MATH3424 HW5

Chow, Hau Cheung Jasper (hcjchow / 20589533)

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Q1a

```
setwd("/Users/jchow/Downloads/MATH3424 R")
cancer_data <- read.csv(file="BreastCancer.txt",header=TRUE)</pre>
#as.numeric(cancer data$Class) # 1 = benign, 2 = malignant
cancer_data$Class <- relevel(cancer_data$Class, ref = "benign")</pre>
cancer_model_1 <- glm(Class ~ ., family="binomial", data=cancer_data)</pre>
summary(cancer_model_1)
##
## Call:
  glm(formula = Class ~ ., family = "binomial", data = cancer_data)
##
## Deviance Residuals:
##
       Min
                 1Q
                      Median
                                   3Q
                                            Max
## -3.4841 -0.1153 -0.0619
                               0.0222
                                         2.4698
##
## Coefficients:
                    Estimate Std. Error z value Pr(>|z|)
##
## (Intercept)
                   -10.10394
                                1.17488 -8.600 < 2e-16 ***
## Cl.thickness
                     0.53501
                                0.14202
                                          3.767 0.000165 ***
## Cell.size
                                0.20908 -0.030 0.976039
                    -0.00628
## Cell.shape
                     0.32271
                                0.23060
                                          1.399 0.161688
                                0.12345
                                           2.678 0.007400 **
## Marg.adhesion
                     0.33064
## Epith.c.size
                     0.09663
                                0.15659
                                           0.617 0.537159
## Bare.nuclei
                     0.38303
                                0.09384
                                           4.082 4.47e-05 ***
## Bl.cromatin
                     0.44719
                                0.17138
                                          2.609 0.009073 **
## Normal.nucleoli
                     0.21303
                                0.11287
                                           1.887 0.059115 .
## Mitoses
                     0.53484
                                0.32877
                                           1.627 0.103788
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
##
  (Dispersion parameter for binomial family taken to be 1)
##
##
       Null deviance: 884.35 on 682 degrees of freedom
## Residual deviance: 102.89 on 673 degrees of freedom
## AIC: 122.89
##
## Number of Fisher Scoring iterations: 8
```

We notice that not all of the regression coefficients are significant at the 0.05 level; only intercept, Cl.thickness, Marg.adhesion, Bare.nuclei and Bl.cromatin are.

Instead of the F-statistic, we use the G-statistic $G = D_0 - D$ where if the null model is correct, $G \sim \chi^2(9)$ since the sample size n = 683 is large.

Since $G = D_0 - D = 884.35 - 102.89 = 781.46$ and as we can see below, the p-value is very close to 0, we reject the null hypothesis and the model is indeed significant.

```
pchisq(781.46, df=9, lower.tail=FALSE)
## [1] 2.079405e-162
rsq_1 \leftarrow 1-(102.89/884.35)
rsq_1
## [1] 0.8836547
Q<sub>1</sub>b
\beta_1 is the estimated coefficient for CL.thickness. So the 95% CI is [0.25666, 0.8133682].
beta_1_hat <- cancer_model_1$coefficients["Cl.thickness"]</pre>
se_beta_1_hat <- 0.14202
alpha <- 0.05
z_alpha_2 <- qnorm(alpha/2, mean=0, sd=1, lower.tail=FALSE)</pre>
beta_1_hat - z_alpha_2*se_beta_1_hat
## Cl.thickness
##
        0.25666
beta_1_hat + z_alpha_2*se_beta_1_hat
## Cl.thickness
##
      0.8133682
# https://stats.idre.ucla.edu/r/dae/logit-regression/
# this gets the CIs based on profiled log-likelihood function
#confint(cancer_model_1, level=0.95, parm="Cl.thickness")
# this gets them with the standard errors (which we want)
confint.default(cancer_model_1, level=0.95, parm="Cl.thickness")
##
                    2.5 %
                              97.5 %
```

To test $H_0: \beta_3 = 0$ versus $H_1: \beta_3 \neq 0$, from the regression output in 1a, we notice that the p-value is 0.161688 (the z-statistic is 1.399) which is larger than our chosen significance of $\alpha = 0.1$. Therefore we fail to reject the null hypothesis that $H_0: \beta_3 = 0$.

Q1c

Cl.thickness 0.256665 0.8133631

```
## Deviance Residuals:
                      Median
##
       Min
                 10
                                    30
                                            Max
##
   -3.2982
           -0.1242 -0.0624
                                0.0234
                                         2.3713
##
##
  Coefficients:
##
                 Estimate Std. Error z value Pr(>|z|)
## (Intercept)
                 -9.74114
                              1.04989
                                       -9.278 < 2e-16 ***
## Cl.thickness
                  0.62576
                              0.13373
                                        4.679 2.88e-06 ***
## Cell.shape
                  0.48994
                              0.15379
                                        3.186 0.001444 **
## Marg.adhesion
                  0.33918
                              0.11221
                                        3.023 0.002505 **
## Bare.nuclei
                  0.37330
                              0.09381
                                        3.979 6.91e-05 ***
## Bl.cromatin
                  0.55731
                              0.16341
                                        3.411 0.000648 ***
##
                   0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for binomial family taken to be 1)
##
##
       Null deviance: 884.35
                                       degrees of freedom
                               on 682
## Residual deviance: 112.57
                               on 677
                                       degrees of freedom
  AIC: 124.57
##
## Number of Fisher Scoring iterations: 8
# p-value for G-test with new model
pchisq(771.78, df=5, lower.tail=FALSE)
## [1] 1.471799e-164
# R^2 for new model
rsq_2 <- 1-(112.57/884.35)
rsq_2
```

[1] 0.8727088

This time we notice all regression coefficients are significant at the 0.05 level and the AIC of the model has increased slightly from 122.89 to 124.57, but not by a significant amount, so the reduced and full model are equally effective for regression. We also observe that G = 884.35 - 112.57 = 771.78 is slightly lower but the p-value is still very low and close to 0, so the model is also significant. The R^2 has decreased slightly from 0.88 to 0.87 but is still very high.

```
#2*(logLik(cancer_model_2)-logLik(cancer_model_1))
residual_dev_diff <- 112.57-102.89
residual_dev_diff
## [1] 9.68
qchisq(0.99, df=4, lower.tail=TRUE)</pre>
```

[1] 13.2767

If we use the hypothesis that H_0 : reduced suitable, H_1 : full suitable, then by examining the statistic $2[L(p+q)-L(p)] \sim \chi^2(q)$ we observe with p=5, p+q=9 that the residual deviance of reduced minus residual deviance of full is 9.68. We notice that at significance level $\alpha=0.01$, the test statistic of 9.68 is smaller than the critical value of 13.2767, so we fail to reject H_0 .

```
Q1d
new <- data.frame(Cl.thickness=6, Cell.shape=3, Marg.adhesion=8, Bare.nuclei=2, Bl.cromatin=5)
# can show log-odds = beta \ O \ hat + innerprod(new, beta i hat) = 1.72983
# where: log-odds = ln(pi/(1-pi)) where pi=probability tumor is malignant
# then to get pi need to do: pi = exp(log-odds)/(1+exp(log-odds))
pi <- predict(cancer_model_2, new, type="response")</pre>
1-pi
##
## 0.1506149
So the probability of this patient's tumour being benign is 1 - \pi = 0.1506149.
Q<sub>1</sub>e
4 variables
cancer_model_3 <- glm(Class ~ Cell.shape + Marg.adhesion + Bare.nuclei + Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 151.3174
cancer_model_3 <- glm(Class ~ Cl.thickness + Marg.adhesion + Bare.nuclei + Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 135.7746
cancer_model_3 <- glm(Class ~ Cl.thickness + Cell.shape + Bare.nuclei + Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 132.7431
cancer_model_3 <- glm(Class ~ Cl.thickness + Cell.shape + Marg.adhesion + Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer model 3)
```

```
## [1] 141.6494
```

[1] 135.7378

3 variables

```
## [1] 205.0236
```

```
cancer_model_3 <- glm(Class ~ Cell.shape + Bare.nuclei + Bl.cromatin,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 156.4433
cancer_model_3 <- glm(Class ~ Cell.shape + Marg.adhesion + Bl.cromatin,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 182.4583
cancer_model_3 <- glm(Class ~ Cell.shape + Marg.adhesion + Bare.nuclei,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 167.7037
# ======
cancer_model_3 <- glm(Class ~ Cl.thickness + Bare.nuclei + Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 150.9232
cancer_model_3 <- glm(Class ~ Cl.thickness + Marg.adhesion + Bl.cromatin,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 174.8883
cancer_model_3 <- glm(Class ~ Cl.thickness + Marg.adhesion + Bare.nuclei,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 163.4411
# ======
cancer_model_3 <- glm(Class ~ Cl.thickness + Cell.shape + Bl.cromatin,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 160.0564
cancer_model_3 <- glm(Class ~ Cl.thickness + Cell.shape + Bare.nuclei,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 148.2462
# ======
cancer_model_3 <- glm(Class ~ Cl.thickness + Cell.shape + Marg.adhesion,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 168.2263
```

2 variables

```
cancer_model_3 <- glm(Class ~ Cl.thickness + Cell.shape,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 206.3944
cancer_model_3 <- glm(Class ~ Cl.thickness + Marg.adhesion,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 260.1006
cancer_model_3 <- glm(Class ~ Cl.thickness + Bare.nuclei,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 204.2192
cancer_model_3 <- glm(Class ~ Cl.thickness + Bl.cromatin,</pre>
                      family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 231.4534
# ======
cancer_model_3 <- glm(Class ~ Cell.shape + Marg.adhesion,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 222.4743
cancer_model_3 <- glm(Class ~ Cell.shape + Bare.nuclei,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 177.3853
cancer_model_3 <- glm(Class ~ Cell.shape + Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 199.0765
# ======
cancer_model_3 <- glm(Class ~ Marg.adhesion + Bare.nuclei,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 257.0478
cancer_model_3 <- glm(Class ~ Marg.adhesion + Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 306.6519
cancer_model_3 <- glm(Class ~ Bare.nuclei + Bl.cromatin,</pre>
```

```
family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 229.2795
```

1 or 0 variables

[1] 886.3502

```
cancer_model_3 <- glm(Class ~ Cell.shape,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 271.5863
cancer_model_3 <- glm(Class ~ Marg.adhesion,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 467.3434
cancer_model_3 <- glm(Class ~ Bare.nuclei,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 344.6277
cancer_model_3 <- glm(Class ~ Bl.cromatin,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 392.2168
cancer_model_3 <- glm(Class ~ Cl.thickness,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
## [1] 462.483
# ======
cancer_model_3 <- glm(Class ~ 1,</pre>
                       family="binomial", data=cancer_data)
AIC(cancer_model_3)
```

As we can see, the 4-variable model with the lowest AIC (132.7431) was with variables "Cl.thickness + Cell.shape + Bare.nuclei + Bl.cromatin." The 3-variable model with lowest AIC (148.2462) was with variables "Cl.thickness + Cell.shape + Bare.nuclei." The 2-variable model with lowest AIC (177.3853) was with variables "Cell.shape + Bare.nuclei." The 1-variable model with lowest AIC (271.5863) was with variable "Cell.shape". As we can see, the more variables we drop, the higher the AIC. In fact, out of all subset models of the variables "Cl.thickness + Cell.shape + Marg.adhesion + Bare.nuclei + Bl.cromatin," the one with the lowest AIC (124.57) is the model with all 5 of those variables, i.e. the model in part (c).