

## VIT-AP UNIVERSITY, ANDHRA PRADESH

### CSE4027 – Data Analytics - Lab Sheet :10

**Academic year:** 2022-2023

**Branch/ Class:** B.Tech

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**School:** SCOPE

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**Reg. no.:**20BCD7171

#### LAB 10

Clustering and Classification Algorithm implementation using R.

1. Use iris dataset for k means clustering in R

```
> setwd("c:/users/jassu/OneDrive/Desktop")
> data(iris)
> str(iris)
'data.frame': 150 obs. of 5 variables:
 $ Sepal.Length: num  5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
 $ Sepal.Width : num  3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
 $ Petal.Length: num  1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
 $ Petal.Width : num  0.2 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
 $ Species      : Factor w/ 3 levels "setosa","versicolor",...: 1 1 1 1 1 1 1 1 1 1 ...
> install.packages("cluster")
WARNING: Rtools is required to build R packages but is not currently installed. Please download and install the appropriate version of Rtools before proceeding:
https://cran.rstudio.com/bin/windows/Rtools/
also installing the dependencies 'gtools', 'gmp'

trying URL 'https://cran.rstudio.com/bin/windows/contrib/4.2/gtools_3.9.3.zip'
Content type 'application/zip' length 359714 bytes (351 KB)
downloaded 351 KB

trying URL 'https://cran.rstudio.com/bin/windows/contrib/4.2/gmp_0.6-8.zip'
Content type 'application/zip' length 737069 bytes (719 KB)
downloaded 719 KB

trying URL 'https://cran.rstudio.com/bin/windows/contrib/4.2/cluster_1.2.7.zip'
Content type 'application/zip' length 1481733 bytes (1.4 MB)
downloaded 1.4 MB

package 'gtools' successfully unpacked and MD5 sums checked
package 'gmp' successfully unpacked and MD5 sums checked
package 'cluster' successfully unpacked and MD5 sums checked

The downloaded binary packages are in
C:\Users\jassu\AppData\Local\Temp\RtmpwNb1no\downloaded_packages
> install.packages("cluster")
WARNING: Rtools is required to build R packages but is not currently installed. Please download and install the appropriate version of Rtools before proceeding:
https://cran.rstudio.com/bin/windows/Rtools/
trying URL 'https://cran.rstudio.com/bin/windows/contrib/4.2/cluster_2.1.4.zip'
Content type 'application/zip' length 586631 bytes (572 KB)
downloaded 572 KB

package 'cluster' successfully unpacked and MD5 sums checked

The downloaded binary packages are in
C:\Users\jassu\AppData\Local\Temp\RtmpwNb1no\downloaded_packages
```

```

> iris_1 <- iris[, -5]
> set.seed(240)
> kmeans.re <- kmeans(iris_1, centers = 3, nstart = 20)
> kmeans.re
K-means clustering with 3 clusters of sizes 50, 62, 38

Cluster means:
  Sepal.Length Sepal.Width Petal.Length Petal.Width
1    5.006000    3.428000    1.462000    0.246000
2    5.901613    2.748387    4.393548    1.433871
3    6.850000    3.073684    5.742105    2.071053

Clustering vector:
 [1] 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
[36] 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 2 2 3 2 2 2 2 2 2 2 2 2 2 2 2 2 2
[71] 2 2 2 2 2 2 3 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 3 2 3 3
[106] 3 2 3 3 3 3 3 3 2 2 3 3 3 2 3 2 3 2 3 2 3 2 3 3 3 3 3 3 2 3 3 3
[141] 3 3 2 3 3 3 2 3 3 2

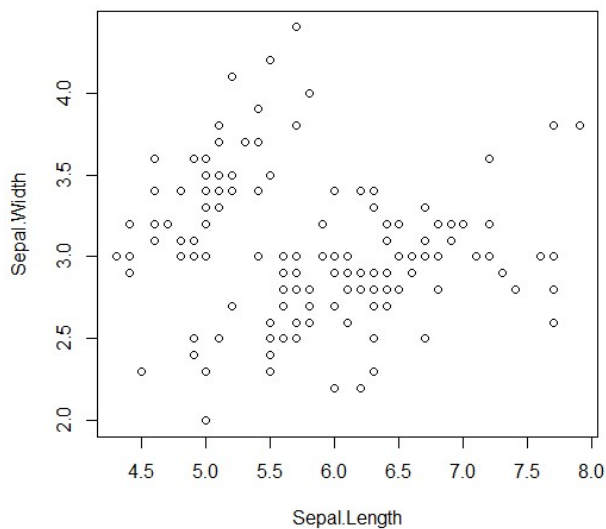
within cluster sum of squares by cluster:
[1] 15.15100 39.82097 23.87947
   (between_SS / total_SS =  88.4 %)

Available components:

[1] "cluster"      "centers"      "totss"        "withinss"
[5] "tot.withinss" "betweenss"    "size"         "iter"
[9] "ifault"
> kmeans.re$cluster
 [1] 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
[36] 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 2 2 3 2 2 2 2 2 2 2 2 2 2 2 2 2 2
[71] 2 2 2 2 2 2 2 3 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 3 2 3 3
[106] 3 2 3 3 3 3 3 3 2 2 3 3 3 2 3 2 3 2 3 2 3 2 3 3 3 3 3 3 2 3 3 3
[141] 3 3 2 3 3 3 2 3 3 2
>
> cm <- table(iris$Species, kmeans.re$cluster)
> cm

      1  2  3
setosa  50  0  0
versicolor  0 48  2
virginica  0 14 36
> plot(iris_1[c("Sepal.Length", "Sepal.Width")])
> |

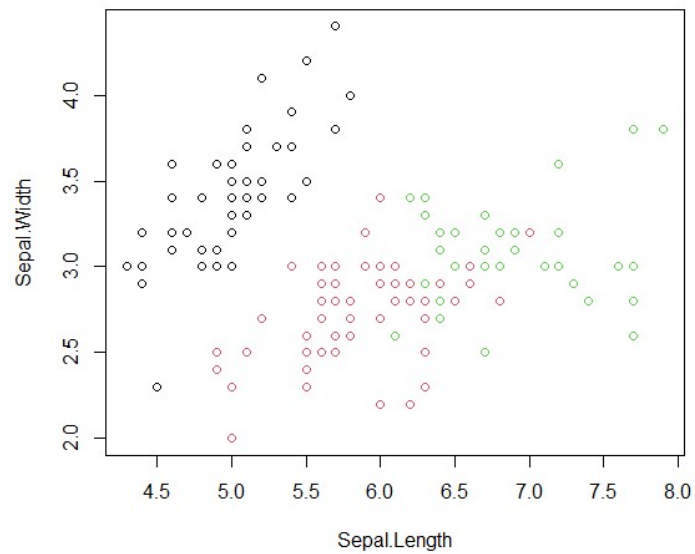
```



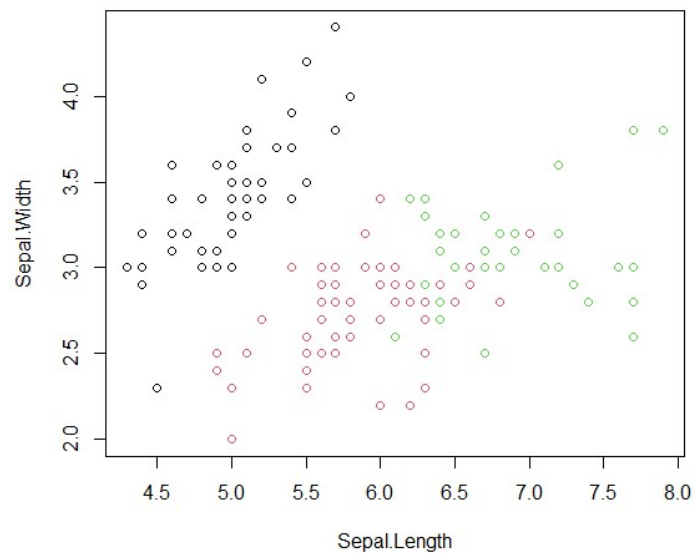
```

> plot(iris_1[c("Sepal.Length", "Sepal.Width")], col = kmeans.re$cluster)
> plot(iris_1[c("Sepal.Length", "Sepal.Width")],
+     col = kmeans.re$cluster,
+     main = "K-means with 3 clusters")

```



**K-means with 3 clusters**



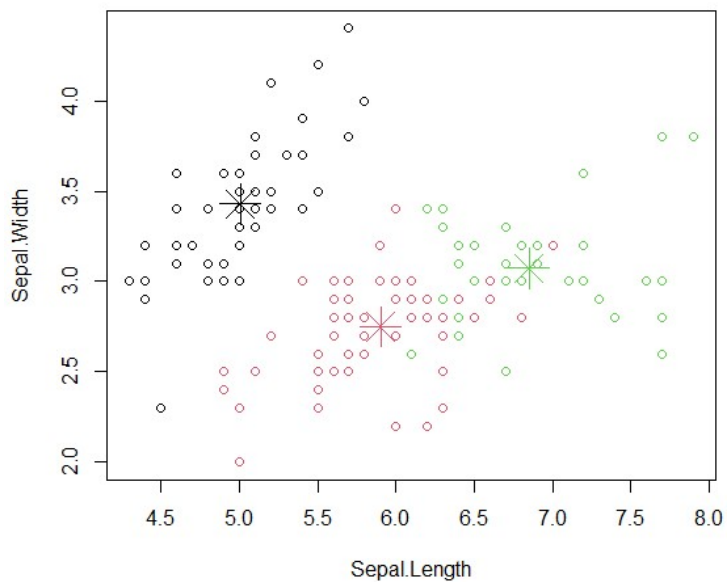
```

> kmeans.re$centers
  Sepal.Length Sepal.Width Petal.Length Petal.Width
1    5.006000    3.428000    1.462000    0.246000
2    5.901613    2.748387    4.393548    1.433871
3    6.850000    3.073684    5.742105    2.071053
> kmeans.re$centers[, c("Sepal.Length", "Sepal.Width")]
  Sepal.Length Sepal.Width
1    5.006000    3.428000
2    5.901613    2.748387
3    6.850000    3.073684
> |

> points(kmeans.re$centers[, c("Sepal.Length", "Sepal.Width")],
+        col = 1:3, pch = 8, cex = 3)
> |

```

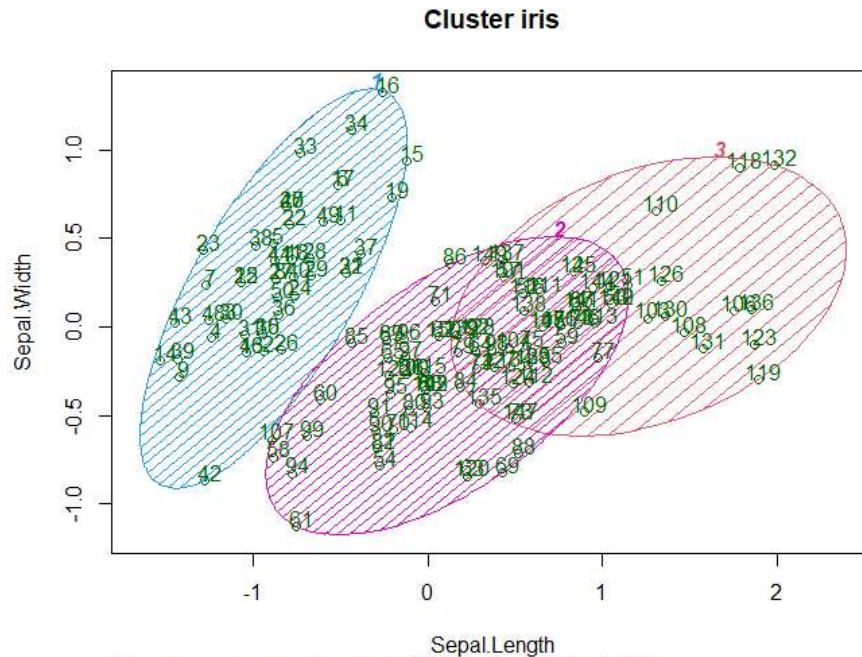
### K-means with 3 clusters



```

> y_kmeans <- kmeans.re$cluster
> clusplot(iris_1[, c("Sepal.Length", "Sepal.Width")],
+          y_kmeans,
+          lines = 0,
+          shade = TRUE,
+          color = TRUE,
+          labels = 2,
+          plotchar = FALSE,
+          span = TRUE,
+          main = paste("Cluster iris"),
+          xlab = 'Sepal.Length',
+          ylab = 'Sepal.Width')
>

```



These two components explain 100 % of the point variability.

2. Use readingSkillsdataset for all classification practice which is default in party package
  - a. Logistic regression

```
> library(caTools)
> library(ROCR)
> split <- sample.split(mtcars, SplitRatio = 0.8)
> split
[1] TRUE TRUE TRUE TRUE TRUE FALSE TRUE FALSE FALSE TRUE TRUE
> split <- sample.split(readingSkills, splitRatio = 0.8)
> split
[1] TRUE TRUE FALSE TRUE
> train_reg <- subset(readingSkills, split == "TRUE")
> test_reg <- subset(readingSkills, split == "FALSE")
>
> logistic_model <- glm( nativespeaker ~ shoesize + disp,
+                        data = train_reg,
+                        family = "binomial")
```



```

> logistic_model <- glm( nativespeaker ~ shoeSize + score,
+                         data = train_reg,
+                         family = "binomial")
> logistic_model

Call:  glm(formula = nativespeaker ~ shoeSize + score, family = "binomial",
          data = train_reg)

Coefficients:
(Intercept)      shoeSize          score 
  32.9899      -2.1367         0.6578 

Degrees of Freedom: 149 Total (i.e. Null); 147 Residual
Null Deviance:      207
Residual Deviance: 104.4      AIC: 110.4
> summary(logistic_model)

Call:
glm(formula = nativespeaker ~ shoeSize + score, family = "binomial",
    data = train_reg)

Deviance Residuals:
    Min       1Q   Median       3Q      Max 
-2.1414 -0.4001  0.1037  0.6069  2.3226 

Coefficients:
              Estimate Std. Error z value Pr(>|z|)
(Intercept)   32.9899     6.3843   5.167 2.37e-07 ***
shoeSize      -2.1367     0.3713  -5.754 8.71e-09 ***
score          0.6578     0.1071   6.142 8.13e-10 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

    Null deviance: 206.98  on 149  degrees of freedom
Residual deviance: 104.35  on 147  degrees of freedom
AIC: 110.35

Number of Fisher Scoring iterations: 6

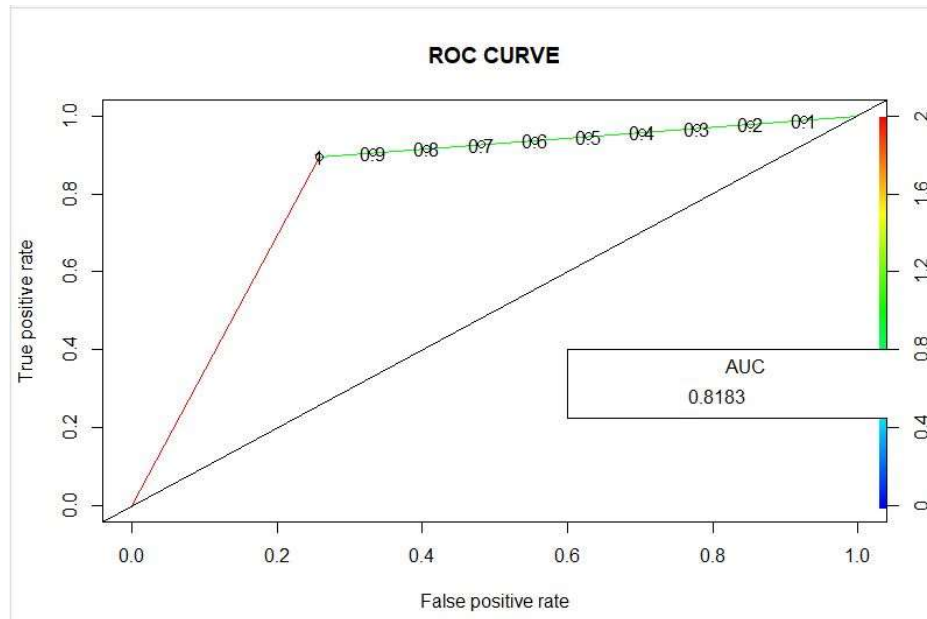
> predict_reg <- predict(logistic_model,
+                         test_reg, type = "response")
> predict_reg
      3      7     11     15
0.6497343204 0.0674429037 0.8001818605 0.1259361937 0.7562228265
      23     27     31     35
0.0390415578 0.2294823509 0.9645698007 0.9909243543 0.9823010644
      43     47     51     55
0.0125358212 0.8807316927 0.8566905201 0.9932261263 0.7389391026
      63     67     71     75
0.0059649782 0.5153077200 0.9926446401 0.0137680617 0.0840078816
      83     87     91     95
0.4937513686 0.5825939761 0.5895929497 0.9979359935 0.5828645787
     103     107     111     115
0.0658866991 0.0664138456 0.8556034606 0.1236914119 0.2364435584
     123     127     131     135
0.8839576303 0.9776036049 0.2323147919 0.9849456681 0.0152679086
     143     147     151     155
0.0381222170 0.9789445806 0.0347475155 0.9998080321 0.1898662649
     163     167     171     175
0.0009417221 0.1363075478 0.0511401642 0.0295756698 0.3720995610
     183     187     191     195
0.9849410286 0.9560153483 0.0598180674 0.9581764030 0.0863175500
> predict_reg <- ifelse(predict_reg > 0.5, 1, 0)
>
> table(test_reg$ nativespeaker, predict_reg)
      predict_reg
      0      1
no  23     8
yes  2    17
> missing_classerr <- mean(predict_reg != test_reg$ nativespeaker)
> print(paste('Accuracy =', 1 - missing_classerr))
[1] "Accuracy = 0"
> ROCPred <- prediction(predict_reg, test_reg$ nativespeaker)
> ROCPer <- performance(ROCPred, measure = "tpr",
+                       x.measure = "fpr")
>
> auc <- performance(ROCPred, measure = "auc")
> auc <- auc@y.values[[1]]
> auc
[1] 0.8183362

```

```

> plot(ROCPer)
> plot(ROCPer, colorize = TRUE,
+       print.cutoffs.at = seq(0.1, by = 0.1),
+       main = "ROC CURVE")
> abline(a = 0, b = 1)
>
> auc <- round(auc, 4)
> legend(.6, .4, auc, title = "AUC", cex = 1)
>

```

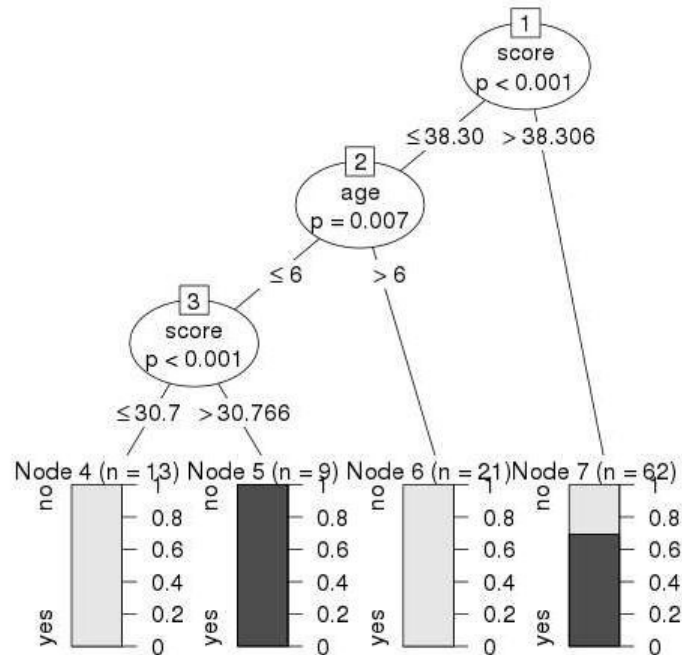


## b. Decision trees

```

> library(party)
> print(head(readingskills))
  nativeSpeaker age shoeSize   score
1         yes    5  24.83189 32.29385
2         yes    6  25.95238 36.63105
3         no   11  30.42170 49.60593
4         yes    7  28.66450 40.28456
5         yes   11  31.88207 55.46085
6         yes   10  30.07843 52.83124
> input.dat <- readingskills[c(1:105),]
> png(file = "decision_tree.png")
> output.tree <- ctree(
+   nativeSpeaker ~ age + shoeSize + score,
+   data = input.dat)
> plot(output.tree)
>
> dev.off()
RStudioGD
  2
>

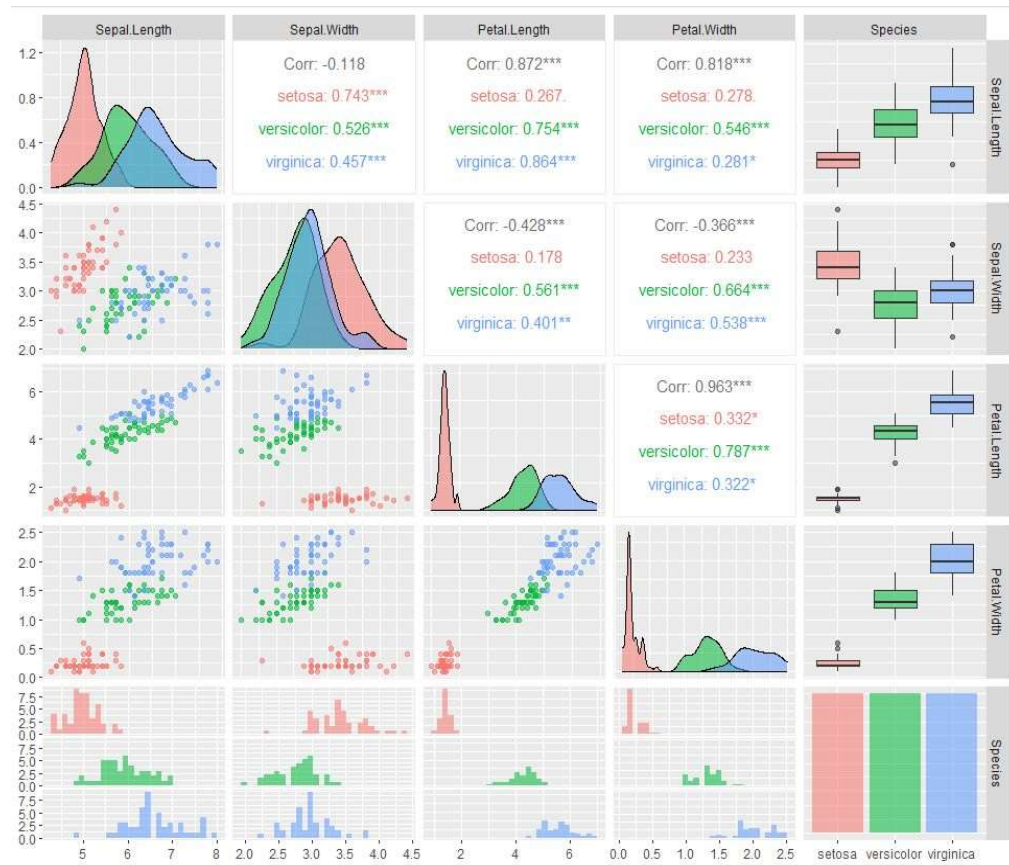
```



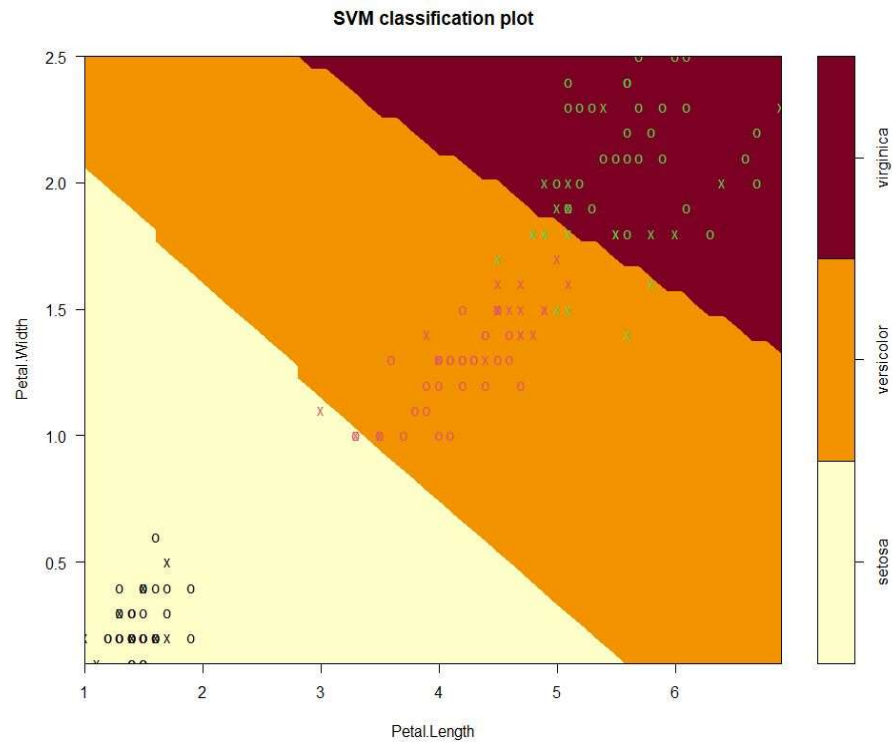
### c. Support Vector Machines (iris data – default)

```
> library("e1071")
> library(ggally)
Loading required package: ggplot2
Registered S3 method overwritten by 'ggally':
  method from
+ .gg      ggplot2
> library(ggplot2)
> data(iris)
> str(iris)
'data.frame':  150 obs. of  5 variables:
 $ Sepal.Length: num  5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
 $ Sepal.Width : num  3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
 $ Petal.Length: num  1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
 $ Petal.Width : num  0.2 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
 $ Species     : Factor w/ 3 levels "setosa","versicolor",...: 1 1 1 1 1 1 1 1 1 1 ...
> head(iris,5)
  Sepal.Length Sepal.Width Petal.Length Petal.Width Species
1          5.1          3.5          1.4          0.2  setosa
2          4.9          3.0          1.4          0.2  setosa
3          4.7          3.2          1.3          0.2  setosa
4          4.6          3.1          1.5          0.2  setosa
5          5.0          3.6          1.4          0.2  setosa
> svm_model <- svm(Species ~., data=iris,
+                  kernel="radial")
> ggpairs(iris, ggplot2::aes(colour = Species, alpha = 0.4))
plot: [5,1] [=====] 84% est: 0s `st
_bin()` using `bins = 30`. Pick better value with `binwidth`.
plot: [5,2] [=====] 88% est: 0s `st
_bin()` using `bins = 30`. Pick better value with `binwidth`.
plot: [5,3] [=====] 92% est: 0s `st
_bin()` using `bins = 30`. Pick better value with `binwidth`.
plot: [5,4] [=====] 96% est: 0s `st
_bin()` using `bins = 30`. Pick better value with `binwidth`.
>
```

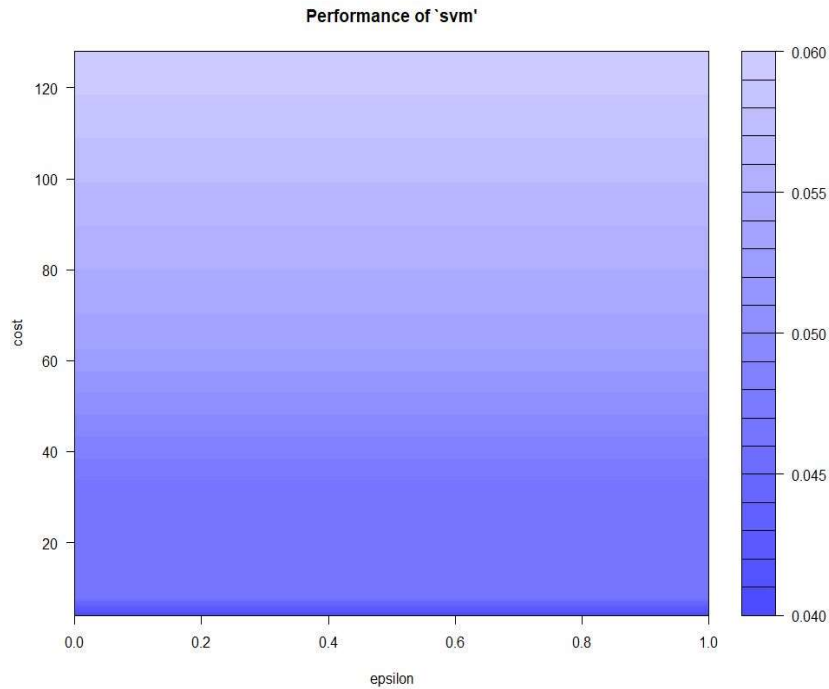




```
> plot(svm_model, data=iris,
+       Petal.Width~Petal.Length,
+       slice = list(Sepal.Width=3, Sepal.Length=4)
+ )
>
```



```
> tab = table(Predicted=pred, Actual = iris$Species)
> tab
      Actual
Predicted setosa versicolor virginica
setosa      50         0         0
versicolor  0         48         2
virginica   0         2         48
> 1-sum(diag(tab)/sum(tab))
[1] 0.02666667
> set.seed(123)
> tmodel=tune(svm,species=., data=iris,
+             ranges=list(epsilon= seq(0,1,0.1), cost = 2^(2:7)))
> plot(tmodel)
>
```



```
> summary(tmodel)

Parameter tuning of 'svm':
- sampling method: 10-fold cross validation

- best parameters:
  epsilon cost
    0      4

- best performance: 0.04

- Detailed performance results:
  epsilon cost      error dispersion
1      0.0      4 0.04000000 0.04661373
2      0.1      4 0.04000000 0.04661373
3      0.2      4 0.04000000 0.04661373
4      0.3      4 0.04000000 0.04661373
5      0.4      4 0.04000000 0.04661373
6      0.5      4 0.04000000 0.04661373
7      0.6      4 0.04000000 0.04661373
8      0.7      4 0.04000000 0.04661373
9      0.8      4 0.04000000 0.04661373
10     0.9      4 0.04000000 0.04661373
11     1.0      4 0.04000000 0.04661373
12     0.0      8 0.04666667 0.06324555
13     0.1      8 0.04666667 0.06324555
14     0.2      8 0.04666667 0.06324555
15     0.3      8 0.04666667 0.06324555
16     0.4      8 0.04666667 0.06324555
17     0.5      8 0.04666667 0.06324555
18     0.6      8 0.04666667 0.06324555
19     0.7      8 0.04666667 0.06324555
20     0.8      8 0.04666667 0.06324555
21     0.9      8 0.04666667 0.06324555
22     1.0      8 0.04666667 0.06324555
23     0.0     16 0.04666667 0.04499657
24     0.1     16 0.04666667 0.04499657
25     0.2     16 0.04666667 0.04499657
26     0.3     16 0.04666667 0.04499657
27     0.4     16 0.04666667 0.04499657
28     0.5     16 0.04666667 0.04499657
29     0.6     16 0.04666667 0.04499657
30     0.7     16 0.04666667 0.04499657
31     0.8     16 0.04666667 0.04499657
32     0.9     16 0.04666667 0.04499657
33     1.0     16 0.04666667 0.04499657
34     0.0     32 0.04666667 0.04499657
35     0.1     32 0.04666667 0.04499657
36     0.2     32 0.04666667 0.04499657
37     0.3     32 0.04666667 0.04499657
38     0.4     32 0.04666667 0.04499657
39     0.5     32 0.04666667 0.04499657
40     0.6     32 0.04666667 0.04499657
41     0.7     32 0.04666667 0.04499657
..
```

```

> mymodel=tmodel$best.model
warning messages:
1: In doTryCatch(return(expr), name, parentenv, handler) :
  display list redraw incomplete
2: In doTryCatch(return(expr), name, parentenv, handler) :
  display list redraw incomplete
3: In doTryCatch(return(expr), name, parentenv, handler) :
  display list redraw incomplete
> summary(mymodel)

Call:
best.tune(METHOD = svm, train.x = Species ~ ., data = iris, ranges = list(epsilon = seq(0,
  1, 0.1), cost = 2^(2:7)))

Parameters:
  SVM-Type:  C-classification
  SVM-Kernel: radial
  cost:      4

Number of Support Vectors:  37

( 6 17 14 )

Number of Classes:  3

Levels:
setosa versicolor virginica

```

```

> mymodel=tmodel$best.model
> summary(mymodel)

Call:
best.tune(METHOD = svm, train.x = Species ~ ., data = iris, ranges = list(epsilon = seq(0,
  1, 0.1), cost = 2^(2:7)))

Parameters:
  SVM-Type:  C-classification
  SVM-Kernel: radial
  cost:      4

Number of Support Vectors:  37

( 6 17 14 )

Number of Classes:  3

Levels:
setosa versicolor virginica

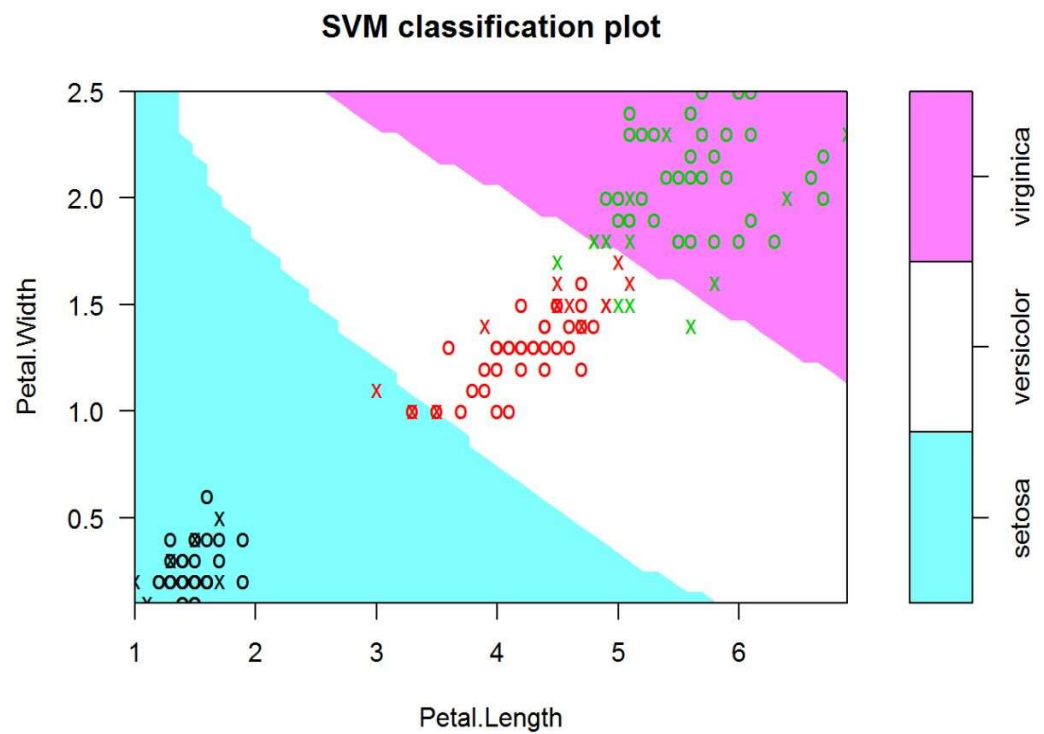
> |

```

```

> plot(mymodel, data=iris,
+       Petal.Width~Petal.Length,
+       slice = list(Sepal.Width=3, Sepal.Length=4)
+ )

```



```
> pred1 = predict(mymodel,iris)
> tab1 = table(Predicted=pred1, Actual = iris$species)
> tab1
```

	Actual		
Predicted	setosa	versicolor	virginica
setosa	50	0	0
versicolor	0	48	0
virginica	0	2	50

```
> 1-sum(diag(tab1)/sum(tab1))
[1] 0.01333333
> |
```

d. Naive Bayes Classifier (use hsbdata.csv)



```
data <- iris      # use the iris dataset
```

```
head(data)        # head() returns the top 6 rows of the datafi
```

A data.frame: 6 × 5

	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
1	5.1	3.5	1.4	0.2	setosa
2	4.9	3.0	1.4	0.2	setosa
3	4.7	3.2	1.3	0.2	setosa
4	4.6	3.1	1.5	0.2	setosa
5	5.0	3.6	1.4	0.2	setosa
6	5.4	3.9	1.7	0.4	setosa

```
In [ ]: summary(data)      # returns the statistical summary of the data columns
```

```
  Sepal.Length  Sepal.Width  Petal.Length  Petal.Width
Min.   :4.300   Min.   :2.000   Min.   :1.000   Min.   :0.100
1st Qu.:5.100   1st Qu.:2.800   1st Qu.:1.600   1st Qu.:0.300
Median :5.800   Median :3.000   Median :4.350   Median :1.300
Mean   :5.843   Mean   :3.057   Mean   :3.758   Mean   :1.199
3rd Qu.:6.400   3rd Qu.:3.300   3rd Qu.:5.100   3rd Qu.:1.800
Max.   :7.900   Max.   :4.400   Max.   :6.900   Max.   :2.500
Species
setosa   :50
versicolor:50
virginica :50
```

```
In [ ]: dim(data)         # returns number of rows and columns in the dataset
```

```
1.150
2.5
```

```
## End of Lecture 11
```

```
In [ ]: # split the data into train-test with a ratio 80:20
split <- sample.split(iris, SplitRatio = 0.8)
train_data <- subset(data, split == "TRUE")
test_data <- subset(data, split == "FALSE")
```

```
In [ ]: # Feature Scaling
train_scale <- scale(train_data[, 1:4])
test_scale <- scale(test_data[, 1:4])
```

```
In [ ]: dim(train_data)
```

```
1.120
2.5
```

```
In [ ]: dim(test_data)
```

```
1.30
2.5
```

```
set.seed(1) # Setting Seed
classifier_naive <- naiveBayes(Species~ ., data = train_data)
classifier_naive
```

Naive Bayes Classifier for Discrete Predictors

Call:  
naiveBayes.default(x = X, y = Y, laplace = laplace)

A-priori probabilities:

Y	setosa	versicolor	virginica
	0.3333333	0.3333333	0.3333333

Conditional probabilities:

		Sepal.Length	
Y		[,1]	[,2]
	setosa	5.020	0.3131314
	versicolor	5.965	0.5206259
	virginica	6.625	0.6581832

		Sepal.Width	
Y		[,1]	[,2]
	setosa	3.4375	0.3739515
	versicolor	2.8000	0.3137858
	virginica	3.0075	0.3237501

	Petal.Length	
Y	[,1]	[,2]
setosa	1.4625	0.1734824
versicolor	4.2625	0.4204317
virginica	5.5650	0.5404414

	Petal.Width	
Y	[,1]	[,2]
setosa	0.2400	0.09001424
versicolor	1.3275	0.20253205
virginica	2.0400	0.27623847

```
In [ ]: summary(classifier_naive)
```

	Length	Class	Mode
apriori	3	table	numeric
tables	4	-none-	list
levels	3	-none-	character
isnumeric	4	-none-	logical
call	4	-none-	call

```
In [ ]: # Predicting on test data'
y_pred <- predict(classifier_naive, newdata = test_data)
```

```
In [ ]: # Confusion Matrix
conf_mat <- table(test_data$Species,y_pred)
print(conf_mat)
```

	y_pred		
	setosa	versicolor	virginica
setosa	10	0	0
versicolor	0	10	0
virginica	0	1	9

```

In [ ]: # Model Evaluation
confusionMatrix(conf_mat)

Confusion Matrix and Statistics

          y_pred
      setosa versicolor virginica
setosa      10         0         0
versicolor  0         10        0
virginica   0          1         9

Overall Statistics

          Accuracy : 0.9667
          95% CI : (0.8278, 0.9992)
      No Information Rate : 0.3667
      P-Value [Acc > NIR] : 4.476e-12

          Kappa : 0.95

      McNemar's Test P-Value : NA

Statistics by Class:

              Class: setosa Class: versicolor Class: virginica
Sensitivity              1.0000              0.9091              1.0000
Specificity              1.0000              1.0000              0.9524
Pos Pred Value           1.0000              1.0000              0.9000

Neg Pred Value           1.0000              0.9500              1.0000
Prevalence                0.3333              0.3667              0.3000
Detection Rate            0.3333              0.3333              0.3000
Detection Prevalence      0.3333              0.3333              0.3333
Balanced Accuracy         1.0000              0.9545              0.9762

Setosa : correctly classified 10.

Versicolor : correctly classified 10, wrongly classified 1.

Virginica : correctly identified 9.

And also, the model achieved an accuracy of 96%

```

e. k-Nearest Neighbour (iris data)

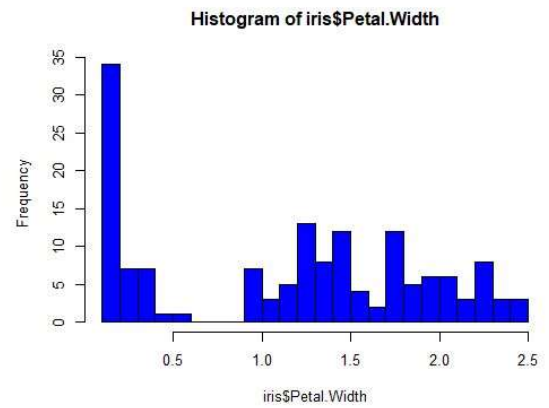
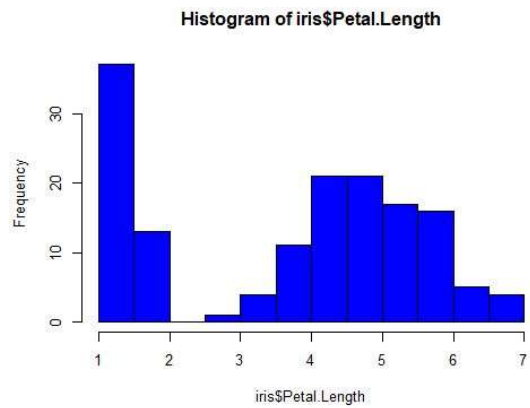
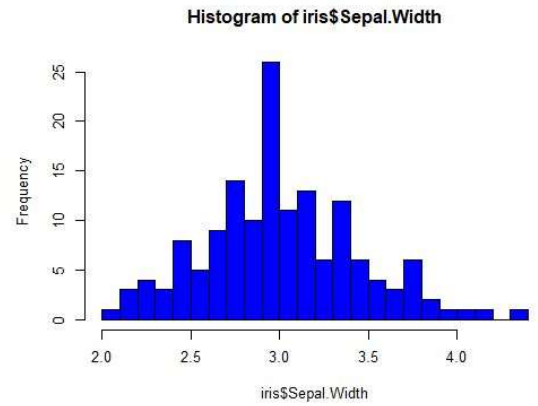
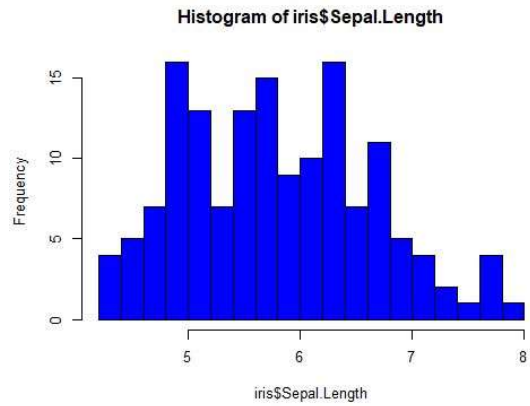
```

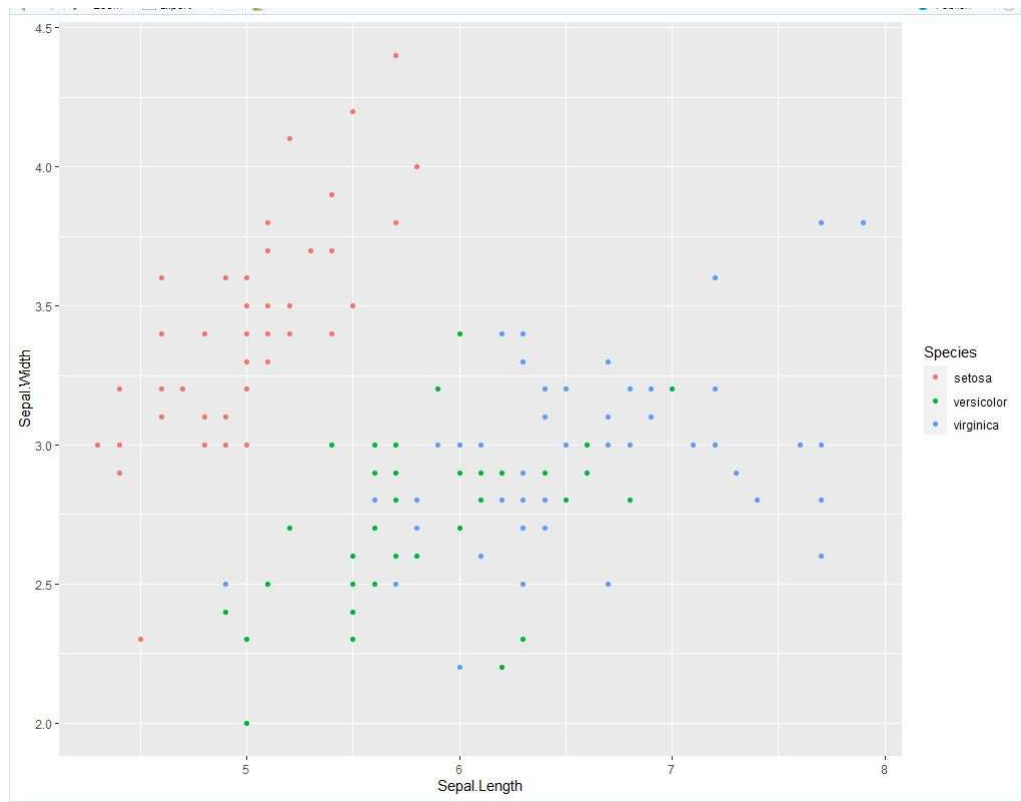
> library(class)
> library(ggplot2)
> library(GGally)
Registered S3 method overwritten by 'GGally':
  method from
+ .gg      ggplot2
>
> summary(iris)
      Sepal.Length      Sepal.width
Min.   :4.300   Min.   :2.000
1st Qu.:5.100   1st Qu.:2.800
Median :5.800   Median :3.000
Mean   :5.843   Mean   :3.057
3rd Qu.:6.400   3rd Qu.:3.300
Max.   :7.900   Max.   :4.400
      Petal.Length      Petal.width
Min.   :1.000   Min.   :0.100
1st Qu.:1.600   1st Qu.:0.300
Median :4.350   Median :1.300
Mean   :3.758   Mean   :1.199
3rd Qu.:5.100   3rd Qu.:1.800
Max.   :6.900   Max.   :2.500
      Species
setosa   :50
versicolor:50
virginica :50

> apply(iris[,1:4], 2, sd)
Sepal.Length Sepal.width Petal.Length
0.8280661    0.4358663    1.7652982
Petal.width
0.7622377
> par(mfrow=c(2,2))
> hist(iris$Sepal.Length, col="blue", breaks=20)
> hist(iris$Sepal.width, col="blue", breaks=20)
> hist(iris$Petal.Length, col="blue", breaks=20)
> hist(iris$Petal.width, col="blue", breaks=20)
> |

```

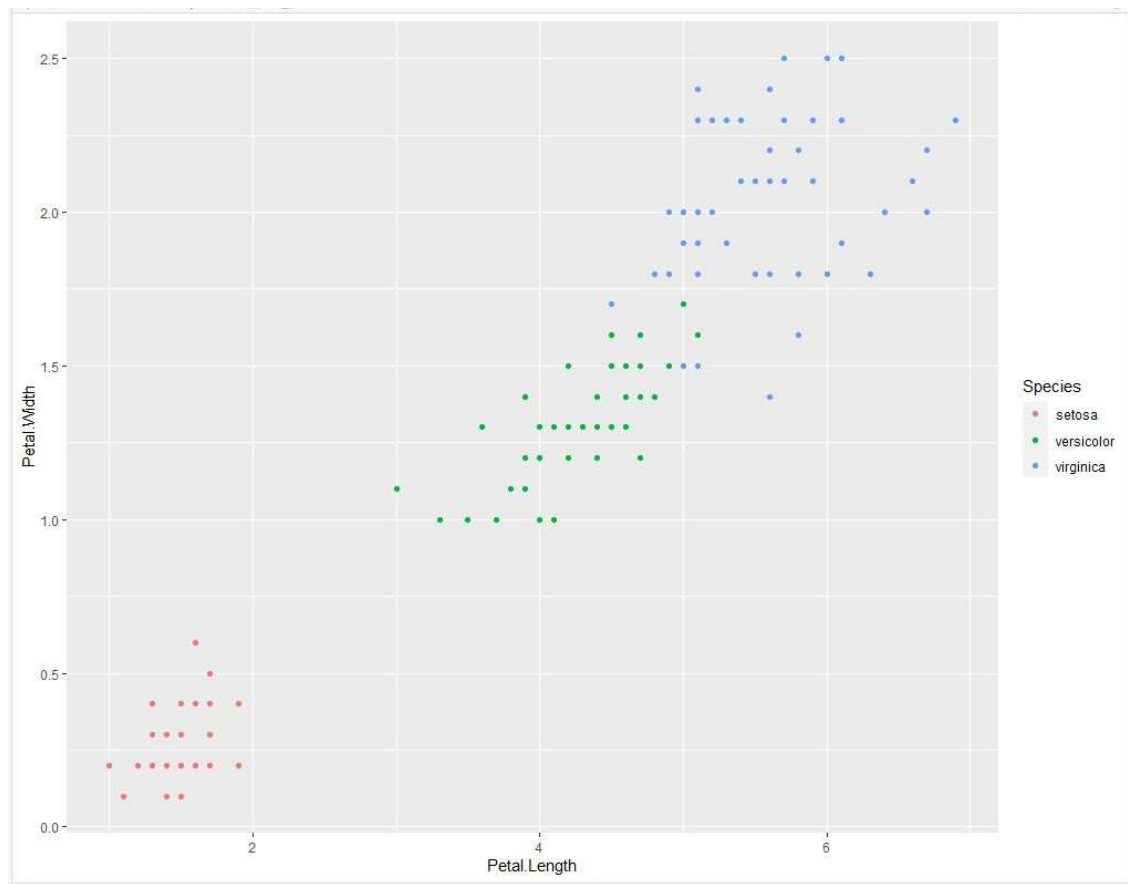




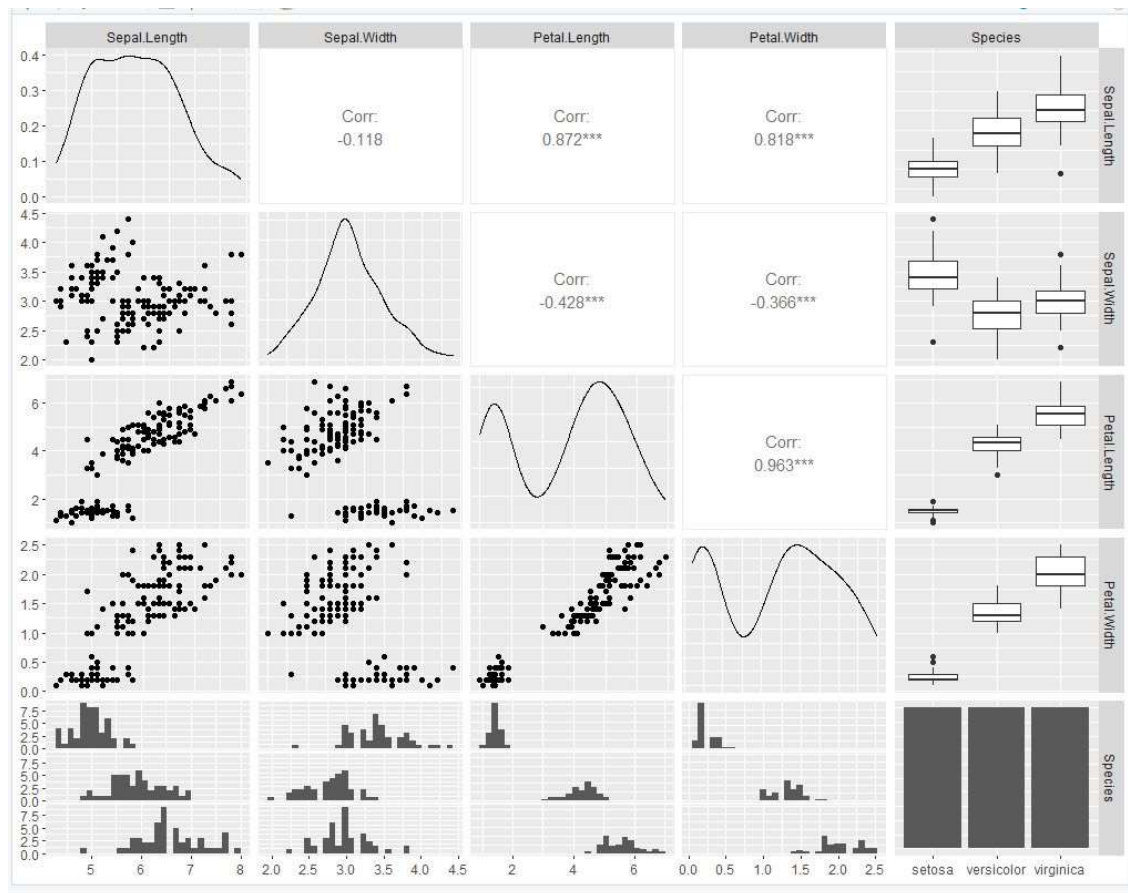


```
> ggplot(data = iris, aes(x = Sepal.Length, y = Sepal.Width, col = Species)) +  
+   geom_point()  
> |
```

```
> ggplot(data = iris, aes(x = Petal.Length, y = Petal.Width, col = Species)) +  
+   geom_point()  
> |
```



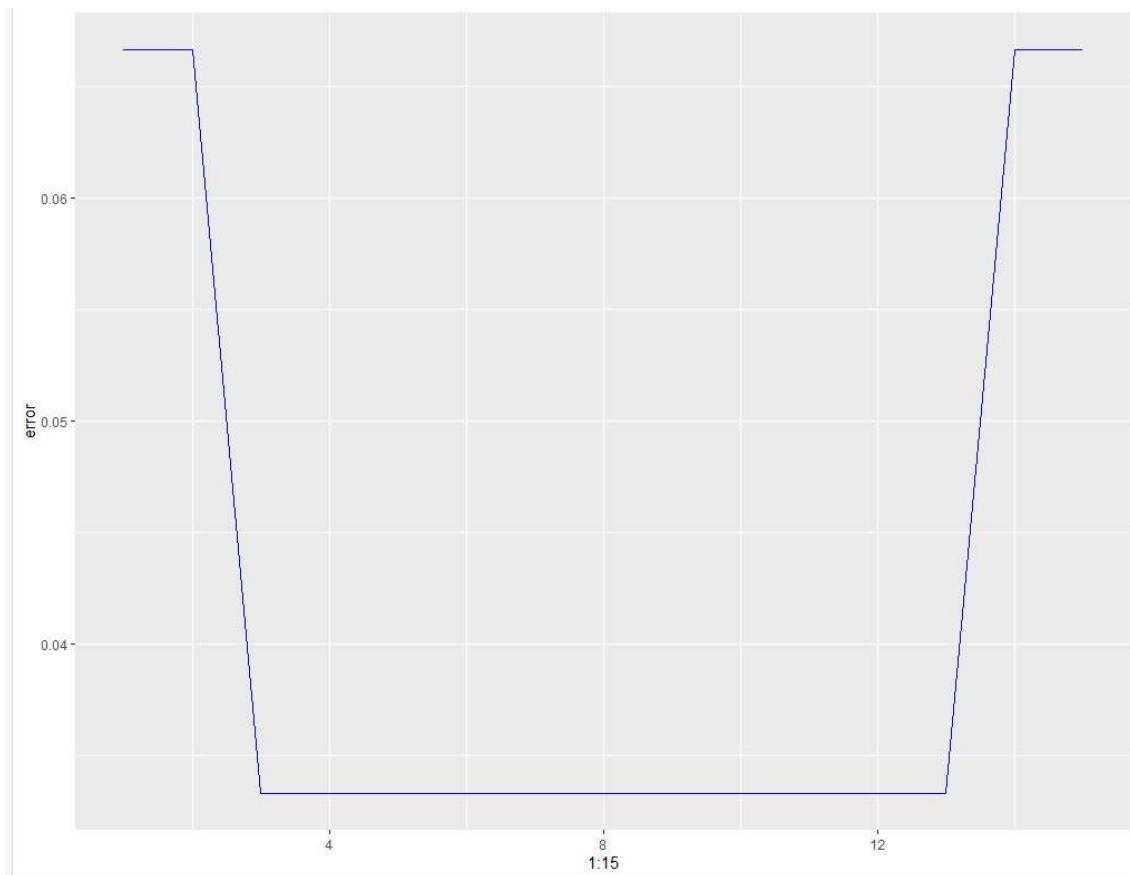
```
> ggpairs(iris)
plot: [5,1] [=====>---] 84% est: 0s `stat_bin()` using `bins = 30`. Pick better
value with `binwidth`.
plot: [5,2] [=====>--] 88% est: 0s `stat_bin()` using `bins = 30`. Pick better
value with `binwidth`.
plot: [5,3] [=====>-] 92% est: 0s `stat_bin()` using `bins = 30`. Pick better
value with `binwidth`.
plot: [5,4] [=====>-] 96% est: 0s `stat_bin()` using `bins = 30`. Pick better
value with `binwidth`.
> |
```



```

> set.seed(12420352)
> iris[,1:4] <- scale(iris[,1:4])
> setosa<- rbind(iris[iris$Species=="setosa",])
> versicolor<- rbind(iris[iris$Species=="versicolor",])
> virginica<- rbind(iris[iris$Species=="virginica",])
>
>
> ind <- sample(1:nrow(setosa), nrow(setosa)*0.8)
> iris.train<- rbind(setosa[ind,], versicolor[ind,], virginica[ind,])
> iris.test<- rbind(setosa[-ind,], versicolor[-ind,], virginica[-ind,])
> iris[,1:4] <- scale(iris[,1:4])
>
> error <- c()
> for (i in 1:15)
+ {
+   knn.fit <- knn(train = iris.train[,1:4], test = iris.test[,1:4], cl = iris.train$Species, k = i)
+   error[i] = 1- mean(knn.fit == iris.test$Species)
+ }
> ggplot(data = data.frame(error), aes(x = 1:15, y = error)) +
+   geom_line(color = "Blue")
>

```



```

> iris_pred <- knn(train = iris.train[,1:4], test = iris.test[,1:4], cl = iris.train$species, k=5)
> table(iris.test$species,iris_pred)
      iris_pred
      setosa versicolor virginica
setosa      10         0         0
versicolor   0         9         1
virginica     0         0        10
>

```

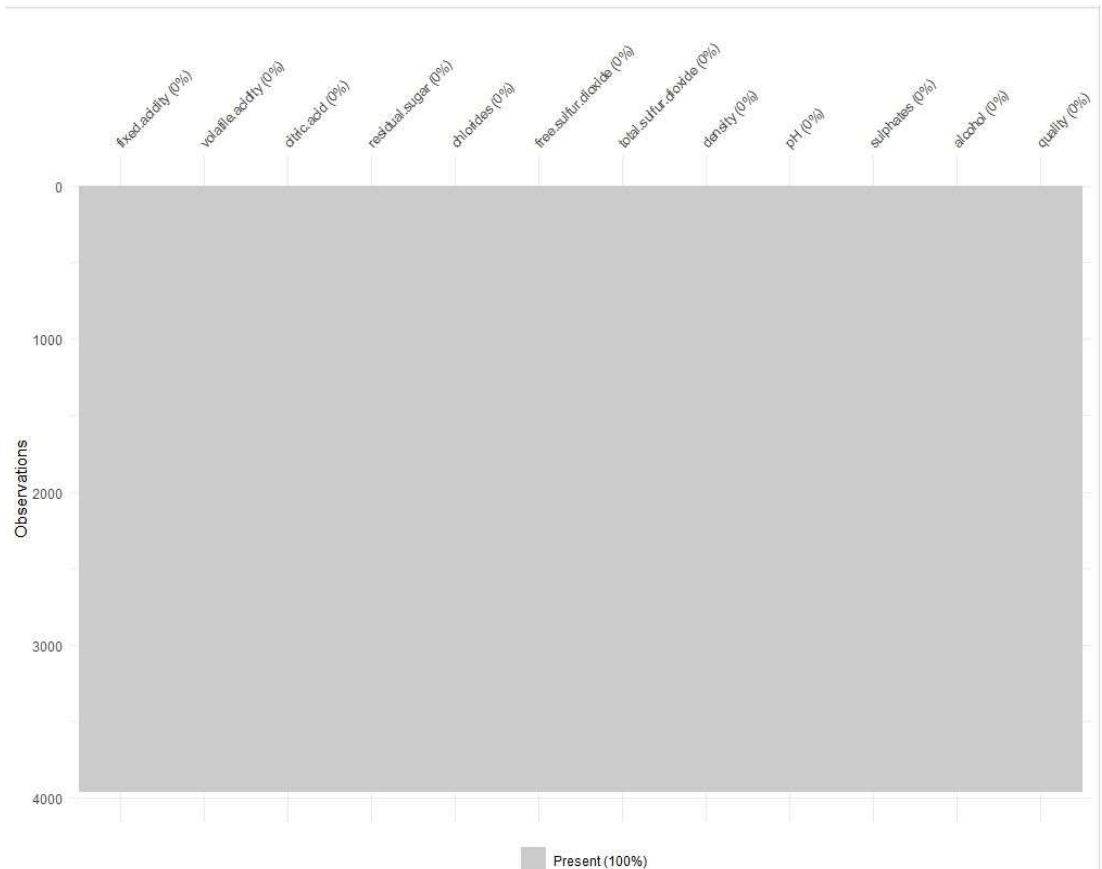
3. Use win equality dataset for all classification practice and use quality as predictor variable
  - a. Logistic regression



```

C:\Users\j\jessu\AppData\Local\Temp\kmpv\mxyg\downloaded_packages
> white_url <- "https://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequality-white.csv"
> whitewine <- read.csv(white_url, header = TRUE, sep = ";")
> white <- whitewine
> str(white)
'data.frame':   4898 obs. of  12 variables:
 $ fixed.acidity   : num  7 6.3 8.1 7.2 7.2 8.1 6.2 7 6.3 8.1 ...
 $ volatile.acidity : num  0.27 0.3 0.28 0.23 0.23 0.28 0.32 0.27 0.3 0.22 ...
 $ citric.acid     : num  0.36 0.34 0.4 0.32 0.32 0.4 0.16 0.36 0.34 0.43 ...
 $ residual.sugar  : num  20.7 1.6 6.9 8.5 8.5 6.9 7 20.7 1.6 1.5 ...
 $ chlorides       : num  0.045 0.049 0.05 0.058 0.058 0.05 0.045 0.045 0.049 0.044 ...
 $ free.sulfur.dioxide : num  45 14 30 47 47 30 30 45 14 28 ...
 $ total.sulfur.dioxide : num  170 132 97 186 186 97 136 170 132 129 ...
 $ density         : num  1.001 0.994 0.995 0.996 0.996 ...
 $ pH              : num  3 3.3 3.26 3.19 3.19 3.26 3.18 3 3.3 3.22 ...
 $ sulphates       : num  0.45 0.49 0.44 0.4 0.4 0.44 0.47 0.45 0.49 0.45 ...
 $ alcohol         : num  8.8 9.5 10.1 9.9 9.9 10.1 9.6 8.8 9.5 11 ...
 $ quality         : int  6 6 6 6 6 6 6 6 6 6 ...
> colnames(white)
[1] "fixed.acidity"      "volatile.acidity"    "citric.acid"
[4] "residual.sugar"     "chlorides"           "free.sulfur.dioxide"
[7] "total.sulfur.dioxide" "density"             "pH"
[10] "sulphates"         "alcohol"             "quality"
> summary(white)
fixed.acidity volatile.acidity citric.acid residual.sugar chlorides
Min.   : 3.800   Min.   :0.0800   Min.   :0.0000   Min.   : 0.600   Min.   :0.00900
1st Qu.: 6.300   1st Qu.:0.2100   1st Qu.:0.2700   1st Qu.: 1.700   1st Qu.:0.03600
Median : 6.800   Median :0.2600   Median :0.3200   Median : 5.200   Median :0.04300
Mean   : 6.855   Mean   :0.2782   Mean   :0.3342   Mean   : 6.391   Mean   :0.04577
3rd Qu.: 7.300   3rd Qu.:0.3200   3rd Qu.:0.3900   3rd Qu.: 9.900   3rd Qu.:0.05000
Max.   :14.200   Max.   :1.1000   Max.   :1.6600   Max.   :65.800   Max.   :0.34600
free.sulfur.dioxide total.sulfur.dioxide density pH sulphates
Min.   : 2.00   Min.   : 9.0   Min.   :0.9871   Min.   :2.720   Min.   :0.2200
1st Qu.:23.00   1st Qu.:108.0   1st Qu.:0.9917   1st Qu.:3.090   1st Qu.:0.4100
Median :34.00   Median :134.0   Median :0.9937   Median :3.180   Median :0.4700
Mean   :35.31   Mean   :138.4   Mean   :0.9940   Mean   :3.188   Mean   :0.4898
3rd Qu.:46.00   3rd Qu.:167.0   3rd Qu.:0.9961   3rd Qu.:3.280   3rd Qu.:0.5500
Max.   :289.00   Max.   :440.0   Max.   :1.0390   Max.   :3.820   Max.   :1.0800
alcohol quality
Min.   : 8.00   Min.   :3.000
1st Qu.: 9.50   1st Qu.:5.000
Median :10.40   Median :6.000
Mean   :10.51   Mean   :5.878
3rd Qu.:11.40   3rd Qu.:6.000
Max.   :14.20   Max.   :9.000
> white <- white[!duplicated(white), ]
> dim(white)
[1] 3961 12
> vis_miss(white)
Warning message:
'gather_()' was deprecated in tidy 1.2.0.
Please use 'gather()' instead.
The deprecated feature was likely used in the visdat package.
Please report the issue at <https://github.com/ropensci/visdat/issues>.

```



```

> sum(is.na(white))
[1] 0
> table(white$quality)
 3    4    5    6    7    8    9
20 153 1175 1788 689 131    5
> round(cor(white, method = "pearson"), 2)

```

	fixed.acidity	volatile.acidity	citric.acid	residual.sugar	chlorides	free.sulfur.dioxide	total.sulfur.dioxide	density	pH	sulphates	alcohol	quality
fixed.acidity	1.00											
volatile.acidity	-0.02	1.00										
citric.acid	0.30	-0.16	1.00									
residual.sugar		0.11	0.11	1.00								
chlorides		0.09	0.13	0.08	1.00							
free.sulfur.dioxide	-0.06	-0.10	0.09	0.31	0.10	1.00						
total.sulfur.dioxide	0.08	0.10	0.12	0.41	0.12	0.41	1.00					
density	0.27	0.06	0.16	0.82	0.16	0.82	0.06	1.00				
pH	-0.43	-0.05	-0.18	-0.17	-0.18	-0.17	-0.05	-0.17	1.00			
sulphates	-0.02	-0.02	0.05	-0.02	0.05	-0.02	-0.02	-0.02	-0.02	1.00		
alcohol	-0.11	0.05	-0.08	-0.40	-0.08	-0.40	-0.08	-0.40	-0.08	-0.40	1.00	
quality	-0.12	-0.19	0.01	-0.12	0.01	-0.12	0.01	-0.12	0.01	-0.12	0.01	1.00

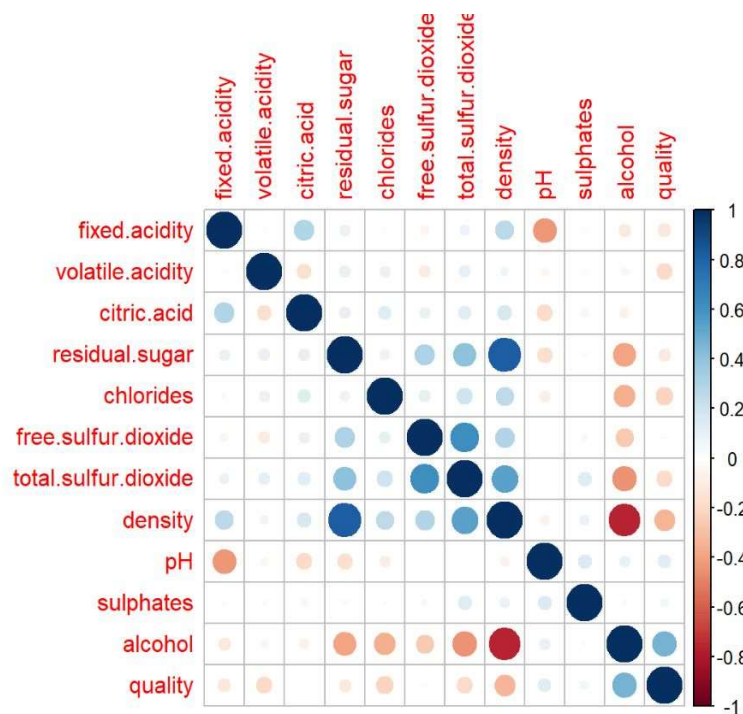
```


```

```

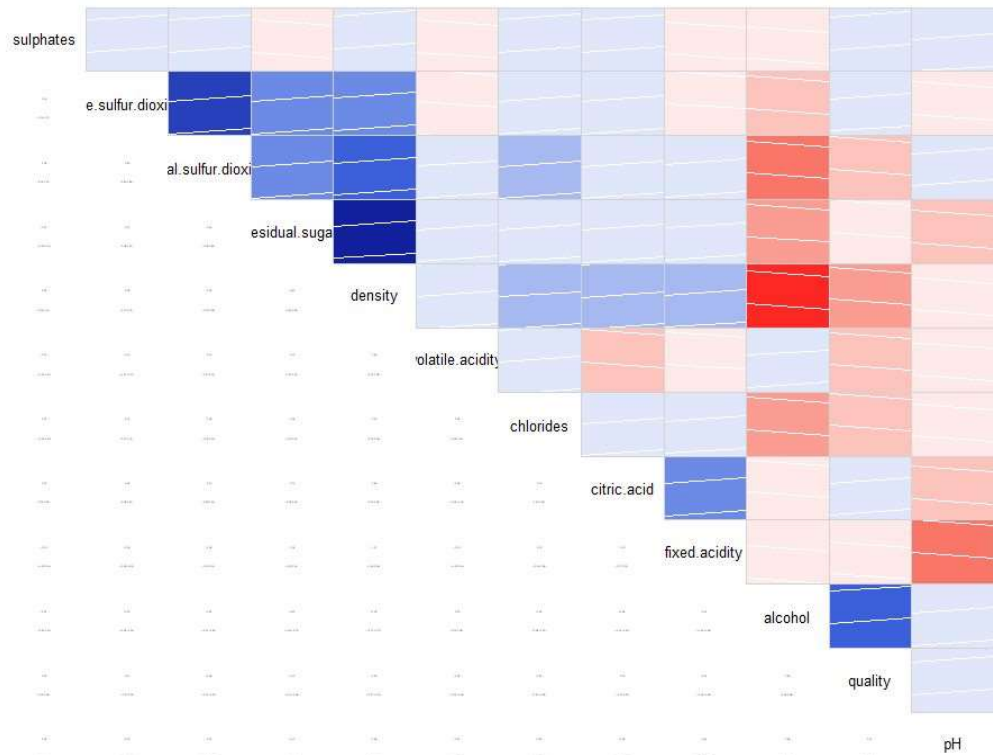
fixed.acidity      free.sulfur.dioxide
volatile.acidity  -0.06
citric.acid        0.09
residual.sugar     0.31
chlorides          0.10
free.sulfur.dioxide 1.00
total.sulfur.dioxide 0.62
density            0.29
pH                 -0.01
sulphates          0.04
alcohol            -0.25
quality            0.01
total.sulfur.dioxide
fixed.acidity      0.08
volatile.acidity   0.10
citric.acid        0.12
residual.sugar     0.41
chlorides          0.19
free.sulfur.dioxide 0.62
total.sulfur.dioxide 1.00
density            0.54
pH                 0.01
sulphates          0.14
alcohol            -0.45
quality            -0.18
density    pH    sulphates
fixed.acidity 0.27 -0.43 -0.02
volatile.acidity 0.06 -0.05 -0.02
citric.acid 0.16 -0.18 0.05
residual.sugar 0.82 -0.17 -0.02
chlorides 0.25 -0.09 0.02
free.sulfur.dioxide 0.29 -0.01 0.04
total.sulfur.dioxide 0.54 0.01 0.14
density 1.00 -0.06 0.08
pH -0.06 1.00 0.14
sulphates 0.08 0.14 1.00
alcohol -0.76 0.09 -0.02
quality -0.34 0.12 0.05
alcohol    quality
fixed.acidity -0.11 -0.12
volatile.acidity -0.05 -0.19
citric.acid -0.08 0.01
residual.sugar -0.40 -0.12
chlorides -0.36 -0.22
free.sulfur.dioxide -0.25 0.01
total.sulfur.dioxide -0.45 -0.18
density -0.76 -0.34
pH 0.09 0.12
sulphates -0.02 0.05
alcohol 1.00 0.46
quality 0.46 1.00
> corrpplot(cor(white))

```

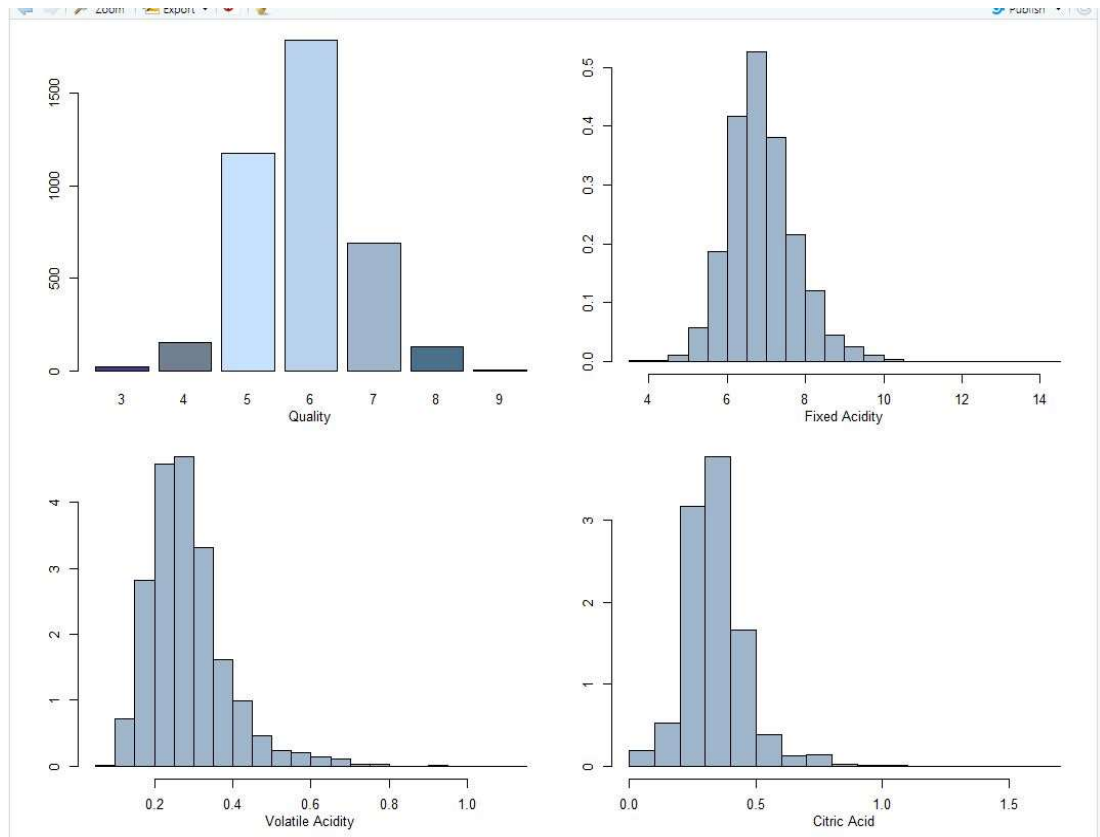


```
> corrgram(white, type="data", lower.panel=panel.conf,
+           upper.panel=panel.shade, main= "Corrgram for wine quality dataset", order=T, cex.labels=1.2)
There were 50 or more warnings (use warnings() to see the first 50)
> |
```

Corrgram for wine quality dataset

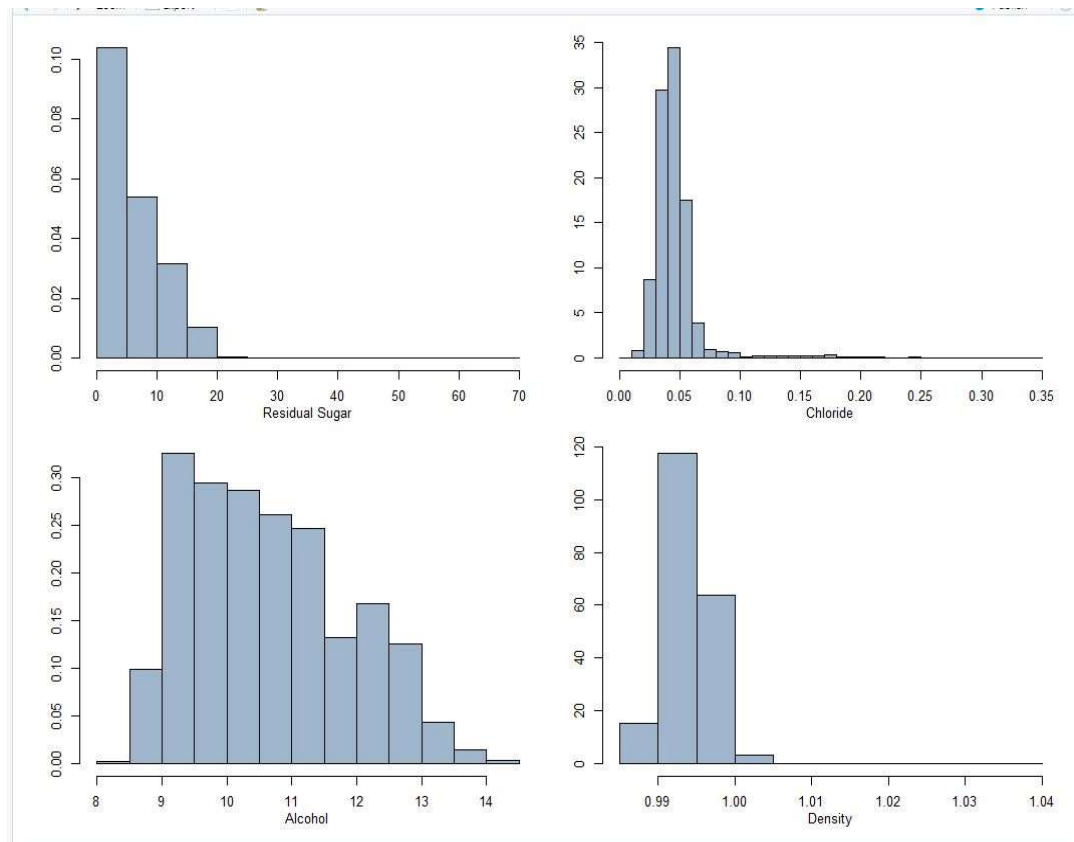


```
> attach(white)
>
> par(mfrow=c(2,2), oma = c(1,1,0,0) + 0.1, mar = c(3,3,1,1) + 0.1)
> barplot(table(quality), col=c("slateblue4", "slategray", "slategray1", "slategray2", "slategray3", "skyblue4"))
> mtext("Quality", side=1, outer=F, line=2, cex=0.8)
>
>
> truehist(fixed.acidity, h = 0.5, col="slategray3")
> mtext("Fixed Acidity", side=1, outer=F, line=2, cex=0.8)
>
> truehist(volatile.acidity, h = 0.05, col="slategray3")
> mtext("Volatile Acidity", side=1, outer=F, line=2, cex=0.8)
>
> truehist(citric.acid, h = 0.1, col="slategray3")
> mtext("Citric Acid", side=1, outer=F, line=2, cex=0.8)
>
```



```
> par(mfrow=c(2,2), oma = c(1,1,0,0) + 0.1, mar = c(3,3,1,1) + 0.1)
>
>
> truehist(residual.sugar, h = 5, col="slategray3")
> mtext("Residual Sugar", side=1, outer=F, line=2, cex=0.8)
>
> truehist(chlorides, h = 0.01, col="slategray3")
> mtext("Chloride", side=1, outer=F, line=2, cex=0.8)
>
> truehist(alcohol, h = 0.5, col="slategray3")
> mtext("Alcohol", side=1, outer=F, line=2, cex=0.8)
>
>
> truehist(density, h = 0.005, col="slategray3")
> mtext("Density", side=1, outer=F, line=2, cex=0.8)
>
```

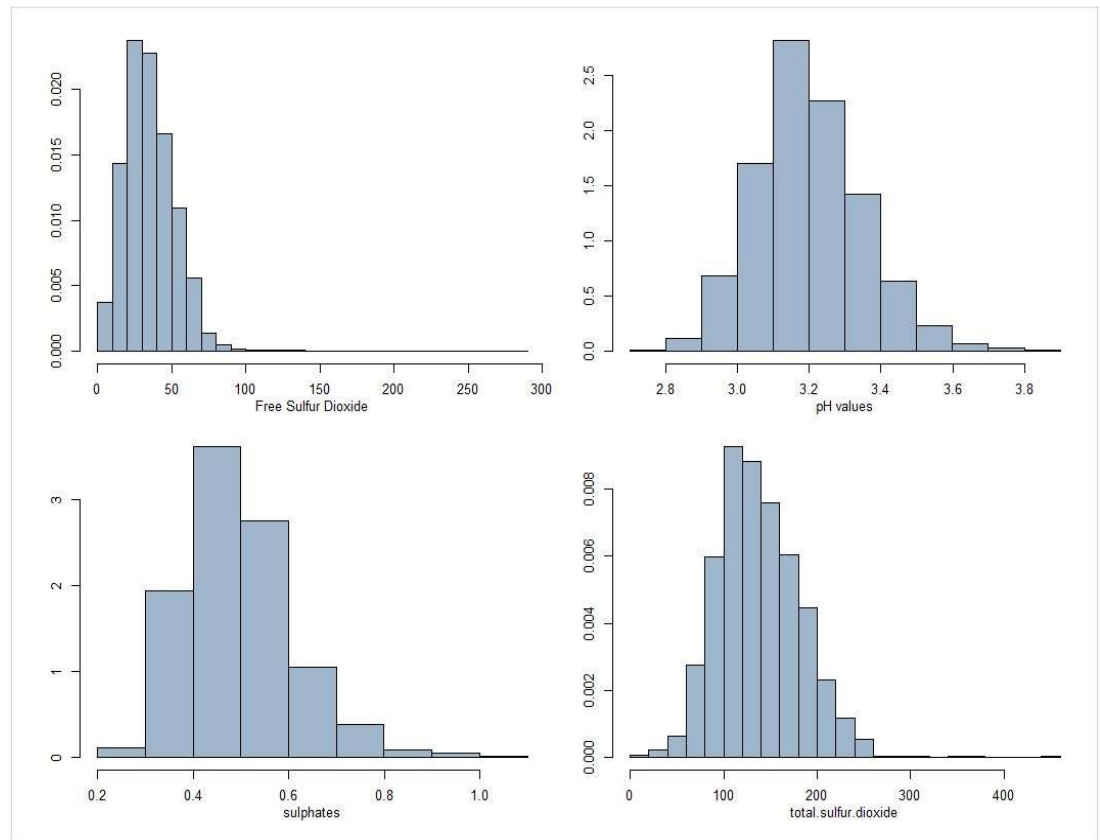




```

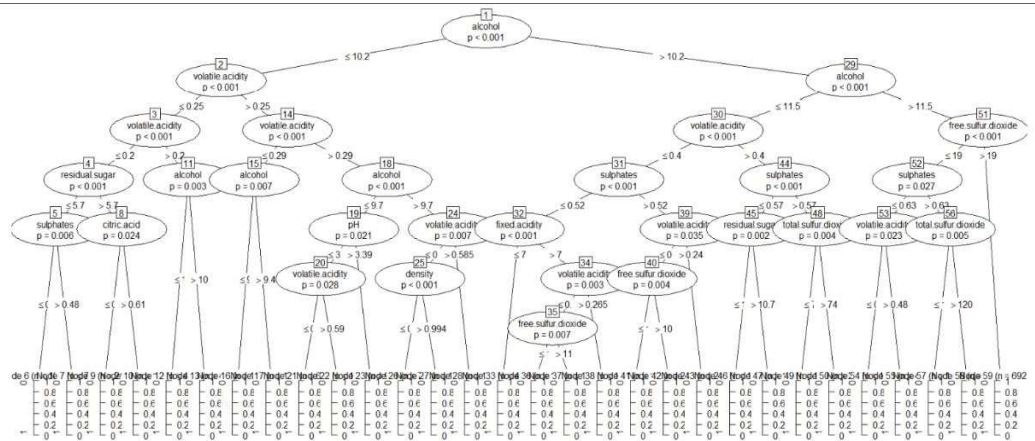
> par(mfrow=c(2,2), oma = c(1,1,0,0) + 0.1, mar = c(3,3,1,1) + 0.1)
> truehist(free.sulfur.dioxide, h = 10, col="slategray3")
> mtext("Free Sulfur Dioxide", side=1, outer=F, line=2, cex=0.8)
> truehist(pH, h = 0.1, col="slategray3")
> mtext("pH values", side=1, outer=F, line=2, cex=0.8)
> truehist(sulphates, h = 0.1, col="slategray3")
> mtext("sulphates", side=1, outer=F, line=2, cex=0.8)
> truehist(total.sulfur.dioxide, h = 20, col="slategray3")
> mtext("total.sulfur.dioxide", side=1, outer=F, line=2, cex=0.8)

```



## b. Decision trees

```
> output.tree <- ctree(
+   quality_bin ~ fixed.acidity + volatile.acidity + citric.acid + residual.sugar + chlorides + free.sulfur.dioxide + total.sulfur.dioxide + density + pH + sulphates + alcohol,
+   data = training_set)
> plot(output.tree)
>
```



### c. Support Vector Machines

```
> set.seed(3033)
> trainset <- createDataPartition(y = dataframe$quality, p = 0.7, list = FALSE)
> training <- dataframe[trainset,]
> testing <- dataframe[-trainset,]
> dim(training)
[1] 4549 12
> dim(testing)
[1] 1948 12
> dim(dataframe)
[1] 6497 12
> anyNA(dataframe)
[1] TRUE
> training <- training[complete.cases(training),]
> testing <- testing[complete.cases(testing),]
> dim(training)
[1] 4531 12
> dim(testing)
+ dim(testing)
Error: unexpected symbol in:
"dim(testing
dim"
> dim(testing)
[1] 1932 12
> anyNA(training)
[1] FALSE
> anyNA(testing)
[1] FALSE
> summary(dataframe)
fixed.acidity    volatile.acidity    citric.acid    residual.sugar    chlorides
Min.   : 3.800    Min.   :0.0800    Min.   :0.0000    Min.   : 0.600    Min.   :0.00900
1st Qu.: 6.400    1st Qu.:0.2300    1st Qu.:0.2500    1st Qu.: 1.800    1st Qu.:0.03800
Median : 7.000    Median :0.2900    Median :0.3100    Median : 3.000    Median :0.04700
Mean   : 7.217    Mean   :0.3397    Mean   :0.3187    Mean   : 5.444    Mean   :0.05604
3rd Qu.: 7.700    3rd Qu.:0.4000    3rd Qu.:0.3900    3rd Qu.: 8.100    3rd Qu.:0.06500
Max.   :15.900    Max.   :1.5800    Max.   :1.6600    Max.   :65.800    Max.   :0.61100
NA's   :10       NA's   :8       NA's   :3       NA's   :2       NA's   :2
free.sulfur.dioxide    total.sulfur.dioxide    density    pH    sulphates
Min.   : 1.00    Min.   : 6.0    Min.   :0.9871    Min.   :2.720    Min.   :0.2200
1st Qu.: 17.00    1st Qu.: 77.0    1st Qu.:0.9923    1st Qu.:3.110    1st Qu.:0.4300
Median : 29.00    Median :118.0    Median :0.9949    Median :3.210    Median :0.5100
Mean   : 30.53    Mean   :115.7    Mean   :0.9947    Mean   :3.218    Mean   :0.5312
3rd Qu.: 41.00    3rd Qu.:156.0    3rd Qu.:0.9970    3rd Qu.:3.320    3rd Qu.:0.6000
Max.   :289.00    Max.   :440.0    Max.   :1.0390    Max.   :4.010    Max.   :2.0000
NA's   :9       NA's   :4
alcohol    quality
Min.   : 8.00    Min.   :3.000
1st Qu.: 9.50    1st Qu.:5.000
Median :10.30    Median :6.000
Mean   :10.49    Mean   :5.818
3rd Qu.:11.30    3rd Qu.:6.000
Max.   :14.90    Max.   :9.000
```

```

> training[["quality"]] = factor(training[["quality"]])
> trctrl <- traincontrol(method = "repeatedcv", number = 10, repeats = 3)
> svm_linear <- train(quality~., -density -fixed.acidity -citric.acid, data = training, method = "svmLinear",
+   trControl = trctrl, preProcess = c("center", "scale"), tuneLength = 10)
1 package is needed and is not installed. (kernlab). would you like to try to install it now?
1: yes
2: no

Selection: svm_linear
Enter an item from the menu, or 0 to exit
Selection: yes
trying URL 'https://cran.rstudio.com/bin/windows/contrib/4.2/kernlab_0.9-31.zip'
Content type 'application/zip' length 2517462 bytes (2.4 MB)
downloaded 2.4 MB

package 'kernlab' successfully unpacked and MD5 sums checked

The downloaded binary packages are in
  C:\Users\sbkum\AppData\Local\Temp\RtmpSPziko\downloaded_packages
Error in train.default(x, y, weights = w, ...) : object 'trctrl' not found
> svm_linear <- train(quality~., -density -fixed.acidity -citric.acid, data = training, method = "svmLinear",
+   trControl = trctrl, preProcess = c("center", "scale"), tuneLength = 10)
Error in train.default(x, y, weights = w, ...) : object 'trctrl' not found
> svm_linear <- train(quality~., -density -fixed.acidity -citric.acid, data = training, method = "svmLinear",
+   trControl = trctrl, preProcess = c("center", "scale"), tuneLength = 10)
> svm_linear
Support Vector Machines with Linear Kernel

4531 samples
  11 predictor
  7 classes: '3', '4', '5', '6', '7', '8', '9'

Pre-processing: centered (8), scaled (8)
Resampling: Cross-validated (10 fold, repeated 3 times)
Summary of sample sizes: 4078, 4078, 4079, 4078, 4078, 4078, ...
Resampling results:

  Accuracy   Kappa
0.5415293  0.2357962

Tuning parameter 'C' was held constant at a value of 1

```

```

> test_pred <- predict(svm_linear,newdata = testing)
> test_pred
[1] 5 6 5 6 6 5 5 6 6 5 5 6 6 6 6 6 6 5 5 6 6 5 5 6 5 5 6 6 6 5 5 6 6 5 5 6 6 5 5 6 6 6 6 5 5 5
[46] 6 6 5 5 5 6 5 5 6 5 5 5 6 6 6 5 6 6 6 6 5 6 5 5 5 6 6 6 6 6 6 5 5 6 6 6 6 6 6 5 6 5 5 5 6 6
[91] 6 5 5 6 5 6 6 6 6 6 6 6 6 6 6 6 6 6 5 6 6 5 6 6 6 5 6 6 6 6 6 5 6 6 6 6 6 5 6 6 5 6 6 5 6
[136] 6 5 6 6 5 6 6 5 5 6 6 6 6 5 6 6 6 6 5 6 6 6 6 5 5 6 5 6 5 6 6 6 5 5 6 5 6 5 5 6 6 5 5
[181] 6 6 5 5 6 6 6 6 6 5 6 5 5 5 6 5 5 6 5 5 6 6 6 5 5 6 6 6 5 5 6 6 6 5 6 6 5 6 6 5 6 6 6 5
[226] 5 5 5 5 6 6 5 5 6 5 6 6 6 6 6 6 6 6 6 6 6 6 5 5 6 5 5 6 6 6 6 6 6 6 6 6 6 6 5 5 6 5 6 5
[271] 6 6 5 5 5 5 5 6 6 5 5 6 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 5 5 6 5 6 6 5 6
[316] 5 6 6 5 5 6 5 6 6 6 6 5 6 6 6 6 5 5 6 6 6 6 5 5 5 6 6 6 6 5 5 6 5 6 6 5 6 5 5 5 5 6 6
[361] 6 6 6 5 6 6 6 5 5 6 6 5 6 5 5 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 5 6 5 6 6 5
[406] 6 5 6 5 6 5 5 6 6 5 6 6 5 6 6 6 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6
[451] 6 5 6 6 5 5 6 5 6 6 5 6 6 6 5 5 6 5 6 6 5 6 6 6 5 6 6 6 6 6 6 6 6 6 5 6 6 6 5 6 6 6 5 6
[496] 6 5 6 6 6 6 5 6 5 6 6 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 5 6 6 6 5
[541] 5 6 6 5 6 5 5 6 5 6 6 6 6 5 5 6 6 6 6 6 5 5 5 6 6 6 6 5 6 6 5 5 5 6 6 5 6 5 6 5 5 5 6 6
[586] 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 5 6 6 6 5 6 6 6 6 6 6 6 5 6 6 6 6 6 5 6 6 6 5
[631] 5 5 5 6 5 5 5 6 6 6 6 6 5 6 6 6 6 6 6 6 6 6 5 5 6 5 5 5 6 6 6 5 5 5 6 6 6 5 5 6 6 6 6 5 5
[676] 6 6 6 6 5 6 6 6 6 6 6 6 6 6 5 5 6 5 6 6 5 6 6 6 5 6 6 6 5 6 6 6 5 6 6 5 5 5 5 5 5 5 6 6 6 5
[721] 6 5 6 5 6 5 6 6 6 5 6 6 6 5 6 6 6 6 5 6 6 5 6 6 5 6 6 5 6 5 6 6 5 6 5 5 5 6 6 5 6 6 6 6
[766] 6 6 5 6 5 6 6 5 6 5 5 6 5 6 6 6 5 6 6 5 6 6 6 6 6 6 5 6 5 6 6 6 5 6 6 6 5 5 6 6 6 6 6 6 6
[811] 6 6 6 6 6 6 6 5 6 6 6 6 6 6 6 5 6 6 6 6 6 6 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6
[856] 6 6 5 6 6 6 6 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6
[901] 6 6 6 6 6 6 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6
[946] 5 6 6 6 6 6 6 6 6 6 6 5 6 6 5 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6
[991] 6 5 5 6 5 6 6 6 6 5
[ reached getOption("max.print") -- omitted 932 entries ]
Levels: 3 4 5 6 7 8 9
> confusionMatrix(table(test_pred,testing$quality))
Confusion Matrix and Statistics

test_pred   3    4    5    6    7    8    9
3      0    0    0    0    0    0    0
4      0    0    0    0    0    0    0
5      9   41  364  197   25    5    0
6      1   29  263  648  300   49    1
7      0    0    0    0    0    0    0
8      0    0    0    0    0    0    0
9      0    0    0    0    0    0    0

Overall Statistics

          Accuracy : 0.5238
          95% CI   : (0.5013, 0.5463)
    No Information Rate : 0.4374
    P-Value [Acc > NIR] : 1.475e-14

          Kappa : 0.2064

  Mcnemar's Test P-Value : NA

Statistics by Class:

Statistics by Class:

              Class: 3 Class: 4 Class: 5 Class: 6 Class: 7 Class: 8 Class: 9
Sensitivity    0.000000  0.000000  0.5805   0.7669   0.0000  0.000000  0.00000000
Specificity    1.000000  1.000000  0.7877   0.4085   1.0000  1.000000  1.00000000
Pos Pred Value      NaN      NaN    0.5679   0.5019      NaN      NaN      NaN

```

#### d. Naive Bayes Classifier



```

> nb = naiveBayes(quality~, -density -fixed.acidity -citric.acid,data = training)
> nb

Naive Bayes Classifier for Discrete Predictors

Call:
naiveBayes.default(x = X, y = Y, laplace = laplace)

A-priori probabilities:
Y
      3      4      5      6      7      8      9
0.0044140366 0.0317810638 0.3312734496 0.4358861179 0.1653056720 0.0304568528 0.0008828073

Conditional probabilities:
volatile.acidity
Y      [,1]      [,2]
3 0.4557500 0.3527832
4 0.4402083 0.2117839
5 0.3908195 0.1755336
6 0.3131899 0.1475216
7 0.2881375 0.1153739
8 0.2865580 0.1122852
9 0.3125000 0.0550000

residual.sugar
Y      [,1]      [,2]
3 5.460000 4.735771
4 4.094792 3.748761
5 5.712692 4.885421
6 5.559291 4.966949
7 4.870427 4.134139
8 5.251087 4.174748
9 4.650000 4.119466

chlorides
Y      [,1]      [,2]
3 0.06290000 0.046070769
4 0.06059722 0.054626042
5 0.06463891 0.043319436
6 0.05404759 0.030506095
7 0.04559947 0.021779886
8 0.04175362 0.016707362
9 0.02650000 0.008266398

free.sulfur.dioxide
Y      [,1]      [,2]
3 50.25000 70.88584
4 20.40972 15.89538
5 30.12592 18.47110
6 30.95038 16.99722
7 30.54272 14.84191
8 34.71014 17.48877
9 35.00000 14.94434

```



	total.sulfur.dioxide	
Y	[,1]	[,2]
3	131.2750	110.22172
4	105.9722	61.14124
5	120.7761	59.99025
6	115.3362	55.92453
7	108.6228	48.56090
8	113.8696	39.45460
9	110.2500	17.42364

	pH	
Y	[,1]	[,2]
3	3.259000	0.19522996
4	3.206181	0.17347133
5	3.212139	0.15551385
6	3.219362	0.16088250
7	3.223632	0.16387651
8	3.222174	0.15720524
9	3.315000	0.09398581

	sulphates	
Y	[,1]	[,2]
3	0.4980000	0.1262245
4	0.5072917	0.1830939
5	0.5267355	0.1467288
6	0.5326025	0.1446391
7	0.5526836	0.1639743
8	0.5088406	0.1632207
9	0.4625000	0.1065755

	alcohol	
Y	[,1]	[,2]
3	10.38000	1.1967500
4	10.16146	0.9930126
5	9.84006	0.8175453
6	10.58143	1.1237238
7	11.33820	1.1967099
8	11.69638	1.2761375
9	12.10000	1.1518102

```
> pred_nb = predict(nb,testing)
> confusionMatrix(table(pred_nb,testing$quality))
Confusion Matrix and Statistics
```

```
> pred_nb = predict(nb,testing)
> confusionMatrix(table(pred_nb,testing$quality))
Confusion Matrix and Statistics
```

pred_nb	3	4	5	6	7	8	9
3	1	4	4	1	1	0	0
4	1	7	18	10	5	1	0
5	6	22	267	154	10	4	0
6	2	32	318	501	146	23	0
7	0	5	20	178	163	26	1
8	0	0	0	1	0	0	0
9	0	0	0	0	0	0	0

overall Statistics

```
> pred_nb = predict(nb,testing)
> confusionMatrix(table(pred_nb,testing$quality))
Confusion Matrix and Statistics
```

pred_nb	3	4	5	6	7	8	9
3	1	4	4	1	1	0	0
4	1	7	18	10	5	1	0
5	6	22	267	154	10	4	0
6	2	32	318	501	146	23	0
7	0	5	20	178	163	26	1
8	0	0	0	1	0	0	0
9	0	0	0	0	0	0	0

overall Statistics

Accuracy : 0.486  
 95% CI : (0.4635, 0.5086)  
 No Information Rate : 0.4374  
 P-value [Acc > NIR] : 9.564e-06

Kappa : 0.2163

McNemar's Test P-value : NA

Statistics by Class:

	Class: 3	Class: 4	Class: 5	Class: 6	Class: 7	Class: 8	Class: 9
Sensitivity	0.1000000	0.100000	0.4258	0.5929	0.50154	0.0000000	0.0000000
Specificity	0.9947971	0.981203	0.8498	0.5207	0.85688	0.9994675	1.0000000
Pos Pred value	0.0909091	0.166667	0.5767	0.4902	0.41476	0.0000000	NaN
Neg Pred value	0.9953149	0.966667	0.7549	0.6220	0.89474	0.9720352	0.9994824
Prevalence	0.0051760	0.036232	0.3245	0.4374	0.16822	0.0279503	0.0005176
Detection Rate	0.0005176	0.003623	0.1382	0.2593	0.08437	0.0000000	0.0000000
Detection Prevalence	0.0056936	0.021739	0.2396	0.5290	0.20342	0.0005176	0.0000000
Balanced Accuracy	0.5473985	0.540602	0.6378	0.5568	0.67921	0.4997338	0.5000000

e. k-Nearest Neighbour

