



Review

Animal models and natural products to investigate *in vivo* and *in vitro* antidiabetic activity

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ABSTRACT

Diabetes mellitus is a chronic disease which has high prevalence. The deficiency in insulin production or impaired insulin function is the underlying cause of this disease. Utilization of plant sources as a cure of diabetes has rich evidence in the history. Recently, the traditional medicinal plants have been investigated scientifically to understand the underlying mechanism behind antidiabetic potential. In this regard, a substantial number of *in vivo* and *in vitro* models have been introduced for investigating the bottom-line mechanism of the antidiabetic effect. A good number of methods have been reported to be used successfully to determine antidiabetic effects of plant extracts or isolated compounds. This review encompasses all the possible methods with a list of medicinal plants which may contribute to discovering a novel drug to treat diabetes more efficaciously with the minimum or no side effects.

1. Introduction

Recently, over a thousand plants have been reported which may possess antidiabetic potentials [1]. However, so far, relatively few traditional medicinal plants have been scientifically evaluated to prove their safety, potential benefits and effectiveness as antidiabetic agent. Moreover, inadequate data exist for most plants to guarantee their quality, efficacy and safety. This article reviews all the possible aspects of antidiabetic research including the animal models used to investigate antidiabetic activity both *in vivo* and *in vitro* along with the medicinal plants reported for exhibiting antidiabetic activity [2,3].

Animal models are one of the major tools to progress with establishing an effective model to investigate the mechanism of action as well as to explore the efficacy of the active principles and plants claimed to show antidiabetic potentials. Furthermore, the disease itself is heterogenic in nature which leads many ways to cause the diabetes as well as the other conditions related to diabetes. Thus, a single animal model to investigate the efficacy of the drug is not possible owing to the existence of different types of diabetes mellitus [4,5].

In this regard, non-diabetic animals and diabetic animals with impaired glucose tolerance are used as normal control and diabetic control respectively to measure the hypoglycaemic condition. Additionally, tumor necrosis factor- α (TNF- α) is used to gain insulin resistant diabetic model [6]. It is a matter of great concern that the active

ingredients of medicinal plants might not be evaluated effectively in lowering the blood sugar level due to the differences in hepatic metabolism system between human and rodents. The metabolism process happens in several steps, so the active principles have to go through all this pathway and the metabolites are the active agents which reach to the body [7–9].

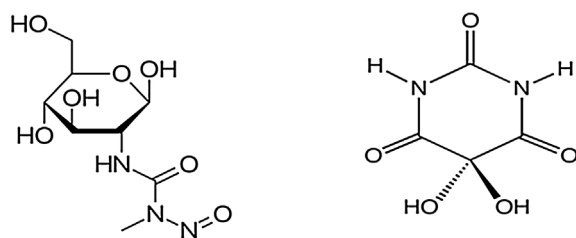
Sensitivity of the same active principles might vary in different species because of the variations in absorption, distribution, metabolism and elimination (ADME) [2,10–12]. Among all the animal models, the rodents have been widely used in diabetes research for various reasons viz., the time of diabetes induction in the rodents is very short and the maintenance cost is relatively low which cuts down the experiment budget significantly. Furthermore, genetic mutations to induce diabetes have been largely reported in favour of rodents which are significantly greater than any other animal groups [13–15].

2. *In vivo* animal experimental models for diabetes mellitus

Diabetes mellitus is induced chemically, surgically or by genetic manipulations (see Table 2). Although, there are some reports that have been shown to use the larger animals to induce diabetes, it is very common to use rodents for the experiments of diabetes [16–18]. Surgical model in large animal has been reported to perform pancreatectomy in dog [19]. In other study, diabetes prone strains have been

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Streptozotocin

Alloxan

Fig. 1. Chemical structures of streptozotocin (STZ) and alloxan (ALX).

introduced to the animals in regard to induce diabetes [20,21].

The classifications of diabetic models can be further narrowed down according to the type of nature of induction of diabetes. The narrowed down animal models are as follows:

1. Spontaneously developed diabetic model (e.g., CBA/Ca mouse, ob/ob mouse, etc.).
2. Experimentally developed diabetic model (e.g., chemically induced by alloxan or streptozotocin; surgically developed by pancreatectomy).

2.1. Chemically induced diabetic model

Streptozotocin (STZ) and alloxan (ALX) (Fig. 1) are widely used to induce diabetes mellitus in animals. Both chemicals can be administered through either intravenous (IV), intraperitoneal (IP) or subcutaneous (SC) ways. The mechanisms of action for both chemicals are quite understandable. Both STZ and ALX are selective cytotoxic agents and consequently destroy the pancreatic beta cells selectively. In short, STZ and ALX are transported to pancreatic beta cells by GLUT2 glucose transporter since both are glucose analogues. The STZ splits into glucose and methyl nitrosourea. Methyl nitrosourea possesses alkylating properties. The alkylation of DNA by methyl nitrosourea leads to the destruction of the beta cells. Contrary, the ALX generates reactive oxygen species (ROS) in the presence of glutathione intracellularly. The ROS cause cyclic redox reaction until hydroxyl radicals are produced

which eventually lead to the destruction of the beta cells [22–24]. Moreover, some studies have suggested that the complete or partial loss of pancreatic beta cells leads to various complications such as hyperglycemia along with glycosuria, polyuria, polydipsia, hyperphagia, and weight loss [25–27].

The streptozotocin induction causes suppression of insulin secretion by the destruction of pancreatic beta cells. Streptozotocin is a nitrosourea analogue which has a hexose moiety linked to N-methyl-N-nitrosourea moiety. Since the nitrosourea is lipophilic, the cellular uptake of STZ into plasma membrane is fast. Additionally, the STZ is selectively accumulated and transported via GLUT2 glucose transporter [28,29]. Consequently, the insulin producing cells could be STZ resistant if the cell does not express any GLUT2 transporter [30–32]. The underlying mechanism of toxic effects of STZ to the pancreatic beta cell is assumed to be taken place by the alkylation of DNA from the interaction of methyl nitrosourea moiety of STZ. The consequence of the alkylation initiates a bunch of events which lead to fragmentation of the DNA [33–36] (Fig. 2).

Another hypothesis for STZ mechanism of action claims that the intracellular nitric oxide (NO) donor is responsible for the diabetogenic effects of STZ [37]. Chemically, STZ could liberate NO as it possesses nitroso group. Moreover, effect of NO is attributed to the elevated action of guanyl cyclase and the formation of cGMP by STZ. Therefore, the most toxic alkylating agent known as methyl methanesulphonate is not a NO donor which may suggest that the NO donating could not be considered as the underlying reason for toxic effects of STZ [38].

Additionally, it has been reported to generate ROS along with superoxide and hydroxyl radicals as byproducts of hydrogen peroxide dismutation in hypoxanthine metabolism. A minor effect from the ROS can be expected during the toxic effects in pancreatic beta cells even though it does not play a vital role [39].

On the other hand, the mechanism of action of alloxan includes the reactive oxygen species (ROS). The dismutation of free radicals causes great increase of calcium concentration inside the cell which leads to rapid destruction of the pancreatic beta cells. Dialuric acid (Fig. 3) is a reduction product of alloxan which helps to generate ROS in a cyclic reaction [40–42]. The dose of alloxan needed to induce diabetes is less compared to STZ. Additionally, there is high possibility of death if it is slightly overdosed [43]. Moreover, the tubular cell necrosis in kidney

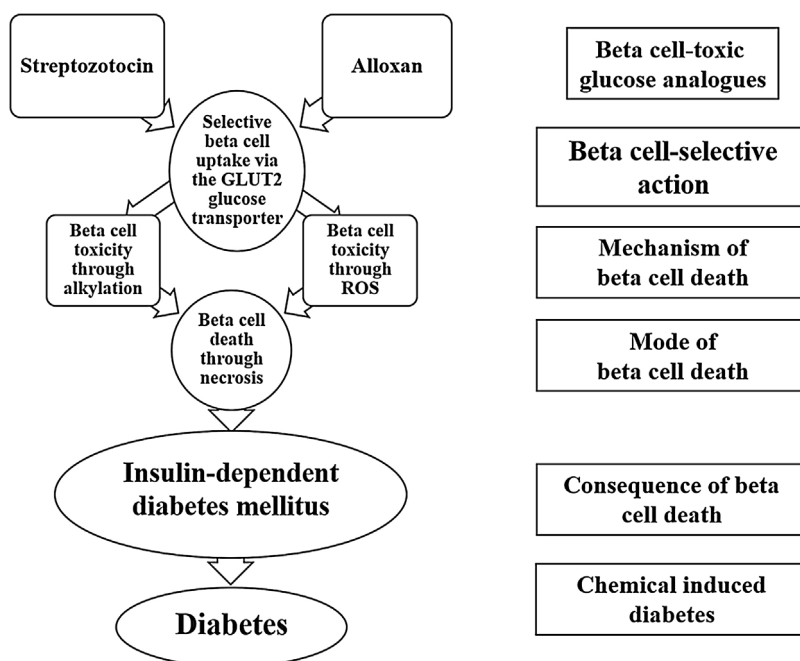


Fig. 2. A graphical illustration of the toxic effects of alloxan and streptozotocin in the beta cells.

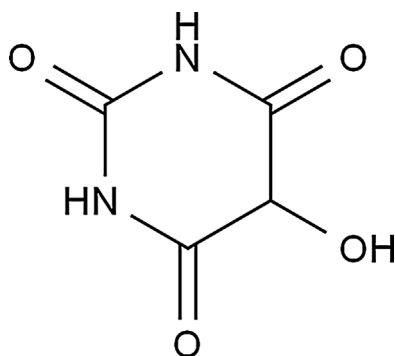


Fig. 3. Structure of dialuric acid i.e., reduction product of alloxan responsible for ROS generation.

has been reported to be observed due to high dose of ALX induction [22]. Since, the correct dosage regime for the induction of diabetes in animals is very important matter for both the STZ and ALZ, scientists have reported the doses extensively in several scientific communications.

The chemical profile, mechanisms of action and doses of streptozotocin and alloxan in various diabetic animal models are presented in Table 3.

2.2. Surgically induction of diabetes

Apart from the chemical induction, the surgical removal of pancreas is an alternative in regard to reduce toxic side effects of chemically induced diabetes. It has been reported to develop diabetic animal model of some species viz., rats, dogs, pigs, etc. [44,4]. However, the limitations of this technique have also been reported by many published studies. The highly complex nature of the technique demands great expertise to perform the surgery. Additionally, the cost of the surgery room set up, large amount of analgesic and antibiotic, post-surgery pancreatic enzyme supplement and risk of animal infection are major drawback of this technique. Moreover, there have been many attempts to perform partial pancreatic surgery which is yet to be developed or acclaimed to achieve desired diabetogenic action [45,46,10,47].

2.3. Spontaneous and transgenic animal models

Several diabetic animal models have so far been developed and reported where no chemical or surgical methods have been used to induce diabetes. In these instances, diabetes is developed either spontaneously or by genetic alteration. Animal models have been listed in Table 1. In some cases, the insulin resistant has been reported as comorbid to obesity, dyslipidemia, and hypertension which helps to investigate the diabetes complications similar to type II diabetes of human being [48,13,8,5].

There are many methods available in these criteria. Each of the models has different underlying mechanism. For instance, db/db mice develops hyperglycemia rapidly, however, Goto–Kakizaki rat which is genetically lean type II diabetic model develops hyperglycemia spontaneously in comparison to glucose-intolerant nondiabetic Wistar rats. BB rats can develop diabetes by 12 weeks whereas NOD mouse usually takes 12–30 weeks to develop hyperglycemia [46,49,50].

2.4. Virus-induced diabetic model

It has been reported that the juvenile diabetes (Type I) has a link with virus infection as well as an autoimmune condition on specific beta cell [51]. A virus named encephalomyocarditis which is a D-variant has been reported to destroy the pancreatic beta cells [52,53]. Two of the virus susceptible animals are male ICR Swiss (adult) mice and male C3H/HeJ (adult) mice whereas the former was found more prone

Table 1
Classification of diabetic models in various animals.

Model category	Obese	Non-obese
(I) Spontaneous or genetically derived diabetic animals	1. CBA/Ca mouse, 2. ob/ob mouse, 3. db/db mouse, 4. KK mouse, 5. KK/Ay mouse, 6. NZO mouse, 7. NONcNZO10 mouse, 8. TSOD mouse, 9. M16 mouse, 10. BHE rat, 11. Zucker fatty rat, 12. WDF/Ta-fa rat (Winstler fatty rat), 13. ZDF rat, 14. obese SHR rats, 15. SHR/N-cp rat, 16. JCR/LA-cp rat, 17. OLETF rat, 18. Obese rhesus monkey	1. NOD mouse (Non obese diabetic mouse), 2. ALS/Lt mouse, 3. (Akita) mutant mouse, 4. GK rat, 5. Cohen diabetic rat, 6. C57BL/6 Torri rat (Non-obese), 7. BB rat (Bio Breeding), 8. WBN/Kob rat, 9. eSS-rat
(II) Diet/nutrition induced diabetic animals	1. C57/BL 6J mouse, 2. Spiny mouse, 3. Sand rat	–
(III) Chemically induced diabetic animals	GTG-treated obese mice	1. Neonatal STZ rat, 2. Low-dose ALX or STZ adult rats and mic
(IV) Transgenic/knock-out diabetic animals	1. β -3 receptor knockout mouse, 2. Uncoupling protein (UCP1) knockout mouse	1. IRS-1, IRS-2, GLUT-4, PTP-1B, and others PPAR-g tissue-specific knockout mouse, 2. Glucokinase or GLUT 2 knockout mice, 3. Human islet amyloid polypeptide (HIP) over expressed rat, 4. RIP-LCMV transgenic mice, 5. Murine Hsp60 transgenic NOD mice
(V) Virus induced diabetic animals	–	1. Male ICR Swiss mice (susceptible to D-variant of encephalomyocarditis virus (EMC-D)), 2. adult C3H/HeJ male mice (Resistant to EMC-D), 3. DBA/2 mice (Susceptible to NDK25, a variant of encephalomyocarditis virus which has been cloned from the M variant of encephalomyocarditis virus)

ALS: alloxan sensitive; ALX: alloxan; BB: Bio Breeding; C3H/HeJ: lipopolysaccharide response, locus; DBA: Dilute Brown Non-Agouti; GLUT: glucose transporter; GK: Goto–Kakizaki; GTG: gold thioglucose; ICR: impaired cytokine response; IRS: insulin receptor substrate; JCR: James C Russel; KK: Kuo Kondo; KK/Ay: yellow KK obese; NZO: New Zealand obese; OLETF: Otuska Long Evans Tokushima fatty; PPAR: peroxisome proliferator activated receptor; PTP: phosphotyrosine phosphatase; SHR/N-cp: spontaneously hypertensive rat/NIH-corpulent; STZ: streptozotocin; TSOD: Tsumara Suzuki obese diabetes; VMH: ventromedial hypothalamus; ZDF: Zucker diabetic fatty.

After [1].

to the induction of diabetes than the later. The experiment was further confirmed by the pre-treatment of a potent immunosuppressant named cyclosporin. The consequences of the induction of immunosuppressant

Table 2

Advantages and disadvantages of various classes of diabetic animal models.

Model category	Advantages	Disadvantages
(I) Spontaneous diabetic animals	<ol style="list-style-type: none"> 1. The spontaneous origin diabetes has many varieties which depicts more precisely the complex nature of diabetes in human being [48,13,8,5]. 2. Many animal methods available according to onset of action or mechanism of action. Some models are very rapid to develop diabetes whereas some models depict variety mechanism of actions including insulin resistant of type II diabetes [46,49,50]. 	<ol style="list-style-type: none"> 1. Apart from showing different mechanism of action, the animal models are highly monogenic when human ADME system is more complex than these models [2,10–12]. 2. The animal model is not very available and post-diabetes maintenance is a major challenge to keep the animals healthy [46,49,50].
(II) Diet/nutrition induced diabetics	<ol style="list-style-type: none"> 1. The experiment is cost effective since diabetes is induced by diet or nutrition which reduces cost of experiment. 2. Safer than chemicals and less toxicity to vital organs. 	<ol style="list-style-type: none"> 1. Long period is needed since the animals are fed over diet to increase blood sugar level.
(III) Chemical induced diabetic animals (STZ or ALX)	<ol style="list-style-type: none"> 1. The toxicity is very selective to pancreatic beta cells which avoids other complications. 2. Mortality rate is relatively low since the correct doses of both STZ and ALX are earlier reported [3,23]. 	<ol style="list-style-type: none"> 1. The treatment is less stable and reversible as the rate of beta cell generation is high. 2. Toxicity in other vital organs is also a major concern.
(IV) Surgical diabetic animals	<ol style="list-style-type: none"> 1. The model is free from toxicity of other vital organs [44,4]. 2. The model is developed rapidly and it can be developed very selectively by only removing the pancreatic beta cells [44,4]. 	<ol style="list-style-type: none"> 1. It is very expensive to set up the whole operation. 2. It demands high level of technical expertise to perform the surgery [45,46,10,47]. 3. The postoperative antibiotic and hormone supplements are very costly and requires high level of monitoring [45,46,10,47].
(V) Transgenic/knock out diabetic animals	<ol style="list-style-type: none"> 1. The model is very selective and able to shed lights on a single gene extensively. 	<ol style="list-style-type: none"> 1. The model is very complex and expensive to develop.

was found to increase in diabetic conditions in both cases [54].

3. *In vitro* animal models

In early studies, insulin-like activity had been measured by the glucose uptake into the fat cells isolated from epididymis of the animals. Adipose tissue of the rat epididymal fat pad had been widely used in this study. The epididymal rat adipose tissue was incubated in glucose containing media and the glucose uptake was measured by the glucose concentration of the media or the oxygen consumption in Warburg vessels. Glucose uptake by incubated rat epididymal adipose tissue was predominantly regulated by the rate of glucose transport across the cell membrane. Another type of experiment was done with the radio labelled glucose where $^{14}\text{CO}_2$ was captured and counted. The $^{14}\text{CO}_2$ produced from the Iodine labelled ^{14}C glucose was measured in

this method [55,56]. For the small amount of insulin quantification, a manometric assay was used to calculate the total gas exchange [57,58]. Later on, some modification in this method was done in various studies. Among them, the isolated fat cells, 3T3-L1 adipocytes and primary cultured adipocytes were found to be very effective in testing insulin-like activity through *in vitro* models [59].

4. Recommendations of animal models for evaluating antidiabetic activities

Both *in vivo* and *in vitro* models have their own advantages over other models. For antidiabetic research, the *in vitro* models are more cell specific and less time consuming. The effects could be easily analysed by various assays. In contrast to that, the *in vivo* methods are more time consuming and costly. Additionally, it needs a lot of expertise to

Table 3

Comparison of streptozotocin and alloxan.

	Streptozotocin	Alloxan
Chemical name	2-Deoxy-2- ([(methylnitrosoamino)carbonyl]amino) D-glucopyranose	2,4,5,6-Tetraoxypyrimidine; 2,4,5,6-pyrimidinetetrone
Chemical characterization	Cytotoxic methylnitrosourea moiety (N-methyl-N-nitrosourea) attached to the glucose (2-deoxyglucose) molecule; glucosamine derivative	Oxygenated pyrimidine derivative; barbituric acid derivative (5ketobarbituric acid)
Chemical properties	<ol style="list-style-type: none"> 1. Hydrophilic, beta cell-toxic glucose analogue 2. Relatively stable at pH 7.4 and 37 °C (at least for up to 1 h) 	<ol style="list-style-type: none"> 1. Very hydrophilic, beta cell-toxic glucose analogue (partition coefficient – 1.8); weak acid 2. Chemically unstable (half-life of 1.5 min at pH 7.4 and 37 °C, decomposing to alloxanic acid); stable at acid pH
Chemical reactivities	<ol style="list-style-type: none"> 1. DNA alkylating agent 2. Protein alkylating agent 3. NO donor 	<ol style="list-style-type: none"> 1. Thiol reagent that is reduced to dialuric acid in the presence of GSH and other thiols 2. A protoxin; intracellular metabolism of this xenobiotic generates toxic ROS through redox cycling with dialuric acid over a long period of time (> 1 h) 3. Compound 305, a non-toxic alloxan-GSH adduct of unknown structure with a characteristic absorbance at a wavelength of 305 nm; a small amount is formed during each redox cycle
Mode of toxicity	DNA alkylation	Generation of ROS
Dose (mg/kg)	Rat: 35–65 (i.v./i.p.) Mice: 100–200 (i.v./i.p.) Hamster: 50 (i.p.) Dog: 20–30 (i.v.) Pig: 100–150 (i.v.) Primates: 50–150 (i.v.)	Rat: 40–200 (i.v./i.p.) Mice: 50–200 (i.v./i.p.) Rabbit: 100–150 (i.v.) Dog: 50–75 (i.v.)

i.v.: intravenous; i.p.: intraperitoneal.

Table 4
Medicinal plants with antidiabetic effects.

No	Plant name	Family	Type of extract	Part used	Activity and references
1	<i>Alangium lamarkii</i>	Alangiaceae	Alcoholic	Leaf	Antidiabetic [84]
2	<i>Albizia odoratissima</i>	Mimosaceae	Methanol	Bark	Antidiabetic [85]
3	<i>Axonopus compressus</i>	Poaceae	Methanol	Leaf	Antidiabetic [86]
4	<i>Berberis vulgaris</i>	Berberidaceae	Aqueous	Root	Hypoglycaemic [87]
5	<i>Brassica juncea</i>	Cruciferae	Aqueous	Seed	Hypoglycemic [88]
6	<i>Caesalpinia digyna</i>	Fabaceae	Methanol	Root	Antidiabetic [89]
7	<i>Catharanthus roseus</i>	Apocynaceae	Methanol	Leaf	Hypoglycemic [90]
8	<i>Centaurium erythraea</i>	Gentianaceae	Aqueous	Leaf	Antidiabetic [91]
9	<i>Chaenomeles sinensis</i>	Rosaceae	Ethyl acetate	Fruit	Antidiabetic [92]
10	<i>Cocos nucifera</i>	Arecaceae	Hydro-methanol	Leaf	Antihyperglycaemic [93]
11	<i>Costus speciosus</i>	Costaceae	Hexane	Rhizome	Antidiabetic [94]
12	<i>Cyclocarya paliurus</i>	Cyclocaryaceae	Aqueous, petroleum ether, chloroform, ethyl acetate & n-butanol	Bark	Hypoglycaemic [95]
13	<i>Dillenia indica</i>	Dilleniaceae	Methanol	Leaf	Antidiabetic [96]
14	<i>Embelia ribes</i>	Myrsinaceae	Hexane	Berries	Antidiabetic [97]
15	<i>Hybanthus enneaspermus</i>	Violaceae	Alcoholic	Whole plant	Antidiabetic [98]
16	<i>Lippa nodiflora</i>	Verbenaceae	Methanol	Whole plant	Antidiabetic and hypolipidemic [99]
17	<i>Lithocarpus polystachyus</i>	Fagaceae	Ethanol & Aqueous	Leaf	Hypoglycaemic [100]
18	<i>Marrubium vulgare</i>	Lamiaceae	Methanol	Aerial parts	Hyperglycaemic and dyslipidemic [101]
19	<i>Ocimum sanctum</i>	Lamiaceae	Hydroalcoholic	Aerial parts	Antidiabetic [102]
20	<i>Opuntia streptacantha</i>	Cactaceae	Ethanol	Leaf	Antihyperglycaemic [103]
21	<i>Psidium guajava</i>	Myrtaceae	Ethanol	Fruit	Antihyperglycaemic [104]
22	<i>Semecarpus anacardium</i>	Anacardiaceae	Milk	Nut	Antidiabetic [105]
23	<i>Prosopis glandulosa</i>	Fabaceae	Gelatine/Jelly	Whole plant	Antidiabetic [106]
24	<i>Ophiopogon japonicus</i>	Asparagaceae	Ethanol	Root	Hypoglycaemic [107]
26	<i>Setaria italica</i>	Poaceae	Aqueous	Seed	Antihyperglycaemic [108]
25	<i>Solanum torvum</i>	Solanaceae	Methanol	Fruit	Antihyperglycaemic [109]
26	<i>Cassia auriculata</i>	Caesalpinaceae	Aqueous	Leaf	Antihyperglycaemic [110]
27	<i>Zygophyllum album</i>	Zygophyllaceae	Ethanol	Whole plant	Antidiabetic [111]
28	<i>Vitex negundo</i>	Lamiaceae	Methanol	Leaf	Antihyperglycaemic [112]
29	<i>Viscum schimperi</i>	Viscaceae	Methanol	Aerial parts	Antihyperglycemic & Hypolipidaemic [113]
30	<i>Symplocos cochinchinensis</i>	Symplocaceae	Hexane	Leaf	Antidiabetic [114]
31	<i>Enicostemma littorale</i>	Gentianaceae	Aqueous	Whole plant	Antidiabetic [115]
32	<i>Vaccinium arctostaphylos</i>	Ericaceae	Ethanol	Fruit	antidiabetic [116]
33	<i>Solanum xanthocarpum</i>	Solanaceae	Aqueous and methanol	Leaf	Antihyperglycaemic [117]

effectively perform the *in vivo* experiments to obtain reproducible data. However, the human body system is a complex system which cannot be revealed by *in vitro* assays only. For that reason, the efficacy of the molecules of samples should also be tested through *in vivo* system for better understanding. In conclusion, the *in vitro* animal models could specially be recommended for mechanism specific research and also primary stage research to find out the exact target molecule or receptor. However, the *in vivo* models are recommended for the further studies to develop a drug as well as to evaluate toxicological profiles.

5. Medicinal plants with antidiabetic effects

Medicinal plants continue to be an important therapeutic support for alleviating disorders of humankind. Following is the list of some important traditional medicinal plants that have recently been scientifically proven for their antidiabetic potential. These plants may further provide lead for the discovery of new plant based antidiabetic agents with safe profiles (Table 4).

6. Role of flavonoids as an antidiabetic agent

Flavonoids are considered as one of the most abundant secondary metabolites of the plant. Approximately, more than 8000 unique flavonoids from different plants have so far been isolated and characterized through various isolation and spectroscopic techniques [60]. In addition to that, the flavonoids are mostly found in fruits, vegetables, nuts, seeds, stem, flowers, etc. [61]. The flavonoids are low molecular weight compounds and have been reported to exert various important pharmacological effects including the cell synthesis [62–65]. Moreover, the scientific claims for flavonoids as a cure of diabetes is noteworthy owing to its wide range of mechanisms of action [66].

Briefly, diabetes mellitus has been stated to hinder the protein, carbohydrate and fat metabolism which result in hyperglycaemia [67]. The first line of therapeutic measurement towards DM is to keep the blood glucose level normal and attain glucose homeostasis. The attempt to achieve glucose homeostasis is quite successful at present but the therapeutic agents are not free from side effects and in some cases, have been proven extremely toxic with high mortality. In this circumstance, the worlds scientists are looking for a better alternative to cure DM [68]. The flavonoids are yet to be commercialized; however, they have gained the prime attention of healthcare professionals for their potentials in the ailment of diabetes [69]. The presence of flavonoids has been confirmed vigorously in the plants which have potential antidiabetic effects. However, a plant contains thousands of compounds and most of the compounds are yet to be isolated and characterized. In this regard, more sincere research efforts must be orchestrated to discover more efficacious antidiabetic agents to help mankind.

Recently, various experimental techniques have been usually used to evaluate the antidiabetic activity of traditional medicinal plants containing flavonoids as one of the major phytoconstituents. Firstly, the experiments are carried out with the crude extract of the medicinal plant to confirm the bioactivity and then subsequently with the isolated flavonoids to confirm active principles responsible for the antidiabetic effect of the particular traditional medicinal plant. Table 5 describes about the research studies in which flavonoids were found as an active principles responsible for the aforementioned effect.

7. Therapeutic approach towards diabetes

Very often the therapeutic approach of diabetes is quite costly and most of the times they are not free from various deleterious effects. The most common and severe side effect associated with antidiabetic agents

Table 5
Some therapeutic approaches in animal model to treat diabetes mellitus by the isolated flavonoids.

Therapeutic approach	Flavonoids	Result
1. <i>Averrhoa carambola</i> isolated flavonoid on glycogen synthesis and insulin secretion	Apigenin-6-C-l-fucopyranoside	Increased insulin secretion and potentiated glucose-induced insulin secretion in hyperglycemic rats [118]
2. <i>Acacia auriculiformis</i> poly flavonoids on antidiabetic factor	Gallic acid, caffeic acid, catechin, rutin, quercetin, myricetin and kaempferol	Significant suppression on α -amylase and -glucosidase [119]
3. Effect of <i>Bauhinia forficata</i> on serum glucose	Kaempferol-3,7-O-(α)-rhamnoside	Hypoglycemic effect was observed in alloxan induced diabetic rats [120]
4. Icarin in diabetic rats on mitochondrial oxidative stress	Icarin	Improvement in overall complication of diabetes in streptozotocin induced rats [121]
5. Kaempferitrin on glucose uptake on skeletal muscles	Kaempferitrin	Insulin-mimetic role of kaempferitrin in glucose homeostasis [122]
6. Isolates from <i>Euonymus alatus</i> in glucose uptake	Kaempferol and quercetin	Potential ameliorated hyperglycemia observed, due to glucose uptake in mature 3T3-L1 adipocytes [123]
7. An isolate exposure on chronic hyperglycemic-cell	Kaempferol	Restoration of high glucose-attenuated intracellular cAMP and ATP production [124]
8. Flavonoid isolated as insulinomimetic on glycemia and glucose uptake	Kaempferitrin	Lowering blood glucose level and ^{14}C uptake of glucose in peripheral muscles of rat [125]
9. Inhibition of intestinal and renal Na-glucose cotransporter	Naringenin	Inhibition of intestinal glucose uptake and renal glucose reabsorption was observed [126]
10. Combined effect of flavonoid in type 2 diabetic rat	Naringin and hesperidin	Synergistic insulin releasing effect [127]
11. Antidiabetic activity of isolated in normoglycemic rats	Naringin	Diminishing the carbohydrate absorption from intestine and reduced post prandial glucose levels [128]
12. Antidiabetic activity of isolated flavonoids from <i>Tetracera indica</i> Merr.	Wogonin, norwogonin and techtochrysin	Enhancement of insulin like activity and insulin sensitizing activity in 3T3-L1 preadipocyte and adipocyte [129]

is hypoglycaemia. Gaining weight is also considered as severe effect particularly associated with sulfonylurea therapy due to excessive insulin release [70]. The risk of developing excessive hypoglycaemic condition is more associated with the use of sitagliptin or linagliptin in combination with sulfonylurea or insulin therapy. Other side effects can also occur due to DPP-4 inhibitors (dipeptidyl peptidase – IV inhibitors) such as – headache, nausea, nasopharyngitis and hypersensitivity or skin reaction [71]. With alpha-glucosidase inhibitors treatment mild symptoms may also occur such as flatulence and diarrhoea. Metformin might cause some mild adverse effects like mild anorexia, nausea, diarrhoea, abdominal discomfort and mostly are gastrointestinal. The existing anti-diabetic drugs have both mild and severe adverse effects like hypoglycaemia, weight gain, gastrointestinal effects, headache, nausea, abdominal discomfort and so on with prolonged use [72]. Thus, it is urgent need to find out more safe and effective classes of anti-diabetic medications to counteract the aforesaid problems associated with current regimens [73]. Furthermore, diabetes is comorbid in nature which makes the treatment more expensive. Consequently, financially well-off people are getting more interested in alternative medicines (e.g., herbal medicines) [74]. Medicine prepared from natural plant is considered to be free of side effects and also less toxic compared to the synthetic medicine [75]. With respect to treat or manage diabetes, several traditional plants have been effectively used by people across the world for centuries. More than 1200 plant species are utilized to treat diabetes, however, a very limited number of them have been reported to exhibit good hypoglycaemic effect while evaluating their antidiabetic properties through experimental animal models and in clinical experiments using human subjects [1]. The medicinal plant species serve as beneficial outcome of oral hypoglycaemic agents to develop new pharmaceutical leads and also provide important sources to existing therapies as a dietary supplement [70].

8. Insulin-like and insulin-sensitizing activity assay

This study is designed to evaluate the insulin-like activity and insulin-sensitizing activity of the extract, fractions, and isolated compounds. Some of the currently available techniques are given in Table 6. In short, the hypoglycemic activity is determined by blood sugar determination of the rodent [76] and also by hypoglycaemic seizures in mice according to European Pharmacopoeia, 3rd edition 1997. In other study, cells and tissues are used to evaluate insulin-like activity by

Table 6
Assay of insulin and insulin-like activity.

Bioactivity	Methods
Hypoglycemic effects	1. Blood sugar determinations in rodent [76,130], 2. Hypoglycemic seizures in mice [131]
Insulin target cells/tissues	1. Epididymal fat pad [77], 2. Fat cells isolated by digestion with collagenase [78], 3. 3T3-L1 adipocytes [79], 4. Assays of glucose transport, (a) Total glucose uptake [132], (b) Transport of 2-deoxy-glucose [80], (c) 2-NBDG uptake [129] 5. Adipocyte plasma [133]

several methods, epididymal fat pad [77], fat cells isolated by digestion with collagenase [78], 3T3-L1 adipocytes [79], glucose transport assays [80], etc.

Various research studies have strongly emphasised that there is a need to discover the better alternative of insulin to reduce the treatment cost as well as safety reasons. The natural products are considered as the potential source of that alternative, but the better alternative is yet to be discovered. In this regard, all these aforementioned animal models (Table 1) have been effectively used to investigate the potential bioactivity of the natural or synthetic products in the absence of insulin [2,10,12].

9. Prospect of medicinal plants as an antidiabetic agent

Since the disease itself is complex so a single animal model is not enough to investigate the underlying cause of diabetes. At the same time, the traditionally used plants cannot be successfully introduced to the market without a thorough investigation of all possible mechanisms reported [3,81,4].

A recent study carried out by Tufts University Centre shows that developing a novel therapeutic drug costs \$2.6 billion and 12–14 years on an average till it receives the Food and Drug administrations (FDA) approval which is 145% increase from 2003. However, to make the medicinal plants commercially available for people is the ultimate goal of natural product research. Therefore, a very comprehensive study is

essential to reach the goal. A thorough literature review of all reported antidiabetic plants is the first stepping stone for natural product research. The plant list should be narrowed down according to certain criteria viz., traditional uses, hypoglycaemic activity proved experimentally, evidence of less toxicity reported scientifically and abundance of plant naturally [82,83].

10. Conclusion

The review has documented a wide range of animal models and medicinal plants used for investigating the antidiabetic activity. The diabetes mellitus is a complex disorder and the human physiology is very complex. Hence, the aim of this literature-based study was to review the existing animal models in the literature along with the potential of medicinal plants with respect to discovering a new alternative to existing drugs. Moreover, the paper also states the drawbacks of utilizing certain animal models and focuses on the specific animal model for suitable investigation. However, the paper suggests more in-depth review on natural product research for the ailment of diabetes that is needed. Finally, the review draws attention of the researchers that a proper documentation of all reported antidiabetic medicinal plants is still warranted in regard to finding out more efficacious antidiabetic agents.

Conflict of interest

The authors have declared that there is no conflict of interest.

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References

- [1] M. Eddouks, A. Lemhadri, J.B. Michel, Hypolipidemic activity of aqueous extract of *Capparis spinosa* L. in normal and diabetic rats, *J. Ethnopharmacol.* 98 (3) (2005) 345–350.
- [2] M. Bnouham, A. Ziyat, H. Mekhfi, A. Tahri, A. Legssyer, Medicinal plants with potential antidiabetic activity—a review of ten years of herbal medicine research (1990–2000), *Int. J. Diabetes Metab.* 14 (1) (2006) 1.
- [3] K. Srinivasan, P. Ramarao, Animal models in type 2 diabetes research: an overview, *Indian J. Med. Res.* 125 (3) (2007) 451.
- [4] L. Vedtofte, T.B. Bodvarsdottir, C.F. Gotfredsen, A.E. Karlens, L.B. Knudsen, R.S. Heller, Liraglutide, but not vildagliptin, restores normoglycaemia and insulin content in the animal model of type 2 diabetes, *Psammomys obesus*, *Regul. Pept.* 160 (1) (2010) 106–114.
- [5] J. Xiao, Y. Lv, S. Lin, L. Jin, Y. Zhang, X. Wang, J. Ma, K. Hu, W. Feng, L. Cai, Cardiac protection by basic fibroblast growth factor from ischemia/reperfusion-induced injury in diabetic rats, *Biol. Pharm. Bull.* 33 (3) (2010) 444–449.
- [6] G.S. Hotamisligil, P. Peraldi, A. Budavari, R. Ellis, IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF- α - and obesity-induced insulin resistance, *Science* 271 (5249) (1996) 665.
- [7] G.Z. Wu, G. Hong, W.P. Zhang, H.B. Zhang, Effect of 1-[4-(2-(4-bromobenzene-sulfonylamino) ethyl) phenylsulfonyl]-3-(trans-4-methylcyclohexyl) urea (i4), a new synthetic sulfonylurea compound, on glucose metabolism in vivo and in vitro, *Arzneimittelforschung* 59 (11) (2009) 550–556.
- [8] S. Ro, C. Park, J. Jin, H. Zheng, P.J. Blair, D. Redelman, S.M. Ward, W. Yan, K.M. Sanders, A model to study the phenotypic changes of interstitial cells of Cajal in gastrointestinal diseases, *Gastroenterology* 138 (3) (2010) 1068–1078 e2.
- [9] D. Wei, J. Li, M. Shen, W. Jia, N. Chen, T. Chen, D. Su, H. Tian, S. Zheng, Y. Dai, Cellular production of n-3 PUFAs and reduction of n-6-to-n-3 ratios in the pancreatic β -cells and islets enhance insulin secretion and confer protection against cytokine-induced cell death, *Diabetes* 59 (2) (2010) 471–478.
- [10] E. Seo, E.-J. Park, Y. Joe, S. Kang, M.-S. Kim, S.-H. Hong, M.-K. Park, D.K. Kim, H. Koh, H.-J. Lee, Overexpression of AMPK α 1 ameliorates fatty liver in hyperlipidemic diabetic rats, *Korean J. Physiol. Pharmacol.* 13 (6) (2009) 449–454.
- [11] D.V. Serreze, M. Niens, J. Kulik, T.P. DiLorenzo, Bridging mice to men: using HLA transgenic mice to enhance the future prediction and prevention of autoimmune type 1 diabetes in humans, *Mouse Models Drug Discov.: Methods Protoc.* (2010) 119–134.
- [12] V. Sordi, R. Melzi, A. Mercalli, R. Formicola, C. Doglioni, F. Tiboni, G. Ferrari, R. Nano, K. Chwalek, E. Lammert, Mesenchymal cells appearing in pancreatic tissue culture are bone marrow-derived stem cells with the capacity to improve transplanted islet function, *Stem Cells* 28 (1) (2010) 140–151.
- [13] S. Sugii, P. Olson, D.D. Sears, M. Saberi, A.R. Atkins, G.D. Barish, S.-H. Hong, G.L. Castro, Y.-Q. Yin, M.C. Nelson, PPAR γ activation in adipocytes is sufficient for systemic insulin sensitization, *Proc. Natl. Acad. Sci.* 106 (52) (2009) 22504–22509.
- [14] A.M. Stranahan, T.V. Arumugam, K. Lee, M.P. Mattson, Mineralocorticoid receptor activation restores medial perforant path LTP in diabetic rats, *Synapse* 64 (7) (2010) 528–532.
- [15] M.L. Sugrue, K.R. Vella, C. Morales, M.E. Lopez, A.N. Hollenberg, The thyrotropin-releasing hormone gene is regulated by thyroid hormone at the level of transcription in vivo, *Endocrinology* 151 (2) (2010) 793–801.
- [16] E. Fernández-Millán, M.N. Gangnerau, L. De Miguel-Santos, S. Calderari, P. Serradas, F. Escrivá, B. Portha, C. Álvarez, Undernutrition of the GK rat during gestation improves pancreatic IGF-2 and beta-cell mass in the fetuses, *Growth Factors* 27 (6) (2009) 409–418.
- [17] H. Matsui-Inohara, H. Uematsu, T. Narita, K. Satoh, H. Yonezawa, K. Kuroda, T. Ito, S. Yoneda, T. Kawarai, H. Sugiyama, E2F-1-deficient NOD/SCID mice developed showing decreased saliva production, *Exp. Biol. Med.* 234 (12) (2009) 1525–1536.
- [18] Y. Matsumoto, K. Torimoto, H. Matsuyoshi, A. Hirayama, K. Fujimoto, N. Yoshimura, Y. Hirao, Long-term effects of diabetes mellitus on voiding function in a new model of type 2 diabetes mellitus, the Spontaneously Diabetic Torii (SDT) rat, *Biomed. Res.* 30 (6) (2009) 331–335.
- [19] G. Kretschmer, D. Sutherland, A. Matas, T. Cain, J. Najarian, Autotransplantation of pancreatic islets without separation of exocrine and endocrine tissue in totally pancreatectomized dogs, *Surgery* 82 (1) (1977) 74–81.
- [20] T. Stewart, B. Hultgren, X. Huang, S. Pitts-Meek, J. Hully, N. MacLachlan, Induction of type I diabetes by interferon- γ in transgenic mice, *Science-New York Then Washington* 260 (1993) 1942.
- [21] I.V. Hutchinson, An endothelin-transforming growth factor beta pathway in the nephrotoxicity of immunosuppressive drugs, *Curr. Opin. Nephrol. Hypertens.* 7 (6) (1998) 665–672.
- [22] T. Szkudelski, The mechanism of alloxan and streptozotocin action in β cells of the rat pancreas, *Physiol. Res.* 50 (6) (2001) 537–546.
- [23] S. Lenzen, The mechanisms of alloxan- and streptozotocin-induced diabetes, *Diabetologia* 51 (2) (2008) 216–226.
- [24] M.S. Islam, d.T. Loots, Experimental rodent models of type 2 diabetes: a review, *Methods Find. Exp. Clin. Pharmacol.* 31 (4) (2009) 249–261.
- [25] J. Han, Y. Liu, Reduction of islet pyruvate carboxylase activity might be related to the development of type 2 diabetes mellitus in agouti-K mice, *J. Endocrinol.* 204 (2) (2010) 143–152.
- [26] A. Higuchi, K. Ohashi, R. Shibata, S. Sono-Romanelli, K. Walsh, N. Ouchi, Thiazolidinediones reduce pathological neovascularization in ischemic retina via an adiponectin-dependent mechanism, *Arterioscler. Thromb. Vasc. Biol.* 30 (1) (2010) 46–53.
- [27] H. Hotta, T. Miura, T. Miki, N. Togashi, T. Maeda, S.J. Kim, M. Tanno, T. Yano, A. Kuno, T. Itoh, Short communication: Angiotensin II type 1 receptor-mediated upregulation of calcineurin activity underlies impairment of cardioprotective signaling in diabetic hearts, *Circ. Res.* 106 (1) (2010) 129–132.
- [28] E. Karunanayake, J.R. Baker, R. Christian, D. Hearse, G. Mellows, Autoradiographic study of the distribution and cellular uptake of (14C)-streptozotocin in the rat, *Diabetologia* 12 (2) (1976) 123–128.
- [29] H. Tjälve, E. Wilander, E.-B. Johansson, Distribution of labelled streptozotocin in mice: uptake and retention in pancreatic islets, *J. Endocrinol.* 69 (3) (1976) 455–NP.
- [30] S.P. Ledoux, G.L. Wilson, Effects of streptozotocin on a clonal isolate of rat insulinoma cells, *Biochim. Biophys. Acta (BBA) Mol. Cell Res.* 804 (4) (1984) 387–392.
- [31] W.J. Schnedl, S. Ferber, J.H. Johnson, C.B. Newgard, STZ transport and cytotoxicity: specific enhancement in GLUT2-expressing cells, *Diabetes* 43 (11) (1994) 1326–1333.
- [32] M. Elsner, B. Guldbakke, M. Tiedge, R. Munday, S. Lenzen, Relative importance of transport and alkylation for pancreatic beta-cell toxicity of streptozotocin, *Diabetologia* 43 (12) (2000) 1528–1533.
- [33] H. Yamamoto, Y. Uchigata, H. Okamoto, Streptozotocin and alloxan induce DNA strand breaks and poly (ADP-ribose) synthetase in pancreatic islets, *Nature* (1981).
- [34] Y. Uchigata, H. Yamamoto, A. Kawamura, H. Okamoto, Protection by superoxide dismutase, catalase, and poly (ADP-ribose) synthetase inhibitors against alloxan- and streptozotocin-induced islet DNA strand breaks and against the inhibition of proinsulin synthesis, *J. Biol. Chem.* 257 (11) (1982) 6084–6088.
- [35] M. Murata, A. Takahashi, I. Saito, S. Kawanishi, Site-specific DNA methylation and apoptosis: induction by diabetogenic streptozotocin, *Biochem. Pharmacol.* 57 (8) (1999) 881–887.
- [36] A.A. Pieper, A. Verma, J. Zhang, S.H. Snyder, Poly(ADP-ribose) polymerase, nitric oxide and cell death, *Trends Pharmacol. Sci.* 20 (4) (1999) 171–181.
- [37] J. Turk, J.A. Corbett, S. Ramanadham, A. Bohrer, M.L. McDaniel, Biochemical evidence for nitric oxide formation from streptozotocin in isolated pancreatic islets, *Biochem. Biophys. Res. Commun.* 197 (3) (1993) 1458–1464.
- [38] C.A. Delaney, A. Dunger, M. Di Matteo, J.M. Cunningham, M.H. Green, I.C. Green, Comparison of inhibition of glucose-stimulated insulin secretion in rat islets of langerhans by streptozotocin and methyl and ethyl nitrosoureas and methanesulphonates: lack of correlation with nitric oxide-releasing or o6-alkylating ability,

- Biochem. Pharmacol. 50 (12) (1995) 2015–2020.
- [39] M. Nukatsuka, Y. Yoshimura, M. Nishida, J. Kawada, Allopurinol protects pancreatic β cells from the cytotoxic effect of streptozotocin: in vitro study, *J. Pharmacobiodyn.* 13 (4) (1990) 259–262.
 - [40] R. Munday, Dialuric acid autooxidation: effects of transition metals on the reaction rate and on the generation of “active oxygen” species, *Biochem. Pharmacol.* 37 (3) (1988) 409–413.
 - [41] C.C. Winterbourn, R. Munday, Glutathione-mediated redox cycling of alloxan: mechanisms of superoxide dismutase inhibition and of metal-catalyzed oh formation, *Biochem. Pharmacol.* 38 (2) (1989) 271–277.
 - [42] C.C. Winterbourn, W.B. Cowden, H.C. Sutton, Auto-oxidation of dialuric acid, divicine and isouramil: superoxide dependent and independent mechanisms, *Biochem. Pharmacol.* 38 (4) (1989) 611–618.
 - [43] C. Liang, K. DeCourcy, M.R. Prater, High-saturated-fat diet induces gestational diabetes and placental vasculopathy in C57BL/6 mice, *Metabolism* 59 (7) (2010) 943–950.
 - [44] Y. Wang, G. Fu, F. Chen, H. Wang, The effect of valsartan and fluvastatin on the connective tissue growth factor expression in experimental diabetic cardiomyopathy, *Zhonghua Nei Ke Za Zhi* 48 (8) (2009) 660–665.
 - [45] S. Bonner-Weir, D. Trent, G. Weir, Partial pancreatectomy in the rat and subsequent defect in glucose-induced insulin release, *J. Clin. Invest.* 71 (6) (1983) 1544.
 - [46] H.G. Vogel, *Antidiabetic Activity*, Springer, 2008, pp. 1323–1607.
 - [47] G. Müller, Methods to Induce Experimental Diabetes Mellitus, in: F. Hock (Ed.), *Drug Discovery and Evaluation: Pharmacological Assays*, Springer, Cham, 2016, pp. 2569–2581.
 - [48] A. Spasov, A. Kucheriavenko, O. Salaznikova, Effect of hypoglycemic drugs on hemorheological parameters, *Eksp. Klin. Farmakol.* 72 (5) (2008) 31–34.
 - [49] M. Pravenec, Use of rat genomics for investigating the metabolic syndrome, *Rat Genomics: Methods Protoc.* (2010) 415–426.
 - [50] S. Renner, C. Fehlings, N. Herbach, A. Hofmann, D.C. von Waldhausen, B. Kessler, K. Ulrichs, I. Chodnevskaja, V. Moskalenko, W. Amselgruber, Glucose intolerance and reduced proliferation of pancreatic β -cells in transgenic pigs with impaired glucose-dependent insulinotropic polypeptide function, *Diabetes* 59 (5) (2010) 1228–1238.
 - [51] J.E. Craighead, Current views on the etiology of insulin-dependent diabetes mellitus, *N. Engl. J. Med.* 299 (26) (1978) 1439–1445.
 - [52] D.J. Giron, S. Cohen, S. Lyons, M. Trombley, C. Gould, Virus-induced diabetes mellitus in ICR Swiss mice is age dependent, *Infect. Immun.* 41 (2) (1983) 834–836.
 - [53] J. Yoon, P. McClintock, T. Onodera, A. Notkins, Virus-induced diabetes mellitus. XVIII. Inhibition by a nondiabetogenic variant of encephalomyocarditis virus, *J. Exp. Med.* 152 (4) (1980) 878–892.
 - [54] C.L. Gould, K.G. McMannama, N.J. Bigley, D.J. Giron, Virus-induced murine diabetes: enhancement by immunosuppression, *Diabetes* 34 (12) (1985) 1217–1221.
 - [55] P. Björntorp, The effect of insulin in vitro on human adipose tissue from normal and diabetic subjects, *J. Intern. Med.* 181 (4) (1967) 389–402.
 - [56] M. Kobayashi, J.M. Olefsky, Effects of streptozotocin-induced diabetes on insulin binding, glucose transport, and intracellular glucose metabolism in isolated rat adipocytes, *Diabetes* 28 (2) (1979) 87–95.
 - [57] E.G. Ball, M.A. Merrill, A manometric assay of insulin and some results of the application of the method to sera and islet containing tissues, *Endocrinology* 69 (3) (1961) 596–607.
 - [58] W.I. Morse, J.S. Soeldner, The measurement of human adipose tissue mass, *Compr. Physiol.* (2011).
 - [59] F. Liu, J.-k. Kim, Y. Li, X.-q. Liu, J. Li, X. Chen, An extract of *Lagerstroemia speciosa* L. has insulin-like glucose uptake-stimulatory and adipocyte differentiation-inhibitory activities in 3T3-L1 cells, *J. Nutr.* 131 (9) (2001) 2242–2247.
 - [60] H.d. Groot, U. Rauen, Tissue injury by reactive oxygen species and the protective effects of flavonoids, *Fundam. Clin. Pharmacol.* 12 (3) (1998) 249–255.
 - [61] N. Cook, S. Samman, Flavonoids-chemistry, metabolism, cardioprotective effects, and dietary sources, *J. Nutr. Biochem.* 7 (2) (1996) 66–76.
 - [62] P.H. Hollman, M. Katan, Dietary flavonoids: intake, health effects and bioavailability, *Food Chem. Toxicol.* 37 (9) (1999) 937–942.
 - [63] K.E. Heim, A.R. Tagliaferro, D.J. Bobilya, Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships, *J. Nutr. Biochem.* 13 (10) (2002) 572–584.
 - [64] T.T. Cushnie, A.J. Lamb, Antimicrobial activity of flavonoids, *Int. J. Antimicrob. Agents* 26 (5) (2005) 343–356.
 - [65] S.P. Fernández, C. Wasowski, L.M. Loscalzo, R.E. Granger, G.A. Johnston, A.C. Paladini, M. Marder, Central nervous system depressant action of flavonoid glycosides, *Eur. J. Pharmacol.* 539 (3) (2006) 168–176.
 - [66] J.C. Patra, B.H. Chua, Artificial neural network-based drug design for diabetes mellitus using flavonoids, *J. Comput. Chem.* 32 (4) (2011) 555–567.
 - [67] K. Alberti, P. Zimmet, New diagnostic criteria and classification of diabetes-again? *Diabet. Med.* 15 (7) (1998) 535–536.
 - [68] M. Aslan, D.D. Orhan, N. Orhan, E. Sezici, E. Yesilada, *In vivo* antidiabetic and antioxidant potential of *Helichrysum plicatum* ssp. *plicatum* capitulum in streptozotocin-induced-diabetic rats, *J. Ethnopharmacol.* 109 (1) (2007) 54–59.
 - [69] S. Mohan, L. Nandhakumar, Role of various flavonoids: hypotheses on novel approach to treat diabetes, *J. Med. Hypotheses Ideas* 8 (1) (2014) 1–6.
 - [70] C.J. Bailey, C. Day, Traditional plant medicines as treatments for diabetes, *Diabetes Care* 12 (8) (1989) 553–564.
 - [71] K. Goossen, S. Graber, Longer term safety of dipeptidyl peptidase-4 inhibitors in patients with type 2 diabetes mellitus: systematic review and meta-analysis, *Diabetes Obes. Metab.* 14 (12) (2012) 1061–1072.
 - [72] M.E. Cobble, A.L. Peters, Clinical practice in type 2 diabetes: after metformin and lifestyle, then what? *J. Fam. Pract.* 58 (11) (2009) S7–S7.
 - [73] J.S. Gabrielsen, Y. Gao, J.A. Simcox, J.Y. Huang, D. Thorup, D. Jones, R.C. Cooksey, D. Gabrielsen, T.D. Adams, S.C. Hunt, P.N. Hopkins, W.T. Cefalu, D.A. McClain, Adipocyte iron regulates adiponectin and insulin sensitivity, *J. Clin. Invest.* 122 (10) (2012) 3529–3540.
 - [74] Q.U. Ahmed, A. Umar, M. Taher, D. Susanti, M.Z.A.M. Amiroudine, J. Latip., Phytochemical investigation of the leaves of *Tetracera scandens* Linn and in vitro antidiabetic activity of hypoletin Proceedings of the International Conference on Science, Technology and Social Sciences (ICSTSS) Springer (2012) 591–608.
 - [75] M. Jung, M. Park, H.C. Lee, Y.H. Kang, E.S. Kang, S.K. Kim, Antidiabetic agents from medicinal plants, *Curr. Med. Chem.* 13 (10) (2006) 1203–1218.
 - [76] S. Sun, N. Liang, H. Piao, H. Yamamoto, Y. Kawashima, F. Cui, Insulin-so (sodium oleate) complex-loaded PLGA nanoparticles: formulation, characterization and in vivo evaluation, *J. Microencapsulation* 27 (6) (2010) 471–478.
 - [77] P.M. Beigelman, I.S. Onoprienko, Insulin-like effects of serum albumin and globulin fractions on glucose uptake by rat epididymal adipose tissue, *Diabetes* 8 (6) (1959) 438–441.
 - [78] M. Rodbell, Metabolism of isolated fat cells. I. Effects of hormones on glucose metabolism and lipolysis, *J. Biol. Chem.* 239 (1964) 375–380.
 - [79] J.C. Santos, R.M. Gotardo, M.T. Brianti, M. Pirace, A. Gambero, M.L. Ribeiro, Effects of yerba maté, a plant extract formulation (“YGD”) and resveratrol in 3T3-L1 adipogenesis, *Molecules* 19 (10) (2014) 16909–16924.
 - [80] S. Gautam, N. Ishrat, R. Singh, T. Narendar, A.K. Srivastava, Aegeline from *Aegle marmelos* stimulates glucose transport via AKT and Rac1 signaling, and contributes to a cytoskeletal rearrangement through PI3K/Rac1, *Eur. J. Pharmacol.* 762 (2015) 419–429.
 - [81] S. Tracy, K. Drescher, J. Jackson, K. Kim, K. Kono, Enteroviruses, type 1 diabetes and hygiene: a complex relationship, *Rev. Med. Virol.* 20 (2) (2010) 106–116.
 - [82] o.D.D. Tufts Center for the Study, T.C. for the Study, Cost to develop and win marketing approval for a new drug is \$ 2.6 billion. http://csdd.tufts.edu/news/complete_story/pr_tufts_csdd_2014_cost_study, 2018 (accessed 26.02.18).
 - [83] A. Henkel, J. Ross, The effect of acquisitions on pharmaceutical drug prices, (2017) https://economics.yale.edu/sites/default/files/files/Undergraduate/Nominated%20Senior%20Essays/2016-17/Alexis%20Henkel_Senior%20Essay.pdf (accessed 28.02.18).
 - [84] R. Kumar, D.K. Patel, S.K. Prasad, D. Laloo, S. Krishnamurthy, S. Hemalatha, Type 2 antidiabetic activity of bergenin from the roots of *Caesalpinia digyna* Rottler, *Fitoterapia* 83 (2) (2012) 395–401.
 - [85] S. Kumar, V. Kumar, O. Prakash, Antidiabetic, hypolipidemic and histopathological analysis of *Dillenia indica* (L.) leaves extract on alloxan induced diabetic rats, *Asian Pac. J. Trop. Med.* 4 (5) (2011) 347–352.
 - [86] B.O. Ibeh, M.I. Ezeaja, Preliminary study of antidiabetic activity of the methanolic leaf extract of *Axonopus compressus* (P. Beauv) in alloxan-induced diabetic rats, *J. Ethnopharmacol.* 138 (3) (2011) 713–716.
 - [87] N. Meliani, M.E.A. Dib, H. Allali, B. Tabti, Hypoglycaemic effect of *Berberis vulgaris* L. in normal and streptozotocin-induced diabetic rats, *Asian Pac. J. Trop. Biomed.* 1 (6) (2011) 468–471.
 - [88] T. Thirumalai, S.V. Therasa, E. Elumalai, E. David, Hypoglycemic effect of *Brassica juncea* (seeds) on streptozotocin induced diabetic male albino rat, *Asian Pac. J. Trop. Biomed.* 1 (4) (2011) 323–325.
 - [89] R. Kumar, D.K. Pate, S.K. Prasad, K. Sairam, S. Hemalatha, Antidiabetic activity of alcoholic leaves extract of *Alangium lamarckii* Thwaites on streptozotocin-nicotinamide induced type 2 diabetic rats, *Asian Pac. J. Trop. Med.* 4 (11) (2011) 904–909.
 - [90] S. Ohadoma, H. Michael, Effects of co-administration of methanol leaf extract of *Catharanthus roseus* on the hypoglycemic activity of metformin and glibenclamide in rats, *Asian Pac. J. Trop. Med.* 4 (6) (2011) 475–477.
 - [91] M. Sefi, H. Fetoui, N. Lachkar, A. Tahraoui, B. Lyousi, T. Boudawara, N. Zeghal, *Centaurea erythraea* (gentianaceae) leaf extract alleviates streptozotocin-induced oxidative stress and β -cell damage in rat pancreas, *J. Ethnopharmacol.* 135 (2) (2011) 243–250.
 - [92] S. Sancheti, S. Sancheti, S.-Y. Seo, Antidiabetic and anticholinesterase effects of ethyl acetate fraction of *Chaenomeles sinensis* (Thouin) Koehne fruits in streptozotocin-induced diabetic rats, *Exp. Toxicol. Pathol.* 65 (1) (2013) 55–60.
 - [93] S. Naskar, U.K. Mazumder, G. Pramanik, M. Gupta, R.S. Kumar, A. Bala, A. Islam, Evaluation of antihyperglycemic activity of *Cocos nucifera* Linn. on streptozotocin induced type 2 diabetic rats, *J. Ethnopharmacol.* 138 (3) (2011) 769–773.
 - [94] J. Eliza, P. Daisy, S. Ignacimuthu, V. Duraipandian, Antidiabetic and antilipidemic effect of eremanthin from *Costus speciosus* (Koen.) Sm., in STZ-induced diabetic rats, *Chem. Biol. Interact.* 182 (1) (2009) 67–72.
 - [95] S. Li, J. Li, X.-L. Guan, J. Li, S.-P. Deng, L.-Q. Li, M.-T. Tang, J.-G. Huang, Z.-Z. Chen, R.-Y. Yang, Hypoglycemic effects and constituents of the barks of *Cyclocarya paliurus* and their inhibiting activities to glucosidase and glycogen phosphorylase, *Fitoterapia* 82 (7) (2011) 1081–1085.
 - [96] D. Kumar, S. Kumar, S. Kohli, R. Arya, J. Gupta, Antidiabetic activity of methanolic bark extract of *Albizia odoratissima* Benth. in alloxan induced diabetic albino mice, *Asian Pac. J. Trop. Med.* 4 (11) (2011) 900–903.
 - [97] S. Mahendran, S. Badami, V. Maithili, Evaluation of antidiabetic effect of embelin from *Embelia ribes* in alloxan induced diabetes in rats, *Biomed. Prev. Nutr.* 1 (1) (2011) 25–31.
 - [98] D. Patel, R. Kumar, S. Prasad, K. Sairam, S. Hemalatha, Antidiabetic and in vitro antioxidant potential of *Hybanthus enneaspermus* (Linn) F. Muell in streptozotocin-induced diabetic rats, *Asian Pac. J. Trop. Biomed.* 1 (4) (2011) 316–322.
 - [99] R. Balamurugan, S. Ignacimuthu, Antidiabetic and hypolipidemic effect of

- methanol extract of *Lippia nodiflora* L. in streptozotocin induced diabetic rats, Asian Pac. J. Trop. Biomed. 1 (1) (2011) S30–S36.
- [100] S.-z. Hou, S.-x. Chen, S. Huang, D.-x. Jiang, C.-J. Zhou, C.-q. Chen, Y.-m. Liang, X.-p. Lai, The hypoglycemic activity of *Lithocarpus polystachyus* Rehd. leaves in the experimental hyperglycemic rats, J. Ethnopharmacol. 138 (1) (2011) 142–149.
- [101] A.A. Elberry, F.M. Harraz, S.A. Ghareib, S.A. Gabr, A.A. Nagy, E. Abdel-Sattar, Methanolic extract of *Marrubium vulgare* ameliorates hyperglycemia and dyslipidemia in streptozotocin-induced diabetic rats, Int. J. Diabetes Mellit. 3 (1) (2015) 37–44.
- [102] R. Patil, R. Patil, B. Ahirwar, D. Ahirwar, Isolation and characterization of anti-diabetic component (bioactivity-guided fractionation) from *Ocimum sanctum* L. (Lamiaceae) aerial part, Asian Pac. J. Trop. Med. 4 (4) (2011) 278–282.
- [103] A. Andrade-Cetto, H. Wiedenfeld, Anti-hyperglycemic effect of *Opuntia streptacantha* Lem, J. Ethnopharmacol. 133 (2) (2011) 940–943.
- [104] C.-S. Huang, M.-C. Yin, L.-C. Chiu, Antihyperglycemic and antioxidative potential of *Psidium guajava* fruit in streptozotocin-induced diabetic rats, Food Chem. Toxicol. 49 (9) (2011) 2189–2195.
- [105] H.B.H. Khan, K.S. Vinayagam, S. Palanivelu, S. Panchanatham, Anti-diabetic effect of *Semecarpus anacardium* Linn nut milk extract in a high fat diet STZ-induced type 2 diabetic rat model, Comp. Clin. Pathol. 21 (6) (2012) 1395–1400.
- [106] C. George, A. Lochner, B. Huisamen, The efficacy of *Prosopis glandulosa* as anti-diabetic treatment in rat models of diabetes and insulin resistance, J. Ethnopharmacol. 137 (1) (2011) 298–304.
- [107] X. Chen, J. Jin, J. Tang, Z. Wang, J. Wang, L. Jin, J. Lu, Extraction, purification, characterization and hypoglycemic activity of a polysaccharide isolated from the root of *Ophiopogon japonicus*, Carbohydr. Polym. 83 (2) (2011) 749–754.
- [108] Y. Sireesha, R.B. Kasetti, S.A. Nabi, S. Swapna, C. Apparao, Antihyperglycemic and hypolipidemic activities of *Setaria italica* seeds in STZ diabetic rats, Pathophysiology 18 (2) (2011) 159–164.
- [109] G.R. Gandhi, S. Ignacimuthu, M.G. Paulraj, P. Sasikumar, Antihyperglycemic activity and antidiabetic effect of methyl caffeate isolated from *Solanum torvum* Swartz. fruit in streptozotocin induced diabetic rats, Eur. J. Pharmacol. 670 (2) (2011) 623–631.
- [110] S. Gupta, S.B. Sharma, U.R. Singh, S.K. Bansal, Salutary effect of *Cassia auriculata* L. leaves on hyperglycemia-induced atherosclerotic environment in streptozotocin rats, Cardiovasc. Toxicol. 11 (4) (2011) 308–315.
- [111] J. El Ghoul, N. Ghanem-Boughanmi, M. Ben-Attia, Biochemical study on the protective effect of ethanolic extract of *Zygophyllum album* on streptozotocin-induced oxidative stress and toxicity in mice, Biomed. Prev. Nutr. 1 (2) (2011) 79–83.
- [112] R. Sundaram, R. Nareesh, P. Shanthi, P. Sachdanandam, Antihyperglycemic effect of iridoid glucoside, isolated from the leaves of *Vitex negundo* in streptozotocin-induced diabetic rats with special reference to glycoprotein components, Phytomedicine 19 (3) (2012) 211–216.
- [113] E.A. Abdel-Sattar, A.A. Elberry, F.M. Harraz, S.A. Ghareib, A.A. Nagy, S.A. Gabr, Antihyperglycemic and hypolipidaemic effects of the methanolic extract of Saudi mistletoe (*Viscum schimperi* Engl.), J. Adv. Res. 2 (2) (2011) 171–177.
- [114] C. Sunil, S. Ignacimuthu, P. Agastian, Antidiabetic effect of *Symplocos cochinchinensis* (Lour.) S. Moore. in type 2 diabetic rats, J. Ethnopharmacol. 134 (2) (2011) 298–304.
- [115] R.D. Sonawane, S.L. Vishwakarma, S. Lakshmi, M. Rajani, H. Padh, R.K. Goyal, Amelioration of STZ-induced type 1 diabetic nephropathy by aqueous extract of *Enicostemma littorale* Blume and swertiamarin in rats, Mol. Cell. Biochem. 340 (1–2) (2010) 1–6.
- [116] A.M. Feshani, S.M. Kouhsari, S. Mohammadi, *Vaccinium arctostaphylos*, a common herbal medicine in Iran: molecular and biochemical study of its antidiabetic effects on alloxan-diabetic wistar rats, J. Ethnopharmacol. 133 (1) (2011) 67–74.
- [117] K. Poongothai, P. Ponnuragan, K.S.Z. Ahmed, B.S. Kumar, S. Sheriff, Antihyperglycemic and antioxidant effects of *Solanum xanthocarpum* leaves (field grown & in vitro raised) extracts on alloxan induced diabetic rats, Asian Pac. J. Trop. Med. 4 (10) (2011) 778–785.
- [118] L.H. Cazarolli, P. Folador, H.H. Moresco, I.M.C. Brighente, M.G. Pizzolatti, F.R.M.B. Silva, Stimulatory effect of apigenin-6-c- β -l-fucopyranoside on insulin secretion and glycogen synthesis, Eur. J. Med. Chem. 44 (11) (2009) 4668–4673.
- [119] A. Sathya, P. Siddhuraju, Role of phenolics as antioxidants, biomolecule protectors and as anti-diabetic factors-evaluation on bark and empty pods of *Acacia auriculiformis*, Asian Pac. J. Trop. Med. 5 (10) (2012) 757–765.
- [120] F.R.M.B. Silva, B. Szpoganicz, M.G. Pizzolatti, M.A.V. Willrich, E. de Sousa, Acute effect of *Bauhinia forficata* on serum glucose levels in normal and alloxan-induced diabetic rats, J. Ethnopharmacol. 83 (1) (2002) 33–37.
- [121] H. Bao, L. Chen, Icarin reduces mitochondrial oxidative stress injury in diabetic rat hearts, Zhongguo Zhong yao za zhi 36 (11) (2011) 1503–1507.
- [122] L.H. Cazarolli, D.F. Pereira, V.D. Kappel, P. Folador, M.d.S.R.B. Figueiredo, M.G. Pizzolatti, F.R.M.B. Silva, Insulin signaling: a potential signaling pathway for the stimulatory effect of kaempferitrin on glucose uptake in skeletal muscle, Eur. J. Pharmacol. 712 (1) (2013) 1–7.
- [123] X.-K. Fang, J. Gao, D.-N. Zhu, Kaempferol and quercetin isolated from *Euonymus alatus* improve glucose uptake of 3T3-L1 cells without adipogenesis activity, Life Sci. 82 (11) (2008) 615–622.
- [124] Y. Zhang, D. Liu, Flavonol kaempferol improves chronic hyperglycemia-impaired pancreatic beta-cell viability and insulin secretory function, Eur. J. Pharmacol. 670 (1) (2011) 325–332.
- [125] A.P. Jorge, H. Horst, E. de Sousa, M.G. Pizzolatti, F.R.M.B. Silva, Insulinomimetic effects of kaempferitrin on glycaemia and on 14C-glucose uptake in rat soleus muscle, Chem. Biol. Interact. 149 (2) (2004) 89–96.
- [126] J.M. Li, C.T. Che, C.B. Lau, P.S. Leung, C.H. Cheng, Inhibition of intestinal and renal Na⁺-glucose cotransporter by naringenin, Int. J. Biochem. Cell Biol. 38 (5) (2006) 985–995.
- [127] A.M. Mahmoud, M.B. Ashour, A. Abdel-Moneim, O.M. Ahmed, Hesperidin and naringin attenuate hyperglycemia-mediated oxidative stress and proinflammatory cytokine production in high fat fed/streptozotocin-induced type 2 diabetic rats, J. Diabetes Complications 26 (6) (2012) 483–490.
- [128] R. Ortiz-Andrade, J. Sánchez-Salgado, G. Navarrete-Vázquez, S. Webster, M. Binnie, S. García-Jiménez, I. León-Rivera, P. Cigarroa-Vázquez, R. Villalobos-Molina, S. Estrada-Soto, Antidiabetic and toxicological evaluations of naringenin in normoglycaemic and NIDDM rat models and its implications on extra-pancreatic glucose regulation, Diabetes Obes. Metab. 10 (11) (2008) 1097–1104.
- [129] M.M. Hasan, Q.U. Ahmed, S.Z.M. Soad, J. Latif, M. Taher, T.M.F. Syafiq, M.N. Sarian, A.M. Alhassan, Z.A. Zakaria, Flavonoids from *Tetracera indica* Merr. induce adipogenesis and exert glucose uptake activities in 3T3-L1 adipocyte cells, BMC Complement. Altern. Med. 17 (1) (2017) 431.
- [130] G. Eneroth, K. Ahlund, Biological assay of insulin by blood sugar determination in mice, Acta Pharm. Suec. 5 (6) (1968) 591–594.
- [131] E. Pharmacopoeia, European Pharmacopoeia, Council of Europe, Strasbourg, 2002.
- [132] A.D. Kohn, S.A. Summers, M.J. Birnbaum, R.A. Roth, Expression of a constitutively active AKT Ser/Thr kinase in 3T3-L1 adipocytes stimulates glucose uptake and glucose transporter 4 translocation, J. Biol. Chem. 271 (49) (1996) 31372–31378.
- [133] M. Karlsson, H. Thorn, A. Danielsson, K.G. Stenkula, A. Ost, J. Gustavsson, F.H. Nystrom, P. Stralfors, Colocalization of insulin receptor and insulin receptor substrate-1 to caveolae in primary human adipocytes – cholesterol depletion blocks insulin signalling for metabolic and mitogenic control, Eur. J. Biochem. 271 (12) (2004) 2471–2479.