



Antidiabetic potential of some less commonly used plants in traditional medicinal systems of India and Nigeria

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ABSTRACT

The incidence of diabetes mellitus continue to rise annually all over the world with India and Nigeria having recorded cases of 65.1 and 3.9 million respectively in 2013 and expected to increase by a large amount in 2035. Hyperglycemia is a pre-condition for the development of diabetic complications and is accompanied by an increase in the production of free radicals. The present available treatment option for diabetes like sulfonylurea, metformin and alpha-glucosidase are restricted by their limited actions, secondary failure rates, and side-effects; and unaffordable to the majority of the population. Hence, the need to screen for more medicinal plants with antidiabetic ability due to the fact that plants are; biodegradable, safe and cheap with fewer side-effects. In this review article, we have presented the current status of diabetes in India and Nigeria and the role of some less commonly used medicinal plants from both countries that have antidiabetic potential.

KEY WORDS: Antidiabetic plants, diabetes, hypoglycaemic activity, medicinal plants, oxidative stress

INTRODUCTION

The World Health Organization (WHO) defines diabetes mellitus (DM) as a degenerative and chronic disease that occurs when the pancreas does not produce enough insulin, or when the body cannot effectively use insulin [1]. It is a disorder of the metabolism of carbohydrates, fats, and lipids, which is characterized by a high fasting blood sugar [2]. It manifests as chronic hyperglycemia and leads to the development of diabetes-specific micro vascular pathology in the retina, glomerulus and peripheral nerve culminating into serious complications affecting the eyes, kidneys and arteries [3,4].

WHO statistics shows that worldwide 347 million people have diabetes and 80% of diabetic deaths occur in low and middle-income countries [1]. According to the International Diabetes Federation, India is ranked second only to China in the list of top ten countries for a number of people with diabetes. [5]. In Africa, it is estimated that about 19.8 million adults have diabetes with Nigeria and South Africa having 3.9 and 2.6 million, respectively. It is estimated that by 2035, the percentage of diabetic patients in Africa would cross an alarming figure of 58% [5].

Type 2 diabetes, is the major form of diabetes accounting for 90-95% of all diabetic cases [6] and nearly half of all patients suffering from the disease are older than 65 years of age [7]. It is a complicated and divergent disease which in addition to blood sugar control requires the management of lipid parameters, blood pressure and thrombotic factors [8].

The treatment for diabetes is both difficult and tedious; it is expensive, costly and not affordable by majority of African and Asian populations [9]. The current treatments for DM include the use of insulin and synthetic drugs such as sulfonylurea, metformin, alpha-glucosidase inhibitors and thiazolidinedione's in addition to lifestyle adjustments. These synthetic drugs are valuable but restricted by their limited action, pharmacokinetic properties, secondary failure rates and accompanying side-effects like hypoglycemia, damage to liver, lactic acidosis, diarrhea, abdominal pain, weight loss and loss of appetite [7,10-12].

Due to the problems associated with the current treatments, a large percentage of diabetics resort to alternative remedies that are purported to improve glycemic control [8]. The WHO estimated that approximately 80% of the world's population rely mainly on traditional medicines for their primary health care [13]. The screening of medicinal plants for novel bioactive compounds is, therefore, an important goal for scientists. Importantly, the plant based drugs are biodegradable, safe, and cheap, having fewer side-effects, in India, China and other ancient traditional medicinal systems in the world, medicinal plants have been the major source of treatment for DM since time immemorial [14-16].

The importance of research on medicinal plants is validated by the fact that a plethora of new drugs have been developed from plants; relevant examples include cromolyn used as bronchodilator, developed from *Ammi visnaga* (L) Lamk;

galegine, from *Galega officinalis* L, which is a model for the synthesis of metformin and other bisguanidine-type antidiabetic drugs, papaverine from *Papaver somniferum* which forms the basis of cerapramil used in the treatment of hypertension, [17]. *Artemisia annua* (Quinhaosu) gave rise to artemisinin, this compound and its analogs are now used as antimalarial therapy in many countries [18]. Paclitaxel (Taxol®), the most exciting plant-derived anticancer drug discovered in recent years, is derived from several key precursors (the baccatins) in the leaves of various *Taxus* species; *Taxus brevifolia* [19].

Although the role of natural product in new drug discovery is encouraging and has frequently resulted in development of new drugs [20], the success of drug discovery depends on evolving stringent criteria to avoid false positive drug candidates. Surfeit of information warrants proper documentation. The evaluation of the scientific efficacy of traditional systems of medicine is an area of great interest especially in developing economies where sometimes the cost of medication may be prohibitive. Excellent reviews [13,21-23] on antidiabetic plants have already been written. This current review aims to bring in focus and document the use of less commonly used antidiabetic plants on which fewer studies have been conducted. Some of these plants, albeit less researched, hold immense potential as antidiabetic therapeutic agents in India and Nigeria.

ANTIDIABETIC PLANTS USED IN NIGERIA AND INDIA

The climatic conditions in Nigeria and India support the growth and thriving of various plant species and hence the use of these plants by the poor population to ameliorate disease conditions. There are about 800 plants that may possess antidiabetic properties according to ethno botanical information [24]. Most of the current drugs available have been directly or indirectly derived from plants. An example is metformin that was derived from the plant *G. officinalis* L.

The plants with antioxidant and antidiabetic potential included in this review are *Azadirachta indica* (AI) A. Juss, *Mangifera indica* (MI) L, *Terminalia arjuna* Roxb. Ex DC, *Terminalia catappa* L, *Terminalia chebula* Retz, *Syzygium cumini* (L) Skeels, *Syzygium aromaticum* (L) Merr. and L.M. Perry, *Vernonia amygdalina* (VA) Delile and *Xylopia aethiopica* (XA) (Dunal) A. Rich.

The general botanical data, taxonomic data, distribution in the world, experimental design, compounds isolated, mechanism of action, the antidiabetic and antioxidant capability of the plants are presented below:

Mangifera indica L. (Common Name: Mango)

Mango is an important species of the family anacardiaceae and the genus *Mangifera*, it is native to South East Asia from where it spread all over the world, it is the most popular fruit in the tropical and subtropical regions of the world. It is the national fruit of India, Pakistan, Philippines and the national tree of Bangladesh [25].

The plant is widely grown in Nigeria, where in addition to the fruit consumption it is used for the treatment and management of diabetes [26]. The peel and pulp of the plant contain carotenoids, and polyphenols such as quercetin, kaempferol, gallic acid, caffeic acid, catechins, tannins, mangiferin, leucocyanidin, epicatechin, quercetin and chromogenic acid [27]. Phenolics have scavenging activity on free radicals mainly due to the presence of hydroxyl groups. Recently, Mohan *et al.* [28] isolated a compound 1, 2, 3, 4, 6-penta-O-gallolyl- β -D-glucose from the methanolic extract fraction of mango that is a potent inhibitor of 11- β -hydroxysteroid hydrogenase enzyme and ameliorates high fat diet (HFD) induced diabetes in C57BL/6 mice.

Mangiferin (1, 3, 6, 7-tetrahydroxy-xanthone-C2- β -D-glucoside) a bioactive compound isolated from MI possesses a wide range of pharmacological actions including being anticancer [29,30], antibacterial [31], anti HIV [32], antioxidants [33], and antidiabetic [34,35].

The administration of mangiferin at a dose of 10 and 20 mg/Kg body weight (i.p.) in type 1 and 2 diabetic rats for 30 days showed significant antidiabetic, hypo-lipidemic, alpha amylase and alpha-glucosidase inhibitory effect [36]. This glucoside has also been shown by Li *et al.* [37] to improve renal function of diabetic nephropathy in rats and its inhibitory effect on overexpression of transforming growth factor- β 1, advanced glycation end and extracellular matrix accumulation, Polyol pathway activation, reactive oxygen species (ROS) generation and mesangial cells proliferation. Miura *et al.* [38] demonstrated that the mangiferin exerts its antidiabetic activity by decreasing the insulin resistance.

The ethanolic extracts of MI showed significant free radical scavenging activity and have cytoprotective (anti-apoptotic) effect; the leaves and fruits extract reduce the absorption of glucose in type 2 diabetes and stimulate glycogenesis in liver causing reduction in blood glucose level [39].

Vernonia amygdalina Delile (Asteraceae)

VA is a perennial shrub-like plant with green leaves growing up to 1.3-3 m high that is native to Africa, widely grown in Nigeria and West Africa. It is reported to contain phytochemicals useful in the treatment and management of certain diseases. It has been introduced into India and is now being cultivated in parts of central and eastern India [40].

VA is rich in amino acids, minerals and vitamins [41]. The decoction from the leaves is often used in the African traditional treatment for the management of diabetes, malaria, infertility, and sexually transmitted diseases [42-47]. The plant is said to have antimalarial compounds like alkaloids, tannins, and saponins [48] and also anticancer properties [49]. In comparison to other plants, VA accounted for 9.2% of medicinal plants used as an alternative medicine in central Nigeria [50]. In Nigeria, a dosage form of freeze-dried aqueous leaf extract of this plant has been developed and formulated, which is suitable for therapeutic use in the management of DM. Mostly in Nigeria, the decoction

from the leaf is often used in combination with that of other plants by traditional healers and medical practitioners to treat diabetes, fever and gastrointestinal problems [51].

The ethanolic extracts of the plant has a strong bioactive compound that has blood sugar lowering action in rats and can serve as an effective antioxidant [52]. Ong *et al.* [53] showed that VA has anti-hyperglycemic effect on streptozotocin (STZ)-induced diabetic rat model and this effect is mediated through the inhibition of key hepatic G6pase, which causes an increase in expression and translocation of GLUT4 in skeletal muscles. The combined leaf extract of *A. indica* (AI) and VA ameliorates hyperglycemia and hepatic oxidative stress in diabetic rats [54] and the methanolic extract of VA has the ability to mitigate cycasin-induced oxidative damage in colonic tissues [55].

The composite decoctions of VA, *Gongronema latifolium* (Benth) and *Occimum gratissimum* (Linn) reduced the postprandial blood glucose concentrations of diabetic subjects [56]. Two flavonoids and terpenoids: Vernolide and edotides have been isolated from the VA plant. Octahydrovernodalin is the most important bitter principle in the plant [57]. Ong *et al.* [58] also isolated four main polyphenols in the ethanolic extract namely dicaffeoyl-quinic acid, chlorogenic acid, 1,5-dicaffeoyl-quinic acid and luteolin-7-O-glucosidase. Dicaffeoyl-quinic acid is the most abundant in the plant. The administration of 400 mg/Kg body weight of VA extract is found to exert most effective anti-hyperglycemic activity [59].

The two major glucose transporters that regulate glucose uptake into the tissues are GLUT1 (non-insulin responsive) and GLUT4 (insulin-responsive). While G6pase is one of the rate-limiting gluconeogenic enzymes that regulate, the synthesis of glucose and results has shown strong suppression of G6pase activity by extracts of VA [60]. VA extract was found also to protect pancreatic β -cells and the polyphenols present are responsible for this action especially dicaffeoyl-quinic acid.

Most of the traditional uses of the plant have been systematically and scientifically validated and the study of oxidative stress in diabetic rats showed that the aqueous extracts of VA decrease the levels of serum malondialdehyde an indication of the antioxidant property of the plant [60].

***Xylopia aethiopica* (Dunal) A. Rich**

XA, also known as the African pepper or Ethiopian pepper, belongs to the family annonaceae and the genus *Xylopia*. It is a tropical, slim, tall and aromatic tree that grows up to 15-30 m. It is found in the west, central and southern Africa in humid forest zones, native to Nigeria, Ghana, Kenya, Ethiopia, Senegal and Uganda.

XA is a common ethno medicine in West Africa where it is used in the treatment of rheumatism and arthritis, cough, stomachache, bronchitis, biliousness and dysentery [61]. The fruit and vegetable have many medicinal properties and contains phytochemicals, vitamins and minerals. Phytochemicals like

flavonoids are potentially anti-allergic, anti-carcinogenic, anti-viral and antioxidants, the ethanolic extract of XA was found to increase steroid hormone [62], the aqueous extract was also shown to have anti-amylase and anti-lipase activity with antioxidant potentials [63].

A poly-herbal formulation sold in Nigeria containing the following: *Stachytarpheta angustifolia*, *Alstonia congensis*, and XA in the ratio 3:2:1 was found to have hypoglycemic and hyperlipidemic activities [64].

***Syzygium aromaticum* (Linn.) Merrill and Perry (Myrtaceae) (Common Name: Cloves)**

S. aromaticum (clove) belongs to the family myrtaceae and the genus *Syzygium*. Native to Indonesia, this plant can grow to a height of 8-12 m, it is an aromatic flower bud commonly used in Africa, Asia and other parts of the world for the preparation of different spicy dishes. In Nigeria most traditional medical practitioners use the fruits and cloves by boiling in water and the decoction is administered to patients for the treatment of cough, chest congestion and catarrh and the compound eugenol present in this plant is responsible for the aroma and has antioxidative and antimycotic ability [65].

A triterpenoid compound extracted from the clove plant named oleanolic acid has potent diuretic/saluretic, anti-hyperlipidemic, antioxidant and hypoglycemic effects [66]. Ngubane *et al.* [67] showed that oleanolic acid exhibited anti-hyperglycemic effect in STZ-induced diabetic rats by the attenuation of the activities of glycogenic enzymes and the compound eugenol present in this plant is responsible for the aroma and has antioxidative and antimycotic ability. The oil from the extract of this plant protects experimental animals from hepato-nephrotoxicity and oxidative stress due to aflatoxins [68].

Clove bud powder (CBP) possesses high phenolic content, free radical scavenging activity and metal chelating and reducing properties, the major phenolic compounds found are Kaempferol, isoquercitrin, gallic acid, ellagic acid, and caffeic acid [69]. Dietary supplementation of CBP in type 2 diabetic rats showed anti-hyperglycemic, hepatoprotective, hypolipidemic and antioxidant activities, by suppressing oxidative stress and delaying carbohydrate digestion [70].

Oleanolic acid (3 β -hydroxy-olea-12-en-28-oic acid) and maslinic acid have been reported to modulate the activity of the intestinal glucose transporters and carbohydrate hydrolyzing enzymes thus reducing postprandial hyperglycaemia and that the ethanolic extract of this plant suppresses elevated blood glucose levels in type 2 diabetic KK-A^y mice [70].

Free and bound phenolic extract of clove bud was found to inhibit carbohydrate hydrolyzing enzymes; alpha-amylase and alpha-glucosidase in a dose-dependent manner (200-800 μ g/ml) [71]. Decreasing the postprandial hyperglycemia peak is very crucial in the treatment of diabetes; there is a strong correlation between the phenolic content of clove and the enzyme inhibitory

activities and with a strong antioxidant property which is the mechanism and the basis for its anti-diabetic action [71].

***Azadirachta indica* A. Juss (Common Name: Neem)**

AI A. Juss is a member of the Meliaceae family and the genus *Azadirachta*. It is a fast-growing tree that can reach up to 15-20 m and can sometimes reach 40 m. The plant is native to India and adapted to sub-arid and sub-humid tropical climates. It is widely grown in India, Pakistan, Indonesia, Sri Lanka, Caribbean, Nigeria, South and Central America. It is called “Dogonyaro” in Nigeria and grown all over the country, especially in the northern region. The plant has been used in the Indian Ayurveda traditional medicine for over 2000 years for the healing of various diseases and ailments [72].

The composite leaf extract of AI and VA at 500 mg/Kg body weight ameliorates hypoglycemia and hepatic oxidative stress in STZ-induced diabetic rats [54]. AI leaves glucosamine an active component of neem leaves is responsible for immunostimulatory activity in albino mice [73].

The chloroform extract of AI administered on murine diabetic model for 21 days significantly reduced the fasting blood sugar and islet regeneration and protection properties [74]. The administration of 500 mg/kg body weight of AI leaf extract and AI bark extract was effective in improving the antioxidant status in cardiac and skeletal muscles [75]. Khosla *et al.* [76] showed that azadirachtin and nimbin are the active ingredients in AI and they have the ability to regenerate the pancreatic beta cell. Recently Tiwari *et al.* [77] showed that the administration of the composite extract of *Aegle marmelos*, AI, *Murraya koengii*, *Occimum sanctum*, and *S. cumini* at 100 mg/Kg body weight caused a significant reduction in the blood sugar level, total cholesterol, triglyceride, low-density lipoproteins and an increase in the level of high-density lipoproteins.

***Syzygium cumini* (L.) Skeels**

This plant belongs to the family myrtaceae and the genus *Syzygium*; it is an evergreen tropical plant native to South East Asia and widely grown in Africa. The fruit of the plant is widely used in cooking as spice and condiments to add flavor to foods.

S. cumini is well-known for its antidiabetic properties; gallic acid, rutin and chlorogenic acid are the main phenolic present in this plant and the extracts of all parts of the plant is used in traditional medicine [78]. Aqueous extract is found to improve endothelial dysfunction, antioxidant, anti-inflammatory and anti-thrombotic properties of adenosine deamine activity in erythrocytes [78].

A dose of 400 mg/kg body weight of aqueous seed extract of *S. cumini* has hypoglycemic, insulin sensitizing and hypo-lipidemic activity in HFD-STZ induced rats due to an increase in peroxisome proliferator-activated receptor (PPAR)_γ and PPAR_α protein expression [79]. The active fraction of *S. cumini* was found to regenerate pancreatic islets and insulin secretion in STZ-induced diabetic mice [80].

Sharma *et al.* [81] demonstrated that the aqueous extract of *S. cumini* seed when given orally to mice at a dose of 250 mg/kg body weight for 21 days effected and repaired the liver damage associated with alloxan diabetes. The extract of this plant inhibits alpha-glucosidase and alpha-amylase, which are the two enzymes responsible for the metabolism of carbohydrate, and this limits the postprandial glucose and consequently controlling diabetes [82]. The seed extract is found to act as a chemo-protective agent against *in vivo* oxidative stress and genomic damage [83].

***Terminalia catappa*. L.**

This plant belongs to the family Combretaceae and to the genus *Terminalia* found growing in the warmer parts of India, Asia, Africa and Australia. The tree is primarily used as an ornamental and as a shade tree; the seeds are edible like almonds. The extracts of the bark and leaves are reported to have anticancer and aphrodisiac capability [84], antioxidant and anti-inflammatory [85] and anti-malarial [86]. This may be as a result of high contents of tannins in the plant making them a good source of antioxidants [87]. Kinoshita *et al.* [88] isolated chebulagic acid and corilagin from the 50% ethanol extract of the plant with a strong free radical scavenging activity and these compounds are found to have hepato-protective and antioxidant actions, by suppressing the generation of ROS followed by the inhibition of apoptosis.

***Terminalia arjuna* (Roxb) Wight and Arn**

This is a plant belonging to the family Combretaceae and genus *Terminalia* commonly called arjuna. It is a large tree found throughout the South Asia region, and it is an exotic tree in India, it can grow up to a height of 25-30 m. The bark and fruits of this plant is used in traditional Indian medicine as an anti-dysenteric, anti-pyretic, astringent, cardiogenic, lithotriptic, anticoagulant, hypolipidemic and anti-microbial, the large amount of flavonoids is responsible for the antioxidant and anti-microbial properties [89]. The bark contains arjunine a lactone, arjunetin, essential oils and reducing sugars. The methanolic extract exhibited analgesic activity and acute anti-inflammatory activity [90]. The extracts of this plant have the presence of alkaloids, triterpenoids, tannins and flavonoids. Gallic acid, apigenin, luteolin, quercetin, epicatechin, ellagic acid and 1-O-galloyl glucose are some of the compounds that have been isolated from this plant [91].

A dose of 250 and 500 mg/kg body weight of *T. arjuna* extract was found to have reno-protective and antioxidant ability in isolated perfused kidneys [92]. The leaf extracts when administered at a dose of 100 and 200 mg/kg body weight orally to STZ-induced diabetic rats was found to significantly normalize blood glucose level and this is due to its antioxidant role [93]. Due to the presence of tannins, saponin, and flavonoids, the bark extract exhibited antidiabetic activity by enhancing the peripheral utilization of glucose by correcting the impaired liver and kidney glycolysis and by limiting gluconeogenic formation, an action similar to that of insulin [94]. Perveen *et al.* [95] showed that

Table 1: Summary of the selected plant species with their active component and their therapeutic effects

Plant species name	Used component	Property/effect	References
MI L.	1, 2, 3, 4, 6-penta-O-gallolyl- β -D-glucose	Inhibits 11- β -hydroxysteroid hydrogenase enzyme. antidiabetic	[23]
	Mangiferin (1, 3, 6, 7-tetrahydrox y-xanthone-C2- β -D-glucoside)	Improve renal function of diabetic nephropathy in rats. Decrease insulin resistance	[29,30,32,33]
VA Delile	Ethanolic extracts	Lowers blood sugar	[48]
	Dicaffeoyl-quinic acid	Protect pancreatic β cells	[55]
XA (Dunal) A. Rich.	Aqueous extract	Anti-amylase, anti-lipase activity with antioxidant potentials	[58]
<i>S. aromaticum</i> (Linn.)	Oleanolic acid	Anti-hyperlipidemic, antioxidant and hypoglycaemic	[6]
Merrill and Perry	Oleanolic acid and maslinic	reducing postprandial hyper-glycaemia	[65]
AI A. Juss	Chloroform extract	Reduced fasting blood sugar, islet regeneration	[70]
	Azadirachtin and Nimbin	Regeneration of pancreatic beta cells	[72]
<i>S. cumini</i> (L.) Skeels	Aqueous seed extract	Hypoglycaemic, insulin sensitising and hypo-lipidemic activity in HFD-STZ induced rats. Repaired liver damage associated with alloxan diabetes	[74,76]
<i>T. catappa</i> . L	Chebularic acid and Corilagin	Antioxidants	[83]
<i>T. arjuna</i> (Roxb)	Leaf extracts	Normalise blood glucose levels	[88]
Wight and Arn	Ethanolic extracts	Inhibit oxidation and lipid degradation	[91]
<i>T. chebula</i> Retz	Methanolic extracts	Plant inhibits lipid peroxide formation and scavenge hydroxyl and superoxide radical	[94]

MI: *Mangifera indica*, VA: *Vernonia amygdalina*, XA: *Xylopia aethiopica*, AI: *Azadirachta indica*, *S. aromaticum*: *Syzygium aromaticum*, *S. cumini*: *Syzygium cumini*, *T. catappa*: *Terminalia catappa*, *T. arjun*: *Terminalia arjuna*, *T. chebula*: *Terminalia chebula*, HFD: High fat diet, STZ: Streptozotocin

the antioxidant activity of *T. arjuna* bark extract is due to the rich concentration of tannins, triterpenoid and saponins like arjunic acid, arjunolic acid, arjungenin, arjunglycosides, gallic acid, ellagic acid, oligomeric proanthocyanidins, and that the antidiabetic activity is due to the stimulation of β -cells of the pancreatic islets. The administration of *T. arjuna* ethanolic extracts at a dose of 250 mg/kg body weight per oral was found to reverse diabetic condition by inhibiting oxidation and degradation of lipids [96], and due to the fact that *T. arjuna* extract has the ability to reduce postprandial hyperglycemia an important cardiovascular risk factor in type 2 diabetic patients, it has got a promising anti-hyperglycaemic and hypo-lipidemic effects in type 2 diabetics [97].

***Terminalia chebula* Retz. (Combretaceae)**

This plant belongs to the family combretaceae, and the genus *Terminalia* found growing in the Sub-Himalayan tracts. It is a tall tree plant rising to about 15-25 m. It is a revered plant in India and has been extensively used in the Ayurveda, Unani and Homeopathic medicine. It has a beneficial effect on digestive diseases, urinary diseases, diabetes, skin, heart, irregular fevers, constipation, ulcers, vomiting, colic pain and hemorrhoids [98]. Phyto-constituents present in this plants is hydrolysable tannins like gallic acid, chebulagic acid, punicalagin, chebulamin, corilagin, neochebulini acid, ellagic acid, casuarinas and 2,3,6-tri-O-gallolyl- β -D-glucose, 1,6-di-O-gallolyl-D-glucose and terchebulin. The methanolic extracts from the plant inhibits lipid peroxide formation and scavenge hydroxyl and superoxide radicals [99]. The methanolic extract of *T. chebula* has antioxidant, anti-inflammatory and anticancer ability and the phenolic derivatives, hydrolysable tannins and oleanane type triterpenoids are the active principles [100]. Chebulagic acid from *T. chebula* at 100 mg/kg body weight significantly reduced postprandial blood glucose levels of Sprague-Dawley rats when compared to the control group.

CONCLUSION

The present review presents the current scientific literature with respect to the antidiabetic and antioxidant potential of AI, MI, *T. arjuna*, *T. catappa*, *T. chebula*, *S. cumini*, *S. aromaticum*, VA and XA; summarize in Table 1. These plants are not the most popular when it comes to their use as antidiabetic plants used in traditional medicine, but yet they are widely used in some traditional medicinal system in India and Nigeria. It is hoped that further studies on these plants will target the isolation, purification and characterization of the bioactive compounds, which may lead to the discovery of potent antidiabetic drugs for the management and treatment of diabetes.

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REFERENCES

1. World Health Organization. Definition and diagnosis of diabetes mellitus and intermediate hyperglycaemia: Report of a WHO/IDF consultation. Geneva: World Health Organization; 2006.
2. Modak M, Dixit P, Londhe J, Ghaskadbi S, Devasagayam TP. Indian herbs and herbal drugs used for the treatment of diabetes. *J Clin Biochem Nutr* 2007;40:163-73.
3. Brownlee M. Biochemistry and molecular cell biology of diabetic complications. *Nature* 2001;414:813-20.
4. Mehdi U, Toto RD. Anemia, diabetes, and chronic kidney disease. *Diabetes Care* 2009;32:1320-6.
5. Guariguata L, Nola T, Beagley J, Linnenkamp U, Jacqmain O. IDF Diabetes Atlas. 6th ed. Brussels, Belgium: International Diabetes Federation; 2013.
6. Philippe J, Raccach D. Treating type 2 diabetes: How safe are current therapeutic agents? *Int J Clin Pract* 2009;63:321-32.
7. Viridi J, Sivakami S, Shahani S, Suthar AC, Banavalikar MM, Biyani MK.

- Antihyperglycemic effects of three extracts from *Momordica charantia*. J Ethnopharmacol 2003;88:107-11.
8. Mohammed A, Ibrahim MA, Islam MS. African medicinal plants with antidiabetic potentials: A review. Planta Med 2014;80:354-77.
9. Bosi E. Metformin – The gold standard in type 2 diabetes: What does the evidence tell us? Diabetes Obes Metab 2009;11 Suppl 2:3-8.
10. Asche CV, McAdam-Marx C, Shane-McWhorter L, Sheng X, Plauschnat CA. Association between oral antidiabetic use, adverse events and outcomes in patients with type 2 diabetes. Diabetes Obes Metab 2008;10:638-45.
11. Jacobsen IB, Henriksen JE, Beck-Nielsen H. The effect of metformin in overweight patients with type 1 diabetes and poor metabolic control. Basic Clin Pharmacol Toxicol 2009;105:145-9.
12. Jia W, Gao W, Tang L. Antidiabetic herbal drugs officially approved in China. Phytoter Res 2003;17:1127-34.
13. Demain AL, Sanchez S. Microbial drug discovery: 80 years of progress. J Antibiot (Tokyo) 2009;62:5-16.
14. Rizvi SI, Mishra N. Traditional Indian medicines used for the management of diabetes mellitus. J Diabetes Res 2013;2013:712092.
15. Udayakumar R, Kasthuriangan S, Mariashibu TS, Rajesh M, Anbazhagan VR, Kim SC, *et al.* Hypoglycaemic and hypolipidaemic effects of *Withania somnifera* root and leaf extracts on alloxan-induced diabetic rats. Int J Mol Sci 2009;10:2367-82.
16. Inayat-ur-Rahman, Malik SA, Bashir M, Khan R, Iqbal M. Serum sialic acid changes in non-insulin-dependant diabetes mellitus (NIDDM) patients following bitter melon (*Momordica charantia*) and rosiglitazone (Avandia) treatment. Phytomedicine 2009;16:401-5.
17. Fabricant DS, Farnsworth NR. The value of plants used in traditional medicine for drug discovery. Environ Health Perspect 2001;109 Suppl 1:69-75.
18. Miller LH, Su X. Artemisinin: Discovery from the Chinese herbal garden. Cell 2011;146:855-8.
19. Kingston DG. Taxol and its analogs. In: Cragg GM, Kingston DG, Newman DJ, editors. Anticancer Agents from Natural Products. 2nd ed. Boca Raton, FL: Taylor and Francis; 2012. p. 123-75.
20. Newman DJ, Cragg GM. Natural products as sources of new drugs over the last 25 years. J Nat Prod 2007;70:461-77.
21. Tiwari BK, Pandey KB, Abidi AB, Rizvi SI. Therapeutic potential of Indian medicinal plants in diabetic condition. Ann Phytomed 2013;2:37-43.
22. Mukherjee PK, Maiti K, Mukherjee K, Houghton PJ. Leads from Indian medicinal plants with hypoglycemic potentials. J Ethnopharmacol 2006;106:1-28.
23. Grover JK, Yadav S, Vats V. Medicinal plants of India with anti-diabetic potential. J Ethnopharmacol 2002;81:81-100.
24. Loh SP, Hadira O. *In vitro* inhibitory potential of selected Malaysian plants against key enzymes involved in hyperglycemia and hypertension. Malays J Nutr 2011;17:77-86.
25. Fasoli E, Righetti PG. The peel and pulp of mango fruit: A proteomic samba. Biochim Biophys Acta 2013;1834:2539-45.
26. Etuk, EU, Bello SO, Isezu SA, Mohammed BJ. Medicinal plants used for the treatment of diabetes mellitus in the North Western region of Nigeria. Asian J Exp Biol Sci 2010;1:55-9.
27. Ma X, Wu H, Liu L, Yao Q, Wang S, Zhan R, *et al.* Polyphenolic compounds and antioxidant properties in mango fruits. Sci Hortic 2011;129:102-7.
28. Mohan CG, Viswanatha GL, Savinay G, Rajendra CE, Halemani PD. 1,2,3,4,6 Penta-O-galloyl-β-D-glucose, a bioactivity guided isolated compound from *Mangifera indica* inhibits 11β-HSD-1 and ameliorates high fat diet-induced diabetes in C57BL/6 mice. Phytomedicine 2013;20:417-26.
29. Shoji K, Tsubaki M, Yamazoe Y, Satou T, Itoh T, Kidera Y, *et al.* Mangiferin induces apoptosis by suppressing Bcl-xL and XIAP expressions and nuclear entry of NF-κB in HL-60 cells. Arch Pharm Res 2011;34:469-75.
30. Yoshimi N, Matsunaga K, Katayama M, Yamada Y, Kuno T, Qiao Z, *et al.* The inhibitory effects of mangiferin, a naturally occurring glucosylxanthone, in bowel carcinogenesis of male F344 rats. Cancer Lett 2001;163:163-70.
31. Subbiya A, Mahalakshmi K, Pushpangadan S, Padmavathy K, Vivekanandan P, Sukumaran VG. Antibacterial efficacy of *Mangifera indica* L. kernel and *Ocimum sanctum* L. leaves against *Enterococcus faecalis* dental biofilm. J Conserv Dent 2013;16:454-7.
32. Wang RR, Gao YD, Ma CH, Zhang XJ, Huang CG, Huang JF, *et al.* Mangiferin, an anti-HIV-1 agent targeting protease and effective against resistant strains. Molecules 2011;16:4264-77.
33. Das J, Ghosh J, Roy A, Sil PC. Mangiferin exerts hepatoprotective activity against D-galactosamine induced acute toxicity and oxidative/nitrosative stress via Nrf2-NFκB pathways. Toxicol Appl Pharmacol 2012;260:35-47.
34. Sellamuthu PS, Muniappan BP, Perumal SM, Kandasamy M. Antihyperglycemic effects of mangiferin in STZ-induced diabetic rats. J Health Sci 2009;55:206-14.
35. Ichiki H, Miura T, Kubo M, Ishihara E, Komatsu Y, Tanigawa K, *et al.* New antidiabetic compounds, mangiferin and its glucoside. Biol Pharm Bull 1998;21:1389-90.
36. Dineshkumar B, Mitra A, Manjunatha M. Studies on the antidiabetic and hypolipidemic potentials of mangiferin (Xanthone glucoside) in STZ-induced type1 and type 2 diabetic rat models. Int J Adv Pharm Sci 2010;1:75-85.
37. Li X, Cui X, Sun X, Li X, Zhu Q, Li W. Mangiferin prevents diabetic nephropathy progression in streptozotocin-induced diabetic rats. Phytoter Res 2010;24:893-9.
38. Miura T, Ichiki H, Hashimoto I, Iwamoto N, Kato M, Kubo M, *et al.* Antidiabetic activity of a xanthone compound, mangiferin. Phytomedicine 2001;8:85-7.
39. Ling LT, Yap SA, Radhakrishnan AK, Subramanian T, Cheng HM, Palanisamy UD. Standardised *Mangifera Indica* extract is an ideal antioxidant. Food Chem 2009;113:1154-9.
40. Bhattacharjee B, Lakshminarasimhan P, Bhattacharjee A, Agrawala DK, Pathak MK. *Vernonia amygdalina* Del (Asteraceae) - An African medicinal plants introduced in India. Zoo's Print 2013;28:18-20.
41. Eleyinmi AF, Sporns P, Bressler DC. Nutritional composition of *Gongro-nema latifolium* and *Vernonia amygdalina*. Nutr Food Sci 2008;38:99-109.
42. Borokini TI, Ighere DA, Clement M, Ajiboye TO, Alowonle AA. Ethno biological survey of traditional medicine practice for the treatment of piles and diabetes mellitus in Oyo State. J Med Plants Stud 2013;1:30-40.
43. Farombi EO, Owuoye O. Antioxidative and chemopreventive properties of *Vernonia amygdalina* and *Garcinia biflavonoid*. Int J Environ Res Public Health 2011;8:2533-55.
44. Gbolade AA. Inventory of antidiabetic plants in selected districts of Lagos State, Nigeria. J Ethnopharmacol 2009;121:135-9.
45. Abo KA, Fred-Jaiyesimi AA, Jaiyesimi AE. Ethnobotanical studies of medicinal plants used in the management of diabetes mellitus in South Western Nigeria. J Ethnopharmacol 2008;115:67-71.
46. Erasto P, Van de Venter M, Roux S, Grierson DS, Afolayan AJ. Effect of leaf extract of *Vernonia amygdalina* on glucose utilization in chang liver C2C12 muscle and 3T3-L1 cells. Pharm Biol 2009;47:175-81.
47. Akah PA, Ekekwe RK. Ethno pharmacology of some Asteraceae family used in Nigerian traditional medicine. Fitoterapia 1995;66:351-5.
48. Sharma MC, Sharma S. Pharmacognostic and phytochemical screening of *Vernonia amygdalina* Linn against selected bacterial strains. Middle-East J Sci Res 2010;6:440-4.
49. Wong FC, Woo CC, Hsu A, Tan BK. The anti-cancer activities of *Vernonia amygdalina* extract in human breast cancer cell lines are mediated through caspase-dependent and p53-independent pathways. PLoS One 2013;8:e78021.
50. Amira OC, Okubadejo NU. Frequency of complementary and alternative medicine utilization in hypertensive patients attending an urban tertiary care centre in Nigeria. BMC Complement Altern Med 2007;7:30.
51. Atangwho IJ, Egbung GE, Ahmad M, Yam MF, Asmawi MZ. Antioxidant versus anti-diabetic properties of leaves from *Vernonia amygdalina* Del. growing in Malaysia. Food Chem 2013;141:3428-34.
52. Atangwho IJ, Ebong PE, Eteng MU, Eyong EU, Obi AU. Effect of *Vernonia amygdalina* Del. leaf on kidney function of diabetic rats. Int J Pharmacol 2007;3:143-8.
53. Ong KW, Hsu A, Song L, Huang D, Tan BK. Polyphenols-rich *Vernonia amygdalina* shows anti-diabetic effects in streptozotocin-induced diabetic rats. J Ethnopharmacol 2011;133:598-607.
54. Akinola OB, Omotoso GO, Akinola OS, Dosumu OO, Adewoye ET. Effects of combined leaf extract of *Vernonia amygdalina* and

- Azadirachta indica* on hepatic morphology and hepatotoxicity markers in streptozotocin-induced diabetic rats. *Zhong Xi Yi Jie He Xue Bao* 2011;9:1373-9.
55. Lolodi O, Eriyamremu GE. Effect of methanolic extract of *Vernonia amygdalina* (common bitter leaf) on lipid peroxidation and antioxidant enzymes in rats exposed to cycasin. *Pak J Biol Sci* 2013;16:642-6.
56. Ejike CE, Awazie SO, Nwangozi PA, Godwin CD. Synergistic postprandial blood glucose modulatory properties of *Vernonia amygdalina* (Del.), *Gongronema latifolium* (Benth.) and *Occimum gratissimum* (Linn.) aqueous decoctions. *J Ethnopharmacol* 2013;149:111-6.
57. Yeap SK, Ho WY, Beh BK, Liang WS, Ky H, Yousr AH, *et al.* *Vernonia amygdalina*, an ethno veterinary and ethno medical used green vegetable with multiple bioactivities. *J Med Plants Res* 2010;4:2787-812.
58. Ong KW, Hsu A, Tan BK. Anti-diabetic and anti-lipidemic effects of chlorogenic acid are mediated by ampk activation. *Biochem Pharmacol* 2013;85:1341-51.
59. Ebong PE, Atangwho IJ, Eyong EU, Egbung GE. The antidiabetic efficacy of combined extracts from two continental plants: *Azadirachta indica* (A. Juss) (Neem) and *Vernonia amygdalina* (Del.) (African Bitter Leaf). *Am Biochem Biotech* 2008;4:239-44.
60. Eteng MU, Bassey BJ, Atangwho IJ. Biochemical indices of macrovascular complication in diabetic rat model: Compared effects of *Vernonia amygdalina*, *Catharanthus roseus* and chlpropamide. *Asian J Biochem* 2008;3:228-34.
61. Fleischer TC, Mensah ML, Mensah AY, Komlaga G, Gbedema SY, Skaltsa H. Antimicrobial activity of essential oils of *Xylopi aethiopica*. *Afr J Tradit Complement Altern Med* 2008;5:391-3.
62. Woode E, Alhassan A, Abaidoo CS. Effect of ethanolic fruit extract of *Xylopi aethiopica* on reproductive function of male rats. *Int J Pharm Biomed Res* 2011;2:161-5.
63. Oben J, Etooundi CB, Kuete D, Ngondi JL. Anti-amylase, anti-lipase and antioxidant effect of aqueous extracts of some Cameroonian spices. *J Nat Prod* 2010;3:165-71.
64. Ogbonna SO, Mbaka GO, Adekunle A, Anyika EN, Gbolade OE, Nwakakwa N. Effect of a poly-herbal formulation, Okudiabet, on alloxan-induced diabetic rats. *Agric Biol J North Am* 2010;1:139-45.
65. Ene AC, Atawodi SE. Ethno- medicinal survey of plants used by the Kanuri's of North-eastern Nigeria. *Indian J Tradit Knowl* 2012;11:640-5.
66. Somova LO, Nadar A, Rammanan P, Shode FO. Cardiovascular, antihyperlipidemic and antioxidant effects of oleanolic and ursolic acids in experimental hypertension. *Phytomedicine* 2003;10:115-21.
67. Ngubane PS, Masola B, Musabayane CT. The effects of *Syzygium aromaticum*-derived oleanolic acid on glycogenic enzymes in streptozotocin-induced diabetic rats. *Ren Fail* 2011;33:434-9.
68. Adefegha SA, Oboh G, Adefegha OM, Boligon AA, Athayde ML. Antihyperglycemic, hypolipidemic, hepatoprotective and antioxidative effects of dietary clove (*Syzygium aromaticum*) bud powder in a high-fat diet/streptozotocin-induced diabetes rat model. *J Sci Food Agric* 2014;94:2726-37.
69. Khatthi A, Serumula MR, Myburg RB, Van Heerden FR, Musabayane CT. Effects of *Syzygium aromaticum*-derived triterpenes on postprandial blood glucose in streptozotocin-induced diabetic rats following carbohydrate challenge. *PLoS One* 2013;8:e81632.
70. Kuroda M, Mimaki Y, Ohtomo T, Yamada J, Nishiyama T, Mae T, *et al.* Hypoglycemic effects of clove (*Syzygium aromaticum* flower buds) on genetically diabetic KK-Ay mice and identification of the active ingredients. *J Nat Med* 2012;66:394-9.
71. Adefegha SA, Oboh G. *In vitro* inhibition activity of polyphenol-rich extracts from *Syzygium aromaticum* (L.) Merr. & Perry (Clove) buds against carbohydrate hydrolyzing enzymes linked to type 2 diabetes and Fe(2+)-induced lipid peroxidation in rat pancreas. *Asian Pac J Trop Biomed* 2012;2:774-81.
72. Subapriya R, Nagini S. Medicinal properties of neem leaves: A review. *Curr Med Chem Anticancer Agents* 2005;5:149-6.
73. Kumar VS, Navaratnam V, Ramachandran S. Isolation and characterization of glucosamine from *Azadirachta indica* leaves: An evaluation of immunostimulant activity in mice. *Asian Pac J Trop Biomed* 2012;51:561-7.
74. Bhat M, Kothiwale SK, Tirmale AR, Bhargava SY, Joshi BN. Antidiabetic properties of *Azadirachta indica* and *Bougainvillea spectabilis*: *In vivo* studies in murine diabetes model. *Evid Based Complement Alternat Med* 2011;2011:561625.
75. Shailey S, Basir SF. Protective role of *Azadirachta indica* against oxidative damage in skeletal and cardiac muscle of alloxan diabetic rats. *Int J Pharm Sci* 2012;4:471-7.
76. Khosla P, Bhanwra S, Singh J, Seth S, Srivastava RK. A study of hypoglycaemic effects of *Azadirachta indica* (Neem) in normal and alloxan diabetic rabbits. *Indian J Physiol Pharmacol* 2000;44:69-74.
77. Tiwari BK, Kumar D, Abidi AB, Rizvi SI. Efficacy of composite extract from leaves and fruits of medicinal plants used in traditional diabetic therapy against oxidative stress in alloxan-induced diabetic rats. *ISRN Pharmacol* 2014;2014:608590.
78. De Bona KS, Bonfanti G, Bitencourt PE, Cargnelutti LO, da Silva PS, da Silva TP, *et al.* *Syzygium cumini* is more effective in preventing the increase of erythrocytic ADA activity than phenolic compounds under hyperglycemic conditions *in vitro*. *J Physiol Biochem* 2014;70:321-30.
79. Sharma AK, Bharti S, Kumar K, Krishnamurthy B, Bhatia J, Kumari S, *et al.* *Syzygium cumini* ameliorates insulin resistance and β -cell dysfunction via modulation of PPAR γ , dyslipidemia, oxidative stress and TNF- α in type 2 diabetic rats. *J Pharmacol Sci* 2012;119:205-13.
80. Dusane MB, Joshi BN. Seeds of *Syzygium cumini* (L.) Skeels: Potential for islet regeneration in experimental diabetes. *Zhong Xi Yi Jie He Xue Bao* 2011;9:1380-7.
81. Sharma B, Siddiqui MS, Kumar SS, Ram G, Choudhary M. Liver protective effects of aqueous extracts of *Syzygium cumini* in Swiss albino mice on alloxan-induced diabetes mellitus. *J Pharm Res* 2013;6:853-8.
82. Shinde J, Taldone T, Barletta M, Kunaparaju N, Hu B, Kumar S, *et al.* Alpha-glucosidase inhibitory activity of *Syzygium cumini* (Linn.) Skeels seed kernel *in vitro* and in Goto-Kakizaki (GK) rats. *Carbohydr Res* 2008;343:1278-81.
83. Arun R, Prakash MV, Abraham SK, Premkumar K. Role of *Syzygium cumini* seed extract in the chemoprevention of *in vivo* genomic damage and oxidative stress. *J Ethnopharmacol* 2011;134:329-33.
84. Mininel FJ, Junior CS, Espanha LG, Rosende FA, Varanda EA, Leite CQ, *et al.* Characterization and quantification of compounds in hydro-alcoholic extract of the leaves from *Terminalia catappa* Linn. (Combrataceae) and their mutagenic activity. *Evid Based Complement Alternat Med* 2014;2014:11. doi: 10.1155/2014/676902.
85. Chen PS, Li JH. Chemopreventive effect of punicalagin, a novel tannin component isolated from *Terminalia catappa*, on H-ras-transformed NIH3T3 cells. *Toxicol Lett* 2006;163:44-53.
86. Chanda S, Rakholiya K, Nair CR. Antimicrobial activity of *Terminalia catappa* L. leaf extracts against some clinically important pathogenic microbial strains. *Chin Med* 2011;2:171-7.
87. Chyau CC, Tsai SY, Ko PT, Mau JL. Antioxidant properties of solvent extracts from *Terminalia catappa* leaves. *Food Chem* 2000;78:483-8.
88. Kinoshita S, Inoue Y, Nakama S, Ichiba T, Aniya Y. Antioxidant and hepatoprotective actions of medicinal herb, *Terminalia catappa* L. from Okinawa Island and its tannin corilagin. *Phytomedicine* 2007;14:755-62.
89. Mandal S, Patra A, Samanta A, Roy S, Mandal A, Mahapatra TD, *et al.* Analysis of phytochemical profile of *Terminalia arjuna* bark extract with antioxidative and antimicrobial properties. *Asian Pac J Trop Biomed* 2013;3:960-6.
90. Biswas M, Biswas K, Karan TK, Bhattacharya S, Gosh AK, Haldar PK. Evaluation of analgesic and anti-inflammatory activities of *Terminalia arjuna* leaf. *J Phytol* 2011;3:33-8.
91. Singh PP, Chauhan SM. Activity-guided isolation of antioxidants from the leaves of *Terminalia arjuna*. *Nat Prod Res* 2014;28:760-3.
92. Raj CD, Shabi MM, Jipnomon J, Dhevi R, Gayathri K, Subashini U, *et al.* *Terminalia arjuna*'s antioxidant effect in isolated perfused kidney. *Res Pharm Sci* 2012;7:181-8.
93. Biswas M, Kar B, Bhattacharya S, Kumar RB, Ghosh AK, Haldar PK. Antihyperglycemic activity and antioxidant role of *Terminalia arjuna* leaf in streptozotocin-induced diabetic rats. *Pharm Biol* 2011;49:335-40.
94. Ragavan B, Krishnakumari S. Antidiabetic effect of *T. arjuna* bark extract in alloxan induced diabetic rats. *Indian J Clin Biochem* 2006;21:123-8.
95. Perveen K, Kahn R, Siddiqui WA. Antidiabetic effects afforded by *Terminalia arjuna* in high fat-feed and STZ-induced type 2 diabetic rats. *Int J Diabetes Metab* 2011;19:23-33.
96. Chander R, Singh K, Khanna AK, Kaul SM, Puri A, Saxena R, *et al.*

- Antidyslipidemic and antioxidant activities of different fractions of *Terminalia arjuna* stem bark. Indian J Clin Biochem 2004;19:141-8.
97. Morshed MA, Haque A, Rokeya B, Ali L. Anti-hyperglycaemic and lipid lowering effect of *Terminalia arjuna* bark extracts on STZ-induced type 2 diabetic model rats. Int J Pharm Pharm Sci 2011;3:450-4.
 98. Juang LJ, Sheu SJ, Lin TC. Determination of hydrolyzable tannins in the fruit of *Terminalia chebula* Retz. by high-performance liquid chromatography and capillary electrophoresis. J Sep Sci 2004;27:718-24.
 99. Bag A, Bhattacharyya SK, Chattopadhyay RR. The development of *Terminalia chebula* Retz. (Combretaceae) in clinical research. Asian Pac J Trop Biomed 2013;3:244-52.
 100. Manosroi A, Jantrawut P, Ogihara E, Yamamoto A, Fukatsu M,

Yasukawa K, *et al.* Biological activities of phenolic compounds and triterpenoids from the galls of *Terminalia chebula*. Chem Biodivers 2013;10:1448-63.

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