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Identification of Novel Anti-inflammatory Agents from Ayurvedic Medicine for Prevention of Chronic Diseases:

“Reverse Pharmacology” and “Bedside to Bench” Approach

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Abstract

Inflammation, although first characterized by Cornelius Celsus, a physician in first Century Rome, it was Rudolf Virchow, a German physician in nineteenth century who suggested a link between inflammation and cancer, cardiovascular diseases, diabetes, pulmonary diseases, neurological diseases and other chronic diseases. Extensive research within last three decades has confirmed these observations and identified the molecular basis for most chronic diseases and for the associated inflammation. The transcription factor, Nuclear Factor-kappaB (NF-κB) that controls over 500 different gene products, has emerged as major mediator of inflammation. Thus agents that can inhibit NF-κB and diminish chronic inflammation have potential to prevent or delay the onset of the chronic diseases and further even treat them. In an attempt to identify novel anti-inflammatory agents which are safe and effective, in contrast to high throughput screen, we have turned to “reverse pharmacology” or “bed to benchside” approach. We found that Ayurveda, a science of long life, almost 6000 years old, can serve as a “goldmine” for novel anti-inflammatory agents used for centuries to treat chronic diseases. The current review is an attempt to provide description of various Ayurvedic plants currently used for treatment, their active chemical components, and the inflammatory pathways that they inhibit.

2. Introduction

Current estimates are that it may cost as much as over a billion dollar to develop a drug by a pharmaceutical company. Today’s Magic bullets or targeted therapies are expensive as cost of treating advanced colorectal cancer patient that was \$500 in 1999 is \$250,000 in 2007 as indicated by Leonard Saltz, from Memorial Sloan-Kettering cancer Center, New York. Despite billions that have been spent, the death rate from most cancers has barely budged. For instance glioblastoma, kills almost everyone who gets it, usually in a little over a year. Radiation and chemotherapy regimen has become the standard of care, which comes with a cost range from \$100,000 to \$500,000. It has been estimated that most population in the world can not afford these smart therapies. Besides cost, safety is a major concern. Similarly it is being asked if someone invented a pill to cut a cancer risk in half, would you take it? Although tamoxifen, raloxifen, celecoxib and finasteride have been approved, they are not very well accepted. The reason for this being is the side effects. For instance, among 1,000 women, 19 would be expected to develop breast cancer over the next five years but if those women all took tamoxifen, however, 9 of those women would avoid breast cancer.

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Tamoxifen is expected to cause 21 additional cases of endometrial cancer, a cancer of the uterine lining that is typically treatable when caught early. An additional 21 would develop blood clots, 31 would develop cataracts and 12 would develop sexual problems. More than half of the 1,000 women would naturally develop hormonal symptoms like hot flashes, changes in vaginal discharge or irregular periods, tamoxifen would cause those symptoms in about an additional 120 women. Raloxifene has been shown to significantly reduce breast cancer risk but with fewer side effects.

To identify a drug that is safe, affordable and effective is a challenge to modern medicine today. Why modern drugs are so unsafe? Why these drugs costs so much? Why are these drugs so ineffective? All these questions require serious thinking “out of the box”. For instance the realization that most chronic diseases are multigenic and thus multi-targeted approach, also called promiscuity in drug development, is needed. As many as 500 gene products or proteins or kinases or signaling intermediates have been linked with any given chronic disease. Thus inhibition of a single kinase or a pathway is unlikely to treat the disease. It has been shown that 74% of all drugs approved by the FDA within last decade for cancer are based on natural products. How to design a drug that is safe, multi-targeted and yet affordable, we have turned to traditional medicine such as Ayurveda which is almost six thousand years old. Ayurveda is a traditional healing system originated in India approximately 6,000 years ago. In Sanskrit, Ayu means “Life” and Veda means “knowledge or science.” Ayurveda can be interpreted as the Science of Life. This “Science of Life” is a holistic healing system, which is designed to promote good health and longevity rather than curing a disease (therapy). Three kinds of primary body constitutions or traits (“prakriti”) have been defined based on three “doshas”, viz, Vata, Pitta, and Kapha. Any imbalance in these dosha results into a disease. To restore the balance, the Ayurveda recommends a customized therapy based on the “prakriti” of an individual. Doshas can be influenced by the food one eats, the type of lifestyle one leads. The term “vata” comes from “vaayu” (in Sanskrit), which means air. The oxidative stress could be caused by insufficient air (vaayu) inhaled, and imbalance in metabolism (the two other tridoshas, the pitta and the kapha). According to ‘Charaka Samhita’, there are five categories of vata. The ‘Prana vata’ is related to inhalation of air, whereas, the ‘Udana vata’ is related to exhalation. ‘Vyana Vata’ regulates the heart and circulatory system; Samana Vata regulates digestive tract, and the ‘Apana Vata’ works in elimination of wastes [1]. Since reactive oxygen species produced ROS) in the body are composed of many species, such as, oxygen ions, peroxides, hydroxyl radicals, etc.; one would require a combination of antioxidants to quench them altogether. Plant polyphenolics though are good source of antioxidants, but they differ in their abilities to quench difference species of ROS [2–4]. Therefore, one may need to use a combination of phytochemicals.

Holistic treatment is the hallmark of treatment in Ayurveda. It demands that one herb or one drug would not cure the imbalance of “Dosha”. Therefore, traditionally, in most of the cases, a combination of herbs and plants (which are even part of staple food) are recommended for treatment [5]. This would probably the most ancient recommendation for a “Combinatorial and Mutliti-targeted Therapy”. It is quite possible that a so called crude herbal formulation has a combination of compounds, where one compound either potentiates the effect other, or increases the bioavailibilty, or reduces the toxicity. A best example is the routine use of turmeric in combination with black pepper as a spice. It is now known that the bioavailability of curcumin (active ingredient of turmeric) is increased by piperine (an active compound in black pepper) by preventing the glucuronidation of the curcumin [6]. Experimentation and documentation of more of such scientific information is highly desirable, and scientific researches to substantiate the use of mixtures of plants in Ayurveda (Table 1) are a worthwhile venture.

We describe here almost 200 different plants that have been used in Ayurveda to treat various chronic diseases (Table 2; Fig. 1). The active component from some of these plants that can modify the inflammatory pathways linked to chronic diseases, are also indicated. Some of these active components have been studied by us and others extensively at the preclinical level. This approach we describe as a “reverse pharmacology” [7] or “bed to benchside” approach to validate the knowledge that has been known for long time.

Inflammatory pathways & chronic diseases

Nuclear factor- κ B (NF- κ B), a nuclear transcription factor, was first identified in 1986 by Sen and Baltimore [8]. As its name implies, it is a nuclear factor bound to an enhancer element of the immunoglobulin kappa light chain gene in B cells. First considered a B-cell transcription factor, NF- κ B is now known to comprise a family of ubiquitous proteins. NF- κ B proteins contain a Rel homology domain (DNA-binding domain/dimerization domain) with a nuclear localization sequence; such sequences are conserved from *Drosophila* to man. Class I proteins include p50, p52, p100, and p105. Multiple copies of ankyrin repeats are present in p100 and p105; proteolytic cleavage of p100 forms p52 and that of p105 forms p50. These protein, in turn, form dimers with class II proteins (c-Rel, RelB, and RelA/p65), which exclusively contain C-terminal activation domains. Whereas RelB forms only heterodimers, all the other proteins can form both homo- and heterodimers. NF- κ B is the most common heterodimer formed between Rel A and p50. Dimeric NF- κ B transcription factors bind to the 10-base-pair consensus site GGGPuNNPyPyCC, where Pu is purine, Py is pyrimidine, and N is any base. The individual dimers have distinct DNA-binding specificities for a collection of related B sites.

The various inhibitors of NF- κ B include I κ B α , I κ B β , I κ B γ (derived from the C-terminal of p100), I κ B ϵ , Bcl-3, pp40 (a chicken homologue), cactus (a *Drosophila* homologue), and avian swine fever virus protein p28, p105 and p100 can also function to retain NF- κ B subunits in the cytoplasm. All of these proteins are characterized by the presence of multiple ankyrin repeats. Perhaps the most common and best-understood form of NF- κ B consists of p50, p65, and I κ B α . I κ B α mediates transient gene expression, whereas I κ B β mediates persistent response.

The I κ B proteins are expressed in a tissue-specific manner and have distinct affinities for individual Rel/NF- κ B complexes. I κ Bs contain six or more ankyrin repeats, an N-terminal regulatory domain, and a C-terminal domain that contains a proline-glutamic acid-serine-threonine motif. I κ Bs bind to NF- κ B dimers and sterically block the function of their nuclear localization sequences, thereby causing their cytoplasmic retention. Most agents that activate NF- κ B mediate the phosphorylation-induced degradation of I κ B. On receipt of a signal, phosphorylation of I κ B α takes place on two conserved serine residues (S32 and S36) in the N-terminal regulatory domain. However, another member of the I κ B family, Bcl-3, stimulates transcription after interacting with p50 and p52 subunits of NF- κ B. Several of the I κ B kinases (IKKs) have been characterized, namely, IKK α , IKK β , and IKK γ . Mutation analysis revealed that IKK α and not IKK β mediates proinflammatory signals. Once phosphorylated, the I κ Bs, which are still bound to NF- κ B, almost immediately undergo a second posttranslational modification known as polyubiquitination. The major ubiquitin acceptor sites in human I κ B α are lysines 21 and 22. Protein ubiquitination occurs through the E1 ubiquitin-activating enzyme, the E2 ubiquitin-conjugating enzyme, and the E3 ubiquitin protein ligases. After ubiquitination, I κ Bs are degraded in 26S proteasomes, leading to the release of NF- κ B dimers, which translocate into the nucleus [9]. In contrast, the activation of NF- κ B in response to ultraviolet (UV) radiation is accompanied by I κ B α degradation but not phosphorylation on the N-terminus of I κ B α . Hypoxia or pervanadate treatment stimulates the phosphorylation of I κ B α at tyrosine 42, but other I κ Bs do not have a tyrosine at this position. Phosphorylation on Ser-276 by the catalytic subunit of protein

kinase A contributes to the intrinsic transcriptional capacity of the p65 subunit of NF- κ B. The catalytic subunit of protein kinase A was also found to be associated with NF- κ B and I κ B in the cytoplasm and was able to phosphorylate p65 only after I κ B degradation [7]. In addition, a site-directed mutant of p65 (Ser-276 to Ala) is phosphorylated at Ser 529 in response to tumor necrosis factor (TNF), suggesting that multiple physiologic stimuli modulate p65 through distinct phosphorylation sites to control transcriptional activity. RelA (C-terminus) has been shown to interact with basal transcriptional apparatus proteins such as TATA-binding protein (TBP), transcription factor (TF) IIB and TBP-associated factor (TAF) 105 and with coactivators such as cAMP responsive element binding protein (CBP) and p300, although the actual role of these interactions is not clear [10]. This pathway is well conserved, both in structure and function, from *Drosophila* to humans.

NF- κ B is activated by many divergent stimuli, including proinflammatory cytokines (e.g., TNF- α , interleukin-1 [IL-1]), T- and B-cell mitogens, bacteria, lipopolysaccharide (LPS), viruses, viral proteins, double-stranded RNA, and physical and chemical stresses. Cellular stresses, including ionizing radiation and chemotherapeutic agents, also activate NF- κ B (Fig. 2).

Although much has been learned since the discovery of NF- κ B, the precise mechanism of its activation is still not fully understood. Depending on the stimulus, this mechanism involves overlapping and nonoverlapping steps. For example, TNF, one of the most potent activators of NF- κ B, interacts with the TNF receptor (TNFR) and then recruits a protein called TNFR-associated death domain. This protein binds to TNFR-associated factor (TRAF) 2, which recruits NF- κ B-inducing kinase (NIK), which in turn activates IKK. IKK phosphorylates I κ B α at serines 32 and 36, which leads to ubiquitination at lysines 21 and 22, and this leads to the degradation of I κ B α by the 26S proteasome. This degradation results in translocation of NF- κ B to the nucleus, where it binds to its consensus sequence (5'-GGGACTTTC-3') and activates gene expression. Thus, NF- κ B can be monitored by the I κ B α degradation seen on Western blotting, by the NF- κ B binding to DNA seen on electrophoretic mobility shift assay, or by the NF- κ B-dependent reporter gene expression seen on transient transfection.

Besides the previously described canonical NF- κ B activation pathway, a noncanonical NF- κ B activation pathway is activated by CD40L, lymphotoxin (LT)- β , receptor activator of NF- κ B ligand (RANKL), and B-cell-activating factor of the TNF family (BAFF), all members of the TNF family. This pathway does not involve I κ B α but instead involves direct phosphorylation and ubiquitin-dependent degradation of p100. Current research indicates that NF- κ B activation is highly complex and may involve dozens of different protein kinases. Besides NIK, IKK- α , and IKK- β , NF- κ B activation may also require the involvement of other kinases, such as atypical protein kinase C, protein kinase C- ζ , pp90rsk, double-stranded RNA-dependent protein kinase, cot kinase (also called TPL2), mitogen-activated protein kinase kinase kinase 1, 2, and 3, phosphatidylinositol 3 protein kinase, Akt, mixed lineage kinase 3, hematopoietic progenitor kinase-1, transforming growth factor β -activated kinase 1, and c-raf kinase. These kinases may form a cascade, and different cascades may form depending on the NF- κ B activator. For instance, IKK can be phosphorylated by NIK, mitogen-activated protein kinase kinase kinase, or Akt. Although IKK is required for NF- κ B activation by most agents, a few (such as human epithelial receptor type 2, H₂O₂, pervanadate, x-rays, and γ -radiation) activate NF- κ B through IKK-independent pathways. Although several signaling proteins and protein kinases have been identified recently that mediate NF- κ B activation, more kinases and protein phosphatases remain to be identified. Besides the ubiquitin-dependent 26S proteasome, which has a role in I κ B α degradation, other proteases have also been implicated in NF- κ B activation.

The genetic deletions of different NF- κ B proteins produce numerous phenotypic changes. For instance, deletion of the *rel a* gene induced embryonic lethality in mice, probably due to massive apoptosis in the liver. In addition, the mouse embryo fibroblasts (MEFs) from *rel a*-deletion mice were found to be hypersensitive to TNF-induced apoptosis. These results indicate a negative role for NF- κ B in TNF-induced apoptosis. Furthermore, mice lacking the RelA subunit were brought to term only in a TNFR1-deficient background. These mice lacked lymph nodes, Peyer's patches, and an organized splenic microarchitecture, and they had a profound defect in their T-cell-dependent antigen responses. Analyses of TNFR1/RelA-deficient embryonic tissues and of radiation chimeras suggest that the dependence on RelA is manifested not in hematopoietic cells but rather in radioresistant stromal cells, which are needed for the development of secondary lymphoid organs. In contrast to the deletion of Rel A, the deletion of the I κ B α gene leads to early neonatal lethality caused by inflammatory dermatitis and granulocytosis that are most likely induced by constitutive activation of NF- κ B, leading to expression of the granulocyte colony-stimulating factor. Once NF- κ B is activated, it causes the expression of almost 500 different gene products that includes enzymes, cytokines, adhesion molecules and other signaling intermediates closely linked with inflammation (Table 3).

NF- κ B and Chronic diseases—NF- κ B activation has been implicated in a wide variety of diseases, including cancers, diabetes mellitus, cardiovascular diseases, autoimmune diseases, viral replication, septic shock, neurodegenerative disorders, ataxia telangiectasia (AT), arthritis, asthma, inflammatory bowel disease, and several other inflammatory conditions (Fig. 3). For example, activation of NF- κ B by LPS may contribute to the development of septic shock because NF- κ B activates transcription of the inducible nitric oxide synthase (iNOS) genes known to be involved in septic shock. Similarly, autoimmune diseases such as systemic lupus erythematosus may also involve activation of NF- κ B. Additionally, in chronic Alzheimer's disease, the amyloid β peptide causes production of reactive oxygen intermediates and indirectly activates gene expression through B sites. The influenza virus protein hemagglutinin also activates NF- κ B, and this activation may contribute to viral induction of cytokines and to some of the symptoms associated with influenza. Furthermore, the oxidized lipids from the low density lipoproteins associated with atherosclerosis activate NF- κ B, which then activates other genes, and mice that are susceptible to atherosclerosis exhibit NF- κ B activation when fed an atherogenic diet. Another important contributor to atherosclerosis is thrombin, which stimulates the proliferation of vascular smooth muscle cells through the activation of NF- κ B. Finally, a truncated form of I κ B α was shown to protect AT cells, which express constitutive levels of an NF- κ B-like activity, from ionizing radiation. In light of all these findings, the abnormal activation or expression of NF- κ B is clearly associated with a wide variety of pathologic conditions.

Ayurvedic plants, their active components and their molecular targets—Almost 200 Ayurvedic plants have been identified that exhibit anti-inflammatory activities. The active component from some of these plants is shown in Fig. 4. The molecular targets of these compounds are shown in Table 4. More specific description of these plants, active components and molecular targets are described below:

1. *Abies pindrow*: *A. pindrow*, known as the 'talisapatra' tree in Sanskrit and 'morinda' in Hindi, is found in abundance in the deciduous forests of Himalayas. Its leaves have been used as Ayurvedic remedy for fever, respiratory and inflammatory ailments. Anti-diabetic, anti-inflammatory, analgesic, hypnotic and anti-ulcerogenic activities in rats, hypotensive effect in dogs, and endurance enhancing in swim stress in mice have been reported for extracts and fractions from *A. pindrow* leaves [11]. Pinitol (3-O-methyl-chiroinositol), a

component of *A. pindrow* was reported to suppress NF- κ B activation both induced by inflammatory stimuli and carcinogens and constitutive NF- κ B activation noted in most tumor cells. The suppression of NF- κ B activation by pinitol occurred through inhibition of the activation of I κ B α kinase, leading to sequential suppression of I κ B α phosphorylation and degradation, p65 phosphorylation and nuclear translocation, and NF- κ B-dependent reporter gene expression. The inhibition of NF- κ B activation thereby led to down-regulation of gene products involved in inflammation (cyclooxygenase [COX]-2), proliferation (cyclin D1 and c-myc), invasion (matrix metalloproteinase [MMP]-9), angiogenesis (vascular endothelial growth factor; VEGF), and cell survival (cIAP1, cIAP2, X-linked inhibitor apoptosis protein [XIAP], Bcl-2, and Bcl-xL). Suppression of these gene products by pinitol enhanced the apoptosis induced by TNF and chemotherapeutic agents and suppressed TNF-induced cellular invasion [12].

2. *Abrus precatorius*: Leaves, roots and seeds of *Abrus precatorius*, known commonly as Jequirity, Crab's Eye, Rosary Pea, or Indian licorice are used for medicinal purposes. A tea is made from the leaves and used to treat fevers, coughs and colds. Abruquinones, the isoflavanquinones isolated from the roots have strong anti-inflammatory and antiallergic effects. Wang et al. [13] suggests that the anti-inflammatory effect of abruquinone is mediated partly by suppressing the release of chemical mediators from mast cells and partly by preventing vascular permeability changes caused by mediators.

3. *Abutilon indicum*: In traditional medicine, *A. indicum* is used as a demulcent, aphrodisiac, laxative, diuretic, pulmonary and sedative. The aqueous extract of the plant has antidiabetic properties, which inhibited glucose absorption and stimulated insulin secretion [14].

4. *Acacia arabica*: The gum of *Acacia Arabica* is the source of useful medicaments and used for treating gingivitis and for reducing plaque. The hypoglycemic effect was indicated that the powdered seeds of *Acacia* by initiating the release of insulin from pancreatic beta cells of normal rabbits [15].

5. *Acacia catechu*: Altavilla et al. [16] studied the anti-inflammatory activity of Flavocoxid, a mixed extract containing baicalin and catechin from *Acacia catechu* that acts as a dual inhibitor of cyclooxygenase (COX) and 5-lipoxygenase (LOX) enzymes and showed that Flavocoxid significantly inhibited COX-2, 5-LOX and inducible nitric oxide (NO) synthase (iNOS) expression in LPS-stimulated peritoneal rat macrophages.

6. *Acacia farnesiana*: The bark and the flowers of *Acacia farnesiana* are the parts most used in traditional medicine. Among all the isolated compounds viz., acasiane A & B, farnesirane A and farnesirane B, three diterpenes, two triterpenes, eight flavonoids, and betulinic acid showed moderate anti-inflammatory activities against five human cancer cell lines [17].

7. *Achillea millefolium*: It has seen historical use as a medicine for treatment of inflammatory diseases. It has been used to treat complaints such as inflammation, pain, wounds, hemorrhages, hepato-biliary disorders and gastrointestinal disturbances such as ulcer, liver cirrosis, chronic hepatitis and diabetes. Anti-tumor activity was studied by Tozyo et al. [18] and showed that achimillic acids A, B and C from *A. millefolium* were found to be active against mouse P-388 leukemia cells *in vivo*.

8. *Achyranthes aspera*: *Achyranthes aspera* is used in the indigenous systems of medicine for the treatment of inflammatory conditions and had hypoglycaemic effect. Its extracts are

also showed anti-inflammatory effects in carrageenin-induced paw oedema in rat [19] and exerted anti-carcinogenic effects *in vivo* two-stage mouse skin carcinogenesis [20].

9. *Acorus calamus*: *Acorus calamus* L., sweet flag, is widely employed in modern herbal medicine as an aromatic stimulant and mild tonic. In Ayurveda, it is highly valued as a rejuvenator for the brain and nervous system and as a remedy for digestive disorders. This plant also exerts antidiabetic, anti-adipogenic, and hypolipidemic activities [21]. *A. calamus* also showed anti-inflammatory effects, and it might be mediated by suppression of NF- κ B and interferon regulatory factor 3 (IRF3) [22]. Also, several reports indicated the neuroprotective effects of *A. calamus* in cortex of rat brain [23].

10. *Adhatoda vasica*: The extracts of *Adhatoda vasica* have been used to treat bronchitis, asthma, ulcer and rheumatism. Gibb [24] showed ambroxol, a natural alkaloid found in *A. vasica*, inhibited IgE-dependent basophil mediator release.

11. *Aegle marmelos*: The compounds, 6-methyl-4-chromanone, isolated from *Aegle marmelos* by Nicolis et al [25] showed inhibition of IL-8 in the IB3-1 CF cells *in vitro*. Cardenolide, periplogenin, isolated from the leaves of *Aegle marmelos* protected the doxorubicin induced cardiotoxicity and lipid peroxidation in rats by reversing the increase in serum creatine kinase-MB, glutamate-pyruvate transaminase, and tissue LPO [26]. Subramaniam et al. [27] reported that marmelin, an ethyl acetate fraction of *Aegle marmelos* extracts suppressed TNF- α -mediated activation and translocation of NF- κ B, inhibited AKT and ERK phosphorylation both *in-vitro* and in tumor xenografts.

12. *Allium Sativum*: The possible therapeutic effects of garlic extract in the treatment of IBD patients showed that it reduced the inflammation by inhibiting cell-mediated T-helper-1 and inflammatory cytokines (TNF- α , IL-1 α , IL-6, IL-8, T-cell IFN- γ and IL-2) while upregulating IL-10 production [28]. Zare A et al [29] showed significant decrease in allergic airway inflammation levels in murine models. The water-soluble allyl sulfur-containing compound, S-Allyl-L-cysteine Sulfoxide (ACSO), have antioxidant and anti-inflammatory activities and Hui et al [30] showed it could inhibit proinflammatory cytokine-induced adhesion of monocytes to endothelial cells by inhibiting the MAPK signaling and related ICAM-1 expression. Ban JO [31] found another sulfur compound, thiacremonone inhibiting the NF- κ B activation via interaction with the sulfhydryl group of NF- κ B molecules, thus could be a used for the treatment of inflammatory and arthritic diseases. Keophiphath M et al [32] used 1,2-vinyldithiin on human preadipocytes to reduce Obesity, a state of chronic low-grade inflammation and found to be a novel, antiobesity nutraceutical.

13. *Aloe vera*: It has been used in the treatment of a variety of disorders including wounds and burns. In addition to its wound healing property *Aloe vera*, has also been shown to have antidiabetic and hypoglycemic properties [33]. Emodin is an active component from *A. vera* exerts anti-inflammatory effects. Emodin suppressed the activation of NF- κ B in human umbelical vein endothelial cells (EC) in a dose- and time-dependent manner. Emodin inhibited degradation of I κ B, an inhibitory subunit of NF- κ B. Thus, emodin also downmodulated adhesion molecules like ICAM-1, VCAM-1, and ELAM-1 contain NF- κ B binding sites in their promoter region in EC [34].

14. *Alpinia galanga*: *Alpinia galanga*, a plant in the ginger family, is an herb used in cooking. Grzanna et al. [35] documented that *Alpinia galanga* extract (GE) can inhibit the activation of human monocytic THP-1 cells by different proinflammatory stimuli and reduce the expression of a wide range of inflammation-related genes such as tumor necrosis factor (TNF)- α , interleukin (IL)-1 β , COX-2, macrophage inflammatory proteins (MIP)- α ,

monocyte chemotactic protein (MCP)-1, and chemokine ligand-10 (IP-10), in microglial-like cells in the central nervous system. The active component from this ginger, 1'-acetoxychavicol acetate (ACA), has been shown to inhibit phorbol ester-induced skin tumor promotion, azoxymethane-induced colonic aberrant crypt foci, estrogen-related endometrial carcinogenesis, hepatic focal lesions, rat oral carcinogenesis, and N-nitrosomethylbenzylamine-induced rat esophageal tumorigenesis. Ichikawa et al. [36] reported that ACA suppressed NF- κ B activation induced by a wide variety of inflammatory and carcinogenic agents, doxorubicin, and cigarette smoke condensate. Suppression was not cell type specific, because both inducible and constitutive NF- κ B activations were blocked by ACA. ACA did not interfere with the binding of NF- κ B to the DNA, but, rather, inhibited I κ B kinase activation, I κ B α phosphorylation, I κ B α degradation, p65 phosphorylation, and subsequent p65 nuclear translocation. ACA also inhibited NF- κ B-dependent reporter gene expression activated by TNF, TNF-receptor (TNFR)-1, TNFR-associated death domain protein (TRADD), TNFR-associated factor-2 (TARF-2), and I κ B kinase, but not that activated by p65. Consequently, ACA suppressed the expression of TNF-induced NF- κ B-regulated proliferative (e.g., cyclin D1 and c-Myc), antiapoptotic (survivin, IAP1, IAP2, XIAP, Bcl-2, Bcl-xL, Bfl-1/A1, and FLIP), and metastatic (COX-2-2, ICAM-1, VEGF and MMP-9) gene products. ACA also enhanced the apoptosis induced by TNF and chemotherapeutic agents and suppressed invasion. Thus, ACA suppressed RANKL signaling had a potential to suppress bone loss. ACA inhibited RANKL signaling and consequent osteoclastogenesis in RAW 264.7 cells, a murine monocytic cell line through suppression of RANKL-induced NF- κ B signaling pathway. ACA also inhibited the osteoclastogenesis induced by human cancer cell lines such as breast cancer, multiple myeloma, and head and neck squamous cell carcinoma [37].

15. *Anacyclus pyrethrum*: The root of *Anacyclus pyrethrum* or Mount Atlas daisy is mostly used in Siddha medicine. The fractions from *A. pyrethrum* showed a marked stimulating effect on the reticulo-endothelial system and increased the number of peritoneal exudate cells, and spleen cells of mice [38].

16. *Andrographis paniculata*: *A. paniculata*, literally 'king of bitters' is used in traditional Siddha and Ayurvedic systems of medicine as well as in tribal medicine in India and some other countries for multiple clinical applications, such as rheumatoid arthritis and inflammatory symptoms of sinusitis. Andrographolide, a diterpenoid lactone, and the major active principle isolated from the plant *A. paniculata*, has been shown to possess a strong anti-inflammatory activity through suppression of inflammatory mediators such as NF- κ B, TNF- α , IL-6, MIP-2, iNOS and COX-2 [39]. The anti-diabetic potential of the plant extract was shown by evoked insulin secretion [40].

17. *Areca catechu*: Betel nut, a partial muscarinic agonist, is one of the mostly widely used substances across Asia has been hypothesized to have beneficial effects on both positive and negative symptoms of schizophrenia. The extracts from this plant has shown to hypotensive properties through its ability to inhibit the pressor responses to both angiotensin I and II [41].

18. *Argyria speciosa*: *Argyria speciosa* is an important 'rasayana' herb in Indian System of medicine that possessed a strong antioxidant, anti-inflammatory and anti-arthritis activity. The ethanolic extract significantly inhibited paw edema induced by carrageenan and Freund's complete adjuvant and prevented accumulation of inflammatory cells in carrageenan-induced peritonitis [19].

19. *Asparagus adscendens*: The plant is a rich source of potential anti-diabetic agents. It has been reported to stimulate insulin secretion, enhance insulin action and to inhibit starch digestion [42].

20. *Asparagus racemosus*: Commonly mentioned as a rasayana in the ayurveda, the plant is considered to be of medicinal importance because of the presence of steroidal saponins and sapogenins in various parts of the plant. It has also been used for nervous disorders, inflammation, liver diseases and certain infectious diseases. The immunomodulating property of the plant has been shown to protect the rat and mice against abdominal sepsis. A recent study showed that potent phytochemicals present in the roots of the plant viz., phytosterols, saponins, polyphenols, flavonoids and ascorbic acid has the ability to regulate cholesterol metabolism and to improve antioxidant status in hypercholesteremic rats [43].

21. *Azadirachta indica*: The plant is known for its medicinal properties since ancient time. A number of phytochemical isolated chiefly from the leaves of the plant has been shown to possess immunomodulatory, anti-inflammatory, antihyperglycaemic, antiulcer, antimalarial, antifungal, antibacterial, antiviral, antioxidant, antimutagenic and anticarcinogenic properties. A recent report indicated that azadirachtin obtained from the plant possess anti-tumor property and has the potential to target NF- κ B [44]. Nimbolide, a limonoid derived from the leaves and flowers of the plant has been shown to exhibit numerous biological activities including anti-cancer [45].

22. *Bacopa monnieri*: In the Indian system of medicine the plant is known as Brahmi. The administration of extract from the plant has been reported to significantly improve short-term and long-term memory. Bacoside-A has also been reported to prevent the occurrence of seizures and to reduce impaired peripheral nervous system in epileptic rats [46]. The methanolic extract as well as Bacoside-A isolated from the plant has been reported to possess wound-healing activity in Swiss albino rats [47].

23. *Bambusa arundinacea*: The leaves of the plant have been shown useful in inflammatory conditions, have the ability to heal the wound and have also been shown to check diarrhea in cattle. Manna, a crystalline substance obtained from the plant has been shown useful in ayurvedic medicine for ptosis and paralytic complaints. The methanol extract from the plant has been shown to possess antiinflammatory effect on carrageenin-induced oedema in rats [48].

24. *Bauhinia variegata*: The powdered bark from the plant has been traditionally used in ayurvedic medicines as a tonic to the liver. The ethanolic extract and the roseoside (major constituent) from the plant have been reported to enhance insulin release in insulin secreting cell line [49]. The extract from the plant has been reported to exert anticarcinogenic and antimutagenic activity in swiss albino mice. The ethanol extract from the plant has also shown potential to possess chemopreventive property against N-nitrosodiethylamine induced liver tumor and human cancer cell lines [50].

25. *Berberis aristata*: *Berberis aristata* DC (Berberidaceae) known, as 'daruharidra' is an evergreen, spinescent shrub with known antichlamydial, antiplatelet, antimicrobial and hepatoprotective activity. Its root mainly contains berberine chloride, palmatine chloride. Root bark extract of the plant is taken twice a day for 1–2 weeks by the tribal people in Sikkim (a north-east state of India) and Darjeeling Himalayan region to treat diabetes. Both the herbs are well known for their anti-inflammatory activity. Berberine has also been found to be effective in experimental herpetic uveitis [51]. Berberine was also shown to abolish NF- κ B activation induced by various inflammatory agents and carcinogens. This alkaloid also suppressed constitutive NF- κ B activation found in certain tumor cells. Suppression of

NF- κ B activation occurred through the inhibition of phosphorylation and degradation of I κ B α by the inhibition of I κ B kinase (IKK) activation, leading to suppression of phosphorylation and nuclear translocation of p65, and finally to inhibition of NF- κ B reporter activity. Inhibition of IKK by berberine was direct and could be reversed by reducing agents. Site-specific mutagenesis suggested the involvement of cysteine residue 179 in IKK. Berberine also suppressed the expression of NF- κ B-regulated gene products involved in antiapoptosis (Bcl-xL, Survivin, IAP1, IAP2, and cFLIP), proliferation (cyclin D1), inflammation (COX-2), and invasion (MMP-9). Suppression of antiapoptotic gene products correlated with enhancement of apoptosis induced by TNF and chemotherapeutic agents and with inhibition of TNF-induced cellular invasion [52]. Thus this indicates that this medicinal plant exhibits activities against inflammation linked to most chronic diseases

26. *Bergenia ligulata*: *Bergenia ligulata* are popularly known in India as Pashanbhedha. *Bergenia ligulata* (family, Saxifragaceae) has been used for centuries in South Asia, mainly India and Pakistan, for a wide range of ailments. The roots of *B. ligulata* have been used for the therapy of urinary stones, chronic ulcers, viral hepatitis, and benign prostatic hypertrophy. In addition, *B. ligulata* has anti-inflammatory and cytoprotective properties. However, the most important activities are its diuretic and lithotriptic effects.

27. *Boerhaavia diffusa*: *Boerhaavia diffusa* L. is commonly known as 'Punarnava' and its various parts are used in the treatment of cancer, jaundice, dyspepsia, inflammation, enlargement of spleen, abdominal pain and as an anti-stress agent [53, 54]. Administration of aqueous methanol extract of *Boerhaavia diffusa* was found to be effective in reducing the metastases formation by B167-10 melanoma cells and Punarnavine, an alkaloid from *Boerhaavia diffusa* enhanced the immune response against metastatic progression of B16F-10 melanoma cells in mice ([55]

28. *Boswellia serrata*: Extracts from Indian Ayurvedic medicinal plant *Boswellia serrata* (BE) contains beta boswellic acid, a pentacyclic triterpene and the active component of the gum resin (also called frankincense in European pharmacopeia) secreted by the bark of the tree. BE has been used for centuries in traditional Ayurvedic medicine for a wide variety of inflammatory diseases including inflammatory bowel disease [56] and rheumatoid arthritis [57]. BE has been shown to inhibit leukotriene biosynthesis from endogenous arachidonic acid in intact peripheral mononuclear neutrophils through the inhibition of 5-lipoxygenase (LOX), with IC₅₀ as low as 1.5 μ M [58]. Other pentacyclic triterpenes (amyrin and ursolic acid) lack this activity. Further studies revealed that the pentacyclic triterpene ring structure, hydrophilic group on C4 ring A, and 11-keto function are all essential for 5-LOX inhibitory activity [59]. By photoaffinity labeling, it was shown that BE binds to 5-LOX at a site distinct from substrate binding site [60]. BE has also been shown to inhibit leukocyte elastase with an IC₅₀ of 15 μ M [61], topoisomerase (topo) I and II α 3 with affinity constant (KD) of 70.6 nM and 7.6 nM, respectively [62]. BE has been shown to inhibit the growth of a wide variety of tumor cells including glioma [63], colon cancer [64, 65], leukemia cells [66–70], human melanoma [71], hepatoma [72] and prostate cancer cells [73]. The apoptotic effects of BE are mediated through various mechanisms including inhibition of topoisomerase I and II without inhibiting DNA fragmentation [63, 69] and downregulation of cyclin D1, bcl-2, and bcl-xl. Apoptotic effects of BE in hepatoma and colon cancer cells were found to be mediated through caspase-8 activation [64, 72]. Recently BE was reported to induce death receptor (DR)-5 but not DR-4 or Fas through increased expression levels of CAAT/enhancer binding protein homologous protein (CHOP), which led to the activation of caspase-8 in prostate cancer cells [74]. BE also downregulated the expression of androgen receptor through modulation of Sp1 binding activity in prostate cancer cells [75].

The secretion and activity of matrix metalloproteinases (MMPs) from human fibrosarcoma HT-1080 cells was also found to be suppressed by BE [71]. The anti-inflammatory effects of this agent are further demonstrated by studies that showed that LPS-induced TNF production is blocked by BE [76]. Anti-proliferative and anti-inflammatory effects of BE are also mediated through the suppression of the NF- κ B pathway [77] and STAT3 pathway [78]. Microarray analysis revealed that BE modulated 113 of 552 genes induced by TNF- α in human endothelial cells including MMP-3, MMP-10 and MMP-12 [79], and protected animals against experimental arthritis [80].

Numerous animal studies have been performed with BE. In guinea pigs, BE modulated the biosynthesis of leukotrienes and the course of experimental autoimmune encephalomyelitis (EAE) [81]. Topical application of a methanolic extract of *Boswellia serrata* (BE) to the backs of mice markedly inhibited TPA-induced increase in skin inflammation, epidermal proliferation, the number of epidermal cell layers, and tumor promotion in 7,12-dimethylbenz[a]anthracene (DMBA)-initiated mice [66]. BE potentially attenuated experimental ileitis (inflammation of the ileum) in rats [82], an experimental model of inflammatory bowel disease (IBD). In another study, Anthoni et al [83] examined the mechanisms by which BE mediated its effects in experimental colitis. They showed that BE conferred protection in experimental murine colitis induced by dextran sodium sulfate (DSS). Clinical measurements of disease activity and histology were used to assess disease progression, and intravital microscopy was employed to monitor the adhesion of leukocytes and platelets in postcapillary venules of the inflamed colon. BE treatment significantly blunted disease activity as assessed both grossly and by histology. By using in vivo Matrigel™ plug assay, it was shown that BE inhibited bFGF-induced angiogenesis [84]. Also, Wistar rats treated with BE 14 days after inoculation of C6 tumor cells into their right caudate nuclei survived more than twice as long as untreated mice. Furthermore, when treatment was started immediately after implantation and stopped after 14 days, a higher dose of BE produced significantly smaller tumors with greater apoptotic fractions than untreated mice, suggesting that it might have both therapeutic and chemopreventive effects [85]. Toxicity studies with BE in rats and primates showed no pathological changes in hematological, biochemical, or histological parameters at doses up to 1000 mg/kg. The LD50 has been established at >2 g/kg [86].

29. *Bryonia laciniosa*: *Bryonia laciniosa* leaves extract have been used in traditional folk medicine to treat numerous diseases. The methanol extract of *B. laciniosa* exhibited analgesic and antipyretic activity in the tested experimental animal models. The extract showed inhibition on the hind paw oedema in rats caused by histamine and serotonin respectively [87].

30. *Butea monosperma*: *Butea monosperma* (Lam.) (family: Fabaceae) also known as flame of the forest or Palasa in Sanskrit, and in the traditional system of medicine known as 'Ayurveda', *Butea monosperma* has been used in the treatment of a variety of ailments including liver disorders. Nearly every part of *Butea monosperma* has been used as tonic, astringent, aphrodisiac and diuretic. The main constituent of its flower is butrin (7,3',4'-trihydroxyflavanone-7,3'-diglucoside) and isobutrin (3,4,2',4'-tetrahydroxychalcone-3,4'-diglucoside) have been shown to be hepatoprotective [88].

31. *Caesalpinia bonducella*: *C. bonducella* FLEMING (Caesalpinaceae) plant is well known for its medicinal and therapeutic values in Indian Ayurveda. It possesses the anti-inflammatory, analgesic activities against ascites carcinoma and prevention of autoimmune diseases. It has also potent antipyretic and antinociceptive activities [89].

32. *Caesalpinia digyna*: Several members of the species of genus *Caesalpinia* are used traditionally for a wide variety of ethnomedical properties such as anti-inflammatory, antidiabetic, antioxidant and hepatoprotective [89]. The plant is one of the ingredients of an indigenous drug preparation “Geriforte”, which has been used for curing senile prurites with excellent results. The methanol extract of *Caesalpinia digyna* root exhibited strong scavenging effect on free radical and inhibition of lipid peroxidation [89].

33. *Callicarpa macrophylla*: *C. macrophylla* is used for treatment of rheumatic joints. Betulinic acid (BA), a pure compound from *C. macrophylla*, has been reported to be a selective inducer of apoptosis in tumor cells. It also exhibits anti-inflammatory and immunomodulatory properties. BA has been reported to suppress the activation of NF- κ B activation through suppression of I κ B kinase, thus abrogate the phosphorylation and degradation of I κ B α . Treatment of cells with this triterpinoid also suppressed NF- κ B-dependent reporter gene expression and the production of NF- κ B-regulated gene products such as COX-2 and MMP-9 induced by inflammatory stimuli. Furthermore, BA enhanced TNF-induced apoptosis [90]. It also inhibits constitutive activation of STAT3 and STAT3-regulated gene products such as Bcl-xL, Bcl-2, cyclin D1 and survivin [91].

34. *Calotropis procera*: The latex of the plant *Calotropis procera* has been reported to exhibit potent anti-inflammatory activity against carrageenin and formalin that are known to release various mediators. Its anti-inflammatory effect is caused by inhibiting PGE2 [92].

35. *Capparis spinosa*: *Capparis spinosa* has been employed as a flavoring in cooking and as a diuretic, hypertensive, and tonic (e.g., as a poultice) since ancient times. The ethanol extract from fruits of *C. spinosa* (ECS) significantly reduced the production of O₂⁻, H₂O₂, and ROS. ECS exhibits a notable activity in protecting against oxidative stress and interrupting of ROS-ERK1/2-Ha-Ras signal loop, suggesting its potential protective effects against skin sclerosis [93].

36. *Carum copticum*: *Carum copticum*. Linn. (Family: Umbelliferae) is popularly known as Ajowan. As a traditional medicine, the seeds of this plant are made into a decoction and used for curing diarrhoeas, amoebiasis, febrile conditions and stomach disorders. It is much valued for its antispasmodic, antiseptic properties and effects against curing dyspepsia and disorders of inflammation [94]. In the Unani system, ajowan is used as an enhancer of body's resistance. And Antiinflammatory effects of the total alcoholic extract (TAE) and total aqueous extract (TAQ) in 100 mg/kg doses from the seeds of *Carum copticum*. Linn were significantly had in acute rat model (carrageenan induced rat paw oedema) and a sub acute rat model (cotton pellet induced granuloma).

37. *Casearia esculenta*: *Casearia esculenta* Roxb. (Flacourtiaceae) has been a popular remedy for the treatment of diabetes mellitus and is one of the major ingredients of D-400, the largest selling antidiabetic drug in India (Himalaya Drug Company, Bangalore). The root extract from *C. esculenta* has been reported to reduce blood sugar level in animal model, and show antihyperglycemic property in a STZ-induced diabetic rats [95].

38. *Cassia angustifolia*: *Cassia angustifolia* are widely used against skin disorders in traditional Chinese medicine. And also has anti-inflammatory activity [96].

39. *Cassia fistula*: *Cassia fistula* linn (Caesalpinaceae) tree is one of the most widespread in the forests of India. The whole plant possesses medicinal properties useful in the treatment of skin diseases, inflammatory diseases, rheumatism, anorexia and jaundice. The hepatoprotective activity and the hypoglycaemic activity have been reported [97]. And anti-

inflammatory and antioxidant activities of the aqueous and methanolic extracts of the *Cassia fistula* Linn. bark were confirmed in Wistar albino rats (both acute and chronic models).

40. *Cassia occidentalis*: *Cassia occidentalis* L. is used to cure various diseases. This weed has been known to exert antimicrobial (antibacterial, antifungal, laxative, analgesic, chloretic and diuretic properties), hepatoprotective, anti-inflammatory, antimutagenic and anticarcinogenic activity. Anti-inflammatory effects of these extracts to lower the lipid peroxide content, γ -glutamyl transpeptidase and phospholipase A2 activity in the exudates of cotton pellet granuloma, resulting in the reduced availability of arachidonic acid, a precursor of prostaglandin biosynthesis, and/or by stabilization of the lysosomal membrane system. And target for anticarcinogenic activity is Lck (p56lck) protein tyrosine kinase [98].

41. *Cassia tora*: *Cassia tora* L. has been also prescribed in oriental herb medicine to treat night blindness, hypertension, hypercholesterolemia, constipation, hypoglycemic, hypolipidemic, antimutagenic, anticlastogenicity, and antihepatotoxic activities. Several polyherbal formulations including *C. tora* seeds are available at Chinese markets for preventing the formation of atherosclerosis plaques. Recently, it was reported that emodin and obtusifolin in *Cassia tora* L. might be the components having antidiabetic functions since they exhibited a significant inhibitory activity on advanced glycation end products formation [99].

42. *Cedrus deodara*: The wood of *C. deodara* has been used since ancient days in Ayurvedic medical practice for the treatment of inflammations and rheumatoid arthritis, anti-cancer activity, potent disinfectant, anti-fungal properties, and analgesic activity [100].

43. *Celastrus paniculatus*: The oil obtained from the seeds of *Celastrus paniculatus* Willd. (Celastraceae) is largely used in Ayurvedic medicine for sedative action, as an anti-rheumatic agent, alleviation of intestinal spasms, analgesic and anti-inflammatory activities and anti-diarrhoea [101].

44. *Cichorium intybus*: Chicory roots have been used as a digestive aid, diuretic, laxative, and mild sedative. Additionally, hepatoprotective agents have been described in the seeds. Its aqueous, ethanolic, and methanolic extracts have been shown to affect cholesterol uptake and tumor development in mice [102], prevent immunotoxicity induced by ethanol, and have anti-inflammatory properties *in vitro* and *in vivo* [103].

45. *Cinnamomum camphora*: *Cinnamomum camphora* Sieb (Lauraceae) has long been prescribed in traditional medicine for the treatment of inflammation-related diseases such as rheumatism, sprains, bronchitis and muscle pains. *C. camphora* has anti-inflammatory mechanisms blocked the production of IL-1, IL-6 and the TNF- α from RAW264.7 cells and NO, PGE2 production in lipopolysaccharide (LPS)/interferon (IFN)- γ -activated macrophages [104].

46. *Cinnamomum cassia*: *Cinnamomum cassia* is used to cure various diseases. This weed has been known to has antimicrobial, laxative, analgesic, chloretic, diuretic and and antidiabetic activity [105].

47. *Cinnamomum zeylanicum*: *C. zeylanicum* is described as having stimulant, antifatulent, antiemetic and antidiarrhoeal properties. The principle constituent of cinnamomum bark is the volatile oil which contains cinnamic aldehyde, eugenol and terpenes. The cinnamon oil from *C. zeylanicum* ameliorated early stage diabetic nephropathy in alloxan-induced diabetic nephropathy [106].

48. *Citrullus colocynthis*: This cucurbitaceae is widely used in Tunisian folk medicine and it possesses therapeutic activities against a wide range of ailments including inflammatory disorders, arthritis and gout [107].

49. *Commiphora wightii*: Guggulsterone [4,17(20)-pregnadiene-3,16-dione] is a plant sterol derived from the gum resin (guggulu) of the tree *Commiphora mukul*. The resin of the *C mukul* tree has been used in Ayurvedic medicine for centuries to treat such ailments as obesity, bone fractures, arthritis, inflammation, cardiovascular disease, and lipid disorders [108, 109]. This steroid has been shown to bind to the farnesoid \times receptor [110] and modulate expression of proteins with antiapoptotic, cell survival, cell proliferation angiogenic, and metastatic activities in tumor cells. Guggulsterone mediates gene expression through regulation of various transcription factors, including NF- κ B [111], STAT-3 [112] and various steroid receptors such as androgen receptor and glucocorticoid receptors [113].

Gujral et al demonstrated the anti-arthritic and anti-inflammatory activity of gum guggul [114]. Sharma et al showed its activity in experimental arthritis induced by mycobacterial adjuvant [115]. The effectiveness of guggul for treating osteoarthritis of the knee has also been demonstrated [116]. Recent studies have shown that guggulsterone is an antagonist for bile acid receptor farnesoid \times receptor (FXR) [110, 117]. Other studies have shown that guggulsterone enhances transcription of the bile salt export pump [118] Thus guggulsterone is an important regulator of cholesterol homeostasis. Meselhy et al showed that guggulsterone can suppress inducible nitric oxide synthetase (iNOS) expression induced by LPS in macrophages [119]. Because NF- κ B has been implicated in obesity, inflammation, hyperlipidemia, atherosclerosis, and osteoarthritis and in the LPS-induced expression of iNOS, we speculated that guggulsterone mediates its effects, at least in part, through suppression of NF- κ B activation. We have shown that guggulsterone will downregulate NF- κ B activation and potentiate apoptosis induced by TNF, taxol and doxorubicin in human myeloid tumor cells [111]. Our laboratory recently showed that guggulsterone suppressed DNA binding of NF- κ B induced by TNF, phorbol ester, okadaic acid, cigarette smoke condensate, hydrogen peroxide, and interleukin 1 (14). Guggulsterone also suppressed constitutive NF- κ B activation expressed in many tumor cells. Through inhibition of I κ B α kinase activation, this steroid blocked I κ B α phosphorylation and degradation, thus suppressing p65 phosphorylation and nuclear translocation [111]. Guggulsterone downregulates expression of cyclooxygenase (COX)-2, matrix metalloprotease (MMP)-9, cyclin D1 expression, VEGF and of antiapoptotic gene products (IAP1, XIAP, Bfl-1/A1, Bcl-2, cFLIP, and survivin through the downregulation of NF- κ B activation (14). Others have shown that guggulsterone alone will induce apoptosis in acute myeloid leukemia [120] and in prostate cancer cells [121] through the activation of caspases.

Guggulsterone inhibits osteoclastogenesis induced by NF- κ B ligand (RANKL), and by breast tumor cells. Because guggulsterone can suppress the NF- κ B activation induced by various carcinogens, our laboratory investigated whether guggulsterone could modulate RANKL signaling and osteoclastogenesis induced by RANKL or tumor cells (see appendix; [122]). We found that treatment of monocytes with guggulsterone suppressed RANKL-activated NF- κ B activation (as indicated by gel-shift assay) and that this suppression correlated with inhibition of I κ B α kinase and phosphorylation and degradation of I κ B α , an inhibitor of NF- κ B. Guggulsterone also suppressed the differentiation of monocytes to osteoclasts in a dose- and time-dependent manner. Finally, differentiation to osteoclasts induced by coincubating human breast tumor cells (MDA-MB-468) or human multiple myeloma (U266) cells with monocytes was also completely suppressed by guggulsterone. Collectively, our results indicate that guggulsterone suppresses RANKL and tumor cell-induced osteoclastogenesis by suppressing the activation of NF- κ B.

50. *Convolvulus pluricaulis*: *Convolvulus pluricaulis* (CP) is known as Shankpushpi (or shankapushpi), an herb that has been used in India for hundreds of years for nervous disorders such as stress, anxiety and insomnia. CP has an antiulcerogenic effect due to augmentation of mucosal defensive factors such as mucin secretion, lifespan of mucosal cells and glycoproteins rather than on the offensive factors such as acid-pepsin [123].

51. *Crataeva nurvula*: *Crataeva nurvula* has an antioxidant potential. SOD mimetic activity was found to be in *Crataeva nurvula*. Lipid peroxidation inhibitory potential was found to be in *Crataeva nurvula* and also showed a comparatively high NO quenching capacity [124].

52. *Crocus sativus*: *Crocus sativus* L. (saffron) is used in folk medicine, for example as an antiedematogenic agent. Aqueous and ethanolic extracts of saffron stigma and petal have an antinociceptive effect, as well as acute and/or chronic anti-inflammatory activity [125].

53. *Cuminum cyminum*: Cumin seeds (*Cuminum cyminum* L.) are largely used as a condiment or spice in Indian food. They are also medicinally useful to correct hoarseness of voice, gonorrhea, dyspepsia, and chronic diarrhea. Cumin supplementation significantly reduced the incidence and number of tumors in the colon. Cumin prevented the accumulation of lipids in tissues and optimized the excretion of fecal sterols and bile acids [126].

54. *Curcuma amada*: *C. amada* belonging to the family of Zingiberaceae, popularly known as mango ginger, has been known for its potent antioxidant activity. Mango ginger (*C. amada*) contains significant amounts of phenolics as both free and bound forms. Both free and bound phenolic fractions of mango ginger were found to be antioxidant and effective in inhibiting H⁺,K⁺-ATPase activity and *H. pylori* growth [127].

55. *Curcuma longa*: *Curcuma longa* (turmeric) has a long history of use in Ayurvedic medicine as a treatment for inflammatory conditions. Turmeric constituents include the three curcuminoids: curcumin (diferuloylmethane; the primary constituent and the one responsible for its vibrant yellow color), demethoxycurcumin, and bisdemethoxycurcumin. Curcumin (diferuloylmethane), an anti-inflammatory agent used in traditional medicine, has been shown to suppress cellular transformation, proliferation, invasion, angiogenesis, and metastasis. Curcumin suppressed TNF-induced NF-κB activation and NF-κB-dependent reporter gene expression. Such TNF-induced NF-κB-regulated gene products involved in cellular proliferation (COX-2, cyclin D1, and c-myc), antiapoptosis (IAP1, IAP2, XIAP, Bcl-2, Bcl-xL, Bfl-1/A1, TRAF1, and cellular cFLIP), and metastasis (VEGF, MMP-9, ICAM-1) were also downregulated by curcumin. COX-2 promoter activity induced by TNF was abrogated by curcumin [128, 129]. Bharti et al [130] reported that curcumin inhibited IL-6-induced STAT3 phosphorylation and consequent STAT3 nuclear translocation. Various activities of curcumin against different chronic diseases has been extensively reviewed by us and others [131–144].

56. *Curcuma zedoaria*: *Curcuma zedoaria* Rosc is a perennial herb found in tropical countries, such as India, Japan and Thailand. Various parts of this plant are used in Ayurveda and other folk medicines for the treatment of different ailments such as diarrhea, cancer. *C. zedoaria* has been used for phytochemical and pharmacological medicine [145].

57. *Cymbopogon citratus*: *Cymbopogon citratus*, commonly called as lemongrass, is a natural herb that contains citral and is a widely used herb as a food flavoring, as a perfume, and for its analgesic and anti-inflammatory purposes. Lemon grass intake ameliorated ileitis through decreasing lymphocyte migration by inhibiting beta7-expression in SAMP1/Yit

mice [146] The extract also showed reduction in the release of pro-inflammatory mediators TNF-alpha and NO significantly indicating an anti-inflammatory effect [147].

58. *Cymbopogon martini*: Antifungal efficacy of essential oils (EO) of *Cymbopogon martini* has been well documented. EOs displayed strong antifungal effects. EOs has been used for treatment of dermatophyte infections and may be recommended as an alternative to synthetic drug for topical application [148].

59. *Cyperus rotundus*: *Cyperus rotundus* (Family Cyperaceae) is used both as a functional food and as a drug. The extract exhibited high reduction capability and powerful free radical scavenging, especially against 1,1-diphenyl-2-picrylhydrazyl (DPPH) and superoxide anions as well as a moderate effect on NO [149].

60. *Cyperus scariosus*: *Cyperus scariosus*, Br. (Syn: *C. pertenuis*, Roxb.; family: Cyperaceae) is a delicate grass, growing luxuriously in damp. The brown coloured plant rhizomes have a folkloric reputation as cordial, tonic, emmenagogue, vermifuge, diuretic, diaphoretic and desiccant. It remains an important component of several prescriptions used in the native system of medicine to treat a variety of diseases including diarrhoea, epilepsy, gonorrhoea, syphilis and liver damage. The hepatoprotective activity of aqueous-methanolic extract of *C. scariosus* was investigated against acetaminophen and CCl₄-induced hepatic damage [150].

61. *Daemonorops draco*: Dragon's blood is a non-specific name for red resinous exudations from quite different plant species endemic to various regions around the globe that belong to the genera *Dracaena* (Africa) and *Daemonorops* (South-East Asia), more rarely also to the genera *Pterocarpus* and *Croton* (both South America). Dragon's blood is used for medicinal purposes where it is endemic [151].

62. *Datura metel*: *Datura metel* L. of Solanaceae family is a sub-glabrous shrubby herb found to exist throughout the world. It is frequently used in traditional systems of medicines as narcotic, anodyne and antispasmodic. The leaves of *D. metel* are reported to have anticholinergic activity and are used to relieve the spasm of bronchioles in asthma [152].

63. *Didymocarpus pedicellata*: *Didymocarpus pedicellata* R. Br. (Gesneriaceae) is widely used in traditional Indian medicines against renal afflictions. *D. pedicellata* extract was found to possess a high content of total polyphenolics, exhibit potent reducing power and significantly scavenge free radicals including several reactive oxygen species (ROS) and reactive nitrogen species (RNS). The extract also significantly and dose-dependently protected against Fe-NTA plus H₂O₂-mediated damage to lipids and DNA [153].

64. *Dolichos biflorus*: *Dolichos biflorus* has traditionally been used to dissolve existing renal calculi, provide symptom relief, and prevent recurrence. *D. biflorus* has been demonstrated that it inhibits calcium phosphate crystallization. *D. biflorus* causes the urinary magnesium increase that considered an inhibitor of stone formation [154].

65. *Dysoxylum binectariferum*: The fruit of this plant has anti-inflammatory, diuretic, and CNS depressant activities. The stem bark contains an alkaloid, rohitukine, which exhibited anti-inflammatory and immunomodulatory property. Flavopiridol, synthetic flavone derived from rohitukine, is known as potent inhibitor of several cyclin-dependent kinases (CDK) and undergoes Phase III clinical trial, currently. Flavopiridol has been reported that it suppressed NF-κB in a dose- and time-dependent manner in several cell types. This effect was mediated through inhibition of NF-κB signaling pathway. Flavopiridol also inhibited the expression of the TNF-induced NF-κB-regulated gene products cyclin D1, COX-2, and MMP-9 [155].

Therefore, flavopiridol suppressed TNF-induced activation of activator protein-1 (AP-1) through suppression of various mitogen-activated protein kinases, including c-Jun NH(2)-terminal kinase (JNK), p38 mitogen-activated protein kinase (MAPK), and p44/p42 MAPK. It is noteworthy that this flavone also suppressed the expression of various antiapoptotic proteins, such as IAP-1, IAP-2, XIAP, Bcl-2, Bcl-xL, and TRAF-1. Flavopiridol also inhibited the TNF-induced induction of intercellular adhesion molecule-1, c-Myc, and c-Fos, all known to mediate tumorigenesis [156].

66. *Eclipta alba*: The hepatoprotective effect of the ethanol/water (1:1) extract of *Eclipta alba* (Ea) has been studied at subcellular levels in rats against CCl₄-induced hepatotoxicity. Ea significantly counteracted CCl₄-induced inhibition of the hepatic microsomal drug metabolising enzyme amidopyrine N-demethylase and membrane bound glucose 6-phosphatase. The loss of hepatic lysosomal acid phosphatase and alkaline phosphatase by CCl₄ was significantly restored by Ea. The study shows that hepatoprotective activity of Ea is by regulating the levels of hepatic microsomal drug metabolising enzymes [157].

67. *Elettaria cardamomum*: *Elettaria cardamomum* (Cardamom) was shown to play a wide range of health-promoting roles against various conditions such as constipation, colic, diarrhea, dyspepsia, vomiting, headache, epilepsy, and cardiovascular diseases. Cardamom was reported to exhibit spasmogenic, spasmolytic, blood pressure-lowering, vasodilator, diuretic, and sedative activities [158]. Recently, experimental evidence suggests that cardamom extracts display anti-cancer activities [159]. It has been reported that aqueous suspensions of cardamom have protective effects on experimentally induced colon carcinogenesis by virtue of their anti-inflammatory, anti-proliferative and pro-apoptotic activity.

68. *Embelia ribes*: The fruit of the *Embelia ribes* Burm. plant (Myrsinaceae) (called false black pepper in English, Vidanda in Sanskrit, and Babrang in Hindi languages) has been used to treat fever, inflammatory diseases, and a variety of gastrointestinal ailments for thousands of years. Embelin from *E. ribes* has been shown to have antitumor, anti-inflammatory, and analgesic properties. More recently, active principle, embelin was also identified as a cell-permeable, small molecular weight inhibitor of the X chromosome-linked inhibitor-of-apoptosis protein (XIAP), an antiapoptotic protein, through structure-based computational screening of a traditional herbal medicine three-dimensional structure database of 8221 individual traditional herbal products [160]. Embelin also inhibited activity through modulation of NF- κ B activation. Embelin inhibited both inducible and constitutive NF- κ B activation was abrogated by embelin. Thus, embelin inhibited sequentially the TNF-induced activation of the I κ B kinase, I κ Ba phosphorylation, I κ Ba degradation, and p65 phosphorylation and nuclear translocation. Furthermore, embelin down-regulated gene products involved in cell survival, proliferation, invasion, and metastasis of the tumor. This down-regulation was associated with enhanced apoptosis by cytokine and chemotherapeutic agents [161].

69. *Emblica officinalis*: *Emblica officinalis* (Family: Euphorbiaceae) indigenous to India, is valued for its unique tannins and flavanoids, which contain very powerful antioxidant properties and used for the treatment of a number of diseases, such as dyslipidemia and atherosclerosis, as hepatoprotective, radioprotective, antibacterial, antitumor, and anti-inflammatory agents [162].

70. *Eugenia jambolana*: *Eugenia jambolana* Lam. (Myrtaceae), popularly known as *Jamun*, is being widely used to treat liver dysfunctions and diabetes by the traditional practitioners for over many centuries. This plant has been reported to have both antidiabetic as well as ulcer protective effects. The extract of jamun pulp showed hypoglycemic activity

by stimulating insulin secretion [163]. The chemical constituents of the seed of *E. jambolana* Lam. are gallic acid, ellagic acid, corilagin, ellagitannins, isoquercetin, quercetin, caffeic acid, ferulic acid, guaiacol, resorcinaldimethyl ether, lignaglucoiside, veratrole, β -sitosterol, palmitic acid etc.

71. *Evolvulus alsinoides*: *Evolvulus alsinoides* (shankhpushpi which is considered as Medhya Rasayana) is an Ayurvedic drug used for its action on the central nervous system, especially for boosting memory and improving intellect. In the Ayurvedic system of medicine, the whole herb of 'Shankhpushpi' has been employed clinically for centuries for its memory potentiating, anxiolytic and tranquilizing properties. The crude extracts of *E. alsinoides* showed a marked reduction in inflammation and edema in adjuvant induced arthritic rat model [164].

72. *Fagonia cretica*: *Fagonia cretica* linn is tropical herbs and have been extensively used in the treatment of various types of haematological, hepatic, neurological and inflammatory conditions. The antioxidant and antibacterial properties of *Fagonia cretica* have also been well documented. *F. cretica* also overcome the oxidative stress mediated injury during ischemic neuronal injury via modulating the antioxidant pool of the cells [165].

73. *Ferula assafoetida*: *Ferula assafoetida* is used as a food spice in many Asian countries and for the treatment of asthma, bronchitis, ulcer, kidney stone, pain, and cancer in traditional herbal medicine. *F. assafoetida* L. was reported to have antitumor, antimutagenic and antiviral activities. Farnesiferol C is one of the sesquiterpene coumarin compounds isolated from the resin of *Ferula assafoetida* L., which have antitumor and antiangiogenic activity [166]. Farnesiferol C inhibits proliferation and angiogenesis by decreasing expression of CD31 and VEGF. It decreased the binding of VEGF to VEGFR1/Flt-1, but not to VEGFR2/KDR/Flk-1. It also decreased the phosphorylation of most of the kinases downstream of VEGFR2: focal adhesion kinase, Src, extracellular signal-regulated kinase 1/2, p38 mitogen-activated protein kinase, and c-jun-NH2-kinase without affecting AKT.

74. *Ficus bengalensis*: *Ficus bengalensis* Linn. Family: (Moraceae) is a very large tree distributed throughout India. It is commonly known as 'Bargad' in Hindi or 'Indian Banyan tree' and considered as holy tree of India. Information based on ethnomedicinal survey reveals that the herbal preparations of different parts of *Ficus bengalensis* had been considered as effective economical and safe treatments for curing various diseases, such as diarrhoea, respiratory disorders, certain skin diseases and diabetes. Extracts of *Ficus bengalensis* bark was also found to reduce allergy and stress in asthmatic condition in milk-induced leucocytosis and milk-induced eosinophilia [167].

75. *Foeniculum vulgare*: *Foeniculum vulgare* Mill (Fennel) belonging to the Family Apiaceae (Umbelliferae) is a perennial herb native to the Mediterranean region. Dried fruits of Fennel possess a fragrant odour and a pleasant aromatic taste. They are used for flavouring soups, meat dishes, sauces and confectionary items. The fruits are aromatic, stimulant, carminative and are considered to be useful in diseases of the chest, spleen and kidney. Anethole, a chief constituent fennel, has been shown to block both inflammation and carcinogenesis, but just how these effects are mediated is not known. One possibility is TNF-mediated signaling, which has also been associated with both inflammation and carcinogenesis. Anethole suppressed TNF-induced NF- κ B activation through inhibition of I κ B α phosphorylation and degradation. Anethole also blocked the NF- κ B activation induced by a variety of other inflammatory agents. Besides NF- κ B, anethole also suppressed TNF-induced activation of the transcription factor AP-1, c-jun N-terminal kinase and MAPK-kinase. In addition, anethole abrogated TNF-induced apoptosis as measured by both caspase

activation and cell viability [168]. The antitumor activity of anethole against Ehrlich ascites carcinoma has been reported [169].

76. *Garcinia cambogia*: *Garcinia cambogia*, an edible fruit native to southeastern Asia, contains large quantities of hydroxy citric acid (HCA), which has been shown to inhibit ATP citrate, suppress de novo fatty acid synthesis and food intake, and consequently decrease body weight gain. *G. cambogia* also acts as an antiulcerogenic agent by decreasing the acidity and to increasing the mucosal defence in the gastric areas [170]. Gambogic acid (GA) its active component has been shown to possess anticancer activity through targeting to histone acetyltransferases [171]. Pandey et al [172] reported that GA enhanced apoptosis induced by TNF and chemotherapeutic agents, inhibited the expression of gene products involved in antiapoptosis (IAP1 and IAP2, Bcl-2, Bcl-xL, and TRAF1), proliferation (cyclin D1 and c-Myc), invasion (COX-2 and MMP-9), and angiogenesis (VEGF), all of which are known to be regulated by NF- κ B. GA suppressed NF- κ B activation induced by various inflammatory agents and carcinogens and this, accompanied by the inhibition of TAK1/TAB1-mediated IKK activation, inhibited I κ B α phosphorylation and degradation, suppressed p65 phosphorylation and nuclear translocation, and finally abrogated NF- κ B-dependent reporter gene expression. GA also significantly inhibited HUVEC proliferation, migration, invasion, tube formation, and microvessel growth. In a xenograft prostate tumor model, GA effectively inhibited tumor angiogenesis and suppressed tumor growth with low side effects using metronomic chemotherapy with GA. Therefore, GA inhibited the activations of VEGFR2 and its downstream protein kinases, such as c-Src, focal adhesion kinase, and AKT[173].

77. *Gaultheria yunnanensis*: Among various species of *Gaultheria*, *Gaultheria yunnanensis* are used widely in the south of China to treat rheumatoid arthritis. *G. yunnanensis* displays considerable effects against Freund's complete adjuvant induced arthritis in rats, which is in concordance with clinical practice. n-Butanol extracts and both of the eluants with water and 30% ethanol produce a significant decrease in the paw edema. 30% ethanol eluants show a stronger activity than others. It also possesses analgesic and anti-inflammatory activities, which may be mediated, at least partly, through the suppression of inflammatory mediators [174].

78. *Glycyrrhiza glabra*: *Glycyrrhiza glabra* L. is one such medicinal plant whose dried roots and stolons form an important component of various Ayurvedic formulations. There are number of reports of *G. glabra* with anti-inflammatory, anticancer, antihepatotoxic, antimicrobial, antioxidant, anti-genotoxic, hepatoprotective, cytoprotective and cytotoxic activities. Other than these, licorice extract also has antidepressant properties. This antidepressant-like effect of liquorice extract is mediated by increase of brain norepinephrine and dopamine, but not by increase of serotonin [175].

79. *Gmelina arborea*: *Gmelina arborea*'s decoction is used as a diuretic for loosening phlegm, as an appetite stimulant and in the treatment of various stomach disorders, fevers, skin problems and liver disorders. *G. arborea* is an important ingredient of generic Ayurvedic formulation "*Dashamularishta*" prescribed for several gynaecological disorders and used in several commercial ayurvedic preparations. *In vitro* studies on bark and fruit extracts showed antioxidant activity and protected liver slice culture cells by alleviating oxidative stress-induced damage to liver cells. *Ex vivo* studies of the extract on perfused isolated rabbit jejunum and *in vivo* studies based on castor oil-induced model proved to have activity against diarrhea in mice but at low doses [176].

80. *Gossypium herbaceum*: The extracts of *Gossypium herbaceum* have antimicrobial, antimutagenic and hepatoprotective properties. Gossypol is a yellow pigment, present in

Gossypium herbaceum plants, which has drawn the attention of many scientist because of its wide biological activities such as contraceptive [177] and anticancer [178].

81. *Gymnema sylvestre*: *Gymnema sylvestre* is a plant used in India and parts of Asia as a natural treatment for diabetes or “sweet urine.” The hypoglycemic action of *Gymnema* leaves was first documented in the late 1920s. *Gymnema* is reported to increase glucose uptake and utilization and improve the function of pancreatic beta cells. *Gymnema* may also decrease glucose absorption in the gastrointestinal tract. Phytochemically the plant has been reported to contain gymnemagenin the sapogenin, gymnemic acid-III, -IV, -V, -VIII, and -IX, were isolated in pure states from the hot water extract of leaves of *G. sylvestre* [179]. Glycoside of gymnemic acid may block the absorption of sugar from the intestine and sweet taste of sugars. Plant extract also increases the number of insulin producing cells in pancreas and balance insulin level.

82. *Hajarala yahuda*: It is one of the main ingredients used in the ayurvedic preparation ashmarihar ras. This preparation contains other ingredients like Yava-kshara, Muli-kshara, and Shveta-parpati, and it is sold as a diuretic (http://divyaproducts.com/index.php?dispatch=products.view&product_id=9).

83. *Hebenaria intermedia*: It is an endangered medicinal orchid known as ‘Ruddhi’ in ayurveda and distributed in grassy slopes between 2000–3000 m in Himalayan region. It is one among the constituents of “Chyawanprash” an anti aging supplements, which is purely herbal in nature.

84. *Hemidesmus indicus*: It has been widely used in treatment of various diseases including leprosy, leucoderma, leucorrhoea, syphilis, chronic rheumatism, asthma, bronchitis, gravel and other urinary diseases. The active principles of *Hemidesmus indicus*, 2-hydroxy 4-methoxy benzoic acid and pregnane glycoside, showed antihyperlipidaemic and antidyslipidemic effects [180, 181].

85. *Holarrhena antidysenterica*: Most of the study on this plant shows its antimicrobial effects and there are no literatures available on chronic diseases. Arseculeratne et al. [182] showed that rats treated with extracts of this plant produced liver lesions, disruption of the centrilobular veins, haemorrhage in the centrilobular sinusoids, focal hepatocellular necrosis and histopathology in the lungs and kidneys. These symptoms were compatible with the action of pyrrolizidine alkaloids.

86. *Hordeum vulgare*: Germinated barley foodstuffs (GBF), which are derived from brewer’s spent grain are a highly safe food substance. In an acute experimental colitis model, GBF increases butyrate production in the lower intestine and prevents mucosal damage and bloody diarrhoea. Phenolic extracts from whole barley kernel possess high antioxidant, antiradical, and antiproliferative potentials [183].

87. *Indigofera tinctoria*: True indigo the common name for *Indigofera tinctoria* is the original sources of indigo dye and is obtained from its leaves. Indirubin, an active principle from indigo has been demonstrated that it had anti-inflammatory and anti-cancer activity through suppression of transcription factor NF- κ B [184]. Sethi et al. [184] reported that indirubin suppressed NF- κ B activation induced by various inflammatory agents and carcinogens. Indirubin also blocked the phosphorylation and degradation of I κ B α through the inhibition of activation of I κ B kinase and phosphorylation and nuclear translocation of p65. NF- κ B reporter activity induced by TNFR1, TNF receptor-associated death domain, TRAF2, TAK1, NF- κ B-inducing kinase, and IKK β was inhibited by indirubin but not that induced by p65 transfection. Thus, indirubin inhibited the expression of NF- κ B-regulated

gene products involved in antiapoptosis (IAP1, IAP2, Bcl-2, Bcl-xL, and TRAF1), proliferation (cyclin D1 and c-Myc), and invasion (COX-2 and MMP-9). This correlated with enhancement of the apoptosis induced by TNF and the chemotherapeutic agent taxol in human leukemic KBM-5 cells. Indirubin also suppressed cytokine-induced cellular invasion.

88. *Inula racemosa*: It is an ornamental plant of the Asteraceae family that is used both internally, as well as externally in ayurveda. Externally it is used to dress the wounds and ulcers as the herb possesses antiseptic, antibacterial and antifungal activity and internally it is used in anorexia (loss of appetite) and dyspepsia (indigestion), cough, hiccup, bronchial asthma, reducing the excessive body fats, wound healing, amenorrhea as well as dysmenorrhea. *In vivo* study the extracts of *Inula racemosa* decreased total cholesterol, triglycerides, low-density lipoprotein cholesterol and the atherogenic index, and increased high-density lipoprotein cholesterol compared with the positive control by scavenging thiobarbituric acid reactive substances and modulating levels of reduced glutathione in liver, and superoxide dismutase and glutathione peroxidase in heart [185].

89. *Ipomoea digitata*: Vidhari Kand (*Ipomoea digitata*) comes in the plant family of sweet potato. In Ayurveda it is used as a general tonic. The leaves contain organic acids, isobutyric, (S)-2-methylbutyric, tiglic, n-decanoic, n-dodecanoic, cinnamic acids, and glycosidic acids, quamoclinic acid A and operculinic acid A [186].

90. *Ipomoea nil*: *Ipomoea nil* is a species of morning glory known as white-edge morning glory, ivy morning glory, and Japanese morning glory. Much of the medicinal uses of this plant remain unknown. The plant is much grown for its beautiful flowers and their fruits are consumed. There are many species of ipomoea, which has laxative or purgative properties. The seeds contains diterpene glycosides, and they posses moderate to mild anti cancer property against various cancer cell lines [187].

91. *Lavandula stoechas*: *L. stoechas* occurs naturally in the Mediterranean region. It has been used for a long time in traditional medicine as an anticonvulsant and antispasmodic. There are very limited literatures available about its medicinal properties and most of them describe about the isolation and constituents of essential oils from the lavender and its antimicrobial property. The roots contains triterpenes, 18-hydroxy-27-norolean-12,14-dien-30-al-28-oic acid and 3 beta-hydroxy-1-oxo-olean-12-ene-30-al-28-oic acid which has been evaluated for the anticancer property [188]. Apart from that there are no literatures available about its use in preventing chronic diseases.

92. *Leucas cephalotes*: *Leucas cephalotes*, the flowering annual herb, is a common weed, which is used in ayurveda to treat several ailments including diabetes. The ethanolic extract of leaves regulates carbohydrate and lipid metabolism and improves antioxidant status in insulin-dependent and non-insulin-dependent diabetes mellitus rats through modulating hepatic glycogen, blood urea and creatinine contents, and hexokinase and glucose-6-phosphatase activities [189].

93. *Malaxis acuminata*: There are no literatures available about the medicinal uses of this plant. This plant is otherwise called as Jeevak and found in India, China, and South-East Asia, at elevations up to to 1400 m. It is a small to medium sized, hot to warm growing terrestrial or lithophytic orchid. Its pseudobulbs are sweet, refrigerant, aphrodisiac, febrifuge and tonic. They are useful in haematemesis, fever, seminal weakness, burning sensations, dipsia, emaciation, tuberculosis and general debility (<http://www.flowersofindia.net/catalog/slides/Jeevak.html>).

94. *Mangifera indica*: Mango is rich in a variety of phytochemicals and nutrients such as vitamins A, C, B6, K and E, polyphenols, omega-3 and -6 poly unsaturated fatty acids and provitamin A carotenoids. The mango contains a triterpene called lupeol which shows strong anti-cancer activity by disrupting survivin/cFLIP activation, modulation of expression levels of cyclins-A, -B1, -D1, -D2, -E2, cyclin-dependent kinase (cdk)-2, CDK-inhibitor p21 and finally inducing G2/M cell cycle arrest [190]. Mangiferin, a xanthone glucoside, isolated from the leaves of *Mangifera indica* possesses significant antidiabetic, antihyperlipidemic and antiatherogenic properties [191].

95. *Mentha piperita*: Peppermint has a long tradition of medicinal use, with archaeological evidence placing its use as far back as ten thousand years ago. Many literatures show its anticancer and radioprotective potentials *in vivo* [192, 193]. It also possesses anti-nociceptive effect against acetic acid-induced writhing and hot plate-induced thermal stimulation and also anti-inflammatory effect against xylene-induced ear oedema and cotton-pellet granuloma [194].

96. *Mesua ferrea*: The xanthenes, mesuaxanthone-A, mesuaxanthone-B and euxanthone, that have been isolated from *Mesua ferrea* exhibits anti-inflammatory activity in normal and adrenalectomised rats as tested by carrageenin induced hind paw oedema, cotton pellet granuloma and granuloma pouch techniques. *M. ferrea* also possesses antioxidant [195], but the molecular mechanisms for its activities are yet to be elucidated.

97. *Mimusops elengi*: The bark, flowers, fruits and seeds of *Mimusops elengi* are generally used as astringent, cooling, anthelmintic, tonic, and febrifuge. It is mainly used in dental ailments like bleeding gums, pyorrhea, dental caries and loose teeth. Ethyl acetate extract possesses anti-ulcer activity against experimental gastric ulcers by decreasing the gastric acid secretory activity along with strengthening of mucosal defensive mechanisms [196].

98. *Momordica charantia*: *M. charantia* is depicted that it is helpful in treating wound, ulcer, dysmenorrhea, eczema, gout, jaundice, kidney stone, leprosy, leucorrhea, piles, pneumonia, psoriasis, rheumatism and scabies. Earlier studies performed with MC extract have demonstrated its antidiabetic, antiviral, antitumor, antileukemic, antibacterial, antihelminthic, antimutagenic, antimycobacterial, antioxidant, antiulcer, anti-inflammatory, hypocholesterolemic, hypotriglyceridemic, hypotensive, immunostimulant and insecticidal properties [197, 198].

99. *Moringa oleifera*: Various parts of this plant such as the leaves, roots, seed, bark, fruit, flowers and immature pods act as cardiac and circulatory stimulants, possess antitumor, antipyretic, antiepileptic, antiinflammatory, antiulcer, antispasmodic, diuretic, antihypertensive, cholesterol lowering, antioxidant, antidiabetic, hepatoprotective, antibacterial and antifungal activities. Ethanolic extract of seeds showed protection against acetylcholine-induced broncho-constriction and airway inflammation [199].

100. *Mucuna pruriens*: *Mucuna pruriens* seeds contain high concentrations of levodopa, a direct precursor of the neurotransmitter dopamine. It has long been used in traditional Ayurvedic Indian medicine for diseases including Parkinson's disease. This plant also showed the hypoglycaemic activities in alloxan diabetic rats [200].

101. *Nigella sativa*: *N. sativa* has been used in the treatment of a variety of illnesses, including bronchial asthma, headache, dysentery, infections, obesity, back pain, hypertension, gastrointestinal problems, and eczema. Several components of black cumin have been identified, including thymoquinone, thymol, thymohydroquinone, and dithymoquinone. Thymoquinone (TQ), the most abundant component of black seed oil, has

been reported to exhibit antioxidant, antiinflammatory, and chemopreventive effects. For instance, TQ has been shown to suppress the proliferation of various tumor cells, including colorectal carcinoma, breast adenocarcinoma, osteosarcoma, ovarian carcinoma, myeloblastic leukemia, and pancreatic carcinoma, although it is minimally toxic to normal cells. TQ, the active principle of *N. sativa*, has been reported that it suppressed NF- κ B activation induced by various carcinogens and inflammatory stimuli. The suppression of NF- κ B signaling pathway led down-regulated the expression of NF- κ B-regulated antiapoptotic (IAP1, IAP2, XIAP Bcl-2, Bcl-xL, and survivin), proliferative (cyclin D1, cyclooxygenase-2, and c-Myc), and angiogenic (matrix metalloproteinase-9 and vascular endothelial growth factor) gene products [10]. This led to potentiation of apoptosis induced by tumor necrosis factor and chemotherapeutic agents. Indeed, Yi et al [201] reported that TQ effectively inhibited human umbilical vein endothelial cell migration, invasion, and tube formation. TQ inhibited cell proliferation and suppressed the activation of AKT and extracellular signal-regulated kinase. Therefore, TQ blocked angiogenesis in both *in vitro* and *in vivo*, prevented tumor angiogenesis in a xenograft human prostate cancer (PC3) model in mouse, and inhibited human prostate tumor growth at low dosage with almost no chemotoxic side effects. Thymoquinone inhibited vascular endothelial growth factor-induced extracellular signal-regulated kinase activation but showed no inhibitory effects on vascular endothelial growth factor receptor 2 activation. Overall, these results indicate that TQ could be used as a potential drug candidate for cancer therapy.

102. *Nardostachys jatamansi*: *Nardostachys jatamansi* Jones DC (commonly named as jatamansi) is used for treatment of mental disorders, insomnia, hyperlipidemia, hypertension and heart diseases. It has protective effect in parkinsonism, epilepsy, cerebral ischemia, liver damage. Various sesquiterpenes (such as Jatamansic acid and Jatamansone) have been reported to be present in the rhizomes of the plant [202].

103. *Nelumbo nucifera*: Lotus has been known for a long time in Ayurvedic literature as an antipyretic, diuretic, as an astringent remedy in diarrhoea and as an aphrodisiac. The dry powder prepared from its flowers has recently been used in the treatment of Diabetes mellitus by Ayurvedic physicians. Armpavine (Arm), an active compound from *N. nucifera*, has been shown to exert immunosuppressive effects both *in vitro* and *in vivo* through inhibition of NF- κ B activation pathways [203]. *In vitro*, Arm suppressed NF- κ B activation and MAPK (p38, ERK1/2, and JNK) phosphorylations and *in vivo*, Arm attenuated the mRNA expression levels of col1 α 2, TGF- β 1, TIMP-1, ICAM-1, iNOS, and IL-6 genes. Kim et al [204] has also shown the downregulation of iNOS and TNF alpha expression via NF- κ B modulation another active compound, kaempferol.

104. *Nyctanthes arbortristis*: Traditionally, the flowers of *Nyctanthes arbortristis* are known to be effective as stomachic, carminative, astringent, antibilious, expectorant, hair tonic and are used in the treatment of piles and various skin diseases. The bark is used for the treatment of bronchitis and snakebite. *N. arbortristis* exhibits potential anti-inflammatory and antinociceptive activity by inhibiting histamine and serotonin induced edema formation and its analgesic may be due to inhibition of the action of prostaglandins [205]. Oral administration of leaf and fruit extracts from *N. arbortristis* reduced the expression of various cytokines, such as TNF- α , IL-1 β and IL-6, in arthritic mice [205].

105. *Ocimum sanctum*: *Ocimum sanctum* commonly known as tulsi in Ayurvedic medicine has demonstrated various medicinal values predominantly by its antioxidant property. Different parts of the plant are traditionally used in Ayurveda and Siddha systems for the treatment of diverse ailments like infections, skin diseases, hepatic disorders and as an antidote for snake bite and scorpion sting. The plant has also demonstrated antidiabetic [206] and hypolipidemic effect [207]. Ursolic acid (UA), a pentacyclic triterpene acid from

O. sanctum has been reported to suppress NF- κ B activation induced by various carcinogens including TNF, phorbol ester, okadaic acid, H₂O₂, and cigarette smoke. Ursolic acid inhibited degradation and phosphorylation of I κ B α , I κ B kinase activation, p65 phosphorylation, p65 nuclear translocation, and NF- κ B-dependent reporter gene expression. The inhibition of NF- κ B activation correlated with suppression of NF- κ B-dependent cyclin D1, COX-2, and MMP-9 expression [208]. UA also inhibited both constitutive and IL-6-inducible STAT3 activation in a dose- and time-dependent manner in multiple myeloma cells. The suppression was mediated through the inhibition of activation of upstream kinases c-Src, JAK-1, JAK-2, and ERK1/2. Ursolic acid down-regulated the expression of STAT3-regulated gene products such as cyclin D1, Bcl-2, Bcl-xL, survivin, Mcl-1, and vascular endothelial growth factor. Finally, ursolic acid inhibited proliferation and induced apoptosis and the accumulation of cells in G1-G0 phase of cell cycle. This triterpenoid also significantly potentiated the apoptotic effects of thalidomide and bortezomib in multiple myeloma cells [209].

106. Operculina turpethum: *O. turpethum* (Family: Convolvulaceae), commonly known as trivrit or nishot in the western part of India and adjoining Pakistan, is a plant with immense ethno-medicinal value. *O. turpethum* extract is used to treat wide range of ailments. For instance, it is used to relieve periodic fevers, constipation, flatulence and colic obesity, to treat anaemia, splenomegaly, raised lipid levels and obesity. Turpethinic acids (A, B, C, D, and E) are isolated from the resin of the plant and lupeol, betulin, and β -sitosterol are isolated from the stem. Recently, their structurally related compounds like lupeol, betulin and sitosterol have been identified to possess a variety of pharmacological activities such as hepatoprotective, anticancer, and anti-inflammatory effects [210].

107. Orchis mascula: *O. mascula* has been proved to have antihypertensive, antidyslipidemic and endothelial modulating activities [211]. These effects are mediated through multiple pathways that include direct vasodilation by calcium channel blockade and reduction of plasma lipids by inhibition of biosynthesis, absorption and secretion.

108. Oroxyllum indicum: In Indian traditional medicine, the roots as well as stem bark of *Oroxyllum indicum* (Family: Bignoniaceae), commonly known as 'Syonaka', has been used for centuries for the treatment of various gastric disorders. *Oroxyllum indicum* used in Bangladeshi folk medicine has been studied for its anticancer potential. Baicalein from *O. indicum* showed anti-cancer potential in leukemia cells through inducing cell cycle arrest and apoptosis [212].

109. Pandanus tectorius: *Pandanus tectorius* is a species of Pandanus (screw pine) that is native to Malesia, eastern Australia, and the Pacific Islands. The fruit can be eaten raw or cooked and is a major source of food in Micronesia, especially in the atolls. In Kiribati, pandanus leaves are used in treatments for cold/flu, hepatitis, dysuria, asthma, boils, and cancer, while the roots are used in a decoction to treat hemorrhoids. In Hawai'i the main parts used in making traditional medicines are the fruits, male flowers, and aerial roots [213]. These are used individually or in combination with other ingredients to treat a wide range of illnesses, including digestive and respiratory disorders.

110. Phyllanthus amarus: *Phyllanthus amarus* is traditionally used to treat flu, dropsy, diabetes, and jaundice, but it has also been reported to inhibit hepatocellular carcinoma development. *P. amarus* has potent free radical scavenging activity and could scavenge superoxides and hydroxyl radicals and inhibit lipid peroxides. Moreover, Kassuya *et al.*, have shown that the extract from *P. amarus* or some of the purified lignans such as phylltetralin, nirtetralin and niranthin exhibit *in vivo* and *in vitro* anti-inflammatory

properties. These anti-inflammatory properties are probably mediated through its direct ability to interact with platelet activating factor receptor binding sites [214].

111. *Phyllanthus niruri*: *Phyllanthus niruri* is a widespread tropical plant commonly found in coastal areas, including South East Asia, Southern India and China. Extracts of this herb have shown promise in treating a wide range of human diseases, such as dysentery, influenza, vaginitis, tumours, diabetes, diuretics, jaundice, kidney stones and dyspepsia. The plant is also useful for treating hepatotoxicity, hepatitis B, hyperglycaemia and viral and bacterial diseases. *P. niruri* has been used in Ayurvedic medicine for over 2000 years.

112. *Picrorhiza kurroa*: The extracts from roots and rhizomes of this plant (commonly called as katuka, kutki, or kutaki) are used to treat a variety of ailments, including fever, hepatitis, allergies, asthma, and other inflammatory diseases. Picroliv, an iridoid glycoside derived from *P. kurroa*, interfered the activation of NF- κ B signal cascade. Picroliv abrogated TNF-induced activation of NF- κ B thorough inhibition of I κ B kinase, leading to inhibition of phosphorylation and degradation of I κ B α . It also inhibited phosphorylation and nuclear translocation of p65. Further, picroliv directly inhibits the binding of p65 to DNA, which was reversed by the treatment with reducing agents, suggesting a role for a cysteine residue in interaction with picroliv. Mutation of Cys(38) in p65 to serine abolished this effect of picroliv. NF- κ B inhibition by picroliv leads to suppression of NF- κ B-regulated proteins, including those linked with cell survival (inhibitor of apoptosis protein 1, Bcl-2, Bcl-xL, survivin, and TNF receptor-associated factor 2), proliferation (cyclin D1 and cyclooxygenase-2), angiogenesis (vascular endothelial growth factor), and invasion (intercellular adhesion molecule-1 and matrix metalloproteinase-9). Suppression of these proteins enhanced apoptosis induced by TNF [215].

113. *Pinus roxburghii*: *Pinus roxburghii* (Chir Pine) named after William Roxburgh, is a pine native to the Himalaya. The turpentine obtained from the resin is antiseptic, diuretic, rubefacient and vermifuge. It is a valuable remedy used internally in the treatment of kidney and bladder complaints and is used both internally and as a rub and steam bath in the treatment of rheumatic affections. It is also very beneficial to the respiratory system and so is useful in treating diseases of the mucous membranes and respiratory complaints such as coughs, colds, influenza and TB. Externally it is a very beneficial treatment for a variety of skin complaints, wounds, sores, burns, boils etc and is used in the form of liniment plasters, poultices, herbal steam baths and inhalers. The wood is diaphoretic and stimulant. It is useful in treating burning of the body, cough, fainting and ulcers.

114. *Piper chaba*: The plant *Piper chaba* is a climbing, glabrous shrub available in various parts of India and Malay Islands. Furthermore, *P. chaba* is commonly used as pepper in the southern part of Bangladesh. Various parts of this plant have been extensively used in different traditional formulations including ayurveda. The root is useful against asthma, bronchitis, and consumption. The fruit is thermogenic, anthelmintic, expectorant, carminative and improves appetite and taste and is also used against asthma, bronchitis, fever, inflammation, piles, pain in the abdomen and at the anus. The fruit has stimulant and carminative properties, and is used in haemorrhoidal affections. Stem is used to alley post-delivery pain in mothers and also useful in rheumatic pains and diarrhea.

115. *Piper longum*: *Piper longum* (Long pepper) is a flowering vine in the family Piperaceae, cultivated for its fruit, which is usually dried and used as a spice and seasoning. The fruits contain the alkaloid piperine. *P. longum* is a component of Indian traditional medicine reported to be used as a remedy for treating gonorrhea, menstrual pain, tuberculosis, sleeping problems, respiratory tract, infection, chronic gut-related pain and arthritic conditions. Piper extracts and piperine possess inhibitory activities on prostaglandin

and leukotrienes COX-1 inhibitory effect, as well as on NF- κ B activation, and thus exhibit anti-inflammatory activity [216, 217].

116. *Piper nigrum*: Black pepper (*Piper nigrum*) is commonly used as a spice in human diets, but it is also used as a medicine, a preservative, and a perfume in many Asian countries. An extract of the active phenolic component, piperine, is well known to provide beneficial physiological effects. It stimulates the digestive enzymes of pancreas, protects against oxidative damage, lowers lipid peroxidation, and enhances the bioavailability of a number of therapeutic drugs. In addition, its anti-inflammatory activities have been demonstrated in rat models of carrageenan-induced rat paw edema, cotton pellet-induced granuloma, and a croton oil-induced granuloma pouch. Constituents of the piper species have shown *in vitro* inhibitory activity against the enzymes responsible for leukotriene and prostaglandin biosynthesis, 5-lipoxygenase and COX-1, respectively [217]. These effects of piperine seem to be beneficial for inflammatory diseases that are accompanied by severe pain; for example, rheumatoid arthritis.

117. *Pistacia integerrima*: *Pistacia integerrima* (Anacardiaceae) is a moderate size deciduous tree with a short stout bole widely distributed in the sub-alpine regions of Himalaya ranging from: Indus to Kumaun. The plant has been used in traditional medicine for rheumatic pain, analgesic and antipyretic effects, and analgesic and anti-inflammatory activities [218].

118. *Pluchea lanceolata*: *Pluchea lanceolata* is widely used for rheumatism and allied disorders, diseases of the abdomen, dyspepsia, bronchitis and inflammation. The decoction of *P. lanceolata* has been used traditionally for the treatment of arthritis. Flavanols isolated from this plant, such as quercetin and rutin are known for their antihistamine, anti-inflammatory and antiviral activities. Quercetin has been shown to mediate down-regulation of mutant p53 in human breast cancer cell lines [219] and to inhibit chemically induced carcinogenesis.

119. *Plumbago zeylanica*: The root of *Plumbago zeylanica* (also called Chitrak), a major source of plumbagin, has been used in the Indian medicine since the period of Charaka, from 750 BC, as an antiatherogenic, cardiogenic, hepatoprotective, and neuroprotective agent [220]. Plumbagin has been shown to exert anticancer and antiproliferative activities in animal models as well as in cells in culture. Sandur et al., suggest that plumbagin may be effective against cancer not only by suppressing invasion but also by inhibiting angiogenesis and inflammation through inhibition of the NF- κ B signaling pathway [221]. Recently, plumbagin shown that it inhibited both constitutive and interleukin 6-inducible STAT3 phosphorylation in human multiple myeloma cells and this correlated with the inhibition of c-Src, Janus-activated kinase (JAK)1, and JAK2 activation. Thus, the inhibition of STAT3 signaling pathway by plumbagin leading the chemosensitization of cancer cells [222].

120. *Polygonatum verticillatum*: *P. verticillatum* is a perennial rhizomatous herb with an extensive range through the northern Hemisphere from Europe to the Himalayas to Siberia. The syrup of the fresh rhizome of this plant is used in the treatment of pain, pyrexia, burning sensation and for phthisis. Other ethnobotanical uses of the plant include as emollient, aphrodisiac, vitiated condition of pitta and vata, appetizer and tonic, galactagogue (increases milk release) and weakness. Recently, this plant also reported its antinociceptive activity [223].

121. *Pongamia pinnata*: *Pongamia pinnata* is a medium sized glabrous tree, found throughout India and further distributed eastwards, mainly in the littoral regions of South Eastern Asia and Australia. The seed and seed oil of this plant have been used for treating

various inflammatory and infectious diseases such as leucoderma, leprosy, lumbago, muscular and articular rheumatism. The leaves are hot, digestive, laxative, anthelmintic and cure piles, wounds and other inflammations [224].

122. *Prunus amygdalus*: *Prunus amygdalus* is a species of tree native to the Middle East. Claimed health benefits of this plant include improved complexion, improved movement of food through the colon (feces) and the prevention of cancer. In Ayurveda, *P.amygdalus* is considered a nutritive for the brain and nervous system. Recent studies have shown that the constituents of this tree have anti-inflammatory, immunity boosting, and anti-hepatotoxicity effects [225].

123. *Pseudarthria viscida*: In Ayurvedic medicines, this plant is namely, 'Dashamoola' 'Mahanarayana Taila' and 'Dhantara Taila'. It is used to treat vitiated conditions of pita and vata, asthma, tuberculosis, helminthiasis, dyspepsia, diarrheas, neurasthenia, diabetes, cardiopathy, hyperthermia and general debility and as a nasal drop in headache. The ethanol-induced gastric ulceration in mice was significantly reduced by oral administration of *P. viscida* extract [226].

124. *Psoralea corylifolia*: The seeds of *Psoralea corylifolia* (Babchi) contain a variety of coumarins including psoralen with medicinal uses. Psoralen plus UVA (PUVA) is used as a very effective treatment modality for various diseases, including psoriasis, eczema, vitiligo and cutaneous T-cell lymphoma. PUVA-induced immune suppression and/or apoptosis are proposed to be responsible for the therapeutic potential [227].

125. *Pterocarpus marsupium*: Parts of this plant (heart wood, leaves, flowers) have long been used for their medicinal properties in Ayurveda. The heartwood is used as an astringent and in the treatment of inflammation and diabetes. A multicentric study has shown that a preparation from the plant is effective in reducing levels of blood glucose and glycosylated haemoglobin in patients with non-insulin-dependent diabetes mellitus. Extract from the plant has been reported to selectively inhibit COX-2 [228].

126. *Pterocarpus santalinus*: It is indigenous to the Indian Peninsula and is chiefly of importance from its yielding the red dye-wood known as red saunders. The heartwood obtained from the plant has various medicinal properties, as a cooling agent, antipyretic, anti-inflammatory, anthelmintic, tonic, hemorrhage, dysentery, aphrodisiac, to treat eye diseases, mental aberrations and ulcers. In a recent study, ethanolic extract obtained from the bark of plant was found to decrease hyperglycemia by increasing glycolysis and decreasing gluconeogenesis in streptozotocin-induced diabetic rats [229].

127. *Pueraria tuberosa*: It is a perennial climber, growing throughout tropical parts of India. In the Ayurvedic system of medicine, the plant is used as a drug of choice to manage pain, inflammation and other related diseases. The Chayawanprash, one of the popular ayurvedic formulations with powerful anti-oxidant potential contains the plant extract as active component.

128. *Punica granatum* (Pomegranate): The tree represents a phytochemical reservoir of heuristic medicinal value. The seed, juice, peel, leaf, flower, bark, and roots of the plant have pharmacologic activity. The juice and peel possess anticancer activities, including interference with tumor cell proliferation, cell cycle, invasion and angiogenesis. Some of the known compounds with anti-cancer and anti-inflammatory activity obtained from the plant include -tocopherol, ursolic acid, punicic acid, hydroxycinnamic acids, quercetin, ellagitannins, flavonols, flavones, apigenin, maslinic acid and asiatic acid. Ellagitannins, and punicalagin from pomegranate have been reported to inhibit cancer cell proliferation and

apoptosis through the modulation of NF- κ B signaling pathway and suppression of NF- κ B-regulated gene expression. Juice (PJ), total pomegranate tannin extract (TPT) and punicalagin from pomegranate significantly suppressed TNF α -induced COX-2 protein expression. Additionally, PJ reduced phosphorylation of the p65 subunit and inhibited NF- κ B DNA-binding. TPT and punicalagin, also, suppressed NF- κ B DNA-binding, whereas ellagic acid was ineffective. PJ also abolished TNF α -induced AKT activation, needed for NF- κ B activity. Therefore, the polyphenolic phytochemicals in the pomegranate can play an important role in the modulation of inflammatory cell signaling in colon cancer cells [230].

129. *Putranjiva roxburghii*: It is a moderate-sized, evergreen tree, growing up to 12 m in height. The pollen from the plant has been found an important aeroallergen for type I hypersensitivity. The leaf extract from the plant has shown potential as antinociceptive, antipyretic, and anti-inflammatory activities in mice [231].

130. *Quercus infectoria*: It is a small tree, most abundant in Asia Minor, and extends up to middle Asia. The galls of the plant possess pleiotropic therapeutic activities, with particular efficacy against inflammatory diseases. Oral administration of gall extract from the plant has been reported to significantly inhibit carrageenan, histamine, serotonin and prostaglandin E2 (PGE2) induced paw oedemas, while topical application of gall extract has the ability to inhibit phorbol-12-myristate-13-acetate (PMA) induced ear inflammation in various *in vivo* and *in vitro* experimental models [232].

131. *Raphanus sativus*: It is a cruciferous plant, rich on flavonoids, isothiocyanates, and phenolic acids and has potential as anti-inflammatory and immunomodulatory agent both *in vitro* and *in vivo*. The granules from the plant have shown potential to act as anti-inflammatory agent in a high fat diet fed rat model [233].

132. *Rauwolfia serpentina*: *R. serpentina* is a tropical woody plant of the Apocyanaceae family indigenous to Asia, South America and Africa. Extracts of different parts of the plant had been used in Hindu medicine for snakebite, insomnia, insanity and many other diseases. It is one of the fundamental herbs used in traditional Chinese medicine. It is also one of the main ingredients in the ayurvedic formulation 'Divya Mukta Vati' used to cure high blood pressure. The plant contains a number of bioactive components, including ajmaline, deserpidine, rescinamine, serpentinine and reserpine. Reserpine, an alkaloid from *Rauwolfia serpentina*, was widely used for its antihypertensive action and single dose of *R. serpentina* formulation is effective, showed by Leary et al [234] in a double-blind placebo-controlled investigation. In animal models reserpine has been reported to significantly reduce inflammation [235, 236].

133. *Ricinus communis*: Commonly known as castor oil plant, it is indigenous to the southeastern Mediterranean Basin, Eastern Africa, and India. The seed from the plant is a rich source of triglycerides (mainly ricinolein) and ricin. The oil obtained from the seed of the plant has been used as a laxative, purgative, and cathartic in Unani, Ayurvedic and other ethnomedical systems. Traditional Ayurvedic medicine considers castor oil the king of medicines for curing arthritic diseases. In a study carried out in guinea-pig eyelid, ricinolein was found to possess both pro-inflammatory and anti-inflammatory properties that were observed upon acute and repeated application of the compound, respectively [237].

134. *Rosa centifolia*: The plant is particular to the French city of Grasse. It is widely cultivated for its singular fragrance - clear and sweet, with light notes of honey and green earth. The plant is one of the ingredients in ayurvedic formulations 'Divya udarakalpa curna', 'Divya Curna', 'Divya peya', 'Himalaya abana'. The flowers are commercially

harvested for the production of rose oil, which is commonly used in perfumery. In a study, the plant was found effective in decreasing the severity of menstrual cramps in women[238].

135. *Rosa damascena*: The plant is a rose hybrid, derived from *Rosa gallica* and *Rosa moschata* and grows as deciduous shrub. *R. damascena* has been reported to exert antidiabetic, antilytic and to use for treating ophthalmic disorders. The roxyloside A (flavonoid) obtained from the plant was shown potential of inhibiting 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase system and was proposed to be effective in improving the cardiovascular system [239]. The oil from the plant has been quoted extensively in ancient literature as being an anxiolytic treatment.

136. *Roscoea alpina*: It is a common Himalayan wildflower and occurs most frequently in open woodlands and on rocky slopes. It is one of the main ingredients in the ayurvedic formulation 'Divya pidantaka taila' used for relieving joint pain, cervical spondylitis, trauma, oedema and inflammation.

137. *Rubia cordifolia*: It is a flowering plant belonging to the family Rubiaceae. It is a common medicinal plant used in the preparation of different formulations in Ayurveda. The root of the plant is commonly known as Manjistha and its dried samples are sold in the market under the name Manjith. The roots of the plant are used as anti-inflammatory, haemostatic, antidysentric, antipyretic, analgesic and the anthelmintic agent. It is also used in the cure of leucoderma, ulcers, urinary discharges, jaundice, and piles. Mollugin, one of the active compounds obtained from the plant was recently shown to inhibit TNF--induced expression of inflammatory molecules by inhibiting NF- κ B activation in colon cancer cells [240].

138. *Rumex maritimus*: It is one of the ingredients in the ayurvedic formulation 'Himalayan Diabecon' that has potential to lower blood sugar levels and minimizing long-term diabetic complications. Leaves are applied to burns; seeds are tonic and remove pain from the back. The methanol extract from the root of the plant has shown antidiarrhoeal activity in mice [241].

139. *Salvadora persica*: The plant has been used for centuries as a natural toothbrush and contains a number of medically beneficial properties including abrasives, antiseptics and astringent. The plant has been shown to contain trimethylamine salvadorine, chloride, fluoride, silica, sulphur, vitamin C, resins and traces of tannins, saponins, flavonoids and sterol. The plant has also been shown to possess antiulcer activity in an experimental rat model [242].

140. *Santalum album*: It is a small tropical tree of the Santalaceae family and has been utilised, cultivated and traded for many years, some cultures placing great significance on its fragrant and medicinal qualities. Plant has been the primary source of sandalwood. The wood from the plant has been shown effective against kidney and urinary disorders. The sandalwood oil obtained from the plant has been used as aromatherapy agent to relieve anxiety, stress, and depression [243].

141. *Sapindus trifoliatus*: A thick aqueous solution of the pericarp from the plant is used for the treatment of hemicrania, hysteria or epilepsy in folklore medicine. The aqueous extract of the plant has shown antihyperalgesic activity by antagonising dopamine D2 activity in mice [244]. The ethanolic extract from the seeds of the plant has shown anti-inflammatory activity in wistar rats [245].

142. *Saraca asoca*: *Saraca asoca* (local names: Ashok, Anganapriya, etc.) is a medicinal plant whose bark is astringent and used in menorrhagia, bleeding haemorrhoids, haemorrhagic dysentery, bone fractures, strangury, vesical calculi, hyperdyspsia, and inflammation. This plant is known to contain tannins, flavonoids, proanthocyanidins and leucoanthocyanidins in the bark. Two procyanidin dimers from *S. asoca* have shown the inhibitory effect of prostaglandin H2 (PGH2) synthetase [246].

143. *Saraca indica*: It is known to be useful for uterine disease, internal piles, diabetes, dyspepsia, indigestion, burning sensation, blood disorders, fractures, tumors, bites, ulcerations, and skin discoloration. The lectin saracin, purified from *Saraca indica* seed integument, has been found to agglutinate human lymphocytes and erythrocytes [247].

144. *Saussurea lappa*: *S. lappa* is considered as antiseptic, astringent, diuretic, aphrodisiac, antispasmodic, antihelmentic, and sedative and are used for the treatment of asthma, dyspepsia, rheumatism, cough, throat infections, tuberculosis, leprosy, malaria, convulsions, fever, helminth infestation and many other diseases. It is also used as a part of preparation for the treatment of various liver disorders. Recently, this plant is reported that it had antiulcer, anti-inflammatory and antiarthritic activities [19, 248].

145. *Saxifraga ligulata*: This plant is reported to be helpful in dissolving kidney stones and urinary antiseptic in action. In lower doses, the extract is mildly diuretic.

146. *Sesamum indicum*: The seeds of *S. indicum* (sesame) are used as a demulcent in respiratory affections, infantile cholera, diarrhea, dysentery and other bowel affections and bladder diseases. The seed powder is known to be benefit in amenorrhea, dysmenorrhea, ulcers and bleeding piles. The active component sesaminol is reported to its antioxidative, neuroprotective effects. Other lignan from sesame, sesamol, has been offered protection against increased blood pressure, hyperlipidaemia [249]. Recently, Harikumar et al [250] found that sesamin, lignan from *S. indicum*, inhibited the proliferation of a wide variety of tumor cells including leukemia, multiple myeloma, and cancers of the colon, prostate, breast, pancreas, and lung. Sesamin also potentiated TNF-induced apoptosis and this correlated with the suppression of gene products linked to cell survival (Bcl-2 and survivin), proliferation (cyclin D1), inflammation (COX-2), invasion (MMP-9, ICAM-1), and angiogenesis (VEGF). Sesamin downregulated constitutive and inducible NF- κ B activation induced by various inflammatory stimuli and carcinogens, and inhibited the degradation of I κ B α , through the suppression of phosphorylation of I κ B α and inhibition of activation of I κ B kinase, thus resulting in the suppression of p65 phosphorylation and nuclear translocation.

147. *Sida cordifolia*: It is reported to possess analgesic, anti-inflammatory, anticancer, diuretic, laxative, hypoglycemic and hepatoprotective activities. Further, studies showed that aqueous fraction of hydroalcoholic extract of leaves induce vasorelaxation, hypotension and bradycardia. This plant also used for treatment of Parkinson's disease and as an anti-rheumatic agent and CNS depressant. The plant alkaloid cryptolepine from *S. cordifolia* has been reported to induces cell cycle arrest in a human osteosarcoma cell line [251]

148. *Solanum indicum* (syn. *Solanum anguivi*): *S. indicum* has been used on treatment of hypertension and diabetics. The steroidal glycosides from fruits have been claimed in folk medicine to have an antihypertensive effect [252] and anticancer effect [253].

149. *Solanum nigrum*: This plant is known as 'Black nightshade' that have been extensively used to cure liver disorders, chronic skin ailments (psoriasis and ringworm), inflammatory conditions, painful periods, fevers, diarrhoea, eye diseases, hydrophobia, etc.

The plant contains glycoalkaloids (solanine, solamargine, solanigrine and solasodine), steroidal glycosides (β -solamargine, solasonine and α,β -solansodamine), diosgenin, gitogenin, tannin and polyphenolic compounds. The fruits are commonly used as hepatoprotective agents [254], which also afford protection against free radical mediated damage.

150. *Solanum xanthocarpum*: *S. xanthocarpum* are known for several medicinal uses like anthelmintic, antipyretic, laxative, antiinflammatory, antiasthmatic and aphrodisiac activities. The stem, flowers and fruits are prescribed for relief in burning sensation in the feet accompanied by vesicular eruptions. The hot aqueous extract of dried fruits is used for treating cough, fever and heart diseases. Results from pilot study with patient having allergy and asthma showed that *S. xanthocarpum* relived the bronchial asthma by bronchodilating effect, reducing the bronchial mucosal edema, and/or reducing in the secretions within the airway lumen [255].

151. *Sphaeranthus indicus*: *S. indicus*, commonly called Gorakhmundi (in Hindi), is plant is known to possess various medicinal properties. It is reported to use in epileptic convulsions, mental illnesses and hemicranias. It is also used to treat vitiated conditions of jaundice, diabetes, leprosy, fever, pectoralgia, cough, gastropathy, hernia, haemorrhoids, helminthiasis, dyspepsia, skin diseases and as a nerve tonic. The oil from the plant root has been used to treat scrofula and as an aphrodisiac, while the external application of the herb paste has been reported to use treatment for pruritus, oedema, arthritis, filariasis, gout and cervical adenopathy [256].

152. *Stereospermum suaveolens*: *Stereospermum suaveolens* is a medicinal tree species and various parts of the plants are used by traditional healers, rural communities and pharmaceutical companies as a remedy for vomiting, eructation, piles, acidity, diarrhoea, gonorrhoea, loss of taste, malaria and other fevers. Balasubramanian et al [257] showed that ethanol extract of *Stereospermum suaveolens* possesses maximum anti-inflammatory activity in various rat models of carrageenan-, dextran-, and histamine-induced hind paw edema, and cotton pellet-induced granuloma formation in a dose-dependent manner.

153. *Strychnos nuxvomica*: The dried seeds of *S. nuxvomica* have been claimed to improve blood circulation and relieve rheumatic pain. Historically, this plant has been widely used in treating diseases, such as tumor and rheumatic arthritis. It also has been used in gastro-hepatic disease and as analgesic, stimulant. Indeed, it also used to treatment of paralysis, diabetes, gonorrhea, anemia and bronchitis etc. Brucine and brucine N-oxide from this plant are reported to exert anti-inflammatory effects through reduction of prostaglandin E2 release [258].

154. *Swertia chirata*: This plant is known as 'Chirata', and multifarious therapeutic value. It is used as an antimalarial, a bitter stomachic, anthelmintic, and as a remedy for scanty urine, epilepsy, ulcer, bronchial asthma and certain type of mental disorder. Studies on the biological activities of *S. chirata* extract reveal that this bitter plant possesses antioxidant, antidiabetic, antimicrobial, anticholinergic and chemopreventive activity. It contains mangiferin along with various secoiridoid glycosides (i.e. amarogentin, amaroswerin, sweroside and swertiamarin). The secoiridoid glycoside, amarogentin, is reported suppressing COX-2 [259] and also possesses various biological activities such as chemopreventive, antibacterial, anticholinergic and antihepatitis activity.

155. *Symplocos crataegoides* (syn. *Symplocos paniculata*): *S. crataegoides* has been used for treatment of menorrhagia, eye diseases, bowel complaints, dysentery, inflammations, vaginal discharges, leprosy and ulcers, as a gargle for giving firmness to spongy and

bleeding gums. And its bark juice is also used to sprains and muscular swellings. The ursane-type triterpenes from this plant showed that the inhibitory effect on protein tyrosine phosphatase 1B (PTP1B) has been proposed as a therapy for treatment of type 2 diabetes and obesity [260].

156. *Syzygium aromaticum*: Traditionally, it has been used to treat respiratory and digestive ailments. Cloves are used as a carminative, to increase hydrochloric acid in the stomach and to improve peristalsis. Cloves are also said to be a natural anthelmintic. The aqueous clove infusion was showed antiseptic and antibiotic properties, clove is used to treat toothache and as an ingredient in a popular toothpaste and mouthwash in India. An anti-herpes virus compound eugenin was purified from clove, which could inhibit viral DNA synthesis. In addition water extracts of clove have shown the inhibitory effect on hepatitis C virus protease. Eugenol, the principle component of clove has been shown to oxygen radical scavenging activity and antitumor potentials targeting COX-2, cMyc, H-ras [261]

157. *Syzygium cumini*: The bark of the plant is astringent to the bowels, sweet, refrigerant, carminative, diuretic, digestive, antihelminthic, febrifuge, constipating, stomachic and antibacterial. The fruits and seeds are used to treat diabetes, pharyngitis, splenopathy, urethrorrhea and ringworm infection. The leaves have been extensively used to treat diabetes, constipation, leucorrhoea, stomachalgia, fever, gastropathy, strangury, and dermatopathy and to inhibit blood discharges in the faeces. This plant has been also reported to poses acetyl oleanolic acid, triterpenoids, ellagic acid, isoquercitin, quercetin, kaempferol and myricetin in different concentrations. Recently, *S. cumini* also shown that the protective effect against radiation-induced DNA damage in human peripheral blood lymphocytes [262].

158. *Tamarix gallica*: This plant is commonly known as “Jhau”, a shrub of the family Tamaricaceae. The fruits and leaves have been employed in traditional medicine as an astringent for dysentery and chronic diarrhea, aperitif, stimulant of perspiration, diuretic, active against leucoderma, spleen trouble and eye diseases. Indeed, methanolic extract of this halophyte was investigated for its anti-carcinogenic and chemopreventive activities by evaluating the levels of hepatic antioxidant defense [263].

159. *Terminalia arjuna*: Arjuna bark (*Terminalia arjuna*) is a medicinal plant of the genus Terminalia, widely used in functional heart problems including angina, hypertension and deposits in arteries. It has been also useful in treatment of angina pectoris, heart failure, coronary artery disease and hypercholesterolemia. *In vivo* study showed its anti-inflammatory, immunomodulatory and antinociceptive activity in mice and rats [264]. In clinical study, terminalia had shown decreases platelet activation which ultimately leads in antithrombotic properties [265].

160. *Terminalia belerica*: *Terminalia belerica* has been used for its lowering serum glucose level and antioxidant activity by reducing lipid peroxidation, scavenge hydroxyl radical and superoxide radicals. This plant has been used for treatment of digestive and liver disorders. It can significantly reduce the total cholesterol, low density lipoprotein (LDL), very low density lipoprotein (VLDL), high density lipoprotein (HDL) and free fatty acid in experimentally induced hypercholesteremic rats [180, 266].

161. *Terminalia chebula* Retz: *Terminalia chebula* Retz. (Combretaceae) have been known from ancient times and were described by Charaka in his text “Charaka Samhita”. It contains the active ingredient chebulagic acid. Aqueous extract of Terminalia chebula had shown numbers of activity such as digestive, allergic and infectious diseases like cough and skin disorders, antimicrobial activity anti-diabetic and antioxidant properties [267]. It also

reported to have strong anti-anaphylactic actions, anti-inflammatory and analgesic properties [268].

162. *Thymus vulgaris*: *Thymus vulgaris* (Lamiaceae) is aromatic plants, which is distributed in subtropical countries. It contains acetophenone glycosides, phenols, thymol (40%) and carvacrol (15%). It is mainly used in smooth muscle relaxing effect, asthma, bronchitis and other respiratory diseases [269]. Thymol significantly reduced the level of DNA damage induced in K562 cells by the strong oxidant H₂O₂ [270].

163. *Tinospora cordifolia*: *Tinospora cordifolia* mainly contain different type of aporphine alkaloids and clerodane diterpenes. It is also used as an adjuvant for the prevention and/or management of insulin resistance and disorders related to it. Epoxy clerodane diterpene which obtained from it, also used against diethylnitrosamine-induced hepatocellular carcinoma [271]. Octacosanol isolated from *Tinospora cordifolia* downregulates VEGF gene expression by inhibiting matrix metalloproteinases and nuclear translocation of NF- κ B and its DNA binding activity [272]. This plant is also used as antidiabetic in streptozotocin-induced diabetes mellitus mouse model [273].

164. *Trachyspermum ammi*: *T. ammi* is used for relieving kidney stone pains and urolithiasis. So far, its diuretic properties have been reported widely in literature and it is actively used in various drug formulations of kidney stone treatments. *Trachyspermum ammi* shown prevention in DMBA- and B(a)-P induced skin and forestomach papillomagenesis [274].

165. *Tribulus terrestris*: *Tribulus terrestris* had shown chemopreventive effect against 7,12- dimethylbenz (a) anthracene induced skin papillomagenesis in mice by oral gavage for 7 days [275]. It also showed protective effects in diabetes mellitus [276]. *In vivo* study suggested that methanolic and aqueous extracts of *T. terrestris* having antihypertensive and vasodilator effects [277].

166. *Trigonella foenum-graecum*: Diosgenin, a steroidal saponin present in fenugreek (*Trigonella foenum graecum*) and other plants, has been shown to suppress inflammation, inhibit proliferation, and induce apoptosis in a variety of tumor cells. It down-regulate TNF-induced expression of NF- κ B-regulated gene products involved in cell proliferation (cyclin D1, COX-2, c-myc), antiapoptosis (IAP1, Bcl-2, Bcl-xL, Bfl-1/A1, TRAF1 and cFLIP), and invasion (MMP-9) [278]. It also inhibits STAT3 signaling pathway leading to suppression of proliferation and chemosensitization of human hepatocellular carcinoma cells [279]. 4-hydroxyisoleucine, an unusual amino acid isolated from *Trigonella foenum-graecum* seeds was characterized in type II diabetes [280].

167. *Uraria lagopoides*: It exhibits as antiviral compound and completely inhibits the growth of Reo virus [281]. Alcohol and aqueous extract of aerial parts of *Uraria lagopoides* showed anti-inflammatory activity in the rat paw edema test.

168. *Valeriana officinalis*: *Valeriana officinalis* (Valerianaceae) has been used for treating mild nervous tension and temporary sleeping problems. In traditional European medicine it has been also reported as an antiinflammatory remedy. Ethanolic extract of the underground parts of *V. officinalis* showed inhibitory activity against NF- κ B on HeLa cells [282].

169. *Valeriana wallichii*: *V. wallichii* shown antidepressant effect, and used in gastrointestinal and cardiovascular disorders, as antispasmodic and hypotensive. This action is through K(ATP) channel activation [283]. It is also used in treating anxiety, tremors, inflammations of the joints, chorea, neurosis, and dysmenorrheal in Indian ayurveda.

170. *Vanda roxburghii*: Extract of *V. roxburghii* has wound-healing potential in rats. It augments the uterine contractions and is a bronchodilator, digestant and blood purifier. It is used in diseases like gout, rheumatic disorders, asthma, abdominal pain, fever and edema [284].

171. *Vernonia cinerea*: *V. cinerea* exhibits anti-cancer and anti-helminthic, anti-diuretic, anti-inflammatory, analgesic, anti-pyretic and anti-bacterial activities. Treatment of *V. cinerea* methanolic extract also showed an anti-inflammatory effect though enhancing phagocytic activity of peritoneal macrophages. Moreover the extract downregulated inflammatory mediators such as iNOS and COX-2, and decreased secretion of TNF α , IL-1 β and IL-6 in LPS-treated macrophages [285].

172. *Viola odorata*: The leaf of this plant is used for treatment of tumor. Cyclotide from *V. odorata* showed cytotoxic effects against various drug-resistant tumor cells [286].

173. *Vitex negundo*: This plant showed anti-asthma, expectorant, antiseptis, hypoxia tolerance enhancement and also shown to could decrease blood glucose level. Vitexins have cytotoxic effect on various types of cancer cell lines and also have antitumor activity on tumor xenograft models including breast, prostate, liver, and cervical cancers [287].

174. *Withania somnifera*: The plant *Withania somnifera* Dunal (Ashwagandha), also known as Indian ginseng, is widely used in the Ayurvedic system of medicine to treat tumors, inflammation, arthritis, asthma, and hypertension. Chemical investigation of the roots and leaves of this plant has yielded bioactive withanolides. Withanolides suppressed NF- κ B activation induced by a variety of inflammatory and carcinogenic agents; including TNF- α , IL-1 β , doxorubicin, and cigarette smoke condensate. It also suppressed both inducible and constitutive NF- κ B activation. The suppression occurred through the inhibition of inhibitory subunit of I κ B α kinase activation, I κ B α phosphorylation, I κ B α degradation, p65 phosphorylation, and subsequent p65 nuclear translocation. Consequently, withanolide suppressed the expression of TNF-induced NF- κ B-regulated gene products such as IAP-1, Bfl-1/A1, and FADD-like IL-1 β -converting enzyme-inhibitory protein, COX-2 and ICAM-1, enhanced the apoptosis induced by TNF and chemotherapeutic agents, and suppressed cellular TNF-induced invasion and receptor activator of NF- κ B ligand-induced osteoclastogenesis [288]. Withanolide sulfoxide is another active compound of this plant inhibits COX-2 expression [289]. Withaferin-A (WA) is a bioactive compound derived from *W. somnifera*, which showed ant-tumor activity through inhibition of Notch-1 signaling and down regulates prosurvival pathways, such as Akt/NF- κ B/Bcl-2 [290].

175. *Zingiber officinale*: Traditionally, ginger has been used to treat a wide range of ailments including gastrointestinal disorders, such as stomachaches, abdominal spasm, nausea, and vomiting, as well as in arthritis and motion sickness. Phytochemical studies showed that the plant is rich in a large number of substances, including gingerols and shogaols. These compounds display diverse biological activities such as antioxidant, anti-inflammatory, and anticarcinogenic properties. They also exhibit a spasmolytic activity, which is mediated via blocking Ca²⁺ channels. A number of recent studies have renewed interest in ginger for the treatment of chronic inflammatory conditions [291].

Conclusions

Overall from this description, it is clear that reverse pharmacology approach to examine the plants for drug development is a viable approach. To fully validate this approach, further clinical trials are needed to examine their potential. It is anticipated that this approach will not be as expensive as currently used and the compounds/drugs isolated will be safe. Also

one must question why using a single chemical compound is preferred as a drug as compare to extracts from the whole plants. Benefits of a single chemical entity may be in convenience to understand its molecular mechanism. However, it may not be beneficial to the patient when examined, in part, due to the possibility of development of resistance to a single chemical entity. It is possible that when whole plant extract or combination of plant extracts are used, it may exhibit improved bioavailability and lower toxicity, as compared to single chemical entity.

References

1. Valiathan, MS. Legacy of Charaka. Chennai, India: Orient Longman; 2003.
2. Loo G. Redox-sensitive mechanisms of phytochemical-mediated inhibition of cancer cell proliferation (review). *J Nutr Biochem*. 2003; 14:64–73. [PubMed: 12667597]
3. Surh YJ. Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer*. 2003; 3:768–80. [PubMed: 14570043]
4. Stevenson DE, Hurst RD. Polyphenolic phytochemicals--just antioxidants or much more? *Cell Mol Life Sci*. 2007; 64:2900–16. [PubMed: 17726576]
5. Garodia P, Ichikawa H, Malani N, Sethi G, Aggarwal BB. From ancient medicine to modern medicine: ayurvedic concepts of health and their role in inflammation and cancer. *J Soc Integr Oncol*. 2007; 5:25–37. [PubMed: 17309811]
6. Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB. Bioavailability of curcumin: problems and promises. *Mol Pharm*. 2007; 4:807–18. [PubMed: 17999464]
7. Vaidya A. Reverse pharmacological correlates of ayurvedic drug actions. *Indian J Pharmacol*. 2006; 38:311–5.
8. Aggarwal BB. Nuclear factor-kappaB: the enemy within. *Cancer Cell*. 2004; 6:203–8. [PubMed: 15380510]
9. Ahn KS, Sethi G, Aggarwal BB. Nuclear factor-kappa B: from clone to clinic. *Curr Mol Med*. 2007; 7:619–37. [PubMed: 18045141]
10. Sethi G, Ahn KS, Aggarwal BB. Targeting nuclear factor-kappa B activation pathway by thymoquinone: role in suppression of antiapoptotic gene products and enhancement of apoptosis. *Mol Cancer Res*. 2008; 6:1059–70. [PubMed: 18567808]
11. Singh RK, Pandey BL, Tripathi M, Pandey VB. Anti-inflammatory effect of (+)-pinitol. *Fitoterapia*. 2001; 72:168–70. [PubMed: 11223227]
12. Sethi G, Ahn KS, Sung B, Aggarwal BB. Pinitol targets nuclear factor-kappaB activation pathway leading to inhibition of gene products associated with proliferation, apoptosis, invasion, and angiogenesis. *Mol Cancer Ther*. 2008; 7:1604–14. [PubMed: 18566231]
13. Wang JP, Hsu MF, Chang LC, Kuo JS, Kuo SC. Inhibition of plasma extravasation by abruquinone A, a natural isoflavanquinone isolated from *Abrus precatorius*. *Eur J Pharmacol*. 1995; 273:73–81. [PubMed: 7537681]
14. Krisanapun C, Peungvicha P, Tamsiririrukkul R, Wongkrajang Y. Aqueous extract of *Abutilon indicum* Sweet inhibits glucose absorption and stimulates insulin secretion in rodents. *Nutr Res*. 2009; 29:579–87. [PubMed: 19761892]
15. Wadood A, Wadood N, Shah SA. Effects of *Acacia arabica* and *Caralluma edulis* on blood glucose levels of normal and alloxan diabetic rabbits. *J Pak Med Assoc*. 1989; 39:208–12. [PubMed: 2509753]
16. Altavilla D, Squadrito F, Bitto A, Polito F, Burnett BP, Di Stefano V, et al. Flavocoxid, a dual inhibitor of cyclooxygenase and 5-lipoxygenase, blunts pro-inflammatory phenotype activation in endotoxin-stimulated macrophages. *Br J Pharmacol*. 2009; 157:1410–8. [PubMed: 19681869]
17. Lin AS, Lin CR, Du YC, Lubken T, Chiang MY, Chen IH, et al. Acasiane A and B and farnesirane A and B, diterpene derivatives from the roots of *Acacia farnesiana*. *Planta Med*. 2009; 75:256–61. [PubMed: 19101886]
18. Tozjo T, Yoshimura Y, Sakurai K, Uchida N, Takeda Y, Nakai H, et al. Novel antitumor sesquiterpenoids in *Achillea millefolium*. *Chem Pharm Bull (Tokyo)*. 1994; 42:1096–100. [PubMed: 8069962]

19. Gokhale AB, Damre AS, Kulkarni KR, Saraf MN. Preliminary evaluation of anti-inflammatory and anti-arthritic activity of *S. lappa*, *A. speciosa* and *A. aspera*. *Phytomedicine*. 2002; 9:433–7. [PubMed: 12222664]
20. Chakraborty A, Brantner A, Mukainaka T, Nobukuni Y, Kuchide M, Konoshima T, et al. Cancer chemopreventive activity of *Achyranthes aspera* leaves on Epstein-Barr virus activation and two-stage mouse skin carcinogenesis. *Cancer Lett*. 2002; 177:1–5. [PubMed: 11809524]
21. Wu HS, Zhu DF, Zhou CX, Feng CR, Lou YJ, Yang B, et al. Insulin sensitizing activity of ethyl acetate fraction of *Acorus calamus* L. in vitro and in vivo. *J Ethnopharmacol*. 2009; 123:288–92. [PubMed: 19429374]
22. Kim H, Han TH, Lee SG. Anti-inflammatory activity of a water extract of *Acorus calamus* L. leaves on keratinocyte HaCaT cells. *J Ethnopharmacol*. 2009; 122:149–56. [PubMed: 19146941]
23. Hazra R, Ray K, Guha D. Inhibitory role of *Acorus calamus* in ferric chloride-induced epileptogenesis in rat. *Hum Exp Toxicol*. 2007; 26:947–53. [PubMed: 18375638]
24. Gibbs BF. Differential modulation of IgE-dependent activation of human basophils by ambroxol and related secretolytic analogues. *Int J Immunopathol Pharmacol*. 2009; 22:919–27. [PubMed: 20074455]
25. Nicolis E, Lampronti I, Dececchi MC, Borgatti M, Tamanini A, Bezzerri V, et al. Modulation of expression of IL-8 gene in bronchial epithelial cells by 5-methoxypsoralen. *Int Immunopharmacol*. 2009; 9:1411–22. [PubMed: 19720161]
26. Panda S, Kar A. Periplogenin-3-O- -D-glucopyranosyl -(1-->6)- -D-glucopyranosyl- -(1-->4)-D-cymaropyranoside, isolated from *Aegle marmelos* protects doxorubicin induced cardiovascular problems and hepatotoxicity in rats. *Cardiovasc Ther*. 2009; 27:108–16. [PubMed: 19426248]
27. Subramaniam D, Giridharan P, Murmu N, Shankaranarayanan NP, May R, Houchen CW, et al. Activation of apoptosis by 1-hydroxy-5,7-dimethoxy-2-naphthalene-carboxaldehyde, a novel compound from *Aegle marmelos*. *Cancer Res*. 2008; 68:8573–81. [PubMed: 18922933]
28. Hodge G, Hodge S, Han P. *Allium sativum* (garlic) suppresses leukocyte inflammatory cytokine production in vitro: potential therapeutic use in the treatment of inflammatory bowel disease. *Cytometry*. 2002; 48:209–15. [PubMed: 12210145]
29. Zare A, Farzaneh P, Pourpak Z, Zahedi F, Moin M, Shahabi S, et al. Purified aged garlic extract modulates allergic airway inflammation in BALB/c mice. *Iran J Allergy Asthma Immunol*. 2008; 7:133–41. [PubMed: 18780948]
30. Hui C, Like W, Yan F, Tian X, Qiuyan W, Lifeng H. S-allyl-L-cysteine sulfoxide inhibits tumor necrosis factor- α induced monocyte adhesion and intercellular cell adhesion molecule-1 expression in human umbilical vein endothelial cells. *Anat Rec (Hoboken)*. 293:421–30. [PubMed: 20091890]
31. Ban JO, Oh JH, Kim TM, Kim DJ, Jeong HS, Han SB, et al. Anti-inflammatory and arthritic effects of thiacremonone, a novel sulfur compound isolated from garlic via inhibition of NF- κ B. *Arthritis Res Ther*. 2009; 11:R145. [PubMed: 19788760]
32. Keophiphath M, Priem F, Jacquemond-Collet I, Clement K, Lacasa D. 1,2-vinyldithiin from garlic inhibits differentiation and inflammation of human preadipocytes. *J Nutr*. 2009; 139:2055–60. [PubMed: 19759245]
33. Chithra P, Sajithlal GB, Chandrakasan G. Influence of *Aloe vera* on the glycosaminoglycans in the matrix of healing dermal wounds in rats. *J Ethnopharmacol*. 1998; 59:179–86. [PubMed: 9507902]
34. Kumar A, Dhawan S, Aggarwal BB. Emodin (3-methyl-1,6,8-trihydroxyanthraquinone) inhibits TNF-induced NF- κ B activation, IkappaB degradation, and expression of cell surface adhesion proteins in human vascular endothelial cells. *Oncogene*. 1998; 17:913–8. [PubMed: 9780008]
35. Grzanna R, Phan P, Polotsky A, Lindmark L, Frondoza CG. Ginger extract inhibits beta-amyloid peptide-induced cytokine and chemokine expression in cultured THP-1 monocytes. *J Altern Complement Med*. 2004; 10:1009–13. [PubMed: 15673995]
36. Ichikawa H, Takada Y, Murakami A, Aggarwal BB. Identification of a novel blocker of I kappa B alpha kinase that enhances cellular apoptosis and inhibits cellular invasion through suppression of NF-kappa B-regulated gene products. *J Immunol*. 2005; 174:7383–92. [PubMed: 15905586]

37. Ichikawa H, Murakami A, Aggarwal BB. 1'-Acetoxychavicol acetate inhibits RANKL-induced osteoclastic differentiation of RAW 264. 7 monocytic cells by suppressing nuclear factor-kappaB activation. *Mol Cancer Res.* 2006; 4:275–81. [PubMed: 16603641]
38. Bendjeddou D, Lalaoui K, Satta D. Immunostimulating activity of the hot water-soluble polysaccharide extracts of *Anacyclus pyrethrum*, *Alpinia galanga* and *Citrullus colocynthis*. *J Ethnopharmacol.* 2003; 88:155–60. [PubMed: 12963136]
39. Chao WW, Kuo YH, Lin BF. Anti-inflammatory activity of new compounds from *Andrographis paniculata* by NF-kappaB transactivation inhibition. *J Agric Food Chem.* 58:2505–12. [PubMed: 20085279]
40. Wibudi A, Kiranadi B, Manalu W, winarto A, Suyono S. The traditional plant, *Andrographis paniculata* (Sambiloto), exhibits insulin-releasing actions in vitro. *Acta Med Indones.* 2008; 40:63–8. [PubMed: 18560025]
41. Inokuchi J, Okabe H, Yamauchi T, Nagamatsu A, Nonaka G, Nishioka I. Antihypertensive substance in seeds of *Areca catechu* L. *Life Sci.* 1986; 38:1375–82. [PubMed: 3007909]
42. Mathews JN, Flatt PR, Abdel-Wahab YH. *Asparagus adscendens* (Shweta musali) stimulates insulin secretion, insulin action and inhibits starch digestion. *Br J Nutr.* 2006; 95:576–81. [PubMed: 16512944]
43. Visavadiya NP, Narasimhacharya AV. *Asparagus* root regulates cholesterol metabolism and improves antioxidant status in hypercholesteremic rats. *Evid Based Complement Alternat Med.* 2009; 6:219–26. [PubMed: 18955232]
44. Thoh M, Kumar P, Nagarajaram HA, Manna SK. Azadirachtin interacts with the tumor necrosis factor (TNF) binding domain of its receptors and inhibits TNF-induced biological responses. *J Biol Chem.* 285:5888–95. [PubMed: 20018848]
45. Harish Kumar G, Vidya Priyadarsini R, Vinothini G, Vidjaya Letchoumy P, Nagini S. The neem limonoids azadirachtin and nimbolide inhibit cell proliferation and induce apoptosis in an animal model of oral oncogenesis. *Invest New Drugs.* 2009
46. Mathew J, Paul J, Nandhu MS, Paulose CS. Increased excitability and metabolism in pilocarpine induced epileptic rats: Effect of *Bacopa monnieri*. *Fitoterapia.*
47. Sharath R, Harish BG, Krishna V, Sathyanarayana BN, Swamy HM. Wound healing and protease inhibition activity of Bacoside-A, isolated from *Bacopa monnieri* wettest. *Phytother Res.*
48. Muniappan M, Sundararaj T. Antiinflammatory and antiulcer activities of *Bambusa arundinacea*. *J Ethnopharmacol.* 2003; 88:161–7. [PubMed: 12963137]
49. Frankish N, de Sousa Menezes F, Mills C, Sheridan H. Enhancement of Insulin Release from the beta-Cell Line INS-1 by an Ethanolic Extract of *Bauhinia variegata* and Its Major Constituent Roseoside. *Planta Med.*
50. Rajkapoor B, Jayakar B, Muruges N, Sakthisekaran D. Chemoprevention and cytotoxic effect of *Bauhinia variegata* against N-nitrosodiethylamine induced liver tumors and human cancer cell lines. *J Ethnopharmacol.* 2006; 104:407–9. [PubMed: 16257158]
51. Mohan M, Sekhar GC, Mahajan VM. Berberine: an indigenous drug in experimental herpetic uveitis. *Indian J Ophthalmol.* 1983; 31:65–8. [PubMed: 6662570]
52. Pandey MK, Sung B, Kunnumakkara AB, Sethi G, Chaturvedi MM, Aggarwal BB. Berberine modifies cysteine 179 of IkappaBalpha kinase, suppresses nuclear factor-kappaB-regulated antiapoptotic gene products, and potentiates apoptosis. *Cancer Res.* 2008; 68:5370–9. [PubMed: 18593939]
53. Kirtikar, KR; Basu, BD. *Boerhaavia diffusa*. Indian Medicinal Plants. Allahabad, India: Lalit Mohan Basu Publications; 1956.
54. Leslie Taylor, ND. *Boerhaavia diffusa*. The Healing Power of Rainforest Herbs. New York, USA: Square one Publishers; 2005.
55. Manu KA, Kuttan G. Effect of Punarnavine, an alkaloid from *Boerhaavia diffusa*, on cell-mediated immune responses and TIMP-1 in B16F-10 metastatic melanoma-bearing mice. *Immunopharmacol Immunotoxicol.* 2007; 29:569–86. [PubMed: 18075866]
56. Gupta I, Parihar A, Malhotra P, Singh GB, Ludtke R, Safayhi H, et al. Effects of *Boswellia serrata* gum resin in patients with ulcerative colitis. *Eur J Med Res.* 1997; 2:37–43. [PubMed: 9049593]

57. Ammon HP. Salai Guggal - *Boswellia serrata*: from a herbal medicine to a non-redox inhibitor of leukotriene biosynthesis. *Eur J Med Res.* 1996; 1:369–70. [PubMed: 9360935]
58. Safayhi H, Sailer ER, Ammon HP. Mechanism of 5-lipoxygenase inhibition by acetyl-11-keto-beta-boswellic acid. *Mol Pharmacol.* 1995; 47:1212–6. [PubMed: 7603462]
59. Sailer ER, Subramanian LR, Rall B, Hoernlein RF, Ammon HP, Safayhi H. Acetyl-11-keto-beta-boswellic acid (AKBA): structure requirements for binding and 5-lipoxygenase inhibitory activity. *Br J Pharmacol.* 1996; 117:615–8. [PubMed: 8646405]
60. Sailer ER, Schweizer S, Boden SE, Ammon HP, Safayhi H. Characterization of an acetyl-11-keto-beta-boswellic acid and arachidonate-binding regulatory site of 5-lipoxygenase using photoaffinity labeling. *Eur J Biochem.* 1998; 256:364–8. [PubMed: 9760176]
61. Safayhi H, Rall B, Sailer ER, Ammon HP. Inhibition by boswellic acids of human leukocyte elastase. *J Pharmacol Exp Ther.* 1997; 281:460–3. [PubMed: 9103531]
62. Syrovets T, Buchele B, Gedig E, Slupsky JR, Simmet T. Acetyl-boswellic acids are novel catalytic inhibitors of human topoisomerases I and II α . *Mol Pharmacol.* 2000; 58:71–81. [PubMed: 10860928]
63. Glaser T, Winter S, Groscurth P, Safayhi H, Sailer ER, Ammon HP, et al. Boswellic acids and malignant glioma: induction of apoptosis but no modulation of drug sensitivity. *Br J Cancer.* 1999; 80:756–65. [PubMed: 10360653]
64. Liu JJ, Nilsson A, Oredsson S, Badmaev V, Zhao WZ, Duan RD. Boswellic acids trigger apoptosis via a pathway dependent on caspase-8 activation but independent on Fas/Fas ligand interaction in colon cancer HT-29 cells. *Carcinogenesis.* 2002; 23:2087–93. [PubMed: 12507932]
65. Liu JJ, Huang B, Hooi SC. Acetyl-keto-beta-boswellic acid inhibits cellular proliferation through a p21-dependent pathway in colon cancer cells. *Br J Pharmacol.* 2006; 148:1099–107. [PubMed: 16783403]
66. Huang MT, Badmaev V, Ding Y, Liu Y, Xie JG, Ho CT. Anti-tumor and anti-carcinogenic activities of triterpenoid, beta-boswellic acid. *Biofactors.* 2000; 13:225–30. [PubMed: 11237186]
67. Shao Y, Ho CT, Chin CK, Badmaev V, Ma W, Huang MT. Inhibitory activity of boswellic acids from *Boswellia serrata* against human leukemia HL-60 cells in culture. *Planta Med.* 1998; 64:328–31. [PubMed: 9619114]
68. Xia L, Chen D, Han R, Fang Q, Waxman S, Jing Y. Boswellic acid acetate induces apoptosis through caspase-mediated pathways in myeloid leukemia cells. *Mol Cancer Ther.* 2005; 4:381–8. [PubMed: 15767547]
69. Hoernlein RF, Orlikowsky T, Zehrer C, Niethammer D, Sailer ER, Simmet T, et al. Acetyl-11-keto-beta-boswellic acid induces apoptosis in HL-60 and CCRF-CEM cells and inhibits topoisomerase I. *J Pharmacol Exp Ther.* 1999; 288:613–9. [PubMed: 9918566]
70. Bhushan S, Kumar A, Malik F, Andotra SS, Sethi VK, Kaur IP, et al. A triterpenediol from *Boswellia serrata* induces apoptosis through both the intrinsic and extrinsic apoptotic pathways in human leukemia HL-60 cells. *Apoptosis.* 2007; 12:1911–26. [PubMed: 17636381]
71. Zhao W, Entschladen F, Liu H, Niggemann B, Fang Q, Zaenker KS, et al. Boswellic acid acetate induces differentiation and apoptosis in highly metastatic melanoma and fibrosarcoma cells. *Cancer Detect Prev.* 2003; 27:67–75. [PubMed: 12600419]
72. Liu JJ, Nilsson A, Oredsson S, Badmaev V, Duan RD. Keto- and acetyl-keto-boswellic acids inhibit proliferation and induce apoptosis in Hep G2 cells via a caspase-8 dependent pathway. *Int J Mol Med.* 2002; 10:501–5. [PubMed: 12239601]
73. Syrovets T, Gschwend JE, Buchele B, Laumonnier Y, Zugmaier W, Genze F, et al. Inhibition of I κ B kinase activity by acetyl-boswellic acids promotes apoptosis in androgen-independent PC-3 prostate cancer cells in vitro and in vivo. *J Biol Chem.* 2005; 280:6170–80. [PubMed: 15576374]
74. Lu M, Xia L, Hua H, Jing Y. Acetyl-keto-beta-boswellic acid induces apoptosis through a death receptor 5-mediated pathway in prostate cancer cells. *Cancer Res.* 2008; 68:1180–6. [PubMed: 18281494]
75. Yuan HQ, Kong F, Wang XL, Young CY, Hu XY, Lou HX. Inhibitory effect of acetyl-11-keto-beta-boswellic acid on androgen receptor by interference of Sp1 binding activity in prostate cancer cells. *Biochem Pharmacol.* 2008; 75:2112–21. [PubMed: 18430409]

76. Syrovets T, Buchele B, Krauss C, Laumonnier Y, Simmet T. Acetyl-boswellic acids inhibit lipopolysaccharide-mediated TNF-alpha induction in monocytes by direct interaction with IkappaB kinases. *J Immunol.* 2005; 174:498–506. [PubMed: 15611276]
77. Takada Y, Ichikawa H, Badmaev V, Aggarwal BB. Acetyl-11-keto-beta-boswellic acid potentiates apoptosis, inhibits invasion, and abolishes osteoclastogenesis by suppressing NF-kappa B and NF-kappa B-regulated gene expression. *J Immunol.* 2006; 176:3127–40. [PubMed: 16493072]
78. Kunnumakkara AB, Nair AS, Sung B, Pandey MK, Aggarwal BB. Boswellic acid blocks signal transducers and activators of transcription 3 signaling, proliferation, and survival of multiple myeloma via the protein tyrosine phosphatase SHP-1. *Mol Cancer Res.* 2009; 7:118–28. [PubMed: 19147543]
79. Roy S, Khanna S, Shah H, Rink C, Phillips C, Preuss H, et al. Human genome screen to identify the genetic basis of the anti-inflammatory effects of Boswellia in microvascular endothelial cells. *DNA Cell Biol.* 2005; 24:244–55. [PubMed: 15812241]
80. Roy S, Khanna S, Krishnaraju AV, Subbaraju GV, Yasmin T, Bagchi D, et al. Regulation of vascular responses to inflammation: inducible matrix metalloproteinase-3 expression in human microvascular endothelial cells is sensitive to antiinflammatory Boswellia. *Antioxid Redox Signal.* 2006; 8:653–60. [PubMed: 16677108]
81. Wildfeuer A, Neu IS, Safayhi H, Metzger G, Wehrmann M, Vogel U, et al. Effects of boswellic acids extracted from a herbal medicine on the biosynthesis of leukotrienes and the course of experimental autoimmune encephalomyelitis. *Arzneimittelforschung.* 1998; 48:668–74. [PubMed: 9689425]
82. Kriegelstein CF, Anthoni C, Rijcken EJ, Laukötter M, Spiegel HU, Boden SE, et al. Acetyl-11-keto-beta-boswellic acid, a constituent of a herbal medicine from *Boswellia serrata* resin, attenuates experimental ileitis. *Int J Colorectal Dis.* 2001; 16:88–95. [PubMed: 11355324]
83. Anthoni C, Laukötter MG, Rijcken E, Vowinkel T, Mennigen R, Müller S, et al. Mechanisms underlying the anti-inflammatory actions of boswellic acid derivatives in experimental colitis. *Am J Physiol Gastrointest Liver Physiol.* 2006; 290:G1131–7. [PubMed: 16423918]
84. Singh SK, Bhusari S, Singh R, Saxena A, Mondhe D, Qazi GN. Effect of acetyl 11-keto beta-boswellic acid on metastatic growth factor responsible for angiogenesis. *Vascul Pharmacol.* 2007; 46:333–7. [PubMed: 17257903]
85. Winking M, Sarikaya S, Rahmanian A, Jodicke A, Boker DK. Boswellic acids inhibit glioma growth: a new treatment option? *J Neurooncol.* 2000; 46:97–103. [PubMed: 10894362]
86. Singh GB, Atal CK. Pharmacology of an extract of salai guggal ex-*Boswellia serrata*, a new non-steroidal anti-inflammatory agent. *Agents Actions.* 1986; 18:407–12. [PubMed: 3751752]
87. Gupta M, Mazumdar UK, Sivakumar T, Vamsi ML, Karki SS, Sambathkumar R, et al. Evaluation of anti-inflammatory activity of chloroform extract of *Bryonia laciniosa* in experimental animal models. *Biol Pharm Bull.* 2003; 26:1342–4. [PubMed: 12951483]
88. Wagner H, Geyer B, Fiebig M, Kiso Y, Hikino H. Isobutrin and butrin, the antihepatotoxic principles of *Butea monosperma* flowers. *Planta Med.* 1986:77–9.
89. Srinivasan R, Chandrasekar MJ, Nanjan MJ, Suresh B. Antioxidant activity of *Caesalpinia digyna* root. *J Ethnopharmacol.* 2007; 113:284–91. [PubMed: 17686593]
90. Takada Y, Aggarwal BB. Betulinic acid suppresses carcinogen-induced NF-kappa B activation through inhibition of I kappa B alpha kinase and p65 phosphorylation: abrogation of cyclooxygenase-2 and matrix metalloproteinase-9. *J Immunol.* 2003; 171:3278–86. [PubMed: 12960358]
91. Pandey MK, Sung B, Aggarwal BB. Betulinic acid suppresses STAT3 activation pathway through induction of protein tyrosine phosphatase SHP-1 in human multiple myeloma cells. *Int J Cancer.* 2009
92. Arya S, Kumar VL. Antiinflammatory efficacy of extracts of latex of *Calotropis procera* against different mediators of inflammation. *Mediators Inflamm.* 2005; 2005:228–32. [PubMed: 16192673]
93. Cao YL, Li X, Zheng M. *Capparis spinosa* protects against oxidative stress in systemic sclerosis dermal fibroblasts. *Arch Dermatol Res.* 2009

94. Zahin M, Ahmad I, Aqil F. Antioxidant and antimutagenic activity of *Carum copticum* fruit extracts. *Toxicol In Vitro*.
95. Prakasam A, Sethupathy S, Pugalendi KV. Influence of *Casearia esculenta* root extract on protein metabolism and marker enzymes in streptozotocin-induced diabetic rats. *Pol J Pharmacol*. 2004; 56:587–93. [PubMed: 15591647]
96. Cuellar MJ, Giner RM, Recio MC, Manez S, Rios JL. Topical anti-inflammatory activity of some Asian medicinal plants used in dermatological disorders. *Fitoterapia*. 2001; 72:221–9. [PubMed: 11295297]
97. Bhakta T, Banerjee S, Mandal SC, Maity TK, Saha BP, Pal M. Hepatoprotective activity of *Cassia fistula* leaf extract. *Phytomedicine*. 2001; 8:220–4. [PubMed: 11417916]
98. Sadique J, Chandra T, Thenmozhi V, Elango V. Biochemical modes of action of *Cassia occidentalis* and *Cardiospermum halicacabum* in inflammation. *J Ethnopharmacol*. 1987; 19:201–12. [PubMed: 3613609]
99. Jang DS, Lee GY, Kim YS, Lee YM, Kim CS, Yoo JL, et al. Anthraquinones from the seeds of *Cassia tora* with inhibitory activity on protein glycation and aldose reductase. *Biol Pharm Bull*. 2007; 30:2207–10. [PubMed: 17978503]
100. Shinde UA, Phadke AS, Nair AM, Mungantiwar AA, Dikshit VJ, Saraf MN. Studies on the anti-inflammatory and analgesic activity of *Cedrus deodara* (Roxb.) Loud. wood oil. *J Ethnopharmacol*. 1999; 65:21–7. [PubMed: 10350366]
101. Ahmad F, Khan RA, Rasheed S. Preliminary screening of methanolic extracts of *Celastrus paniculatus* and *Tecomella undulata* for analgesic and anti-inflammatory activities. *J Ethnopharmacol*. 1994; 42:193–8. [PubMed: 7934089]
102. Hazra B, Sarkar R, Bhattacharyya S, Roy P. Tumour inhibitory activity of chicory root extract against Ehrlich ascites carcinoma in mice. *Fitoterapia*. 2002; 73:730–3. [PubMed: 12490244]
103. Cavin C, Delannoy M, Malnoe A, Debeve E, Touche A, Courtois D, et al. Inhibition of the expression and activity of cyclooxygenase-2 by chicory extract. *Biochem Biophys Res Commun*. 2005; 327:742–9. [PubMed: 15649409]
104. Lee HJ, Hyun EA, Yoon WJ, Kim BH, Rhee MH, Kang HK, et al. In vitro anti-inflammatory and anti-oxidative effects of *Cinnamomum camphora* extracts. *J Ethnopharmacol*. 2006; 103:208–16. [PubMed: 16182479]
105. Verspohl EJ, Bauer K, Neddermann E. Antidiabetic effect of *Cinnamomum cassia* and *Cinnamomum zeylanicum* in vivo and in vitro. *Phytother Res*. 2005; 19:203–6. [PubMed: 15934022]
106. Mishra A, Bhatti R, Singh A, Singh Ishar MP. Ameliorative effect of the cinnamon oil from *Cinnamomum zeylanicum* upon early stage diabetic nephropathy. *Planta Med*. 76:412–7. [PubMed: 19876811]
107. Marzouk B, Marzouk Z, Decor R, Edziri H, Haloui E, Fenina N, et al. Antibacterial and anticandidal screening of Tunisian *Citrullus colocynthis* Schrad. from Medenine. *J Ethnopharmacol*. 2009; 125:344–9. [PubMed: 19397972]
108. Urizar NL, Moore DD. GUGULIPID: a natural cholesterol-lowering agent. *Annu Rev Nutr*. 2003; 23:303–13. [PubMed: 12626688]
109. Sinal CJ, Gonzalez FJ. Guggulsterone: an old approach to a new problem. *Trends Endocrinol Metab*. 2002; 13:275–6. [PubMed: 12163224]
110. Urizar NL, Liverman AB, Dodds DT, Silva FV, Ordentlich P, Yan Y, et al. A natural product that lowers cholesterol as an antagonist ligand for FXR. *Science*. 2002; 296:1703–6. [PubMed: 11988537]
111. Shishodia S, Aggarwal BB. Guggulsterone inhibits NF-kappaB and IkappaBalpha kinase activation, suppresses expression of anti-apoptotic gene products, and enhances apoptosis. *J Biol Chem*. 2004; 279:47148–58. [PubMed: 15322087]
112. Ahn KS, Sethi G, Sung B, Goel A, Ralhan R, Aggarwal BB. Guggulsterone, a farnesoid X receptor antagonist, inhibits constitutive and inducible STAT3 activation through induction of a protein tyrosine phosphatase SHP-1. *Cancer Res*. 2008; 68:4406–15. [PubMed: 18519703]

113. Burris TP, Montrose C, Houck KA, Osborne HE, Bocchinfuso WP, Yaden BC, et al. The hypolipidemic natural product guggulsterone is a promiscuous steroid receptor ligand. *Mol Pharmacol.* 2005; 67:948–54. [PubMed: 15602004]
114. Gujral ML, Sareen K, Tangri KK, Amma MK, Roy AK. Antiarthritic and anti-inflammatory activity of gum guggul (*Balsamodendron mukul* Hook). *Indian J Physiol Pharmacol.* 1960; 4:267–73. [PubMed: 13709695]
115. Sharma JN. Comparison of the anti-inflammatory activity of *Commiphora mukul* (an indigenous drug) with those of phenylbutazone and ibuprofen in experimental arthritis induced by mycobacterial adjuvant. *Arzneimittelforschung.* 1977; 27:1455–7. [PubMed: 578471]
116. Singh BB, Mishra LC, Vinjamury SP, Aquilina N, Singh VJ, Shepard N. The effectiveness of *Commiphora mukul* for osteoarthritis of the knee: an outcomes study. *Altern Ther Health Med.* 2003; 9:74–9. [PubMed: 12776478]
117. Cui J, Huang L, Zhao A, Lew JL, Yu J, Sahoo S, et al. Guggulsterone is a farnesoid X receptor antagonist in coactivator association assays but acts to enhance transcription of bile salt export pump. *J Biol Chem.* 2003; 278:10214–20. [PubMed: 12525500]
118. Wu J, Xia C, Meier J, Li S, Hu X, Lala DS. The hypolipidemic natural product guggulsterone acts as an antagonist of the bile acid receptor. *Mol Endocrinol.* 2002; 16:1590–7. [PubMed: 12089353]
119. Meselhy MR. Inhibition of LPS-induced NO production by the oleogum resin of *Commiphora wightii* and its constituents. *Phytochemistry.* 2003; 62:213–8. [PubMed: 12482459]
120. Samudio I, Konopleva M, Safe S, McQueen T, Andreeff M. Guggulsterones induce apoptosis and differentiation in acute myeloid leukemia: identification of isomer-specific antileukemic activities of the pregnadienedione structure. *Mol Cancer Ther.* 2005; 4:1982–92. [PubMed: 16373713]
121. Singh SV, Zeng Y, Xiao D, Vogel VG, Nelson JB, Dhir R, et al. Caspase-dependent apoptosis induction by guggulsterone, a constituent of Ayurvedic medicinal plant *Commiphora mukul*, in PC-3 human prostate cancer cells is mediated by Bax and Bak. *Mol Cancer Ther.* 2005; 4:1747–54. [PubMed: 16275996]
122. Ichikawa H, Aggarwal BB. Guggulsterone inhibits osteoclastogenesis induced by receptor activator of nuclear factor-kappaB ligand and by tumor cells by suppressing nuclear factor-kappaB activation. *Clin Cancer Res.* 2006; 12:662–8. [PubMed: 16428513]
123. Kumar V. Potential medicinal plants for CNS disorders: an overview. *Phytother Res.* 2006; 20:1023–35. [PubMed: 16909441]
124. Kumari A, Kakkar P. Screening of antioxidant potential of selected barks of Indian medicinal plants by multiple in vitro assays. *Biomed Environ Sci.* 2008; 21:24–9. [PubMed: 18478975]
125. Hosseinzadeh H, Younesi HM. Antinociceptive and anti-inflammatory effects of *Crocus sativus* L. stigma and petal extracts in mice. *BMC Pharmacol.* 2002; 2:7. [PubMed: 11914135]
126. Nalini N, Manju V, Menon VP. Effect of spices on lipid metabolism in 1,2-dimethylhydrazine-induced rat colon carcinogenesis. *J Med Food.* 2006; 9:237–45. [PubMed: 16822210]
127. Siddaraju MN, Dharmesh SM. Inhibition of gastric H(+), K(+)-ATPase and *Helicobacter pylori* growth by phenolic antioxidants of *Curcuma amada*. *J Agric Food Chem.* 2007; 55:7377–86. [PubMed: 17725316]
128. Aggarwal S, Ichikawa H, Takada Y, Sandur SK, Shishodia S, Aggarwal BB. Curcumin (diferuloylmethane) down-regulates expression of cell proliferation and antiapoptotic and metastatic gene products through suppression of IkappaBalpha kinase and Akt activation. *Mol Pharmacol.* 2006; 69:195–206. [PubMed: 16219905]
129. Singh S, Aggarwal BB. Activation of transcription factor NF-kappa B is suppressed by curcumin (diferuloylmethane) [corrected]. *J Biol Chem.* 1995; 270:24995–5000. [PubMed: 7559628]
130. Bharti AC, Donato N, Aggarwal BB. Curcumin (diferuloylmethane) inhibits constitutive and IL-6-inducible STAT3 phosphorylation in human multiple myeloma cells. *J Immunol.* 2003; 171:3863–71. [PubMed: 14500688]
131. Aggarwal BB, Harikumar KB. Potential therapeutic effects of curcumin, the anti-inflammatory agent, against neurodegenerative, cardiovascular, pulmonary, metabolic, autoimmune and neoplastic diseases. *Int J Biochem Cell Biol.* 2009; 41:40–59. [PubMed: 18662800]

132. Aggarwal BB, Kumar A, Bharti AC. Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res.* 2003; 23:363–98. [PubMed: 12680238]
133. Aggarwal BB, Sundaram C, Malani N, Ichikawa H. Curcumin: the Indian solid gold. *Adv Exp Med Biol.* 2007; 595:1–75. [PubMed: 17569205]
134. Bright JJ. Curcumin and autoimmune disease. *Adv Exp Med Biol.* 2007; 595:425–51. [PubMed: 17569223]
135. Das T, Sa G, Saha B, Das K. Multifocal signal modulation therapy of cancer: ancient weapon, modern targets. *Mol Cell Biochem.* 336:85–95. [PubMed: 19826768]
136. Goel A, Kunnumakkara AB, Aggarwal BB. Curcumin as “Curecumin”: from kitchen to clinic. *Biochem Pharmacol.* 2008; 75:787–809. [PubMed: 17900536]
137. Hanai H, Sugimoto K. Curcumin has bright prospects for the treatment of inflammatory bowel disease. *Curr Pharm Des.* 2009; 15:2087–94. [PubMed: 19519446]
138. Hatcher H, Planalp R, Cho J, Torti FM, Torti SV. Curcumin: from ancient medicine to current clinical trials. *Cell Mol Life Sci.* 2008; 65:1631–52. [PubMed: 18324353]
139. Jurenka JS. Anti-inflammatory properties of curcumin, a major constituent of *Curcuma longa*: a review of preclinical and clinical research. *Altern Med Rev.* 2009; 14:141–53. [PubMed: 19594223]
140. Maheshwari RK, Singh AK, Gaddipati J, Srimal RC. Multiple biological activities of curcumin: a short review. *Life Sci.* 2006; 78:2081–7. [PubMed: 16413584]
141. Menon VP, Sudheer AR. Antioxidant and anti-inflammatory properties of curcumin. *Adv Exp Med Biol.* 2007; 595:105–25. [PubMed: 17569207]
142. Ravindran J, Prasad S, Aggarwal BB. Curcumin and cancer cells: how many ways can curry kill tumor cells selectively? *AAPS J.* 2009; 11:495–510. [PubMed: 19590964]
143. Srivastava G, Mehta JL. Currying the heart: curcumin and cardioprotection. *J Cardiovasc Pharmacol Ther.* 2009; 14:22–7. [PubMed: 19153099]
144. Surh YJ, Chun KS. Cancer chemopreventive effects of curcumin. *Adv Exp Med Biol.* 2007; 595:149–72. [PubMed: 17569209]
145. Lobo R, Prabhu KS, Shirwaikar A. *Curcuma zedoaria* Rosc. (white turmeric): a review of its chemical, pharmacological and ethnomedicinal properties. *J Pharm Pharmacol.* 2009; 61:13–21. [PubMed: 19126292]
146. Watanabe C, Hokari R, Komoto S, Kurihara C, Okada Y, Matsunaga H, et al. Lemon grass (*Cymbopogon citratus*) ameliorates murine spontaneous ileitis by decreasing lymphocyte recruitment to the inflamed intestine. *Microcirculation.* 17:321–32. [PubMed: 20618690]
147. Tiwari M, Dwivedi U, Kakkar P. Suppression of oxidative stress and pro-inflammatory mediators by *Cymbopogon citratus* D Stapf extract in lipopolysaccharide stimulated murine alveolar macrophages. *Food Chem Toxicol.*
148. Prasad CS, Shukla R, Kumar A, Dubey NK. In vitro and in vivo antifungal activity of essential oils of *Cymbopogon martini* and *Chenopodium ambrosioides* and their synergism against dermatophytes. *Mycoses.* 2009
149. Natarajan KS, Narasimhan M, Shanmugasundaram KR, Shanmugasundaram ER. Antioxidant activity of a salt-spice-herbal mixture against free radical induction. *J Ethnopharmacol.* 2006; 105:76–83. [PubMed: 16337350]
150. Gilani AU, Janbaz KH. Studies on protective effect of *Cyperus scariosus* extract on acetaminophen and CCl₄-induced hepatotoxicity. *Gen Pharmacol.* 1995; 26:627–31. [PubMed: 7789738]
151. Baumer U, Dietemann P. Identification and differentiation of dragon’s blood in works of art using gas chromatography/mass spectrometry. *Anal Bioanal Chem.*
152. Rajesh, Sharma GL. Studies on antimycotic properties of *Datura metel*. *J Ethnopharmacol.* 2002; 80:193–7. [PubMed: 12007710]
153. Kaur G, Lone IA, Athar M, Alam MS. Protective effect of *Didymocarpus pedicellata* on ferric nitrilotriacetate (Fe-NTA) induced renal oxidative stress and hyperproliferative response. *Chem Biol Interact.* 2007; 165:33–44. [PubMed: 17140554]

154. Kieley S, Dwivedi R, Monga M. Ayurvedic medicine and renal calculi. *J Endourol.* 2008; 22:1613–6. [PubMed: 18620498]
155. Takada Y, Aggarwal BB. Flavopiridol inhibits NF-kappaB activation induced by various carcinogens and inflammatory agents through inhibition of IkappaBalpha kinase and p65 phosphorylation: abrogation of cyclin D1, cyclooxygenase-2, and matrix metalloproteinase-9. *J Biol Chem.* 2004; 279:4750–9. [PubMed: 14630924]
156. Takada Y, Sethi G, Sung B, Aggarwal BB. Flavopiridol suppresses tumor necrosis factor-induced activation of activator protein-1, c-Jun N-terminal kinase, p38 mitogen-activated protein kinase (MAPK), p44/p42 MAPK, and Akt, inhibits expression of antiapoptotic gene products, and enhances apoptosis through cytochrome c release and caspase activation in human myeloid cells. *Mol Pharmacol.* 2008; 73:1549–57. [PubMed: 18287248]
157. Saxena AK, Singh B, Anand KK. Hepatoprotective effects of *Eclipta alba* on subcellular levels in rats. *J Ethnopharmacol.* 1993; 40:155–61. [PubMed: 8145570]
158. Jamal A, Javed K, Aslam M, Jafri MA. Gastroprotective effect of cardamom, *Elettaria cardamomum* Maton. fruits in rats. *J Ethnopharmacol.* 2006; 103:149–53. [PubMed: 16298093]
159. Majdalawieh AF, Carr RI. In vitro investigation of the potential immunomodulatory and anti-cancer activities of black pepper (*Piper nigrum*) and cardamom (*Elettaria cardamomum*). *J Med Food.* 13:371–81. [PubMed: 20210607]
160. Nikolovska-Coleska Z, Xu L, Hu Z, Tomita Y, Li P, Roller PP, et al. Discovery of embelin as a cell-permeable, small-molecular weight inhibitor of XIAP through structure-based computational screening of a traditional herbal medicine three-dimensional structure database. *J Med Chem.* 2004; 47:2430–40. [PubMed: 15115387]
161. Ahn KS, Sethi G, Aggarwal BB. Embelin, an inhibitor of X chromosome-linked inhibitor-of-apoptosis protein, blocks nuclear factor-kappaB (NF-kappaB) signaling pathway leading to suppression of NF-kappaB-regulated antiapoptotic and metastatic gene products. *Mol Pharmacol.* 2007; 71:209–19. [PubMed: 17028156]
162. Jindal A, Soyad D, Sharma A, Goyal PK. Protective effect of an extract of *Embolia officinalis* against radiation-induced damage in mice. *Integr Cancer Ther.* 2009; 8:98–105. [PubMed: 19223372]
163. Achrekar S, Kaklij GS, Pote MS, Kelkar SM. Hypoglycemic activity of *Eugenia jambolana* and *Ficus bengalensis*: mechanism of action. *In Vivo.* 1991; 5:143–7. [PubMed: 1768783]
164. Ganju L, Karan D, Chanda S, Srivastava KK, Sawhney RC, Selvamurthy W. Immunomodulatory effects of agents of plant origin. *Biomed Pharmacother.* 2003; 57:296–300. [PubMed: 14499177]
165. Rawal AK, Muddeshwar MG, Biswas SK. *Rubia cordifolia*, *Fagonia cretica* linn and *Tinospora cordifolia* exert neuroprotection by modulating the antioxidant system in rat hippocampal slices subjected to oxygen glucose deprivation. *BMC Complement Altern Med.* 2004; 4:11. [PubMed: 15310392]
166. Lee JH, Choi S, Lee Y, Lee HJ, Kim KH, Ahn KS, et al. Herbal compound farnesiferol C exerts antiangiogenic and antitumor activity and targets multiple aspects of VEGFR1 (Flt1) or VEGFR2 (Flk1) signaling cascades. *Mol Cancer Ther.* 9:389–99. [PubMed: 20103598]
167. Taur DJ, Nirmal SA, Patil RY, Kharya MD. Antistress and antiallergic effects of *Ficus bengalensis* bark in asthma. *Nat Prod Res.* 2007; 21:1266–70. [PubMed: 18075889]
168. Chaiy GB, Manna SK, Chaturvedi MM, Aggarwal BB. Anethole blocks both early and late cellular responses transduced by tumor necrosis factor: effect on NF-kappaB, AP-1, JNK, MAPKK and apoptosis. *Oncogene.* 2000; 19:2943–50. [PubMed: 10871845]
169. al-Harbi MM, Qureshi S, Raza M, Ahmed MM, Giangreco AB, Shah AH. Influence of anethole treatment on the tumour induced by Ehrlich ascites carcinoma cells in paw of Swiss albino mice. *Eur J Cancer Prev.* 1995; 4:307–18. [PubMed: 7549823]
170. Mahendran P, Vanisree AJ, Shyamala Devi CS. The antiulcer activity of *Garcinia cambogia* extract against indomethacin-induced gastric ulcer in rats. *Phytother Res.* 2002; 16:80–3. [PubMed: 11807973]
171. Balasubramanyam K, Altaf M, Varier RA, Swaminathan V, Ravindran A, Sadhale PP, et al. Polyisoprenylated benzophenone, garcinol, a natural histone acetyltransferase inhibitor, represses

chromatin transcription and alters global gene expression. *J Biol Chem*. 2004; 279:33716–26. [PubMed: 15155757]

172. Pandey MK, Sung B, Ahn KS, Kunnumakkara AB, Chaturvedi MM, Aggarwal BB. Gambogic acid, a novel ligand for transferrin receptor, potentiates TNF-induced apoptosis through modulation of the nuclear factor-kappaB signaling pathway. *Blood*. 2007; 110:3517–25. [PubMed: 17673602]
173. Yi T, Yi Z, Cho SG, Luo J, Pandey MK, Aggarwal BB, et al. Gambogic acid inhibits angiogenesis and prostate tumor growth by suppressing vascular endothelial growth factor receptor 2 signaling. *Cancer Res*. 2008; 68:1843–50. [PubMed: 18339865]
174. Zhang B, Li JB, Zhang DM, Ding Y, Du GH. Analgesic and anti-inflammatory activities of a fraction rich in gaultherin isolated from *Gaultheria yunnanensis* (FRANCH) REHDER. *Biol Pharm Bull*. 2007; 30:465–9. [PubMed: 17329839]
175. Dhingra D, Sharma A. Antidepressant-like activity of *Glycyrrhiza glabra* L. in mouse models of immobility tests *Prog Neuropsychopharmacol. Biol Psychiatry*. 2006; 30:449–54.
176. Agunu A, Yusuf S, Andrew GO, Zezi AU, Abdurahman EM. Evaluation of five medicinal plants used in diarrhoea treatment in Nigeria. *J Ethnopharmacol*. 2005; 101:27–30. [PubMed: 15908152]
177. Ojha P, Dhar JD, Dwivedi AK, Singh RL, Gupta G. Effect of antispermatogenic agents on cell marker enzymes of rat Sertoli cells in vitro. *Contraception*. 2006; 73:102–6. [PubMed: 16371305]
178. Mi JX, Wang GF, Wang HB, Sun XQ, Ni XY, Zhang XW, et al. Synergistic antitumoral activity and induction of apoptosis by novel pan Bcl-2 proteins inhibitor apogossypolone with adriamycin in human hepatocellular carcinoma. *Acta Pharmacol Sin*. 2008; 29:1467–77. [PubMed: 19026166]
179. Liu HM, Kiuchi F, Tsuda Y. Isolation and structure elucidation of gymnemic acids, antisweet principles of *Gymnema sylvestre*. *Chem Pharm Bull (Tokyo)*. 1992; 40:1366–75. [PubMed: 1327559]
180. Saravanan N, Nalini N. Effect of 2-hydroxy 4-methoxy benzoic acid on an experimental model of hyperlipidaemia, induced by chronic ethanol treatment. *J Pharm Pharmacol*. 2007; 59:1537–42. [PubMed: 17976265]
181. Sethi A, Bhatia A, Srivastava S, Bhatia G, Khan MM, Khanna AK, et al. Pregnane glycoside from *Hemidesmus indicus* as a potential anti-oxidant and anti-dyslipidemic agent. *Nat Prod Res*. :1–7.
182. Arseculeratne SN, Gunatilaka AA, Panabokke RG. Studies on medicinal plants of Sri Lanka: occurrence of pyrrolizidine alkaloids and hepatotoxic properties in some traditional medicinal herbs. *J Ethnopharmacol*. 1981; 4:159–77. [PubMed: 7311596]
183. Madhujith T, Shahidi F. Antioxidative and antiproliferative properties of selected barley (*Hordeum vulgare* L) cultivars and their potential for inhibition of low-density lipoprotein (LDL) cholesterol oxidation. *J Agric Food Chem*. 2007; 55:5018–24. [PubMed: 17542605]
184. Sethi G, Ahn KS, Sandur SK, Lin X, Chaturvedi MM, Aggarwal BB. Indirubin enhances tumor necrosis factor-induced apoptosis through modulation of nuclear factor-kappa B signaling pathway. *J Biol Chem*. 2006; 281:23425–35. [PubMed: 16785236]
185. Mangathayaru K, Kuruvilla S, Balakrishna K, Venkatesh J. Modulatory effect of *Inula racemosa* Hook. f. (Asteraceae) on experimental atherosclerosis in guinea-pigs. *J Pharm Pharmacol*. 2009; 61:1111–8. [PubMed: 19703356]
186. Ono M, Fukuda H, Murata H, Miyahara K. Resin glycosides from the leaves and stems of *Ipomoea digitata*. *J Nat Med*. 2009; 63:176–80. [PubMed: 19153807]
187. Kim KH, Choi SU, Lee KR. Diterpene glycosides from the seeds of *Pharbitis nil*. *J Nat Prod*. 2009; 72:1121–7. [PubMed: 19435339]
188. Topcu G, Ayral MN, Aydin A, Goren AC, Chai HB, Pezzuto JM. Triterpenoids of the roots of *Lavandula stoechas* ssp. *stoechas*. *Pharmazie*. 2001; 56:892–5. [PubMed: 11817178]
189. Bavarva JH, Narasimhacharya AV. *Leucas cephalotes* regulates carbohydrate and lipid metabolism and improves antioxidant status in IDDM and NIDDM rats. *J Ethnopharmacol*. 127:98–102. [PubMed: 19799987]

190. Saleem M, Murtaza I, Witkowsky O, Kohl AM, Maddodi N. Lupeol triterpene, a novel diet-based microtubule targeting agent: disrupts survivin/cFLIP activation in prostate cancer cells. *Biochem Biophys Res Commun.* 2009; 388:576–82. [PubMed: 19683515]
191. Muruganandan S, Srinivasan K, Gupta S, Gupta PK, Lal J. Effect of mangiferin on hyperglycemia and atherogenicity in streptozotocin diabetic rats. *J Ethnopharmacol.* 2005; 97:497–501. [PubMed: 15740886]
192. Samarth RM, Panwar M, Kumar M, Kumar A. Radioprotective influence of *Mentha piperita* (Linn) against gamma irradiation in mice: Antioxidant and radical scavenging activity. *Int J Radiat Biol.* 2006; 82:331–7. [PubMed: 16782650]
193. Samarth RM, Goyal PK, Kumar A. Protection of swiss albino mice against whole-body gamma irradiation by *Mentha piperita* (Linn.) *Phytother Res.* 2004; 18:546–50. [PubMed: 15305314]
194. Atta AH, Alkofahi A. Anti-nociceptive and anti-inflammatory effects of some Jordanian medicinal plant extracts. *J Ethnopharmacol.* 1998; 60:117–24. [PubMed: 9582001]
195. Yadav AS, Bhatnagar D. Inhibition of iron induced lipid peroxidation and antioxidant activity of Indian spices and *Acacia* in vitro. *Plant Foods Hum Nutr.* 65:18–24. [PubMed: 20033297]
196. Shah PJ, Gandhi MS, Shah MB, Goswami SS, Santani D. Study of *Mimusops elengi* bark in experimental gastric ulcers. *J Ethnopharmacol.* 2003; 89:305–11. [PubMed: 14611897]
197. Grover JK, Yadav SP. Pharmacological actions and potential uses of *Momordica charantia*: a review. *J Ethnopharmacol.* 2004; 93:123–32. [PubMed: 15182917]
198. Schmourlo G, Mendonca-Filho RR, Alviano CS, Costa SS. Screening of antifungal agents using ethanol precipitation and bioautography of medicinal and food plants. *J Ethnopharmacol.* 2005; 96:563–8. [PubMed: 15619579]
199. Mahajan SG, Mehta AA. Effect of *Moringa oleifera* Lam. seed extract on ovalbumin-induced airway inflammation in guinea pigs. *Inhal Toxicol.* 2008; 20:897–909. [PubMed: 18686107]
200. Kar A, Choudhary BK, Bandyopadhyay NG. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. *J Ethnopharmacol.* 2003; 84:105–8. [PubMed: 12499084]
201. Yi T, Cho SG, Yi Z, Pang X, Rodriguez M, Wang Y, et al. Thymoquinone inhibits tumor angiogenesis and tumor growth through suppressing AKT and extracellular signal-regulated kinase signaling pathways. *Mol Cancer Ther.* 2008; 7:1789–96. [PubMed: 18644991]
202. Chatterjee A, Basak B, Saha M, Dutta U, Mukhopadhyay C, Banerji J, et al. Structure and stereochemistry of nardostachysin, a new terpenoid ester constituent of the rhizomes of *Nardostachys jatamansi*. *J Nat Prod.* 2000; 63:1531–3. [PubMed: 11087600]
203. Weng TC, Shen CC, Chiu YT, Lin YL, Kuo CD, Huang YT. Inhibitory effects of artemepavine against hepatic fibrosis in rats. *J Biomed Sci.* 2009; 16:78. [PubMed: 19723340]
204. Kim HK, Park HR, Lee JS, Chung TS, Chung HY, Chung J. Down-regulation of iNOS and TNF- α expression by kaempferol via NF- κ B inactivation in aged rat gingival tissues. *Biogerontology.* 2007; 8:399–408. [PubMed: 17278014]
205. Das S, Sasmal D, Basu SP. Anti-inflammatory and antinociceptive activity of arbortristoside-A. *J Ethnopharmacol.* 2008; 116:198–203. [PubMed: 18178352]
206. Grover JK, Yadav S, Vats V. Medicinal plants of India with anti-diabetic potential. *J Ethnopharmacol.* 2002; 81:81–100. [PubMed: 12020931]
207. Sarkar A, Lavania SC, Pandey DN, Pant MC. Changes in the blood lipid profile after administration of *Ocimum sanctum* (Tulsi) leaves in the normal albino rabbits. *Indian J Physiol Pharmacol.* 1994; 38:311–2. [PubMed: 7883302]
208. Shishodia S, Majumdar S, Banerjee S, Aggarwal BB. Ursolic acid inhibits nuclear factor- κ B activation induced by carcinogenic agents through suppression of IkappaB α kinase and p65 phosphorylation: correlation with down-regulation of cyclooxygenase 2, matrix metalloproteinase 9, and cyclin D1. *Cancer Res.* 2003; 63:4375–83. [PubMed: 12907607]
209. Pathak AK, Bhutani M, Nair AS, Ahn KS, Chakraborty A, Kadara H, et al. Ursolic acid inhibits STAT3 activation pathway leading to suppression of proliferation and chemosensitization of human multiple myeloma cells. *Mol Cancer Res.* 2007; 5:943–55. [PubMed: 17855663]

210. Ahmad R, Ahmed S, Khan NU, Hasnain AU. Operculina turpethum attenuates N-nitrosodimethylamine induced toxic liver injury and clastogenicity in rats. *Chem Biol Interact.* 2009; 181:145–53. [PubMed: 19589336]
211. Aziz N, Mehmood MH, Siddiqi HS, Mandukhail SU, Sadiq F, Maan W, et al. Antihypertensive, antidyslipidemic and endothelial modulating activities of *Orchis mascula*. *Hypertens Res.* 2009; 32:997–1003. [PubMed: 19745827]
212. Roy MK, Nakahara K, Na TV, Trakoontivakorn G, Takenaka M, Isobe S, et al. Baicalein, a flavonoid extracted from a methanolic extract of *Oroxylum indicum* inhibits proliferation of a cancer cell line in vitro via induction of apoptosis. *Pharmazie.* 2007; 62:149–53. [PubMed: 17341037]
213. Tan MA, Takayama H, Aimi N, Kitajima M, Franzblau SG, Nonato MG. Antitubercular triterpenes and phytosterols from *Pandanus tectorius* Soland. var. *laevis*. *J Nat Med.* 2008; 62:232–5. [PubMed: 18404330]
214. Kassuya CA, Silvestre A, Menezes-de-Lima O Jr, Marotta DM, Rehder VL, Calixto JB. Antiinflammatory and antiallodynic actions of the lignan niranthin isolated from *Phyllanthus amarus*. Evidence for interaction with platelet activating factor receptor. *Eur J Pharmacol.* 2006; 546:182–8. [PubMed: 16925995]
215. Anand P, Kunnumakkara AB, Harikumar KB, Ahn KS, Badmaev V, Aggarwal BB. Modification of cysteine residue in p65 subunit of nuclear factor-kappaB (NF-kappaB) by picroliv suppresses NF-kappaB-regulated gene products and potentiates apoptosis. *Cancer Res.* 2008; 68:8861–70. [PubMed: 18974130]
216. Singh N, Kumar S, Singh P, Raj HG, Prasad AK, Parmar VS, et al. Piper longum Linn. Extract inhibits TNF-alpha-induced expression of cell adhesion molecules by inhibiting NF-kappaB activation and microsomal lipid peroxidation. *Phytomedicine.* 2008; 15:284–91. [PubMed: 17689945]
217. Stohr JR, Xiao PG, Bauer R. Constituents of Chinese Piper species and their inhibitory activity on prostaglandin and leukotriene biosynthesis in vitro. *J Ethnopharmacol.* 2001; 75:133–9. [PubMed: 11297843]
218. Ahmad NS, Waheed A, Farman M, Qayyum A. Analgesic and anti-inflammatory effects of *Pistacia integerrima* extracts in mice. *J Ethnopharmacol.*
219. Avila MA, Velasco JA, Cansado J, Notario V. Quercetin mediates the down-regulation of mutant p53 in the human breast cancer cell line MDA-MB468. *Cancer Res.* 1994; 54:2424–8. [PubMed: 8162591]
220. Tilak JC, Adhikari S, Devasagayam TP. Antioxidant properties of *Plumbago zeylanica*, an Indian medicinal plant and its active ingredient, plumbagin. *Redox Rep.* 2004; 9:219–27. [PubMed: 15479566]
221. Sandur SK, Ichikawa H, Sethi G, Ahn KS, Aggarwal BB. Plumbagin (5-hydroxy-2-methyl-1,4-naphthoquinone) suppresses NF-kappaB activation and NF-kappaB-regulated gene products through modulation of p65 and IkappaBalpha kinase activation, leading to potentiation of apoptosis induced by cytokine and chemotherapeutic agents. *J Biol Chem.* 2006; 281:17023–33. [PubMed: 16624823]
222. Sandur SK, Pandey MK, Sung B, Aggarwal BB. 5-hydroxy-2-methyl-1,4-naphthoquinone, a vitamin K3 analogue, suppresses STAT3 activation pathway through induction of protein tyrosine phosphatase, SHP-1: potential role in chemosensitization. *Mol Cancer Res.* 8:107–18. [PubMed: 20068065]
223. Khan H, Saeed M, Gilani AU, Khan MA, Dar A, Khan I. The antinociceptive activity of *Polygonatum verticillatum* rhizomes in pain models. *J Ethnopharmacol.* 127:521–7. [PubMed: 19853648]
224. Badole SL, Bodhankar SL. Investigation of antihyperglycaemic activity of aqueous and petroleum ether extract of stem bark of *Pongamia pinnata* on serum glucose level in diabetic mice. *J Ethnopharmacol.* 2009; 123:115–20. [PubMed: 19429349]
225. Sang S, Kikuzaki H, Lapsley K, Rosen RT, Nakatani N, Ho CT. Sphingolipid and other constituents from almond nuts (*Prunus amygdalus* Batsch). *J Agric Food Chem.* 2002; 50:4709–12. [PubMed: 12137501]

226. Babu TD, Sasidharan N, Vijayan FP, Padikkala J. Comparative phytochemical and biological analysis to detect the genuineness of substitutes of the plant Moovila in drug preparations. *J Basic Clin Physiol Pharmacol*. 2008; 19:119–30. [PubMed: 19024929]
227. Wolf P, Nghiem DX, Walterscheid JP, Byrne S, Matsumura Y, Bucana C, et al. Platelet-activating factor is crucial in psoralen and ultraviolet A-induced immune suppression, inflammation, and apoptosis. *Am J Pathol*. 2006; 169:795–805. [PubMed: 16936256]
228. Hougee S, Faber J, Sanders A, de Jong RB, van den Berg WB, Garssen J, et al. Selective COX-2 inhibition by a *Pterocarpus marsupium* extract characterized by pterostilbene, and its activity in healthy human volunteers. *Planta Med*. 2005; 71:387–92. [PubMed: 15931573]
229. Kondeti VK, Badri KR, Maddirala DR, Thur SK, Fatima SS, Kasetti RB, et al. Effect of *Pterocarpus santalinus* bark, on blood glucose, serum lipids, plasma insulin and hepatic carbohydrate metabolic enzymes in streptozotocin-induced diabetic rats. *Food Chem Toxicol*. 48:1281–7. [PubMed: 20178824]
230. Adams LS, Seeram NP, Aggarwal BB, Takada Y, Sand D, Heber D. Pomegranate juice, total pomegranate ellagitannins, and punicalagin suppress inflammatory cell signaling in colon cancer cells. *J Agric Food Chem*. 2006; 54:980–5. [PubMed: 16448212]
231. Reanmongkol W, Noppapan T, Subhadhirasakul S. Antinociceptive, antipyretic, and anti-inflammatory activities of *Putranjiva roxburghii* Wall. leaf extract in experimental animals. *J Nat Med*. 2009; 63:290–6. [PubMed: 19387768]
232. Kaur G, Hamid H, Ali A, Alam MS, Athar M. Antiinflammatory evaluation of alcoholic extract of galls of *Quercus infectoria*. *J Ethnopharmacol*. 2004; 90:285–92. [PubMed: 15013194]
233. Sipos P, Hagymasi K, Lugasi A, Feher E, Blazovics A. Effects of black radish root (*Raphanus sativus* L. var *niger*) on the colon mucosa in rats fed a fat rich diet. *Phytother Res*. 2002; 16:677–9. [PubMed: 12410553]
234. Leary WP, Reyes AJ, van der Byl K. Effects of single doses of two antihypertensive rauwolfia-diuretic combinations on urinary excretion. *S Afr Med J*. 1986; 70:95–8. [PubMed: 3088739]
235. Lam FY, Ferrell WR. Neurogenic component of different models of acute inflammation in the rat knee joint. *Ann Rheum Dis*. 1991; 50:747–51. [PubMed: 1722965]
236. Mokhtarian F, Griffin DE. The role of mast cells in virus-induced inflammation in the murine central nervous system. *Cell Immunol*. 1984; 86:491–500. [PubMed: 6329524]
237. Vieira C, Fetzter S, Sauer SK, Evangelista S, Averbek B, Kress M, et al. Pro- and anti-inflammatory actions of ricinoleic acid: similarities and differences with capsaicin. *Naunyn Schmiedebergs Arch Pharmacol*. 2001; 364:87–95. [PubMed: 11534859]
238. Han SH, Hur MH, Buckle J, Choi J, Lee MS. Effect of aromatherapy on symptoms of dysmenorrhea in college students: A randomized placebo-controlled clinical trial. *J Altern Complement Med*. 2006; 12:535–41. [PubMed: 16884344]
239. Kwon EK, Lee DY, Lee H, Kim DO, Baek NI, Kim YE, et al. Flavonoids from the buds of *Rosa damascena* inhibit the activity of 3-hydroxy-3-methylglutaryl-coenzyme A reductase and angiotensin I-converting enzyme. *J Agric Food Chem*. 58:882–6. [PubMed: 20038104]
240. Kim KJ, Lee JS, Kwak MK, Choi HG, Yong CS, Kim JA, et al. Anti-inflammatory action of mollugin and its synthetic derivatives in HT-29 human colonic epithelial cells is mediated through inhibition of NF-kappaB activation. *Eur J Pharmacol*. 2009; 622:52–7. [PubMed: 19765578]
241. Rouf AS, Islam MS, Rahman MT. Evaluation of antidiarrhoeal activity *Rumex maritimus* root. *J Ethnopharmacol*. 2003; 84:307–10. [PubMed: 12648830]
242. Sanogo R, Monforte MT, Daquino A, Rossitto A, Maur DD, Galati EM. Antiulcer activity of *Salvadora persica* L: structural modifications. *Phytomedicine*. 1999; 6:363–6. [PubMed: 11962545]
243. Setzer WN. Essential oils and anxiolytic aromatherapy. *Nat Prod Commun*. 2009; 4:1305–16. [PubMed: 19831048]
244. Arulmozhi DK, Veeranjanyulu A, Bodhankar SL, Arora SK. Effect of *Sapindus trifoliatus* on hyperalgesic in vivo migraine models. *Braz J Med Biol Res*. 2005; 38:469–75. [PubMed: 15761628]

245. Arul B, Kothai R, Jacob P, Sangameswaran B, Sureshkumar K. Anti-inflammatory activity of *Sapindus trifoliatus* Linn. *J Herb Pharmacother*. 2004; 4:43–50. [PubMed: 15927924]
246. Middelkoop TB, Labadie RP. The action of *Saraca asoca* Roxb. de Wilde bark on the PGH2 synthetase enzyme complex of the sheep vesicular gland. *Z Naturforsch C*. 1985; 40:523–6. [PubMed: 3931371]
247. Ghosh S, Majumder M, Majumder S, Ganguly NK, Chatterjee BP. Saracin: A lectin from *Saraca indica* seed integument induces apoptosis in human T-lymphocytes. *Arch Biochem Biophys*. 1999; 371:163–8. [PubMed: 10545202]
248. Li Y, Xu C, Zhang Q, Liu JY, Tan RX. In vitro anti-*Helicobacter pylori* action of 30 Chinese herbal medicines used to treat ulcer diseases. *J Ethnopharmacol*. 2005; 98:329–33. [PubMed: 15814268]
249. Sankar D, Sambandam G, Ramakrishna Rao M, Pugalendi KV. Modulation of blood pressure, lipid profiles and redox status in hypertensive patients taking different edible oils. *Clin Chim Acta*. 2005; 355:97–104. [PubMed: 15820483]
250. Harikumar KB, Sung B, Tharakan ST, Pandey MK, Joy B, Guha S, et al. Sesamin Manifests Chemopreventive Effects through the Suppression of NF- κ B-Regulated Cell Survival, Proliferation, Invasion, and Angiogenic Gene Products. *Mol Cancer Res*.
251. Matsui TA, Sowa Y, Murata H, Takagi K, Nakanishi R, Aoki S, et al. The plant alkaloid cryptolepine induces p21WAF1/CIP1 and cell cycle arrest in a human osteosarcoma cell line. *Int J Oncol*. 2007; 31:915–22. [PubMed: 17786325]
252. Bahgat A, Abdel-Aziz H, Raafat M, Mahdy A, El-Khatib AS, Ismail A, et al. *Solanum indicum* ssp. *distichum* extract is effective against L-NAME-induced hypertension in rats. *Fundam Clin Pharmacol*. 2008; 22:693–9. [PubMed: 19049674]
253. Ma P, Cao TT, Gu GF, Zhao X, Du YG, Zhang Y. Inducement effect of synthetic indiosides from *Solanum indicum* L. on apoptosis of human hepatocarcinoma cell line Bel-7402 and its mechanism. *Ai Zheng*. 2006; 25:438–42. [PubMed: 16613676]
254. Raju K, Anbuganapathi G, Gokulakrishnan V, Rajkapoor B, Jayakar B, Manian S. Effect of dried fruits of *Solanum nigrum* LINN against CCl4-induced hepatic damage in rats. *Biol Pharm Bull*. 2003; 26:1618–9. [PubMed: 14600413]
255. Govindan S, Viswanathan S, Vijayasekaran V, Alagappan R. A pilot study on the clinical efficacy of *Solanum xanthocarpum* and *Solanum trilobatum* in bronchial asthma. *J Ethnopharmacol*. 1999; 66:205–10. [PubMed: 10433479]
256. Prabhu KS, Lobo R, Shirwaikar A. Antidiabetic properties of the alcoholic extract of *Sphaeranthus indicus* in streptozotocin-nicotinamide diabetic rats. *J Pharm Pharmacol*. 2008; 60:909–16. [PubMed: 18549678]
257. Balasubramanian T, Chatterjee TK, Sarkar M, Meena SL. Anti-inflammatory effect of *Stereospermum suaveolens* ethanol extract in rats. *Pharm Biol*. 48:318–23. [PubMed: 20645819]
258. Yin W, Deng XK, Yin FZ, Zhang XC, Cai BC. The cytotoxicity induced by brucine from the seed of *Strychnos nux-vomica* proceeds via apoptosis and is mediated by cyclooxygenase 2 and caspase 3 in SMMC 7221 cells. *Food Chem Toxicol*. 2007; 45:1700–8. [PubMed: 17449162]
259. Saha P, Mandal S, Das A, Das S. Amarogentin can reduce hyperproliferation by downregulation of Cox-II and upregulation of apoptosis in mouse skin carcinogenesis model. *Cancer Lett*. 2006; 244:252–9. [PubMed: 16517061]
260. Na M, Yang S, He L, Oh H, Kim BS, Oh WK, et al. Inhibition of protein tyrosine phosphatase 1B by ursane-type triterpenes isolated from *Symplocos paniculata*. *Planta Med*. 2006; 72:261–3. [PubMed: 16534732]
261. Banerjee S, Panda CK, Das S. Clove (*Syzygium aromaticum* L), a potential chemopreventive agent for lung cancer. *Carcinogenesis*. 2006; 27:1645–54. [PubMed: 16501250]
262. Jagetia GC, Baliga MS. *Syzygium cumini* (Jamun) reduces the radiation-induced DNA damage in the cultured human peripheral blood lymphocytes: a preliminary study. *Toxicol Lett*. 2002; 132:19–25. [PubMed: 12084616]
263. Sehrawat A, Sultana S. Evaluation of possible mechanisms of protective role of *Tamarix gallica* against DEN initiated and 2-AAF promoted hepatocarcinogenesis in male Wistar rats. *Life Sci*. 2006; 79:1456–65. [PubMed: 16698044]

264. Halder S, Bharal N, Mediratta PK, Kaur I, Sharma KK. Anti-inflammatory, immunomodulatory and antinociceptive activity of *Terminalia arjuna* Roxb bark powder in mice and rats. *Indian J Exp Biol.* 2009; 47:577–83. [PubMed: 19761042]
265. Malik N, Dhawan V, Bahl A, Kaul D. Inhibitory effects of *Terminalia arjuna* on platelet activation in vitro in healthy subjects and patients with coronary artery disease. *Platelets.* 2009; 20:183–90. [PubMed: 19437336]
266. Shaila HP, Udupa AL, Udupa SL. Preventive actions of *Terminalia belerica* in experimentally induced atherosclerosis. *Int J Cardiol.* 1995; 49:101–6. [PubMed: 7628881]
267. Shinde SL, Junne SB, Wadje SS, Baig MM. The diversity of antibacterial compounds of *Terminalia* species (Combretaceae). *Pak J Biol Sci.* 2009; 12:1483–6. [PubMed: 20180323]
268. Shin TY, Jeong HJ, Kim DK, Kim SH, Lee JK, Chae BS, et al. Inhibitory action of water soluble fraction of *Terminalia chebula* on systemic and local anaphylaxis. *J Ethnopharmacol.* 2001; 74:133–40. [PubMed: 11167031]
269. Vigo E, Cepeda A, Gualillo O, Perez-Fernandez R. In-vitro anti-inflammatory effect of *Eucalyptus globulus* and *Thymus vulgaris*: nitric oxide inhibition in J774A. 1 murine macrophages. *J Pharm Pharmacol.* 2004; 56:257–63. [PubMed: 15005885]
270. Horvathova E, Turcaniova V, Slamenova D. Comparative study of DNA-damaging and DNA-protective effects of selected components of essential plant oils in human leukemic cells K562. *Neoplasma.* 2007; 54:478–83. [PubMed: 17949230]
271. Dhanasekaran M, Baskar AA, Ignacimuthu S, Agastian P, Duraipandiyan V. Chemopreventive potential of Epoxy clerodane diterpene from *Tinospora cordifolia* against diethylnitrosamine-induced hepatocellular carcinoma. *Invest New Drugs.* 2009; 27:347–55. [PubMed: 18853103]
272. Thippeswamy G, Sheela ML, Salimath BP. Octacosanol isolated from *Tinospora cordifolia* downregulates VEGF gene expression by inhibiting nuclear translocation of NF- κ B and its DNA binding activity. *Eur J Pharmacol.* 2008; 588:141–50. [PubMed: 18513715]
273. Umamaheswari S, Mainzen Prince PS. Antihyperglycaemic effect of 'Ilogen-Excel', an ayurvedic herbal formulation in streptozotocin-induced diabetes mellitus. *Acta Pol Pharm.* 2007; 64:53–61. [PubMed: 17665851]
274. Singh B, Kale RK. Chemomodulatory effect of *Trachyspermum ammi* on murine skin and forestomach papillomagenesis. *Nutr Cancer.* 62:74–84. [PubMed: 20043262]
275. Kumar M, Soni AK, Shukla S, Kumar A. Chemopreventive potential of *Tribulus terrestris* against 7,12- dimethylbenz (a) anthracene induced skin papillomagenesis in mice. *Asian Pac J Cancer Prev.* 2006; 7:289–94. [PubMed: 16839225]
276. Amin A, Lotfy M, Shafiullah M, Adeghate E. The protective effect of *Tribulus terrestris* in diabetes. *Ann N Y Acad Sci.* 2006; 1084:391–401. [PubMed: 17151317]
277. Phillips OA, Mathew KT, Oriowo MA. Antihypertensive and vasodilator effects of methanolic and aqueous extracts of *Tribulus terrestris* in rats. *J Ethnopharmacol.* 2006; 104:351–5. [PubMed: 16289603]
278. Shishodia S, Aggarwal BB. Diosgenin inhibits osteoclastogenesis, invasion, and proliferation through the downregulation of Akt, I kappa B kinase activation and NF-kappa B-regulated gene expression. *Oncogene.* 2006; 25:1463–73. [PubMed: 16331273]
279. Li F, Fernandez PP, Rajendran P, Hui KM, Sethi G. Diosgenin, a steroidal saponin, inhibits STAT3 signaling pathway leading to suppression of proliferation and chemosensitization of human hepatocellular carcinoma cells. *Cancer Lett.* 292:197–207. [PubMed: 20053498]
280. Singh AB, Tamarkar AK, Narender T, Srivastava AK. Antihyperglycaemic effect of an unusual amino acid (4-hydroxyisoleucine) in C57BL/KsJ-db/db mice. *Nat Prod Res.* 24:258–65. [PubMed: 20140804]
281. Jabbar S, Khan MT, Choudhuri MS, Sil BK. Bioactivity studies of the individual ingredients of the Dashamularishta. *Pak J Pharm Sci.* 2004; 17:9–17. [PubMed: 16414581]
282. Jacobo-Herrera NJ, Vartiainen N, Bremner P, Gibbons S, Koistinaho J, Heinrich M. NF-kappaB modulators from *Valeriana officinalis*. *Phytother Res.* 2006; 20:917–9. [PubMed: 16909443]
283. Gilani AH, Khan AU, Jabeen Q, Subhan F, Ghafar R. Antispasmodic and blood pressure lowering effects of *Valeriana wallichii* are mediated through K⁺ channel activation. *J Ethnopharmacol.* 2005; 100:347–52. [PubMed: 16002246]

284. Nayak BS, Suresh R, Rao AV, Pillai GK, Davis EM, Ramkissoon V, et al. Evaluation of wound healing activity of *Vanda roxburghii* R. Br(Orchidaceae): a preclinical study in a rat model. *Int J Low Extrem Wounds*. 2005; 4:200–4. [PubMed: 16286371]
285. Pratheeshkumar P, Kuttan G. Modulation of immune response by *Vernonia cinerea* L. inhibits the proinflammatory cytokine profile, iNOS, and COX-2 expression in LPS-stimulated macrophages. *Immunopharmacol Immunotoxicol*.
286. Lindholm P, Goransson U, Johansson S, Claeson P, Gullbo J, Larsson R, et al. Cyclotides: a novel type of cytotoxic agents. *Mol Cancer Ther*. 2002; 1:365–9. [PubMed: 12477048]
287. Zhou Y, Liu YE, Cao J, Zeng G, Shen C, Li Y, et al. Vitexins, nature-derived lignan compounds, induce apoptosis and suppress tumor growth. *Clin Cancer Res*. 2009; 15:5161–9. [PubMed: 19671865]
288. Ichikawa H, Takada Y, Shishodia S, Jayaprakasam B, Nair MG, Aggarwal BB. Withanolides potentiate apoptosis, inhibit invasion, and abolish osteoclastogenesis through suppression of nuclear factor-kappaB (NF-kappaB) activation and NF-kappaB-regulated gene expression. *Mol Cancer Ther*. 2006; 5:1434–45. [PubMed: 16818501]
289. Mulabagal V, Subbaraju GV, Rao CV, Sivaramakrishna C, Dewitt DL, Holmes D, et al. Withanolide sulfoxide from *Aswagandha* roots inhibits nuclear transcription factor-kappa-B, cyclooxygenase and tumor cell proliferation. *Phytother Res*. 2009; 23:987–92. [PubMed: 19152372]
290. Koduru S, Kumar R, Srinivasan S, Evers MB, Damodaran C. Notch-1 inhibition by Withaferin-A: a therapeutic target against colon carcinogenesis. *Mol Cancer Ther*. 9:202–10. [PubMed: 20053782]
291. El-Abhar HS, Hammad LN, Gawad HS. Modulating effect of ginger extract on rats with ulcerative colitis. *J Ethnopharmacol*. 2008; 118:367–72. [PubMed: 18571884]
292. Kuo SC, Chen SC, Chen LH, Wu JB, Wang JP, Teng CM. Potent antiplatelet, anti-inflammatory and antiallergic isoflavanquinones from the roots of *Abrus precatorius*. *Planta Med*. 1995; 61:307–12. [PubMed: 7480175]
293. Ramnath V, Rekha PS, Kuttan G, Kuttan R. Regulation of Caspase-3 and Bcl-2 Expression in Dalton's Lymphoma Ascites Cells by Abrin. *Evid Based Complement Alternat Med*. 2009; 6:233–8. [PubMed: 18955274]
294. Ramnath V, Kuttan G, Kuttan R. Antitumour effect of abrin on transplanted tumours in mice. *Indian J Physiol Pharmacol*. 2002; 46:69–77. [PubMed: 12024960]
295. Ghosh D, Maiti TK. Immunomodulatory and anti-tumor activities of native and heat denatured *Abrus* agglutinin. *Immunobiology*. 2007; 212:589–99. [PubMed: 17678717]
296. Bhutia SK, Mallick SK, Maiti S, Maiti TK. Inhibitory effect of *Abrus* abrin-derived peptide fraction against Dalton's lymphoma ascites model. *Phytomedicine*. 2009; 16:377–85. [PubMed: 18706794]
297. Clark DT, Gazi MI, Cox SW, Eley BM, Tinsley GF. The effects of *Acacia arabica* gum on the in vitro growth and protease activities of periodontopathic bacteria. *J Clin Periodontol*. 1993; 20:238–43. [PubMed: 8473532]
298. Morgan SL, Baggott JE, Moreland L, Desmond R, Kendrach AC. The safety of flavocoxid, a medical food, in the dietary management of knee osteoarthritis. *J Med Food*. 2009; 12:1143–8. [PubMed: 19857081]
299. Haidara K, Zamir L, Shi QW, Batist G. The flavonoid Casticin has multiple mechanisms of tumor cytotoxicity action. *Cancer Lett*. 2006; 242:180–90. [PubMed: 16387422]
300. Benedek B, Kopp B. *Achillea millefolium* L. s.l. revisited: recent findings confirm the traditional use. *Wien Med Wochenschr*. 2007; 157:312–4. [PubMed: 17704978]
301. Cavalcanti AM, Baggio CH, Freitas CS, Rieck L, de Sousa RS, Da Silva-Santos JE, et al. Safety and antiulcer efficacy studies of *Achillea millefolium* L. after chronic treatment in Wistar rats. *J Ethnopharmacol*. 2006; 107:277–84. [PubMed: 16647233]
302. Vetrichelvan T, Jegadeesan M. Effect of alcohol extract of *Achyranthes aspera* Linn. on acute and subacute inflammation. *Phytother Res*. 2003; 17:77–9. [PubMed: 12557252]

303. Shukla PK, Khanna VK, Ali MM, Maurya R, Khan MY, Srimal RC. Neuroprotective effect of *Acorus calamus* against middle cerebral artery occlusion-induced ischaemia in rat. *Hum Exp Toxicol*. 2006; 25:187–94. [PubMed: 16696294]
304. Shrivastava N, Srivastava A, Banerjee A, Nivsarkar M. Anti-ulcer activity of *Adhatoda vasica* Nees. *J Herb Pharmacother*. 2006; 6:43–9. [PubMed: 17182484]
305. Jahangir T, Sultana S. Tumor Promotion and Oxidative Stress in Ferric Nitrilotriacetate-Mediated Renal Carcinogenesis: Protection by *Adhatoda vasica*. *Toxicol Mech Methods*. 2007; 17:421–30. [PubMed: 20020945]
306. Claeson UP, Malmfors T, Wikman G, Bruhn JG. *Adhatoda vasica*: a critical review of ethnopharmacological and toxicological data. *J Ethnopharmacol*. 2000; 72:1–20. [PubMed: 10967448]
307. Maity P, Hansda D, Bandyopadhyay U, Mishra DK. Biological activities of crude extracts and chemical constituents of *Bael*, *Aegle marmelos* (L). *Corr Indian J Exp Biol*. 2009; 47:849–61.
308. Khan TH, Sultana S. Antioxidant and hepatoprotective potential of *Aegle marmelos* Correa. against CCl₄-induced oxidative stress and early tumor events. *J Enzyme Inhib Med Chem*. 2009; 24:320–7. [PubMed: 18830880]
309. Aviello G, Abenavoli L, Borrelli F, Capasso R, Izzo AA, Lembo F, et al. Garlic: empiricism or science? *Nat Prod Commun*. 2009; 4:1785–96. [PubMed: 20120123]
310. Butt MS, Sultan MT, Iqbal J. Garlic: nature's protection against physiological threats. *Crit Rev Food Sci Nutr*. 2009; 49:538–51. [PubMed: 19484634]
311. Iciek M, Kwiecien I, Wlodek L. Biological properties of garlic and garlic-derived organosulfur compounds. *Environ Mol Mutagen*. 2009; 50:247–65. [PubMed: 19253339]
312. Pittler MH, Ernst E. Clinical effectiveness of garlic (*Allium sativum*). *Mol Nutr Food Res*. 2007; 51:1382–5. [PubMed: 17918163]
313. Lei YP, Chen HW, Sheen LY, Lii CK. Diallyl disulfide and diallyl trisulfide suppress oxidized LDL-induced vascular cell adhesion molecule and E-selectin expression through protein kinase A- and B-dependent signaling pathways. *J Nutr*. 2008; 138:996–1003. [PubMed: 18492825]
314. Lee HS, Lee CH, Tsai HC, Salter DM. Inhibition of cyclooxygenase 2 expression by diallyl sulfide on joint inflammation induced by urate crystal and IL-1 β . *Osteoarthritis Cartilage*. 2009; 17:91–9. [PubMed: 18573668]
315. Lang A, Lahav M, Sakhnini E, Barshack I, Fidler HH, Avidan B, et al. Allicin inhibits spontaneous and TNF- α induced secretion of proinflammatory cytokines and chemokines from intestinal epithelial cells. *Clin Nutr*. 2004; 23:199–208. [PubMed: 15380914]
316. Son EW, Mo SJ, Rhee DK, Pyo S. Inhibition of ICAM-1 expression by garlic component, allicin, in gamma-irradiated human vascular endothelial cells via downregulation of the JNK signaling pathway. *Int Immunopharmacol*. 2006; 6:1788–95. [PubMed: 17052669]
317. Guo J, Xiao B, Liu Q, Gong Z, Le Y. Suppression of C-myc expression associates with anti-proliferation of aloe-emodin on gastric cancer cells. *Cancer Invest*. 2008; 26:369–74. [PubMed: 18443957]
318. Lin JG, Chen GW, Li TM, Chouh ST, Tan TW, Chung JG. Aloe-emodin induces apoptosis in T24 human bladder cancer cells through the p53 dependent apoptotic pathway. *J Urol*. 2006; 175:343–7. [PubMed: 16406939]
319. Feily A, Namazi MR. Aloe vera in dermatology: a brief review. *G Ital Dermatol Venereol*. 2009; 144:85–91. [PubMed: 19218914]
320. Eamlamnam K, Patumraj S, Visdopas N, Thong-Ngam D. Effects of Aloe vera and sucralfate on gastric microcirculatory changes, cytokine levels and gastric ulcer healing in rats. *World J Gastroenterol*. 2006; 12:2034–9. [PubMed: 16610053]
321. Akev N, Turkay G, Can A, Gurel A, Yildiz F, Yardibi H, et al. Effect of Aloe vera leaf pulp extract on Ehrlich ascites tumours in mice. *Eur J Cancer Prev*. 2007; 16:151–7. [PubMed: 17297391]
322. Eshun K, He Q. Aloe vera: a valuable ingredient for the food, pharmaceutical and cosmetic industries--a review. *Crit Rev Food Sci Nutr*. 2004; 44:91–6. [PubMed: 15116756]
323. Vogler BK, Ernst E. Aloe vera: a systematic review of its clinical effectiveness. *Br J Gen Pract*. 1999; 49:823–8. [PubMed: 10885091]

324. Ye Y, Li B. 1'S-1'-acetoxychavicol acetate isolated from *Alpinia galanga* inhibits human immunodeficiency virus type 1 replication by blocking Rev transport. *J Gen Virol.* 2006; 87:2047–53. [PubMed: 16760408]
325. Matsuda H, Morikawa T, Managi H, Yoshikawa M. Antiallergic principles from *Alpinia galanga*: structural requirements of phenylpropanoids for inhibition of degranulation and release of TNF- α and IL-4 in RBL-2H3 cells. *Bioorg Med Chem Lett.* 2003; 13:3197–202. [PubMed: 12951092]
326. Morikawa T, Ando S, Matsuda H, Kataoka S, Muraoka O, Yoshikawa M. Inhibitors of nitric oxide production from the rhizomes of *Alpinia galanga*: structures of new 8–9' linked neolignans and sesquieolignan. *Chem Pharm Bull (Tokyo).* 2005; 53:625–30. [PubMed: 15930771]
327. Bao Z, Guan S, Cheng C, Wu S, Wong SH, Kemeny DM, et al. A novel antiinflammatory role for andrographolide in asthma via inhibition of the nuclear factor-kappaB pathway. *Am J Respir Crit Care Med.* 2009; 179:657–65. [PubMed: 19201922]
328. Burgos RA, Seguel K, Perez M, Meneses A, Ortega M, Guarda MI, et al. Andrographolide inhibits IFN- γ and IL-2 cytokine production and protects against cell apoptosis. *Planta Med.* 2005; 71:429–34. [PubMed: 15931581]
329. Yang L, Wu D, Luo K, Wu S, Wu P. Andrographolide enhances 5-fluorouracil-induced apoptosis via caspase-8-dependent mitochondrial pathway involving p53 participation in hepatocellular carcinoma (SMMC-7721) cells. *Cancer Lett.* 2009; 276:180–8. [PubMed: 19097688]
330. Shi MD, Lin HH, Lee YC, Chao JK, Lin RA, Chen JH. Inhibition of cell-cycle progression in human colorectal carcinoma Lovo cells by andrographolide. *Chem Biol Interact.* 2008; 174:201–10. [PubMed: 18619950]
331. Zhou J, Lu GD, Ong CS, Ong CN, Shen HM. Andrographolide sensitizes cancer cells to TRAIL-induced apoptosis via p53-mediated death receptor 4 up-regulation. *Mol Cancer Ther.* 2008; 7:2170–80. [PubMed: 18645026]
332. Zhao F, He EQ, Wang L, Liu K. Anti-tumor activities of andrographolide, a diterpene from *Andrographis paniculata*, by inducing apoptosis and inhibiting VEGF level. *J Asian Nat Prod Res.* 2008; 10:467–73. [PubMed: 18464090]
333. Harjotaruno S, Widyawaruyanti A, Sismindari, Zaini NC. Apoptosis Inducing Effect of Andrographolide on TD-47 Human Breast Cancer Cell Line. *Afr J Tradit Complement Altern Med.* 2007; 4:345–51. [PubMed: 20161898]
334. Sheeja K, Guruvayoorappan C, Kuttan G. Antiangiogenic activity of *Andrographis paniculata* extract and andrographolide. *Int Immunopharmacol.* 2007; 7:211–21. [PubMed: 17178389]
335. Trivedi NP, Rawal UM, Patel BP. Potency of andrographolide as an antitumor compound in BHC-induced liver damage. *Integr Cancer Ther.* 2009; 8:177–89. [PubMed: 19679627]
336. Abu-Ghefreh AA, Canatan H, Ezeamuzie CI. In vitro and in vivo anti-inflammatory effects of andrographolide. *Int Immunopharmacol.* 2009; 9:313–8. [PubMed: 19110075]
337. Iruretagoyena MI, Tobar JA, Gonzalez PA, Sepulveda SE, Figueroa CA, Burgos RA, et al. Andrographolide interferes with T cell activation and reduces experimental autoimmune encephalomyelitis in the mouse. *J Pharmacol Exp Ther.* 2005; 312:366–72. [PubMed: 15331658]
338. Sheeja K, Kuttan G. Activation of cytotoxic T lymphocyte responses and attenuation of tumor growth in vivo by *Andrographis paniculata* extract and andrographolide. *Immunopharmacol Immunotoxicol.* 2007; 29:81–93. [PubMed: 17464769]
339. Sheeja K, Kuttan G. Modulation of natural killer cell activity, antibody-dependent cellular cytotoxicity, and antibody-dependent complement-mediated cytotoxicity by andrographolide in normal and Ehrlich ascites carcinoma-bearing mice. *Integr Cancer Ther.* 2007; 6:66–73. [PubMed: 17351028]
340. Liu J, Wang ZT, Ji LL, Ge BX. Inhibitory effects of neoandrographolide on nitric oxide and prostaglandin E2 production in LPS-stimulated murine macrophage. *Mol Cell Biochem.* 2007; 298:49–57. [PubMed: 17109078]
341. Ji LL, Wang Z, Dong F, Zhang WB, Wang ZT. Andrograpanin, a compound isolated from anti-inflammatory traditional Chinese medicine *Andrographis paniculata*, enhances chemokine SDF-1 α -induced leukocytes chemotaxis. *J Cell Biochem.* 2005; 95:970–8. [PubMed: 15937916]

342. Liu J, Wang ZT, Ge BX. Andrograpanin, isolated from *Andrographis paniculata*, exhibits anti-inflammatory property in lipopolysaccharide-induced macrophage cells through down-regulating the p38 MAPKs signaling pathways. *Int Immunopharmacol*. 2008; 8:951–8. [PubMed: 18486905]
343. Chao WW, Kuo YH, Hsieh SL, Lin BF. Inhibitory effects of ethyl acetate extract of *Andrographis paniculata* on NF- κ B trans-activation activity and LPS-induced acute inflammation in mice. *Evid Based Complement Alternat Med*. 2009
344. Chandrasekaran CV, Gupta A, Agarwal A. Effect of an extract of *Andrographis paniculata* leaves on inflammatory and allergic mediators in vitro. *J Ethnopharmacol*. 129:203–7. [PubMed: 20307638]
345. Burgos RA, Hancke JL, Bertoglio JC, Aguirre V, Arriagada S, Calvo M, et al. Efficacy of an *Andrographis paniculata* composition for the relief of rheumatoid arthritis symptoms: a prospective randomized placebo-controlled trial. *Clin Rheumatol*. 2009; 28:931–46. [PubMed: 19408036]
346. Gabrielian ES, Shukarian AK, Goukasova GI, Chandanian GL, Panossian AG, Wikman G, et al. A double blind, placebo-controlled study of *Andrographis paniculata* fixed combination Kan Jang in the treatment of acute upper respiratory tract infections including sinusitis. *Phytomedicine*. 2002; 9:589–97. [PubMed: 12487322]
347. Lee KK, Choi JD. The effects of areca catechu L extract on anti-inflammation and anti-melanogenesis. *Int J Cosmet Sci*. 1999; 21:275–84. [PubMed: 18503445]
348. Habbu P, Shastry R, Mahadevan KM, Joshi H, Das S. Hepatoprotective and antioxidant effects of *Argyrea speciosa* in rats. *Afr J Tradit Complement Altern Med*. 2008; 5:158–64. [PubMed: 20161932]
349. Gokhale AB, Damre AS, Saraf MN. Investigations into the immunomodulatory activity of *Argyrea speciosa*. *J Ethnopharmacol*. 2003; 84:109–14. [PubMed: 12499085]
350. Bachhav RS, Gulecha VS, Upasani CD. Analgesic and anti-inflammatory activity of *Argyrea speciosa* root. *Indian J Pharmacol*. 2009; 41:158–61. [PubMed: 20523865]
351. Habbu PV, Mahadevan KM, Kulkarni PV, Daulatsingh C, Veerapur VP, Shastry RA. Adaptogenic and in vitro antioxidant activity of flavanoids and other fractions of *Argyrea speciosa* (Burm.f) Boj. in acute and chronic stress paradigms in rodents. *Indian J Exp Biol*. 48:53–60. [PubMed: 20358867]
352. Kanwar AS, Bhutani KK. Effects of *Chlorophytum arundinaceum*, *Asparagus adscendens* and *Asparagus racemosus* on pro-inflammatory cytokine and corticosterone levels produced by stress. *Phytother Res*.
353. Visavadiya NP, Soni B, Madamwar D. Suppression of reactive oxygen species and nitric oxide by *Asparagus racemosus* root extract using in vitro studies. *Cell Mol Biol (Noisy-le-grand)*. 2009; 55(Suppl):OL1083–95. [PubMed: 19267991]
354. Singh GK, Garabadu D, Muruganandam AV, Joshi VK, Krishnamurthy S. Antidepressant activity of *Asparagus racemosus* in rodent models. *Pharmacol Biochem Behav*. 2009; 91:283–90. [PubMed: 18692086]
355. Agrawal A, Sharma M, Rai SK, Singh B, Tiwari M, Chandra R. The effect of the aqueous extract of the roots of *Asparagus racemosus* on hepatocarcinogenesis initiated by diethylnitrosamine. *Phytother Res*. 2008; 22:1175–82. [PubMed: 18729252]
356. Goyal RK, Singh J, Lal H. *Asparagus racemosus*--an update. *Indian J Med Sci*. 2003; 57:408–14. [PubMed: 14515032]
357. Bopana N, Saxena S. *Asparagus racemosus*--ethnopharmacological evaluation and conservation needs. *J Ethnopharmacol*. 2007; 110:1–15. [PubMed: 17240097]
358. Priyadarsini RV, Manikandan P, Kumar GH, Nagini S. The neem limonoids azadirachtin and nimbolide inhibit hamster cheek pouch carcinogenesis by modulating xenobiotic-metabolizing enzymes, DNA damage, antioxidants, invasion and angiogenesis. *Free Radic Res*. 2009; 43:492–504. [PubMed: 19391054]
359. Gangar SC, Sandhir R, Koul A. Anti-clastogenic activity of *Azadirachta indica* against benzo(a)pyrene in murine forestomach tumorigenesis bioassay. *Acta Pol Pharm*. 67:381–90. [PubMed: 20635534]

360. Subapriya R, Nagini S. Medicinal properties of neem leaves: a review. *Curr Med Chem Anticancer Agents*. 2005; 5:149–6. [PubMed: 15777222]
361. Maity P, Biswas K, Chattopadhyay I, Banerjee RK, Bandyopadhyay U. The use of neem for controlling gastric hyperacidity and ulcer. *Phytother Res*. 2009; 23:747–55. [PubMed: 19140119]
362. Akihisa T, Noto T, Takahashi A, Fujita Y, Banno N, Tokuda H, et al. Melanogenesis inhibitory, anti-inflammatory, and chemopreventive effects of limonoids from the seeds of *Azadirachta indica* A. Juss. (neem). *J Oleo Sci*. 2009; 58:581–94. [PubMed: 19844073]
363. Bose A, Chakraborty K, Sarkar K, Goswami S, Chakraborty T, Pal S, et al. Neem leaf glycoprotein induces perforin-mediated tumor cell killing by T and NK cells through differential regulation of IFN γ signaling. *J Immunother*. 2009; 32:42–53. [PubMed: 19307993]
364. Ramzanighara A, Ezzatighadi F, Rai DV, Dhawan DK. Effect of Neem (*Azadirachta indica*) on serum glycoprotein contents of rats administered 1,2 dimethylhydrazine. *Toxicol Mech Methods*. 2009; 19:298–301. [PubMed: 19778220]
365. Vinothini G, Manikandan P, Anandan R, Nagini S. Chemoprevention of rat mammary carcinogenesis by *Azadirachta indica* leaf fractions: modulation of hormone status, xenobiotic-metabolizing enzymes, oxidative stress, cell proliferation and apoptosis. *Food Chem Toxicol*. 2009; 47:1852–63. [PubMed: 19427891]
366. Ghosh D, Bose A, Haque E, Baral R. Neem (*Azadirachta indica*) leaf preparation prevents leukocyte apoptosis mediated by cisplatin plus 5-fluorouracil treatment in Swiss mice. *Chemotherapy*. 2009; 55:137–44. [PubMed: 19346744]
367. Bhat M, Kothiwale SK, Tirmale AR, Bhargava SY, Joshi BN. Antidiabetic Properties of *Azadirachta indica* and *Bougainvillea spectabilis*: In Vivo Studies in Murine Diabetes Model. *Evid Based Complement Alternat Med*. 2009
368. Koul A, Bharrhan S, Singh B, Rishi P. Potential of *Azadirachta indica* against *Salmonella typhimurium*-induced inflammation in BALB/c mice. *Inflammopharmacology*. 2009; 17:29–36. [PubMed: 19127350]
369. Manikandan P, Vidhya Letchoumy P, Prathiba D, Nagini S. Combinatorial chemopreventive effect of *Azadirachta indica* and *Ocimum sanctum* on oxidant-antioxidant status, cell proliferation, apoptosis and angiogenesis in a rat forestomach carcinogenesis model. *Singapore Med J*. 2008; 49:814–22. [PubMed: 18946617]
370. Zhou Y, Shen YH, Zhang C, Su J, Liu RH, Zhang WD. Triterpene saponins from *Bacopa monnieri* and their antidepressant effects in two mice models. *J Nat Prod*. 2007; 70:652–5. [PubMed: 17343408]
371. Limpeanchob N, Jaipan S, Rattanakaruna S, Phrompittayarat W, Ingkaninan K. Neuroprotective effect of *Bacopa monnieri* on beta-amyloid-induced cell death in primary cortical culture. *J Ethnopharmacol*. 2008; 120:112–7. [PubMed: 18755259]
372. Uabundit N, Wattanathorn J, Mucimapura S, Ingkaninan K. Cognitive enhancement and neuroprotective effects of *Bacopa monnieri* in Alzheimer's disease model. *J Ethnopharmacol*. 127:26–31. [PubMed: 19808086]
373. Krishnakumar A, Abraham PM, Paul J, Paulose CS. Down-regulation of cerebellar 5-HT(2C) receptors in pilocarpine-induced epilepsy in rats: therapeutic role of *Bacopa monnieri* extract. *J Neurol Sci*. 2009; 284:124–8. [PubMed: 19439326]
374. Agrawal RC, Pandey S. Evaluation of anticarcinogenic and antimutagenic potential of *Bauhinia variegata* extract in Swiss albino mice. *Asian Pac J Cancer Prev*. 2009; 10:913–6. [PubMed: 20104989]
375. Bodakhe SH, Ram A. Hepatoprotective properties of *Bauhinia variegata* bark extract. *Yakugaku Zasshi*. 2007; 127:1503–7. [PubMed: 17827931]
376. Raj Kapoor B, Jayakar B, Muruges N. Antitumor activity of *Bauhinia variegata* on Dalton's ascitic lymphoma. *J Ethnopharmacol*. 2003; 89:107–9. [PubMed: 14522440]
377. Rao Y, Fang SH, Tzeng YM. Antiinflammatory activities of flavonoids and a triterpene caffeate isolated from *Bauhinia variegata*. *Phytother Res*. 2008; 22:957–62. [PubMed: 18384188]
378. Kulkarni SK, Dhir A. On the mechanism of antidepressant-like action of berberine chloride. *Eur J Pharmacol*. 2008; 589:163–72. [PubMed: 18585703]

379. Gupta SK, Agarwal R, Srivastava S, Agarwal P, Agrawal SS, Saxena R, et al. The anti-inflammatory effects of *Curcuma longa* and *Berberis aristata* in endotoxin-induced uveitis in rabbits. *Invest Ophthalmol Vis Sci*. 2008; 49:4036–40. [PubMed: 18421073]
380. Rajbhandari M, Wegner U, Schopke T, Lindequist U, Mentel R. Inhibitory effect of *Bergenia ligulata* on influenza virus A. *Pharmazie*. 2003; 58:268–71. [PubMed: 12749411]
381. Olaleye MT, Akinmoladun AC, Ogunboye AA, Akindahunsi AA. Antioxidant activity and hepatoprotective property of leaf extracts of *Boerhaavia diffusa* Linn against acetaminophen-induced liver damage in rats. *Food Chem Toxicol*. 48:2200–5. [PubMed: 20553784]
382. Sreeja S, Sreeja S. An in vitro study on antiproliferative and antiestrogenic effects of *Boerhaavia diffusa* L. extracts. *J Ethnopharmacol*. 2009; 126:221–5.
383. Manu KA, Kuttan G. *Boerhaavia diffusa* stimulates cell-mediated immune response by upregulating IL-2 and downregulating the pro-inflammatory cytokines and GM-CSF in B16F-10 metastatic melanoma bearing mice. *J Exp Ther Oncol*. 2008; 7:17–29. [PubMed: 18472639]
384. Kaur M, Goel RK. Anti-convulsant Activity of *Boerhaavia diffusa*: Plausible Role of Calcium Channel Antagonism. *Evid Based Complement Alternat Med*. 2009
385. Pari L, Amarnath Satheesh M. Antidiabetic effect of *Boerhaavia diffusa*: effect on serum and tissue lipids in experimental diabetes. *J Med Food*. 2004; 7:472–6. [PubMed: 15671692]
386. Pari L, Amarnath Satheesh M. Antidiabetic activity of *Boerhaavia diffusa* L: effect on hepatic key enzymes in experimental diabetes. *J Ethnopharmacol*. 2004; 91:109–13. [PubMed: 15036478]
387. Leyon PV, Lini CC, Kuttan G. Inhibitory effect of *Boerhaavia diffusa* on experimental metastasis by B16F10 melanoma in C57BL/6 mice. *Life Sci*. 2005; 76:1339–49. [PubMed: 15670614]
388. Bharali R, Azad MR, Tabassum J. Chemopreventive action of *Boerhaavia diffusa* on DMBA-induced skin carcinogenesis in mice. *Indian J Physiol Pharmacol*. 2003; 47:459–64. [PubMed: 15266960]
389. Manu KA, Kuttan G. Punarnavine induces apoptosis in B16F-10 melanoma cells by inhibiting NF-kappaB signaling. *Asian Pac J Cancer Prev*. 2009; 10:1031–7. [PubMed: 20192578]
390. Manu KA, Kuttan G. Anti-metastatic potential of Punarnavine, an alkaloid from *Boerhaavia diffusa* Linn. *Immunobiology*. 2009; 214:245–55. [PubMed: 19171408]
391. Pandey R, Maurya R, Singh G, Sathiamoorthy B, Naik S. Immunosuppressive properties of flavonoids isolated from *Boerhaavia diffusa* Linn. *Int Immunopharmacol*. 2005; 5:541–53. [PubMed: 15683850]
392. Gayathri B, Manjula N, Vinaykumar KS, Lakshmi BS, Balakrishnan A. Pure compound from *Boswellia serrata* extract exhibits anti-inflammatory property in human PBMCs and mouse macrophages through inhibition of TNFalpha, IL-1beta, NO and MAP kinases. *Int Immunopharmacol*. 2007; 7:473–82. [PubMed: 17321470]
393. Pang X, Yi Z, Zhang X, Sung B, Qu W, Lian X, et al. Acetyl-11-keto-beta-boswellic acid inhibits prostate tumor growth by suppressing vascular endothelial growth factor receptor 2-mediated angiogenesis. *Cancer Res*. 2009; 69:5893–900. [PubMed: 19567671]
394. Sharma ML, Bani S, Singh GB. Anti-arthritis activity of boswellic acids in bovine serum albumin (BSA)-induced arthritis. *Int J Immunopharmacol*. 1989; 11:647–52. [PubMed: 2807636]
395. Madisch A, Miehlke S, Eichele O, Mrwa J, Bethke B, Kuhlisch E, et al. *Boswellia serrata* extract for the treatment of collagenous colitis. A double-blind, randomized, placebo-controlled, multicenter trial. *Int J Colorectal Dis*. 2007; 22:1445–51. [PubMed: 17764013]
396. Cuaz-Perolin C, Billiet L, Bauge E, Copin C, Scott-Algara D, Genze F, et al. Antiinflammatory and antiatherogenic effects of the NF-kappaB inhibitor acetyl-11-keto-beta-boswellic acid in LPS-challenged ApoE^{-/-} mice. *Arterioscler Thromb Vasc Biol*. 2008; 28:272–7. [PubMed: 18032778]
397. Singh S, Khajuria A, Taneja SC, Khajuria RK, Singh J, Johri RK, et al. The gastric ulcer protective effect of boswellic acids, a leukotriene inhibitor from *Boswellia serrata*, in rats. *Phytomedicine*. 2008; 15:408–15. [PubMed: 18424019]
398. Sharma R, Singh S, Singh GD, Khajuria A, Sidiq T, Singh SK, et al. In vivo genotoxicity evaluation of a plant based antiarthritic and anticancer therapeutic agent Boswellic acids in rodents. *Phytomedicine*. 2009; 16:1112–8. [PubMed: 19679457]

399. Rasheed Z, Akhtar N, Khan A, Khan KA, Haqqi TM. Butrin, isobutrin, and butein from medicinal plant *Butea monosperma* selectively inhibit nuclear factor-kappaB in activated human mast cells: suppression of tumor necrosis factor-alpha, interleukin (IL)-6, and IL-8. *J Pharmacol Exp Ther.* 333:354–63. [PubMed: 20164300]
400. Panda S, Jafri M, Kar A, Meheta BK. Thyroid inhibitory, antiperoxidative and hypoglycemic effects of stigmasterol isolated from *Butea monosperma*. *Fitoterapia.* 2009; 80:123–6. [PubMed: 19105977]
401. Choedon T, Shukla SK, Kumar V. Chemopreventive and anti-cancer properties of the aqueous extract of flowers of *Butea monosperma*. *J Ethnopharmacol.* 129:208–13. [PubMed: 20307637]
402. Sharma N, Garg V. Antidiabetic and antioxidant potential of ethanolic extract of *Butea monosperma* leaves in alloxan-induced diabetic mice. *Indian J Biochem Biophys.* 2009; 46:99–105. [PubMed: 19374261]
403. Sharma N, Shukla S. Hepatoprotective potential of aqueous extract of *Butea monosperma* against CCl₄ induced damage in rats. *Exp Toxicol Pathol.*
404. Shahavi VM, Desai SK. Anti-inflammatory activity of *Butea monosperma* flowers. *Fitoterapia.* 2008; 79:82–5. [PubMed: 17904309]
405. Sehrawat A, Sultana S. Chemoprevention by *Butea monosperma* of hepatic carcinogenesis and oxidative damage in male wistar rats. *Asian Pac J Cancer Prev.* 2006; 7:140–8. [PubMed: 16629533]
406. Sumitra M, Manikandan P, Suguna L. Efficacy of *Butea monosperma* on dermal wound healing in rats. *Int J Biochem Cell Biol.* 2005; 37:566–73. [PubMed: 15618014]
407. Kasture VS, Kasture SB, Chopde CT. Anticonvulsive activity of *Butea monosperma* flowers in laboratory animals. *Pharmacol Biochem Behav.* 2002; 72:965–72. [PubMed: 12062587]
408. Shukla S, Mehta A, Mehta P, Vyas SP, Shukla S, Bajpai VK. Studies on anti-inflammatory, antipyretic and analgesic properties of *Caesalpinia bonducella* F. seed oil in experimental animal models. *Food Chem Toxicol.* 48:61–4. [PubMed: 19766160]
409. Chakrabarti S, Biswas TK, Seal T, Rokeya B, Ali L, Azad Khan AK, et al. Antidiabetic activity of *Caesalpinia bonducella* F. in chronic type 2 diabetic model in Long-Evans rats and evaluation of insulin secretagogue property of its fractions on isolated islets. *J Ethnopharmacol.* 2005; 97:117–22. [PubMed: 15652285]
410. Gupta M, Mazumder UK, Kumar RS, Sivakumar T, Vamsi ML. Antitumor activity and antioxidant status of *Caesalpinia bonducella* against Ehrlich ascites carcinoma in Swiss albino mice. *J Pharmacol Sci.* 2004; 94:177–84. [PubMed: 14978356]
411. Mathur R, Gupta SK, Mathur SR, Velpandian T. Anti-tumor studies with extracts of *Calotropis procera* (Ait.) R.Br. root employing Hep2 cells and their possible mechanism of action. *Indian J Exp Biol.* 2009; 47:343–8. [PubMed: 19579799]
412. Bharti S, Wahane VD, Kumar VL. Protective effect of *Calotropis procera* latex extracts on experimentally induced gastric ulcers in rat. *J Ethnopharmacol.* 127:440–4. [PubMed: 19853030]
413. Ramachandra Setty S, Quereshi AA, Viswanath Swamy AH, Patil T, Prakash T, Prabhu K, et al. Hepatoprotective activity of *Calotropis procera* flowers against paracetamol-induced hepatic injury in rats. *Fitoterapia.* 2007; 78:451–4. [PubMed: 17600635]
414. Kumar VL, Roy S. *Calotropis procera* latex extract affords protection against inflammation and oxidative stress in Freund's complete adjuvant-induced monoarthritis in rats. *Mediators Inflamm.* 2007; 2007:47523. [PubMed: 17497032]
415. Kumar VL, Sehgal R. *Calotropis procera* latex-induced inflammatory hyperalgesia - effect of bradyzide and morphine. *Auton Autacoid Pharmacol.* 2007; 27:143–9. [PubMed: 17584444]
416. Lam SK, Ng TB. A protein with antiproliferative, antifungal and HIV-1 reverse transcriptase inhibitory activities from caper (*Capparis spinosa*) seeds. *Phytomedicine.* 2009; 16:444–50. [PubMed: 19019643]
417. Gilani AH, Jabeen Q, Ghayur MN, Janbaz KH, Akhtar MS. Studies on the antihypertensive, antispasmodic, bronchodilator and hepatoprotective activities of the *Carum copticum* seed extract. *J Ethnopharmacol.* 2005; 98:127–35. [PubMed: 15763373]

418. Deeptha K, Kamaleeswari M, Sengottuvelan M, Nalini N. Dose dependent inhibitory effect of dietary caraway on 1,2-dimethylhydrazine induced colonic aberrant crypt foci and bacterial enzyme activity in rats. *Invest New Drugs*. 2006; 24:479–88. [PubMed: 16598436]
419. Dashti-Rahmatabadi MH, Hejazian SH, Morshedi A, Rafati A. The analgesic effect of *Carum copticum* extract and morphine on phasic pain in mice. *J Ethnopharmacol*. 2007; 109:226–8. [PubMed: 17005345]
420. Chandramohan G, Al-Numair KS, Sridevi M, Pugalendi KV. Antihyperlipidemic activity of 3-hydroxymethyl xylitol, a novel antidiabetic compound isolated from *Casearia esculenta* (Roxb.) root, in streptozotocin-diabetic rats. *J Biochem Mol Toxicol*. 24:95–101. [PubMed: 20146230]
421. Prakasam A, Sethupathy S, Pugalendi KV. Antiperoxidative and antioxidant effects of *Casearia esculenta* root extract in streptozotocin-induced diabetic rats. *Yale J Biol Med*. 2005; 78:15–23. [PubMed: 16197726]
422. Gupta M, Mazumder UK, Rath N, Mukhopadhyay DK. Antitumor activity of methanolic extract of *Cassia fistula* L. seed against Ehrlich ascites carcinoma. *J Ethnopharmacol*. 2000; 72:151–6. [PubMed: 10967466]
423. el-Saadany SS, el-Massry RA, Labib SM, Sitohy MZ. The biochemical role and hypocholesterolaemic potential of the legume *Cassia fistula* in hypercholesterolaemic rats. *Nahrung*. 1991; 35:807–15. [PubMed: 1780005]
424. Pradeep K, Mohan CV, Gobianand K, Karthikeyan S. Effect of *Cassia fistula* Linn. leaf extract on diethylnitrosamine induced hepatic injury in rats. *Chem Biol Interact*. 2007; 167:12–8. [PubMed: 17289008]
425. Sharma N, Trikha P, Athar M, Raisuddin S. In vitro inhibition of carcinogen-induced mutagenicity by *Cassia occidentalis* and *Emblica officinalis*. *Drug Chem Toxicol*. 2000; 23:477–84. [PubMed: 10959548]
426. Jafri MA, Jalis Subhani M, Javed K, Singh S. Hepatoprotective activity of leaves of *Cassia occidentalis* against paracetamol and ethyl alcohol intoxication in rats. *J Ethnopharmacol*. 1999; 66:355–61. [PubMed: 10473185]
427. Bin-Hafeez B, Ahmad I, Haque R, Raisuddin S. Protective effect of *Cassia occidentalis* L. on cyclophosphamide-induced suppression of humoral immunity in mice. *J Ethnopharmacol*. 2001; 75:13–8. [PubMed: 11282437]
428. Yadav JP, Arya V, Yadav S, Panghal M, Kumar S, Dhankhar S. *Cassia occidentalis* L.: a review on its ethnobotany, phytochemical and pharmacological profile. *Fitoterapia*. 81:223–30. [PubMed: 19796670]
429. Dhanasekaran M, Ignacimuthu S, Agastian P. Potential hepatoprotective activity of ononitol monohydrate isolated from *Cassia tora* L. on carbon tetrachloride induced hepatotoxicity in wistar rats. *Phytomedicine*. 2009; 16:891–5. [PubMed: 19345078]
430. Rejiya CS, Cibir TR, Abraham A. Leaves of *Cassia tora* as a novel cancer therapeutic--an in vitro study. *Toxicol In Vitro*. 2009; 23:1034–8. [PubMed: 19540331]
431. Lim SHHK. Hypoglycemic effect of fractions of *Cassia tora* extract in Streptozotocin-induced diabetic Rats. *Journal of the Korean Society of Food Science and Nutrition*. 1997; 13:23–9.
432. Cho IJ, Lee C, Ha TY. Hypolipidemic effect of soluble fiber isolated from seeds of *Cassia tora* Linn. in rats fed a high-cholesterol diet. *J Agric Food Chem*. 2007; 55:1592–6. [PubMed: 17300158]
433. Nam J, Choi H. Effect of butanol fraction from *Cassia tora* L. seeds on glycemic control and insulin secretion in diabetic rats. *Nutr Res Pract*. 2008; 2:240–6. [PubMed: 20016725]
434. Singh SK, Shanmugavel M, Kampasi H, Singh R, Mondhe DM, Rao JM, et al. Chemically standardized isolates from *Cedrus deodara* stem wood having anticancer activity. *Planta Med*. 2007; 73:519–26. [PubMed: 17534788]
435. Shinde UA, Kulkarni KR, Phadke AS, Nair AM, Mungantiwar AA, Dikshit VJ, et al. Mast cell stabilizing and lipoxygenase inhibitory activity of *Cedrus deodara* (Roxb.) Loud. wood oil. *Indian J Exp Biol*. 1999; 37:258–61. [PubMed: 10641156]
436. Godkar PB, Gordon RK, Ravindran A, Doctor BP. *Celastrus paniculatus* seed oil and organic extracts attenuate hydrogen peroxide- and glutamate-induced injury in embryonic rat forebrain neuronal cells. *Phytomedicine*. 2006; 13:29–36. [PubMed: 16360930]

437. Kumar MH, Gupta YK. Antioxidant property of *Celastrus paniculatus* willd : a possible mechanism in enhancing cognition. *Phytomedicine*. 2002; 9:302–11. [PubMed: 12120811]
438. Lee KT, Kim JI, Park HJ, Yoo KO, Han YN, Miyamoto K. Differentiation-inducing effect of magnolialide, a 1 beta-hydroxyeudesmanolide isolated from *Cichorium intybus*, on human leukemia cells. *Biol Pharm Bull*. 2000; 23:1005–7. [PubMed: 10963313]
439. Ahmed B, Khan S, Masood MH, Siddique AH. Anti-hepatotoxic activity of cichotyboside, a sesquiterpene glycoside from the seeds of *Cichorium intybus*. *J Asian Nat Prod Res*. 2008; 10:223–31. [PubMed: 18335337]
440. Kim M. The water-soluble extract of chicory reduces cholesterol uptake in gut-perfused rats. *Nutrition Research*. 2000; 20:1017–26.
441. Pushparaj PN, Low HK, Manikandan J, Tan BK, Tan CH. Anti-diabetic effects of *Cichorium intybus* in streptozotocin-induced diabetic rats. *J Ethnopharmacol*. 2007; 111:430–4. [PubMed: 17197141]
442. Koppikar SJ, Choudhari AS, Suryavanshi SA, Kumari S, Chattopadhyay S, Kaul-Ghanekar R. Aqueous cinnamon extract (ACE-c) from the bark of *Cinnamomum cassia* causes apoptosis in human cervical cancer cell line (SiHa) through loss of mitochondrial membrane potential. *BMC Cancer*. 10:210. [PubMed: 20482751]
443. Lim CS, Kim EY, Lee HS, Soh Y, Sohn Y, Kim SY, et al. Protective effects of *Cinnamomum cassia* Blume in the fibrogenesis of activated HSC-T6 cells and dimethylnitrosamine-induced acute liver injury in SD rats. *Biosci Biotechnol Biochem*. 74:477–83. [PubMed: 20208363]
444. Kirkham S, Akilen R, Sharma S, Tsiami A. The potential of cinnamon to reduce blood glucose levels in patients with type 2 diabetes and insulin resistance. *Diabetes Obes Metab*. 2009; 11:1100–13. [PubMed: 19930003]
445. Dugoua JJ, Seely D, Perri D, Cooley K, Forelli T, Mills E, et al. From type 2 diabetes to antioxidant activity: a systematic review of the safety and efficacy of common and cassia cinnamon bark. *Can J Physiol Pharmacol*. 2007; 85:837–47. [PubMed: 18066129]
446. Lee CW, Lee SH, Lee JW, Ban JO, Lee SY, Yoo HS, et al. 2-hydroxycinnamaldehyde inhibits SW620 colon cancer cell growth through AP-1 inactivation. *J Pharmacol Sci*. 2007; 104:19–28. [PubMed: 17510524]
447. Ng LT, Wu SJ. Antiproliferative Activity of *Cinnamomum cassia* Constituents and Effects of Pifithrin-Alpha on Their Apoptotic Signaling Pathways in Hep G2 Cells. *Evid Based Complement Alternat Med*. 2009
448. Ka H, Park HJ, Jung HJ, Choi JW, Cho KS, Ha J, et al. Cinnamaldehyde induces apoptosis by ROS-mediated mitochondrial permeability transition in human promyelocytic leukemia HL-60 cells. *Cancer Lett*. 2003; 196:143–52. [PubMed: 12860272]
449. Cao H, Urban JF Jr, Anderson RA. Cinnamon polyphenol extract affects immune responses by regulating anti- and proinflammatory and glucose transporter gene expression in mouse macrophages. *J Nutr*. 2008; 138:833–40. [PubMed: 18424588]
450. Kwon HK, Jeon WK, Hwang JS, Lee CG, So JS, Park JA, et al. Cinnamon extract suppresses tumor progression by modulating angiogenesis and the effector function of CD8+ T cells. *Cancer Lett*. 2009; 278:174–82. [PubMed: 19203831]
451. Qin B, Dawson H, Polansky MM, Anderson RA. Cinnamon extract attenuates TNF-alpha-induced intestinal lipoprotein ApoB48 overproduction by regulating inflammatory, insulin, and lipoprotein pathways in enterocytes. *Horm Metab Res*. 2009; 41:516–22. [PubMed: 19593846]
452. Kanuri G, Weber S, Volynets V, Spruss A, Bischoff SC, Bergheim I. Cinnamon extract protects against acute alcohol-induced liver steatosis in mice. *J Nutr*. 2009; 139:482–7. [PubMed: 19126670]
453. Moselhy SS, Ali HK. Hepatoprotective effect of cinnamon extracts against carbon tetrachloride induced oxidative stress and liver injury in rats. *Biol Res*. 2009; 42:93–8. [PubMed: 19621136]
454. Abdel-Hassan IA, Abdel-Barry JA, Tariq Mohammeda S. The hypoglycaemic and antihyperglycaemic effect of *Citrullus colocynthis* fruit aqueous extract in normal and alloxan diabetic rabbits. *J Ethnopharmacol*. 2000; 71:325–30. [PubMed: 10904181]
455. Daradka H, Almasad MM, Qazan W, El-Banna NM, Samara OH. Hypolipidaemic effects of *Citrullus colocynthis* L. in rabbits. *Pak J Biol Sci*. 2007; 10:2768–71. [PubMed: 19070101]

456. Huseini HF, Darvishzadeh F, Heshmat R, Jafariazar Z, Raza M, Larijani B. The clinical investigation of *Citrullus colocynthis* (L) schrad fruit in treatment of Type II diabetic patients: a randomized, double blind, placebo-controlled clinical trial. *Phytother Res.* 2009; 23:1186–9. [PubMed: 19170143]
457. Marzouk B, Marzouk Z, Haloui E, Fenina N, Bouraoui A, Aouni M. Screening of analgesic and anti-inflammatory activities of *Citrullus colocynthis* from southern Tunisia. *J Ethnopharmacol.* 128:15–9. [PubMed: 19962436]
458. Manjula N, Gayathri B, Vinaykumar KS, Shankernarayanan NP, Vishwakarma RA, Balakrishnan A. Inhibition of MAP kinases by crude extract and pure compound isolated from *Commiphora mukul* leads to down regulation of TNF-alpha, IL-1beta and IL-2. *Int Immunopharmacol.* 2006; 6:122–32. [PubMed: 16399617]
459. Shishodia S, Sethi G, Ahn KS, Aggarwal BB. Guggulsterone inhibits tumor cell proliferation, induces S-phase arrest, and promotes apoptosis through activation of c-Jun N-terminal kinase, suppression of Akt pathway, and downregulation of antiapoptotic gene products. *Biochem Pharmacol.* 2007; 74:118–30. [PubMed: 17475222]
460. Mencarelli A, Renga B, Palladino G, Distrutti E, Fiorucci S. The plant sterol guggulsterone attenuates inflammation and immune dysfunction in murine models of inflammatory bowel disease. *Biochem Pharmacol.* 2009; 78:1214–23. [PubMed: 19555671]
461. Sharma B, Salunke R, Srivastava S, Majumder C, Roy P. Effects of guggulsterone isolated from *Commiphora mukul* in high fat diet induced diabetic rats. *Food Chem Toxicol.* 2009; 47:2631–9. [PubMed: 19635521]
462. Nohr LA, Rasmussen LB, Straand J. Resin from the mukul myrrh tree, guggul, can it be used for treating hypercholesterolemia? A randomized, controlled study. *Complement Ther Med.* 2009; 17:16–22. [PubMed: 19114224]
463. Yu BZ, Kaimal R, Bai S, El Sayed KA, Tatulian SA, Apitz RJ, et al. Effect of guggulsterone and cembranoids of *Commiphora mukul* on pancreatic phospholipase A(2): role in hypocholesterolemia. *J Nat Prod.* 2009; 72:24–8. [PubMed: 19102680]
464. Panda S, Kar A. Guggulu (*Commiphora mukul*) potentially ameliorates hypothyroidism in female mice. *Phytother Res.* 2005; 19:78–80. [PubMed: 15798994]
465. Deng R. Therapeutic effects of guggul and its constituent guggulsterone: cardiovascular benefits. *Cardiovasc Drug Rev.* 2007; 25:375–90. [PubMed: 18078436]
466. Shishodia S, Harikumar KB, Dass S, Ramawat KG, Aggarwal BB. The guggul for chronic diseases: ancient medicine, modern targets. *Anticancer Res.* 2008; 28:3647–64. [PubMed: 19189646]
467. Agarwal RC, Singh SP, Saran RK, Das SK, Sinha N, Asthana OP, et al. Clinical trial of guggulipid--a new hypolipidemic agent of plant origin in primary hyperlipidemia. *Indian J Med Res.* 1986; 84:626–34. [PubMed: 3552974]
468. Szapary PO, Wolfe ML, Bloedon LT, Cucchiara AJ, DerMarderosian AH, Cirigliano MD, et al. Guggulipid for the treatment of hypercholesterolemia: a randomized controlled trial. *JAMA.* 2003; 290:765–72. [PubMed: 12915429]
469. Tripathi YB, Reddy MM, Pandey RS, Subhashini J, Tiwari OP, Singh BK, et al. Anti-inflammatory properties of BHUx, a polyherbal formulation to prevent atherosclerosis. *Inflammopharmacology.* 2004; 12:131–52. [PubMed: 15265316]
470. Chauhan CK, Joshi MJ, Vaidya AD. Growth inhibition of struvite crystals in the presence of herbal extract *Commiphora wightii*. *J Mater Sci Mater Med.* 2009; 20 (Suppl 1):S85–92. [PubMed: 18568390]
471. Bihagi SW, Sharma M, Singh AP, Tiwari M. Neuroprotective role of *Convolvulus pluricaulis* on aluminium induced neurotoxicity in rat brain. *J Ethnopharmacol.* 2009; 124:409–15. [PubMed: 19505562]
472. Dhingra D, Valecha R. Evaluation of the antidepressant-like activity of *Convolvulus pluricaulis choisy* in the mouse forced swim and tail suspension tests. *Med Sci Monit.* 2007; 13:BR155–61. [PubMed: 17599020]
473. Sairam K, Rao CV, Goel RK. Effect of *Convolvulus pluricaulis Choisy* on gastric ulceration and secretion in rats. *Indian J Exp Biol.* 2001; 39:350–4. [PubMed: 11491580]

474. Aung HH, Wang CZ, Ni M, Fishbein A, Mehendale SR, Xie JT, et al. Crocin from *Crocus sativus* possesses significant anti-proliferation effects on human colorectal cancer cells. *Exp Oncol.* 2007; 29:175–80. [PubMed: 18004240]
475. Imenshahidi M, Hosseinzadeh H, Javadpour Y. Hypotensive effect of aqueous saffron extract (*Crocus sativus* L.) and its constituents, safranal and crocin, in normotensive and hypertensive rats. *Phytother Res.* 24:990–4. [PubMed: 20013822]
476. Asdaq SM, Inamdar MN. Potential of *Crocus sativus* (saffron) and its constituent, crocin, as hypolipidemic and antioxidant in rats. *Appl Biochem Biotechnol.* 162:358–72. [PubMed: 19672721]
477. Xu GL, Li G, Ma HP, Zhong H, Liu F, Ao GZ. Preventive effect of crocin in inflamed animals and in LPS-challenged RAW 264. 7 cells. *J Agric Food Chem.* 2009; 57:8325–30. [PubMed: 19754168]
478. Hosseinzadeh H, Modaghegh MH, Saffari Z. *Crocus sativus* L. (Saffron) extract and its active constituents (crocin and safranal) on ischemia-reperfusion in rat skeletal muscle. *Evid Based Complement Alternat Med.* 2009; 6:343–50. [PubMed: 18955256]
479. Schmidt M, Betti G, Hensel A. Saffron in phytotherapy: pharmacology and clinical uses. *Wien Med Wochenschr.* 2007; 157:315–9. [PubMed: 17704979]
480. Abdullaev FI, Espinosa-Aguirre JJ. Biomedical properties of saffron and its potential use in cancer therapy and chemoprevention trials. *Cancer Detect Prev.* 2004; 28:426–32. [PubMed: 15582266]
481. Nair SC, Kurumboor SK, Hasegawa JH. Saffron chemoprevention in biology and medicine: a review. *Cancer Biother.* 1995; 10:257–64. [PubMed: 8590890]
482. Tavakkol-Afshari J, Brook A, Mousavi SH. Study of cytotoxic and apoptogenic properties of saffron extract in human cancer cell lines. *Food Chem Toxicol.* 2008; 46:3443–7. [PubMed: 18790714]
483. Ochiai T, Shimeno H, Mishima K, Iwasaki K, Fujiwara M, Tanaka H, et al. Protective effects of carotenoids from saffron on neuronal injury in vitro and in vivo. *Biochim Biophys Acta.* 2007; 1770:578–84. [PubMed: 17215084]
484. Akhondzadeh S, Shafiee Sabet M, Harirchian MH, Togha M, Cheraghmakani H, Razeghi S, et al. A 22-week, multicenter, randomized, double-blind controlled trial of *Crocus sativus* in the treatment of mild-to-moderate Alzheimer's disease. *Psychopharmacology (Berl).* 207:637–43. [PubMed: 19838862]
485. Wang Y, Han T, Zhu Y, Zheng CJ, Ming QL, Rahman K, et al. Antidepressant properties of bioactive fractions from the extract of *Crocus sativus* L. *J Nat Med.* 64:24–30. [PubMed: 19787421]
486. Sambaiah K, Srinivasan K. Effect of cumin, cinnamon, ginger, mustard and tamarind in induced hypercholesterolemic rats. *Nahrung.* 1991; 35:47–51. [PubMed: 1865890]
487. Gagandeep, Dhanalakshmi S, Mendiz E, Rao AR, Kale RK. Chemopreventive effects of *Cuminum cyminum* in chemically induced forestomach and uterine cervix tumors in murine model systems. *Nutr Cancer.* 2003; 47:171–80. [PubMed: 15087270]
488. Janahmadi M, Niazi F, Danyali S, Kamalinejad M. Effects of the fruit essential oil of *Cuminum cyminum* Linn. (Apiaceae) on pentylenetetrazol-induced epileptiform activity in F1 neurones of *Helix aspersa*. *J Ethnopharmacol.* 2006; 104:278–82. [PubMed: 16226415]
489. Shirke SS, Jagtap AG. Effects of methanolic extract of *Cuminum cyminum* on total serum cholesterol in ovariectomized rats. *Indian J Pharmacol.* 2009; 41:92–3. [PubMed: 20336228]
490. Jagtap AG, Patil PB. Antihyperglycemic activity and inhibition of advanced glycation end product formation by *Cuminum cyminum* in streptozotocin induced diabetic rats. *Food Chem Toxicol.* 48:2030–6. [PubMed: 20451573]
491. Jatoi SA, Kikuchi A, Gilani SA, Watanabe KN. Phytochemical, pharmacological and ethnobotanical studies in mango ginger (*Curcuma amada* Roxb; Zingiberaceae). *Phytother Res.* 2007; 21:507–16. [PubMed: 17397131]
492. Arafa HM. Uroprotective effects of curcumin in cyclophosphamide-induced haemorrhagic cystitis paradigm. *Basic Clin Pharmacol Toxicol.* 2009; 104:393–9. [PubMed: 19413659]

493. Mandal MN, Patlolla JM, Zheng L, Agbaga MP, Tran JT, Wicker L, et al. Curcumin protects retinal cells from light-and oxidant stress-induced cell death. *Free Radic Biol Med*. 2009; 46:672–9. [PubMed: 19121385]
494. Martelli L, Ragazzi E, di Mario F, Martelli M, Castagliuolo I, Dal Maschio M, et al. A potential role for the vanilloid receptor TRPV1 in the therapeutic effect of curcumin in dinitrobenzene sulphonic acid-induced colitis in mice. *Neurogastroenterol Motil*. 2007; 19:668–74. [PubMed: 17640182]
495. Huang MT, Lysz T, Ferraro T, Abidi TF, Laskin JD, Conney AH. Inhibitory effects of curcumin on in vitro lipoxygenase and cyclooxygenase activities in mouse epidermis. *Cancer Res*. 1991; 51:813–9. [PubMed: 1899046]
496. Pari L, Tewas D, Eckel J. Role of curcumin in health and disease. *Arch Physiol Biochem*. 2008; 114:127–49. [PubMed: 18484280]
497. Yoshioka T, Fujii E, Endo M, Wada K, Tokunaga Y, Shiba N, et al. Antiinflammatory potency of dehydrocurdione, a zedoary-derived sesquiterpene. *Inflamm Res*. 1998; 47:476–81. [PubMed: 9892041]
498. Oh OJ, Min HY, Lee SK. Inhibition of inducible prostaglandin E2 production and cyclooxygenase-2 expression by curdione from *Curcuma zedoaria*. *Archives of pharmacol research*. 2007; 30:1236–9. [PubMed: 18038902]
499. Seo WG, Hwang JC, Kang SK, Jin UH, Suh SJ, Moon SK, et al. Suppressive effect of *Zedoariae rhizoma* on pulmonary metastasis of B16 melanoma cells. *Journal of ethnopharmacology*. 2005; 101:249–57. [PubMed: 16023317]
500. Makabe H, Maru N, Kuwabara A, Kamo T, Hirota M. Anti-inflammatory sesquiterpenes from *Curcuma zedoaria*. *Natural product research*. 2006; 20:680–5. [PubMed: 16901812]
501. Matsuda H, Tewtrakul S, Morikawa T, Nakamura A, Yoshikawa M. Anti-allergic principles from Thai zedoary: structural requirements of curcuminoids for inhibition of degranulation and effect on the release of TNF-alpha and IL-4 in RBL-2H3 cells. *Bioorganic & medicinal chemistry*. 2004; 12:5891–8. [PubMed: 15498665]
502. Matsuda H, Ninomiya K, Morikawa T, Yoshikawa M. Inhibitory effect and action mechanism of sesquiterpenes from *Zedoariae Rhizoma* on D-galactosamine/lipopolysaccharide-induced liver injury. *Bioorganic & medicinal chemistry letters*. 1998; 8:339–44. [PubMed: 9871681]
503. Kim KI, Kim JW, Hong BS, Shin DH, Cho HY, Kim HK, et al. Antitumor, genotoxicity and anticlastogenic activities of polysaccharide from *Curcuma zedoaria*. *Molecules and cells*. 2000; 10:392–8. [PubMed: 10987135]
504. Katsukawa M, Nakata R, Takizawa Y, Hori K, Takahashi S, Inoue H. Citral, a component of lemongrass oil, activates PPARalpha and gamma and suppresses COX-2 expression. *Biochim Biophys Acta*.
505. Lee HJ, Jeong HS, Kim DJ, Noh YH, Yuk DY, Hong JT. Inhibitory effect of citral on NO production by suppression of iNOS expression and NF-kappa B activation in RAW264. 7 cells. *Arch Pharm Res*. 2008; 31:342–9. [PubMed: 18409048]
506. Figueirinha A, Cruz MT, Francisco V, Lopes MC, Batista MT. Anti-inflammatory activity of *Cymbopogon citratus* leaf infusion in lipopolysaccharide-stimulated dendritic cells: contribution of the polyphenols. *J Med Food*. 13:681–90. [PubMed: 20438326]
507. Carbajal D, Casaco A, Arruzazabala L, Gonzalez R, Tolon Z. Pharmacological study of *Cymbopogon citratus* leaves. *J Ethnopharmacol*. 1989; 25:103–7. [PubMed: 2716341]
508. Sforzin JM, Amaral JT, Fernandes A Jr, Sousa JP, Bastos JK. Lemongrass effects on IL-1beta and IL-6 production by macrophages. *Nat Prod Res*. 2009; 23:1151–9. [PubMed: 19662581]
509. Silva MR, Ximenes RM, da Costa JG, Leal LK, de Lopes AA, Viana GS. Comparative anticonvulsant activities of the essential oils (EOs) from *Cymbopogon winterianus* Jowitt and *Cymbopogon citratus* (DC) Stapf. in mice. *Naunyn Schmiedeberg Arch Pharmacol*. 381:415–26. [PubMed: 20237771]
510. Kilani S, Ben Sghaier M, Limem I, Bouhlel I, Boubaker J, Bhouri W, et al. In vitro evaluation of antibacterial, antioxidant, cytotoxic and apoptotic activities of the tubers infusion and extracts of *Cyperus rotundus*. *Bioresource technology*. 2008; 99:9004–8. [PubMed: 18538563]

511. Kilani-Jaziri S, Neffati A, Limem I, Boubaker J, Skandrani I, Sghair MB, et al. Relationship correlation of antioxidant and antiproliferative capacity of *Cyperus rotundus* products towards K562 erythroleukemia cells. *Chemico-biological interactions*. 2009; 181:85–94. [PubMed: 19446539]
512. Raut NA, Gaikwad NJ. Antidiabetic activity of hydro-ethanolic extract of *Cyperus rotundus* in alloxan induced diabetes in rats. *Fitoterapia*. 2006; 77:585–8. [PubMed: 17056202]
513. Muthu AK, Sethupathy S, Manavalan R, Karar PK. Hypolipidemic effect of methanolic extract of *Dolichos biflorus* Linn. in high fat diet fed rats. *Indian journal of experimental biology*. 2005; 43:522–5. [PubMed: 15991577]
514. Bijarnia RK, Kaur T, Singla SK, Tandon C. A novel calcium oxalate crystal growth inhibitory protein from the seeds of *Dolichos biflorus* (L). *The protein journal*. 2009; 28:161–8. [PubMed: 19488841]
515. Singh B, Saxena AK, Chandan BK, Agarwal SG, Anand KK. In vivo hepatoprotective activity of active fraction from ethanolic extract of *Eclipta alba* leaves. *Indian journal of physiology and pharmacology*. 2001; 45:435–41. [PubMed: 11883149]
516. Jayathirtha MG, Mishra SH. Preliminary immunomodulatory activities of methanol extracts of *Eclipta alba* and *Centella asiatica*. *Phytomedicine*. 2004; 11:361–5. [PubMed: 15185851]
517. Ananthi J, Prakasam A, Pugalendi KV. Antihyperglycemic activity of *Eclipta alba* leaf on alloxan-induced diabetic rats. *The Yale journal of biology and medicine*. 2003; 76:97–102. [PubMed: 15369623]
518. Thakur VD, Mengi SA. Neuropharmacological profile of *Eclipta alba* (Linn). *Hassk Journal of ethnopharmacology*. 2005; 102:23–31.
519. Rangineni V, Sharada D, Saxena S. Diuretic, hypotensive, and hypocholesterolemic effects of *Eclipta alba* in mild hypertensive subjects: a pilot study. *Journal of medicinal food*. 2007; 10:143–8. [PubMed: 17472478]
520. Sengupta A, Ghosh S, Bhattacharjee S. Dietary cardamom inhibits the formation of azoxymethane-induced aberrant crypt foci in mice and reduces COX-2 and iNOS expression in the colon. *Asian Pac J Cancer Prev*. 2005; 6:118–22. [PubMed: 16101317]
521. Bhattacharjee S, Rana T, Sengupta A. Inhibition of lipid peroxidation and enhancement of GST activity by cardamom and cinnamon during chemically induced colon carcinogenesis in Swiss albino mice. *Asian Pac J Cancer Prev*. 2007; 8:578–82. [PubMed: 18260732]
522. Gilani AH, Jabeen Q, Khan AU, Shah AJ. Gut modulatory, blood pressure lowering, diuretic and sedative activities of cardamom. *Journal of ethnopharmacology*. 2008; 115:463–72. [PubMed: 18037596]
523. Xu M, Cui J, Fu H, Proksch P, Lin W, Li M. Embelin derivatives and their anticancer activity through microtubule disassembly. *Planta medica*. 2005; 71:944–8. [PubMed: 16254827]
524. Chitra M, Sukumar E, Suja V, Devi CS. Antitumor, anti-inflammatory and analgesic property of embelin, a plant product. *Chemotherapy*. 1994; 40:109–13. [PubMed: 7510605]
525. Sreepriya M, Bali G. Chemopreventive effects of embelin and curcumin against N-nitrosodiethylamine/phenobarbital-induced hepatocarcinogenesis in Wistar rats. *Fitoterapia*. 2005; 76:549–55. [PubMed: 16009505]
526. Singh D, Singh R, Singh P, Gupta RS. Effects of Embelin on Lipid Peroxidation and Free Radical Scavenging Activity against Liver Damage in Rats. *Basic & clinical pharmacology & toxicology*. 2009
527. Mahendran S, Thippeswamy BS, Veerapur VP, Badami S. Anticonvulsant activity of embelin isolated from *Embelia ribes*. *Phytomedicine*.
528. Bhandari U, Ansari MN. Antihyperglycaemic activity of aqueous extract of *Embelia ribes* Burm in streptozotocin-induced diabetic rats. *Indian journal of experimental biology*. 2008; 46:607–13. [PubMed: 18814490]
529. Bhandari U, Ansari MN, Islam F. Cardioprotective effect of aqueous extract of *Embelia ribes* Burm fruits against isoproterenol-induced myocardial infarction in albino rats. *Indian journal of experimental biology*. 2008; 46:35–40. [PubMed: 18697569]
530. Nazam Ansari M, Bhandari U, Islam F, Tripathi CD. Evaluation of antioxidant and neuroprotective effect of ethanolic extract of *Embelia ribes* Burm in focal cerebral ischemia/

- reperfusion-induced oxidative stress in rats. *Fundamental & clinical pharmacology*. 2008; 22:305–14. [PubMed: 18485149]
531. Nicolis E, Lampronti I, Dehecchi MC, Borgatti M, Tamanini A, Bianchi N, et al. Pyrogallol, an active compound from the medicinal plant *Emblica officinalis*, regulates expression of pro-inflammatory genes in bronchial epithelial cells. *International immunopharmacology*. 2008; 8:1672–80. [PubMed: 18760383]
 532. Yang CJ, Wang CS, Hung JY, Huang HW, Chia YC, Wang PH, et al. Pyrogallol induces G2-M arrest in human lung cancer cells and inhibits tumor growth in an animal model. *Lung cancer (Amsterdam, Netherlands)*. 2009; 66:162–8.
 533. Sultana S, Ahmed S, Jahangir T. *Emblica officinalis* and hepatocarcinogenesis: a chemopreventive study in Wistar rats. *Journal of ethnopharmacology*. 2008; 118:1–6. [PubMed: 18467048]
 534. Saini A, Sharma S, Chhibber S. Protective efficacy of *Emblica officinalis* against *Klebsiella pneumoniae* induced pneumonia in mice. *The Indian journal of medical research*. 2008; 128:188–93. [PubMed: 19001683]
 535. Kumar NP, Annamalai AR, Thakur RS. Antinociceptive property of *Emblica officinalis* Gaertn (Amla) in high fat diet-fed/low dose streptozotocin induced diabetic neuropathy in rats. *Indian journal of experimental biology*. 2009; 47:737–42. [PubMed: 19957886]
 536. Muthuraman A, Sood S, Singla SK. The antiinflammatory potential of phenolic compounds from *Emblica officinalis* L. in rat. *Inflammopharmacology*.
 537. Li L, Adams LS, Chen S, Killian C, Ahmed A, Seeram NP. *Eugenia jambolana* Lam. berry extract inhibits growth and induces apoptosis of human breast cancer but not non-tumorigenic breast cells. *Journal of agricultural and food chemistry*. 2009; 57:826–31. [PubMed: 19166352]
 538. Grover JK, Rathi SS, Vats V. Amelioration of experimental diabetic neuropathy and gastropathy in rats following oral administration of plant (*Eugenia jambolana*, *Mucuna pruriens* and *Tinospora cordifolia*) extracts. *Indian J Exp Biol*. 2002; 40:273–6. [PubMed: 12635695]
 539. Sharma SB, Nasir A, Prabhu KM, Murthy PS. Antihyperglycemic effect of the fruit-pulp of *Eugenia jambolana* in experimental diabetes mellitus. *Journal of ethnopharmacology*. 2006; 104:367–73. [PubMed: 16386863]
 540. Sharma B, Balomajumder C, Roy P. Hypoglycemic and hypolipidemic effects of flavonoid rich extract from *Eugenia jambolana* seeds on streptozotocin induced diabetic rats. *Food Chem Toxicol*. 2008; 46:2376–83. [PubMed: 18474411]
 541. Chaturvedi A, Bhawani G, Agarwal PK, Goel S, Singh A, Goel RK. Antidiabetic and antiulcer effects of extract of *Eugenia jambolana* seed in mild diabetic rats: study on gastric mucosal offensive acid-pepsin secretion. *Indian journal of physiology and pharmacology*. 2009; 53:137–46. [PubMed: 20112817]
 542. Tanwar RS, Sharma SB, Singh UR, Prabhu KM. Attenuation of renal dysfunction by anti-hyperglycemic compound isolated from fruit pulp of *Eugenia jambolana* in streptozotocin-induced diabetic rats. *Indian journal of biochemistry & biophysics*. 47:83–9.
 543. Siripurapu KB, Gupta P, Bhatia G, Maurya R, Nath C, Palit G. Adaptogenic and anti-amnesic properties of *Evolvulus alsinoides* in rodents. *Pharmacology, biochemistry, and behavior*. 2005; 81:424–32.
 544. Gupta P, Akanksha, Siripurapu KB, Ahmad A, Palit G, Arora A, et al. Anti-stress constituents of *Evolvulus alsinoides*: an ayurvedic crude drug. *Chemical & pharmaceutical bulletin*. 2007; 55:771–5.
 545. Nahata A, Patil UK, Dixit VK. Effect of *Evolvulus alsinoides* Linn. on learning behavior and memory enhancement activity in rodents. *Phytother Res*. 24:486–93. [PubMed: 19610035]
 546. Lee CL, Chiang LC, Cheng LH, Liaw CC, Abd El-Razek MH, Chang FR, et al. Influenza A (H1N1) Antiviral and Cytotoxic Agents from *Ferula assa-foetida*. *Journal of natural products*. 2009; 72:1568–72. [PubMed: 19691312]
 547. Mallikarjuna GU, Dhanalakshmi S, Raisuddin S, Rao AR. Chemomodulatory influence of *Ferula asafoetida* on mammary epithelial differentiation, hepatic drug metabolizing enzymes, antioxidant profiles and N-methyl-N-nitrosourea-induced mammary carcinogenesis in rats. *Breast cancer research and treatment*. 2003; 81:1–10. [PubMed: 14531492]

548. Fatehi M, Farifteh F, Fatehi-Hassanabad Z. Antispasmodic and hypotensive effects of *Ferula asafoetida* gum extract. *Journal of ethnopharmacology*. 2004; 91:321–4. [PubMed: 15120456]
549. Abu-Zaiton AS. Anti-diabetic activity of *Ferula assafoetida* extract in normal and alloxan-induced diabetic rats. *Pakistan journal of biological sciences: PJBS*. 13:97–100. [PubMed: 20415145]
550. Geetha BS, Mathew BC, Augusti KT. Hypoglycemic effects of leucodelphinidin derivative isolated from *Ficus bengalensis* (Linn). *Indian journal of physiology and pharmacology*. 1994; 38:220–2. [PubMed: 7814088]
551. Daniel RS, Devi KS, Augusti KT, Sudhakaran Nair CR. Mechanism of action of antiatherogenic and related effects of *Ficus bengalensis* Linn. flavonoids in experimental animals. *Indian journal of experimental biology*. 2003; 41:296–303. [PubMed: 15255637]
552. Shukla R, Gupta S, Gambhir JK, Prabhu KM, Murthy PS. Antioxidant effect of aqueous extract of the bark of *Ficus bengalensis* in hypercholesterolaemic rabbits. *Journal of ethnopharmacology*. 2004; 92:47–51. [PubMed: 15099846]
553. Singh RK, Mehta S, Jaiswal D, Rai PK, Watal G. Antidiabetic effect of *Ficus bengalensis* aerial roots in experimental animals. *Journal of ethnopharmacology*. 2009; 123:110–4. [PubMed: 19429348]
554. Thakare VN, Suralkar AA, Deshpande AD, Naik SR. Stem bark extraction of *Ficus bengalensis* Linn for anti-inflammatory and analgesic activity in animal models. *Indian journal of experimental biology*. 48:39–45. [PubMed: 20358865]
555. Tognolini M, Ballabeni V, Bertoni S, Bruni R, Impicciatore M, Barocelli E. Protective effect of *Foeniculum vulgare* essential oil and anethole in an experimental model of thrombosis. *Pharmacol Res*. 2007; 56:254–60. [PubMed: 17709257]
556. Miguel MG, Cruz C, Faleiro L, Simoes MT, Figueiredo AC, Barroso JG, et al. *Foeniculum vulgare* essential oils: chemical composition, antioxidant and antimicrobial activities. *Natural product communications*. 5:319–28. [PubMed: 20334152]
557. Choi EM, Hwang JK. Antiinflammatory, analgesic and antioxidant activities of the fruit of *Foeniculum vulgare*. *Fitoterapia*. 2004; 75:557–65. [PubMed: 15351109]
558. Joshi H, Parle M. Cholinergic basis of memory-strengthening effect of *Foeniculum vulgare* Linn. *Journal of medicinal food*. 2006; 9:413–7. [PubMed: 17004908]
559. Agarwal R, Gupta SK, Agrawal SS, Srivastava S, Saxena R. Oculohypotensive effects of *foeniculum vulgare* in experimental models of glaucoma. *Indian journal of physiology and pharmacology*. 2008; 52:77–83. [PubMed: 18831355]
560. Singh B, Kale RK. Chemomodulatory action of *Foeniculum vulgare* (Fennel) on skin and forestomach papillomagenesis, enzymes associated with xenobiotic metabolism and antioxidant status in murine model system. *Food Chem Toxicol*. 2008; 46:3842–50. [PubMed: 18976688]
561. Pan MH, Chang WL, Lin-Shiau SY, Ho CT, Lin JK. Induction of apoptosis by garcinol and curcumin through cytochrome c release and activation of caspases in human leukemia HL-60 cells. *Journal of agricultural and food chemistry*. 2001; 49:1464–74. [PubMed: 11312881]
562. Prasad S, Ravindran J, Sung B, Pandey MK, Aggarwal BB. Garcinol potentiates TRAIL-induced apoptosis through modulation of death receptors and antiapoptotic proteins. *Molecular cancer therapeutics*. 9:856–68. [PubMed: 20371723]
563. Ahmad A, Wang Z, Ali R, Maitah MY, Kong D, Banerjee S, et al. Apoptosis-inducing effect of garcinol is mediated by NF-kappaB signaling in breast cancer cells. *Journal of cellular biochemistry*. 109:1134–41. [PubMed: 20108249]
564. Yoshida K, Tanaka T, Hirose Y, Yamaguchi F, Kohno H, Toida M, et al. Dietary garcinol inhibits 4-nitroquinoline 1-oxide-induced tongue carcinogenesis in rats. *Cancer letters*. 2005; 221:29–39. [PubMed: 15797624]
565. Padhye S, Ahmad A, Oswal N, Sarkar FH. Emerging role of Garcinol, the antioxidant chalcone from *Garcinia indica* Choisy and its synthetic analogs. *Journal of hematology & oncology*. 2009; 2:38. [PubMed: 19725977]
566. Heymsfield SB, Allison DB, Vasselli JR, Pietrobello A, Greenfield D, Nunez C. *Garcinia cambogia* (hydroxycitric acid) as a potential antiobesity agent: a randomized controlled trial. *Jama*. 1998; 280:1596–600. [PubMed: 9820262]

567. Kolodziejczyk J, Masullo M, Olas B, Piacente S, Wachowicz B. Effects of garcinol and guttiferone K isolated from *Garcinia cambogia* on oxidative/nitrative modifications in blood platelets and plasma. *Platelets*. 2009; 20:487–92. [PubMed: 19852687]
568. Lau GT, Ye L, Leung LK. The licorice flavonoid isoliquiritigenin suppresses phorbol ester-induced cyclooxygenase-2 expression in the non-tumorigenic MCF-10A breast cell line. *Planta medica*. 76:780–5. [PubMed: 20033868]
569. Wan XY, Luo M, Li XD, He P. Hepatoprotective and anti-hepatocarcinogenic effects of glycyrrhizin and matrine. *Chemico-biological interactions*. 2009; 181:15–9. [PubMed: 19426721]
570. Yu XQ, Xue CC, Zhou ZW, Li CG, Du YM, Liang J, et al. In vitro and in vivo neuroprotective effect and mechanisms of glabridin, a major active isoflavan from *Glycyrrhiza glabra* (licorice). *Life sciences*. 2008; 82:68–78. [PubMed: 18048062]
571. Visavadiya NP, Soni B, Dalwadi N. Evaluation of antioxidant and anti-atherogenic properties of *Glycyrrhiza glabra* root using in vitro models. *International journal of food sciences and nutrition*. 2009; 60 (Suppl 2):135–49. [PubMed: 19384750]
572. Shin YW, Bae EA, Lee B, Lee SH, Kim JA, Kim YS, et al. In vitro and in vivo antiallergic effects of *Glycyrrhiza glabra* and its components. *Planta medica*. 2007; 73:257–61. [PubMed: 17327992]
573. Sheela ML, Ramakrishna MK, Salimath BP. Angiogenic and proliferative effects of the cytokine VEGF in Ehrlich ascites tumor cells is inhibited by *Glycyrrhiza glabra*. *International immunopharmacology*. 2006; 6:494–8. [PubMed: 16428085]
574. Visavadiya NP, Narasimhacharya AV. Hypocholesterolaemic and antioxidant effects of *Glycyrrhiza glabra* (Linn) in rats. *Molecular nutrition & food research*. 2006; 50:1080–6. [PubMed: 17054099]
575. Gholap S, Kar A. Effects of *Inula racemosa* root and *Gymnema sylvestre* leaf extracts in the regulation of corticosteroid induced diabetes mellitus: involvement of thyroid hormones. *Pharmazie*. 2003; 58:413–5. [PubMed: 12857006]
576. Daisy P, Eliza J, Mohamed Farook KA. A novel dihydroxy gymnemic triacetate isolated from *Gymnema sylvestre* possessing normoglycemic and hypolipidemic activity on STZ-induced diabetic rats. *Journal of ethnopharmacology*. 2009; 126:339–44. [PubMed: 19703537]
577. Yadav M, Lavania A, Tomar R, Prasad GB, Jain S, Yadav H. Complementary and comparative study on hypoglycemic and antihyperglycemic activity of various extracts of *Eugenia jambolana* seed, *Momordica charantia* fruits, *Gymnema sylvestre*, and *Trigonella foenum graecum* seeds in rats. *Applied biochemistry and biotechnology*. 160:2388–400. [PubMed: 19904502]
578. Kanetkar P, Singhal R, Kamat M. *Gymnema sylvestre*: A Memoir. *Journal of clinical biochemistry and nutrition*. 2007; 41:77–81. [PubMed: 18193099]
579. Leach MJ. *Gymnema sylvestre* for diabetes mellitus: a systematic review. *Journal of alternative and complementary medicine* (New York, NY. 2007; 13:977–83.
580. Alam MI, Gomes A. Viper venom-induced inflammation and inhibition of free radical formation by pure compound (2-hydroxy-4-methoxy benzoic acid) isolated and purified from anantamul (*Hemidesmus indicus* R. BR) root extract. *Toxicon*. 1998; 36:207–15. [PubMed: 9604294]
581. Saravanan S, Srikumar R, Manikandan S, Jeya Parthasarathy N, Sheela Devi R. Hypolipidemic effect of triphala in experimentally induced hypercholesteremic rats. *Yakugaku Zasshi*. 2007; 127:385–8. [PubMed: 17268159]
582. Gayathri M, Kannabiran K. 2-hydroxy 4-methoxy benzoic acid isolated from roots of *Hemidesmus indicus* ameliorates liver, kidney and pancreas injury due to streptozotocin-induced diabetes in rats. *Indian J Exp Biol*. 48:159–64. [PubMed: 20455325]
583. Sultana S, Alam A, Khan N, Sharma S. Inhibition of cutaneous oxidative stress and two-stage skin carcinogenesis by *Hemidesmus indicus* (L) in Swiss albino mice. *Indian J Exp Biol*. 2003; 41:1416–23. [PubMed: 15320495]
584. Kavitha D, Shilpa PN, Devaraj SN. Antibacterial and antidiarrhoeal effects of alkaloids of *Holarrhena antidysenterica* WALL. *Indian journal of experimental biology*. 2004; 42:589–94. [PubMed: 15260110]

585. Jeong JB, Hong SC, Jeong HJ. 3,4-dihydroxybenzaldehyde purified from the barley seeds (*Hordeum vulgare*) inhibits oxidative DNA damage and apoptosis via its antioxidant activity. *Phytomedicine*. 2009; 16:85–94. [PubMed: 19022639]
586. Akramiene D, Grazeliene G, Didziapetriene J, Kevelaitis E. Treatment of Lewis lung carcinoma by photodynamic therapy and glucan from barley. *Medicina (Kaunas)*. 2009; 45:480–5. [PubMed: 19605969]
587. Singh B, Chandan BK, Sharma N, Bhardwaj V, Satti NK, Gupta VN, et al. Isolation, structure elucidation and in vivo hepatoprotective potential of trans-tetracos-15-enoic acid from *Indigofera tinctoria* Linn. *Phytother Res*. 2006; 20:831–9. [PubMed: 16841368]
588. Singh B, Saxena AK, Chandan BK, Bhardwaj V, Gupta VN, Suri OP, et al. Hepatoprotective activity of indigtone--a bioactive fraction from *Indigofera tinctoria* Linn. *Phytother Res*. 2001; 15:294–7. [PubMed: 11406850]
589. Puri A, Khaliq T, Rajendran SM, Bhatia G, Chandra R, Narender T. Antidyslipidemic activity of *Indigofera tinctoria*. *J Herb Pharmacother*. 2007; 7:59–64. [PubMed: 17594987]
590. Sreepriya M, Devaki T, Balakrishna K, Apparanantham T. Effect of *Indigofera tinctoria* Linn on liver antioxidant defense system during D-galactosamine/endotoxin-induced acute hepatitis in rodents. *Indian J Exp Biol*. 2001; 39:181–4. [PubMed: 11480218]
591. Pal HC, Sehar I, Bhushan S, Gupta BD, Saxena AK. Activation of caspases and poly (ADP-ribose) polymerase cleavage to induce apoptosis in leukemia HL-60 cells by *Inula racemosa*. *Toxicol In Vitro*.
592. Srivastava S, Gupta PP, Prasad R, Dixit KS, Palit G, Ali B, et al. Evaluation of antiallergic activity (type I hypersensitivity) of *Inula racemosa* in rats. *Indian J Physiol Pharmacol*. 1999; 43:235–41. [PubMed: 10365318]
593. Ko SG, Koh SH, Jun CY, Nam CG, Bae HS, Shin MK. Induction of apoptosis by *Saussurea lappa* and *Pharbitis nil* on AGS gastric cancer cells. *Biol Pharm Bull*. 2004; 27:1604–10. [PubMed: 15467204]
594. Gamez MJ, Jimenez J, Risco S, Zarzuelo A. Hypoglycemic activity in various species of the genus *Lavandula*. Part 1: *Lavandula stoechas* L and *Lavandula multifida* L. *Pharmazie*. 1987; 42:706–7. [PubMed: 3438332]
595. Gilani AH, Aziz N, Khan MA, Shaheen F, Jabeen Q, Siddiqui BS, et al. Ethnopharmacological evaluation of the anticonvulsant, sedative and antispasmodic activities of *Lavandula stoechas* L. *J Ethnopharmacol*. 2000; 71:161–7. [PubMed: 10904159]
596. Lemus-Molina Y, Sanchez-Gomez MV, Delgado-Hernandez R, Matute C. *Mangifera indica* L. extract attenuates glutamate-induced neurotoxicity on rat cortical neurons. *Neurotoxicology*. 2009; 30:1053–8. [PubMed: 19591864]
597. Garrido G, Gonzalez D, Lemus Y, Garcia D, Lodeiro L, Quintero G, et al. In vivo and in vitro anti-inflammatory activity of *Mangifera indica* L. extract (VIMANG). *Pharmacol Res*. 2004; 50:143–9. [PubMed: 15177302]
598. Noratto GD, Bertoldi MC, Krenek K, Talcott ST, Stringheta PC, Mertens-Talcott SU. Anticarcinogenic effects of polyphenolics from mango (*Mangifera indica*) varieties. *J Agric Food Chem*. 58:4104–12. [PubMed: 20205391]
599. Pourahmad J, Eskandari MR, Shakibaei R, Kamalinejad M. A search for hepatoprotective activity of fruit extract of *Mangifera indica* L. against oxidative stress cytotoxicity. *Plant Foods Hum Nutr*. 65:83–9. [PubMed: 20204522]
600. Kumar S, Maheshwari KK, Singh V. Effects of *Mangifera indica* fruit extract on cognitive deficits in mice. *J Environ Biol*. 2009; 30:563–6. [PubMed: 20120497]
601. Severi JA, Lima ZP, Kushima H, Brito AR, Santos LC, Vilegas W, et al. Polyphenols with antiulcerogenic action from aqueous decoction of mango leaves (*Mangifera indica* L). *Molecules*. 2009; 14:1098–110. [PubMed: 19305363]
602. Akila M, Devaraj H. Synergistic effect of tincture of *Crataegus* and *Mangifera indica* L. extract on hyperlipidemic and antioxidant status in atherogenic rats. *Vascul Pharmacol*. 2008; 49:173–7. [PubMed: 18755296]

603. Parmar HS, Kar A. Possible amelioration of atherogenic diet induced dyslipidemia, hypothyroidism and hyperglycemia by the peel extracts of *Mangifera indica*, *Cucumis melo* and *Citrullus vulgaris* fruits in rats. *Biofactors*. 2008; 33:13–24. [PubMed: 19276533]
604. Pardo-Andreu GL, Philip SJ, Riano A, Sanchez C, Viada C, Nunez-Selles AJ, et al. *Mangifera indica* L. (Vimang) protection against serum oxidative stress in elderly humans. *Arch Med Res*. 2006; 37:158–64. [PubMed: 16314203]
605. Ojewole JA. Antiinflammatory, analgesic and hypoglycemic effects of *Mangifera indica* Linn. (Anacardiaceae) stem-bark aqueous extract. *Methods Find Exp Clin Pharmacol*. 2005; 27:547–54. [PubMed: 16273134]
606. Sangeetha KN, Sujatha S, Muthusamy VS, Anand S, Nithya N, Velmurugan D, et al. 3beta-taraxerol of *Mangifera indica*, a PI3K dependent dual activator of glucose transport and glycogen synthesis in 3T3-L1 adipocytes. *Biochim Biophys Acta*. 1800:359–66. [PubMed: 20026188]
607. Carvalho AC, Guedes MM, de Souza AL, Trevisan MT, Lima AF, Santos FA, et al. Gastroprotective effect of mangiferin, a xanthonoid from *Mangifera indica*, against gastric injury induced by ethanol and indomethacin in rodents. *Planta Med*. 2007; 73:1372–6. [PubMed: 17918041]
608. Kumar A, Samarth RM, Yasmeen S, Sharma A, Sugahara T, Terado T, et al. Anticancer and radioprotective potentials of *Mentha piperita*. *Biofactors*. 2004; 22:87–91. [PubMed: 15630259]
609. Sharma A, Sharma MK, Kumar M. Protective effect of *Mentha piperita* against arsenic-induced toxicity in liver of Swiss albino mice. *Basic Clin Pharmacol Toxicol*. 2007; 100:249–57. [PubMed: 17371529]
610. Samarth RM. Protection against radiation induced hematopoietic damage in bone marrow of Swiss albino mice by *Mentha piperita* (Linn). *J Radiat Res (Tokyo)*. 2007; 48:523–8. [PubMed: 17938557]
611. McKay DL, Blumberg JB. A review of the bioactivity and potential health benefits of peppermint tea (*Mentha piperita* L). *Phytother Res*. 2006; 20:619–33. [PubMed: 16767798]
612. Shkurupii VA, Odintsova OA, Kazarinova NV, Tkachenko KG. [Use of essential oil of peppermint (*Mentha piperita*) in the complex treatment of patients with infiltrative pulmonary tuberculosis]. *Probl Tuberk Bolezn Legk*. 2006:43–5.
613. de Sousa AA, Soares PM, de Almeida AN, Maia AR, de Souza EP, Assreuy AM. Antispasmodic effect of *Mentha piperita* essential oil on tracheal smooth muscle of rats. *J Ethnopharmacol*. 130:433–6. [PubMed: 20488237]
614. Inoue T, Sugimoto Y, Masuda H, Kamei C. Antiallergic effect of flavonoid glycosides obtained from *Mentha piperita* L. *Biol Pharm Bull*. 2002; 25:256–9. [PubMed: 11853178]
615. Dar A, Behbahanian S, Malik A, Jahan N. Hypotensive effect of the methanolic extract of *Mimusops elengi* in normotensive rats. *Phytomedicine*. 1999; 6:373–8. [PubMed: 11962547]
616. Kobori M, Nakayama H, Fukushima K, Ohnishi-Kameyama M, Ono H, Fukushima T, et al. Bitter gourd suppresses lipopolysaccharide-induced inflammatory responses. *J Agric Food Chem*. 2008; 56:4004–11. [PubMed: 18489106]
617. Akihisa T, Higo N, Tokuda H, Ukiya M, Akazawa H, Tochigi Y, et al. Cucurbitane-type triterpenoids from the fruits of *Momordica charantia* and their cancer chemopreventive effects. *J Nat Prod*. 2007; 70:1233–9. [PubMed: 17685651]
618. Fan JM, Zhang Q, Xu J, Zhu S, Ke T, Gao de F, et al. Inhibition on Hepatitis B virus in vitro of recombinant MAP30 from bitter melon. *Mol Biol Rep*. 2009; 36:381–8. [PubMed: 18058255]
619. Lii CK, Chen HW, Yun WT, Liu KL. Suppressive effects of wild bitter gourd (*Momordica charantia* Linn. var. *abbreviata* ser.) fruit extracts on inflammatory responses in RAW264. 7 macrophages. *J Ethnopharmacol*. 2009; 122:227–33. [PubMed: 19330915]
620. Ray RB, Raychoudhuri A, Steele R, Nerurkar P. Bitter melon (*Momordica charantia*) extract inhibits breast cancer cell proliferation by modulating cell cycle regulatory genes and promotes apoptosis. *Cancer Res*. 70:1925–31. [PubMed: 20179194]
621. Uebanso T, Arai H, Taketani Y, Fukaya M, Yamamoto H, Mizuno A, et al. Extracts of *Momordica charantia* suppress postprandial hyperglycemia in rats. *J Nutr Sci Vitaminol (Tokyo)*. 2007; 53:482–8. [PubMed: 18202535]

622. Shih CC, Lin CH, Lin WL. Effects of *Momordica charantia* on insulin resistance and visceral obesity in mice on high-fat diet. *Diabetes Res Clin Pract.* 2008; 81:134–43. [PubMed: 18550200]
623. Teoh SL, Latiff AA, Das S. The effect of topical extract of *Momordica charantia* (bitter gourd) on wound healing in nondiabetic rats and in rats with diabetes induced by streptozotocin. *Clin Exp Dermatol.* 2009; 34:815–22. [PubMed: 19508570]
624. Alam S, Asad M, Asdaq SM, Prasad VS. Antiulcer activity of methanolic extract of *Momordica charantia* L. in rats. *J Ethnopharmacol.* 2009; 123:464–9. [PubMed: 19501279]
625. Nerurkar P, Ray RB. Bitter melon: antagonist to cancer. *Pharm Res.* 27:1049–53. [PubMed: 20198408]
626. Klomann SD, Mueller AS, Pallauf J, Krawinkel MB. Antidiabetic effects of bitter gourd extracts in insulin-resistant db/db mice. *Br J Nutr.* :1–8.
627. Leung L, Birtwhistle R, Kotecha J, Hannah S, Cuthbertson S. Anti-diabetic and hypoglycaemic effects of *Momordica charantia* (bitter melon): a mini review. *Br J Nutr.* 2009; 102:1703–8. [PubMed: 19825210]
628. Mahajan SG, Mali RG, Mehta AA. Protective Effect of Ethanolic Extract of Seeds of *Moringa oleifera* Lam. Against Inflammation Associated with Development of Arthritis in Rats. *J Immunotoxicol.* 2007; 4:39–47. [PubMed: 18958711]
629. Mahajan SG, Banerjee A, Chauhan BF, Padh H, Nivsarkar M, Mehta AA. Inhibitory effect of n-butanol fraction of *Moringa oleifera* Lam. seeds on ovalbumin-induced airway inflammation in a guinea pig model of asthma. *Int J Toxicol.* 2009; 28:519–27. [PubMed: 19966143]
630. Mahajan SG, Mehta AA. Immunosuppressive activity of ethanolic extract of seeds of *Moringa oleifera* Lam. in experimental immune inflammation. *J Ethnopharmacol.* 130:183–6. [PubMed: 20435128]
631. Nandave M, Ojha SK, Joshi S, Kumari S, Arya DS. *Moringa oleifera* leaf extract prevents isoproterenol-induced myocardial damage in rats: evidence for an antioxidant, antiperoxidative, and cardioprotective intervention. *J Med Food.* 2009; 12:47–55. [PubMed: 19298195]
632. Mishra D, Gupta R, Pant SC, Kushwah P, Satish HT, Flora SJ. Co-administration of monoisoamyl dimercaptosuccinic acid and *Moringa oleifera* seed powder protects arsenic-induced oxidative stress and metal distribution in mice. *Toxicol Mech Methods.* 2009; 19:169–82. [PubMed: 19778263]
633. Hamza AA. Ameliorative effects of *Moringa oleifera* Lam seed extract on liver fibrosis in rats. *Food Chem Toxicol.* 48:345–55. [PubMed: 19854235]
634. Anwar F, Latif S, Ashraf M, Gilani AH. *Moringa oleifera*: a food plant with multiple medicinal uses. *Phytother Res.* 2007; 21:17–25. [PubMed: 17089328]
635. Rath SS, Grover JK, Vats V. The effect of *Momordica charantia* and *Mucuna pruriens* in experimental diabetes and their effect on key metabolic enzymes involved in carbohydrate metabolism. *Phytother Res.* 2002; 16:236–43. [PubMed: 12164268]
636. Katzenschlager R, Evans A, Manson A, Patsalos PN, Ratnaraj N, Watt H, et al. *Mucuna pruriens* in Parkinson's disease: a double blind clinical and pharmacological study. *J Neurol Neurosurg Psychiatry.* 2004; 75:1672–7. [PubMed: 15548480]
637. Bhaskar A, Vidhya VG, Ramya M. Hypoglycemic effect of *Mucuna pruriens* seed extract on normal and streptozotocin-diabetic rats. *Fitoterapia.* 2008; 79:539–43. [PubMed: 18672037]
638. Kasture S, Pontis S, Pinna A, Schintu N, Spina L, Longoni R, et al. Assessment of symptomatic and neuroprotective efficacy of *Mucuna pruriens* seed extract in rodent model of Parkinson's disease. *Neurotox Res.* 2009; 15:111–22. [PubMed: 19384573]
639. Lieu CA, Kunselman AR, Manyam BV, Venkiteswaran K, Subramanian T. A water extract of *Mucuna pruriens* provides long-term amelioration of parkinsonism with reduced risk for dyskinesias. *Parkinsonism Relat Disord.* 16:458–65. [PubMed: 20570206]
640. Ahmad M, Yousuf S, Khan MB, Hoda MN, Ahmad AS, Ansari MA, et al. Attenuation by *Nardostachys jatamansi* of 6-hydroxydopamine-induced parkinsonism in rats: behavioral, neurochemical, and immunohistochemical studies. *Pharmacol Biochem Behav.* 2006; 83:150–60. [PubMed: 16500697]

641. Dhingra D, Goyal PK. Inhibition of MAO and GABA: probable mechanisms for antidepressant-like activity of *Nardostachys jatamansi* DC. in mice. *Indian J Exp Biol.* 2008; 46:212–8. [PubMed: 18512329]
642. Bae GS, Park HJ, Kim DY, Song JM, Kim TH, Oh HJ, et al. *Nardostachys jatamansi* protects against cerulein-induced acute pancreatitis. *Pancreas.* 39:520–9. [PubMed: 19940795]
643. Song MY, Bae UJ, Lee BH, Kwon KB, Seo EA, Park SJ, et al. *Nardostachys jatamansi* extract protects against cytokine-induced beta-cell damage and streptozotocin-induced diabetes. *World J Gastroenterol.* 16:3249–57. [PubMed: 20614480]
644. Liu CP, Tsai WJ, Shen CC, Lin YL, Liao JF, Chen CF, et al. Inhibition of (S)-armepavine from *Nelumbo nucifera* on autoimmune disease of MRL/MpJ-lpr/lpr mice. *Eur J Pharmacol.* 2006; 531:270–9. [PubMed: 16413531]
645. Ka SM, Kuo YC, Ho PJ, Tsai PY, Hsu YJ, Tsai WJ, et al. (S)-armepavine from Chinese medicine improves experimental autoimmune crescentic glomerulonephritis. *Rheumatology (Oxford).*
646. Mukherjee PK, Mukherjee D, Maji AK, Rai S, Heinrich M. The sacred lotus (*Nelumbo nucifera*) - phytochemical and therapeutic profile. *J Pharm Pharmacol.* 2009; 61:407–22. [PubMed: 19298686]
647. Sugimoto Y, Furutani S, Nishimura K, Itoh A, Tanahashi T, Nakajima H, et al. Antidepressant-like effects of neferine in the forced swimming test involve the serotonin1A (5-HT1A) receptor in mice. *Eur J Pharmacol.* 634:62–7. [PubMed: 20176013]
648. Shim SY, Choi JS, Byun DS. Kaempferol isolated from *Nelumbo nucifera* stamens negatively regulates FcεpsilonRI expression in human basophilic KU812F cells. *J Microbiol Biotechnol.* 2009; 19:155–60. [PubMed: 19307764]
649. Xiao JH, Zhang JH, Chen HL, Feng XL, Wang JL. Inhibitory effects of isoliensinine on bleomycin-induced pulmonary fibrosis in mice. *Planta Med.* 2005; 71:225–30. [PubMed: 15770542]
650. Sohn DH, Kim YC, Oh SH, Park EJ, Li X, Lee BH. Hepatoprotective and free radical scavenging effects of *Nelumbo nucifera*. *Phytomedicine.* 2003; 10:165–9. [PubMed: 12725571]
651. Mani SS, Subramanian IP, Pillai SS, Muthusamy K. Evaluation of Hypoglycemic Activity of Inorganic Constituents in *Nelumbo nucifera* Seeds on Streptozotocin-Induced Diabetes in Rats. *Biol Trace Elem Res.*
652. Jafri SH, Glass J, Shi R, Zhang S, Prince M, Kleiner-Hancock H. Thymoquinone and cisplatin as a therapeutic combination in lung cancer: In vitro and in vivo. *J Exp Clin Cancer Res.* 29:87. [PubMed: 20594324]
653. Nader MA, el-Agamy DS, Suddek GM. Protective effects of propolis and thymoquinone on development of atherosclerosis in cholesterol-fed rabbits. *Arch Pharm Res.* 33:637–43. [PubMed: 20422375]
654. Helal GK. Thymoquinone supplementation ameliorates acute endotoxemia-induced liver dysfunction in rats. *Pak J Pharm Sci.* 23:131–7. [PubMed: 20363688]
655. Chehl N, Chipitsyna G, Gong Q, Yeo CJ, Arafat HA. Anti-inflammatory effects of the *Nigella sativa* seed extract, thymoquinone, in pancreatic cancer cells. *HPB (Oxford).* 2009; 11:373–81. [PubMed: 19768141]
656. Mohamed A, Waris HM, Ramadan H, Quereshi M, Kalra J. Amelioration of chronic relapsing experimental autoimmune encephalomyelitis (cr-eae) using thymoquinone - biomed 2009. *Biomed Sci Instrum.* 2009; 45:274–9. [PubMed: 19369775]
657. Juhas S, Cikos S, Czikkova S, Vesela J, Il'kova G, Hajek T, et al. Effects of borneol and thymoquinone on TNBS-induced colitis in mice. *Folia Biol (Praha).* 2008; 54:1–7. [PubMed: 18226358]
658. El Mezayen R, El Gazzar M, Nicolls MR, Marecki JC, Dreskin SC, Nomiya H. Effect of thymoquinone on cyclooxygenase expression and prostaglandin production in a mouse model of allergic airway inflammation. *Immunol Lett.* 2006; 106:72–81. [PubMed: 16762422]
659. Tekeoglu I, Dogan A, Demiralp L. Effects of thymoquinone (volatile oil of black cumin) on rheumatoid arthritis in rat models. *Phytother Res.* 2006; 20:869–71. [PubMed: 16835876]
660. Ali BH, Blunden G. Pharmacological and toxicological properties of *Nigella sativa*. *Phytother Res.* 2003; 17:299–305. [PubMed: 12722128]

661. Salem ML. Immunomodulatory and therapeutic properties of the *Nigella sativa* L. seed. *Int Immunopharmacol.* 2005; 5:1749–70. [PubMed: 16275613]
662. Ragheb A, Attia A, Eldin WS, Elbarbry F, Gazarin S, Shoker A. The protective effect of thymoquinone, an anti-oxidant and anti-inflammatory agent, against renal injury: a review. *Saudi J Kidney Dis Transpl.* 2009; 20:741–52. [PubMed: 19736468]
663. Ghannadi A, Hajhashemi V, Jafarabadi H. An investigation of the analgesic and anti-inflammatory effects of *Nigella sativa* seed polyphenols. *J Med Food.* 2005; 8:488–93. [PubMed: 16379560]
664. Majdalawieh AF, Hmaidan R, Carr RI. *Nigella sativa* modulates splenocyte proliferation, Th1/Th2 cytokine profile, macrophage function and NK anti-tumor activity. *J Ethnopharmacol.*
665. Bashir MU, Qureshi HJ. Analgesic Effect of *Nigella sativa* Seeds Extract on Experimentally Induced Pain in Albino Mice. *J Coll Physicians Surg Pak.* 20:464–7. [PubMed: 20642947]
666. Isik H, Cevikbas A, Gurer US, Kiran B, Uresin Y, Rayaman P, et al. Potential adjuvant effects of *Nigella sativa* seeds to improve specific immunotherapy in allergic rhinitis patients. *Med Princ Pract.* 19:206–11. [PubMed: 20357504]
667. Boskabady MH, Mohsenpoor N, Takaloo L. Antiasthmatic effect of *Nigella sativa* in airways of asthmatic patients. *Phytomedicine.* 17:707–13. [PubMed: 20149611]
668. Akhondian J, Parsa A, Rakhshande H. The effect of *Nigella sativa* L. (black cumin seed) on intractable pediatric seizures. *Med Sci Monit.* 2007; 13:CR555–9. [PubMed: 18049435]
669. Rathore B, Paul B, Chaudhury BP, Saxena AK, Sahu AP, Gupta YK. Comparative studies of different organs of *Nyctanthes arbortristis* in modulation of cytokines in murine model of arthritis. *Biomed Environ Sci.* 2007; 20:154–9. [PubMed: 17624191]
670. Gupta P, Bajpai SK, Chandra K, Singh KL, Tandon JS. Antiviral profile of *Nyctanthes arbortristis* L. against encephalitis causing viruses. *Indian J Exp Biol.* 2005; 43:1156–60. [PubMed: 16359127]
671. Sen P, Maiti PC, Puri S, Ray A, Audulov NA, Valdman AV. Mechanism of anti-stress activity of *Ocimum sanctum* Linn, eugenol and *Tinospora malabarica* in experimental animals. *Indian J Exp Biol.* 1992; 30:592–6. [PubMed: 1459632]
672. Kath RK, Gupta RK. Antioxidant activity of hydroalcoholic leaf extract of *ocimum sanctum* in animal models of peptic ulcer. *Indian J Physiol Pharmacol.* 2006; 50:391–6. [PubMed: 17402269]
673. Prakash P, Gupta N. Therapeutic uses of *Ocimum sanctum* Linn (Tulsi) with a note on eugenol and its pharmacological actions: a short review. *Indian J Physiol Pharmacol.* 2005; 49:125–31. [PubMed: 16170979]
674. Mondal S, Mirdha BR, Mahapatra SC. The science behind sacredness of Tulsi (*Ocimum sanctum* Linn). *Indian J Physiol Pharmacol.* 2009; 53:291–306. [PubMed: 20509321]
675. Magesh V, Lee JC, Ahn KS, Lee HJ, Lee EO, et al. *Ocimum sanctum* induces apoptosis in A549 lung cancer cells and suppresses the in vivo growth of Lewis lung carcinoma cells. *Phytother Res.* 2009; 23:1385–91. [PubMed: 19277950]
676. Kaur G, Jaggi AS, Singh N. Exploring the potential effect of *Ocimum sanctum* in vincristine-induced neuropathic pain in rats. *J Brachial Plex Peripher Nerve Inj.* 5:3. [PubMed: 20181005]
677. Suanarunsawat T, Devakul Na Ayutthaya W, Songsak T, Thirawarapan S, Pongshompoo S. Antioxidant Activity and Lipid-Lowering Effect of Essential Oils Extracted from *Ocimum sanctum* L. Leaves in Rats Fed with a High Cholesterol Diet. *J Clin Biochem Nutr.* 46:52–9. [PubMed: 20104265]
678. Jyoti S, Satendra S, Sushma S, Anjana T, Shashi S. Antistressor activity of *Ocimum sanctum* (Tulsi) against experimentally induced oxidative stress in rabbits. *Methods Find Exp Clin Pharmacol.* 2007; 29:411–6. [PubMed: 17922070]
679. Anbuselvam C, Vijayavel K, Balasubramanian MP. Protective effect of *Operculina turpethum* against 7,12-dimethyl benz(a)anthracene induced oxidative stress with reference to breast cancer in experimental rats. *Chem Biol Interact.* 2007; 168:229–36. [PubMed: 17531963]
680. Jiawajinda S, Santisopasri V, Murakami A, Kim OK, Kim HW, Ohigashi H. Suppressive Effects of Edible Thai Plants on Superoxide and Nitric Oxide Generation. *Asian Pac J Cancer Prev.* 2002; 3:215–23. [PubMed: 12718578]

681. Siriwatanametanon N, Fiebich BL, Efferth T, Prieto JM, Heinrich M. Traditionally used Thai medicinal plants: in vitro anti-inflammatory, anticancer and antioxidant activities. *J Ethnopharmacol.* 130:196–207. [PubMed: 20435130]
682. Laupattarakasem P, Houghton PJ, Hoult JR, Itharat A. An evaluation of the activity related to inflammation of four plants used in Thailand to treat arthritis. *J Ethnopharmacol.* 2003; 85:207–15. [PubMed: 12639742]
683. Chirdchupunseree H, Pramyothin P. Protective activity of phyllanthin in ethanol-treated primary culture of rat hepatocytes. *J Ethnopharmacol.* 128:172–6. [PubMed: 20064596]
684. Rajeshkumar NV, Kuttan R. Phyllanthus amarus extract administration increases the life span of rats with hepatocellular carcinoma. *J Ethnopharmacol.* 2000; 73:215–9. [PubMed: 11025159]
685. Kiemer AK, Hartung T, Huber C, Vollmar AM. Phyllanthus amarus has anti-inflammatory potential by inhibition of iNOS, COX-2, and cytokines via the NF-kappaB pathway. *J Hepatol.* 2003; 38:289–97. [PubMed: 12586294]
686. Kassuya CA, Leite DF, de Melo LV, Rehder VL, Calixto JB. Anti-inflammatory properties of extracts, fractions and lignans isolated from Phyllanthus amarus. *Planta Med.* 2005; 71:721–6. [PubMed: 16142635]
687. Kassuya CA, Silvestre AA, Rehder VL, Calixto JB. Anti-allodynic and anti-oedematogenic properties of the extract and lignans from Phyllanthus amarus in models of persistent inflammatory and neuropathic pain. *Eur J Pharmacol.* 2003; 478:145–53. [PubMed: 14575799]
688. Xin-Hua W, Chang-Qing L, Xing-Bo G, Lin-Chun F. A comparative study of Phyllanthus amarus compound and interferon in the treatment of chronic viral hepatitis B. *Southeast Asian J Trop Med Public Health.* 2001; 32:140–2. [PubMed: 11485076]
689. Adeneye AA, Amole OO, Adeneye AK. Hypoglycemic and hypocholesterolemic activities of the aqueous leaf and seed extract of Phyllanthus amarus in mice. *Fitoterapia.* 2006; 77:511–4. [PubMed: 16905277]
690. Raphael KR, Sabu M, Kumar KH, Kuttan R. Inhibition of N-Methyl N'-nitro-N-nitrosoguanidine (MNNG) induced gastric carcinogenesis by Phyllanthus amarus extract. *Asian Pac J Cancer Prev.* 2006; 7:299–302. [PubMed: 16839226]
691. Mellinger CG, Carbonero ER, Noleto GR, Cipriani TR, Oliveira MB, Gorin PA, et al. Chemical and biological properties of an arabinogalactan from Phyllanthus niruri. *J Nat Prod.* 2005; 68:1479–83. [PubMed: 16252911]
692. Bagalkotkar G, Sagineedu SR, Saad MS, Stanslas J. Phytochemicals from Phyllanthus niruri Linn. and their pharmacological properties: a review. *J Pharm Pharmacol.* 2006; 58:1559–70. [PubMed: 17331318]
693. Venkateswaran PS, Millman I, Blumberg BS. Effects of an extract from Phyllanthus niruri on hepatitis B and woodchuck hepatitis viruses: in vitro and in vivo studies. *Proc Natl Acad Sci U S A.* 1987; 84:274–8. [PubMed: 3467354]
694. Chatterjee M, Sil PC. Hepatoprotective effect of aqueous extract of Phyllanthus niruri on nimesulide-induced oxidative stress in vivo. *Indian J Biochem Biophys.* 2006; 43:299–305. [PubMed: 17133737]
695. Sharma I, Gusain D, Dixit VP. Hypolipidaemic and antiatherosclerotic effects of plumbagin in rabbits. *Indian J Physiol Pharmacol.* 1991; 35:10–4. [PubMed: 1917004]
696. Murugaiyah V, Chan KL. Mechanisms of antihyperuricemic effect of Phyllanthus niruri and its lignan constituents. *J Ethnopharmacol.* 2009; 124:233–9. [PubMed: 19397979]
697. Nworu CS, Akah PA, Okoye FB, Proksch P, Esimone CO. The effects of Phyllanthus niruri aqueous extract on the activation of murine lymphocytes and bone marrow-derived macrophages. *Immunol Invest.* 39:245–67. [PubMed: 20380522]
698. Dhuley JN. Effect of picroliv administration on hepatic microsomal mixed function oxidases and glutathione-conjugating enzyme system in cholestatic rats. *Hindustan Antibiot Bull.* 2005; 47–48:13–9.
699. Vivekanandan P, Gobianand K, Priya S, Vijayalakshmi P, Karthikeyan S. Protective effect of picroliv against hydrazine-induced hyperlipidemia and hepatic steatosis in rats. *Drug Chem Toxicol.* 2007; 30:241–52. [PubMed: 17613009]

700. Gupta A, Khajuria A, Singh J, Bedi KL, Satti NK, Dutt P, et al. Immunomodulatory activity of biopolymeric fraction RLJ-NE-205 from *Picrorhiza kurroa*. *Int Immunopharmacol*. 2006; 6:1543–9. [PubMed: 16919826]
701. Rastogi R, Saksena S, Garg NK, Kapoor NK, Agarwal DP, Dhawan BN. Picroliv protects against alcohol-induced chronic hepatotoxicity in rats. *Planta Med*. 1996; 62:283–5. [PubMed: 8693047]
702. Verma PC, Basu V, Gupta V, Saxena G, Rahman LU. Pharmacology and chemistry of a potent hepatoprotective compound Picroliv isolated from the roots and rhizomes of *Picrorhiza kurroa* royle ex benth (kutki). *Curr Pharm Biotechnol*. 2009; 10:641–9. [PubMed: 19619118]
703. Zhang Y, DeWitt DL, Murugesan S, Nair MG. Novel lipid-peroxidation- and cyclooxygenase-inhibitory tannins from *Picrorhiza kurroa* seeds. *Chem Biodivers*. 2004; 1:426–41. [PubMed: 17191857]
704. Morikawa T, Matsuda H, Yamaguchi I, Pongpiriyadacha Y, Yoshikawa M. New amides and gastroprotective constituents from the fruit of *Piper chaba*. *Planta Med*. 2004; 70:152–9. [PubMed: 14994194]
705. Matsuda H, Ninomiya K, Morikawa T, Yasuda D, Yamaguchi I, Yoshikawa M. Hepatoprotective amide constituents from the fruit of *Piper chaba*: Structural requirements, mode of action, and new amides. *Bioorg Med Chem*. 2009; 17:7313–23. [PubMed: 19775895]
706. Pradeep CR, Kuttan G. Effect of piperine on the inhibition of nitric oxide (NO) and TNF-alpha production. *Immunopharmacol Immunotoxicol*. 2003; 25:337–46. [PubMed: 19180797]
707. Bae GS, Kim MS, Jung WS, Seo SW, Yun SW, Kim SG, et al. Inhibition of lipopolysaccharide-induced inflammatory responses by piperine. *Eur J Pharmacol*. 642:154–62. [PubMed: 20621590]
708. Lee SA, Hong SS, Han XH, Hwang JS, Oh GJ, Lee KS, et al. Piperine from the fruits of *Piper longum* with inhibitory effect on monoamine oxidase and antidepressant-like activity. *Chem Pharm Bull (Tokyo)*. 2005; 53:832–5. [PubMed: 15997146]
709. Iwashita M, Oka N, Ohkubo S, Saito M, Nakahata N. Piperlongumine, a constituent of *Piper longum* L. inhibits rabbit platelet aggregation as a thromboxane A(2) receptor antagonist. *Eur J Pharmacol*. 2007; 570:38–42. [PubMed: 17618620]
710. Devan P, Bani S, Suri KA, Satti NK, Qazi GN. Immunomodulation exhibited by piperinic acid through suppression of proinflammatory cytokines. *Int Immunopharmacol*. 2007; 7:889–99. [PubMed: 17499191]
711. Sunila ES, Kuttan G. *Piper longum* inhibits VEGF and proinflammatory cytokines and tumor-induced angiogenesis in C57BL/6 mice. *Int Immunopharmacol*. 2006; 6:733–41. [PubMed: 16546703]
712. Kumar A, Panghal S, Mallapur SS, Kumar M, Ram V, Singh BK. Antiinflammatory Activity of *Piper longum* Fruit Oil. *Indian J Pharm Sci*. 2009; 71:454–6. [PubMed: 20502557]
713. Subramanian U, Poongavanam S, Vanisree AJ. Studies on the neuroprotective role of *Piper longum* in C6 glioma induced rats. *Invest New Drugs*. 28:615–23. [PubMed: 19730792]
714. Wakade AS, Shah AS, Kulkarni MP, Juvekar AR. Protective effect of *Piper longum* L. on oxidative stress induced injury and cellular abnormality in adriamycin induced cardiotoxicity in rats. *Indian J Exp Biol*. 2008; 46:528–33. [PubMed: 18807757]
715. Agrawal AK, Rao CV, Sairam K, Joshi VK, Goel RK. Effect of *Piper longum* Linn, *Zingiber officinalis* Linn and *Ferula* species on gastric ulceration and secretion in rats. *Indian J Exp Biol*. 2000; 38:994–8. [PubMed: 11324171]
716. Chonpathompikunlert P, Wattanathorn J, Muchimapura S. Piperine, the main alkaloid of Thai black pepper, protects against neurodegeneration and cognitive impairment in animal model of cognitive deficit like condition of Alzheimer's disease. *Food Chem Toxicol*. 48:798–802. [PubMed: 20034530]
717. Vijayakumar RS, Nalini N. Piperine, an active principle from *Piper nigrum*, modulates hormonal and apo lipoprotein profiles in hyperlipidemic rats. *J Basic Clin Physiol Pharmacol*. 2006; 17:71–86. [PubMed: 16910313]
718. Pattanaik S, Hota D, Prabhakar S, Kharbanda P, Pandhi P. Effect of piperine on the steady-state pharmacokinetics of phenytoin in patients with epilepsy. *Phytother Res*. 2006; 20:683–6. [PubMed: 16767797]

719. Srinivasan K. Black pepper and its pungent principle-piperine: a review of diverse physiological effects. *Crit Rev Food Sci Nutr.* 2007; 47:735–48. [PubMed: 17987447]
720. Wei K, Dou DQ, Pei YP, Chen YJ. Comparison of the chemical constituents and pharmacological action of *Piper nigrum* Linn. with *P. methysticum* forst. *Zhongguo Zhong Yao Za Zhi.* 2002; 27:328–33. [PubMed: 12814095]
721. Hirata N, Naruto S, Inaba K, Itoh K, Tokunaga M, Iinuma M, et al. Histamine release inhibitory activity of *Piper nigrum* leaf. *Biol Pharm Bull.* 2008; 31:1973–6. [PubMed: 18827366]
722. Ahmad NS, Farman M, Najmi MH, Mian KB, Hasan A. Pharmacological basis for use of *Pistacia integerrima* leaves in hyperuricemia and gout. *J Ethnopharmacol.* 2008; 117:478–82. [PubMed: 18420362]
723. Jahangir T, Sultana S. Modulatory effects of *Pluchea lanceolata* against chemically induced oxidative damage, hyperproliferation and two-stage renal carcinogenesis in Wistar rats. *Mol Cell Biochem.* 2006; 291:175–85. [PubMed: 16767495]
724. Bhagwat DP, Kharya MD, Bani S, Kaul A, Kour K, Chauhan PS, et al. Immunosuppressive properties of *Pluchea lanceolata* leaves. *Indian J Pharmacol.* 42:21–6. [PubMed: 20606832]
725. Yang SJ, Chang SC, Wen HC, Chen CY, Liao JF, Chang CH. Plumbagin activates ERK1/2 and Akt via superoxide, Src and PI3-kinase in 3T3-L1 cells. *Eur J Pharmacol.* 638:21–8. [PubMed: 20420821]
726. Checker R, Sharma D, Sandur SK, Khanam S, Poduval TB. Anti-inflammatory effects of plumbagin are mediated by inhibition of NF-kappaB activation in lymphocytes. *Int Immunopharmacol.* 2009; 9:949–58. [PubMed: 19374955]
727. Parimala R, Sachdanandam P. Effect of Plumbagin on some glucose metabolising enzymes studied in rats in experimental hepatoma. *Mol Cell Biochem.* 1993; 125:59–63. [PubMed: 8264573]
728. Tsai WJ, Chen YC, Wu MH, Lin LC, Chuang KA, Chang SC, et al. Seselin from *Plumbago zeylanica* inhibits phytohemagglutinin (PHA)-stimulated cell proliferation in human peripheral blood mononuclear cells. *J Ethnopharmacol.* 2008; 119:67–73. [PubMed: 18577441]
729. Chen YC, Tsai WJ, Wu MH, Lin LC, Kuo YC. Suberosin inhibits proliferation of human peripheral blood mononuclear cells through the modulation of the transcription factors NF-AT and NF-kappaB. *Br J Pharmacol.* 2007; 150:298–312. [PubMed: 17179947]
730. Tamrakar AK, Yadav PP, Tiwari P, Maurya R, Srivastava AK. Identification of pongamol and karanjin as lead compounds with antihyperglycemic activity from *Pongamia pinnata* fruits. *J Ethnopharmacol.* 2008; 118:435–9. [PubMed: 18572336]
731. Srinivasan K, Muruganandan S, Lal J, Chandra S, Tandan SK, Prakash VR. Evaluation of anti-inflammatory activity of *Pongamia pinnata* leaves in rats. *J Ethnopharmacol.* 2001; 78:151–7. [PubMed: 11694360]
732. Punitha R, Manoharan S. Antihyperglycemic and antilipidperoxidative effects of *Pongamia pinnata* (Linn) Pierre flowers in alloxan induced diabetic rats. *J Ethnopharmacol.* 2006; 105:39–46. [PubMed: 16271443]
733. Raghavendra M, Trigunayat A, Singh RK, Mitra S, Goel RK, Acharya SB. Effect of ethanolic extract of root of *Pongamia pinnata* (L) pierre on oxidative stress, behavioral and histopathological alterations induced by cerebral ischemia--reperfusion and long-term hypoperfusion in rats. *Indian J Exp Biol.* 2007; 45:868–76. [PubMed: 17948735]
734. Prabha T, Dorababu M, Goel S, Agarwal PK, Singh A, Joshi VK, et al. Effect of methanolic extract of *Pongamia pinnata* Linn seed on gastro-duodenal ulceration and mucosal offensive and defensive factors in rats. *Indian J Exp Biol.* 2009; 47:649–59. [PubMed: 19775071]
735. Badole SL, Bodhankar SL. Antidiabetic activity of cycloart-23-ene-3beta, 25-diol (B2) isolated from *Pongamia pinnata* (L. Pierre) in streptozotocin-nicotinamide induced diabetic mice. *Eur J Pharmacol.* 632:103–9. [PubMed: 20122920]
736. Choi JH, Rho MC, Lee SW, Choi JN, Kim K, Song GY, et al. Bavachin and isobavachalcone, acyl-coenzyme A: cholesterol acyltransferase inhibitors from *Psoralea corylifolia*. *Arch Pharm Res.* 2008; 31:1419–23. [PubMed: 19023538]

737. Xu Q, Pan Y, Yi LT, Li YC, Mo SF, Jiang FX, et al. Antidepressant-like effects of psoralen isolated from the seeds of *Psoralea corylifolia* in the mouse forced swimming test. *Biol Pharm Bull.* 2008; 31:1109–14. [PubMed: 18520040]
738. Cho H, Jun JY, Song EK, Kang KH, Baek HY, Ko YS, et al. Bakuchiol: a hepatoprotective compound of *Psoralea corylifolia* on tacrine-induced cytotoxicity in Hep G2 cells. *Planta Med.* 2001; 67:750–1. [PubMed: 11731920]
739. Wu CZ, Hong SS, Cai XF, Dat NT, Nan JX, Hwang BY, et al. Hypoxia-inducible factor-1 and nuclear factor-kappaB inhibitory meroterpenoid analogues of bakuchiol, a constituent of the seeds of *Psoralea corylifolia*. *Bioorg Med Chem Lett.* 2008; 18:2619–23. [PubMed: 18359631]
740. Chen Y, Wang HD, Xia X, Kung HF, Pan Y, Kong LD. Behavioral and biochemical studies of total furocoumarins from seeds of *Psoralea corylifolia* in the chronic mild stress model of depression in mice. *Phytomedicine.* 2007; 14:523–9. [PubMed: 17085027]
741. Zhao G, Zheng XW, Qin GW, Gai Y, Jiang ZH, Guo LH. In vitro dopaminergic neuroprotective and in vivo antiparkinsonian-like effects of Delta 3,2-hydroxybakuchiol isolated from *Psoralea corylifolia* (L). *Cell Mol Life Sci.* 2009; 66:1617–29. [PubMed: 19322517]
742. An HJ, Seo MJ, Choi IY, Park RK, Jeong S, Lee JY, et al. Induction of nitric oxide & tumour necrosis factor-alpha by *Psoralea corylifolia*. *Indian J Med Res.* 2008; 128:752–8. [PubMed: 19246800]
743. Latha PG, Evans DA, Panikkar KR, Jayavardhanan KK. Immunomodulatory and antitumour properties of *Psoralea corylifolia* seeds. *Fitoterapia.* 2000; 71:223–31. [PubMed: 10844159]
744. Jahromi MA, Ray AB. Antihyperlipidemic effect of flavonoids from *Pterocarpus marsupium*. *J Nat Prod.* 1993; 56:989–94. [PubMed: 8377021]
745. Dhanabal SP, Kokate CK, Ramanathan M, Kumar EP, Suresh B. Hypoglycaemic activity of *Pterocarpus marsupium* Roxb. *Phytother Res.* 2006; 20:4–8. [PubMed: 16397913]
746. Cho JY, Park J, Kim PS, Yoo ES, Baik KU, Park MH. Savinin, a lignan from *Pterocarpus santalinus* inhibits tumor necrosis factor-alpha production and T cell proliferation. *Biol Pharm Bull.* 2001; 24:167–71. [PubMed: 11217086]
747. Narayan S, Devi RS, Srinivasan P, Shyamala Devi CS. *Pterocarpus santalinus*: a traditional herbal drug as a protectant against ibuprofen induced gastric ulcers. *Phytother Res.* 2005; 19:958–62. [PubMed: 16317653]
748. Dey D, Pal BC, Biswas T, Roy SS, Bandyopadhyay A, Mandal SK, et al. A Lupinoid prevented fatty acid induced inhibition of insulin sensitivity in 3T3 L1 adipocytes. *Mol Cell Biochem.* 2007; 300:149–57. [PubMed: 17149545]
749. Santosh N, Mohan K, Royana S, Yamini TB. Hepatotoxicity of tubers of Indian Kudzu (*Pueraria tuberosa*) in rats. *Food Chem Toxicol.* 48:1066–71. [PubMed: 20122980]
750. Dell'agli M, Galli GV, Bulgari M, Basilico N, Romeo S, Bhattacharya D, et al. Ellagitannins of the fruit rind of pomegranate (*Punica granatum*) antagonize in vitro the host inflammatory response mechanisms involved in the onset of malaria. *Malar J.* 9:208. [PubMed: 20642847]
751. Katz SR, Newman RA, Lansky EP. *Punica granatum*: heuristic treatment for diabetes mellitus. *J Med Food.* 2007; 10:213–7. [PubMed: 17651054]
752. Lansky EP, Newman RA. *Punica granatum* (pomegranate) and its potential for prevention and treatment of inflammation and cancer. *J Ethnopharmacol.* 2007; 109:177–206. [PubMed: 17157465]
753. Jurenka JS. Therapeutic applications of pomegranate (*Punica granatum* L): a review. *Altern Med Rev.* 2008; 13:128–44. [PubMed: 18590349]
754. Lee SI, Kim BS, Kim KS, Lee S, Shin KS, Lim JS. Immune-suppressive activity of punicalagin via inhibition of NFAT activation. *Biochem Biophys Res Commun.* 2008; 371:799–803. [PubMed: 18466764]
755. Afaq F, Saleem M, Krueger CG, Reed JD, Mukhtar H. Anthocyanin- and hydrolyzable tannin-rich pomegranate fruit extract modulates MAPK and NF-kappaB pathways and inhibits skin tumorigenesis in CD-1 mice. *Int J Cancer.* 2005; 113:423–33. [PubMed: 15455341]
756. Shukla M, Gupta K, Rasheed Z, Khan KA, Haqqi TM. Bioavailable constituents/metabolites of pomegranate (*Punica granatum* L) preferentially inhibit COX2 activity ex vivo and IL-1beta-

induced PGE2 production in human chondrocytes in vitro. *J Inflamm (Lond)*. 2008; 5:9. [PubMed: 18554383]

757. Ahmed S, Wang N, Hafeez BB, Cheruvu VK, Haqqi TM. Punica granatum L. extract inhibits IL-1 β -induced expression of matrix metalloproteinases by inhibiting the activation of MAP kinases and NF- κ B in human chondrocytes in vitro. *J Nutr*. 2005; 135:2096–102. [PubMed: 16140882]
758. Toklu HZ, Sehirli O, Ozyurt H, Mayadagli AA, Eksioglu-Demiralp E, Cetinel S, et al. Punica granatum peel extract protects against ionizing radiation-induced enteritis and leukocyte apoptosis in rats. *J Radiat Res (Tokyo)*. 2009; 50:345–53. [PubMed: 19478462]
759. Jung KH, Kim MJ, Ha E, Uhm YK, Kim HK, Chung JH, et al. Suppressive effect of Punica granatum on the production of tumor necrosis factor (Tnf) in BV2 microglial cells. *Biol Pharm Bull*. 2006; 29:1258–61. [PubMed: 16755029]
760. Huang TH, Peng G, Kota BP, Li GQ, Yamahara J, Roufogalis BD, et al. Anti-diabetic action of Punica granatum flower extract: activation of PPAR- γ and identification of an active component. *Toxicol Appl Pharmacol*. 2005; 207:160–9. [PubMed: 16102567]
761. Pithayanukul P, Nithitanakool S, Bavovada R. Hepatoprotective potential of extracts from seeds of Areca catechu and nutgalls of Quercus infectoria. *Molecules*. 2009; 14:4987–5000. [PubMed: 20032872]
762. Wang LS, Sun XD, Cao Y, Wang L, Li FJ, Wang YF. Antioxidant and pro-oxidant properties of acylated pelargonidin derivatives extracted from red radish (*Raphanus sativus* var. niger, Brassicaceae). *Food Chem Toxicol*.
763. Chaturvedi P. Inhibitory Response of *Raphanus sativus* on Lipid Peroxidation in Albino Rats. *Evid Based Complement Alternat Med*. 2008; 5:55–9. [PubMed: 18317549]
764. Salah-Abbes JB, Abbes S, Ouane S, Houas Z, Abdel-Wahhab MA, Bacha H, et al. Tunisian radish extract (*Raphanus sativus*) enhances the antioxidant status and protects against oxidative stress induced by zearalenone in Balb/c mice. *J Appl Toxicol*. 2008; 28:6–14. [PubMed: 17385802]
765. Gutierrez RM, Perez RL. *Raphanus sativus* (Radish): their chemistry and biology. *ScientificWorldJournal*. 2004; 4:811–37. [PubMed: 15452648]
766. Ilavarasan R, Mallika M, Venkataraman S. Anti-inflammatory and free radical scavenging activity of *Ricinus communis* root extract. *J Ethnopharmacol*. 2006; 103:478–80. [PubMed: 16310994]
767. Shokeen P, Anand P, Murali YK, Tandon V. Antidiabetic activity of 50% ethanolic extract of *Ricinus communis* and its purified fractions. *Food Chem Toxicol*. 2008; 46:3458–66. [PubMed: 18790711]
768. Lomash V, Parihar SK, Jain NK, Katiyar AK. Effect of *Solanum nigrum* and *Ricinus communis* extracts on histamine and carrageenan-induced inflammation in the chicken skin. *Cell Mol Biol (Noisy-le-grand)*. 56(Suppl):OL1239–51. [PubMed: 20158977]
769. Mahmood N, Piacente S, Pizza C, Burke A, Khan AI, Hay AJ. The anti-HIV activity and mechanisms of action of pure compounds isolated from *Rosa damascena*. *Biochem Biophys Res Commun*. 1996; 229:73–9. [PubMed: 8954085]
770. Ramezani R, Moghimi A, Rakhshandeh H, Ejtehadi H, Kheirabadi M. The effect of *Rosa damascena* essential oil on the amygdala electrical kindling seizures in rat. *Pak J Biol Sci*. 2008; 11:746–51. [PubMed: 18819571]
771. Gholamhoseinian A, Fallah H, Sharifi far F. Inhibitory effect of methanol extract of *Rosa damascena* Mill. flowers on alpha-glucosidase activity and postprandial hyperglycemia in normal and diabetic rats. *Phytomedicine*. 2009; 16:935–41. [PubMed: 19380218]
772. Rao GM, Rao CV, Pushpangadan P, Shirwaikar A. Hepatoprotective effects of rubiadin, a major constituent of *Rubia cordifolia* Linn. *J Ethnopharmacol*. 2006; 103:484–90. [PubMed: 16213120]
773. Jain A, Basal E. Inhibition of *Propionibacterium acnes*-induced mediators of inflammation by Indian herbs. *Phytomedicine*. 2003; 10:34–8. [PubMed: 12622461]
774. Patil RA, Jagdale SC, Kasture SB. Antihyperglycemic, antistress and nootropic activity of roots of *Rubia cordifolia* Linn. *Indian J Exp Biol*. 2006; 44:987–92. [PubMed: 17176672]

775. Joy J, Nair CK. Amelioration of cisplatin induced nephrotoxicity in Swiss albino mice by *Rubia cordifolia* extract. *J Cancer Res Ther.* 2008; 4:111–5. [PubMed: 18923202]
776. Islam MS, Rahman MT, Rouf AS, Rahman F. Evaluation of neuropharmacological effects of *Rumex maritimus* Linn. (Polygonaceae) root extracts. *Pharmazie.* 2003; 58:738–41. [PubMed: 14609288]
777. Dwivedi C, Abu-Ghazaleh A. Chemopreventive effects of sandalwood oil on skin papillomas in mice. *Eur J Cancer Prev.* 1997; 6:399–401. [PubMed: 9370104]
778. Matsuo Y, Mimaki Y. Lignans from *Santalum album* and their cytotoxic activities. *Chem Pharm Bull (Tokyo).* 58:587–90. [PubMed: 20410650]
779. Arulmozhi V, Krishnaveni M, Karthishwaran K, Dhamodharan G, Mirunalini S. Antioxidant and antihyperlipidemic effect of *Solanum nigrum* fruit extract on the experimental model against chronic ethanol toxicity. *Pharmacogn Mag.* 6:42–50. [PubMed: 20548935]
780. Kim EJ, Lim SS, Park SY, Shin HK, Kim JS, Park JH. Apoptosis of DU145 human prostate cancer cells induced by dehydrocostus lactone isolated from the root of *Saussurea lappa*. *Food Chem Toxicol.* 2008; 46:3651–8. [PubMed: 18848968]
781. Cho MK, Jang YP, Kim YC, Kim SG. Arctigenin, a phenylpropanoid dibenzylbutyrolactone lignan, inhibits MAP kinases and AP-1 activation via potent MKK inhibition: the role in TNF- α inhibition. *Int Immunopharmacol.* 2004; 4:1419–29. [PubMed: 15313439]
782. Cho JY, Baik KU, Jung JH, Park MH. In vitro anti-inflammatory effects of cynaropicrin, a sesquiterpene lactone, from *Saussurea lappa*. *Eur J Pharmacol.* 2000; 398:399–407. [PubMed: 10862830]
783. Cho JY, Kim AR, Jung JH, Chun T, Rhee MH, Yoo ES. Cytotoxic and pro-apoptotic activities of cynaropicrin, a sesquiterpene lactone, on the viability of leukocyte cancer cell lines. *Eur J Pharmacol.* 2004; 492:85–94. [PubMed: 15178350]
784. Kang JS, Yoon YD, Lee KH, Park SK, Kim HM. Costunolide inhibits interleukin-1 β expression by down-regulation of AP-1 and MAPK activity in LPS-stimulated RAW 264. 7 cells. *Biochem Biophys Res Commun.* 2004; 313:171–7. [PubMed: 14672714]
785. Jainu M, Devi CS. Antiulcerogenic and ulcer healing effects of *Solanum nigrum* (L) on experimental ulcer models: possible mechanism for the inhibition of acid formation. *J Ethnopharmacol.* 2006; 104:156–63. [PubMed: 16202548]
786. Kumar P, Kalonia H, Kumar A. Protective effect of sesamol against 3-nitropropionic acid-induced cognitive dysfunction and altered glutathione redox balance in rats. *Basic Clin Pharmacol Toxicol.* 107:577–82. [PubMed: 20102363]
787. Kumar PP, Kuttan G. *Vernonia cinerea* L. scavenges free radicals and regulates nitric oxide and proinflammatory cytokines profile in carrageenan induced paw edema model. *Immunopharmacol Immunotoxicol.* 2009; 31:94–102. [PubMed: 19234957]
788. Visavadiya NP, Narasimhacharya AV. Sesame as a hypocholesterolaemic and antioxidant dietary component. *Food Chem Toxicol.* 2008; 46:1889–95. [PubMed: 18353516]
789. Visavadiya NP, Soni B, Dalwadi N. Free radical scavenging and antiatherogenic activities of *Sesamum indicum* seed extracts in chemical and biological model systems. *Food Chem Toxicol.* 2009; 47:2507–15. [PubMed: 19607871]
790. Xu H, Yang X, Yang J, Qi W, Liu C, Yang Y. Antitumor effect of alcohol extract from *Sesamum indicum* flower on S180 and H22 experimental tumor. *Zhong Yao Cai.* 2003; 26:272–3. [PubMed: 14528695]
791. Takeuchi H, Mooi LY, Inagaki Y, He P. Hypoglycemic effect of a hot-water extract from defatted sesame (*Sesamum indicum* L) seed on the blood glucose level in genetically diabetic KK-Ay mice. *Biosci Biotechnol Biochem.* 2001; 65:2318–21. [PubMed: 11758931]
792. Franzotti EM, Santos CV, Rodrigues HM, Mourao RH, Andrade MR, Antoniolli AR. Anti-inflammatory, analgesic activity and acute toxicity of *Sida cordifolia* L. (Malva-branca). *J Ethnopharmacol.* 2000; 72:273–7. [PubMed: 10967481]
793. Medeiros IA, Santos MR, Nascimento NM, Duarte JC. Cardiovascular effects of *Sida cordifolia* leaves extract in rats. *Fitoterapia.* 2006; 77:19–27. [PubMed: 16257496]

794. Philip BK, Muralidharan A, Natarajan B, Varadamurthy S, Venkataraman S. Preliminary evaluation of anti-pyretic and anti-ulcerogenic activities of *Sida cordifolia* methanolic extract. *Fitoterapia*. 2008; 79:229–31. [PubMed: 18325683]
795. Sumanth M, Mustafa SS. Antistress, Adoptogenic Activity of *Sida cordifolia* Roots in Mice. *Indian J Pharm Sci*. 2009; 71:323–4. [PubMed: 20490305]
796. Swathy SS, Panicker S, Nithya RS, Anuja MM, Rejitha S, Indira M. Antiperoxidative and Antiinflammatory Effect of *Sida Cordifolia* Linn. on Quinolinic Acid Induced Neurotoxicity. *Neurochem Res*.
797. Reyes BA, Bautista ND, Tanquilut NC, Anunciado RV, Leung AB, Sanchez GC, et al. Anti-diabetic potentials of *Momordica charantia* and *Andrographis paniculata* and their effects on estrous cyclicity of alloxan-induced diabetic rats. *J Ethnopharmacol*. 2006; 105:196–200. [PubMed: 16298503]
798. Heo KS, Lee SJ, Ko JH, Lim K, Lim KT. Glycoprotein isolated from *Solanum nigrum* L. inhibits the DNA-binding activities of NF-kappaB and AP-1, and increases the production of nitric oxide in TPA-stimulated MCF-7 cells. *Toxicol In Vitro*. 2004; 18:755–63. [PubMed: 15465640]
799. Yang MY, Hsu LS, Peng CH, Shi YS, Wu CH, Wang CJ. Polyphenol-rich extracts from *Solanum nigrum* attenuated PKC alpha-mediated migration and invasion of hepatocellular carcinoma cells. *J Agric Food Chem*. 58:5806–14. [PubMed: 20349911]
800. Hsieh CC, Fang HL, Lina WC. Inhibitory effect of *Solanum nigrum* on thioacetamide-induced liver fibrosis in mice. *J Ethnopharmacol*. 2008; 119:117–21. [PubMed: 18606216]
801. Wannang NN, Anuka JA, Kwanashie HO, Gyang SS, Auta A. Anti-seizure activity of the aqueous leaf extract of *Solanum nigrum* linn (solanaceae) in experimental animals. *Afr Health Sci*. 2008; 8:74–9. [PubMed: 19357754]
802. Li J, Li QW, Gao DW, Han ZS, Lu WZ. Antitumor and immunomodulating effects of polysaccharides isolated from *Solanum nigrum* Linne. *Phytother Res*. 2009; 23:1524–30. [PubMed: 19449342]
803. Zakaria ZA, Sulaiman MR, Morsid NA, Aris A, Zainal H, Pojan NH, et al. Antinociceptive, anti-inflammatory and antipyretic effects of *Solanum nigrum* aqueous extract in animal models. *Methods Find Exp Clin Pharmacol*. 2009; 31:81–8. [PubMed: 19455262]
804. Kar DM, Maharana L, Pattnaik S, Dash GK. Studies on hypoglycaemic activity of *Solanum xanthocarpum* Schrad. & Wendl. fruit extract in rats. *J Ethnopharmacol*. 2006; 108:251–6. [PubMed: 16829003]
805. Fonseca LC, Dadarkar SS, Lobo AS, Suthar AC, Chauhan VS, Chandrababu S, et al. 7-hydroxyfrullanolide, a sesquiterpene lactone, inhibits pro-inflammatory cytokine production from immune cells and is orally efficacious in animal models of inflammation. *Eur J Pharmacol*.
806. Bafna AR, Mishra SH. Protective effect of bioactive fraction of *Sphaeranthus indicus* Linn. against cyclophosphamide induced suppression of humoral immunity in mice. *J Ethnopharmacol*. 2006; 104:426–9. [PubMed: 16289412]
807. Chandrashekhar VM, Muchandi AA, Sudi SV, Ganapthy S. Hepatoprotective activity of *Stereospermum suaveolens* against CCl4-induced liver damage in albino rats. *Pharm Biol*. 48:524–8. [PubMed: 20645794]
808. Yin W, Wang TS, Yin FZ, Cai BC. Analgesic and anti-inflammatory properties of brucine and brucine N-oxide extracted from seeds of *Strychnos nux-vomica*. *J Ethnopharmacol*. 2003; 88:205–14. [PubMed: 12963144]
809. Deng XK, Yin W, Li WD, Yin FZ, Lu XY, Zhang XC, et al. The anti-tumor effects of alkaloids from the seeds of *Strychnos nux-vomica* on HepG2 cells and its possible mechanism. *J Ethnopharmacol*. 2006; 106:179–86. [PubMed: 16442763]
810. Rao PS, Ramanadham M, Prasad MN. Anti-proliferative and cytotoxic effects of *Strychnos nux-vomica* root extract on human multiple myeloma cell line - RPMI 8226. *Food Chem Toxicol*. 2009; 47:283–8. [PubMed: 19027818]
811. Saha P, Mandal S, Das A, Das PC, Das S. Evaluation of the anticarcinogenic activity of *Swertia chirata* Buch. Ham, an Indian medicinal plant, on DMBA-induced mouse skin carcinogenesis model. *Phytother Res*. 2004; 18:373–8. [PubMed: 15173996]

812. Rafatullah S, Tariq M, Mossa JS, al-Yahya MA, al-Said MS, Ageel AM. Protective effect of *Swertia chirata* against indomethacin and other ulcerogenic agent-induced gastric ulcers. *Drugs Exp Clin Res.* 1993; 19:69–73. [PubMed: 8223145]
813. Srivastava KC, Malhotra N. Acetyl eugenol, a component of oil of cloves (*Syzygium aromaticum* L) inhibits aggregation and alters arachidonic acid metabolism in human blood platelets. *Prostaglandins Leukot Essent Fatty Acids.* 1991; 42:73–81. [PubMed: 2011614]
814. Banerjee S, Das S. Anticarcinogenic effects of an aqueous infusion of cloves on skin carcinogenesis. *Asian Pac J Cancer Prev.* 2005; 6:304–8. [PubMed: 16235990]
815. Tanko Y, Mohammed A, Okasha MA, Umar AH, Magaji RA. Anti-nociceptive and anti-inflammatory activities of ethanol extract of *syzygium aromaticum* flower bud in Wistar rats and mice. *Afr J Tradit Complement Altern Med.* 2008; 5:209–12. [PubMed: 20161939]
816. Chaieb K, Hajlaoui H, Zmantar T, Kahla-Nakbi AB, Rouabhia M, Mahdouani K, et al. The chemical composition and biological activity of clove essential oil, *Eugenia caryophyllata* (*Syzygium aromaticum* L. Myrtaceae): a short review. *Phytother Res.* 2007; 21:501–6. [PubMed: 17380552]
817. Mandal S, Barik B, Mallick C, De D, Ghosh D. Therapeutic effect of ferulic acid, an ethereal fraction of ethanolic extract of seed of *Syzygium cumini* against streptozotocin-induced diabetes in male rat. *Methods Find Exp Clin Pharmacol.* 2008; 30:121–8. [PubMed: 18560627]
818. Helmstadter A. *Syzygium cumini* (L) SKEELS (Myrtaceae) against diabetes--125 years of research. *Pharmazie.* 2008; 63:91–101. [PubMed: 18380393]
819. Muruganandan S, Pant S, Srinivasan K, Chandra S, Tandan SK, Lal J, et al. Inhibitory role of *Syzygium cumini* on autacoid-induced inflammation in rats. *Indian J Physiol Pharmacol.* 2002; 46:482–6. [PubMed: 12683225]
820. Muruganandan S, Srinivasan K, Chandra S, Tandan SK, Lal J, Raviprakash V. Anti-inflammatory activity of *Syzygium cumini* bark. *Fitoterapia.* 2001; 72:369–75. [PubMed: 11395258]
821. Brito FA, Lima LA, Ramos MF, Nakamura MJ, Cavalher-Machado SC, Siani AC, et al. Pharmacological study of anti-allergic activity of *Syzygium cumini* (L) Skeels. *Braz J Med Biol Res.* 2007; 40:105–15. [PubMed: 17225003]
822. Parmar J, Sharma P, Verma P, Goyal PK. Chemopreventive action of *Syzygium cumini* on DMBA-induced skin papillomagenesis in mice. *Asian Pac J Cancer Prev.* 11:261–5. [PubMed: 20593968]
823. Sehrawat A, Sultana S. *Tamarix gallica* ameliorates thioacetamide-induced hepatic oxidative stress and hyperproliferative response in Wistar rats. *J Enzyme Inhib Med Chem.* 2006; 21:215–23. [PubMed: 16789436]
824. Sun FY, Chen XP, Wang JH, Qin HL, Yang SR, Du GH. Arjunic acid, a strong free radical scavenger from *Terminalia arjuna*. *Am J Chin Med.* 2008; 36:197–207. [PubMed: 18306462]
825. Kuo PL, Hsu YL, Lin TC, Chang JK, Lin CC. Induction of cell cycle arrest and apoptosis in human non-small cell lung cancer A549 cells by casuarinin from the bark of *Terminalia arjuna* Linn. *Anticancer Drugs.* 2005; 16:409–15. [PubMed: 15746577]
826. Chen CH, Liu TZ, Kuo TC, Lu FJ, Chen YC, Chang-Chien YW, et al. Casuarinin protects cultured MDCK cells from hydrogen peroxide-induced oxidative stress and DNA oxidative damage. *Planta Med.* 2004; 70:1022–6. [PubMed: 15549656]
827. Ali A, Kaur G, Hamid H, Abdullah T, Ali M, Niwa M, et al. Terminoside A, a new triterpene glycoside from the bark of *Terminalia arjuna* inhibits nitric oxide production in murine macrophages. *J Asian Nat Prod Res.* 2003; 5:137–42. [PubMed: 12765198]
828. Gupta R, Singhal S, Goyle A, Sharma VN. Antioxidant and hypocholesterolaemic effects of *Terminalia arjuna* tree-bark powder: a randomised placebo-controlled trial. *J Assoc Physicians India.* 2001; 49:231–5. [PubMed: 11225136]
829. Devi RS, Kist M, Vani G, Devi CS. Effect of methanolic extract of *Terminalia arjuna* against *Helicobacter pylori* 26695 lipopolysaccharide-induced gastric ulcer in rats. *J Pharm Pharmacol.* 2008; 60:505–14. [PubMed: 18380924]
830. Kumar S, Enjamoori R, Jaiswal A, Ray R, Seth S, Maulik SK. Catecholamine-induced myocardial fibrosis and oxidative stress is attenuated by *Terminalia arjuna* (Roxb.) *J Pharm Pharmacol.* 2009; 61:1529–36. [PubMed: 19903379]

831. Dwivedi S. Terminalia arjuna Wight & Arn--a useful drug for cardiovascular disorders. *J Ethnopharmacol.* 2007; 114:114–29. [PubMed: 17875376]
832. Tariq M, Hussain SJ, Asif M, Jahan M. Protective effect of fruit extracts of *Emblica officinalis* (Gaertn.) & *Terminalia belerica* (Roxb) in experimental myocardial necrosis in rats. *Indian J Exp Biol.* 1977; 15:485–6. [PubMed: 598885]
833. Shaila HP, Udupa SL, Udupa AL. Hypolipidemic activity of three indigenous drugs in experimentally induced atherosclerosis. *Int J Cardiol.* 1998; 67:119–24. [PubMed: 9891944]
834. Srikumar R, Parthasarathy NJ, Manikandan S, Narayanan GS, Sheeladevi R. Effect of Triphala on oxidative stress and on cell-mediated immune response against noise stress in rats. *Mol Cell Biochem.* 2006; 283:67–74. [PubMed: 16444587]
835. Sabu MC, Kuttan R. Antidiabetic and antioxidant activity of *Terminalia belerica*. *Roxb Indian J Exp Biol.* 2009; 47:270–5.
836. Hazra B, Sarkar R, Biswas S, Mandal N. Comparative study of the antioxidant and reactive oxygen species scavenging properties in the extracts of the fruits of *Terminalia chebula*, *Terminalia belerica* and *Emblica officinalis*. *BMC Complement Altern Med.* 10:20. [PubMed: 20462461]
837. Reddy DB, Reddy TC, Jyotsna G, Sharan S, Priya N, Lakshmipathi V, et al. Chebulagic acid, a COX-LOX dual inhibitor isolated from the fruits of *Terminalia chebula* Retz. induces apoptosis in COLO-205 cell line. *J Ethnopharmacol.* 2009; 124:506–12. [PubMed: 19481594]
838. Lee SI, Hyun PM, Kim SH, Kim KS, Lee SK, Kim BS, et al. Suppression of the onset and progression of collagen-induced arthritis by chebulagic acid screened from a natural product library. *Arthritis Rheum.* 2005; 52:345–53. [PubMed: 15641090]
839. Sabu MC, Kuttan R. Anti-diabetic activity of medicinal plants and its relationship with their antioxidant property. *J Ethnopharmacol.* 2002; 81:155–60. [PubMed: 12065146]
840. Amit A, Saxena VS, Pratibha N, D'Souza P, Bagchi M, Bagchi D, et al. Mast cell stabilization, lipoxygenase inhibition, hyaluronidase inhibition, antihistaminic and antispasmodic activities of Aller-7, a novel botanical formulation for allergic rhinitis. *Drugs Exp Clin Res.* 2003; 29:107–15. [PubMed: 14708456]
841. Prasad L, Husain Khan T, Jahangir T, Sultana S. Chemomodulatory effects of *Terminalia chebula* against nickel chloride induced oxidative stress and tumor promotion response in male Wistar rats. *J Trace Elem Med Biol.* 2006; 20:233–9. [PubMed: 17098582]
842. Patel CD, Modi VD, Chakraborty BS, Mathuria N, Dadhaniya P, Borade PA, et al. Effect of a novel antiinflammatory polyherbal preparation (Sudarshanam oil) on hematological parameters in Wistar rats. *Acta Pol Pharm.* 67:277–81. [PubMed: 20524430]
843. Jukic M, Politeo O, Maksimovic M, Milos M. In vitro acetylcholinesterase inhibitory properties of thymol, carvacrol and their derivatives thymoquinone and thymohydroquinone. *Phytother Res.* 2007; 21:259–61. [PubMed: 17186491]
844. Okazaki K, Kawazoe K, Takaishi Y. Human platelet aggregation inhibitors from thyme (*Thymus vulgaris* L). *Phytother Res.* 2002; 16:398–9. [PubMed: 12112303]
845. Singh N, Singh SM, Shrivastava P. Effect of *Tinospora cordifolia* on the antitumor activity of tumor-associated macrophages-derived dendritic cells. *Immunopharmacol Immunotoxicol.* 2005; 27:1–14. [PubMed: 15803856]
846. Bafna PA, Balaraman R. Anti-ulcer and anti-oxidant activity of pepticare, a herbomineral formulation. *Phytomedicine.* 2005; 12:264–70. [PubMed: 15898703]
847. Desai VR, Ramkrishnan R, Chintalwar GJ, Sainis KB. G1–4A, an immunomodulatory polysaccharide from *Tinospora cordifolia*, modulates macrophage responses and protects mice against lipopolysaccharide induced endotoxic shock. *Int Immunopharmacol.* 2007; 7:1375–86. [PubMed: 17673153]
848. Patel SS, Shah RS, Goyal RK. Antihyperglycemic, antihyperlipidemic and antioxidant effects of Dihar, a polyherbal ayurvedic formulation in streptozotocin induced diabetic rats. *Indian J Exp Biol.* 2009; 47:564–70. [PubMed: 19761040]
849. Akhtar S. Use of *Tinospora cordifolia* in HIV infection. *Indian J Pharmacol.* 42:57. [PubMed: 20606842]

850. Kaur T, Bijarnia RK, Singla SK, Tandon C. In vivo efficacy of *Trachyspermum ammi* anticalcifying protein in urolithiatic rat model. *J Ethnopharmacol.* 2009; 126:459–62. [PubMed: 19781619]
851. Zhang S, Li H, Yang SJ. Tribulosin protects rat hearts from ischemia/reperfusion injury. *Acta Pharmacol Sin.* 31:671–8. [PubMed: 20453871]
852. Sun B, Qu W, Bai Z. The inhibitory effect of saponins from *Tribulus terrestris* on Bcap-37 breast cancer cell line in vitro. *Zhong Yao Cai.* 2003; 26:104–6. [PubMed: 12795220]
853. Liu XM, Huang QF, Zhang YL, Lou JL, Liu HS, Zheng H. Effects of *Tribulus terrestris* L. saponin on apoptosis of cortical neurons induced by hypoxia-reoxygenation in rats. *Zhong Xi Yi Jie He Xue Bao.* 2008; 6:45–50. [PubMed: 18184546]
854. Heidari MR, Mehrabani M, Pardakhty A, Khazaeli P, Zahedi MJ, Yakhchali M, et al. The analgesic effect of *Tribulus terrestris* extract and comparison of gastric ulcerogenicity of the extract with indomethacin in animal experiments. *Ann N Y Acad Sci.* 2007; 1095:418–27. [PubMed: 17404054]
855. Tuncer MA, Yamacı B, Sati L, Caylı S, Acar G, Altug T, et al. Influence of *Tribulus terrestris* extract on lipid profile and endothelial structure in developing atherosclerotic lesions in the aorta of rabbits on a high-cholesterol diet. *Acta Histochem.* 2009; 111:488–500. [PubMed: 19269683]
856. Raju J, Patlolla JM, Swamy MV, Rao CV. Diosgenin, a steroid saponin of *Trigonella foenum graecum* (Fenugreek), inhibits azoxymethane-induced aberrant crypt foci formation in F344 rats and induces apoptosis in HT-29 human colon cancer cells. *Cancer Epidemiol Biomarkers Prev.* 2004; 13:1392–8. [PubMed: 15298963]
857. Uemura T, Hirai S, Mizoguchi N, Goto T, Lee JY, Taketani K, et al. Diosgenin present in fenugreek improves glucose metabolism by promoting adipocyte differentiation and inhibiting inflammation in adipose tissues. *Mol Nutr Food Res.*
858. Srichamroen A, Thomson AB, Field CJ, Basu TK. In vitro intestinal glucose uptake is inhibited by galactomannan from Canadian fenugreek seed (*Trigonella foenum graecum* L.) in genetically lean and obese rats. *Nutr Res.* 2009; 29:49–54. [PubMed: 19185777]
859. Pandian RS, Anuradha CV, Viswanathan P. Gastroprotective effect of fenugreek seeds (*Trigonella foenum graecum*) on experimental gastric ulcer in rats. *J Ethnopharmacol.* 2002; 81:393–7. [PubMed: 12127242]
860. Tahiliani P, Kar A. The combined effects of *Trigonella* and *Allium* extracts in the regulation of hyperthyroidism in rats. *Phytomedicine.* 2003; 10:665–8. [PubMed: 14692727]
861. Raju J, Bird RP. Alleviation of hepatic steatosis accompanied by modulation of plasma and liver TNF- α levels by *Trigonella foenum graecum* (fenugreek) seeds in Zucker obese (fa/fa) rats. *Int J Obes (Lond).* 2006; 30:1298–307. [PubMed: 16477270]
862. Hannan JM, Ali L, Rokeya B, Khaleque J, Akhter M, Flatt PR, et al. Soluble dietary fibre fraction of *Trigonella foenum-graecum* (fenugreek) seed improves glucose homeostasis in animal models of type 1 and type 2 diabetes by delaying carbohydrate digestion and absorption, and enhancing insulin action. *Br J Nutr.* 2007; 97:514–21. [PubMed: 17313713]
863. Vyas S, Agrawal RP, Solanki P, Trivedi P. Analgesic and anti-inflammatory activities of *Trigonella foenum-graecum* (seed) extract. *Acta Pol Pharm.* 2008; 65:473–6. [PubMed: 19051589]
864. Gupta SK, Kalaiselvan V, Srivastava S, Saxena R, Agrawal SS. *Trigonella foenum-graecum* (Fenugreek) Protects Against Selenite-Induced Oxidative Stress in Experimental Cataractogenesis. *Biol Trace Elem Res.* 2009
865. Reddy RL, Srinivasan K. Fenugreek seeds reduce atherogenic diet-induced cholesterol gallstone formation in experimental mice. *Can J Physiol Pharmacol.* 2009; 87:933–43. [PubMed: 19935901]
866. Varjas T, Nowrasteh G, Budan F, Horvath G, Cseh J, Gyongyi Z, et al. The effect of fenugreek on the gene expression of arachidonic acid metabolizing enzymes. *Phytother Res.*
867. Rehni AK, Pantlya HS, Shri R, Singh M. Effect of chlorophyll and aqueous extracts of *Bacopa monniera* and *Valeriana wallichii* on ischaemia and reperfusion-induced cerebral injury in mice. *Indian J Exp Biol.* 2007; 45:764–9. [PubMed: 17907741]

868. Subhan F, Karim N, Gilani AH, Sewell RD. Terpenoid content of *Valeriana wallichii* extracts and antidepressant-like response profiles. *Phytother Res.* 24:686–91. [PubMed: 19943315]
869. Prasad DN, Achari G. A study of anti-arthritic action of *Vanda roxburghii* in albino rats. *J Indian Med Assoc.* 1966; 46:234–7. [PubMed: 5906161]
870. Latha RM, Geetha T, Varalakshmi P. Effect of *Vernonia cinerea* Less flower extract in adjuvant-induced arthritis. *Gen Pharmacol.* 1998; 31:601–6. [PubMed: 9792223]
871. Mazumder UK, Gupta M, Manikandan L, Bhattacharya S, Halder PK, Roy S. Evaluation of anti-inflammatory activity of *Vernonia cinerea* Less. extract in rats. *Phytomedicine.* 2003; 10:185–8. [PubMed: 12725574]
872. Iwalewa EO, Iwalewa OJ, Adeboye JO. Analgesic, antipyretic, anti-inflammatory effects of methanol, chloroform and ether extracts of *Vernonia cinerea* less leaf. *J Ethnopharmacol.* 2003; 86:229–34. [PubMed: 12738092]
873. Leelarungrayub D, Pratanaphon S, Pothongsunon P, Sriboonreung T, Yankai A, Bloomer RJ. *Vernonia cinerea* Less. supplementation and strenuous exercise reduce smoking rate: relation to oxidative stress status and beta-endorphin release in active smokers. *J Int Soc Sports Nutr.* 7:21. [PubMed: 20500899]
874. Gerlach SL, Rathinakumar R, Chakravarty G, Goransson U, Wimley WC, Darwin SP, et al. Anticancer and chemosensitizing abilities of cycloviolacin O2 from *Viola odorata* and psyle cyclotides from *psychotria leptothyrsa*. *Biopolymers.*
875. Ebrahimzadeh MA, Nabavi SM, Nabavi SF, Bahramian F, Bekhradnia AR. Antioxidant and free radical scavenging activity of *H. officinalis* L. var. *angustifolius*, *V. odorata*, *B. hyrcana* and *C. speciosum*. *Pak J Pharm Sci.* 23:29–34. [PubMed: 20067863]
876. Zheng CJ, Huang BK, Wang Y, Ye Q, Han T, Zhang QY, et al. Anti-inflammatory diterpenes from the seeds of *Vitex negundo*. *Bioorg Med Chem.* 18:175–81. [PubMed: 19931461]
877. Chawla AS, Sharma AK, Handa SS, Dhar KL. Chemical investigation and anti-inflammatory activity of *Vitex negundo* seeds. *J Nat Prod.* 1992; 55:163–7. [PubMed: 1624939]
878. Dharmasiri MG, Jayakody JR, Galhena G, Liyanage SS, Ratnasooriya WD. Anti-inflammatory and analgesic activities of mature fresh leaves of *Vitex negundo*. *J Ethnopharmacol.* 2003; 87:199–206. [PubMed: 12860308]
879. Tandon VR, Gupta RK. *Vitex negundo* Linn (VN) leaf extract as an adjuvant therapy to standard anti-inflammatory drugs. *Indian J Med Res.* 2006; 124:447–50. [PubMed: 17159267]
880. Rooban BN, Lija Y, Biju PG, Sasikala V, Sahasranamam V, Abraham A. *Vitex negundo* attenuates calpain activation and cataractogenesis in selenite models. *Exp Eye Res.* 2009; 88:575–82. [PubMed: 19094987]
881. Maitra R, Porter MA, Huang S, Gilmour BP. Inhibition of NF κ B by the natural product Withaferin A in cellular models of Cystic Fibrosis inflammation. *J Inflamm (Lond).* 2009; 6:15. [PubMed: 19439083]
882. Park HJ, Rayalam S, Della-Fera MA, Ambati S, Yang JY, Baile CA. Withaferin A induces apoptosis and inhibits adipogenesis in 3T3-L1 adipocytes. *Biofactors.* 2008; 33:137–48. [PubMed: 19346589]
883. Mishra LC, Singh BB, Dagenais S. Scientific basis for the therapeutic use of *Withania somnifera* (ashwagandha): a review. *Altern Med Rev.* 2000; 5:334–46. [PubMed: 10956379]
884. Kour K, Pandey A, Suri KA, Satti NK, Gupta KK, Bani S. Restoration of stress-induced altered T cell function and corresponding cytokines patterns by Withanolide A. *Int Immunopharmacol.* 2009; 9:1137–44. [PubMed: 19524704]
885. Jayaprakasam B, Padmanabhan K, Nair MG. Withanamides in *Withania somnifera* fruit protect PC-12 cells from beta-amyloid responsible for Alzheimer's disease. *Phytother Res.* 24:859–63. [PubMed: 19957250]
886. Sumantran VN, Chandwaskar R, Joshi AK, Boddul S, Patwardhan B, Chopra A, et al. The relationship between chondroprotective and antiinflammatory effects of *Withania somnifera* root and glucosamine sulphate on human osteoarthritic cartilage in vitro. *Phytother Res.* 2008; 22:1342–8. [PubMed: 18697233]

887. Bhatnagar M, Sisodia SS, Bhatnagar R. Antiulcer and antioxidant activity of *Asparagus racemosus* Willd and *Withania somnifera* Dunal in rats. *Ann N Y Acad Sci.* 2005; 1056:261–78. [PubMed: 16387694]
888. Udayakumar R, Kasthuriangan S, Mariashibu TS, Rajesh M, Anbazhagan VR, Kim SC, et al. Hypoglycaemic and hypolipidaemic effects of *Withania somnifera* root and leaf extracts on alloxan-induced diabetic rats. *Int J Mol Sci.* 2009; 10:2367–82. [PubMed: 19564954]
889. Jeyanthi T, Subramanian P. Protective effect of *Withania somnifera* root powder on lipid peroxidation and antioxidant status in gentamicin-induced nephrotoxic rats. *J Basic Clin Physiol Pharmacol.* 21:61–78. [PubMed: 20506689]
890. Kim JK, Kim Y, Na KM, Surh YJ, Kim TY. [6]-Gingerol prevents UVB-induced ROS production and COX-2 expression in vitro and in vivo. *Free Radic Res.* 2007; 41:603–14. [PubMed: 17454143]
891. Dugasani S, Pichika MR, Nadarajah VD, Balijepalli MK, Tandra S, Korlakunta JN. Comparative antioxidant and anti-inflammatory effects of [6]-gingerol, [8]-gingerol, [10]-gingerol and [6]-shogaol. *J Ethnopharmacol.* 127:515–20. [PubMed: 19833188]
892. Pan MH, Hsieh MC, Hsu PC, Ho SY, Lai CS, Wu H, et al. 6-Shogaol suppressed lipopolysaccharide-induced up-expression of iNOS and COX-2 in murine macrophages. *Mol Nutr Food Res.* 2008; 52:1467–77. [PubMed: 18683823]
893. Pan MH, Hsieh MC, Kuo JM, Lai CS, Wu H, Sang S, et al. 6-Shogaol induces apoptosis in human colorectal carcinoma cells via ROS production, caspase activation, and GADD 153 expression. *Mol Nutr Food Res.* 2008; 52:527–37. [PubMed: 18384088]
894. Sang S, Hong J, Wu H, Liu J, Yang CS, Pan MH, et al. Increased growth inhibitory effects on human cancer cells and anti-inflammatory potency of shogaols from *Zingiber officinale* relative to gingerols. *J Agric Food Chem.* 2009; 57:10645–50. [PubMed: 19877681]
895. Brown AC, Shah C, Liu J, Pham JT, Zhang JG, Jadus MR. Ginger's (*Zingiber officinale* Roscoe) inhibition of rat colonic adenocarcinoma cells proliferation and angiogenesis in vitro. *Phytother Res.* 2009; 23:640–5. [PubMed: 19117330]
896. Guahk GH, Ha SK, Jung HS, Kang C, Kim CH, Kim YB, et al. *Zingiber officinale* protects HaCaT cells and C57BL/6 mice from ultraviolet B-induced inflammation. *J Med Food.* 13:673–80. [PubMed: 20521990]
897. Lantz RC, Chen GJ, Sarihan M, Solyom AM, Jolad SD, Timmermann BN. The effect of extracts from ginger rhizome on inflammatory mediator production. *Phytomedicine.* 2007; 14:123–8. [PubMed: 16709450]
898. Al-Amin ZM, Thomson M, Al-Qattan KK, Peltonen-Shalaby R, Ali M. Anti-diabetic and hypolipidaemic properties of ginger (*Zingiber officinale*) in streptozotocin-induced diabetic rats. *Br J Nutr.* 2006; 96:660–6. [PubMed: 17010224]
899. Ahui ML, Champy P, Ramadan A, Pham Van L, Araujo L, Brou Andre K, et al. Ginger prevents Th2-mediated immune responses in a mouse model of airway inflammation. *Int Immunopharmacol.* 2008; 8:1626–32. [PubMed: 18692598]
900. Habib SH, Makpol S, Abdul Hamid NA, Das S, Ngah WZ, Yusof YA. Ginger extract (*Zingiber officinale*) has anti-cancer and anti-inflammatory effects on ethionine-induced hepatoma rats. *Clinics (Sao Paulo).* 2008; 63:807–13. [PubMed: 19061005]
901. Lakshmi BV, Sudhakar M. Attenuation of acute and chronic restraint stress-induced perturbations in experimental animals by *Zingiber officinale* Roscoe. *Food Chem Toxicol.* 48:530–5. [PubMed: 19909780]
902. Grzanna R, Lindmark L, Frondoza CG. Ginger--an herbal medicinal product with broad anti-inflammatory actions. *J Med Food.* 2005; 8:125–32. [PubMed: 16117603]
903. Kundu JK, Na HK, Surh YJ. Ginger-derived phenolic substances with cancer preventive and therapeutic potential. *Forum Nutr.* 2009; 61:182–92. [PubMed: 19367122]
904. Anggakusuma, Yanti; Hwang, JK. Effects of macelignan isolated from *Myristica fragrans* Hoult. on UVB-induced matrix metalloproteinase-9 and cyclooxygenase-2 in HaCaT cells. *J Dermatol Sci.* 57:114–22. [PubMed: 19914807]

905. Ma J, Hwang YK, Cho WH, Han SH, Hwang JK, Han JS. Macelignan attenuates activations of mitogen-activated protein kinases and nuclear factor kappa B induced by lipopolysaccharide in microglial cells. *Biol Pharm Bull.* 2009; 32:1085–90. [PubMed: 19483320]
906. Akev N, Turkay G, Can A, Gurel A, Yildiz F, Yardibi H, et al. Tumour preventive effect of Aloe vera leaf pulp lectin (Aloctin I) on Ehrlich ascites tumours in mice. *Phytother Res.* 2007; 21:1070–5. [PubMed: 17685385]
907. Shishodia S, Sethi G, Aggarwal BB. Curcumin: getting back to the roots. *Ann N Y Acad Sci.* 2005; 1056:206–17. [PubMed: 16387689]
908. Hong J, Sang S, Park HJ, Kwon SJ, Suh N, Huang MT, et al. Modulation of arachidonic acid metabolism and nitric oxide synthesis by garcinol and its derivatives. *Carcinogenesis.* 2006; 27:278–86. [PubMed: 16093250]
909. Koeberle A, Northoff H, Werz O. Identification of 5-lipoxygenase and microsomal prostaglandin E2 synthase-1 as functional targets of the anti-inflammatory and anti-carcinogenic garcinol. *Biochem Pharmacol.* 2009; 77:1513–21. [PubMed: 19426689]
910. Kim SO, Kundu JK, Shin YK, Park JH, Cho MH, Kim TY, et al. [6]-Gingerol inhibits COX-2 expression by blocking the activation of p38 MAP kinase and NF-kappaB in phorbol ester-stimulated mouse skin. *Oncogene.* 2005; 24:2558–67. [PubMed: 15735738]
911. Liu KL, Chen HW, Wang RY, Lei YP, Sheen LY, Lii CK. DATS reduces LPS-induced iNOS expression, NO production, oxidative stress, and NF-kappaB activation in RAW 264. 7 macrophages. *J Agric Food Chem.* 2006; 54:3472–8. [PubMed: 16637709]
912. Genovese T, Menegazzi M, Mazzon E, Crisafulli C, Di Paola R, Dal Bosco M, et al. Glycyrrhizin reduces secondary inflammatory process after spinal cord compression injury in mice. *Shock.* 2009; 31:367–75. [PubMed: 18665052]
913. Ram A, Mabalirajan U, Das M, Bhattacharya I, Dinda AK, Gangal SV, et al. Glycyrrhizin alleviates experimental allergic asthma in mice. *Int Immunopharmacol.* 2006; 6:1468–77. [PubMed: 16846841]
914. Chiou WF, Chen CF, Lin JJ. Mechanisms of suppression of inducible nitric oxide synthase (iNOS) expression in RAW 264. 7 cells by andrographolide. *Br J Pharmacol.* 2000; 129:1553–60. [PubMed: 10780958]
915. Cho JY, Park J, Yoo ES, Baik KU, Jung JH, Lee J, et al. Inhibitory effect of sesquiterpene lactones from *Saussurea lappa* on tumor necrosis factor-alpha production in murine macrophage-like cells. *Planta Med.* 1998; 64:594–7. [PubMed: 9810262]
916. Oh GS, Pae HO, Chung HT, Kwon JW, Lee JH, Kwon TO, et al. Dehydrocostus lactone enhances tumor necrosis factor-alpha-induced apoptosis of human leukemia HL-60 cells. *Immunopharmacol Immunotoxicol.* 2004; 26:163–75. [PubMed: 15209353]
917. Jin M, Lee HJ, Ryu JH, Chung KS. Inhibition of LPS-induced NO production and NF-kappaB activation by a sesquiterpene from *Saussurea lappa*. *Arch Pharm Res.* 2000; 23:54–8. [PubMed: 10728658]
918. Lampronti I, Khan MT, Bianchi N, Ather A, Borgatti M, Vizziello L, et al. Bangladeshi medicinal plant extracts inhibiting molecular interactions between nuclear factors and target DNA sequences mimicking NF-kappaB binding sites. *Med Chem.* 2005; 1:327–33. [PubMed: 16789890]
919. Murakami A, Ohigashi H. Cancer-preventive anti-oxidants that attenuate free radical generation by inflammatory cells. *Biol Chem.* 2006; 387:387–92. [PubMed: 16606336]
920. Joo HY, Lim K, Lim KT. Phytyglycoprotein (150 kDa) isolated from *Solanum nigrum* Linne has a preventive effect on dextran sodium sulfate-induced colitis in A/J mouse. *J Appl Toxicol.* 2009; 29:207–13. [PubMed: 18988204]
921. Lee YY, Yang SF, Ho WH, Lee YH, Hung SL. Eugenol modulates cyclooxygenase-2 expression through the activation of nuclear factor kappa B in human osteoblasts. *J Endod.* 2007; 33:1177–82. [PubMed: 17889685]
922. Nair PK, Melnick SJ, Ramachandran R, Escalon E, Ramachandran C. Mechanism of macrophage activation by (1,4)-alpha-D-glucan isolated from *Tinospora cordifolia*. *Int Immunopharmacol.* 2006; 6:1815–24. [PubMed: 17052672]

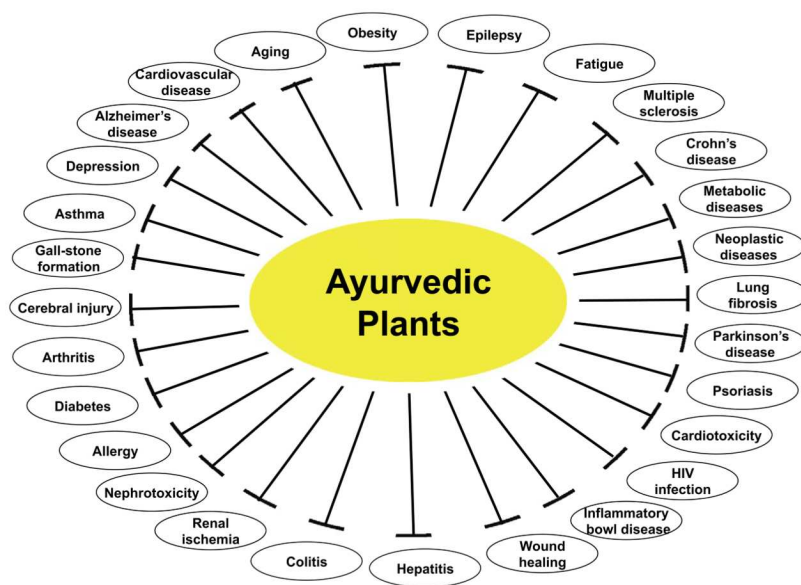


Fig. 1.
The use of Ayurvedic plants for treatment of various chronic diseases.

Inflammatory Network

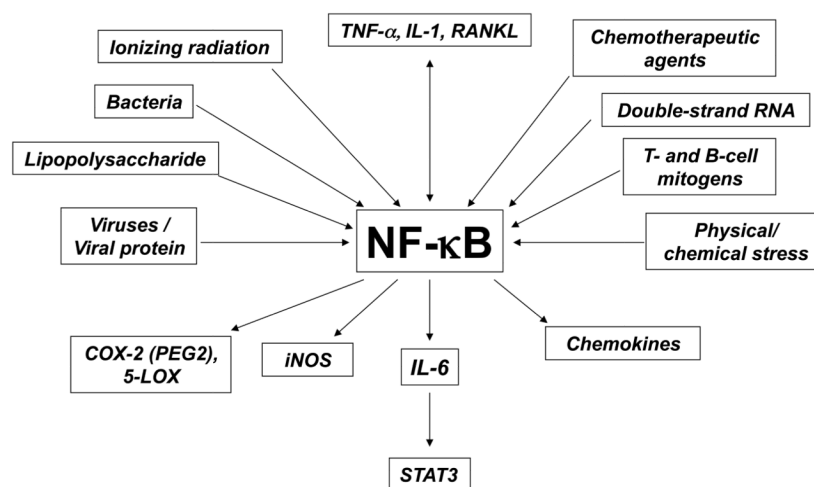


Fig. 2.
Activation of inflammatory network by various agents.

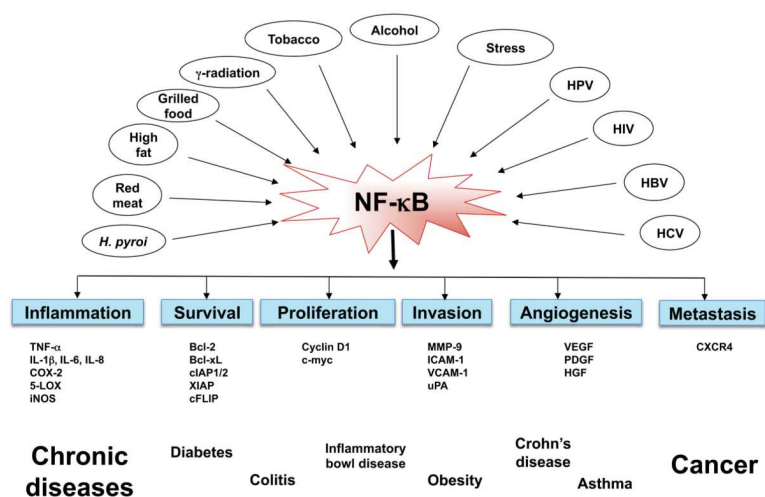


Fig. 3. Activation of inflammatory network by life style factors and its contribution to chronic diseases.

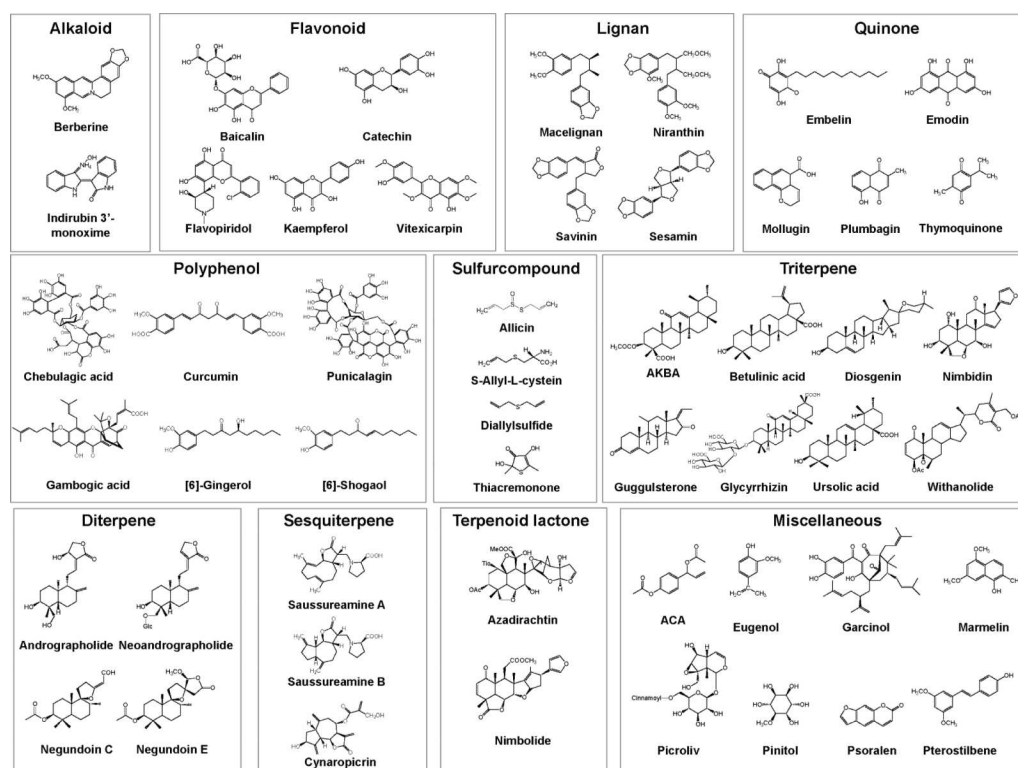


Fig. 4.
Structures of active phytochemicals derived from Ayurvedic plants.

Table 1

Ayurvedic formulations and their uses*

Formulation	Plants used	Uses
Amrutanjana Balm	<i>Cinnamomum camphora</i> , <i>C. zeylanicum</i> , <i>Cymbopogon citrates</i> , <i>Eucalyptus polybractea</i> , <i>Gaultheria</i> sp., <i>Mentha arvensis</i> , <i>M. piperita</i> , <i>Rosa</i> sp., <i>Thymus vulgaris</i>	Cures pain, sprain, cold and sinus
Ayurslim	<i>Commiphora wightii</i>, <i>Garcinia cambogia</i>, <i>G. sylvestre</i>, <i>Terminalia chebula</i>, <i>Trigonella foenum-graecum</i>	Burns fat and reduces cholesterol
Dabur Chyawanprash	<i>Phyllanthus emblica</i> , <i>Piper longum</i> , <i>Pistacia integerrima</i> <i>Pueraria tuberosa</i> , <i>Tinospora cordifolia</i>	Protects from infections, coughs, cold and stress; adjuvant pulmonary tuberculosis therapy
Divya Arshkalpa Vati	<i>Aloe vera</i>, <i>Azadirachta indica</i>, <i>Berberis aristata</i>, <i>C. camphora</i>, <i>Daemenorops draco</i>, <i>Sapindus</i> sp., <i>Solanum nigrum</i>, <i>T. chebula</i>	Cures piles, burning sensation and colic pain
Divya Ashmarihara Kvath	<i>Bergenia ligulata</i> , <i>Boerhavia diffusa</i> , <i>Crataeva nurvula</i> <i>Dolichos biflorus</i> , <i>Tribulus terrestris</i>	Diuretic, anti-oedemous; dissolves kidney, urinary & gall bladder stones
Divya Ashmarihara Rasa	<i>Hajarala yahuda</i> , <i>Hordeum vulgare</i> , <i>Raphanus sativus</i>	Diuretic, dissolves calculi, relieves pains, anti-oedemous.
Divya Danta Manjana	<i>Acacia arabica</i> , <i>Anacyclus pyrethrum</i> , <i>A. indica</i> , <i>C. camphora</i> <i>M. piperita</i> , <i>Mimusops elengi</i> , <i>P. longum</i> , <i>Quercus infectoria</i> , <i>Syzygium aromaticum</i> , <i>Zanthoxylum alatum</i>	Strengthens the gums and stops bleeding
Divya Gaisahara Choorna	<i>Citrus limon</i> , <i>Cuminum cymimum</i> , <i>Ferula foetida</i> , <i>Piper nigra</i> , <i>T. chebula</i> , <i>Trachyspermum ammi</i>	Reduces gas, acidity, flatulence, colic pain & anorexia
Divya Kayakalpa Kvatha	<i>Acacia catechu</i>, <i>A. indica</i>, <i>B. aristata</i>, <i>Cassia tora</i>, <i>Cedrus deodara</i>, <i>Curcuma longa</i>, <i>Leucas cephalotes</i>, <i>Picrorhiza kurroa</i>, <i>Pongamia pinnata</i>, <i>Psorlia corylifolia</i>, <i>Pterocarpus santalinus</i>, <i>Rubia cordifolia</i>, <i>Swertia chirata</i> <i>T. cordifolia</i>	Cures eczema, leprosy, filariasis; helps in reducing obesity
Divya Kayakalp Vati	<i>A. catechu</i>, <i>A. indica</i>, <i>B. aristata</i>, <i>C. tora</i>, <i>C. deodara</i>, <i>Citrullus colocynthis</i>, <i>C. longa</i>, <i>Embllica officinalis</i>, <i>Leucas cephalotes</i>, <i>Nigella sativa</i>, <i>P. kurroa</i>, <i>Pongamia pinnata</i> <i>P. santalinus</i>, <i>R. cordifolia</i>, <i>Sarsa parilla</i>, <i>Solanum xantho-carpum</i>, <i>S. chirata</i>, <i>Terminalia belerica</i>, <i>T. chebula</i>, <i>T. cordifolia</i>	Purifies blood, removes acne, pimples; cures from ring-worms, itches, pruritus, eczema, leucoderma & psoriasis
Divya Madhu-kalpa Vati	<i>Acacia arabica</i> , <i>Aconitum heterophilum</i> , <i>Aegle marmelos</i>, <i>Andrographis paniculata</i>, <i>A. indica</i>, <i>C. longa</i>, <i>C. zedoaria</i> <i>Embllica officinalis</i> , <i>Ficus bengalensis</i> , <i>G. sylvestre</i> , <i>Holarrhena antidysenterica</i> , <i>Momordica charantia</i> , <i>N. sativa</i> , <i>P. kurroa</i> , <i>S. chirata</i> , <i>Syzyium cumini</i> , <i>T. belerica</i> , <i>T. chebula</i>, <i>T. cordifolia</i>, <i>T. terrestris</i>, <i>T. foenum-graecum</i>, <i>Withania somnifera</i>	Balance insulin secretion, strengthens immune system
Divya Medha Kwatha	<i>Bacopa monnieri</i> , <i>Celatrus paniculatus</i> , <i>Convovulus pluricaulis</i> , <i>Foeniculum vulgare</i> , <i>Lavandula stoechas</i> , <i>Nardostachys jatamansi</i> , <i>Onosma bracteatum</i> , <i>W. somnifera</i>	Cures chronic headache, migraine, sleeplessness and depression
Divya Mukta Vati	<i>Acorus calamus</i> , <i>B. monnieri</i> , <i>C. paniculatus</i> , <i>C. pluricaulis</i> , <i>Inula racemosa</i> , <i>L. stoechas</i> , <i>N. jatamansi</i> , <i>Rauwolfia serpentina</i> , <i>T. arjuna</i> , <i>W. somnifera</i>	Cures high blood pressure, insomnia, palpitation, chest pain
Divya Pidantaka Kvatha	<i>Cyperus rotundus</i> , <i>Nyctanthes arbortristis</i> , <i>Piper chaba</i> , <i>P. longum</i> , <i>Pluchea lanceolata</i> , <i>Ricinus communis</i> , <i>T. ammi</i> , <i>Vitex negundo</i>, <i>W. somnifera</i>, <i>Z. officinale</i>	Useful in joint pain, sciatica, osteo-arthritis, gout, rheumatoid arthritis, muscular and skeletal pains and oedema.
Divya Udaramrita Vati	<i>A. marmelos</i>, <i>A. vera</i>, <i>B. diffusa</i>, <i>E. officinalis</i>, <i>Mangifera indica</i>, <i>Operculina turpethum</i>, <i>Phyllanthus niruri</i>, <i>P. kurroa</i>, <i>Plumbago zeylanica</i>, <i>S. nigrum</i>, <i>T. belerica</i>, <i>T. ammi</i>	Cures jaundice, anaemia, chronic fever, diarrhoea and abdominal pain
Divya Yauvanamrita Vati	<i>Anacyclus pyrethrum</i> , <i>Asparagus racemosus</i> , <i>Castorium</i> , <i>Crocus sativus</i> , <i>Mucuna pruriens</i> , <i>Myristica fragrans</i> <i>Sida cordifolia</i>	Strengthens heart and brain, promotes luster and youthness and cures impotency.
Divya Udarakalpa Curna	<i>Cassia angustifolia</i> , <i>F. vulgare</i>, <i>Glycyrrhiza glabra</i>, <i>Rosa centifolia</i>, <i>T. chebula</i>	Stimulates digestion and removes constipation
Divya Kayakalpa Taila	<i>A. indica</i>, <i>B. aristata</i>, <i>C. tora</i>, <i>C. deodara</i>, <i>C. longa</i>, <i>E. officinalis</i>, <i>Leucas cephalotes</i>, <i>N. sativa</i>, <i>P. kurroa</i>, <i>P. pinnata</i>, <i>Psorlia corylifolia</i>, <i>P. santalinus</i>, <i>R. cordifolia</i>, <i>Saphindus trifoliatus</i>, <i>Sarsa parilla</i>, <i>Sesamum indicum</i>, <i>Solanum indicum</i>, <i>S. chirata</i>, <i>T. chebula</i>, <i>T. cordifolia</i>	Cures skin diseases like ring worm, itching, sun burning, eczema, leucoderma, psoriasis, urticaria and skin allergy

Formulation	Plants used	Uses
Divya Kesa Taila	<i>Abrus precatorius</i> , <i>B. monnieri</i> , B. aristata , Callicarpa macrophylla , <i>C. rotundus</i> , <i>Eclipta alba</i> , <i>E. officinalis</i> , <i>Fagonia cretica</i> , Indigofera tinctoria , <i>Mesua ferrea</i> , <i>N. jatamansi</i> , Nelumbo nucifera , <i>Onosma ehioides</i> , <i>Pandanus tectorius</i> , P. santalinus , <i>S. cordifolia</i> , <i>Symplocos crataegoides</i>	Cures untimely hair fall, dandruff, alopecia, premature graying of hair
Divya Churna	F. vulgare , <i>Ipomoea nil</i> , <i>R. centifolia</i> , T. chebula , Z. officinale	Cures abdominal pain, flatulence, heaviness & nausea
Divya Peya (Herbal Tea)	<i>Adhatoda vasita</i> , <i>B. monnieri</i> , B. diffusa , <i>C. zeylanicum</i> , <i>C. pluricaulis</i> , <i>Cymbopogon martini</i> , <i>C. rotundus</i> , <i>Elettaria cardamomum</i> , <i>Ephedra gerardiana</i> , F. vulgare , M. fragrans , Nelumbo nucifera , Ocimum sanctum , <i>P. niruri</i> , <i>P. chaba</i> , <i>P. longum</i> , <i>P. nigrum</i> , P. zeylanica , P. santalinus , <i>R. centifolia</i> , <i>Santalum album</i> , S. aromaticum , <i>T. arjuna</i> , T. cordifolia , <i>Viola odorata</i> , W. somnifera , Z. officinale	Controls cholesterol and protects from heart diseases; promotes immunity, stimulates digestion
Divya Dhara	<i>C. camphora</i> , <i>M. piperita</i> , S. aromaticum , <i>T. ammi</i>	Cures asthma, cholera, ear-diseases, epistaxis, trauma, urticaria, cough, colic pain and flatulence
Divya Pidantaka Rasa	A. marmelos , <i>Clerodendron phlomoides</i> , Commiphora mukul , <i>C. rotundus</i> , <i>Gmelina arborea</i> , <i>Moringa oleifera</i> , <i>Oroxylum indicum</i> , <i>P. lanceolata</i> , <i>Pseudarthria viscida</i> , <i>S. indicum</i> , <i>Stereospermum suaveolens</i> , <i>Strychnos nuxvomica</i> , T. cordifolia , <i>T. ammi</i> , <i>T. terrestris</i> , <i>Urtica lagopoides</i> , V. negundo , W. somnifera	Useful in joint pain, arthritis, lumbar pain, cervical spondylitis and sciatica
Divya Pidantaka Taila	<i>Aconitum ferox</i> , <i>A. calamus</i> , A. marmelos , Allium sativum , <i>Anethum sowa</i> , <i>A. racemosus</i> , B. aristata , <i>Butea mono-sperma</i> , <i>Calotropis procera</i> , <i>C. paniculatus</i> , <i>C. zeylanicum</i> , <i>C. phlomoides</i> , C. longa , <i>Datura metel</i> , <i>E. alba</i> , F. vulgare , G. glabra , <i>G. arborea</i> , <i>Hebenaria intermedia</i> , <i>I. racemosa</i> , <i>Lilium polyphyllum</i> , <i>Malaxis acuminata</i> , <i>M. ferrea</i> , <i>N. jatamansi</i> , <i>Oroxylum indicum</i> , <i>Paderia foetida</i> , <i>P. chaba</i> , <i>P. longum</i> , <i>P. lanceolata</i> , P. zeylanica , <i>Polygonatum verticillatum</i> , <i>Pseudarthria viscida</i> , <i>R. communis</i> , <i>Roscoeia alpina</i> , R. cordifolia , <i>Sesamum indicum</i> , <i>S. indicum</i> , <i>Stereospermum suaveolens</i> , <i>Strychnos nuxvomica</i> , <i>T. ammi</i> , <i>T. terrestris</i> , <i>Urtica lagopoides</i> , <i>Valeriana wallichii</i> , V. negundo , Z. officinale	Relieves pain of lumbar region, knee-joints, cervical spondylitis, oedema & inflammation
Divya Madhunasini Vati	<i>A. arabica</i> , <i>A. heterophilum</i> , A. marmelos , A. paniculata , A. indica , C. longa , <i>C. zedoaria</i> , <i>E. officinalis</i> , <i>F. bengalensis</i> , <i>G. sylvestre</i> , <i>Holarrhena antidysenterica</i> , <i>M. charantia</i> , <i>N. sativa</i> , P. kurroa , <i>S. chirata</i> , <i>Syzygium cumini</i> , <i>T. belerica</i> , T. chebula , T. cordifolia , <i>T. terrestris</i> , T. foenum-graecum , W. somnifera	Balance insulin secretion, strengthens immune system
Divya Medha Vati	<i>A. calamus</i> , <i>B. monnieri</i> , <i>C. paniculatus</i> , <i>C. pluricaulis</i> , <i>I. racemosa</i> , <i>Lavandula stoechas</i> , <i>N. jatamansi</i> , W. somnifera	Cures mental disorders, depression and epileptic fits
Divya Amirta Rasayana	<i>A. racemosus</i> , <i>B. monnieri</i> , <i>Bambusa arundinacea</i> , <i>C. zeylanicum</i> , <i>C. pluricaulis</i> , <i>C. sativus</i> , <i>E. cardamomum</i> , <i>E. officinalis</i> , <i>M. prurita</i> , <i>Prunus amygdalus</i>	Rejuvenates and nourishes the brain and body
Divya Medohara Vati	B. diffusa , C. mukul , <i>E. officinalis</i> , E. ribes , <i>Operculina turpethum</i> , P. kurroa , <i>T. belerica</i> , T. chebula	Thyroid disorders, rheumatic arthritis, joint pains, pain to lumbar region and knee joints.
Divya Svasari Rasa	<i>A. ferox</i> , <i>Anacyclus pyrethrum</i> , <i>Capparis moonii</i> , <i>C. zeylanicum</i> , G. glabra , <i>P. longum</i> , <i>P. nigrum</i> , <i>Pistacia integerrima</i> , S. aromaticum , Z. officinale	Bronchitis, cough, coryza, cold, asthma and sinusitis
Divya Strirasayana Vati	<i>A. racemosus</i> , <i>Bambusa arundinacea</i> , B. aristata , <i>Bryonia laciniata</i> , <i>C. deodara</i> , C. mukul , <i>E. officinalis</i> , G. glabra , <i>M. ferrea</i> , N. nucifera , <i>Putranjiva roxburghii</i> , <i>S. album</i> , <i>Saraca asoca</i> , <i>S. cordifolia</i> , <i>T. belerica</i> , T. chebula , W. somnifera	Leucorrhoea, menorrhagia, irregularity in menstruation
Divya Hridayamrita Vati	B. diffusa , C. mukul , <i>C. rotundus</i> , <i>P. lanceolatus</i> , P. zeylanica , <i>T. arjuna</i> , T. cordifolia , V. negundo	Removes the arterial block, angina pain and palpitation
Divya Silajita Rasayana Vati	<i>E. officinalis</i> , <i>P. niruri</i> , <i>T. belerica</i> , T. chebula , W. somnifera	Diabetes & leucorrhoea
Divya Sarvakalpa Kvatha	B. diffusa , <i>Cassia fistula</i> , <i>P. niruri</i> , S. nigrum	Cures jaundice, oedema, oliguria, oedema

Formulation	Plants used	Uses
Divya Kanti Lepa	A. catechu , <i>C. camphora</i> , <i>Curcuma amada</i> , C. longa , M. fragrans , R. cordifolia , <i>S. album</i> , <i>Valeriana wallichii</i>	Cures skin disorders viz pimples, acne, wrinkles on face
Divya Vatari Churna	<i>M. oleifera</i> , P. kurroa , T. foenum-graecum , W. somnifera , Z. officinale	Cures rheumatoid arthritis, sciatica, pain in back and lumbar region
Himalaya Abana	<i>A. calamus</i> , <i>A. racemosus</i> , B. diffusa , <i>C. copticum</i> , <i>Celastrus paniculatus</i> , <i>Centella asiatica</i> , <i>Cinnamomum cassia</i> , C. wightii , <i>C. pluricaulis</i> , <i>C. sativus</i> , <i>C. rotundus</i> , <i>E. alba</i> , <i>E. cardamomum</i> , E. ribes , <i>E. officinalis</i> , F. vulgare , G. glabra , <i>N. jatamansi</i> , <i>Nepeta hindostana</i> , O. sanctum , <i>P. longum</i> , <i>Rosa damascena</i> , <i>R. centifolia</i> , <i>S. album</i> , S. aromaticum , <i>T. arjuna</i> , T. chebula , T. cordifolia , W. somnifera , Z. officinale	Hyperlipidemia and hypertension, adjuvant in angina therapy
Himalaya Cystone	<i>Achyranthes aspera</i> , <i>Cyperus scariosus</i> , <i>Didymocarpus pedicellata</i> , <i>Onosma bracteatum</i> , R. cordifolia , <i>Saxifraga ligulata</i> , <i>Vernonia cinerea</i>	Prevents urinary infections & stone formation
Himalaya Dental Cream	<i>A. arabica</i> , A. catechu , <i>A. farnesiana</i> , <i>Pistacia</i> , <i>C. copticum</i> , E. ribes , <i>E. officinalis</i> , <i>Mimusops elengi</i> , Punica granatum <i>Salvadora persica</i> , <i>T. bellerica</i> , T. chebula , <i>V. negundo</i> , <i>Zanthoxylum alatum</i>	Stops gum bleeding, boils and sores
Himalaya Diabecon	<i>Abutilon indicum</i> , A. vera , <i>A. racemosus</i> , B. aristata , B. diffusa , <i>Casearia esculenta</i> , C. wightii , C. longa , <i>Eugenia jambolana</i> , G. glabra , <i>G. arborea</i> , <i>Gossypium herbaceum</i> <i>G. sylvestre</i> , <i>M. charantia</i> , O. sanctum , Phyllanthus amarus , <i>P. nigrum</i> , Pterocarpus marsupium , <i>Rumex maritimus</i> , <i>Sphaeranthus indicus</i> , <i>S. chirata</i> , T. cordifolia , <i>T. terrestris</i> , <i>Trikatu</i> , <i>Triphala</i>	Reduce blood sugar levels and diabetic complications
Himalaya Geriforte	<i>Achillea millefolium</i> , <i>Argyria speciosa</i> , <i>Asparagus adscendens</i> , <i>A. racemosus</i> , B. aristata , <i>Caesalpinia digyna</i> , <i>Capparis spinosa</i> , <i>C. copticum</i> , <i>Cassia occidentalis</i> , <i>Centella asiatica</i> , <i>Cichorium intybus</i> , <i>C. sativus</i> , C. longa , <i>E. alba</i> , <i>E. cardamomum</i> , G. glabra , <i>M. pruriens</i> , M. fragrans , <i>P. longum</i> , S. aromaticum , <i>Tamarix gallica</i> , <i>T. arjuna</i> , T. chebula	Slows down aging, reduces stress and enhances immunity
Himalaya Himplasia	A. catechu , <i>A. racemosus</i> , <i>Caesalpinia bonducella</i> , <i>Tribulus terrestris</i>	Inhibits prostatic stromal proliferation & improves fertility
Himalaya Liv52	<i>A. millefolium</i> , <i>C. spinosa</i> , <i>C. occidentalis</i> , <i>Cichorium intybus</i> , S. nigrum , <i>Tamarix gallica</i> , <i>T. arjuna</i>	Pre- and early cirrhosis, viral hepatitis & alcoholic liver disease
Himalaya Menosan	<i>A. racemosa</i> , <i>Centella asiatica</i> , G. glabra , <i>Saraca indica</i> , <i>S. cordifolia</i> , T. chebula	Regulates overall hormonal balance & urinary tract function
Himalaya Mentat	<i>A. calamus</i> , <i>B. monnieri</i> , <i>C. paniculatus</i> , <i>C. asiatica</i> , E. ribes , <i>E. cardamomum</i> , <i>E. officinalis</i> , <i>Evolvulus alsinoides</i> , F. vulgare , <i>Ipomoea digitata</i> , <i>M. pruriens</i> , M. fragrans , <i>N. jatamansi</i> , <i>Orchis mascula</i> , <i>Oroxylum indicum</i> , <i>P. amygdalum</i> , S. aromaticum , <i>T. arjuna</i> , <i>T. bellirica</i> , T. chebula , T. cordifolia , <i>Valeriana sp.</i> , W. somnifera , Z. officinale	Adjuvant in Parkinson's and Alzheimers diseases
Himalaya Pilex	<i>Bauhinia variegata</i> , B. aristata , <i>C. fistula</i> , C. wightii , <i>E. officinalis</i> , Melia azadirachta , <i>M. ferrea</i> , <i>T. bellirica</i> , T. chebula	Treats piles, cures hemorrhoids, treats varicose veins
Himalaya Purim	A. paniculata , <i>Pistacia</i> , <i>C. fistula</i> , <i>Crataeva magna</i> , C. longa , <i>E. alba</i> , E. ribes , <i>E. officinalis</i> , P. kurroa , Psoralea corylifolia , Saussurea lappa , <i>T. bellerica</i> , T. chebula , T. cordifolia	Regulates detoxification and cleansing
Himalaya Reosto	C. wightii , <i>Sida cordifolia</i> , <i>T. arjuna</i> , <i>Vanda roxburghii</i> , W. somnifera	Reverses hypogonadism and osteoporosis
Himalaya Rumalaya Forte	Alpinia galanga , Boswellia serrata , C. wightii , G. glabra , T. cordifolia , <i>Tribulus terrestris</i>	Relieves pain from arthritis and traumatic inflammation
Rumalaya Gel	B. serrata , <i>Cedrus deodara</i> , <i>Cinnamomum zeylanicum</i> , <i>Gaultheria fragrantissima</i> , <i>M. arvensis</i> , <i>Pinus roxburghii</i> , V. negundo , Z. officinale	Analgesic, relieves pain, joint mobility
Himalaya Septilin	C. mukul , <i>E. officinalis</i> , G. glabra , <i>Moringa pterygosperma</i> , R. cordifolia , T. cordifolia	Strengthens immune system & body's defense mechanisms
Himalaya Triphala	<i>E. officinalis</i> , <i>T. bellirica</i> , T. chebula	Reduces high blood pressure and hypertension; effective in irritable bowel syndrome, ulcerative colitis and tumor

Boldface indicates anti-inflammatory activity shown in Table 4.

* The information is derived from websites <http://www.divyayoga.com>, <http://www.himalayahealthcare.com>, and <http://amrutanjan.com>, to illustrate as examples of Ayurvedic formulations, with no connection or affiliations to them of the authors.

Table 2

A list of Ayurvedic plants, their active components and their role in chronic diseases

Plant	Active compounds	In-vitro	In-vivo	Review
<i>Abrus precatorius</i>	Abruquinones	[243]	[11]	
	Abrin	[244]	[245]	
	Abrus agglutinin	[246]	[246]	
	ABP		[247]	
<i>Abutilon indicum</i>	Extract		[12]	
<i>Acacia arabica</i>	Gum	[248]		
<i>Acacia catechu</i>	Flavocoxid	[14]	[249]	
<i>Acacia farnesiana</i>	Acasiane, Farnesirane	[15]		
<i>Achillea millefolium</i>	Casticin	[250]		[251]
	Extract		[252]	
<i>Achyranthes aspera</i>	Extract		[17, 18, 253]	
<i>Acorus calamus</i>	Extract	[20]	[21, 254]	
<i>Adhatoda vasica</i>	Extract		[255, 256]	[257]
	Ambroxol	[22]		
<i>Aegle marmelos</i>	Marmelin	[25]		[258]
	Extract		[259]	
<i>Allium sativum</i>	Thiocremonone	[29]		
	1,2-vinyldithiin	[30]		[260–263]
	Diallyl sulfide/trisulfide	[264, 265]		
	Allicin	[266, 267]		
<i>Aloe vera</i>	Extract		[26, 27]	
	Aloe-emodin	[268, 269]		[270]
	Extract		[271, 272]	[273, 274]
<i>Alpinia galanga</i>	Acetoxychavicol acetate	[275–277]		
<i>Andrographis paniculata</i>	Andrographolide	[278–285]	[286–290]	
	Neoandrographolide	[291]		
	Andrograpanin	[292, 293]		
	Extract		[294, 295]	
	Composition		[296, 297]	
<i>Areca catechu</i>	Extract	[39, 298]		
<i>Argyria speciosa</i>	Extract		[17, 299–302]	
<i>Asparagus adscendens</i>	Extract	[40]	[303]	
<i>Asparagus racemosus</i>	Extract	[304]	[41, 305, 306]	[307, 308]
<i>Azadirachta indica</i>	Azadirachtin/	[43, 309]		[310–312]
	Nimbolide	[42, 313]		
	Extract	[314]	[312, 315–320]	
<i>Bacopa monnieri</i>	Bacoside-A		[44, 45]	
	Triterpene saponins		[321]	
	Extract	[322]	[323, 324]	

Plant	Active compounds	In-vitro	In-vivo	Review
<i>Bambusa arundinacea</i>	Extract		[46]	
<i>Bauhinia variegata</i>	Extract	[47, 48]	[325–327]	
	Flavonoids	[328]		
<i>Berberis aristata</i>	Berberine	[50]	[329, 330]	
<i>Bergenia ligulata</i>	Extract	[331]		
<i>Boerhaavia diffusa</i>	Extract	[332–335]	[334, 336–339]	
	Punarnavine		[340, 341]	
	Flavonoids	[342]		
<i>Boswellia serrata</i>	AKBA	[61, 343–345]	[54–56, 58–60, 346–348]	
<i>Bryonia laciniosa</i>	Extract		[62]	
<i>Butea monosperma</i>	Butrin/isobutrin/butein	[349]		
	Stigmsterol		[350]	
	Extract	[351]	[352–357]	
<i>Caesalpinia bonducella</i>	Oil/Extract		[358–360]	
<i>Caesalpinia digyna</i>	Extract		[64]	
<i>Calotropis procera</i>	Latex extract	[361]	[362–365]	
<i>Capparis spinosa</i>	Extract	[68, 366]		
<i>Carum copticum</i>	Extract		[367–369]	
<i>Casearia esculenta</i>	3-hydroxymethyl xylitol		[370]	
	Extract		[371]	
<i>Cassia angustifolia</i>	Extract		[71]	
<i>Cassia fistula</i>	Extract		[72, 372–374]	
<i>Cassia occidentalis</i>	Extract	[375]	[73, 376, 377]	[378]
<i>Cassia tora</i>	Ononitol monohydrate		[379]	
	Extract	[380]	[381–383]	
<i>Cedrus deodara</i>	Extract/oil	[384]	[75, 385]	
<i>Celastrus paniculatus</i>	Extract	[386]	[76, 387]	
<i>Cichorium intybus</i>	Magnolialide	[388]		
	Cichotyboside		[389]	
	Extract		[77, 390, 391]	
<i>Cinnamomum camphora</i>	Extract	[79]	[247]	
<i>Cinnamomum cassia</i>	Extract	[392]	[80, 393, 394]	[395]
	Cinnamaldehyde	[396–398]		
<i>Cinnamomum zeylanicum</i>	Extract	[399–401]	[402, 403]	
	Oil		[81]	
<i>Citrullus colocynthis</i>	Extract		[404–407]	
<i>Commiphora mukul</i>	Guggulsterone	[408–410]	[411–416]	[417–419]
	Gugulipid		[420, 421]	
	BHUx		[422]	
<i>Commiphora wightii</i>	Extract	[423, 424]		
<i>Convolvulus pluricaulis</i>	Extract		[425–427]	
<i>Crocus sativus</i>	Safranal, Crocin	[428]	[429–432]	[433–435]

Plant	Active compounds	In-vitro	In-vivo	Review
	Extract	[436, 437]	[438, 439]	
<i>Cuminum cymimum</i>	Extract/oil		[91, 440–444]	
<i>Curcuma amada</i>	Extract		[92]	[445]
<i>Curcuma longa</i>	Curcumin	[93–95]	[446–449]	[450–464]
<i>Curcuma zedoaria</i>	Curdione	[465, 466]		[96]
	Extract	[467–469]	[470, 471]	
<i>Cymbopogon citratus</i>	Citral	[472, 473]		
	Extract/oil	[98, 474]	[97, 475–477]	
<i>Cyperus rotundus</i>	Extract	[478, 479]	[480]	
<i>Cyperus scariosus</i>	Extract		[101]	
<i>Didymocarpus pedicellata</i>	Extract	[104]		
<i>Dolichos biflorus</i>	Extract		[481, 482]	
<i>Eclipta alba</i>	Extract		[483–487]	
<i>Elettaria cardamomum</i>	Extract	[110]	[109, 488–490]	
<i>Embelia ribes</i>	Embelin	[491]	[492–495]	
	Extract		[496–498]	
<i>Emblica officinalis</i>	Pyrogallol	[499]	[500]	
	Extract		[20, 501–504]	
<i>Eugenia jambolana</i>	Extract	[505]	[506–510]	
<i>Evolvulus alsinoides</i>	Extract		[511–513]	
<i>Fagonia cretica</i>	Extract		[116]	
<i>Ferula assafoetida</i>	Gum/Extract	[514]	[515–517]	
<i>Ficus bengalensis</i>	Leucodelphinidin	[518]		
	Extract		[118, 519–522]	
<i>Foeniculum vulgare</i>	Anethole	[120]	[523]	
	Extract	[524]	[523, 525–528]	
<i>Garcinia cambogia</i>	Garcinol	[529–531]	[532]	[533]
	Extract		[121, 534, 535]	
<i>Glycyrrhiza glabra</i>	Isoliquiritigenin	[536]		
	Glycyrrhizin		[537]	
	Glabridin	[538]	[538]	
	Extract	[539–541]	[126, 540, 542]	
<i>Gymnema sylvestre</i>	Extract		[543–545]	[546, 547]
<i>Hemidesmus indicus</i>	HMBA	[548]	[549, 550]	
	Extract		[551]	
<i>Holarrhena antidysenterica</i>	Extract		[552]	
<i>Hordeum vulgare</i>	Extract	[134, 553]	[554]	
<i>Indigofera tinctoria</i>	TCA		[555]	
	Indigtone		[556]	
	Extract		[557, 558]	
<i>Inula racemosa</i>	Extract	[559]	[136, 543, 560]	
<i>Ipomoea nil</i>	Extract	[561]		

Plant	Active compounds	In-vitro	In-vivo	Review
<i>Lavendula stoechas</i>	Extract	[562, 563]		
<i>Leucas cephalotes</i>	Extract		[140]	
<i>Mangifera indica</i>	Extract	[564–567]	[565, 568–573]	
	3beta-taraxerol	[574]		
	Mangiferin		[575]	
<i>Mentha piperita</i>	Extract	[576]	[577, 578]	[579]
	Oil	[580, 581]		
	Flavonoid		[582]	
<i>Mimusops elengi</i>	Extract		[147, 583]	
<i>Momordica charantia</i>	Extract	[584–588]	[589–594]	[148, 595]
<i>Moringa oleifera</i>	Extract		[596–601]	[602]
<i>Mucuna pruriens</i>	Extract		[603–607]	
<i>Nardostachys jatamansi</i>	Extract		[608–611]	
<i>Nelumbo nucifera</i>	(S)-armepavine		[154, 612, 613]	[614]
	Neferine		[615]	
	Kaempferol	[616]		
	Isoliensinine	[617]		
	Extract		[618–620]	
<i>Nigella sativa</i>	Thymoquinone	[621]	[621–628]	[629–631]
	Polyphenols		[632]	
	Extract	[633]	[624, 634–637]	
<i>Nyctanthes arbortristis</i>	arbortristoside-A		[156]	
	Extract		[638, 639]	
<i>Ocimum sanctum</i>	Eugenol		[640, 641]	[642, 643]
	Extract/oil	[644]	[320, 645–647]	
<i>Operculina turpethum</i>	Extract		[161, 648]	
<i>Orchis mascula</i>	Extract		[162]	
<i>Oroxylum indicum</i>	Baicalein	[163]		
	Extract	[649, 650]	[651]	
<i>Phyllanthus amarus</i>	Niranthin		[165]	
	Phyllanthin	[652]		
	Extract/lignan		[653–659]	
<i>Phyllanthus niruri</i>	Arabinogalactan	[660]		[661]
	Extract/lignan	[662]	[662–666]	
<i>Picrorhiza kurroa</i>	Picroliv		[667–670]	[671]
	Extract	[166, 672]		
<i>Piper chaba</i>	Amides/Extracts		[673, 674]	
<i>Piper longum</i>	Piperine	[675]	[675–677]	
	Piperlongumine	[678]		
	Piperinic acid	[679]	[679, 680]	
	Extract/oil	[167]	[681–684]	
<i>Piper nigrum</i>	Piperine		[685–687]	[688, 689]

Plant	Active compounds	In-vitro	In-vivo	Review
	Extract	[110]	[690]	
<i>Pistacia integerrima</i>	Extract		[169, 691]	
<i>Pluchea lanceolata</i>	Extract		[692, 693]	
<i>Plumbago zeylanica</i>	Plumbagin	[172, 694, 695]	[664, 696]	
	Seselin	[697]		
	Suberosin	[698]		
<i>Pongamia pinnata</i>	Pongamol, Karanjin		[699]	
	Extract		[700–704]	
<i>Psoralea corylifolia</i>	Bavachin	[705]		
	Psoralen		[706]	
	Bakuchiol	[707, 708]		
	Furocoumarins		[709]	
	Extract	[710, 711]	[710, 712]	
<i>Pterocarpus marsupium</i>	Pterostilbene		[179]	
	Flavanoids		[713]	
	Extract		[714]	
<i>Pterocarpus santalinus</i>	Savinin	[715]		
	Extract		[180, 716]	
<i>Pueraria tuberosa</i>	Lupinoside		[717]	
	Extract		[718]	
<i>Punica granatum</i>	Ellagitannins		[181, 719]	[720–722]
	Punicalagin	[723]	[181, 723]	
	Anthocyanin		[724]	
	Extract	[725, 726]	[727–729]	
<i>Putranjiva roxburghii</i>	Extract		[182]	
<i>Quercus infectoria</i>	Extract		[183, 730]	
<i>Raphanus sativus</i>	Extract	[731]	[732, 733]	[734]
<i>Ricinus communis</i>	Ricinoleic acid		[188]	
	Extract		[735–737]	
<i>Rosa damascena</i>	Flavonoids	[190]		
	Extract/oil		[738–740]	
<i>Rubia cordifolia</i>	Rubiadin		[741]	
	Mollugin	[191]		
	Extract		[742–744]	
<i>Rumex maritimus</i>	Extract		[745]	
<i>Salvadora persica</i>	Extract		[193]	
<i>Santalum album</i>	Oil		[746]	
	Lignan	[747]		
<i>Saphindus trifoliatus</i>	Extract		[196, 748]	
<i>Saraca indica</i>	Saracin	[198]		
<i>Saussurea lappa</i>	Extract	[749]	[17]	
	Lactone	[750]		

Plant	Active compounds	In-vitro	In-vivo	Review
<i>Sesamum indicum</i>	Arctigenin	[751]		
	Cynaropicrin	[752, 753]		
	Costunolide	[754]		
	Sesaminol		[755, 756]	
	Sesamin	[201]		
	Sesamol		[757]	
<i>Sida cordifolia</i>	Extract		[758–761]	
	Extract		[762–766]	
<i>Solanum indicum</i>	Extract	[767]	[203]	
<i>Solanum nigrum</i>	Extract	[768, 769]	[748, 755, 770–773]	
<i>Solanum xanthocarpum</i>	Extract		[206, 774]	
<i>Sphaeranthus indicus</i>	7-hydroxyfrullanolide	[775]		
	Extract		[207, 776]	
<i>Stereospermum suaveolen</i>	Extract		[208, 777]	
<i>Strychnos nuxvomica</i>	Brucine	[209]	[778]	
	Extract	[779, 780]		
<i>Swertia chirata</i>	Amarogentin		[210]	
	Extract		[781, 782]	
<i>Symplocos crataegoides</i>	Triterpenes	[211]		
<i>Syzygium aromaticum</i>	Acetyl eugenol	[783]		
<i>Syzygium cumini</i>	Extract		[212, 784, 785]	[786]
	ferulic acid		[787] [788]	
	Extract		[789–792]	
<i>Tamarix gallica</i>	Extract		[214, 793]	
<i>Terminalia arjuna</i>	Arjunic acid		[794]	
	Casuarinin	[795, 796]		
	Terminoside A	[797]		
	Extract		[215, 216, 798–800]	[801]
<i>Terminalia belerica</i>	Extract		[549, 802–806]	
<i>Terminalia chebula</i>	Chebularic acid	[807]	[808]	
	Extract		[549, 803, 806, 809–812]	
<i>Thymus vulgaris</i>	Thymol, Carvacrol	[813]		
	Extract	[220]	[814]	
<i>Tinospora cordifolia</i>	Extract	[815]	[222, 506, 816–819]	
<i>Trachyspermum ammi</i>	Extract		[225, 820]	
<i>Tribulus terrestris</i>	Tribulosin		[821]	
	Saponion	[822]	[823]	
	Extract		[226, 228, 824, 825]	
<i>Trigonella foenum-graecum</i>	Diosgenin	[826]	[229, 826, 827]	
	Galactomannan		[828]	
	Extract		[829–836]	
<i>Valeriana wallichii</i>	Extract		[837, 838]	

Plant	Active compounds	In-vitro	In-vivo	Review
<i>Vanda roxburghii</i>	Extract		[235, 839]	
<i>Vernonia cinerea</i>	Extract	[236]	[757, 840–843]	
<i>Viola odorata</i>	Cycloviolacin	[844]		
	Extract	[845]		
<i>Vitex negundo</i>	Vitexins		[238]	
	Diterpenes	[846]		
	Extract		[847–850]	
<i>Withania somnifera</i>	Withaferin-A	[851, 852]	[241]	[853]
	Withanolide sulfoxide	[240]	[854]	
	Withanamides	[855]		
	Extract	[856]	[857–859]	
<i>Zingiber officinale</i>	[6]-Gingerol	[860]	[860, 861]	
	6-Shogaol	[862–864]		
	Extract	[865, 866]	[867–871]	[872, 873]

ABP, abrin-derived peptide; AKBA, Acetyl-11-keto- β -boswellic acid; TCA, trans-tetracos-15-enoic acid; HMBA, 2-hydroxy-4-methoxy benzoic acid

Table 3List of inflammatory gene products regulated by NF- κ B*

Enzymes	Stress response genes	Gro b	CD21	Lox-1	Clone 330
11bHSD2	12-LOX	Gro g	CD38	Ly49	Clone 68
17bHSD	5-LOX	Gro-1	CD3g	Mdr1	Connexin32
ABC Transporters	COX-2	ICOS	CD40	mGlu2	Cyclin D1
ADH	Cu/Zn SOD	IFN-g	CD48	Mu-OR	Cyclin D2
AID	CYP2C11	IiGp1	CD83	NMDA-RS 2A	Cyclin D3
alpha 1ACT	CYP2E1	IL-1 RA	CD86	NP Y-Y1 R	DIF2
AMACR	CYP7b	IL-10	CD98	NR-1	DMT1
ARFRP1	FHC	IL-11	CXCR	Oxytocin R	Elafin
ASS	GCL	IL-12A	CXCR2	PAF-R1	Endothelin 1
Aromatase	GCLM	IL-12B	F11-R	P-gp	Ephrin-A1
ART2.1	HSP90-a	IL-13	Fc e R II	RAGE	epsilon-Globin
BACE-1	iNOS	IL-15	FcRn	Transcription factors	Factor VIII
Btk	MAP4K1	IL-17	HLA-G	/Regulators	FHC
Cathepsin B	Mx1	IL-1a	ICOS	A20	Gadd45b
Cathepsin L	DTD	IL-1b	Ig C g1	ABIN-3	Galectin 3
cdk6	cPLA2	IL-2	Ig e heavy chain	AR	Galpha i2
CGT	SEN2	IL-23A	Ig g1	Bcl-3	GBP-1
CHI3L1	SEPS1	IL-27	Ig g4	BMI-1	GIF
CRAD1	SOD1	IL-6	Ig k light chain	C/EBPd	Gro-1
CRAD2	SOD2	IL-8	IL-2 R a-chain	CDX1	GS3686
Collagenase 1	Early response genes	IL-9	Invariant Chain II	c-fos	HK protein
DDH	B94	IP-10	Kinin B1 Receptor	c-myb	HCCS1
DNASIL2	Egr-1	KC	MHC-I (H-2Kb)	c-myc	HMG14
DYPD	p22/PRG1	LIX	MHC-I HLA-B7	c-rel	IBABP
EL	p62	Lymphotoxin a	Nod2	DC-SCRIPT	IMP2
ENO2	TIEG	Lymphotoxin b	PGRP-S	Dmp1	K15 Keratin
gamma-GCS	Viruses	MCP-1/JE	Polymeric Ig R	E2F3a	K3 Keratin
GAD67	AV	MIG	T-CR b chain	Elf3	K6b Keratin
GCL	AVV	MIP-1a,b	T-CR/CD3g	ELYS	Lactoferrin

Enzymes	Stress response genes	Gro b	CD21	Lox-1	Clone 330
GCLM	BLV	MIP-2	TLR-2	ETR101	Laminin-B2
GCLC	CMV	MIP-3a/CCL20	TLR9	Gata-3	Lipocalin-2
GD3-synthase	EBV	mob-1	TNF-Receptor	GCR	MCT1
Gelatinase B	HBV	NAP-78	TREM-1	HIF-1a	Mir125b
G6PD	HIV-1	RANTES	b-2 Microglobulin	HOXA9	Mir146a, b
Glc-6-Pase	HPV-16	T-CA gene 3	Growth factors/ligands	IkB-a	Mir155
GnRH II	HSV	TNFalpha	Activin A	IkB-e	MNE1
gp91 phox	JCV	TNFBeta	Angiopoietin	IRF-1	Mts1
Granzyme B	SIV	TRAIL	BCAP	IRF-2	Mucin
GSTP1-1	SV-40	Treefoil factor-3	BDNF	IRF-4	MBP
H+-K+ATPase a2	Apoptosis Regulators	VEGI	BLNK	IRF-7	Naf1
Heparanase	ASC	Cell adhesion molecules	BLyS	jmjD3	NGL
HO-1	B7-H1	CD44	BMP-2	junB	NLF1
Has	Bax	DC-SIGN	BMP-4	Lef1	p11
IDO	Bcl-2	ELAM-1	CGRP	LZIP	p21-CIP1
iNOS	Bcl-xL	Endoglin	EPO	Mail	PA28 a
ITD-2	Bfl1/A1	Fibronectin	FGF8	nfkbl	PA28 b
L-PGDS	Bim	ICAM-1	FLRG	nfkbl2	PAI-1
Lysozyme	BNIP3	MadCAM-1	G-CSF	NLRP2	Pax8
MMP-3	Caspase-11	NCAM	GM-CSF	NURR1	PCBD
MMp-9	CD95 (Fas)	P-selectin	HGF/SF	Osterix	Perforin
MKP-1	c-FLIP	Tenascin-C	IGFBP-1	p53	PGK1
MLCK	CIDEA	VCAM-1	IGFBP-2	PR	POMC
Mthfr	FAPase-1	Acute phase proteins	M-CSF	PU.1	PSG
GnT-I	Fas-Ligand	Angiotensinogen	Midkine	relb	PDYN
n-NOS	IAPs	b-defensin-2	NGF	Snail	PSA
PDE7A1	IEX-1L	C4b BP	NK-1R	Sox9	PTEN
PGES	Nr13	CF-kB	NK4	Stat5a	RAG-1
PIK3CA	TRAF-1	CF-C4	Nrg1	Tfec	RAG-2
PIM-1	TRAF-2	C-RP	OPN	Twist	RbAp48
PKAalpha	TRAF-2 BP	Hepcidin	PDGF B chain	WT1	RICK

Enzymes	Stress response genes	Gro b	CD21	Lox-1	Clone 330
PKCdelta	XIAP	LPS BP	PIGF	YY1	S100A6
PLCdelta	Cytokines/Chemokines	Pentraxin-3	Proenkephalin	Miscellaneous	Serpine2
Plk3	aka (LAG-1)	SAA1	Prolactin	AGP	SH3BGRL
PP5	BAFF	SAA2	SCF	a1-AT	SK2 channels
PTGIS	b-Interferon	SAA3	THBS1	a2(I) collagen	Skp2
PTHrP	BLIMP-1	Tissue factor-1	THBS2	ABCG5	Spergen-1
PTP1B	CCL15	UPA	VEGF C	ABCG8	SWS1
RACK1	CCL17	Antigen presentation	WNT10B	AbetaH-J-J	Syncytin-1
REV3	CCL19	Complement B	Cell-surface receptors	AFP	Syndecan-4
Serpin 2A	CCL20	Complement component 3	A1 AR	AMH	TASK-2
SIAT1	CCL22	Complement Receptor 2	A2A	APOBEC2	TAUT
Slfn-2	CCL23	Peptide Transporter TAP1	a2B-AR	Apo CIII	TFPI-2
SNARK	CCL28	Proteasome Subunit LMP2	ABCA1	Apo D	Transferrin
sGC-1	CCL5	Tapasin	ABCC6	Apo E	TRIF
SSAT	CINC-1	Immunoreceptors	ADAM19	AQP4	TRPC1
SUV3	CXCL 11	B7.1 (CD80)	AS Na-channel	b-amyloid	UBE2M
TERT	CXCL5	BRL-1	Bradykinin B1-R	Biglycan	UCP-2
TG	CXCL6	CCR5	CD23	BRCA2	Uroplakin Ib
TTG	EBI3/IL-27B	CCR7	CD69	Calsarcin-1	Vimentin
type II-sPLA(2)	Eotaxin	CD134	DOR	Caveolin-1	zeta-Globin
		CD137	EGFR	Claudin-2	
UPase	Fractalkine	CD154	ErbB2	Clone 156	
XDH	Gro a		Gal1 R		

* Adopted from www.nf-kb.org

Table 4

Selected Ayurvedic Plants, Their Phytochemicals and Their Molecular Targets

Ayurvedic Plants Alkaloid	Active Compounds	Molecular Targets	References
<i>Berberis aristata</i>	Berberine	NF-κB, COX-2	[50]
<i>Indigofera tinctoria</i>	Indirubin	NF-κB, COX-2	[135]
Flavonoid			
<i>Acacia catechu</i>	Baicalin, catechin	COX-2, 5-LOX, iNOS	[14]
<i>Dysoxylum binectariferum</i>	Flavopiridol	NF-κB, COX-2	[107]
<i>Nelumbo nucifera</i>	Kaempferol	NF-κB, COX-2, iNOS	[155]
Lignan			
<i>Myristica fragrans</i>	Macelignan	NF-κB, COX-2	[874, 875]
<i>Phyllanthus amarus</i>	Niranthin	PAFR	[165]
<i>Pterocarpus santalinus</i>	Savinin	TNF-α	[715]
Quinone			
<i>Embelia ribes</i>	Embelin	NF-κB, COX-2	[112]
<i>Aloe vera</i>	Emodin, lectin	NF-κB, TNF-α	[32, 876]
<i>Rubia cordifolia</i>	Mollugin	NF-κB	[191]
<i>Plumbago zeylanica</i>	Plumbagin	NF-κB, COX-2, STAT-3	[172, 173]
<i>Nigella sativa</i>	Thymoquinone	NF-κB, TNF-α, IL-1β, COX-2	[8, 624]
Polyphenol			
<i>Terminalia chebula</i>	Chebulagic acid	COX-1, COX-2, 5-LOX	[807]
<i>Curcuma longa</i>	Curcumin	NF-κB, COX-2, STAT-3	[94, 95, 877]
<i>Punica granatum</i>	Punicalagin	NF-κB, COX-2	[181]
<i>Garcinia cambogia</i>	Garcinol, gambogic acid	NF-κB, COX-2, 5-LOX, iNOS	[123, 878, 879]
<i>Zingiber officinale</i>	[6]-Gingerol, shogaol	NF-κB, COX-2	[862, 880]
Sulfur Compound			
<i>Allium sativum</i>	S-Allyl-L-cysteine, allicin, diallyl sulfide, thiacecremonone	NF-κB, TNF-α, iNOS, COX-2, IL-6, MCP-1, IL-8, IL-10, MIG, IL-1β	[29, 264, 265, 267, 881]
Triterpine			
<i>Boswellia serrata</i>	Boswellic acid	NF-κB, COX-2, STAT-3, 5-LOX	[61, 882]
<i>Callicarpa macrophylla</i>	Betulinic acid	NF-κB, COX-2, STAT-3	[65, 66]
<i>Trigonella foenum-graecum</i>	Diosgenin	NF-κB, COX-2, STAT-3	[229, 230]
<i>Commiphora mukul</i>	Guggulsterone	NF-κB, STAT-3	[83, 85]
<i>Commiphora wightii</i>	Guggulsterone	NF-κB, STAT-3	[83, 85]
<i>Glycyrrhiza glabra</i>	Glycyrrhizin	iNOS, NF-κB, IL-4, IL-5, IFN-γ	[883, 884]
<i>Ocimum sanctum</i>	Ursolic acid	NF-κB, COX-2, STAT-3	[159, 160]
<i>Withania somnifera</i>	Withanolides	NF-κB, COX-2	[239, 240]
Diterpine			
<i>Andrographis paniculata</i>	Andrographolide, neoandrographolide	NF-κB, TNF-α, IL-6, iNOS, IFN-γ, IL-12p70, COX-2	[278, 279, 283, 291, 293, 294, 885]
<i>Vitex negundo</i>	Negundo C, E	iNOS, COX-2	[846]
Sesquiterpine			
<i>Saussurea lappa</i>	Cynaropicrin, saussureamines A, B	TNF-α, NF-κB, iNOS	[886-888]

Ayurvedic Plants Alkaloid	Active Compounds	Molecular Targets	References
Terpenoid Lactone			
<i>Azadirachta indica</i>	Azadirachtin	NF- κ B, STAT-3	[42]
	Nimbidin, nimbolide	PGE2, IL-1, NF- κ B	[43, 183]
Miscellaneous			
<i>Abies pindrow</i>	Pinitol	NF- κ B, COX-2	[10]
<i>Aegle marmelos</i>	Marmelin	COX-2, IL-8, TNF- α	[889]
<i>Alpinia galanga</i>	ACA	NF- κ B, COX-2, iNOS	[34, 890]
<i>Boerhaavia diffusa</i>	Punarnavine	NF- κ B	[53]
<i>Foeniculum vulgare</i>	Anethole	NF- κ B, COX-2	[119]
<i>Picrorhiza kurroa</i>	Picroliv	NF- κ B, COX-2	[166]
<i>Psoralea corylifolia</i>	Psoralen	IL-10	[178]
<i>Pterocarpus marsupium</i>	Pterostilbene	COX-2	[179]
<i>Solanum nigrum</i>	Phytoglycoprotein	NF- κ B, iNOS, COX-2	[891]
<i>Syzygium aromaticum</i>	Eugenol	NF- κ B, COX-2	[892]
<i>Tinospora cordifolia</i>	(1,4)- α -D-glucan	NF- κ B	[893]

5-LOX, 5-lipoxygenase; COX-2, cyclooxygenase-2; IL-1, interleukin (IL)-1; iNOS, inducible nitrogen oxide synthase; IP-10, IFN- γ -induced protein-10; MCP-1, monocyte chemoattractant protein-1; MIG, monokine induced by IFN- γ ; MIP- α , macrophage inflammatory protein-1 α ; NF- κ B, nuclear factor-kappaB; PAFR, platelet-activating factor-receptor; PGE2, prostaglandin E2; STAT-3, signal transducer and activator of transcription 3; TNF- α , tumor necrosis factor- α