# Class 8 Breast Cancer Mini Project

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Before we dive into breast cancer project, we will finish class 7 (where we left off) first.

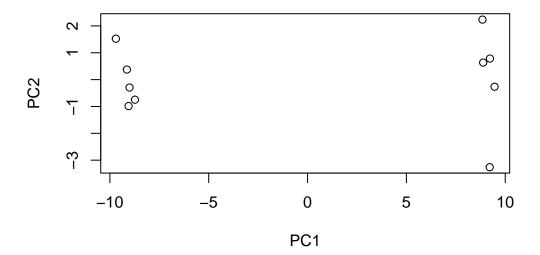
```
url2 <- "https://tinyurl.com/expression-CSV"</pre>
  rna.data <- read.csv(url2, row.names=1)</pre>
  head(rna.data)
       wt1 wt2 wt3 wt4 wt5 ko1 ko2 ko3 ko4 ko5
gene1 439 458 408 429 420 90 88 86
gene2 219 200
               204 210 187 427 423 434 433 426
gene3 1006 989 1030 1017 973 252 237 238 226 210
gene4
               829 856 760 849 856 835 885 894
      783 792
      181 249
               204 244 225 277 305 272 270 279
gene5
      460 502 491 491 493 612 594 577 618 638
gene6
```

Q10: How many genes and samples are in this data set? 6 genes and 10 samples

#### ##Run PCA

```
## Again we have to take the transpose of our data
pca <- prcomp(t(rna.data), scale=TRUE)

## Simple unpolished plot of pc1 and pc2
plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2")</pre>
```



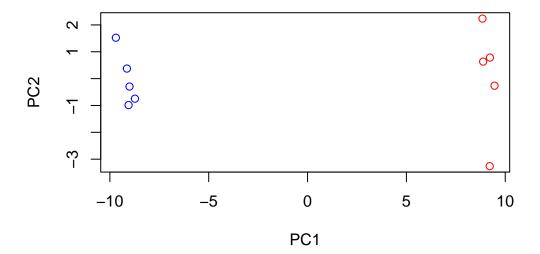
#### summary(pca)

```
Importance of components:
```

```
PC2
                                         PC3
                                                 PC4
                                                         PC5
                          PC1
                                                                 PC6
                                                                         PC7
Standard deviation
                       9.6237 1.5198 1.05787 1.05203 0.88062 0.82545 0.80111
Proportion of Variance 0.9262 0.0231 0.01119 0.01107 0.00775 0.00681 0.00642
Cumulative Proportion 0.9262 0.9493 0.96045 0.97152 0.97928 0.98609 0.99251
                           PC8
                                   PC9
                                            PC10
Standard deviation
                       0.62065 0.60342 3.457e-15
Proportion of Variance 0.00385 0.00364 0.000e+00
Cumulative Proportion 0.99636 1.00000 1.000e+00
```

```
# We have 5wt and 5ko samples
mycols <- c(rep("blue",5), rep("red",5))
mycols</pre>
```

[1] "blue" "blue" "blue" "blue" "red" "red" "red" "red" plot(pca\$x[,1], pca\$x[,2], xlab="PC1", ylab="PC2", col = mycols)



I could examine which genes contribute to this first PC

```
head(sort(abs(pca$rotation[,1]), decreasing = T))
```

gene100 gene66 gene45 gene68 gene98 gene60 0.1038708 0.1038455 0.1038402 0.1038395 0.1038372 0.1038055

#Analysis of Breast Cancer FNA data

```
# Save your input data file into your Project directory
fna.data <- "WisconsinCancer.csv"

# Complete the following code to input the data and store as wisc.df
wisc.df <- read.csv(fna.data, row.names=1)
head(wisc.df)</pre>
```

	diagnosis	radius_mean	texture_mean	perimeter_mean	area_mean
842302	M	17.99	10.38	122.80	1001.0
842517	M	20.57	17.77	132.90	1326.0
84300903	M	19.69	21.25	130.00	1203.0
84348301	M	11.42	20.38	77.58	386.1

84358402	М	20.29	14.34	135.1	.0 1297.0	
843786	M	12.45	15.70	82.5		
010100	smoothness_mean					nts mean
842302	0.11840	-	.27760	0.3001		0.14710
842517	0.08474		.07864	0.0869		0.07017
84300903	0.10960		. 15990	0.1974		0.12790
84348301	0.14250		. 28390	0.2414		0.10520
84358402	0.10030		. 13280	0.1980		0.10430
843786	0.12780		. 17000	0.1578		0.08089
	symmetry_mean fr				exture_se pe	
842302	0.2419	_	0.07871	1.0950	0.9053	8.589
842517	0.1812		0.05667	0.5435	0.7339	3.398
84300903	0.2069		0.05999	0.7456	0.7869	4.585
84348301	0.2597		0.09744	0.4956	1.1560	3.445
84358402	0.1809		0.05883	0.7572	0.7813	5.438
843786	0.2087		0.07613	0.3345	0.8902	2.217
	area_se smoothne	ess_se compa	actness_se	concavity_s	se concave.po	oints_se
842302	153.40 0.0	006399	0.04904	0.0537	'3	0.01587
842517	74.08 0.0	005225	0.01308	0.0186	80	0.01340
84300903	94.03 0.0	006150	0.04006	0.0383	32	0.02058
84348301	27.23 0.0	009110	0.07458	0.0566	51	0.01867
84358402	94.44 0.0	011490	0.02461	0.0568	88	0.01885
843786	27.19 0.0	007510	0.03345	0.0367	2	0.01137
	symmetry_se frac	ctal_dimensi	ion_se radi	ius_worst te	exture_worst	
842302	0.03003	0.0	006193	25.38	17.33	
842517	0.01389	0.0	003532	24.99	23.41	
84300903	0.02250	0.0	004571	23.57	25.53	
84348301	0.05963	0.0	009208	14.91	26.50	
84358402	0.01756		005115	22.54	16.67	
843786	0.02165		005082	15.47	23.75	
	${\tt perimeter\_worst}$	_	smoothness	-	<del>-</del>	
842302	184.60	2019.0		0.1622	0.66	
842517	158.80	1956.0		0.1238	0.186	56
84300903	152.50	1709.0		0.1444	0.424	
84348301	98.87	567.7		0.2098	0.866	
84358402	152.20	1575.0		0.1374	0.20	
843786	103.40	741.6		0.1791	0.524	49
	concavity_worst	concave.poi		•		
842302	0.7119		0.2654		:601	
842517	0.2416		0.1860		2750	
84300903	0.4504		0.2430		8613	
84348301	0.6869		0.2575		638	
84358402	0.4000		0.1625	0.2	2364	

843786	0.5355	0.1741	0.3985
	<pre>fractal_dimension_worst</pre>		
842302	0.11890		
842517	0.08902		
84300903	0.08758		
84348301	0.17300		
84358402	0.07678		
843786	0.12440		

```
diagnosis <- as.factor(wisc.df$diagnosis)</pre>
```

Note that the first column here wisc.df\$diagnosis is a pathologist provided expert diagnosis.

Now I want to make sure I remove that column from my sataset for analysis

```
wisc.data <- wisc.df[,-1]
head(wisc.data)</pre>
```

	radius_mean text	ure_mean	perimete	er_mean a	area_mean	smooth	ness_mean
842302	17.99	10.38		122.80	1001.0		0.11840
842517	20.57	17.77		132.90	1326.0		0.08474
84300903	19.69	21.25		130.00	1203.0		0.10960
84348301	11.42	20.38		77.58	386.1		0.14250
84358402	20.29	14.34		135.10	1297.0		0.10030
843786	12.45	15.70		82.57	477.1		0.12780
	compactness_mean	concavi	ty_mean o	concave.p	ooints_mea	n symme	etry_mean
842302	0.27760	)	0.3001		0.1471	.0	0.2419
842517	0.07864	Ŀ	0.0869		0.0701	.7	0.1812
84300903	0.15990	)	0.1974		0.1279	0	0.2069
84348301	0.28390	)	0.2414		0.1052	20	0.2597
84358402	0.13280	)	0.1980		0.1043	30	0.1809
843786	0.17000	)	0.1578		0.0808	9	0.2087
	fractal_dimension	n_mean ra	adius_se	texture_	_se perime	ter_se	area_se
842302	(	0.07871	1.0950	0.90	)53	8.589	153.40
842517	(	0.05667	0.5435	0.73	339	3.398	74.08
84300903	(	0.05999	0.7456	0.78	369	4.585	94.03
84348301	(	.09744	0.4956	1.15	560	3.445	27.23
84358402	(	0.05883	0.7572	0.78	313	5.438	94.44
843786	(	0.07613	0.3345	0.89	902	2.217	27.19
	smoothness_se co	mpactness	s_se cond	cavity_se	e concave.	points	_se
842302	0.006399	0.04	4904	0.05373	3	0.01	587
842517	0.005225	0.0	1308	0.01860	)	0.013	340

84300903	0.006150	0.0400	0.0	03832	0.02058
84348301	0.009110	0.0745	0.0	05661	0.01867
84358402	0.011490	0.0246	0.0	05688	0.01885
843786	0.007510	0.0334	15 0.0	03672	0.01137
	symmetry_se frac	ctal_dimensi	ion_se rad:	ius_worst	texture_worst
842302	0.03003	0.0	006193	25.38	17.33
842517	0.01389	0.0	003532	24.99	23.41
84300903	0.02250	0.0	04571	23.57	25.53
84348301	0.05963	0.0	009208	14.91	26.50
84358402	0.01756	0.0	005115	22.54	16.67
843786	0.02165	0.0	05082	15.47	23.75
	<pre>perimeter_worst</pre>	area_worst	smoothness	s_worst co	ompactness_worst
842302	184.60	2019.0		0.1622	0.6656
842517	158.80	1956.0		0.1238	0.1866
84300903	152.50	1709.0		0.1444	0.4245
84348301	98.87	567.7		0.2098	0.8663
84358402	152.20	1575.0		0.1374	0.2050
843786	103.40	741.6		0.1791	0.5249
	<pre>concavity_worst</pre>	concave.poi	ints_worst	symmetry	_worst
842302	0.7119		0.2654	(	0.4601
842517	0.2416		0.1860	(	0.2750
84300903	0.4504		0.2430	(	0.3613
84348301	0.6869		0.2575	(	0.6638
84358402	0.4000		0.1625	(	0.2364
843786	0.5355		0.1741	(	0.3985
	fractal_dimension	on_worst			
842302		0.11890			
842517		0.08902			
84300903		0.08758			
84348301		0.17300			
84358402		0.07678			
843786		0.12440			

Q1. How many observations are in this dataset?

nrow(wisc.df)

[1] 569

There are 569 observations

Q2. How many of the observations have a malignant diagnosis?

#### table(wisc.df\$diagnosis) В М 357 212 There are 212 malignant diagnosis. Q3. How many variables/features in the data are suffixed with \_mean? colnames(wisc.data) [1] "radius\_mean" "texture\_mean" [3] "perimeter\_mean" "area mean" [5] "smoothness\_mean" "compactness\_mean" [7] "concavity\_mean" "concave.points\_mean" [9] "symmetry\_mean" "fractal\_dimension\_mean" [11] "radius\_se" "texture\_se" [13] "perimeter\_se" "area\_se" [15] "smoothness\_se" "compactness\_se" [17] "concavity\_se" "concave.points\_se" [19] "symmetry\_se" "fractal\_dimension\_se" [21] "radius\_worst" "texture\_worst" [23] "perimeter\_worst" "area\_worst" [25] "smoothness\_worst" "compactness\_worst" [27] "concavity\_worst" "concave.points\_worst" [29] "symmetry\_worst" "fractal\_dimension\_worst" grep("\_mean", colnames(wisc.data), value=TRUE)

There are 10 variables/features in the data that are suffixed with "\_mean"

"texture\_mean"

"smoothness\_mean"

"concave.points\_mean"

"perimeter\_mean"

"symmetry\_mean"

"compactness\_mean"

[1] "radius\_mean"

[7] "concavity\_mean"

[10] "fractal\_dimension\_mean"

[4] "area\_mean"

### **Principal Component Analysis**

Here we will use prcomp() on the wisc.data object - the one without the diagnosis column.

First, we have to decide whether to use the scale=TRUE argument when we run prcomp().

We can look at the means and sd of each column. If they are similar then we are all good to go. If not we should use scale=TRUE

#### colMeans(wisc.data)

perimeter_mean	texture_mean	radius_mean
9.196903e+01	1.928965e+01	1.412729e+01
${\tt compactness\_mean}$	${\tt smoothness\_mean}$	area_mean
1.043410e-01	9.636028e-02	6.548891e+02
symmetry_mean	concave.points_mean	concavity_mean
1.811619e-01	4.891915e-02	8.879932e-02
texture_se	radius_se	fractal_dimension_mean
1.216853e+00	4.051721e-01	6.279761e-02
smoothness_se	area_se	perimeter_se
7.040979e-03	4.033708e+01	2.866059e+00
concave.points_se	concavity_se	compactness_se
1.179614e-02	3.189372e-02	2.547814e-02
radius_worst	fractal_dimension_se	symmetry_se
1.626919e+01	3.794904e-03	2.054230e-02
area_worst	perimeter_worst	texture_worst
8.805831e+02	1.072612e+02	2.567722e+01
concavity_worst	compactness_worst	smoothness_worst
2.721885e-01	2.542650e-01	1.323686e-01
${\tt fractal\_dimension\_worst}$	symmetry_worst	concave.points_worst
8.394582e-02	2.900756e-01	1.146062e-01

### apply(wisc.data,2,sd)

radius_mean 3.524049e+00	texture_mean 4.301036e+00	perimeter_mean 2.429898e+01
area_mean	smoothness_mean	compactness_mean
3.519141e+02	1.406413e-02	5.281276e-02
concavity_mean	concave.points_mean	$symmetry_mean$
7.971981e-02	3.880284e-02	2.741428e-02
fractal_dimension_mean	radius_se	texture_se
7.060363e-03	2.773127e-01	5.516484e-01

perimeter_se	area_se	smoothness_se
2.021855e+00	4.549101e+01	3.002518e-03
compactness_se	concavity_se	concave.points_se
1.790818e-02	3.018606e-02	6.170285e-03
symmetry_se	fractal_dimension_se	radius_worst
8.266372e-03	2.646071e-03	4.833242e+00
texture_worst	perimeter_worst	area_worst
6.146258e+00	3.360254e+01	5.693570e+02
smoothness_worst	compactness_worst	concavity_worst
2.283243e-02	1.573365e-01	2.086243e-01
<pre>concave.points_worst</pre>	symmetry_worst	<pre>fractal_dimension_worst</pre>
6.573234e-02	6.186747e-02	1.806127e-02

These are very different so we should scale=TRUE.

```
wisc.pr <- prcomp(wisc.data, scale=TRUE)
summary(wisc.pr)</pre>
```

#### Importance of components:

```
PC2
                          PC1
                                         PC3
                                                  PC4
                                                          PC5
                                                                  PC6
                                                                          PC7
Standard deviation
                       3.6444 2.3857 1.67867 1.40735 1.28403 1.09880 0.82172
Proportion of Variance 0.4427 0.1897 0.09393 0.06602 0.05496 0.04025 0.02251
Cumulative Proportion 0.4427 0.6324 0.72636 0.79239 0.84734 0.88759 0.91010
                           PC8
                                  PC9
                                         PC10
                                                PC11
                                                         PC12
                                                                 PC13
                                                                         PC14
Standard deviation
                       0.69037 0.6457 0.59219 0.5421 0.51104 0.49128 0.39624
Proportion of Variance 0.01589 0.0139 0.01169 0.0098 0.00871 0.00805 0.00523
Cumulative Proportion
                       0.92598 0.9399 0.95157 0.9614 0.97007 0.97812 0.98335
                          PC15
                                  PC16
                                          PC17
                                                   PC18
                                                           PC19
                                                                   PC20
Standard deviation
                       0.30681 0.28260 0.24372 0.22939 0.22244 0.17652 0.1731
Proportion of Variance 0.00314 0.00266 0.00198 0.00175 0.00165 0.00104 0.0010
Cumulative Proportion 0.98649 0.98915 0.99113 0.99288 0.99453 0.99557 0.9966
                          PC22
                                  PC23
                                         PC24
                                                  PC25
                                                          PC26
                                                                  PC27
                                                                          PC28
Standard deviation
                       0.16565 0.15602 0.1344 0.12442 0.09043 0.08307 0.03987
Proportion of Variance 0.00091 0.00081 0.0006 0.00052 0.00027 0.00023 0.00005
Cumulative Proportion
                       0.99749 0.99830 0.9989 0.99942 0.99969 0.99992 0.99997
                          PC29
                                  PC30
Standard deviation
                       0.02736 0.01153
Proportion of Variance 0.00002 0.00000
Cumulative Proportion 1.00000 1.00000
```

Q4. From your results, what proportion of the original variance is captured by the first principal components (PC1)?

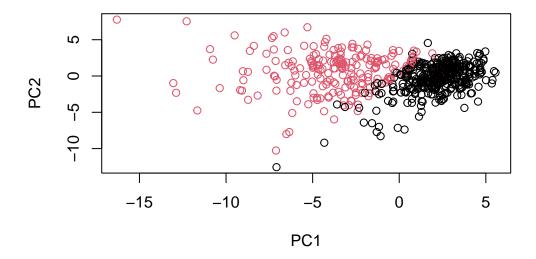
- 44.47% is captured by PC1
- Q5. How many principal components (PCs) are required to describe at least 70% of the original variance in the data?
- 3 PCs are required to describe at least 70% of original variance in the data set
- Q6. How many principal components (PCs) are required to describe at least 90% of the original variance in the data?
- 7 PCs are required to describe at least 90% of the original variance in the data

#### Plotting the PCA results

```
#biplot(wisc.pr)
```

Q7. What stands out to you about this plot? Is it easy or difficult to understand? Why? It's a hot mess, difficult to understand because it's meant for small data sets.

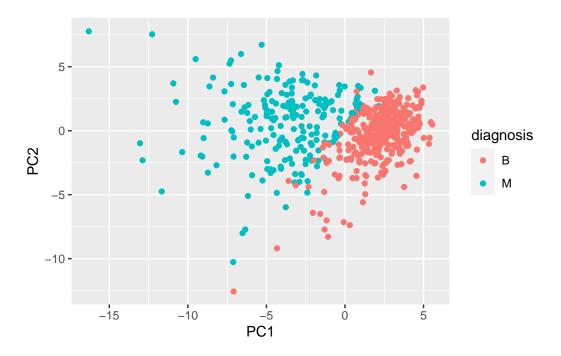
We need to make our own plot.



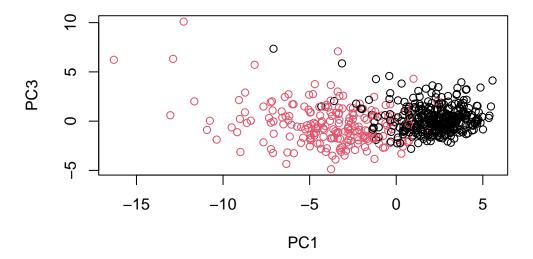
```
library(ggplot2)

pc <- as.data.frame(wisc.pr$x)

ggplot(pc) +
   aes(PC1, PC2, col=diagnosis) +
   geom_point()</pre>
```



Q8. Generate a similar plot for principal components 1 and 3. What do you notice about these plots?



There is a distinct separation between benign and malignant tumors. Both plots look similar to each other. Cells with similar characteristics should cluster together.

#### **Communicating PCA results**

Q9. For the first principal component, what is the component of the loading vector (i.e. wisc.pr\$rotation[,1]) for the feature concave.points\_mean?

```
wisc.pr$rotation["concave.points_mean",1]
```

[1] -0.2608538

Q10. What is the minimum number of principal components required to explain 80% of the variance of the data?

5 PCs are required to explain 80% of the variance of the data

```
tbl <- summary(wisc.pr)
which(tbl$importance[3,] > 0.8)[1]
```

PC5

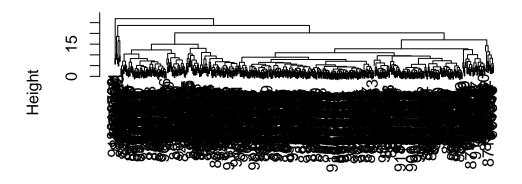
5

# **Hierarchial Clustering**

The main function for Hierarchial Clustering is called hclust() which takes a distance matrix as input.

```
d <- dist(scale(wisc.data))
wisc.hclust <- hclust(d)
plot(wisc.hclust)</pre>
```

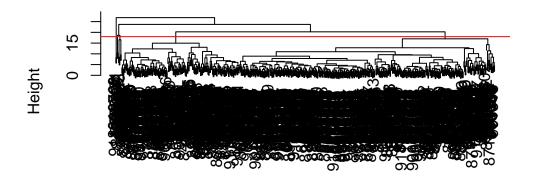
# **Cluster Dendrogram**



d hclust (\*, "complete")

```
plot(wisc.hclust)
abline(h=18, col="red")
```

# **Cluster Dendrogram**



d hclust (\*, "complete")

```
grps <- cutree(wisc.hclust, h=18)
table(grps)</pre>
```

Come back here later to see how our cluster grps correspond to M or B groups.

Q11. Using the plot() and abline() functions, what is the height at which the clustering model has 4 clusters?

Around 18

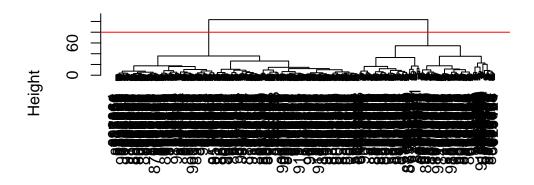
#### 5. Combining Methods

Here we will perform clustering on our PCA results rather than the original data.

In other words we will cluster using wisc.pr\$x - our new better variable or PCs. We can chose as many or as few PCs to use as we like. It is your call!

```
d.pc <- dist(wisc.pr$x[, 1:3])
wisc.pr.hclust <- hclust( d.pc, method="ward.D2")
plot(wisc.pr.hclust)
abline(h=80, col="red")</pre>
```

# **Cluster Dendrogram**



d.pc hclust (\*, "ward.D2")

```
grps <- cutree(wisc.pr.hclust, h=80)
table(grps)</pre>
```

grps 1 2 203 366

We can use table() function to make a cross-table as well as just a count table.

```
table(diagnosis)
```

diagnosis B M 357 212

```
table(grps, diagnosis)
```

```
diagnosis
grps B M
1 24 179
2 333 33
```

- Q12. Can you find a better cluster vs diagnoses match by cutting into a different number of clusters between 2 and 10?
- Q13. Which method gives your favorite results for the same data.dist dataset? Explain your reasoning.

Maybe the average just because it's simple and straight-forward and is the only one that I really remember.

Write a note here about how to read this cross-table result. The results indicate that our cluster 1 mostly captures cancer (M) and our cluster 2 mostly captures healthy (B) samples/individuals.

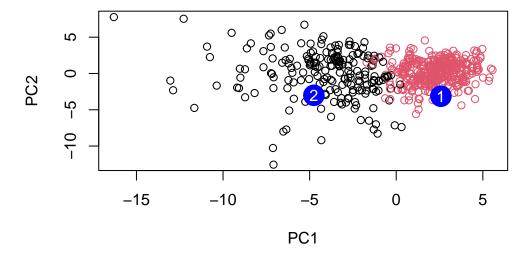
#### ##7. Prediction

```
#url <- "new_samples.csv"
url <- "https://tinyurl.com/new-samples-CSV"
new <- read.csv(url)
npc <- predict(wisc.pr, newdata=new)
npc</pre>
```

```
PC1
                     PC2
                                PC3
                                           PC4
                                                     PC5
                                                                 PC6
                                                                            PC7
     2.576616 -3.135913
                         1.3990492 -0.7631950 2.781648 -0.8150185 -0.3959098
[2,] -4.754928 -3.009033 -0.1660946 -0.6052952 -1.140698 -1.2189945
                                                                      0.8193031
            PC8
                      PC9
                                PC10
                                          PC11
                                                     PC12
[1,] -0.2307350 0.1029569 -0.9272861 0.3411457 0.375921 0.1610764 1.187882
[2,] -0.3307423 0.5281896 -0.4855301 0.7173233 -1.185917 0.5893856 0.303029
                                                          PC19
          PC15
                     PC16
                                 PC17
                                             PC18
[1,] 0.3216974 -0.1743616 -0.07875393 -0.11207028 -0.08802955 -0.2495216
[2,] 0.1299153
               0.1448061 -0.40509706
                                      0.06565549
                                                   0.25591230 -0.4289500
                      PC22
                                 PC23
                                            PC24
                                                         PC25
[1,] 0.1228233 0.09358453 0.08347651 0.1223396
                                                  0.02124121
                                                              0.078884581
[2,] -0.1224776 0.01732146 0.06316631 -0.2338618 -0.20755948 -0.009833238
             PC27
                         PC28
                                      PC29
                                                   PC30
     0.220199544 -0.02946023 -0.015620933
                                            0.005269029
[1,]
[2,] -0.001134152  0.09638361  0.002795349 -0.019015820
```

And plot this up

```
plot(wisc.pr$x[,1:2], col=grps)
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
text(npc[,1], npc[,2], c(1,2), col="white")
```



Q18. Which of these new patients should we prioritize for follow up based on your results? Patient 2