mycobacteria". In: *Annu Rev Biochem* 64, pp. 29–63.

Brewer, W. J., E. M. Walton, A. M. Xet-Mull, and D. M. Tobin (2022a). "Data release in re: Macrophage NFATC2 Signaling Drives Angiogenic Signaling During Mycobacterial Infection". In: *Zenodo*.

Brewer, W. J., A. M. Xet-Mull, A. Yu, M. I. Sweeney, E. M. Walton, and D. M. Tobin (2022b). "Macrophage NFATC2 Mediates Angiogenic Signaling During Mycobacterial Infection". In: *Cell Rep* 41.111817.

Briolat, V., L. Jouneau, R. Carvalho, N. Palha, C. Langevin, P. Herbomel, O. Schwartz, H. P. Spaink, J. P. Levraud, and P. Boudinot (2014). "Contrasted innate responses to two viruses in zebrafish: insights into the ancestral repertoire of vertebrate IFN-stimulated genes". In: *J Immunol* 192.9, pp. 4328–41.

Britto, D. D., B. Wyroba, W. Chen, R. A. Lockwood, K. B. Tran, P. R. Shepherd, C. J. Hall, K. E. Crosier, P. S. Crosier, and J. W. Astin (2018). "Macrophages enhance Vegfa-driven angiogenesis in an embryonic zebrafish tumour xenograft model". In: *Dis Model Mech* 11.12.

– This captivating study identified a critical role for macrophages in mediating VEGFA-dependent angiogenesis from VEGFA-high tumors, suggesting that these macrophages were involved not in the production of VEGFA, but in providing some other cue for the vessels. Intravital microscopy was able to uncover direct macrophage-endothelial interactions that guided endothelial cells along the VEGFA gradient toward the site of the tumor. Ablation of macrophages, even in the context of VEGFA-high tumors, resulted in impaired angiogenesis. This is an provocative study on how macrophages may, through physical interactions, influence angiogenesis independent of direct production of pro-angiogenic chemokines.

Brontë, C. (1847). *Jane Eyre*. Dean Press.

Brown, G. D. (2006). "Dectin-1: a signalling non-TLR pattern-recognition receptor". In: *Nat Rev Immunol* 6.1, pp. 33–43.

Brown, G. D., J. Herre, D. L. Williams, J. A. Willment, A. S. Marshall, and S. Gordon (2003). “Dectin-1 mediates the biological effects of beta-glucans”. In: *J Exp Med* 197.9, pp. 1119–24.

Brown, G. D., P. R. Taylor, D. M. Reid, J. A. Willment, D. L. Williams, L. Martinez-Pomares, S. Y. Wong, and S. Gordon (2002). “Dectin-1 is a major beta-glucan receptor on macrophages”. In: *J Exp Med* 196.3, pp. 407–12.

Brown, G. D., J. A. Willment, and L. Whitehead (2018). “C-type lectins in immunity and homeostasis”. In: *Nat Rev Immunol* 18.6, pp. 374–389.

Brown, J., C. A. O’Callaghan, A. S. Marshall, R. J. Gilbert, C. Siebold, S. Gordon, G. D. Brown, and E. Y. Jones (2007). “Structure of the fungal beta-glucan-binding immune receptor dectin-1: implications for function”. In: *Protein Sci* 16.6, pp. 1042–52.

Brown, K. H., K. P. Dobrinski, A. S. Lee, O. Gokcumen, R. E. Mills, X. Shi, W. W. Chong, J. Y. Chen, P. Yoo, S. David, S. M. Peterson, T. Raj, K. W. Choy, B. E. Stranger, R. E. Williamson, L. I. Zon, J. L. Freeman, and C. Lee (2012). “Extensive genetic diversity and substructuring among zebrafish strains revealed through copy number variant analysis”. In: *Proc Natl Acad Sci U S A* 109.2, pp. 529–34.

Brown, L., J. M. Wolf, R. Prados-Rosales, and A. Casadevall (2015). “Through the wall: extracellular vesicles in Gram-positive bacteria, mycobacteria and fungi”. In: *Nat Rev Microbiol* 13.10, pp. 620–30.

Bruni, G., A. J. Rennekamp, A. Velenich, M. McCarroll, L. Gendelev, E. Fertsch, J. Taylor, P. Lakhani, D. Lensen, T. Evron, P. J. Lorello, X. P. Huang, S. Kolczewski, G. Carey, B. J. Caldarone, E. Prinssen, B. L. Roth, M. J. Keiser, R. T. Peterson, and D. Kokel (2016). “Zebrafish behavioral profiling identifies multitarget antipsychotic-like compounds”. In: *Nat Chem Biol* 12.7, pp. 559–66.

Bryant, C. D. (2011). “The blessings and curses of C57BL/6 substrains in mouse genetic studies”. In: *Ann N Y Acad Sci* 1245, pp. 31–3.

Bryant, J. M., D. M. Grogono, D. Rodriguez-Rincon, I. Everall, K. P. Brown, P. Moreno, D. Verma, E. Hill, J. Drijkoningen, P. Gilligan, C. R. Esther, P. G. Noone, O. Giddings, S. C. Bell, R. Thomson, C. E. Wainwright, C. Coulter, S. Pandey, M. E. Wood, R. E. Stockwell, K. A. Ramsay, L. J. Sherrard, T. J. Kidd, N. Jabbour, G. R. Johnson, L. D. Knibbs, L. Morawska, P. D. Sly, A. Jones, D. Bilton, I. Laurenson, M. Ruddy, S. Bourke, I. C. Bowler, S. J. Chapman, A. Clayton, M. Cullen,

T. Daniels, O. Dempsey, M. Denton, M. Desai, R. J. Drew, F. Edenborough, J. Evans, J. Folb, H. Humphrey, B. Isalska, S. Jensen-Fangel, B. Jonsson, A. M. Jones, T. L. Katzenstein, T. Lillebaek, G. MacGregor, S. Mayell, M. Millar, D. Modha, E. F. Nash, C. O'Brien, D. O'Brien, C. Ohri, C. S. Pao, D. Peckham, F. Perrin, A. Perry, T. Pressler, L. Prtak, T. Qvist, A. Robb, H. Rodgers, K. Schaffer, N. Shafi, J. van Ingen, M. Walshaw, D. Watson, N. West, J. Whitehouse, C. S. Haworth, S. R. Harris, D. Ordway, J. Parkhill, and R. A. Floto (2016). "Emergence and spread of a human-transmissible multidrug-resistant nontuberculous mycobacterium". In: *Science* 354.6313, pp. 751–757.

Bryson, B. D., T. R. Rosebrock, F. G. Tafesse, C. Y. Itoh, A. Nibasumba, G. H. Babunovic, B. Corleis, C. Martin, C. Keegan, P. Andrade, S. Realegeno, D. Kwon, R. L. Modlin, and S. M. Fortune (2019). "Heterogeneous GM-CSF signaling in macrophages is associated with control of *Mycobacterium tuberculosis*". In: *Nat Commun* 10.1, p. 2329.

Buchholz, M., A. Schatz, M. Wagner, P. Michl, T. Linhart, G. Adler, T. M. Gress, and V. Ellenrieder (2006). "Overexpression of c-myc in pancreatic cancer caused by ectopic activation of NFATc1 and the Ca²⁺/calcineurin signaling pathway". In: *EMBO J* 25.15, pp. 3714–24.

Bunduc, C. M., W. Bitter, and E. N. G. Houben (2020). "Structure and Function of the Mycobacterial Type VII Secretion Systems". In: *Annu Rev Microbiol* 74, pp. 315–335.

Bushdid, P. B., H. Osinska, R. R. Waclaw, J. D. Molkentin, and K. E. Yutzey (2003). "NFATc3 and NFATc4 are required for cardiac development and mitochondrial function". In: *Circ Res* 92.12, pp. 1305–13.

Bussmann, J., F. L. Bos, A. Urasaki, K. Kawakami, H. J. Duckers, and S. Schulte-Merker (2010). "Arteries provide essential guidance cues for lymphatic endothelial cells in the zebrafish trunk". In: *Development* 137.16, pp. 2653–7.

Bussmann, J., N. Lawson, L. Zon, S. Schulte-Merker, and Committee Zebrafish Nomenclature (2008). "Zebrafish VEGF receptors: a guideline to nomenclature". In: *PLoS Genet* 4.5, e1000064.

Bustamante, J., S. Boisson-Dupuis, L. Abel, and J. L. Casanova (2014). "Mendelian susceptibility to mycobacterial disease: genetic, immunological, and clinical features of inborn errors of IFN-gamma immunity". In: *Semin Immunol* 26.6, pp. 454–70.

Byrne, G. I., L. K. Lehmann, and G. J. Landry (1986). “Induction of tryptophan catabolism is the mechanism for gamma-interferon-mediated inhibition of intracellular Chlamydia psittaci replication in T24 cells”. In: *Infect Immun* 53.2, pp. 347–51.

Cadena, A. M., J. L. Flynn, and S. M. Fortune (2016). “The Importance of First Impressions: Early Events in *Mycobacterium tuberculosis* Infection Influence Outcome”. In: *mBio* 7.2, e00342–16.

Cadena, A. M., S. M. Fortune, and J. L. Flynn (2017). “Heterogeneity in tuberculosis”. In: *Nat Rev Immunol* 17.11, pp. 691–702.

Cagan, R. L., L. I. Zon, and R. M. White (2019). “Modeling Cancer with Flies and Fish”. In: *Dev Cell* 49.3, pp. 317–324.

Cambau, E. and M. Drancourt (2014). “Steps towards the discovery of *Mycobacterium tuberculosis* by Robert Koch, 1882”. In: *Clin Microbiol Infect* 20.3, pp. 196–201.

Cambier, C. J., S. M. Banik, J. A. Buonomo, and C. R. Bertozzi (2020). “Spreading of a mycobacterial cell-surface lipid into host epithelial membranes promotes infectivity”. In: *Elife* 9.

– After identifying a critical role for PDIM in moderating TLR2-dependent immune responses, CJ went on to identify entirely novel biochemistry wherein mycobacterial PDIM physically spreads across host lipid membranes to mute the ability of host macrophages to respond and kill the bacteria. This model of lipids being able to directly disrupt immunity through engagement with the membrane itself is a compelling model that may be evocative of mechanisms at play in other diseases, but this will require extensive additional study.

Cambier, C. J., S. Falkow, and L. Ramakrishnan (2014). “Host evasion and exploitation schemes of *Mycobacterium tuberculosis*”. In: *Cell* 159.7, pp. 1497–509.

Cambier, C. J., S. M. O’Leary, M. P. O’Sullivan, J. Keane, and L. Ramakrishnan (2017). “Phenolic Glycolipid Facilitates Mycobacterial Escape from Microbicidal Tissue-Resident Macrophages”. In: *Immunity* 47.3, 552–565 e4.

Cambier, C. J., K. K. Takaki, R. P. Larson, R. E. Hernandez, D. M. Tobin, K. B. Urdahl, C. L. Cosma, and L. Ramakrishnan (2014). “Mycobacteria manipulate macrophage recruitment through coordinated use of membrane lipids”. In: *Nature* 505.7482, pp. 218–22.

– This study is a creative look at how a particular cell wall lipid of pathogenic mycobacteria, phthiocerol dimycoceroserate (PDIM) masks immunostimulatory bacterial ligands to prevent host TLR2-dependent responses and avoid a hostile immune environment primed by commensal species within the lung. This role for PDIM is revealing as Bowdish, Sakamoto, Kim, Kroos, Mukhopadhyay, Leifer, Tryggvason, Gordon, and Russell (2009) found that TDM-mediated responses were dependent on TLR2, but if PDIM is masking TLR2 ligands, then that may be a critical component in shifting the reliance toward C-type lectin responses that are less efficiently inhibited by PDIM. This also adds dramatically to the overall understanding of how cell wall lipids influence the immune response.

Campuzano, A., N. Castro-Lopez, K. L. Wozniak, C. M. Leopold Wager, and Jr. Wormley F. L. (2017). “Dectin-3 Is Not Required for Protection against *Cryptococcus neoformans* Infection”. In: *PLoS One* 12.1, e0169347.

Candela, M., E. Biagi, M. Centanni, S. Turroni, M. Vici, F. Musiani, B. Vitali, S. Bergmann, S. Hammerschmidt, and P. Brigidi (2009). “Bifidobacterial enolase, a cell surface receptor for human plasminogen involved in the interaction with the host”. In: *Microbiology (Reading)* 155.Pt 10, pp. 3294–3303.

Caneparo, V., S. Landolfo, M. Gariglio, and M. De Andrea (2018). “The Absent in Melanoma 2-Like Receptor IFN-Inducible Protein 16 as an Inflammasome Regulator in Systemic Lupus Erythematosus: The Dark Side of Sensing Microbes”. In: *Front Immunol* 9, p. 1180.

Cante-Barrett, K., M. M. Winslow, and G. R. Crabtree (2007). “Selective role of NFATc3 in positive selection of thymocytes”. In: *J Immunol* 179.1, pp. 103–10.

– This publication explains the reason why NFATC2 null mice do not have any defect in T cell development and this is due to a specific activity of NFATC3 in regulating the positive selection of CD4/CD8 double positive thymocytes. In the absence of NFATC3, the cells stall in positive selection. This study also sought to explain the role for NFATC1 in these cells and found that NFATC1 and NFATC3 cooperate to allow T cell positive selection and transition to single positive cells. The results also evoke specific and differentiable roles for the NFAT isoforms in inducing these responses as this paper utilized a constitutively active NFATC1 to rescue some, but not all of the effect of NFATC3 knockout.

Cardoso, M. S., T. M. Silva, M. Resende, R. Appelberg, and M. Borges (2015). “Lack of the Transcription Factor Hypoxia-Inducible Factor 1alpha (HIF-1alpha)

in Macrophages Accelerates the Necrosis of *Mycobacterium avium*-Induced Granulomas". In: *Infect Immun* 83.9, pp. 3534–44.

Carlone, G. M., M. J. Valadez, and M. J. Pickett (1982). "Methods for distinguishing gram-positive from gram-negative bacteria". In: *J Clin Microbiol* 16.6, pp. 1157–9.

Carlson, J. C., M. Cantu Gutierrez, B. Lozzi, E. Huang-Hobbs, W. D. Turner, B. Tepe, Y. Zhang, A. M. Herman, G. Rao, C. J. Creighton, J. D. Wythe, and B. Deneen (2021). "Identification of diverse tumor endothelial cell populations in malignant glioma". In: *Neuro Oncol* 23.6, pp. 932–944.

Carmeliet, P., V. Ferreira, G. Breier, S. Pollefeyt, L. Kieckens, M. Gertsenstein, M. Fahrig, A. Vandenhoeck, K. Harpal, C. Eberhardt, C. Declercq, J. Pawling, L. Moons, D. Collen, W. Risau, and A. Nagy (1996). "Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele". In: *Nature* 380.6573, pp. 435–9.

Carmeliet, P. and R. K. Jain (2011). "Molecular mechanisms and clinical applications of angiogenesis". In: *Nature* 473.7347, pp. 298–307.

Carmi, Y., E. Voronov, S. Dotan, N. Lahat, M. A. Rahat, M. Fogel, M. Huszar, M. R. White, C. A. Dinarello, and R. N. Apte (2009). "The role of macrophage-derived IL-1 in induction and maintenance of angiogenesis". In: *J Immunol* 183.7, pp. 4705–14.

Carpenter, A. E., T. R. Jones, M. R. Lamprecht, C. Clarke, I. H. Kang, O. Friman, D. A. Guertin, J. H. Chang, R. A. Lindquist, J. Moffat, P. Golland, and D. M. Sabatini (2006). "CellProfiler: image analysis software for identifying and quantifying cell phenotypes". In: *Genome Biol* 7.10, R100.

Carpenter, C. M., A. W. Naylor-Foote, G. V. Taplin, C. A. Lawrence, and C. L. Drake (1959). "Preliminary report on vaccines prepared from gamma-irradiated *Mycobacterium tuberculosis* and *Brucella suis*". In: *Am Rev Tuberc* 79.3, pp. 374–7.

Carranza, C., E. Juarez, M. Torres, J. J. Ellner, E. Sada, and S. K. Schwander (2006). "*Mycobacterium tuberculosis* growth control by lung macrophages and CD8 cells from patient contacts". In: *Am J Respir Crit Care Med* 173.2, pp. 238–45.

Carroll, P., L. J. Schreuder, J. Muwanguzi-Karugaba, S. Wiles, B. D. Robertson, J. Ripoll, T. H. Ward, G. J. Bancroft, U. E. Schaible, and T. Parish (2010). "Sensitive

detection of gene expression in mycobacteria under replicating and non-replicating conditions using optimized far-red reporters”. In: *PLoS One* 5.3, e9823.

Carvalho, L. D., L. K. Teixeira, N. Carrossini, A. T. Caldeira, K. M. Ansel, A. Rao, and J. P. Viola (2007). “The NFAT1 transcription factor is a repressor of cyclin A2 gene expression”. In: *Cell Cycle* 6.14, pp. 1789–95.

Casadevall, A. and L. A. Pirofski (1999). “Host-pathogen interactions: redefining the basic concepts of virulence and pathogenicity”. In: *Infect Immun* 67.8, pp. 3703–13.

– (2003). “The damage-response framework of microbial pathogenesis”. In: *Nat Rev Microbiol* 1.1, pp. 17–24.

Casonato, S., R. Provvedi, E. Dainese, G. Palu, and R. Manganelli (2014). “*Mycobacterium tuberculosis* requires the ECF sigma factor SigE to arrest phagosome maturation”. In: *PLoS One* 9.9, e108893.

Cassol, E., L. Cassetta, M. Alfano, and G. Poli (2010). “Macrophage polarization and HIV-1 infection”. In: *J Leukoc Biol* 87.4, pp. 599–608.

Catar, R., J. Witowski, P. Wagner, I. Annett Schramm, E. Kawka, A. Philippe, D. Dragun, and A. Jorres (2013). “The proto-oncogene c-Fos transcriptionally regulates VEGF production during peritoneal inflammation”. In: *Kidney Int* 84.6, pp. 1119–28.

Cattin, A. L., J. J. Burden, L. Van Emmenis, F. E. Mackenzie, J. J. Hoving, N. Garcia Calavia, Y. Guo, M. McLaughlin, L. H. Rosenberg, V. Quereda, D. Jamecna, I. Napoli, S. Parrinello, T. Enver, C. Ruhrberg, and A. C. Lloyd (2015). “Macrophage-Induced Blood Vessels Guide Schwann Cell-Mediated Regeneration of Peripheral Nerves”. In: *Cell* 162.5, pp. 1127–39.

Cavalcanti, Y. V., M. C. Brelaz, J. K. Neves, J. C. Ferraz, and V. R. Pereira (2012). “Role of TNF-Alpha, IFN-Gamma, and IL-10 in the Development of Pulmonary Tuberculosis”. In: *Pulm Med* 2012, p. 745483.

Centers for Disease Control (2016). *History of World TB Day*.

Cervantes, J. L. (2017). “MyD88 in *Mycobacterium tuberculosis* infection”. In: *Med Microbiol Immunol* 206.3, pp. 187–193.

Cesca, M., F. Bizzaro, M. Zucchetti, and R. Giavazzi (2013). “Tumor delivery of chemotherapy combined with inhibitors of angiogenesis and vascular targeting agents”. In: *Front Oncol* 3, p. 259.

Cha, S. B., W. S. Kim, J. S. Kim, H. Kim, K. W. Kwon, S. J. Han, S. Y. Eum, S. N. Cho, and S. J. Shin (2015). “Repeated Aerosolized-Boosting with Gamma-Irradiated *Mycobacterium bovis* BCG Confers Improved Pulmonary Protection against the Hypervirulent *Mycobacterium tuberculosis* Strain HN878 in Mice”. In: *PLoS One* 10.10, e0141577.

Chabon, J. J., L. Gebreab, R. Kumar, E. Debella, T. Tanaka, D. Koyanagi, A. Rodriguez Garcia, L. Sanders, M. Perez, R. M. Tudor, and B. B. Graham (2014). “Role of vascular endothelial growth factor signaling in *Schistosoma*-induced experimental pulmonary hypertension”. In: *Pulm Circ* 4.2, pp. 289–99.

Chakraborty, S. and K. Y. Rhee (2015). “Tuberculosis Drug Development: History and Evolution of the Mechanism-Based Paradigm”. In: *Cold Spring Harb Perspect Med* 5.8, a021147.

Chakravarty, S. D., G. Zhu, M. C. Tsai, V. P. Mohan, S. Marino, D. E. Kirschner, L. Huang, J. Flynn, and J. Chan (2008). “Tumor necrosis factor blockade in chronic murine tuberculosis enhances granulomatous inflammation and disorganizes granulomas in the lungs”. In: *Infect Immun* 76.3, pp. 916–26.

Chalke, H. D. (1962). “The Impact of Tuberculosis on History, Literature and Art”. In: *Med Hist* 6.4, pp. 301–18.

– This is an interesting look at how tuberculosis has influenced Western cultural development and is worth a read to garner a better understanding of the impact of tuberculosis in the 1800s.

Chan, J., P. E. Bayliss, J. M. Wood, and T. M. Roberts (2002). “Dissection of angiogenic signaling in zebrafish using a chemical genetic approach”. In: *Cancer Cell* 1.3, pp. 257–67.

Chan, J., T. Fujiwara, P. Brennan, M. McNeil, S. J. Turco, J. C. Sible, M. Snapper, P. Aisen, and B. R. Bloom (1989). “Microbial glycolipids: possible virulence factors that scavenge oxygen radicals”. In: *Proc Natl Acad Sci U S A* 86.7, pp. 2453–7.

Chandra, P., S. J. Grigsby, and J. A. Philips (2022). “Immune evasion and provocation by *Mycobacterium tuberculosis*”. In: *Nat Rev Microbiol*.

Chang, C. P., J. R. Neilson, J. H. Bayle, J. E. Gestwicki, A. Kuo, K. Stankunas, I. A. Graef, and G. R. Crabtree (2004). “A field of myocardial-endocardial NFAT signaling underlies heart valve morphogenesis”. In: *Cell* 118.5, pp. 649–63.

– *The identification of NFAT binding sites within the VEGFA promoter is an important component of understanding how NFAT is regulating this chemokine. This is the first and, to my knowledge, only study that identified a functional role for a particular NFAT binding motif in the VEGFA promoter. The particular motif identified was actually inhibitory toward expression, suggesting that NFAT can act to both inhibit and promote transcriptional responses, a hypothesis borne out in diverse other contexts.*

Chang, S. W., W. S. Pan, D. Lozano Beltran, L. Oleyda Baldelomar, M. A. Solano, I. Tuero, J. S. Friedland, F. Torrico, and R. H. Gilman (2013). “Gut hormones, appetite suppression and cachexia in patients with pulmonary TB”. In: *PLoS One* 8.1, e54564.

Chang, W. (2022). “extrafont: Tools for Using Fonts”. In.

Chanput, W., V. Peters, and H. Wickers (2015). “THP-1 and U937 Cells”. In: ed. by K. Verhoeckx, P. Cotter, I. Lopez-Exposito, C. Kleiveland, T. Lea, A. Mackie, T. Requena, D. Swiatecka, and H. Wickers. Cham (CH), pp. 147–159.

Chapman, J. S. (1971). “The ecology of the atypical mycobacteria”. In: *Arch Environ Health* 22.1, pp. 41–6.

Chatterjee, D. (1997). “The mycobacterial cell wall: structure, biosynthesis and sites of drug action”. In: *Curr Opin Chem Biol* 1.4, pp. 579–88.

Chavakis, E. and S. Dimmeler (2002). “Regulation of endothelial cell survival and apoptosis during angiogenesis”. In: *Arterioscler Thromb Vasc Biol* 22.6, pp. 887–93.

Chaves, E. (1966). “The pathology of the arterial pulmonary vasculature in manson’s schistosomiasis”. In: *Dis Chest* 50.1, pp. 72–7.

Chavez, M. N., G. Aedo, F. A. Fierro, M. L. Allende, and J. T. Egana (2016). “Zebrafish as an Emerging Model Organism to Study Angiogenesis in Development and Regeneration”. In: *Front Physiol* 7, p. 56.

Chavkin, N. W. and K. K. Hirschi (2020). “Single Cell Analysis in Vascular Biology”. In: *Front Cardiovasc Med* 7, p. 42.

Chawla, A., Y. Barak, L. Nagy, D. Liao, P. Tontonoz, and R. M. Evans (2001). “PPAR-gamma dependent and independent effects on macrophage-gene expression in lipid metabolism and inflammation”. In: *Nat Med* 7.1, pp. 48–52.

Cheer, S. M. and G. L. Plosker (2001). “Tacrolimus ointment. A review of its therapeutic potential as a topical therapy in atopic dermatitis”. In: *Am J Clin Dermatol* 2.6, pp. 389–406.

Chen, F., W. A. M. Elgaher, M. Winterhoff, K. Bussow, F. H. Waqas, E. Graner, Y. Pires-Afonso, L. Casares Perez, L. de la Vega, N. Sahini, L. Czichon, W. Zobl, T. Zillinger, M. Shehata, S. Pleschka, H. Bahre, C. Falk, A. Michelucci, S. Schuchardt, W. Blankenfeldt, A. K. H. Hirsch, and F. Pessler (2022). “Citraconate inhibits ACOD1 (IRG1) catalysis, reduces interferon responses and oxidative stress, and modulates inflammation and cell metabolism”. In: *Nat Metab* 4.5, pp. 534–546.

Chen, J., Y. Amasaki, Y. Kamogawa, M. Nagoya, N. Arai, K. Arai, and S. Miyatake (2003). “Role of NFATx (NFAT4/NFATc3) in expression of immunoregulatory genes in murine peripheral CD4+ T cells”. In: *J Immunol* 170.6, pp. 3109–17.

Chen, J. N., P. Haffter, J. Odenthal, E. Vogelsang, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, R. N. Kelsh, M. C. Mullins, and C. Nusslein-Volhard (1996). “Mutations affecting the cardiovascular system and other internal organs in zebrafish”. In: *Development* 123, pp. 293–302.

Chen, L., J. N. Glover, P. G. Hogan, A. Rao, and S. C. Harrison (1998). “Structure of the DNA-binding domains from NFAT, Fos and Jun bound specifically to DNA”. In: *Nature* 392.6671, pp. 42–8.

Chen, W., X. Qiang, Y. Wang, S. Zhu, J. Li, A. Babaev, H. Yang, J. Gong, L. Becker, P. Wang, K. J. Tracey, and H. Wang (2020). “Identification of tetranectin-targeting monoclonal antibodies to treat potentially lethal sepsis”. In: *Sci Transl Med* 12.539.

Cheng, S. C., J. Quintin, R. A. Cramer, K. M. Shepardson, S. Saeed, V. Kumar, E. J. Giamparellos-Bourboulis, J. H. Martens, N. A. Rao, A. Aghajanirefah, G. R. Manjeri, Y. Li, D. C. Ifrim, R. J. Arts, B. M. van der Veer, P. M. Deen, C. Logie, L. A. O’Neill, P. Willems, F. L. van de Veerdonk, J. W. van der Meer, A. Ng,

L. A. Joosten, C. Wijmenga, H. G. Stunnenberg, R. J. Xavier, and M. G. Netea (2014). “mTOR- and HIF-1alpha-mediated aerobic glycolysis as metabolic basis for trained immunity”. In: *Science* 345.6204, p. 1250684.

Cheng, Y. and J. S. Schorey (2018). “*Mycobacterium tuberculosis*-induced IFN-beta production requires cytosolic DNA and RNA sensing pathways”. In: *J Exp Med* 215.11, pp. 2919–2935.

Chernow, R. (2010). *Washington: A Life*. New York: Penguin Press.

Chinot, O. L., W. Wick, W. Mason, R. Henriksson, F. Saran, R. Nishikawa, A. F. Carpentier, K. Hoang-Xuan, P. Kavan, D. Cernea, A. A. Brandes, M. Hilton, L. Abrey, and T. Cloughesy (2014). “Bevacizumab plus radiotherapy-temozolomide for newly diagnosed glioblastoma”. In: *N Engl J Med* 370.8, pp. 709–22.

Chiplunkar, S. S., C. A. Silva, L. E. Bermudez, and L. Danelishvili (2019). “Characterization of membrane vesicles released by *Mycobacterium avium* in response to environment mimicking the macrophage phagosome”. In: *Future Microbiol* 14.4, pp. 293–313.

Cho, K. F., T. C. Branon, N. D. Udeshi, S. A. Myers, S. A. Carr, and A. Y. Ting (2020). “Proximity labeling in mammalian cells with TurboID and split-TurboID”. In: *Nat Protoc* 15.12, pp. 3971–3999.

Chow, C. W., M. Rincon, J. Cavanagh, M. Dickens, and R. J. Davis (1997). “Nuclear accumulation of NFAT4 opposed by the JNK signal transduction pathway”. In: *Science* 278.5343, pp. 1638–41.

– While the nuclear import of NFAT was known to be dependent on calcineurin, mechanisms of export remained elusive. This foundational report identified JNK as a critical kinase able to phosphorylate NFAT and induce nuclear export. While other kinases have been identified since, this added needed regulatory nuance to the NFAT pathway as such a sensitive pathway should be able to be dynamically activated and deactivated posttranscriptionally and leave the cell primed to respond again if necessary. It also explained the cycles of import and export observable by live cell microscopy, providing a mechanism to a previously observed phenomenon.

Chow, C. W., M. Rincon, and R. J. Davis (1999). “Requirement for transcription factor NFAT in interleukin-2 expression”. In: *Mol Cell Biol* 19.3, pp. 2300–7.

Chung, A. S. and N. Ferrara (2011). “Developmental and pathological angiogenesis”. In: *Annu Rev Cell Dev Biol* 27, pp. 563–84.

Chung, K. and K. Deisseroth (2013). “CLARITY for mapping the nervous system”. In: *Nat Methods* 10.6, pp. 508–13.

– CLARITY has revolutionized our ability to understand a vast range of biological processes and was absolutely essential to our studies on the adult granuloma as it is all but impossible to quantitate vascular association in thin tissue sections, but being able to capture these interactions in their native context allowed us to deeply understand our phenotype in the most complete way.

Churchyard, G., P. Kim, N. S. Shah, R. Rustomjee, N. Gandhi, B. Mathema, D. Dowdy, A. Kasmar, and V. Cardenas (2017). “What We Know About Tuberculosis Transmission: An Overview”. In: *J Infect Dis* 216.suppl_6, S629–S635.

Chuvpilo, S., A. Avots, F. Berberich-Siebelt, J. Glockner, C. Fischer, A. Kerstan, C. Escher, I. Inashkina, F. Hlubek, E. Jankevics, T. Brabletz, and E. Serfling (1999). “Multiple NF-ATc isoforms with individual transcriptional properties are synthesized in T lymphocytes”. In: *J Immunol* 162.12, pp. 7294–301.

Cicchese, J. M., V. Dartois, D. E. Kirschner, and J. J. Linderman (2020). “Both Pharmacokinetic Variability and Granuloma Heterogeneity Impact the Ability of the First-Line Antibiotics to Sterilize Tuberculosis Granulomas”. In: *Front Pharmacol* 11, p. 333.

Cikos, S., A. Bukovska, and J. Koppel (2007). “Relative quantification of mRNA: comparison of methods currently used for real-time PCR data analysis”. In: *BMC Mol Biol* 8, p. 113.

Claesson-Welsh, L. (2015). “Vascular permeability—the essentials”. In: *Ups J Med Sci* 120.3, pp. 135–43.

Clark, A. N., R. Youkey, X. Liu, L. Jia, R. Blatt, Y. J. Day, G. W. Sullivan, J. Linden, and A. L. Tucker (2007). “A1 adenosine receptor activation promotes angiogenesis and release of VEGF from monocytes”. In: *Circ Res* 101.11, pp. 1130–8.

Clark, K. J., D. P. Argue, A. M. Petzold, and S. C. Ekker (2012). “zfishbook: connecting you to a world of zebrafish revertible mutants”. In: *Nucleic Acids Res* 40.Database issue, pp. D907–11.

Clark, S., Y. Hall, and A. Williams (2014). “Animal models of tuberculosis: Guinea pigs”. In: *Cold Spring Harb Perspect Med* 5.5, a018572.

Clarke, Erik and Scott Sherrill-Mix (2017). “ggbeeswarm: Categorical Scatter (Violin Point) Plots”. In.

Clarke, T. B. (2014). “Early innate immunity to bacterial infection in the lung is regulated systemically by the commensal microbiota via NOD-like receptor ligands”. In: *Infect Immun* 82.11, pp. 4596–606.

Clatworthy, A. E., J. S. Lee, M. Leibman, Z. Kostun, A. J. Davidson, and D. T. Hung (2009). “*Pseudomonas aeruginosa* infection of zebrafish involves both host and pathogen determinants”. In: *Infect Immun* 77.4, pp. 1293–303.

Clay, H., J. M. Davis, D. Beery, A. Huttenlocher, S. E. Lyons, and L. Ramakrishnan (2007). “Dichotomous role of the macrophage in early *Mycobacterium marinum* infection of the zebrafish”. In: *Cell Host Microbe* 2.1, pp. 29–39.

– This study affirmed that macrophages are important protective cells against mycobacterial infection, which had become a bit of a debate in the literature. After depletion of macrophages, mycobacteria grew unrestricted but were limited in their growth by phagocytosis within macrophages. This gave impetus to the discovery of macrophage factors responsible for both bacterial restriction and factors that bacteria subvert to block these protective responses.

Clayton, K. L., D. R. Collins, J. Lengieza, M. Ghebremichael, F. Dotiwala, J. Lieberman, and B. D. Walker (2018). “Resistance of HIV-infected macrophages to CD8(+) T lymphocyte-mediated killing drives activation of the immune system”. In: *Nat Immunol* 19.5, pp. 475–486.

Clayton, K. L., J. V. Garcia, J. E. Clements, and B. D. Walker (2017). “HIV Infection of Macrophages: Implications for Pathogenesis and Cure”. In: *Pathog Immun* 2.2, pp. 179–192.

Cobbs, S. L. and J. L. Gooch (2007). “NFATc is required for TGFbeta-mediated transcriptional regulation of fibronectin”. In: *Biochem Biophys Res Commun* 362.2, pp. 288–94.

Coelho, C. (2022). “Itaconate or how I learned to stop avoiding the study of immunometabolism”. In: *PLoS Pathog* 18.3, e1010361.

- Coelho, C., A. L. Bocca, and A. Casadevall (2014). “The intracellular life of *Cryptococcus neoformans*”. In: *Annu Rev Pathol* 9, pp. 219–38.
- Coenjaerts, F. E., M. van der Flier, P. N. Mwinzi, A. E. Brouwer, J. Scharringa, W. S. Chaka, M. Aarts, A. Rajanuwong, D. A. van de Vijver, T. S. Harrison, and A. I. Hoepelman (2004). “Intrathecal production and secretion of vascular endothelial growth factor during Cryptococcal Meningitis”. In: *J Infect Dis* 190.7, pp. 1310–7.
- Cohen, S. B., B. H. Gern, J. L. Delahaye, K. N. Adams, C. R. Plumlee, J. K. Winkler, D. R. Sherman, M. Y. Gerner, and K. B. Urdahl (2018). “Alveolar Macrophages Provide an Early *Mycobacterium tuberculosis* Niche and Initiate Dissemination”. In: *Cell Host Microbe* 24.3, 439–446 e4.
- Colella, M., F. Grisan, V. Robert, J. D. Turner, A. P. Thomas, and T. Pozzan (2008). “Ca²⁺ oscillation frequency decoding in cardiac cell hypertrophy: role of calcineurin/NFAT as Ca²⁺ signal integrators”. In: *Proc Natl Acad Sci U S A* 105.8, pp. 2859–64.
- Collin, M. and V. Bigley (2018). “Human dendritic cell subsets: an update”. In: *Immunology* 154.1, pp. 3–20.
- Colombo, M., L. Marongiu, F. Mingozzi, R. Marzi, C. Cigni, F. A. Facchini, R. Rotem, M. Valache, G. Stucchi, G. Rocca, L. Gornati, M. A. Rizzuto, L. Salvioni, I. Zanoni, A. Gori, D. Prosperi, and F. Granucci (2022). “Specific immunosuppressive role of nanodrugs targeting calcineurin in innate myeloid cells”. In: *iScience* 25.10, p. 105042.
– *Consistent with the growing interest in NFAT over the past two years, this offers a method and a rationale for myeloid-targeted NFAT inhibition through use of nanodrugs. This should serve as a model going forward for how to target this pathway in a restricted subset of cells.*
- Combs, M. D. and K. E. Yutzey (2009). “VEGF and RANKL regulation of NFATc1 in heart valve development”. In: *Circ Res* 105.6, pp. 565–74.
- Compans, R. W. and P. C. Roberts (1994). “Viruses as model systems in cell biology”. In: *Methods Cell Biol* 43 Pt A, pp. 3–42.
- Comroe J. H., Jr. (1978). “Pay dirt: the story of streptomycin. Part I. From Waksman to Waksman”. In: *Am Rev Respir Dis* 117.4, pp. 773–81.

Conboy, I. M., D. Manoli, V. Mhaiskar, and P. P. Jones (1999). “Calcineurin and vacuolar-type H⁺-ATPase modulate macrophage effector functions”. In: *Proc Natl Acad Sci U S A* 96.11, pp. 6324–9.

– This publication outlined an antagonistic relationship between NFAT and NF-κB signaling within macrophages in response to LPS, a phenomenon that urgently needs to be integrated into existing models for the activity of both pathways. This is the first publication to investigate the intersection between these pathway and this negative feedback phenomenon is an interesting mechanism to limit inflammation after LPS detection.

Cooper, A. M., D. K. Dalton, T. A. Stewart, J. P. Griffin, D. G. Russell, and I. M. Orme (1993). “Disseminated tuberculosis in interferon gamma gene-disrupted mice”. In: *J Exp Med* 178.6, pp. 2243–7.

Cooper, A. M., J. Magram, J. Ferrante, and I. M. Orme (1997). “Interleukin 12 (IL-12) is crucial to the development of protective immunity in mice intravenously infected with *Mycobacterium tuberculosis*”. In: *J Exp Med* 186.1, pp. 39–45.

Corcoran, S. E. and L. A. O’Neill (2016). “HIF1alpha and metabolic reprogramming in inflammation”. In: *J Clin Invest* 126.10, pp. 3699–3707.

Cordelières, F. P. (2005). *Metamorph nd & ROI files importer (nd stack builder)*. URL: https://imagejdocu.list.lu/plugin/inputoutput/nd_stacks_builder/start.

Corleis, B. and A. Dorhoi (2020). “Early dynamics of innate immunity during pulmonary tuberculosis”. In: *Immunol Lett* 221, pp. 56–60.

Correa, A. F., A. M. Bailao, I. M. Bastos, I. M. Orme, C. M. Soares, A. Kipnis, J. M. Santana, and A. P. Junqueira-Kipnis (2014). “The endothelin system has a significant role in the pathogenesis and progression of *Mycobacterium tuberculosis* infection”. In: *Infect Immun* 82.12, pp. 5154–65.

Cortie, K. de, N. S. Russell, R. P. Coppes, F. A. Stewart, and M. Scharpfenecker (2014). “Bone marrow-derived macrophages incorporate into the endothelium and influence vascular and renal function after irradiation”. In: *Int J Radiat Biol* 90.9, pp. 769–77.

Cosma, C. L., O. Humbert, and L. Ramakrishnan (2004). “Superinfect-

ing mycobacteria home to established tuberculous granulomas". In: *Nat Immunol* 5.8, pp. 828–35.

– *Is the granuloma a host-protective or a host-maladaptive response? Given the findings presented in this paper on mycobacteria homing to existing granulomas and entering the necrotic core unscathed, it was an early suggestion that the granuloma was the basic site of mycobacterial subversion of host immune responses and set off a series of studies on this phenomenon, including a contemporary study (Volkman, Clay, Beery, Chang, Sherman, and Ramakrishnan, 2004) on this precise effect and how mycobacteria actively facilitate granuloma formation for their own benefit.*

Cosma, C. L., L. E. Swaim, H. Volkman, L. Ramakrishnan, and J. M. Davis (2006). "Zebrafish and frog models of *Mycobacterium marinum* infection". In: *Curr Protoc Microbiol* Chapter 10, Unit 10B 2.

Couper, K. N., D. G. Blount, and E. M. Riley (2008). "IL-10: the master regulator of immunity to infection". In: *J Immunol* 180.9, pp. 5771–7.

Courtney, R., D. C. Ngo, N. Malik, K. Ververis, S. M. Tortorella, and T. C. Karagiannis (2015). "Cancer metabolism and the Warburg effect: the role of HIF-1 and PI3K". In: *Mol Biol Rep* 42.4, pp. 841–51.

Courtwright, A., S. Siamakpour-Reihani, J. L. Arbiser, N. Banet, E. Hilliard, L. Fried, C. Livasy, D. Ketelsen, D. B. Nepal, C. M. Perou, C. Patterson, and N. Klauber-Demore (2009). "Secreted frizzle-related protein 2 stimulates angiogenesis via a calcineurin/NFAT signaling pathway". In: *Cancer Res* 69.11, pp. 4621–8.

Cousins, F. L., A. A. Murray, J. P. Scanlon, and P. T. Saunders (2016). "Hypoxyprobe reveals dynamic spatial and temporal changes in hypoxia in a mouse model of endometrial breakdown and repair". In: *BMC Res Notes* 9, p. 30.

Covarrubias, A. J., H. I. Aksoylar, and T. Horng (2015). "Control of macrophage metabolism and activation by mTOR and Akt signaling". In: *Semin Immunol* 27.4, pp. 286–96.

Covassin, L. D., J. A. Villefranc, M. C. Kacergis, B. M. Weinstein, and N. D. Lawson (2006). "Distinct genetic interactions between multiple Vegf receptors are required for development of different blood vessel types in zebrafish". In: *Proc Natl Acad Sci U S A* 103.17, pp. 6554–9.

Covian, C., A. Fernandez-Fierro, A. Retamal-Diaz, F. E. Diaz, A. E. Vasquez, M. K. Lay, C. A. Riedel, P. A. Gonzalez, S. M. Bueno, and A. M. Kalergis (2019). “BCG-Induced Cross-Protection and Development of Trained Immunity: Implication for Vaccine Design”. In: *Front Immunol* 10, p. 2806.

Cox, J. S., B. Chen, M. McNeil, and Jr. Jacobs W. R. (1999). “Complex lipid determines tissue-specific replication of *Mycobacterium tuberculosis* in mice”. In: *Nature* 402.6757, pp. 79–83.

Crabtree, G. R. and E. N. Olson (2002). “NFAT signaling: choreographing the social lives of cells”. In: *Cell* 109 Suppl, S67–79.

– This older review analyzed the diverse roles that had been uncovered for NFAT signaling until 2002, which encompassed a rather wide scope of tissues and functions. This vividly demonstrated the importance of this pathway in nearly every tissue and more and more roles have been uncovered in the two decades since.

Cramer, T. and R. S. Johnson (2003). “A novel role for the hypoxia inducible transcription factor HIF-1alpha: critical regulation of inflammatory cell function”. In: *Cell Cycle* 2.3, pp. 192–3.

Creagh, E. M. and L. A. O’Neill (2006). “TLRs, NLRs and RLRs: a trinity of pathogen sensors that co-operate in innate immunity”. In: *Trends Immunol* 27.8, pp. 352–7.

Cribbs, S. K., J. Lennox, A. M. Caliendo, L. A. Brown, and D. M. Guidot (2015). “Healthy HIV-1-infected individuals on highly active antiretroviral therapy harbor HIV-1 in their alveolar macrophages”. In: *AIDS Res Hum Retroviruses* 31.1, pp. 64–70.

Cronan, M. R. (2022). “In the Thick of It: Formation of the Tuberculous Granuloma and Its Effects on Host and Therapeutic Responses”. In: *Front Immunol* 13, p. 820134.

Cronan, M. R., R. W. Beerman, A. F. Rosenberg, J. W. Saelens, M. G. Johnson, S. H. Oehlers, D. M. Sisk, K. L. Jurcic Smith, N. A. Medvitz, S. E. Miller, L. A. Trinh, S. E. Fraser, J. F. Madden, J. Turner, J. E. Stout, S. Lee, and D. M. Tobin (2016). “Macrophage Epithelial Reprogramming Underlies Mycobacterial Granuloma Formation and Promotes Infection”. In: *Immunity* 45.4, pp. 861–876.

Cronan, M. R., E. J. Hughes, W. J. Brewer, G. Viswanathan, E. G. Hunt, B. Singh, S. Mehra, S. H. Oehlers, S. G. Gregory, D. Kaushal, and D. M. Tobin (2021). “A non-canonical type 2 immune response coordinates tuberculous granuloma formation and epithelialization”. In: *Cell* 184.7, 1757–1774 e14.

Cronan, M. R., M. A. Matty, A. F. Rosenberg, L. Blanc, C. J. Pyle, S. T. Espenschied, J. F. Rawls, V. Dartois, and D. M. Tobin (2018). “An explant technique for high-resolution imaging and manipulation of mycobacterial granulomas”. In: *Nat Methods* 15.12, pp. 1098–1107.

Cronan, M. R., A. F. Rosenberg, S. H. Oehlers, J. W. Saelens, D. M. Sisk, K. L. Jurcic Smith, S. Lee, and D. M. Tobin (2015). “CLARITY and PACT-based imaging of adult zebrafish and mouse for whole-animal analysis of infections”. In: *Dis Model Mech* 8.12, pp. 1643–50.

Cronan, M. R. and D. M. Tobin (2014). “Fit for consumption: zebrafish as a model for tuberculosis”. In: *Dis Model Mech* 7.7, pp. 777–84.

Cudkowicz, L. (1952). “The blood supply of the lung in pulmonary tuberculosis”. In: *Thorax* 7.3, pp. 270–6.

– This paper contains a number of radiographs that clearly show the vascular involvement of tuberculosis and was an early description of this phenomenon. While they had no means by which to dissect the cause or contributions of this vasculature to the disease, it is a reminder of the relevance of angiogenesis to human tuberculosis disease and that it is not just an abstract phenomenon observable in the zebrafish.

Cui, P., P. Zhang, L. Yuan, L. Wang, X. Guo, G. Cui, Y. Zhang, M. Li, X. Zhang, X. Li, Y. Yin, and Z. Yu (2021). “HIF-1alpha Affects the Neural Stem Cell Differentiation of Human Induced Pluripotent Stem Cells via MFN2-Mediated Wnt/beta-Catenin Signaling”. In: *Front Cell Dev Biol* 9, p. 671704.

Cursiefen, C., L. Chen, L. P. Borges, D. Jackson, J. Cao, C. Radziejewski, P. A. D’Amore, M. R. Dana, S. J. Wiegand, and J. W. Streilein (2004). “VEGF-A stimulates lymphangiogenesis and hemangiogenesis in inflammatory neovascularization via macrophage recruitment”. In: *J Clin Invest* 113.7, pp. 1040–50.

Dallenga, T., U. Repnik, B. Corleis, J. Eich, R. Reimer, G. W. Griffiths, and U. E. Schaible (2017). “*M. tuberculosis*-Induced Necrosis of Infected Neutrophils Pro-

motes Bacterial Growth Following Phagocytosis by Macrophages". In: *Cell Host Microbe* 22.4, 519–530 e3.

D'Amato, R. J., M. S. Loughnan, E. Flynn, and J. Folkman (1994). "Thalidomide is an inhibitor of angiogenesis". In: *Proc Natl Acad Sci U S A* 91.9, pp. 4082–5.

D'Ambrosio, L., R. Centis, G. Sotgiu, E. Pontali, A. Spanevello, and G. B. Migliori (2015). "New anti-tuberculosis drugs and regimens: 2015 update". In: *ERJ Open Res* 1.1.

Dambuza, I. M. and G. D. Brown (2015). "C-type lectins in immunity: recent developments". In: *Curr Opin Immunol* 32, pp. 21–7.

Dana, H., B. Mohar, Y. Sun, S. Narayan, A. Gordus, J. P. Hasseman, G. Tsegaye, G. T. Holt, A. Hu, D. Walpita, R. Patel, J. J. Macklin, C. I. Bargmann, M. B. Ahrens, E. R. Schreiter, V. Jayaraman, L. L. Looger, K. Svoboda, and D. S. Kim (2016). "Sensitive red protein calcium indicators for imaging neural activity". In: *Elife* 5.

Daniel, C., K. Gerlach, M. Vath, M. F. Neurath, and B. Weigmann (2014). "Nuclear factor of activated T cells - a transcription factor family as critical regulator in lung and colon cancer". In: *Int J Cancer* 134.8, pp. 1767–75.

Danilchanka, O., J. Sun, M. Pavlenok, C. Maueroder, A. Speer, A. Siroy, J. Marrero, C. Trujillo, D. L. Mayhew, K. S. Doornbos, L. E. Munoz, M. Herrmann, S. Ehrt, C. Berens, and M. Niederweis (2014). "An outer membrane channel protein of *Mycobacterium tuberculosis* with exotoxin activity". In: *Proc Natl Acad Sci U S A* 111.18, pp. 6750–5.

Darveau, R. P., T. T. Pham, K. Lemley, R. A. Reife, B. W. Bainbridge, S. R. Coats, W. N. Howald, S. S. Way, and A. M. Hajjar (2004). "*Porphyromonas gingivalis* lipopolysaccharide contains multiple lipid A species that functionally interact with both toll-like receptors 2 and 4". In: *Infect Immun* 72.9, pp. 5041–51.

Darwich, L., G. Coma, R. Pena, R. Bellido, E. J. Blanco, J. A. Este, F. E. Borras, B. Clotet, L. Ruiz, A. Rosell, F. Andreo, R. M. Parkhouse, and M. Bofill (2009). "Secretion of interferon-gamma by human macrophages demonstrated at the single-cell level after costimulation with interleukin (IL)-12 plus IL-18". In: *Immunology* 126.3, pp. 386–93.

Das, N., D. T. Murray, and T. A. Cross (2013). “Lipid bilayer preparations of membrane proteins for oriented and magic-angle spinning solid-state NMR samples”. In: *Nat Protoc* 8.11, pp. 2256–70.

Das, R., M. S. Koo, B. H. Kim, S. T. Jacob, S. Subbian, J. Yao, L. Leng, R. Levy, C. Murchison, W. J. Burman, C. C. Moore, W. M. Scheld, J. R. David, G. Kaplan, J. D. MacMicking, and R. Bucala (2013). “Macrophage migration inhibitory factor (MIF) is a critical mediator of the innate immune response to *Mycobacterium tuberculosis*”. In: *Proc Natl Acad Sci U S A* 110.32, E2997–3006.

Das, S., B. M. F. Pettersson, P. R. K. Behra, A. Mallick, M. Cheramie, M. Ramesh, L. Shirreff, T. DuCote, S. Dasgupta, D. G. Ennis, and L. A. Kirsebom (2018). “Extensive genomic diversity among *Mycobacterium marinum* strains revealed by whole genome sequencing”. In: *Sci Rep* 8.1, p. 12040.

Datta, M., L. E. Via, W. S. Kamoun, C. Liu, W. Chen, G. Seano, D. M. Weiner, D. Schimel, K. England, J. D. Martin, X. Gao, L. Xu, 3rd Barry C. E., and R. K. Jain (2015). “Anti-vascular endothelial growth factor treatment normalizes tuberculosis granuloma vasculature and improves small molecule delivery”. In: *Proc Natl Acad Sci U S A* 112.6, pp. 1827–32.

– *Concurrent with the work in Oehlers, Cronan, Scott, Thomas, Okuda, Walton, Beerman, Crosier, and Tobin (2015), Datta et al. clearly demonstrated the utility of VEGFA inhibition in enhancing drug delivery to the granuloma in a process mirroring that of cancer, where bevacizumab is able to normalize blood vessels in the proximity of the tumor to facilitate drug delivery. This is an excellent companion piece and very nicely investigates the endothelial cell biology in the proximity of both human and rabbit granulomas, giving us perhaps the best ever images of these vessels on histology.*

Datta, S. K., S. Okamoto, T. Hayashi, S. S. Shin, I. Mihajlov, A. Fermin, D. G. Guiney, J. Fierer, and E. Raz (2006). “Vaccination with irradiated *Listeria* induces protective T cell immunity”. In: *Immunity* 25.1, pp. 143–52.

Dauros-Singorenko, P., C. Blenkiron, A. Phillips, and S. Swift (2018). “The functional RNA cargo of bacterial membrane vesicles”. In: *FEMS Microbiol Lett* 365.5.

Davidson, A. E., D. Balciunas, D. Mohn, J. Shaffer, S. Hermanson, S. Sivasubbu, M. P. Cliff, P. B. Hackett, and S. C. Ekker (2003). “Efficient gene delivery and gene expression in zebrafish using the Sleeping Beauty transposon”. In: *Dev Biol* 263.2, pp. 191–202.

- Davies, L. C., S. J. Jenkins, J. E. Allen, and P. R. Taylor (2013a). “Tissue-resident macrophages”. In: *Nat Immunol* 14.10, pp. 986–95.
- Davies, L. C., M. Rosas, S. J. Jenkins, C. T. Liao, M. J. Scurr, F. Brombacher, D. J. Fraser, J. E. Allen, S. A. Jones, and P. R. Taylor (2013b). “Distinct bone marrow-derived and tissue-resident macrophage lineages proliferate at key stages during inflammation”. In: *Nat Commun* 4, p. 1886.
- Davies, P. and J. Grange (2001a). “The genetics of host resistance and susceptibility to tuberculosis”. In: *Ann N Y Acad Sci* 953, pp. 151–6.
- Davies, P. D. and J. M. Grange (2001b). “Factors affecting susceptibility and resistance to tuberculosis”. In: *Thorax* 56 Suppl 2, pp. ii23–9.
- Davis, A., G. Meintjes, and R. J. Wilkinson (2018). “Treatment of Tuberculous Meningitis and Its Complications in Adults”. In: *Curr Treat Options Neurol* 20.3, p. 5.
- Davis, J. M., H. Clay, J. L. Lewis, N. Ghori, P. Herbomel, and L. Ramakrishnan (2002).** “Real-time visualization of *Mycobacterium*-macrophage interactions leading to initiation of granuloma formation in zebrafish embryos”. In: *Immunity* 17.6, pp. 693–702.
- This is the study that founded the zebrafish-*Mycobacterium marinum* model of infection. Little else needs to be said.
- Davis, J. M., M. Huang, M. R. Botts, C. M. Hull, and A. Huttenlocher (2016). “A Zebrafish Model of Cryptococcal Infection Reveals Roles for Macrophages, Endothelial Cells, and Neutrophils in the Establishment and Control of Sustained Fungemia”. In: *Infect Immun* 84.10, pp. 3047–62.
- Davis, J. M. and L. Ramakrishnan (2009).** “The role of the granuloma in expansion and dissemination of early tuberculous infection”. In: *Cell* 136.1, pp. 37–49.
- Day, Carolyn (2017). *Consumptive chic : a history of beauty, fashion, and disease*. London ; New York: Bloomsbury Academic, an imprint of Bloomsbury Publishing Plc.
- Day, T. A., J. E. Mittler, M. R. Nixon, C. Thompson, M. D. Miner, M. J. Hickey, R. P. Liao, J. M. Pang, D. M. Shayakhmetov, and D. R. Sherman (2014). “My-

cobacterium tuberculosis strains lacking surface lipid phthiocerol dimycocerosate are susceptible to killing by an early innate host response". In: *Infect Immun* 82.12, pp. 5214–22.

Dayan, F., N. M. Mazure, M. C. Brahimi-Horn, and J. Pouyssegur (2008). "A dialogue between the hypoxia-inducible factor and the tumor microenvironment". In: *Cancer Microenviron* 1.1, pp. 53–68.

de Chastellier, C. (2009). "The many niches and strategies used by pathogenic mycobacteria for survival within host macrophages". In: *Immunobiology* 214.7, pp. 526–42.

De Maeyer, R. P. H. and E. S. Chambers (2021). "The impact of ageing on monocytes and macrophages". In: *Immunol Lett* 230, pp. 1–10.

De Palma, M., C. Murdoch, M. A. Venneri, L. Naldini, and C. E. Lewis (2007). "Tie2-expressing monocytes: regulation of tumor angiogenesis and therapeutic implications". In: *Trends Immunol* 28.12, pp. 519–24.

De Palma, M. and L. Naldini (2011). "Angiopoietin-2 TIEs up macrophages in tumor angiogenesis". In: *Clin Cancer Res* 17.16, pp. 5226–32.

De Rosa, L., R. Di Stasi, and L. D. D'Andrea (2018). "Pro-angiogenic peptides in biomedicine". In: *Arch Biochem Biophys* 660, pp. 72–86.

De Simone, A., M. N. Evanitsky, L. Hayden, B. D. Cox, J. Wang, V. A. Tornini, J. Ou, A. Chao, K. D. Poss, and S. Di Talia (2021). "Control of osteoblast regeneration by a train of Erk activity waves". In: *Nature* 590.7844, pp. 129–133.

Debinski, W., B. Slagle-Webb, M. G. Achen, S. A. Stacke, E. Tulchinsky, G. Y. Gillespie, and D. M. Gibo (2001). "VEGF-D is an X-linked/AP-1 regulated putative onco-angiogen in human glioblastoma multiforme". In: *Mol Med* 7.9, pp. 598–608.

Dedrick, R. M., K. G. Freeman, J. A. Nguyen, A. Bahadirli-Talbott, B. E. Smith, A. E. Wu, A. S. Ong, C. T. Lin, L. C. Ruppel, N. M. Parrish, G. F. Hatfull, and K. A. Cohen (2021). "Potent antibody-mediated neutralization limits bacteriophage treatment of a pulmonary *Mycobacterium abscessus* infection". In: *Nat Med* 27.8, pp. 1357–1361.

Dedrick, R. M., C. A. Guerrero-Bustamante, R. A. Garlena, D. A. Russell, K. Ford, K. Harris, K. C. Gilmour, J. Soothill, D. Jacobs-Sera, R. T. Schooley, G. F. Hatfull, and H. Spencer (2019). “Engineered bacteriophages for treatment of a patient with a disseminated drug-resistant *Mycobacterium abscessus*”. In: *Nat Med* 25.5, pp. 730–733.

Dedrick, R. M., B. E. Smith, M. Cristinziano, K. G. Freeman, D. Jacobs-Sera, Y. Belessis, A. Whitney Brown, K. A. Cohen, R. M. Davidson, D. van Duin, A. Gainey, C. B. Garcia, C. R. Robert George, G. Haidar, W. Ip, J. Iredell, A. Khatami, J. S. Little, K. Malmivaara, B. J. McMullan, D. E. Michalik, A. Moscatelli, J. A. Nick, M. G. Tupayachi Ortiz, H. M. Polenakovic, P. D. Robinson, M. Skurnik, D. A. Solomon, J. Soothill, H. Spencer, P. Wark, A. Worth, R. T. Schooley, C. A. Benson, and G. F. Hatfull (2022). “Phage Therapy of *Mycobacterium* Infections: Compassionate-use of Phages in Twenty Patients with Drug-Resistant Mycobacterial Disease”. In: *Clin Infect Dis*.

Deerhake, M. E., K. Danzaki, M. Inoue, E. D. Cardakli, T. Nonaka, N. Aggarwal, W. E. Barclay, R. R. Ji, and M. L. Shinohara (2021). “Dectin-1 limits autoimmune neuroinflammation and promotes myeloid cell-astrocyte crosstalk via Card9-independent expression of Oncostatin M”. In: *Immunity* 54.3, 484–498 e8.

– *The first in this recent wave of publications on the role of NFAT signaling in myeloid cells, this study set the groundwork for the next generation of study in this pathway. The authors found that, in a model of experimental autoimmune encephalomyelitis, the activation of DECTIN-1 offered protection against disease specifically through the activity of NFAT within myeloid cells while CARD9 activity resulted in enhanced pathology. This revealed not only the Janus faced nature of C-type lectin activation, but ascribed a distinct, protective role to NFAT signaling during this autoimmune context.*

Deisseroth, K. (2015). “Optogenetics: 10 years of microbial opsins in neuroscience”. In: *Nat Neurosci* 18.9, pp. 1213–25.

Delafont, V., F. Mougari, E. Cambau, M. Joyeux, D. Bouchon, Y. Hechard, and L. Moulin (2014). “First evidence of amoebae-mycobacteria association in drinking water network”. In: *Environ Sci Technol* 48.20, pp. 11872–82.

Delling, U., J. Tureckova, H. W. Lim, L. J. De Windt, P. Rotwein, and J. D. Molkentin (2000). “A calcineurin-NFATc3-dependent pathway regulates skeletal muscle differentiation and slow myosin heavy-chain expression”. In: *Mol Cell Biol* 20.17, pp. 6600–11.

Demetz, E., P. Tymoszuk, R. Hilbe, C. Volani, D. Haschka, C. Heim, K. Auer, D. Lener, L. B. Zeiger, C. Pfeifhofer-Obermair, A. Boehm, G. J. Obermair, C. Ablinger, S. Coassini, C. Lamina, J. Kager, V. Petzer, M. Asshoff, A. Schroll, M. Nairz, S. Dichtl, M. Seifert, L. von Raffay, C. Fischer, M. Barros-Pinkelnig, N. Brigo, L. Valente de Souza, S. Sopper, J. Hirsch, M. Gruber, C. Gollmann-Tepelkoylu, J. Holfeld, J. Halper, S. Macheiner, J. Gostner, G. F. Vogel, R. Pechlaner, P. Moser, M. Imboden, P. Marques-Vidal, N. M. Probst-Hensch, H. Meiselbach, K. Strauch, A. Peters, B. Paulweber, J. Willeit, S. Kiechl, F. Kronenberg, I. Theurl, I. Tancevski, and G. Weiss (2020). "The haemochromatosis gene Hfe and Kupffer cells control LDL cholesterol homeostasis and impact on atherosclerosis development". In: *Eur Heart J* 41.40, pp. 3949–3959.

den Dunnen, J., S. I. Gringhuis, and T. B. Geijtenbeek (2009). "Innate signaling by the C-type lectin DC-SIGN dictates immune responses". In: *Cancer Immunol Immunother* 58.7, pp. 1149–57.

Deng, Y., Y. Qian, M. Meng, H. Jiang, Y. Dong, C. Fang, S. He, and L. Yang (2022). "Extensive sequence divergence between the reference genomes of two zebrafish strains, Tuebingen and AB". In: *Mol Ecol Resour* 22.6, pp. 2148–2157.

Derbyshire, Keith M., Todd A. Gray, Graham F. Hatfull, and William R. Jacobs Jr (2014). "Distributive Conjugal Transfer: New Insights into Horizontal Gene Transfer and Genetic Exchange in Mycobacteria". In: *Microbiology Spectrum* 2.1.

Desvignes, L., A. J. Wolf, and J. D. Ernst (2012). "Dynamic roles of type I and type II IFNs in early infection with *Mycobacterium tuberculosis*". In: *J Immunol* 188.12, pp. 6205–15.

Deyerle, K. L., J. E. Sims, S. K. Dower, and M. A. Bothwell (1992). "Pattern of IL-1 receptor gene expression suggests role in noninflammatory processes". In: *J Immunol* 149.5, pp. 1657–65.

Diacovich, L. and J. P. Gorvel (2010). "Bacterial manipulation of innate immunity to promote infection". In: *Nat Rev Microbiol* 8.2, pp. 117–28.

Diallo, T., M. Adjibimey, R. Ruslami, A. Trajman, O. Sow, J. Obeng Baah, G. B. Marks, R. Long, K. Elwood, D. Zielinski, M. Gninafon, D. A. Wulandari, L. Apriani, C. Valiquette, F. Fregonese, K. Hornby, P. Z. Li, P. C. Hill, K. Schwartzman, A. Benedetti, and D. Menzies (2018). "Safety and Side Effects of Rifampin versus Isoniazid in Children". In: *N Engl J Med* 379.5, pp. 454–463.

Dickinson, Mary S., Miriam Kutsch, Linda Sistemich, Dulcemaria Hernandez, Anthony S. Piro, David Needham, Cammie F. Lesser, Christian Herrmann, and Jörn Coers (2022). “GBP2 aggregates LPS and activates the caspase-4 inflammasome independent of the bacterial encapsulation factor GBP1”. In: *bioRxiv*.

Diehl, S., C. W. Chow, L. Weiss, A. Palmetshofer, T. Twardzik, L. Rounds, E. Serfling, R. J. Davis, J. Anguita, and M. Rincon (2002). “Induction of NFATc2 expression by interleukin 6 promotes T helper type 2 differentiation”. In: *J Exp Med* 196.1, pp. 39–49.

– This nice study delineated a pathway downstream of IL-6R activation that results in the production of IL-4 and differentiation of T cells into T_H2 cells.

Dietrich, T., J. Onderka, F. Bock, F. E. Kruse, D. Vossmeyer, R. Stragies, G. Zahn, and C. Cursiefen (2007). “Inhibition of inflammatory lymphangiogenesis by integrin alpha5 blockade”. In: *Am J Pathol* 171.1, pp. 361–72.

Dillon, N. A., N. D. Peterson, H. A. Feaga, K. C. Keiler, and A. D. Baughn (2017). “Anti-tubercular Activity of Pyrazinamide is Independent of trans-Translation and RpsA”. In: *Sci Rep* 7.1, p. 6135.

Dimmeler, S., J. Haendeler, and A. M. Zeiher (2002). “Regulation of endothelial cell apoptosis in atherosclerosis”. In: *Curr Opin Lipidol* 13.5, pp. 531–6.

Dineen, S. P., K. D. Lynn, S. E. Holloway, A. F. Miller, J. P. Sullivan, D. S. Shames, A. W. Beck, C. C. Barnett, J. B. Fleming, and R. A. Brekken (2008). “Vascular endothelial growth factor receptor 2 mediates macrophage infiltration into orthotopic pancreatic tumors in mice”. In: *Cancer Res* 68.11, pp. 4340–6.

Divangahi, M., P. Aaby, S. A. Khader, L. B. Barreiro, S. Bekkering, T. Chavakis, R. van Crevel, N. Curtis, A. R. DiNardo, J. Dominguez-Andres, R. Duivenvoorden, S. Fanucchi, Z. Fayad, E. Fuchs, M. Hamon, K. L. Jeffrey, N. Khan, L. A. B. Joosten, E. Kaufmann, E. Latz, G. Matarese, J. W. M. van der Meer, M. Mhlanga, Sjcfm Moorlag, W. J. M. Mulder, S. Naik, B. Novakovic, L. O’Neill, J. Ochando, K. Ozato, N. P. Riksen, R. Sauerwein, E. R. Sherwood, A. Schlitzer, J. L. Schultze, M. H. Sieweke, C. S. Benn, H. Stunnenberg, J. Sun, F. L. van de Veerdonk, S. Weis, D. L. Williams, R. Xavier, and M. G. Netea (2021). “Trained immunity, tolerance, priming and differentiation: distinct immunological processes”. In: *Nat Immunol* 22.1, pp. 2–6.

Divangahi, M., S. M. Behar, and H. Remold (2013). “Dying to live: how the death modality of the infected macrophage affects immunity to tuberculosis”. In: *Adv Exp Med Biol* 783, pp. 103–20.

Dixon, D. R. and R. P. Darveau (2005). “Lipopolysaccharide heterogeneity: innate host responses to bacterial modification of lipid a structure”. In: *J Dent Res* 84.7, pp. 584–95.

Dockterman, J. and J. Coers (2022). “How did we get here? Insights into mechanisms of immunity-related GTPase targeting to intracellular pathogens”. In: *Curr Opin Microbiol* 69, p. 102189.

Dodd, R. B. and K. Drickamer (2001). “Lectin-like proteins in model organisms: implications for evolution of carbohydrate-binding activity”. In: *Glycobiology* 11.5, 71R–9R.

Dolmetsch, R. E., R. S. Lewis, C. C. Goodnow, and J. I. Healy (1997). “Differential activation of transcription factors induced by Ca²⁺ response amplitude and duration”. In: *Nature* 386.6627, pp. 855–8.

– *An early study on the relationship between NF-κB and NFAT, they found that NF-κB was sensitive to transient changes in intracellular calcium while NFAT required longer-lived calcium increases to be induced. Whether this pattern is general to all NF-κB and NFAT family members is unknown, but would be an intriguing study to differentiate the behavior of these different isoforms.*

Dominguez-Medina, C. C., M. Perez-Toledo, A. E. Schager, J. L. Marshall, C. N. Cook, S. Bobat, H. Hwang, B. J. Chun, E. Logan, J. A. Bryant, W. M. Channell, F. C. Morris, S. E. Jossi, A. Alshayea, A. E. Rossiter, P. A. Barrow, W. G. Horsnell, C. A. MacLennan, I. R. Henderson, J. H. Lakey, J. C. Gumbart, C. Lopez-Macias, V. N. Bavro, and A. F. Cunningham (2020). “Outer membrane protein size and LPS O-antigen define protective antibody targeting to the Salmonella surface”. In: *Nat Commun* 11.1, p. 851.

Donald, P. R. and A. H. Diacon (2015). “Para-aminosalicylic acid: the return of an old friend”. In: *Lancet Infect Dis* 15.9, pp. 1091–1099.

Dong, C., R. J. Davis, and R. A. Flavell (2002). “MAP kinases in the immune response”. In: *Annu Rev Immunol* 20, pp. 55–72.

- Donnachie, E., E. P. Fedotova, and S. A. Hwang (2016). “Trehalose 6,6-Dimycolate from *Mycobacterium tuberculosis* Induces Hypercoagulation”. In: *Am J Pathol* 186.5, pp. 1221–33.
- Donnem, T., A. R. Reynolds, E. A. Kuczynski, K. Gatter, P. B. Vermeulen, R. S. Kerbel, A. L. Harris, and F. Pezzella (2018). “Non-angiogenic tumours and their influence on cancer biology”. In: *Nat Rev Cancer* 18.5, pp. 323–336.
- Dor, Y., V. Djonov, R. Abramovitch, A. Itin, G. I. Fishman, P. Carmeliet, G. Goelman, and E. Keshet (2002). “Conditional switching of VEGF provides new insights into adult neovascularization and pro-angiogenic therapy”. In: *EMBO J* 21.8, pp. 1939–47.
- Dorhoi, A., C. Desel, V. Yeremeev, L. Pradl, V. Brinkmann, H. J. Mollenkopf, K. Hanke, O. Gross, J. Ruland, and S. H. Kaufmann (2010). “The adaptor molecule CARD9 is essential for tuberculosis control”. In: *J Exp Med* 207.4, pp. 777–92.
– *CARD9’s essentiality for control of tuberculosis indicates an important role for C-type lectin signaling in host defense, but the precise contributions of discrete C-type lectin receptors in inducing this phenotype remain mysterious. It may be that cutting off the root is needed to fully abrogate C-type lectin signaling responses. Whether NFAT is then hyperactivated in the absence of CARD9 is not known but is a compelling possibility that might explain the exacerbated pathology. Simultaneous limitation of NFAT may have provided some degree of protection, but this is unknown from the results presented.*
- Dorman, S. E., C. L. Hatem, S. Tyagi, K. Aird, J. Lopez-Molina, M. L. Pitt, B. C. Zook, Jr. Dannenberg A. M., W. R. Bishai, and Y. C. Manabe (2004). “Susceptibility to tuberculosis: clues from studies with inbred and outbred New Zealand White rabbits”. In: *Infect Immun* 72.3, pp. 1700–5.
- Dorman, S. E., P. Nahid, E. V. Kurbatova, P. P. J. Phillips, K. Bryant, K. E. Dooley, M. Engle, S. V. Goldberg, H. T. T. Phan, J. Hakim, J. L. Johnson, M. Lourens, N. A. Martinson, G. Muzanyi, K. Narunsky, S. Nerette, N. V. Nguyen, T. H. Pham, S. Pierre, A. E. Purfield, W. Samaneka, R. M. Savic, I. Sanne, N. A. Scott, J. Shenje, E. Sizemore, A. Vernon, Z. Waja, M. Weiner, S. Swindells, R. E. Chaisson, Aids Clinical Trials Group, and Consortium Tuberculosis Trials (2021). “Four-Month Rifapentine Regimens with or without Moxifloxacin for Tuberculosis”. In: *N Engl J Med* 384.18, pp. 1705–1718.
- Driever, W., L. Solnica-Krezel, A. F. Schier, S. C. Neuhauss, J. Malicki, D. L. Stemple, D. Y. Stainier, F. Zwartkruis, S. Abdelilah, Z. Rangini, J. Belak, and C. Boggs

(1996). “A genetic screen for mutations affecting embryogenesis in zebrafish”. In: *Development* 123, pp. 37–46.

Driever, W., D. Stemple, A. Schier, and L. Solnica-Krezel (1994). “Zebrafish: genetic tools for studying vertebrate development”. In: *Trends Genet* 10.5, pp. 152–9.

Driver, E. R., G. J. Ryan, D. R. Hoff, S. M. Irwin, R. J. Basaraba, I. Kramnik, and A. J. Lenaerts (2012). “Evaluation of a mouse model of necrotic granuloma formation using C3HeB/FeJ mice for testing of drugs against *Mycobacterium tuberculosis*”. In: *Antimicrob Agents Chemother* 56.6, pp. 3181–95.

Drobniewski, F. A., A. L. Pozniak, and A. H. Uttley (1995). “Tuberculosis and AIDS”. In: *J Med Microbiol* 43.2, pp. 85–91.

Drummond, R. A. and G. D. Brown (2011). “The role of Dectin-1 in the host defence against fungal infections”. In: *Curr Opin Microbiol* 14.4, pp. 392–9.

– (2013). “Signalling C-type lectins in antimicrobial immunity”. In: *PLoS Pathog* 9.7, e1003417.

Drummond, R. A., L. M. Franco, and M. S. Lionakis (2018). “Human CARD9: A Critical Molecule of Fungal Immune Surveillance”. In: *Front Immunol* 9, p. 1836.

Drummond, R. A. and M. S. Lionakis (2016). “Mechanistic Insights into the Role of C-Type Lectin Receptor/CARD9 Signaling in Human Antifungal Immunity”. In: *Front Cell Infect Microbiol* 6, p. 39.

Drummond, R. A., S. Saijo, Y. Iwakura, and G. D. Brown (2011). “The role of Syk/CARD9 coupled C-type lectins in antifungal immunity”. In: *Eur J Immunol* 41.2, pp. 276–81.

Du, M., X. Wang, L. Yuan, B. Liu, X. Mao, D. Huang, L. Yang, K. Huang, F. Zhang, Y. Wang, X. Luo, C. Wang, J. Peng, M. Liang, D. Huang, and K. Huang (2020). “Targeting NFATc4 attenuates non-alcoholic steatohepatitis in mice”. In: *J Hepatol* 73.6, pp. 1333–1346.

– *Non-transcriptional role for NFAT proteins is an intriguing additional layer of mechanistic regulation but little work has been done to identify these protein:protein interactions or their functional consequences. This very nice study found that NFATC4 bound PPAR α in the nucleus to inhibit its activity and that knockdown of*

NFATC4 led to reductions in hepatic steatosis in a mouse model of non-alcoholic steatohepatitis. This liberation of PPAR α resulted in improved lipid metabolism that was protective against disease.

Du, M., L. Yang, B. Liu, L. Yang, X. Mao, M. Liang, and K. Huang (2021). “Inhibition of NFAT suppresses foam cell formation and the development of diet-induced atherosclerosis”. In: *FASEB J* 35.10, e21951.

– A complement to the previous study from the same group, this one found that NFATC2 bound to PPAR γ to inhibit its activity and that NFATC2 activity was essential for the formation of foam cells or foamy macrophages, which are prevalent during atherosclerosis as well as in tuberculosis. This argues for a model where NFAT-PPAR interactions are important determinants of various lipid metabolism responses and integrates NFAT into a more general model of metabolism within macrophages.

Duan, J., H. Hu, L. Feng, X. Yang, and Z. Sun (2017). “Silica nanoparticles inhibit macrophage activity and angiogenesis via VEGFR2-mediated MAPK signaling pathway in zebrafish embryos”. In: *Chemosphere* 183, pp. 483–490.

Dubos, René J. and Jean Dubos (1987). *The White Plague: Tuberculosis, Man, and Society.* New Brunswick: Rutgers University Press.

Dubovsky, H. (1988). “The history of para-aminosalicylic acid (PAS), the first tuberculosis anti-microbial agent, and streptomycin (SM): a comparative study), the first tuberculosis anti-microbial agent, and streptomycin (SM): a comparative study”. In: *Adler Mus Bull* 14.1, pp. 7–11.

Dudiki, T., J. Meller, G. Mahajan, H. Liu, I. Zhevlakova, S. Stefl, C. Witherow, E. Podrez, C. R. Kothapalli, and T. V. Byzova (2020). “Microglia control vascular architecture via a TGFbeta1 dependent paracrine mechanism linked to tissue mechanics”. In: *Nat Commun* 11.1, p. 986.

Dumm, R. E., J. K. Fiege, B. M. Waring, C. T. Kuo, R. A. Langlois, and N. S. Heaton (2019). “Non-lytic clearance of influenza B virus from infected cells preserves epithelial barrier function”. In: *Nat Commun* 10.1, p. 779.

Dumm, R. E., S. A. Wellford, E. A. Moseman, and N. S. Heaton (2020). “Heterogeneity of Antiviral Responses in the Upper Respiratory Tract Mediates Differential Non-lytic Clearance of Influenza Viruses”. In: *Cell Rep* 32.9, p. 108103.

- Duong, L., H. G. Radley, B. Lee, D. E. Dye, F. J. Pixley, M. D. Grounds, D. J. Nelson, and C. Jackaman (2021). “Macrophage function in the elderly and impact on injury repair and cancer”. In: *Immun Ageing* 18.1, p. 4.
- Duong, T., P. Koopman, and M. Francois (2012). “Tumor lymphangiogenesis as a potential therapeutic target”. In: *J Oncol* 2012, p. 204946.
- Duran, C. L., L. Borriello, G. S. Karagiannis, D. Entenberg, M. H. Oktay, and J. S. Condeelis (2021). “Targeting Tie2 in the Tumor Microenvironment: From Angiogenesis to Dissemination”. In: *Cancers (Basel)* 13.22.
- Dusseau, J. W., P. M. Hutchins, and D. S. Malbasa (1986). “Stimulation of angiogenesis by adenosine on the chick chorioallantoic membrane”. In: *Circ Res* 59.2, pp. 163–70.
- Dutta, S. and Y. Ahmad (2011). “The efficacy and safety of tacrolimus in rheumatoid arthritis”. In: *Ther Adv Musculoskeletal Dis* 3.6, pp. 283–91.
- Duval, H., M. Harris, J. Li, N. Johnson, and C. Print (2003). “New insights into the function and regulation of endothelial cell apoptosis”. In: *Angiogenesis* 6.3, pp. 171–83.
- Dvinge, H. and P. Bertone (2009). “HTqPCR: high-throughput analysis and visualization of quantitative real-time PCR data in R”. In: *Bioinformatics* 25.24, pp. 3325–6.
- Dziadek, J., S. A. Rutherford, M. V. Madiraju, M. A. L. Atkinson, and M. Rajagopalan (2003). “Conditional expression of *Mycobacterium smegmatis* ftsZ, an essential cell division gene”. In: *Microbiology (Reading)* 149.Pt 6, pp. 1593–1603.
- Eagle, H. (1954). “The multiple mechanisms of penicillin resistance”. In: *J Bacteriol* 68.5, pp. 610–6.
- Echeverria-Valencia, G., M. Silva-Miranda, E. Ekaza, A. J. Vallecillo, C. Parada, I. Sada-Ovalle, F. Altare, and C. Espitia (2019). “Interaction of mycobacteria with Plasmin(ogen) affects phagocytosis and granuloma development”. In: *Tuberculosis (Edinb)* 117, pp. 36–44.
- Eerola, S. K., N. M. Santio, S. Rinne, P. Kouvonnen, G. L. Corthals, M. Scaravilli, G. Scala, A. Serra, D. Greco, P. Ruusuvuori, L. Latonen, E. M. Rainio, T. Visakorpi,

and P. J. Koskinen (2019). “Phosphorylation of NFATC1 at PIM1 target sites is essential for its ability to promote prostate cancer cell migration and invasion”. In: *Cell Commun Signal* 17.1, p. 148.

Eferl, R. and E. F. Wagner (2003). “AP-1: a double-edged sword in tumorigenesis”. In: *Nat Rev Cancer* 3.11, pp. 859–68.

Ehlers, M. L., B. Celona, and B. L. Black (2014). “NFATc1 controls skeletal muscle fiber type and is a negative regulator of MyoD activity”. In: *Cell Rep* 8.6, pp. 1639–1648.

Ehlers, S. and U. E. Schaible (2012). “The granuloma in tuberculosis: dynamics of a host-pathogen collusion”. In: *Front Immunol* 3, p. 411.

Ehrt, S., K. Rhee, and D. Schnappinger (2015). “Mycobacterial genes essential for the pathogen’s survival in the host”. In: *Immunol Rev* 264.1, pp. 319–26.

Ehrt, S., D. Schnappinger, and K. Y. Rhee (2018). “Metabolic principles of persistence and pathogenicity in *Mycobacterium tuberculosis*”. In: *Nat Rev Microbiol* 16.8, pp. 496–507.

Eisen, J. S. (2020). “History of zebrafish research”. In: Elsevier, pp. 3–14.

Ekins, S. (2014). “Hacking into the granuloma: could antibody antibiotic conjugates be developed for TB?” In: *Tuberculosis (Edinb)* 94.6, pp. 715–6.

Eklund, L., J. Kangas, and P. Saharinen (2017). “Angiopoietin-Tie signalling in the cardiovascular and lymphatic systems”. In: *Clin Sci (Lond)* 131.1, pp. 87–103.

El-Brolosy, M. A., Z. Kontarakis, A. Rossi, C. Kuenne, S. Gunther, N. Fukuda, K. Kikhi, G. L. M. Boezio, C. M. Takacs, S. L. Lai, R. Fukuda, C. Gerri, A. J. Giraldez, and D. Y. R. Stainier (2019). “Genetic compensation triggered by mutant mRNA degradation”. In: *Nature* 568.7751, pp. 193–197.

– Organisms are naturally robust to genetic perturbation and, as Didier Stainier has beautifully outlined over the past few years, one of the ways that they are able to tolerate loss of function mutations is through compensatory expression of genes related to those lost by mutation. This work found that nonsense-mediated mRNA decay of the mutated allele drive an RNA-dependent upregulation of transcription, although the precise mechanism remains unknown.

El-Brolosy, M. A. and D. Y. R. Stainier (2017). “Genetic compensation: A phenomenon in search of mechanisms”. In: *PLoS Genet* 13.7, e1006780.

El-Etr, S. H., L. Yan, and J. D. Cirillo (2001). “Fish monocytes as a model for mycobacterial host-pathogen interactions”. In: *Infect Immun* 69.12, pp. 7310–7.

El Kasmi, K. C., J. E. Qualls, J. T. Pesce, A. M. Smith, R. W. Thompson, M. Henao-Tamayo, R. J. Basaraba, T. Konig, U. Schleicher, M. S. Koo, G. Kaplan, K. A. Fitzgerald, E. I. Tuomanen, I. M. Orme, T. D. Kanneganti, C. Bogdan, T. A. Wynn, and P. J. Murray (2008). “Toll-like receptor-induced arginase 1 in macrophages thwarts effective immunity against intracellular pathogens”. In: *Nat Immunol* 9.12, pp. 1399–406.

Elbein, A. D., Y. T. Pan, I. Pastuszak, and D. Carroll (2003). “New insights on trehalose: a multifunctional molecule”. In: *Glycobiology* 13.4, 17R–27R.

Elder, M. J., S. J. Webster, R. Chee, D. L. Williams, J. S. Hill Gaston, and J. C. Goodall (2017). “beta-Glucan Size Controls Dectin-1-Mediated Immune Responses in Human Dendritic Cells by Regulating IL-1beta Production”. In: *Front Immunol* 8, p. 791.

Elder, M. J., S. J. Webster, T. J. Fitzmaurice, A. S. D. Shaunak, M. Steinmetz, R. Chee, Z. Mallat, E. S. Cohen, D. L. Williams, J. S. H. Gaston, and J. C. Goodall (2019). “Dendritic Cell-Derived TSLP Negatively Regulates HIF-1alpha and IL-1beta During Dectin-1 Signaling”. In: *Front Immunol* 10, p. 921.

Elks, P. M., S. Brizee, M. van der Vaart, S. R. Walmsley, F. J. van Eeden, S. A. Renshaw, and A. H. Meijer (2013). “Hypoxia inducible factor signaling modulates susceptibility to mycobacterial infection via a nitric oxide dependent mechanism”. In: *PLoS Pathog* 9.12, e1003789.

Elks, P. M., F. J. van Eeden, G. Dixon, X. Wang, C. C. Reyes-Aldasoro, P. W. Ingham, M. K. Whyte, S. R. Walmsley, and S. A. Renshaw (2011). “Activation of hypoxia-inducible factor-1alpha (Hif-1alpha) delays inflammation resolution by reducing neutrophil apoptosis and reverse migration in a zebrafish inflammation model”. In: *Blood* 118.3, pp. 712–22.

Elks, P. M., S. A. Renshaw, A. H. Meijer, S. R. Walmsley, and F. J. van Eeden (2015). “Exploring the HIFs, buts and maybes of hypoxia signalling in disease: lessons from zebrafish models”. In: *Dis Model Mech* 8.11, pp. 1349–60.

- Ellertsdottir, E., A. Lenard, Y. Blum, A. Krudewig, L. Herwig, M. Affolter, and H. G. Belting (2010). “Vascular morphogenesis in the zebrafish embryo”. In: *Dev Biol* 341.1, pp. 56–65.
- Ellis, D. (1995). “Clinical use of tacrolimus (FK-506) in infants and children with renal transplants”. In: *Pediatr Nephrol* 9.4, pp. 487–94.
- Elloumi, H. Z., N. Maharshak, K. N. Rao, T. Kobayashi, H. S. Ryu, M. Muhlbauer, F. Li, C. Jobin, and S. E. Plevy (2012). “A cell permeable peptide inhibitor of NFAT inhibits macrophage cytokine expression and ameliorates experimental colitis”. In: *PLoS One* 7.3, e34172.
- Elshabrawy, H. A., Z. Chen, M. V. Volin, S. Ravella, S. Virupannavar, and S. Shahrara (2015). “The pathogenic role of angiogenesis in rheumatoid arthritis”. In: *Angiogenesis* 18.4, pp. 433–48.
- Emal, D., E. Rampanelli, N. Claessen, F. J. Bemelman, J. C. Leemans, S. Florquin, and M. C. Dessing (2019). “Calcineurin inhibitor tacrolimus impairs host immune response against urinary tract infection”. In: *Sci Rep* 9.1, p. 106.
- Engeszer, R. E., L. B. Patterson, A. A. Rao, and D. M. Parichy (2007). “Zebrafish in the wild: a review of natural history and new notes from the field”. In: *Zebrafish* 4.1, pp. 21–40.
- Eom, D. S. and D. M. Parichy (2017). “A macrophage relay for long-distance signaling during postembryonic tissue remodeling”. In: *Science* 355.6331, pp. 1317–1320.
- Epelman, S., K. J. Lavine, and G. J. Randolph (2014). “Origin and functions of tissue macrophages”. In: *Immunity* 41.1, pp. 21–35.
- Ernst, J. D. (1998). “Macrophage receptors for *Mycobacterium tuberculosis*”. In: *Infect Immun* 66.4, pp. 1277–81.
- (2012). “The immunological life cycle of tuberculosis”. In: *Nat Rev Immunol* 12.8, pp. 581–91.
- Escoll, P. and C. Buchrieser (2019). “Metabolic reprogramming: an innate cellular defence mechanism against intracellular bacteria?” In: *Curr Opin Immunol* 60, pp. 117–123.

- Escoll, P., M. Rolando, L. Gomez-Valero, and C. Buchrieser (2013). “From amoeba to macrophages: exploring the molecular mechanisms of *Legionella pneumophila* infection in both hosts”. In: *Curr Top Microbiol Immunol* 376, pp. 1–34.
- Esher, S. K., K. S. Ost, M. A. Kohlbrenner, K. M. Pianalto, C. L. Telzrow, A. Campuzano, C. B. Nichols, C. Munro, Jr. Wormley F. L., and J. A. Alspaugh (2018). “Defects in intracellular trafficking of fungal cell wall synthases lead to aberrant host immune recognition”. In: *PLoS Pathog* 14.6, e1007126.
- Espenschied, S. T., M. R. Cronan, M. A. Matty, O. Mueller, M. R. Redinbo, D. M. Tobin, and J. F. Rawls (2019). “Epithelial delamination is protective during pharmaceutical-induced enteropathy”. In: *Proc Natl Acad Sci U S A* 116.34, pp. 16961–16970.
- Evans, M. R. and M. H. Wansbrough-Jones (1996). “Alveolar macrophage activation in HIV infection”. In: *J Infect* 33.2, pp. 91–4.
- Ewart, K. V. and G. L. Fletcher (1993). “Herring antifreeze protein: primary structure and evidence for a C-type lectin evolutionary origin”. In: *Mol Mar Biol Biotechnol* 2.1, pp. 20–7.
- Ewart, K. V., Z. Li, D. S. Yang, G. L. Fletcher, and C. L. Hew (1998). “The ice-binding site of Atlantic herring antifreeze protein corresponds to the carbohydrate-binding site of C-type lectins”. In: *Biochemistry* 37.12, pp. 4080–5.
- Falkinham III, J. O. (2009). “The biology of environmental mycobacteria”. In: *Environ Microbiol Rep* 1.6, pp. 477–87.
- Falkow, S. (1988). “Molecular Koch’s postulates applied to microbial pathogenicity”. In: *Rev Infect Dis* 10 Suppl 2, S274–6.
- (2004). “Molecular Koch’s postulates applied to bacterial pathogenicity—a personal recollection 15 years later”. In: *Nat Rev Microbiol* 2.1, pp. 67–72.
- Famelis, N., A. Rivera-Calzada, G. Degliesposti, M. Wingender, N. Mietrach, J. M. Skehel, R. Fernandez-Leiro, B. Bottcher, A. Schlosser, O. Llorca, and S. Geibel (2019). “Architecture of the mycobacterial type VII secretion system”. In: *Nature* 576.7786, pp. 321–325.

Fan, Jingwen, Victoria L. Hale, Lindsey T Lelieveld, Laura J. Whitworth, Elisabeth M. Busch-Nentwich, Mark Troll, Paul H. Edelstein, Timothy M. Cox, Francisco J. Roca, Johannes M.F.G. Aerts, and Lalita Ramakrishnan (2022). “Gaucher Disease Protects Against Tuberculosis”. In: *bioRxiv*.

Fan, Y., P. Xie, T. Zhang, H. Zhang, D. Gu, M. She, and H. Li (2008). “Regulation of the stability and transcriptional activity of NFATc4 by ubiquitination”. In: *FEBS Lett* 582.29, pp. 4008–14.

Fantin, A., J. M. Vieira, G. Gestri, L. Denti, Q. Schwarz, S. Prykhozhij, F. Peri, S. W. Wilson, and C. Ruhrberg (2010). “Tissue macrophages act as cellular chaperones for vascular anastomosis downstream of VEGF-mediated endothelial tip cell induction”. In: *Blood* 116.5, pp. 829–40.

Fazio, M., J. Ablain, Y. Chuan, D. M. Langenau, and L. I. Zon (2020). “Zebrafish patient avatars in cancer biology and precision cancer therapy”. In: *Nat Rev Cancer* 20.5, pp. 263–273.

Fearnley, G. W., G. A. Smith, I. Abdul-Zani, N. Yuldasheva, N. A. Mughal, S. Homer-Vanniasinkam, M. T. Kearney, I. C. Zachary, D. C. Tomlinson, M. A. Harrison, S. B. Wheatcroft, and S. Ponnambalam (2016). “VEGF-A isoforms program differential VEGFR2 signal transduction, trafficking and proteolysis”. In: *Biol Open* 5.5, pp. 571–83.

Feinberg, H., S. A. Jegouzo, T. J. Rowntree, Y. Guan, M. A. Brash, M. E. Taylor, W. I. Weis, and K. Drickamer (2013). “Mechanism for recognition of an unusual mycobacterial glycolipid by the macrophage receptor mincle”. In: *J Biol Chem* 288.40, pp. 28457–65.

Feinberg, H., N. D. Rambaruth, S. A. Jegouzo, K. M. Jacobsen, R. Djurhuus, T. B. Poulsen, W. I. Weis, M. E. Taylor, and K. Drickamer (2016). “Binding Sites for Acylated Trehalose Analogs of Glycolipid Ligands on an Extended Carbohydrate Recognition Domain of the Macrophage Receptor Minicle”. In: *J Biol Chem* 291.40, pp. 21222–21233.

Feldmann, M. (2002). “Development of anti-TNF therapy for rheumatoid arthritis”. In: *Nat Rev Immunol* 2.5, pp. 364–71.

Feng, Y., A. Dorhoi, H. J. Mollenkopf, H. Yin, Z. Dong, L. Mao, J. Zhou, A. Bi, S. Weber, J. Maertzdorf, G. Chen, Y. Chen, and S. H. Kaufmann (2014). “Platelets direct monocyte differentiation into epithelioid-like multinucleated giant foam cells

with suppressive capacity upon mycobacterial stimulation". In: *J Infect Dis* 210.11, pp. 1700–10.

Fenton, M. J., M. W. Vermeulen, S. Kim, M. Burdick, R. M. Strieter, and H. Kornfeld (1997). "Induction of gamma interferon production in human alveolar macrophages by *Mycobacterium tuberculosis*". In: *Infect Immun* 65.12, pp. 5149–56.

Ferrara, N., K. J. Hillan, H. P. Gerber, and W. Novotny (2004). "Discovery and development of bevacizumab, an anti-VEGF antibody for treating cancer". In: *Nat Rev Drug Discov* 3.5, pp. 391–400.

– This time capsule on the development of bevacizumab by the discoverer of VEGFA is a worthy read for the sake of understanding the basic and translational understanding of VEGFA-targeting therapy in 2004. The promise of bevacizumab was remarkable and the timeline of only 15 years from basic discovery to FDA-approved treatment is a remarkable testament to the utility of basic science in improving human health and is, perhaps, the most remarkable story of its kind in all of medicine.

Ferrari, G., B. D. Cook, V. Terushkin, G. Pintucci, and P. Mignatti (2009). "Transforming growth factor-beta 1 (TGF-beta1) induces angiogenesis through vascular endothelial growth factor (VEGF)-mediated apoptosis". In: *J Cell Physiol* 219.2, pp. 449–58.

Ferrero, G., E. Gomez, S. Lyer, M. Rovira, M. Misericocchi, D. M. Langenau, J. Y. Bertrand, and V. Wittamer (2020). "The macrophage-expressed gene (mpeg) 1 identifies a subpopulation of B cells in the adult zebrafish". In: *J Leukoc Biol* 107.3, pp. 431–443.

Feyaerts, D., C. Urbschat, B. Gaudilliere, and I. A. Stelzer (2022). "Establishment of tissue-resident immune populations in the fetus". In: *Semin Immunopathol*.

Fiedler, U., M. Scharpfenecker, S. Koidl, A. Hegen, V. Grunow, J. M. Schmidt, W. Kriz, G. Thurston, and H. G. Augustin (2004). "The Tie-2 ligand angiopoietin-2 is stored in and rapidly released upon stimulation from endothelial cell Weibel-Palade bodies". In: *Blood* 103.11, pp. 4150–6.

Fiege, J. K., I. A. Stone, R. E. Dumm, B. M. Waring, B. T. Fife, J. Agudo, B. D. Brown, N. S. Heaton, and R. A. Langlois (2019). "Long-term surviving influenza infected cells evade CD8+ T cell mediated clearance". In: *PLoS Pathog* 15.9, e1008077.

- Figueiredo, B. C., A. A. Da'dara, S. C. Oliveira, and P. J. Skelly (2015). "Schistosomes Enhance Plasminogen Activation: The Role of Tegumental Enolase". In: *PLoS Pathog* 11.12, e1005335.
- Finco, T. S. and A. S. Baldwin (1995). "Mechanistic aspects of NF-kappa B regulation: the emerging role of phosphorylation and proteolysis". In: *Immunity* 3.3, pp. 263–72.
- Finethy, R., J. Dockterman, M. Kutsch, N. Orench-Rivera, G. D. Wallace, A. S. Piro, S. Luoma, A. K. Haldar, S. Hwang, J. Martinez, M. J. Kuehn, G. A. Taylor, and J. Coers (2020). "Dynamin-related Irgm proteins modulate LPS-induced caspase-11 activation and septic shock". In: *EMBO Rep* 21.11, e50830.
- Finkenzeller, G., A. Technau, and D. Marme (1995). "Hypoxia-induced transcription of the vascular endothelial growth factor gene is independent of functional AP-1 transcription factor". In: *Biochem Biophys Res Commun* 208.1, pp. 432–9.
- Finlay, B. B. and G. McFadden (2006). "Anti-immunology: evasion of the host immune system by bacterial and viral pathogens". In: *Cell* 124.4, pp. 767–82.
- Fisher, W. G., P. C. Yang, R. K. Medikonduri, and M. S. Jafri (2006). "NFAT and NFκB activation in T lymphocytes: a model of differential activation of gene expression". In: *Ann Biomed Eng* 34.11, pp. 1712–28.
- Fishman, J. A. (2007). "Infection in solid-organ transplant recipients". In: *N Engl J Med* 357.25, pp. 2601–14.
- Fleming, Alexander (1941). "Penicillin". In: *BMJ* 2.4210, pp. 386–386.
- Flockhart, R. J., J. L. Armstrong, N. J. Reynolds, and P. E. Lovat (2009). "NFAT signalling is a novel target of oncogenic BRAF in metastatic melanoma". In: *Br J Cancer* 101.8, pp. 1448–55.
- Floden, A. M., J. A. Watt, and C. A. Brissette (2011). "*Borrelia burgdorferi* enolase is a surface-exposed plasminogen binding protein". In: *PLoS One* 6.11, e27502.
- Florio, T. J., R. K. Lokareddy, D. P. Yeggoni, R. S. Sankhala, C. A. Ott, R. E. Gillilan, and G. Cingolani (2022). "Differential recognition of canonical NF-κB dimers by Importin α3". In: *Nat Commun* 13.1, p. 1207.

- Flynn, J. L. and J. Chan (2001). "Immunology of tuberculosis". In: *Annu Rev Immunol* 19, pp. 93–129.
- (2005). "What's good for the host is good for the bug". In: *Trends Microbiol* 13.3, pp. 98–102.
- Flynn, J. L., J. Chan, and P. L. Lin (2011). "Macrophages and control of granulomatous inflammation in tuberculosis". In: *Mucosal Immunol* 4.3, pp. 271–8.
- Flynn, J. L., J. Chan, K. J. Triebold, D. K. Dalton, T. A. Stewart, and B. R. Bloom (1993).** "An essential role for interferon gamma in resistance to *Mycobacterium tuberculosis* infection". In: *J Exp Med* 178.6, pp. 2249–54.
- Flynn, J. L., M. M. Goldstein, J. Chan, K. J. Triebold, K. Pfeffer, C. J. Lowenstein, R. Schreiber, T. W. Mak, and B. R. Bloom (1995).** "Tumor necrosis factor-alpha is required in the protective immune response against *Mycobacterium tuberculosis* in mice". In: *Immunity* 2.6, pp. 561–72.
- Forkner, C. E. (1930). "The Origin and Fate of Two Types of Multi-Nucleated Giant Cells in the Circulating Blood". In: *J Exp Med* 52.2, pp. 279–97.
- Fortune, S. M. and E. J. Rubin (2007). "The complex relationship between mycobacteria and macrophages: it's not all bliss". In: *Cell Host Microbe* 2.1, pp. 5–6.
- Fortune, S. M., A. Solache, A. Jaeger, P. J. Hill, J. T. Belisle, B. R. Bloom, E. J. Rubin, and J. D. Ernst (2004). "*Mycobacterium tuberculosis* inhibits macrophage responses to IFN-gamma through myeloid differentiation factor 88-dependent and -independent mechanisms". In: *J Immunol* 172.10, pp. 6272–80.
- Fougere, M., B. Gaudineau, J. Barbier, F. Guaddachi, J. P. Feugeas, D. Auboeuf, and S. Jauliac (2010). "NFAT3 transcription factor inhibits breast cancer cell motility by targeting the Lipocalin 2 gene". In: *Oncogene* 29.15, pp. 2292–301.
- Franchi, L., N. Warner, K. Viani, and G. Nunez (2009). "Function of Nod-like receptors in microbial recognition and host defense". In: *Immunol Rev* 227.1, pp. 106–28.

Franks, M. E., G. R. Macpherson, and W. D. Figg (2004). "Thalidomide". In: *Lancet* 363.9423, pp. 1802–11.

Fremond, C. M., D. Togbe, E. Doz, S. Rose, V. Vasseur, I. Maillet, M. Jacobs, B. Ryffel, and V. F. Quesniaux (2007). "IL-1 receptor-mediated signal is an essential component of MyD88-dependent innate response to *Mycobacterium tuberculosis* infection". In: *J Immunol* 179.2, pp. 1178–89.

Fremond, C. M., V. Yeremeev, D. M. Nicolle, M. Jacobs, V. F. Quesniaux, and B. Ryffel (2004). "Fatal *Mycobacterium tuberculosis* infection despite adaptive immune response in the absence of MyD88". In: *J Clin Invest* 114.12, pp. 1790–9.

Frentzas, S., E. Simoneau, V. L. Bridgeman, P. B. Vermeulen, S. Foo, E. Kostaras, M. Nathan, A. Wotherspoon, Z. H. Gao, Y. Shi, G. Van den Eynden, F. Daley, C. Peckitt, X. Tan, A. Salman, A. Lazaris, P. Gazinska, T. J. Berg, Z. Eltahir, L. Ritsma, J. Van Rheenen, A. Khashper, G. Brown, H. Nystrom, M. Sund, S. Van Laere, E. Loyer, L. Dirix, D. Cunningham, P. Metrakos, and A. R. Reynolds (2016). "Vessel co-option mediates resistance to anti-angiogenic therapy in liver metastases". In: *Nat Med* 22.11, pp. 1294–1302.

Freudenberg, M. A., S. Tchaptchet, S. Keck, G. Fejer, M. Huber, N. Schutze, B. Beutler, and C. Galanos (2008). "Lipopolysaccharide sensing an important factor in the innate immune response to Gram-negative bacterial infections: benefits and hazards of LPS hypersensitivity". In: *Immunobiology* 213.3-4, pp. 193–203.

Fric, J., C. X. Lim, E. G. Koh, B. Hofmann, J. Chen, H. S. Tay, S. A. Mohammad Isa, A. Mortellaro, C. Ruedl, and P. Ricciardi-Castagnoli (2012a). "Calcineurin/NFAT signalling inhibits myeloid haematopoiesis". In: *EMBO Mol Med* 4.4, pp. 269–82.

Fric, J., T. Zelante, A. Y. Wong, A. Mertes, H. B. Yu, and P. Ricciardi-Castagnoli (2012b). "NFAT control of innate immunity". In: *Blood* 120.7, pp. 1380–9.

– A phenomenal review of the diverse roles of NFAT in innate immunity up to that time. This was foundational for me starting out in this project by providing an overview of the known roles of this pathway in determining diverse immune responses, specifically those downstream of C-type lectin responses. Essential reading.

Fric, Jan, Teresa Zelante, and P. Ricciardi-Castagnoli (2014). "Phagocytosis of particulate antigens - all roads lead to calcineurin/NFAT signaling pathway". In: *Front Immunol* 4.513.

- Friedrich, D., R. A. Fecher, J. Rupp, and Jr. Deepe G. S. (2017). “Impact of HIF-1alpha and hypoxia on fungal growth characteristics and fungal immunity”. In: *Microbes Infect* 19.3, pp. 204–209.
- Friedrich, D., D. Zapf, B. Lohse, R. A. Fecher, Jr. Deepe G. S., and J. Rupp (2019). “The HIF-1alpha/LC3-II Axis Impacts Fungal Immunity in Human Macrophages”. In: *Infect Immun* 87.7.
- Frith, John A. (2014). “History of tuberculosis. Part 1 - phthisis, consumption and the white plague”. In: *Journal of Military and Veterans' Health* 22, pp. 29–35.
- Frutos, S. de, J. M. Diaz, C. H. Nitta, M. L. Sherpa, and L. V. Bosc (2011). “Endothelin-1 contributes to increased NFATc3 activation by chronic hypoxia in pulmonary arteries”. In: *Am J Physiol Cell Physiol* 301.2, pp. C441–50.
- Fu, C. L., J. I. Odegaard, D. R. Herbert, and M. H. Hsieh (2012). “A novel mouse model of *Schistosoma haematobium* egg-induced immunopathology”. In: *PLoS Pathog* 8.3, e1002605.
- Fu, M. S., L. C. Liporagi-Lopes, S. R. Junior Dos Santos, J. L. Tenor, J. R. Perfect, C. A. Cuomo, and A. Casadevall (2021). “Amoeba Predation of *Cryptococcus neoformans* Results in Pleiotropic Changes to Traits Associated with Virulence”. In: *mBio* 12.2.
- Fujita, Keiko and Masumi Akita (2017). “Tumor Angiogenesis: A Focus on the Role of Cancer Stem Cells”. In: IntechOpen. Chap. 12.
- Fujita, Y., T. Doi, K. Sato, and I. Yano (2005). “Diverse humoral immune responses and changes in IgG antibody levels against mycobacterial lipid antigens in active tuberculosis”. In: *Microbiology (Reading)* 151.Pt 6, pp. 2065–2074.
- Fujiwara, N., S. Oka, M. Ide, K. Kashima, T. Honda, and I. Yano (1999). “Production and partial characterization of antibody to cord factor (trehalose 6,6'-dimycolate) in mice”. In: *Microbiol Immunol* 43.8, pp. 785–93.
- Fuller, G. L., J. A. Williams, M. G. Tomlinson, J. A. Eble, S. L. Hanna, S. Pohlmann, K. Suzuki-Inoue, Y. Ozaki, S. P. Watson, and A. C. Pearce (2007). “The C-type lectin receptors CLEC-2 and Dectin-1, but not DC-SIGN, signal via a novel YXXL-dependent signaling cascade”. In: *J Biol Chem* 282.17, pp. 12397–409.

- Furin, J., E. Lessem, and V. Cox (2017). “Recommending prolonged bedaquiline use for the treatment of highly resistant strains of tuberculosis”. In: *Eur Respir J* 50.5.
- Furukawa, A., J. Kamishikiryo, D. Mori, K. Toyonaga, Y. Okabe, A. Toji, R. Kanda, Y. Miyake, T. Ose, S. Yamasaki, and K. Maenaka (2013). “Structural analysis for glycolipid recognition by the C-type lectins Mincle and MCL”. In: *Proc Natl Acad Sci U S A* 110.43, pp. 17438–43.
- Furutani-Seiki, M., Y. J. Jiang, M. Brand, C. P. Heisenberg, C. Houart, D. Beuchle, F. J. van Eeden, M. Granato, P. Haffter, M. Hammerschmidt, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, and C. Nusslein-Volhard (1996). “Neural degeneration mutants in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 229–39.
- Gagneux, S. (2009). “Fitness cost of drug resistance in *Mycobacterium tuberculosis*”. In: *Clin Microbiol Infect* 15 Suppl 1, pp. 66–8.
- (2018). “Ecology and evolution of *Mycobacterium tuberculosis*”. In: *Nat Rev Microbiol* 16.4, pp. 202–213.
- Gagneux, S., K. DeRiemer, T. Van, M. Kato-Maeda, B. C. de Jong, S. Narayanan, M. Nicol, S. Niemann, K. Kremer, M. C. Gutierrez, M. Hilty, P. C. Hopewell, and P. M. Small (2006a). “Variable host-pathogen compatibility in *Mycobacterium tuberculosis*”. In: *Proc Natl Acad Sci U S A* 103.8, pp. 2869–73.
- Gagneux, S., C. D. Long, P. M. Small, T. Van, G. K. Schoolnik, and B. J. Bohannan (2006b). “The competitive cost of antibiotic resistance in *Mycobacterium tuberculosis*”. In: *Science* 312.5782, pp. 1944–6.
- Galagan, J. E. (2014). “Genomic insights into tuberculosis”. In: *Nat Rev Genet* 15.5, pp. 307–20.
- Galagan, J. E., K. Minch, M. Peterson, A. Lyubetskaya, E. Azizi, L. Sweet, A. Gomes, T. Rustad, G. Dolganov, I. Glotova, T. Abeel, C. Mahwinney, A. D. Kennedy, R. Allard, W. Brabant, A. Krueger, S. Jaini, B. Honda, W. H. Yu, M. J. Hickey, J. Zucker, C. Garay, B. Weiner, P. Sisk, C. Stolte, J. K. Winkler, Y. Van de Peer, P. Iazzetti, D. Camacho, J. Dreyfuss, Y. Liu, A. Dorhoi, H. J. Mollenkopf, P. Drogaris, J. Lamontagne, Y. Zhou, J. Piquenot, S. T. Park, S. Raman, S. H. Kaufmann, R. P. Mohney, D. Chelsky, D. B. Moody, D. R. Sherman, and G. K. Schoolnik (2013). “The *Mycobacterium tuberculosis* regulatory network and hypoxia”. In: *Nature* 499.7457, pp. 178–83.

- Gale, G. L. (1957). "Vascular changes in the lungs in pulmonary tuberculosis". In: *Am Rev Tuberc* 75.3, pp. 410–9.
- Galvan-Pena, S. and L. A. O'Neill (2014). "Metabolic reprogramming in macrophage polarization". In: *Front Immunol* 5, p. 420.
- Ganchua, S. K. C., A. G. White, E. C. Klein, and J. L. Flynn (2020). "Lymph nodes—The neglected battlefield in tuberculosis". In: *PLoS Pathog* 16.8, e1008632.
- Garcia, A. and H. Singh (2013). "Bevacizumab and ovarian cancer". In: *Ther Adv Med Oncol* 5.2, pp. 133–41.
- Garcia-Barros, M., F. Paris, C. Cordon-Cardo, D. Lyden, S. Rafii, A. Haimovitz-Friedman, Z. Fuks, and R. Kolesnick (2003). "Tumor response to radiotherapy regulated by endothelial cell apoptosis". In: *Science* 300.5622, pp. 1155–9.
- Garcia-Vallejo, J. J. and Y. van Kooyk (2009). "Endogenous ligands for C-type lectin receptors: the true regulators of immune homeostasis". In: *Immunol Rev* 230.1, pp. 22–37.
- (2013). "The physiological role of DC-SIGN: a tale of mice and men". In: *Trends Immunol* 34.10, pp. 482–6.
- Gardner, Victor, Chikezie O. Madu, and Yi Lu (2017). "Anti-VEGF Therapy in Cancer: A Double-Edged Sword". In: IntechOpen. Chap. 19.
- Garg, R., D. Tripathi, S. Kant, H. Chandra, R. Bhatnagar, and N. Banerjee (2015). "The conserved hypothetical protein Rv0574c is required for cell wall integrity, stress tolerance, and virulence of *Mycobacterium tuberculosis*". In: *Infect Immun* 83.1, pp. 120–9.
- Garry, D. J. (2016). "Etv2 is a master regulator of hematoendothelial lineages". In: *Trans Am Clin Climatol Assoc* 127, pp. 212–223.
- Gatfield, J. and J. Pieters (2000). "Essential role for cholesterol in entry of mycobacteria into macrophages". In: *Science* 288.5471, pp. 1647–50.
- Gazova, I., L. Lefevre, S. J. Bush, S. Clohisey, E. Arner, M. de Hoon, J. Severin, L. van Duin, R. Andersson, A. Lengeling, D. A. Hume, and K. M. Summers (2020). "The Transcriptional Network That Controls Growth Arrest and Macrophage Dif-

ferentiation in the Human Myeloid Leukemia Cell Line THP-1". In: *Front Cell Dev Biol* 8, p. 498.

Gearing, L. J., H. E. Cumming, R. Chapman, A. M. Finkel, I. B. Woodhouse, K. Luu, J. A. Gould, S. C. Forster, and P. J. Hertzog (2019). "CiiDER: A tool for predicting and analysing transcription factor binding sites". In: *PLoS One* 14.9, e0215495.

Geeraerts, X., J. Fernandez-Garcia, F. J. Hartmann, K. E. de Goede, L. Martens, Y. Elkrim, A. Debraekeleer, B. Stijlemans, A. Vandekeere, G. Rinaldi, R. De Rycke, M. Planque, D. Broekaert, E. Meinstre, E. Clappaert, P. Bardet, A. Murgaski, C. Gysemans, F. A. Nana, Y. Saeys, S. C. Bendall, D. Laoui, J. Van den Bossche, S. M. Fendt, and J. A. Van Ginderachter (2021). "Macrophages are metabolically heterogeneous within the tumor microenvironment". In: *Cell Rep* 37.13, p. 110171.

Geijtenbeek, T. B., A. Engering, and Y. Van Kooyk (2002). "DC-SIGN, a C-type lectin on dendritic cells that unveils many aspects of dendritic cell biology". In: *J Leukoc Biol* 71.6, pp. 921–31.

Geijtenbeek, T. B. and S. I. Gringhuis (2009). "Signalling through C-type lectin receptors: shaping immune responses". In: *Nat Rev Immunol* 9.7, pp. 465–79.

Geijtenbeek, T. B., S. J. Van Vliet, E. A. Koppel, M. Sanchez-Hernandez, C. M. Vandebroucke-Grauls, B. Appelmelk, and Y. Van Kooyk (2003). "Mycobacteria target DC-SIGN to suppress dendritic cell function". In: *J Exp Med* 197.1, pp. 7–17.

*– This first of its kind publication on the ability of *Mycobacterium tuberculosis* to undermine C-type lectin signaling responses, specifically DC-SIGN. Binding of mycobacterial mannose-capped lipoarabinomannan to DC-SIGN blocked dendritic cell maturation and subsequent adaptive immune responses. This provides a model for different ways that mycobacteria can subvert host pattern recognition receptor responses to their own benefit.*

Geisel, R. E., K. Sakamoto, D. G. Russell, and E. R. Rhoades (2005). "*In vivo* activity of released cell wall lipids of *Mycobacterium bovis* Bacillus Calmette-Guerin is due principally to trehalose mycolates". In: *J Immunol* 174.8, pp. 5007–15.

Geissmann, Frederic (2010). "Development of Monocytes, Macrophages, and Dendritic Cells". In: *Science*.

Gengenbacher, M. and S. H. Kaufmann (2012). “*Mycobacterium tuberculosis*: success through dormancy”. In: *FEMS Microbiol Rev* 36.3, pp. 514–32.

Gerber, H. P., A. K. Malik, G. P. Solar, D. Sherman, X. H. Liang, G. Meng, K. Hong, J. C. Marsters, and N. Ferrara (2002). “VEGF regulates haematopoietic stem cell survival by an internal autocrine loop mechanism”. In: *Nature* 417.6892, pp. 954–8.

German, N., D. Doyscher, and C. Rensing (2013). “Bacterial killing in macrophages and amoeba: do they all use a brass dagger?” In: *Future Microbiol* 8.10, pp. 1257–64.

Gerri, C., R. Marin-Juez, M. Marass, A. Marks, H. M. Maischein, and D. Y. R. Stainier (2017). “Hif-1alpha regulates macrophage-endothelial interactions during blood vessel development in zebrafish”. In: *Nat Commun* 8, p. 15492.

— *Loss of function in HIF-1α is known to cause severe vascular defects but the underlying cause of this angiogenic failure was unknown. It turns out that many of these effects are mediated by HIF-1α activity within macrophages, where it physically mediates the growth of angiogenic vessels and facilitates vessel repair after injury. This adds a critical dimension to our understanding of the role of macrophages in angiogenesis and developed a useful toolset for further study. This nicely pairs with the findings from Britto, Wyroba, Chen, Lockwood, Tran, Shepherd, Hall, Crosier, Crosier, and Astin (2018) and Liu, Wu, Yang, Gao, Li, Yang, and Luo (2016), which identified similar macrophage-endothelial interactions during angiogenesis.*

Getahun, A. and J. C. Cambier (2015). “Of ITIMs, ITAMs, and ITAMIs: revisiting immunoglobulin Fc receptor signaling”. In: *Immunol Rev* 268.1, pp. 66–73.

Ghodbane, R., F. M. Medie, H. Lepidi, C. Nappez, and M. Drancourt (2014). “Long-term survival of tuberculosis complex mycobacteria in soil”. In: *Microbiology (Reading)* 160.Pt 3, pp. 496–501.

Ghosh, G., V. Y. Wang, D. B. Huang, and A. Fusco (2012). “NF-kappaB regulation: lessons from structures”. In: *Immunol Rev* 246.1, pp. 36–58.

Ghotra, V. P., S. He, G. van der Horst, S. Nijhoff, H. de Bont, A. Lekkerkerker, R. Janssen, G. Jenster, G. J. van Leenders, A. M. Hoogland, E. I. Verhoef, Z. Baranski, J. Xiong, B. van de Water, G. van der Pluijm, B. E. Snaar-Jagalska,

and E. H. Danen (2015). “SYK is a candidate kinase target for the treatment of advanced prostate cancer”. In: *Cancer Res* 75.1, pp. 230–40.

Gideon, H. P., T. K. Hughes, C. N. Tzouanas, 2nd Wadsworth M. H., A. A. Tu, T. M. Gierahn, J. M. Peters, F. F. Hopkins, J. R. Wei, C. Kummerlowe, N. L. Grant, K. Nargan, J. Y. Phuah, H. J. Borish, P. Maiello, A. G. White, C. G. Winchell, S. K. Nyquist, S. K. C. Ganchua, A. Myers, K. V. Patel, C. L. Ameel, C. T. Cochran, S. Ibrahim, J. A. Tomko, L. J. Frye, J. M. Rosenberg, A. Shih, M. Chao, E. Klein, C. A. Scanga, J. Ordovas-Montanes, B. Berger, J. T. Mattila, R. Madansein, J. C. Love, P. L. Lin, A. Leslie, S. M. Behar, B. Bryson, J. L. Flynn, S. M. Fortune, and A. K. Shalek (2022). “Multimodal profiling of lung granulomas in macaques reveals cellular correlates of tuberculosis control”. In: *Immunity* 55.5, 827–846 e10.

Gilbert, M. R., J. J. Dignam, T. S. Armstrong, J. S. Wefel, D. T. Blumenthal, M. A. Vogelbaum, H. Colman, A. Chakravarti, S. Pugh, M. Won, R. Jeraj, P. D. Brown, K. A. Jaeckle, D. Schiff, V. W. Stieber, D. G. Brachman, M. Werner-Wasik, I. W. Tremont-Lukats, E. P. Sulman, K. D. Aldape, Jr. Curran W. J., and M. P. Mehta (2014). “A randomized trial of bevacizumab for newly diagnosed glioblastoma”. In: *N Engl J Med* 370.8, pp. 699–708.

Giles, R.H. (1802–1881). *A girl reads to a convalescent while a nurse brings in the patient's medicine*.

Gilleron, M., J. Nigou, D. Nicolle, V. Quesniaux, and G. Puzo (2006). “The acylation state of mycobacterial lipomannans modulates innate immunity response through toll-like receptor 2”. In: *Chem Biol* 13.1, pp. 39–47.

Gilmore, S. A., M. W. Schelle, C. M. Holsclaw, C. D. Leigh, M. Jain, J. S. Cox, J. A. Leary, and C. R. Bertozzi (2012). “Sulfolipid-1 biosynthesis restricts *Mycobacterium tuberculosis* growth in human macrophages”. In: *ACS Chem Biol* 7.5, pp. 863–70.

Gilmore, T. D. (2006). “Introduction to NF-kappaB: players, pathways, perspectives”. In: *Oncogene* 25.51, pp. 6680–4.

Ginhoux, F. and M. Guilliams (2016). “Tissue-Resident Macrophage Ontogeny and Homeostasis”. In: *Immunity* 44.3, pp. 439–449.

Ginhoux, F. and S. Jung (2014). “Monocytes and macrophages: developmental pathways and tissue homeostasis”. In: *Nat Rev Immunol* 14.6, pp. 392–404.

Girish, V. and A. Vijayalakshmi (2004). “Affordable image analysis using NIH Image/ImageJ”. In: *Indian J Cancer* 41.1, p. 47.

Glasauer, S. M. and S. C. Neuhauss (2014). “Whole-genome duplication in teleost fishes and its evolutionary consequences”. In: *Mol Genet Genomics* 289.6, pp. 1045–60.

Gleeson, L. E., F. J. Sheedy, E. M. Palsson-McDermott, D. Triglia, S. M. O’Leary, M. P. O’Sullivan, L. A. O’Neill, and J. Keane (2016). “Cutting Edge: *Mycobacterium tuberculosis* Induces Aerobic Glycolysis in Human Alveolar Macrophages That Is Required for Control of Intracellular Bacillary Replication”. In: *J Immunol* 196.6, pp. 2444–9.

Glickman, M. S. (2003). “The mmaA2 gene of *Mycobacterium tuberculosis* encodes the distal cyclopropane synthase of the alpha-mycolic acid”. In: *J Biol Chem* 278.10, pp. 7844–9.

Glickman, M. S., S. M. Cahill, and Jr. Jacobs W. R. (2001). “The *Mycobacterium tuberculosis* cmaA2 gene encodes a mycolic acid trans-cyclopropane synthetase”. In: *J Biol Chem* 276.3, pp. 2228–33.

Glickman, M. S., J. S. Cox, and W. R. Jacobs (2000). “A novel mycolic acid cyclopropane synthetase is required for cording, persistence, and virulence of *Mycobacterium tuberculosis*”. In: *Molecular Cell* 5.4, pp. 717–727.

– This is the first published description of pcaA and set the stage for all of the work to come on the role of cyclopropane modification of TDM in modulating host immune responses.

Glickman, M. S. and Jr. Jacobs W. R. (2001). “Microbial pathogenesis of *Mycobacterium tuberculosis*: dawn of a discipline”. In: *Cell* 104.4, pp. 477–85.

Glickman, M. S. and N. Schluger (2016). “Adding Insult to Injury: Exacerbating TB Risk with Smoking”. In: *Cell Host Microbe* 19.4, pp. 432–3.

Glickman, Michael S. (2008). “Cording, Cord Factors, and Trehalose Dimycolate”. In: John Wiley & Sons, Ltd. Chap. 5, pp. 63–73.

- Go, W. Y., X. Liu, M. A. Roti, F. Liu, and S. N. Ho (2004). “NFAT5/TonEBP mutant mice define osmotic stress as a critical feature of the lymphoid microenvironment”. In: *Proc Natl Acad Sci U S A* 101.29, pp. 10673–8.
- Goel, H. L. and A. M. Mercurio (2013). “VEGF targets the tumour cell”. In: *Nat Rev Cancer* 13.12, pp. 871–82.
- Goel, S., A. H. Wong, and R. K. Jain (2012). “Vascular normalization as a therapeutic strategy for malignant and nonmalignant disease”. In: *Cold Spring Harb Perspect Med* 2.3, a006486.
- Goetz, Thomas (2014). *The Remedy: Robert Koch, Arthur Conan Doyle, and the Quest to Cure Tuberculosis*. New York, New York: Gotham Books.
- Goldman, A. L. and S. S. Braman (1972). “Isoniazid: a review with emphasis on adverse effects”. In: *Chest* 62.1, pp. 71–7.
- Gomes, M. C. and S. Mostowy (2020). “The Case for Modeling Human Infection in Zebrafish”. In: *Trends Microbiol* 28.1, pp. 10–18.
- Gomez-Abenza, E., S. Ibanez-Molero, D. Garcia-Moreno, I. Fuentes, L. I. Zon, M. C. Mione, M. L. Cayuela, C. Gabellini, and V. Mulero (2019). “Zebrafish modeling reveals that SPINT1 regulates the aggressiveness of skin cutaneous melanoma and its crosstalk with tumor immune microenvironment”. In: *J Exp Clin Cancer Res* 38.1, p. 405.
- Gong, L., M. Xu, W. Xu, W. Tang, J. Lu, W. Jiang, F. Xie, L. Ding, and X. Qian (2021). “Efficacy and safety of tacrolimus monotherapy versus tacrolimus-corticosteroid combination therapy for idiopathic membranous nephropathy: A meta-analysis”. In: *Medicine (Baltimore)* 100.51, e28225.
- Goodridge, H. S., C. N. Reyes, C. A. Becker, T. R. Katsumoto, J. Ma, A. J. Wolf, N. Bose, A. S. Chan, A. S. Magee, M. E. Danielson, A. Weiss, J. P. Vasilakos, and D. M. Underhill (2011). “Activation of the innate immune receptor Dectin-1 upon formation of a ‘phagocytic synapse’”. In: *Nature* 472.7344, pp. 471–5.
- Goodridge, H. S., T. Shimada, A. J. Wolf, Y. M. Hsu, C. A. Becker, X. Lin, and D. M. Underhill (2009). “Differential use of CARD9 by dectin-1 in macrophages and dendritic cells”. In: *J Immunol* 182.2, pp. 1146–54.

Goodridge, H. S., R. M. Simmons, and D. M. Underhill (2007). “Dectin-1 stimulation by *Candida albicans* yeast or zymosan triggers NFAT activation in macrophages and dendritic cells”. In: *J Immunol* 178.5, pp. 3107–15.

– While *CARD9* was of clear importance in determining responses downstream of C-type lectin receptors, other phenomena could not be explained solely by the loss of *CARD9*, implying the existence of alternative response pathways. This study found a critical role for NFAT signaling in the induction of antifungal responses and defined a limited set of genes that were specifically regulated by NFAT in myeloid cells, allowing for future studies to expand on this set and to more comprehensively define the role of NFAT in mediating C-type lectin receptor responses.

Goodridge, Helen S. and David M. Underhill (2008). “Fungal Recognition by TLR2 and Dectin-1”. In: ed. by Stefan Bauer and Gunther Hartmann. Berlin, Heidelberg: Springer Berlin Heidelberg, pp. 87–109.

Gopal, P., M. Yee, J. Sarathy, J. L. Low, J. P. Sarathy, F. Kaya, V. Dartois, M. Gengenbacher, and T. Dick (2016). “Pyrazinamide Resistance Is Caused by Two Distinct Mechanisms: Prevention of Coenzyme A Depletion and Loss of Virulence Factor Synthesis”. In: *ACS Infect Dis* 2.9, pp. 616–626.

Gordon, M. S. and D. Cunningham (2005). “Managing patients treated with bevacizumab combination therapy”. In: *Oncology* 69 Suppl 3, pp. 25–33.

Gordon, S. and A. Pluddemann (2017). “Tissue macrophages: heterogeneity and functions”. In: *BMC Biol* 15.1, p. 53.

Gordon, S. and P. R. Taylor (2005). “Monocyte and macrophage heterogeneity”. In: *Nat Rev Immunol* 5.12, pp. 953–64.

Gore, A. V., K. Monzo, Y. R. Cha, W. Pan, and B. M. Weinstein (2012). “Vascular development in the zebrafish”. In: *Cold Spring Harb Perspect Med* 2.5, a006684.

Gosselin, D., V. M. Link, C. E. Romanoski, G. J. Fonseca, D. Z. Eichenfield, N. J. Spann, J. D. Stender, H. B. Chun, H. Garner, F. Geissmann, and C. K. Glass (2014). “Environment drives selection and function of enhancers controlling tissue-specific macrophage identities”. In: *Cell* 159.6, pp. 1327–40.

Gosset, N., A. A. Bankier, and R. L. Eisenberg (2009). “Tree-in-bud pattern”. In: *AJR Am J Roentgenol* 193.6, W472–7.

- Gotham, D., L. McKenna, M. Frick, and E. Lessem (2020). “Public investments in the clinical development of bedaquiline”. In: *PLoS One* 15.9, e0239118.
- Gough, P. and I. A. Myles (2020). “Tumor Necrosis Factor Receptors: Pleiotropic Signaling Complexes and Their Differential Effects”. In: *Front Immunol* 11, p. 585880.
- Goumans, M. J., Z. Liu, and P. ten Dijke (2009). “TGF-beta signaling in vascular biology and dysfunction”. In: *Cell Res* 19.1, pp. 116–27.
- Goyal, S., T. E. Klassert, and H. Slevogt (2016). “C-type lectin receptors in tuberculosis: what we know”. In: *Med Microbiol Immunol* 205.6, pp. 513–535.
- Grabher, C., J. S. Joly, and J. Wittbrodt (2004). “Highly efficient zebrafish transgenesis mediated by the meganuclease I-SceI”. In: *Methods Cell Biol* 77, pp. 381–401.
- Grace, A. G., A. Mittal, S. Jain, J. P. Tripathy, S. Satyanarayana, P. Tharyan, and R. Kirubakaran (2019). “Shortened treatment regimens versus the standard regimen for drug-sensitive pulmonary tuberculosis”. In: *Cochrane Database Syst Rev* 12, p. CD012918.
- Graef, I. A., F. Chen, L. Chen, A. Kuo, and G. R. Crabtree (2001). “Signals transduced by Ca(2+)/calcineurin and NFATc3/c4 pattern the developing vasculature”. In: *Cell* 105.7, pp. 863–75.
- Graef, I. A., P. G. Mermelstein, K. Stankunas, J. R. Neilson, K. Deisseroth, R. W. Tsien, and G. R. Crabtree (1999). “L-type calcium channels and GSK-3 regulate the activity of NF-ATc4 in hippocampal neurons”. In: *Nature* 401.6754, pp. 703–8.
- Graham, F. L., J. Smiley, W. C. Russell, and R. Nairn (1977). “Characteristics of a human cell line transformed by DNA from human adenovirus type 5”. In: *J Gen Virol* 36.1, pp. 59–74.
- Granato, M., F. J. van Eeden, U. Schach, T. Trowe, M. Brand, M. Furutani-Seiki, P. Haffter, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, and C. Nusslein-Volhard (1996). “Genes controlling and mediating locomotion behavior of the zebrafish embryo and larva”. In: *Development* 123, pp. 399–413.

Graney, P. L., S. Ben-Shaul, S. Landau, A. Bajpai, B. Singh, J. Eager, A. Cohen, S. Levenberg, and K. L. Spiller (2020). “Macrophages of diverse phenotypes drive vascularization of engineered tissues”. In: *Sci Adv* 6.18, eaay6391.

Granucci, F., S. Feau, V. Angeli, F. Trottein, and P. Ricciardi-Castagnoli (2003). “Early IL-2 production by mouse dendritic cells is the result of microbial-induced priming”. In: *J Immunol* 170.10, pp. 5075–81.

Granucci, F., C. Vizzardelli, N. Pavelka, S. Feau, M. Persico, E. Virzi, M. Rescigno, G. Moro, and P. Ricciardi-Castagnoli (2001). “Inducible IL-2 production by dendritic cells revealed by global gene expression analysis”. In: *Nat Immunol* 2.9, pp. 882–8.

Gratacap, R. L. and R. T. Wheeler (2014). “Utilization of zebrafish for intravital study of eukaryotic pathogen-host interactions”. In: *Dev Comp Immunol* 46.1, pp. 108–15.

Graversen, J. H., R. H. Lorentsen, C. Jacobsen, S. K. Moestrup, B. W. Sigurskjold, H. C. Thogersen, and M. Etzerodt (1998). “The plasminogen binding site of the C-type lectin tetranectin is located in the carbohydrate recognition domain, and binding is sensitive to both calcium and lysine”. In: *J Biol Chem* 273.44, pp. 29241–6.

Gray, E. E., D. Winship, J. M. Snyder, S. J. Child, A. P. Geballe, and D. B. Stetson (2016). “The AIM2-like Receptors Are Dispensable for the Interferon Response to Intracellular DNA”. In: *Immunity* 45.2, pp. 255–66.

Gray, M. J., J. Zhang, L. M. Ellis, G. L. Semenza, D. B. Evans, S. S. Watowich, and G. E. Gallick (2005). “HIF-1alpha, STAT3, CBP/p300 and Ref-1/APE are components of a transcriptional complex that regulates Src-dependent hypoxia-induced expression of VEGF in pancreatic and prostate carcinomas”. In: *Oncogene* 24.19, pp. 3110–20.

Gray, S. F., R. S. Smith, N. J. Reynolds, and E. W. Williams (1990). “Fish tank granuloma”. In: *BMJ* 300.6731, pp. 1069–70.

Greenberger, S., I. Adini, E. Boscolo, J. B. Mulliken, and J. Bischoff (2010). “Targeting NF-kappaB in infantile hemangioma-derived stem cells reduces VEGF-A expression”. In: *Angiogenesis* 13.4, pp. 327–35.

Greenblatt, M. B., A. Aliprantis, B. Hu, and L. H. Glimcher (2010). “Cal-

cineurin regulates innate antifungal immunity in neutrophils". In: *J Exp Med* 207.5, pp. 923–31.

– This study looked at the role of calcineurin within neutrophils and found that the activity of calcineurin was essential for effective antimicrobial responses downstream of C-type lectin receptors and independently of MYD88. This is a nice study on a relatively underappreciated myeloid cell population and found a similar set of regulated genes to those found by Goodridge, Simmons, and Underhill (2007).

Greenblatt, M. B., S. Y. Ritter, J. Wright, K. Tsang, D. Hu, L. H. Glimcher, and A. O. Aliprantis (2013). "NFATc1 and NFATc2 repress spontaneous osteoarthritis". In: *Proc Natl Acad Sci U S A* 110.49, pp. 19914–9.

Gridelli, C., J. de Castro Carpeno, A. C. Dingemans, F. Griesinger, F. Grossi, C. Langer, Y. Ohe, K. Syrigos, N. Thatcher, A. Das-Gupta, M. Truman, M. Donica, V. Smoljanovic, and J. Bennouna (2018). "Safety and Efficacy of Bevacizumab Plus Standard-of-Care Treatment Beyond Disease Progression in Patients With Advanced Non-Small Cell Lung Cancer: The AvaALL Randomized Clinical Trial". In: *JAMA Oncol* 4.12, e183486.

Gringhuis, S. I., J. den Dunnen, M. Litjens, M. van der Vlist, and T. B. Geijtenbeek (2009). "Carbohydrate-specific signaling through the DC-SIGN signalosome tailors immunity to *Mycobacterium tuberculosis*, HIV-1 and Helicobacter pylori". In: *Nat Immunol* 10.10, pp. 1081–8.

Gross, O., A. Gewies, K. Finger, M. Schafer, T. Sparwasser, C. Peschel, I. Forster, and J. Ruland (2006). "Card9 controls a non-TLR signalling pathway for innate anti-fungal immunity". In: *Nature* 442.7103, pp. 651–6.

Gross, O., C. J. Thomas, G. Guarda, and J. Tschopp (2011). "The inflammasome: an integrated view". In: *Immunol Rev* 243.1, pp. 136–51.

Grunwald, D. J. and J. S. Eisen (2002). "Headwaters of the zebrafish – emergence of a new model vertebrate". In: *Nat Rev Genet* 3.9, pp. 717–24.

Guan, Q., M. Garbati, S. Mfarrej, T. AlMutairi, T. Laval, A. Singh, S. Fagbo, A. Smyth, J. A. Browne, M. A. urRahman, A. Alruwaili, A. Hoosen, C. J. Meehan, C. Nakajima, Y. Suzuki, C. Demangel, A. Bhatt, S. V. Gordon, F. AlAsmari, and A. Pain (2021). "Insights into the ancestry evolution of the *Mycobacterium*

- tuberculosis* complex from analysis of *Mycobacterium riyadhense*". In: *NAR Genom Bioinform* 3.3, lqab070.
- Guan, R. and R. A. Mariuzza (2007). "Peptidoglycan recognition proteins of the innate immune system". In: *Trends Microbiol* 15.3, pp. 127–34.
- Guerrini, V. and M. L. Gennaro (2019). "Foam Cells: One Size Doesn't Fit All". In: *Trends Immunol* 40.12, pp. 1163–1179.
- Guha, M. and N. Mackman (2001). "LPS induction of gene expression in human monocytes". In: *Cell Signal* 13.2, pp. 85–94.
- Guichard, A., B. Cruz-Moreno, B. Aguilar, N. M. van Sorge, J. Kuang, A. A. Kurkciyan, Z. Wang, S. Hang, G. P. Pineton de Chambrun, D. F. McCole, P. Watnick, V. Nizet, and E. Bier (2013). "Cholera toxin disrupts barrier function by inhibiting exocyst-mediated trafficking of host proteins to intestinal cell junctions". In: *Cell Host Microbe* 14.3, pp. 294–305.
- Guimaraes, G. R., P. P. Almeida, L. de Oliveira Santos, L. P. Rodrigues, J. L. de Carvalho, and M. Boroni (2021). "Hallmarks of Aging in Macrophages: Consequences to Skin Inflammaging". In: *Cells* 10.6.
- Guirado, E. and L. S. Schlesinger (2013). "Modeling the *Mycobacterium tuberculosis* Granuloma - the Critical Battlefield in Host Immunity and Disease". In: *Front Immunol* 4, p. 98.
- Guirado, E., L. S. Schlesinger, and G. Kaplan (2013). "Macrophages in tuberculosis: friend or foe". In: *Semin Immunopathol* 35.5, pp. 563–83.
- Gunawan, F., A. Gentile, S. Gauvrit, D. Y. R. Stainier, and A. Bensimon-Brito (2020). "Nfatc1 Promotes Interstitial Cell Formation During Cardiac Valve Development in Zebrafish". In: *Circ Res* 126.8, pp. 968–984.
- Guo, S., J. Li, Z. Huang, T. Yue, J. Zhu, X. Wang, Y. Liu, P. Wang, and S. Chen (2022). "The CBS-H2S axis promotes liver metastasis of colon cancer by upregulating VEGF through AP-1 activation". In: *Br J Cancer* 126.7, pp. 1055–1066.
- Guo, X., H. Xue, Q. Shao, J. Wang, X. Guo, X. Chen, J. Zhang, S. Xu, T. Li, P. Zhang, X. Gao, W. Qiu, Q. Liu, and G. Li (2016). "Hypoxia promotes glioma-

- associated macrophage infiltration via periostin and subsequent M2 polarization by upregulating TGF-beta and M-CSFR”. In: *Oncotarget* 7.49, pp. 80521–80542.
- Gupta, A. K., A. Adamiak, and M. Chow (2002). “Tacrolimus: a review of its use for the management of dermatoses”. In: *J Eur Acad Dermatol Venereol* 16.2, pp. 100–14.
- Gupta, K. and J. Zhang (2005). “Angiogenesis: a curse or cure?” In: *Postgrad Med J* 81.954, pp. 236–42.
- Gupta, M. N., S. Pandey, N. Z. Ehtesham, and S. E. Hasnain (2019). “Medical implications of protein moonlighting”. In: *Indian J Med Res* 149.3, pp. 322–325.
- Gupta, P., G. P. Jadaun, R. Das, U. D. Gupta, K. Srivastava, A. Chauhan, V. D. Sharma, D. S. Chauhan, and V. M. Katoch (2006). “Simultaneous ethambutol & isoniazid resistance in clinical isolates of *Mycobacterium tuberculosis*”. In: *Indian J Med Res* 123.2, pp. 125–30.
- Gupta, S. and G. M. Rodriguez (2018). “Mycobacterial extracellular vesicles and host pathogen interactions”. In: *Pathog Dis* 76.4.
- Guryev, V., M. J. Koudijs, E. Berezikov, S. L. Johnson, R. H. Plasterk, F. J. van Eeden, and E. Cuppen (2006). “Genetic variation in the zebrafish”. In: *Genome Res* 16.4, pp. 491–7.
- Gutfreund, K., W. Bienias, A. Szewczyk, and A. Kaszuba (2013). “Topical calcineurin inhibitors in dermatology. Part I: Properties, method and effectiveness of drug use”. In: *Postepy Dermatol Alergol* 30.3, pp. 165–9.
- Gwack, Y., S. Sharma, J. Nardone, B. Tanasa, A. Iuga, S. Srikanth, H. Okamura, D. Bolton, S. Feske, P. G. Hogan, and A. Rao (2006). “A genome-wide *Drosophila* RNAi screen identifies DYRK-family kinases as regulators of NFAT”. In: *Nature* 441.7093, pp. 646–50.
- Gyles, C. L. (2007). “Shiga toxin-producing *Escherichia coli*: an overview”. In: *J Anim Sci* 85.13 Suppl, E45–62.
- Gyorfy, Z., E. Duda, and C. Vizler (2013). “Interactions between LPS moieties and macrophage pattern recognition receptors”. In: *Vet Immunol Immunopathol* 152.1-2, pp. 28–36.

Habeck, H., J. Odenthal, B. Walderich, H. Maischein, S. Schulte-Merker, and consortium Tubingen screen (2002). “Analysis of a zebrafish VEGF receptor mutant reveals specific disruption of angiogenesis”. In: *Curr Biol* 12.16, pp. 1405–12.

Haffter, P., M. Granato, M. Brand, M. C. Mullins, M. Hammerschmidt, D. A. Kane, J. Odenthal, F. J. van Eeden, Y. J. Jiang, C. P. Heisenberg, R. N. Kelsh, M. Furutani-Seiki, E. Vogelsang, D. Beuchle, U. Schach, C. Fabian, and C. Nusslein-Volhard (1996). “The identification of genes with unique and essential functions in the development of the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 1–36.

Hagar, J. A., D. A. Powell, Y. Aachoui, R. K. Ernst, and E. A. Miao (2013). “Cyttoplasmic LPS activates caspase-11: implications in TLR4-independent endotoxic shock”. In: *Science* 341.6151, pp. 1250–3.

Hagedorn, M., K. H. Rohde, D. G. Russell, and T. Soldati (2009). “Infection by tubercular mycobacteria is spread by nonlytic ejection from their amoeba hosts”. In: *Science* 323.5922, pp. 1729–33.

Haibe, Y., M. Kreidieh, H. El Hajj, I. Khalifeh, D. Mukherji, S. Temraz, and A. Shamseddine (2020). “Resistance Mechanisms to Anti-angiogenic Therapies in Cancer”. In: *Front Oncol* 10, p. 221.

Haiko, P., T. Makinen, S. Keskitalo, J. Taipale, M. J. Karkkainen, M. E. Baldwin, S. A. Stacker, M. G. Achen, and K. Alitalo (2008). “Deletion of vascular endothelial growth factor C (VEGF-C) and VEGF-D is not equivalent to VEGF receptor 3 deletion in mouse embryos”. In: *Mol Cell Biol* 28.15, pp. 4843–50.

Haldar, A. K., C. Foltz, R. Finethy, A. S. Piro, E. M. Feeley, D. M. Pilla-Moffett, M. Komatsu, E. M. Frickel, and J. Coers (2015). “Ubiquitin systems mark pathogen-containing vacuoles as targets for host defense by guanylate binding proteins”. In: *Proc Natl Acad Sci U S A* 112.41, E5628–37.

Hall, C., M. V. Flores, T. Storm, K. Crosier, and P. Crosier (2007). “The zebrafish lysozyme C promoter drives myeloid-specific expression in transgenic fish”. In: *BMC Dev Biol* 7, p. 42.

Hall, K. L., L. D. Volk-Draper, M. J. Flister, and S. Ran (2012). “New model of macrophage acquisition of the lymphatic endothelial phenotype”. In: *PLoS One* 7.3, e31794.

Hall, T. J., M. P. Mullen, G. P. McHugo, K. E. Killick, S. C. Ring, D. P. Berry, C. N. Correia, J. A. Browne, S. V. Gordon, and D. E. MacHugh (2021). “Integrative genomics of the mammalian alveolar macrophage response to intracellular mycobacteria”. In: *BMC Genomics* 22.1, p. 343.

Halloum, I., S. Carrere-Kremer, M. Blaise, A. Viljoen, A. Bernut, V. Le Moigne, C. Vilchez, Y. Guerardel, G. Lutfalla, J. L. Herrmann, Jr. Jacobs W. R., and L. Kremer (2016). “Deletion of a dehydratase important for intracellular growth and cording renders rough *Mycobacterium abscessus* avirulent”. In: *Proc Natl Acad Sci U S A* 113.29, E4228–37.

Hambleton, S., S. Salem, J. Bustamante, V. Bigley, S. Boisson-Dupuis, J. Azevedo, A. Fortin, M. Haniffa, L. Ceron-Gutierrez, C. M. Bacon, G. Menon, C. Trouillet, D. McDonald, P. Carey, F. Ginhoux, L. Alsina, T. J. Zumwalt, X. F. Kong, D. Kumararatne, K. Butler, M. Hubeau, J. Feinberg, S. Al-Muhsen, A. Cant, L. Abel, D. Chaussabel, R. Doffinger, E. Talesnik, A. Grumach, A. Duarte, K. Abarca, D. Moraes-Vasconcelos, D. Burk, A. Berghuis, F. Geissmann, M. Collin, J. L. Casanova, and P. Gros (2011). “IRF8 mutations and human dendritic-cell immunodeficiency”. In: *N Engl J Med* 365.2, pp. 127–38.

Hamilton, J. R., D. Sachs, J. K. Lim, R. A. Langlois, P. Palese, and N. S. Heaton (2016). “Club cells surviving influenza A virus infection induce temporary nonspecific antiviral immunity”. In: *Proc Natl Acad Sci U S A* 113.14, pp. 3861–6.

Hammerschmidt, M., F. Pelegri, M. C. Mullins, D. A. Kane, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, P. Haffter, C. P. Heisenberg, Y. J. Jiang, R. N. Kelsh, J. Odenthal, R. M. Warga, and C. Nusslein-Volhard (1996a). “Mutations affecting morphogenesis during gastrulation and tail formation in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 143–51.

Hammerschmidt, M., F. Pelegri, M. C. Mullins, D. A. Kane, F. J. van Eeden, M. Granato, M. Brand, M. Furutani-Seiki, P. Haffter, C. P. Heisenberg, Y. J. Jiang, R. N. Kelsh, J. Odenthal, R. M. Warga, and C. Nusslein-Volhard (1996b). “*dino* and *mercedes*, two genes regulating dorsal development in the zebrafish embryo”. In: *Development* 123, pp. 95–102.

Hammond, F. R., A. Lewis, and P. M. Elks (2020). “If it’s not one thing, HIF’s another: immunoregulation by hypoxia inducible factors in disease”. In: *FEBS J* 287.18, pp. 3907–3916.

- Hammond, T. R., D. Robinton, and B. Stevens (2018). "Microglia and the Brain: Complementary Partners in Development and Disease". In: *Annu Rev Cell Dev Biol* 34, pp. 523–544.
- Han, L., M. Zhang, X. Liang, X. Jia, J. Jia, M. Zhao, and Y. Fan (2017). "Interleukin-33 promotes inflammation-induced lymphangiogenesis via ST2/TRAF6-mediated Akt/eNOS/NO signalling pathway". In: *Sci Rep* 7.1, p. 10602.
- Hanna, G. J., O. Neill A, J. M. Cutler, M. Flynn, T. Vijaykumar, J. R. Clark, L. J. Wirth, J. H. Lorch, J. C. Park, J. K. Mito, J. G. Lohr, J. Kaufman, N. S. Burr, L. I. Zon, and R. I. Haddad (2021). "A phase II trial of all-trans retinoic acid (ATRA) in advanced adenoid cystic carcinoma". In: *Oral Oncol* 119, p. 105366.
- Hannanta-Anan, P. and B. Y. Chow (2016). "Optogenetic Control of Calcium Oscillation Waveform Defines NFAT as an Integrator of Calcium Load". In: *Cell Syst* 2.4, pp. 283–8.
- Hansen, M., J. Peltier, B. Killy, B. Amin, B. Bodendorfer, A. Hartlova, S. Uebel, M. Bosmann, J. Hofmann, C. Buttner, A. B. Ekici, M. Kuttke, H. Franzky, C. Foged, S. Beer-Hammer, G. Schabbauer, M. Trost, and R. Lang (2019). "Macrophage Phosphoproteome Analysis Reveals MINCLE-dependent and -independent Mycobacterial Cord Factor Signaling". In: *Mol Cell Proteomics* 18.4, pp. 669–685.
- Hara, H., C. Ishihara, A. Takeuchi, T. Imanishi, L. Xue, S. W. Morris, M. Inui, T. Takai, A. Shibuya, S. Saijo, Y. Iwakura, N. Ohno, H. Koseki, H. Yoshida, J. M. Penninger, and T. Saito (2007). "The adaptor protein CARD9 is essential for the activation of myeloid cells through ITAM-associated and Toll-like receptors". In: *Nat Immunol* 8.6, pp. 619–29.
- Hara, M., T. Nagasaki, K. Shiga, and H. Takeyama (2016). "Suppression of Cancer -associated Fibroblasts and Endothelial Cells by Itraconazole in Bevacizumab -resistant Gastrointestinal Cancer". In: *Anticancer Res* 36.1, pp. 169–77.
- Harding, J., A. Ritter, A. Rayasam, Z. Fabry, and M. Sandor (2015).** "Lymphangiogenesis is induced by mycobacterial granulomas via vascular endothelial growth factor receptor-3 and supports systemic T-cell responses against mycobacterial antigen". In: *Am J Pathol* 185.2, pp. 432–45.
- Harding, J. S., M. Herbath, Y. Chen, A. Rayasam, A. Ritter, B. Csoka, G. Hasko, I. P. Michael, Z. Fabry, A. Nagy, and M. Sandor (2019).** "VEGF-

A from Granuloma Macrophages Regulates Granulomatous Inflammation by a Non-angiogenic Pathway during Mycobacterial Infection". In: *Cell Rep* 27.7, 2119–2131 e6.

– The non-angiogenic roles of VEGFA are an interesting dimension to this pathway that is often neglected, but this nice study looked at how VEGFA modulated macrophage responses to tuberculosis and found that VEGFA stimulation impaired their antimicrobial responses, arguing that, in addition to the impact that this cytokine has on the vasculature, that it is also involved in impeding macrophage killing of bacteria in a doubly beneficial activity for the pathogen. VEGFA increases myeloid cell recruitment and enhances local inflammation and blockade improves mouse survival. Interestingly, given the lack of an angiogenic response in the mouse model of tuberculosis, this actually facilitates functional dissection of these processes and reveals an interesting potential approach to understanding the relative contributions of angiogenesis and macrophage-targeted activities of VEGFA.

Hardison, S. E. and G. D. Brown (2012). "C-type lectin receptors orchestrate anti-fungal immunity". In: *Nat Immunol* 13.9, pp. 817–22.

Harland, C. W., D. Rabuka, C. R. Bertozzi, and R. Parthasarathy (2008). "The *Mycobacterium tuberculosis* virulence factor trehalose dimycolate imparts desiccation resistance to model mycobacterial membranes". In: *Biophys J* 94.12, pp. 4718–24.

Harper, J., C. Skerry, S. L. Davis, R. Tasneen, M. Weir, I. Kramnik, W. R. Bishai, M. G. Pomper, E. L. Nuermberger, and S. K. Jain (2012). "Mouse model of necrotic tuberculosis granulomas develops hypoxic lesions". In: *J Infect Dis* 205.4, pp. 595–602.

Harris, T. N., K. R. Szempruch, R. E. Dupuis, P. Serrano Rodriguez, and A. H. Toledo (2021). "Alemtuzumab in Renal Retransplantation - Transplant Outcomes and Associated Infections". In: *Transplant Proc* 53.10, pp. 2888–2894.

Hashimoto, D., A. Chow, C. Noizat, P. Teo, M. B. Beasley, M. Leboeuf, C. D. Becker, P. See, J. Price, D. Lucas, M. Greter, A. Mortha, S. W. Boyer, E. C. Forsberg, M. Tanaka, N. van Rooijen, A. Garcia-Sastre, E. R. Stanley, F. Ginhoux, P. S. Frenette, and M. Merad (2013). "Tissue-resident macrophages self-maintain locally throughout adult life with minimal contribution from circulating monocytes". In: *Immunity* 38.4, pp. 792–804.

Hashish, E., A. Merwad, S. Elgaml, A. Amer, H. Kamal, A. Elsadek, A. Marei, and M. Sitohy (2018). "*Mycobacterium marinum* infection in fish and man: epidemiology, pathophysiology and management; a review". In: *Vet Q* 38.1, pp. 35–46.

- Hason, M. and P. Bartunek (2019). “Zebrafish Models of Cancer-New Insights on Modeling Human Cancer in a Non-Mammalian Vertebrate”. In: *Genes (Basel)* 10.11.
- Hato, T., M. Tabata, and Y. Oike (2008). “The role of angiopoietin-like proteins in angiogenesis and metabolism”. In: *Trends Cardiovasc Med* 18.1, pp. 6–14.
- Hattori, Y., D. Morita, N. Fujiwara, D. Mori, T. Nakamura, H. Harashima, S. Yamasaki, and M. Sugita (2014). “Glycerol monomycolate is a novel ligand for the human, but not mouse macrophage inducible C-type lectin, Mincle”. In: *J Biol Chem* 289.22, pp. 15405–12.
- Hawgood, B. J. (2007). “Albert Calmette (1863-1933) and Camille Guerin (1872-1961): the C and G of BCG vaccine”. In: *J Med Biogr* 15.3, pp. 139–46.
- Hayashi, J. M., K. Richardson, E. S. Melzer, S. J. Sandler, B. B. Aldridge, M. S. Siegrist, and Y. S. Morita (2018). “Stress-Induced Reorganization of the Mycobacterial Membrane Domain”. In: *mBio* 9.1.
- He, W., A. Heinz, D. Jahn, and K. Hiller (2021). “Complexity of macrophage metabolism in infection”. In: *Curr Opin Biotechnol* 68, pp. 231–239.
- Heath, Victoria L., Roy Bicknell, and Victoria A. Simms (2017). “Development of an ImageJ-based method for analysing the developing zebrafish vasculature”. In: *Vascular Cell* 9.1.
- Heaton, N. S., R. A. Langlois, D. Sachs, J. K. Lim, P. Palese, and B. R. tenOever (2014). “Long-term survival of influenza virus infected club cells drives immunopathology”. In: *J Exp Med* 211.9, pp. 1707–14.
- Heguy, A., C. T. Baldari, G. Macchia, J. L. Telford, and M. Melli (1992). “Amino acids conserved in interleukin-1 receptors (IL-1Rs) and the *Drosophila* toll protein are essential for IL-1R signal transduction”. In: *J Biol Chem* 267.4, pp. 2605–9.
- Heisenberg, C. P., M. Brand, Y. J. Jiang, R. M. Warga, D. Beuchle, F. J. van Eeden, M. Furutani-Seiki, M. Granato, P. Haffter, M. Hammerschmidt, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, and C. Nusslein-Volhard (1996). “Genes involved in forebrain development in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 191–203.

Heitmann, L., H. Schoenen, S. Ehlers, R. Lang, and C. Holscher (2013). “Mincle is not essential for controlling *Mycobacterium tuberculosis* infection”. In: *Immunobiology* 218.4, pp. 506–16.

– This publication, in addition to Behler, Steinwede, Balboa, Ueberberg, Maus, Kirchhof, Yamasaki, Welte, and Maus (2012) and Lee, Kang, Yan, Lee, Jeon, Cho, and Kim (2012), investigated the role of MINCLE in controlling mycobacterial infections, but the three studies are difficult or impossible to reconcile as this study from Heitmann, Schoenen, Ehlers, Lang, and Holscher (2013) found that Mincle knockout mice had no increased susceptibility to infection while the other two studies found that Mincle was essential for control of bacterial growth. Further study is clearly needed to resolve the role of this receptor in the response to mycobacteria.

Henderson, B., P. A. Lund, and A. R. Coates (2010). “Multiple moonlighting functions of mycobacterial molecular chaperones”. In: *Tuberculosis (Edinb)* 90.2, pp. 119–24.

Henderson, J. W. (1997). “The yellow brick road to penicillin: a story of serendipity”. In: *Mayo Clin Proc* 72.7, pp. 683–7.

Heninger, E., L. H. Hogan, J. Karman, S. Macvilay, B. Hill, J. P. Woods, and M. Sandor (2006). “Characterization of the *Histoplasma capsulatum*-induced granuloma”. In: *J Immunol* 177.5, pp. 3303–13.

Henry, S. C., X. G. Daniell, A. R. Burroughs, M. Indaram, D. N. Howell, J. Coers, M. N. Starnbach, J. P. Hunn, J. C. Howard, C. G. Feng, A. Sher, and G. A. Taylor (2009). “Balance of Irgm protein activities determines IFN-gamma-induced host defense”. In: *J Leukoc Biol* 85.5, pp. 877–85.

Heppleston, A. G. (1949). “Quantitative air-borne tuberculosis in the rabbit; the cause of human type infection”. In: *J Exp Med* 89.6, pp. 597–610.

Herath, T. D., R. P. Darveau, C. J. Seneviratne, C. Y. Wang, Y. Wang, and L. Jin (2013). “Tetra- and penta-acylated lipid A structures of *Porphyromonas gingivalis* LPS differentially activate TLR4-mediated NF-kappaB signal transduction cascade and immuno-inflammatory response in human gingival fibroblasts”. In: *PLoS One* 8.3, e58496.

Herbein, G., A. Coaquette, D. Perez-Bercoff, and G. Pancino (2002). “Macrophage activation and HIV infection: can the Trojan horse turn into a fortress?” In: *Curr Mol Med* 2.8, pp. 723–38.

- Herbein, G. and A. Varin (2010). “The macrophage in HIV-1 infection: from activation to deactivation?” In: *Retrovirology* 7, p. 33.
- Herbst, S., U. E. Schaible, and B. E. Schneider (2011). “Interferon gamma activated macrophages kill mycobacteria by nitric oxide induced apoptosis”. In: *PLoS One* 6.5, e19105.
- Herbst, S., A. Shah, M. Carby, G. Chusney, N. Kikkeri, A. Dorling, E. Bignell, S. Shaunak, and D. Armstrong-James (2013). “A new and clinically relevant murine model of solid-organ transplant aspergillosis”. In: *Dis Model Mech* 6.3, pp. 643–51.
- Herbst, S., A. Shah, M. Mazon Moya, V. Marzola, B. Jensen, A. Reed, M. A. Birrell, S. Saijo, S. Mostowy, S. Shaunak, and D. Armstrong-James (2015). “Phagocytosis-dependent activation of a TLR9-BTK-calcineurin-NFAT pathway co-ordinates innate immunity to *Aspergillus fumigatus*”. In: *EMBO Mol Med* 7.3, pp. 240–58.
- Hershberg, R. (2016). “Human host range of *Mycobacterium tuberculosis*”. In: *Nat Genet* 48.12, pp. 1453–1454.
- Hershkovitz, I., H. D. Donoghue, D. E. Minnikin, H. May, O. Y. Lee, M. Feldman, E. Galili, M. Spigelman, B. M. Rothschild, and G. K. Bar-Gal (2015). “Tuberculosis origin: The Neolithic scenario”. In: *Tuberculosis (Edinb)* 95 Suppl 1, S122–6.
- Hett, E. C., M. C. Chao, and E. J. Rubin (2010). “Interaction and modulation of two antagonistic cell wall enzymes of mycobacteria”. In: *PLoS Pathog* 6.7, e1001020.
- Hett, E. C. and E. J. Rubin (2008). “Bacterial growth and cell division: a mycobacterial perspective”. In: *Microbiol Mol Biol Rev* 72.1, 126–56, table of contents.
- Higgins, D. and J. Dworkin (2012). “Recent progress in *Bacillus subtilis* sporulation”. In: *FEMS Microbiol Rev* 36.1, pp. 131–48.
- Higgins, J. P., K. Soares-Weiser, J. A. Lopez-Lopez, A. Kakourou, K. Chaplin, H. Christensen, N. K. Martin, J. A. Sterne, and A. L. Reingold (2016). “Association of BCG, DTP, and measles containing vaccines with childhood mortality: systematic review”. In: *BMJ* 355, p. i5170.
- Hinshaw, H. C., W. H. Feldman, and K. H. Pfuetze (1946). “Treatment of tuberculosis with streptomycin; a summary of observations on one hundred cases”. In: *J Am Med Assoc* 132.13, pp. 778–82.

- Hinshaw, H. C., M. M. Pyle, and W. H. Feldman (1947). “Streptomycin in tuberculosis”. In: *Am J Med* 2.5, pp. 429–35.
- Hmama, Z., S. Pena-Diaz, S. Joseph, and Y. Av-Gay (2015). “Immunoevasion and immunosuppression of the macrophage by *Mycobacterium tuberculosis*”. In: *Immunol Rev* 264.1, pp. 220–32.
- Ho, S. N., D. J. Thomas, L. A. Timmerman, X. Li, U. Francke, and G. R. Crabtree (1995). “NFATc3, a lymphoid-specific NFATc family member that is calcium-regulated and exhibits distinct DNA binding specificity”. In: *J Biol Chem* 270.34, pp. 19898–907.
- Ho, V. C. and G. H. Fong (2015). “Vasculogenesis and Angiogenesis in VEGF Receptor-1 Deficient Mice”. In: *Methods Mol Biol* 1332, pp. 161–76.
- Hodge, M. R., A. M. Ranger, F. Charles de la Brousse, T. Hoey, M. J. Grusby, and L. H. Glimcher (1996). “Hyperproliferation and dysregulation of IL-4 expression in NF-ATp-deficient mice”. In: *Immunity* 4.4, pp. 397–405.
- Hodge, S., G. Hodge, J. Ahern, H. Jersmann, M. Holmes, and P. N. Reynolds (2007). “Smoking alters alveolar macrophage recognition and phagocytic ability: implications in chronic obstructive pulmonary disease”. In: *Am J Respir Cell Mol Biol* 37.6, pp. 748–55.
- Hogan, B. M., F. L. Bos, J. Bussmann, M. Witte, N. C. Chi, H. J. Duckers, and S. Schulte-Merker (2009). “Ccbe1 is required for embryonic lymphangiogenesis and venous sprouting”. In: *Nat Genet* 41.4, pp. 396–8.
- Hogan, B. M. and S. Schulte-Merker (2017). “How to Plumb a Pisces: Understanding Vascular Development and Disease Using Zebrafish Embryos”. In: *Dev Cell* 42.6, pp. 567–583.
- Holden, L. A. and K. H. Brown (2018). “Baseline mRNA expression differs widely between common laboratory strains of zebrafish”. In: *Sci Rep* 8.1, p. 4780.
- Holmgren, J. (1981). “Actions of cholera toxin and the prevention and treatment of cholera”. In: *Nature* 292.5822, pp. 413–17.
- Holscher, C., N. Reiling, U. E. Schaible, A. Holscher, C. Bathmann, D. Korbel, I. Lenz, T. Sonntag, S. Kroger, S. Akira, H. Mossmann, C. J. Kirschning, H. Wagner,

- M. Freudenberg, and S. Ehlers (2008). “Containment of aerogenic *Mycobacterium tuberculosis* infection in mice does not require MyD88 adaptor function for TLR2, -4 and -9”. In: *Eur J Immunol* 38.3, pp. 680–94.
- Holtet, T. L., J. H. Graversen, I. Clemmensen, H. C. Thogersen, and M. Etzerodt (1997). “Ttranectin, a trimeric plasminogen-binding C-type lectin”. In: *Protein Sci* 6.7, pp. 1511–5.
- Honan, A. M. and Z. Chen (2021). “Stromal Cells Underlining the Paths From Autoimmunity, Inflammation to Cancer With Roles Beyond Structural and Nutritional Support”. In: *Front Cell Dev Biol* 9, p. 658984.
- Honda, J. R., R. Virdi, and E. D. Chan (2018). “Global Environmental Nontuberculous Mycobacteria and Their Contemporaneous Man-Made and Natural Niches”. In: *Front Microbiol* 9, p. 2029.
- Hong, H., L. Jiang, Y. Lin, C. He, G. Zhu, Q. Du, X. Wang, F. She, and Y. Chen (2016). “TNF-alpha promotes lymphangiogenesis and lymphatic metastasis of gallbladder cancer through the ERK1/2/AP-1/VEGF-D pathway”. In: *BMC Cancer* 16, p. 240.
- Hong, S. S., H. Lee, and K. W. Kim (2004). “HIF-1alpha: a valid therapeutic target for tumor therapy”. In: *Cancer Res Treat* 36.6, pp. 343–53.
- Hooijberg, E., A. Q. Bakker, J. J. Ruizendaal, and H. Spits (2000). “NFAT-controlled expression of GFP permits visualization and isolation of antigen-stimulated primary human T cells”. In: *Blood* 96.2, pp. 459–66.
- Hopkins, D. R. (2013). “Disease eradication”. In: *N Engl J Med* 368.1, pp. 54–63.
- Horsley, V., A. O. Aliprantis, L. Polak, L. H. Glimcher, and E. Fuchs (2008). “NFATc1 balances quiescence and proliferation of skin stem cells”. In: *Cell* 132.2, pp. 299–310.
- Horsley, V., B. B. Friday, S. Matteson, K. M. Kegley, J. Gephart, and G. K. Pavlath (2001). “Regulation of the growth of multinucleated muscle cells by an NFATC2-dependent pathway”. In: *J Cell Biol* 153.2, pp. 329–38.
- Horsley, V. and G. K. Pavlath (2002). “NFAT: ubiquitous regulator of cell differentiation and adaptation”. In: *J Cell Biol* 156.5, pp. 771–4.

- Hortle, E., K. E. Johnson, M. D. Johansen, T. Nguyen, J. A. Shavit, W. J. Britton, D. M. Tobin, and S. H. Oehlers (2019). "Thrombocyte Inhibition Restores Protective Immunity to Mycobacterial Infection in Zebrafish". In: *J Infect Dis* 220.3, pp. 524–534.
- Hoshijima, K., M. J. Juryne, and D. J. Grunwald (2016). "Precise Editing of the Zebrafish Genome Made Simple and Efficient". In: *Dev Cell* 36.6, pp. 654–67.
- Hoshijima, K., M. J. Juryne, D. Klatt Shaw, A. M. Jacobi, M. A. Behlke, and D. J. Grunwald (2019). "Highly Efficient CRISPR-Cas9-Based Methods for Generating Deletion Mutations and F0 Embryos that Lack Gene Function in Zebrafish". In: *Dev Cell* 51.5, 645–657 e4.
- Hosoi, T., Y. Imai, and T. Irimura (1998). "Coordinated binding of sugar, calcium, and antibody to macrophage C-type lectin". In: *Glycobiology* 8.8, pp. 791–8.
- Hosseini, R., G. E. M. Lamers, E. Bos, P. C. W. Hogendoorn, A. J. Koster, A. H. Meijer, H. P. Spaink, and M. J. M. Schaaf (2021). "The adapter protein Myd88 plays an important role in limiting mycobacterial growth in a zebrafish model for tuberculosis". In: *Virchows Arch* 479.2, pp. 265–275.
- Hou, X., J. Chen, Y. Luo, F. Liu, G. Xu, and Y. Gao (2013). "Silencing of STIM1 attenuates hypoxia-induced PASMCs proliferation via inhibition of the SOC/ Ca²⁺/NFAT pathway". In: *Respir Res* 14, p. 2.
- Houben, E. N., K. V. Korotkov, and W. Bitter (2014). "Take five - Type VII secretion systems of Mycobacteria". In: *Biochim Biophys Acta* 1843.8, pp. 1707–16.
- Houben, E. N., L. Nguyen, and J. Pieters (2006). "Interaction of pathogenic mycobacteria with the host immune system". In: *Curr Opin Microbiol* 9.1, pp. 76–85.
- Housden, B. E., M. Muhar, M. Gemberling, C. A. Gersbach, D. Y. Stainier, G. Seydoux, S. E. Mohr, J. Zuber, and N. Perrimon (2017). "Loss-of-function genetic tools for animal models: cross-species and cross-platform differences". In: *Nat Rev Genet* 18.1, pp. 24–40.
- Hoving, J. C., G. J. Wilson, and G. D. Brown (2014). "Signalling C-type lectin receptors, microbial recognition and immunity". In: *Cell Microbiol* 16.2, pp. 185–94.

- Howard, N. C. and S. A. Khader (2020). "Immunometabolism during *Mycobacterium tuberculosis* Infection". In: *Trends Microbiol* 28.10, pp. 832–850.
- Howe, K. et al. (2013). "The zebrafish reference genome sequence and its relationship to the human genome". In: *Nature* 496.7446, pp. 498–503.
- Hsu, K., D. Traver, J. L. Kutok, A. Hagen, T. X. Liu, B. H. Paw, J. Rhodes, J. N. Berman, L. I. Zon, J. P. Kanki, and A. T. Look (2004). "The pu.1 promoter drives myeloid gene expression in zebrafish". In: *Blood* 104.5, pp. 1291–7.
- Hsu, Y. M., Y. Zhang, Y. You, D. Wang, H. Li, O. Duramad, X. F. Qin, C. Dong, and X. Lin (2007). "The adaptor protein CARD9 is required for innate immune responses to intracellular pathogens". In: *Nat Immunol* 8.2, pp. 198–205.
- Hu, G., M. Guo, J. Xu, F. Wu, J. Fan, Q. Huang, G. Yang, Z. Lv, X. Wang, and Y. Jin (2019). "Nanoparticles Targeting Macrophages as Potential Clinical Therapeutic Agents Against Cancer and Inflammation". In: *Front Immunol* 10, p. 1998.
- Huang, C., W. Y. Ma, M. I. Dawson, M. Rincon, R. A. Flavell, and Z. Dong (1997). "Blocking activator protein-1 activity, but not activating retinoic acid response element, is required for the antitumor promotion effect of retinoic acid". In: *Proc Natl Acad Sci U S A* 94.11, pp. 5826–30.
- Huang, H., A. Bhat, G. Woodnutt, and R. Lappe (2010). "Targeting the ANGPT-TIE2 pathway in malignancy". In: *Nat Rev Cancer* 10.8, pp. 575–85.
- Huang, H., E. J. Chang, J. Ryu, Z. H. Lee, Y. Lee, and H. H. Kim (2006). "Induction of c-Fos and NFATc1 during RANKL-stimulated osteoclast differentiation is mediated by the p38 signaling pathway". In: *Biochem Biophys Res Commun* 351.1, pp. 99–105.
- Huang, L. E., J. Gu, M. Schau, and H. F. Bunn (1998). "Regulation of hypoxia-inducible factor 1alpha is mediated by an O₂-dependent degradation domain via the ubiquitin-proteasome pathway". In: *Proc Natl Acad Sci U S A* 95.14, pp. 7987–92.
- Huang, T., W. Gu, E. Liu, B. Wang, G. Wang, F. Dong, F. Guo, W. Jiao, Y. Sun, X. Wang, S. Li, and G. Xu (2022). "miR-301b-5p and its target gene nfatc2ip regulate inflammatory responses in the liver of rainbow trout (*Oncorhynchus mykiss*) under high temperature stress". In: *Ecotoxicol Environ Saf* 242, p. 113915.

- Huang, T., Z. Xie, J. Wang, M. Li, N. Jing, and L. Li (2011). “Nuclear factor of activated T cells (NFAT) proteins repress canonical Wnt signaling via its interaction with Dishevelled (Dvl) protein and participate in regulating neural progenitor cell proliferation and differentiation”. In: *J Biol Chem* 286.43, pp. 37399–405.
- Huang, X., Y. Li, M. Fu, and H. B. Xin (2018). “Polarizing Macrophages *in vitro*”. In: *Methods Mol Biol* 1784, pp. 119–126.
- Huang, Y., Q. Li, Z. Feng, and L. Zheng (2020). “STIM1 controls calcineurin / Akt / mTOR / NFATC2-mediated osteoclastogenesis induced by RANKL/M-CSF”. In: *Exp Ther Med* 20.2, pp. 736–747.
- Huberts, D. H. and I. J. van der Klei (2010). “Moonlighting proteins: an intriguing mode of multitasking”. In: *Biochim Biophys Acta* 1803.4, pp. 520–5.
- Hummer, A. M., B. Abanades, and C. M. Deane (2022). “Advances in computational structure-based antibody design”. In: *Curr Opin Struct Biol* 74, p. 102379.
- Hunter, R. L., S. A. Hwang, C. Jagannath, and J. K. Actor (2016). “Cord factor as an invisibility cloak? A hypothesis for asymptomatic TB persistence”. In: *Tuberculosis (Edinb)* 101S, S2–S8.
- Hunter, R. L., C. Jagannath, and J. K. Actor (2007). “Pathology of postprimary tuberculosis in humans and mice: contradiction of long-held beliefs”. In: *Tuberculosis (Edinb)* 87.4, pp. 267–78.
- Hunter, R. L., M. Olsen, C. Jagannath, and J. K. Actor (2006). “Trehalose 6,6'-dimycolate and lipid in the pathogenesis of caseating granulomas of tuberculosis in mice”. In: *Am J Pathol* 168.4, pp. 1249–61.
- Hunter, R. L., N. Venkataprasad, and M. R. Olsen (2006). “The role of trehalose dimycolate (cord factor) on morphology of virulent *M. tuberculosis* *in vitro*”. In: *Tuberculosis (Edinb)* 86.5, pp. 349–56.
- Hussell, T. and T. J. Bell (2014). “Alveolar macrophages: plasticity in a tissue-specific context”. In: *Nat Rev Immunol* 14.2, pp. 81–93.
- Hustedt, N. and D. Durocher (2016). “The control of DNA repair by the cell cycle”. In: *Nat Cell Biol* 19.1, pp. 1–9.

Huynh, K. K., S. A. Joshi, and E. J. Brown (2011). “A delicate dance: host response to mycobacteria”. In: *Curr Opin Immunol* 23.4, pp. 464–72.

Ichino, N., M. R. Serres, R. M. Urban, M. D. Urban, A. J. Treichel, K. J. Schaefbauer, L. E. Tallant, G. K. Varshney, K. J. Skuster, M. S. McNulty, C. L. Daby, Y. Wang, H. K. Liao, S. El-Rass, Y. Ding, W. Liu, J. L. Anderson, M. D. Wishman, A. Sabharwal, L. A. Schimmenti, S. Sivasubbu, D. Balciunas, M. Hammerschmidt, S. A. Farber, X. Y. Wen, X. Xu, M. McGrail, J. J. Essner, S. M. Burgess, K. J. Clark, and S. C. Ekker (2020). “Building the vertebrate codex using the gene breaking protein trap library”. In: *Elife* 9.

Idh, J., B. Andersson, M. Lerm, J. Raffetseder, D. Eklund, H. Woksepp, J. Werngren, M. Mansjo, T. Sundqvist, O. Stendahl, and T. Schon (2017). “Reduced susceptibility of clinical strains of *Mycobacterium tuberculosis* to reactive nitrogen species promotes survival in activated macrophages”. In: *PLoS One* 12.7, e0181221.

Imhof, B. A. and M. Aurrand-Lions (2004). “Adhesion mechanisms regulating the migration of monocytes”. In: *Nat Rev Immunol* 4.6, pp. 432–44.

Incio, J., J. Tam, N. N. Rahbari, P. Suboj, D. T. McManus, S. M. Chin, T. D. Vardam, A. Batista, S. Babykutty, K. Jung, A. Khachatryan, T. Hato, J. A. Ligibel, I. E. Krop, S. B. Puchner, C. L. Schlett, U. Hoffmman, M. Ancukiewicz, M. Shibuya, P. Carmeliet, R. Soares, D. G. Duda, R. K. Jain, and D. Fukumura (2016). “PIGF/VEGFR-1 Signaling Promotes Macrophage Polarization and Accelerated Tumor Progression in Obesity”. In: *Clin Cancer Res* 22.12, pp. 2993–3004.

Indigo, J., R. L. Hunter, and J. K. Actor (2002). “Influence of trehalose 6,6'-dimycolate (TDM) during mycobacterial infection of bone marrow macrophages”. In: *Microbiology (Reading)* 148.Pt 7, pp. 1991–1998.

– (2003). “Cord factor trehalose 6,6'-dimycolate (TDM) mediates trafficking events during mycobacterial infection of murine macrophages”. In: *Microbiology (Reading)* 149.Pt 8, pp. 2049–2059.

Ishikawa, E., T. Ishikawa, Y. S. Morita, K. Toyonaga, H. Yamada, O. Takeuchi, T. Kinoshita, S. Akira, Y. Yoshikai, and S. Yamasaki (2009). “Direct recognition of the mycobacterial glycolipid, trehalose dimycolate, by C-type lectin Mincle”. In: *J Exp Med* 206.13, pp. 2879–88.

– This study definitively shows that MINCLE binds to TDM and is the definitive receptor for this important mycobacterial ligand.

Ishikawa, E., D. Mori, and S. Yamasaki (2017). “Recognition of Mycobacterial Lipids by Immune Receptors”. In: *Trends Immunol* 38.1, pp. 66–76.

Ishikawa, T., F. Itoh, S. Yoshida, S. Saijo, T. Matsuzawa, T. Gonoi, T. Saito, Y. Okawa, N. Shibata, T. Miyamoto, and S. Yamasaki (2013). “Identification of distinct ligands for the C-type lectin receptors Mincle and Dectin-2 in the pathogenic fungus *Malassezia*”. In: *Cell Host Microbe* 13.4, pp. 477–88.

Isiaku, A. I., Z. Zhang, V. Pazhakh, H. R. Manley, E. R. Thompson, L. C. Fox, S. Yerneni, P. Blomberg, and G. J. Lieschke (2021). “Transient, flexible gene editing in zebrafish neutrophils and macrophages for determination of cell-autonomous functions”. In: *Dis Model Mech* 14.7.

Italiani, P. and D. Boraschi (2014). “From Monocytes to M1/M2 Macrophages: Phenotypical vs. Functional Differentiation”. In: *Front Immunol* 5, p. 514.

Itatani, Y., K. Kawada, T. Yamamoto, and Y. Sakai (2018). “Resistance to Anti-Angiogenic Therapy in Cancer-Alterations to Anti-VEGF Pathway”. In: *Int J Mol Sci* 19.4.

Iwamoto, F. M., L. E. Abrey, K. Beal, P. H. Gutin, M. K. Rosenblum, V. E. Reuter, L. M. DeAngelis, and A. B. Lassman (2009). “Patterns of relapse and prognosis after bevacizumab failure in recurrent glioblastoma”. In: *Neurology* 73.15, pp. 1200–6.

Iwasaki, A. and R. Medzhitov (2010). “Regulation of adaptive immunity by the innate immune system”. In: *Science* 327.5963, pp. 291–5.

Izquierdo Lafuente, B., R. Ummels, C. Kuijl, W. Bitter, and A. Speer (2021). “*Mycobacterium tuberculosis* Toxin CpnT Is an ESX-5 Substrate and Requires Three Type VII Secretion Systems for Intracellular Secretion”. In: *mBio* 12.2.

Izumi, Y., S. Kim, M. Namba, H. Yasumoto, H. Miyazaki, M. Hoshiga, Y. Kaneda, R. Morishita, Y. Zhan, and H. Iwao (2001). “Gene transfer of dominant-negative mutants of extracellular signal-regulated kinase and c-Jun NH₂-terminal kinase prevents neointimal formation in balloon-injured rat artery”. In: *Circ Res* 88.11, pp. 1120–6.

- Jackson, M. (2014). "The mycobacterial cell envelope-lipids". In: *Cold Spring Harb Perspect Med* 4.10.
- Jain, J., P. G. McCaffrey, Z. Miner, T. K. Kerppola, J. N. Lambert, G. L. Verdine, T. Curran, and A. Rao (1993). "The T-cell transcription factor NFATp is a substrate for calcineurin and interacts with Fos and Jun". In: *Nature* 365.6444, pp. 352–5.
- Jakab, M., T. Rostalski, K. H. Lee, C. Mogler, and H. G. Augustin (2022). "Tie2 Receptor in Tumor-Infiltrating Macrophages Is Dispensable for Tumor Angiogenesis and Tumor Relapse after Chemotherapy". In: *Cancer Res* 82.7, pp. 1353–1364.
- Jambusaria, A., Z. Hong, L. Zhang, S. Srivastava, A. Jana, P. T. Toth, Y. Dai, A. B. Malik, and J. Rehman (2020).** "Endothelial heterogeneity across distinct vascular beds during homeostasis and inflammation". In: *Elife* 9.
- Janeway, Charles (2005). *Immunobiology : the immune system in health and disease*. 6th. New York: Garland Science.
- Jang, G. H., I. S. Park, J. H. Yang, J. Bischoff, and Y. M. Lee (2010). "Differential functions of genes regulated by VEGF-NFATc1 signaling pathway in the migration of pulmonary valve endothelial cells". In: *FEBS Lett* 584.1, pp. 141–6.
- Jang, J. G. and J. H. Chung (2020). "Diagnosis and treatment of multidrug-resistant tuberculosis". In: *Yeungnam Univ J Med* 37.4, pp. 277–285.
- Jankute, M., J. A. Cox, J. Harrison, and G. S. Besra (2015).** "Assembly of the Mycobacterial Cell Wall". In: *Annu Rev Microbiol* 69, pp. 405–23.
- Jao, L. E., S. R. Wente, and W. Chen (2013). "Efficient multiplex biallelic zebrafish genome editing using a CRISPR nuclease system". In: *Proc Natl Acad Sci U S A* 110.34, pp. 13904–9.
- Jappinen, N., I. Felix, E. Lokka, S. Tyystjarvi, A. Pynttari, T. Lahtela, H. Gerke, K. Elimä, P. Rantakari, and M. Salmi (2019). "Fetal-derived macrophages dominate in adult mammary glands". In: *Nat Commun* 10.1, p. 281.
- Jarlier, V. and H. Nikaido (1994). "Mycobacterial cell wall: structure and role in natural resistance to antibiotics". In: *FEMS Microbiol Lett* 123.1-2, pp. 11–8.

Jauliac, S., C. Lopez-Rodriguez, L. M. Shaw, L. F. Brown, A. Rao, and A. Toker (2002). “The role of NFAT transcription factors in integrin-mediated carcinoma invasion”. In: *Nat Cell Biol* 4.7, pp. 540–4.

Jayachandran, R., J. Gatfield, J. Massner, I. Albrecht, B. Zanolari, and J. Pieters (2008). “RNA interference in J774 macrophages reveals a role for coronin 1 in mycobacterial trafficking but not in actin-dependent processes”. In: *Mol Biol Cell* 19.3, pp. 1241–51.

Jayachandran, R., V. Sundaramurthy, B. Combaluzier, P. Mueller, H. Korf, K. Huygen, T. Miyazaki, I. Albrecht, J. Massner, and J. Pieters (2007). “Survival of mycobacteria in macrophages is mediated by coronin 1-dependent activation of calcineurin”. In: *Cell* 130.1, pp. 37–50.

– One of, if not the, earliest studies on the role of calcineurin in regulating myeloid responses to mycobacteria, this work found that mycobacteria specifically induce the recruitment of CORO1A to the phagosome which blocks lysosomal fusion (could there be a receptor within the phagosome mediating this?). They found that CORO1A activates calcineurin, which is responsible for blocking this phagosomal-lysosomal fusion event and pharmacological inhibition of calcineurin resulted in fusion and mycobacterial killing. This is an intriguing role that further supports the use of calcineurin inhibitors in the treatment of mycobacterial infections.

Jegouzo, S. A., E. C. Harding, O. Acton, M. J. Rex, A. J. Fadden, M. E. Taylor, and K. Drickamer (2014). “Defining the conformation of human mincle that interacts with mycobacterial trehalose dimycolate”. In: *Glycobiology* 24.12, pp. 1291–300.

Jeon, S. H., B. C. Chae, H. A. Kim, G. Y. Seo, D. W. Seo, G. T. Chun, N. S. Kim, S. W. Yie, W. H. Byeon, S. H. Eom, K. S. Ha, Y. M. Kim, and P. H. Kim (2007). “Mechanisms underlying TGF-beta1-induced expression of VEGF and Flk-1 in mouse macrophages and their implications for angiogenesis”. In: *J Leukoc Biol* 81.2, pp. 557–66.

Jessie, B. and H. M. Dobrovolny (2021). “The role of syncytia during viral infections”. In: *J Theor Biol* 525, p. 110749.

Jetten, N., S. Verbruggen, M. J. Gijbels, M. J. Post, M. P. De Winther, and M. M. Donners (2014). “Anti-inflammatory M2, but not pro-inflammatory M1 macrophages promote angiogenesis *in vivo*”. In: *Angiogenesis* 17.1, pp. 109–18.

- Ji, D. X., K. C. Witt, D. I. Kotov, S. R. Margolis, A. Louie, V. Chevee, K. J. Chen, M. M. Gaidt, H. S. Dhaliwal, A. Y. Lee, S. L. Nishimura, D. S. Zamboni, I. Kramnik, D. A. Portnoy, K. H. Darwin, and R. E. Vance (2021). “Role of the transcriptional regulator SP140 in resistance to bacterial infections via repression of type I interferons”. In: *Elife* 10.
- Jia, J., J. Fu, and H. Tang (2021). “Activation and Evasion of RLR Signaling by DNA Virus Infection”. In: *Front Microbiol* 12, p. 804511.
- Jiang, Y. J., M. Brand, C. P. Heisenberg, D. Beuchle, M. Furutani-Seiki, R. N. Kelsh, R. M. Warga, M. Granato, P. Haffter, M. Hammerschmidt, D. A. Kane, M. C. Mullins, J. Odenthal, F. J. van Eeden, and C. Nusslein-Volhard (1996). “Mutations affecting neurogenesis and brain morphology in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 205–16.
- Jiang, Z., P. Georgel, X. Du, L. Shamel, S. Sovath, S. Mudd, M. Huber, C. Kalis, S. Keck, C. Galanos, M. Freudenberg, and B. Beutler (2005). “CD14 is required for MyD88-independent LPS signaling”. In: *Nat Immunol* 6.6, pp. 565–70.
- Jin, S. W., D. Beis, T. Mitchell, J. N. Chen, and D. Y. Stainier (2005). “Cellular and molecular analyses of vascular tube and lumen formation in zebrafish”. In: *Development* 132.23, pp. 5199–209.
- Jinich, Adrian, Sakila Z. Nazia, Andrea V. Tellez, Amy M. Wu, Clare M. Smith, and Kyu Rhee (2022). “Genome-wide co-essentiality analysis in *Mycobacterium tuberculosis* reveals an itaconate defense enzyme module”. In: *bioRxiv*.
- Johansen, M. D., J. A. Kasparian, E. Hortle, W. J. Britton, A. C. Purdie, and S. H. Oehlers (2018). “*Mycobacterium marinum* infection drives foam cell differentiation in zebrafish infection models”. In: *Dev Comp Immunol* 88, pp. 169–172.
- Johnson, C. J., J. M. Davis, A. Huttenlocher, J. F. Kernien, and J. E. Nett (2018). “Emerging Fungal Pathogen *Candida auris* Evades Neutrophil Attack”. In: *mBio* 9.4.
- Johnson, E. N., Y. M. Lee, T. L. Sander, E. Rabkin, F. J. Schoen, S. Kaushal, and J. Bischoff (2003a). “NFATc1 mediates vascular endothelial growth factor-induced proliferation of human pulmonary valve endothelial cells”. In: *J Biol Chem* 278.3, pp. 1686–92.

Johnson, G. B., G. J. Brunn, A. H. Tang, and J. L. Platt (2003b). “Evolutionary clues to the functions of the Toll-like family as surveillance receptors”. In: *Trends Immunol* 24.1, pp. 19–24.

Jones, E. A., J. Conover, and A. J. Symes (2000). “Identification of a novel gp130-responsive site in the vasoactive intestinal peptide cytokine response element”. In: *J Biol Chem* 275.46, pp. 36013–20.

Jones, E. A., D. Sun, L. Kobierski, and A. J. Symes (2003). “NFAT4 is expressed in primary astrocytes and activated by glutamate”. In: *J Neurosci Res* 72.2, pp. 191–7.

Jong, J. L. de, C. E. Burns, A. T. Chen, E. Pugach, E. A. Mayhall, A. C. Smith, H. A. Feldman, Y. Zhou, and L. I. Zon (2011). “Characterization of immune-matched hematopoietic transplantation in zebrafish”. In: *Blood* 117.16, pp. 4234–42.

Jordan, T. C., S. H. Burnett, S. Carson, S. M. Caruso, K. Clase, R. J. DeJong, J. J. Dennehy, D. R. Denver, D. Dunbar, S. C. Elgin, A. M. Findley, C. R. Gissendanner, U. P. Golebiewska, N. Guild, G. A. Hartzog, W. H. Grillo, G. P. Hollowell, L. E. Hughes, A. Johnson, R. A. King, L. O. Lewis, W. Li, F. Rosenzweig, M. R. Rubin, M. S. Saha, J. Sandoz, C. D. Shaffer, B. Taylor, L. Temple, E. Vazquez, V. C. Ware, L. P. Barker, K. W. Bradley, D. Jacobs-Sera, W. H. Pope, D. A. Russell, S. G. Cresawn, D. Lopatto, C. P. Bailey, and G. F. Hatfull (2014). “A broadly implementable research course in phage discovery and genomics for first-year undergraduate students”. In: *mBio* 5.1, e01051–13.

Joshi, J. M. (2011). “Tuberculosis chemotherapy in the 21 century: Back to the basics”. In: *Lung India* 28.3, pp. 193–200.

Joshi, N., J. M. Walter, and A. V. Misharin (2018). “Alveolar Macrophages”. In: *Cell Immunol* 330, pp. 86–90.

Josko, J. and M. Mazurek (2004). “Transcription factors having impact on vascular endothelial growth factor (VEGF) gene expression in angiogenesis”. In: *Med Sci Monit* 10.4, RA89–98.

Juffermans, N. P., S. Florquin, L. Camoglio, A. Verbon, A. H. Kolk, P. Speelman, S. J. van Deventer, and T. van Der Poll (2000). “Interleukin-1 signaling is essential for host defense during murine pulmonary tuberculosis”. In: *J Infect Dis* 182.3, pp. 902–8.

Jumper, J., R. Evans, A. Pritzel, T. Green, M. Figurnov, O. Ronneberger, K. Tunyasuvunakool, R. Bates, A. Zidek, A. Potapenko, A. Bridgland, C. Meyer, S. A. A. Kohl, A. J. Ballard, A. Cowie, B. Romera-Paredes, S. Nikolov, R. Jain, J. Adler, T. Back, S. Petersen, D. Reiman, E. Clancy, M. Zielinski, M. Steinegger, M. Paccholska, T. Berghammer, S. Bodenstein, D. Silver, O. Vinyals, A. W. Senior, K. Kavukcuoglu, P. Kohli, and D. Hassabis (2021). "Highly accurate protein structure prediction with AlphaFold". In: *Nature* 596.7873, pp. 583–589.

June, C. H. (2007). "Adoptive T cell therapy for cancer in the clinic". In: *J Clin Invest* 117.6, pp. 1466–76.

Jung, H. M., D. Castranova, M. R. Swift, V. N. Pham, M. Venero Galanternik, S. Isogai, M. G. Butler, T. S. Mulligan, and B. M. Weinstein (2017). "Development of the larval lymphatic system in zebrafish". In: *Development* 144.11, pp. 2070–2081.

Kabadi, A. M., D. G. Ousterout, I. B. Hilton, and C. A. Gersbach (2014). "Multiplex CRISPR/Cas9-based genome engineering from a single lentiviral vector". In: *Nucleic Acids Res* 42.19, e147.

Kabbinavar, F. F., J. Hambleton, R. D. Mass, H. I. Hurwitz, E. Bergsland, and S. Sarkar (2005). "Combined analysis of efficacy: the addition of bevacizumab to fluorouracil/leucovorin improves survival for patients with metastatic colorectal cancer". In: *J Clin Oncol* 23.16, pp. 3706–12.

Kahn, B. M., A. Lucas, R. G. Alur, M. D. Wengyn, G. W. Schwartz, J. Li, K. Sun, H. C. Maurer, K. P. Olive, R. B. Faryabi, and B. Z. Stanger (2021). "The vascular landscape of human cancer". In: *J Clin Invest* 131.2.

Kahnert, A., P. Seiler, M. Stein, S. Bandermann, K. Hahnke, H. Mollenkopf, and S. H. Kaufmann (2006). "Alternative activation deprives macrophages of a coordinated defense program to *Mycobacterium tuberculosis*". In: *Eur J Immunol* 36.3, pp. 631–47.

Kalinda, A. S. and C. C. Aldrich (2012). "Pyrazinamide: a frontline drug used for tuberculosis. Molecular mechanism of action resolved after 50 years?" In: *ChemMedChem* 7.4, pp. 558–60.

Kalucka, J., L. Bierhansl, B. Wielockx, P. Carmeliet, and G. Eelen (2017). "Interaction of endothelial cells with macrophages-linking molecular and metabolic signaling". In: *Pflugers Arch* 469.3-4, pp. 473–483.

Kambouchner, M., D. Pirici, J. F. Uhl, L. Mogoanta, D. Valeyre, and J. F. Bernaudin (2011). “Lymphatic and blood microvasculature organisation in pulmonary sarcoid granulomas”. In: *Eur Respir J* 37.4, pp. 835–40.

Kamei, M., W. B. Saunders, K. J. Bayless, L. Dye, G. E. Davis, and B. M. Weinstein (2006). “Endothelial tubes assemble from intracellular vacuoles *in vivo*”. In: *Nature* 442.7101, pp. 453–6.

Kamentsky, L., T. R. Jones, A. Fraser, M. A. Bray, D. J. Logan, K. L. Madden, V. Ljosa, C. Rueden, K. W. Eliceiri, and A. E. Carpenter (2011). “Improved structure, function and compatibility for CellProfiler: modular high-throughput image analysis software”. In: *Bioinformatics* 27.8, pp. 1179–80.

Kaminuma, O., F. Kitamura, N. Kitamura, T. Hiroi, H. Miyoshi, A. Miyawaki, and S. Miyatake (2008). “Differential contribution of NFATc2 and NFATc1 to TNF-alpha gene expression in T cells”. In: *J Immunol* 180.1, pp. 319–26.

– The domain-specific features of the NFAT isoforms that allow them to regulate the expression of specific genes is of clear basic scientific interest. Here, NFATC1 and NFATC2 were compared in their ability to induce TNF transcription and, while both bound to the TNF promoter, only NFATC2 was able to induce expression due to distinct features in the C-terminal domain. These factors may influence the ability of NFATC2 to induce transcription of VEGFA, but a more comprehensive biochemical approach is needed to understand these protein:protein interactions important for the induction of angiogenesis.

Kan-Sutton, C., C. Jagannath, and Jr. Hunter R. L. (2009). “Trehalose 6,6'-dimycolate on the surface of *Mycobacterium tuberculosis* modulates surface marker expression for antigen presentation and costimulation in murine macrophages”. In: *Microbes Infect* 11.1, pp. 40–8.

Kane, D. A., M. Hammerschmidt, M. C. Mullins, H. M. Maischein, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, P. Haffter, C. P. Heisenberg, Y. J. Jiang, R. N. Kelsh, J. Odenthal, R. M. Warga, and C. Nusslein-Volhard (1996a). “The zebrafish epiboly mutants”. In: *Development* 123, pp. 47–55.

Kane, D. A., H. M. Maischein, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, P. Haffter, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, R. N. Kelsh, M. C. Mullins, J. Odenthal, R. M. Warga, and C. Nusslein-Volhard (1996b). “The zebrafish early arrest mutants”. In: *Development* 123, pp. 57–66.

- Kaneko, H., H. Yamada, S. Mizuno, T. Udagawa, Y. Kazumi, K. Sekikawa, and I. Sugawara (1999). “Role of tumor necrosis factor-alpha in *Mycobacterium*-induced granuloma formation in tumor necrosis factor-alpha-deficient mice”. In: *Lab Invest* 79.4, pp. 379–86.
- Kang, B. K. and L. S. Schlesinger (1998). “Characterization of mannose receptor-dependent phagocytosis mediated by *Mycobacterium tuberculosis* lipoarabinomannan”. In: *Infect Immun* 66.6, pp. 2769–77.
- Kang, D., G. Liu, A. Lundstrom, E. Gelius, and H. Steiner (1998). “A peptidoglycan recognition protein in innate immunity conserved from insects to humans”. In: *Proc Natl Acad Sci U S A* 95.17, pp. 10078–82.
- Kang, Y. J., B. Kusler, M. Otsuka, M. Hughes, N. Suzuki, S. Suzuki, W. C. Yeh, S. Akira, J. Han, and P. P. Jones (2007). “Calcineurin negatively regulates TLR-mediated activation pathways”. In: *J Immunol* 179.7, pp. 4598–607.
- Kanno, E., K. Kawakami, H. Tanno, A. Suzuki, N. Sato, A. Masaki, A. Imamura, N. Takagi, T. Miura, H. Yamamoto, K. Ishii, H. Hara, Y. Imai, R. Maruyama, and M. Tachi (2017). “Contribution of CARD9-mediated signalling to wound healing in skin”. In: *Exp Dermatol* 26.11, pp. 1097–1104.
- Kanther, M. and J. F. Rawls (2010). “Host-microbe interactions in the developing zebrafish”. In: *Curr Opin Immunol* 22.1, pp. 10–9.
- Kanther, M., X. Sun, M. Muhlbauer, L. C. Mackey, 3rd Flynn E. J., M. Bagnat, C. Jobin, and J. F. Rawls (2011). “Microbial colonization induces dynamic temporal and spatial patterns of NF-kappaB activation in the zebrafish digestive tract”. In: *Gastroenterology* 141.1, pp. 197–207.
- Kaplan, G., F. A. Post, A. L. Moreira, H. Wainwright, B. N. Kreiswirth, M. Tanverdi, B. Mathema, S. V. Ramaswamy, G. Walther, L. M. Steyn, 3rd Barry C. E., and L. G. Bekker (2003). “*Mycobacterium tuberculosis* growth at the cavity surface: a microenvironment with failed immunity”. In: *Infect Immun* 71.12, pp. 7099–108.
- Kar, P., G. R. Mirams, H. C. Christian, and A. B. Parekh (2016).** “Control of NFAT Isoform Activation and NFAT-Dependent Gene Expression through Two Coincident and Spatially Segregated Intracellular Ca(2+) Signals”. In: *Mol Cell* 64.4, pp. 746–759.

Kar, P. and A. B. Parekh (2015). “Distinct spatial Ca²⁺ signatures selectively activate different NFAT transcription factor isoforms”. In: *Mol Cell* 58.2, pp. 232–43.

– While there had been work prior to this, this was the first comprehensive effort to understand the distinguishing features of the various NFAT isoforms and thoroughly dissected the kinetics of NFATC2 and NFATC3 activation and discovered that NFATC3 is a rapidly switching isoform sensitive to small changes in calcium while NFATC2 steadily accumulates over time and is induced by sustained activity. This is consistent with a role for NFATC2 in the granuloma, where signaling is constantly induced and might suggest a role for NFATC3 in earlier stages of infection.

Kar, P., K. Samanta, H. Kramer, O. Morris, D. Bakowski, and A. B. Parekh (2014). “Dynamic assembly of a membrane signaling complex enables selective activation of NFAT by Orai1”. In: *Curr Biol* 24.12, pp. 1361–1368.

Karar, J. and A. Maity (2011). “PI3K/AKT/mTOR Pathway in Angiogenesis”. In: *Front Mol Neurosci* 4, p. 51.

Karlstrom, R. O., T. Trowe, S. Klostermann, H. Baier, M. Brand, A. D. Crawford, B. Grunewald, P. Haffter, H. Hoffmann, S. U. Meyer, B. K. Muller, S. Richter, F. J. van Eeden, C. Nusslein-Volhard, and F. Bonhoeffer (1996). “Zebrafish mutations affecting retinotectal axon pathfinding”. In: *Development* 123, pp. 427–38.

Karpurapu, M., Y. G. Lee, Z. Qian, J. Wen, M. N. Ballinger, L. Rusu, S. Chung, J. Deng, F. Qian, B. F. Reader, T. S. Nirujogi, G. Y. Park, D. Pei, and J. W. Christman (2018). “Inhibition of nuclear factor of activated T cells (NFAT) c3 activation attenuates acute lung injury and pulmonary edema in murine models of sepsis”. In: *Oncotarget* 9.12, pp. 10606–10620.

Karra, R., M. J. Foglia, W. Y. Choi, C. Belliveau, P. DeBenedittis, and K. D. Poss (2018). “Vegfaa instructs cardiac muscle hyperplasia in adult zebrafish”. In: *Proc Natl Acad Sci U S A* 115.35, pp. 8805–8810.

Kasai, F., N. Hirayama, M. Fukushima, A. Kohara, and Y. Nakamura (2022). “THP-1 reference data: Proposal of an *in vitro* branched evolution model for cancer cell lines”. In: *Int J Cancer* 151.3, pp. 463–472.

Katzmarski, N., J. Dominguez-Andres, B. Cirovic, G. Renieris, E. Ciarlo, D. Le Roy, K. Lepikhov, K. Kattler, G. Gasparoni, K. Handler, H. Theis, M. Beyer, J. W. M. van der Meer, L. A. B. Joosten, J. Walter, J. L. Schultze, T. Roger, E. J.

Giamarellos-Bourboulis, A. Schlitzer, and M. G. Netea (2021). “Transmission of trained immunity and heterologous resistance to infections across generations”. In: *Nat Immunol* 22.11, pp. 1382–1390.

Katzmarski, N., J. Dominguez-Andres, B. Cirovic, G. Renieris, E. Ciarlo, D. Le Roy, K. Lepikhov, K. Kattler, G. Gasparoni, K. Handler, H. Theis, M. Beyer, J. W. M. van der Meer, L. A. B. Joosten, J. Walter, J. L. Schultze, T. Roger, E. J. Giamarellos-Bourboulis, A. Schlitzer, and M. G. Netea (2022). “Reply to: ‘Lack of evidence for intergenerational inheritance of immune resistance to infections’”. In: *Nat Immunol* 23.2, pp. 208–209.

Kauffman, C. A. (2007). “Histoplasmosis: a clinical and laboratory update”. In: *Clin Microbiol Rev* 20.1, pp. 115–32.

Kaufman, C. K., C. Mosimann, Z. P. Fan, S. Yang, A. J. Thomas, J. Ablain, J. L. Tan, R. D. Fogley, E. van Rooijen, E. J. Hagedorn, C. Ciarlo, R. M. White, D. A. Matos, A. C. Puller, C. Santoriello, E. C. Liao, R. A. Young, and L. I. Zon (2016). “A zebrafish melanoma model reveals emergence of neural crest identity during melanoma initiation”. In: *Science* 351.6272, aad2197.

Kaufmann, E., M. Landekic, J. Downey, J. Chronopoulos, S. Teimouri Nezhad, K. Tran, D. C. Vinh, L. B. Barreiro, and M. Divangahi (2022). “Lack of evidence for intergenerational inheritance of immune resistance to infections”. In: *Nat Immunol* 23.2, pp. 203–207.

Kaufmann, S. H. (2002). “Protection against tuberculosis: cytokines, T cells, and macrophages”. In: *Ann Rheum Dis* 61 Suppl 2, pp. ii54–8.

— *An early review on the factors influencing granuloma formation and maintenance. This is a very nice snapshot of the state of knowledge at that time and remains an interesting read as the conclusions have been validated and challenged over the last two decades.*

Kaufmann, S. H., C. Lange, M. Rao, K. N. Balaji, M. Lotze, M. Schito, A. I. Zumla, and M. Maeurer (2014). “Progress in tuberculosis vaccine development and host-directed therapies—a state of the art review”. In: *Lancet Respir Med* 2.4, pp. 301–20.

Kawahara, T., E. Kashiwagi, H. Ide, Y. Li, Y. Zheng, H. Ishiguro, and H. Miyamoto (2015a). “The role of NFATc1 in prostate cancer progression: cyclosporine A and

tacrolimus inhibit cell proliferation, migration, and invasion”. In: *Prostate* 75.6, pp. 573–84.

Kawahara, T., E. Kashiwagi, H. Ide, Y. Li, Y. Zheng, Y. Miyamoto, G. J. Netto, H. Ishiguro, and H. Miyamoto (2015b). “Cyclosporine A and tacrolimus inhibit bladder cancer growth through down-regulation of NFATc1”. In: *Oncotarget* 6.3, pp. 1582–93.

Kawai, T., O. Adachi, T. Ogawa, K. Takeda, and S. Akira (1999). “Unresponsiveness of MyD88-deficient mice to endotoxin”. In: *Immunity* 11.1, pp. 115–22.

Kawai, T. and S. Akira (2007). “Signaling to NF-kappaB by Toll-like receptors”. In: *Trends Mol Med* 13.11, pp. 460–9.

- (2009). “The roles of TLRs, RLRs and NLRs in pathogen recognition”. In: *Int Immunol* 21.4, pp. 317–37.
- (2010). “The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors”. In: *Nat Immunol* 11.5, pp. 373–84.

Kawasaki, T. and T. Kawai (2014). “Toll-like receptor signaling pathways”. In: *Front Immunol* 5, p. 461.

Kayagaki, N., M. T. Wong, I. B. Stowe, S. R. Ramani, L. C. Gonzalez, S. Akashi-Takamura, K. Miyake, J. Zhang, W. P. Lee, A. Muszynski, L. S. Forsberg, R. W. Carlson, and V. M. Dixit (2013). “Noncanonical inflammasome activation by intracellular LPS independent of TLR4”. In: *Science* 341.6151, pp. 1246–9.

Kazazi-Hyseni, F., J. H. Beijnen, and J. H. Schellens (2010). “Bevacizumab”. In: *Oncologist* 15.8, pp. 819–25.

Ke, Q., J. Li, J. Ding, M. Ding, L. Wang, B. Liu, M. Costa, and C. Huang (2006). “Essential role of ROS-mediated NFAT activation in TNF-alpha induction by crystalline silica exposure”. In: *Am J Physiol Lung Cell Mol Physiol* 291.2, pp. L257–64.

Keane, J., M. K. Balcewicz-Sablinska, H. G. Remold, G. L. Chupp, B. B. Meek, M. J. Fenton, and H. Kornfeld (1997). “Infection by *Mycobacterium tuberculosis* promotes human alveolar macrophage apoptosis”. In: *Infect Immun* 65.1, pp. 298–304.

Keane, J., H. G. Remold, and H. Kornfeld (2000). “Virulent *Mycobacterium tuberculosis* strains evade apoptosis of infected alveolar macrophages”. In: *J Immunol* 164.4, pp. 2016–20.

Keating, G. M. (2014). “Bevacizumab: a review of its use in advanced cancer”. In: *Drugs* 74.16, pp. 1891–1925.

Kegley, K. M., J. Gephart, G. L. Warren, and G. K. Pavlath (2001). “Altered primary myogenesis in NFATC3(−/−) mice leads to decreased muscle size in the adult”. In: *Dev Biol* 232.1, pp. 115–26.

Kelly, B. and L. A. O’Neill (2015). “Metabolic reprogramming in macrophages and dendritic cells in innate immunity”. In: *Cell Res* 25.7, pp. 771–84.

Kelsh, R. N., M. Brand, Y. J. Jiang, C. P. Heisenberg, S. Lin, P. Haffter, J. Odenthal, M. C. Mullins, F. J. van Eeden, M. Furutani-Seiki, M. Granato, M. Hammer-schmidt, D. A. Kane, R. M. Warga, D. Beuchle, L. Vogelsang, and C. Nusslein-Volhard (1996). “Zebrafish pigmentation mutations and the processes of neural crest development”. In: *Development* 123, pp. 369–89.

Kemp, S. F. and R. F. Lockey (2002). “Anaphylaxis: a review of causes and mechanisms”. In: *J Allergy Clin Immunol* 110.3, pp. 341–8.

Kenyon, A., D. Gavriouchkina, J. Zorman, G. Napolitani, V. Cerundolo, and T. Sauka-Spengler (2017). “Active nuclear transcriptome analysis reveals inflammasome-dependent mechanism for early neutrophil response to *Mycobacterium marinum*”. In: *Sci Rep* 7.1, p. 6505.

Keragala, C. B. and R. L. Medcalf (2021). “Plasminogen: an enigmatic zymogen”. In: *Blood* 137.21, pp. 2881–2889.

Kerrigan, A. M. and G. D. Brown (2011). “Syk-coupled C-type lectins in immunity”. In: *Trends Immunol* 32.4, pp. 151–6.

Kerscher, B., I. M. Dambuza, M. Christofi, D. M. Reid, S. Yamasaki, J. A. Willment, and G. D. Brown (2016a). “Signalling through MyD88 drives surface expression of the mycobacterial receptors MCL (Clecsf8, Clec4d) and Mincle (Clec4e) following microbial stimulation”. In: *Microbes Infect* 18.7-8, pp. 505–9.

Kerscher, B., G. J. Wilson, D. M. Reid, D. Mori, J. A. Taylor, G. S. Besra, S. Yamasaki, J. A. Willment, and G. D. Brown (2016b). “Mycobacterial receptor, Clec4d (CLECSF8, MCL), is coregulated with Mincle and upregulated on mouse myeloid cells following microbial challenge”. In: *Eur J Immunol* 46.2, pp. 381–9.

Keskin, D., J. Kim, V. G. Cooke, C. C. Wu, H. Sugimoto, C. Gu, M. De Palma, R. Kalluri, and V. S. LeBleu (2015). “Targeting vascular pericytes in hypoxic tumors increases lung metastasis via angiopoietin-2”. In: *Cell Rep* 10.7, pp. 1066–81.

Keunen, O., M. Johansson, A. Oudin, M. Sanzey, S. A. Rahim, F. Fack, F. Thorsen, T. Taxt, M. Bartos, R. Jirik, H. Miletic, J. Wang, D. Stieber, L. Stuhr, I. Moen, C. B. Rygh, R. Bjerkvig, and S. P. Niclou (2011). “Anti-VEGF treatment reduces blood supply and increases tumor cell invasion in glioblastoma”. In: *Proc Natl Acad Sci U S A* 108.9, pp. 3749–54.

Keyes, B. E., J. P. Segal, E. Heller, W. H. Lien, C. Y. Chang, X. Guo, D. S. Oristian, D. Zheng, and E. Fuchs (2013). “Nfatc1 orchestrates aging in hair follicle stem cells”. In: *Proc Natl Acad Sci U S A* 110.51, E4950–9.

Khalaf, H., J. Jass, and P. E. Olsson (2013). “The role of calcium, NF-kappaB and NFAT in the regulation of CXCL8 and IL-6 expression in Jurkat T-cells”. In: *Int J Biochem Mol Biol* 4.3, pp. 150–6.

Khameneh, H. J., A. W. Ho, R. Spreafico, H. Derkx, H. Q. Quek, and A. Mortellaro (2017). “The Syk-NFAT-IL-2 Pathway in Dendritic Cells Is Required for Optimal Sterile Immunity Elicited by Alum Adjuvants”. In: *J Immunol* 198.1, pp. 196–204.

Khan, N., J. Downey, J. Sanz, E. Kaufmann, B. Blankenhaus, A. Pacis, E. Pernet, E. Ahmed, S. Cardoso, A. Nijnik, B. Mazer, C. Sassetti, M. A. Behr, M. P. Soares, L. B. Barreiro, and M. Divangahi (2020). “*M. tuberculosis* Reprograms Hematopoietic Stem Cells to Limit Myelopoiesis and Impair Trained Immunity”. In: *Cell* 183.3, 752–770 e22.

Khan, T. A., H. Mazhar, S. Saleha, H. N. Tipu, N. Muhammad, and M. N. Abbas (2016). “Interferon-Gamma Improves Macrophages Function against *M. tuberculosis* in Multidrug-Resistant Tuberculosis Patients”. In: *Chemother Res Pract* 2016, p. 7295390.

Kiani, A., F. J. Garcia-Cozar, I. Habermann, S. Laforsch, T. Aebscher, G. Ehninger, and A. Rao (2001). “Regulation of interferon-gamma gene expression by nuclear factor of activated T cells”. In: *Blood* 98.5, pp. 1480–8.

- Kiani, A., I. Habermann, M. Haase, S. Feldmann, S. Boxberger, M. A. Sanchez-Fernandez, C. Thiede, M. Bornhauser, and G. Ehninger (2004). “Expression and regulation of NFAT (nuclear factors of activated T cells) in human CD34+ cells: down-regulation upon myeloid differentiation”. In: *J Leukoc Biol* 76.5, pp. 1057–65.
- Kiani, A., J. P. Viola, A. H. Lichtman, and A. Rao (1997). “Down-regulation of IL-4 gene transcription and control of Th2 cell differentiation by a mechanism involving NFAT1”. In: *Immunity* 7.6, pp. 849–60.
- Kieser, K. J. and E. J. Rubin (2014). “How sisters grow apart: mycobacterial growth and division”. In: *Nat Rev Microbiol* 12.8, pp. 550–62.
- Kim, B. H., A. R. Shenoy, P. Kumar, C. J. Bradfield, and J. D. MacMicking (2012). “IFN-inducible GTPases in host cell defense”. In: *Cell Host Microbe* 12.4, pp. 432–44.
- Kim, D. H., J. Kim, J. C. Marques, A. Grama, D. G. C. Hildebrand, W. Gu, J. M. Li, and D. N. Robson (2017). “Pan-neuronal calcium imaging with cellular resolution in freely swimming zebrafish”. In: *Nat Methods* 14.11, pp. 1107–1114.
- Kim, H. B., M. Kong, T. M. Kim, Y. H. Suh, W. H. Kim, J. H. Lim, J. H. Song, and M. H. Jung (2006). “NFATc4 and ATF3 negatively regulate adiponectin gene expression in 3T3-L1 adipocytes”. In: *Diabetes* 55.5, pp. 1342–52.
- Kim, H. B., A. Kumar, L. Wang, G. H. Liu, S. R. Keller, Jr. Lawrence J. C., B. N. Finck, and T. E. Harris (2010a). “Lipin 1 represses NFATc4 transcriptional activity in adipocytes to inhibit secretion of inflammatory factors”. In: *Mol Cell Biol* 30.12, pp. 3126–39.
- Kim, J. H. and N. Kim (2014). “Regulation of NFATc1 in Osteoclast Differentiation”. In: *J Bone Metab* 21.4, pp. 233–41.
- Kim, J. H., S. R. Lee, L. H. Li, H. J. Park, J. H. Park, K. Y. Lee, M. K. Kim, B. A. Shin, and S. Y. Choi (2011). “High cleavage efficiency of a 2A peptide derived from porcine teschovirus-1 in human cell lines, zebrafish and mice”. In: *PLoS One* 6.4, e18556.
- Kim, K., S. H. Lee, J. Ha Kim, Y. Choi, and N. Kim (2008). “NFATc1 induces osteoclast fusion via up-regulation of Atp6v0d2 and the dendritic cell-specific transmembrane protein (DC-STAMP)”. In: *Mol Endocrinol* 22.1, pp. 176–85.

- Kim, M. J., H. C. Wainwright, M. Locketz, L. G. Bekker, G. B. Walther, C. Ditzich, A. Visser, W. Wang, F. F. Hsu, U. Wiehart, L. Tsanova, G. Kaplan, and D. G. Russell (2010b). “Caseation of human tuberculosis granulomas correlates with elevated host lipid metabolism”. In: *EMBO Mol Med* 2.7, pp. 258–74.
- Kim, U. K., E. Jorgenson, H. Coon, M. Leppert, N. Risch, and D. Drayna (2003). “Positional cloning of the human quantitative trait locus underlying taste sensitivity to phenylthiocarbamide”. In: *Science* 299.5610, pp. 1221–5.
- Kimmel, C. B. (1989). “Genetics and early development of zebrafish”. In: *Trends Genet* 5.8, pp. 283–8.
- Kimmel, C. B., W. W. Ballard, S. R. Kimmel, B. Ullmann, and T. F. Schilling (1995). “Stages of embryonic development of the zebrafish”. In: *Dev Dyn* 203.3, pp. 253–310.
- Kimmel, C. B., D. S. Sepich, and B. Trevarrow (1988). “Development of segmentation in zebrafish”. In: *Development* 104 Suppl, pp. 197–207.
- Kinoshita, S., L. Su, M. Amano, L. A. Timmerman, H. Kaneshima, and G. P. Nolan (1997). “The T cell activation factor NF-ATc positively regulates HIV-1 replication and gene expression in T cells”. In: *Immunity* 6.3, pp. 235–44.
- Kiran, D., B. K. Podell, M. Chambers, and R. J. Basaraba (2016). “Host-directed therapy targeting the *Mycobacterium tuberculosis* granuloma: a review”. In: *Semin Immunopathol* 38.2, pp. 167–83.
- Kiriakidis, S., E. Andreakos, C. Monaco, B. Foxwell, M. Feldmann, and E. Paleolog (2003). “VEGF expression in human macrophages is NF-kappaB-dependent: studies using adenoviruses expressing the endogenous NF-kappaB inhibitor IkappaBalpha and a kinase-defective form of the IkappaB kinase 2”. In: *J Cell Sci* 116.Pt 4, pp. 665–74.
- Kiss, M., S. Van Gassen, K. Movahedi, Y. Saeys, and D. Laoui (2018). “Myeloid cell heterogeneity in cancer: not a single cell alike”. In: *Cell Immunol* 330, pp. 188–201.
– *A useful review of the known heterogeneity in myeloid cell populations and informative as to how those specifically connect to cancer. It is useful to think about how those findings translate to the context of the granuloma where such heterogeneity has become ever more obvious but the consequences remain poorly understood.*

Kitamura, N. and O. Kaminuma (2021). "Isoform-Selective NFAT Inhibitor: Potential Usefulness and Development". In: *Int J Mol Sci* 22.5.

Klare, I., A. C. Rodloff, J. Wagner, W. Witte, and R. Hakenbeck (1992). "Overproduction of a penicillin-binding protein is not the only mechanism of penicillin resistance in *Enterococcus faecium*". In: *Antimicrob Agents Chemother* 36.4, pp. 783–7.

Klein, M., S. Klein-Hessling, A. Palmetshofer, E. Serfling, C. Tertilt, T. Bopp, V. Heib, M. Becker, C. Taube, H. Schild, E. Schmitt, and M. Stassen (2006). "Specific and redundant roles for NFAT transcription factors in the expression of mast cell-derived cytokines". In: *J Immunol* 177.10, pp. 6667–74.

Klein-Hessling, S., K. Muhammad, M. Klein, T. Pusch, R. Rudolf, J. Floter, M. Qureischi, A. Beilhack, M. Vaeth, C. Kummerow, C. Backes, R. Schoppmeyer, U. Hahn, M. Hoth, T. Bopp, F. Berberich-Siebelt, A. Patra, A. Avots, N. Muller, A. Schulze, and E. Serfling (2017). "NFATc1 controls the cytotoxicity of CD8(+) T cells". In: *Nat Commun* 8.1, p. 511.

Knapik, E. W., A. Goodman, O. S. Atkinson, C. T. Roberts, M. Shiozawa, C. U. Sim, S. Weksler-Zangen, M. R. Trolliet, C. Futrell, B. A. Innes, G. Koike, M. G. McLaughlin, L. Pierre, J. S. Simon, E. Vilallonga, M. Roy, P. W. Chiang, M. C. Fishman, W. Driever, and H. J. Jacob (1996). "A reference cross DNA panel for zebrafish (*Danio rerio*) anchored with simple sequence length polymorphisms". In: *Development* 123, pp. 451–60.

Knight, M., J. Braverman, K. Asfaha, K. Gronert, and S. Stanley (2018). "Lipid droplet formation in *Mycobacterium tuberculosis* infected macrophages requires IFN-gamma/HIF-1alpha signaling and supports host defense". In: *PLoS Pathog* 14.1, e1006874.

Koch, Robert (1876). "Untersuchungen über Bakterien: V. Die Ätiologie der Milzbrand-Krankheit, begründet auf die Entwicklungsgeschichte des *Bacillus anthracis*". In: *Cohns Beiträge zur Biologie der Pflanzen* 2.2, pp. 277–310.

– (1882). "Die Ätiologie der Tuberkulose". In: 19.

Koch, S. and L. Claesson-Welsh (2012). "Signal transduction by vascular endothelial growth factor receptors". In: *Cold Spring Harb Perspect Med* 2.7, a006502.

- Kochakpour, N. and P. B. Moens (2008). “Sex-specific crossover patterns in Zebrafish (*Danio rerio*)”. In: *Heredity (Edinb)* 100.5, pp. 489–95.
- Koga, T., M. Inui, K. Inoue, S. Kim, A. Suematsu, E. Kobayashi, T. Iwata, H. Ohnishi, T. Matozaki, T. Kodama, T. Taniguchi, H. Takayanagi, and T. Takai (2004). “Costimulatory signals mediated by the ITAM motif cooperate with RANKL for bone homeostasis”. In: *Nature* 428.6984, pp. 758–63.
- Kokel, D., J. Bryan, C. Laggner, R. White, C. Y. Cheung, R. Mateus, D. Healey, S. Kim, A. A. Werdich, S. J. Haggarty, C. A. Macrae, B. Shoichet, and R. T. Peterson (2010). “Rapid behavior-based identification of neuroactive small molecules in the zebrafish”. In: *Nat Chem Biol* 6.3, pp. 231–237.
- Kokel, D. and R. T. Peterson (2011). “Using the zebrafish photomotor response for psychotropic drug screening”. In: *Methods Cell Biol* 105, pp. 517–24.
- Kolliniati, O., E. Ieronymaki, E. Vergadi, and C. Tsatsanis (2022). “Metabolic Regulation of Macrophage Activation”. In: *J Innate Immun* 14.1, pp. 51–68.
- Kontarakis, Z. and D. Y. R. Stainier (2020). “Genetics in Light of Transcriptional Adaptation”. In: *Trends Genet* 36.12, pp. 926–935.
- Kontoyiannis, D. P. (2010). “Manipulation of host angioneogenesis: A critical link for understanding the pathogenesis of invasive mold infections?” In: *Virulence* 1.3, pp. 192–6.
- Koppel, E. A., I. S. Ludwig, M. S. Hernandez, T. L. Lowary, R. R. Gadikota, A. B. Tuzikov, C. M. Vandenbroucke-Grauls, Y. van Kooyk, B. J. Appelman, and T. B. Geijtenbeek (2004). “Identification of the mycobacterial carbohydrate structure that binds the C-type lectins DC-SIGN, L-SIGN and SIGNR1”. In: *Immunobiology* 209.1-2, pp. 117–27.
- Korbut, R., F. Mehrdana, P. W. Kania, M. H. Larsen, D. Frees, I. Dalgaard, and Lv Jorgensen (2016). “Antigen Uptake during Different Life Stages of Zebrafish (*Danio rerio*) Using a GFP-Tagged *Yersinia ruckeri*”. In: *PLoS One* 11.7, e0158968.
- Korf, J., A. Stoltz, J. Verschoor, P. De Baetselier, and J. Grootenhuis (2005). “The *Mycobacterium tuberculosis* cell wall component mycolic acid elicits pathogen-associated host innate immune responses”. In: *Eur J Immunol* 35.3, pp. 890–900.

- Kossack, M. E. and B. W. Draper (2019). “Genetic regulation of sex determination and maintenance in zebrafish (*Danio rerio*)”. In: *Curr Top Dev Biol* 134, pp. 119–149.
- Koul, A., T. Herget, B. Klebl, and A. Ullrich (2004). “Interplay between mycobacteria and host signalling pathways”. In: *Nat Rev Microbiol* 2.3, pp. 189–202.
- Kozma, M., A. Meszaros, A. Nyul-Toth, K. Molnar, L. Costea, Z. Hernadi, C. Fazakas, A. E. Farkas, I. Wilhelm, and I. A. Krizbai (2021). “Cerebral Pericytes and Endothelial Cells Communicate through Inflammasome-Dependent Signals”. In: *Int J Mol Sci* 22.11.
- Kreier, F. (2022). “Just 14 cases: Guinea worm disease nears eradication”. In: *Nature*.
- Kremer, L., Y. Guerardel, S. S. Gurcha, C. Locht, and G. S. Besra (2002). “Temperature-induced changes in the cell-wall components of *Mycobacterium thermoresistibile*”. In: *Microbiology (Reading)* 148.Pt 10, pp. 3145–3154.
- Krishnamurty, A. T. and S. J. Turley (2020). “Lymph node stromal cells: cartographers of the immune system”. In: *Nat Immunol* 21.4, pp. 369–380.
- Kroidl, I., M. I. M. Ahmed, S. Horn, C. Polyak, A. Esber, A. Parikh, L. A. Eller, H. Kibuuka, M. Semwogerere, B. Mwesigwa, P. Naluyima, J. M. Kasumba, J. Maswai, J. Owuoth, V. Sing’oei, E. Rono, R. Loose, M. Hoelscher, J. Ake, C. Geldmacher, and Africos Study Group (2022). “Assessment of tuberculosis disease activity in people infected with *Mycobacterium tuberculosis* and living with HIV: A longitudinal cohort study”. In: *EClinicalMedicine* 49, p. 101470.
- Krokowski, S., D. Lobato-Marquez, A. Chastanet, P. M. Pereira, D. Angelis, D. Galea, G. Larrouy-Maumus, R. Henriques, E. T. Spiliotis, R. Carballido-Lopez, and S. Mostowy (2018). “Septins Recognize and Entrap Dividing Bacterial Cells for Delivery to Lysosomes”. In: *Cell Host Microbe* 24.6, 866–874 e4.
- Kroll, F., G. T. Powell, M. Ghosh, G. Gestri, P. Antinucci, T. J. Hearn, H. Tunbak, S. Lim, H. W. Dennis, J. M. Fernandez, D. Whitmore, E. Dreosti, S. W. Wilson, E. J. Hoffman, and J. Rihel (2021). “A simple and effective F0 knockout method for rapid screening of behaviour and other complex phenotypes”. In: *Elife* 10.
- Krueger, J., D. Liu, K. Scholz, A. Zimmer, Y. Shi, C. Klein, A. Siekmann, S. Schulte-Merker, M. Cudmore, A. Ahmed, and F. le Noble (2011). “Flt1 acts as a negative

regulator of tip cell formation and branching morphogenesis in the zebrafish embryo". In: *Development* 138.10, pp. 2111–20.

Kruize, Z. and N. A. Kootstra (2019). "The Role of Macrophages in HIV-1 Persistence and Pathogenesis". In: *Front Microbiol* 10, p. 2828.

Krzywinska, E., J. Krzywinski, and J. S. Schorey (2004). "Naturally occurring horizontal gene transfer and homologous recombination in *Mycobacterium*". In: *Microbiology (Reading)* 150.Pt 6, pp. 1707–1712.

Kuchler, A. M., E. Gjini, J. Peterson-Maduro, B. Cancilla, H. Wolburg, and S. Schulte-Merker (2006). "Development of the zebrafish lymphatic system requires VEGFC signaling". In: *Curr Biol* 16.12, pp. 1244–8.

Kugler, E. C., J. Frost, V. Silva, K. Plant, K. Chhabria, T. J. A. Chico, and P. A. Armitage (2022). "Zebrafish vascular quantification: a tool for quantification of three-dimensional zebrafish cerebrovascular architecture by automated image analysis". In: *Development* 149.3.

Kuhlicke, J., J. S. Frick, J. C. Morote-Garcia, P. Rosenberger, and H. K. Eltzschig (2007). "Hypoxia inducible factor (HIF)-1 coordinates induction of Toll-like receptors TLR2 and TLR6 during hypoxia". In: *PLoS One* 2.12, e1364.

Kujawski, S., W. Lin, F. Kitte, M. Bormel, S. Fuchs, G. Arulmozhivarman, S. Vogt, D. Theil, Y. Zhang, and C. L. Antos (2014). "Calcineurin regulates coordinated outgrowth of zebrafish regenerating fins". In: *Dev Cell* 28.5, pp. 573–87.

– A recent study but still one of the earliest to look at the effects of calcineurin inhibition in the zebrafish. This work looked at fin regeneration and found that tacrolimus treatment induced enhanced fin regrowth, suggesting that calcineurin is important for regulating regeneration and that inhibition may be one method to enhance pro-growth responses.

Kulkarni, R. M., J. M. Greenberg, and A. L. Akeson (2009). "NFATc1 regulates lymphatic endothelial development". In: *Mech Dev* 126.5-6, pp. 350–65.

Kumar, R., J. F. DuMond, S. H. Khan, E. B. Thompson, Y. He, M. B. Burg, and J. D. Ferraris (2020a). "NFAT5, which protects against hypertonicity, is activated by that stress via structuring of its intrinsically disordered domain". In: *Proc Natl Acad Sci U S A* 117.33, pp. 20292–20297.

- Kumar, S., J. T. Koehn, M. Gonzalez-Juarrero, D. C. Crans, and D. C. Crick (2020b). “*Mycobacterium tuberculosis* Survival in J774A.1 Cells Is Dependent on MenJ Moonlighting Activity, Not Its Enzymatic Activity”. In: *ACS Infect Dis* 6.10, pp. 2661–2671.
- Kuri, P., N. L. Schieber, T. Thumberger, J. Wittbrodt, Y. Schwab, and M. Leptin (2017). “Dynamics of *in vivo* ASC speck formation”. In: *J Cell Biol* 216.9, pp. 2891–2909.
- Kurita, K., S. M. Burgess, and N. Sakai (2004). “Transgenic zebrafish produced by retroviral infection of *in vitro*-cultured sperm”. In: *Proc Natl Acad Sci U S A* 101.5, pp. 1263–7.
- Kusner, D. J. and J. A. Barton (2001). “ATP stimulates human macrophages to kill intracellular virulent *Mycobacterium tuberculosis* via calcium-dependent phagosome-lysosome fusion”. In: *J Immunol* 167.6, pp. 3308–15.
- Kutsch, M., L. Sistemich, C. F. Lesser, M. B. Goldberg, C. Herrmann, and J. Coers (2020). “Direct binding of polymeric GBP1 to LPS disrupts bacterial cell envelope functions”. In: *Embo J* 39.13, e104926.
- Kwan, K. M., E. Fujimoto, C. Grabher, B. D. Mangum, M. E. Hardy, D. S. Campbell, J. M. Parant, H. J. Yost, J. P. Kanki, and C. B. Chien (2007). “The Tol2kit: a multisite gateway-based construction kit for Tol2 transposon transgenesis constructs”. In: *Dev Dyn* 236.11, pp. 3088–99.**
- Kwon, H. S., D. R. Kim, E. G. Yang, Y. K. Park, H. C. Ahn, S. J. Min, and D. R. Ahn (2012). “Inhibition of VEGF transcription through blockade of the hypoxia inducible factor-1alpha-p300 interaction by a small molecule”. In: *Bioorg Med Chem Lett* 22.16, pp. 5249–52.
- Kwon, V., P. Cai, C. T. Dixon, V. Hamlin, C. G. Spencer, A. M. Rojas, M. Hamilton, and C. E. Shiao (2022). “Peripheral NOD-like receptor deficient inflammatory macrophages trigger neutrophil infiltration into the brain disrupting daytime locomotion”. In: *Commun Biol* 5.1, p. 464.
- La Porta, S., L. Roth, M. Singhal, C. Mogler, C. Spegg, B. Schieb, X. Qu, R. H. Adams, H. S. Baldwin, S. Savant, and H. G. Augustin (2018). “Endothelial Tie1-mediated angiogenesis and vascular normalization promote tumor progression and metastasis”. In: *J Clin Invest* 128.2, pp. 834–845.

- Lagay, F. (2002). "The legacy of humoral medicine". In: *Virtual Mentor* 4.7.
- Lai, Y. S., R. Wahyuningtyas, S. P. Aui, and K. T. Chang (2019). "Autocrine VEGF signalling on M2 macrophages regulates PD-L1 expression for immunomodulation of T cells". In: *J Cell Mol Med* 23.2, pp. 1257–1267.
- Lamagna, C., M. Aurrand-Lions, and B. A. Imhof (2006). "Dual role of macrophages in tumor growth and angiogenesis". In: *J Leukoc Biol* 80.4, pp. 705–13.
- Lamont, E. A. and A. D. Baughn (2019). "Impact of the host environment on the antitubercular action of pyrazinamide". In: *EBioMedicine* 49, pp. 374–380.
- Lamont, E. A., N. A. Dillon, and A. D. Baughn (2020). "The Bewildering Antitubercular Action of Pyrazinamide". In: *Microbiol Mol Biol Rev* 84.2.
- Lang, R. (2013). "Recognition of the mycobacterial cord factor by Minclle: relevance for granuloma formation and resistance to tuberculosis". In: *Front Immunol* 4, p. 5.
- Langenau, D. M., D. Traver, A. A. Ferrando, J. L. Kutok, J. C. Aster, J. P. Kanki, S. Lin, E. Prochownik, N. S. Trede, L. I. Zon, and A. T. Look (2003). "Myc-induced T cell leukemia in transgenic zebrafish". In: *Science* 299.5608, pp. 887–90.
- Langston, P. K., M. Shibata, and T. Horng (2017). "Metabolism Supports Macrophage Activation". In: *Front Immunol* 8, p. 61.
- Larsen, M. H., K. Lacourciere, T. M. Parker, A. Kraigsley, J. M. Achkar, L. B. Adams, K. M. Dupnik, L. Hall-Stoodley, T. Hartman, C. Kanipe, S. L. Kurtz, M. A. Miller, L. C. M. Salvador, J. S. Spencer, and R. T. Robinson (2020). "The Many Hosts of Mycobacteria 8 (MHM8): A conference report". In: *Tuberculosis (Edinb)* 121, p. 101914.
- Latty, S. L., J. Sakai, L. Hopkins, B. Verstak, T. Paramo, N. A. Berglund, E. Cammarota, P. Cicuta, N. J. Gay, P. J. Bond, D. Klenerman, and C. E. Bryant (2018). "Activation of Toll-like receptors nucleates assembly of the MyDDosome signaling hub". In: *Elife* 7.
- Laval, T., L. Chaumont, and C. Demangel (2021). "Not too fat to fight: The emerging role of macrophage fatty acid metabolism in immunity to *Mycobacterium tuberculosis*". In: *Immunol Rev* 301.1, pp. 84–97.

- Lavin, Y., A. Mortha, A. Rahman, and M. Merad (2015). “Regulation of macrophage development and function in peripheral tissues”. In: *Nat Rev Immunol* 15.12, pp. 731–44.
- Laviron, M. and A. Boissonnas (2019). “Ontogeny of Tumor-Associated Macrophages”. In: *Front Immunol* 10, p. 1799.
- Lawir, D. F., I. Hess, K. Sikora, N. Iwanami, I. Siamishi, M. Schorpp, and T. Boehm (2019). “Evolutionary transition from degenerate to nonredundant cytokine signaling networks supporting intrathymic T cell development”. In: *Proc Natl Acad Sci U S A*.
- Lawson, N. D. and B. M. Weinstein (2002). “*In vivo* imaging of embryonic vascular development using transgenic zebrafish”. In: *Dev Biol* 248.2, pp. 307–18.
- Lay, G., Y. Poquet, P. Salek-Peyron, M. P. Puissegur, C. Botanch, H. Bon, F. Levillain, J. L. Duteyrat, J. F. Emile, and F. Altare (2007). “Langhans giant cells from *M. tuberculosis*-induced human granulomas cannot mediate mycobacterial uptake”. In: *J Pathol* 211.1, pp. 76–85.
- Layne, J. J., M. E. Werner, D. C. Hill-Eubanks, and M. T. Nelson (2008). “NFATc3 regulates BK channel function in murine urinary bladder smooth muscle”. In: *Am J Physiol Cell Physiol* 295.3, pp. C611–23.
- Lazarevic, V., A. J. Zullo, M. N. Schweitzer, T. L. Staton, E. M. Gallo, G. R. Crabtree, and L. H. Glimcher (2009). “The gene encoding early growth response 2, a target of the transcription factor NFAT, is required for the development and maturation of natural killer T cells”. In: *Nat Immunol* 10.3, pp. 306–13.
- Le Guen, L., T. Karpanen, D. Schulte, N. C. Harris, K. Koltowska, G. Roukens, N. I. Bower, A. van Impel, S. A. Stacker, M. G. Achen, S. Schulte-Merker, and B. M. Hogan (2014). “Ccbe1 regulates Vegfc-mediated induction of Vegfr3 signaling during embryonic lymphangiogenesis”. In: *Development* 141.6, pp. 1239–49.
- Lebeer, S., J. Vanderleyden, and S. C. De Keersmaecker (2010). “Host interactions of probiotic bacterial surface molecules: comparison with commensals and pathogens”. In: *Nat Rev Microbiol* 8.3, pp. 171–84.
- Lee, C. G., K. H. Kang, J. S. So, H. K. Kwon, J. S. Son, M. K. Song, A. Sahoo, H. J. Yi, K. C. Hwang, T. Matsuyama, K. Yui, and S. H. Im (2009). “A distal cis-regulatory

element, CNS-9, controls NFAT1 and IRF4-mediated IL-10 gene activation in T helper cells". In: *Mol Immunol* 46.4, pp. 613–21.

Lee, J., S. G. Lee, K. K. Kim, Y. J. Lim, J. A. Choi, S. N. Cho, C. Park, and C. H. Song (2019). "Characterisation of genes differentially expressed in macrophages by virulent and attenuated *Mycobacterium tuberculosis* through RNA-Seq analysis". In: *Sci Rep* 9.1, p. 4027.

Lee, J. H., D. H. Bhang, A. Beede, T. L. Huang, B. R. Stripp, K. D. Bloch, A. J. Waggers, Y. H. Tseng, S. Ryeom, and C. F. Kim (2014). "Lung stem cell differentiation in mice directed by endothelial cells via a BMP4-NFATc1-thrombospondin-1 axis". In: *Cell* 156.3, pp. 440–55.

Lee, W. B., J. S. Kang, J. J. Yan, M. S. Lee, B. Y. Jeon, S. N. Cho, and Y. J. Kim (2012). "Neutrophils Promote Mycobacterial Trehalose Dimycolate-Induced Lung Inflammation via the Mincle Pathway". In: *PLoS Pathog* 8.4, e1002614.

Leemans, J. C., N. P. Juffermans, S. Florquin, N. van Rooijen, M. J. Vervoordeldonk, A. Verbon, S. J. van Deventer, and T. van der Poll (2001). "Depletion of alveolar macrophages exerts protective effects in pulmonary tuberculosis in mice". In: *J Immunol* 166.7, pp. 4604–11.

– *The depletion of alveolar macrophages was shown to be protective against tuberculosis disease, suggesting that these susceptible cell types were actively inhibiting protective responses during infection and that mycobacteria had uncovered exquisite mechanisms to use macrophages to grow in excess of their ability to outside of macrophages, making them properly macrophage-trophic pathogens.*

Leemans, J. C., T. Thepen, S. Weijer, S. Florquin, N. van Rooijen, J. G. van de Winkel, and T. van der Poll (2005). "Macrophages play a dual role during pulmonary tuberculosis in mice". In: *J Infect Dis* 191.1, pp. 65–74.

– *Given Leemans' previous findings on the maladaptive role of alveolar macrophages, they went on to investigate other macrophage populations and found that non-selective ablation of macrophages improved disease outcomes while specific ablation of activated macrophages resulted in worsened outcomes. They propose a dual role where quiescent macrophages are harmful in disease but, after activation, macrophages are able to provide some protection against disease.*

- Lehmann, R. and C. Nusslein-Volhard (1986). "Abdominal segmentation, pole cell formation, and embryonic polarity require the localized activity of oskar, a maternal gene in *Drosophila*". In: *Cell* 47.1, pp. 141–52.
- Lei, B., C. J. Wei, and S. C. Tu (2000). "Action mechanism of antitubercular isoniazid. Activation by *Mycobacterium tuberculosis* KatG, isolation, and characterization of inha inhibitor". In: *J Biol Chem* 275.4, pp. 2520–6.
- Leibert, E., M. Danckers, and W. N. Rom (2014). "New drugs to treat multidrug-resistant tuberculosis: the case for bedaquiline". In: *Ther Clin Risk Manag* 10, pp. 597–602.
- Leibovich, S. J., P. J. Polverini, H. M. Shepard, D. M. Wiseman, V. Shively, and N. Nuseir (1987). "Macrophage-induced angiogenesis is mediated by tumour necrosis factor-alpha". In: *Nature* 329.6140, pp. 630–2.
- Lele, Z. and P. H. Krone (1996). "The zebrafish as a model system in developmental, toxicological and transgenic research". In: *Biotechnol Adv* 14.1, pp. 57–72.
- Lenaerts, A., 3rd Barry C. E., and V. Dartois (2015). "Heterogeneity in tuberculosis pathology, microenvironments and therapeutic responses". In: *Immunol Rev* 264.1, pp. 288–307.
- Lerner, T. R., C. J. Queval, A. Fearns, U. Repnik, G. Griffiths, and M. G. Gutierrez (2018). "Phthiocerol dimycocerosates promote access to the cytosol and intracellular burden of *Mycobacterium tuberculosis* in lymphatic endothelial cells". In: *BMC Biol* 16.1, p. 1.
- Lerner, T. R., C. J. Queval, R. P. Lai, M. R. Russell, A. Fearn, D. J. Greenwood, L. Collinson, R. J. Wilkinson, and M. G. Gutierrez (2020). "*Mycobacterium tuberculosis* cords within lymphatic endothelial cells to evade host immunity". In: *JCI Insight* 5.10.
- Lerner, T. R., C. de Souza Carvalho-Wodarz, U. Repnik, M. R. Russell, S. Borel, C. R. Diedrich, M. Rohde, H. Wainwright, L. M. Collinson, R. J. Wilkinson, G. Griffiths, and M. G. Gutierrez (2016). "Lymphatic endothelial cells are a replicative niche for *Mycobacterium tuberculosis*". In: *J Clin Invest* 126.3, pp. 1093–108.
- Lerut, J., J. Mathys, C. Verbaandert, S. Talpe, O. Ciccarelli, J. Lemaire, E. Bonaccorsi-Riani, V. Vanthuyne, N. Hetsch, F. Roggen, C. D. Reyck, P. Goffette,

D. Latinne, G. Orlando, J. Rahier, C. Sempoux, P. Wallemacq, P. F. Laterre, and P. Gianello (2008). “Tacrolimus monotherapy in liver transplantation: one-year results of a prospective, randomized, double-blind, placebo-controlled study”. In: *Ann Surg* 248.6, pp. 956–67.

Lesley, R. and L. Ramakrishnan (2008). “Insights into early mycobacterial pathogenesis from the zebrafish”. In: *Curr Opin Microbiol* 11.3, pp. 277–83.

Leung, D. W., G. Cachianes, W. J. Kuang, D. V. Goeddel, and N. Ferrara (1989). “Vascular endothelial growth factor is a secreted angiogenic mitogen”. In: *Science* 246.4935, pp. 1306–9.

Lev, S., D. Desmarini, M. Chayakulkeeree, T. C. Sorrell, and J. T. Djordjevic (2012). “The Crz1/Sp1 transcription factor of *Cryptococcus neoformans* is activated by calcineurin and regulates cell wall integrity”. In: *PLoS One* 7.12, e51403.

Levin-Reisman, I., I. Ronin, O. Gefen, I. Braniss, N. Shores, and N. Q. Balaban (2017). “Antibiotic tolerance facilitates the evolution of resistance”. In: *Science* 355.6327, pp. 826–830.

Levraud, J. P., N. Palha, C. Langevin, and P. Boudinot (2014). “Through the looking glass: witnessing host-virus interplay in zebrafish”. In: *Trends Microbiol* 22.9, pp. 490–7.

Lewis, A. and P. M. Elks (2019). “Hypoxia Induces Macrophage tnfa Expression via Cyclooxygenase and Prostaglandin E2 *in vivo*”. In: *Front Immunol* 10, p. 2321.

Lewis, C. E., M. De Palma, and L. Naldini (2007). “Tie2-expressing monocytes and tumor angiogenesis: regulation by hypoxia and angiopoietin-2”. In: *Cancer Res* 67.18, pp. 8429–32.

Ley, K. (2017). “M1 Means Kill; M2 Means Heal”. In: *J Immunol* 199.7, pp. 2191–2193.

Li, D., X. P. Li, H. X. Wang, Q. Y. Shen, X. P. Li, L. Wen, X. J. Qin, Q. L. Jia, H. F. Kung, and Y. Peng (2012). “VEGF induces angiogenesis in a zebrafish embryo glioma model established by transplantation of human glioma cells”. In: *Oncol Rep* 28.3, pp. 937–42.

- Li, L., K. M. Candelario, K. Thomas, R. Wang, K. Wright, A. Messier, and L. A. Cunningham (2014). “Hypoxia inducible factor-1alpha (HIF-1alpha) is required for neural stem cell maintenance and vascular stability in the adult mouse SVZ”. In: *J Neurosci* 34.50, pp. 16713–9.
- Li, M., X. Z. Fang, Y. F. Zheng, Y. B. Xie, X. D. Ma, X. T. Liu, Y. Xia, and D. H. Shao (2019). “Transient receptor potential vanilloid 4 is a critical mediator in LPS mediated inflammation by mediating calcineurin/NFATc3 signaling”. In: *Biochem Biophys Res Commun* 513.4, pp. 1005–1012.
- Li, P., Z. Cong, Y. Qiang, L. Xiong, L. Tang, Y. Zhang, H. Wu, J. Yi, H. Jing, D. Li, and Y. Shen (2018a). “Clinical significance of CCBE1 expression in lung cancer”. In: *Mol Med Rep* 17.2, pp. 2107–2112.
- Li, Q., Y. Xie, Z. Cui, H. Huang, C. Yang, B. Yuan, P. Shen, and C. Shi (2021). “Activation of hypoxia-inducible factor 1 (Hif-1) enhanced bactericidal effects of macrophages to *Mycobacterium tuberculosis*”. In: *Tuberculosis (Edinb)* 126, p. 102044.
- Li, Y., S. Ali, J. Clarke, and S. Cha (2017). “Bevacizumab in Recurrent Glioma: Patterns of Treatment Failure and Implications”. In: *Brain Tumor Res Treat* 5.1, pp. 1–9.
- Li, Z., X. Zhang, Z. Guo, Y. Zhong, P. Wang, J. Li, Z. Li, and P. Liu (2018b). “SIRT6 Suppresses NFATc4 Expression and Activation in Cardiomyocyte Hypertrophy”. In: *Front Pharmacol* 9, p. 1519.
- Liang, D., J. R. Chang, A. J. Chin, A. Smith, C. Kelly, E. S. Weinberg, and R. Ge (2001). “The role of vascular endothelial growth factor (VEGF) in vasculogenesis, angiogenesis, and hematopoiesis in zebrafish development”. In: *Mech Dev* 108.1-2, pp. 29–43.
- Liang, L., H. Liu, J. Yue, L. R. Liu, M. Han, L. L. Luo, Y. L. Zhao, and H. Xiao (2017). “Association of Single-Nucleotide Polymorphism in the Hepcidin Promoter Gene with Susceptibility to Extrapulmonary Tuberculosis”. In: *Genet Test Mol Biomarkers* 21.6, pp. 351–356.
- Liao, W., B. W. Bisgrove, H. Sawyer, B. Hug, B. Bell, K. Peters, D. J. Grunwald, and D. Y. Stainier (1997). “The zebrafish gene cloche acts upstream of a flk-1 homologue to regulate endothelial cell differentiation”. In: *Development* 124.2, pp. 381–9.

Liao, W., C. Y. Ho, Y. L. Yan, J. Postlethwait, and D. Y. Stainier (2000). “Hhex and scl function in parallel to regulate early endothelial and blood differentiation in zebrafish”. In: *Development* 127.20, pp. 4303–13.

Lima, V. M., V. L. Bonato, K. M. Lima, S. A. Dos Santos, R. R. Dos Santos, E. D. Goncalves, L. H. Faccioli, I. T. Brandao, J. M. Rodrigues-Junior, and C. L. Silva (2001). “Role of trehalose dimycolate in recruitment of cells and modulation of production of cytokines and NO in tuberculosis”. In: *Infect Immun* 69.9, pp. 5305–12.

Lin, J., J. Ren, D. S. Gao, Y. Dai, and L. Yu (2021). “The Emerging Application of Itaconate: Promising Molecular Targets and Therapeutic Opportunities”. In: *Front Chem* 9, p. 669308.

Lin, P. L., C. B. Ford, M. T. Coleman, A. J. Myers, R. Gawande, T. Ioerger, J. Sacchettini, S. M. Fortune, and J. L. Flynn (2014). “Sterilization of granulomas is common in active and latent tuberculosis despite within-host variability in bacterial killing”. In: *Nat Med* 20.1, pp. 75–9.

Lin, P. L., S. Pawar, A. Myers, A. Pegu, C. Fuhrman, T. A. Reinhart, S. V. Capuano, E. Klein, and J. L. Flynn (2006). “Early events in *Mycobacterium tuberculosis* infection in cynomolgus macaques”. In: *Infect Immun* 74.7, pp. 3790–803.

Lin, X. and J. Heitman (2006). “The biology of the *Cryptococcus neoformans* species complex”. In: *Annu Rev Microbiol* 60, pp. 69–105.

Lin, Z., I. Murtaza, K. Wang, J. Jiao, J. Gao, and P. F. Li (2009). “miR-23a functions downstream of NFATc3 to regulate cardiac hypertrophy”. In: *Proc Natl Acad Sci U S A* 106.29, pp. 12103–8.

Linehan, E. and D. C. Fitzgerald (2015). “Ageing and the immune system: focus on macrophages”. In: *Eur J Microbiol Immunol (Bp)* 5.1, pp. 14–24.

Linell, F. and A. Norden (1954). “*Mycobacterium balnei*, a new acid-fast bacillus occurring in swimming pools and capable of producing skin lesions in humans”. In: *Acta Tuberc Scand Suppl* 33, pp. 1–84.

Liongue, C., C. J. Hall, B. A. O’Connell, P. Crosier, and A. C. Ward (2009). “Zebrafish granulocyte colony-stimulating factor receptor signaling promotes myelopoiesis and myeloid cell migration”. In: *Blood* 113.11, pp. 2535–46.

Lipsick, J. S. (2010). “The C-MYB story—is it definitive?” In: *Proc Natl Acad Sci U S A* 107.40, pp. 17067–8.

Liu, C., C. Wu, Q. Yang, J. Gao, L. Li, D. Yang, and L. Luo (2016). “Macrophages Mediate the Repair of Brain Vascular Rupture through Direct Physical Adhesion and Mechanical Traction”. In: *Immunity* 44.5, pp. 1162–76.

– The physical force able to exerted by macrophages is an interesting and under-studied phenomenon, but this work presents a model where macrophages pull on the endothelium to guide it toward functional repair after disruption within the brain. This physical role for macrophages is interesting in the context of a disease like tuberculosis, which is defined by the involvement of macrophages.

Liu, C. J., Y. C. Cheng, K. W. Lee, H. H. Hsu, C. H. Chu, F. J. Tsai, C. H. Tsai, C. Y. Chu, J. Y. Liu, W. W. Kuo, and C. Y. Huang (2008a). “Lipopolysaccharide induces cellular hypertrophy through calcineurin/NFAT-3 signaling pathway in H9c2 myocardial cells”. In: *Mol Cell Biochem* 313.1-2, pp. 167–78.

Liu, F., S. Kambakam, M. P. Almeida, Z. Ming, J. M. Welker, W. A. Wierson, L. E. Schultz-Rogers, S. C. Ekker, K. J. Clark, J. J. Essner, and M. McGrail (2022a). “Cre/lox regulated conditional rescue and inactivation with zebrafish UFlip alleles generated by CRISPR-Cas9 targeted integration”. In: *Elife* 11.

Liu, H., Q. Sun, S. Chen, L. Chen, W. Jia, J. Zhao, and X. Sun (2021a). “DYRK1A activates NFATC1 to increase glioblastoma migration”. In: *Cancer Med* 10.18, pp. 6416–6427.

Liu, H., L. Yang, M. Qi, and J. Zhang (2018). “NFAT1 enhances the effects of tumor-associated macrophages on promoting malignant melanoma growth and metastasis”. In: *Biosci Rep* 38.6.

Liu, J. F., S. H. Zhao, and S. S. Wu (2013). “Depleting NFAT1 expression inhibits the ability of invasion and migration of human lung cancer cells”. In: *Cancer Cell Int* 13.1, p. 41.

Liu, Q., Y. Chen, M. Auger-Messier, and J. D. Molkentin (2012). “Interaction between NFκB and NFAT coordinates cardiac hypertrophy and pathological remodeling”. In: *Circ Res* 110.8, pp. 1077–86.

Liu, R., V. Muliadi, W. Mou, H. Li, J. Yuan, J. Holmberg, B. J. Chambers, N. Ullah, J. Wurth, M. Alzrigat, S. Schlisio, B. Carow, L. G. Larsson, and M. E. Rottenberg

(2022b). “HIF-1 stabilization in T cells hampers the control of *Mycobacterium tuberculosis* infection”. In: *Nat Commun* 13.1, p. 5093.

Liu, T., L. Zhang, D. Joo, and S. C. Sun (2017). “NF-kappaB signaling in inflammation”. In: *Signal Transduct Target Ther* 2.

Liu, X., J. W. Guo, X. C. Lin, Y. H. Tuo, W. L. Peng, S. Y. He, Z. Q. Li, Y. C. Ye, J. Yu, F. R. Zhang, M. M. Ma, J. Y. Shang, X. F. Lv, A. D. Zhou, Y. Ouyang, C. Wang, R. P. Pang, J. X. Sun, J. S. Ou, J. G. Zhou, and S. J. Liang (2021b). “Macrophage NFATc3 prevents foam cell formation and atherosclerosis: evidence and mechanisms”. In: *Eur Heart J* 42.47, pp. 4847–4861.

Liu, Y., Z. Li, Q. Lin, J. Kosinski, J. Seetharaman, J. M. Bujnicki, J. Sivaraman, and C. L. Hew (2007). “Structure and evolutionary origin of Ca(2+)-dependent herring type II antifreeze protein”. In: *PLoS One* 2.6, e548.

Liu, Y., Y. Liu, and M. Wang (2017). “Design, Optimization and Application of Small Molecule Biosensor in Metabolic Engineering”. In: *Front Microbiol* 8, p. 2012.

Liu, Z., F. Lebrin, J. A. Maring, S. van den Driesche, S. van der Brink, M. van Dinther, M. Thorikay, S. Martin, K. Kobayashi, L. J. Hawinkels, L. A. van Meeteren, E. Pardali, J. Korving, M. Letarte, H. M. Arthur, C. Theuer, M. J. Goumans, C. Mummery, and P. ten Dijke (2014). “Endoglin is dispensable for vasculogenesis, but required for vascular endothelial growth factor-induced angiogenesis”. In: *PLoS One* 9.1, e86273.

Liu, Z., J. Lee, S. Krummey, W. Lu, H. Cai, and M. J. Lenardo (2011). “The kinase LRRK2 is a regulator of the transcription factor NFAT that modulates the severity of inflammatory bowel disease”. In: *Nat Immunol* 12.11, pp. 1063–70.

Livak, K. J. and T. D. Schmittgen (2001). “Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method”. In: *Methods* 25.4, pp. 402–8.

Lobato-Pascual, A., P. C. Saether, S. Fossum, E. Dissen, and M. R. Daws (2013). “Mincl, the receptor for mycobacterial cord factor, forms a functional receptor complex with MCL and FcepsilonRI-gamma”. In: *Eur J Immunol* 43.12, pp. 3167–74.

– *This work established a role for MCL/MINCLE heterodimers in mediating signaling from these two closely related C-type lectin receptors and implied potential*

higher order regulatory mechanisms governing the responses of these receptors based on their hetero-/homo-dimerization or monomeric state.

Loddenkemper, R., M. Lipman, and A. Zumla (2015). “Clinical Aspects of Adult Tuberculosis”. In: *Cold Spring Harb Perspect Med* 6.1, a017848.

Loeffler, D. A., S. K. Lundy, K. P. Singh, H. C. Gerard, A. P. Hudson, and D. L. Boros (2002). “Soluble egg antigens from *Schistosoma mansoni* induce angiogenesis-related processes by up-regulating vascular endothelial growth factor in human endothelial cells”. In: *J Infect Dis* 185.11, pp. 1650–6.

Loh, C., K. T. Shaw, J. Carew, J. P. Viola, C. Luo, B. A. Perrino, and A. Rao (1996). “Calcineurin binds the transcription factor NFAT1 and reversibly regulates its activity”. In: *J Biol Chem* 271.18, pp. 10884–91.

Long, Y., J. Yan, G. Song, X. Li, X. Li, Q. Li, and Z. Cui (2015). “Transcriptional events co-regulated by hypoxia and cold stresses in Zebrafish larvae”. In: *BMC Genomics* 16, p. 385.

Loo, Y. M. and Jr. Gale M. (2011). “Immune signaling by RIG-I-like receptors”. In: *Immunity* 34.5, pp. 680–92.

Looney, M., R. Lorenc, M. K. Halushka, and P. C. Karakousis (2021). “Key Macrophage Responses to Infection With *Mycobacterium tuberculosis* Are Co-Regulated by microRNAs and DNA Methylation”. In: *Front Immunol* 12, p. 685237.

Lopez-Rodriguez, C., J. Aramburu, L. Jin, A. S. Rakeman, M. Michino, and A. Rao (2001). “Bridging the NFAT and NF-kappaB families: NFAT5 dimerization regulates cytokine gene transcription in response to osmotic stress”. In: *Immunity* 15.1, pp. 47–58.

Losslein, A. K., F. Lohrmann, L. Scheuermann, K. Gharun, J. Neuber, J. Kolter, A. J. Forde, C. Kleimeyer, Y. Y. Poh, M. Mack, A. Triantafyllopoulou, M. D. Dunlap, S. A. Khader, M. Seidl, A. Holscher, C. Holscher, X. L. Guan, A. Dorhoi, and P. Henneke (2021). “Monocyte progenitors give rise to multinucleated giant cells”. In: *Nat Commun* 12.1, p. 2027.

Love, M. I., W. Huber, and S. Anders (2014). “Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2”. In: *Genome Biol* 15.12, p. 550.

- Lovewell, R. R., C. E. Baer, B. B. Mishra, C. M. Smith, and C. M. Sassetti (2021). “Granulocytes act as a niche for *Mycobacterium tuberculosis* growth”. In: *Mucosal Immunol* 14.1, pp. 229–241.
- Lowbury, E. J. (1958). “Symposium on epidemiological risks of antibiotics: hospital infection”. In: *Proc R Soc Med* 51.10, pp. 807–10.
- Lu, X., M. Nagata, and S. Yamasaki (2018). “Mincle: 20 years of a versatile sensor of insults”. In: *Int Immunol* 30.6, pp. 233–239.
- Lu, Y. C., W. C. Yeh, and P. S. Ohashi (2008). “LPS/TLR4 signal transduction pathway”. In: *Cytokine* 42.2, pp. 145–151.
- Lucerna, M., A. Zernecke, R. de Nooijer, S. C. de Jager, I. Bot, C. van der Lans, I. Kholova, E. A. Liehn, T. J. van Berkel, S. Yla-Herttuala, C. Weber, and E. A. Biessen (2007). “Vascular endothelial growth factor-A induces plaque expansion in ApoE knock-out mice by promoting *de novo* leukocyte recruitment”. In: *Blood* 109.1, pp. 122–9.
- Luddecke, J., L. Francois, P. Spat, B. Watzer, T. Chilczuk, G. Poschet, R. Hell, B. Radlwimmer, and K. Forchhammer (2017). “PII Protein-Derived FRET Sensors for Quantification and Live-Cell Imaging of 2-Oxoglutarate”. In: *Sci Rep* 7.1, p. 1437.
- Lugo-Villarino, G., K. M. Balla, D. L. Stachura, K. Banuelos, M. B. Werneck, and D. Traver (2010). “Identification of dendritic antigen-presenting cells in the zebrafish”. In: *Proc Natl Acad Sci U S A* 107.36, pp. 15850–5.
- Lugo-Villarino, G. and O. Neyrolles (2014). “Of clots and granulomas: platelets are new players in immunity to tuberculosis”. In: *J Infect Dis* 210.11, pp. 1687–90.
- Lukiw, W. J., A. Ottlecz, G. Lambrou, M. Grueninger, J. Finley, H. W. Thompson, and N. G. Bazan (2003). “Coordinate activation of HIF-1 and NF-kappaB DNA binding and COX-2 and VEGF expression in retinal cells by hypoxia”. In: *Invest Ophthalmol Vis Sci* 44.10, pp. 4163–70.
- Lund, M. E., J. To, B. A. O’Brien, and S. Donnelly (2016). “The choice of phorbol 12-myristate 13-acetate differentiation protocol influences the response of THP-1 macrophages to a pro-inflammatory stimulus”. In: *J Immunol Methods* 430, pp. 64–70.

- Luo, J. J., W. P. Bian, Y. Liu, H. Y. Huang, Q. Yin, X. J. Yang, and D. S. Pei (2018). “CRISPR/Cas9-based genome engineering of zebrafish using a seamless integration strategy”. In: *FASEB J* 32.9, pp. 5132–5142.
- Luoma, J. I. and L. Zirpel (2008). “Deafferentation-induced activation of NFAT (nuclear factor of activated T-cells) in cochlear nucleus neurons during a developmental critical period: a role for NFATc4-dependent apoptosis in the CNS”. In: *J Neurosci* 28.12, pp. 3159–69.
- Luque, A., A. Gonzalez Granja, L. Gonzalez, and C. Tafalla (2014). “Establishment and characterization of a rainbow trout heart endothelial cell line with susceptibility to viral hemorrhagic septicemia virus (VHSV)”. In: *Fish Shellfish Immunol* 38.1, pp. 255–64.
- Lyadova, I. V. and A. V. Panteleev (2015). “Th1 and Th17 Cells in Tuberculosis: Protection, Pathology, and Biomarkers”. In: *Mediators Inflamm* 2015, p. 854507.
- Lyakh, L., P. Ghosh, and N. R. Rice (1997). “Expression of NFAT-family proteins in normal human T cells”. In: *Mol Cell Biol* 17.5, pp. 2475–84.
- Ma, Z., P. Zhu, M. Pang, L. Guo, N. Chang, J. Zheng, X. Zhu, C. Gao, H. Huang, Z. Cui, J. W. Xiong, J. Peng, and J. Chen (2017). “A novel inducible mutagenesis screen enables to isolate and clone both embryonic and adult zebrafish mutants”. In: *Sci Rep* 7.1, p. 10381.
- MacDonald, I. J., S. C. Liu, C. M. Su, Y. H. Wang, C. H. Tsai, and C. H. Tang (2018). “Implications of Angiogenesis Involvement in Arthritis”. In: *Int J Mol Sci* 19.7.
- Macedo, L., G. Pinhal-Enfield, V. Alshits, G. Elson, B. N. Cronstein, and S. J. Leibovich (2007). “Wound healing is impaired in MyD88-deficient mice: a role for MyD88 in the regulation of wound healing by adenosine A2A receptors”. In: *Am J Pathol* 171.6, pp. 1774–88.
- MacGilvary, N. J. and S. Tan (2018). “Fluorescent *Mycobacterium tuberculosis* reporters: illuminating host-pathogen interactions”. In: *Pathog Dis* 76.3.
- Macian, F. (2005). “NFAT proteins: key regulators of T-cell development and function”. In: *Nat Rev Immunol* 5.6, pp. 472–84.

Macian, F., C. Lopez-Rodriguez, and A. Rao (2001). “Partners in transcription: NFAT and AP-1”. In: *Oncogene* 20.19, pp. 2476–89.

MacMicking, J., Q. W. Xie, and C. Nathan (1997). “Nitric oxide and macrophage function”. In: *Annu Rev Immunol* 15, pp. 323–50.

MacMicking, J. D. (2004). “IFN-inducible GTPases and immunity to intracellular pathogens”. In: *Trends Immunol* 25.11, pp. 601–9.

– (2012). “Interferon-inducible effector mechanisms in cell-autonomous immunity”. In: *Nat Rev Immunol* 12.5, pp. 367–82.

– (2014). “Cell-autonomous effector mechanisms against *Mycobacterium tuberculosis*”. In: *Cold Spring Harb Perspect Med* 4.10.

MacMicking, J. D., C. Nathan, G. Hom, N. Chartrain, D. S. Fletcher, M. Trumbauer, K. Stevens, Q. W. Xie, K. Sokol, N. Hutchinson, and et al. (1995). “Altered responses to bacterial infection and endotoxic shock in mice lacking inducible nitric oxide synthase”. In: *Cell* 81.4, pp. 641–50.

MacMicking, J. D., R. J. North, R. LaCourse, J. S. Mudgett, S. K. Shah, and C. F. Nathan (1997). “Identification of nitric oxide synthase as a protective locus against tuberculosis”. In: *Proc Natl Acad Sci U S A* 94.10, pp. 5243–8.

MacRae, C. A. and R. T. Peterson (2003). “Zebrafish-based small molecule discovery”. In: *Chem Biol* 10.10, pp. 901–8.

– (2015). “Zebrafish as tools for drug discovery”. In: *Nat Rev Drug Discov* 14.10, pp. 721–31.

Madacki, J., M. Orgeur, G. Mas Fiol, W. Frigui, L. Ma, and R. Brosch (2021). “ESX-1-Independent Horizontal Gene Transfer by *Mycobacterium tuberculosis* Complex Strains”. In: *mBio* 12.3.

Madan-Lala, R., K. V. Peixoto, F. Re, and J. Rengarajan (2011). “*Mycobacterium tuberculosis* Hip1 dampens macrophage proinflammatory responses by limiting toll-like receptor 2 activation”. In: *Infect Immun* 79.12, pp. 4828–38.

Madden, K., Y. C. Liang, N. Rajabalee, G. G. Alvarez, and J. Sun (2022). “Surveying the Epigenetic Landscape of Tuberculosis in Alveolar Macrophages”. In: *Infect Immun* 90.5, e0052221.

Maddrey, W. C. and J. K. Boitnott (1973). “Isoniazid hepatitis”. In: *Ann Intern Med* 79.1, pp. 1–12.

Madigan, C. A., C. J. Cambier, K. M. Kelly-Scumpia, P. O. Scumpia, T. Y. Cheng, J. Zailaa, B. R. Bloom, D. B. Moody, S. T. Smale, A. Sagasti, R. L. Modlin, and L. Ramakrishnan (2017). “A Macrophage Response to *Mycobacterium leprae* Phenolic Glycolipid Initiates Nerve Damage in Leprosy”. In: *Cell* 170.5, 973–985 e10.

Maeda, N., J. Nigou, J. L. Herrmann, M. Jackson, A. Amara, P. H. Lagrange, G. Puzo, B. Gicquel, and O. Neyrolles (2003). “The cell surface receptor DC-SIGN discriminates between *Mycobacterium* species through selective recognition of the mannose caps on lipoarabinomannan”. In: *J Biol Chem* 278.8, pp. 5513–6.

Mahajan, R. (2013). “Bedaquiline: First FDA-approved tuberculosis drug in 40 years”. In: *Int J Appl Basic Med Res* 3.1, pp. 1–2.

Mahamed, D., M. Boulle, Y. Ganga, C. Mc Arthur, S. Skroch, L. Oom, O. Catinas, K. Pillay, M. Naicker, S. Rampersad, C. Mathonsi, J. Hunter, E. B. Wong, M. Suleman, G. Sreejit, A. S. Pym, G. Lustig, and A. Sigal (2017). “Intracellular growth of *Mycobacterium tuberculosis* after macrophage cell death leads to serial killing of host cells”. In: *Elife* 6.

Makinen, T., L. Jussila, T. Veikkola, T. Karpanen, M. I. Kettunen, K. J. Pulkkanen, R. Kauppinen, D. G. Jackson, H. Kubo, S. Nishikawa, S. Yla-Herttuala, and K. Alitalo (2001). “Inhibition of lymphangiogenesis with resulting lymphedema in transgenic mice expressing soluble VEGF receptor-3”. In: *Nat Med* 7.2, pp. 199–205.

Makino, H., S. Seki, Y. Yahara, S. Shiozawa, Y. Aikawa, H. Motomura, M. Nogami, K. Watanabe, T. Sainoh, H. Ito, N. Tsumaki, Y. Kawaguchi, M. Yamazaki, and T. Kimura (2017). “A selective inhibition of c-Fos/activator protein-1 as a potential therapeutic target for intervertebral disc degeneration and associated pain”. In: *Sci Rep* 7.1, p. 16983.

Maldonado, R. F., I. Sa-Correia, and M. A. Valvano (2016). “Lipopolysaccharide modification in Gram-negative bacteria during chronic infection”. In: *FEMS Microbiol Rev* 40.4, pp. 480–93.

- Malecic, N. and H. S. Young (2017). “Excessive angiogenesis associated with psoriasis as a cause for cardiovascular ischaemia”. In: *Exp Dermatol* 26.4, pp. 299–304.
- Malek, T. R. and A. L. Bayer (2004). “Tolerance, not immunity, crucially depends on IL-2”. In: *Nat Rev Immunol* 4.9, pp. 665–74.
- Malicki, J., S. C. Neuhauss, A. F. Schier, L. Solnica-Krezel, D. L. Stemple, D. Y. Stainier, S. Abdelilah, F. Zwartkruis, Z. Rangini, and W. Driever (1996a). “Mutations affecting development of the zebrafish retina”. In: *Development* 123, pp. 263–73.
- Malicki, J., A. F. Schier, L. Solnica-Krezel, D. L. Stemple, S. C. Neuhauss, D. Y. Stainier, S. Abdelilah, Z. Rangini, F. Zwartkruis, and W. Driever (1996b). “Mutations affecting development of the zebrafish ear”. In: *Development* 123, pp. 275–83.
- Malik, A. and T. D. Kanneganti (2018). “Function and regulation of IL-1alpha in inflammatory diseases and cancer”. In: *Immunol Rev* 281.1, pp. 124–137.
- Malik, Z. A., G. M. Denning, and D. J. Kusner (2000). “Inhibition of Ca(2+) signalling by *Mycobacterium tuberculosis* is associated with reduced phagosome-lysosome fusion and increased survival within human macrophages”. In: *J Exp Med* 191.2, pp. 287–302.
- Manabe, Y. C. and W. R. Bishai (2000). “Latent *Mycobacterium tuberculosis*-persistence, patience, and winning by waiting”. In: *Nat Med* 6.12, pp. 1327–9.
- Mancini, M. and A. Toker (2009). “NFAT proteins: emerging roles in cancer progression”. In: *Nat Rev Cancer* 9.11, pp. 810–20.
- Mann, E. S., S. J. Fogarty, and M. C. Kincaid (2000). “Choroidal neovascularization with granulomatous inflammation in ocular histoplasmosis syndrome”. In: *Am J Ophthalmol* 130.2, pp. 247–50.
- Manne, B. K., R. Badolia, C. Dangelmaier, J. A. Eble, W. Ellmeier, M. Kahn, and S. P. Kunapuli (2015). “Distinct pathways regulate Syk protein activation downstream of immune tyrosine activation motif (ITAM) and hemITAM receptors in platelets”. In: *J Biol Chem* 290.18, pp. 11557–68.

- Manukyan, M., K. Triantafilou, M. Triantafilou, A. Mackie, N. Nilsen, T. Espevik, K. H. Wiesmuller, A. J. Ulmer, and H. Heine (2005). “Binding of lipopeptide to CD14 induces physical proximity of CD14, TLR2 and TLR1”. In: *Eur J Immunol* 35.3, pp. 911–21.
- Manzanillo, P. S., M. U. Shiloh, D. A. Portnoy, and J. S. Cox (2012). “*Mycobacterium tuberculosis* activates the DNA-dependent cytosolic surveillance pathway within macrophages”. In: *Cell Host Microbe* 11.5, pp. 469–80.
- Marakalala, M. J., L. M. Graham, and G. D. Brown (2010). “The role of Syk/CARD9-coupled C-type lectin receptors in immunity to *Mycobacterium tuberculosis* infections”. In: *Clin Dev Immunol* 2010, p. 567571.
- Marakalala, M. J., R. Guler, L. Matika, G. Murray, M. Jacobs, F. Brombacher, A. G. Rothfuchs, A. Sher, and G. D. Brown (2011). “The Syk/CARD9-coupled receptor Dectin-1 is not required for host resistance to *Mycobacterium tuberculosis* in mice”. In: *Microbes Infect* 13.2, pp. 198–201.
- Marakalala, M. J. and H. Ndlovu (2017). “Signaling C-type lectin receptors in antimycobacterial immunity”. In: *PLoS Pathog* 13.6, e1006333.
- Marass, M., A. Beisaw, C. Gerri, F. Luzzani, N. Fukuda, S. Gunther, C. Kuenne, S. Reischauer, and D. Y. R. Stainier (2019). “Genome-wide strategies reveal target genes of Npas4l associated with vascular development in zebrafish”. In: *Development* 146.11.
- Marconcini, L., S. Marchio, L. Morbidelli, E. Cartocci, A. Albini, M. Ziche, F. Busolino, and S. Oliviero (1999). “c-fos-induced growth factor/vascular endothelial growth factor D induces angiogenesis *in vivo* and *in vitro*”. In: *Proc Natl Acad Sci U S A* 96.17, pp. 9671–6.
- Mariam, D. H., Y. Mengistu, S. E. Hoffner, and D. I. Andersson (2004). “Effect of rpoB mutations conferring rifampin resistance on fitness of *Mycobacterium tuberculosis*”. In: *Antimicrob Agents Chemother* 48.4, pp. 1289–94.
- Mariani, F., D. Goletti, A. Ciaramella, A. Martino, V. Colizzi, and M. Fraziano (2001). “Macrophage response to *Mycobacterium tuberculosis* during HIV infection: relationships between macrophage activation and apoptosis”. In: *Curr Mol Med* 1.2, pp. 209–16.

- Mariner, J. C., J. A. House, C. A. Mebus, A. E. Sollod, D. Chibeu, B. A. Jones, P. L. Roeder, B. Admassu, and G. G. van 't Klooster (2012). "Rinderpest eradication: appropriate technology and social innovations". In: *Science* 337.6100, pp. 1309–12.
- Martidis, A., D. G. Miller, T. A. Ciulla, R. P. Danis, and R. S. Moorthy (1999). "Corticosteroids as an antiangiogenic agent for histoplasmosis-related subfoveal choroidal neovascularization". In: *J Ocul Pharmacol Ther* 15.5, pp. 425–8.
- Martignoni, M. E., P. Kunze, and H. Friess (2003). "Cancer cachexia". In: *Mol Cancer* 2, p. 36.
- Martin, C. J., M. G. Booty, T. R. Rosebrock, C. Nunes-Alves, D. M. Desjardins, I. Keren, S. M. Fortune, H. G. Remold, and S. M. Behar (2012). "Efferocytosis is an innate antibacterial mechanism". In: *Cell Host Microbe* 12.3, pp. 289–300.
- Martin, J., S. Collot-Teixeira, L. McGregor, and J. L. McGregor (2007). "The dialogue between endothelial cells and monocytes/macrophages in vascular syndromes". In: *Curr Pharm Des* 13.17, pp. 1751–9.
- Martinez, F. O. and S. Gordon (2014).** "The M1 and M2 paradigm of macrophage activation: time for reassessment". In: *F1000Prime Rep* 6, p. 13.
- Martinez, G. J., R. M. Pereira, T. Aijo, E. Y. Kim, F. Marangoni, M. E. Pipkin, S. Togher, V. Heissmeyer, Y. C. Zhang, S. Crotty, E. D. Lamperti, K. M. Ansel, T. R. Mempel, H. Lahdesmaki, P. G. Hogan, and A. Rao (2015). "The transcription factor NFAT promotes exhaustion of activated CD8(+) T cells". In: *Immunity* 42.2, pp. 265–278.
- Martinez, L., R. Verma, J. Croda, Jr. Horsburgh C. R., K. S. Walter, N. Degner, K. Middelkoop, A. Koch, S. Hermans, D. F. Warner, R. Wood, F. Cobelens, and J. R. Andrews (2019). "Detection, survival and infectious potential of *Mycobacterium tuberculosis* in the environment: a review of the evidence and epidemiological implications". In: *Eur Respir J* 53.6.
- Martinon, F. and J. Tschopp (2005). "NLRs join TLRs as innate sensors of pathogens". In: *Trends Immunol* 26.8, pp. 447–54.
- Masoud, G. N. and W. Li (2015). "HIF-1alpha pathway: role, regulation and intervention for cancer therapy". In: *Acta Pharm Sin B* 5.5, pp. 378–89.

- Mass, E. (2018). "Delineating the origins, developmental programs and homeostatic functions of tissue-resident macrophages". In: *Int Immunol* 30.11, pp. 493–501.
- Master, S. S., S. K. Rampini, A. S. Davis, C. Keller, S. Ehlers, B. Springer, G. S. Timmins, P. Sander, and V. Deretic (2008). "Mycobacterium tuberculosis prevents inflammasome activation". In: *Cell Host Microbe* 3.4, pp. 224–32.
- Masud, S., V. Torraca, and A. H. Meijer (2017). "Modeling Infectious Diseases in the Context of a Developing Immune System". In: *Curr Top Dev Biol* 124, pp. 277–329.
- Masuda, E. S., Y. Naito, H. Tokumitsu, D. Campbell, F. Saito, C. Hannum, K. Arai, and N. Arai (1995). "NFATx, a novel member of the nuclear factor of activated T cells family that is expressed predominantly in the thymus". In: *Mol Cell Biol* 15.5, pp. 2697–706.
- Mathema, B., J. R. Andrews, T. Cohen, M. W. Borgdorff, M. Behr, J. R. Glynn, R. Rustomjee, B. J. Silk, and R. Wood (2017). "Drivers of Tuberculosis Transmission". In: *J Infect Dis* 216.suppl_6, S644–S653.
- Matsumoto, M., M. Kogawa, S. Wada, H. Takayanagi, M. Tsujimoto, S. Katayama, K. Hisatake, and Y. Nogi (2004). "Essential role of p38 mitogen-activated protein kinase in cathepsin K gene expression during osteoclastogenesis through association of NFATc1 and PU.1". In: *J Biol Chem* 279.44, pp. 45969–79.
- Matsumoto, M., T. Tanaka, T. Kaisho, H. Sanjo, N. G. Copeland, D. J. Gilbert, N. A. Jenkins, and S. Akira (1999). "A novel LPS-inducible C-type lectin is a transcriptional target of NF-IL6 in macrophages". In: *J Immunol* 163.9, pp. 5039–48.
- Matsunaga, I. and D. B. Moody (2009). "Mincl is a long sought receptor for mycobacterial cord factor". In: *J Exp Med* 206.13, pp. 2865–8.
- Matsuoka, R. L., M. Marass, A. Avdesh, C. S. Helker, H. M. Maischein, A. S. Grosse, H. Kaur, N. D. Lawson, W. Herzog, and D. Y. Stainier (2016). "Radial glia regulate vascular patterning around the developing spinal cord". In: *Elife* 5.
- Matsuoka, R. L. and D. Y. R. Stainier (2018). "Recent insights into vascular development from studies in zebrafish". In: *Curr Opin Hematol* 25.3, pp. 204–211.

- Matteelli, A., A. Roggi, and A. C. Carvalho (2014). “Extensively drug-resistant tuberculosis: epidemiology and management”. In: *Clin Epidemiol* 6, pp. 111–8.
- Mattonet, K., F. W. Riemslagh, S. Guenther, K. D. Prummel, G. Kesavan, S. Hans, I. Ebersberger, M. Brand, A. Burger, S. Reischauer, C. Mosimann, and D. Y. R. Stainier (2022). “Endothelial versus pronephron fate decision is modulated by the transcription factors Cloche/Npas4l, Tal1, and Lmo2”. In: *Sci Adv* 8.35, eabn2082.
- Matty, M. A., D. R. Knudsen, E. M. Walton, R. W. Beerman, M. R. Cronan, C. J. Pyle, R. E. Hernandez, and D. M. Tobin (2019). “Potentiation of P2RX7 as a host-directed strategy for control of mycobacterial infection”. In: *Elife* 8.
- Matty, M. A., F. J. Roca, M. R. Cronan, and D. M. Tobin (2015). “Adventures within the speckled band: heterogeneity, angiogenesis, and balanced inflammation in the tuberculous granuloma”. In: *Immunol Rev* 264.1, pp. 276–87.
- Maueroder, C., R. A. Chaurio, T. Dumych, M. Podolska, M. D. Lootsik, S. Culemann, R. P. Friedrich, R. Bilyy, C. Alexiou, G. Schett, C. Berens, M. Herrmann, and L. E. Munoz (2016). “A blast without power - cell death induced by the tuberculosis-necrotizing toxin fails to elicit adequate immune responses”. In: *Cell Death Differ* 23.6, pp. 1016–25.
- Mayer-Barber, K. D., B. B. Andrade, S. D. Oland, E. P. Amaral, D. L. Barber, J. Gonzales, S. C. Derrick, R. Shi, N. P. Kumar, W. Wei, X. Yuan, G. Zhang, Y. Cai, S. Babu, M. Catalfamo, A. M. Salazar, L. E. Via, 3rd Barry C. E., and A. Sher (2014). “Host-directed therapy of tuberculosis based on interleukin-1 and type I interferon crosstalk”. In: *Nature* 511.7507, pp. 99–103.
- Mazurek, J., L. Ignatowicz, G. Kallenius, S. B. Svenson, A. Pawlowski, and B. Hamarsur (2012). “Divergent effects of mycobacterial cell wall glycolipids on maturation and function of human monocyte-derived dendritic cells”. In: *PLoS One* 7.8, e42515.
- McAlister, V. C., E. Haddad, E. Renouf, R. A. Malthaner, M. S. Kjaer, and L. L. Gluud (2006). “Cyclosporin versus tacrolimus as primary immunosuppressant after liver transplantation: a meta-analysis”. In: *Am J Transplant* 6.7, pp. 1578–85.
- McBride, William Griffith (1961). “Thalidomide and congenital abnormalities”. In: *Lancet* 2.1358, pp. 90927–8.

McCaffrey, E. F., M. Donato, L. Keren, Z. Chen, A. Delmastro, M. B. Fitzpatrick, S. Gupta, N. F. Greenwald, A. Baranski, W. Graf, R. Kumar, M. Bosse, C. C. Fullaway, P. K. Ramdial, E. Forgo, V. Jovic, D. Van Valen, S. Mehra, S. A. Khader, S. C. Bendall, M. van de Rijn, D. Kalman, D. Kaushal, R. L. Hunter, N. Banaei, A. J. C. Steyn, P. Khatri, and M. Angelo (2022). “The immunoregulatory landscape of human tuberculosis granulomas”. In: *Nat Immunol* 23.2, pp. 318–329.

McCaffrey, P. G., B. A. Perrino, T. R. Soderling, and A. Rao (1993). “NF-ATp, a T lymphocyte DNA-binding protein that is a target for calcineurin and immunosuppressive drugs”. In: *J Biol Chem* 268.5, pp. 3747–52.

– *The earliest mechanistic dissection of NFATC2 from Anjana Rao which identified its regulation by phosphorylation and calcineurin and was instrumental to our understanding of the regulatory mechanisms of the entire NFAT family.*

McCann, J. R., J. A. McDonough, M. S. Pavelka, and M. Braunstein (2007). “Beta-lactamase can function as a reporter of bacterial protein export during *Mycobacterium tuberculosis* infection of host cells”. In: *Microbiology (Reading)* 153.Pt 10, pp. 3350–3359.

McCarter, J. and E. G. Hastings (1935). “The Morphology of the Mycobacteria”. In: *J Bacteriol* 29.5, pp. 503–13.

McConnell, A. M. and L. I. Zon (2021). “Dissecting melanocytes to predict melanoma”. In: *Nat Cell Biol* 23.9, pp. 930–931.

McCormack, P. L. and S. J. Keam (2008). “Bevacizumab: a review of its use in metastatic colorectal cancer”. In: *Drugs* 68.4, pp. 487–506.

McCullagh, K. J., E. Calabria, G. Pallafacchina, S. Ciciliot, A. L. Serrano, C. Argentini, J. M. Kalhovde, T. Lomo, and S. Schiaffino (2004). “NFAT is a nerve activity sensor in skeletal muscle and controls activity-dependent myosin switching”. In: *Proc Natl Acad Sci U S A* 101.29, pp. 10590–5.

McDonough, K. A., Y. Kress, and B. R. Bloom (1993). “Pathogenesis of tuberculosis: interaction of *Mycobacterium tuberculosis* with macrophages”. In: *Infect Immun* 61.7, pp. 2763–73.

- McElvania Tekippe, E., I. C. Allen, P. D. Hulseberg, J. T. Sullivan, J. R. McCann, M. Sandor, M. Braunstein, and J. P. Ting (2010). "Granuloma formation and host defense in chronic *Mycobacterium tuberculosis* infection requires PYCARD/ASC but not NLRP3 or caspase-1". In: *PLoS One* 5.8, e12320.
- McGrath, M., N. C. Gey van Pittius, P. D. van Helden, R. M. Warren, and D. F. Warner (2014). "Mutation rate and the emergence of drug resistance in *Mycobacterium tuberculosis*". In: *J Antimicrob Chemother* 69.2, pp. 292–302.
- McGreal, E. P., J. L. Miller, and S. Gordon (2005). "Ligand recognition by antigen-presenting cell C-type lectin receptors". In: *Curr Opin Immunol* 17.1, pp. 18–24.
- McGuire, V. A. and J. S. Arthur (2015). "Subverting Toll-Like Receptor Signaling by Bacterial Pathogens". In: *Front Immunol* 6, p. 607.
- McQuin, C., A. Goodman, V. Chernyshev, L. Kamentsky, B. A. Cimini, K. W. Karhohs, M. Doan, L. Ding, S. M. Rafelski, D. Thirstrup, W. Wiegraebe, S. Singh, T. Becker, J. C. Caicedo, and A. E. Carpenter (2018). "CellProfiler 3.0: Next-generation image processing for biology". In: *PLoS Biol* 16.7, e2005970.
- Medeiros, R. C., K. D. Girardi, F. K. Cardoso, B. S. Mietto, T. G. Pinto, L. S. Gomez, L. S. Rodrigues, M. Gandini, J. J. Amaral, S. L. Antunes, S. Corte-Real, P. S. Rosa, M. C. Pessolani, J. A. Nery, E. N. Sarno, L. R. Batista-Silva, M. Sola-Penna, M. F. Oliveira, M. O. Moraes, and F. A. Lara (2016). "Subversion of Schwann Cell Glucose Metabolism by *Mycobacterium leprae*". In: *J Biol Chem* 291.41, pp. 21375–21387.
- Meeker, N. D., S. A. Hutchinson, L. Ho, and N. S. Trede (2007). "Method for isolation of PCR-ready genomic DNA from zebrafish tissues". In: *Biotechniques* 43.5, pp. 610, 612, 614.
- Mehla, K. and P. K. Singh (2019). "Metabolic Regulation of Macrophage Polarization in Cancer". In: *Trends Cancer* 5.12, pp. 822–834.
- Mehrotra, P., S. V. Jamwal, N. Saquib, N. Sinha, Z. Siddiqui, V. Manivel, S. Chatterjee, and K. V. Rao (2014). "Pathogenicity of *Mycobacterium tuberculosis* is expressed by regulating metabolic thresholds of the host macrophage". In: *PLoS Pathog* 10.7, e1004265.

Mendoza-Coronel, E. and M. Castanon-Arreola (2016). “Comparative evaluation of *in vitro* human macrophage models for mycobacterial infection study”. In: *Pathog Dis* 74.6.

Meng, X., B. Grottsch, Y. Luo, K. X. Knaup, M. S. Wiesener, X. X. Chen, J. Jantsch, S. Fillatreau, G. Schett, and A. Bozec (2018). “Hypoxia-inducible factor-1alpha is a critical transcription factor for IL-10-producing B cells in autoimmune disease”. In: *Nat Commun* 9.1, p. 251.

Meniche, X., R. Otten, M. S. Siegrist, C. E. Baer, K. C. Murphy, C. R. Bertozzi, and C. M. Sassetti (2014). “Subpolar addition of new cell wall is directed by DivIVA in mycobacteria”. In: *Proc Natl Acad Sci U S A* 111.31, E3243–51.

Menzies, B. E. and I. Kourteva (1998). “Internalization of *Staphylococcus aureus* by endothelial cells induces apoptosis”. In: *Infect Immun* 66.12, pp. 5994–8.

Merad, M., F. Ginhoux, and M. Collin (2008). “Origin, homeostasis and function of Langerhans cells and other langerin-expressing dendritic cells”. In: *Nat Rev Immunol* 8.12, pp. 935–47.

Mercurio, A. M., E. A. Lipscomb, and R. E. Bachelder (2005). “Non-angiogenic functions of VEGF in breast cancer”. In: *J Mammary Gland Biol Neoplasia* 10.4, pp. 283–90.

Meredith, H. R., J. K. Srimani, A. J. Lee, A. J. Lopatkin, and L. You (2015). “Collective antibiotic tolerance: mechanisms, dynamics and intervention”. In: *Nat Chem Biol* 11.3, pp. 182–8.

Merker, M., C. Blin, S. Mona, N. Duforet-Frebourg, S. Lecher, E. Willery, M. G. Blum, S. Rusch-Gerdes, I. Mokrousov, E. Aleksic, C. Allix-Beguec, A. Antierens, E. Augustynowicz-Kopec, M. Ballif, F. Barletta, H. P. Beck, 3rd Barry C. E., M. Bonnet, E. Borroni, I. Campos-Herrero, D. Cirillo, H. Cox, S. Crowe, V. Crudu, R. Diel, F. Drobniowski, M. Fauville-Dufaux, S. Gagneux, S. Ghebremichael, M. Hanekom, S. Hoffner, W. W. Jiao, S. Kalon, T. A. Kohl, I. Kontsevaya, T. Lillebaek, S. Maeda, V. Nikolayevskyy, M. Rasmussen, N. Rastogi, S. Samper, E. Sanchez-Padilla, B. Savic, I. C. Shamputa, A. Shen, L. H. Sng, P. Stakenas, K. Toit, F. Varaine, D. Vukovic, C. Wahl, R. Warren, P. Supply, S. Niemann, and T. Wirth (2015). “Evolutionary history and global spread of the *Mycobacterium tuberculosis* Beijing lineage”. In: *Nat Genet* 47.3, pp. 242–9.

Mesange, P., V. Poindessous, M. Sabbah, A. E. Escargueil, A. de Gramont, and A. K. Larsen (2014). “Intrinsic bevacizumab resistance is associated with prolonged activation of autocrine VEGF signaling and hypoxia tolerance in colorectal cancer cells and can be overcome by nintedanib, a small molecule angiokinase inhibitor”. In: *Oncotarget* 5.13, pp. 4709–21.

Metushi, I., J. Utrecht, and E. Phillips (2016). “Mechanism of isoniazid-induced hepatotoxicity: then and now”. In: *Br J Clin Pharmacol* 81.6, pp. 1030–6.

Metushi, I. G., P. Cai, X. Zhu, T. Nakagawa, and J. P. Utrecht (2011). “A fresh look at the mechanism of isoniazid-induced hepatotoxicity”. In: *Clin Pharmacol Ther* 89.6, pp. 911–4.

Meyer, A. and M. Schartl (1999). “Gene and genome duplications in vertebrates: the one-to-four (-to-eight in fish) rule and the evolution of novel gene functions”. In: *Curr Opin Cell Biol* 11.6, pp. 699–704.

Mezouar, S., I. Diarra, J. Roudier, B. Desnues, and J. L. Mege (2019). “Tumor Necrosis Factor-Alpha Antagonist Interferes With the Formation of Granulomatous Multinucleated Giant Cells: New Insights Into *Mycobacterium tuberculosis* Infection”. In: *Front Immunol* 10, p. 1947.

Michaelsen, S. R., M. Staberg, H. Pedersen, K. E. Jensen, W. Majewski, H. Broholm, M. K. Nedergaard, C. Meulengracht, T. Urup, M. Villingshoj, S. Lukacova, J. Skjoth-Rasmussen, J. Brennum, A. Kjaer, U. Lassen, M. T. Stockhausen, H. S. Poulsen, and P. Hamerlik (2018). “VEGF-C sustains VEGFR2 activation under bevacizumab therapy and promotes glioblastoma maintenance”. In: *Neuro Oncol* 20.11, pp. 1462–1474.

Middleton, J. R., P. Sen, M. Lange, J. Salaki, R. Kapila, and D. B. Louria (1977). “Death-producing hemoptysis in tuberculosis”. In: *Chest* 72.5, pp. 601–4.

Mihatsch, M. J., M. Kyo, K. Morozumi, Y. Yamaguchi, V. Nickeleit, and B. Ryffel (1998). “The side-effects of ciclosporine-A and tacrolimus”. In: *Clin Nephrol* 49.6, pp. 356–63.

Millard, J. D., E. A. Mackay, L. J. Bonnett, and G. R. Davies (2019). “The impact of inclusion, dose and duration of pyrazinamide (PZA) on efficacy and safety outcomes in tuberculosis: systematic review and meta-analysis protocol”. In: *Syst Rev* 8.1, p. 329.

Miller, James Alexander (July 1938). “Some Unsolved Problems Of Tuberculosis”. In: *Journal of the American Medical Association* 111.2, pp. 111–117.

Miller, S. I., R. K. Ernst, and M. W. Bader (2005). “LPS, TLR4 and infectious disease diversity”. In: *Nat Rev Microbiol* 3.1, pp. 36–46.

Mills, C. D. (2015). “Anatomy of a discovery: M1 and M2 macrophages”. In: *Front Immunol* 6, p. 212.

Mills, C. D., K. Kincaid, J. M. Alt, M. J. Heilman, and A. M. Hill (2000). “M-1/M-2 macrophages and the Th1/Th2 paradigm”. In: *J Immunol* 164.12, pp. 6166–73.

– *This is the original article to have proposed the M1/M2 model I deride throughout this document. This is not to say that the model is useless, but that it is a distraction from proper understanding of macrophage biology.*

Mills, C. D. and K. Ley (2014). “M1 and M2 macrophages: the chicken and the egg of immunity”. In: *J Innate Immun* 6.6, pp. 716–26.

Minematsu, H., M. J. Shin, A. B. Celil Aydemir, K. O. Kim, S. A. Nizami, G. J. Chung, and F. Y. Lee (2011). “Nuclear presence of nuclear factor of activated T cells (NFAT) c3 and c4 is required for Toll-like receptor-activated innate inflammatory response of monocytes/macrophages”. In: *Cell Signal* 23.11, pp. 1785–93.

Minnikin, D. E., L. Kremer, L. G. Dover, and G. S. Besra (2002). “The methyl-branched fortifications of *Mycobacterium tuberculosis*”. In: *Chem Biol* 9.5, pp. 545–53.

Mishra, A., S. Akhtar, C. Jagannath, and A. Khan (2017a). “Pattern recognition receptors and coordinated cellular pathways involved in tuberculosis immunopathogenesis: Emerging concepts and perspectives”. In: *Mol Immunol* 87, pp. 240–248.

Mishra, B. B., R. R. Lovewell, A. J. Olive, G. Zhang, W. Wang, E. Eugenin, C. M. Smith, J. Y. Phuah, J. E. Long, M. L. Dubuke, S. G. Palace, J. D. Goguen, R. E. Baker, S. Nambi, R. Mishra, M. G. Booty, C. E. Baer, S. A. Shaffer, V. Dartois, B. A. McCormick, X. Chen, and C. M. Sassetti (2017b). “Nitric oxide prevents a pathogen-permissive granulocytic inflammation during tuberculosis”. In: *Nat Microbiol* 2, p. 17072.

- Miyake, Y., K. Toyonaga, D. Mori, S. Kakuta, Y. Hoshino, A. Oyamada, H. Yamada, K. Ono, M. Suyama, Y. Iwakura, Y. Yoshikai, and S. Yamasaki (2013). “C-type lectin MCL is an FcRgamma-coupled receptor that mediates the adjuvanticity of mycobacterial cord factor”. In: *Immunity* 38.5, pp. 1050–62.
- Miyazaki, Y., H. Imoto, L. C. Chen, and T. J. Wandless (2012). “Destabilizing domains derived from the human estrogen receptor”. In: *J Am Chem Soc* 134.9, pp. 3942–5.
- Mocsai, A., J. Ruland, and V. L. Tybulewicz (2010). “The SYK tyrosine kinase: a crucial player in diverse biological functions”. In: *Nat Rev Immunol* 10.6, pp. 387–402.
- Moench, C., A. P. Barreiros, M. Schuchmann, F. Bittinger, J. Thiesen, G. Hommel, I. Kraemer, and G. Otto (2007). “Tacrolimus monotherapy without steroids after liver transplantation—a prospective randomized double-blinded placebo-controlled trial”. In: *Am J Transplant* 7.6, pp. 1616–23.
- Mogues, T., M. Etzerodt, C. Hall, G. Engelich, J. H. Graversen, and K. L. Hartshorn (2004). “Ttranectin Binds to the Kringle 1-4 Form of Angiostatin and Modifies Its Functional Activity”. In: *J Biomed Biotechnol* 2004.2, pp. 73–78.
- Mojsilovic, S. S., S. Mojsilovic, V. H. Villar, and J. F. Santibanez (2021). “The Metabolic Features of Tumor-Associated Macrophages: Opportunities for Immunotherapy?” In: *Anal Cell Pathol (Amst)* 2021, p. 5523055.
- Moleri, S., G. Cappellano, G. Gaudenzi, S. Cermenati, F. Cotelli, D. S. Horner, and M. Beltrame (2011). “The HMGB protein gene family in zebrafish: Evolution and embryonic expression patterns”. In: *Gene Expr Patterns* 11.1-2, pp. 3–11.
- Moliva, J. I., A. P. Hossfeld, S. Sidiki, C. H. Canan, V. Dwivedi, G. Beamer, J. Turner, and J. B. Torrelles (2019). “Selective delipidation of *Mycobacterium bovis* BCG enables direct pulmonary vaccination and enhances protection against *Mycobacterium tuberculosis*”. In: *Mucosal Immunol* 12.3, pp. 805–815.
- Molloy, A., P. Laochumroonvorapong, and G. Kaplan (1994).** “Apoptosis, but not necrosis, of infected monocytes is coupled with killing of intracellular *Bacillus Calmette-Guerin*”. In: *J Exp Med* 180.4, pp. 1499–509.

- Monack, D. M., A. Mueller, and S. Falkow (2004). “Persistent bacterial infections: the interface of the pathogen and the host immune system”. In: *Nat Rev Microbiol* 2.9, pp. 747–65.
- Monroe, J. G. (2006). “ITAM-mediated tonic signalling through pre-BCR and BCR complexes”. In: *Nat Rev Immunol* 6.4, pp. 283–94.
- Monroy, V., A. Amador, B. Ruiz, P. Espinoza-Cueto, W. Xolalpa, R. Mancilla, and C. Espitia (2000). “Binding and activation of human plasminogen by *Mycobacterium tuberculosis*”. In: *Infect Immun* 68.7, pp. 4327–30.
- Montemagno, C. and G. Pages (2020). “Resistance to Anti-angiogenic Therapies: A Mechanism Depending on the Time of Exposure to the Drugs”. In: *Front Cell Dev Biol* 8, p. 584.
- Montesinos, M. C., A. Desai, J. F. Chen, H. Yee, M. A. Schwarzschild, J. S. Fink, and B. N. Cronstein (2002). “Adenosine promotes wound healing and mediates angiogenesis in response to tissue injury via occupancy of A(2A) receptors”. In: *Am J Pathol* 160.6, pp. 2009–18.
- Monticelli, S. and A. Rao (2002). “NFAT1 and NFAT2 are positive regulators of IL-4 gene transcription”. In: *Eur J Immunol* 32.10, pp. 2971–8.
– *An elegant study from Anjana Rao on the different roles of NFATC1 and NFATC2 and uncovered conflictual activity wherein NFATC1 repressed the transcription of IL-4 while NFATC2 activated transcription but that these effects were mediated by higher order regulatory mechanisms as constitutively active versions of these proteins were able to induce similar IL-4 transcription.*
- Monticelli, S., D. C. Solymar, and A. Rao (2004). “Role of NFAT proteins in IL-13 gene transcription in mast cells”. In: *J Biol Chem* 279.35, pp. 36210–8.
- Montoya-Rosales, A., P. Castro-Garcia, F. Torres-Juarez, J. A. Enciso-Moreno, and B. Rivas-Santiago (2016). “Glucose levels affect LL-37 expression in monocyte-derived macrophages altering the *Mycobacterium tuberculosis* intracellular growth control”. In: *Microb Pathog* 97, pp. 148–53.
- Moorlag, Sjcfm, N. Khan, B. Novakovic, E. Kaufmann, T. Jansen, R. van Crevel, M. Divangahi, and M. G. Netea (2020). “beta-Glucan Induces Protective Trained Immunity against *Mycobacterium tuberculosis* Infection: A Key Role for IL-1”. In: *Cell Rep* 31.7, p. 107634.

Mootoo, A., E. Stylianou, M. A. Arias, and R. Reljic (2009). “TNF-alpha in tuberculosis: a cytokine with a split personality”. In: *Inflamm Allergy Drug Targets* 8.1, pp. 53–62.

Moreno, M., V. Fernandez, J. M. Monllau, V. Borrell, C. Lerin, and N. de la Iglesia (2015). “Transcriptional Profiling of Hypoxic Neural Stem Cells Identifies Calcineurin-NFATc4 Signaling as a Major Regulator of Neural Stem Cell Biology”. In: *Stem Cell Reports* 5.2, pp. 157–65.

Moreno-Mateos, M. A., C. E. Vejnar, J. D. Beaudoin, J. P. Fernandez, E. K. Mis, M. K. Khokha, and A. J. Giraldez (2015). “CRISPRscan: designing highly efficient sgRNAs for CRISPR-Cas9 targeting *in vivo*”. In: *Nat Methods* 12.10, pp. 982–8.

Morgens, D. W., M. Wainberg, E. A. Boyle, O. Ursu, C. L. Araya, C. K. Tsui, M. S. Haney, G. T. Hess, K. Han, E. E. Jeng, A. Li, M. P. Snyder, W. J. Greenleaf, A. Kundaje, and M. C. Bassik (2017). “Genome-scale measurement of off-target activity using Cas9 toxicity in high-throughput screens”. In: *Nat Commun* 8, p. 15178.

Mosavari, N., A. Karimi, K. Tadayon, G. Shahhosseini, A. Zavarani Hosseini, and M. Babaie (2021). “Evaluation of Heating and Irradiation Methods for Production of Purified Protein Derivative (PPD) of *Mycobacterium tuberculosis*”. In: *Arch Razi Inst* 75.4, pp. 439–449.

Motta, V., F. Soares, T. Sun, and D. J. Philpott (2015). “NOD-like receptors: versatile cytosolic sentinels”. In: *Physiol Rev* 95.1, pp. 149–78.

Moutel, S., N. Bery, V. Bernard, L. Keller, E. Lemesre, A. de Marco, L. Ligat, J. C. Rain, G. Favre, A. Olichon, and F. Perez (2016). “NaLi-H1: A universal synthetic library of humanized nanobodies providing highly functional antibodies and intrabodies”. In: *Elife* 5.

Mtetwa, H. N., I. D. Amoah, S. Kumari, F. Bux, and P. Reddy (2022). “The source and fate of *Mycobacterium tuberculosis* complex in wastewater and possible routes of transmission”. In: *BMC Public Health* 22.1, p. 145.

Mueller, S. N. and R. N. Germain (2009). “Stromal cell contributions to the homeostasis and functionality of the immune system”. In: *Nat Rev Immunol* 9.9, pp. 618–29.

Muhammad, K., H. Alrefai, R. Marienfeld, D. A. Pham, K. Murti, A. K. Patra, A. Avots, V. Bukur, U. Sahin, E. Kondo, S. Klein-Hessling, and E. Serfling (2014). “NF- κ B factors control the induction of NFATc1 in B lymphocytes”. In: *Eur J Immunol* 44.11, pp. 3392–402.

– This work found that NF- κ B p50 and c-Rel were able to induce the transcription of NFATC1 to regulate the activity of peripheral B lymphocytes. This is yet another description of NF- κ B regulation of NFATC1 transcription and adds another layer of the interaction between these two closely related protein families.

Muhsin, M., J. Graham, and P. Kirkpatrick (2004). “Bevacizumab”. In: *Nat Rev Drug Discov* 3.12, pp. 995–6.

Mukhopadhyay, S. and A. L. Katzenstein (2010). “Biopsy findings in acute pulmonary histoplasmosis: unusual histologic features in 4 cases mimicking lymphomatoid granulomatosis”. In: *Am J Surg Pathol* 34.4, pp. 541–6.

Mukhtar, M., H. Ali, N. Ahmed, R. Munir, S. Talib, A. S. Khan, and R. Ambrus (2020). “Drug delivery to macrophages: a review of nano-therapeutics targeted approach for inflammatory disorders and cancer”. In: *Expert Opin Drug Deliv* 17.9, pp. 1239–1257.

Muller, M. R. and A. Rao (2010). “NFAT, immunity and cancer: a transcription factor comes of age”. In: *Nat Rev Immunol* 10.9, pp. 645–56.

Mullins, M. C., M. Hammerschmidt, P. Haffter, and C. Nusslein-Volhard (1994). “Large-scale mutagenesis in the zebrafish: in search of genes controlling development in a vertebrate”. In: *Curr Biol* 4.3, pp. 189–202.

Mullins, M. C., M. Hammerschmidt, D. A. Kane, J. Odenthal, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, P. Haffter, C. P. Heisenberg, Y. J. Jiang, R. N. Kelsh, and C. Nusslein-Volhard (1996). “Genes establishing dorsoventral pattern formation in the zebrafish embryo: the ventral specifying genes”. In: *Development* 123, pp. 81–93.

Mullins, M. C., J. Navajas Acedo, R. Priya, L. Solnica-Krezel, and S. W. Wilson (2021). “The zebrafish issue: 25 years on”. In: *Development* 148.24.

Murakami, M., S. Iwai, S. Hiratsuka, M. Yamauchi, K. Nakamura, Y. Iwakura, and M. Shibuya (2006). “Signaling of vascular endothelial growth factor receptor-1

tyrosine kinase promotes rheumatoid arthritis through activation of monocytes/-macrophages". In: *Blood* 108.6, pp. 1849–56.

Murakami, M., Y. Zheng, M. Hirashima, T. Suda, Y. Morita, J. Ooehara, H. Ema, G. H. Fong, and M. Shibuya (2008). "VEGFR1 tyrosine kinase signaling promotes lymphangiogenesis as well as angiogenesis indirectly via macrophage recruitment". In: *Arterioscler Thromb Vasc Biol* 28.4, pp. 658–64.

Murdoch, C. C., S. T. Espenschied, M. A. Matty, O. Mueller, D. M. Tobin, and J. F. Rawls (2019). "Intestinal Serum amyloid A suppresses systemic neutrophil activation and bactericidal activity in response to microbiota colonization". In: *PLoS Pathog* 15.3, e1007381.

Murdoch, C. C. and J. F. Rawls (2019). "Commensal Microbiota Regulate Vertebrate Innate Immunity-Insights From the Zebrafish". In: *Front Immunol* 10, p. 2100.

Murray, B. E. and B. Mederski-Samaroj (1983). "Transferable beta-lactamase. A new mechanism for *in vitro* penicillin resistance in *Streptococcus faecalis*". In: *J Clin Invest* 72.3, pp. 1168–71.

Murray, J. F. (2004). "A century of tuberculosis". In: *Am J Respir Crit Care Med* 169.11, pp. 1181–6.

Murray, P. J., J. E. Allen, S. K. Biswas, E. A. Fisher, D. W. Gilroy, S. Goerdt, S. Gordon, J. A. Hamilton, L. B. Ivashkiv, T. Lawrence, M. Locati, A. Mantovani, F. O. Martinez, J. L. Mege, D. M. Mosser, G. Natoli, J. P. Saeij, J. L. Schultze, K. A. Shirey, A. Sica, J. Suttles, I. Udalova, J. A. van Ginderachter, S. N. Vogel, and T. A. Wynn (2014). "Macrophage activation and polarization: nomenclature and experimental guidelines". In: *Immunity* 41.1, pp. 14–20.

Murray, P. J. and T. A. Wynn (2011). "Protective and pathogenic functions of macrophage subsets". In: *Nat Rev Immunol* 11.11, pp. 723–37.

Murray, Patrick Desmond Fitzgerald (1932). "The development *in vitro* of the blood of the early chick embryo". In: *Proceedings of the Royal Society of London. Series B, Containing Papers of a Biological Character* 111.773, pp. 497–521.

Mwandumba, H. C., D. G. Russell, M. H. Nyirenda, J. Anderson, S. A. White, M. E. Molyneux, and S. B. Squire (2004). "*Mycobacterium tuberculosis* resides in nonacid-

ified vacuoles in endocytically competent alveolar macrophages from patients with tuberculosis and HIV infection". In: *J Immunol* 172.7, pp. 4592–8.

Myllymaki, H., C. A. Bauerlein, and M. Ramet (2016). "The Zebrafish Breathes New Life into the Study of Tuberculosis". In: *Front Immunol* 7, p. 196.

Myllymaki, H., M. Niskanen, K. E. Oksanen, E. Sherwood, M. Ahava, M. Parikka, and M. Ramet (2017). "Identification of novel antigen candidates for a tuberculosis vaccine in the adult zebrafish (*Danio rerio*)". In: *PLoS One* 12.7, e0181942.

Na, Y. R., M. Stakenborg, S. H. Seok, and G. Matteoli (2019). "Macrophages in intestinal inflammation and resolution: a potential therapeutic target in IBD". In: *Nat Rev Gastroenterol Hepatol* 16.9, pp. 531–543.

Nacev, B. A., P. Grassi, A. Dell, S. M. Haslam, and J. O. Liu (2011). "The antifungal drug itraconazole inhibits vascular endothelial growth factor receptor 2 (VEGFR2) glycosylation, trafficking, and signaling in endothelial cells". In: *J Biol Chem* 286.51, pp. 44045–44056.

Nahid, P., S. R. Mase, G. B. Migliori, G. Sotgiu, G. H. Bothamley, J. L. Brozek, A. Cattamanchi, J. P. Cegielski, L. Chen, C. L. Daley, T. L. Dalton, R. Duarte, F. Fregonese, Jr. Horsburgh C. R., F. Ahmad Khan, F. Kheir, Z. Lan, A. Lardizabal, M. Lauzardo, J. M. Mangan, S. M. Marks, L. McKenna, D. Menzies, C. D. Mitnick, D. M. Nilsen, F. Parvez, C. A. Peloquin, A. Raftery, H. S. Schaaf, N. S. Shah, J. R. Starke, J. W. Wilson, J. M. Wortham, T. Chorba, and B. Seaworth (2019). "Treatment of Drug-Resistant Tuberculosis. An Official ATS/CDC/ERS/IDSA Clinical Practice Guideline". In: *Am J Respir Crit Care Med* 200.10, e93–e142.

Nahrendorf, M. and F. K. Swirski (2016). "Abandoning M1/M2 for a Network Model of Macrophage Function". In: *Circ Res* 119.3, pp. 414–7.

– *Presents an entirely new model for understanding macrophage biology within a network of possible activation states and is likely to be useful in the description of actual macrophage activity during homeostasis, development, and disease.*

Nair, S., J. P. Huynh, V. Lampropoulou, E. Loginicheva, E. Esaulova, A. P. Gounder, A. C. M. Boon, E. A. Schwarzkopf, T. R. Bradstreet, B. T. Edelson, M. N. Artyomov, C. L. Stallings, and M. S. Diamond (2018). "Irg1 expression in myeloid cells prevents immunopathology during *M. tuberculosis* infection". In: *J Exp Med* 215.4, pp. 1035–1045.

- Nakade, S., T. Tsubota, Y. Sakane, S. Kume, N. Sakamoto, M. Obara, T. Daimon, H. Sezutsu, T. Yamamoto, T. Sakuma, and K. T. Suzuki (2014). “Microhomology-mediated end-joining-dependent integration of donor DNA in cells and animals using TALENs and CRISPR/Cas9”. In: *Nat Commun* 5, p. 5560.
- Nakai, J., M. Ohkura, and K. Imoto (2001). “A high signal-to-noise Ca(2+) probe composed of a single green fluorescent protein”. In: *Nat Biotechnol* 19.2, pp. 137–41.
- Nakamura, K., T. Kinjo, S. Saijo, A. Miyazato, Y. Adachi, N. Ohno, J. Fujita, M. Kaku, Y. Iwakura, and K. Kawakami (2007). “Dectin-1 is not required for the host defense to *Cryptococcus neoformans*”. In: *Microbiol Immunol* 51.11, pp. 1115–9.
- Nakamura, R., A. Sene, A. Santeford, A. Gdoura, S. Kubota, N. Zapata, and R. S. Apte (2015). “IL10-driven STAT3 signalling in senescent macrophages promotes pathological eye angiogenesis”. In: *Nat Commun* 6, p. 7847.
- Nakata, T., M. Yasuda, M. Fujita, H. Kataoka, K. Kiura, H. Sano, and K. Shibata (2006). “CD14 directly binds to triacylated lipopeptides and facilitates recognition of the lipopeptides by the receptor complex of Toll-like receptors 2 and 1 without binding to the complex”. In: *Cell Microbiol* 8.12, pp. 1899–909.
- Nakaya, Y., J. Lilue, S. Stavrou, E. A. Moran, and S. R. Ross (2017). “AIM2-Like Receptors Positively and Negatively Regulate the Interferon Response Induced by Cytosolic DNA”. In: *mBio* 8.4.
- Nam, E. H., S. R. Park, and P. H. Kim (2010). “TGF-beta1 induces mouse dendritic cells to express VEGF and its receptor (Flt-1) under hypoxic conditions”. In: *Exp Mol Med* 42.9, pp. 606–13.
- Nankivell, B. J., C. H. P’Ng, P. J. O’Connell, and J. R. Chapman (2016). “Calcineurin Inhibitor Nephrotoxicity Through the Lens of Longitudinal Histology: Comparison of Cyclosporine and Tacrolimus Eras”. In: *Transplantation* 100.8, pp. 1723–31.
- Napione, Lucia, Maria Alvaro, and Federico Bussolino (2017). “VEGF-Mediated Signal Transduction in Tumor Angiogenesis”. In: IntechOpen. Chap. 13.
- Nasevicius, A., J. Larson, and S. C. Ekker (2000). “Distinct requirements for zebrafish angiogenesis revealed by a VEGF-A morphant”. In: *Yeast* 17.4, pp. 294–301.

- Nataraj, V., C. Varela, A. Javid, A. Singh, G. S. Besra, and A. Bhatt (2015). “Mycolic acids: deciphering and targeting the Achilles’ heel of the tubercle bacillus”. In: *Mol Microbiol* 98.1, pp. 7–16.
- Nathan, C. F., H. W. Murray, M. E. Wiebe, and B. Y. Rubin (1983). “Identification of interferon-gamma as the lymphokine that activates human macrophage oxidative metabolism and antimicrobial activity”. In: *J Exp Med* 158.3, pp. 670–89.
- Nazarova, E. V., C. R. Montague, L. Huang, T. La, D. Russell, and B. C. Van der Ven (2019). “The genetic requirements of fatty acid import by *Mycobacterium tuberculosis* within macrophages”. In: *Elife* 8.
- Neff, C. P., S. M. Atif, E. C. Logue, J. Siebert, C. Gorg, J. Lavelle, S. Fiorillo, H. Twigg, T. B. Campbell, A. P. Fontenot, and B. E. Palmer (2020). “HIV Infection Is Associated with Loss of Anti-Inflammatory Alveolar Macrophages”. In: *J Immunol* 205.9, pp. 2447–2455.
- Negishi-Koga, T. and H. Takayanagi (2009). “Ca²⁺-NFATc1 signaling is an essential axis of osteoclast differentiation”. In: *Immunol Rev* 231.1, pp. 241–56.
- Nehvi, I. B., N. Quadir, M. Khubaib, J. A. Sheikh, M. Shariq, K. Mohareer, S. Banerjee, S. A. Rahman, N. Z. Ehtesham, and S. E. Hasnain (2022). “ArgD of *Mycobacterium tuberculosis* is a functional N-acetylmornithine aminotransferase with moonlighting function as an effective immune modulator”. In: *Int J Med Microbiol* 312.1, p. 151544.
- Neri, D. and R. Bicknell (2005). “Tumour vascular targeting”. In: *Nat Rev Cancer* 5.6, pp. 436–46.
- Netea, M. G., J. Dominguez-Andres, L. B. Barreiro, T. Chavakis, M. Divangahi, E. Fuchs, L. A. B. Joosten, J. W. M. van der Meer, M. M. Mhlanga, W. J. M. Mulder, N. P. Riksen, A. Schlitzer, J. L. Schultze, C. Stabell Benn, J. C. Sun, R. J. Xavier, and E. Latz (2020). “Defining trained immunity and its role in health and disease”. In: *Nat Rev Immunol* 20.6, pp. 375–388.
- Neuhauss, S. C., L. Solnica-Krezel, A. F. Schier, F. Zwartkruis, D. L. Stemple, J. Malicki, S. Abdelilah, D. Y. Stainier, and W. Driever (1996). “Mutations affecting craniofacial development in zebrafish”. In: *Development* 123, pp. 357–67.
- Neuwirth, Erich (2022). “RColorBrewer: ColorBrewer Palettes”. In.

Neyrolles, O. and C. Guilhot (2011). “Recent advances in deciphering the contribution of *Mycobacterium tuberculosis* lipids to pathogenesis”. In: *Tuberculosis (Edinb)* 91.3, pp. 187–95.

Nguyen, L. and J. Pieters (2009). “Mycobacterial subversion of chemotherapeutic reagents and host defense tactics: challenges in tuberculosis drug development”. In: *Annu Rev Pharmacol Toxicol* 49, pp. 427–53.

Nguyen, T. V. A., R. M. Anthony, A. L. Banuls, T. V. A. Nguyen, D. H. Vu, and J. C. Alffenaar (2018). “Bedaquiline Resistance: Its Emergence, Mechanism, and Prevention”. In: *Clin Infect Dis* 66.10, pp. 1625–1630.

Ni, T. T., J. Lu, M. Zhu, L. A. Maddison, K. L. Boyd, L. Huskey, B. Ju, D. Hesselson, T. P. Zhong, P. S. Page-McCaw, D. Y. Stainier, and W. Chen (2012). “Conditional control of gene function by an invertible gene trap in zebrafish”. In: *Proc Natl Acad Sci U S A* 109.38, pp. 15389–94.

Nicenboim, J., G. Malkinson, T. Lupo, L. Asaf, Y. Sela, O. Mayseless, L. Gibbs-Bar, N. Senderovich, T. Hashimshony, M. Shin, A. Jerafi-Vider, I. Avraham-David, V. Krupalnik, R. Hofi, G. Almog, J. W. Astin, O. Golani, S. Ben-Dor, P. S. Crosier, W. Herzog, N. D. Lawson, J. H. Hanna, I. Yanai, and K. Yaniv (2015). “Lymphatic vessels arise from specialized angioblasts within a venous niche”. In: *Nature* 522.7554, pp. 56–61.

Nicholas, S. A., A. E. Oniku, and V. V. Sumbayev (2010). “Myeloid cell death associated with Toll-like receptor 7/8-mediated inflammatory response. Implication of ASK1, HIF-1 alpha, IL-1 beta and TNF-alpha”. In: *Mol Immunol* 48.1-3, pp. 240–7.

Nielsen, B. B., J. S. Kastrup, H. Rasmussen, T. L. Holtet, J. H. Graversen, M. Etzerodt, H. C. Thogersen, and I. K. Larsen (1997). “Crystal structure of tetranectin, a trimeric plasminogen-binding protein with an alpha-helical coiled coil”. In: *FEBS Lett* 412.2, pp. 388–96.

Nielsen, H., I. Clemmensen, and A. Kharazmi (1993). “Tetranectin: a novel secretory protein from human monocytes”. In: *Scand J Immunol* 37.1, pp. 39–42.

Nieves-Cintron, M., G. C. Amberg, M. F. Navedo, J. D. Molkentin, and L. F. Santana (2008). “The control of Ca²⁺ influx and NFATc3 signaling in arterial smooth muscle during hypertension”. In: *Proc Natl Acad Sci U S A* 105.40, pp. 15623–8.

Nieves-Cintron, M., G. C. Amberg, C. B. Nichols, J. D. Molkentin, and L. F. Santana (2007). “Activation of NFATc3 down-regulates the beta1 subunit of large conductance, calcium-activated K⁺ channels in arterial smooth muscle and contributes to hypertension”. In: *J Biol Chem* 282.5, pp. 3231–40.

Nishi, K., T. Oda, S. Takabuchi, S. Oda, K. Fukuda, T. Adachi, G. L. Semenza, K. Shingu, and K. Hirota (2008). “LPS induces hypoxia-inducible factor 1 activation in macrophage-differentiated cells in a reactive oxygen species-dependent manner”. In: *Antioxid Redox Signal* 10.5, pp. 983–95.

Nkwouano, V., S. Witkowski, N. Rehberg, R. Kalscheuer, N. Nausch, E. Mayatepek, and M. Jacobsen (2017). “A novel mycobacterial *in vitro* infection assay identifies differences of induced macrophage apoptosis between CD4+ and CD8+ T cells”. In: *PLoS One* 12.2, e0171817.

Noguchi, H., M. Matsushita, T. Okitsu, A. Moriwaki, K. Tomizawa, S. Kang, S. T. Li, N. Kobayashi, S. Matsumoto, K. Tanaka, N. Tanaka, and H. Matsui (2004). “A new cell-permeable peptide allows successful allogeneic islet transplantation in mice”. In: *Nat Med* 10.3, pp. 305–9.

Noll, H. (1956). “The chemistry of cord factor, a toxic glycolipid of *M. tuberculosis*”. In: *Bibl Tuberc* 10, pp. 149–83.

Noll, H., H. Bloch, J. Asselineau, and E. Lederer (1956). “The chemical structure of the cord factor of *Mycobacterium tuberculosis*”. In: *Biochim Biophys Acta* 20.2, pp. 299–309.

Norrmén, C., K. I. Ivanov, J. Cheng, N. Zanger, M. Delorenzi, M. Jaquet, N. Miura, P. Puolakkainen, V. Horsley, J. Hu, H. G. Augustin, S. Yla-Herttuala, K. Alitalo, and T. V. Petrova (2009). “FOXC2 controls formation and maturation of lymphatic collecting vessels through cooperation with NFATc1”. In: *J Cell Biol* 185.3, pp. 439–57.

North, R. J. and Y. J. Jung (2004). “Immunity to tuberculosis”. In: *Annu Rev Immunol* 22, pp. 599–623.

Northrop, J. P., S. N. Ho, L. Chen, D. J. Thomas, L. A. Timmerman, G. P. Nolan, A. Admon, and G. R. Crabtree (1994). “NF-AT components define a family of transcription factors targeted in T-cell activation”. In: *Nature* 369.6480, pp. 497–502.

- Noy, R. and J. W. Pollard (2014). “Tumor-associated macrophages: from mechanisms to therapy”. In: *Immunity* 41.1, pp. 49–61.
- Ntellias, P., L. Mavroeidis, S. Gkoura, I. Gazouli, A. L. Amylidi, A. Papadaki, G. Zarkavelis, D. Mauri, G. Karpathiou, E. Kolettas, A. Batistatou, and G. Pentheroudakis (2020). “Old Player-New Tricks: Non Angiogenic Effects of the VEGF/VEGFR Pathway in Cancer”. In: *Cancers (Basel)* 12.11.
- Nunes, P., D. Cornut, V. Bochet, U. Hasler, M. Oh-Hora, J. M. Waldburger, and N. Demaurex (2012). “STIM1 juxtaposes ER to phagosomes, generating Ca(2)(+) hotspots that boost phagocytosis”. In: *Curr Biol* 22.21, pp. 1990–7.
- Nusslein-Volhard, C. (2012). “The zebrafish issue of Development”. In: *Development* 139.22, pp. 4099–103.
- Odegaard, J. I. and A. Chawla (2011). “Alternative macrophage activation and metabolism”. In: *Annu Rev Pathol* 6, pp. 275–97.
- Odenthal, J., P. Haffter, E. Vogelsang, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, R. N. Kelsh, M. C. Mullins, R. M. Warga, M. L. Allende, E. S. Weinberg, and C. Nusslein-Volhard (1996a). “Mutations affecting the formation of the notochord in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 103–15.
- Odenthal, J., K. Rossnagel, P. Haffter, R. N. Kelsh, E. Vogelsang, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, M. C. Mullins, and C. Nusslein-Volhard (1996b). “Mutations affecting xanthophore pigmentation in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 391–8.
- Odermatt, P. D., M. T. M. Hannebelle, H. A. Eskandarian, A. P. Nievergelt, J. D. McKinney, and G. E. Fantner (2020). “Overlapping and essential roles for molecular and mechanical mechanisms in mycobacterial cell division”. In: *Nat Phys* 16.1, pp. 57–62.
- Odero, M. D., N. J. Zeleznik-Le, V. Chinwalla, and J. D. Rowley (2000). “Cytogenetic and molecular analysis of the acute monocytic leukemia cell line THP-1 with an MLL-AF9 translocation”. In: *Genes Chromosomes Cancer* 29.4, pp. 333–8.

Oeckinghaus, A. and S. Ghosh (2009). “The NF-kappaB family of transcription factors and its regulation”. In: *Cold Spring Harb Perspect Biol* 1.4, a000034.

Oehlers, S. H., M. R. Cronan, R. W. Beerman, M. G. Johnson, J. Huang, C. D. Kontos, J. E. Stout, and D. M. Tobin (2017). “Infection-Induced Vascular Permeability Aids Mycobacterial Growth”. In: *J Infect Dis* 215.5, pp. 813–817.

Oehlers, S. H., M. R. Cronan, N. R. Scott, M. I. Thomas, K. S. Okuda, E. M. Walton, R. W. Beerman, P. S. Crosier, and D. M. Tobin (2015). “Interception of host angiogenic signalling limits mycobacterial growth”. In: *Nature* 517.7536, pp. 612–5.

– This paper from the Tobin Lab was the first in a pioneering series of publications in the field of tuberculosis pathogenesis at large to address the nature of the angiogenesis phenomenon during infection. This has set the stage for the primary work I have presented in Chapter 3 and is foundational to our understanding of mycobacterial engagement with the vasculature.

Oestreich, K. J., H. Yoon, R. Ahmed, and J. M. Boss (2008). “NFATc1 regulates PD-1 expression upon T cell activation”. In: *J Immunol* 181.7, pp. 4832–9.

O’Garra, A., P. S. Redford, F. W. McNab, C. I. Bloom, R. J. Wilkinson, and M. P. Berry (2013). “The immune response in tuberculosis”. In: *Annu Rev Immunol* 31, pp. 475–527.

Ogle, D., J. Doll, P. Wheeler, and A. Dinno (2022). “FSA: Simple Fisheries Stock Assessment Methods”. In.

Ogryzko, N. V., A. Lewis, H. L. Wilson, A. H. Meijer, S. A. Renshaw, and P. M. Elks (2019). “Hif-1alpha-Induced Expression of IL-1beta Protects against Mycobacterial Infection in Zebrafish”. In: *J Immunol* 202.2, pp. 494–502.

Oh, C. W., J. Hoover-Plow, and E. F. Plow (2003). “The role of plasminogen in angiogenesis *in vivo*”. In: *J Thromb Haemost* 1.8, pp. 1683–7.

Ohno, H., G. Zhu, V. P. Mohan, D. Chu, S. Kohno, Jr. Jacobs W. R., and J. Chan (2003). “The effects of reactive nitrogen intermediates on gene expression in *Mycobacterium tuberculosis*”. In: *Cell Microbiol* 5.9, pp. 637–48.

Ohtsubo, K. and J. D. Marth (2006). “Glycosylation in cellular mechanisms of health and disease”. In: *Cell* 126.5, pp. 855–67.

- Oikawa, T., A. Nakamura, N. Onishi, T. Yamada, K. Matsuo, and H. Saya (2013). “Acquired expression of NFATc1 downregulates E-cadherin and promotes cancer cell invasion”. In: *Cancer Res* 73.16, pp. 5100–9.
- Okikawa, S., Y. Morine, Y. Saito, S. Yamada, K. Tokuda, H. Teraoku, K. Miyazaki, S. Yamashita, T. Ikemoto, S. Imura, and M. Shimada (2022). “Inhibition of the VEGF signaling pathway attenuates tumor-associated macrophage activity in liver cancer”. In: *Oncol Rep* 47.4.
- Okuda, K. S., J. W. Astin, J. P. Misa, M. V. Flores, K. E. Crosier, and P. S. Crosier (2012). “lyve1 expression reveals novel lymphatic vessels and new mechanisms for lymphatic vessel development in zebrafish”. In: *Development* 139.13, pp. 2381–91.
- Okuda, K. S., J. P. Misa, S. H. Oehlers, C. J. Hall, F. Ellett, S. Alasmari, G. J. Lieschke, K. E. Crosier, P. S. Crosier, and J. W. Astin (2015). “A zebrafish model of inflammatory lymphangiogenesis”. In: *Biol Open* 4.10, pp. 1270–80.
- O’Leary, B., R. S. Finn, and N. C. Turner (2016). “Treating cancer with selective CDK4/6 inhibitors”. In: *Nat Rev Clin Oncol* 13.7, pp. 417–30.
- Olive, A. J., C. M. Smith, M. C. Kiritsy, and C. M. Sassetti (2018). “The Phagocyte Oxidase Controls Tolerance to *Mycobacterium tuberculosis* Infection”. In: *J Immunol* 201.6, pp. 1705–1716.
- Olsson, A. K., A. Dimberg, J. Kreuger, and L. Claesson-Welsh (2006). “VEGF receptor signalling - in control of vascular function”. In: *Nat Rev Mol Cell Biol* 7.5, pp. 359–71.
- Onder, L., U. Morbe, N. Pikor, M. Novkovic, H. W. Cheng, T. Hehlgans, K. Pfeffer, B. Becher, A. Waisman, T. Rulicke, J. Gommerman, C. G. Mueller, S. Sawa, E. Scandella, and B. Ludewig (2017). “Lymphatic Endothelial Cells Control Initiation of Lymph Node Organogenesis”. In: *Immunity* 47.1, 80–92 e4.
- O’Neill, L. A., D. Golenbock, and A. G. Bowie (2013). “The history of Toll-like receptors - redefining innate immunity”. In: *Nat Rev Immunol* 13.6, pp. 453–60.
- O’Neill, L. A. J. and M. G. Netea (2020). “BCG-induced trained immunity: can it offer protection against COVID-19?” In: *Nat Rev Immunol* 20.6, pp. 335–337.

- Onyewu, C., Jr. Wormley F. L., J. R. Perfect, and J. Heitman (2004). “The calcineurin target, Crz1, functions in azole tolerance but is not required for virulence of *Candida albicans*”. In: *Infect Immun* 72.12, pp. 7330–3.
- Oosterhof, N., E. Boddeke, and T. J. van Ham (2015). “Immune cell dynamics in the CNS: Learning from the zebrafish”. In: *Glia* 63.5, pp. 719–35.
- Opazo, J. C., G. T. Butts, M. F. Nery, J. F. Storz, and F. G. Hoffmann (2013). “Whole-genome duplication and the functional diversification of teleost fish hemoglobins”. In: *Mol Biol Evol* 30.1, pp. 140–53.
- Orme, I. M. (1998). “The immunopathogenesis of tuberculosis: a new working hypothesis”. In: *Trends Microbiol* 6.3, pp. 94–7.
- (2014). “A new unifying theory of the pathogenesis of tuberculosis”. In: *Tuberculosis (Edinb)* 94.1, pp. 8–14.
- Orme, I. M. and R. J. Basaraba (2014). “The formation of the granuloma in tuberculosis infection”. In: *Semin Immunol* 26.6, pp. 601–9.
- Orme, I. M., R. T. Robinson, and A. M. Cooper (2015). “The balance between protective and pathogenic immune responses in the TB-infected lung”. In: *Nat Immunol* 16.1, pp. 57–63.
- Osada-Oka, M., N. Goda, H. Saiga, M. Yamamoto, K. Takeda, Y. Ozeki, T. Yamaguchi, T. Soga, Y. Tateishi, K. Miura, D. Okuzaki, K. Kobayashi, and S. Matsumoto (2019). “Metabolic adaptation to glycolysis is a basic defense mechanism of macrophages for *Mycobacterium tuberculosis* infection”. In: *Int Immunol* 31.12, pp. 781–793.
- Osborne, R. (2013). “First novel anti-tuberculosis drug in 40 years”. In: *Nat Biotechnol* 31.2, pp. 89–91.
- Osherov, N. and R. Ben-Ami (2016). “Modulation of Host Angiogenesis as a Microbial Survival Strategy and Therapeutic Target”. In: *PLoS Pathog* 12.4, e1005479.
- Osoegawa, K., M. Tateno, P. Y. Woon, E. Frengen, A. G. Mammoser, J. J. Catanese, Y. Hayashizaki, and P. J. de Jong (2000). “Bacterial artificial chromosome libraries for mouse sequencing and functional analysis”. In: *Genome Res* 10.1, pp. 116–28.

- Oukka, M., I. C. Ho, F. C. de la Brousse, T. Hoey, M. J. Grusby, and L. H. Glimcher (1998). “The transcription factor NFAT4 is involved in the generation and survival of T cells”. In: *Immunity* 9.3, pp. 295–304.
- Ouyang, W., S. Rutz, N. K. Crellin, P. A. Valdez, and S. G. Hymowitz (2011). “Regulation and functions of the IL-10 family of cytokines in inflammation and disease”. In: *Annu Rev Immunol* 29, pp. 71–109.
- Pabinger, S., S. Rodiger, A. Kriegner, K. Vierlinger, and A. Weinhausel (2014). “A survey of tools for the analysis of quantitative PCR (qPCR) data”. In: *Biomol Detect Quantif* 1.1, pp. 23–33.
- Pacheco, S. A., F. F. Hsu, K. M. Powers, and G. E. Purdy (2013). “MmpL11 protein transports mycolic acid-containing lipids to the mycobacterial cell wall and contributes to biofilm formation in *Mycobacterium smegmatis*”. In: *J Biol Chem* 288.33, pp. 24213–22.
- Pack, M., L. Solnica-Krezel, J. Malicki, S. C. Neuhauss, A. F. Schier, D. L. Stemple, W. Driever, and M. C. Fishman (1996). “Mutations affecting development of zebrafish digestive organs”. In: *Development* 123, pp. 321–8.
- Packer, J. S., Q. Zhu, C. Huynh, P. Sivaramakrishnan, E. Preston, H. Dueck, D. Stefanik, K. Tan, C. Trapnell, J. Kim, R. H. Waterston, and J. I. Murray (2019). “A lineage-resolved molecular atlas of *C. elegans* embryogenesis at single-cell resolution”. In: *Science* 365.6459.
- Pagan, A. J., L. J. Lee, J. Edwards-Hicks, C. B. Moens, D. M. Tobin, E. M. Busch-Nentwich, E. L. Pearce, and L. Ramakrishnan (2022). “mTOR-regulated mitochondrial metabolism limits *Mycobacterium*-induced cytotoxicity”. In: *Cell* 185.20, 3720–3738 e13.
– This brand new work identified an interesting role for mTOR in moderating ESAT6-induced macrophage necrosis through regulation of mitochondrial oxidative phosphorylation. Based on our general understandings of macrophage biology, this argues for a hypersusceptible role for “M1” macrophages during infection as these are glycolytically shifted and incapable of oxidative phosphorylation.
- Pagan, A. J. and L. Ramakrishnan (2018).** “The Formation and Function of Granulomas”. In: *Annu Rev Immunol* 36, pp. 639–665.

– The definitive granuloma review and essential reading for all interested. Indispensable to the field at large in synthesizing the complex factors that contribute to the unique biology of the granuloma in tuberculosis disease.

Pagan, A. J., C. T. Yang, J. Cameron, L. E. Swaim, F. Ellett, G. J. Lieschke, and L. Ramakrishnan (2015). “Myeloid Growth Factors Promote Resistance to Mycobacterial Infection by Curtailing Granuloma Necrosis through Macrophage Replenishment”. In: *Cell Host Microbe* 18.1, pp. 15–26.

Pahl, H. L. (1999). “Activators and target genes of Rel/NF-kappaB transcription factors”. In: *Oncogene* 18.49, pp. 6853–66.

Pai, M., M. A. Behr, D. Dowdy, K. Dheda, M. Divangahi, C. C. Boehme, A. Ginsberg, S. Swaminathan, M. Spigelman, H. Getahun, D. Menzies, and M. Raviglione (2016). “Tuberculosis”. In: *Nat Rev Dis Primers* 2, p. 16076.

Pai, M., T. Kasaeva, and S. Swaminathan (2022). “Covid-19’s Devastating Effect on Tuberculosis Care - A Path to Recovery”. In: *N Engl J Med* 386.16, pp. 1490–1493.

Pajuelo, D., N. Gonzalez-Juarbe, and M. Niederweis (2020). “NAD hydrolysis by the tuberculosis necrotizing toxin induces lethal oxidative stress in macrophages”. In: *Cell Microbiol* 22.1, e13115.

Pajuelo, D., N. Gonzalez-Juarbe, U. Tak, J. Sun, C. J. Orihuela, and M. Niederweis (2018). “NAD(+) Depletion Triggers Macrophage Necroptosis, a Cell Death Pathway Exploited by *Mycobacterium tuberculosis*”. In: *Cell Rep* 24.2, pp. 429–440.

Pajuelo, D., U. Tak, L. Zhang, O. Danilchanka, A. D. Tischler, and M. Niederweis (2021). “Toxin secretion and trafficking by *Mycobacterium tuberculosis*”. In: *Nat Commun* 12.1, p. 6592.

Palazon, A., A. W. Goldrath, V. Nizet, and R. S. Johnson (2014). “HIF transcription factors, inflammation, and immunity”. In: *Immunity* 41.4, pp. 518–28.

Paleolog, E. M. (2002). “Angiogenesis in rheumatoid arthritis”. In: *Arthritis Res* 4 Suppl 3, S81–90.

Palmer, M. V., M. D. Welsh, and J. M. Hostetter (2011). “Mycobacterial diseases of animals”. In: *Vet Med Int* 2011, p. 292469.

- Palmer, T. (2017). "Structural biology: Mycobacterial ESX secrets revealed". In: *Nat Microbiol* 2, p. 17074.
- Palmieri, E. M., C. McGinity, D. A. Wink, and D. W. McVicar (2020). "Nitric Oxide in Macrophage Immunometabolism: Hiding in Plain Sight". In: *Metabolites* 10.11.
- Panagos, P. G., K. P. Dobrinski, X. Chen, A. W. Grant, D. Traver, J. Y. Djeu, S. Wei, and J. A. Yoder (2006). "Immune-related, lectin-like receptors are differentially expressed in the myeloid and lymphoid lineages of zebrafish". In: *Immunogenetics* 58.1, pp. 31–40.
- Pandey, A., C. Shen, S. Feng, and S. M. Man (2021). "Cell biology of inflammasome activation". In: *Trends Cell Biol* 31.11, pp. 924–939.
- Pandey, A. K. and C. M. Sasse (2008). "Mycobacterial persistence requires the utilization of host cholesterol". In: *Proc Natl Acad Sci U S A* 105.11, pp. 4376–80.
- Papadopoulos, N., J. Martin, Q. Ruan, A. Rafique, M. P. Rosconi, E. Shi, E. A. Pyles, G. D. Yancopoulos, N. Stahl, and S. J. Wiegand (2012). "Binding and neutralization of vascular endothelial growth factor (VEGF) and related ligands by VEGF Trap, ranibizumab and bevacizumab". In: *Angiogenesis* 15.2, pp. 171–85.
- Papon, N., G. Nevez, S. Le Gal, C. Vigneau, F. Robert-Gangneux, J. P. Bouchara, O. A. Cornely, D. W. Denning, and J. P. Gangneux (2021). "Fungal infections in transplant recipients: pros and cons of immunosuppressive and antimicrobial treatment". In: *Lancet Microbe* 2.1, e6–e8.
- Parichy, D. M. (2015). "Advancing biology through a deeper understanding of zebrafish ecology and evolution". In: *Elife* 4.
- Parisot, T. J. (1958). "Tuberculosis of fish; a review of the literature with a description of the disease in salmonoid fish". In: *Bacteriol Rev* 22.4, pp. 240–5.
- Park, J., D. S. Levic, K. D. Sumigray, J. Bagwell, O. Eroglu, C. L. Block, C. Eroglu, R. Barry, C. R. Lickwar, J. F. Rawls, S. A. Watts, T. Lechler, and M. Bagnat (2019). "Lysosome-Rich Enterocytes Mediate Protein Absorption in the Vertebrate Gut". In: *Dev Cell* 51.1, 7–20 e6.
- Park, J. S., I. K. Kim, S. Han, I. Park, C. Kim, J. Bae, S. J. Oh, S. Lee, J. H. Kim, D. C. Woo, Y. He, H. G. Augustin, I. Kim, D. Lee, and G. Y. Koh (2016).

“Normalization of Tumor Vessels by Tie2 Activation and Ang2 Inhibition Enhances Drug Delivery and Produces a Favorable Tumor Microenvironment”. In: *Cancer Cell* 30.6, pp. 953–967.

Park, K. S., J. Lee, S. C. Jang, S. R. Kim, M. H. Jang, J. Lotvall, Y. K. Kim, and Y. S. Gho (2013). “Pulmonary inflammation induced by bacteria-free outer membrane vesicles from *Pseudomonas aeruginosa*”. In: *Am J Respir Cell Mol Biol* 49.4, pp. 637–45.

Park, Y. J., S. A. Yoo, M. Kim, and W. U. Kim (2020). “The Role of Calcium-Calcineurin-NFAT Signaling Pathway in Health and Autoimmune Diseases”. In: *Front Immunol* 11, p. 195.

Park-Windhol, C. and P. A. D’Amore (2016). “Disorders of Vascular Permeability”. In: *Annu Rev Pathol* 11, pp. 251–81.

Parker, L. and D. Y. Stainier (1999). “Cell-autonomous and non-autonomous requirements for the zebrafish gene cloche in hematopoiesis”. In: *Development* 126.12, pp. 2643–51.

Patin, E. C., A. C. Geffken, S. Willcocks, C. Leschczyk, A. Haas, F. Nimmerjahn, R. Lang, T. H. Ward, and U. E. Schaible (2017). “Trehalose dimycolate interferes with FcγR-mediated phagosome maturation through Mincle, SHP-1 and FcγRIIB signalling”. In: *PLoS One* 12.4, e0174973.

Patin, E. C., S. J. Orr, and U. E. Schaible (2017). “Macrophage Inducible C-Type Lectin As a Multifunctional Player in Immunity”. In: *Front Immunol* 8, p. 861.

Patton, E. E., H. R. Widlund, J. L. Kutok, K. R. Kopani, J. F. Amatruda, R. D. Murphey, S. Berghmans, E. A. Mayhall, D. Traver, C. D. Fletcher, J. C. Aster, S. R. Granter, A. T. Look, C. Lee, D. E. Fisher, and L. I. Zon (2005). “BRAF mutations are sufficient to promote nevi formation and cooperate with p53 in the genesis of melanoma”. In: *Curr Biol* 15.3, pp. 249–54.

Patton, E. E., L. I. Zon, and D. M. Langenau (2021). “Zebrafish disease models in drug discovery: from preclinical modelling to clinical trials”. In: *Nat Rev Drug Discov* 20.8, pp. 611–628.

Paulson, T. (2013). “Epidemiology: A mortal foe”. In: *Nature* 502.7470, S2–3.

- Pavlath, G. K. and V. Horsley (2003). “Cell fusion in skeletal muscle—central role of NFATC2 in regulating muscle cell size”. In: *Cell Cycle* 2.5, pp. 420–3.
- Pazhakh, V. and G. J. Lieschke (2018). “Hematopoietic growth factors: the scenario in zebrafish”. In: *Growth Factors* 36.5-6, pp. 196–212.
- Peace, C. G. and L. A. O’Neill (2022). “The role of itaconate in host defense and inflammation”. In: *J Clin Invest* 132.2.
- Peddireddy, V., S. N. Doddam, and N. Ahmed (2017). “Mycobacterial Dormancy Systems and Host Responses in Tuberculosis”. In: *Front Immunol* 8, p. 84.
- Pedelacq, J. D., S. Cabantous, T. Tran, T. C. Terwilliger, and G. S. Waldo (2006). “Engineering and characterization of a superfolder green fluorescent protein”. In: *Nat Biotechnol* 24.1, pp. 79–88.
- Pedigo, C. E., G. M. Ducasa, F. Leclercq, A. Sloan, A. Mitrofanova, T. Hashmi, J. Molina-David, M. Ge, M. I. Lassenius, C. Forsblom, M. Lehto, P. H. Groop, M. Kretzler, S. Eddy, S. Martini, H. Reich, P. Wahl, G. Ghiggeri, C. Faul, 3rd Burke G. W., O. Kretz, T. B. Huber, A. J. Mendez, S. Merscher, and A. Fornoni (2016). “Local TNF causes NFATc1-dependent cholesterol-mediated podocyte injury”. In: *J Clin Invest* 126.9, pp. 3336–50.
- Pena, J. C. and W. Z. Ho (2015). “Monkey models of tuberculosis: lessons learned”. In: *Infect Immun* 83.3, pp. 852–62.
- Peng, S. L., A. J. Gerth, A. M. Ranger, and L. H. Glimcher (2001). “NFATc1 and NFATc2 together control both T and B cell activation and differentiation”. In: *Immunity* 14.1, pp. 13–20.
- Pepin, F., N. Bertos, J. Laferriere, S. Sadekova, M. Souleimanova, H. Zhao, G. Finak, S. Meterissian, M. T. Hallett, and M. Park (2012). “Gene-expression profiling of microdissected breast cancer microvasculature identifies distinct tumor vascular subtypes”. In: *Breast Cancer Res* 14.4, R120.
- Pereira, A. C., B. Ramos, A. C. Reis, and M. V. Cunha (2020). “Non-Tuberculous Mycobacteria: Molecular and Physiological Bases of Virulence and Adaptation to Ecological Niches”. In: *Microorganisms* 8.9.

- Pereira, T. A., G. Xie, S. S. Choi, W. K. Syn, I. Voieta, J. Lu, I. S. Chan, M. Swiderska, K. B. Amaral, C. M. Antunes, W. E. Secor, R. P. Witek, J. R. Lambertucci, F. L. Pereira, and A. M. Diehl (2013). “Macrophage-derived Hedgehog ligands promotes fibrogenic and angiogenic responses in human schistosomiasis mansoni”. In: *Liver Int* 33.1, pp. 149–61.
- Perez, R. L., J. Roman, S. Roser, C. Little, M. Olsen, J. Indrigo, R. L. Hunter, and J. K. Actor (2000). “Cytokine message and protein expression during lung granuloma formation and resolution induced by the mycobacterial cord factor trehalose-6,6'-dimycolate”. In: *J Interferon Cytokine Res* 20.9, pp. 795–804.
- Perkins, J. R., J. M. Dawes, S. B. McMahon, D. L. Bennett, C. Orengo, and M. Kohl (2012). “ReadqPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data”. In: *BMC Genomics* 13, p. 296.
- Pernas, L. (2021). “Cellular metabolism in the defense against microbes”. In: *J Cell Sci* 134.5.
- Perotti, V., P. Baldassari, A. Molla, C. Vegetti, I. Bersani, A. Maurichi, M. Santinami, A. Anichini, and R. Mortarini (2016). “NFATc2 is an intrinsic regulator of melanoma dedifferentiation”. In: *Oncogene* 35.22, pp. 2862–72.
- Perry, R. D. and J. D. Fetherston (1997). “*Yersinia pestis*—etiologic agent of plague”. In: *Clin Microbiol Rev* 10.1, pp. 35–66.
- Pestel, J., R. Ramadass, S. Gauvrit, C. Helker, W. Herzog, and D. Y. Stainier (2016). “Real-time 3D visualization of cellular rearrangements during cardiac valve formation”. In: *Development* 143.12, pp. 2217–27.
- Peters, W., J. G. Cyster, M. Mack, D. Schlondorff, A. J. Wolf, J. D. Ernst, and I. F. Charo (2004). “CCR2-dependent trafficking of F4/80dim macrophages and CD11cdim/intermediate dendritic cells is crucial for T cell recruitment to lungs infected with *Mycobacterium tuberculosis*”. In: *J Immunol* 172.12, pp. 7647–53.
- Peters, W. and J. D. Ernst (2003). “Mechanisms of cell recruitment in the immune response to *Mycobacterium tuberculosis*”. In: *Microbes Infect* 5.2, pp. 151–8.

- Peters, W., H. M. Scott, H. F. Chambers, J. L. Flynn, I. F. Charo, and J. D. Ernst (2001). "Chemokine receptor 2 serves an early and essential role in resistance to *Mycobacterium tuberculosis*". In: *Proc Natl Acad Sci U S A* 98.14, pp. 7958–63.
- Peterson, R. T. and C. A. Macrae (2012). "Systematic approaches to toxicology in the zebrafish". In: *Annu Rev Pharmacol Toxicol* 52, pp. 433–53.
- Petit, J., E. C. Bailey, R. T. Wheeler, C. A. F. de Oliveira, M. Forlenza, and G. F. Wiegertjes (2019). "Studies Into beta-Glucan Recognition in Fish Suggests a Key Role for the C-Type Lectin Pathway". In: *Front Immunol* 10, p. 280.
- Peuker, K., A. Strigli, D. V. F. Tauriello, A. Hendricks, W. von Schonfels, G. Burmeister, M. Brosch, A. Herrmann, S. Kruger, J. Nitsche, L. Juznic, M. M. Geissler, A. Hiergeist, A. Gessner, J. Wirbel, R. P. Ponnudurai, A. Tunger, R. Wehner, D. E. Stange, J. Weitz, D. E. Aust, G. B. Baretton, M. Schmitz, C. Rocken, J. Hampe, S. Hinz, G. Zeller, T. Chavakis, C. Schafmayer, E. Batlle, and S. Zeissig (2022). "Microbiota-dependent activation of the myeloid calcineurin-NFAT pathway inhibits B7H3- and B7H4-dependent anti-tumor immunity in colorectal cancer".** In: *Immunity* 55.4, 701–717 e7.
- This complex integration of microbiota, immune cell signaling, and colorectal cancer reveals an important role for microbiome-dependent activation of NFAT within myeloid cells to drive IL-6 production that increases susceptibility to colorectal cancer. Inhibition of this pathway results in improved adaptive immune responses, arguing that NFAT signaling inhibits effective antitumor immunity and supporting approaches that target myeloid NFAT signaling in improving anti-tumor chemotherapy.*
- Pfuetze, K. H. and M. M. Pyle (1949). "Streptomycin in the treatment of tuberculosis". In: *J Am Med Assoc* 139.10, pp. 634–9.
- Pham, P. H., W. W. L. Tong, E. Misk, G. Jones, J. S. Lumsden, and N. C. Bols (2017). "Atlantic salmon endothelial cells from the heart were more susceptible than fibroblasts from the bulbus arteriosus to four RNA viruses but protected from two viruses by dsRNA pretreatment". In: *Fish Shellfish Immunol* 70, pp. 214–227.
- Phan, A. T., A. W. Goldrath, and C. K. Glass (2017). "Metabolic and Epigenetic Coordination of T Cell and Macrophage Immunity". In: *Immunity* 46.5, pp. 714–729.

- Phennicie, R. T., M. J. Sullivan, J. T. Singer, J. A. Yoder, and C. H. Kim (2010). “Specific resistance to *Pseudomonas aeruginosa* infection in zebrafish is mediated by the cystic fibrosis transmembrane conductance regulator”. In: *Infect Immun* 78.11, pp. 4542–50.
- Philip, A. M., Y. Wang, A. Mauro, S. El-Rass, J. C. Marshall, W. L. Lee, A. S. Slutsky, C. C. dosSantos, and X. Y. Wen (2017). “Development of a zebrafish sepsis model for high-throughput drug discovery”. In: *Mol Med* 23, pp. 134–148.
- Philips, J. A. and J. D. Ernst (2012). “Tuberculosis pathogenesis and immunity”. In: *Annu Rev Pathol* 7, pp. 353–84.
- Pierre, K. B., C. M. Jones, J. M. Pierce, I. B. Nicoud, T. M. Earl, and R. S. Chari (2009). “NFAT4 deficiency results in incomplete liver regeneration following partial hepatectomy”. In: *J Surg Res* 154.2, pp. 226–33.
- Pieters, J. (2008). “*Mycobacterium tuberculosis* and the macrophage: maintaining a balance”. In: *Cell Host Microbe* 3.6, pp. 399–407.
- Pieters, J. and J. Gatfield (2002). “Hijacking the host: survival of pathogenic mycobacteria inside macrophages”. In: *Trends Microbiol* 10.3, pp. 142–6.
- Pilla, D. M., J. A. Hagar, A. K. Haldar, A. K. Mason, D. Degrandi, K. Pfeffer, R. K. Ernst, M. Yamamoto, E. A. Miao, and J. Coers (2014). “Guanylate binding proteins promote caspase-11-dependent pyroptosis in response to cytoplasmic LPS”. In: *Proc Natl Acad Sci U S A* 111.16, pp. 6046–51.
- Pimentel, H., N. L. Bray, S. Puente, P. Melsted, and L. Pachter (2017). “Differential analysis of RNA-seq incorporating quantification uncertainty”. In: *Nat Methods* 14.7, pp. 687–690.
- Pineros, A. R., L. W. Campos, D. M. Fonseca, T. B. Bertolini, A. F. Gembre, R. Q. Prado, J. C. Alves-Filho, S. G. Ramos, M. Russo, and V. L. Bonato (2017). “M2 macrophages or IL-33 treatment attenuate ongoing *Mycobacterium tuberculosis* infection”. In: *Sci Rep* 7, p. 41240.
- Piotrowski, T., T. F. Schilling, M. Brand, Y. J. Jiang, C. P. Heisenberg, D. Beuchle, H. Grandel, F. J. van Eeden, M. Furutani-Seiki, M. Granato, P. Haffter, M. Hammerschmidt, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, R. M. Warga,

and C. Nusslein-Volhard (1996). “Jaw and branchial arch mutants in zebrafish II: anterior arches and cartilage differentiation”. In: *Development* 123, pp. 345–56.

Pisu, D., L. Huang, J. K. Grenier, and D. G. Russell (2020). “Dual RNA-Seq of Mtb-Infected Macrophages *in vivo* Reveals Ontologically Distinct Host-Pathogen Interactions”. In: *Cell Rep* 30.2, 335–350 e4.

Pitarque, S., J. L. Herrmann, J. L. Duteyrat, M. Jackson, G. R. Stewart, F. Lecointe, B. Payre, O. Schwartz, D. B. Young, G. Marchal, P. H. Lagrange, G. Puzo, B. Gicquel, J. Nigou, and O. Neyrolles (2005). “Deciphering the molecular bases of *Mycobacterium tuberculosis* binding to the lectin DC-SIGN reveals an underestimated complexity”. In: *Biochem J* 392.Pt 3, pp. 615–24.

Platnich, J. M. and D. A. Muruve (2019). “NOD-like receptors and inflammasomes: A review of their canonical and non-canonical signaling pathways”. In: *Arch Biochem Biophys* 670, pp. 4–14.

Pletz, M. W., S. Hagel, and C. Forstner (2017). “Who benefits from antimicrobial combination therapy?” In: *Lancet Infect Dis* 17.7, pp. 677–678.

Ploemacher, T., W. R. Faber, H. Menke, V. Rutten, and T. Pieters (2020). “Reservoirs and transmission routes of leprosy; A systematic review”. In: *PLoS Negl Trop Dis* 14.4, e0008276.

Plumlee, C. R., F. J. Duffy, B. H. Gern, J. L. Delahaye, S. B. Cohen, C. R. Stoltzfus, T. R. Rustad, S. G. Hansen, M. K. Axthelm, L. J. Picker, J. D. Aitchison, D. R. Sherman, V. V. Ganusov, M. Y. Gerner, D. E. Zak, and K. B. Urdahl (2021). “Ultra-low Dose Aerosol Infection of Mice with *Mycobacterium tuberculosis* More Closely Models Human Tuberculosis”. In: *Cell Host Microbe* 29.1, 68–82 e5.

Poeck, H. and J. Ruland (2010). “ITAM receptor signaling and the NLRP3 inflammasome in antifungal immunity”. In: *J Clin Immunol* 30.4, pp. 496–501.

Polena, H., F. Boudou, S. Tilleul, N. Dubois-Colas, C. Lecointe, N. Rakotosamimanana, M. Pelizzola, S. F. Andriamandimby, V. Raharimanga, P. Charles, J. L. Herrmann, P. Ricciardi-Castagnoli, V. Rasolofolo, B. Gicquel, and L. Tailleux (2016). “*Mycobacterium tuberculosis* exploits the formation of new blood vessels for its dissemination”. In: *Sci Rep* 6, p. 33162.

– The mechanisms utilized by mycobacteria to disseminate to distal tissues have remained somewhat mysterious for a period of time, but dissemination along blood vessels is a logical analogy to phenotypes seen in cancer. This study found that infected macrophages produced VEGFA in a region of difference 1 (RD1)-dependent manner. They found that inhibition of VEGFA resulted in restricted dissemination from the initial site of infection and reduced bacterial burden. Study of these phenomena in the mouse raise the possibility that these phenotypes are regulated by alterations in macrophage biology, an effect consistent with Harding, Herbath, Chen, Rayasam, Ritter, Csoka, Hasko, Michael, Fabry, Nagy, and Sandor (2019).

Poli, V., M. Di Gioia, M. Sola-Visner, F. Granucci, 3rd Frelinger A. L., A. D. Michelson, and I. Zanoni (2022). “Inhibition of transcription factor NFAT activity in activated platelets enhances their aggregation and exacerbates gram-negative bacterial septicemia”. In: *Immunity* 55.2, 224–236 e5.

– Roles for NFAT in cytosolic protein:protein interactions is a fascinating possibility and one that is difficult to address in the nucleated cells that make up the majority of somatic cells. This study interrogated the role of NFAT in platelets, which lack a nucleus, in regulating responses to Gram-negative sepsis. This involves a set of interactions between platelets and neutrophils and applied transgenic expression of the VIVIT peptide within platelets. Inhibition of NFAT in platelets exacerbated hypercoagulation and resulted in increased disease severity while also increasing coagulation in human platelets from individuals with hypocoagulation disorders. This is a very innovative publication on the role of NFAT in a new tissue type and provides a new transgenic mouse likely to be applied to diverse tissues in the future.

Pollard, J. W. (2009). “Trophic macrophages in development and disease”. In: *Nat Rev Immunol* 9.4, pp. 259–70.

Polverini, P. J., P. S. Cotran, Jr. Gimbrone M. A., and E. R. Unanue (1977). “Activated macrophages induce vascular proliferation”. In: *Nature* 269.5631, pp. 804–6.

– This is the first description of macrophages in the induction of angiogenesis and is the foundation of all future studies on this topic.

Pompa, J. L. de la, L. A. Timmerman, H. Takimoto, H. Yoshida, A. J. Elia, E. Samper, J. Potter, A. Wakeham, L. Marengere, B. L. Langille, G. R. Crabtree, and T. W. Mak (1998). “Role of the NF-ATc transcription factor in morphogenesis of cardiac valves and septum”. In: *Nature* 392.6672, pp. 182–6.

- Pont, S. and A. B. Blanc-Potard (2021). “Zebrafish Embryo Infection Model to Investigate *Pseudomonas aeruginosa* Interaction With Innate Immunity and Validate New Therapeutics”. In: *Front Cell Infect Microbiol* 11, p. 745851.
- Porcheray, F., B. Samah, C. Leone, N. Dereuddre-Bosquet, and G. Gras (2006). “Macrophage activation and human immunodeficiency virus infection: HIV replication directs macrophages towards a pro-inflammatory phenotype while previous activation modulates macrophage susceptibility to infection and viral production”. In: *Virology* 349.1, pp. 112–20.
- Porter, C. M. and N. A. Clipstone (2002). “Sustained NFAT signaling promotes a Th1-like pattern of gene expression in primary murine CD4+ T cells”. In: *J Immunol* 168.10, pp. 4936–45.
- Potente, M., H. Gerhardt, and P. Carmeliet (2011). “Basic and therapeutic aspects of angiogenesis”. In: *Cell* 146.6, pp. 873–87.
- Prados-Rosales, R., A. Baena, L. R. Martinez, J. Luque-Garcia, R. Kalscheuer, U. Veeraraghavan, C. Camara, J. D. Nosanchuk, G. S. Besra, B. Chen, J. Jimenez, A. Glatman-Freedman, Jr. Jacobs W. R., S. A. Porcelli, and A. Casadevall (2011). “Mycobacteria release active membrane vesicles that modulate immune responses in a TLR2-dependent manner in mice”. In: *J Clin Invest* 121.4, pp. 1471–83.
- Prajsnar, T. K., V. T. Cunliffe, S. J. Foster, and S. A. Renshaw (2008). “A novel vertebrate model of *Staphylococcus aureus* infection reveals phagocyte-dependent resistance of zebrafish to non-host specialized pathogens”. In: *Cell Microbiol* 10.11, pp. 2312–25.
- Prasad, K., M. B. Singh, and H. Ryan (2016). “Corticosteroids for managing tuberculous meningitis”. In: *Cochrane Database Syst Rev* 4, p. CD002244.
- Prasanthi, K and DS Murty (2014). “A Brief Review on Ecology and Evolution of Mycobacteria”. In: *Mycobacterial Diseases* 4.6.
- Price, J. V. and R. E. Vance (2014). “The macrophage paradox”. In: *Immunity* 41.5, pp. 685–93.
- Prill, K. and J. F. Dawson (2020). “Homology-Directed Repair in Zebrafish: Witchcraft and Wizardry?” In: *Front Mol Biosci* 7, p. 595474.

Prosser, G., J. Brandenburg, N. Reiling, 3rd Barry C. E., R. J. Wilkinson, and K. A. Wilkinson (2017). “The bacillary and macrophage response to hypoxia in tuberculosis and the consequences for T cell antigen recognition”. In: *Microbes Infect* 19.3, pp. 177–192.

Pu, W., C. Zhao, J. Wazir, Z. Su, M. Niu, S. Song, L. Wei, L. Li, X. Zhang, X. Shi, and H. Wang (2021). “Comparative transcriptomic analysis of THP-1-derived macrophages infected with *Mycobacterium tuberculosis* H37Rv, H37Ra and BCG”. In: *J Cell Mol Med* 25.22, pp. 10504–10520.

Puthenveetil, A. and S. Dubey (2020). “Metabolic reprogramming of tumor-associated macrophages”. In: *Ann Transl Med* 8.16, p. 1030.

Pym, A. S., A. H. Diacon, S. J. Tang, F. Conradie, M. Danilovits, C. Chuchottaworn, I. Vasilyeva, K. Andries, N. Bakare, T. De Marez, M. Haxaire-Theeuwes, N. Lounis, P. Meyvisch, B. Van Baelen, R. P. van Heeswijk, B. Dannemann, and Tmc C. Study Group (2016). “Bedaquiline in the treatment of multidrug- and extensively drug-resistant tuberculosis”. In: *Eur Respir J* 47.2, pp. 564–74.

Qi, C., J. Zhang, X. Chen, J. Wan, J. Wang, P. Zhang, and Y. Liu (2017). “Hypoxia stimulates neural stem cell proliferation by increasing HIF1alpha expression and activating Wnt/beta-catenin signaling”. In: *Cell Mol Biol (Noisy-le-grand)* 63.7, pp. 12–19.

Qian, C. N., D. Huang, B. Wondergem, and B. T. Teh (2009). “Complexity of tumor vasculature in clear cell renal cell carcinoma”. In: *Cancer* 115.10 Suppl, pp. 2282–9.

Qian, F., F. Zhen, C. Ong, S. W. Jin, H. Meng Soo, D. Y. Stainier, S. Lin, J. Peng, and Z. Wen (2005). “Microarray analysis of zebrafish cloche mutant using amplified cDNA and identification of potential downstream target genes”. In: *Dev Dyn* 233.3, pp. 1163–72.

Qu, Z., J. Zhou, Y. Zhou, Y. Xie, Y. Jiang, J. Wu, Z. Luo, G. Liu, L. Yin, and X. L. Zhang (2020). “Mycobacterial EST12 activates a RACK1-NLRP3-gasdermin D pyroptosis-IL-1beta immune pathway”. In: *Sci Adv* 6.43.

Quadrato, G., M. Benevento, S. Alber, C. Jacob, E. M. Floriddia, T. Nguyen, M. Y. Elnaggar, C. M. Pedroarena, J. D. Molkentin, and S. Di Giovanni (2012). “Nuclear factor of activated T cells (NFATc4) is required for BDNF-dependent survival of adult-born neurons and spatial memory formation in the hippocampus”. In: *Proc Natl Acad Sci U S A* 109.23, E1499–508.

- Qualls, J. E. and P. J. Murray (2016). "Immunometabolism within the tuberculosis granuloma: amino acids, hypoxia, and cellular respiration". In: *Semin Immunopathol* 38.2, pp. 139–52.
- Quenard, F., P. E. Fournier, M. Drancourt, and P. Brouqui (2017). "Role of second-line injectable antituberculosis drugs in the treatment of MDR/XDR tuberculosis". In: *Int J Antimicrob Agents* 50.2, pp. 252–254.
- Quigley, J., V. K. Hughitt, C. A. Velikovsky, R. A. Mariuzza, N. M. El-Sayed, and V. Briken (2017). "The Cell Wall Lipid PDIM Contributes to Phagosomal Escape and Host Cell Exit of *Mycobacterium tuberculosis*". In: *mBio* 8.2.
- R Core Team (2022). "R: A Language and Environment for Statistical Computing". In.
- Rabold, K., M. G. Netea, G. J. Adema, and R. T. Netea-Maier (2017). "Cellular metabolism of tumor-associated macrophages - functional impact and consequences". In: *FEBS Lett* 591.19, pp. 3022–3041.
- Raetz, C. R., C. M. Reynolds, M. S. Trent, and R. E. Bishop (2007). "Lipid A modification systems in gram-negative bacteria". In: *Annu Rev Biochem* 76, pp. 295–329.
- Rahbari, N. N., D. Kedrin, J. Incio, H. Liu, W. W. Ho, H. T. Nia, C. M. Edrich, K. Jung, J. Daubriac, I. Chen, T. Heishi, J. D. Martin, Y. Huang, N. Maimon, C. Reissfelder, J. Weitz, Y. Boucher, J. W. Clark, A. J. Grodzinsky, D. G. Duda, R. K. Jain, and D. Fukumura (2016). "Anti-VEGF therapy induces ECM remodeling and mechanical barriers to therapy in colorectal cancer liver metastases". In: *Sci Transl Med* 8.360, 360ra135.
- Rahi, A., S. K. Matta, A. Dhiman, J. Garhyan, M. Gopalani, S. Chandra, and R. Bhatnagar (2017). "Enolase of *Mycobacterium tuberculosis* is a surface exposed plasminogen binding protein". In: *Biochim Biophys Acta Gen Subj* 1861.1 Pt A, pp. 3355–3364.
- Rahman, M. A., B. M. Cumming, K. W. Addicott, H. T. Pacl, S. L. Russell, K. Nargan, T. Naidoo, P. K. Ramdial, J. H. Adamson, R. Wang, and A. J. C. Steyn (2020). "Hydrogen sulfide dysregulates the immune response by suppressing central carbon metabolism to promote tuberculosis". In: *Proc Natl Acad Sci U S A* 117.12, pp. 6663–6674.

Rajaram, M. V. S., E. Arnett, A. K. Azad, E. Guirado, B. Ni, A. D. Gerberick, L. Z. He, T. Keler, L. J. Thomas, W. P. Lafuse, and L. S. Schlesinger (2017). “*M. tuberculosis*-Initiated Human Mannose Receptor Signaling Regulates Macrophage Recognition and Vesicle Trafficking by FcRgamma-Chain, Grb2, and SHP-1”. In: *Cell Rep* 21.1, pp. 126–140.

Ramakrishnan, L. (1997). “Images in clinical medicine. *Mycobacterium marinum* infection of the hand”. In: *N Engl J Med* 337.9, p. 612.

- (2012). “Revisiting the role of the granuloma in tuberculosis”. In: *Nat Rev Immunol* 12.5, pp. 352–66.
- (2013a). “Looking within the zebrafish to understand the tuberculous granuloma”. In: *Adv Exp Med Biol* 783, pp. 251–66.
- (2013b). “The zebrafish guide to tuberculosis immunity and treatment”. In: *Cold Spring Harb Symp Quant Biol* 78, pp. 179–92.

Ramakrishnan, L. and S. Falkow (1994). “*Mycobacterium marinum* persists in cultured mammalian cells in a temperature-restricted fashion”. In: *Infect Immun* 62.8, pp. 3222–9.

Ramakrishnan, L., N. A. Federspiel, and S. Falkow (2000). “Granuloma-specific expression of *Mycobacterium* virulence proteins from the glycine-rich PE-PGRS family”. In: *Science* 288.5470, pp. 1436–9.

Ramakrishnan, L., R. H. Valdivia, J. H. McKerrow, and S. Falkow (1997). “*Mycobacterium marinum* causes both long-term subclinical infection and acute disease in the leopard frog (*Rana pipiens*)”. In: *Infect Immun* 65.2, pp. 767–73.

Rambaruth, N. D., S. A. Jegouzo, H. Marlor, M. E. Taylor, and K. Drickamer (2015). “Mouse mincle: characterization as a model for human mincle and evolutionary implications”. In: *Molecules* 20.4, pp. 6670–82.

Ramiro-Diaz, J. M., W. Giermakowska, J. M. Weaver, N. L. Jernigan, and L. V. Gonzalez Bosc (2014). “Mechanisms of NFATc3 activation by increased superoxide and reduced hydrogen peroxide in pulmonary arterial smooth muscle”. In: *Am J Physiol Cell Physiol* 307.10, pp. C928–38.

– This work demonstrates important roles for hypoxia in regulating NFAT responses and provides an intellectual foundation for the study of hypoxia as a physical phenomenon and HIF-1 α as a genetic response with NFAT-dependent responses.

Ramsey, K. M., W. Chen, J. D. Marion, S. Bergqvist, and E. A. Komives (2019). “Exclusivity and Compensation in NF κ B Dimer Distributions and I κ B Inhibition”. In: *Biochemistry* 58.21, pp. 2555–2563.

Randhawa, P. S., T. E. Starzl, and A. J. Demetris (1997). “Tacrolimus (FK506)-Associated Renal Pathology”. In: *Adv Anat Pathol* 4.4, pp. 265–276.

Randow, F., J. D. MacMicking, and L. C. James (2013). “Cellular self-defense: how cell-autonomous immunity protects against pathogens”. In: *Science* 340.6133, pp. 701–6.

Ranea, J. A., A. Grant, J. M. Thornton, and C. A. Orengo (2005). “Microeconomic principles explain an optimal genome size in bacteria”. In: *Trends Genet* 21.1, pp. 21–5.

Ranger, A. M., L. C. Gerstenfeld, J. Wang, T. Kon, H. Bae, E. M. Gravallese, M. J. Glimcher, and L. H. Glimcher (2000). “The nuclear factor of activated T cells (NFAT) transcription factor NFATp (NFATc2) is a repressor of chondrogenesis”. In: *J Exp Med* 191.1, pp. 9–22.

Ranger, A. M., M. J. Grusby, M. R. Hodge, E. M. Gravallese, F. C. de la Brousse, T. Hoey, C. Mickanin, H. S. Baldwin, and L. H. Glimcher (1998a). “The transcription factor NF-ATc is essential for cardiac valve formation”. In: *Nature* 392.6672, pp. 186–90.

Ranger, A. M., M. Oukka, J. Rengarajan, and L. H. Glimcher (1998b). “Inhibitory function of two NFAT family members in lymphoid homeostasis and Th2 development”. In: *Immunity* 9.5, pp. 627–35.

Ranieri, G., M. Mammi, E. Donato Di Paola, E. Russo, L. Gallelli, R. Citraro, C. D. Gadaleta, I. Marech, M. Ammendola, and G. De Sarro (2014). “Pazopanib a tyrosine kinase inhibitor with strong anti-angiogenetic activity: a new treatment for metastatic soft tissue sarcoma”. In: *Crit Rev Oncol Hematol* 89.2, pp. 322–9.

Ranjan, R., J. Deng, S. Chung, Y. G. Lee, G. Y. Park, L. Xiao, M. Joo, J. W. Christman, and M. Karpurapu (2014). “The transcription factor nuclear factor of

activated T cells c3 modulates the function of macrophages in sepsis". In: *J Innate Immun* 6.6, pp. 754–64.

Ransom, D. G., P. Haffter, J. Odenthal, A. Brownlie, E. Vogelsang, R. N. Kelsh, M. Brand, F. J. van Eeden, M. Furutani-Seiki, M. Granato, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, M. C. Mullins, and C. Nusslein-Volhard (1996). "Characterization of zebrafish mutants with defects in embryonic hematopoiesis". In: *Development* 123, pp. 311–9.

Rao, A., C. Luo, and P. G. Hogan (1997). "Transcription factors of the NFAT family: regulation and function". In: *Annu Rev Immunol* 15, pp. 707–47.

Rao, V., N. Fujiwara, S. A. Porcelli, and M. S. Glickman (2005). "*Mycobacterium tuberculosis* controls host innate immune activation through cyclopropane modification of a glycolipid effector molecule". In: *J Exp Med* 201.4, pp. 535–43.

– *This study found that the loss of pcaA reduced the inflammatory response during infection, arguing that the addition of cis-cyclopropanation to TDM is a pro-inflammatory modification. This alludes to the fact that inflammatory responses are bacteria-beneficial components of the immune response and is a foundation for the study of TDM modification in the angiogenic responses as was done in Walton, Cronan, Cambier, Rossi, Marass, Foglia, Brewer, Poss, Stainier, Bertozzi, and Tobin (2018).*

Rao, V., F. Gao, B. Chen, Jr. Jacobs W. R., and M. S. Glickman (2006). "Trans-cyclopropanation of mycolic acids on trehalose dimycolate suppresses *Mycobacterium tuberculosis* -induced inflammation and virulence". In: *J Clin Invest* 116.6, pp. 1660–7.

– *Paralleling the effects of cis-cyclopropanation on TDM, trans-cyclopropanation after addition by cmaA2 is a hypoinflammatory modification and unleashes a potent inflammatory response in its absence. This develops a model for TDM modifications balancing pro- and anti-inflammatory effects, perhaps dynamically over the course of infection as bacterial needs change.*

Rathinam, V. A. K., Y. Zhao, and F. Shao (2019). "Innate immunity to intracellular LPS". In: *Nat Immunol* 20.5, pp. 527–533.

Rawls, J. F., M. A. Mahowald, R. E. Ley, and J. I. Gordon (2006). "Reciprocal gut microbiota transplants from zebrafish and mice to germ-free recipients reveal host habitat selection". In: *Cell* 127.2, pp. 423–33.

- Rawls, J. F., B. S. Samuel, and J. I. Gordon (2004). “Gnotobiotic zebrafish reveal evolutionarily conserved responses to the gut microbiota”. In: *Proc Natl Acad Sci U S A* 101.13, pp. 4596–601.
- Ray, S., D. Tillo, S. R. Durell, S. Khund-Sayeed, and C. Vinson (2021). “REL Domain of NFATc2 Binding to Five Types of DNA Using Protein Binding Microarrays”. In: *ACS Omega* 6.6, pp. 4147–4154.
- Reade, A., L. B. Motta-Mena, K. H. Gardner, D. Y. Stainier, O. D. Weiner, and S. Woo (2017). “TAEL: a zebrafish-optimized optogenetic gene expression system with fine spatial and temporal control”. In: *Development* 144.2, pp. 345–355.
- Redelman-Sidi, G. (2020). “Could BCG be used to protect against COVID-19?” In: *Nat Rev Urol* 17.6, pp. 316–317.
- Redelman-Sidi, G., M. S. Glickman, and B. H. Bochner (2014). “The mechanism of action of BCG therapy for bladder cancer—a current perspective”. In: *Nat Rev Urol* 11.3, pp. 153–62.
- Reed, G. H., J. O. Kent, and C. T. Wittwer (2007). “High-resolution DNA melting analysis for simple and efficient molecular diagnostics”. In: *Pharmacogenomics* 8.6, pp. 597–608.
- Rehwinkel, J. and M. U. Gack (2020). “RIG-I-like receptors: their regulation and roles in RNA sensing”. In: *Nat Rev Immunol* 20.9, pp. 537–551.
- Reid, D. M., N. A. Gow, and G. D. Brown (2009). “Pattern recognition: recent insights from Dectin-1”. In: *Curr Opin Immunol* 21.1, pp. 30–7.
- Reischauer, S., O. A. Stone, A. Villasenor, N. Chi, S. W. Jin, M. Martin, M. T. Lee, N. Fukuda, M. Marass, A. Witty, I. Fiddes, T. Kuo, W. S. Chung, S. Salek, R. Lerrigo, J. Alsio, S. Luo, D. Tworus, S. M. Augustine, S. Muceniek, B. Nystedt, A. J. Giraldez, G. P. Schroth, O. Andersson, and D. Y. Stainier (2016). “Cloche is a bHLH-PAS transcription factor that drives haemato-vascular specification”. In: *Nature* 535.7611, pp. 294–8.
- Rengarajan, J., B. Tang, and L. H. Glimcher (2002). “NFATc2 and NFATc3 regulate T(H)2 differentiation and modulate TCR-responsiveness of naive T(H)cells”. In: *Nat Immunol* 3.1, pp. 48–54.

- Rens, C., J. D. Chao, D. L. Sexton, E. I. Tocheva, and Y. Av-Gay (2021). “Roles for phthiocerol dimycocerosate lipids in *Mycobacterium tuberculosis* pathogenesis”. In: *Microbiology (Reading)* 167.3.
- Renshaw, S. A. and N. S. Trede (2012). “A model 450 million years in the making: zebrafish and vertebrate immunity”. In: *Dis Model Mech* 5.1, pp. 38–47.
- Retzinger, G. S. (1987). “Dissemination of beads coated with trehalose 6,6'-dimycolate: a possible role for coagulation in the dissemination process”. In: *Exp Mol Pathol* 46.2, pp. 190–8.
- Retzinger, G. S., S. C. Meredith, R. L. Hunter, K. Takayama, and F. J. Kezdy (1982). “Identification of the physiologically active state of the mycobacterial glycolipid trehalose 6,6'-dimycolate and the role of fibrinogen in the biologic activities of trehalose 6,6'-dimycolate monolayers”. In: *J Immunol* 129.2, pp. 735–44.
- Reva, O., I. Korotetskiy, and A. Ilin (2015). “Role of the horizontal gene exchange in evolution of pathogenic Mycobacteria”. In: *BMC Evol Biol* 15 Suppl 1, S2.
- Reynaud, E. G., U. Krzic, K. Greger, and E. H. Stelzer (2008). “Light sheet-based fluorescence microscopy: more dimensions, more photons, and less photodamage”. In: *HFSP J* 2.5, pp. 266–75.
- Rhoades, E., F. Hsu, J. B. Torrelles, J. Turk, D. Chatterjee, and D. G. Russell (2003). “Identification and macrophage-activating activity of glycolipids released from intracellular *Mycobacterium bovis* BCG”. In: *Mol Microbiol* 48.4, pp. 875–88.
- Riabov, V., A. Gudima, N. Wang, A. Mickley, A. Orekhov, and J. Kzhyshkowska (2014). “Role of tumor associated macrophages in tumor angiogenesis and lymphangiogenesis”. In: *Front Physiol* 5, p. 75.
- Ribatti, D., B. Nico, E. Crivellato, and A. Vacca (2007). “Macrophages and tumor angiogenesis”. In: *Leukemia* 21.10, pp. 2085–9.
- Ribeiro, A. L. and O. K. Okamoto (2015). “Combined effects of pericytes in the tumor microenvironment”. In: *Stem Cells Int* 2015, p. 868475.
- Rice, N. R., M. L. MacKichan, and A. Israel (1992). “The precursor of NF-kappa B p50 has I kappa B-like functions”. In: *Cell* 71.2, pp. 243–53.

Richardson, M. B. and S. J. Williams (2014). “MCL and Mincle: C-Type Lectin Receptors That Sense Damaged Self and Pathogen-Associated Molecular Patterns”. In: *Front Immunol* 5, p. 288.

Rihel, J., D. A. Prober, A. Arvanites, K. Lam, S. Zimmerman, S. Jang, S. J. Haggerty, D. Kokel, L. L. Rubin, R. T. Peterson, and A. F. Schier (2010). “Zebrafish behavioral profiling links drugs to biological targets and rest/wake regulation”. In: *Science* 327.5963, pp. 348–51.

Rinne, A., K. Banach, and L. A. Blatter (2009). “Regulation of nuclear factor of activated T cells (NFAT) in vascular endothelial cells”. In: *J Mol Cell Cardiol* 47.3, pp. 400–10.

– *An early pioneering study on the different induction dynamics of NFAT isoforms, this time within vascular endothelial cells. This work found that NFATC1 was induced in a persistent manner by exposure to ATP, NFATC3 was rapidly shuttled between the nucleus and the cytosol. This feature of NFATC3 appears to be broadly conserved between cell types as this precise behavior has been described in endothelial cells, neurons, and mast cells.*

Ristow, L. C. and J. M. Davis (2021). “The granuloma in cryptococcal disease”. In: *PLoS Pathog* 17.3, e1009342.

– *An excellent review of the cryptococcal granuloma and how it compares to other granulomas from Muse Davis, who has been studying granulomas for two solid decades, first in tuberculosis and now in Cryptococcus.*

Rivera, L. B. and G. Bergers (2015). “CANCER. Tumor angiogenesis, from foe to friend”. In: *Science* 349.6249, pp. 694–5.

Robbs, B. K., A. L. Cruz, M. B. Werneck, G. P. Mognol, and J. P. Viola (2008). “Dual roles for NFAT transcription factor genes as oncogenes and tumor suppressors”. In: *Mol Cell Biol* 28.23, pp. 7168–81.

Robinson, C. M., D. O’Dee, T. Hamilton, and G. J. Nau (2010). “Cytokines involved in interferon-gamma production by human macrophages”. In: *J Innate Immun* 2.1, pp. 56–65.

Robinson, Henry Peach (1858). *Fading Away*.

Robinson, M. J., D. Sancho, E. C. Slack, S. LeibundGut-Landmann, and C. Reis e Sousa (2006). “Myeloid C-type lectins in innate immunity”. In: *Nat Immunol* 7.12, pp. 1258–65.

Robinson, R. T. and A. R. Huppler (2017). “The Goldilocks model of immune symbiosis with Mycobacteria and *Candida* colonizers”. In: *Cytokine* 97, pp. 49–65.

Roca, F. J., L. J. Whitworth, H. A. Prag, M. P. Murphy, and L. Ramakrishnan (2022). “Tumor necrosis factor induces pathogenic mitochondrial ROS in tuberculosis through reverse electron transport”. In: *Science* 376.6600, eabh2841.

Roca, F. J., L. J. Whitworth, S. Redmond, A. A. Jones, and L. Ramakrishnan (2019). “TNF Induces Pathogenic Programmed Macrophage Necrosis in Tuberculosis through a Mitochondrial-Lysosomal-Endoplasmic Reticulum Circuit”. In: *Cell* 178.6, 1344–1361 e11.

– *This role for mitochondrial reactive oxygen species in mediating TNF- α -induced necrosis is an interesting intracellular signaling pathways that integrates diverse aspects of cell biology in coordinating this response to mycobacterial infection and prevention of this mitochondrially-induced necrosis protects against infection.*

Rock, F. L., G. Hardiman, J. C. Timans, R. A. Kastlein, and J. F. Bazan (1998). “A family of human receptors structurally related to *Drosophila* Toll”. In: *Proc Natl Acad Sci U S A* 95.2, pp. 588–93.

Rodrigues, L. C., V. K. Diwan, and J. G. Wheeler (1993). “Protective effect of BCG against tuberculous meningitis and miliary tuberculosis: a meta-analysis”. In: *Int J Epidemiol* 22.6, pp. 1154–8.

Roehrl, M. H., S. Kang, J. Aramburu, G. Wagner, A. Rao, and P. G. Hogan (2004). “Selective inhibition of calcineurin-NFAT signaling by blocking protein-protein interaction with small organic molecules”. In: *Proc Natl Acad Sci U S A* 101.20, pp. 7554–9.

Roehrl, M. H., J. Y. Wang, and G. Wagner (2004). “Discovery of small-molecule inhibitors of the NFAT–calcineurin interaction by competitive high-throughput fluorescence polarization screening”. In: *Biochemistry* 43.51, pp. 16067–75.

Rogers, N. C., E. C. Slack, A. D. Edwards, M. A. Nolte, O. Schulz, E. Schweighoffer, D. L. Williams, S. Gordon, V. L. Tybulewicz, G. D. Brown, and C. Reis e Sousa (2005). “Syk-dependent cytokine induc-

tion by Dectin-1 reveals a novel pattern recognition pathway for C type lectins”. In: *Immunity* 22.4, pp. 507–17.

Rojanathammanee, L., A. M. Floden, G. D. Manocha, and C. K. Combs (2015). “Attenuation of microglial activation in a mouse model of Alzheimer’s disease via NFAT inhibition”. In: *J Neuroinflammation* 12, p. 42.

Romanchikova, N., V. Ivanova, C. Scheller, E. Jankevics, C. Jassoy, and E. Serfling (2003). “NFAT transcription factors control HIV-1 expression through a binding site downstream of TAR region”. In: *Immunobiology* 208.4, pp. 361–5.

Rombough, P. and H. Drader (2009). “Hemoglobin enhances oxygen uptake in larval zebrafish (*Danio rerio*) but only under conditions of extreme hypoxia”. In: *J Exp Biol* 212.Pt 6, pp. 778–84.

Romero, M. M., J. I. Basile, B. Lopez, V. Ritacco, L. Barrera, C. Sasiain Mdel, and M. Aleman (2014). “Outbreaks of *Mycobacterium tuberculosis* MDR strains differentially induce neutrophil respiratory burst involving lipid rafts, p38 MAPK and Syk”. In: *BMC Infect Dis* 14, p. 262.

Rong, L. and A. S. Perelson (2010). “Treatment of hepatitis C virus infection with interferon and small molecule direct antivirals: viral kinetics and modeling”. In: *Crit Rev Immunol* 30.2, pp. 131–48.

Rosenbloom, Raymond, Igor Gavrilash, Kerstin Seidel, Igor Kramnik, and Nicholas A Crossland (2021). “Dissemination and progression of pulmonary *Mycobacterium avium* infection in mouse model is associated with type 2 macrophage activation”. In: *bioRxiv*.

Rosowski, E. E., B. P. Knox, L. S. Archambault, A. Huttenlocher, N. P. Keller, R. T. Wheeler, and J. M. Davis (2018). “The Zebrafish as a Model Host for Invasive Fungal Infections”. In: *J Fungi (Basel)* 4.4.

Ross, J. M., Y. Xie, Y. Wang, J. K. Collins, C. Horst, J. B. Doody, P. Lindstedt, J. R. Ledesma, A. E. Shapiro, P. S. I. Hay, H. H. Kyu, and A. D. Flaxman (2021). “Estimating the population at high risk for tuberculosis through household exposure in high-incidence countries: a model-based analysis”. In: *EClinicalMedicine* 42, p. 101206.

Rossi, A., S. Gauvrit, M. Marass, L. Pan, C. B. Moens, and D. Y. R. Stainier (2016). “Regulation of Vegf signaling by natural and synthetic ligands”. In: *Blood* 128.19, pp. 2359–2366.

Rossi, A., Z. Kontarakis, C. Gerri, H. Nolte, S. Holper, M. Kruger, and D. Y. Stainier (2015). “Genetic compensation induced by deleterious mutations but not gene knockdowns”. In: *Nature* 524.7564, pp. 230–3.

– This paper gave birth to the concept of genetic compensation, which allows organisms to genetically adapt to loss of function mutations in otherwise critical host genes. While it was difficult to divine the precise contributors, even this initial publication suggested that nonsense-mediated mRNA decay was critical for these compensatory responses, findings that have borne out over time and have given rise to new recommendations for the most effective study of loss of function in various genes, where simple indels from CRISPR may be difficult to interpret due to these compensatory mechanisms and approaches removing the entire locus or at least the majority of the promoter and transcriptional start sites are needed.

Rothfuchs, A. G., A. Bafica, C. G. Feng, J. G. Egen, D. L. Williams, G. D. Brown, and A. Sher (2007). “Dectin-1 interaction with *Mycobacterium tuberculosis* leads to enhanced IL-12p40 production by splenic dendritic cells”. In: *J Immunol* 179.6, pp. 3463–71.

Rotty, J. D., H. E. Brighton, S. L. Craig, S. B. Asokan, N. Cheng, J. P. Ting, and J. E. Bear (2017). “Arp2/3 Complex Is Required for Macrophage Integrin Functions but Is Dispensable for FcR Phagocytosis and *in vivo* Motility”. In: *Dev Cell* 42.5, 498–513 e6.

Roux, K. J., D. I. Kim, M. Raida, and B. Burke (2012). “A promiscuous biotin ligase fusion protein identifies proximal and interacting proteins in mammalian cells”. In: *J Cell Biol* 196.6, pp. 801–10.

Rozwarski, D. A., G. A. Grant, D. H. Barton, Jr. Jacobs W. R., and J. C. Sacchettini (1998). “Modification of the NADH of the isoniazid target (InhA) from *Mycobacterium tuberculosis*”. In: *Science* 279.5347, pp. 98–102.

RStudio Team (2022). “RStudio: Integrated Development Environment for R”. In.

Rubin, E. J. (2009). “The granuloma in tuberculosis—friend or foe?” In: *N Engl J Med* 360.23, pp. 2471–3.

Rudd, P. M., T. Elliott, P. Cresswell, I. A. Wilson, and R. A. Dwek (2001). “Glycosylation and the immune system”. In: *Science* 291.5512, pp. 2370–6.

Rueden, C. T., M. C. Hiner, 3rd Evans E. L., M. A. Pinkert, A. M. Lucas, A. E. Carpenter, B. A. Cimini, and K. W. Eliceiri (2022). “PyImageJ: A library for integrating ImageJ and Python”. In: *Nat Methods*.

Rueden, C. T., J. Schindelin, M. C. Hiner, B. E. DeZonia, A. E. Walter, E. T. Arena, and K. W. Eliceiri (2017). “ImageJ2: ImageJ for the next generation of scientific image data”. In: *BMC Bioinformatics* 18.1, p. 529.

Ruhl, C. R., B. L. Pasko, H. S. Khan, L. M. Kindt, C. E. Stamm, L. H. Franco, C. C. Hsia, M. Zhou, C. R. Davis, T. Qin, L. Gautron, M. D. Burton, G. L. Mejia, D. K. Naik, G. Dussor, T. J. Price, and M. U. Shiloh (2020). “*Mycobacterium tuberculosis* Sulfolipid-1 Activates Nociceptive Neurons and Induces Cough”. In: *Cell* 181.2, 293–305 e11.

Ruiz, P., F. Rodriguez-Cano, F. J. Zerolo, and M. Casal (2003). “Streptomycin as second-line chemotherapy for tuberculosis”. In: *Rev Esp Quimioter* 16.2, pp. 188–94.

Runft, D. L., K. C. Mitchell, B. H. Abuaita, J. P. Allen, S. Bajer, K. Ginsburg, M. N. Neely, and J. H. Withey (2014). “Zebrafish as a natural host model for *Vibrio cholerae* colonization and transmission”. In: *Appl Environ Microbiol* 80.5, pp. 1710–7.

Russell, D. G. (2007). “Who puts the tubercle in tuberculosis?” In: *Nat Rev Microbiol* 5.1, pp. 39–47.

Russell, D. G., 3rd Barry C. E., and J. L. Flynn (2010). “Tuberculosis: what we don’t know can, and does, hurt us”. In: *Science* 328.5980, pp. 852–6.

Russell, D. G., P. J. Cardona, M. J. Kim, S. Allain, and F. Altare (2009). “Foamy macrophages and the progression of the human tuberculosis granuloma”. In: *Nat Immunol* 10.9, pp. 943–8.

Russell, D. G., L. Huang, and B. C. VanderVen (2019). “Immunometabolism at the interface between macrophages and pathogens”. In: *Nat Rev Immunol* 19.5, pp. 291–304.

- Russell, S. L., D. A. Lamprecht, T. Mandizvo, T. T. Jones, V. Naidoo, K. W. Addicott, C. Moodley, B. Ngcobo, D. K. Crossman, G. Wells, and A. J. C. Steyn (2019). “Compromised Metabolic Reprogramming Is an Early Indicator of CD8(+) T Cell Dysfunction during Chronic *Mycobacterium tuberculosis* Infection”. In: *Cell Rep* 29.11, 3564–3579 e5.
- Rustad, T. R., A. M. Sherrid, K. J. Minch, and D. R. Sherman (2009). “Hypoxia: a window into *Mycobacterium tuberculosis* latency”. In: *Cell Microbiol* 11.8, pp. 1151–9.
- Ruyra, A., M. Cano-Sarabia, P. Garcia-Valtanen, D. Yero, I. Gibert, S. A. Mackenzie, A. Estepa, D. Maspoch, and N. Roher (2014). “Targeting and stimulation of the zebrafish (*Danio rerio*) innate immune system with LPS/dsRNA-loaded nanoliposomes”. In: *Vaccine* 32.31, pp. 3955–62.
- Ryll, R., Y. Kumazawa, and I. Yano (2001). “Immunological properties of trehalose dimycolate (cord factor) and other mycolic acid-containing glycolipids—a review”. In: *Microbiol Immunol* 45.12, pp. 801–11.
- Sabath, L. D., N. Wheeler, M. Laverdiere, D. Blazevic, and B. J. Wilkinson (1977). “A new type of penicillin resistance of *Staphylococcus aureus*”. In: *Lancet* 1.8009, pp. 443–7.
- Sabbaghi, F., L. Ullner, T. Bohn, J. Hahlbrock, T. Bopp, E. Schmitt, M. Klein, and M. Stassen (2021). “In Activated Murine Mast Cells, NFATc2 Is Critical for the Production of Autocrine IL-3, Thereby Promoting the Expression of IL-9”. In: *J Immunol* 206.1, pp. 67–76.
- Saber, T., D. J. Veale, E. Balogh, J. McCormick, S. NicAnUltaigh, M. Connolly, and U. Fearon (2011). “Toll-like receptor 2 induced angiogenesis and invasion is mediated through the Tie2 signalling pathway in rheumatoid arthritis”. In: *PLoS One* 6.8, e23540.
- Sabin, F (1920). “Studies on the origin of blood-vessels and of red blood corpuscles as seen in the living blastoderm of chicks during the second day of incubation. Carnegie inst. of Washington publ”. In: *Contributions to Embryology Bd* 9, p. 213.
- Saelens, J. W., M. I. Sweeney, G. Viswanathan, A. M. Xet-Mull, K. L. Jurcic Smith, D. M. Sisk, D. D. Hu, R. M. Cronin, E. J. Hughes, W. J. Brewer, J. Coers, M. M. Champion, P. A. Champion, C. B. Lowe, C. M. Smith, S. Lee, J. E. Stout, and

D. M. Tobin (2022). “An ancestral mycobacterial effector promotes dissemination of infection”. In: *Cell* 185.24, P4507–4525.E18.

Saelens, J. W., G. Viswanathan, and D. M. Tobin (2019). “Mycobacterial Evolution Intersects With Host Tolerance”. In: *Front Immunol* 10, p. 528.

Safi, H., S. Lingaraju, A. Amin, S. Kim, M. Jones, M. Holmes, M. McNeil, S. N. Peterson, D. Chatterjee, R. Fleischmann, and D. Alland (2013). “Evolution of high-level ethambutol-resistant tuberculosis through interacting mutations in decaprenylphosphoryl-beta-D-arabinose biosynthetic and utilization pathway genes”. In: *Nat Genet* 45.10, pp. 1190–7.

Saharinen, P., L. Eklund, and K. Alitalo (2017). “Therapeutic targeting of the angiopoietin-TIE pathway”. In: *Nat Rev Drug Discov* 16.9, pp. 635–661.

Saita, N., N. Fujiwara, I. Yano, K. Soejima, and K. Kobayashi (2000). “Trehalose 6,6'-dimycolate (cord factor) of *Mycobacterium tuberculosis* induces corneal angiogenesis in rats”. In: *Infect Immun* 68.10, pp. 5991–7.

– One of the two seminal papers in 2000 that established a pro-angiogenic role for TDM. This study adopted a rat corneal model of angiogenesis and inoculated the cornea with TDM to assess the angiogenic response and found that macrophage production of VEGFA and IL-8 (CXCL8) were essential for angiogenesis. This set an excellent foundation for later study by Walton, Cronan, Cambier, Rossi, Marass, Foglia, Brewer, Poss, Stainier, Bertozzi, and Tobin (2018) on the specific aspects of TDM able to modulate host angiogenesis.

Sakabe, M., J. Fan, Y. Odaka, N. Liu, A. Hassan, X. Duan, P. Stump, L. Byerly, M. Donaldson, J. Hao, M. Fruttiger, Q. R. Lu, Y. Zheng, R. A. Lang, and M. Xin (2017). “YAP/TAZ-CDC42 signaling regulates vascular tip cell migration”. In: *Proc Natl Acad Sci U S A* 114.41, pp. 10918–10923.

Sakaguchi, I., N. Ikeda, M. Nakayama, Y. Kato, I. Yano, and K. Kaneda (2000). “Trehalose 6,6'-dimycolate (Cord factor) enhances neovascularization through vascular endothelial growth factor production by neutrophils and macrophages”. In: *Infect Immun* 68.4, pp. 2043–52.

– One of the two seminal papers in 2000 that established a pro-angiogenic role for TDM. This study utilized air pouches in a mouse model inoculated with Rhodococcus or Mycobacterium TDM and found that VEGFA⁺ neutrophils were recruited to the site and produced a variety of other cytokines. Blockade of VEGFA led to

inhibition of TDM-induced granuloma formation, an early indication of the importance of vascularization in the process of forming the granuloma itself. They also applied a cell culture model of macrophage and neutrophil exposure to TDM and found similar results regarding VEGFA release, which may be a useful model for future study of this pathway.

Sakamoto, K., R. E. Geisel, M. J. Kim, B. T. Wyatt, L. B. Sellers, S. T. Smiley, A. M. Cooper, D. G. Russell, and E. R. Rhoades (2010). “Fibrinogen regulates the cytotoxicity of mycobacterial trehalose dimycolate but is not required for cell recruitment, cytokine response, or control of mycobacterial infection”. In: *Infect Immun* 78.3, pp. 1004–11.

Sakamoto, K., M. J. Kim, E. R. Rhoades, R. E. Allavena, S. Ehrt, H. C. Wainwright, D. G. Russell, and K. H. Rohde (2013). “Mycobacterial trehalose dimycolate reprograms macrophage global gene expression and activates matrix metalloproteinases”. In: *Infect Immun* 81.3, pp. 764–76.

Sakuma, K., J. Nishikawa, R. Nakao, K. Watanabe, T. Totsuka, H. Nakano, M. Sano, and M. Yasuhara (2003). “Calcineurin is a potent regulator for skeletal muscle regeneration by association with NFATc1 and GATA-2”. In: *Acta Neuropathol* 105.3, pp. 271–80.

Salah, I. B., E. Ghigo, and M. Drancourt (2009). “Free-living amoebae, a training field for macrophage resistance of mycobacteria”. In: *Clin Microbiol Infect* 15.10, pp. 894–905.

Saleem, Amer and Mohammed Azher (2013). “The next Pandemic-Tuberculosis: The oldest disease of mankind rising one more time”. In: *British Journal of Medical Practitioners BJMP* 66, p. 21.

Salter, Jim (2016). *blindanalysis: v1.0*.

Salton, M. R. (1953). “Studies of the bacterial cell wall. IV. The composition of the cell walls of some Gram-positive and Gram-negative bacteria”. In: *Biochim Biophys Acta* 10.4, pp. 512–23.

Sampath, P., K. Moideen, U. D. Ranganathan, and R. Bethunaickan (2018). “Monocyte Subsets: Phenotypes and Function in Tuberculosis Infection”. In: *Front Immunol* 9, p. 1726.

- Sancho, D. and C. Reis e Sousa (2012). "Signaling by myeloid C-type lectin receptors in immunity and homeostasis". In: *Annu Rev Immunol* 30, pp. 491–529.
- Sanderson, L. E., A. T. Chien, J. W. Astin, K. E. Crosier, P. S. Crosier, and C. J. Hall (2015). "An inducible transgene reports activation of macrophages in live zebrafish larvae". In: *Dev Comp Immunol* 53.1, pp. 63–9.
- Sanjana, N. E., O. Shalem, and F. Zhang (2014). "Improved vectors and genome-wide libraries for CRISPR screening". In: *Nat Methods* 11.8, pp. 783–784.
- Sankaran, V. G., J. S. Weissman, and L. I. Zon (2022). "Cellular barcoding to decipher clonal dynamics in disease". In: *Science* 378.6616, eabm5874.
- Santhakumar, K., E. C. Judson, P. M. Elks, S. McKee, S. Elworthy, E. van Rooijen, S. S. Walmsley, S. A. Renshaw, S. S. Cross, and F. J. van Eeden (2012). "A zebrafish model to study and therapeutically manipulate hypoxia signaling in tumorigenesis". In: *Cancer Res* 72.16, pp. 4017–27.
- Saraiva, M. and A. O'Garra (2010). "The regulation of IL-10 production by immune cells". In: *Nat Rev Immunol* 10.3, pp. 170–81.
- Sarathy, J. P. and V. Dartois (2020). "Caseum: a Niche for *Mycobacterium tuberculosis* Drug-Tolerant Persisters". In: *Clin Microbiol Rev* 33.3.
- Sarathy, J. P., G. Gruber, and T. Dick (2019). "Re-Understanding the Mechanisms of Action of the Anti-Mycobacterial Drug Bedaquiline". In: *Antibiotics (Basel)* 8.4.
- Sarsani, V. K., N. Raghupathy, I. T. Fiddes, J. Armstrong, F. Thibaud-Nissen, O. Zinder, M. Bolisetty, K. Howe, D. Hinerfeld, X. Ruan, L. Rowe, M. Barter, G. Ananda, B. Paten, G. M. Weinstock, G. A. Churchill, M. V. Wiles, V. A. Schneider, A. Srivastava, and L. G. Reinholdt (2019). "The Genome of C57BL/6J "Eve", the Mother of the Laboratory Mouse Genome Reference Strain". In: *G3 (Bethesda)* 9.6, pp. 1795–1805.
- Sasindran, S. J. and J. B. Torrelles (2011). "*Mycobacterium tuberculosis* Infection and Inflammation: what is Beneficial for the Host and for the Bacterium?" In: *Front Microbiol* 2, p. 2.
- Sassetti, C. M. and E. J. Rubin (2003). "Genetic requirements for mycobacterial survival during infection". In: *Proc Natl Acad Sci U S A* 100.22, pp. 12989–94.

- Savant, S., S. La Porta, A. Budnik, K. Busch, J. Hu, N. Tisch, C. Korn, A. F. Valls, A. V. Benest, D. Terhardt, X. Qu, R. H. Adams, H. S. Baldwin, C. Ruiz de Almodovar, H. R. Rodewald, and H. G. Augustin (2015). “The Orphan Receptor Tie1 Controls Angiogenesis and Vascular Remodeling by Differentially Regulating Tie2 in Tip and Stalk Cells”. In: *Cell Rep* 12.11, pp. 1761–73.
- Saxena, M. and G. Yeretssian (2014). “NOD-Like Receptors: Master Regulators of Inflammation and Cancer”. In: *Front Immunol* 5, p. 327.
- Scanga, C. A., A. Bafica, C. G. Feng, A. W. Cheever, S. Hieny, and A. Sher (2004). “MyD88-deficient mice display a profound loss in resistance to *Mycobacterium tuberculosis* associated with partially impaired Th1 cytokine and nitric oxide synthase 2 expression”. In: *Infect Immun* 72.4, pp. 2400–4.
- Schabbing, R. W., A. Garcia, and R. L. Hunter (1994). “Characterization of the trehalose 6,6'-dimycolate surface monolayer by scanning tunneling microscopy”. In: *Infect Immun* 62.2, pp. 754–6.
- Schamberg, Jay F. (1899). “The Nature of the Leprosy of the Bible. From a Medical and Biblical Point of View”. In: *The Biblical World* 13.3, pp. 162–169.
- Schatz, V., Y. Strussmann, A. Mahnke, G. Schley, M. Waldner, U. Ritter, J. Wild, C. Willam, N. Dehne, B. Brune, J. M. McNiff, O. R. Colegio, C. Bogdan, and J. Jantsch (2016). “Myeloid Cell-Derived HIF-1alpha Promotes Control of *Leishmania major*”. In: *J Immunol* 197.10, pp. 4034–4041.
- Schier, A. F., S. C. Neuhauss, M. Harvey, J. Malicki, L. Solnica-Krezel, D. Y. Stainier, F. Zwartkruis, S. Abdelilah, D. L. Stemple, Z. Rangini, H. Yang, and W. Driever (1996). “Mutations affecting the development of the embryonic zebrafish brain”. In: *Development* 123, pp. 165–78.
- Schild, Y., A. Mohamed, E. J. Wootton, A. Lewis, and P. M. Elks (2020). “Hif-1alpha stabilisation is protective against infection in zebrafish comorbid models”. In: *FEBS J* 287.18, pp. 3925–3943.
- Schilling, T. F., T. Piotrowski, H. Grandel, M. Brand, C. P. Heisenberg, Y. J. Jiang, D. Beuchle, M. Hammerschmidt, D. A. Kane, M. C. Mullins, F. J. van Eeden, R. N. Kelsh, M. Furutani-Seiki, M. Granato, P. Haffter, J. Odenthal, R. M. Warga, T. Trowe, and C. Nüsslein-Volhard (1996). “Jaw and branchial arch mutants in zebrafish I: branchial arches”. In: *Development* 123, pp. 329–44.

- Schindelin, J., I. Arganda-Carreras, E. Frise, V. Kaynig, M. Longair, T. Pietzsch, S. Preibisch, C. Rueden, S. Saalfeld, B. Schmid, J. Y. Tinevez, D. J. White, V. Hartenstein, K. Eliceiri, P. Tomancak, and A. Cardona (2012). “Fiji: an open-source platform for biological-image analysis”. In: *Nat Methods* 9.7, pp. 676–82.
- Schlesinger, L. S. (1993). “Macrophage phagocytosis of virulent but not attenuated strains of *Mycobacterium tuberculosis* is mediated by mannose receptors in addition to complement receptors”. In: *J Immunol* 150.7, pp. 2920–30.
- Schmidt, D., B. Textor, O. T. Pein, A. H. Licht, S. Andrecht, M. Sator-Schmitt, N. E. Fusenig, P. Angel, and M. Schorpp-Kistner (2007). “Critical role for NF-kappaB-induced JunB in VEGF regulation and tumor angiogenesis”. In: *EMBO J* 26.3, pp. 710–9.
- Schneider, C. A., W. S. Rasband, and K. W. Eliceiri (2012).** “NIH Image to ImageJ: 25 years of image analysis”. In: *Nat Methods* 9.7, pp. 671–5.
- Schoenen, H., A. Huber, N. Sonda, S. Zimmermann, J. Jantsch, B. Lepenies, V. Bronte, and R. Lang (2014). “Differential control of Mincle-dependent cord factor recognition and macrophage responses by the transcription factors C/EBPbeta and HIF1alpha”. In: *J Immunol* 193.7, pp. 3664–75.
- Schorey, J. S. and C. Lawrence (2008). “The pattern recognition receptor Dectin-1: from fungi to mycobacteria”. In: *Curr Drug Targets* 9.2, pp. 123–9.
- Schrager, L. K., J. Vekemans, N. Drager, D. M. Lewinsohn, and O. F. Olesen (2020). “The status of tuberculosis vaccine development”. In: *Lancet Infect Dis* 20.3, e28–e37.
- Schuermann, A., C. S. Helker, and W. Herzog (2014).** “Angiogenesis in zebrafish”. In: *Semin Cell Dev Biol* 31, pp. 106–14.
- Schulze, M., F. Belema-Bedada, A. Technau, and T. Braun (2005). “Mesenchymal stem cells are recruited to striated muscle by NFAT/IL-4-mediated cell fusion”. In: *Genes Dev* 19.15, pp. 1787–98.
- Schwegmann, A. and F. Brombacher (2008). “Host-directed drug targeting of factors hijacked by pathogens”. In: *Sci Signal* 1.29, re8.

- Scott, L. J., K. McKeage, S. J. Keam, and G. L. Plosker (2003). "Tacrolimus: a further update of its use in the management of organ transplantation". In: *Drugs* 63.12, pp. 1247–97.
- Secanella-Fandos, S., E. Noguera-Ortega, F. Olivares, M. Luquin, and E. Julian (2014). "Killed but metabolically active *Mycobacterium bovis* Bacillus Calmette-Guerin retains the antitumor ability of live Bacillus Calmette-Guerin". In: *J Urol* 191.5, pp. 1422–8.
- Seeliger, J. C., C. M. Holsclaw, M. W. Schelle, Z. Botyanszki, S. A. Gilmore, S. E. Tully, M. Niederweis, B. F. Cravatt, J. A. Leary, and C. R. Bertozzi (2012). "Elucidation and chemical modulation of sulfolipid-1 biosynthesis in *Mycobacterium tuberculosis*". In: *J Biol Chem* 287.11, pp. 7990–8000.
- Segerstrom, L., D. Fuchs, U. Backman, K. Holmquist, R. Christofferson, and F. Azarbayjani (2006). "The anti-VEGF antibody bevacizumab potently reduces the growth rate of high-risk neuroblastoma xenografts". In: *Pediatr Res* 60.5, pp. 576–81.
- Segre, J. A. (2013). "What does it take to satisfy Koch's postulates two centuries later? Microbial genomics and *Propionibacteria acnes*". In: *J Invest Dermatol* 133.9, pp. 2141–2.
- Seifert, A., S. Rau, G. Kullertz, B. Fischer, and A. N. Santos (2009). "TCDD induces cell migration via NFATc1/ATX-signaling in MCF-7 cells". In: *Toxicol Lett* 184.1, pp. 26–32.
- Selwyn, P. A., P. Alcabes, D. Hartel, D. Buono, E. E. Schoenbaum, R. S. Klein, K. Davenny, and G. H. Friedland (1992). "Clinical manifestations and predictors of disease progression in drug users with human immunodeficiency virus infection". In: *N Engl J Med* 327.24, pp. 1697–703.
- Senavirathna, L. K., C. Huang, X. Yang, M. C. Munteanu, R. Sathiaseelan, D. Xu, C. A. Henke, and L. Liu (2018). "Hypoxia induces pulmonary fibroblast proliferation through NFAT signaling". In: *Sci Rep* 8.1, p. 2709.
- SenGupta, S., L. E. Hittle, R. K. Ernst, S. M. Uriarte, and T. C. Mitchell (2016). "A *Pseudomonas aeruginosa* hepta-acylated lipid A variant associated with cystic fibrosis selectively activates human neutrophils". In: *J Leukoc Biol* 100.5, pp. 1047–1059.

- Serhan, C. N. and N. Chiang (2008). "Endogenous pro-resolving and anti-inflammatory lipid mediators: a new pharmacologic genus". In: *Br J Pharmacol* 153 Suppl 1, S200–15.
- Serrano-Gomez, D., J. A. Leal, and A. L. Corbi (2005). "DC-SIGN mediates the binding of Aspergillus fumigatus and keratinophytic fungi by human dendritic cells". In: *Immunobiology* 210.2-4, pp. 175–83.
- Sertori, R., R. Jones, F. Basheer, L. Rivera, S. Dawson, S. Loke, S. Heidary, A. Dhillon, C. Liongue, and A. C. Ward (2022). "Generation and Characterization of a Zebrafish IL-2Rgammac SCID Model". In: *Int J Mol Sci* 23.4.
- Sertori, R., C. Liongue, F. Basheer, K. L. Lewis, P. Rasighaemi, D. de Coninck, D. Traver, and A. C. Ward (2016). "Conserved IL-2Rgammac Signaling Mediates Lymphopoiesis in Zebrafish". In: *J Immunol* 196.1, pp. 135–43.
- Sesler, C. L. and M. Zayzafoon (2013). "NFAT signaling in osteoblasts regulates the hematopoietic niche in the bone microenvironment". In: *Clin Dev Immunol* 2013, p. 107321.
- Seweryn, E., J. Pietkiewicz, A. Szamborska, and A. Gamian (2007). "Enolase on the surface of prokaryotic and eukaryotic cells is a receptor for human plasminogen". In: *Postepy Hig Med Dosw (Online)* 61, pp. 672–82.
- Shaku, M., C. Ealand, and B. D. Kana (2020). "Cell Surface Biosynthesis and Remodeling Pathways in Mycobacteria Reveal New Drug Targets". In: *Front Cell Infect Microbiol* 10, p. 603382.
- Shan, S., S. Luo, Z. Yang, J. Hong, Y. Su, F. Ding, L. Fu, C. Li, P. Chen, J. Ma, X. Shi, Q. Zhang, B. Berger, L. Zhang, and J. Peng (2022). "Deep learning guided optimization of human antibody against SARS-CoV-2 variants with broad neutralization". In: *Proc Natl Acad Sci U S A* 119.11, e2122954119.
- Shariati, F., J. L. Perez-Arellano, C. Carranza, J. Lopez-Aban, B. Vicente, M. Arefi, and A. Muro (2011). "Evaluation of the role of angiogenic factors in the pathogenesis of schistosomiasis". In: *Exp Parasitol* 128.1, pp. 44–9.
- Sharma, A., S. Sinha, and N. Shrivastava (2022). "Therapeutic Targeting Hypoxia-Inducible Factor (HIF-1) in Cancer: Cutting Gordian Knot of Cancer Cell Metabolism". In: *Front Genet* 13, p. 849040.

- Sharma, D., A. R. Cukras, E. J. Rogers, D. R. Southworth, and R. Green (2007). “Mutational analysis of S12 protein and implications for the accuracy of decoding by the ribosome”. In: *J Mol Biol* 374.4, pp. 1065–76.
- Sharma, S., G. M. Findlay, H. S. Bandukwala, S. Oberdoerffer, B. Baust, Z. Li, V. Schmidt, P. G. Hogan, D. B. Sacks, and A. Rao (2011). “Dephosphorylation of the nuclear factor of activated T cells (NFAT) transcription factor is regulated by an RNA-protein scaffold complex”. In: *Proc Natl Acad Sci U S A* 108.28, pp. 11381–6.
- Sharrock, A. V., S. P. McManaway, M. H. Rich, J. S. Mumm, I. F. Hermans, M. Tercel, F. B. Pruijn, and D. F. Ackerley (2021). “Engineering the *Escherichia coli* Nitroreductase NfsA to Create a Flexible Enzyme-Prodrug Activation System”. In: *Front Pharmacol* 12, p. 701456.
- Shaw, J. P., P. J. Utz, D. B. Durand, J. J. Toole, E. A. Emmel, and G. R. Crabtree (1988). “Identification of a putative regulator of early T cell activation genes”. In: *Science* 241.4862, pp. 202–5.
- Shaw, M. H., T. Reimer, Y. G. Kim, and G. Nunez (2008). “NOD-like receptors (NLRs): bona fide intracellular microbial sensors”. In: *Curr Opin Immunol* 20.4, pp. 377–82.
- Shen, Y., R. Jin, X. Liang, Z. Deng, J. He, Y. Ding, F. Ding, L. Lu, F. Liu, and J. Yang (2022). “Angiogenesis modulation-mediated inhibitory effects of tacrolimus on hypertrophic scar formation”. In: *Microvasc Res*, p. 104446.**
- Shi, J., Y. Zhao, Y. Wang, W. Gao, J. Ding, P. Li, L. Hu, and F. Shao (2014). “Inflammatory caspases are innate immune receptors for intracellular LPS”. In: *Nature* 514.7521, pp. 187–92.
- Shi, L., Q. Jiang, Y. Bushkin, S. Subbian, and S. Tyagi (2019). “Biphasic Dynamics of Macrophage Immunometabolism during *Mycobacterium tuberculosis* Infection”. In: *mBio* 10.2.
- Shi, S., C. Nathan, D. Schnappinger, J. Drenkow, M. Fuortes, E. Block, A. Ding, T. R. Gingeras, G. Schoolnik, S. Akira, K. Takeda, and S. Ehrt (2003). “MyD88 primes macrophages for full-scale activation by interferon-gamma yet mediates few responses to *Mycobacterium tuberculosis*”. In: *J Exp Med* 198.7, pp. 987–97.

- Shi, W., X. Zhang, X. Jiang, H. Yuan, J. S. Lee, 3rd Barry C. E., H. Wang, W. Zhang, and Y. Zhang (2011). “Pyrazinamide inhibits trans-translation in *Mycobacterium tuberculosis*”. In: *Science* 333.6049, pp. 1630–2.
- Shiau, C. E., Z. Kaufman, A. M. Meireles, and W. S. Talbot (2015). “Differential requirement for *irf8* in formation of embryonic and adult macrophages in zebrafish”. In: *PLoS One* 10.1, e0117513.
- Shibuya, K., A. Hirata, J. Omura, M. Sugamata, S. Katori, N. Saito, N. Murata, A. Morita, K. Takahashi, C. Hasegawa, A. Mitsuda, T. Hatori, and H. Nonaka (2005). “Granuloma and cryptococcosis”. In: *J Infect Chemother* 11.3, pp. 115–22.
- Shibuya, M. (2011). “Vascular Endothelial Growth Factor (VEGF) and Its Receptor (VEGFR) Signaling in Angiogenesis: A Crucial Target for Anti- and Pro-Angiogenic Therapies”. In: *Genes Cancer* 2.12, pp. 1097–105.
- Shibuya, M. and L. Claesson-Welsh (2006). “Signal transduction by VEGF receptors in regulation of angiogenesis and lymphangiogenesis”. In: *Exp Cell Res* 312.5, pp. 549–60.
- Shih, S. C. and K. P. Claffey (2001). “Role of AP-1 and HIF-1 transcription factors in TGF-beta activation of VEGF expression”. In: *Growth Factors* 19.1, pp. 19–34.
- Shin, M., T. J. Beane, A. Quillien, I. Male, L. J. Zhu, and N. D. Lawson (2016). “Vegfa signals through ERK to promote angiogenesis, but not artery differentiation”. In: *Development* 143.20, pp. 3796–3805.
- Shin, M., I. Male, T. J. Beane, J. A. Villefranc, F. O. Kok, L. J. Zhu, and N. D. Lawson (2017). “Vegfc acts through ERK to induce sprouting and differentiation of trunk lymphatic progenitors”. In: *Development* 144.3, p. 531.
- Shin, M., T. Nozaki, F. Idrizi, S. Isogai, K. Ogasawara, K. Ishida, S. Yuge, B. Roscoe, S. A. Wolfe, S. Fukuhara, N. Mochizuki, T. Deguchi, and N. D. Lawson (2019). “Valves Are a Conserved Feature of the Zebrafish Lymphatic System”. In: *Dev Cell* 51.3, 374–386 e5.
- Shiokawa, M., S. Yamasaki, and S. Saijo (2017). “C-type lectin receptors in anti-fungal immunity”. In: *Curr Opin Microbiol* 40, pp. 123–130.

Shortman, K. and Y. J. Liu (2002). "Mouse and human dendritic cell subtypes". In: *Nat Rev Immunol* 2.3, pp. 151–61.

Shoshan, E., R. R. Braeuer, T. Kamiya, A. K. Mobley, L. Huang, M. E. Vasquez, G. Velazquez-Torres, N. Chakravarti, C. Ivan, V. Prieto, G. J. Villares, and M. Bar-Eli (2016). "NFAT1 Directly Regulates IL8 and MMP3 to Promote Melanoma Tumor Growth and Metastasis". In: *Cancer Res* 76.11, pp. 3145–55.

Shrivastava, P. and T. Bagchi (2013). "IL-10 modulates *in vitro* multinucleate giant cell formation in human tuberculosis". In: *PLoS One* 8.10, e77680.

Shwartz, A., W. Goessling, and C. Yin (2019). "Macrophages in Zebrafish Models of Liver Diseases". In: *Front Immunol* 10, p. 2840.

Sica, A., L. Dorman, V. Viggiano, M. Cippitelli, P. Ghosh, N. Rice, and H. A. Young (1997). "Interaction of NF-kappaB and NFAT with the interferon-gamma promoter". In: *J Biol Chem* 272.48, pp. 30412–20.

Siddiqi, O. K., M. Ghebremichael, K. Musukuma, X. Dang, E. Mubanga, G. L. Birbeck, S. Love, C. Buback, B. Kosloff, H. Ayles, M. Atadzhanyan, and I. J. Koralnik (2021). "LTA4H Prevalence and Mortality in Adult Zambians with Tuberculous Meningitis". In: *Ann Neurol* 90.6, pp. 994–998.

Siggers, T., A. B. Chang, A. Teixeira, D. Wong, K. J. Williams, B. Ahmed, J. Ragoussis, I. A. Udalova, S. T. Smale, and M. L. Bulyk (2011). "Principles of dimer-specific gene regulation revealed by a comprehensive characterization of NF-kappaB family DNA binding". In: *Nat Immunol* 13.1, pp. 95–102.

Silva, C. L. and L. H. Faccioli (1988). "Tumor necrosis factor (cachectin) mediates induction of cachexia by cord factor from mycobacteria". In: *Infect Immun* 56.12, pp. 3067–71.

Simeone, R., A. Bobard, J. Lippmann, W. Bitter, L. Majlessi, R. Brosch, and J. Enninga (2012). "Phagosomal rupture by *Mycobacterium tuberculosis* results in toxicity and host cell death". In: *PLoS Pathog* 8.2, e1002507.

Simmons, K. B., L. B. Haddad, K. Nanda, and K. M. Curtis (2018a). "Drug interactions between non-rifamycin antibiotics and hormonal contraception: a systematic review". In: *Am J Obstet Gynecol* 218.1, 88–97 e14.

Simmons, K. B., L. B. Haddad, K. Nanda, and K. M. Curtis (2018b). “Drug interactions between rifamycin antibiotics and hormonal contraception: a systematic review”. In: *Bjog* 125.7, pp. 804–811.

Simonett, S. P., S. Shin, J. A. Herring, R. Bacher, L. A. Smith, C. Dong, M. E. Rabaglia, D. S. Stapleton, K. L. Schueler, J. Choi, M. N. Bernstein, D. R. Turkewitz, C. Perez-Cervantes, J. Spaeth, R. Stein, J. S. Tessem, C. Kendziorski, S. Keles, I. P. Moskowitz, M. P. Keller, and A. D. Attie (2021). “Identification of direct transcriptional targets of NFATC2 that promote beta cell proliferation”. In: *J Clin Invest* 131.21.

Simons, M., E. Gordon, and L. Claesson-Welsh (2016). “Mechanisms and regulation of endothelial VEGF receptor signalling”. In: *Nat Rev Mol Cell Biol* 17.10, pp. 611–25.

Simpson, B. W. and M. S. Trent (2019). “Pushing the envelope: LPS modifications and their consequences”. In: *Nat Rev Microbiol* 17.7, pp. 403–416.

Singh, J. A., A. Hossain, A. Kotb, and G. Wells (2016b). “Risk of serious infections with immunosuppressive drugs and glucocorticoids for lupus nephritis: a systematic review and network meta-analysis”. In: *BMC Med* 14.1, p. 137.

Singh, S. B., A. S. Davis, G. A. Taylor, and V. Deretic (2006). “Human IRGM induces autophagy to eliminate intracellular mycobacteria”. In: *Science* 313.5792, pp. 1438–41.

Singh, S. B., W. Ornatowski, I. Vergne, J. Naylor, M. Delgado, E. Roberts, M. Ponpuak, S. Master, M. Pilli, E. White, M. Komatsu, and V. Deretic (2010). “Human IRGM regulates autophagy and cell-autonomous immunity functions through mitochondria”. In: *Nat Cell Biol* 12.12, pp. 1154–65.

Sinsimer, D., G. Huet, C. Manca, L. Tsanova, M. S. Koo, N. Kurepina, B. Kana, B. Mathema, S. A. Marras, B. N. Kreiswirth, C. Guilhot, and G. Kaplan (2008). “The phenolic glycolipid of *Mycobacterium tuberculosis* differentially modulates the early host cytokine response but does not in itself confer hypervirulence”. In: *Infect Immun* 76.7, pp. 3027–36.

– *The role of the phenolic glycolipid in tuberculosis is still somewhat controversial, but this equivocating study found that PGL alters host cytokine production but is not required for virulence. However, these in vitro studies were unable to fully dissect these effects and later work would clarify some of the functions of PGL and why*

it is so prevalent in circulating strains but has been lost in some sublineages of the laboratory reference strain H37Rv.

Skolnick, J. L., B. S. Stoler, D. B. Katz, and W. H. Anderson (1976). “Rifampin, oral contraceptives, and pregnancy”. In: *Jama* 236.12, p. 1382.

Smith, C. M., R. E. Baker, M. K. Proulx, B. B. Mishra, J. E. Long, S. W. Park, H. N. Lee, M. C. Kiritsy, M. M. Bellerose, A. J. Olive, K. C. Murphy, K. Papavinasasundaram, F. J. Boehm, C. J. Reames, R. K. Meade, B. K. Hampton, C. L. Linnertz, G. D. Shaw, P. Hock, T. A. Bell, S. Ehrt, D. Schnappinger, F. Pardo-Manuel de Villena, M. T. Ferris, T. R. Ioerger, and C. M. Sasetti (2022). “Host-pathogen genetic interactions underlie tuberculosis susceptibility in genetically diverse mice”. In: *Elife* 11.

Smith, C. M., M. K. Proulx, A. J. Olive, D. Laddy, B. B. Mishra, C. Moss, N. M. Gutierrez, M. M. Bellerose, P. Barreira-Silva, J. Y. Phuah, R. E. Baker, S. M. Behar, H. Kornfeld, T. G. Evans, G. Beamer, and C. M. Sasetti (2016). “Tuberculosis Susceptibility and Vaccine Protection Are Independently Controlled by Host Genotype”. In: *mBio* 7.5.

Smith, K. A. (2012). “Louis pasteur, the father of immunology?” In: *Front Immunol* 3, p. 68.

Soares-Silva, M., F. F. Diniz, G. N. Gomes, and D. Bahia (2016). “The Mitogen-Activated Protein Kinase (MAPK) Pathway: Role in Immune Evasion by Trypanosomatids”. In: *Front Microbiol* 7, p. 183.

Sofroniew, Nicholas, Talley Lambert, Kira Evans, Juan Nunez-Iglesias, Grzegorz Bokota, Philip Winston, Gonzalo Peña-Castellanos, Kevin Yamauchi, Matthias Bussonnier, Draga Doncila Pop, Ahmet Can Solak, Ziyang Liu, Pam Wadhwa, Alister Burt, Genevieve Buckley, Andrew Sweet, Lukasz Migas, Volker Hilsenstein, Lorenzo Gaivas, Jordão Bragantini, Jaime Rodríguez-Guerra, Hector Muñoz, Jeremy Freeman, Peter Boone, Alan Lowe, Christoph Gohlke, Loic Royer, Andrea PIERRÉ, Hagai Har-Gil, and Abigail McGovern (Nov. 2022). *napari: a multidimensional image viewer for Python*. Version v0.4.17rc8.

Sohn, H. W., H. Gu, and S. K. Pierce (2003). “Cbl-b negatively regulates B cell antigen receptor signaling in mature B cells through ubiquitination of the tyrosine kinase Syk”. In: *J Exp Med* 197.11, pp. 1511–24.

- Somashekar, B. S., A. G. Amin, C. D. Rithner, J. Troudt, R. Basaraba, A. Izzo, D. C. Crick, and D. Chatterjee (2011). “Metabolic profiling of lung granuloma in *Mycobacterium tuberculosis* infected guinea pigs: ex vivo ^1H magic angle spinning NMR studies”. In: *J Proteome Res* 10.9, pp. 4186–95.
- Sonanez-Organis, J. G., J. A. Godoy-Lugo, M. L. Hernandez-Palomares, D. Rodriguez-Martinez, J. A. Rosas-Rodriguez, G. Gonzalez-Ochoa, A. Virgen-Ortiz, and R. M. Ortiz (2016). “HIF-1alpha and PPARgamma during physiological cardiac hypertrophy induced by pregnancy: Transcriptional activities and effects on target genes”. In: *Gene* 591.2, pp. 376–81.
- Song, J., W. Chen, X. Cui, Z. Huang, D. Wen, Y. Yang, W. Yu, L. Cui, and C. Y. Liu (2020). “CCBE1 promotes tumor lymphangiogenesis and is negatively regulated by TGFbeta signaling in colorectal cancer”. In: *Theranostics* 10.5, pp. 2327–2341.
- Sorkin, E., H. Erlenmeyer, and H. Bloch (1952). “Purification of a lipid material ('cord factor') obtained from young cultures of tubercle bacilli”. In: *Nature* 170.4316, p. 124.
- Soroldoni, D., B. M. Hogan, and A. C. Oates (2009). “Simple and efficient transgenesis with meganuclease constructs in zebrafish”. In: *Methods Mol Biol* 546, pp. 117–30.
- Souders, C. R. and A. T. Smith (1952). “The clinical significance of hemoptysis”. In: *N Engl J Med* 247.21, pp. 790–3.
- Soza-Ried, C., I. Hess, N. Netuschil, M. Schorpp, and T. Boehm (2010). “Essential role of c-myb in definitive hematopoiesis is evolutionarily conserved”. In: *Proc Natl Acad Sci U S A* 107.40, pp. 17304–8.
- Spargo, B. J., L. M. Crowe, T. Ioneda, B. L. Beaman, and J. H. Crowe (1991). “Cord factor (alpha,alpha-trehalose 6,6'-dimycolate) inhibits fusion between phospholipid vesicles”. In: *Proc Natl Acad Sci U S A* 88.3, pp. 737–40.
- Spyropoulos, B., P. B. Moens, J. Davidson, and J. A. Lowden (1981). “Heterozygote advantage in Tay-Sachs carriers?” In: *Am J Hum Genet* 33.3, pp. 375–80.
- Srivastava, S., J. D. Ernst, and L. Desvignes (2014). “Beyond macrophages: the diversity of mononuclear cells in tuberculosis”. In: *Immunol Rev* 262.1, pp. 179–92.

Stachura, D. L., O. Svoboda, C. A. Campbell, R. Espin-Palazon, R. P. Lau, L. I. Zon, P. Bartunek, and D. Traver (2013). “The zebrafish granulocyte colony-stimulating factors (Gcsfs): 2 paralogous cytokines and their roles in hematopoietic development and maintenance”. In: *Blood* 122.24, pp. 3918–28.

Stacker, S. A., S. P. Williams, T. Karnezis, R. Shayan, S. B. Fox, and M. G. Achen (2014). “Lymphangiogenesis and lymphatic vessel remodelling in cancer”. In: *Nat Rev Cancer* 14.3, pp. 159–72.

Stainier, D. Y., B. Fouquet, J. N. Chen, K. S. Warren, B. M. Weinstein, S. E. Meiler, M. A. Mohideen, S. C. Neuhauss, L. Solnica-Krezel, A. F. Schier, F. Zwartkruis, D. L. Stemple, J. Malicki, W. Driever, and M. C. Fishman (1996). “Mutations affecting the formation and function of the cardiovascular system in the zebrafish embryo”. In: *Development* 123, pp. 285–92.

Stainier, D. Y., Z. Kontarakis, and A. Rossi (2015). “Making sense of anti-sense data”. In: *Dev Cell* 32.1, pp. 7–8.

Stainier, D. Y., B. M. Weinstein, 3rd Detrich H. W., L. I. Zon, and M. C. Fishman (1995). “Cloche, an early acting zebrafish gene, is required by both the endothelial and hematopoietic lineages”. In: *Development* 121.10, pp. 3141–50.

– Didier Stainier’s description of the naturally occurring cloche mutation was a pioneering study on the common origins of the hematopoietic and endothelial cell lineages and established a platform upon which two decades of research had to be done to identify the functional basis of this mutation and how it contributed to the development of these two critical organ systems.

Stainier, D. Y. R., E. Raz, N. D. Lawson, S. C. Ekker, R. D. Burdine, J. S. Eisen, P. W. Ingham, S. Schulte-Merker, D. Yelon, B. M. Weinstein, M. C. Mullins, S. W. Wilson, L. Ramakrishnan, S. L. Amacher, S. C. F. Neuhauss, A. Meng, N. Mochizuki, P. Panula, and C. B. Moens (2017). “Guidelines for morpholino use in zebrafish”. In: *PLoS Genet* 13.10, e1007000.

Stamm, C. E., A. C. Collins, and M. U. Shiloh (2015). “Sensing of *Mycobacterium tuberculosis* and consequences to both host and bacillus”. In: *Immunol Rev* 264.1, pp. 204–19.

- Stamm, L. M. and E. J. Brown (2004). “*Mycobacterium marinum*: the generalization and specialization of a pathogenic mycobacterium”. In: *Microbes Infect* 6.15, pp. 1418–28.
- Stancu, C. and A. Sima (2001). “Statins: mechanism of action and effects”. In: *J Cell Mol Med* 5.4, pp. 378–87.
- Stanley, S. A., S. Raghavan, W. W. Hwang, and J. S. Cox (2003). “Acute infection and macrophage subversion by *Mycobacterium tuberculosis* require a specialized secretion system”. In: *Proc Natl Acad Sci U S A* 100.22, pp. 13001–6.
- Stark, K., A. Eckart, S. Haidari, A. Tirmiceriu, M. Lorenz, M. L. von Bruhl, F. Gartner, A. G. Khandoga, K. R. Legate, R. Pless, I. Hepper, K. Lauber, B. Walzog, and S. Massberg (2013). “Capillary and arteriolar pericytes attract innate leukocytes exiting through venules and ‘instruct’ them with pattern-recognition and motility programs”. In: *Nat Immunol* 14.1, pp. 41–51.
- Starr, Michael Seth (2016). *Ringo: With a Little Help*. Milwaukee, WI: Backbeat Books.
- Staskawicz, B. J., F. M. Ausubel, B. J. Baker, J. G. Ellis, and J. D. Jones (1995). “Molecular genetics of plant disease resistance”. In: *Science* 268.5211, pp. 661–7.
- Steele, M. A. and R. M. Des Prez (1988). “The role of pyrazinamide in tuberculosis chemotherapy”. In: *Chest* 94.4, pp. 845–50.
- Stefanec, T. (2000). “Endothelial apoptosis: could it have a role in the pathogenesis and treatment of disease?” In: *Chest* 117.3, pp. 841–54.
- Steimle, A., I. B. Autenrieth, and J. S. Frick (2016). “Structure and function: Lipid A modifications in commensals and pathogens”. In: *Int J Med Microbiol* 306.5, pp. 290–301.
- Stemple, D. L., L. Solnica-Krezel, F. Zwartkruis, S. C. Neuhauss, A. F. Schier, J. Malicki, D. Y. Stainier, S. Abdelilah, Z. Rangini, E. Mountcastle-Shah, and W. Driever (1996). “Mutations affecting development of the notochord in zebrafish”. In: *Development* 123, pp. 117–28.
- Stern, H. M. and L. I. Zon (2003). “Cancer genetics and drug discovery in the zebrafish”. In: *Nat Rev Cancer* 3.7, pp. 533–9.

- Stevenson, A. S., M. F. Gomez, D. C. Hill-Eubanks, and M. T. Nelson (2001). “NFAT4 movement in native smooth muscle. A role for differential Ca(2+) signaling”. In: *J Biol Chem* 276.18, pp. 15018–24.
- Stewart, G. R., B. D. Robertson, and D. B. Young (2003). “Tuberculosis: a problem with persistence”. In: *Nat Rev Microbiol* 1.2, pp. 97–105.
- Stinear, T. P., G. A. Jenkin, P. D. Johnson, and J. K. Davies (2000). “Comparative genetic analysis of *Mycobacterium ulcerans* and *Mycobacterium marinum* reveals evidence of recent divergence”. In: *J Bacteriol* 182.22, pp. 6322–30.
- Stinear, T. P., T. Seemann, P. F. Harrison, G. A. Jenkin, J. K. Davies, P. D. Johnson, Z. Abdellah, C. Arrowsmith, T. Chillingworth, C. Churcher, K. Clarke, A. Cronin, P. Davis, I. Goodhead, N. Holroyd, K. Jagels, A. Lord, S. Moule, K. Mungall, H. Norbertczak, M. A. Quail, E. Rabbinowitsch, D. Walker, B. White, S. Whitehead, P. L. Small, R. Brosch, L. Ramakrishnan, M. A. Fischbach, J. Parkhill, and S. T. Cole (2008). “Insights from the complete genome sequence of *Mycobacterium marinum* on the evolution of *Mycobacterium tuberculosis*”.** In: *Genome Res* 18.5, pp. 729–41.
- Stirling, D. R., M. J. Swain-Bowden, A. M. Lucas, A. E. Carpenter, B. A. Cimini, and A. Goodman (2021). “CellProfiler 4: improvements in speed, utility and usability”. In: *BMC Bioinformatics* 22.1, p. 433.
- Stokes, R. W. and D. Doxsee (1999). “The receptor-mediated uptake, survival, replication, and drug sensitivity of *Mycobacterium tuberculosis* within the macrophage-like cell line THP-1: a comparison with human monocyte-derived macrophages”. In: *Cell Immunol* 197.1, pp. 1–9.
- Stratman, A. N., S. A. Pezoa, O. M. Farrelly, D. Castranova, 3rd Dye L. E., M. G. Butler, H. Sidik, W. S. Talbot, and B. M. Weinstein (2017). “Interactions between mural cells and endothelial cells stabilize the developing zebrafish dorsal aorta”. In: *Development* 144.1, pp. 115–127.
- Stratmann, A., T. Acker, A. M. Burger, K. Amann, W. Risau, and K. H. Plate (2001). “Differential inhibition of tumor angiogenesis by tie2 and vascular endothelial growth factor receptor-2 dominant-negative receptor mutants”. In: *Int J Cancer* 91.3, pp. 273–82.

- Streisinger, G., C. Walker, N. Dower, D. Knauber, and F. Singer (1981). “Production of clones of homozygous diploid zebra fish (*BrachyDanio rerio*)”. In: *Nature* 291.5813, pp. 293–6.
- Stuart, G. W., J. V. McMurray, and M. Westerfield (1988). “Replication, integration and stable germ-line transmission of foreign sequences injected into early zebrafish embryos”. In: *Development* 103.2, pp. 403–12.
- Stunault, M. I., G. Bories, R. R. Guinamard, and S. Ivanov (2018). “Metabolism Plays a Key Role during Macrophage Activation”. In: *Mediators Inflamm* 2018, p. 2426138.
- Sugawara, I., T. Udagawa, S. C. Hua, M. Reza-Gholizadeh, K. Otomo, Y. Saito, and H. Yamada (2002). “Pulmonary granulomas of guinea pigs induced by inhalation exposure of heat-treated BCG Pasteur, purified trehalose dimycolate and methyl ketomycolate”. In: *J Med Microbiol* 51.2, pp. 131–137.
- Sugawara, I., H. Yamada, S. Mizuno, K. Takeda, and S. Akira (2003). “Mycobacterial infection in MyD88-deficient mice”. In: *Microbiol Immunol* 47.11, pp. 841–7.
- Sugden, W. W., R. Meissner, T. Aegeater-Wilmsen, R. Tsaryk, E. V. Leonard, J. Bussmann, M. J. Hamm, W. Herzog, Y. Jin, L. Jakobsson, C. Denz, and A. F. Siekmann (2017). “Endoglin controls blood vessel diameter through endothelial cell shape changes in response to haemodynamic cues”. In: *Nat Cell Biol* 19.6, pp. 653–665.
- Sugimoto, K., S. P. Hui, D. Z. Sheng, and K. Kikuchi (2017). “Dissection of zebrafish shha function using site-specific targeting with a Cre-dependent genetic switch”. In: *Elife* 6.
- Sugita, T., M. Tajima, T. Ito, M. Saito, R. Tsuboi, and A. Nishikawa (2005). “Antifungal activities of tacrolimus and azole agents against the eleven currently accepted Malassezia species”. In: *J Clin Microbiol* 43.6, pp. 2824–9.
- Sukumar, N., S. Tan, B. B. Aldridge, and D. G. Russell (2014). “Exploitation of *Mycobacterium tuberculosis* reporter strains to probe the impact of vaccination at sites of infection”. In: *PLoS Pathog* 10.9, e1004394.

- Sullivan, C., M. A. Matty, D. Jurczyszak, K. A. Gabor, P. J. Millard, D. M. Tobin, and C. H. Kim (2017). "Infectious disease models in zebrafish". In: *Methods Cell Biol* 138, pp. 101–136.
- Sun, D., Y. Heianza, X. Li, X. Shang, S. R. Smith, G. A. Bray, F. M. Sacks, and L. Qi (2018). "Genetic, epigenetic and transcriptional variations at NFATC2IP locus with weight loss in response to diet interventions: The POUNDS Lost Trial". In: *Diabetes Obes Metab* 20.9, pp. 2298–2303.
- Sun, J., A. Siroy, R. K. Lokareddy, A. Speer, K. S. Doornbos, G. Cingolani, and M. Niederweis (2015). "The tuberculosis necrotizing toxin kills macrophages by hydrolyzing NAD". In: *Nat Struct Mol Biol* 22.9, pp. 672–8.
- Sunderkotter, C., K. Steinbrink, M. Goebeler, R. Bhardwaj, and C. Sorg (1994). "Macrophages and angiogenesis". In: *J Leukoc Biol* 55.3, pp. 410–22.
- Suurvali, J., A. R. Whiteley, Y. Zheng, K. Gharbi, M. Leptin, and T. Wiehe (2020). "The Laboratory Domestication of Zebrafish: From Diverse Populations to Inbred Substrains". In: *Mol Biol Evol* 37.4, pp. 1056–1069.
- Suzuki, T., M. Hayashi, T. Komatsu, A. Tanioka, M. Nagasawa, K. Tanimura-Inagaki, M. S. Rahman, S. Masuda, K. Yusa, J. Sakai, H. Shibata, and T. Inagaki (2021). "Measurement of the nuclear concentration of alpha-ketoglutarate during adipocyte differentiation by using a fluorescence resonance energy transfer-based biosensor with nuclear localization signals". In: *Endocr J* 68.12, pp. 1429–1438.
- Svedberg, F. R., S. L. Brown, M. Z. Krauss, L. Campbell, C. Sharpe, M. Clausen, G. J. Howell, H. Clark, J. Madsen, C. M. Evans, T. E. Sutherland, A. C. Ivens, D. J. Thornton, R. K. Grencis, T. Hussell, D. M. Cunoosamy, P. C. Cook, and A. S. MacDonald (2019). "The lung environment controls alveolar macrophage metabolism and responsiveness in type 2 inflammation". In: *Nat Immunol* 20.5, pp. 571–580.
- Swaim, L. E., L. E. Connolly, H. E. Volkman, O. Humbert, D. E. Born, and L. Ramakrishnan (2006).** "*Mycobacterium marinum* infection of adult zebrafish causes caseating granulomatous tuberculosis and is moderated by adaptive immunity". In: *Infect Immun* 74.11, pp. 6108–17.
- Symes, A., T. Gearan, and J. S. Fink (1998). "NFAT interactions with the vasoactive intestinal peptide cytokine response element". In: *J Neurosci Res* 52.1, pp. 93–104.

- Szabo, C. (2007). "Hydrogen sulphide and its therapeutic potential". In: *Nat Rev Drug Discov* 6.11, pp. 917–35.
- Sztal, T. E. and D. Y. R. Stainier (2020). "Transcriptional adaptation: a mechanism underlying genetic robustness". In: *Development* 147.15.
- Szuhai, K., M. Ijszenga, D. de Jong, A. Karseladze, H. J. Tanke, and P. C. Hogendoorn (2009). "The NFATc2 gene is involved in a novel cloned translocation in a Ewing sarcoma variant that couples its function in immunology to oncology". In: *Clin Cancer Res* 15.7, pp. 2259–68.
- Tabata, H., H. Morita, H. Kaji, K. Tohyama, and Y. Tohyama (2020). "Syk facilitates phagosome-lysosome fusion by regulating actin-remodeling in complement-mediated phagocytosis". In: *Sci Rep* 10.1, p. 22086.
- Tailleux, L., N. Pham-Thi, A. Bergeron-Lafaurie, J. L. Herrmann, P. Charles, O. Schwartz, P. Scheinmann, P. H. Lagrange, J. de Blic, A. Tazi, B. Gicquel, and O. Neyrolles (2005). "DC-SIGN induction in alveolar macrophages defines privileged target host cells for mycobacteria in patients with tuberculosis". In: *PLoS Med* 2.12, e381.
- Tailleux, L., O. Schwartz, J. L. Herrmann, E. Pivert, M. Jackson, A. Amara, L. Legres, D. Dreher, L. P. Nicod, J. C. Gluckman, P. H. Lagrange, B. Gicquel, and O. Neyrolles (2003). "DC-SIGN is the major *Mycobacterium tuberculosis* receptor on human dendritic cells". In: *J Exp Med* 197.1, pp. 121–7.
- Tak, U., T. Dokland, and M. Niederweis (2021). "Pore-forming Esx proteins mediate toxin secretion by *Mycobacterium tuberculosis*". In: *Nat Commun* 12.1, p. 394.
- Tak, U., J. Vlach, A. Garza-Garcia, D. William, O. Danilchanka, L. P. S. de Carvalho, J. S. Saad, and M. Niederweis (2019). "The tuberculosis necrotizing toxin is an NAD(+) and NADP(+) glycohydrolase with distinct enzymatic properties". In: *J Biol Chem* 294.9, pp. 3024–3036.
- Takaki, K., J. M. Davis, K. Winglee, and L. Ramakrishnan (2013). "Evaluation of the pathogenesis and treatment of *Mycobacterium marinum* infection in zebrafish". In: *Nat Protoc* 8.6, pp. 1114–24.

- Takaki, K. K., G. Rinaldi, M. Berriman, A. J. Pagan, and L. Ramakrishnan (2021). “*Schistosoma mansoni* Eggs Modulate the Timing of Granuloma Formation to Promote Transmission”. In: *Cell Host Microbe* 29.1, 58–67 e5.
- Takayama, K., C. Wang, and G. S. Besra (2005). “Pathway to synthesis and processing of mycolic acids in *Mycobacterium tuberculosis*”. In: *Clin Microbiol Rev* 18.1, pp. 81–101.
- Takayanagi, H., S. Kim, T. Koga, H. Nishina, M. Isshiki, H. Yoshida, A. Saiura, M. Isobe, T. Yokochi, J. Inoue, E. F. Wagner, T. W. Mak, T. Kodama, and T. Taniguchi (2002). “Induction and activation of the transcription factor NFATc1 (NFAT2) integrate RANKL signaling in terminal differentiation of osteoclasts”. In: *Dev Cell* 3.6, pp. 889–901.
- Takeda, K. and S. Akira (2004). “TLR signaling pathways”. In: *Semin Immunol* 16.1, pp. 3–9.
- Tammela, T., G. Zarkada, H. Nurmi, L. Jakobsson, K. Heinolainen, D. Tvorogov, W. Zheng, C. A. Franco, A. Murtomaki, E. Aranda, N. Miura, S. Yla-Herttuala, M. Fruttiger, T. Makinen, A. Eichmann, J. W. Pollard, H. Gerhardt, and K. Alitalo (2011). “VEGFR-3 controls tip to stalk conversion at vessel fusion sites by reinforcing Notch signalling”. In: *Nat Cell Biol* 13.10, pp. 1202–13.
- Tamura, R., T. Tanaka, K. Miyake, K. Yoshida, and H. Sasaki (2017). “Bevacizumab for malignant gliomas: current indications, mechanisms of action and resistance, and markers of response”. In: *Brain Tumor Pathol* 34.2, pp. 62–77.
- Tamura, T., P. Thotakura, T. S. Tanaka, M. S. Ko, and K. Ozato (2005). “Identification of target genes and a unique cis element regulated by IRF-8 in developing macrophages”. In: *Blood* 106.6, pp. 1938–47.
- Tan, S., N. Sukumar, R. B. Abramovitch, T. Parish, and D. G. Russell (2013). “*Mycobacterium tuberculosis* responds to chloride and pH as synergistic cues to the immune status of its host cell”. In: *PLoS Pathog* 9.4, e1003282.
- Tan, Y., I. Zanoni, T. W. Cullen, A. L. Goodman, and J. C. Kagan (2015). “Mechanisms of Toll-like Receptor 4 Endocytosis Reveal a Common Immune-Evasion Strategy Used by Pathogenic and Commensal Bacteria”. In: *Immunity* 43.5, pp. 909–22.

Tang, J., G. Lin, W. Y. Langdon, L. Tao, and J. Zhang (2018). “Regulation of C-Type Lectin Receptor-Mediated Antifungal Immunity”. In: *Front Immunol* 9, p. 123.

Tang, Q., N. S. Abdelfattah, J. S. Blackburn, J. C. Moore, S. A. Martinez, F. E. Moore, R. Lobbardi, I. M. Tenente, M. S. Ignatius, J. N. Berman, R. S. Liwski, Y. Houvras, and D. M. Langenau (2014). “Optimized cell transplantation using adult *rag2* mutant zebrafish”. In: *Nat Methods* 11.8, pp. 821–4.

Tannahill, G. M., A. M. Curtis, J. Adamik, E. M. Palsson-McDermott, A. F. McGettrick, G. Goel, C. Frezza, N. J. Bernard, B. Kelly, N. H. Foley, L. Zheng, A. Gardet, Z. Tong, S. S. Jany, S. C. Corr, M. Haneklaus, B. E. Caffrey, K. Pierce, S. Walmley, F. C. Beasley, E. Cummins, V. Nizet, M. Whyte, C. T. Taylor, H. Lin, S. L. Masters, E. Gottlieb, V. P. Kelly, C. Clish, P. E. Auron, R. J. Xavier, and L. A. O’Neill (2013). “Succinate is an inflammatory signal that induces IL-1beta through HIF-1alpha”. In: *Nature* 496.7444, pp. 238–42.

Tanne, A., B. Ma, F. Boudou, L. Tailleux, H. Botella, E. Badell, F. Levillain, M. E. Taylor, K. Drickamer, J. Nigou, K. M. Dobos, G. Puzo, D. Vestweber, M. K. Wild, M. Marcinko, P. Sobieszczuk, L. Stewart, D. Lebus, B. Gicquel, and O. Neyrolles (2009). “A murine DC-SIGN homologue contributes to early host defense against *Mycobacterium tuberculosis*”. In: *J Exp Med* 206.10, pp. 2205–20.

Tavares, A. H., P. H. Burgel, and A. L. Bocca (2015). “Turning Up the Heat: Inflammasome Activation by Fungal Pathogens”. In: *PLoS Pathog* 11.7, e1004948.

Taylor, C. T. and C. C. Scholz (2022). “The effect of HIF on metabolism and immunity”. In: *Nat Rev Nephrol* 18.9, pp. 573–587.

Teixeira, L. K., B. P. Fonseca, A. Vieira-de Abreu, B. A. Barboza, B. K. Robbs, P. T. Bozza, and J. P. Viola (2005). “IFN-gamma production by CD8+ T cells depends on NFAT1 transcription factor and regulates Th differentiation”. In: *J Immunol* 175.9, pp. 5931–9.

Telenti, A., P. Imboden, F. Marchesi, D. Lowrie, S. Cole, M. J. Colston, L. Matter, K. Schopfer, and T. Bodmer (1993). “Detection of rifampicin-resistance mutations in *Mycobacterium tuberculosis*”. In: *Lancet* 341.8846, pp. 647–50.

Tenor, J. L., S. H. Oehlers, J. L. Yang, D. M. Tobin, and J. R. Perfect (2015). “Live Imaging of Host-Parasite Interactions in a Zebrafish Infection Model Reveals Cryptococcal Determinants of Virulence and Central Nervous System Invasion”. In: *mBio* 6.5, e01425–15.

- Teran, G., H. Li, S. B. Catrina, R. Liu, S. Brighenti, X. Zheng, J. Grunler, S. Nylen, B. Carow, and M. E. Rottenberg (2022). “High Glucose and Carbonyl Stress Impair HIF-1-Regulated Responses and the Control of *Mycobacterium tuberculosis* in Macrophages”. In: *mBio*, e0108622.
- Tesh, V. L. and A. D. O’Brien (1991). “The pathogenic mechanisms of Shiga toxin and the Shiga-like toxins”. In: *Mol Microbiol* 5.8, pp. 1817–22.
- Tewari, K. S., M. W. Sill, 3rd Long H. J., R. T. Penson, H. Huang, L. M. Ramondetta, L. M. Landrum, A. Oaknin, T. J. Reid, M. M. Leitao, H. E. Michael, and B. J. Monk (2014). “Improved survival with bevacizumab in advanced cervical cancer”. In: *N Engl J Med* 370.8, pp. 734–43.
- Thackaberry, E. A., D. M. Gabaldon, M. K. Walker, and S. M. Smith (2002). “Aryl hydrocarbon receptor null mice develop cardiac hypertrophy and increased hypoxia-inducible factor-1alpha in the absence of cardiac hypoxia”. In: *Cardiovasc Toxicol* 2.4, pp. 263–74.
- Thangarajah, H., D. Yao, E. I. Chang, Y. Shi, L. Jazayeri, I. N. Vial, R. D. Galiano, X. L. Du, R. Grogan, M. G. Galvez, M. Januszyk, M. Brownlee, and G. C. Gurtner (2009). “The molecular basis for impaired hypoxia-induced VEGF expression in diabetic tissues”. In: *Proc Natl Acad Sci U S A* 106.32, pp. 13505–10.
- The 1000 Genomes Project Consortium, A. Auton, L. D. Brooks, R. M. Durbin, E. P. Garrison, H. M. Kang, J. O. Korbel, J. L. Marchini, S. McCarthy, G. A. McVean, and G. R. Abecasis (2015). “A global reference for human genetic variation”. In: *Nature* 526.7571, pp. 68–74.
- Theret, M., R. Mounier, and F. Rossi (2019). “The origins and non-canonical functions of macrophages in development and regeneration”. In: *Development* 146.9.
- Thermes, V., C. Grabher, F. Ristoratore, F. Bourrat, A. Choulika, J. Wittbrodt, and J. S. Joly (2002). “I-SceI meganuclease mediates highly efficient transgenesis in fish”. In: *Mech Dev* 118.1-2, pp. 91–8.
- Thewes, S. (2014). “Calcineurin-Crz1 signaling in lower eukaryotes”. In: *Eukaryot Cell* 13.6, pp. 694–705.
- Thirunavukkarasu, M., B. Juhasz, L. Zhan, V. P. Menon, A. Tosaki, H. Otani, and N. Maulik (2007). “VEGFR1 (Flt-1+/-) gene knockout leads to the disruption

of VEGF-mediated signaling through the nitric oxide/heme oxygenase pathway in ischemic preconditioned myocardium". In: *Free Radic Biol Med* 42.10, pp. 1487–95.

Thisse, C. and L. I. Zon (2002). "Organogenesis–heart and blood formation from the zebrafish point of view". In: *Science* 295.5554, pp. 457–62.

Thoen, C. O., A. G. Karlson, and E. M. Himes (1981). "Mycobacterial infections in animals". In: *Rev Infect Dis* 3.5, pp. 960–72.

Thomas, H. R., S. M. Percival, B. K. Yoder, and J. M. Parant (2014). "High-throughput genome editing and phenotyping facilitated by high resolution melting curve analysis". In: *PLoS One* 9.12, e114632.

Thomas, J. P., C. O. Baughn, R. G. Wilkinson, and R. G. Shepherd (1961). "A new synthetic compound with antituberculous activity in mice: ethambutol (dextro-2,2'-(ethylenediimino)-di-l-butanol)". In: *Am Rev Respir Dis* 83, pp. 891–3.

Thompson, A. A., R. S. Dickinson, F. Murphy, J. P. Thomson, H. M. Marriott, A. Tavares, J. Willson, L. Williams, A. Lewis, A. Mirchandani, P. Dos Santos Coelho, C. Doherty, E. Ryan, E. Watts, N. M. Morton, S. Forbes, R. H. Stimson, A. G. Hameed, N. Arnold, J. A. Preston, A. Lawrie, V. Finisguerra, M. Mazzone, P. Sadiku, J. Goveia, F. Taverna, P. Carmeliet, S. J. Foster, E. R. Chilvers, A. S. Cowburn, D. H. Dockrell, R. S. Johnson, R. R. Meehan, M. K. Whyte, and S. R. Walmsley (2017). "Hypoxia determines survival outcomes of bacterial infection through HIF-1alpha dependent re-programming of leukocyte metabolism". In: *Sci Immunol* 2.8.

Thompson, A. A., P. M. Elks, H. M. Marriott, S. Eamsamarn, K. R. Higgins, A. Lewis, L. Williams, S. Parmar, G. Shaw, E. E. McGrath, F. Formenti, F. J. Van Eeden, V. L. Kinnula, C. W. Pugh, I. Sabroe, D. H. Dockrell, E. R. Chilvers, P. A. Robbins, M. J. Percy, M. C. Simon, R. S. Johnson, S. A. Renshaw, M. K. Whyte, and S. R. Walmsley (2014). "Hypoxia-inducible factor 2alpha regulates key neutrophil functions in humans, mice, and zebrafish". In: *Blood* 123.3, pp. 366–76.

Thuong, N. T., S. J. Dunstan, T. T. Chau, V. Thorsson, C. P. Simmons, N. T. Quyen, G. E. Thwaites, N. Thi Ngoc Lan, M. Hibberd, Y. Y. Teo, M. Seielstad, A. Aderem, J. J. Farrar, and T. R. Hawn (2008). "Identification of tuberculosis susceptibility genes with human macrophage gene expression profiles". In: *PLoS Pathog* 4.12, e1000229.

Thuong, N. T. T., D. Heemskerk, T. T. B. Tram, L. T. P. Thao, L. Ramakrishnan, V. T. N. Ha, N. D. Bang, T. T. H. Chau, N. H. Lan, M. Caws, S. J. Dunstan, N. V. V. Chau, M. Wolbers, N. T. H. Mai, and G. E. Thwaites (2017). “Leukotriene A4 Hydrolase Genotype and HIV Infection Influence Intracerebral Inflammation and Survival From Tuberculous Meningitis”. In: *J Infect Dis* 215.7, pp. 1020–1028.

Thurston, G. and C. Daly (2012). “The complex role of angiopoietin-2 in the angiopoietin-tie signaling pathway”. In: *Cold Spring Harb Perspect Med* 2.9, a006550.

Thwaites, G. E., R. van Toorn, and J. Schoeman (2013). “Tuberculous meningitis: more questions, still too few answers”. In: *Lancet Neurol* 12.10, pp. 999–1010.

Tidu, F., M. De Zuani, S. S. Jose, K. Bendickova, L. Kubala, F. Caruso, F. Cavalieri, G. Forte, and J. Fric (2021). “NFAT signaling in human mesenchymal stromal cells affects extracellular matrix remodeling and antifungal immune responses”. In: *iScience* 24.6, p. 102683.

– *A new study on the role of mesenchymal stromal cell NFAT activation in controlling antifungal immune responses and that NFAT regulates production of important extracellular matrix genes in these populations and mesenchymal stromal cell NFAT is able to regulate cytokine production by myeloid cells after exposure to the fungal ligand zymosan. This integrates aspects of the stromal environment with innate immune responses and adds additional layers of complexity to these responses. It also indicates the stromal-myeloid interactions may be important components for in vitro modeling of certain aspects of innate immunity.*

Tiemersma, E. W., M. J. van der Werf, M. W. Borgdorff, B. G. Williams, and N. J. Nagelkerke (2011). “Natural history of tuberculosis: duration and fatality of untreated pulmonary tuberculosis in HIV negative patients: a systematic review”. In: *PLoS One* 6.4, e17601.

Timmins, G. S. and V. Deretic (2006). “Mechanisms of action of isoniazid”. In: *Mol Microbiol* 62.5, pp. 1220–7.

Ting, L. M., A. C. Kim, A. Cattamanchi, and J. D. Ernst (1999). “*Mycobacterium tuberculosis* inhibits IFN-gamma transcriptional responses without inhibiting activation of STAT1”. In: *J Immunol* 163.7, pp. 3898–906.

Tisdale, M. J. (2002). “Cachexia in cancer patients”. In: *Nat Rev Cancer* 2.11, pp. 862–71.

- Tobin, D. M. (2015). "Host-Directed Therapies for Tuberculosis". In: *Cold Spring Harb Perspect Med* 5.10.
- Tobin, D. M. and L. Ramakrishnan (2008). "Comparative pathogenesis of *Mycobacterium marinum* and *Mycobacterium tuberculosis*". In: *Cell Microbiol* 10.5, pp. 1027–39.
- Tobin, D. M., F. J. Roca, S. F. Oh, R. McFarland, T. W. Vickery, J. P. Ray, D. C. Ko, Y. Zou, N. D. Bang, T. T. Chau, J. C. Vary, T. R. Hawn, S. J. Dunstan, J. J. Farrar, G. E. Thwaites, M. C. King, C. N. Serhan, and L. Ramakrishnan (2012). "Host genotype-specific therapies can optimize the inflammatory response to mycobacterial infections". In: *Cell* 148.3, pp. 434–46.
- Tobin, D. M., Jr. Vary J. C., J. P. Ray, G. S. Walsh, S. J. Dunstan, N. D. Bang, D. A. Hagge, S. Khadge, M. C. King, T. R. Hawn, C. B. Moens, and L. Ramakrishnan (2010). "The lta4h locus modulates susceptibility to mycobacterial infection in zebrafish and humans". In: *Cell* 140.5, pp. 717–30.
- Tocqueville, Alexis de (1835). *Democracy in America*. A Vintage book, K-5A–K-5B. New York, Vintage Books.
- Toledo-Pereyra, L. H. (2010). "Joseph Lister's surgical revolution". In: *J Invest Surg* 23.5, pp. 241–3.
- Tomita, M., M. I. Reinhold, J. D. Molkentin, and M. C. Naski (2002). "Calcineurin and NFAT4 induce chondrogenesis". In: *J Biol Chem* 277.44, pp. 42214–8.
- Torgerson, T. R., A. Genin, C. Chen, M. Zhang, B. Zhou, S. Anover-Sombke, M. B. Frank, I. Dozmorov, E. Ocheltree, P. Kulmala, M. Centola, H. D. Ochs, A. D. Wells, and R. Q. Cron (2009). "FOXP3 inhibits activation-induced NFAT2 expression in T cells thereby limiting effector cytokine expression". In: *J Immunol* 183.2, pp. 907–15.
- Torraca, V. and S. Mostowy (2018). "Zebrafish Infection: From Pathogenesis to Cell Biology". In: *Trends Cell Biol* 28.2, pp. 143–156.
- Torraca, V., C. Tulotta, B. E. Snaar-Jagalska, and A. H. Meijer (2017).** "The chemokine receptor CXCR4 promotes granuloma formation by sustaining a mycobacteria-induced angiogenesis programme". In: *Sci Rep* 7, p. 45061.

– A first non-VEGFA model for granuloma angiogenesis where CXCR4 signaling is necessary for both the establishment of the granuloma itself and the angiogenic response. Integrating these phenomena with VEGFA signaling would be a nice addition to our understanding of the processes necessary and sufficient to mediate granuloma formation and the contributions of the angiogenic response to mycobacterial growth.

Torrelles, J. B. and L. S. Schlesinger (2010). “Diversity in *Mycobacterium tuberculosis* mannosylated cell wall determinants impacts adaptation to the host”. In: *Tuberculosis (Edinb)* 90.2, pp. 84–93.

Tothova, J., B. Blaauw, G. Pallafacchina, R. Rudolf, C. Argentini, C. Reggiani, and S. Schiaffino (2006). “NFATc1 nucleocytoplasmic shuttling is controlled by nerve activity in skeletal muscle”. In: *J Cell Sci* 119.Pt 8, pp. 1604–11.

Tracey, K. J., S. F. Lowry, and A. Cerami (1988). “Cachectin: a hormone that triggers acute shock and chronic cachexia”. In: *J Infect Dis* 157.3, pp. 413–20.

Trauer, J. M., A. Kawai, A. K. Coussens, M. Datta, B. M. Williams, E. S. McBryde, and R. Ragonnet (2021). “Timing of *Mycobacterium tuberculosis* exposure explains variation in BCG effectiveness: a systematic review and meta-analysis”. In: *Thorax* 76.11, pp. 1131–1141.

Trede, N. S., D. M. Langenau, D. Traver, A. T. Look, and L. I. Zon (2004). “The use of zebrafish to understand immunity”. In: *Immunity* 20.4, pp. 367–79.

Triantafilou, M. and K. Triantafilou (2002). “Lipopolysaccharide recognition: CD14, TLRs and the LPS-activation cluster”. In: *Trends Immunol* 23.6, pp. 301–4.

Trowe, T., S. Klostermann, H. Baier, M. Granato, A. D. Crawford, B. Grunewald, H. Hoffmann, R. O. Karlstrom, S. U. Meyer, B. Muller, S. Richter, C. Nusslein-Volhard, and F. Bonhoeffer (1996). “Mutations disrupting the ordering and topographic mapping of axons in the retinotectal projection of the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 439–50.

Tsai, M. C., S. Chakravarty, G. Zhu, J. Xu, K. Tanaka, C. Koch, J. Tufariello, J. Flynn, and J. Chan (2006). “Characterization of the tuberculous granuloma in murine and human lungs: cellular composition and relative tissue oxygen tension”. In: *Cell Microbiol* 8.2, pp. 218–32.

– A nice comparative study on the hypoxia present in human and murine “granulomas” and a nice model to follow in understanding these processes in the zebrafish.

Tsatsaronis, J. A., S. Franch-Arroyo, U. Resch, and E. Charpentier (2018). “Extracellular Vesicle RNA: A Universal Mediator of Microbial Communication?” In: *Trends Microbiol* 26.5, pp. 401–410.

Tsou, R., C. Fathke, L. Wilson, K. Wallace, N. Gibran, and F. Isik (2002). “Retroviral delivery of dominant-negative vascular endothelial growth factor receptor type 2 to murine wounds inhibits wound angiogenesis”. In: *Wound Repair Regen* 10.4, pp. 222–9.

Tsuchiya, S., M. Yamabe, Y. Yamaguchi, Y. Kobayashi, T. Konno, and K. Tada (1980). “Establishment and characterization of a human acute monocytic leukemia cell line (THP-1)”. In: *Int J Cancer* 26.2, pp. 171–6.

Tullius, M. V., C. A. Harmston, C. P. Owens, N. Chim, R. P. Morse, L. M. McMath, A. Iniguez, J. M. Kimmey, M. R. Sawaya, J. P. Whitelegge, M. A. Horwitz, and C. W. Goulding (2011). “Discovery and characterization of a unique mycobacterial heme acquisition system”. In: *Proc Natl Acad Sci U S A* 108.12, pp. 5051–6.

Turgut, B., M. Guler, N. Akpolat, T. Demir, and U. Celiker (2011). “The impact of tacrolimus on vascular endothelial growth factor in experimental corneal neovascularization”. In: *Curr Eye Res* 36.1, pp. 34–40.

Turner, O. C., R. J. Basaraba, and I. M. Orme (2003). “Immunopathogenesis of pulmonary granulomas in the guinea pig after infection with *Mycobacterium tuberculosis*”. In: *Infect Immun* 71.2, pp. 864–71.

Turner, Oliver C., Randall J. Basaraba, Anthony A. Frank, and Ian M. Orme (2003). “Granuloma Formation in Mouse and Guinea Pig Models of Experimental Tuberculosis”. In: John Wiley & Sons, Ltd. Chap. 3, pp. 65–84.

Turner, R. D., C. Chiu, G. J. Churchyard, H. Esmail, D. M. Lewinsohn, N. R. Gandhi, and K. P. Fnelly (2017). “Tuberculosis Infectiousness and Host Susceptibility”. In: *J Infect Dis* 216.suppl_6, S636–S643.

Tweedie, S., B. Braschi, K. Gray, T. E. M. Jones, R. L. Seal, B. Yates, and E. A. Bruford (2021). “Genenames.org: the HGNC and VGNC resources in 2021”. In: *Nucleic Acids Res* 49.D1, pp. D939–D946.

- Tyrkalska, S. D., A. B. Perez-Oliva, L. Rodriguez-Ruiz, F. J. Martinez-Morcillo, F. Alcaraz-Perez, F. J. Martinez-Navarro, C. Lachaud, N. Ahmed, T. Schroeder, I. Pardo-Sanchez, S. Candel, A. Lopez-Munoz, A. Choudhuri, M. P. Rossmann, L. I. Zon, M. L. Cayuela, D. Garcia-Moreno, and V. Mulero (2019). “Inflammasome Regulates Hematopoiesis through Cleavage of the Master Erythroid Transcription Factor GATA1”. In: *Immunity* 51.1, 50–63 e5.
- Tzelepis, F., J. Blagih, N. Khan, J. Gillard, L. Mendonca, D. G. Roy, E. H. Ma, P. Joubert, R. G. Jones, and M. Divangahi (2018). “Mitochondrial cyclophilin D regulates T cell metabolic responses and disease tolerance to tuberculosis”. In: *Sci Immunol* 3.23.
- Ubil, E., J. Duan, I. C. Pillai, M. Rosa-Garrido, Y. Wu, F. Bargiacchi, Y. Lu, S. Stanbouly, J. Huang, M. Rojas, T. M. Vondriska, E. Stefani, and A. Deb (2014). “Mesenchymal-endothelial transition contributes to cardiac neovascularization”. In: *Nature* 514.7524, pp. 585–90.
- Ueda, Y., H. Kayama, S. G. Jeon, T. Kusu, Y. Isaka, H. Rakugi, M. Yamamoto, and K. Takeda (2010). “Commensal microbiota induce LPS hyporesponsiveness in colonic macrophages via the production of IL-10”. In: *Int Immunopharmacol* 22.12, pp. 953–62.
- Ufimtseva, E. G., N. I. Eremeeva, E. M. Petrunina, T. V. Umpeleva, S. I. Bayborodin, D. V. Vakhrusheva, and S. N. Skornyakov (2018). “*Mycobacterium tuberculosis* cording in alveolar macrophages of patients with pulmonary tuberculosis is likely associated with increased mycobacterial virulence”. In: *Tuberculosis (Edinb)* 112, pp. 1–10.
- Ulengin-Talkish, I. and M. S. Cyert (2022). “A cellular atlas of calcineurin signaling”. In: *Biochim Biophys Acta Mol Cell Res* 1870.1, p. 119366.
- Ulrich, J. D., M. S. Kim, P. R. Houlihan, L. P. Shutov, D. P. Mohapatra, S. Strack, and Y. M. Usachev (2012). “Distinct activation properties of the nuclear factor of activated T-cells (NFAT) isoforms NFATc3 and NFATc4 in neurons”. In: *J Biol Chem* 287.45, pp. 37594–609.
– *The first of the modern wave of studies investigating the variable kinetics of NFAT protein activation, this time in neurons comparing the activation of NFATC3 and NFATC4 and finding that NFATC3 is rapidly activated (consistent with later studies) and NFATC4 has a slower activation mechanism that steadily accumulates over time (similar to NFATC2). However, this resulted in NFATC3 being the only*

functionally relevant isoform in this context due to the extensive delay in NFATC4-dependent responses.

Ulrichs, T. and S. H. Kaufmann (2006). “New insights into the function of granulomas in human tuberculosis”. In: *J Pathol* 208.2, pp. 261–9.

Unanue, E. R. and P. M. Allen (1987). “The basis for the immunoregulatory role of macrophages and other accessory cells”. In: *Science* 236.4801, pp. 551–7.

Underhill, D. M., A. Ozinsky, K. D. Smith, and A. Aderem (1999). “Toll-like receptor-2 mediates mycobacteria-induced proinflammatory signaling in macrophages”. In: *Proc Natl Acad Sci U S A* 96.25, pp. 14459–63.

Upadhyay, S., E. Mittal, and J. A. Philips (2018). “Tuberculosis and the art of macrophage manipulation”. In: *Pathog Dis* 76.4.

Vaeth, M. and S. Feske (2018). “NFAT control of immune function: New Frontiers for an Abiding Trooper”. In: *F1000Res* 7, p. 260.

Valdes-Tresanco, M. S., A. Molina-Zapata, A. G. Pose, and E. Moreno (2022). “Structural Insights into the Design of Synthetic Nanobody Libraries”. In: *Molecules* 27.7.

van der Peet, P. L., C. Gunawan, S. Torigoe, S. Yamasaki, and S. J. Williams (2015). “Corynomycolic acid-containing glycolipids signal through the pattern recognition receptor Mincl”. In: *Chem Commun (Camb)* 51.24, pp. 5100–3.

van der Sar, A. M., B. J. Appelmelk, C. M. Vandenbroucke-Grauls, and W. Bitter (2004). “A star with stripes: zebrafish as an infection model”. In: *Trends Microbiol* 12.10, pp. 451–7.

van der Sar, A. M., R. J. Musters, F. J. van Eeden, B. J. Appelmelk, C. M. Vandenbroucke-Grauls, and W. Bitter (2003). “Zebrafish embryos as a model host for the real time analysis of *Salmonella typhimurium* infections”. In: *Cell Microbiol* 5.9, pp. 601–11.

van Eeden, F. J., M. Granato, U. Schach, M. Brand, M. Furutani-Seiki, P. Haffter, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, R. M. Warga, M. L. Allende, E. S. Weinberg, and C. Nüsslein-Volhard (1996a). “Mutations affecting somite formation and patterning in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 153–64.

- van Eeden, F. J., M. Granato, U. Schach, M. Brand, M. Furutani-Seiki, P. Haffter, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, R. M. Warga, and C. Nusslein-Volhard (1996b). “Genetic analysis of fin formation in the zebrafish, *Danio rerio*”. In: *Development* 123, pp. 255–62.
- van Kooyk, Y. and T. B. Geijtenbeek (2003). “DC-SIGN: escape mechanism for pathogens”. In: *Nat Rev Immunol* 3.9, pp. 697–709.
- van Lessen, M., S. Shibata-Germanos, A. van Impel, T. A. Hawkins, J. Rihel, and S. Schulte-Merker (2017). “Intracellular uptake of macromolecules by brain lymphatic endothelial cells during zebrafish embryonic development”. In: *Elife* 6.
- Van Meter, M. E. and E. S. Kim (2010). “Bevacizumab: current updates in treatment”. In: *Curr Opin Oncol* 22.6, pp. 586–91.
- van Puffelen, J. H., S. T. Keating, E. Oosterwijk, A. G. van der Heijden, M. G. Netea, L. A. B. Joosten, and S. H. Vermeulen (2020). “Trained immunity as a molecular mechanism for BCG immunotherapy in bladder cancer”. In: *Nat Rev Urol* 17.9, pp. 513–525.
- van Rossum, Guido (1995). *Python reference manual*.
- van Sandwijk, M. S., F. J. Bemelman, and I. J. Ten Berge (2013). “Immunosuppressive drugs after solid organ transplantation”. In: *Neth J Med* 71.6, pp. 281–9.
- van Toorn, R., S. D. Zaharie, J. A. Seddon, M. van der Kuip, A. Marceline van Furth, J. F. Schoeman, and R. S. Solomons (2021). “The use of thalidomide to treat children with tuberculosis meningitis: A review”. In: *Tuberculosis (Edinb)* 130, p. 102125.
- Vanaja, S. K., A. J. Russo, B. Behl, I. Banerjee, M. Yankova, S. D. Deshmukh, and V. A. K. Rathinam (2016). “Bacterial Outer Membrane Vesicles Mediate Cytosolic Localization of LPS and Caspase-11 Activation”. In: *Cell* 165.5, pp. 1106–1119.
- Vance, J., A. Santos, L. Sadofsky, A. Morice, and J. Cervantes (2019). “Effect of High Glucose on Human Alveolar Macrophage Phenotype and Phagocytosis of Mycobacteria”. In: *Lung* 197.1, pp. 89–94.

Vanegas, G., W. Quinones, C. Carrasco-Lopez, J. L. Concepcion, F. Albericio, and L. Avilan (2007). “Enolase as a plasminogen binding protein in *Leishmania mexicana*”. In: *Parasitol Res* 101.6, pp. 1511–6.

Vanhollebeke, B., O. A. Stone, N. Bostaille, C. Cho, Y. Zhou, E. Maquet, A. Gauquier, P. Cabochette, S. Fukuhara, N. Mochizuki, J. Nathans, and D. Y. Stainier (2015). “Tip cell-specific requirement for an atypical Gpr124- and Reck-dependent Wnt/beta-catenin pathway during brain angiogenesis”. In: *Elife* 4.

Varol, C., A. Mildner, and S. Jung (2015). “Macrophages: development and tissue specialization”. In: *Annu Rev Immunol* 33, pp. 643–75.

Vashishta, A., A. Habas, P. Pruunsild, J. J. Zheng, T. Timmusk, and M. Hetman (2009). “Nuclear factor of activated T-cells isoform c4 (NFATc4/NFAT3) as a mediator of antiapoptotic transcription in NMDA receptor-stimulated cortical neurons”. In: *J Neurosci* 29.48, pp. 15331–40.

Vasudevan, S. O., A. J. Russo, P. Kumari, S. K. Vanaja, and V. A. Rathinam (2022). “A TLR4-independent critical role for CD14 in intracellular LPS sensing”. In: *Cell Rep* 39.5, p. 110755.

– *This work demonstrates the TLR4-independent roles for CD14 in mediating responses to LPS intracellularly, adding to the possible pathways by which this common bacterial ligand can be sensed and responded to and providing further evidence of the CD14-dependent responses to LPS being far more diverse than those dependent upon TLR4 sensu stricto.*

Veatch, A. V. and D. Kaushal (2018). “Opening Pandora’s Box: Mechanisms of *Mycobacterium tuberculosis* Resuscitation”. In: *Trends Microbiol* 26.2, pp. 145–157.

Vega, A., P. Chacon, J. Monteseirin, R. El Bekay, G. Alba, J. Martin-Nieto, and F. Sobrino (2007). “Expression of the transcription factor NFAT2 in human neutrophils: IgE-dependent, Ca²⁺- and calcineurin-mediated NFAT2 activation”. In: *J Cell Sci* 120.Pt 14, pp. 2328–37.

– *A very clear and comprehensive study on the role of NFATC1 in neutrophils and provided a nice intellectual foundation for any subsequent work on this topic.*

Veloso, F. J., M. Trombetta-Lima, V. Anschau, M. C. Sogayar, and R. G. Correa (2019). “NOD-like receptors: major players (and targets) in the interface between innate immunity and cancer”. In: *Biosci Rep* 39.4.

Venero Galanternik, M., D. Castranova, A. V. Gore, N. H. Blewett, H. M. Jung, A. N. Stratman, M. R. Kirby, J. Iben, M. F. Miller, K. Kawakami, R. J. Maraia, and B. M. Weinstein (2017). “A novel perivascular cell population in the zebrafish brain”. In: *Elife* 6.

Venkatasubramanian, S., D. Tripathi, T. Tucker, P. Paidipally, S. Cheekatla, E. Welch, A. Raghunath, A. Jeffers, A. R. Tsvinnereim, M. E. Schechter, B. B. Andrade, N. Mackman, S. Idell, and R. Vankayalapati (2016). “Tissue factor expression by myeloid cells contributes to protective immune response against *Mycobacterium tuberculosis* infection”. In: *Eur J Immunol* 46.2, pp. 464–79.

Verrall, A. J., M. G. Netea, B. Alisjahbana, P. C. Hill, and R. van Crevel (2014). “Early clearance of *Mycobacterium tuberculosis*: a new frontier in prevention”. In: *Immunology* 141.4, pp. 506–13.

Via, L. E., A. V. Tsytsovova, R. Rajsbaum, J. V. Falvo, and A. E. Goldfeld (2012). “The transcription factor NFATp plays a key role in susceptibility to TB in mice”. In: *PLoS One* 7.7, e41427.

– This is the first study of a specific NFAT isoform in regulating host immunity to mycobacteria and found that loss of NFATC2 resulted in increased bacterial burden and impaired adaptive immune responses. This is somewhat inhibited by it having been done in the mouse model, which exhibits extensive engagement between the mycobacteria and T cells which is often lacking in human and zebrafish granulomas, but provides a standard to compare against for the work I have completed here.

Vihma, H., M. Luhakooder, P. Pruunsild, and T. Timmusk (2016). “Regulation of different human NFAT isoforms by neuronal activity”. In: *J Neurochem* 137.3, pp. 394–408.

Vihma, H., P. Pruunsild, and T. Timmusk (2008). “Alternative splicing and expression of human and mouse NFAT genes”. In: *Genomics* 92.5, pp. 279–91.

Vilchez, C. and Jr. Jacobs W. R. (2007). “The mechanism of isoniazid killing: clarity through the scope of genetics”. In: *Annu Rev Microbiol* 61, pp. 35–50.

Vilchez, C., F. Wang, M. Arai, M. H. Hazbon, R. Colangeli, L. Kremer, T. R. Weisbrod, D. Alland, J. C. Sacchettini, and Jr. Jacobs W. R. (2006). “Transfer of a point mutation in *Mycobacterium tuberculosis inhA* resolves the target of isoniazid”. In: *Nat Med* 12.9, pp. 1027–9.

- Villagracia, A. R., H. L. Ong, F. M. Lagua, and G. Alea (2020). “Chemical reactivity and bioactivity properties of pyrazinamide analogs of acetylsalicylic acid and salicylic acid using conceptual density functional theory”. In: *Helijon* 6.6, e04239.
- Villanueva, M. T. (2016). “Angiogenesis: Going with the flow”. In: *Nat Rev Cancer* 16.12, p. 751.
- Villarreal, D. O., R. J. Siefert, and D. B. Weiner (2015). “Alarmin IL-33 elicits potent TB-specific cell-mediated responses”. In: *Hum Vaccin Immunother* 11.8, pp. 1954–60.
- Villefranc, J. A., S. Nicoli, K. Bentley, M. Jeltsch, G. Zarkada, J. C. Moore, H. Gerhardt, K. Alitalo, and N. D. Lawson (2013). “A truncation allele in vascular endothelial growth factor c reveals distinct modes of signaling during lymphatic and vascular development”. In: *Development* 140.7, pp. 1497–506.
- Viola, A., F. Munari, R. Sanchez-Rodriguez, T. Scolaro, and A. Castegna (2019). “The Metabolic Signature of Macrophage Responses”. In: *Front Immunol* 10, p. 1462.
- Viziteu, E., C. Grandmougin, H. Goldschmidt, A. Seckinger, D. Hose, B. Klein, and J. Moreaux (2016). “Chetomin, targeting HIF-1alpha/p300 complex, exhibits antitumour activity in multiple myeloma”. In: *Br J Cancer* 114.5, pp. 519–23.
- Vogeli, K. M., S. W. Jin, G. R. Martin, and D. Y. Stainier (2006). “A common progenitor for haematopoietic and endothelial lineages in the zebrafish gastrula”. In: *Nature* 443.7109, pp. 337–9.
- *This work provides conclusive evidence of the long-implied existence of the hemangioblast, a common precursor to both the endothelial and hematopoietic lineages.*
- Voldoire, E., F. Brunet, M. Naville, J. N. Volff, and D. Galiana (2017). “Expansion by whole genome duplication and evolution of the sox gene family in teleost fish”. In: *PLoS One* 12.7, e0180936.
- Volkman, H. E., H. Clay, D. Beery, J. C. Chang, D. R. Sherman, and L. Ramakrishnan (2004). “Tuberculous granuloma formation is enhanced by a *Mycobacterium* virulence determinant”. In: *PLoS Biol* 2.11, e367.
- *Given what we know now about the importance of the granuloma in promoting bacterial growth, these findings are obvious because of how foundational they are to the field at large. These findings on the importance of the region of difference 1*

(*RD1*) in inducing granuloma formation both explain the attenuation of *BCG* and conclusively demonstrate that mycobacteria induce granuloma formation for their own benefit.

Volkman, H. E., T. C. Pozos, J. Zheng, J. M. Davis, J. F. Rawls, and L. Ramakrishnan (2010). “Tuberculous granuloma induction via interaction of a bacterial secreted protein with host epithelium”. In: *Science* 327.5964, pp. 466–9.

– *Expression of MMP9 by granuloma-adjacent epithelial cells results in increased bacterial burden, demonstrating another role for the granuloma in promoting mycobacterial growth through enhanced recruitment of macrophages. Inhibition of MMP9, for the very first time, impaired both granuloma formation and bacterial growth, allowing for the functional differentiation of the granuloma and host bacterial restriction.*

Voskuil, M. I., I. L. Bartek, K. Visconti, and G. K. Schoolnik (2011). “The response of *Mycobacterium tuberculosis* to reactive oxygen and nitrogen species”. In: *Front Microbiol* 2, p. 105.

Vymazal, O., K. Bendickova, M. De Zuani, M. Vlkova, M. Hortová-Kohoutkova, and J. Fric (2021). “Immunosuppression Affects Neutrophil Functions: Does Calcineurin-NFAT Signaling Matter?” In: *Front Immunol* 12, p. 770515.

Wade, M. M. and Y. Zhang (2004). “Anaerobic incubation conditions enhance pyrazinamide activity against *Mycobacterium tuberculosis*”. In: *J Med Microbiol* 53.Pt 8, pp. 769–773.

Wagener, M., J. C. Hoving, H. Ndlovu, and M. J. Marakalala (2018). “Dectin-1-Syk-CARD9 Signaling Pathway in TB Immunity”. In: *Front Immunol* 9, p. 225.

Walczak-Drzewiecka, A., M. Ratajewski, W. Wagner, and J. Dastych (2008). “HIF-1alpha is up-regulated in activated mast cells by a process that involves calcineurin and NFAT”. In: *J Immunol* 181.3, pp. 1665–72.

– *One of the only published connections between NFAT and HIF-1α signaling, this establishes a model wherein NFAT activation results in increased transcription of HIF1A and resulting increases in VEGFA production. This argues that the induction of angiogenesis by NFAT may be an indirect interaction, although additional work is needed to dissect these phenomena to understand the layers of regulation possible between C-type lectin activation and VEGFA induction.*

Wall, Larry, Tom Christiansen, and Jon Orwant (2000). *Programming Perl*. O'Reilly Media, Inc.

Wallis, R. S. and R. Hafner (2015). “Advancing host-directed therapy for tuberculosis”. In: *Nat Rev Immunol* 15.4, pp. 255–63.

Walsh, N. M., M. Wuthrich, H. Wang, B. Klein, and C. M. Hull (2017). “Characterization of C-type lectins reveals an unexpectedly limited interaction between *Cryptococcus neoformans* spores and Dectin-1”. In: *PLoS One* 12.3, e0173866.

Walton, E. M., M. R. Cronan, R. W. Beerman, and D. M. Tobin (2015). “The Macrophage-Specific Promoter *mfap4* Allows Live, Long-Term Analysis of Macrophage Behavior during Mycobacterial Infection in Zebrafish”. In: *PLoS One* 10.10, e0138949.

Walton, E. M., M. R. Cronan, C. J. Cambier, A. Rossi, M. Marass, M. D. Foglia, W. J. Brewer, K. D. Poss, D. Y. R. Stainier, C. R. Bertozzi, and D. M. Tobin (2018). “Cyclopropane Modification of Trehalose Dimycolate Drives Granuloma Angiogenesis and Mycobacterial Growth through Vegf Signaling”. In: *Cell Host Microbe* 24.4, 514–525 e6.

– My work was born whole from this study to which I made minor contributions. This work identified an important modification on a critical mycobacterial glycolipid (cis-cyclopropanation of trehalose dimycolate) as being necessary and sufficient for the induction of angiogenesis during mycobacterial infection and connected a known maladaptive response to a specific mechanism utilized by pathogenic mycobacteria to enhance this phenomenon. This begged the question that my work sought to answer: how are host macrophages detecting and responding to TDM?

Wang, J., Y. Zhang, L. Liu, Z. Cui, R. Shi, J. Hou, Z. Liu, L. Yang, L. Wang, and Y. Li (2020a). “NFAT2 overexpression suppresses the malignancy of hepatocellular carcinoma through inducing Egr2 expression”. In: *BMC Cancer* 20.1, p. 966.

Wang, K., B. Long, J. Zhou, and P. F. Li (2010). “miR-9 and NFATc3 regulate myocardin in cardiac hypertrophy”. In: *J Biol Chem* 285.16, pp. 11903–12.

Wang, S., X. Kang, S. Cao, H. Cheng, D. Wang, and J. Geng (2012). “Calcineurin/NFATc1 pathway contributes to cell proliferation in hepatocellular carcinoma”. In: *Dig Dis Sci* 57.12, pp. 3184–8.

- Wang, S., J. Lu, Q. You, H. Huang, Y. Chen, and K. Liu (2016). “The mTOR/AP-1/VEGF signaling pathway regulates vascular endothelial cell growth”. In: *Oncotarget* 7.33, pp. 53269–53276.
- Wang, X., A. M. Bove, G. Simone, and B. Ma (2020b). “Molecular Bases of VEGFR-2-Mediated Physiological Function and Pathological Role”. In: *Front Cell Dev Biol* 8, p. 599281.
- Wang, Y., A. Y. Hsu, E. M. Walton, S. J. Park, R. Syahirah, T. Wang, W. Zhou, C. Ding, A. P. Lemke, G. Zhang, D. M. Tobin, and Q. Deng (2021). “A robust and flexible CRISPR/Cas9-based system for neutrophil-specific gene inactivation in zebrafish”. In: *J Cell Sci* 134.8.
- Warner, D. F. and V. Mizrahi (2007). “The survival kit of *Mycobacterium tuberculosis*”. In: *Nat Med* 13.3, pp. 282–4.
- Wassermann, R., M. F. Gulen, C. Sala, S. G. Perin, Y. Lou, J. Rybniker, J. L. Schmid-Burgk, T. Schmidt, V. Hornung, S. T. Cole, and A. Ablasser (2015). “*Mycobacterium tuberculosis* Differentially Activates cGAS- and Inflammasome-Dependent Intracellular Immune Responses through ESX-1”. In: *Cell Host Microbe* 17.6, pp. 799–810.
- Watanabe, S., M. Alexander, A. V. Misharin, and G. R. S. Budinger (2019). “The role of macrophages in the resolution of inflammation”. In: *J Clin Invest* 129.7, pp. 2619–2628.
- Watrus, S. J., M. L. Smith, C. P. Rodrigues, E. J. Hagedorn, J. W. Kim, B. Budnik, and L. I. Zon (2022). “Quality assurance of hematopoietic stem cells by macrophages determines stem cell clonality”. In: *Science* 377.6613, pp. 1413–1419.
- Weber, G. M., Y. Ju, and K. Borner (2020). “Considerations for Using the Vasculature as a Coordinate System to Map All the Cells in the Human Body”. In: *Front Cardiovasc Med* 7, p. 29.
- Wehrli, W. (1983). “Rifampin: mechanisms of action and resistance”. In: *Rev Infect Dis* 5 Suppl 3, S407–11.
- Weichand, B., R. Popp, S. Dziumbla, J. Mora, E. Strack, E. Elwakeel, A. C. Frank, K. Scholich, S. Pierre, S. N. Syed, C. Olesch, J. Ringleb, B. Oren, C. Doring, R. Savai, M. Jung, A. von Knethen, B. Levkau, I. Fleming, A. Weigert, and B.

Brune (2017). “S1PR1 on tumor-associated macrophages promotes lymphangiogenesis and metastasis via NLRP3/IL-1beta”. In: *J Exp Med* 214.9, pp. 2695–2713.

Weider, M., L. J. Starost, K. Groll, M. Kuspert, E. Sock, M. Wedel, F. Frob, C. Schmitt, T. Baroti, A. C. Hartwig, S. Hillgartner, S. Piefke, T. Fadler, M. Ehrlich, C. Ehlert, M. Stehling, S. Albrecht, A. Jabali, H. R. Scholer, J. Winkler, T. Kuhlmann, and M. Wegner (2018). “Nfat/calcineurin signaling promotes oligodendrocyte differentiation and myelination by transcription factor network tuning”. In: *Nat Commun* 9.1, p. 899.

Weigmann, B., H. A. Lehr, G. Yancopoulos, D. Valenzuela, A. Murphy, S. Stevens, J. Schmidt, P. R. Galle, S. Rose-John, and M. F. Neurath (2008). “The transcription factor NFATc2 controls IL-6-dependent T cell activation in experimental colitis”. In: *J Exp Med* 205.9, pp. 2099–110.

Wein, T. and R. Sorek (2022). “Bacterial origins of human cell-autonomous innate immune mechanisms”. In: *Nat Rev Immunol* 22.10, pp. 629–638.

Weinstein, B. M., A. F. Schier, S. Abdelilah, J. Malicki, L. Solnica-Krezel, D. L. Stemple, D. Y. Stainier, F. Zwartkruis, W. Driever, and M. C. Fishman (1996). “Hematopoietic mutations in the zebrafish”. In: *Development* 123, pp. 303–9.

Weis, S. M. and D. A. Cheresh (2011). “Tumor angiogenesis: molecular pathways and therapeutic targets”. In: *Nat Med* 17.11, pp. 1359–70.

Weiser, J. N., D. M. Ferreira, and J. C. Paton (2018). “*Streptococcus pneumoniae*: transmission, colonization and invasion”. In: *Nat Rev Microbiol* 16.6, pp. 355–367.

Welch, M. D. (2015). “Why should cell biologists study microbial pathogens?” In: *Mol Biol Cell* 26.24, pp. 4295–301.

Wells, C. A., J. A. Salvage-Jones, X. Li, K. Hitchens, S. Butcher, R. Z. Murray, A. G. Beckhouse, Y. L. Lo, S. Manzanero, C. Cobbold, K. Schroder, B. Ma, S. Orr, L. Stewart, D. Lebus, P. Sobieszczuk, D. A. Hume, J. Stow, H. Blanchard, and R. B. Ashman (2008). “The macrophage-inducible C-type lectin, micle, is an essential component of the innate immune response to *Candida albicans*”. In: *J Immunol* 180.11, pp. 7404–13.

Welsh, K. J., R. L. Hunter, and J. K. Actor (2013). “Trehalose 6,6'-dimycolate—a coat to regulate tuberculosis immunopathogenesis”. In: *Tuberculosis (Edinb)* 93 Suppl, S3–9.

Welti, J., S. Loges, S. Dimmeler, and P. Carmeliet (2013). “Recent molecular discoveries in angiogenesis and antiangiogenic therapies in cancer”. In: *J Clin Invest* 123.8, pp. 3190–200.

Wenes, M., M. Shang, M. Di Matteo, J. Goveia, R. Martin-Perez, J. Serneels, H. Prenen, B. Ghesquiere, P. Carmeliet, and M. Mazzone (2016). “Macrophage Metabolism Controls Tumor Blood Vessel Morphogenesis and Metastasis”. In: *Cell Metab* 24.5, pp. 701–715.

– During pathology, macrophages and endothelial cells engage in an arms race to sequester the limited supply of glucose within tissues and the result of this is that glycolytic macrophages rapidly “waste” most of the glucose, which results in endothelial starvation and vascular regression. This is one explanation for why inflamed tissues have delayed revascularization and developed a correspondence between tumor-associated macrophage metabolism and angiogenesis in a fascinating competition for resources in the tumor microenvironment.

Weng, W., H. Nagai, S. Hamidi, and G. Sheng (2020). “NPAS4L is involved in avian hemangioblast specification”. In: *Haematologica* 105.11, pp. 2647–2650.

Werninghaus, K., A. Babiak, O. Gross, C. Holscher, H. Dietrich, E. M. Agger, J. Mages, A. Mocsai, H. Schoenen, K. Finger, F. Nimmerjahn, G. D. Brown, C. Kirschning, A. Heit, P. Andersen, H. Wagner, J. Ruland, and R. Lang (2009). “Adjuvanticity of a synthetic cord factor analogue for subunit *Mycobacterium tuberculosis* vaccination requires FcRgamma-Syk-Card9-dependent innate immune activation”. In: *J Exp Med* 206.1, pp. 89–97.

Wessjohann, L. A., W. Brandt, and T. Thiemann (2003). “Biosynthesis and metabolism of cyclopropane rings in natural compounds”. In: *Chem Rev* 103.4, pp. 1625–48.

Westblade, L. F., J. Errington, and T. Dorr (2020). “Antibiotic tolerance”. In: *PLoS Pathog* 16.10, e1008892.

Wevers, B. A., T. M. Kaptein, E. M. Zijlstra-Willems, B. Theelen, T. Boekhout, T. B. Geijtenbeek, and S. I. Gringhuis (2014). “Fungal engagement of the C-type lectin mincle suppresses dectin-1-induced antifungal immunity”. In: *Cell Host Microbe* 15.4, pp. 494–505.

White, R., K. Rose, and L. Zon (2013). “Zebrafish cancer: the state of the art and the path forward”. In: *Nat Rev Cancer* 13.9, pp. 624–36.

Whitehouse, G., E. Gray, S. Mastoridis, E. Merritt, E. Kodela, J. H. M. Yang, R. Danger, M. Mairal, S. Christakoudi, J. J. Lozano, I. C. Macdougall, T. I. M. Tree, A. Sanchez-Fueyo, and M. Martinez-Llordella (2017). “IL-2 therapy restores regulatory T-cell dysfunction induced by calcineurin inhibitors”. In: *Proc Natl Acad Sci U S A* 114.27, pp. 7083–7088.

Whitfield, T. T., M. Granato, F. J. van Eeden, U. Schach, M. Brand, M. Furutani-Seiki, P. Haffter, M. Hammerschmidt, C. P. Heisenberg, Y. J. Jiang, D. A. Kane, R. N. Kelsh, M. C. Mullins, J. Odenthal, and C. Nusslein-Volhard (1996). “Mutations affecting development of the zebrafish inner ear and lateral line”. In: *Development* 123, pp. 241–54.

Whiting, G. C., J. T. Evans, S. Patel, and S. H. Gillespie (2002). “Purification of native alpha-enolase from *Streptococcus pneumoniae* that binds plasminogen and is immunogenic”. In: *J Med Microbiol* 51.10, pp. 837–843.

Wick, W., T. Gorlia, M. Bendszus, M. Taphoorn, F. Sahm, I. Harting, A. A. Brandes, W. Taal, J. Domont, A. Idbaih, M. Campone, P. M. Clement, R. Stupp, M. Fabbro, E. Le Rhun, F. Dubois, M. Weller, A. von Deimling, V. Golfinopoulos, J. C. Bromberg, M. Platten, M. Klein, and M. J. van den Bent (2017). “Lomustine and Bevacizumab in Progressive Glioblastoma”. In: *N Engl J Med* 377.20, pp. 1954–1963.

Wickham, H. (2009). *ggplot2*. Springer.

- (2016). *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York.
- (2022). *reshape: Flexibly Reshape Data*.

Wickham, H., W. Chang, L. Henry, T. L. Pedersen, K. Takahashi, C. Wilke, K. Woo, H. Yutani, and D. Dunnington (2022a). “ggplot2: Create Elegant Data Visualisations Using the Grammar of Graphics”. In.

Wickham, H., R. François, L. Henry, and K. Müller (2022b). “dplyr: A Grammar of Data Manipulation”. In.

Wickham, H. and D. Seidel (2022). “scales: Scale Functions for Visualization”. In.

Wierenga, E. A. and G. Messer (2000). “Regulation of interleukin 4 gene transcription: alterations in atopic disease?” In: *Am J Respir Crit Care Med* 162.3 Pt 2, S81–5.

Wierson, W. A., J. M. Welker, M. P. Almeida, C. M. Mann, D. A. Webster, M. E. Torrie, T. J. Weiss, S. Kambakam, M. K. Vollbrecht, M. Lan, K. C. McKeighan, J. Levey, Z. Ming, A. Wehmeier, C. S. Mikelson, J. A. Haltom, K. M. Kwan, C. B. Chien, D. Balciunas, S. C. Ekker, K. J. Clark, B. R. Webber, B. S. Moriarity, S. L. Solin, D. F. Carlson, D. L. Dobbs, M. McGrail, and J. Essner (2020). “Efficient targeted integration directed by short homology in zebrafish and mammalian cells”. In: *Elife* 9.

Wik, J. A. and B. S. Skalhegg (2022). “T Cell Metabolism in Infection”. In: *Front Immunol* 13, p. 840610.

Wilburn, K. M., R. K. Meade, E. M. Heckenberg, J. Dockterman, J. Coers, C. M. Sassetti, A. J. Olive, and C. M. Smith (2022). “Differential requirement for IRGM proteins during tuberculosis infection in mice”. In: *bioRxiv*.

Wild, R., A. Klems, M. Takamiya, Y. Hayashi, U. Strahle, K. Ando, N. Mochizuki, A. van Impel, S. Schulte-Merker, J. Krueger, L. Preau, and F. le Noble (2017). “Neuronal sFlt1 and Vegfaa determine venous sprouting and spinal cord vascularization”. In: *Nat Commun* 8, p. 13991.

Wilkins, B. J., Y. S. Dai, O. F. Bueno, S. A. Parsons, J. Xu, D. M. Plank, F. Jones, T. R. Kimball, and J. D. Molkentin (2004). “Calcineurin/NFAT coupling participates in pathological, but not physiological, cardiac hypertrophy”. In: *Circ Res* 94.1, pp. 110–8.

Wilkins, B. J., L. J. De Windt, O. F. Bueno, J. C. Braz, B. J. Glascock, T. F. Kimball, and J. D. Molkentin (2002). “Targeted disruption of NFATc3, but not NFATc4, reveals an intrinsic defect in calcineurin-mediated cardiac hypertrophic growth”. In: *Mol Cell Biol* 22.21, pp. 7603–13.

Wilkinson, R. J., U. Rohlwink, U. K. Misra, R. van Crevel, N. T. H. Mai, K. E. Dooley, M. Caws, A. Figaji, R. Savic, R. Solomons, G. E. Thwaites, and Consortium Tuberculous Meningitis International Research (2017). “Tuberculous meningitis”. In: *Nat Rev Neurol* 13.10, pp. 581–598.

Wilkinson, R. N. and F. J. van Eeden (2014). “The zebrafish as a model of vascular development and disease”. In: *Prog Mol Biol Transl Sci* 124, pp. 93–122.

Williams, N. C. and L. A. J. O'Neill (2018). "A Role for the Krebs Cycle Intermediate Citrate in Metabolic Reprogramming in Innate Immunity and Inflammation". In: *Front Immunol* 9, p. 141.

Williams, S. J. (2017). "Sensing Lipids with Mincle: Structure and Function". In: *Front Immunol* 8, p. 1662.

Willment, J. A. and G. D. Brown (2008). "C-type lectin receptors in antifungal immunity". In: *Trends Microbiol* 16.1, pp. 27–32.

Wilman, W., S. Wrobel, W. Bielska, P. Deszynski, P. Dudzic, I. Jaszczyzyn, J. Kaniewski, J. Mlokosiewicz, A. Rouyan, T. Satlawa, S. Kumar, V. Greiff, and K. Krawczyk (2022). "Machine-designed biotherapeutics: opportunities, feasibility and advantages of deep learning in computational antibody discovery". In: *Brief Bioinform* 23.4.

Wilson, C. A., S. K. High, B. M. McCluskey, A. Amores, Y. L. Yan, T. A. Titus, J. L. Anderson, P. Batzel, 3rd Carvan M. J., M. Schartl, and J. H. Postlethwait (2014). "Wild sex in zebrafish: loss of the natural sex determinant in domesticated strains". In: *Genetics* 198.3, pp. 1291–308.

Wilson, G. J., M. J. Marakalala, J. C. Hoving, A. van Laarhoven, R. A. Drummond, B. Kerscher, R. Keeton, E. van de Vosse, T. H. Ottenhoff, T. S. Plantinga, B. Alisjahbana, D. Govender, G. S. Besra, M. G. Netea, D. M. Reid, J. A. Willment, M. Jacobs, S. Yamasaki, R. van Crevel, and G. D. Brown (2015). "The C-type lectin receptor CLECSF8/CLEC4D is a key component of anti-mycobacterial immunity". In: *Cell Host Microbe* 17.2, pp. 252–9.

— A really nice complementary work to that in Behler, Maus, Bohling, Knippenberg, Kirchhof, Nagata, Jonigk, Izquierdo, Magel, Welte, Yamasaki, and Maus (2015), this study examined in greater detail the broader role for MCL in executing host defense against mycobacteria and found a critical role for this receptor in host protection. This suggests that MCL is a more critical component of the antimycobacterial defense program than is MINCLE despite the attention often given MINCLE as the “primary” TDM receptor. This study is evocative of the need for a MINCLE/MCL locus deletion mouse to really understand the nature of TDM detection during infection.

Wilson, J. L., H. K. Mayr, and T. Weichhart (2019). "Metabolic Programming of Macrophages: Implications in the Pathogenesis of Granulomatous Disease". In: *Front Immunol* 10, p. 2265.

Winslow, M. M., M. Pan, M. Starbuck, E. M. Gallo, L. Deng, G. Karsenty, and G. R. Crabtree (2006). “Calcineurin/NFAT signaling in osteoblasts regulates bone mass”. In: *Dev Cell* 10.6, pp. 771–82.

Wirnsberger, G., F. Zwolanek, T. Asaoka, I. Kozieradzki, L. Tortola, R. A. Wimmer, A. Kavirayani, F. Fresser, G. Baier, W. Y. Langdon, F. Ikeda, K. Kuchler, and J. M. Penninger (2016). “Inhibition of CBLB protects from lethal *Candida albicans* sepsis”. In: *Nat Med* 22.8, pp. 915–23.

Wiszniaak, S. and Q. Schwarz (2021). “Exploring the Intracrine Functions of VEGF-A”. In: *Biomolecules* 11.1.

Wolf, A. J. and D. M. Underhill (2018). “Peptidoglycan recognition by the innate immune system”. In: *Nat Rev Immunol* 18.4, pp. 243–254.

Wong, B. W., X. Wang, A. Zecchin, B. Thienpont, I. Cornelissen, J. Kalucka, M. Garcia-Caballero, R. Missiaen, H. Huang, U. Bruning, S. Blacher, S. Vinckier, J. Goveia, M. Knobloch, H. Zhao, C. Dierkes, C. Shi, R. Hagerling, V. Moral-Darde, S. Wyns, M. Lippens, S. Jessberger, S. M. Fendt, A. Luttun, A. Noel, F. Kiefer, B. Ghesquiere, L. Moons, L. Schoonjans, M. Dewerchin, G. Eelen, D. Lambrechts, and P. Carmeliet (2017). “The role of fatty acid beta-oxidation in lymphangiogenesis”. In: *Nature* 542.7639, pp. 49–54.

Wong, K. W. (2017). “The Role of ESX-1 in *Mycobacterium tuberculosis* Pathogenesis”. In: *Microbiol Spectr* 5.3.

Woodruff, P. G., L. L. Koth, Y. H. Yang, M. W. Rodriguez, S. Favoreto, G. M. Dolganov, A. C. Paquet, and D. J. Erle (2005). “A distinctive alveolar macrophage activation state induced by cigarette smoking”. In: *Am J Respir Crit Care Med* 172.11, pp. 1383–92.

Woods, P. S., L. M. Kimmig, K. A. Sun, A. Y. Meliton, O. R. Shamaa, Y. Tian, R. Cetin-Atalay, W. W. Sharp, R. B. Hamanaka, and G. M. Mutlu (2022). “HIF-1alpha induces glycolytic reprogramming in tissue-resident alveolar macrophages to promote cell survival during acute lung injury”. In: *Elife* 11.

World Health Organization (2021). *Global Tuberculosis Report 2022*. Tech. rep. World Health Organization.

- Worrell, J. C. and M. K. L. MacLeod (2021). “Stromal-immune cell crosstalk fundamentally alters the lung microenvironment following tissue insult”. In: *Immunology* 163.3, pp. 239–249.
- Wright, S. D., R. A. Ramos, P. S. Tobias, R. J. Ulevitch, and J. C. Mathison (1990). “CD14, a receptor for complexes of lipopolysaccharide (LPS) and LPS binding protein”. In: *Science* 249.4975, pp. 1431–3.
- Wu, B., H. S. Baldwin, and B. Zhou (2013). “Nfatc1 directs the endocardial progenitor cells to make heart valve primordium”. In: *Trends Cardiovasc Med* 23.8, pp. 294–300.
- Wu, B., Z. Zhang, W. Lui, X. Chen, Y. Wang, A. A. Chamberlain, R. A. Moreno-Rodriguez, R. R. Markwald, B. P. O’Rourke, D. J. Sharp, D. Zheng, J. Lenz, H. S. Baldwin, C. P. Chang, and B. Zhou (2012). “Endocardial cells form the coronary arteries by angiogenesis through myocardial-endocardial VEGF signaling”. In: *Cell* 151.5, pp. 1083–96.
- Wu, L., D. Zhang, L. Zhou, Y. Pei, Y. Zhuang, W. Cui, and J. Chen (2019). “FUN14 domain-containing 1 promotes breast cancer proliferation and migration by activating calcium-NFATC1-BMI1 axis”. In: *EBioMedicine* 41, pp. 384–394.
- Wu, R., F. Chen, N. Wang, D. Tang, and R. Kang (2020). “ACOD1 in immunometabolism and disease”. In: *Cell Mol Immunol* 17.8, pp. 822–833.
- Wu, R., X. Wang, Y. Shao, Y. Jiang, Y. Zhou, and C. Lu (2021). “NFATc4 mediates ethanol-triggered hepatocyte senescence”. In: *Toxicol Lett* 350, pp. 10–21.
- Wu, R. S., II Lam, H. Clay, D. N. Duong, R. C. Deo, and S. R. Coughlin (2018). “A Rapid Method for Directed Gene Knockout for Screening in G0 Zebrafish”. In: *Dev Cell* 46.1, 112–125 e4.
- Wu, Y., L. Chen, P. G. Scott, and E. E. Tredget (2007). “Mesenchymal stem cells enhance wound healing through differentiation and angiogenesis”. In: *Stem Cells* 25.10, pp. 2648–59.
- Wynn, T. A., A. Chawla, and J. W. Pollard (2013). “Macrophage biology in development, homeostasis and disease”. In: *Nature* 496.7446, pp. 445–55.

Xanthoudakis, S., J. P. Viola, K. T. Shaw, C. Luo, J. D. Wallace, P. T. Bozza, D. C. Luk, T. Curran, and A. Rao (1996). “An enhanced immune response in mice lacking the transcription factor NFAT1”. In: *Science* 272.5263, pp. 892–5.

– The relative (anti-)inflammatory character of the different NFAT isoforms is an intriguing question that bridges structural biology with differential expression and ambiguous biochemical features of the individual isoforms, but this pioneering work found that loss of NFATC2 results in enhanced inflammatory responses, suggesting that NFATC2 either inhibits inflammation or induces anti-inflammatory responses, making it a ”type II” transcription factor.

Xiao, Y., M. Qureischi, L. Dietz, M. Vaeth, S. D. Vallabhapurapu, S. Klein-Hessling, M. Klein, C. Liang, A. Konig, E. Serfling, A. Mottok, T. Bopp, A. Rosenwald, M. Buttmann, I. Berberich, A. Beilhack, and F. Berberich-Siebelt (2021). “Lack of NFATc1 SUMOylation prevents autoimmunity and alloreactivity”. In: *J Exp Med* 218.1.

Xiao, Z. J., J. Liu, S. Q. Wang, Y. Zhu, X. Y. Gao, V. P. Tin, J. Qin, J. W. Wang, and M. P. Wong (2017). “NFATc2 enhances tumor-initiating phenotypes through the NFATc2/SOX2/ALDH axis in lung adenocarcinoma”. In: *Elife* 6.

Xie, T. X., Z. Xia, N. Zhang, W. Gong, and S. Huang (2010). “Constitutive NF-kappaB activity regulates the expression of VEGF and IL-8 and tumor angiogenesis of human glioblastoma”. In: *Oncol Rep* 23.3, pp. 725–32.

Xiong, J. W. (2008). “Molecular and developmental biology of the hemangioblast”. In: *Dev Dyn* 237.5, pp. 1218–31.

Xu, G., H. Liu, X. Jia, X. Wang, and P. Xu (2021a). “Mechanisms and detection methods of *Mycobacterium tuberculosis* rifampicin resistance: The phenomenon of drug resistance is complex”. In: *Tuberculosis (Edinb)* 128, p. 102083.

Xu, H., J. Zhu, S. Smith, J. Foldi, B. Zhao, A. Y. Chung, H. Outtz, J. Kitajewski, C. Shi, S. Weber, P. Saftig, Y. Li, K. Ozato, C. P. Blobel, L. B. Ivashkiv, and X. Hu (2012). “Notch-RBP-J signaling regulates the transcription factor IRF8 to promote inflammatory macrophage polarization”. In: *Nat Immunol* 13.7, pp. 642–50.

Xu, J. and Y. Zhang (2010). “How significant is a protein structure similarity with TM-score = 0.5?” In: *Bioinformatics* 26.7, pp. 889–95.

- Xu, L., D. G. Duda, E. di Tomaso, M. Ancukiewicz, D. C. Chung, G. Y. Lauwers, R. Samuel, P. Shellito, B. G. Czito, P. C. Lin, M. Poleski, R. Bentley, J. W. Clark, C. G. Willett, and R. K. Jain (2009a). “Direct evidence that bevacizumab, an anti-VEGF antibody, up-regulates SDF1alpha, CXCR4, CXCL6, and neuropilin 1 in tumors from patients with rectal cancer”. In: *Cancer Res* 69.20, pp. 7905–10.
- Xu, S., J. Huo, M. Gunawan, I. H. Su, and K. P. Lam (2009b). “Activated dectin-1 localizes to lipid raft microdomains for signaling and activation of phagocytosis and cytokine production in dendritic cells”. In: *J Biol Chem* 284.33, pp. 22005–22011.
- Xu, S., J. Huo, K. G. Lee, T. Kurosaki, and K. P. Lam (2009c). “Phospholipase Cgamma2 is critical for Dectin-1-mediated Ca²⁺ flux and cytokine production in dendritic cells”. In: *J Biol Chem* 284.11, pp. 7038–46.
- Xu, S., P. Shu, S. Zou, X. Shen, Y. Qu, Y. Zhang, K. Sun, and J. Zhang (2018a). “NFATc1 is a tumor suppressor in hepatocellular carcinoma and induces tumor cell apoptosis by activating the FasL-mediated extrinsic signaling pathway”. In: *Cancer Med* 7.9, pp. 4701–4717.
- Xu, X., N. Li, Y. Wang, J. Yu, and J. Mi (2021b). “Calcium channel TRPV6 promotes breast cancer metastasis by NFATC2IP”. In: *Cancer Lett* 519, pp. 150–160.
- Xu, X., J. Xu, J. Wu, Y. Hu, Y. Han, Y. Gu, K. Zhao, Q. Zhang, X. Liu, J. Liu, B. Liu, and X. Cao (2018b). “Phosphorylation-Mediated IFN-gammaR2 Membrane Translocation Is Required to Activate Macrophage Innate Response”. In: *Cell* 175.5, 1336–1351 e17.
- Yadav, M. and J. S. Schorey (2006). “The beta-glucan receptor dectin-1 functions together with TLR2 to mediate macrophage activation by mycobacteria”. In: *Blood* 108.9, pp. 3168–75.
- Yahia-Cherbal, H., M. Rybczynska, D. Lovecchio, T. Stephen, C. Lescale, K. Placek, J. Larghero, L. Rogge, and E. Bianchi (2019). “NFAT primes the human RORC locus for RORgammat expression in CD4(+) T cells”. In: *Nat Commun* 10.1, p. 4698.
- Yamamoto, M., S. Sato, H. Hemmi, K. Hoshino, T. Kaisho, H. Sanjo, O. Takeuchi, M. Sugiyama, M. Okabe, K. Takeda, and S. Akira (2003). “Role of adaptor TRIF in the MyD88-independent toll-like receptor signaling pathway”. In: *Science* 301.5633, pp. 640–3.

Yamasaki, S. (2013). "Signaling while eating: MCL is coupled with Mincl". In: *Eur J Immunol* 43.12, pp. 3156–8.

Yamasaki, S., E. Ishikawa, M. Sakuma, H. Hara, K. Ogata, and T. Saito (2008). "Mincl is an ITAM-coupled activating receptor that senses damaged cells". In: *Nat Immunol* 9.10, pp. 1179–88.

– *Ten years after its discovery, the ligand for MINCLE remained unknown, but this seminal paper identified key dimensions of the signaling cascade downstream of MINCLE and identified its first ligand: SAP130, a nuclear protein indicative of host cell damage.*

Yamasaki, S., M. Matsumoto, O. Takeuchi, T. Matsuzawa, E. Ishikawa, M. Sakuma, H. Tateno, J. Uno, J. Hirabayashi, Y. Mikami, K. Takeda, S. Akira, and T. Saito (2009). "C-type lectin Mincl is an activating receptor for pathogenic fungus, *Malassezia*". In: *Proc Natl Acad Sci U S A* 106.6, pp. 1897–902.

Yamazaki, T. and Y. S. Mukouyama (2018). "Tissue Specific Origin, Development, and Pathological Perspectives of Pericytes". In: *Front Cardiovasc Med* 5, p. 78.

Yan, C., D. C. Brunson, Q. Tang, D. Do, N. A. Iftimia, J. C. Moore, M. N. Hayes, A. M. Welker, E. G. Garcia, T. D. Dubash, X. Hong, B. J. Drapkin, D. T. Myers, S. Phat, A. Volorio, D. L. Marvin, M. Ligorio, L. Dershowitz, K. M. McCarthy, M. N. Karabacak, J. A. Fletcher, D. C. Sgroi, J. A. Iafrate, S. Maheswaran, N. J. Dyson, D. A. Haber, J. F. Rawls, and D. M. Langenau (2019). "Visualizing Engrafted Human Cancer and Therapy Responses in Immunodeficient Zebrafish". In: *Cell* 177.7, 1903–1914 e14.

Yan, C., Q. Yang, S. Zhang, D. G. Millar, E. J. Alpert, D. Do, A. Veloso, D. C. Brunson, B. J. Drapkin, M. Stanzione, I. Scarfo, J. C. Moore, S. Iyer, Q. Qin, Y. Wei, K. M. McCarthy, J. F. Rawls, N. J. Dyson, M. Cobbold, M. V. Maus, and D. M. Langenau (2021). "Single-cell imaging of T cell immunotherapy responses *in vivo*". In: *J Exp Med* 218.10.

Yan, J. and T. Horng (2020). "Lipid Metabolism in Regulation of Macrophage Functions". In: *Trends Cell Biol* 30.12, pp. 979–989.

Yanagi, S., R. Inatome, J. Ding, H. Kitaguchi, V. L. Tybulewicz, and H. Yamamura (2001). "Syk expression in endothelial cells and their morphologic defects in embryonic Syk-deficient mice". In: *Blood* 98.9, pp. 2869–71.

Yang, B., J. B. Treweek, R. P. Kulkarni, B. E. Deverman, C. K. Chen, E. Lubeck, S. Shah, L. Cai, and V. Gradinaru (2014a). “Single-cell phenotyping within transparent intact tissue through whole-body clearing”. In: *Cell* 158.4, pp. 945–958.

Yang, C. T., C. J. Cambier, J. M. Davis, C. J. Hall, P. S. Crosier, and L. Ramakrishnan (2012). “Neutrophils exert protection in the early tuberculous granuloma by oxidative killing of mycobacteria phagocytosed from infected macrophages”. In: *Cell Host Microbe* 12.3, pp. 301–12.

– *The function of neutrophils during tuberculosis infection remains controversial with this publication finding a protective role for neutrophils during the early phases of the immune response. Others have found deleterious roles for neutrophils at other stages, but the overall role and any prospects for modulating neutrophil behavior to improve disease outcomes remains unclear.*

Yang, J., X. Wang, G. Fuh, L. Yu, E. Wakshull, M. Khosraviani, E. S. Day, B. De meule, J. Liu, S. J. Shire, N. Ferrara, and S. Yadav (2014b). “Comparison of binding characteristics and *in vitro* activities of three inhibitors of vascular endothelial growth factor A”. In: *Mol Pharm* 11.10, pp. 3421–30.

Yang, J. D., D. Mott, R. Sutiwisesak, Y. J. Lu, F. Raso, B. Stowell, G. H. Babunovic, J. Lee, S. M. Carpenter, S. S. Way, S. M. Fortune, and S. M. Behar (2018). “*Mycobacterium tuberculosis*-specific CD4+ and CD8+ T cells differ in their capacity to recognize infected macrophages”. In: *PLoS Pathog* 14.5, e1007060.

Yang, M., J. H. Shao, Y. J. Miao, W. Cui, Y. F. Qi, J. H. Han, X. Lin, and J. Du (2014c). “Tumor cell-activated CARD9 signaling contributes to metastasis-associated macrophage polarization”. In: *Cell Death Differ* 21.8, pp. 1290–302.

Yang, Y., M. R. Chung, S. Zhou, X. Gong, H. Xu, Y. Hong, A. Jin, X. Huang, W. Zou, Q. Dai, and L. Jiang (2019). “STAT3 controls osteoclast differentiation and bone homeostasis by regulating NFATc1 transcription”. In: *J Biol Chem* 294.42, pp. 15395–15407.

Yang, Y., K. Kulka, R. C. Montelaro, T. A. Reinhart, J. Sissons, A. Aderem, and A. K. Ojha (2014d). “A hydrolase of trehalose dimycolate induces nutrient influx and stress sensitivity to balance intracellular growth of *Mycobacterium tuberculosis*”. In: *Cell Host Microbe* 15.2, pp. 153–63.

Yang, Y., P. Xie, Y. Li, Y. Bi, and D. B. Prusky (2022). “Updating Insights into the Regulatory Mechanisms of Calcineurin-Activated Transcription Factor Crz1 in Pathogenic Fungi”. In: *J Fungi (Basel)* 8.10.

Yaniv, K., S. Isogai, D. Castranova, L. Dye, J. Hitomi, and B. M. Weinstein (2006). “Live imaging of lymphatic development in the zebrafish”. In: *Nat Med* 12.6, pp. 711–6.

Yarilina, A., K. Xu, J. Chen, and L. B. Ivashkiv (2011). “TNF activates calcium-nuclear factor of activated T cells (NFAT)c1 signaling pathways in human macrophages”. In: *Proc Natl Acad Sci U S A* 108.4, pp. 1573–8.

Yeager, R. L., W. G. Munroe, and F. I. Dessau (1952). “Pyrazinamide (aldinamide) in the treatment of pulmonary tuberculosis”. In: *Am Rev Tuberc* 65.5, pp. 523–46.

Yin, L., L. A. Maddison, M. Li, N. Kara, M. C. LaFave, G. K. Varshney, S. M. Burgess, J. G. Patton, and W. Chen (2015). “Multiplex Conditional Mutagenesis Using Transgenic Expression of Cas9 and sgRNAs”. In: *Genetics* 200.2, pp. 431–41.

Yissachar, N., T. Sharar Fischler, A. A. Cohen, S. Reich-Zeliger, D. Russ, E. Shifrut, Z. Porat, and N. Friedman (2013). “Dynamic response diversity of NFAT isoforms in individual living cells”. In: *Mol Cell* 49.2, pp. 322–30.

Yoder, J. A., R. T. Litman, M. G. Mueller, S. Desai, K. P. Dobrinski, J. S. Montgomery, M. P. Buzzeo, T. Ota, C. T. Amemiya, N. S. Trede, S. Wei, J. Y. Djeu, S. Humphray, K. Jekosch, J. A. Hernandez Prada, D. A. Ostrov, and G. W. Litman (2004). “Resolution of the novel immune-type receptor gene cluster in zebrafish”. In: *Proc Natl Acad Sci U S A* 101.44, pp. 15706–11.

Yokobori, N., C. A. Sabio y Garcia, L. Geffner, P. Schierloh, B. Lopez, V. Ritacco, L. Barrera, S. de la Barrera, and M. del Carmen Saisiain (2012). “Differential induction of macrophage cell death by antigens of a clustered and a non-clustered multidrug-resistant *Mycobacterium tuberculosis* strain from Haarlem family”. In: *FEMS Immunol Med Microbiol* 66.3, pp. 363–71.

Yona, S., K. W. Kim, Y. Wolf, A. Mildner, D. Varol, M. Breker, D. Strauss-Ayali, S. Viukov, M. Guilliams, A. Misharin, D. A. Hume, H. Perlman, B. Malissen, E. Zelzer, and S. Jung (2013). “Fate mapping reveals origins and dynamics of

monocytes and tissue macrophages under homeostasis". In: *Immunity* 38.1, pp. 79–91.

– *A very nice examination of the developmental origins of tissue resident macrophages that identified the fetal origin for many of these long-lived macrophages with occasional contributions from circulating monocytes, explaining one dimension of macrophage heterogeneity.*

Yonucu, S., D. Yiotalmaz, C. Phipps, M. B. Unlu, and M. Kohandel (2017). "Quantifying the effects of antiangiogenic and chemotherapy drug combinations on drug delivery and treatment efficacy". In: *PLoS Comput Biol* 13.9, e1005724.

Yoshitomi, Y., T. Ikeda, H. Saito-Takatsuji, and H. Yonekura (2021). "Emerging Role of AP-1 Transcription Factor JunB in Angiogenesis and Vascular Development". In: *Int J Mol Sci* 22.6.

Yu, B. X., J. N. Yuan, F. R. Zhang, Y. Y. Liu, T. T. Zhang, K. Li, X. F. Lv, J. G. Zhou, L. Y. Huang, J. Y. Shang, and S. J. Liang (2018). "Inhibition of Orai1-mediated Ca(2+) entry limits endothelial cell inflammation by suppressing calcineurin-NFATc4 signaling pathway". In: *Biochem Biophys Res Commun* 495.2, pp. 1864–1870.

Yu, J., V. Tran, M. Li, X. Huang, C. Niu, D. Wang, J. Zhu, J. Wang, Q. Gao, and J. Liu (2012). "Both phthiocerol dimycocerosates and phenolic glycolipids are required for virulence of *Mycobacterium marinum*". In: *Infect Immun* 80.4, pp. 1381–9.

Yu, Q., Y. Wang, L. Dong, Y. He, R. Liu, Q. Yang, Y. Cao, Y. Wang, A. Jia, Y. Bi, and G. Liu (2020). "Regulations of Glycolytic Activities on Macrophages Functions in Tumor and Infectious Inflammation". In: *Front Cell Infect Microbiol* 10, p. 287.

Yu, X., J. Feng, L. Huang, H. Gao, J. Liu, S. Bai, B. Wu, and J. Xie (2019). "Molecular Basis Underlying Host Immunity Subversion by *Mycobacterium tuberculosis* PE/PPE Family Molecules". In: *DNA Cell Biol* 38.11, pp. 1178–1187.

Yu, Y., K. Dai, Z. Gao, W. Tang, T. Shen, Y. Yuan, J. Wang, and C. Liu (2021). "Sulfated polysaccharide directs therapeutic angiogenesis via endogenous VEGF secretion of macrophages". In: *Sci Adv* 7.7.

Yutani, H. (2022). "gghighlight: Highlight Lines and Points in ggplot2". In.

Zahra, F. T., M. S. Sajib, and C. M. Mikelis (2021). “Role of bFGF in Acquired Resistance upon Anti-VEGF Therapy in Cancer”. In: *Cancers (Basel)* 13.6.

Zanoni, I., M. Foti, P. Ricciardi-Castagnoli, and F. Granucci (2005). “TLR-dependent activation stimuli associated with Th1 responses confer NK cell stimulatory capacity to mouse dendritic cells”. In: *J Immunol* 175.1, pp. 286–92.

Zanoni, I. and F. Granucci (2012). “Regulation and dysregulation of innate immunity by NFAT signaling downstream of pattern recognition receptors (PRRs)”. In: *Eur J Immunol* 42.8, pp. 1924–31.

– One of three notable reviews on the subject of NFAT signaling in myeloid cells (the others being Fric, Zelante, Wong, Mertes, Yu, and Ricciardi-Castagnoli (2012b) and Bendickova, Tidu, and Fric (2017)), this one focuses on the roles of NFAT in mediating innate responses that are able to interface with adaptive immunity with a focus on Ivan Zanoni’s own work on LPS-CD14 interactions driving NFAT activation, expanding the model for receptors able to active NFAT signaling beyond C-type lectins.

Zanoni, I., R. Ostuni, G. Capuano, M. Collini, M. Caccia, A. E. Ronchi, M. Rocchetti, F. Mingozzi, M. Foti, G. Chirico, B. Costa, A. Zaza, P. Ricciardi-Castagnoli, and F. Granucci (2009). “CD14 regulates the dendritic cell life cycle after LPS exposure through NFAT activation”. In: *Nature* 460.7252, pp. 264–8.

– While TLRs do not activate NFAT, LPS does. This paradox is resolved through the capacity for the LPS co-receptor, CD14, to directly activate NFAT independently of TLR4 and MYD88. CD14 is localized to lipid rafts and associates with PLC γ 2 to induce the activation of NFAT. This resolves a fundamental question and opens up new opportunities for CD14 to activate NFAT in other contexts, including perhaps in the study from Bowdish, Sakamoto, Kim, Kroos, Mukhopadhyay, Leifer, Tryggvason, Gordon, and Russell (2009), where TDM-mediated responses were shown to be dependent on CD14.

Zelante, T., A. Y. Wong, A. Mencarelli, S. Foo, F. Zolezzi, B. Lee, M. Poidinger, P. Ricciardi-Castagnoli, and J. Fric (2017). “Impaired calcineurin signaling in myeloid cells results in downregulation of pentraxin-3 and increased susceptibility to aspergillosis”. In: *Mucosal Immunol* 10.2, pp. 470–480.

Zelensky, A. N. and J. E. Gready (2005). “The C-type lectin-like domain superfamily”. In: *FEBS J* 272.24, pp. 6179–217.

- Zenaro, E., M. Donini, and S. Dusi (2009). “Induction of Th1/Th17 immune response by *Mycobacterium tuberculosis*: role of dectin-1, Mannose Receptor, and DC-SIGN”. In: *J Leukoc Biol* 86.6, pp. 1393–401.
- Zenk, S. F., S. Hauck, D. Mayer, M. Grieshaber, and S. Stenger (2021). “Stabilization of Hypoxia-Inducible Factor Promotes Antimicrobial Activity of Human Macrophages Against *Mycobacterium tuberculosis*”. In: *Front Immunol* 12, p. 678354.
- Zhan, Q., F. Miao, R. Huang, X. Zhou, M. Ge, and X. Liang (2019). “Efficacy and safety of bevacizumab combined with chemotherapy in symptomatic brain metastases from lung adenocarcinoma: a retrospective analysis”. In: *J Thorac Dis* 11.11, pp. 4725–4734.
- Zhanel, G. G., S. Siemens, K. Slayter, and L. Mandell (1999). “Antibiotic and oral contraceptive drug interactions: Is there a need for concern?” In: *Can J Infect Dis* 10.6, pp. 429–33.
- Zhang, C., N. Wang, H. Y. Tan, W. Guo, F. Chen, Z. Zhong, K. Man, S. W. Tsao, L. Lao, and Y. Feng (2020a). “Direct inhibition of the TLR4/MyD88 pathway by geniposide suppresses HIF-1alpha-independent VEGF expression and angiogenesis in hepatocellular carcinoma”. In: *Br J Pharmacol* 177.14, pp. 3240–3257.
- Zhang, H., X. Xie, X. Zhu, J. Zhu, C. Hao, Q. Lu, L. Ding, Y. Liu, L. Zhou, Y. Liu, C. Huang, C. Wen, and Q. Ye (2005). “Stimulatory cross-talk between NFAT3 and estrogen receptor in breast cancer cells”. In: *J Biol Chem* 280.52, pp. 43188–97.
- Zhang, L., R. Li, W. Shi, X. Liang, S. Liu, Z. Ye, C. Yu, Y. Chen, B. Zhang, W. Wang, Y. Lai, J. Ma, Z. Li, and X. Tan (2013). “NFAT2 inhibitor ameliorates diabetic nephropathy and podocyte injury in db/db mice”. In: *Br J Pharmacol* 170.2, pp. 426–39.
- Zhang, L., Y. Zhao, Y. Gao, L. Wu, R. Gao, Q. Zhang, Y. Wang, C. Wu, F. Wu, S. S. Gurcha, N. Veerapen, S. M. Batt, W. Zhao, L. Qin, X. Yang, M. Wang, Y. Zhu, B. Zhang, L. Bi, X. Zhang, H. Yang, L. W. Guddat, W. Xu, Q. Wang, J. Li, G. S. Besra, and Z. Rao (2020b). “Structures of cell wall arabinosyltransferases with the anti-tuberculosis drug ethambutol”. In: *Science* 368.6496, pp. 1211–1219.
- Zhang, N., J. Li, R. Luo, J. Jiang, and J. A. Wang (2008a). “Bone marrow mesenchymal stem cells induce angiogenesis and attenuate the remodeling of diabetic cardiomyopathy”. In: *Exp Clin Endocrinol Diabetes* 116.2, pp. 104–11.

- Zhang, Y., D. Chen, L. Zhao, W. Li, Y. Ni, Y. Chen, and H. Li (2019). “Nfatc4 Deficiency Attenuates Ototoxicity by Suppressing Tnf-Mediated Hair Cell Apoptosis in the Mouse Cochlea”. In: *Front Immunol* 10, p. 1660.
- Zhang, Y., J. Gaekwad, M. A. Wolfert, and G. J. Boons (2008b). “Synthetic tetraacylated derivatives of lipid A from *Porphyromonas gingivalis* are antagonists of human TLR4”. In: *Org Biomol Chem* 6.18, pp. 3371–81.
- Zhang, Y., H. Huang, M. Coleman, A. Ziemys, P. Gopal, S. M. Kazmi, and R. A. Brekken (2021a). “VEGFR2 activity on myeloid cells mediates immune suppression in the tumor microenvironment”. In: *JCI Insight* 6.23.
- Zhang, Y., W. Qin, X. Lu, J. Xu, H. Huang, H. Bai, S. Li, and S. Lin (2017). “Programmable base editing of zebrafish genome using a modified CRISPR-Cas9 system”. In: *Nat Commun* 8.1, p. 118.
- Zhang, Y., W. Shi, W. Zhang, and D. Mitchison (2014). “Mechanisms of Pyrazinamide Action and Resistance”. In: *Microbiol Spectr* 2.4, MGM2–0023–2013.
- Zhang, Y., M. M. Wade, A. Scorpio, H. Zhang, and Z. Sun (2003). “Mode of action of pyrazinamide: disruption of *Mycobacterium tuberculosis* membrane transport and energetics by pyrazinoic acid”. In: *J Antimicrob Chemother* 52.5, pp. 790–5.
- Zhang, Yan, Márton Rózsa, Yajie Liang, Daniel Bushey, Ziqiang Wei, Jihong Zheng, Daniel Reep, Gerard Joey Broussard, Arthur Tsang, Getahun Tsegaye, Sujatha Narayan, Christopher J. Obara, Jing-Xuan Lim, Ronak Patel, Rongwei Zhang, Misha B. Ahrens, Glenn C. Turner, Samuel S.-H. Wang, Wyatt L. Korff, Eric R. Schreiter, Karel Svoboda, Jeremy P. Hasseman, Ilya Kolb, and Loren L. Looger (2021b). “Fast and sensitive GCaMP calcium indicators for imaging neural populations”. In: *bioRxiv*.
- Zhao, M., Z. Yu, Z. Li, J. Tang, X. Lai, and L. Liu (2017). “Expression of angiogenic growth factors VEGF, bFGF and ANG1 in colon cancer after bevacizumab treatment *in vitro*: A potential self-regulating mechanism”. In: *Oncol Rep* 37.1, pp. 601–607.
- Zhao, X. Q., L. L. Zhu, Q. Chang, C. Jiang, Y. You, T. Luo, X. M. Jia, and X. Lin (2014). “C-type lectin receptor dectin-3 mediates trehalose 6,6'-dimycolate (TDM)-induced Mincle expression through CARD9/Bcl10/MALT1-dependent nuclear factor (NF)-kappaB activation”. In: *J Biol Chem* 289.43, pp. 30052–62.

Zhao, Y. R., H. Liu, L. M. Xiao, C. G. Jin, Z. P. Zhang, and C. G. Yang (2018). “The clinical significance of CCBE1 expression in human colorectal cancer”. In: *Cancer Manag Res* 10, pp. 6581–6590.

Zhao, Z., X. Hou, X. Yin, Y. Li, R. Duan, B. F. Boyce, and Z. Yao (2015). “TNF Induction of NF-kappaB RelB Enhances RANKL-Induced Osteoclastogenesis by Promoting Inflammatory Macrophage Differentiation but also Limits It through Suppression of NFATc1 Expression”. In: *PLoS One* 10.8, e0135728.

Zhong, Y., A. Kinio, and M. Saleh (2013). “Functions of NOD-Like Receptors in Human Diseases”. In: *Front Immunol* 4, p. 333.

Zhu, L. L., T. M. Luo, X. Xu, Y. H. Guo, X. Q. Zhao, T. T. Wang, B. Tang, Y. Y. Jiang, J. F. Xu, X. Lin, and X. M. Jia (2016). “E3 ubiquitin ligase Cbl-b negatively regulates C-type lectin receptor-mediated antifungal innate immunity”. In: *J Exp Med* 213.8, pp. 1555–70.

Zhu, L. L., X. Q. Zhao, C. Jiang, Y. You, X. P. Chen, Y. Y. Jiang, X. M. Jia, and X. Lin (2013). “C-type lectin receptors Dectin-3 and Dectin-2 form a heterodimeric pattern-recognition receptor for host defense against fungal infection”. In: *Immunity* 39.2, pp. 324–34.

Zhu, X., Y. Zhao, Y. Jiang, T. Qin, J. Chen, X. Chu, Q. Yi, S. Gao, and S. Wang (2017). “Dectin-1 signaling inhibits osteoclastogenesis via IL-33-induced inhibition of NFATc1”. In: *Oncotarget* 8.32, pp. 53366–53374.

Zhu, X. W. and J. S. Friedland (2006). “Multinucleate giant cells and the control of chemokine secretion in response to *Mycobacterium tuberculosis*”. In: *Clin Immunol* 120.1, pp. 10–20.

Zhu, X. W., N. M. Price, R. H. Gilman, S. Recarvarren, and J. S. Friedland (2007). “Multinucleate giant cells release functionally unopposed matrix metalloproteinase-9 *in vitro* and *in vivo*”. In: *J Infect Dis* 196.7, pp. 1076–9.

Zimmerli, S., M. Majeed, M. Gustavsson, O. Stendahl, D. A. Sanan, and J. D. Ernst (1996). “Phagosome-lysosome fusion is a calcium-independent event in macrophages”. In: *J Cell Biol* 132.1-2, pp. 49–61.

Zon, L. I. and R. T. Peterson (2005). “*In vivo* drug discovery in the zebrafish”. In: *Nat Rev Drug Discov* 4.1, pp. 35–44.

Zou, J. and C. J. Secombes (2016). “The Function of Fish Cytokines”. In: *Biology (Basel)* 5.2.

Zumla, A., M. Rao, R. S. Wallis, S. H. Kaufmann, R. Rustomjee, P. Mwaba, C. Vilaplana, D. Yeboah-Manu, J. Chakaya, G. Ippolito, E. Azhar, M. Hoelscher, M. Maeurer, and consortium Host-Directed Therapies Network (2016). “Host-directed therapies for infectious diseases: current status, recent progress, and future prospects”. In: *Lancet Infect Dis* 16.4, e47–63.

Biography

Jared was born in the mountains of Appalachia in Barbourville, KY in 1994. After graduating valedictorian (in a class of 57) from Barbourville High School, he enrolled at Transylvania University in Lexington, KY where he received a Bachelor of Arts degree in Biology and Political Science, *summa cum laude* in May 2016. In the fall of 2016, he began his Ph.D. at Duke University in the Molecular Genetics and Microbiology department, having been awarded a James B. Duke Fellowship. He then joined the lab of David Tobin in the summer of 2017. He attended the Cold Spring Harbor course “Programming for Biologists” in the fall of 2017, where he learned Python. In his time at Duke, he has contributed to a number of publications, including “Cyclopropane Modification of Trehalose Dimycolate Drives Granuloma Angiogenesis and Mycobacterial Growth through Vegf Signaling” by Walton et al. (2018), “A Non-Canonical Type 2 Immune Response Coordinates Tuberculous Granuloma Formation and Epithelialization” by Cronan and Hughes et al. (2021), “An Ancestral Mycobacterial Effector Promotes Dissemination of Infection” by Saelens and Sweeney et al. (2022) and “Macrophage NFATC2 Mediates Angiogenic Signaling During Mycobacterial Infection” (2022), his primary thesis publication. He was awarded a Ruth L. Kirschstein National Research Service Award F31 fellowship from the National Heart, Lung, and Blood Institute to assist in the completion of his thesis work.